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Hydrothermal synthesis of modified lignin-based carbon dots derived from biomass waste for fluorescence determination of valsartan

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Recently, carbon dots (CDs) have been extensively investigated as potential tools for numerous applications. Modified lignin-based CDs have been synthesized and used in the field of drug detection. They were found to be highly selective and sensitive to valsartan (VAL). Using a simple hydrothermal method, phosphorus and chlorine co-doped CDs were synthesized using lignin extracted from date seeds. The fluorescence properties of the synthesized CDs are influenced by several factors, which were investigated in detail. The optimal synthesis conditions were 1.50 g of lignin, 18 mL of 2 M NaOH, 1 mM H_3PO_4, 3 mM HCl and the mixture was heated at 220 °C for 16 hours. The synthesized lignin-based CDs have excellent FL properties and are well soluble in water with reasonable stability. Characterization of the prepared CDs revealed that they have various functional groups with a graphene oxide-like structure. The developed CDs show a good quantum yield of 37.7%. The FL of the CDs is quenched by VAL at λ_{em} 313 nm after λ_{ex} at 275 nm by a combination of static and dynamic quenching mechanisms. The response of VAL was linear in the range of 4.0–100.0 μg mL^{-1}. The detection and quantification limits of VAL were 1.23 and 3.71 μg mL^{-1}, respectively. The nanoprobe was successfully used to analyze VAL in drug samples and provided satisfactory results.

1. Introduction

Carbon dots (CDs), which are spherical, zero-dimensional carbon-based substances with diameters of less than ten nanometers, are also referred to as carbon quantum dots in some literature studies. CDs are a unique type of "core–shell" nanostructure, usually consisting of a conjugated sp^2 core and containing groups based on carbon, nitrogen or oxygen.[1] It is important to note that, unlike conventional quantum dots, CDs are usually not crystalline; instead, amorphous CDs are usually formed when organic precursors carbonize. The main building blocks of CDs are carbon sources that have undergone various forms of surface passivation through functionalization or modification.[2]

Considerable progress has been made in the development of synthesis strategies, the mechanistic understanding of CDs, and the investigation of their potential uses. CDs are doped with heteroatoms, such as oxygen, nitrogen, sulfur, phosphorus, and boron, depending on the initial material used in their fabrication. Furthermore, the surface of CDs has been modified with polymers to introduce functional groups onto it.[3]

The properties of CDs include photophysical and photochemical stability comparable to that of widely recognized and commercially available quantum dots. They also have the

significant advantages of being readily soluble in water and exhibiting minimal toxicity, no more toxicity than the chosen molecules for surface functionalization, and favorable biocompatibility. In addition, CDs have been shown to exhibit highly tunable fluorescence (FL) emission and excitation as well as nonblinking FL emission.[4] Due to their outstanding properties, CDs have been explored for a wide range of attractive and/or novel applications. Some examples of these applications include photocatalysis,[5] bioimaging,[6] FL probing and analysis.[7]

Recently, numerous techniques have been developed for the production of CDs. These include laser ablation,[8] chemical oxidation,[9] electrochemical exfoliation,[10] arc discharge,[11] microwave pyrolysis,[12] combustion/thermal methods,[13] solvothermal methods,[14] and hydrothermal synthesis.[15]

Among these techniques, hydrothermal synthesis is a straightforward approach with no special equipment requirements and is therefore an interesting and affordable method for synthesizing CDs.[16] The preparation of CDs using a hydrothermal method offers the advantages of being nontoxic and inexpensive. In general, the aqueous solutions of the mixtures were sealed with Teflon in an oven and subjected to a hydrothermal reaction at high pressure and temperature.[17]

Biomass is an effective carbon source for the production of biomass-based CDs. Utilizing renewable biomass sources for the production of CDs is helpful in converting low-grade biowaste into useful and valuable materials. Moreover, biomass-based carbon sources are more environmentally friendly than

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other carbon sources. Biomass containing heteroatoms is the most effective starting material for the production of CDs, unlike CDs from artificial carbon sources, which require the use of external reagents.[18]

Lignin is one of the most abundant biobased aromatic macromolecules, and is a renewable and green raw material supplied by all agricultural and forestry plants. It can be obtained from lignin-containing biomass waste such as wood, some plant peels, seeds, *etc.*, or can be obtained as a byproduct from the black liquor produced during paper pulping.[19] Date seeds consist mainly of cellulose, hemicellulose and lignin. Depending on the type of date seed, the lignin content can be between 21 and 24%.[20]

Date seeds were selected as a source of lignin because they are abundant as agricultural material in Saudi Arabia, and there are no studies on their use as a source of CDs. Lignin is a perfect carbon source for CDs due to its aromatic conjugate structure, high carbon content, naturally occurring chemical modification and heteroatom doping.[21]

Lignin-based CDs can be synthesized directly from alkali lignin by hydrothermal treatment and doped with various heteroatoms to improve their optical properties.[22] Chen *et al.* fabricated CDs with particle sizes ranging from 2 to 10 nm using lignin and H_2O_2.[23] Jiang *et al.* synthesized lignin-based CDs with a FL quantum yield (QY) of 7.95% by subjecting a mixture of lignin, lactic acid, and betaine to a temperature of 300 °C.[24] Shi *et al.* used alkali lignin, formaldehyde, and ethylenediamine to synthesize lignin-based CDs with an 8.1% FL QY under a nitrogen environment at 300 °C.[25] It is obvious that the FL QY of the reported lignin-based CDs was quite low, and additional research is required to explore their functionalization. A newly developed method for the preparation of lignin-based CDs with a high FL QY (43.9%) was developed by Sun *et al.*[16] The production of lignin-based CDs with environmentally friendly modifiers that enable a reasonable QY is therefore still a challenge.

One of the most significant applications of CDs is as fluorescent sensors, which are used to detect metal ions, anions and molecules. In most cases, the ions or molecules quenched the FL emission of the CDs. Compared to other ions, Fe is one of the most frequently detected ions due to its efficient combination with CDs.[26] Moreover, other anions, such as CrO_4^{2-} and ClO^-,[27] can be detected by FL emission signal quenching of CDs. Furthermore, some molecules and drugs can be sensitively detected by interactions with CDs, such as vitamin B12,[28] ascorbic acid,[29] zoledronic acid,[30] baicalin,[31] tamsulosin hydrochloride,[32] and cholesterol.[33]

Chronic high blood pressure is a common age-related disease that can debilitate the heart and kidneys. The disease also affects the large arteries, microcirculation, endocrine components and central nervous system. Treatment with antihypertensive drugs reduces the problems associated with hypertension.[34] The angiotensin II receptor blocker (ARB) valsartan (VAL) works by blocking an internal chemical that constricts blood vessels. VAL lowers blood pressure and relieves blood vessels. The heart is supplied with more oxygen and blood when blood pressure is lowered.[35] The detection of VAL is crucial in various domains of society and can be utilized to further evaluate a patient seeking assessment. Numerous techniques (spectroscopic, electrochemical and chromatographic methods) have been documented for the quantification of VAL in biological fluids, pharmacological preparations and bulk forms.[36–38]

The aim of the present study was to evaluate the environmentally friendly synthesis of CDs using lignin extracted from date seeds as a precursor under different conditions. Simple and accurate fluorometric probe is proposed in the presence of the prepared CDs for the determination of VAL in bulk powder and in their commercial products. The fluorometric method is performed by measuring the influence of the added CDs on the intensity of the peaks of the proposed method. It should be mentioned that the developed nanoprobe is simple to make, highly selective, and environmentally benign. Furthermore, it relies on uncomplicated and easily obtainable tools, harmless substances, and gentle synthetic conditions. In addition, compared to other electrochemical or chromatographic procedures, it is also accurate and precise and does not require considerable technical expertise.

