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Ortho-quinone methide driven synthesis of kynurenic acid lactams†

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Lactam formation of different KYNA amides and Mannich bases mediated by *ortho*-quinone methide has been investigated. The efficiency of the two routes of the cyclization process was revealed and the influence of diverse amide side chains was explored. In this regard compounds bearing a tertiary amine function in the amide side chain resulted in the formation of the lactam product, while the formation of dimer derivatives was observed in the case of other KYNA amides. Furthermore, derivatives bearing different substituents on the KYNA B ring were synthesized and their effects on the ring-closure reaction were investigated.

1 Introduction

Kynurenic acid (KYNA) and 4-hydroxyquinoline-2-carboxylic acid are endogenous substances, produced during tryptophan metabolism (TRP) through the pathway that is also responsible for the production of nicotinamide adenine dinucleotide (NAD⁺) and NAD phosphate (NADP⁺).^{1,2} It is one of the few endogenous excitatory amino acid receptor blockers with a broad spectrum of antagonistic properties in supra-physiological concentrations. KYNA is well established to have high affinity for NMDA receptors and has been shown to have a positive modulatory binding site of higher affinity at the α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA).³

In recent years there has been increasing interest in the synthesis and pharmacological studies of KYNA derivatives. This is due to their neuroprotective ability as it can prevent neuronal loss following excitotoxic, ischemia-induced, and infectious neuronal injuries.^{4,5} Several papers have been published on different alterations of the KYNA skeleton. Substitutions at positions 5–8 were achieved by starting from the corresponding anilines using the modified Conrad-Limpach method.^{6–8} The substituents in positions 2 and 4 have also been

investigated in a broad spectrum ranging from alkoxy groups on C-4 or amides on C-2.^{8–14} Based on the evaluations of previous KYNA amides, a tertiary nitrogen is needed for a biological activity concerning the central nervous system.^{15–18} Derivatives bearing such functional groups can be synthesized by various methods, such as amidation of the carboxylic function^{12–14} or the transformation of the 4-hydroxy group.^{8–10} Another possibility can be the substitution of KYNA using the modified Mannich reaction (mMr). In this one-pot reaction KYNA used as a CH acid can be reacted with different types of secondary amines and aldehydes giving an aminoalkylated derivative with the desired cationic center.^{19,20}

These cationic centers, beside bearing a function possibly required for transport/passage through the blood–brain–barrier, present a new reaction route to synthesize further KYNA derivatives. Depending on the amines used, these groups can function as leaving groups forming an *ortho*-quinone methide (*o*Qm) intermediate. It is well known that these derivatives bear high reactivity as they contain a methide function²¹ that may react with a well-selected structural element. Under specific reaction conditions, the aminoalkylated KYNA amides undergo intramolecular ring closure resulting in the formation of pyrroloquinolinone derivatives. Synthesis of similar compounds have already been discussed in the literature: using oxidative methods²² or transforming naturally occurring KYNA derivatives;²³ however, this instance is the first to utilize a reaction *via* *o*Qm (Fig. 1). This structure bears information about the reactivity of specific *o*Qm and they have high resemblance to biologically active compounds, including camptothecin derivatives currently in medical use²⁴ and analogues acting on benzodiazepine receptors.²⁵ Compounds, bearing the 4-quinolone scaffold have also been investigated as potential analgesics, due to their NMDA receptor binding capabilities²⁶ (Fig. 1). Consequently, a suitable synthetic procedure was planned.

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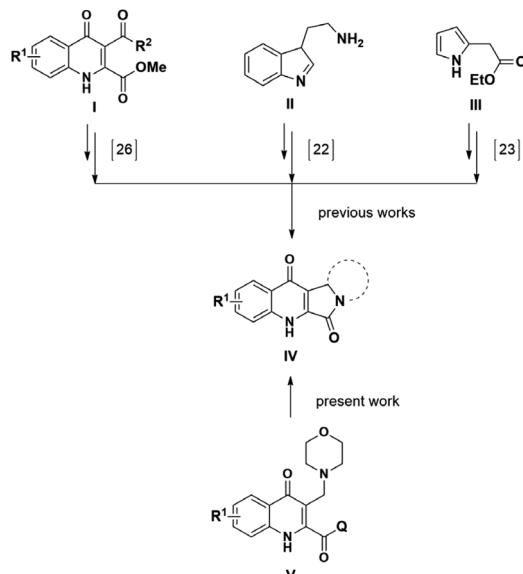


Fig. 1 Summary of previous and recent work.

2 Results and discussion

The first KYNA lactam (**4**) was observed as a side product during the scale-up morpholinomethylation of KYNA amide **1** in mM. Taking into consideration the present compounds in the reaction media and the possible intermediates, compound **4** could have been formed *via* two distinct routes: (A) from starting compound **1** and paraformaldehyde with a one-step ring closure or (B) through the aminoalkylation of **1** resulting in a Mannich base (**2**) that can subsequently act as an intermediate in a two-step reaction route (Scheme 1).

Since aminoalkylated KYNA amide **2** was in excess when starting the work-up of the reaction discussed above, route (B) was investigated first. Different solvents (toluene, DCM, 1,4-dioxane, acetonitrile, isopropyl alcohol, EtOH and MeOH) were tested showing the highest conversion in MeOH at 100 °C in 1.5 h. In the next step, the optimization of the reaction was

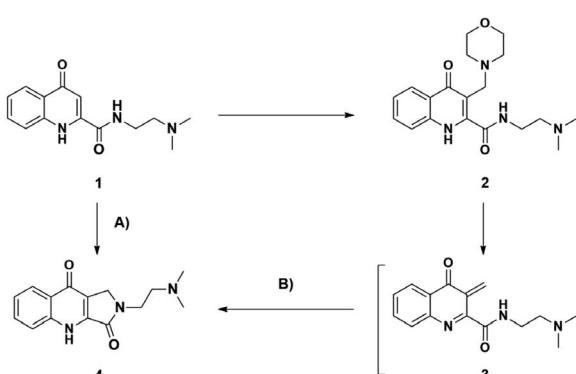
Table 1 Screening for optimal temperature

Entry	Reaction route	Temperature (°C)	Conversion ^a (%)
1	(A)	40	0
2		80	0
3		100	0
4		120	0
5		140	60
6		150	90
7		160	99
8	(B)	40	0
9		80	0
10		100	30
11		120	45
12		140	70
13		150	99
14		160	99

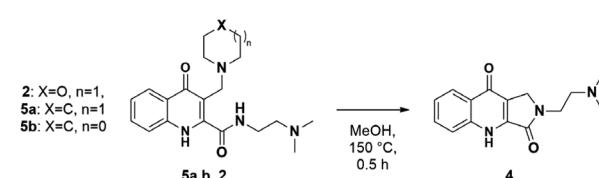
^a Starting from **1** or **2** towards the formation of lactam (**4**), calculated from NMR spectra.

carried out at different temperatures. At lower temperature intervals no conversion was detected (Table 1, entries 8, 9). At higher temperatures increasing conversions were measured (entries 10–12) eventually reaching a plateau at 150 °C after 1.5 h (entry 13). Even though the reaction took place with the same conversion at 160 °C as well (entry 14), 150 °C was chosen as optimal for the preceding reactions. As NMR tracking of the reactions carried out at higher temperatures (entry 13 and 14) showed an almost full transformation of **2** to **4**, it is surmised that the reaction fully takes place *via* oQm 3. However, since the reaction matrix for aminoalkylation also had the starting materials for route (A) as well, a one-pot reaction was planned with entry 13 parameters starting from **1** and utilising morpholine and paraformaldehyde. The aim of this reaction was to decide whether a direct reaction could be achieved *via* route (B) under the optimized conditions, when starting from the prerequisites of the Mannich base instead of isolated compound **2**. According to crude NMR spectra, the reaction showed only minimal formation of the desired lactam, leading to Mannich-base **2** as the main product. Then reactions were carried out *via* route (A) showing a similar temperature-dependent transformation (Table 1, entries 1–7), but with lower or no conversion at similar temperatures (entries 3 vs. 10, 4 vs. 11, 5 vs. 12, 6 vs. 13) reaching the same plateau at 160 °C (entry 7).

