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# Synthesis, DFT studies on a series of tunable quinoline derivatives†

Nagesh Dhanaji Chavan and Vijayaparthasarathi Vijayakumar \*

The synthesis, Density Functional Theory (DFT) calculations, and photo physical characteristics of a range of quinoline derivatives have been described in the present work. Initially, the innovative derivatives are synthesized through the cyclization of 2-amino-5-nitrobenzophenone with either acetyl acetone or ethyl acetoacetate, followed by reducing the nitro group to an amine. Subsequently, these compounds undergo an acid-amine cross-coupling reaction. The investigation shows the DFT and photo physical properties of these substances. It is noteworthy that compound **6z** exhibits the most remarkable Stokes shift among the fluorophores investigated. Furthermore, the research also provides insights into the electrophilicity index, Electronegativity, chemical potential, chemical hardness and softness properties. These properties are determined by utilizing Density Functional Theory (DFT) calculations and evaluating electron potential efficiency and using computational methods Time-Dependent Density Functional Theory (TD-DFT) to predict absorption spectra in molecules at the B3LYP/6-31G'(d,p) level/basis.

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## 1 Introduction

A range of naturally occurring compounds and chemically significant molecules contain a quinoline-amide moiety exhibiting diverse biological activities. Quinoline derivatives have been used in various biological applications including anti-bacterial,<sup>1</sup> antimalarial,<sup>2</sup> anticancer (specifically against breast cancer), antiproliferative, and anti-epidermal growth factor receptor (EGFR)<sup>3</sup> activities. They also have properties as anti-microbial,<sup>4</sup> anti-inflammatory,<sup>5</sup> antioxidant,<sup>6</sup> antitubercular,<sup>7</sup> antimalarial, antiplasmodial,<sup>8</sup> and antiprotozoal drugs.<sup>9</sup> In addition, quinoline aryl amides have been evaluated for their antidiabetic activity, specifically their inhibition of  $\alpha$ -glucosidase and  $\alpha$ -amylase enzymes,<sup>10</sup> as well as their antioxidant, antifungal, and antibacterial properties.<sup>11</sup> The development of an amide bond through the coupling of an amine and a carboxylic acid is a widely utilized chemical reaction in drug development.<sup>12</sup> This reaction is of considerable significance due to the abundance of carboxylic acids and amine derivatives facilitating the synthesis of novel medicinal compounds typically this reaction proceeds in the presence of an acid-activating reagent or catalyst.<sup>13</sup> Numerous reagents and protocols have been documented for this transformation including the use of

acyl chlorides and various *N*-acylation techniques such as mixed anhydride methods,<sup>14–16</sup> carbodiimides (with or without additives),<sup>17,18</sup> polymer-supported coupling agents,<sup>19</sup> native chemical ligations,<sup>20</sup> and more researchers have employed various reagents with suitable bases for this coupling process, including POCl<sub>3</sub>, DCC/DMAP, SOCl<sub>2</sub>, T<sub>3</sub>P, EDC.HCl, HATU, and other coupling agents.<sup>21,22</sup> Particularly the most favourable results were obtained using HATU/DIPEA, with minimal side effects.

Organic fluorophores with rapid response times enabling real-time event monitoring with high spatial resolution, have gained significant popularity in research.<sup>23</sup> In physiological research, the use of fluorescent dyes that are easy to functionalize and demonstrate efficacy in biological investigations is a critical consideration.<sup>24,25</sup> Fluorescent probes particularly those based on quinoline structures are crucial for bio-imaging investigations.<sup>26,27</sup> Fluorescence-based assays are key for studying DNA structure, hybridization and interactions with molecules, the dyes are used in DNA labelling for cell imaging, gel analysis, and PCR quantification shown in Fig. 1.<sup>28–34</sup> Quinoline moieties are important in fluorescent sensors for their rapid modification of water solubility, chemical stability and biocompatibility.<sup>35</sup> Quinoline constitutes the core structure

Department of Chemistry, School of Advanced Sciences, Vellore Institute of Technology, Vellore 632016, India. E-mail: chavannagesh605@gmail.com; vvijayakumar@vit.ac.in; kvpsvijayakumar@gmail.com

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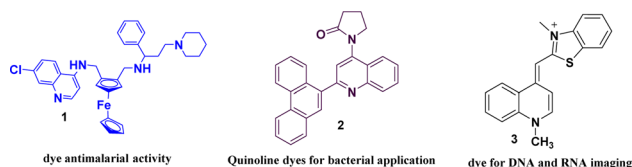


Fig. 1 Some examples of quinoline-based dyes.



of quinine an initial and well-characterized small-molecule fluorophore<sup>36,37</sup> and has been examined as a compelling molecular probe due to the nitrogen atom serving as a locus for tracking interactions with target analytes *via* alterations in fluorescence emission.<sup>38–40</sup> Quinoline derivatives have electron donation and intense  $\pi$ – $\pi^*$  transitions making them useful in selective fluorophores.<sup>41</sup> These derivatives have been observed to effectively detect hydrazine<sup>42</sup> effectively and have recently been used to detect various anions and cations.<sup>43–47</sup> Selecting an optimal fluorescence probe requires consideration of quantum yield, absorption/emission wavelengths, photo stability, chemical stability, and Stokes shift. New probes are continually being developed.<sup>48</sup> Fluorescent quinolines are effective in identifying bacteria and metal ions in chemo-sensing.<sup>49,50</sup> They exhibit intramolecular charge transfer (ICT) due to their electron-accepting properties and a distinct dipole moment.<sup>51,52</sup> The use of quinoline derivatives, such as 8-hydroxyquinolines, as chelators is common. These compounds are widely employed as  $\text{Zn}^{2+}$  sensors due to their excellent properties.<sup>53–56</sup> In 1968, 8-hydroxyquinoline was the first fluorescent zinc ion indicator used to detect zinc in human plasma.<sup>57</sup> When these compounds are excited by light, their electronic properties change, resulting in fluorescence with a significant shift in the emitted light's wavelength.<sup>58</sup> This substantial shift is advantageous in biological applications such as fluorescence microscopy as it helps minimize colour distortion and enhance resolution.<sup>59</sup> Compounds with large spectral shifts are also useful in Förster resonance energy transfer (FRET) microscopy, allowing direct excitation of acceptor molecules by neighbouring donor molecules.<sup>60</sup> These compounds have gained significant interest due to their potential applications in photo-electronic systems, as non-linear optical materials, light emitters, or in organic light-emitting diodes (OLED).<sup>61–63</sup> They are sensitive to external factors such as solvent polarity or viscosity and are commonly used as chemical detectors.<sup>64,65</sup> While Schiff bases are not commonly known for their fluorescent properties, they are often recognized for their antifungal, antibacterial, antiproliferative, anti-inflammatory, antimalarial, and antiviral properties.<sup>66–68</sup> Additionally, Schiff bases are employed as sensors for metal ions or pH levels.<sup>69,70</sup> Computational chemistry, driven by advancements in technology and algorithms, highlights the use of Density Functional Theory (DFT) for understanding electronic structures and reactivity in molecular systems and predicting molecular characteristics.<sup>71,72</sup> DFT uses reactivity descriptors such as electronegativity and hardness to better understand and predict molecular behaviour and attributes in chemistry and materials science.<sup>73,74</sup>

In this present work a new series of quinoline derivatives have been synthesized by cyclizing 2-amino-5-nitrobenzophenone with acetyl acetone or ethyl acetoacetate, followed by reducing the nitro group to an amine (Schemes 1 and 2) and then subjecting it to an acid-amine cross-coupling reaction. Using different types of carboxylic acid and coupling reagents such as HATU and DIPEA, 49 derivatives were synthesized, with yields ranging from 50–90%. All compounds were confirmed through various analytical techniques such as  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR, DEPT-135, HRMS, and IR, in addition to examining the structural

properties, chemical stability, and reactivity of target molecule.<sup>31</sup> compound recorded UV-vis and FL spectra and calculated stokes shift, molar extinction coefficient and quantum yield. All compounds were optimized by density functional theory (DFT) and investigated to understand the stability of the most active derivatives. We performed DFT calculations, frontier molecular orbital analysis (FMO), molecular electrostatic potential and TD DFT recorded theoretical absorbance UV-vis spectra.

## 2 Results and discussion

### 2.1. Chemistry

The synthesis of quinoline-amide derivatives **6a–6z** and **6aa–6aw** shown in Fig. 2 and 3 proceeds through several steps as outlined in Schemes 1 and 2. Initially, the reaction of (2-amino-5-nitrophenyl)(phenyl)methanone (**1**) with pentane-2,4-dione (**2**) and ethyl 3-oxobutanoate (**2a**) yields intermediates 1-(2-methyl-6-nitro-4-phenylquinolin-3-yl)ethan-1-one (**3**) and ethyl 2-methyl-6-nitro-4-phenylquinoline-3-carboxylate (**3a**). Subsequent reduction of the nitro group in compound **3** affords 1-(6-amino-2-methyl-4-phenylquinolin-3-yl)ethan-1-one (**4**). The target quinoline-amide derivatives (**6a–6z** and **6aa–6aw**) are then synthesized by reacting compound **4** and ethyl 6-amino-2-methyl-4-phenylquinoline-3-carboxylate (**4a**) with various carboxylic acids. This multi-step process involves condensation reactions, nitro group reduction, and coupling reactions to yield the desired products. The chemical structures of all compounds (**6a–6z** and **6aa–6aw**) were thoroughly analyzed using various spectroscopic techniques including  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR, DEPT-135, IR, and HRMS. Additionally, for compounds **6a**, **6b**, **6c**, **6d**, **6l**, **6m**, **6p**, and **6t**, 2D NMR techniques such as HSQC (Heteronuclear Single Quantum Coherence) and H, H–COSY (Correlation Spectroscopy) were employed to further elucidate their structures. The IR spectral analysis of compounds **6a–6z** and **6aa–6aw** revealed distinct features. Specifically, two bands in the range of 3583–3489  $\text{cm}^{-1}$  and 3394–3209  $\text{cm}^{-1}$  were identified, corresponding to the –NH stretching vibrations. Additionally, characteristic bands of carbonyl (C=O) groups were observed in the spectra, appearing in the range of 1710–1702  $\text{cm}^{-1}$  and 1691–1678  $\text{cm}^{-1}$ . Furthermore, the presence of C=C bonds was evidenced by bands in the region of 1555–1541  $\text{cm}^{-1}$  and 1537–1526  $\text{cm}^{-1}$ . The  $^1\text{H}$  NMR spectral analysis of compounds **6a–6w** and **6aa–6aw** revealed several key signals indicative of their structural features. Signals corresponding to the NH (amide) groups of the quinoline moiety were observed in the region of  $\delta$  11.15–10.33 ppm, highlighting their presence in the molecules. Additionally, the methyl group at the 2nd position of the quinoline exhibited a singlet signal in the range of  $\delta$  2.55–2.59 ppm. Notably, signals for the 5th, 7th, and 8th positions of the quinoline ring were detected at specific chemical shift regions: 7.85–7.80 ppm (d,  $J$  = 1.60 Hz), 8.01–7.97 ppm (dd,  $J$  = 2.00 and 8.80 Hz), and 7.98–7.95 ppm (d,  $J$  = 9.20 Hz), respectively. Moreover, in compounds **6n** and **6m**, the presence of trans protons was confirmed by the coupling constant ( $J$ ) value, with one proton appearing as a doublet in the region of 6.82–6.63 ppm and another proton displaying signals at 7.55–7.51 ppm with  $J$  = 15.6 Hz. This *trans* proton confirmation was further supported by 2D NMR H, H–COSY analysis. The