2. Experimental

2.1. Materials and chemicals

Pure-quality reagents and chemicals were purchased from commercial suppliers and utilized without additional purification. Sulfuric acid (H_2SO_4, 95–98%), hydrochloric acid (HCl 37%), phosphoric acid (H_3PO_3, 85%), and sodium hydroxide (NaOH, 99.0%) were obtained from Sigma-Aldrich, (Hamburg, Germany). Ethanol (99.9%), sodium dihydrogen phosphate (99.5%), and citric acid were obtained from BDH Laboratory supplies (London, UK). Lignin was isolated from date seeds (Al-Qassim, Saudi Arabia). Tabuk Pharmaceutical Co. (Tabuk, Saudi Arabia) provided the pure form of valsartan. Pharmaceutical preparations, specifically Diovan® 160 mg VAL per tablet manufactured by Novartis Pharmaceutical Corporation in Switzerland, were acquired from local medication stores in Riyadh, Saudi Arabia. Deionized (DI) water was used to prepare the solutions and during the experiments. Polyethersulfone (PES) membrane filters with a pore size of 0.22 μm and dialysis tubing with a cutoff molecular weight of 1000 Da (MD44) were obtained from Real Laboratory Supplies Store, China.

2.2. Instruments

Multiple spectroscopic and microscopic approaches were employed to validate the synthesis of CDs. Photoluminescence spectroscopy, Fourier transform infrared (FTIR) spectroscopy, UV-vis spectroscopy, energy dispersive X-ray analysis (EDX), and X-ray diffraction (XRD) were performed using a RF-5301pc Shimadzu luminescence spectrometer with a 150 W xenon arc lamp, a PerkinElmer FTIR spectrophotometer, an Ultrospec 2100 Biochrom spectrophotometer, a JSM-7610F JEOL EDX, and a Siemens D-5000 diffractometer, respectively. The synthesized CDs were analyzed using a transmission electron microscope (TEM-1400, JEOL Ltd) and a scanning electron microscope

(SEM-2100F, JEOL Ltd) to obtain microscopic information. The pH measurements were conducted with a pH-meter (HANNA-Instruments HI 2211, Italy).

2.3. Extraction of lignin from waste date seeds biomass

The date seeds (DS) were washed with DI water to remove dirt particles and then dried in a closed room at room temperature. These seeds were ground using a high-quality stainless-steel mill until they yielded a brown powder, which was sieved to obtain a particle size of approximately 177 microns, then washed with DI water, oven-dried and stored in a glass bottle in a desiccator for further use.

Lignin was extracted from DS by a combination of ultrasonic irradiation and chemical methods at high temperatures and alkaline concentrations to shorten the extraction time,[39] followed by the precipitation of lignin under acidic conditions.[40]

Lignin was first extracted from DS (10 g) in 200 mL of 2 M NaOH solution at 80 °C for 1.5 h under sonication. Then, the mixture was stirred for another 1.5 hours at 90 °C. After the mixture cooled to room temperature, it was vacuum filtered and washed with 0.1 M NaOH to obtain the residual lignin on the cellulose surface. The hemicellulose was then isolated from the filtrate by precipitating the acidified filtrate in ethanol. Dilute sulfuric acid (20% v/v) was slowly added to the black filtrate with stirring to lower the pH.

At pH 6, the color of the solution obviously changed from black to cloudy brown. The mixture was stirred for 20 minutes to promote the growth of the precipitating particles. Then, it was transferred to a water bath at 65 °C and stirred for 45 minutes with the dropwise addition of an ethanol solution (50% v/v) as co-solvent. The solution was then centrifuged (4500 rpm) for 30 minutes. The filtrate, which was rich in lignin, was collected. The ethanol in the filtrate was removed using a rotary evaporator (60 °C). The alkali-soluble lignin was recovered by precipitation at pH 1.5 with sulfuric acid (20% v/v). The precipitated lignin was then centrifuged and washed with dilute acid (0.01 M) to remove the coprecipitated impurities. The resulting alkali lignin was then dried at 60 °C for 48 hours before further use.

2.4. Preparation and optimization conditions of lignin-based CDs

Approximately 1.0 g of alkali lignin was dispersed in 18 mL of different solvents (water, 0.1 M NaOH, DMSO, 60% ethanol, and 80% acetone), and the mixture of each sample was subjected to sonication for 30 minutes. Subsequently, the mixtures were transferred to a Teflon-lined autoclave with a capacity of 25 mL and subjected to a temperature of 200 °C for 12 hours. Once the mixture had cooled to the temperature of the surroundings, it was subjected to centrifugation at a speed of 6000 rpm for 15 minutes. Next, the solution was filtered through a 0.22 μm PES membrane. The liquid portions were transferred to a dialysis bag with a molecular weight cutoff of 1000 Da and subjected to dialysis against DI water for 12 hours. The samples were then kept at 4 °C.

The effect of using different concentrations of NaOH was investigated. For this purpose, 1.0 g of alkali lignin was dissolved in 18 mL of different NaOH concentrations from 0 to 4.0 M. The mixtures of NaOH and alkali lignin were then mixed well, transferred to a 25 mL Teflon-lined autoclave and heated at 200 °C for 12 hours. After cooling to room temperature, the samples were acidified with HCl to pH = 2 to precipitate unreacted lignin, sonicated for 30 min, centrifuged (6000 rpm for 15 min), and filtered through a 0.22 μm PES membrane. The supernatants were transferred to a dialysis bag (MWCO 1000 Da), dialyzed against DI water for 48 hours to remove ultrasmall molecules and ions, and then stored at 4 °C.[41]

Different amounts of lignin ranging from 0.1 g to 3.0 g were dissolved in 18 mL of NaOH (2 M) and ultrasonicated for 30 minutes. The mixtures were then transferred to a 25 mL Teflon autoclave and heated at 200 °C for 12 hours. The mixtures were then cooled to room temperature, and the purification process was completed as previously described. Five samples of approximately 1.5 g of lignin were prepared at different temperatures (140, 160, 180, 200 and 220 °C) for 12 hours in a Teflon-lined autoclave using 18 mL of NaOH (2 M) as a solvent. The purification procedure was carried out as previously described.

To study the effect of time, five samples of 1.5 g of lignin were dissolved in 18 mL of NaOH (2 M) and treated in a Teflon-lined autoclave at 220 °C for 8–24 hours. After cooling to room temperature, purification was performed as previously described.

Varying amounts of each modifier (phosphoric acid or hydrochloric acid) were dissolved in NaOH (2 M), added to 1.5 g of lignin, and mixed with NaOH to a final volume of 18 mL. The suspension of each sample was sonicated for 30 minutes. The mixtures were then transferred to a 25 mL Teflon-lined autoclave and heated to 220 °C for 16 hours. After cooling to room temperature, the purification process was carried out as previously described. The solid products were obtained by freeze-drying for further characterization.

