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Metal-based non-enzymatic systems for cholesterol detection: mechanisms, features, and performance

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Metal based catalysts and electrodes are versatile tools known for their redox properties, catalytic efficiency, and stability under various conditions. Despite the absence of significant scientific hurdles, the utilization of these methods in cholesterol detection, particularly in non-enzymatic approaches, has been relatively underexplored. To this end, there is a pressing need to delve deeper into existing metal-based systems used in non-enzymatic cholesterol sensing, with the goal of fostering the development of innovative practical solutions. Various electrode systems, such as those employing Ni, Ti, Cu, Zn, W, Mn, and Fe, have already been reported for non-enzymatic cholesterol detection, some of them elucidated sensing mechanisms and potential in physiological detection. A detailed mechanistic understanding of oxide-based cholesterol sensors, along with the methodologies for constructing such systems, holds promise of advancing the exploration of practical applications. This review aims to provide a broad perspective on metal oxide systems and their characteristics that are conducive to non-enzymatic cholesterol sensing. It is intended to serve as a springboard with offering a guide to the design and development of efficient and sensitive electrochemical cholesterol sensors.

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Introduction

Cholesterol is essential for human life, yet when its levels exceed a critical threshold within the body, it can become a substantial hazard to human well-being. Maintaining a controlled cholesterol intake is vital for supporting crucial physiological functions. Nonetheless, lifestyle choices and dietary habits can contribute to an excessive accumulation of cholesterol in the bloodstream, giving rise to significant health issues. Timely identification of cholesterol levels in the human body, along with appropriate interventions, is of great importance for preserving health and safeguarding human lives. In addition to traditional chemical techniques, enzymatic assay, enzymatic calorimetry, microphotometry, gas chromatography, liquid chromatography, and mass spectrometry are some of the methods used to detect cholesterol.^{1–5} The majority of them require expensive, time-consuming, and complex techniques for the effective and precise detection of cholesterol. At the same time electrochemical sensors are excellent substitutes

over traditional analytical methods due to their quick response, high reproducibility, low limit of detection (LOD), and wide concentration range of application.^{6,7}

When using an electrochemical sensor, the species on the electrode's surface interact chemically with the analyte, in this case cholesterol, and produces an electrical signal of either current, potential, conductivity, or resistivity. Based on their operating principles, these sensors are known as amperometric, potentiometric, conductometric, or impedimetric sensors, respectively.⁸ A component of the electrochemical biosensor, *i.e.*, the transducer, then transforms the signal into a detectable format.

Many different materials have been investigated for the construction of enzymatic, non-enzymatic, and redox-mediated electrochemical biosensors.^{9–16} Non-enzymatic sensors are receiving considerable attention due to their excellent characteristics, such as the ability to incorporate metals, metal oxides, and composites as electrocatalysts, and strong interaction between the modified electrode and the analyte. This results in high conductivity, biocompatibility, and facile device miniaturization, while these structures are generally unaffected by atmospheric conditions. Conductive polymer composites,^{17–19} carbonaceous materials,^{20–23} metal nanoparticles,^{24–29} metal sulphides,^{30–32} metal oxides,^{29–36} and their composites are typical choices of active material for electrochemical biosensors including cholesterol sensors.³⁷

Nanomaterials, as the electroactive component in electrochemical sensors, exhibit diverse mechanisms when employed

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for cholesterol detection. However, unlike enzymatic sensors, active electrodes are the only essential component in non-enzymatic devices. The integration of nanomaterials into sensor systems offers numerous advantages, like customized electrical and optical characteristics, and enhanced sensitivity due to their high surface area. Polyaniline (PANI) or polypyrrole (PPy) are predominantly reported as polymeric constituents in the literature, such as in CuO-PANI with a fibrous murexide matrix,¹⁴ PANI/MWCNTs/starch modified carbon paste,¹⁵ mesoporous PANI nanofiber decorated graphene microflowers,³⁸ carboxy-functionalized graphene oxide (fGO) decorated with magnetite (Fe₃O₄) nanoparticles on PANI,³⁹ Ru-phosphate (Ru-Pi)/PPy modified carbon fiber paper,⁴⁰ Au@NiO decorated PPy composite.⁴¹ Additionally, alternative polymer-based materials, such as poly(ionic liquid) poly(vinylbutylimidazolium)-cobalt polyoxometalate supported on carbon,¹⁶ poly(3,4-ethylenedioxythiophene) incorporating taurine,⁴² have also been investigated in this context. Carbon-based materials encompass β -cyclodextrin (β -CD)-functionalized graphene and reduced graphene oxide (rGO),⁴³ a graphene oxide-derived molecular imprinted polymer (GO-MIP) serving as the active material,⁴⁴ a network of platinum nanoparticle-functionalized carbon nanotubes (CNTs) assembled through layer-by-layer techniques,⁴⁵ CNTs derived from coconut oil,⁴⁶ multi-walled carbon nanotubes (MWCNTs) that have been functionalized with gold nanoparticles (Au NPs),⁴⁷ and a disposable screen-printed carbon electrode (SPCE) constructed from MWCNTs and β -CD.⁴⁸

Here we review non-enzymatic electrochemical cholesterol sensors constructed of metal compounds and composites and focusing on the mechanistic understanding of the interaction between the electrode materials and the analyte. The goal of this review is to facilitate future advancements in the field by offering valuable insights into the underlying physical-chemical processes, and to aid with the development of innovative and high-performance sensor chips.

Chemistry of electrochemical cholesterol sensors

An electrochemical biosensor operates by detecting the presence or concentration of a specific biological target (analyte) through interactions with specialized materials in its recognition layer. These interactions generate signals that convey information about the analyte. The recognition layer typically comprises an electrode with functional groups capable of binding to the analyte. For instance, in cholesterol detection the electrode surface may feature enzymes that interact with cholesterol molecules or a substance capable of initiating a redox process upon contact with cholesterol. Redox mediators can also be incorporated to enhance the sensitivity of the biosensor when cholesterol molecules attach to the recognition layer.^{49–51} Subsequently, a transducer transforms the resulting physical-chemical or electrochemical interactions into electrical signals. Finally, a processor processes and converts these signals into a digital output, which can be easily displayed and interpreted.

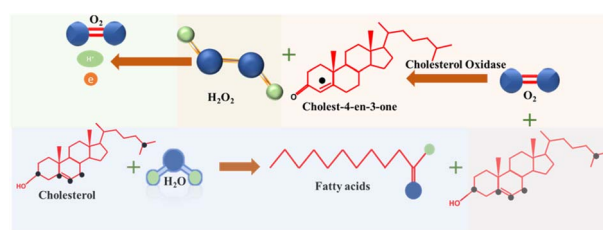
Sensing mechanism in different types of cholesterol sensors

The foundation of electrochemical cholesterol sensing involved the integration of enzymes, such as cholesterol oxidase (ChOx) and cholesterol esterase (ChE), within the recognition layer. These enzymes were chosen for their capacity to either bind to or undergo reactions with the target cholesterol molecules as part of the sensing process.

In enzymatic cholesterol sensors ChE initiates a hydrolysis reaction, breaking down cholesterol ester into free cholesterol and fatty acids, which in turn provides the total cholesterol content in the sample. Simultaneously, a cholesterol molecule undergoes oxidation through the interaction with ChOx, resulting in the production of cholest-4-ene-3-one. Notably, hydrogen peroxide (H₂O₂) generated as a byproduct during this redox reaction serves as redox agent for the electrochemical detection of cholesterol at the sensor, eliminating the need for additional redox mediators. *E.g.*, in amperometric sensors current is measured when H₂O₂ is oxidized at an applied potential, which in turn correlates with cholesterol concentration. However, in some instances, the current response associated with dioxygen electroreduction may also be used. Chemical reactions taking place in enzymatic cholesterol sensing are depicted in Scheme 1.⁸ A significant anodic potential of approximately +0.7 V *vs.* RHE is required for the oxidation of H₂O₂, which represents a significant drawback of this technique. This issue is addressed by incorporating redox mediators alongside the enzymes to facilitate electron transfer.

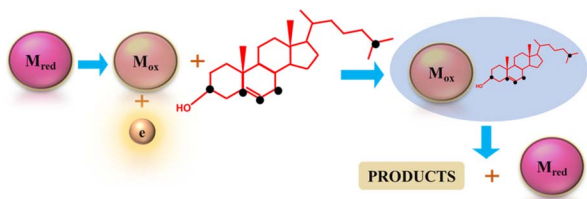
In the redox mediator based enzymatic detection techniques, ferrocene,⁷ methylene blue,⁸ Prussian blue,⁵² hydroquinone,⁵³ poly(*o*-phenylenediamine),⁵⁴ and (hydroxymethyl) ferrocene⁵⁵ were reported to be implemented in the system. Here, the regeneration of the reduced form of the redox mediators by electrical potential is utilized for the detection process. First the oxidized mediator forms an adduct with cholesterol. The latter then gets oxidized into various products, while the redox mediator is electrochemically reduced to generate the signal as shown in Scheme 2.⁷

Horseradish peroxidase (HRP) based bienzymatic mediator-free systems are also capable of sensing cholesterol to some extent.^{56–58} While these enzymatic systems exhibit specificity, sensitivity, and speed in cholesterol sensing, they do face certain limitations. The latter primarily revolves around high costs and limited tolerance to variations in environmental



Scheme 1 Reactions involved in enzymatic detection of cholesterol.





Scheme 2 Reactions involved in the redox mediator-based detection of cholesterol.

factors, such as pH and temperature. These shortcomings pose challenges when considering the practicality and economic viability of these systems in real-world applications. Consequently, numerous researchers have undertaken efforts to develop simpler, more efficient, and cost-effective non-enzymatic cholesterol sensors.

A non-enzymatic cholesterol sensor comprises a recognition layer devoid of enzymes that interacts with cholesterol molecules. The fundamental role of this layer remains consistent across various materials employed for this purpose, which primarily is to serve as cholesterol oxidants facilitated by electrical processes. These non-enzymatic systems necessitate the use of a conductive electrolyte containing cholesterol to obtain an electrical response during the interaction between the electrode and the analyte. Numerous electrolyte combinations were explored with cholesterol based on their chemical and electrochemical properties. Also, various electrode types, ranging from simple bare electrodes to modified nanostructures, were investigated as recognition layers in non-enzymatic systems.

The resulting products and the pathways of electro-oxidation can vary depending on the conditions and materials, since cholesterol contains multiple sites for oxidation. The sites

susceptible to oxidation, as depicted in Fig. 1, lead to a diverse array of products during electro-oxidation.⁵⁹ However, the chemistry of all the products has not been completely confirmed, yet.