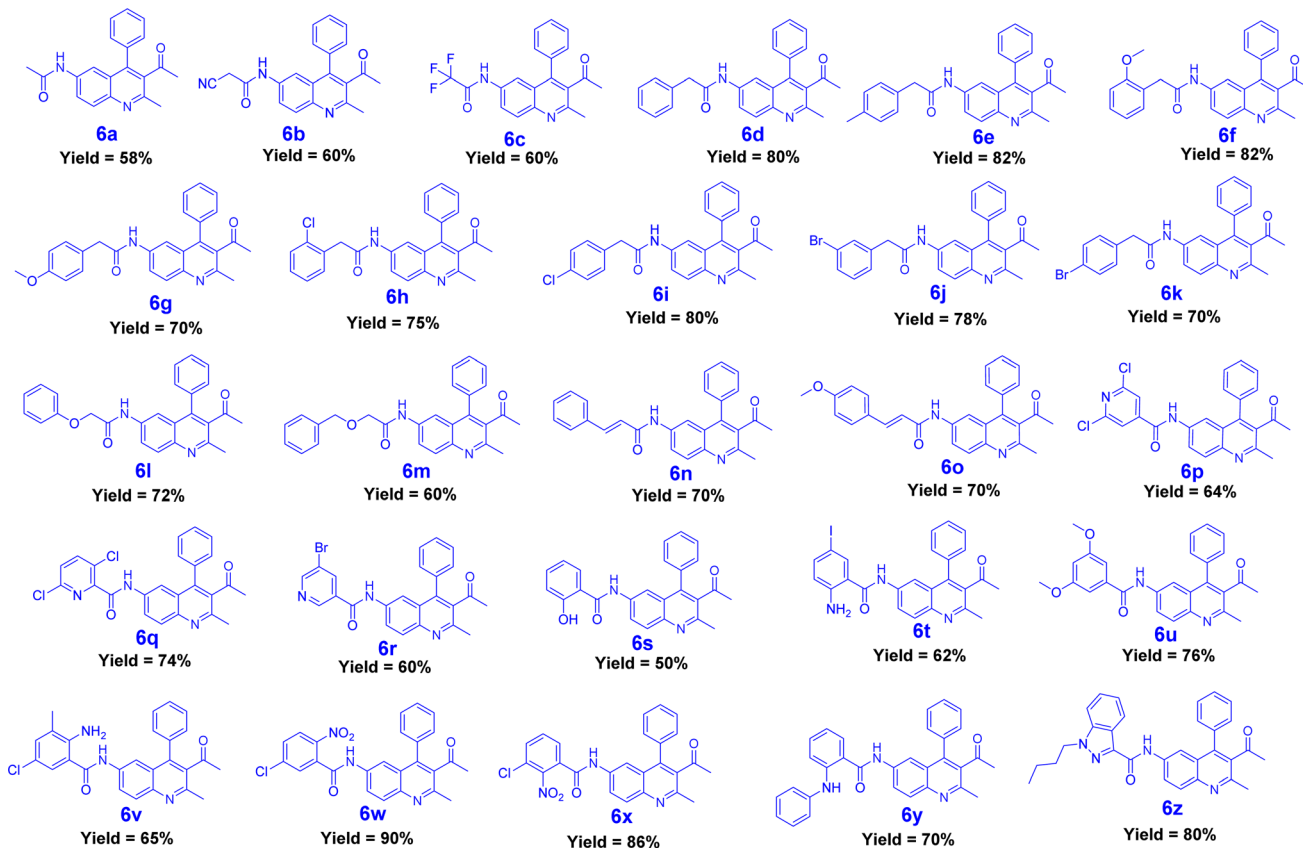


Fig. 2 Synthesized derivatives (6a–6z).

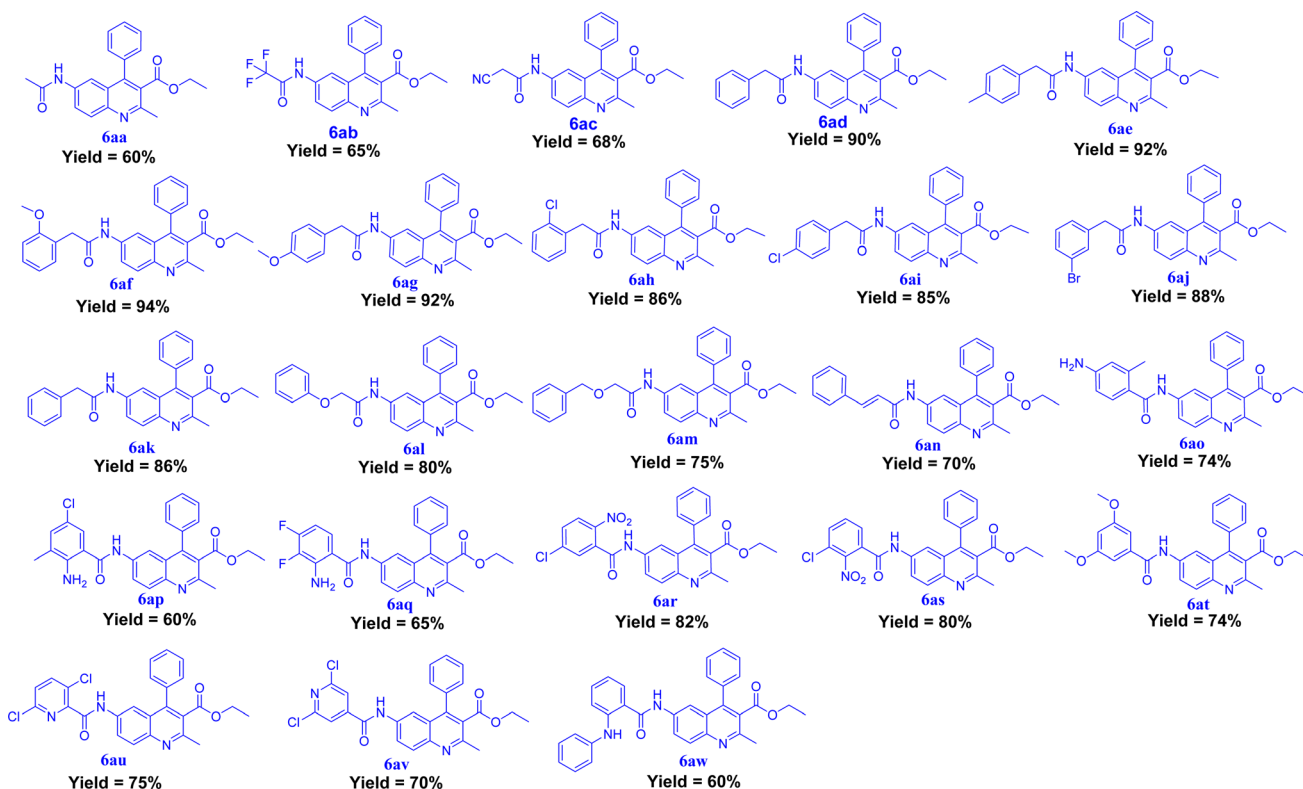
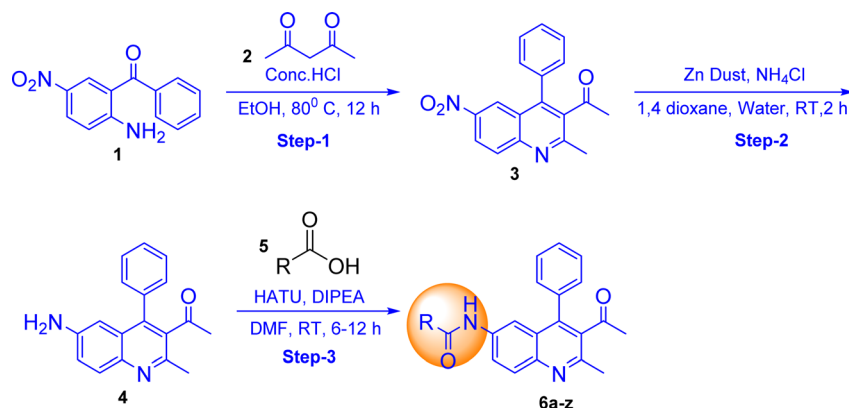
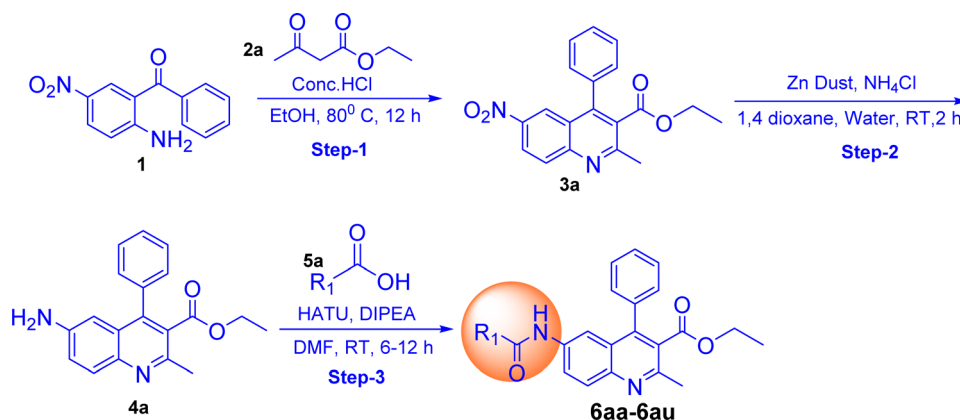


Fig. 3 Synthesized derivatives (6aa–6aw).