Next, the reaction was repeated using different Mannich bases to find the best leaving group. This included two secondary amines (pyrrolidine and piperidine) used for the



Scheme 1 Possible reaction routes toward the formation of lactam derivative **4**. (A) direct cyclization of **1** with paraformaldehyde (B) aminoalkylation with paraformaldehyde and morpholine followed by intramolecular cyclization.



Scheme 2 Reaction of alternative Mannich bases.



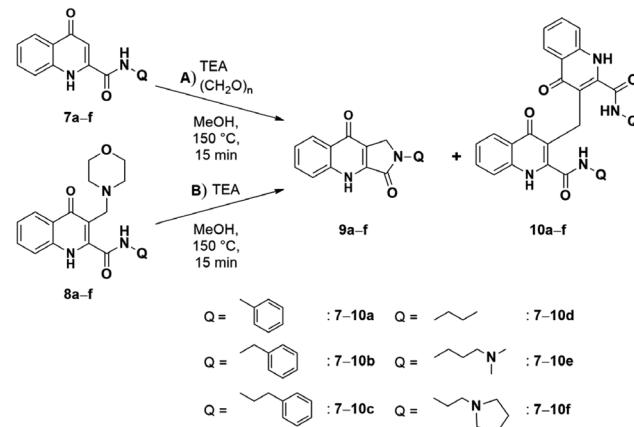
Table 2 Exemplary catalysts (in 0.75 equivalent) in the case of both reaction routes

Entry	Reaction route	Additive	Conversion (%)
1	(A)	AcOH	0
2		<i>p</i> -TsOH	0
3		TEA	99 ^a
4		DMAP	30
5	(B)	Et ₃ N	99 ^b

^a 30 min reaction time. ^b 15 min reaction time.

mMr and subsequently for route (B) (Scheme 2). Based on our previous work concerning C–C bond formations on the KYNA structure starting from Mannich bases, primary amines were omitted from the tests as they proved to be poor leaving groups.²⁷ Reactions were carried out with aminoalkylated derivatives 2 and 5a,b. The results showed that higher conversions could be achieved from 5a,b in only 30 min (99% from 5a,b presumably due to the higher *K*_b of the amine moieties *versus* 85% from 2). However, the workup was hindered by the presence of a strong base. Additional steps were necessary for their removal and the overall yields were lower in these cases (56% for 5a, 54% for 5b *versus* 76% for 2).

A further difference between reaction route (A) and (B), beside the possible *o*Qm intermediate, might be in the elimination of morpholine in the case of (B) that can create basic conditions. Based on this information and on results of our previous research, where Mannich bases reacted in a similar manner,²⁷ we planned to test acidic and basic additives in route (A) (Table 2). Reactions applying acidic catalysts (AcOH or *p*-toluenesulfonic acid [*p*-TsOH]) showed inhibition of the reaction, similar to the basic *N,N*-dimethylaminopyridine (DMAP), though the latter had only partial inhibition of the reaction. In the case of triethylamine (TEA), high conversion in shorter reaction (30 min.) was observed. It is notable that TEA promoted the formation of 4 not only in reaction route (A) but also in the case of (B) in a reaction time of 15 min.



Scheme 4 Scope and limitations investigation *via* amide derivatives.

The scope and limitations of the reaction were also investigated using different amides bearing alkyl, aryl, and aminoalkyl functions. The relevant amide derivatives and the corresponding Mannich bases were synthesized according to the methods described herein (Scheme 3). The ring-closure reactions were conducted applying the optimized reaction conditions and in the case of route (B) starting from the corresponding Mannich base (Scheme 4, Table 3).

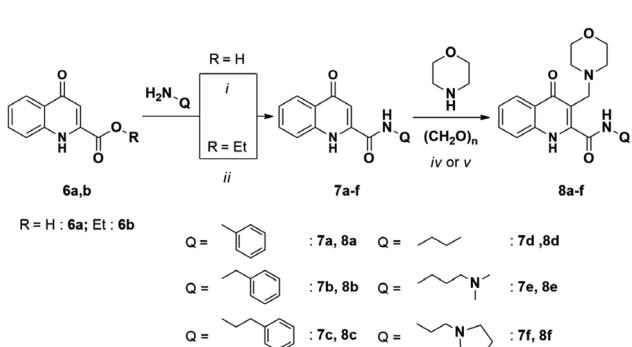
Based on the experimental results it is hypothesized that an aminoalkyl functional group is crucial for the formation of the lactam ring in both reaction routes. Note, however, that there is still a shift towards route (B) (Table 3, entries 3–8 *vs.* 15 and 16). Furthermore, a side chain too long is also less favored indicating a possible electrostatic stabilizing effect through a seven-membered ring (entries 7 and 8). Results in entries 15 and 16 are in accordance with this, since *n*-propylamine is a smaller

Table 3 Yields and products of both reaction routes in the case of exemplary amides (reaction time = 0.25 h)

Entry	Q-	Reaction route	Conversion ^a (%)	Product
1	H	(A)	—	—
2		(B)	99	— ^b
3	— ^c	(A)	70	4
4	— ^c	(B)	99	4
5	— ^c	(A)	99	9f
6	— ^c	(B)	99	9f
7	— ^c	(A)	65	— ^b
8	— ^c	(B)	80	9e
9	— ^c	(A)	0	—
10	— ^c	(B)	65	10a
11	— ^c	(A)	0	—
12	— ^c	(B)	99	10b
13	— ^c	(A)	0	—
14	— ^c	(B)	99	— ^b
15	— ^c	(A)	— ^c	10d
16	— ^c	(B)	— ^c	10d

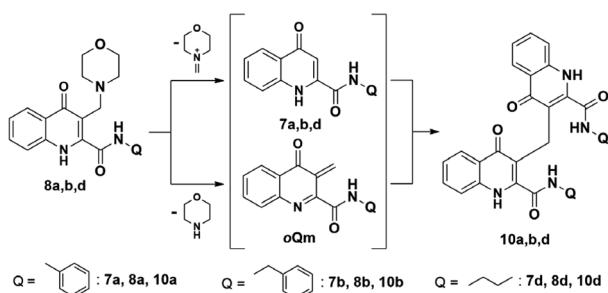
^a Starting from 7a-f or 8a-f towards the formation of lactams or dimers (4, 9e-f or 10a-d), determined from the NMR spectra.

^b Multicomponent reaction mixture. ^c Conversions could not be determined, because products are highly insoluble in DMSO.

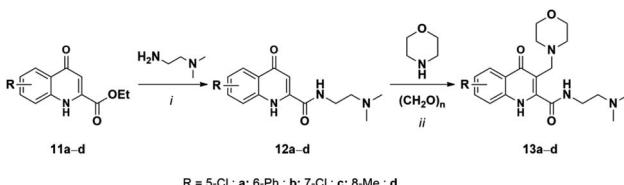


Scheme 3 Synthesis of different alkyl, aryl, and aminoalkyl amide derivatives and their subsequent transformation to Mannich bases. (i) Equimolar diisopropylcarbodiimide (DIC) and 1-hydroxybenzotriazole (1-HOBT) as catalysts at r.t. in DMF (7a–c) (ii) neat conditions, r.t. (7d) (iv) 1,4-Dioxane, reflux (8a–d) (v) acetonitrile, reflux (8e).

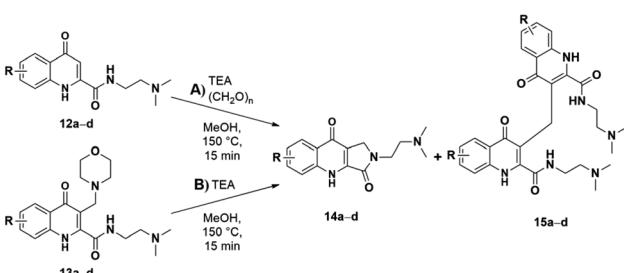




Scheme 5 Possible reaction mechanism for the formation of dimer derivatives starting out from Mannich derivatives.



Scheme 6 Synthesis of B-ring-substituted KYNA amides and Mannich bases. (i) *N,N*-Dimethylethane-1,2-diamine in EtOH, reflux (ii) 1,4-dioxane, reflux.