Metal-based systems for non-enzymatic detection of cholesterol

Cholesterol sensing can be achieved using a bare Pt or C electrode in combination with an organic medium.^{60–63} However, the resulting product and cholesterol oxidation efficiency vary due to differences in electrode characteristics and the composition of the medium.^{60–75} Table 1 overviews non-enzymatic cholesterol oxidation and detection efficiencies (LOD, LOQ), product distributions at the indicated potentials using metal-based electrodes with various supporting electrolytes. Certain data points are not available (NA), as some studies focus on electro-oxidation mechanisms, while others address the direct sensing characteristics. It is also worth noting, that although according to the IUPAC definition, the LOQ/LOD ratio must be $10/3 \approx 3.33$, this relationship is not fulfilled for some of the reported performance metrics in Table 1. These data are marked by asterisks.

Metal oxide-based non-enzymatic cholesterol sensors – material development, characteristics, and performance

Rengaraj *et al.* innovatively engineered a flowerlike NiO structure atop high-quality graphene to enable non-enzymatic cholesterol sensing.⁶⁷ Graphene is the optimal support in biosensor applications because of its exceptional carrier

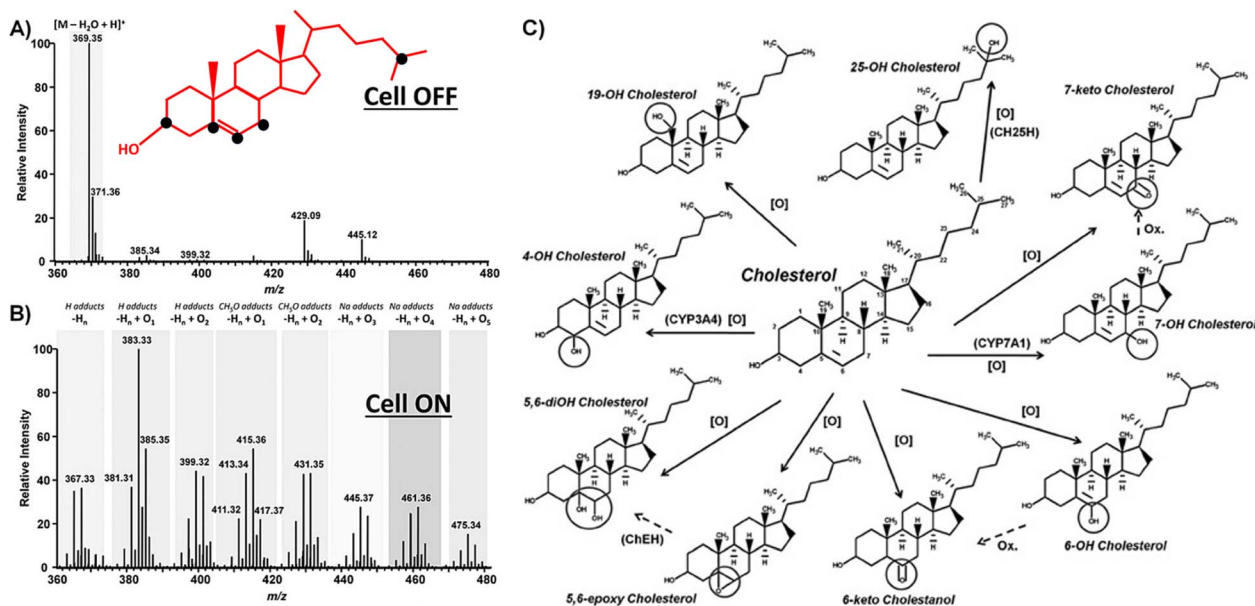


Fig. 1 Oxidizable sites (black dots) in the cholesterol molecule along with the MS spectra of cholesterol (100 mmol L^{-1}) before (A) and after EC-oxidation (B). The corresponding potential products of cholesterol electrooxidation. (C) Reprinted with permission from ref. 59. Copyright 2016 John Wiley and Sons and Copyright Clearance Center (LN: 5833140916083).



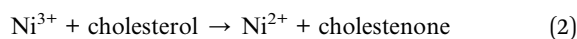
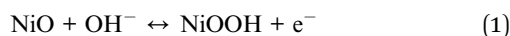


Table 1 Details of non-enzymatic cholesterol detectors using bare metal or modified electrodes^a

Electrode	Supporting electrolytes (media)	Cholesterol oxidation products	Oxidation potential (V)	Sensitivity	LOD	LOQ	Ref.
Pt	Glacial acetic acid, sodium perchlorate and sodium acetate	7 α -Acetoxycholesterol and 7 β -acetoxycholesterol	1.7 (vs. SCE)	NA	NA	NA	60
Pt	TBATPB in dichloromethane and cholesterol in glacial acetic acid/acetonitrile	Cholesterol dichlorides/chlorohydrine/dicholesteryl ether/cholesterol acetate/3 β -acetylamino derivative of cholesterol	1.75 (vs. Ag/0.1 M AgNO ₃)	NA	NA	NA	61
Pencil Pb	Acetonitrile, acetone and lithium perchlorate	Cholesta-4,6-dien-3-one	1.45 (vs. SCE)	1455.22 μ A mM ⁻¹ cm ⁻²	0.625 mM*	9.375 mM*	64
Au/Pt	Triton X-100, 2-propanol and PBS	NA	0.3–0.2 (vs. Ag/AgCl)	226.2 μ A mM ⁻¹ cm ⁻²	0.015 mM*	5 mM*	65
Cu ₂ S/Cu	Acetate buffers containing cholesterol	NA	NA	62.5 μ A mM ⁻¹ cm ⁻²	0.1 μ M*	6.8 mM*	66
NiO/graphene	PBS containing isopropanol and Triton X-100	5-Cholesten-3-one 4-Cholesten-3-one	0.55 (vs. Ag/AgCl)	40.6 μ A mM ⁻¹ cm ⁻²	0.13 μ M	40 μ M	67
(MnO ₂ /GR/PGE) ZnO NRs Ag NPs–ZnO NRs	PBS PBS	Cholestenone Oxysterols (7 β -ketocholesterol, 7 α -hydroxycholesterol and 7 β -hydroxycholesterol)	–0.24 (vs. SCE) +0.5 (vs. Ag/AgCl)	63 869 μ A mM ⁻¹ cm ⁻² 4.2 μ A mM ⁻¹ cm ⁻² 135.5 μ A mM ⁻¹ cm ⁻²	0.42 nM 1.78 mM 0.184 mM	1.2 nM 9 mM	68 69
PMO–NiO/MoS ₂ /SPCE	PBS	Oxysterols	–0.27 (vs. NA)	7.95 $\times 10^{-6}$ A mg ⁻¹ dL ⁻¹ cm ⁻²	0.24 mg dL ⁻¹ *	15 mg dL ⁻¹ *	70
TiO ₂ /GO	PBS	NA	0.619 (vs. SCE)	NA	0.05 μ M*	0.1 μ M*	71
Cu/Ni dispersed carbon nanofiber/polymer	PBS	NA	0.308 (Ag/AgCl (0.3 M KCl))	226.30 μ A mM ⁻¹ cm ²	2 μ g dL ⁻¹	6.2 μ g dL ⁻¹	72
nanocomposite SiO ₂ /Fe ₂ O ₃ /MWCNT	NA	NA	–0.6 (vs. Ag/AgCl)	NA	5 μ M	NA	75
Polyindole/WC/SS	NA	NA	0.45 (vs. Ag/AgCl)	NA	1.23 $\times 10^{-6}$ mol L ⁻¹	NA	76
Cu ₂ O–TiO ₂	PBS and isopropanol	Oxysterols (7 β -ketocholesterol, 7 α -hydroxycholesterol and 7 β -hydroxycholesterol)	–1.04 V (vs. Ag/AgCl)	6034.04 μ A mM ⁻¹ cm ⁻²	0.05 μ M*	622 μ M*	77

^a * – the IUPAC definition of LOQ/LOD = 10/3 \approx 3.33 is not fulfilled for these reported values.

mobility, ambipolar behavior, and unique energy structure. The single atom-thick carbon layer was deposited onto copper *via* chemical vapor deposition using methane and oxygen as reagents at $\sim 1000^\circ\text{C}$, and subsequently transferred onto glassy carbon electrode (GCE) by the PMMA method. NiO was then electrodeposited onto the graphene flakes from an electrolyte containing $\text{Ni}(\text{NO}_3)_2$ with acetate buffering as it is outlined in Fig. 2A. After annealing under air at 270°C the performance of the developed electrodes was assessed in the absence of any enzymes in a solution of isopropanol and Triton X-100 containing varying concentration of cholesterol in phosphate buffer (PBS) and 1 M KOH by cyclic voltammetry (CV) and chronoamperometry (CA). The fabricated sensor exhibited a detection range spanning from $2\ \mu\text{M}$ to $40\ \mu\text{M}$ cholesterol concentrations with LOD of $0.13\ \mu\text{M}$ and a rapid response time of 5 seconds. The attractive performance can be attributed to the electrocatalytic properties of NiO and the large specific surface area provided by graphene. The reaction mechanism involves the electrochemical conversion of Ni^{2+} to Ni^{3+} by hydroxyl ions in the surrounding medium, which is subsequently followed by the oxidation of cholesterol.⁶⁷



This process results in a linear increase in the current associated with cholesterol oxidation as cholesterol concentration rises, and the electrodes exhibit a high sensitivity of $40.6\ \text{mA}\ \text{mM}^{-1}\ \text{cm}^{-2}$ with good stability throughout the study.

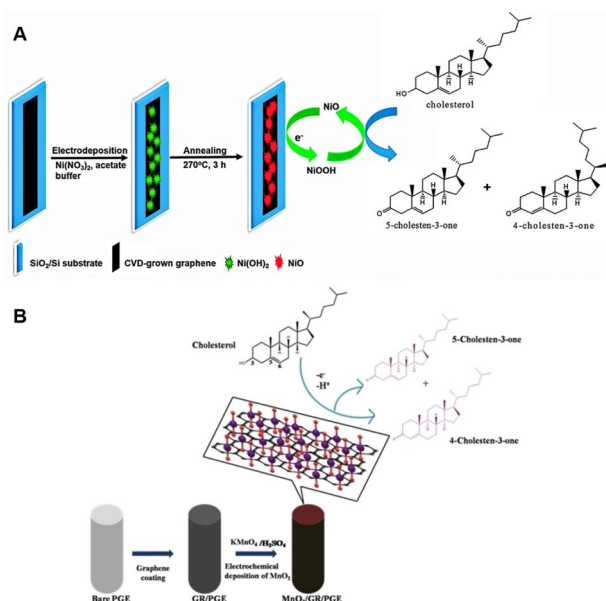


Fig. 2 Fabrication of a NiO/graphene composite electrode for cholesterol sensing (A). Reprinted with permission from ref. 67. Copyright 2015 Royal Society of Chemistry (LN: 1507244-1), and the schematics of the electrocatalytic oxidation of cholesterol at $\text{MnO}_2/\text{GR}/\text{PGE}$ (B), reprinted with permission from ref. 68. Copyright 2020 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim (LN: 5833150494623).