Scheme 1 (R = substituted group of carboxylic acid).

Scheme 2 ( $R_1$  = substituted group of carboxylic acid).

confirmation of quinoline amide carbon formation was further substantiated through  $^{13}\text{C}$  NMR studies. Specifically, the signal corresponding to the  $\text{H}_2\text{N}-\text{C}=\text{O}$  carbon in compounds **6a–6z** and **6aa–6aw** was detected in the range of 175–182 ppm, consistent with the expected chemical shift for this functional group. In DEPT-135 spectra, all carbon signals observed were found to be by the expected chemical shifts for the respective carbon environments within compounds **6a–6z** and **6aa–6aw**. Detailed reaction procedures and spectral data for the target compounds are provided in the Experimental and ESI† sections, respectively. Based on the yield of the compounds we selected two representative samples; one with a higher yield and the other one with a lower yield from out of the two schemes (Schemes 1 and 2) and did optimization in different bases and solvents. The optimization process for the reactions of compounds **6a** and **6af** was carried out by introducing variations in the solvents and bases utilized. It is worth mentioning that employing the base DIPEA and the solvent DMF resulted in favourable outcomes, as evidenced by the satisfactory yield of 58% achieved for compound **6a** and the high yield of 94% demonstrated by compound **6af**. In addition to these positive results, alternative bases such as triethylamine and pyridine, as well as solvents including chloroform, and dichloromethane, also exhibited promising results for compound **6af**. However, it is important to

note that these alternatives failed to yield satisfactory results for compound **6a**. The selection of the appropriate base and solvent is of utmost importance in determining the efficacy of the reaction, and in this case, it is evident that DIPEA and DMF emerged as the preferred combination for both compounds. However, THF did not support successful reactions when utilized with the aforementioned bases (Table 1).

## 2.2. Density functional theory (DFT)

Density Functional Theory (DFT) is a highly effective theoretical approach with many applications. It can be used to determine compound's kinetic and thermodynamic stability, perform structural calculations, gain insights into mechanisms, analyze molecular interactions, and evaluate the optical and electronic properties of atoms and molecules. All theoretical calculations were carried out for this study using the Gaussian 09 software program.<sup>75</sup> The compound's geometry was optimized using DFT at the B3LYP/6–31 G'(d,p) basic level set, and visualization was facilitated with the assistance of GaussView 5.0.<sup>76</sup>

## 2.3. Frontier molecular orbital (FMO)

Frontier Molecular Orbital (FMO) analysis plays an essential role in analyzing the chemical stability of molecules by studying the energy Levels of their frontier molecular orbitals. In this



**Table 1** Optimization of scheme compound **6a** and **6af** different base and solvent. (Qui-amine = quinoline amine, 4 = 1-(6-amino-2-methyl-4-phenylquinolin-3-yl)ethan-1-one, **4a** = ethyl 6-amino-2-methyl-4-phenylquinoline-3-carboxylate, acid = substituted carboxylic acid.)

Sr. no	Compounds	Base	HATU	Qui-amine	Acid	Solvents	Yield (%)
1	<b>6a</b>	DIPEA	1.5–2.0 equiv.	<b>4</b>	CH <sub>3</sub> -COOH	DMF	58
2	<b>6a</b>	DIPEA	1.5–2.0 equiv.	<b>4</b>	CH <sub>3</sub> -COOH	CHCl <sub>3</sub>	45
3	<b>6a</b>	DIPEA	1.5–2.0 equiv.	<b>4</b>	CH <sub>3</sub> -COOH	DCM	48
4	<b>6a</b>	DIPEA	1.5–2.0 equiv.	<b>4</b>	CH <sub>3</sub> -COOH	THF	—
5	<b>6a</b>	Et <sub>3</sub> N	1.5–2.0 equiv.	<b>4</b>	CH <sub>3</sub> -COOH	DMF	46
6	<b>6a</b>	Et <sub>3</sub> N	1.5–2.0 equiv.	<b>4</b>	CH <sub>3</sub> -COOH	CHCl <sub>3</sub>	40
7	<b>6a</b>	Et <sub>3</sub> N	1.5–2.0 equiv.	<b>4</b>	CH <sub>3</sub> -COOH	DCM	32
8	<b>6a</b>	Et <sub>3</sub> N	1.5–2.0 equiv.	<b>4</b>	CH <sub>3</sub> -COOH	THF	—
9	<b>6a</b>	Pyridine	1.5–2.0 equiv.	<b>4</b>	CH <sub>3</sub> -COOH	DMF	56
10	<b>6a</b>	Pyridine	1.5–2.0 equiv.	<b>4</b>	CH <sub>3</sub> -COOH	CHCl <sub>3</sub>	44
11	<b>6a</b>	Pyridine	1.5–2.0 equiv.	<b>4</b>	CH <sub>3</sub> -COOH	DCM	50
12	<b>6a</b>	Pyridine	1.5–2.0 equiv.	<b>4</b>	CH <sub>3</sub> -COOH	THF	—
13	<b>6af</b>	DIPEA	1.5–2.0 equiv.	<b>4a</b>	4-OMe-Ph-CH <sub>2</sub> -COOH	DMF	94
14	<b>6af</b>	DIPEA	1.5–2.0 equiv.	<b>4a</b>	4-OMe-Ph-CH <sub>2</sub> -COOH	CHCl <sub>3</sub>	75
15	<b>6af</b>	DIPEA	1.5–2.0 equiv.	<b>4a</b>	4-OMe-Ph-CH <sub>2</sub> -COOH	DCM	78
16	<b>6af</b>	DIPEA	1.5–2.0 equiv.	<b>4a</b>	4-OMe-Ph-CH <sub>2</sub> -COOH	THF	—
17	<b>6af</b>	Et <sub>3</sub> N	1.5–2.0 equiv.	<b>4a</b>	4-OMe-Ph-CH <sub>2</sub> -COOH	DMF	80
18	<b>6af</b>	Et <sub>3</sub> N	1.5–2.0 equiv.	<b>4a</b>	4-OMe-Ph-CH <sub>2</sub> -COOH	CHCl <sub>3</sub>	72
19	<b>6af</b>	Et <sub>3</sub> N	1.5–2.0 equiv.	<b>4a</b>	4-OMe-Ph-CH <sub>2</sub> -COOH	DCM	66
20	<b>6af</b>	Et <sub>3</sub> N	1.5–2.0 equiv.	<b>4a</b>	4-OMe-Ph-CH <sub>2</sub> -COOH	THF	—
21	<b>6af</b>	Pyridine	1.5–2.0 equiv.	<b>4a</b>	4-OMe-Ph-CH <sub>2</sub> -COOH	DMF	70
22	<b>6af</b>	Pyridine	1.5–2.0 equiv.	<b>4a</b>	4-OMe-Ph-CH <sub>2</sub> -COOH	CHCl <sub>3</sub>	78
23	<b>6af</b>	Pyridine	1.5–2.0 equiv.	<b>4a</b>	4-OMe-Ph-CH <sub>2</sub> -COOH	DCM	66
24	<b>6af</b>	Pyridine	1.5–2.0 equiv.	<b>4a</b>	4-OMe-Ph-CH <sub>2</sub> -COOH	THF	—

study, the molecular geometry of all the synthesized quinoline derivatives **6a–6z** and **6aa–6aw** was successfully optimized, resulting in local minima for these compounds. The energy gap ( $\Delta E$ ) between the HOMO and LUMO determines chemical reactivity and kinetic stability. A large  $\Delta E$  indicates low reactivity and high stability, whereas a smaller gap promotes electronic charge transfer, making the molecule more polarized and reactive, also known as a “soft” molecule. A high  $\Delta E$  indicates a “hard” molecule with lower reactivity and greater stability. Fig. 4 shows the frontier diagram and calculated energy gaps for compounds **6a**, **6e**, **6n**, **6q**, **6t**, **6ab**, **6al**, **6aq**, **6ar**, and **6aw**: 3.720, 3.756, 3.597, 3.382, 3.640, 3.658, 3.938, 3.793, 3.760, and 1.878 eV, respectively. In the case of HOMO, while the charge density for **6a** and **6ab** is accumulated in the quinoline ring, amide group and to a lesser extent on the aryl ring at 4th position, for **6e** and **6al**, the charge density is accumulated in the quinoline group along with the aryl ring also. However, in the case of LUMO, the charge densities for **6a**, **6ab**, **6e** and **6al** are spread out only on the quinoline ring. In the LUMO the charge density for **6n** is accumulated in the quinoline moiety but in the case of **6a** and **6ab** LUMO, charge density is accumulated in the quinoline moiety and also on the carbonyl group at the 3rd position to a lesser extent. For **6q** the HOMO revealed that the charge density is accumulated in the quinoline moiety and amide group but in the LUMO the charge density accumulated on the amide group and substituted pyridine ring with Cl group because charge density is spread from the quinoline through the amide bond towards the pyridine ring. For **6t** the HOMO illustrated that the charge density is accumulated on the amide, aryl ring since aryl carries electron withdrawing  $-\text{NH}_2$  while its

LUMO shows that the charge density accumulated on the quinoline ring. In the case of **6y** and **6aw**, the HOMO shows that the charge density is accumulated on amide and diphenyl amine moiety also while their LUMO indicates that charge density accumulated on the quinoline ring itself. The HOMO of **6aq** shows that the charge density is accumulated on amide, substituted aryl ring and also on quinoline ring to a lesser extent; the LUMO shows that the charge density accumulated on quinoline ring only. HOMO–LUMO visualizations and energy values for **6a–6z** and **6aa–6aw** are given in the ESI in Fig. S7.1–S7.49 † (FMO diagrams) and Table 2 (calculation), respectively. Excitation energy is the energy required to move an electron from its ground state to an excited state. Excitation energies are often calculated in the context of DFT calculations to understand electronic transitions, optical absorption spectra, and other electron-related phenomena. Molecules with smaller energy gaps are more easily “activated” and can readily participate in chemical reactions. The reactivity of molecules depends on their energy gap. Molecules with a smaller energy gap require less energy to cross the energy barrier and are therefore more reactive than those with a larger energy gap. The comparison of energy gaps in Table 2 indicates that electron-donating functional groups, including  $-\text{NH}_2$ ,  $-\text{OH}$ , 2-OMe,  $-\text{CH}_3$ , pyridine present in various positions on the molecules such as **6t** (3.640 eV), **6v** (3.402 eV), **6s** (3.809 eV), **6f** (3.696 eV), **6ao** (3.610 eV), **6ap** (3.474 eV), **6aq** (3.761 eV), and **6aw** (3.395 eV), can effectively reduce the energy gap. These groups exert their influence by altering the electronic structure of the molecules, thereby increasing electron density and stabilizing energy levels. Alternatively, electron-withdrawing groups such