2.5. Quantum yield measurements

The relative quantum yield (QY) of the CDs can be calculated as follows (1):

$$\varphi_X = \varphi_{std} \times \frac{I_X}{I_{std}} \times \frac{A_{std}}{A_X} \times \left(\frac{\eta_X}{\eta_{std}}\right)^2 \tag{1}$$

where φ_x is the function that represents the FL QY and X are the CDs. The standard compound chosen was quinine sulfate, which was dissolved in a solution of 0.1 M H_2SO_4 ($\varphi_{std} = 0.54$). The refractive index, denoted by η, is equal to 1.33 for an aqueous solution. A represents the light absorption at the specific excitation wavelength of 350 nm. I represents the integrated FL intensity over the FL emission spectrum. To minimize the impact of reabsorption effects, the absorbance at an excitation wavelength of 350 nm was adjusted to a value of 0.1.[42]

2.6. VAL determination using modified CDs

The stock solution of the modified CDs was prepared by suspending 1.0 g of CDs in 100 mL of DI water. The resulting

suspension was sonicated for 10 minutes and stored in a refrigerator at 4 °C. A stock solution (1000 μg mL^{-1}) of pure VAL was prepared by dissolving 0.1 g in 100 mL of ethanol. Aliquots of VAL (4.0–100.0 μg mL^{-1}) were added to a series of 5 mL volumetric flasks. Approximately 1.5 mL (pH 8) of citrate–phosphate buffer was added to 0.10 mL of CDs for the VAL solutions and made up to the mark with DI water. The FL spectra of the prepared solutions were measured at λ_{em} 313 nm after λ_{ex} at 275 nm (ex : em slit = 3 : 5) after 5 min at 10 °C.

Working solutions of VAL medicinal products were prepared by finely grinding 10 tablets of Diovan® 160 mg VAL per tablet. An equivalent amount (0.1 g) of each powder was diluted with 100 mL of DI water, sonicated for 10 minutes and then filtered. Analytical working samples were obtained by further dilution with the same solvent. The nominal content of the drug under study was estimated using a regression equation.

3. Results and discussion

3.1. Synthesis of lignin-based CDs

CDs were synthesized by a simple hydrothermal method using lignin extracted from DS as a carbon source and heteroatoms as modification precursors. Lignin was chosen as the starting material because it is the second most abundant component of plant material, and this biopolymer is rich in hydroxyl groups, which improves the FL emission of hydroxyl group-coated CDs. Scheme 1 shows the overall steps for the formation and purification of CDs. The synthesized CDs exhibit excellent dispersibility and high stability in water.

The mechanism of the synthesis of CDs from lignin can be explained by four main steps (Scheme 2): depolymerization, carbonization, passivation and surface functionalization. Depolymerization occurs by splitting the lignin into smaller fragments. Further exposure to high temperatures and high pressure leads to carbonization (growth of CDs). Passivation is a necessary subsequent process, as pure carbon nanoparticles typically lack fluorescence. The chemical treatment of different compounds on the surface is a crucial stage in the manufacturing of CDs. This procedure involves applying an insulating layer to the surface, which reduces the impact of surface imperfections, trapping, and direct quenching caused
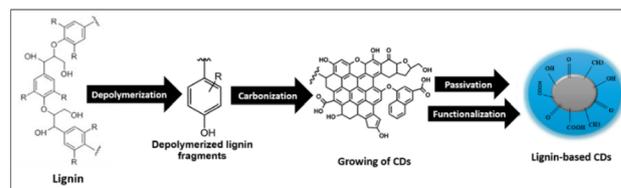

Scheme 2 The possible mechanism for the synthesis of lignin-based CDs.

by the environment. These factors contributed to an increase in FL emissions. Due to the absence of a passivating agent in this work, Bajpai *et al.* suggested that the residual polymer chains encased the carbon nanoparticles and produced hydroxyl groups on the surface, resulting in the generation of the FL.[43] Functionalization is the last necessary step in the production of carbon dots to ensure their effectiveness. The exceptional optical characteristics of CDs are a direct consequence of the passivation of carboxyl or hydroxyl groups by capping materials.[44]

Modified lignin-based CDs were successfully synthesized by hydrothermal treatment of two modifiers (HCl and H_3PO_4) with alkali lignin. The FL QY of the modified CDs was calculated using quinine sulfate as a reference at a λ_{ex} of 350 nm and was found to be 37.7%.

3.2. Effect of preparation conditions on FL intensity

The FL properties of the prepared lignin-based CDs are influenced by several factors, such as the type of solvent, the amount of lignin, the retention time, the hydrothermal temperature, and the amount of additive. The FL intensities were measured at an excitation wavelength of 350 nm to evaluate the FL performance of the CDs. Since alkali lignin is insoluble in water and exhibits good chemical stability under mild hydrothermal treatment, the carbonization process is expected to be limited in neutral aqueous medium. Therefore, the FL intensity is low, as shown in Fig. 1. Although a polar organic solvent (DMSO) can dissolve lignin, neglected CDs form, as seen in DLS (Fig. 2), with low FL intensity. The use of 60% ethanol or 80% acetone was not successful under the conditions used, despite the high FL
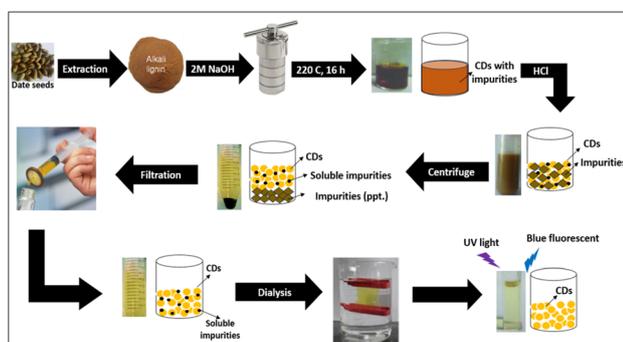
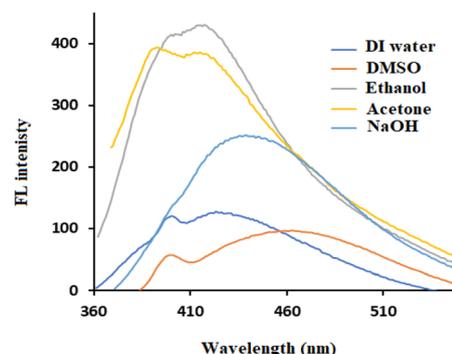Scheme 1 Schematic illustration of the formation and purification of CDs.

Fig. 1 FL spectra of lignin after hydrothermal treatment in different solvents at 200 °C for 12 h.

intensity (Fig. 1), but DLS showed that most of the particles formed had sizes of approximately 100–1000 nm (Fig. 2). The acid groups of the phenolic hydroxyl groups in lignin increase its solubility in aqueous sodium hydroxide (0.1 M NaOH), which is used as a solvent for lignin. The CDs prepared in this medium showed acceptable FL intensity with a particle size of approximately 10 nm (Fig. 1 and 2).

Different concentrations of NaOH were used to produce CDs, as shown in Fig. 3a. The greatest increase in the FL intensity clearly occurred when 2 M NaOH was used, and this concentration was chosen for further preparation of the CDs. TEM analysis (Fig. 3b) showed that when 2 M NaOH was used as a solvent, the hydrothermal treatment produced uniform particles in the size range of 3–5 nm. The hydroxyl group (OH^-) from NaOH contributes to the formation of a protective shell, which decreases the effective diameter of the CDs. As previously reported,[45] hydroxyl-coated CDs synthesized by hydrothermal reactions have a strong electron donating ability, with a positive effect on FL emission.