Scheme 7 Reaction of B-ring-substituted KYNA amides in lactam cyclization.

side-chain in comparison to the aromatic substituents, but also lack a tertiary amine function. It is interesting to note that even though the pyrrolidine function at the end of the alkylamide side chain is bulkier, it facilitates the reaction in route (A) in comparison to the dimethylamine counterpart (entry 5 vs. 3). In

the case of aromatic functional groups it is surmised that they have steric hindering effects. Note, that they do not have the tertiary nitrogen atom and, consequently, lactam formation cannot take place (Table 3, entries 9–14). Furthermore, in the case of a directly connected phenyl group, the negative inductive effect of the aromatic system can also be implied (Table 3, entries 9 and 10).

It is interesting to mention that the formation of a dimer derivatives (**10a–d**) of two KYNA amides connected through a methylene bridge at C-3 was also observed. These compounds were formed in the case of the derivatives bearing no tertiary nitrogen in their amide functions. In case of starting out from amide derivative **7d** the formation of such compounds is in accordance with dimers of hydroxy-quinoline derivatives reported previously.^{28,29} However, in the case of the Mannich derivatives **8a,b,d** it is surmised that the reaction may take place *via* a retro-Mannich pathway. In this, through the elimination of a morpholine the *oQm* intermediate is formed that reacts through an electrophilic addition with the starting amide that is formed *via* the elimination of the morpholinomethyl moiety as an iminium cation (Scheme 5). A more complex but also possible pathway was hypothesized by Funk *et al.* that also depends on the formation of an *oQm* intermediate and suggests the temporary dearomatization of the *oQm*-Mannich base conjugate.³⁰

To further investigate how different substituents may affect the lactam formation, the reaction was extended to KYNA derivatives substituted at the B ring. Different alkyl, aryl, and halogen analogues with these functional groups being in various positions were synthesized and used (Schemes 6, 7 and Table 4). *N¹,N¹*-Dimethylethane-1,2-diamide was chosen to build the amide side chain, because it shows a lesser stabilizing effect compared to 2-(pyrrolidine-1-yl)ethanamine thus giving additional possibility to the modifying effects of the B-ring substituents.

In the case of the aryl analogue no conversion was observed in the crude NMR spectra. Consequently, it is surmised that the phenyl substituent of the B ring inhibits the reaction altogether. It is worth mentioning that this inhibition, while not relevant in the case of the ester derivative,¹⁹ was observed in the amino-alkylation of the amide derivative, hindering its conversion and altogether inhibiting the subsequent intramolecular ring closure (Table 4, entries 3 and 4). While the methyl substituent

Table 4 Comparison of different B-ring substituents

Entry	R-	Route	Time (min)	Conversion ^a (%)	Products ^b	Yield (%)
1	5-Cl	(A)	15	99	14a : 15a (1 : 1)	—
2		(B)	15	99	14a : 15a (1 : 1)	28 : 19 ^c
3	6-Ph	(A)	—	0	—	—
4		(B)	—	0	—	—
5	7-Cl	(A)	15	99	14c : 15c (1 : 1)	—
6		(B)	15	99	14c : 15c (2 : 1)	23 : 14 ^c
7	8-Me	(A)	30	99	14d : 15d (1 : 0)	—
8		(B)	15	99	14d : 15d (1 : 0)	47 : ^c

^a Starting from **12a–d** or **13a–d** towards the formation of lactams or dimers (**14a,c,d** or **15a,c**), determined from NMR spectra. ^b Determined from NMR spectra. ^c Reaction used for work-up.

showed no difference (entries 7 *vs.* 8, 30 min. *vs.* 15 min. until full conversion) compared to the unsubstituted **1** and **2** derivatives, halogen substituents increased the reactivity of the aminoalkylated KYNA derivative and subsequently the reactivity of the *o*Qm intermediate (entries 1, 2, 5, and 6). However, this resulted not only in a faster reaction toward the desired lactam derivatives even from amides, but also increased the quantity of the dimer side product.

3 Conclusions

The synthesis of different KYNA lactam derivatives was investigated. The ring-closing synthesis was explored *via* two routes: (A) reacting the amide with paraformaldehyde, or (B) using the Mannich-base of the corresponding amide. The latter process led to better conversions, possibly due to the formation of a highly reactive *o*Qm intermediate. Next, different aminoalkylated Mannich bases were investigated to test the role of the leaving group. Despite showing lower conversion, the morpholinomethylated derivative proved to be ideal compared to the pyrrolidino and piperidino analogues, due to the more favorable work-up process. The scope and limitations were further investigated using different alkyl-, aryl-, and tertiary nitrogen-containing amides. During these reactions, the alkyl-amine derivatives proved to be the most reactive ones, achieving full conversion in 15 min. The cause of this may be a stabilizing interaction mediated by the cationic center in the amide side chain. Alkyl- and arylamides, not bearing the aforementioned tertiary nitrogen functional group in the amide side chain, underwent a different reaction, which resulted in the formation of bis-quinoline-2-carboxamide structured derivatives. To further study the reactivity of the KYNA amides, we investigated how different B-ring substituents affect lactam formation. Using alkyl, aryl, and halogen derivatives we concluded, that chlorination of the B ring has a positive effect on the conversion, promoting the formation of the lactam, but also the dimer derivative. Alkyl substitution of the KYNA amide had little to no effect on the reactivity, while phenyl substitution heavily inhibits the reaction.

4 Materials and methods

¹H and ¹³C-NMR spectra were recorded in DMSO-d₆ and CDCl₃ solutions in 5 mm tubes at room temperature (r.t.) on a Bruker Ascend 500 spectrometer with a 5 mm BBO Prodigy Probe or on a Bruker AVANCE III 600 MHz spectrometer equipped with a 5 mm CP-TCI triple-resonance cryoprobe (Bruker Biospin, Karlsruhe, Baden Württemberg, Germany) at 500 (1H) and 125 (13C) MHz or 600 (1H) and 150 (13C) MHz, with the deuterium signal of the solvent as the lock and TMS as internal standard (1H, 13C).

Melting points were determined on a Hinotek X-4 melting point apparatus. Merck Kieselgel 60F₂₅₄ plates were used for TLC.

Reactions were conducted in a CEM Focused Microwave Synthesis System, Discover SP (CEM, Charlotte, North Carolina, USA).

HRMS flow injection analysis was performed with Thermo Scientific Q Exactive Plus hybrid quadrupole-Orbitrap (Thermo Fischer Scientific, Waltham, MA, USA) mass spectrometer coupled to a Waters Acquity I-Class UPLC™ (Waters, Manchester, UK).

The synthesis of compounds **7e,f** and **8f** was carried out according to literature method.^{14,31}

4.1. Synthesis

4.1.1. General procedure for the synthesis of amide derivatives. Method I: 4-hydroxyquinoline-2-carboxylic acid (**6a**) (300 mg, 1.587 mmol) and amine derivative (3.147 mmol aniline, phenylmethanamine or 2-phenylethaneamine, respectively) was added to a 100 mL round-bottom flask and dissolved in DMF (15 mL). The reaction was stirred and cooled to 0 °C, then 1-hydroxylbenzotriazole (205.8 mg, 1.523 mmol) and diisopropylcarbodiimide (317 mg, 2.512 mmol) were added to the solution. After reaching r.t. the reaction was stirred for an additional 24 h. The mixture was filtered, and the filtrate was concentrated under vacuum. The desired product was crystallized using EtOAc (10 mL), filtered, and washed with 2 × 5 mL EtOAc.

Method II: ethyl 4-hydroxyquinoline-2-carboxylate (**6b**) (100 mg, 0.460 mmol) was added to an excess of propylamine in a pressure-resistant 10 mL vessel and closed. The reaction was stirred for 1 h and, after the completion of the reaction monitored by TLC (eluent: DCM : MeOH 4 : 1), it was diluted with 3 mL DCM. The formed precipitate was filtered and washed with 2 × 5 mL DCM.

Method III: the appropriate B-ring-substituted ester (**11a-d**, 0.340 mmol) and 36 mg, 0.408 mmol N¹,N¹-dimethylethane-1,2-diamine was added to a round-bottom flask with 20 mL EtOH. The reaction mixture was treated under reflux conditions for 8 h and the solvent was evaporated under vacuum. The residue was crystallized by adding 5 mL of EtOAc and the formed precipitate was filtered and washed with 2 × 5 mL EtOAc.