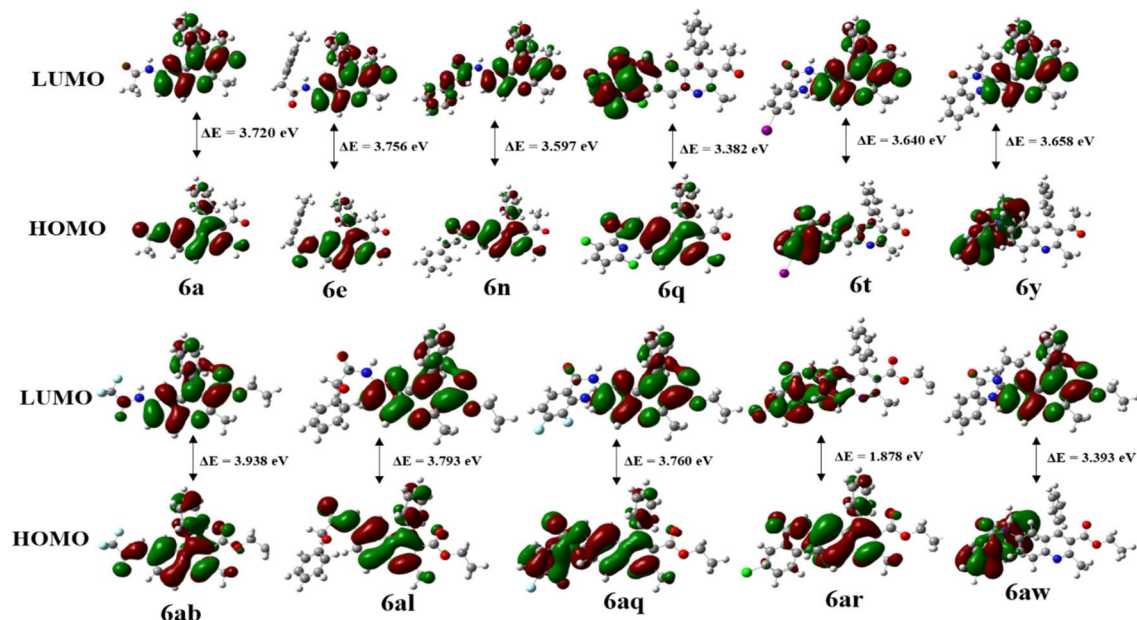


Fig. 4 Frontier Molecular Orbitals of compounds 6a, 6e, 6n, 6q, 6t, 6y, 6ab, 6al, 6aq, 6ar and 6aw.

Table 2 FMO parameters HOMO, LUMO and  $\Delta E$  energies(eV) at the level B3LYP/6-31 G'(d,p)

S No.	Compound code	HOMO	LUMO	$\Delta E$	S No.	Code	HOMO	LUMO	$\Delta E$
1	6a	-5.954	-2.234	3.720	26	6z	-5.597	-1.939	3.658
2	6b	-6.173	-2.341	3.832	27	6aa	-5.856	-2.114	3.742
3	6c	-6.317	-2.377	3.940	28	6ab	-6.198	-2.259	3.939
4	6d	-5.837	-2.084	3.753	29	6ac	-6.067	-2.225	3.842
5	6e	-5.818	-2.071	3.747	30	6ad	-5.747	-1.961	3.786
6	6f	-5.755	-2.059	3.696	31	6ae	-5.736	-1.968	3.768
7	6g	-5.893	-2.159	3.734	32	6af	-5.682	-1.977	3.705
8	6h	-5.893	-2.063	3.830	33	6ag	-5.629	-1.823	3.806
9	6i	-5.91	-2.114	3.796	34	6ah	-5.639	-1.801	3.838
10	6j	-5.901	-2.106	3.795	35	6ai	-5.806	-1.991	3.815
11	6k	-5.903	-2.108	3.795	36	6aj	-5.782	-1.967	3.815
12	6l	-5.964	-2.191	3.773	37	6ak	-5.798	-1.985	3.813
13	6m	-5.912	-2.156	3.756	38	6al	-5.866	-2.072	3.794
14	6n	-5.756	-2.159	3.597	39	6am	-5.817	-2.039	3.778
15	6o	-5.702	-2.122	3.580	40	6an	-5.669	-2.071	3.598
16	6p	-6.211	-2.603	3.608	41	6ao	-5.494	-1.884	3.610
17	6q	-5.794	-2.417	3.377	42	6ap	-5.623	-2.149	3.474
18	6r	-6.065	-2.315	3.750	43	6aq	-5.925	-2.164	3.761
19	6s	-6.001	-2.192	3.809	44	6ar	-5.701	-3.822	1.879
20	6t	-5.772	-2.132	3.640	45	6as	-5.563	-3.721	1.842
21	6u	-5.851	-2.177	3.674	46	6at	-5.761	-2.056	3.705
22	6v	-5.67	-2.268	3.402	47	6au	-5.81	-2.415	3.395
23	6w	-5.844	-3.908	1.936	48	6av	-6.107	-2.551	3.556
24	6x	-5.779	-3.851	1.928	49	6aw	-5.341	-1.946	3.395
25	6y	-5.415	-2.069	3.346					

as the CN group in positions **6b** (3.832 eV) and **6ac** (3.842 eV), CF<sub>3</sub> groups in positions **6c** (3.940 eV) and **6ab** (3.939), and NO<sub>2</sub> groups in positions **6w** (1.936 eV), **6x** (1.928 eV), **6ar** (1.879 eV), and **6as** (1.842 eV), exert a notable decrease in the energy gap of the respective molecules. By withdrawing electrons from the phenyl ring, these groups effectively destabilize energy levels, leading to a significant reduction in the energy gap. Consequently, this decrease in the energy gap enhances the chemical

reactivity of the molecules, rendering them more reactive in various chemical processes.

#### 2.4 Molecular electrostatic potential (MEP)

Molecular Electrostatic Potential (MEP) is a useful tool for studying molecular interactions and reactions, especially in biological contexts where electron-rich sites such as oxygen or nitrogen act as donors and hydrogen atoms or groups act as



acceptors in hydrogen bonding. MEP images provide insights into electron density distribution, which helps in identifying favorable interaction sites. By recognizing electron-rich or electron-deficient regions, MEP can predict reactivity patterns, guiding research and development efforts in chemistry by shedding light on the mechanics of electrophilic and nucleophilic reactions. This facilitates rational drug design and synthetic strategies.<sup>77–79</sup> MEP surface analysis predicts molecular reactivity by mapping electron-rich (red) and electron-poor (blue) regions and helps identify sites prone to electrophilic or nucleophilic attack, crucial for optimizing chemical reactions and molecular properties design.<sup>80</sup> The MEP diagram is a valuable tool for understanding the charge distribution within the molecule. It allows us to predict the electrophilic (electron-deficient) and nucleophilic (electron-rich) active sites within the complex based on the electron density.<sup>81</sup> The MEP analysis of all the synthesized compounds **6a–6z** and **6aa–6aw** is shown in the ESI Fig. S8.† In Fig. 5, the electron density for compounds **6a**, **6e**, **6n**, **6q**, **6t**, **6y**, **6ab**, **6al**, **6aq**, **6ar**, and **6aw** has been visually represented where it is colour-coded to provide a scale. Among the FMO distribution pattern of all the compounds **6a–6z** and **6aa–6aw**, it was observed that compound **6c** had a higher energy gap, which could be due to the strong dipole-dipole movements and van der Waals forces of dispersion that nitriles

exhibit. The electronic chemical potential is defined as the variation in energy of a system to the number of electrons ( $N$ ) at a constant external potential. It is a measure of the system's ability to exchange electron density with the surrounding environment in its lowest energy state.<sup>82,83</sup> Some reagents involved in acid amine cross-coupling reactions have their electronic chemical potential listed in the ESI† (Table 2), along with some electron-donating and electron-withdrawing groups ( $\mu$ ) in Table S2.† Although reagents such as compound **6w** with a value of 4.87 eV act as strong electron-donating molecules, as the electron-withdrawing character of the substituent present increases, the electronic chemical potential of the corresponding quinoline derivative decreases this results in an increase in the polar character of **6t**, **6v**, **6ap**, **6aq**, and **6ao** (**6w** < **6x** < **6ar** < **6as**). Compounds **6i**, **6h**, **6ai**, and **6j** have the same molecular formula, but the position of Cl is distinct, resulting in a slight change in the Stokes shift and energy ( $\Delta E$ ) difference. In compounds **6j**, **6k**, **6aj**, and **6ak**, the bromine substituent is either in the *meta* or *para* position, but their shifts are similar.

## 2.5 TD-DFT (UV-vis)

Density functional theory (DFT) is a quantum mechanical approach that posits that all system properties stem from