The FL property of CDs is affected by lignin dosage, as shown in Fig. 4a. The intensity of the emission peak of CDs increased when the mass of lignin increased from 0.25 g to 1.5 g, which was due to the increase in the concentration of CDs formed. No further improvement in intensity occurred when the amount of lignin increased to 3.0 g. This could be due to the limited reaction space in the 18 mL solution, which is consistent with the results of Zhu et al.[29] These results indicate that the lignin dosage is a crucial factor for the maximum emission peak.
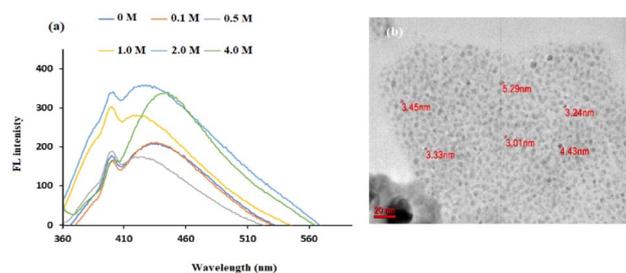

Fig. 3 (a) FL spectra of lignin after hydrothermal treatment with different concentrations of NaOH at 200 °C for 12 h (b) TEM image when 2 M NaOH was used as the solvent at 200 °C for 12 h.

The hydrothermal temperature of the synthesis plays an important role in the quality of the CDs produced. As shown in Fig. 4b, the lignin-based CDs were prepared over a wide temperature range from 140 to 220 °C. The maximum autoclave temperature (i.e., 220 °C) resulted in the highest photoluminescence signal. The enhancement of the FL signal with increasing temperature was due to increased functionalization of the CDs, which added more additional electrons to the π-system. As a result, at 220 °C, the CDs had more excited electrons at the π*-energy level than did the CDs at lower temperatures.[30]

The reaction time was adjusted from 8 to 24 hours to maximize the photoelectric performance. The color of synthesized CDs solutions became darker with increasing heating time from 8 to 16 hours, suggesting an increase in the CDs concentrations.
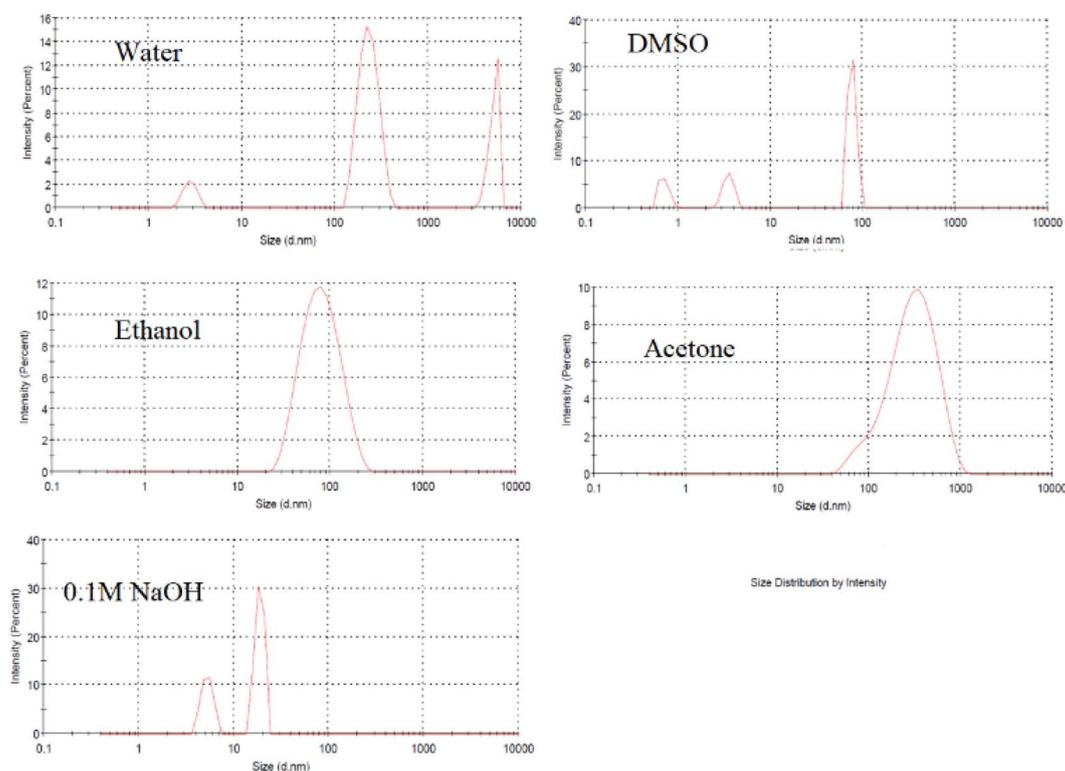

Fig. 2 Size distribution of lignin after hydrothermal treatment in different solvents at 200 °C for 12 h.
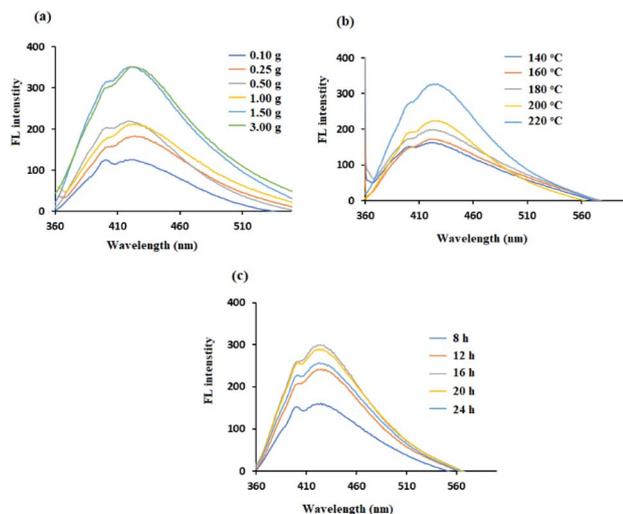
Fig. 4 FL spectra of (a) lignin-based CDs with different amounts of lignin at 200 °C for 12 h, (b) lignin-based CDs with different temperatures for 12 h, and (c) lignin-based CDs with different durations of hydrothermal treatment at 220 °C.

Thereafter, the color became lighter with longer heating times (16–24 h). This observation is consistent with the results in Fig. 4c, where the maximum increase in the FL intensity occurs at 16 hours. These results are in agreement with previous studies and suggest that inadequate reaction occurs during short heating periods, but excessive carbonization leads to the agglomeration of CDs at longer heating times.[31]

It was reported that the doping with chlorine can generate more energy levels, resulting in additional electron transitions in band structures of CDs. On the other hand, phosphorus was selected for modification by H_3PO_4 regardless of its FL intensity since the first experiments showed its sensitivity to the studied drug. Moreover, the combination of H_3PO_4 with HCl (to prepare PClCDs) reduces photobleaching compared to that of PCDs or ClCDs when H_3PO_4 or HCl is used alone, which was also reported by Wang *et al.*[46] The amounts of dopant precursors were varied to optimize the optical properties of the CDs. As shown in Fig. 5a, different amounts of H_3PO_4 were used, and as the amount increased, the FL decreased, possibly due to the decrease in pH, which affects the solubility of lignin. It also appears that the shift to longer wavelengths (from 425 to 447 nm) is due to the addition of acid as a result of new fluorescent
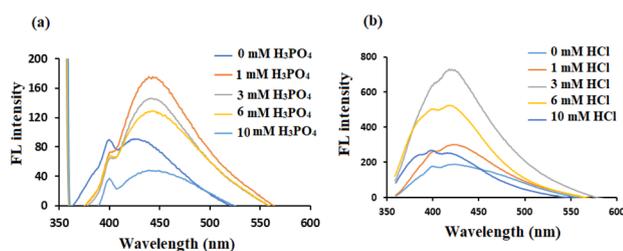
centers. The FL intensity increased when the amount of HCl increased from 0.00 to 3 mmol (Fig. 5b), which was selected as the optimal HCl dosage per 1.50 g of lignin. The FL intensity then decreased as the amount of HCl increased to 10 mM.