4.1.1.1 4-Oxo-N-phenyl-1,4-dihydroquinoline-2-carboxamide (7a**).** Yield: 195 mg, 47%. Mp.: decomposition over 300 °C. HRMS calcd for [M + H⁺] *m/z* = 265.0972, found *m/z* = 265.0965; ¹H NMR (500 MHz, DMSO-d₆): 6.91 (1H, s); 7.18 (1H, t, *J* = 7.01 Hz); 7.37–7.42 (3H, m); 7.67–7.76 (1H, m); 7.78–7.88 (2H, m); 7.90–8.00 (1H, m); 8.07–8.15 (1H, m); 10.69 (1H, s); 11.97 (1H, s); ¹³C NMR (125 MHz, DMSO-d₆): 106.4; 108.1; 119.9; 121.2; 124.2; 125.1; 129.3; 132.9; 138.6; 142.3; 161.6; 178.2 (Fig. S1, S2 and S71†).

4.1.1.2 N-Benzyl-4-oxo-1,4-dihydroquinoline-2-carboxamide (7b**).** Yield: 300 mg, 68%. Mp.: 216.9–217.8 °C. HRMS calcd for [M + H⁺] *m/z* = 279.1128, found *m/z* = 279.1123; ¹H NMR (600 MHz, DMSO-d₆): 4.53 (2H, d, *J* = 5.65 Hz); 6.75 (1H, s); 7.26–7.29 (1H, m); 7.36 (5H, d, *J* = 4.29 Hz); 7.69 (1H, s); 7.94 (1H, d, *J* = 7.63 Hz); 8.07 (1H, d, *J* = 7.63 Hz); 9.62 (1H, s); 11.87 (1H, s); ¹³C NMR (150 MHz, DMSO-d₆): 43.4; 107.5; 120.0; 124.2; 125.1; 126.0; 127.5; 127.9; 128.9; 132.8; 139.1; 140.3; 141.9; 162.5; 178.2 (Fig. S3, S4 and S72†).

4.1.1.3 4-Oxo-N-phenethyl-1,4-dihydroquinoline-2-carboxamide (7c**).** Yield: 191 mg, 41%. Mp.: decomposition over



300 °C. HRMS calcd for $[M + H^+]$ $m/z = 293.1285$, found $m/z = 293.1280$; ^1H NMR (600 MHz, DMSO-d₆): 2.89 (2H, t, $J = 7.38$ Hz); 3.56 (2H, q, $J = 6.64$ Hz); 6.66 (1H, s); 7.22 (1H, t, $J = 6.64$ Hz); 7.27 (2H, d, $J = 6.64$ Hz); 7.29–7.38 (3H, m); 7.64–7.73 (1H, m); 7.94 (1H, d, $J = 8.32$ Hz); 8.07 (1H, d, $J = 6.54$ Hz); 9.11 (1H, s); 11.81 (1H, s); ^{13}C NMR (150 MHz, DMSO-d₆): 35.1; 41.5; 107.3; 120.0; 124.1; 125.1; 126.0; 126.7; 128.9; 129.2; 132.8; 139.7; 140.3; 142.0; 162.3; 178.2 (Fig. S5, S6 and S73†).

4.1.1.4 4-Oxo-N-propyl-1,4-dihydroquinoline-2-carboxamide (7d). Yield: 92 mg, 87%. Mp.: 161.0–161.8 °C. HRMS calcd for $[M + H^+]$ $m/z = 231.1128$, found $m/z = 231.1125$; ^1H NMR (600 MHz, DMSO-d₆): 0.91 (3H, t, $J = 7.45$ Hz); 1.54–1.60 (2H, m); 3.25–3.30 (2H, q); 6.71 (1H, s); 7.31–7.39 (1H, m); 7.64–7.72 (1H, m); 7.95 (1H, d, $J = 8.43$ Hz); 8.07 (1H, d, $J = 7.49$ Hz); 9.01 (1H, s); 11.82 (1H, s); ^{13}C NMR (150 MHz, DMSO-d₆): 11.9; 22.5; 41.7; 107.2; 120.0; 124.1; 125.1; 125.9; 132.7; 140.3; 142.1; 162.3; 178.2 (Fig. S7, S8 and S74†).

4.1.1.5 5-Chloro-N-(2-(dimethylamino)ethyl)-4-oxo-1,4-dihydroquinoline-2-carboxamide (12a). Yield: 45 mg, 45%. Mp.: decomposition over 275 °C. HRMS calcd for $[M + H^+]$ $m/z = 294.1004$, found $m/z = 294.0999$; ^1H NMR (600 MHz, DMSO-d₆): 2.19 (6H, s); 2.44 (2H, t, $J = 6.61$ Hz); 3.39 (2H, q, $J = 6.09$ Hz); 6.71 (1H, brs); 7.31 (1H, s); 7.56 (1H, t, $J = 8.17$ Hz); 7.90 (1H, d, $J = 8.43$ Hz); 8.91 (1H, s); ^{13}C NMR (150 MHz, MeOH-d₄): 13.7; 36.6; 57.2; 108.1; 121.5; 126.6; 131.5; 132.1; 143.3; 162.2 (Fig. S9–13 and S75†).

4.1.1.6 N-(2-(Dimethylamino)ethyl)-4-oxo-6-phenyl-1,4-dihydroquinoline-2-carboxamide (12b). Yield: 96 mg, 84%. Mp.: decomposition over 260 °C. HRMS calcd for $[M + H^+]$ $m/z = 336.1707$, found $m/z = 336.1702$; ^1H NMR (600 MHz, DMSO-d₆): 2.21 (6H, s); 2.45 (2H, t, $J = 6.61$ Hz); 3.41 (2H, q, $J = 6.09$ Hz); 6.81 (1H, s); 7.38 (1H, t, $J = 7.28$ Hz); 7.50 (2H, t, $J = 7.82$ Hz); 7.75 (1H, d, $J = 7.34$ Hz); 7.90–7.99 (2H, m); 8.33 (1H, s); 8.84 (1H, brs); ^{13}C NMR (150 MHz, CDCl₃): 37.3; 45.1; 56.7; 106.8; 118.9; 123.8; 127.1; 127.7; 128.9; 129.1; 131.8; 132.2; 137.7; 139.5; 160.7 (Fig. S14–18 and S76†).

4.1.1.7 7-Chloro-N-(2-(dimethylamino)ethyl)-4-oxo-1,4-dihydroquinoline-2-carboxamide (12c). Yield: 45 mg, 45%. Mp.: decomposition over 160 °C. HRMS calcd for $[M + H^+]$ $m/z = 294.1004$, found $m/z = 294.1000$; ^1H NMR (600 MHz, MeOH-d₄): 2.44 (6H, s); 2.73 (2H, t, $J = 6.7$ Hz); 3.61 (2H, t, $J = 6.52$ Hz); 6.90 (1H, s); 7.41 (1H, d, $J = 8.77$ Hz); 7.88 (1H, s); 8.20 (1H, d, $J = 8.76$ Hz); ^{13}C NMR (150 MHz, MeOH-d₄): 36.4; 43.6; 57.1; 105.9; 120.1; 123.3; 124.6; 126.0; 137.6; 142.6; 145.1; 163.1; 176.8 (Fig. S19–22 and S77†).

4.1.1.8 N-(2-(Dimethylamino)ethyl)-8-methyl-4-oxo-1,4-dihydroquinoline-2-carboxamide (12d). Yield: 57 mg, 61%. Mp.: 158.9–160.3 °C. HRMS calcd for $[M + H^+]$ $m/z = 274.1550$, found $m/z = 274.1547$; ^1H NMR (600 MHz, CDCl₃): 2.31 (6H, s); 2.54–2.63 (5H, m); 3.54–3.62 (2H, m); 6.81 (1H, s); 7.28–7.34 (1H, m); 7.49–7.55 (1H, m); 8.18–8.24 (1H, m); 9.46 (1H, brs); ^{13}C NMR (150 MHz, CDCl₃): 16.7; 37.3; 45.0; 56.7; 106.7; 124.0; 124.1; 125.7; 133.6; 137.5; 137.5; 138.7; 161.3; 180.0 (Fig. S23, S24 and S78†).

4.1.2 General procedure for the synthesis of the Mannich bases. Method IV: Amide derivative (7a–d (0.435 mmol), paraformaldehyde (14.3 mg, 0.478 mmol), and morpholine

(41.6 mg, 0.478 mmol) were added to a round-bottom flask. The mixture was treated under reflux conditions in 1,4-dioxane (10 mL) for 8 h. After completion of the reaction, the solvent was removed under reduced pressure and the desired product was crystallized using EtOAc (5 mL), filtered, and washed with 2 × 5 mL EtOAc.