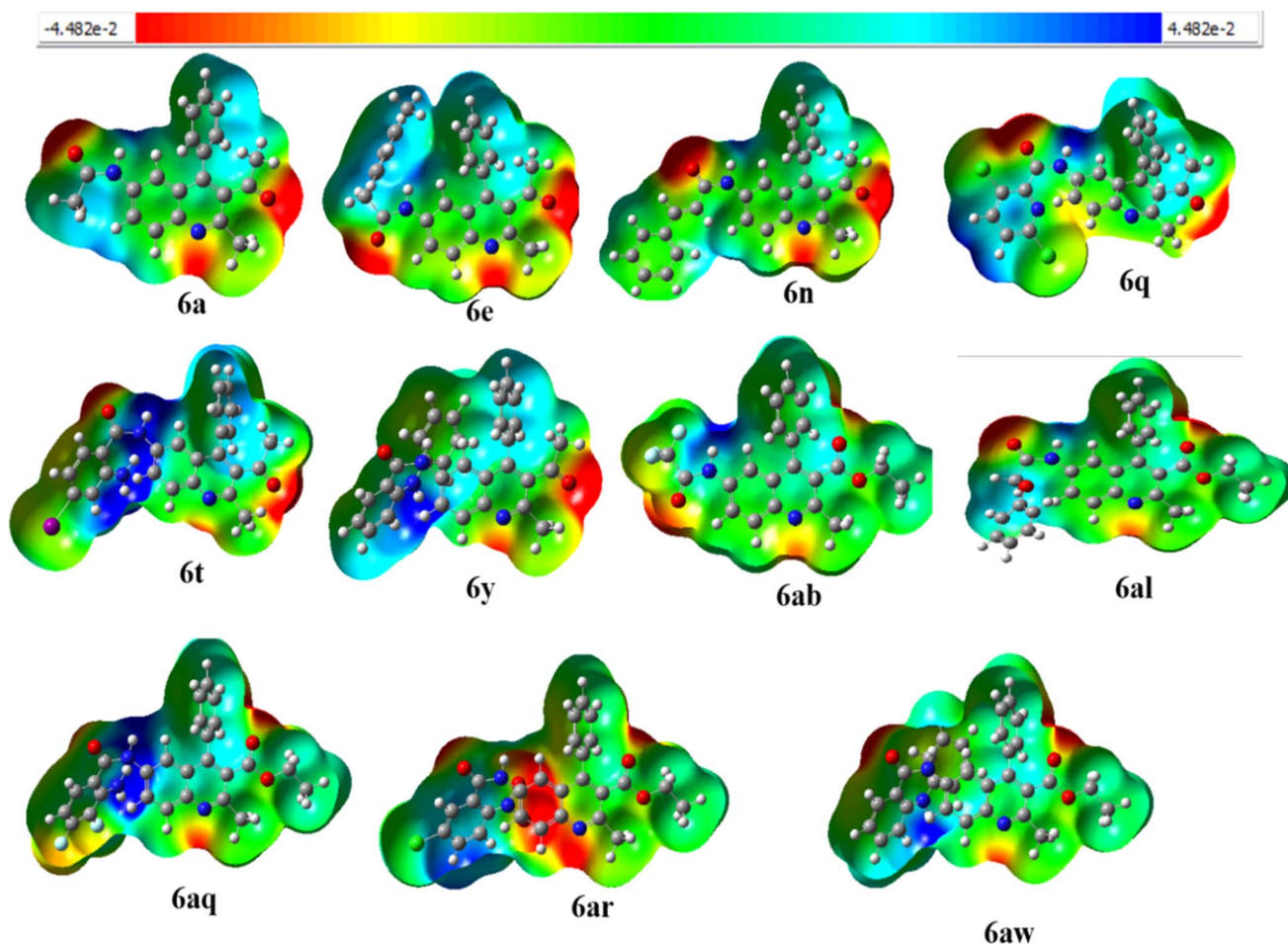


Fig. 5 Molecular Electrostatic Potential (MEP) surface of compound **6a**, **6e**, **6n**, **6q**, **6t**, **6y**, **6ab**, **6al**, **6aq**, **6ar**, **6aw**.

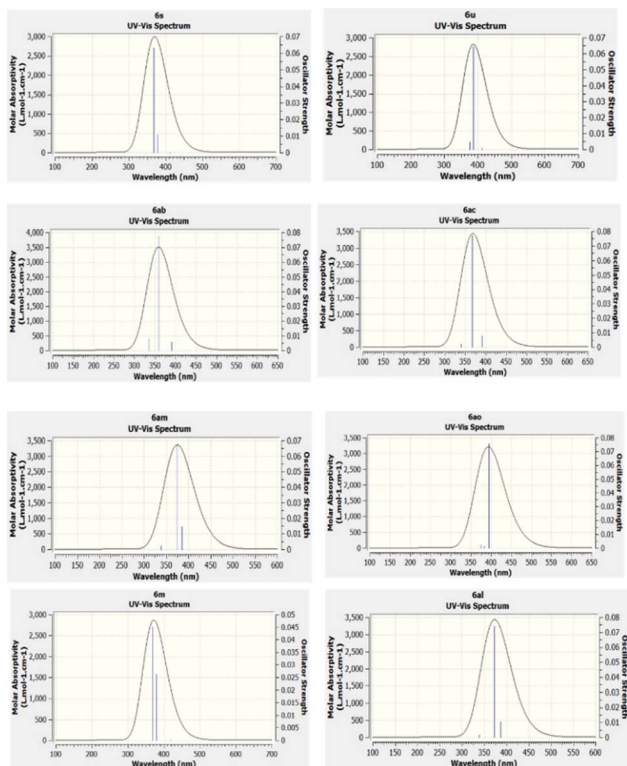
charge density functions. This framework facilitates precise calculations elucidating a compound's structure, energy, and molecular attributes. Time-dependent density functional theory (TD-DFT) emerges as a result of leveraging DFT for excited states, enabling the computation of electronic spectra for absorption.<sup>84,85</sup> TD-DFT empowers investigations into various processes involving excited states, offering insights into phenomena such as molecular transitions and optical properties. This methodological synergy between DFT and TD-DFT furnishes a robust toolkit for comprehensively analyzing molecular behaviour, thereby advancing our understanding of chemical reactivity, spectroscopy, and materials science applications. TD-DFT is a prevalent computational method used to predict absorption spectra in molecules. UV-vis spectra were calculated using the TD-DFT approach employing the B3LYP functional and 6-31G' (d,p) basis sets. These calculations furnish comprehensive data enabling the elucidation of theoretical UV-vis absorption profiles, aiding in the comprehension of molecular absorptivity data. By leveraging GaussView 5, pertinent information about UV-vis spectra was extracted and depicted in Fig. 6. The TD-DFT-derived UV-vis prediction results exhibit a satisfactory correspondence with experimental data, as evidenced by the comparison presented in Table 3. This convergence underscores the utility and reliability of the TD-DFT methodology in simulating molecular absorption spectra, thereby facilitating a deeper understanding of molecular properties and behaviour.<sup>86</sup> Our study analyzed the UV-vis and

**Table 3** Theoretical  $\lambda_{\text{max}}$ , experimental  $\lambda_{\text{max}}$ , oscillation strength and energy (eV)

Sr. No.	Experimental $\lambda_{\text{max}}$ nm	Theoretical $\lambda_{\text{max}}$ nm	Oscillation strength, $f$	Energy eV
1	<b>6ab</b> (372.65)	359.51	0.0777	3.4486
2	<b>6ac</b> (339.11)	339.97	0.002	3.6469
3	<b>6al</b> (337.72)	338.72	0.0018	3.6604
4	<b>6am</b> (338.18)	338.24	0.0023	3.6656
5	<b>6ao</b> (370.32)	374.17	0.0023	3.3135
6	<b>6m</b> (337.12)	368.49	0.042	3.3647
7	<b>6s</b> (352.83)	368.85	0.0632	3.3614
8	<b>6u</b> (352.63)	375.31	0.0045	3.3035

emission spectra of 31 compounds. We then focused on eight compounds that span a range of low and high wavelengths, as shown in Table 3. Among these eight compounds, compound **6am** showed the smallest variance, with a theoretical  $\lambda_{\text{max}}$  of 338.23 nm and an experimental  $\lambda_{\text{max}}$  of 338.18 nm, resulting in a marginal difference of 0.05 nm. Similarly, compound **6al** exhibited a modest difference, with a theoretical  $\lambda_{\text{max}}$  of 338.23 nm and an experimental  $\lambda_{\text{max}}$  of 337.72 nm, indicating a variance of 0.51 nm. Compound **6ac** demonstrated a minimal difference, with theoretical and experimental  $\lambda_{\text{max}}$  values of 339.96 nm and 339.11 nm, respectively, producing a disparity of 0.85 nm. On the other hand, compound **6m** exhibited the most significant difference, with a theoretical  $\lambda_{\text{max}}$  of 368.50 nm and an experimental  $\lambda_{\text{max}}$  of 337.12 nm, resulting in a notable discrepancy of 31.37 nm. Compounds **6u** and **6s** showed comparable results, with theoretical  $\lambda_{\text{max}}$  values of 368.63 nm and 368.83 nm, and experimental  $\lambda_{\text{max}}$  values of 352.63 nm and 352.83 nm, respectively, indicating differences of approximately 15.87 nm and 16.2 nm. Compound **6ab** and **6ao** also showed variances, with differences of 13.14 nm and 3.85 nm, respectively, between theoretical and experimental  $\lambda_{\text{max}}$  values. These findings highlight the importance of meticulous analysis to reconcile theoretical predictions with experimental observations in spectral studies. The theoretical and experimental UV-vis spectra and  $\lambda_{\text{max}}$ , oscillation strength  $f$ , energy, of all 31 compounds are shown in ESI Fig. S9 and Table S2,<sup>†</sup> respectively. The UV-vis spectra data for the compounds studied revealed a notable correlation between wavelength and energy gap, as evidenced by the generated graphs. It was observed that as the wavelength approached the visible light range (around 400 nm), the energy gap decreased. For instance, compound **6al** exhibited a calculated energy gap of 3.6604 eV and a  $\lambda_{\text{max}}$  of 337.72 nm, consistent with its UV absorption profile, which indicates activity within the UVC range (180–360 nm). Conversely, compound **6ab**, as displayed an energy gap of 3.1748 eV and a  $\lambda_{\text{max}}$  of 372.65 nm, suggesting its primary UV absorption occurs within the UVA range (320–400 nm). This correlation underscores the relationship between molecular structure and spectral behaviour.

The variation between simulated and experimental spectral data, with respect to oscillator strength and energy is given in Table 3. The spectral analysis revealed distinct medium energy band  $n-\pi^*$  transitions for various compounds: **6ab** exhibited absorption peaks at 390.51 nm, 359.51 nm, and 334.90 nm;



**Fig. 6** Theoretical UV-vis spectra of compounds by TD-DFT at level using the B3LYP/6-31G'(d,p). Blue lines demonstrate the main electronic transitions (singlet state).





**6ac** displayed transitions at 391.08 nm, 367.69 nm, and 339.97 nm; for compound **6al** absorption bands at 387.18 nm, 372.38 nm, and 338.72 nm; **6am** absorption peaks at 385.51 nm, 373.92 nm, and 338.24 nm; **6ao** presented absorptions peaks at 394.79 nm, 383.47 nm, and 374.17 nm; **6m** demonstrated transitions at 418.60 nm, 378.81 nm, and 368.49 nm; for compound **6s** absorptions peaks at 412.70 nm, 379.63 nm, and 368.85 nm; and **6u** exhibited peaks at 411.77 nm, 387.23 nm, and 375.31 nm. These findings elucidate the distinct electronic transitions within the compounds. Additionally, compounds **6ab**, **6ac**, **6al**, **6am**, **6ao**, **6m**, **6s**, and **6u** notably exhibited major contributions above 50% in their respective spectra, indicative of their significant roles in the observed electronic transitions. All 31 compounds calculated (three excited state) transition state from that selected major electronic transition contribution shown in Table S3† and FMO diagram density surface of the selected major contributions of compounds frontier molecular orbitals involved in electronic transitions of at Basic set level of B3LYP/6-31 + G'(d,p) using iso-surface value of 0.02 au shown in Table S4.† Lower energy (LE) and higher energy (HE) bands, that is, S0 → S1 and S0 → S2 transitions, indicating the ICT process.