In the study of Rison *et al.* graphene-modified pencil graphite electrode (PGE) was decorated with MnO_2 nanoclusters.⁶⁸ Initially, graphene was synthesized through the exfoliation of 60 mesh particle size graphite in acidic medium, followed by heating and exfoliation in a microwave oven. The resulting graphene was further purified using a solvent mixture containing acetone, dimethylsulfoxide, and water. For sensor electrode fabrication, a pre-treated pencil graphite electrode (PGE) was brush-coated with a paste composed of graphene, polyvinylidene chloride (PVDF), and *N*-methyl-2-pyrrolidene (NMP). MnO_2 was then electrodeposited onto the heat-treated graphene-coated PGE from $\text{KMnO}_4/\text{H}_2\text{SO}_4$ solution. Electrochemical behavior was investigated at various pHs, and cholesterol sensing performance were assessed over several cycles by performing CV and Differential Pulse Voltammetry (DPV) in PBS buffer and with interfering substances modeling human blood serum. Enhanced electron transfer ability was observed due to the presence of catalytic MnO_2 as evidenced by the elevated anodic peak current at $-0.2\ \text{V}$ (vs. SCE) when compared to that of the bare electrode. LOD and LOQ were found to be $4.2 \times 10^{-10}\ \text{M}$ and $12 \times 10^{-10}\ \text{M}$, respectively. Furthermore, the cholesterol sensing process on $\text{MnO}_2/\text{G}/\text{PGE}$ is diffusion-controlled involving a two-electron transfer mechanism resulting in the oxidation of cholesterol to cholestenone as outlined in Fig. 2B.

ZnO seeds were precipitated directly onto a Pt interdigitated electrode by annealing $\text{Zn}(\text{CH}_3\text{COO})_2$, where then ZnO nanorods were grown from those seeds *via* hydrothermal synthesis. The as-fabricated sensor displayed a cholesterol sensitivity of $4.2\ \mu\text{A}\ \text{mM}^{-1}\ \text{cm}^{-2}$. This was further improved by Ag nanoparticle deposition ($\text{AgNPs}/\text{ZnO-NR}$) reaching a sensitivity of $135.5\ \mu\text{A}\ \text{mM}^{-1}\ \text{cm}^{-2}$ along with a remarkably low LOD of $0.184\ \text{mM}$. A reduction in charge transfer resistance (R_{ct}) was observed in $\text{AgNP}/\text{ZnO-NRs}$ by EIS, leading to improved sensing performance through the enhanced electron transfer.⁶⁹

Ariyanta *et al.* immobilized a nanocomposite of NiO and MoS_2 on a screen-printed carbon electrode (SPCE), which was subsequently modified by electrochemically polymerizing poly(methyl orange) (MO) from a MO/PBS solution as a cholesterol-identifying agent. Calibration was done in Triton-X/PBS solutions, and pretreated milk and yogurt samples were electrochemically tested *via* CV and DPV. Negative potential at the n-type semiconductor MoS_2 increased the electron density in the conduction band, which in turn facilitated electron transfer to the O_2 molecule converting O_2 to O_2^- . The latter then reacted with H_2O to produce H_2O_2 and hydroxyl radicals, thus initiating cholesterol oxidation to oxysterol. The sensor exhibited a detection range spanning from $25.86\ \mu\text{M}$ to $0.39\ \text{mM}$ with LOD and LOQ values of $0.24\ \text{mg}\ \text{dL}^{-1}$ and $0.81\ \text{mg}\ \text{dL}^{-1}$ at a reasonably high sensitivity of $30.63\ \mu\text{A}\ \text{mM}^{-1}\ \text{cm}^{-2}$, respectively. The obtained cholesterol concentrations were in accordance with those indicated on the packaging, further validating the fabricated sensor electrode.⁷⁰

A gold-decorated nickel oxide ($\text{Au}@\text{NiO}$) on PPy nanocomposite was deposited onto glassy carbon electrode ($\text{Au}@\text{NiO}/\text{PPy-GCE}$) and studied in electrochemical cholesterol sensing *via* CV and DPV. The results revealed a linear detection range spanning from 1.0×10^{-5} to $1.0 \times 10^{-4}\ \text{M}$ at a sensitivity

of $7.6 \mu\text{A } \mu\text{M}^{-1} \text{ cm}^{-2}$ and an impressive LOD of $5.8 \times 10^{-7} \text{ M}$. Crucially, interference with various substances such as glucose, dopamine (DA), NaCl, uracil, uric acid, and ascorbic acid did not affect the determination accuracy. This high selectivity was attributed to the combination of exceptional electrocatalytic activity and the vast specific surface area.⁴¹

A cholesterol sensing electrode was developed specifically for the purpose of investigating a variety of food products, such as vanilla essence, vanilla-flavored biscuits, ice cream, and cake mix. First, TiO_2 was directly grown onto rGO through a hydrothermal process (rGO- TiO_2), then polypyrrole was deposited by *in situ* chemical oxidative polymerization. The constructed rGO- TiO_2 /PPy nanocomposite was applied onto GCE and its cholesterol sensing performance was studied by electrochemical measurements, including CV and CA. This electrode exhibited a remarkably low LOD and LOQ of $0.05 \mu\text{M}$ and $0.1 \mu\text{M}$, respectively.⁷¹

Bairagi and Verma fabricated a poly methyl orange (PMO) dendritic film/Cu/Ni/carbon nanofiber (CNF) electrode for non-enzymatic cholesterol sensing. Cu/Ni bimetal nanoparticles were deposited onto activated carbon fiber (ACF),^{73,74} which was transformed into a Cu/Ni/CNF nanocomposite in a CVD process. The ball-milled composite was cast into a polyvinylacetate (PVAc) metal-carbon-polymer composite film and a dendritic PMO nanofilm was grown through electrochemical polymerization of methyl orange. Here, PVAc served as a binder, and the composite is the active sensing element. The electrode was tested in electrochemical cholesterol sensing in Triton-X/PBS (pH = 7.0) and in human blood samples *via* CV, DPV, and EIS. Impedance spectra showed the diffusion-controlled cholesterol transport toward the electrode surface, and an electrochemical surface area of 3.784 cm^2 and an average catalytic rate constant of $195 \text{ mol}^{-1} \text{ L s}^{-1}$. The sensor demonstrated a linear response from a cholesterol concentration of 0.04 to 600 mg dL^{-1} at a sensitivity of $226.30 \mu\text{A mM}^{-1} \text{ cm}^{-2}$. The LOD and the LOQ were determined to be 0.00204 and $0.0062 \text{ mg dL}^{-1}$, respectively. Measurements on the clinical samples showed promise in further clinical testing.⁷²⁻⁷⁴

A composite combined glucose and cholesterol biosensor of Fe_3O_4 dispersed on silica-coated (SiO_2) CVD-grown MWNTs was reported. Non-enzymatic cholesterol sensing was performed by applying the material onto a GCE with Nafion binding and protective agent. The active material functioned as an electron transfer mediator resulting in an increased current response in cholesterol oxidation/reduction. The presence of Fe^{3+} and Fe^{2+} ions enabled the detection of H_2O_2 produced during cholesterol oxidation. The composite biosensor exhibited a linear detection range between $10 \mu\text{M}$ and 4 mM , an LOD of $5 \mu\text{M}$, and a rapid response time of 5 seconds. Furthermore, stability tests conducted over a 2 month-period demonstrated a stable response, and a 90% response retention after 15 consecutive measurements for both glucose and cholesterol.⁷⁵

Khaliq *et al.* developed an electrochemical cholesterol sensor based on a hybrid nanostructure composed of Cu_2O and TiO_2 nanotubes. The latter was synthesized from a pretreated Ti foil through a two-step anodization procedure followed by annealing,⁷⁸ while the subsequent Cu_2O nanoparticle deposition was done by cluster beam deposition (CBD). The cholesterol sensing

performance was verified by electroanalytical techniques (CV, EIS, CA *etc.*) in PBS/IPA, and the quantification based on amperometry as shown in Fig. 3a–c. A linear detection range (LDR) spanning from 24.4 to $622 \mu\text{M}$ was found with an LOD of $0.05 \mu\text{M}$ along with a high sensitivity of $6034 \mu\text{A mM}^{-1} \text{ cm}^{-2}$, and a fast response of 3 seconds. The copper's redox behavior was identified as the driving mechanism behind cholesterol sensing as illustrated in Fig. 3d. Cu_2O first adsorbs oxygen (O_2) and then generates O_2^- ions, which subsequently react with water and forming hydroxyl radicals (OH^\cdot). The latter initiates cholesterol auto-oxidation and ultimately leads to electron generation, which in turn provides an electrical signal connected to cholesterol concentration. Furthermore, a comprehensive clinical sample study was undertaken to assess the impact of interfering substances during the sensing process. The performance of the as-fabricated electrodes was comparable to that of certain commercial sensors, indicating the robustness and reliability of the developed system.⁷⁷

Polycrystalline ZnO and $\text{Zn}_2\text{In}_2\text{O}_5$ phases were deposited on a copper substrate and was utilized as a cholesterol biosensor. The electrode was fabricated onto a pre-treated copper substrate through the co-deposition of Zn and In from the aqueous solution of ZnCl_2 , KCl, InF_3 , and H_3BO_3 , followed by hydrothermal oxidation. Cholesterol sensing tests were conducted in PBS buffer employing CV and DPV. During positive potential sweep, oxygen adsorbed on the electrode is converted into ionic species by extracting electrons from the electrode. These mobile oxygen ions (O_2^-/O^-) participate in the reverse sweep, leading to the oxidation of cholesterol to cholestenone and the production of H_2O_2 as a byproduct. Subsequently, an electron is released due to H_2O_2 oxidation, and the transfer of this electron to the electrode reduces a metal ion back to its atomic form. The two-step process is outlined in Scheme 3. The electrode demonstrated a linear detection range for cholesterol concentrations between 0.5 mM and 9 mM , with a sensitivity of $81 \mu\text{A mM}^{-1} \text{ cm}^{-2}$ and a fast response of 1 second.⁷⁹