Method V: *N*-(3-(dimethylamino)propyl)-4-hydroxyquinoline-2-carboxamide (7e) (118.9 mg, 0.435 mmol), paraformaldehyde (15.7 mg, 0.522 mmol), and morpholine (45.5 mg, 0.522 mmol) were added to a round-bottom flask. The mixture was treated under reflux conditions in MeCN (10 mL) for 24 h. After completion of the reaction, the solvent was removed under reduced pressure and the desired product was crystallized using EtOAc (5 mL), filtered, and washed with 2 × 5 mL EtOAc.

4.1.2.1 3-(Morpholinomethyl)-4-oxo-N-phenyl-1,4-dihydroquinoline-2-carboxamide (8a). Yield: 104 mg, 66%. Mp.: decomposition over 230 °C. HRMS calcd for $[M + H^+]$ $m/z = 364.1656$, found $m/z = 364.1651$; ^1H NMR (500 MHz, DMSO-d₆): 2.47 (4H, s); 3.54 (4H, s); 3.70 (2H, s); 7.19 (1H, t, $J = 6.97$ Hz); 7.38 (1H, t, $J = 8.52$ Hz); 7.44 (2H, t, $J = 8.52$ Hz); 7.70 (1H, t, $J = 8.52$ Hz); 7.73 (2H, d, $J = 7.75$ Hz); 7.83 (1H, d, $J = 8.52$ Hz); 8.13 (1H, d, $J = 7.75$ Hz); 12.02 (2H, s); ^{13}C NMR (125 MHz, DMSO-d₆): 51.1; 52.5; 66.4; 113.5; 120.9; 124.4; 125.1; 125.7; 129.6; 132.7; 138.5; 162.1; 171.4 (Fig. S25, S26 and S79†).

4.1.2.2 N-Benzyl-3-(morpholinomethyl)-4-oxo-1,4-dihydroquinoline-2-carboxamide (8b). Yield: 106 mg, 65%. Mp.: decomposition over 200 °C. HRMS calcd for $[M + H^+]$ $m/z = 378.1812$, found $m/z = 378.1808$; ^1H NMR (500 MHz, DMSO-d₆): 2.22–2.28 (4H, m); 3.16–3.24 (4H, m); 3.55 (2H, s); 4.59 (2H, d, $J = 5.21$ Hz); 7.30–7.35 (2H, m); 7.38 (2H, t, $J = 7.42$ Hz); 7.43 (2H, d, $J = 7.42$ Hz); 7.66 (1H, t, $J = 7.63$ Hz); 7.89 (1H, d, $J = 8.48$ Hz); 8.08 (1H, d, $J = 7.93$ Hz); 11.21 (1H, t, $J = 5.25$ Hz); 11.76 (1H, s); ^{13}C NMR (125 MHz, DMSO-d₆): 44.1; 50.5; 51.9; 66.3; 113.4; 119.6; 124.2; 124.4; 125.7; 128.0; 128.7; 129.1; 132.6; 138.6; 139.7; 144.0; 163.2; 177.7 (Fig. S27, S28 and S80†).

4.1.2.3 3-(Morpholinomethyl)-4-oxo-N-phenethyl-1,4-dihydroquinoline-2-carboxamide (8c). Yield: 150 mg, 75%. Mp.: decomposition over 300 °C. HRMS calcd for $[M + H^+]$ $m/z = 392.1969$, found $m/z = 392.1960$; ^1H NMR (500 MHz, MeOH-d₄): 2.23–2.32 (4H, m); 3.02 (2H, t, $J = 6.51$ Hz); 3.36–3.48 (4H, m); 3.56 (2H, s); 3.76 (2H, t, $J = 6.51$ Hz); 7.25 (1H, t, $J = 7.17$ Hz); 7.29 (2H, d, $J = 7.17$ Hz); 7.34 (2H, t, $J = 7.17$ Hz); 7.44 (1H, t, $J = 7.17$ Hz); 7.74 (1H, t, $J = 7.17$ Hz); 7.81 (1H, d, $J = 8.76$ Hz); 8.24 (1H, d, $J = 7.96$ Hz); ^{13}C NMR (125 MHz, MeOH-d₄): 34.8; 41.8; 49.9; 51.3; 66.4; 113.6; 118.6; 124.1; 124.3; 125.2; 126.3; 128.4; 128.8; 132.7; 139.5; 162.7; 182.0 (Fig. S29, S30 and S81†).

4.1.2.4 3-(Morpholinomethyl)-4-oxo-N-propyl-1,4-dihydroquinoline-2-carboxamide (8d). Yield: 106 mg, 74%. Mp.: decomposition over 190 °C. HRMS calcd for $[M + H^+]$ $m/z = 330.1812$, found $m/z = 330.1807$; ^1H NMR (500 MHz, MeOH-d₄): 1.06 (3H, t, $J = 7.35$ Hz); 1.69–1.77 (2H, m); 2.58–2.70 (4H, m); 3.47 (2H, t, $J = 6.97$ Hz); 3.67–3.73 (4H, m); 3.78 (2H, s); 7.44 (1H, t, $J = 7.43$ Hz); 7.73 (1H, t, $J = 8.35$ Hz); 7.79 (1H, d, $J = 8.35$ Hz); 8.26 (1H, d, $J = 8.15$ Hz); ^{13}C NMR (125 MHz, MeOH-d₄): 10.7; 22.4; 41.7; 51.7; 66.5; 96.9; 124.3; 125.1; 132.6; 163.4; 178.2 (Fig. S31, S32 and S82†).



4.1.2.5 *N*-(3-(Dimethylamino)propyl)-3-(morpholinomethyl)-4-oxo-1,4-dihydroquinoline-2-carboxamide (8e). Yield: 126 mg, 78%. Mp.: decomposition over 200 °C. HRMS calcd for $[M + H^+]$ $m/z = 373.2234$, found $m/z = 373.2230$; ^1H NMR (500 MHz, DMSO-d₆): 1.70–1.75 (2H, m); 2.15 (6H, s); 2.31 (2H, t, $J = 7.04$ Hz); 2.44–2.52 (4H, m); 3.38–3.41 (2H, m); 3.53–3.66 (6H, m); 7.34 (1H, t, $J = 8.04$ Hz); 7.66 (1H, t, $J = 8.04$ Hz); 7.87 (1H, d, $J = 8.31$ Hz); 8.09 (1H, d, $J = 8.31$ Hz); 10.71 (1H, brs); 11.76 (1H, brs); ^{13}C NMR (125 MHz, DMSO-d₆): 27.4; 38.3; 45.6; 50.8; 52.2; 57.1; 66.6; 113.3; 119.5; 124.1; 124.3; 125.7; 132.6; 139.7; 144.3; 163.2; 177.6 (Fig. S33, S34 and S83†).

4.1.2.6 5-Chloro-*N*-(2-(dimethylamino)ethyl)-3-(morpholinomethyl)-4-oxo-1,4-dihydroquinoline-2-carboxamide (13a). Yield: 128 mg, 75%. Mp.: decomposition over 177 °C. HRMS calcd for $[M + H^+]$ $m/z = 393.1688$, found $m/z = 393.1684$; ^1H NMR (600 MHz, DMSO-d₆): 2.18 (6H, s); 2.44 (2H, t, $J = 6.15$ Hz); 2.46–2.50 (4H, m); 3.48 (5.89 Hz); 3.53–3.61 (6H, m); 7.29 (1H, d, $J = 7.45$ Hz); 7.54 (1H, t, $J = 8.19$ Hz); 7.84 (1H, d, $J = 8.19$ Hz); 10.74 (1H, brs); 11.70 (1H, brs); ^{13}C NMR (150 MHz, DMSO-d₆): 38.0; 45.6; 50.6; 52.2; 58.7; 66.6; 115.1; 119.1; 120.4; 126.7; 132.3; 132.5; 142.5; 143.3; 162.9; 176.8 (Fig. S35–38 and S84†).