3.3. Stability of synthesized lignin-based CDs

The synthesized lignin-based CDs have outstanding FL characteristics and are highly soluble in water. To increase the range of their applications, it is necessary to look at their stability. The stability of the lignin-based CDs was thoroughly examined by observing their FL properties in a salt solution, at different pH values (ranging from 2 to 12, adjusted using 6 M of HCl and NaOH), under a UV lamp, and after extended periods of storage. As depicted in Fig. 6a, the FL of the CDs remains nearly constant when exposed to a high concentration NaCl solution (2.0 mol L^{-1}), indicating that the CDs are capable of withstanding high levels of ionic strength. The emission intensities of the CDs exhibit a high level of stability throughout the pH range of 5.0–9.0 (Fig. 6b) but decrease in strongly acidic or alkaline environments. The FL intensity decreases at high pH levels due to the increased proportion of negative charges and the inhibition of the π–π* and n–π* transitions.[5] Under acidic conditions, the presence of adjacent hydroxyl groups on the surfaces of CDs facilitates the formation of hydrogen bonds, leading to their aggregation. Consequently, the FL emission of CDs is suppressed.[47]

Following continuous exposure to a UV lamp for 120 minutes, as shown in Fig. 6c, there was no discernible alteration in the FL intensity of the synthesized CDs. This indicates that the CDs exhibit a high level of photostability. The CDs powder obtained by lyophilization can be easily dispersed in DI water without experiencing any aggregation. This property is advantageous for transportation and storage purposes. After being stored in a refrigerator at 4 °C for approximately 30 days, the CDs solution remained visually consistent, with no signs of

Fig. 5 FL spectra of modified lignin-based CDs with different amounts of dopant precursors (a) H_3PO_4 and (b) HCl.
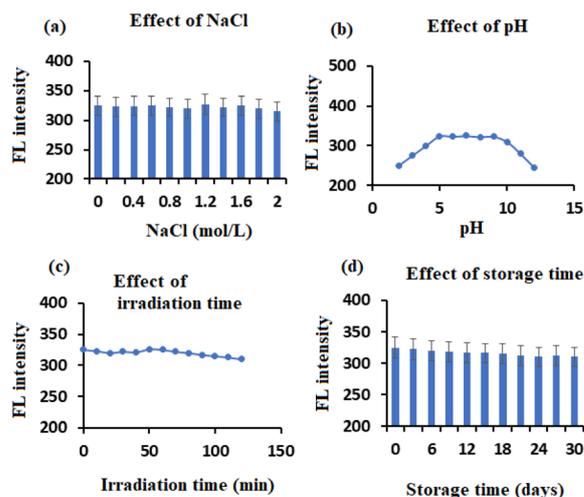

Fig. 6 Factors affecting the stability of synthesized lignin-based CDs: (a) effect of NaCl, (b) effect of pH, (c) effect of irradiation time, and (d) effect of storage time.

separation or precipitation. Additionally, the FL intensity remains nearly unaltered, as shown in Fig. 6d. The results provide strong evidence of the excellent stability of the synthesized lignin-based CDs, which is advantageous for future applications.

3.4. Characterization of the prepared CDs

The formation of PClCDs was confirmed using a variety of spectroscopic and microscopic techniques, including UV-vis spectroscopy, photoluminescence, FT-IR spectroscopy, particle size analysis, EDX analysis, XRD analysis, SEM, and TEM. Fig. 7a shows a TEM image of PClCDs. It can be clearly seen that the samples are uniformly dispersed and have no obvious aggregation. Considering that TEM can provide the most accurate result for determining the size of nanoparticles, TEM was used to measure the size of the synthesized CDs using ImageJ software. For this purpose, particle size distribution curves were generated from different TEM images, and a good number of particles were randomly selected. Fig. 7b shows that the particle size distribution of PClCDs is in the range of 1.58–10.60 nm, with an average particle size of 5.08 ± 1.60 nm obtained by statistically analyzing 300 randomly selected particles using ImageJ software and a Gaussian fit curve. The results indicated that the particle size distributions of the synthesized PClCDs were approximately 10 nm or smaller. This suggests that the PClCDs are in the form of carbon quantum dots, which exhibit a quantum size effect and a quantum confinement effect. These effects cause the energy band to change from continuous to discrete at the molecular level. In contrast, the solid-state of PClCDs was observed in the SEM image and AFM 3D image (Fig. 7c and d). It is well known that in nanomaterials such as carbon dots, conjugation leads to interactions between functional groups, which facilitates the formation of aggregates.[48]

UV-vis spectroscopic investigations were used to visualize the optical absorption behavior of the produced CDs. The studies were conducted at a fixed path length of 1.0 cm, covering a range of 200–700 nm. In general, the CDs exhibit
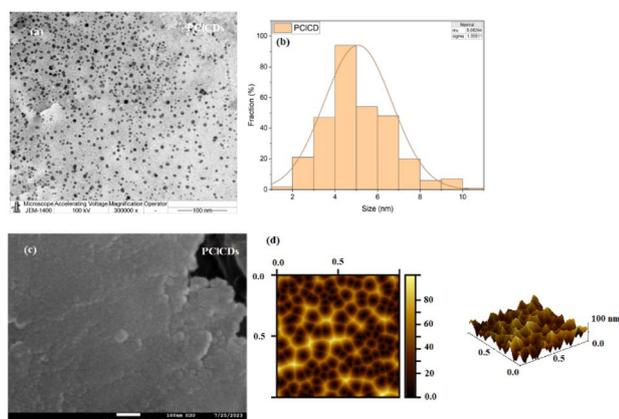

Fig. 7 (a) TEM image, (b) average particle size distribution using ImageJ software, (c) SEM image, and (d) AFM 3D image of PClCDs.
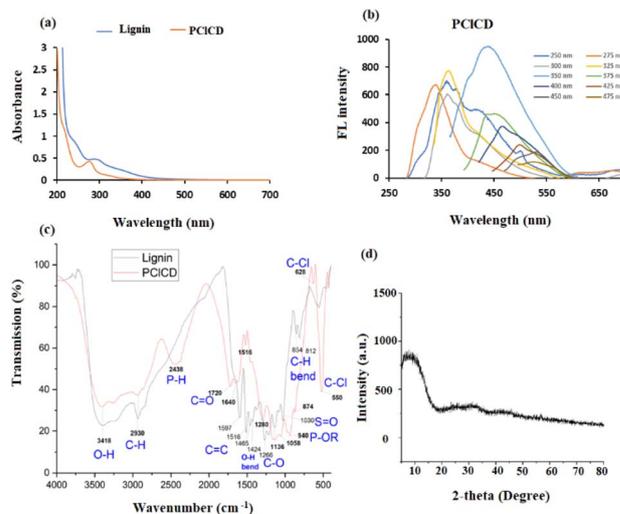