The corresponding major distribution for **6ab** is HOMO-1 → LUMO (86%), HOMO-2 → LUMO (76%), HOMO → LUMO

(68%), for Compound **6ac** HOMO-1 → LUMO (89%), HOMO-2 → LUMO (85%), HOMO → LUMO (80%), Compound **6al** is HOMO-3 → LUMO (90%), HOMO-1 → LUMO (89%), HOMO → LUMO (85%), Compound **6am** is HOMO-2 → LUMO (90%), HOMO-1 → LUMO (86%), HOMO → LUMO (82%), for compound **6ao** is HOMO-1 → LUMO (96%), HOMO → LUMO (93%), HOMO-2 → LUMO (91%), for compound **6m** is HOMO-1 → LUMO (82%), HOMO → LUMO (55%), HOMO-2 → LUMO (53%), for compound **6s** is HOMO → LUMO (78%), HOMO-1 → LUMO (61%), HOMO-2 → LUMO (57%), for compound **6u** is HOMO → LUMO (89%), HOMO-2 → LUMO (62%), HOMO-3 → LUMO (59%) transitions for compounds, which reveals the donor- $\pi$ -acceptor charge transfer character of the compounds.

## 2.6. Absorption and emission

The photophysical study was conducted in methanol absorbance and emission spectra were recorded at room temperature. Observed UV-vis range 305–37 nm and emission range 430–502 nm shown in Table 4. It was observed that the intensity was higher when the ester group was placed at the 3rd position of quinoline compared to the keto function. The fluorescence intensity properties of ketones and esters are complex and dependent on various factors. While ketones can exhibit

**Table 4** Photophysical data of compounds absorption, emission, stokes shift, molar extinction coefficient and quantum yield (the excitation wave length is 350 nm with  $2 \times 10^{-5}$  M concentrations)

Entry	Compound code	$\lambda_{\text{Absorption}}$ (nm)	$\lambda_{\text{Emission}}$ (nm)	Stokes shift $\Delta \times 10^4$	Molar extinction coefficient $\times 10^4$	Quantum yield ( $\Phi$ )
1	<b>6a</b>	341.12	455.19	0.7347		
2	<b>6b</b>	338.22	440.15	0.6858	6.2125	0.0020
3	<b>6d</b>	340.38	462.33	0.7766	4.6795	0.0014
4	<b>6e</b>	342.76	452.81	0.7116	5.1485	0.0019
5	<b>6f</b>	341.45	457.39	0.7444	5.0510	0.0022
6	<b>6g</b>	343.51	450.43	0.6932	3.0205	0.0057
7	<b>6h</b>	340.28	451.36	0.7239	5.3315	0.0020
8	<b>6i</b>	344.68	448.98	0.6748	4.8225	0.0037
9	<b>6j</b>	341.09	450.16	0.7103	4.2445	0.0038
10	<b>6k</b>	340.29	450.38	0.7189	4.8250	0.0015
11	<b>6l</b>	338.87	441.77	0.691	4.4505	0.0030
12	<b>6m</b>	337.12	446.06	0.7252	3.6155	0.0050
13	<b>6s</b>	352.83	452.8	0.6286	5.3030	0.0035
14	<b>6u</b>	352.63	434.54	0.5368	6.2180	0.0005
15	<b>6v</b>	347.33	450.28	0.5959	3.6155	0.00012
16	<b>6z</b>	309.18	457.29	1.0411	7.9805	0.0012
17	<b>6aa</b>	338.87	442.94	0.6961	6.4730	0.0013
18	<b>6ab</b>	372.65	501.58	0.6921	6.5455	0.0021
19	<b>6ac</b>	339.11	411.03	0.5168	5.3670	0.0171
20	<b>6ad</b>	342.78	417.73	0.5259	8.2825	0.0096
21	<b>6ae</b>	341.95	423.86	0.5685	8.3715	0.0103
22	<b>6af</b>	340.42	420.21	0.5602	7.1585	0.0156
23	<b>6ag</b>	342.67	420.73	0.543	8.3370	0.0110
24	<b>6ah</b>	340.22	418.34	0.5488	8.8605	0.0124
25	<b>6ai</b>	340.43	423.52	0.5771	9.5080	0.0062
26	<b>6aj</b>	339.17	422.27	0.5802	8.3370	0.0150
27	<b>6ak</b>	340.54	417.61	0.5431	7.9785	0.0188
28	<b>6al</b>	337.72	416.98	0.5635	9.0055	0.0179
29	<b>6am</b>	338.16	416.04	0.5547	8.1260	0.0053
30	<b>6ao</b>	370.32	502.67	0.7107	9.1775	0.0082
31	<b>6ap</b>	350.27	454.16	0.6545	6.9935	0.0154



fluorescence, their intensity may be reduced by electron-donating or electron-accepting groups. On the other hand, ketones with extended conjugation, such as aromatic ketones, often display increased fluorescence due to electron delocalization. The fluorescence intensity of esters can be notably affected by solvent polarity, with nonpolar solvents sometimes enhancing their fluorescence their intensity may be reduced by electron-donating or electron-accepting groups. On the other hand, ketones with extended conjugation, such as aromatic ketones, often display increased fluorescence due to electron delocalization. The fluorescence intensity of esters can be notably affected by solvent polarity, with nonpolar solvents sometimes enhancing their fluorescence. The presence of specific substituents and the degree of esterification also play a role in determining the fluorescence properties of these compounds, with conjugation and rigidity potentially resulting in stronger fluorescence. Additionally, the ability of esters to undergo intramolecular rotation can impact their fluorescence characteristics. The unique fluorescence behavior of specific ketones and esters is further contributed by individual molecular structures, environmental conditions, and the choice of excitation wavelength. Table 4 presents the photophysical characteristics of the quinoline derivatives that were investigated. It was found that all the derivatives found to exhibit the unique fluorescence and the conversion of amine into amide at the C6 position of quinoline moiety significantly-affected the absorption (Fig. 7) and emission (Fig. 8) in solution (and some of compounds such as **6ae**, **6af**, **6ag**, **6ah**, **6ai** in solid state also Fig. 9). The Fig. 8 has been recorded as unnormalized spectra. The individual emission spectra for all the compounds in solution are available in the ESI.† The quinoline fluorophore **6ab** found to exhibit the absorption maximum at higher wavelength (372.65 nm). The absorption, emission, Stokes shift

molar molar, extinction coefficient and quantum yield for all the derivatives which are exhibiting the fluorescence can be found in Table 4. A larger Stokes shift refers to a greater separation between the wavelengths of excitation and emission. This increased separation reduces overlap between the spectra, leading to more accurate and sensitive fluorescence measurements. It results in better discrimination between the excitation light and the fluorescence signal, which minimizes interference from the background and improves the signal-to-noise ratio. Compounds with high Stokes shifts are valuable for various applications, including fluorescence microscopy, bio-sensing, and fluorescence-based assays. They offer greater flexibility in experimental design and enable the detection of fluorescent signals in complex biological or environmental samples where background fluorescence or scattering is a concern.<sup>87</sup> The compounds **6z**, and **6d** exhibited excellent Stokes shifts of  $1.0411 \times 10^{-4}$  and  $0.7766 \times 10^{-4}$  respectively in methanol solution, while the Stokes shift of compound **6ac** is comparatively low at  $0.5168 \times 10^{-4}$  in the same solution. Some compounds were recorded in solid-state spectra, which showed different absorption and emission wavelengths in the spectra, as well as Stokes shift. The solid-state Stokes shift values were  $0.6226 \times 10^{-4}$  for **6e**,  $0.5226 \times 10^{-4}$  for **6f**, and  $0.7587 \times 10^{-4}$  for **6h**. Photo-physical data of synthesized derivatives, including emission, absorption, and Stokes shift, can be found in Table 4. All compounds exhibited favorable molar extinction coefficients ( $\epsilon$ ), with values ranging from  $3.0205 \times 10^4 \text{ M}^{-1} \text{ cm}^{-1}$  to  $9.5080 \times 10^4 \text{ M}^{-1} \text{ cm}^{-1}$  across entries 1–31 in Table 4. Particularly noteworthy were quinoline derivatives showcasing enhanced aromaticity, notably those bearing ester groups at the third position of quinoline core, in combination with halide (Cl) and amine functionalities, which displayed the highest molar extinction coefficients (Table 4, entries 25 and 30). Assessment of quantum efficiencies was carried out by comparing them to the established quantum efficiencies of quinine sulfate in a 0.5 M  $\text{H}_2\text{SO}_4$  solution ( $\Phi = 0.54$ ) at an excitation wavelength of 340 nm. All the derivatives displayed a noteworthy level of quantum yield, as detailed in (Table 4, entries 1 to 31). The quantum efficiency of the quinoline framework was found to be dependent on the specific electronic characteristics exhibited by the aryl substituents. It was observed that the inclusion of  $\text{Ar-CH}_2$  ring substituents significantly enhanced the quantum yield, in contrast, the presence of  $\text{Ar}$  substituents was found to have a suppressive impact on the quantum yield, as indicated in Table 4.

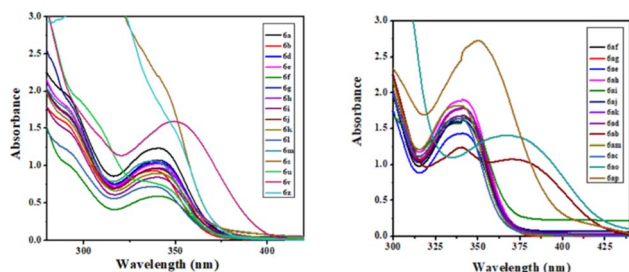


Fig. 7 UV-vis spectra of compounds in MeOH solution.

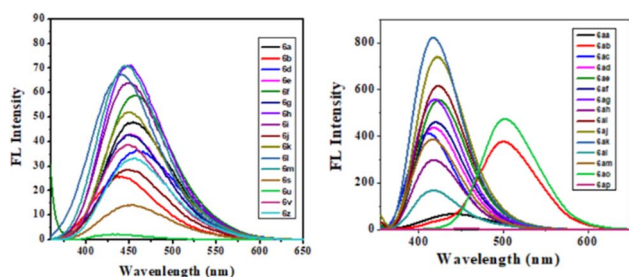


Fig. 8 FL spectra of compounds in MeOH solution.