4.1.2.7 *N*-(2-(Dimethylamino)ethyl)-3-(morpholinomethyl)-4-oxo-6-phenyl-1,4-dihydroquinoline-2-carboxamide (13b). Yield: 51 mg, 27%. Mp.: 179.4–181.3 °C. HRMS calcd for $[M + H^+]$ $m/z = 435.2391$, found $m/z = 435.2385$; ^1H NMR (600 MHz, DMSO-d₆): 2.20 (6H, s); 2.46 (2H, t, $J = 6.21$ Hz); 2.47–2.55 (4H, m); 3.52 (2H, q, $J = 5.88$ Hz); 3.57–3.61 (4H, m); 3.64 (2H, s); 7.40 (1H, t, $J = 6.97$ Hz); 7.51 (2H, t, $J = 7.96$ Hz); 7.74 (2H, d, $J = 8.08$ Hz); 7.98–8.04 (2H, m); 8.33 (1H, d, $J = 2.35$ Hz); 10.81 (1H, brs); 11.88 (1H, brs); ^{13}C NMR (150 MHz, DMSO-d₆): 38.07; 45.7; 50.6; 52.2; 58.7; 66.6; 113.6; 120.5; 122.9; 124.5; 127.1; 128.0; 129.6; 131.4; 136.0; 139.1; 139.8; 144.0; 163.2; 177.7 (Fig. S39–41 and S85†).

4.1.2.8 7-Chloro-*N*-(2-(dimethylamino)ethyl)-3-(morpholinomethyl)-4-oxo-1,4-dihydroquinoline-2-carboxamide (13c). Yield: 101 mg, 59%. Mp.: 158.3–160.4 °C. HRMS calcd for $[M + H^+]$ $m/z = 393.1688$, found $m/z = 393.1684$; ^1H NMR (600 MHz, DMSO-d₆): 2.18 (6H, s); 2.44 (2H, t, $J = 6.31$ Hz); 2.46–2.50 (4H, m); 3.49 (2H, q, $J = 5.72$ Hz); 3.55–3.61 (6H, m); 7.35 (1H, d, $J = 8.52$ Hz); 7.96 (1H, s); 8.08 (1H, d, $J = 8.86$ Hz); 10.78 (1H, brs); 11.85 (1H, brs); ^{13}C NMR (150 MHz, DMSO-d₆): 38.1; 45.6; 50.5; 52.1; 58.7; 66.5; 114.2; 118.8; 122.9; 124.5; 128.1; 137.3; 140.5; 144.5; 163.0; 177.1 (Fig. S42, S43 and S86†).

4.1.2.9 *N*-(2-(Dimethylamino)ethyl)-8-methyl-3-(morpholinomethyl)-4-oxo-1,4-dihydroquinoline-2-carboxamide (13d). Yield: 119 mg, 73%. Mp.: 105.1–106.6 °C. HRMS calcd for $[M + H^+]$ $m/z = 373.2234$, found $m/z = 373.2230$; ^1H NMR (600 MHz, CDCl₃): 2.28 (6H, s); 2.53–2.59 (5H, m); 2.63–2.73 (4H, m); 3.57–3.87 (8H, m); 7.29 (1H, t, $J = 7.51$ Hz); 7.50 (1H, d, $J = 6.72$ Hz); 8.22 (1H, d, $J = 8.26$ Hz); 9.55 (1H, brs); 11.61 (1H, brs); ^{13}C NMR (150 MHz, CDCl₃): 16.4; 38.5; 45.5; 50.5; 51.7; 58.6; 66.8; 114.3; 124.0; 124.3; 124.5; 125.3; 133.4; 136.9; 140.9; 163.3; 179.4 (Fig. S44, S45 and S87†).

4.1.3. General procedure for the synthesis of lactams. Method A: *N*-(2-(dimethylamino)ethyl)-4-hydroxyquinoline-2-carboxamide (1) or its derivatives (7a–f and 12a–d) (0.070

mmol), paraformaldehyde (2.5 mg, 0.084 mmol), and triethylamine (72.6 mg, 0.717 mmol) were put in a pressure-resistant 10 mL vessel. The reaction was heated in microwave reactor at 150 °C in 1.3 mL MeOH for 30 minutes. The solution was filtered in the case of dimer formation (discussed in the next paragraph) and the mother liquor was transferred to a 25 mL round-bottom flask and the solvent was removed under reduced pressure. The sample was purified using column chromatography (DCM : MeOH 4 : 1 as eluent in SiO₂). The fractions were collected and the solvent was removed under reduced pressure. The desired product was crystallized using EtOAc (5 mL), filtered, and washed using 2 × 5 mL EtOAc.

Method B: *N*-(2-(dimethylamino)ethyl)-4-hydroxy-3-(morpholinomethyl)quinoline-2-carboxamide (2) or its derivatives (8a–f and 13a–d) (2.5 mg, 0.070 mmol), triethylamine (72.6 mg, 0.717 mmol) were put in a pressure-resistant 10 mL vessel. The reaction was heated in microwave reactor at 150 °C in MeOH (1.3 mL) for 30 minutes. The solution was filtered in the case of dimer formation (discussed in next paragraph) and the mother liquor was transferred to a 25 mL round-bottom flask and the solvent was removed under reduced pressure. The sample was purified using column chromatography (DCM : MeOH 4 : 1 as eluent in SiO₂). The fractions were collected and a solvent was removed under reduced pressure. The desired product was crystallized using EtOAc (5 mL), filtered, and washed using 2 × 5 mL EtOAc.

4.1.4 2-(2-(Dimethylamino)ethyl)-1*H*-pyrrolo[3,4-*b*]quinoline-3,9(2*H*,4*H*)-dione (4). Yield: 17 mg, 89%. Mp.: decomposition over 230 °C. HRMS calcd for $[M + H^+]$ $m/z = 272.1394$, found $m/z = 272.1389$; ^1H NMR (500 MHz, MeOH-d₄): 2.35 (6H, s); 2.71 (2H, t, $J = 6.54$ Hz); 3.83 (2H, t, $J = 6.63$ Hz); 4.50 (2H, s); 7.46 (1H, t, $J = 7.51$ Hz); 7.74 (1H, t, $J = 7.69$ Hz); 7.81 (1H, d, $J = 8.43$ Hz); 8.36 (1H, d, $J = 8.34$ Hz); ^{13}C NMR (125 MHz, MeOH-d₄): 40.2; 44.1; 46.9; 56.5; 119.5; 120.7; 123.8; 124.7; 126.4; 131.8; 165.1; 173.9 (Fig. S46, S47 and S88†).

4.1.4.1 2-(3-(Dimethylamino)propyl)-1*H*-pyrrolo[3,4-*b*]quinoline-3,9(2*H*,4*H*)-dione (9e). Yield: 14 mg, 72%. Mp.: decomposition over 240 °C. HRMS calcd for $[M + H^+]$ $m/z = 298.1550$, found $m/z = 298.1545$; ^1H NMR (500 MHz, DMSO-d₆): 1.73–1.81 (2H, m); 2.25 (2H, t, $J = 6.99$ Hz); 3.57 (2H, t, $J = 7.30$ Hz); 4.37 (2H, s); 7.38 (1H, t, $J = 7.44$ Hz); 7.71 (1H, t, $J = 8.07$ Hz); 7.80 (1H, d, $J = 8.35$ Hz); 8.18 (1H, d, $J = 8.07$ Hz); 12.59 (1H, brs); ^{13}C NMR (125 MHz, DMSO-d₆): 26.2; 41.2; 45.6; 47.4; 56.9; 119.9; 120.5; 124.0; 125.4; 126.7; 132.6; 140.9; 141.3; 164.0; 173.6 (Fig. S48, S49 and S89†).

4.1.4.2 2-(2-(Pyrrolidin-1-yl)ethyl)-1*H*-pyrrolo[3,4-*b*]quinoline-3,9(2*H*,4*H*)-dione (9f). Yield: 18 mg, 87%. Mp.: decomposition over 220 °C. HRMS calcd for $[M + H^+]$ $m/z = 286.1550$, found $m/z = 286.1546$; ^1H NMR (500 MHz, DMSO-d₆): 1.63–1.73 (4H, m); 2.50–2.56 (4H, m); 2.72 (2H, t, $J = 6.25$ Hz); 3.68 (2H, t, $J = 6.10$ Hz); 4.43 (2H, s); 7.39 (1H, t, $J = 7.62$ Hz); 7.72 (1H, t, $J = 7.62$ Hz); 7.80 (1H, d, $J = 7.62$ Hz); 8.19 (1H, d, $J = 7.62$ Hz); 12.61 (1H, brs); ^{13}C NMR (125 MHz, DMSO-d₆): 23.6; 41.6; 47.6; 53.9; 54.1; 119.9; 124.0; 125.4; 132.6; 140.8; 141.2; 163.9; 173.6 (Fig. S50, S51 and S90†).