Willyam *et al.* constructed a non-enzymatic cholesterol biosensor by combining magnetite (Fe_3O_4) nanoparticles (MNP) and β -cyclodextrin (CD). The composite was synthesized by the co-precipitation of Fe^{2+} and Fe^{3+} in BCD (MNP/BCD) under alkaline conditions, and the resulting precipitate was then separated with an external magnet. The active material was immobilized on SPCE, and cholesterol detection was achieved by adding methylene blue (MB) to the electrode, as sensing mechanism based on the competition of inclusion complex formation between cholesterol and β -cyclodextrin (BCD), and methylene blue (MB), respectively. The electrochemical behavior of the fabricated electrode was studied by CV, DPV, and CA, and the sensor showed a wide linear response range from 0 to $150 \mu\text{M}$ along with a low LOD of $8.11 \mu\text{M}$, $4.77 \mu\text{M}$, and $2.88 \mu\text{M}$ for the anodic and the cathodic CV peaks, and for utilizing amperometry, respectively. The schematic procedure of cholesterol sensing using the developed sensor, the corresponding current responses obtained from CV under various conditions and the amperometric response during cholesterol sensing are shown in Fig. 4. The sensor also displayed excellent repeatability and recovery in cholesterol determination in



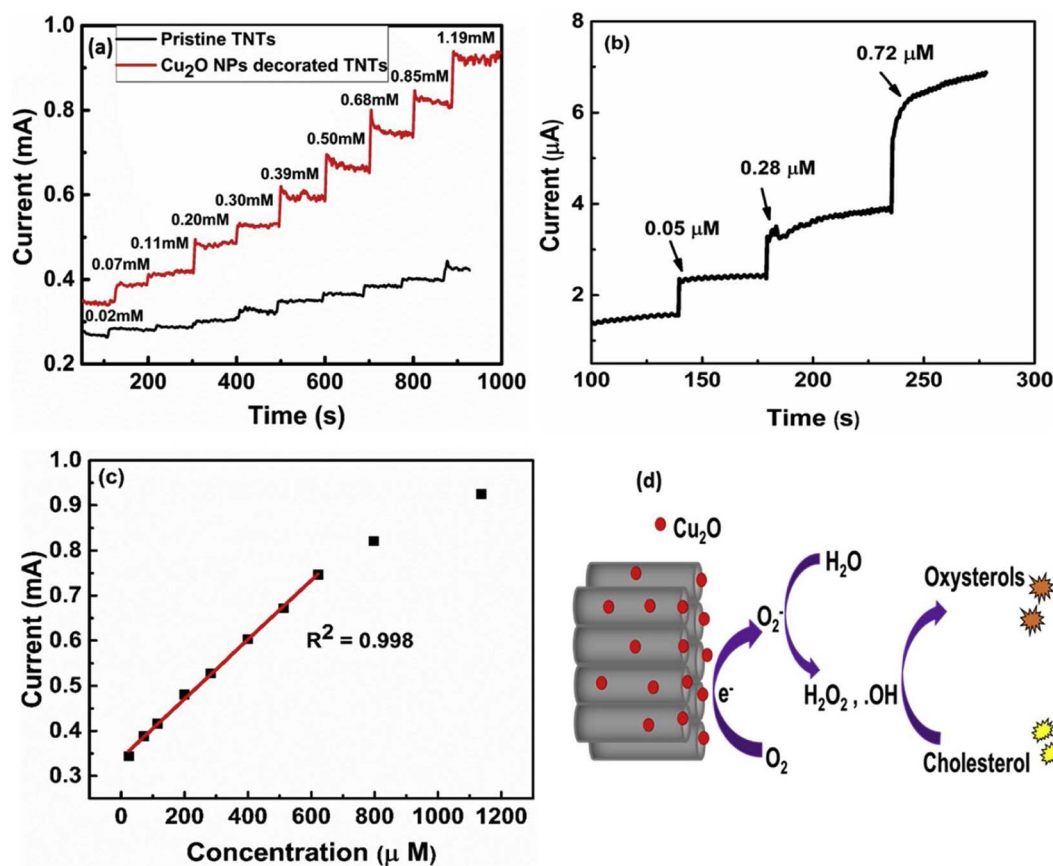
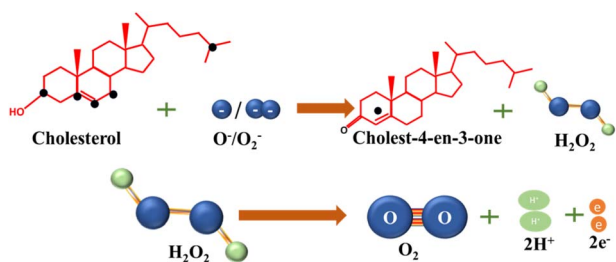


Fig. 3 Amperometric response of the pristine TNTs and Cu₂O decorated TNTs at -0.46 V in 0.1 M PBS at pH 7.0 upon successive addition of cholesterol in the (a) high, and (b) low concentration ranges. (c) The corresponding linear calibration curve at high cholesterol concentration, and (d) schematic of electrochemical oxidation of cholesterol at the surface of the Cu₂O NPs decorated TNTs electrode, reprinted with permission from ref. 77. Copyright 2020 Elsevier and Copyright Clearance Center (LN: 5833130132597).



Scheme 3 Reactions involved during non-enzymatic detection of cholesterol with the participation of oxygen ions.

50 mM PBS solution (pH = 7.4). It is worth noting, that further interference studies demonstrated that the presence of NaCl, CaCl₂, glycine, glucose, and ascorbic acid at concentrations ten times higher than cholesterol did not significantly affect the accuracy of cholesterol measurements.⁸⁰

A copper(II) oxide/(CuO)/PANI/murexide (Mu) composite biosensor was fabricated on glassy carbon electrode *via* electrooxidation and electrodeposition. The electrochemical performance of the sensor was determined by CV, EIS, and linear sweep voltammetry (LSV), and showed high sensitivity and high stability across a broad concentration range from 0.5

to 1 mM. The developed bioanalytical system was applied to determine cholesterol concentration in milk with high recovery rates and high selectivity, demonstrating its potential as a valuable tool in real-life cholesterol detection.¹⁴

A hybrid material comprising poly(ionic liquid) (PVIM) and cobalt polyoxometalate (Co₅POM) anchored on carbonaceous materials (MNCs) was utilized as electrochemical cholesterol sensor, and demonstrated an unprecedentedly low LOD of 1 fM (1×10^{-15} M). The schematics of cholesterol detection, the structural details and some important electrochemical results are shown in Fig. 5. It showed a rapid response time of 5 seconds along with a wide dynamic detection range between 1 fM and 5 mM. The latter consists of two linear regions, one ranging from 1 fM to 200 nM and another spanning from 0.5 μM to 5 mM with a sensitivity of $210 \mu\text{A} \mu\text{M}^{-1} \text{cm}^{-2}$ and $64 \mu\text{A} \mu\text{M}^{-1} \text{cm}^{-2}$, respectively. Importantly, the impressive sensor performance remained unaffected in the presence of interfering substances. To validate its practical utility, the sensor was tested in a human blood serum sample at a physiological pH within a concentration range relevant to human biology. Furthermore, a flexible version of this sensor was also developed by coating PVIM-Co₅POM/MNC onto a filter paper retaining the original high sensitivity and broad detection range, making it a versatile tool for a wide range of applications.⁸¹

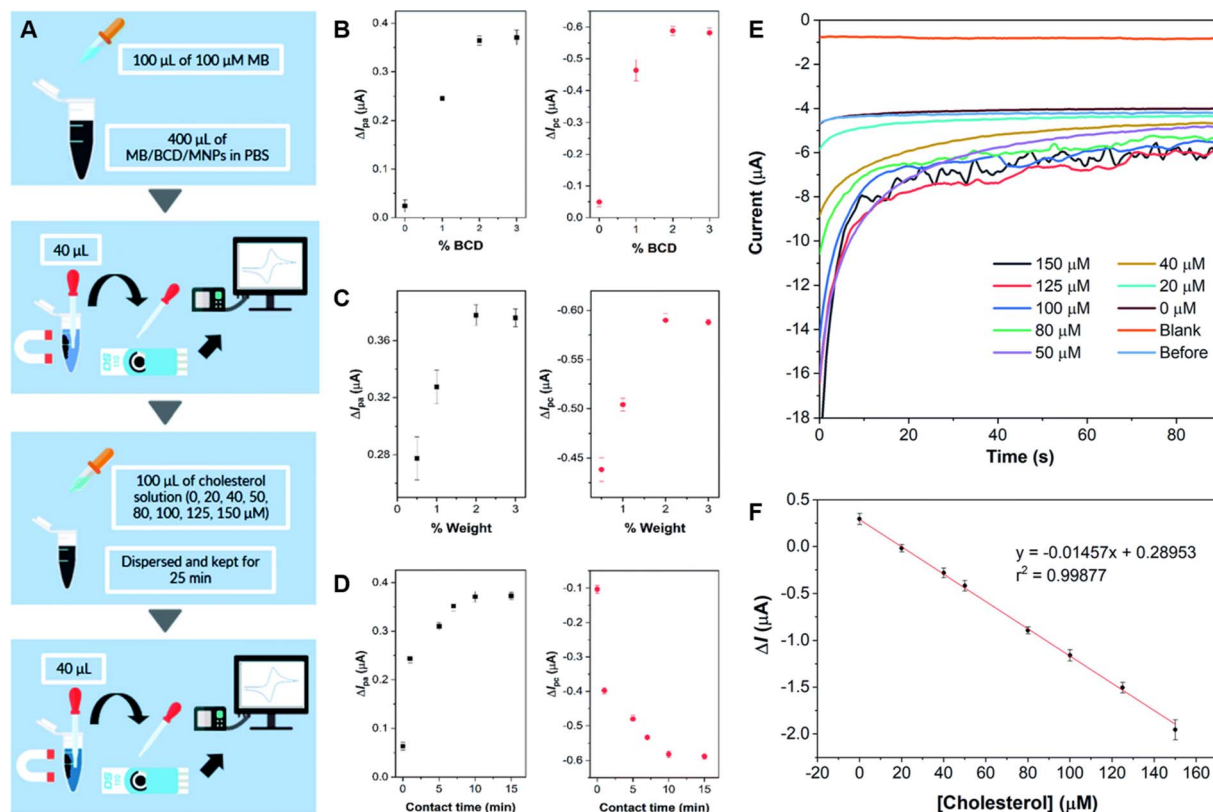


Fig. 4 Schematics of the β -cyclodextrin/ Fe_3O_4 nanocomposite cholesterol sensor fabrication process (A), plots of methylene blue current differences before and after the addition of 100 μM cholesterol in 50 mM PBS (pH 7.4) by cyclic voltammetry on screen-printed carbon electrode using nanocomposites synthesized with different BCD contents in BCD/MNPs (2% w/w) (B), different amounts of 3%-BCD/MNPs (C), and different contact times (D). Amperograms obtained for blank solution (50 mM PBS, pH 7.4), before and after cholesterol addition at concentrations between 0 and 150 μM at a potential of -0.43 V for 90 s under optimum conditions (E), and the corresponding calibration curve of the developed cholesterol sensor (F). Reprinted with permission from ref. 80. Copyright 2020 Royal Society of Chemistry (LN: 1507238-1).