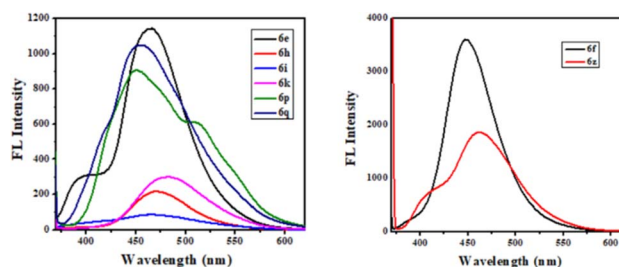


Fig. 9 FL spectra of compounds in solid state.



### 3 Experimental

#### 3.1. General method

The experiments were carried out using round bottom flasks, and all chemical materials and solvents were obtained from commercial sources. The preparation of 1-(6-amino-2-methyl-4-phenylquinolin-3-yl)-ethan-1-one was done following previously reported protocols. The  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were recorded using a Bruker Avance 400 spectrometer and referenced to the residual solvent signal  $\text{CDCl}_3$ : (7.26) for  $^1\text{H}$  and (77.16) for  $^{13}\text{C}$  NMR; dimethyl sulfoxide- $\text{d}_6$  (2.50) for  $^1\text{H}$  and (39.50) for  $^{13}\text{C}$ . The chemical shifts ( $\delta$ ) are given in ppm, and coupling constants ( $J$ ) were measured in Hz. The abbreviations used were: s-singlet, d-doublet, dd-doublet of the doublet, t-triplet, m (multiple), and br-broad. HR ESI-MS values were obtained using Xeo G2-XS QToF (waters) and given in  $m/z$ . Absorption was recorded using a JASCO V-670 spectrometer. Steady-state fluorescence spectra were recorded on the Hitachi F-7000 FL spectrofluorophotometer by exciting at the respective absorption maxima. Column chromatography was performed using silica gel (100–200 mesh) packed in glass columns. Analytical TLC was conducted on Macherey-Nagel 60 F245 aluminium-backed silica gel plates.

The Stoke's shift was calculated using the following equation.

$$\Delta\bar{\nu} = 10^7 / \lambda_{\text{max (absorption)}} - 10^7 / \lambda_{\text{max (emission)}} \quad (1)$$

Quantum yield calculated the following equation.

$$\Phi_{\text{f}} = \Phi_{\text{R}} \cdot A_{\text{R}} / A \cdot I / I_{\text{R}} \cdot n_{\text{R}}^2 / n^2 \quad (2)$$

where  $\Phi$  is the quantum yield,  $I$  is the integrated intensity,  $A$  is the absorbance, and  $n$  is the refractive index. The subscript R refers to quinine sulphate. The molar extinction coefficient ( $\epsilon$ ) was calculated using Beer–Lambert law  $A = \epsilon cl$ . The molar extinction coefficient is defined as the measure of absorbance per unit concentration and path length. Mathematically, it is expressed as  $\epsilon = A / (c \times l)$ , where  $A$  is the absorbance,  $c$  is the concentration of the absorbing species, and  $l$  is the path length of the light through the sample.

#### 3.2. Synthesis of title compounds (acid amine cross-coupling reaction) 6a–6z and 6aa–6aw. General procedure

To perform the reaction, take 1-(6-amino-2-methyl-4-phenylquinolin-3-yl) ethan-1-one (300–500 mg), substituted carboxylic acids (1.5 to 5.0 equivalents), HATU (2.0 to 2.5 equivalents), and DIPEA (2.5 equivalents) in a 50 mL round bottom flask containing 10 mL of DMF of analytical grade. Stir the mixture at room temperature under nitrogen gas for 6–12 hours. Once the reaction is complete, add the reaction mixture to cold water and extract with ethyl acetate ( $2 \times 25$  mL). Wash the organic layer with brine water (25 mL). Concentrate the organic layer under high vacuum to obtain crude compound. Purify the crude compound using column chromatography to obtain pure products **6a–6z** and **6aa–6aw**. You can find all reaction procedures and identification spectral data in the ESI.†

### 4 Conclusion

In Conclusion through the implementation of an acid-amine coupling methodology, a total of 49 new quinoline derivatives have been synthesized. Further modifications were carried out to introduce different substituents at the C6 position of the quinoline scaffold, allowing for the customization of their DFT investigations and photo-physical characteristics. We compared theoretical  $\lambda_{\text{max}}$  value with experimental  $\lambda_{\text{max}}$  value and got very less difference in compounds **6am**. Currently, ongoing research is focused on conducting a comprehensive investigation into this newly discovered class of quinoline compounds.

### Author contributions

The work has been designed and executed by the corresponding author Vijayaparthasarathi Vijayakumar and target molecules are synthesized, characterized and the studies are carried out by the first author Nagesh Dhanaji Chavan.

### Conflicts of interest

The authors declare no conflicts of interest.

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### References

- (a) K. B. Patel and P. Kumari, *J. Mol. Struct.*, 2022, **15**, 1268; (b) N. Sun, R. L. Du, Y. Y. Zheng, B. H. Huang, Q. Guo, R. f. Zhang, K. Y. Wong and Y. Lu, *Eur. J. Med. Chem.*, 2017, **135**, 1–11.
- O. A. Olateju, C. o. Babalola, O. O. Olubiyi, O. A. Kotila, D. A. Kwasi and A. O. Oaikhen, *Front. Microbiol.*, 2021, **12**, 556550.
- (a) K. Govindarao, N. Srinivasan, R. Suresh, R. K. Raheja, S. Annadurai, R. R. Bhandare and A. B. Shaik, *J. Saudi Chem. Soc.*, 2022, **26**(3), 101471; (b) A. Shi, T. A. Nguyen, S. K. Battina, S. Rana, D. J. Takemoto, P. K. Chiang and D. H. Hua, *Bioorg. Med. Chem. Lett.*, 2008, **18**(11), 3364–3368.
- A. Kania, W. Tejchman, A. M. Pawlak, K. Mokrzyński, B. Rózanowski, B. M. Musielak and M. Greczek-Stachura, *Molecules*, 2022, **27**(3), 1069.
- K. L. Ji, W. Liu, W. H. Yin, J. Y. Li and J. M. Yue, *Org. Biomol. Chem.*, 2022, **20**(20), 4176–4182.
- L. Savegnago, A. I. Vieira, N. Seus, B. S. Goldani, M. R. Castro, E. J. Lenardão and D. Alves, *Tetrahedron Lett.*, 2013, **54**(1), 40–44.