4.1.4.3 8-Chloro-2-(2-(dimethylamino)ethyl)-1*H*-pyrrolo[3,4-*b*]quinoline-3,9(2*H*,4*H*)-dione (14a). Yield: 6 mg, 28%. Mp.:



decomposition over 295 °C. HRMS calcd for $[M + H^+]$ $m/z = 306.1004$, found $m/z = 306.0999$; ^1H NMR (600 MHz, DMSO-d₆): 2.22 (6H, s); 2.52–2.58 (2H, m); 3.65 (2H, t, $J = 6.05$ Hz); 4.37 (2H, s); 7.34 (1H, d, $J = 7.44$ Hz); 7.60 (1H, t, $J = 8.01$ Hz); 7.75 (1H, d, $J = 8.05$ Hz); 12.68 (1H, brs); ^{13}C NMR (150 MHz, DMSO-d₆): 40.3; 45.4; 47.6; 57.2; 119.4; 122.7; 126.6; 132.3; 132.7; 140.2; 143.7; 163.7; 173.1 (Fig. S52, S53 and S91†).

4.1.4.4 6-Chloro-2-(2-(dimethylamino)ethyl)-1*H*-pyrrolo[3,4-*b*]quinoline-3,9(2*H*,4*H*)-dione (14c). Yield: 5 mg, 23%. Mp.: decomposition over 295 °C. HRMS calcd for $[M + H^+]$ $m/z = 306.1004$, found $m/z = 306.0999$; ^1H NMR (600 MHz, DMSO-d₆): 2.20 (6H, s); 2.52–2.56 (2H, m); 3.65 (2H, t, $J = 5.86$ Hz); 4.41 (2H, s); 7.41 (1H, d, $J = 8.91$ Hz); 7.82 (1H, s); 8.18 (1H, d, $J = 8.61$ Hz); 12.74 (1H, brs); ^{13}C NMR (150 MHz, DMSO-d₆): 40.3; 45.4; 47.5; 57.3; 119.0; 121.4; 124.4; 125.4; 127.8; 137.2; 141.7; 163.7; 173.0 (Fig. S54–56 and S92†).

4.1.4.5 2-(2-(Dimethylamino)ethyl)-5-methyl-1*H*-pyrrolo[3,4-*b*]quinoline-3,9(2*H*,4*H*)-dione (14d). Yield: 9 mg, 47%. Mp.: decomposition over 230 °C. HRMS calcd for $[M + H^+]$ $m/z = 286.1550$, found $m/z = 286.1545$; ^1H NMR (500 MHz, MeOH-d₄): 2.39 (6H, s); 2.68 (3H, s); 2.76 (2H, t, $J = 5.98$ Hz); 3.85 (2H, t, $J = 6.79$ Hz); 4.51 (2H, s); 7.39 (1H, t, $J = 7.74$ Hz); 7.65 (1H, d, $J = 7.74$ Hz); 8.23 (1H, d, $J = 8.45$ Hz); ^{13}C NMR (125 MHz, MeOH-d₄): 40.2; 44.1; 46.9; 56.4; 120.7; 123.0; 123.9; 133.6; 164.6; 175.0 (Fig. S57, S58 and S93†).

4.1.5. General procedure for the synthesis of dimers. Method A: *N*-(2-(dimethylamino)ethyl)-4-hydroxyquinoline-2-carboxamide (1) or its derivatives (7a–f and 12a–d) (0.070 mmol), paraformaldehyde (2.5 mg, 0.070 mmol), and triethylamine (72.6 mg, 0.717 mmol) were put in a pressure-resistant 10 mL vessel. The reaction was heated in microwave reactor at 150 °C in 1.3 mL MeOH for 2 h. The formed precipitate was filtered and washed with 2 × 2 mL MeOH.

Method B: *N*-(2-(dimethylamino)ethyl)-4-hydroxy-3-(morpholinomethyl)quinoline-2-carboxamide (2) or its derivatives (8a–f and 13a–d) (2.5 mg, 0.070 mmol), triethylamine (72.6 mg, 0.717 mmol) were put in a pressure-resistant 10 mL vessel. The reaction was heated in microwave reactor at 150 °C in MeOH (1.3 mL) for 2 h. The formed precipitate was filtered and washed with 2 × 2 mL MeOH.

4.1.5.1 3,3'-Methylenebis(4-oxo-*N*-phenyl-1,4-dihydroquinoline-2-carboxamide) (10a). Yield: 9 mg, 48%. Mp.: decomposition over 300 °C. HRMS calcd for $[M + H^+]$ $m/z = 541.1870$, found $m/z = 541.1863$; ^1H NMR (500 MHz, DMSO-d₆): 4.06 (2H, s); 7.17 (2H, t, $J = 7.38$ Hz); 7.40 (6H, m); 7.75 (2H, t, $J = 7.64$ Hz); 7.82 (6H, d, $J = 8.18$ Hz); 8.16 (2H, d, $J = 8.18$ Hz); 12.70 (2H, s); 13.38 (2H, s); ^{13}C NMR (125 MHz, DMSO-d₆): 26.7; 114.7; 120.1; 124.0; 124.7; 125.0; 125.2; 129.6; 133.1; 138.7; 145.2; 145.8; 146.4; 161.9; 178.4 (Fig. S59, S60 and S94†).

4.1.5.2 3,3'-Methylenebis(*N*-benzyl-4-oxo-1,4-dihydroquinoline-2-carboxamide) (10b). Yield: 10 mg, 49%. Mp.: decomposition over 300 °C. HRMS calcd for $[M + H^+]$ $m/z = 569.2183$, found $m/z = 569.2178$; ^1H NMR (600 MHz, DMSO-d₆): 3.94 (2H, s); 4.66 (4H, d, $J = 5.90$ Hz); 7.26 (2H, t, $J = 7.13$ Hz); 7.31 (2H, t, $J = 7.56$ Hz); 7.34 (4H, t, $J = 7.56$ Hz); 7.43 (4H, d, $J = 7.56$ Hz); 7.68 (2H, t, $J = 7.56$ Hz); 7.78 (2H, d, $J = 8.64$ Hz); 7.96 (2H, d, $J = 8.10$ Hz); 11.63 (2H, t, $J = 5.89$ Hz); 12.46 (2H, s); ^{13}C

NMR (150 MHz, DMSO-d₆): 26.3; 43.2; 115.0; 119.0; 123.9; 124.3; 125.1; 127.6; 128.0; 128.9; 132.8; 138.8; 139.4; 145.0; 163.8; 178.2 (Fig. S61, S62 and S95†).

4.1.5.3 3,3'-Methylenebis(4-oxo-*N*-propyl-1,4-dihydroquinoline-2-carboxamide) (10d). Yield: 7 mg, 38%. Mp.: decomposition over 300 °C. HRMS calcd for $[M + H^+]$ $m/z = 473.2183$, found $m/z = 473.2178$; ^1H NMR (600 MHz, DMSO-d₆): 0.94 (6H, t, $J = 6.83$ Hz); 1.57–1.63 (4H, m); 3.39 (4H, t, $J = 6.07$ Hz); 3.94 (2H, s); 7.32 (2H, t, $J = 7.83$ Hz); 7.68 (2H, t, $J = 7.83$ Hz); 7.77 (2H, d, $J = 7.83$ Hz); 8.01 (2H, d, $J = 7.83$ Hz); 11.13 (2H, s); 12.40 (2H, s); ^{13}C NMR (150 MHz, DMSO-d₆): 12.1; 22.7; 29.5; 41.4; 114.6; 119.0; 123.9; 124.2; 125.1; 130.1; 132.8; 139.4; 145.4; 163.6; 178.2 (Fig. S63, S64 and S96†).