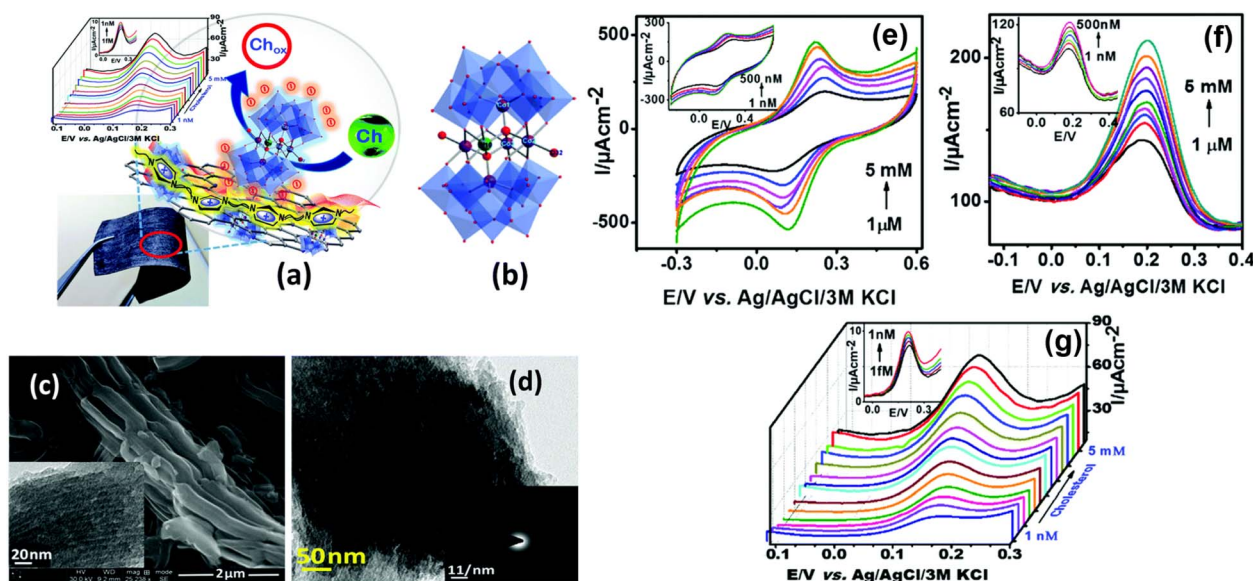


Fig. 5 Schematic representation of cholesterol detection with the PVIM- Co_5POM /MNC composite on a flexible paper electrode (a). Single crystal X-ray structure of $\text{Na}_{12}[\text{WCo}_3(\text{H}_2\text{O})_2(\text{CoW}_9\text{O}_{34})_2]$; Na and H atoms are omitted for clarity (b). FE-SEM image of the MNC (inset: TEM image of the MNC) (c), and TEM image of the PVIM- Co_5POM /MNC composite (inset: SAED pattern of the same structure) (d). CV (e) and DPV (f) of the PVIM- Co_5POM /MNC-600 modified graphite electrode at various concentrations of blood serum in 0.1 M phosphate buffer (pH 7.4) and 1 mM $\text{K}_4[\text{Fe}(\text{CN})_6]$ electrolyte in the presence of 500 μM cholesterol. DPV of the PVIM- Co_5POM /MNC-600 modified flexible paper electrode at various concentrations of cholesterol in 0.1 M NaOH and 1 mM $\text{K}_4[\text{Fe}(\text{CN})_6]$ electrolyte at a scan rate of 10 mV s^{-1} ; CE: Pt wire, RE: Ag/AgCl/3 M KCl (g). Reprinted with permission from ref. 81. Copyright Royal Society of Chemistry (LN: 1507238-1).



Rabbani *et al.* deposited mesoporous NiCo₂S₄ nanoflakes onto nickel foam, and the fabricated electrode demonstrated a high sensitivity of 8623.6 $\mu\text{A mM}^{-1} \text{cm}^{-2}$ across a broad linear range from 0.01 to 0.25 mM along with a considerable low LOD 0.01 μM . The NiCo₂S₄ structure also possesses robust thermal stability and consistent performance over the extended period of 8 weeks. Furthermore, the sensor exhibited high recovery in measurements on real-life blood samples.⁸²

Challenges and future perspectives

In the pursuit of direct cholesterol sensing through electro-oxidation, researchers have to be aware of the oxidation potential of cholesterol, which is typically $\approx 0.4 \text{ V}$ (*vs.* Ag/AgCl/sat'd KCl). It is crucial to take any potential variations due to the choice of reference electrodes and the specific oxidation sites within cholesterol into account. Furthermore, in order to have a mechanistic understanding of our system the redox properties of any functional moieties have to be known. It needs be ensured that these moieties are capable of either directly oxidizing cholesterol or participating in further redox reactions. Depending on the actual sensing layer, this could be reaction with byproducts H₂O₂, O₂, *etc.*, or with intermediates such as oxygen, hydroxide ions, and radicals. Moreover, these moieties should serve as effective current capturers or sensors throughout the redox reaction process. Conducting *in situ* and *operando* analysis on intermediates and products during cholesterol oxidation, along with the investigation on elemental interactions between the electrode components and cholesterol, would significantly advance the field.

Cost-effective, stable, durable, and reproducible metal oxide-based electrodes with high sensitivity can compete with existing enzyme-assisted sensor systems. Tailoring the characteristics of metal-based systems to possess inherent cholesterol-oxidizing capabilities eliminates the need for additional activating agents, and ensures the development of an easily monitorable current signals at the electrode. Additional materials can also be incorporated into the sensor chips either as filters to eliminate interfering components or as supporting reagents to achieve minimal or zero input current. Commercially viable sensor chips utilizing highly active metal(oxide)-based working electrodes with cost-effective counter and reference electrodes on suitable substrate would give further impetus to the continuously developing field of electrochemical cholesterol sensing.

Summary

Non-enzymatic electrochemical cholesterol sensing techniques offer numerous advantages over its enzymatic counterparts, such as being enzyme-free, having a straightforward operating principle, maintaining high stability in various environments, and offering a wide array of synthesis strategies for electrode development. Among the materials explored for non-enzymatic sensors, oxide-based systems are particularly noteworthy due to their sensitivity, stability, and reproducibility. A wide range of material families were already studied, primarily based on the

oxides of Mn, Fe, Ni, Cu, Zn, Mo, Ti, In, and Si. While many research focused on lab-scale experiments, a few studies already incorporated clinical sample analysis as well. The advancement of chip-based non-enzymatic metal oxide-based systems for clinical applications necessitates extensive research into mechanistic aspects of material and electrode development strategies, cholesterol sensing methodologies, and protocols for testing physiological samples. This review aims to provide insights to researchers in academia and industry, with the hope that commercially available cholesterol sensor system will be realized soon.

Abbreviations

PANI	Polyaniline
PPy	Polypyrrole
MWCNT	Multiwalled carbon nanotubes
rGO	Reduced graphene oxide
SPCE	Screen-printed carbon electrode
β -CD	β -Cyclodextrin
ChOx	Cholesterol oxidase
ChE	Cholesterol esterase
HRP	Horseradish peroxidase
PBS	Phosphate buffer saline
CV	Cyclic voltammetry
CA	Chronoamperometry
LSV	Linear sweep voltammetry
EIS	Electrochemical impedance spectroscopy
DPV	Differential pulse voltammetry
LOD	Limit of detection
PGE	Pencil graphite electrode
PVDF	Polyvinylidene chloride (PVDF)
NMP	N-Methyl-2-pyrrolidene
SCE	Saturated calomel electrode
LOQ	Limit of quantification
NR	Nanorods
NP	Nanoparticles
R_{ct}	Charge transfer resistance
MO	Methyl orange
GCE	Glassy carbon electrode
DA	Dopamine
PMO	Poly methyl orange
CNF	Carbon nanofiber
CVD	Chemical vapour deposition
ACF	Activated carbon fibre
PVC	Poly vinyl chloride
TEOS	Tetraethoxysilane
TNT	TiO ₂ nanotubes
CBD	Cluster beam deposition
MNP	Magnetic nanoparticles
Mu	Murexide (Mu)

Data availability

No primary research results, software or code have been included and no new data were generated or analysed as part of this review.



Author contributions

The manuscript was written through contributions of all authors. All authors have given approval to the final version of the manuscript.

Conflicts of interest

The authors declare no competing financial interest.

Appendix: vocabulary

Cholesterol, a compound present in the foods you consume that serves vital functions in human body. Enzymes, biological catalyst accelerate chemical reactions without being consumed or permanently altered in the process. Biosensors is a specialized sensor that utilizes biological molecules or components to detect and measure specific biological or chemical substances, often used for applications in healthcare, environmental monitoring, and biotechnology. Analyte, a specific substance or chemical compound that a sensor or analytical instrument is designed to detect and measure its concentration within a given sample or environment. Electrode, in sensing is a specialized conductor used to detect, measure, or record electrical signals or changes in electrical properties resulting from interactions with the surrounding environment, which is essential for various sensing applications such as electrochemical, pH, or biosensors. The limit of detection (LOD) is the lowest concentration or amount of a substance that can be reliably detected, but not necessarily quantified with a particular analytical method or sensor. The limit of quantification (LOQ) is the lowest concentration or amount of a substance that can be accurately and precisely measured and quantified with a particular analytical method or instrument. According to IUPAC, LOD and LOQ is 3 and 10 times higher than the standard deviation of the blank measurement (background noise), respectively. Sensitivity, in the context of sensing refers to the ability of a sensor to reliably detect even small changes in the quantity or concentration of an analyte in a sample. It is, by definition, the slope of the calibration curve.