Fig. 8 (a) UV-vis absorption spectra of lignin and PClCDs, (b) FL spectra of PClCDs at different λ_{ex} values, (c) FT-IR spectra of lignin and PClCDs, and (d) XRD spectrum of PClCDs.

a largely unremarkable absorption profile, consistent with that of semiconductor nanoparticles with indirect bandgaps.[49] As shown by the orange lines in Fig. 8a, there is a distinct peak for PClCDs at approximately 275 nm, which is due to the $n \rightarrow \pi^*$ transition of the C=O band and the excited defect surface states induced by heteroatoms, as described in the literature.[50] The synthesized CDs exhibited a blue shift to 275 nm in comparison to the UV-vis absorption peak of pure lignin (blue line) at 295 nm. This blue shift was likely caused by the aromatic ring substituents of the CDs, which resulted in an increase in steric hindrance. Consequently, the level of conjugation is reduced, making electron transfer from n to π^* more difficult, so the absorption wavelength eventually decreases. In addition, there is a shoulder peak at 245 nm in lignin and a shift to approximately 222 nm in PClCDs, which is related to $\pi \rightarrow \pi^*$ of aromatic sp^2 hybridization in C=C bonds, which was also noted by Meng et al.[51] The blueshift of this absorption peak can be explained by the quantum confinement effect upon the addition of dopants.

The energy of the bandgap of the synthesized CDs was determined using the formula $E_g = h\nu$, where E_g is the bandgap energy, ν is the frequency and h is a blank constant. The band gap of PClCDs is 4.51 eV, which is comparable to the values in the literature.[52] The band gap of 4.51 eV at 275 nm is caused by the transition from the highest occupied molecular orbital (HOMO) to the lowest unoccupied molecular orbital (LUMO) of the surface state. On the other hand, the band gap of 5.58 eV at 222 nm is likely due to the transition from the HOMO to the LUMO of the carbon core.[53]

The photoluminescence (PL) behavior of the synthesized CDs, a classical feature of carbon dots, was characterized by FL spectroscopy. The PClCDs solutions were yellow in daylight and glowed blue when irradiated with UV light. The FL emission spectra of the CDs were analyzed at different excitation wavelengths from 250 to 475 nm in steps of 25 nm. Fig. 8b shows the

emission spectra of PClCDs at different excitation wavelengths. They exhibit blueshifts (with increasing excitation wavelengths from 250 to 275 nm) and redshifts (with increasing excitation wavelengths from 350 to 475 nm) and a decrease in the emission intensity with increasing excitation wavelength. This behavior was also observed by Hao *et al.*,[50] who explained that the excitation-dependent FL spectra might be related to the π* → n transition of the surface-bound functionalities, which is common in CDs. Moreover, the emission peak at 363 nm did not shift with increasing excitation wavelength from 300 to 325 nm, indicating excitation-independent emission at these wavelengths. The highest emission intensity of PClCDs was observed at an excitation wavelength of approximately 350 nm, while the excitation wavelength of 275 nm is in good agreement with their UV-vis spectra, implying that there are hardly any nonradiative transitions at this wavelength, and they could be used for further studies.

By comparing the QY of PClCDs at 350 nm to that of quinine sulfate (QY = 54% in 0.1 mol per L^{-1} H_2SO_4), it was determined that the QY of PClCDs is 37.7%. This value is greater than the QY obtained from synthesizing CDs using biomass waste.[54] Introducing electron-withdrawing atoms through doping greatly enhances the emission of FL. The P and Cl atoms exhibit strong electron-withdrawing properties, and the combined impact of codoping promotes the creation of bandgaps, leading to a high QY.

Fig. 8c shows the FTIR spectra of lignin and PClCD, which exhibit a broad absorption band at 3418 cm^{-1} corresponding to O–H stretching, indicating that hydroxyl groups were formed on their surfaces, increasing their hydrophilicity. The peak at 2930 cm^{-1} was attributed to C–H stretching in lignin and PClCD. In addition, the strong absorption peaks at 1720 (C=O stretching vibrations), 1640 and 1516 (C=C stretching vibrations), and 1280 and 1136 cm^{-1} (C–O stretching vibrations) indicate that the PClCDs terminated with carboxylic acid groups, further supporting their dispersibility in water. Two new absorption peaks appeared at 2438 cm^{-1} (stretching vibrations of P–H) and 940 cm^{-1} (stretching vibrations of P–OR), indicating successful doping of the CDs with P, while the peaks at 628 cm^{-1} and 550 cm^{-1} (stretching vibrations of C–Cl) indicated successful doping of the CDs with Cl.[46] All the previous results indicate that many functional groups are present on the surface of the synthesized PClCDs, leading to high hydrophilicity and stability in aqueous solution, as well as excellent FL performance, sensor specificity, and good biological compatibility. Moreover, the FTIR spectrum of the developed CDs strongly resembled that of lignin, which can be attributed to the fact that it was used as the starting material.

The crystallographic pattern of the synthesized CDs was analyzed by X-ray diffraction. Most of the prepared CDs show a broad diffraction peak at approximately ∼22°–25°, indicating the formation of amorphous graphitic carbon with an interplanar spacing (*d*-spacing) between 0.2–0.35 nm.[55] The interlayer spacing was calculated according to the Bragg equation as shown below (2 *d* Sin θ = *n*λ), where *n* is the order of reflection (1), λ is the wavelength (0.15418 nm is used in the present study), and *d* is the interplanar spacing. In Fig. 8d, the XRD

pattern of PClCDs shows a broad and weak peak at approximately 8.1°, indicating disordered carbon atoms with an amorphous character, and the position of the peaks can be attributed to the graphene oxide-like structure of PClCDs. Karimi *et al.* prepared GO, which showed an intense characteristic diffraction peak (001) at 2θ = 10.1°, and the GOQDs showed an intense peak (001) at 2θ = 7.3°;[56] these peaks could be shifted here due to the presence of functional groups on the PClCDs structure. The interlayer distance (*d*-spacing) of the PClCDs increased to 1.09 nm due to the GO-like structure, which provides greater uptake of oxygen species and functionalization of the sides and edges of GO sheets by oxygen-containing groups (carboxylic acids, alcohols, epoxides) or adsorbed H_2O or CO_2 molecules between two graphene sheets compared to graphite.[57]