4.1.5.4 3,3'-Methylenebis(5-chloro-*N*-(2-(dimethylamino)ethyl)-4-oxo-1,4-dihydroquinoline-2-carboxamide) (15a). Yield: 4 mg, 19%. Mp.: decomposition over 280 °C. HRMS calcd for $[M + H^+]$ $m/z = 599.1935$, found $m/z = 599.1930$; ^1H NMR (600 MHz, DMSO-d₆): 2.10 (12H, s); 2.42–2.50 (4H, m); 3.48 (4H, q, $J = 5.70$ Hz); 3.97 (2H, s); 7.28 (2H, d, $J = 7.64$ Hz); 7.56 (2H, t, $J = 7.64$ Hz); 7.71 (2H, d, $J = 8.61$ Hz); 10.66 (2H, brs); ^{13}C NMR (150 MHz, DMSO-d₆): 26.2; 37.7; 45.3; 57.9; 117.2; 118.6; 120.0; 126.7; 132.1; 132.3; 142.1; 144.1; 163.4; 177.8 (Fig. S65, S66 and S97†).

4.1.5.5 3,3'-Methylenebis(7-chloro-*N*-(2-(dimethylamino)ethyl)-4-oxo-1,4-dihydroquinoline-2-carboxamide) (15c). Yield: 3 mg, 14%. Mp.: decomposition over 300 °C. HRMS calcd for $[M + H^+]$ $m/z = 599.1935$, found $m/z = 599.1933$; ^1H NMR (600 MHz, DMSO-d₆): 2.11 (12H, s); 2.42–2.50 (4H, m); 3.48–3.52 (4H, m); 4.07 (2H, s); 7.33 (2H, d, $J = 8.86$ Hz); 7.78 (2H, s); 8.01 (2H, d, $J = 8.83$ Hz); 10.84 (2H, brs); ^{13}C NMR (150 MHz, DMSO-d₆): 25.9; 37.7; 45.3; 58.0; 116.1; 118.2; 122.5; 124.6; 127.5; 145.6; 177.8 (Fig. S67–70 and S98†).

Data availability

The data supporting this article have been included as part of the ESI.†

Author contributions

Conceptualization, I. S., B. L.; investigation, A. H., R. B., S. J. R.; writing – original draft preparation, B. L., S. J. R.; writing – review and editing, I. S., B. L. All authors have read and agreed to the published version of the manuscript.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- 1 É. Rózsa, H. Robotka, L. Vécsei and J. Toldi, *J. Neural. Transm.*, 2008, **115**, 1087–1091.
- 2 T. W. Stone, *Expert Opin. Invest. Drugs*, 2001, **10**, 633–645.
- 3 K. Sas, H. Robotka, J. Toldi and L. Vécsei, *J. Neurol. Sci.*, 2007, **257**, 221–239.
- 4 G. Gigler, G. Szénási, A. Simó, G. Lévay, L. G. Hársing Jr, K. Sas, L. Vécsei and J. Toldi, *Eur. J. Pharmacol.*, 2007, **564**, 116–122.
- 5 E. Luchowska, P. Luchowski, A. Sarnowska, M. Wielosz, W. A. Turski and E. M. Urbańska, *Pol. J. Pharmacol.*, 2003, **55**, 443–447.
- 6 B. L. Harrison, B. M. Baron, D. M. Cousino and I. A. McDonald, *J. Med. Chem.*, 1990, **12**, 3130–3132.
- 7 D. Edmont, R. Rocher, C. Plisson and J. Chenault, *Bioorg. Med. Chem. Lett.*, 2000, **16**, 1831–1834.
- 8 F. P. Bonina, L. Arenare, R. Ippolito, G. Boatto, G. Battaglia, V. Bruno and P. de Caprariis, *Int. J. Pharm.*, 2000, **202**, 79–88.
- 9 S. Manfredini, B. Pavan, S. Vertuani, M. Scaglianti, D. Compagnone, C. Biondi, A. Scatturin, S. Tanganelli, L. Ferraro, P. Prasad and A. Dalpiaz, *J. Med. Chem.*, 2002, **45**, 559–562.
- 10 S. Manfredini, S. Vertuani, B. Pavan, F. Vitali, M. Scaglianti, F. Bortolotti, C. Biondi, A. Scatturin, P. Prasad and A. Dalpiaz, *Bioorg. Med. Chem.*, 2004, **12**, 5453–5463.
- 11 A. C. Nichols and K. L. Yielding, *Mol. Chem. Neuropathol.*, 1998, **35**, 1–12.
- 12 A. Brik, Y. C. Lin, J. Elder and C. H. Wong, *Chem. Biol.*, 2002, **9**, 891–896.
- 13 E. Knyihár-Csillik, A. Mihály, B. Krisztin-Péva, H. Robotka, I. Szatmári, F. Fülöp, J. Toldi, B. Csillik and L. Vécsei, *Neurosci. Res.*, 2008, **61**, 429–432.
- 14 F. Fülöp, I. Szatmári, E. Vámos, D. Zádori, J. Toldi and L. Vécsei, *Curr. Med. Chem.*, 2009, **16**, 4828–4842.
- 15 D. Zádori, G. Nyíri, A. Szőnyi, I. Szatmári, F. Fülöp, J. Toldi, T. F. Freund, L. Vécsei and P. Klivényi, *J. Neural. Transm.*, 2011, **118**, 865–875.
- 16 D. Zádori, G. Veres, L. Szalárdy, P. Klivényi, J. Toldi and L. Vécsei, *J. Alzheimer. Dis.*, 2014, **42**, S177–S187.
- 17 R. Greco, C. Demartini, A. M. Zanaboni, E. Redavide, S. Pampalone, J. Toldi, F. Fülöp, F. Blandini, G. Nappi, G. Sandrini, L. Vécsei and C. Tassorelli, *Cephalalgia*, 2017, **37**, 1272–1284.
- 18 A. Fejes-Szabó, Z. Bohár, E. Vámos, G. Nagy-Grócz, L. Tar, G. Veres, D. Zádori, M. Szentirmai, J. Tajti, I. Szatmári, F. Fülöp, J. Toldi, A. Paardutz and L. Vecsei, *J. Neural. Transm.*, 2014, **121**, 725–738.
- 19 B. Lőrinczi, A. Csámpai, F. Fülöp and I. Szatmári, *Molecules*, 2020, **25**, 937.
- 20 B. Lőrinczi, A. Csámpai, F. Fülöp and I. Szatmári, *RSC Adv.*, 2021, **11**, 543–554.
- 21 P. Barta, F. Fülöp and I. Szatmári, *Beilstein J. Org. Chem.*, 2018, **14**, 560–575.
- 22 C. Trarat, S. Giorgi-Renault and H. Henri-Philippe, *Synlett*, 1998, **10**, 1071–1072.
- 23 R. Cincinelli, G. Beretta and S. Dallavalle, *Tetrahedron Lett.*, 2018, **59**, 163–166.
- 24 M. Anzini, A. Cappelli, S. Vomero, M. Seeber, M. C. Menziani, T. Langer, B. Hagen, C. Manzoni and J. J. Bourguignon, *J. Med. Chem.*, 2001, **8**, 1134.
- 25 Y. Q. Liu, W. Q. Li, S. L. Morris-Natschke, K. Qian, L. Yang, G. X. Zhu, X. B. Wu, A. L. Chen, S. Y. Zhang, X. Nan and K. H. Lee, *Med. Res. Rev.*, 2015, **4**, 753.
- 26 A. A. Boteva, I. V. Fefilova, G. A. Triandafilova, V. V. Maslova, S. Y. Solodnikov and O. P. Krasnykh, *Pharm. Chem. J.*, 2019, **53**, 616–619.
- 27 B. Lőrinczi, P. Simon and I. Szatmári, *Int. J. Mol. Sci.*, 2022, **13**, 7152.
- 28 J. H. Burckhalter, V. C. Stephens, H. C. Scarborough Jr, W. S. Brinigar and W. H. Edgerton, *J. Am. Chem. Soc.*, 1954, **19**, 4902–4906.
- 29 M. Movrin, D. Maysinger and E. Marok, *Pharmazie*, 1980, **8**, 458–460.
- 30 P. Funk, K. Motyka, M. Soural, M. Malon, H. Koshino, J. Kusz and J. Hlavac, *PLoS One*, 2017, **12**, e0175364.
- 31 E. Fehér, I. Szatmári, T. Dudás, A. Zalatnai, T. Farkas, B. Lőrinczi, F. Fülöp, L. Vécsei and J. Toldi, *Molecules*, 2019, **19**, 3502.