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References

- 1 L. H. Li, E. P. Dutkiewicz, Y. C. Huang, H. B. Zhou and C. C. Hsu, Analytical methods for cholesterol quantification, *J. Food Drug Anal.*, 2019, 27(2), 375–386, DOI: [10.1016/j.jfda.2018.09.001](https://doi.org/10.1016/j.jfda.2018.09.001).
- 2 C. Wu, D. R. Ifa, N. E. Manicke and R. G. Cooks, Rapid, Direct Analysis of Cholesterol by Charge Labelling in Reactive Desorption Electrospray Ionization, *Anal. Chem.*, 2009, 81, 7618–7624, DOI: [10.1021/ac901003u](https://doi.org/10.1021/ac901003u).
- 3 H. Y. Hsieh, L. H. Li, R. Y. Hsu, W. F. Kao, Y. C. Huang and C. C. Hsu, Quantification of Endogenous Cholesterol in Human Serum on Paper Using Direct Analysis in Real Time Mass Spectrometry, *Anal. Chem.*, 2017, 89(11), 6146–6152, DOI: [10.1021/acs.analchem.7b00943](https://doi.org/10.1021/acs.analchem.7b00943).
- 4 L. Xu, M. Kliman, J. G. Forsythe, K. Zeljka, A. B. Hmelo, N. A. Porter and J. A. McLean, Profiling and Imaging Ion Mobility-Mass Spectrometry Analysis of Cholesterol and 7-Dehydrocholesterol in Cells via Sputtered Silver MALDI, *J. Am. Soc. Mass Spectrom.*, 2016, 27(6), 924–933, DOI: [10.1007/s13361-015-1131-0](https://doi.org/10.1007/s13361-015-1131-0).
- 5 H. Hidaka, N. Hanyu, M. Sugano, K. Kawasaki, K. Yamauchi and T. Katsuyama, Analysis of human serum lipoprotein lipid composition using MALDI-TOF mass spectrometry, *Ann. Clin. Lab. Sci.*, 2007, 37(3), 213–221, <https://pubmed.ncbi.nlm.nih.gov/17709684/>.
- 6 J. Baranwal, B. Barse, G. Gatto, G. Broncova and A. Kumar, Electrochemical Sensors and Their Applications: A Review, *Chemosensors*, 2022, 10(9), 363, DOI: [10.3390/chemosensors10090363](https://doi.org/10.3390/chemosensors10090363).
- 7 H. M. Yadav, J. D. Park, H. C. Kang and J. J. Lee, Recent Development in Nanomaterial-Based Electrochemical Sensors for Cholesterol Detection, *Chemosensors*, 2021, 9(5), 98, DOI: [10.3390/chemosensors9050098](https://doi.org/10.3390/chemosensors9050098).
- 8 O. Domínguez-Renedo, A. M. Navarro-Cuñado and M. A. Alonso-Lomillo, Electrochemical Devices for Cholesterol Detection, *J. Pharm. Biomed. Anal.*, 2023, 224, 115195, DOI: [10.1016/j.jpba.2022.115195](https://doi.org/10.1016/j.jpba.2022.115195).
- 9 U. Saxena, M. Chakraborty and P. Goswami, Covalent Immobilization of Cholesterol Oxidase on Self-Assembled Gold Nanoparticles for Highly Sensitive Amperometric Detection of Cholesterol in Real Samples, *Biosens. Bioelectron.*, 2011, 26, 3037–3043, DOI: [10.1016/j.bios.2010.12.009](https://doi.org/10.1016/j.bios.2010.12.009).
- 10 R. Molaei, R. E. Sabzi, K. Farhadi, F. Kheiri and M. Forough, Amperometric Biosensor for Cholesterol Based on Novel Nanocomposite Array Gold Nanoparticles/Acetone-Extracted Propolis/Multiwall Carbon Nanotubes/Gold, *Micro Nano Lett.*, 2014, 9(2), 100–104, DOI: [10.1049/mnl.2013.0664](https://doi.org/10.1049/mnl.2013.0664).
- 11 R. Ahmad, N. Tripathy and Y. B. Hahn, Wide Linear-Range Detecting High Sensitivity Cholesterol Biosensors Based on Aspect-Ratio Controlled ZnO Nanorods Grown on Silver



- Electrodes, *Sens. Actuators, B*, 2012, **169**, 382–386, DOI: [10.1016/j.snb.2012.05.027](#).
- 12 N. Peñ, G. Ruiz, A. J. Reviejo and J. M. Pingarró, Graphite-Teflon Composite Bienzyme Electrodes for the Determination of Cholesterol in Reversed Micelles Application to Food Samples, *Anal. Chem.*, 2001, **73**, 1190–1195, DOI: [10.1021/ac0012243](#).
 - 13 K. V. Mokwebo, O. S. Oluwafemi and O. A. Arotiba, An Electrochemical Cholesterol Biosensor Based on A CdTe/CdSe/ZnSe Quantum Dots-Poly(Propylene Imine) Dendrimer Nanocomposite Immobilisation Layer, *Sensors*, 2018, **18**(10), 3368, DOI: [10.3390/S18103368](#).
 - 14 C. B. A. Hassine, H. Kahri and H. Barhoumi, A Novel Impedimetric Sensor Based on CuO(NPs)/Polyaniline/Murexide Composite for Cholesterol Detection, *J. Iran. Chem. Soc.*, 2022, **19**(5), 1651–1659, DOI: [10.1007/s13738-021-02404-8](#).
 - 15 V. Gautam, K. P. Singh and V. L. Yadav, Polyaniline/MWCNTs/Starch Modified Carbon Paste Electrode for Non-enzymatic Detection of Cholesterol: Application to Real Sample (Cow Milk), *Anal. Bioanal. Chem.*, 2018, **410**, 2173–2181, DOI: [10.1007/s00216-018-0880-6](#).
 - 16 N. Thakur, M. Kumar, S. Das Adhikary, D. Mandal and T. C. Nagaiah, PVIM-Co₅POM/MNC Composite as a Flexible Electrode for the Ultrasensitive and Highly Selective Non-enzymatic Electrochemical Detection of Cholesterol, *Chem. Commun.*, 2019, **55**(34), 5021–5024, DOI: [10.1039/C9CC01534E](#).
 - 17 Y. Zare and K. Y. Rhee, Effect of Contact Resistance on the Electrical Conductivity of Polymer Graphene Nanocomposites to Optimize the Biosensors Detecting Breast Cancer Cells, *Sci. Rep.*, 2022, **12**, 5406, DOI: [10.1038/s41598-022-09398-0](#).
 - 18 Š. Tumová, R. Malečková, L. Kubáč, J. Akerman, V. Enev, L. Kalina, E. Vojtková, M. Pesková, J. Viteček, M. Vala and M. Weiter, Novel Highly Stable Conductive Polymer Composite PEDOT:DBSA for Bioelectronic Applications, *Polym. J.*, 2023, **55**(9), 983–995, DOI: [10.1038/s41428-023-00784-7](#).
 - 19 P. Tan, H. Wang, F. Xiao, X. Lu, W. Shang, X. Deng, H. Song, Z. Xu, J. Cao, T. Gan, B. Wang and X. Zhou, Solution-Processable, Soft, Self-Adhesive, and Conductive Polymer Composites for Soft Electronics, *Nat. Commun.*, 2022, **13**, 358, DOI: [10.1038/s41467-022-28027-y](#).
 - 20 S. M. Monavari, F. Marsusi, N. Memarian and M. Qasemnazhand, Carbon Nanotubes and Nanobelts as Potential Materials for Biosensor, *Sci. Rep.*, 2023, **13**, 3118, DOI: [10.1038/s41598-023-29862-9](#).
 - 21 X. Liang, N. Li, R. Zhang, P. Yin, C. Zhang, N. Yang, K. Liang and B. Kong, Carbon-Based SERS Biosensor: From Substrate Design to Sensing and Bioapplication, *NPG Asia Mater.*, 2021, **13**, 8, DOI: [10.1038/s41427-020-00278-5](#).
 - 22 S. Liu and X. Guo, Carbon Nanomaterials Field-Effect-Transistor-Based Biosensors, *NPG Asia Mater.*, 2012, **4**, e23, DOI: [10.1038/am.2012.42](#).
 - 23 J. Suriyaprakash, K. Bala, L. Shan, L. Wu and N. Gupta, Molecular Engineered Carbon-Based Sensor for Ultrafast and Specific Detection of Neurotransmitters, *ACS Appl. Mater. Interfaces*, 2021, **13**(51), 51, DOI: [10.1021/acsami.1c18137](#).
 - 24 H. Altug, S. H. Oh, S. A. Maier and J. Homola, Advances and Applications of Nanophotonic Biosensors, *Nat. Nanotechnol.*, 2022, **17**, 5–16, DOI: [10.1038/s41565-021-01045-5](#).
 - 25 J. N. Anker, W. P. Hall, O. Lyandres, N. C. Shah, J. Zhao and R. P. Van Duyne, Biosensing with Plasmonic Nanosensors, *Nat. Mater.*, 2008, **7**(6), 442–453, DOI: [10.1038/nmat2162](#).
 - 26 N. R. Jo and Y. B. Shin, Enhancing Biosensing Sensitivity of Metal Nanostructures through Site-Selective Binding, *Sci. Rep.*, 2020, **10**, 1024, DOI: [10.1038/s41598-020-57791-4](#).
 - 27 X. Ma, S. He, B. Qiu, F. Luo, L. Guo and Z. Lin, Noble Metal Nanoparticle-Based Multicolor Immunoassays: An Approach toward Visual Quantification of the Analytes with the Naked Eye, *ACS Sens.*, 2019, **4**(4), 782–791, DOI: [10.1021/acssensors.9b00438](#).
 - 28 L. Xiao, L. Wei, Y. He and E. S. Yeung, Single Molecule Biosensing Using Color Coded Plasmon Resonant Metal Nanoparticles, *Anal. Chem.*, 2010, **82**(14), 6308–6314, DOI: [10.1021/ac101018v](#).
 - 29 Y. Gao, Y. Zhou and R. Chandrawati, Metal and Metal Oxide Nanoparticles to Enhance the Performance of Enzyme-Linked Immunosorbent Assay (ELISA), *ACS Appl. Nano Mater.*, 2020, **3**(1), 1–21, DOI: [10.1021/acsanm.9b02003](#).
 - 30 C. Li, Y. Wang, H. Jiang and X. Wang, Biosensors Based on Advanced Sulfur-Containing Nanomaterials, *Sensors*, 2020, **20**(12), 3488, DOI: [10.3390/s20123488](#).
 - 31 K. J. Babu, T. Raj Kumar, D. J. Yoo, S.-M. Phang and G. Gnana Kumar, Electrodeposited Nickel Cobalt Sulfide Flowerlike Architectures on Disposable Cellulose Filter Paper for Enzyme-Free Glucose Sensor Applications, *ACS Sustainable Chem. Eng.*, 2018, **6**(12), 16982–16989, DOI: [10.1021/acssuschemeng.8b04340](#).
 - 32 S. Hashmi, M. Singh, P. Weerathunge, E. L. H. Mayes, P. D. Mariathomas, S. N. Prasad, R. Ramanathan and V. Bansal, Cobalt Sulfide Nanosheets as Peroxidase Mimics for Colorimetric Detection of L-Cysteine, *ACS Appl. Nano Mater.*, 2021, **4**(12), 13352–13362, DOI: [10.1021/acsanm.1c02851](#).
 - 33 P. R. Solanki, A. Kaushik, V. V. Agrawal and B. D. Malhotra, Nanostructured metal oxide-based biosensors, *NPG Asia Mater.*, 2011, **3**, 17–24, DOI: [10.1038/asiamat.2010.137](#).
 - 34 H. Abunahla, B. Mohammad, A. Alazzam, M. A. Jaoude, M. Al-Qutayri, S. Abdul Hadi and S. F. Al-Sarawi, MOM-Sense: Metal-Oxide-Metal Elementary Glucose Sensor, *Sci. Rep.*, 2019, **9**, 5524, DOI: [10.1038/s41598-019-41892-w](#).
 - 35 L. Liu, Y. Wang, Y. Liu, S. Wang, T. Li, S. Feng, S. Qin and T. Zhang, Heteronanostructural metal oxide-based gas microsensors, *Microsyst. Nanoeng.*, 2022, **8**, 85, DOI: [10.1038/s41378-022-00410-1](#).
 - 36 Y. Yoon, P. L. Truong, D. Lee and S. H. Ko, Metal-Oxide Nanomaterials Synthesis and Applications in Flexible and Wearable Sensors, *ACS Nanosci. Au*, 2022, **2**(2), 64–92, DOI: [10.1021/acsnanosciencenau.1c00029](#).