XPS analysis was conducted to gain a deeper comprehension of the elemental composition and surface structure of PClCDs. Fig. 9a displays a survey spectrum of the XPS results of PClCDs, revealing four distinct peaks at 529.7, 283.9, 199.4, and 128.6 eV, corresponding to O 1s, C 1s, Cl 2p, and P 2p, respectively. Fig. 9b displays the C 1s spectra of the PClCDs, which can be categorized into three peaks situated at 289.7, 287.2, and 284.1 eV. These peaks correspond to the presence of C=O, C–O, and C=C bonds, respectively. This suggests that the surface of the developed PClCDs had many carboxylic acid, hydroxyl, and carbonyl groups, which aligns with the findings from the FTIR analysis. The O 1s spectrum (Fig. 9c) exhibits two distinct peaks at 533.2 and 530.8 eV, corresponding to the presence of *O–C=
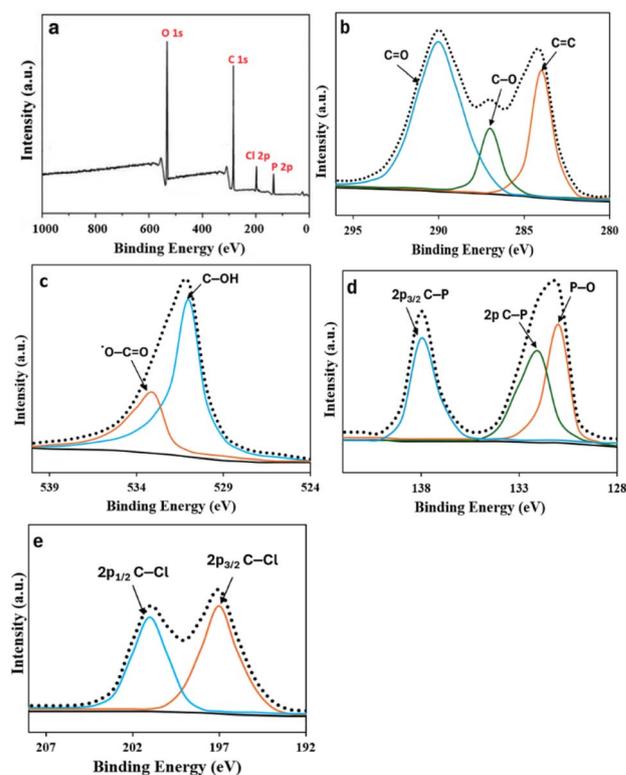

Fig. 9 (a) XPS survey scan, (b) C 1s, (c) O 1s, (d) P 2p, and (e) Cl 2p of PClCDs.

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O and C–OH, respectively. The P 2p spectrum (Fig. 9d) shows three peaks at 138.1, 132.3, and 130.9 eV, corresponding to $2p_{3/2}$ C–P, 2p C–P, and P–O, respectively. The Cl 2p spectrum (Fig. 9e) splits into two distinct peaks at 201.6 and 197.2 eV, corresponding to $2p_{1/2}$ C–Cl and $2p_{3/2}$ C–Cl, respectively.[58,59] In general, the PClCDs were effectively doped with P and Cl heteroatoms and surface-functionalized with many functional groups, enhancing their water solubility and stability.

EDX analysis was performed to determine the chemical composition of the synthesized PClCDs. According to the EDX spectrum, PClCDs contain a reasonable percentage phosphorus and chlorine in addition to carbon, oxygen, and a small amount of sulfur. The weight percentages and atomic percentages of the elements are shown in Fig. 10. The absence of any additional peaks belonging to other elements suggested that the synthesized CDs were pure. The results indicate a high oxygen content, which is consistent with the XRD results of the graphene oxide-like structure and the large d-spacing, which is due to the presence of large amounts of oxygen-containing functional groups originating from the starting material (lignin).

3.5. VAL determination

3.5.1. Spectral characteristics. The VAL drug shows an emission band at 385 nm after excitation at 275 nm. The addition of different concentrations of the VAL drug to PClCDs changed the FL intensity of the CDs to varying degrees, depending on the excitation and emission wavelengths. Notably, the difference in the FL intensity ($F_0 - F$) at an excitation wavelength of 350 nm was not significantly affected by the change in the VAL concentration (F_0 and F are the FL intensities of the CDs in the absence and presence of VAL, respectively). This could be because the groups on the CDs responsible for FL at the excitation wavelength of 350 nm do not interact with the VAL.[60] On the other hand, at an excitation wavelength of 275 nm, FL quenching of CDs emission occurred
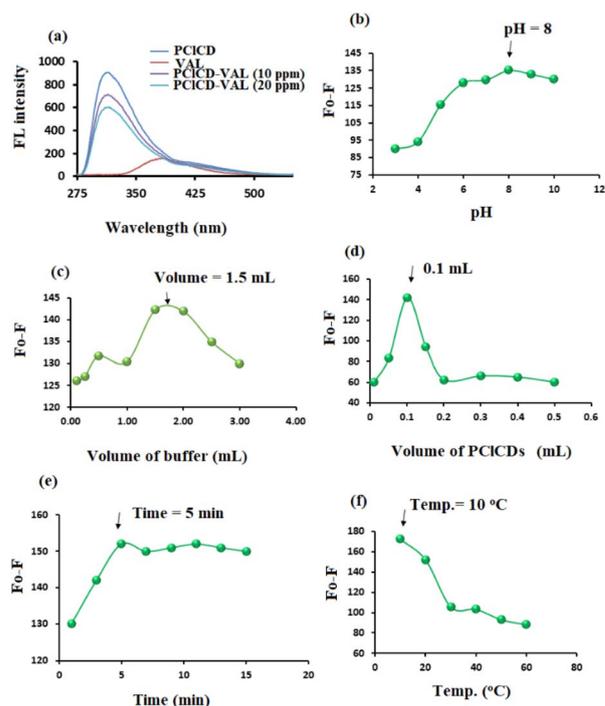

Fig. 11 (a) FL spectra of different concentrations of VAL solutions with and without the presence of PClCDs; effects of (b) pH, (c) buffer volume, (d) PClCDs volume, (e) response time and (f) temperature on the $F_0 - F$ of 10 μg mL^{-1} VAL in the presence of PClCDs.

gradually after the addition of various concentrations of the VAL drug (Fig. 11a). Therefore, 275 nm and 313 nm were chosen for the excitation and emission of the PClCDs-VAL system, respectively.

3.5.2. Optimization of reaction conditions. To improve the sensitivity, the experimental conditions for the proposed method were adjusted, including various parameters such as the effect of pH, the volume of buffer, the volume of CDs, the reaction time, and the temperature. The optimized conditions for the proposed fluorometric method are summarized in Table 1. The evaluation of the effect of pH in the range of 3–10 was performed in the presence of 0.10 mL of CDs and 10 μg per mL^{-1} VAL solution. The obtained results showed that the maximum $F_0 - F$ was observed when using citrate–phosphate buffer with a pH of 8 for PClCDs-VAL (Fig. 11b). In addition, the
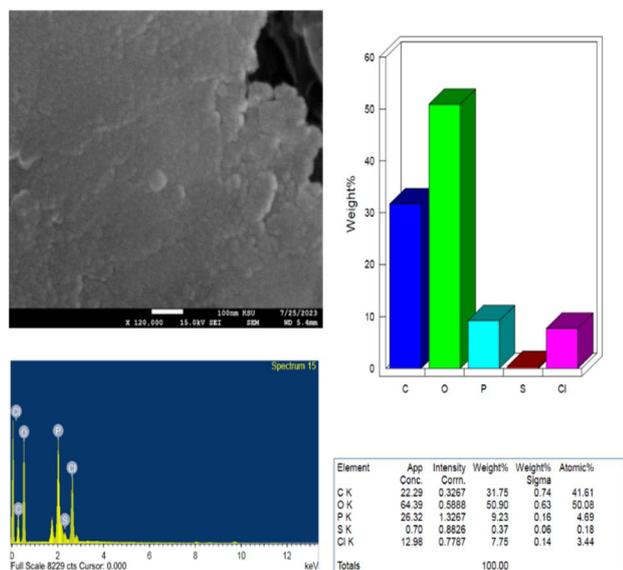

Fig. 10 EDX analysis of the synthesized PClCDs.