- 7 R. S. Keri and S. A. Patil, *Biomed. Pharmacother.*, 2014, **68**(8), 1161–1175.
- 8 Y. Q. Hu, C. Gao, S. Zhang, L. Xu, Z. Xu, L. S. Feng, X. Wu and F. Zhao, *Eur. J. Med. Chem.*, 2017, **139**, 22–47.
- 9 A. Dorababu, *ChemistrySelect*, 2021, **6**(9), 2164–2177.
- 10 L. J. Kumar, Y. Suresh, R. Rajasekaran, S. R. Reddy and V. Vijayakumar, *J. Iran. Chem. Soc.*, 2019, **16**, 1071–1080.
- 11 L. Jyothish Kumar and V. Vijayakumar, *Res. Chem. Intermed.*, 2017, (43), 5691–5705.
- 12 B. Mahjour, Y. Shen and W. Liu, *Nature*, 2020, (580), 71–75.
- 13 R. Dabhi, U. Patel, V. Rathod, S. Shah and J. Maru, *Curr. Chem. Lett.*, 2023, **12**(1), 133–140.
- 14 R. Jasinski, O. M. Demchuk and D. Babyuk, *J. Chem.*, 2017, **2017**, 3617527.
- 15 A. Łapczuk-Krygier, A. Kącka-Zych and K. Kula, *Chem. Lett.*, 2019, (8), 13–38.
- 16 R. Jasinski and E. Dresler, *Organics*, 2020, (1), 49–69.
- 17 J. C. Sheehan and G. P. Hess, *J. Am. Chem. Soc.*, 1955, **77**(4), 1067–1068.
- 18 W. König and R. Geiger, *Chem. Ber.*, 1970, **103**(3), 788–798.
- 19 F. Albericio, J. M. Bofill, A. El-Faham and S. A. Kates, *J. Org. Chem.*, 1998, **63**(26), 9678–9683.
- 20 M. Zhang, P. Vedantham, D. L. Flynn and P. R. Hanson, *J. Org. Chem.*, 2004, **69**(24), 8340–8344.
- 21 V. Bhatt, S. D. Samant and S. Pednekar, *Lett. Org. Chem.*, 2017, **14**(10), 764–768.
- 22 C. A. Montalbetti and V. Falque, *Tetrahedron*, 2005, **61**(46), 10827–10852.
- 23 M. Fernández-Suárez and A. Y. Ting, *Nat. Rev. Mol. Cell Biol.*, 2008, **9**(12), 929–943.
- 24 C. Y. Wu, J. J. Wei, D. H. Tian, Y. Feng, R. H. Miller and Y. M. Wang, *J. Med. Chem.*, 2008, **51**, 6682.
- 25 C. N. Wang, C. Y. Wu, J. Q. Zhu, R. H. Miller and Y. M. Wang, *J. Med. Chem.*, 2011, **54**, 2331.
- 26 R. L. Sheng, P. F. Wang, Y. H. Gao, Y. Wu, W. M. Liu, J. J. Ma, H. P. Li and S. K. Wu, *Org. Lett.*, 2008, **10**, 5015.
- 27 H. S. Jung, P. S. Kwon, J. W. Lee, J. I. Kim, C. S. Hong, J. W. Kim, S. H. Yan, J. Y. Lee, J. H. Lee, T. Joo and J. S. Kim, *J. Am. Chem. Soc.*, 2009, **131**, 2008–2012.
- 28 N. A. Owiti, Z. D. Nagel and B. P. Engelward, *Trends Cancer*, 2021, **7**, 240–248.
- 29 N. R. Gassman and N. W. Holton, *Curr. Opin. Biotechnol.*, 2019, **55**, 30–35.
- 30 A. Zarkov, A. Vasilev, T. Deligeorgiev, S. Stoyanov and M. Nedelcheva-Veleva, *Mol. Imag.*, 2013, **12**(2), 90–99.
- 31 J. Gebhard, L. Hirsch, C. Schwechheimer and H. A. Wagenknecht, *Bioconjugate Chem.*, 2022, **33**, 1634–1642.
- 32 P. J. Kolbeck, W. Vanderlinden, G. Gemmecker, C. Gebhardt, M. Lehmann, A. Lak, T. Nicolaus, T. Cordes and J. Lipfert, *Nucleic Acids Res.*, 2021, **49**, 5143–5158.
- 33 D. M. Kolpashchikov, *Acc. Chem. Res.*, 2019, **52**, 1949–1956.
- 34 E. A. Prasetyanto, H. S. Wasisto and D. Septiadi, *Acta Biomater.*, 2022, **143**, 39–51.
- 35 H. Freiser, *Analyst*, 1952, **77**, 830–845.
- 36 L. D. Lavis and R. T. Raines, *ACS Chem. Biol.*, 2008, **3**(3), 142–155.
- 37 T. S. Kaufman and E. A. Rúveda, *Angew. Chem., Int. Ed.*, 2005, **44**(6), 854–885.
- 38 Y. Laras, V. Hugues, Y. Chandrasekaran, M. Blanchard-Desce, F. C. Acher and N. J. Pietrancosta, *J. Org. Chem.*, 2012, **77**(18), 8294–8302.
- 39 (a) C. Fahrni and T. O'Halloran, *J. Am. Chem. Soc.*, 1999, **121**, 11448–11458; (b) A. Nowakowski and D. Petering, *Inorg. Chem.*, 2011, **50**, 10124–10133; (c) W. Jeffrey, J. Meeusen, H. Tomasiewicz, A. Nowakowski and D. Petering, *Inorg. Chem.*, 2011, **50**, 7563–7573; (d) K. Hanaoka, K. Kikuchi, H. Kojima, Y. Urano and T. Nagano, *J. Am. Chem. Soc.*, 2004, **126**(39), 12470–12476; (e) G. Li, D. Zhu, L. Xue and H. Jiang, *Org. Lett.*, 2013, **15**(19), 5020–5023.
- 40 W. Li, W. Lin, J. Wang and X. Guan, *Org. Lett.*, 2013, **15**(7), 1768–1771.
- 41 S. Bhagat, N. Sharma and T. S. Chundawat, *J. Chem.*, 2012, **2013**, 909217.
- 42 K. Rajalakshmi, S. Muthusamy, H. J. Lee, P. Kannan, D. Zhu, R. S. Lodi, M. Xie, J. Xie, J. W. Song and Y. Xu, *Spectrochim. Acta, Part A*, 2024, **304**, 123282.
- 43 S. Wu, Q. Yan, Y. Wang, Y. Bian, D. Shi and H. Xu, *Microchem. J.*, 2024, **201**, 110597.
- 44 S. Che, L. Yin, M. Chen, F. Fan, A. Xu, C. Zhou, H. Fu and Y. She, *Food Chem.*, 2023, **407**, 135052.
- 45 S. Suguna, K. Velmurugan, D. Parimaladevi, A. Abiram, P. M. Sukitha, V. R. Kannan, R. S. Kumar, A. L. Almansour, K. Perumal, J. Prabhu and R. Nandhakumar, *J. Photochem. Photobiol., A*, 2024, **447**, 115226.
- 46 P. C. Mai, T. C. Ngo, T. H. Nguyen and T. H. Le, *J. Fluoresc.*, 2023, 1–9, DOI: [10.1007/s10895-023-03477-9](https://doi.org/10.1007/s10895-023-03477-9).
- 47 K. Balasubramanian, S. K. Ramaraj, C. Karuppiyah, C. C. Yang and K. Nagaraj, *Inorg. Chem. Commun.*, 2023, **155**, 111069.
- 48 R. W. Sinkeldam, N. J. Graco and Y. Tor, *Chem. Rev.*, 2010, **110**(5), 2579–2619.
- 49 R. Dhanapal, P. T. Perumal, M. Damodiran, C. Ramprasath and N. Mathivanan, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 6494–6497.
- 50 X. Meng, S. X. Wang and M. Z. Zhu, Quinoline-Based Fluorescence Sensors, *Molecular Photochemistry – Various Aspects*, InTech, London, UK, 2012, pp. 1–22, DOI: [10.5772/31771](https://doi.org/10.5772/31771).
- 51 R. Cinar, J. Nordmann, E. Dirksen and T. J. J. Muller, *Org. Biomol. Chem.*, 2013, **11**, 2597.
- 52 M. Malathi, P. S. Mohan, R. J. Butcher and C. K. Venil, *Can. J. Chem.*, 2009, **87**, 1692–1703.
- 53 H. G. Lee, J. H. Lee, S. P. Jang, I. H. Hwang, S. J. Kim, Y. Kim, C. Kim and R. G. Harrison, *Inorg. Chim. Acta.*, 2013, **394**, 542–551.
- 54 L. E. McQuade and S. J. Lippard, *Inorg. Chem.*, 2010, **49**, 9535–9545.
- 55 H. Kim, J. Kang, K. B. Kim, E. J. Song and C. Kim, *Spectrochim. Acta, Part A*, 2014, **118**, 883–887.
- 56 G. Xie, P. Xi, X. Wang, X. Zhao, L. Huang, F. Chen, Y. Wu, X. Yao and Z. Zeng, *Eur. J. Inorg. Chem.*, 2011, **2011**, 2927–2931.
- 57 D. Mahanand and J. C. Houck, *Clin. Chem.*, 1968, **14**, 6–11.





- 58 G. Yang, S. Li, S. Wang, R. Hu, J. Feng, Y. Li and Y. Qian, *Pure Appl. Chem.*, 2013, **85**, 1465–1478.
- 59 M. Fernández-Suárez, A. Y. Ting and M. Fern, *Nat. Rev. Mol. Cell Biol.*, 2008, (9), 929–943.
- 60 C. Santana-Calvo, F. Romero, I. López-González and T. Nishigaki, *BioTechniques*, 2018, (65), 211–218.
- 61 Y. Li, T. Liu, H. Liu, M. Z. Tian and Y. Li, *Accounts Chem. Res.*, 2014, **47**, 1186–1198.
- 62 M. Suresh, P. Kar and A. Das, *Inorg. Chim. Acta.*, 2010, **363**, 2881–2885.
- 63 L. Xu, Y. Y. Zhao, G. Long, Y. Wang, J. Zhao, D. Li, J. Li, R. Ganguly, Y. Li, H. Sun and X. W. Sun, *RSC Adv.*, 2015, **5**, 63080–63086.
- 64 R. Das, D. Guha, S. Mitra, S. Kar, S. Lahiri and S. Mukherjee, *J. Phys. Chem. A*, 1997, **101**, 4042–4047.
- 65 C. V. Kumar and L. M. Tolosa, *J. Chem. Soc. Chem. Commun.*, 1993, **78**(1), 722–724.
- 66 A. Prakash and D. Adhikari, *Int. J. ChemTech Res.*, 2011, **3**, 1891–1896.
- 67 A. Kajal, S. Bala, S. Kamboj, N. Sharma and V. Saini, *J. Catal.*, 2013, **2013**, 1–14.
- 68 K. Brodowska and E. Lodyga-Chruscinska, *Cheminform*, 2015, **46**, 129–134.
- 69 J. C. Qin, T. R. Li, B. D. Wang, Z. Y. Yang and L. Fan, *Synth. Met.*, 2014, **195**, 141–146.
- 70 S. Halder, S. Dey and P. Roy, *RSC Adv.*, 2015, **5**, 54873–54881.
- 71 K. Grychowska, G. Satała, T. Kos, A. Partyka, E. Colacino, S. Chaumont-Dubel, X. Bantreil, A. Wesołowska, M. Pawłowski, J. Martinez and P. Marin, *Chem. Neurosci.*, 2016, **7**(7), 972–983.
- 72 A. L. Bédé, A. B. Assoma, K. D. Yapó, M. G. Koné, S. Koné, M. Koné, B. R. N'Guessan and E. H. Bamba, *Comput. Chem.*, 2018, **6**(03), 57.
- 73 M. Rossi, J. Erlebacher, D. E. Zacharias, H. L. Carrell and B. Iannucci, *Carcinog*, 1991, **12**(12), 2227–2232.
- 74 M. Z. Wang, J. Gao, Y. Chu, J. Niu, M. Chen, Q. Shang, L. H. Peng and Z. H. Jiang, *BMC Complementary Med. Ther.*, 2020, **20**, 1–8.
- 75 M. Frisch, G. Trucks, H. Schlegel, G. Scuseria, M. Robb, J. Cheeseman, G. Scalmani, V. Barone, B. Mennucci and G. Petersson, *Gaussian09, Revision D. 01*, Gaussian, Inc., Wallingford CT, 2009, <http://www.gaussian.com>.
- 76 R. Dennington, T. Keith and J. Millam, *GaussView 5.0*. Gaussian. Inc., Wallingford, 2008.
- 77 B. Kosar and C. Albayrak, *Spectrochim. Acta, Part A*, 2011, **78**(1), 160.
- 78 A. Mermer, H. Bayrak, S. Alyar and M. Alagumuthu, *J. Mol. Struct.*, 2020, (1208), 127891.
- 79 P. Subhapiya, K. Sadasivam, M. L. Madhu Mohan and P. S. Vijayanand, *Spectrochim. Acta, Part A*, 2014, **123**, 511–523.
- 80 K. burke, *J. Chem. Phys.*, 2012, (15), 136.
- 81 L. R. Domingo, M. Ríos-Gutiérrez and P. Perez, *Molecules*, 2016, **21**(6), 748.
- 82 A. E. Reed and F. Weinhold, *J. Chem. Phys.*, 1985, **83**(4), 1736–1740.
- 83 C. B. Santos, C. C. Lobato, M. A. de Sousa, W. J. Macedo and J. C. Carvalho, *Rev. Theor. Sci.*, 2014, **2**, 91–115.
- 84 D. Pankin, M. Smirnov, A. Povolotckaia, A. Povolotskiy, E. Borisov, M. Moskovskiy, A. Gulyaev, S. Gerasimenko, A. Aksenov and M. Litvinov, *Materials*, 2022, (15), 649.
- 85 C. Adamo and D. Jacquemin, *Chem. Soc. Rev.*, 2013, **42**, 845–856.
- 86 R. D. Garcia, V. G. Maltarollo, K. M. Honorio and G. H. Trossini, *J. Mol. Model.*, 2015, (21), 150.
- 87 B. Czaplińska, K. Malarz, A. Mrozek-Wilczkiewicz, A. Korzec and R. Musiol, *Molecules*, 2020, **25**(11), 2488.