- 37 N. Arunadevi, Metal Nanocomposites for Advanced Futuristic Biosensing Applications, *Mater. Lett.*, 2022, **309**, 131320, DOI: [10.1016/j.matlet.2021.131320](https://doi.org/10.1016/j.matlet.2021.131320).
- 38 V. S. Lakshmi, A. Sharma, P. R. Solanki and D. K. Avasthi, Mesoporous Polyaniline Nanofiber Decorated Graphene Micro-Flowers for Enzyme-Less Cholesterol Biosensors, *Nanotechnology*, 2016, **27**(34), 345101, DOI: [10.1088/0957-4484/27/34/345101](https://doi.org/10.1088/0957-4484/27/34/345101).
- 39 R. Batool, M. Asim Akhtar, A. Hayat, D. Han, L. Niu, M. A. Ahmad and M. H. Nawaz, A Nanocomposite Prepared from Magnetite Nanoparticles, Polyaniline and Carboxy-Modified Graphene Oxide for Non-Enzymatic Sensing of Glucose, *Microchim. Acta*, 2019, **186**(5), 267, DOI: [10.1007/s00604-019-3364-2](https://doi.org/10.1007/s00604-019-3364-2).
- 40 K. B. Akshaya, A. Varghese, M. Nidhin and L. George, Amorphous Ru-Pi Nanoclusters Coated on Polypyrrole Modified Carbon Fiber Paper for Non-Enzymatic Electrochemical Determination of Cholesterol, *J. Electrochem. Soc.*, 2019, **166**(12), B3272–B3282, DOI: [10.1149/2.113191jes](https://doi.org/10.1149/2.113191jes).
- 41 A. Nagarajan, V. Sethuraman, T. M. Sridhar and R. Sasikumar, Development of Au@NiO Decorated Polypyrrole Composite for Non-Enzymatic Electrochemical Sensing of Cholesterol, *J. Ind. Eng. Chem.*, 2023, **120**, 460–466, DOI: [10.1016/j.jiec.2022.12.054](https://doi.org/10.1016/j.jiec.2022.12.054).
- 42 P. Thiya, R. Ramya and J. Wilson, Poly(3,4-Ethylenedioxythiophene)/Taurine Biocomposite on Screen Printed Electrode: Non-Enzymatic Cholesterol Biosensor, *Microchem. J.*, 2020, **157**, 105037, DOI: [10.1016/j.microc.2020.105037](https://doi.org/10.1016/j.microc.2020.105037).
- 43 N. Agnihotri, A. D. Chowdhury and A. De, Non-Enzymatic Electrochemical Detection of Cholesterol Using β -Cyclodextrin Functionalized Graphene, *Biosens. Bioelectron.*, 2015, **63**, 212–217, DOI: [10.1016/j.bios.2014.07.037](https://doi.org/10.1016/j.bios.2014.07.037).
- 44 S. Alexander, P. Baraneedharan, S. Balasubrahmanyam and S. Ramaprabhu, Modified Graphene Based Molecular Imprinted Polymer for Electrochemical Non-Enzymatic Cholesterol Biosensor, *Eur. Polym. J.*, 2017, **86**, 106–116, DOI: [10.1016/j.eurpolymj.2016.11.024](https://doi.org/10.1016/j.eurpolymj.2016.11.024).
- 45 J. Yang, H. Lee, M. Cho, J. Nam and Y. Lee, Nonenzymatic Cholesterol Sensor Based on Spontaneous Deposition of Platinum Nanoparticles on Layer-by-Layer Assembled CNT Thin Film, *Sens. Actuators, B*, 2012, **171**–172, 374–379, DOI: [10.1016/j.snb.2012.04.070](https://doi.org/10.1016/j.snb.2012.04.070).
- 46 M. Saha and S. Das, Fabrication of a Nonenzymatic Cholesterol Biosensor Using Carbon Nanotubes from Coconut Oil, *J. Polym. Environ.*, 2015, **23**(3), 374–380, DOI: [10.1007/s10924-015-0690-4](https://doi.org/10.1007/s10924-015-0690-4).
- 47 J. Ji, Z. Zhou, X. Zhao, J. Sun and X. Sun, Electrochemical Sensor Based on Molecularly Imprinted Film at Au Nanoparticles-Carbon Nanotubes Modified Electrode for Determination of Cholesterol, *Biosens. Bioelectron.*, 2015, **66**, 590–595, DOI: [10.1016/j.bios.2014.12.014](https://doi.org/10.1016/j.bios.2014.12.014).
- 48 M. A. H. Nawaz, M. Majdinasab, U. Latif, M. Nasir, G. Gokce, M. W. Anwar and A. Hayat, Development of a Disposable Electrochemical Sensor for Detection of Cholesterol Using Differential Pulse Voltammetry, *J. Pharm. Biomed. Anal.*, 2018, **159**, 398–405, DOI: [10.1016/j.jpba.2018.07.005](https://doi.org/10.1016/j.jpba.2018.07.005).
- 49 J. Wu, H. Liu, W. Chen, B. Ma and H. Ju, Device Integration of Electrochemical Biosensors, *Nat. Rev. Bioeng.*, 2023, **1**, 346–360, DOI: [10.1038/s44222-023-00032-w](https://doi.org/10.1038/s44222-023-00032-w).
- 50 V. Kumar, D. M. N. Kashyap, S. Hebbar, R. Swetha, S. Prasad, T. Kamala, S. S. Srikanta, P. R. Krishnaswamy and B. Navakanta, Aza-Heterocyclic Receptors for Direct Electron Transfer Hemoglobin Biosensor, *Sci. Rep.*, 2017, **7**, 42031, DOI: [10.1038/srep42031](https://doi.org/10.1038/srep42031).
- 51 H. A. Hussein, A. Hanora, S. M. Solymann and R. Y. Hassan, Designing and Fabrication of Electrochemical Nano-Biosensor for the Fast Detection of SARS-CoV-2-RNA, *Sci. Rep.*, 2023, **13**, 5139, DOI: [10.1038/s41598-023-32168-5](https://doi.org/10.1038/s41598-023-32168-5).
- 52 M. A. Komkova, E. E. Karyakina and A. A. Karyakin, Catalytically Synthesized Prussian Blue Nanoparticles Defeating Natural Enzyme Peroxidase, *J. Am. Chem. Soc.*, 2018, **140**(36), 11302–11307, DOI: [10.1021/jacs.8b05223](https://doi.org/10.1021/jacs.8b05223).
- 53 M. M. Rahman, X.-b. Li, J. Kim, B. O. Lim, A. J. S. Ahammad and J.-J. Lee, A Cholesterol Biosensor Based on a Bi-Enzyme Immobilized on Conducting Poly(Thionine) Film, *Sens. Actuators, B*, 2014, **202**, 536–542, DOI: [10.1016/j.snb.2014.05.114](https://doi.org/10.1016/j.snb.2014.05.114).
- 54 D. Koyuncu Zeybek, B. Zeybek, N. Özçiçek Pekmez, Ş. Pekyardımcı and E. Kılıç, Development of an Amperometric Enzyme Electrode Based on Poly(o-phenylenediamine) for the Determination of Total Cholesterol in Serum, *J. Braz. Chem. Soc.*, 2012, **23**(12), 2222–2231, DOI: [10.1590/S0103-50532012001200011](https://doi.org/10.1590/S0103-50532012001200011).
- 55 L. Charpentier and N. El Murr, Amperometric Determination of Cholesterol in Serum with Use of a Renewable Surface Peroxidase Electrode, *Anal. Chim. Acta*, 1995, **318**(1), 89–93, DOI: [10.1016/0003-2670\(95\)00311-8](https://doi.org/10.1016/0003-2670(95)00311-8).
- 56 M. Zhao, Y. Li, X. Ma, M. Xia and Y. Zhang, Adsorption of Cholesterol Oxidase and Entrapment of Horseradish Peroxidase in Metal-Organic Frameworks for the Colorimetric Biosensing of Cholesterol, *Talanta*, 2019, **200**, 293–299, DOI: [10.1016/j.talanta.2019.03.060](https://doi.org/10.1016/j.talanta.2019.03.060).
- 57 O. Demkiv, W. Nogala, N. Stasyuk, N. Grynychshyn, B. Vus and M. Gonchar, The Peroxidase-like Nanocomposites as Hydrogen Peroxide-Sensitive Elements in Cholesterol Oxidase-Based Biosensors for Cholesterol Assay, *J. Funct. Biomater.*, 2023, **14**(6), 315, DOI: [10.3390/jfb14060315/s1](https://doi.org/10.3390/jfb14060315/s1).
- 58 S. Sahu, S. S. Shera and R. M. Banik, Enhanced Reusability of Horseradish Peroxidase Immobilized onto Graphene Oxide/Magnetic Chitosan Beads for Cost-Effective Cholesterol Oxidase Assay, *Open Biotechnol. J.*, 2019, **13**(1), 93–104, DOI: [10.2174/1874070701913010093](https://doi.org/10.2174/1874070701913010093).
- 59 D. Weber, Z. Ni, D. Vetter, R. Hoffmann and M. Fedorova, Electrochemical Oxidation of Cholesterol: An Easy Way to Generate Numerous Oxysterols in Short Reaction Times, *Eur. J. Lipid Sci. Technol.*, 2016, **118**(2), 325–331, DOI: [10.1002/ejlt.201500101](https://doi.org/10.1002/ejlt.201500101).
- 60 J. Kowalski, J. Płoszyńska, A. Sobkowiak, J. W. Morzycki and A. Z. Wilczewska, Direct Electrochemical Acetoxylation of Cholesterol at the Allylic Position, *J. Electroanal. Chem.*, 2005, **585**(2), 275–280, DOI: [10.1016/j.jelechem.2005.09.003](https://doi.org/10.1016/j.jelechem.2005.09.003).



- 61 J. Kowalski, Z. Lotowski, J. W. Morzycki, J. Płoszyńska, A. Sobkowiak and A. Z. Wilczewska, Unusual Electrochemical Oxidation of Cholesterol, *Steroids*, 2008, 73(5), 543–548, DOI: [10.1016/j.steroids.2008.01.014](#).
- 62 L. Benešová, J. Klouda, E. Bláhová, K. Nesměrák, P. Kočovský, J. Nádvorníková, P. Barták, J. Skopalová and K. Schwarzová-Pecková, Non-enzymatic Electrochemical Determination of Cholesterol in Dairy Products on Boron-doped Diamond Electrode, *Food Chem.*, 2022, 393, 133278, DOI: [10.1016/j.foodchem.2022.133278](#).
- 63 Y. Y. Hosokawa, H. Hakamata, T. Murakami, S. Aoyagi, M. Kuroda, Y. Mimaki, A. Ito, S. Morosawa and F. Kusu, Electrochemical Oxidation of Cholesterol in Acetonitrile Leads to the Formation of Cholesta-4,6-dien-3-one, *Electrochim. Acta*, 2009, 54(26), 6412–6416, DOI: [10.1016/j.electacta.2009.06.005](#).
- 64 N. Punnakal, J. Raveendran, S. P. Vasu, B. G. Nair and T. G. S. Babu, Highly Sensitive and Wide-Range Non-Enzymatic Electrochemical Detection of Cholesterol Using Pencil Lead Electrodes, *J. Electrochem. Soc.*, 2021, 168(4), 047515, DOI: [10.1149/1945-7111/ABF8D8](#).
- 65 Y. J. Lee and J. Y. Park, Nonenzymatic Free-Cholesterol Detection via a Modified Highly Sensitive Macroporous Gold Electrode with Platinum Nanoparticles, *Biosens. Bioelectron.*, 2010, 26(4), 1353–1358, DOI: [10.1016/j.bios.2010.07.048](#).
- 66 R. Ji, L. Wang, G. Wang and X. Zhang, Synthesize Thickness Copper (I) Sulfide Nanoplates on Copper Rod and It's Application as Nonenzymatic Cholesterol Sensor, *Electrochim. Acta*, 2014, 130, 239–244, DOI: [10.1016/j.electacta.2014.02.155](#).
- 67 A. Rengaraj, Y. Haldorai and B. Cheol, Electrodeposition of Flower-like Nickel Oxide on CVD-Grown Graphene to Develop an Electrochemical Non-enzymatic Biosensor, *J. Mater. Chem. B*, 2015, 3(30), 6301–6309, DOI: [10.1039/C5TB00908A](#).
- 68 S. Rison, K. B. Akshaya, V. S. Bhat, G. Shanker, T. Maiyalagan, E. K. Joice, G. Hegde and A. Varghese, MnO₂ Nanoclusters Decorated on Graphene Modified Pencil Graphite Electrode for Non-Enzymatic Determination of Cholesterol, *Electroanalysis*, 2020, 32(10), 2128–2136, DOI: [10.1002/elan.202000049](#).
- 69 T. T. N. Anh, H. Lan, L. T. Tam, V-H. Pham and P. D. Tam, Highly Sensitive Nonenzymatic Cholesterol Sensor Based on Zinc Oxide Nanorods, *J. Electronic Mater.*, 2018, 47(11), 6701–6708, DOI: [10.1007/s11664-018-6568-z](#).
- 70 H. A. Ariyanta, T. A. Ivandini and Y. Yulizar, Poly(methyl orange)-modified NiO/MoS₂/SPCE for a Non-enzymatic Detection of Cholesterol, *FlatChem*, 2021, 29, 100285, DOI: [10.1016/j.flatc.2021.100285](#).
- 71 S. Deivanayaki, P. Jayamurugan, S. Ashokan, V. G. Krishnan, B. Yogeswari, M. Ubaidullah, B. Pandit, G. V. S. S. Sarma and H. K. Narsetti, Growth of Non-enzymatic Cholesterol Biosensor Using TiO₂ Decorated Graphene Oxide with Bare GCE and PPy-GCE, *J. Indian Chem. Soc.*, 2023, 100(3), 100906, DOI: [10.1016/j.jics.2023.100906](#).
- 72 P. K. Bairagi and N. Verma, Electrochemically Deposited Dendritic Poly(methyl orange) Nanofilm on Metal-Carbon-Polymer Nanocomposite: A Novel Non-enzymatic Electrochemical Biosensor for Cholesterol, *J. Electroanal. Chem.*, 2018, 814, 134–143, DOI: [10.1016/j.jelechem.2018.02.011](#).
- 73 M. Ashfaq, S. Khan and N. Verma, Synthesis of PVA-CAP-Based Biomaterial *in situ* Dispersed with Cu Nanoparticles and Carbon Micro-Nanofibers for Antibiotic Drug Delivery Applications, *Biochem. Eng. J.*, 2014, 90, 79–89, DOI: [10.1016/j.bej.2014.05.016](#).
- 74 S. Singh, M. Ashfaq, R. K. Singh, H. C. Joshi, A. Srivastava, A. Sharma and N. Verma, Preparation of Surfactant-Mediated Silver and Copper Nanoparticles Dispersed in Hierarchical Carbon Micro-Nanofibers for Antibacterial Applications, *New Biotechnol.*, 2013, 30(6), 656–665, DOI: [10.1016/j.nbt.2013.05.002](#).
- 75 T. T. Baby and S. Ramaprabhu, Non-enzymatic Glucose and Cholesterol Biosensors Based on Silica Coated Nano Iron Oxide Dispersed Multiwalled Carbon Nanotubes, *2011 International Conference on Nanoscience, Technology and Societal Implications, NSTSI11 2011*, DOI: [10.1109/NSTSI.2011.6111981](#).
- 76 S. Sharma, P. Joshi, S. Mehtab, G. H. Zaidi, K. Singhal and T. I. Siddiqi, Development of Non-enzymatic Cholesterol Electrochemical Sensor Based on Polyindole/Tungsten Carbide Nanocomposite, *J. Anal. Testing*, 2020, 4(1), 13–22, DOI: [10.1007/s41664-020-00117-0](#).
- 77 N. Khaliq, M. A. Rasheed, G. Cha, M. Khan, S. Karim, P. Schmuki and G. Ali, Development of Non-enzymatic Cholesterol Biosensor Based on TiO₂ Nanotubes Decorated with Cu₂O Nanoparticles, *Sens. Actuators, B*, 2020, 302, 127200, DOI: [10.1016/j.snb.2019.127200](#).
- 78 G. Ali, S. H. Yoo, J. M. Kum, Y. N. Kim and S. O. Cho, A Novel Route to Large-Scale and Robust Free-Standing TiO₂ Nanotube Membranes Based on N₂ Gas Blowing Combined with Methanol Wetting, *Nanotechnology*, 2011, 22(24), 245602, DOI: [10.1088/0957-4484/22/24/245602](#).
- 79 S. Khan, M. A. Rasheed, A. Shah, A. Mahmood, A. Waheed, S. Karim, M. Khan and G. Ali, Preparation of Oxidized Zn-In Nanostructures for Electrochemical Non-enzymatic Cholesterol Sensing, *Mater. Sci. Semicond. Process.*, 2021, 135, 106101, DOI: [10.1016/j.msssp.2021.106101](#).
- 80 S. J. Willyam, E. Saepudin and T. A. Ivandini, b-Cyclodextrin/Fe₃O₄ Nanocomposites for an Electrochemical Non-enzymatic Cholesterol Sensor, *Anal. Methods*, 2020, 12, 3454, DOI: [10.1039/D0AY00933D](#).
- 81 N. Thakur, M. Kumar, S. Das Adhikary, D. Mandal and T. C. Nagaiah, PVIM-Co₅POM/MNC Composite as a Flexible Electrode for the Ultrasensitive and Highly Selective Non-enzymatic Electrochemical Detection of Cholesterol, *Chem. Commun.*, 2019, 55(34), 5021–5024, DOI: [10.1039/C9CC01534E](#).
- 82 S. S. Rabbani, A. Nisar, A. Zafar, Y. Liu, H. Sun, S. Karim, S. Hussain, A. U. Shah, S. Z. Hussain, N. Mehboob, Y. Yu and M. Ahmad, Mesoporous NiCo₂S₄ Nanoflakes as an Efficient and Durable Electrocatalyst for Non-enzymatic Detection of Cholesterol, *Nanotechnology*, 2022, 33(37), 375502, DOI: [10.1088/1361-6528/ac75fb](#).