Table 1 Optimized parameters for measuring the concentration of VAL using the suggested spectrofluorometric technique in the presence of PClCDs

Parameter	Studied range	PClCDs-VAL
$\lambda_{ex/em}$ (nm)	250–800	275/313
ex : em slit	1.5–10 : 1.5–10	3 : 5
Buffer pH	3–10	8
Buffer volume (mL)	0.25–3.00	1.50
CDs volume (mL)	0.01–0.50	0.10
Time (min)	1–15	5
Temperature (°C)	10–60	10

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effect of buffer volume on the quenching of CDs by VAL was investigated in the range of 0.25 to 3.00 mL (Fig. 11c). The use of 1.50 mL of citrate–phosphate buffer gave the maximum $F_0 - F$ for the PClCDs-VAL system.

A suitable volume of PClCDs in the range of 0.01–0.50 mL was tested by using a 10 μg mL^{-1} VAL solution. The highest value of $F_0 - F$ was obtained after adding 0.10 mL of PClCDs to the VAL solution (Fig. 11d). Different time intervals (1–15 min) were used to study the effect of response time on the quenching of FL of CDs by the addition of 10 μg mL^{-1} VAL solution. Fig. 11e shows that the FL intensity of the PClCDs-VAL system gradually decreased with the addition of VAL. After 5 min, the FL intensity of PClCDs-VAL changed slightly and then tended to remain stable.

The effect of temperature in the range of 10–60 °C on the quenching of FL of CDs by VAL was investigated. The test was performed by cooling the samples and the blank in an ice bath or by heating in a water bath. The best quenching of FL of CDs by VAL occurred at 10 °C for the PClCDs-VAL system. As the temperature increased, the $F_0 - F$ values decreased (Fig. 11f).

3.5.3. Method validation. A method validation was performed to verify the suitability and validity of the suggested analytical approach for detecting the substance being studied with a high level of accuracy and precision. Calibration curve (Fig. 12) for the determination of VAL using the proposed spectrofluorometric system was constructed by plotting the FL difference $F_0 - F$ as a function of medication concentration. The linearity of the graph was observed within the concentration range of 4–100 μg per mL^{-1} for VAL in the presence of PClCDs. The regression equation of the linear relationship was $F_0 - F = 6.1793C + 114.51$. The statistical examination of the acquired results showed strong correlation coefficients (r) and low standard deviations for the intercept (S_a) and slope (S_b), indicating the outstanding linearity of the constructed graph (Table 2).

The LOD and LOQ for VAL determination in the presence of PClCDs using the suggested spectrofluorometric method were calculated using LOD = $3.3S_a$/slope and LOQ = $10S_a$/slope. The LOD was 1.23 μg mL^{-1}, and the LOQ was 3.71 μg mL^{-1} for the PClCDs-VAL system (Table 2).
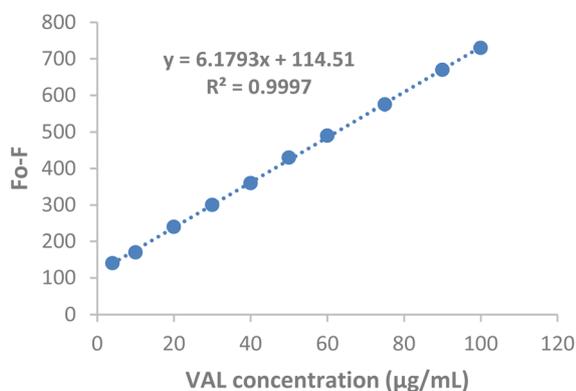
Table 2 The response data acquired by determining the VAL using the proposed spectrofluorometric method in the presence of PClCDs

Parameters	PClCDs-VAL
Concentration range (μg mL^{-1})	4–100
Slope	6.1793
Intercept	114.51
Correlation coefficient (r)	0.9998
Standard deviation of slope (S_b)	0.040
Standard deviation of intercept (S_a)	2.294
LOD (μg mL^{-1})	1.23
LOQ (μg mL^{-1})	3.71

The accuracy of the proposed spectrofluorometric method was evaluated using ten authentic VAL samples, and the results obtained were estimated as the mean recovery percentage ± standard deviation (mean ± SD). Excellent detection with high accuracy was observed, and the percent recovery was reported to be 100.55 ± 1.19% in the presence of PClCDs (Table 3).

The precision of the suggested approach was evaluated by both intraday and interday testing. The study examined three distinct concentrations of the drug, each tested three times ($n = 3$), and the resulting data were analyzed as a percentage of the relative standard deviation (RSD%). As summarized in Table 4, the calculated mean RSD% was 0.87% for the intraday determination of VAL in the presence of PClCDs. Nevertheless, the interday test demonstrated a mean RSD of 0.51% for the previous medicine in the presence of PClCDs. These results are less than 2% and demonstrate the high precision of the proposed method.

The robustness of the spectrofluorometric method for the determination of VAL was investigated by making small changes in the method parameters. The robustness of the methods was checked by changing the pH (±1.0) and the volume of the PClCDs (±0.01 mL). The $F_0 - F$ was not affected

Table 3 Accuracy data obtained from the determination of VAL in bulk powder using the proposed spectrofluorometric method in the presence of PClCDs

	Taken (μg mL^{-1})	Found (μg mL^{-1})	%Recovery
Statistical analysis	4	4.10	102.50
	10	9.98	99.80
	20	20.31	101.54
	30	30.02	100.06
	40	39.73	99.32
	50	51.06	102.11
	60	60.77	101.28
	75	74.52	99.36
	90	89.90	99.88
	100	99.61	99.61
Mean ± SD		100.55 ± 1.19	
n		10	
Variance		1.42	
%SE^a		0.38	
%RSD^b		1.19	

^{a} %SE = SD/$\sqrt{n}$. ^{b} %RSD = (SD/mean) × 100.

Fig. 12 Calibration curve for VAL determination in the presence of PClCDs.

y = 6.1793x + 114.51
R² = 0.9997

Table 4 Intra- and interday precision of the VAL assay by the proposed spectrofluorometric method in the presence of PClCDs

	Taken, $\mu g\ mL^{-1}$	% Recovery ± SD[a]	%RSD[b]	%Error[c]
Intraday precision	4	96.17 ± 1.26	1.31	0.76
	50	99.47 ± 1.03	1.03	0.60
	100	99.59 ± 0.28	0.28	0.16
Interday precision	4	98.67 ± 0.76	0.77	0.45
	50	98.95 ± 0.51	0.51	0.30
	100	99.59 ± 0.26	0.26	0.15

[a] Mean of three determinations. [b] %RSD = (SD/mean) ×100. [c] %Error = %RSD/$\sqrt{n}$.

by these changes. The calculated percent recovery was 99.26 ± 0.80% in the presence of PClCDs. The ruggedness of the spectrofluorometric detection method for the determination of VAL was investigated by measuring the same samples under different conditions, including in different laboratories, by different operators, and with different equipment. The $F_0 - F$ was not affected by these changes. The percent recovery was 99.73 ± 0.50% in the presence of PClCDs.

The selectivity of the proposed spectrofluorometric method for the determination of VAL was evaluated in the presence of different classes of antihypertensive drugs and also in the presence of various coadditives and possible interfering species in pharmaceutical preparations. Fig. 13 shows the changes in fluorescence intensity in the presence of the coadditive species in the PClCDs-VAL system. The results revealed no notable interferences; thus, the proposed spectrofluorometric system for the quantification of VAL can be considered as a selective method.

The proposed method was used to estimate the tested medication in its pharmaceutical preparation (Diovan® 160 mg VAL per tablet), and the results are showed in Table 5. The calculated results were 98.94 ± 0.80 in the presence of PClCDs. The results were examined by statistical analysis using Student's t-test and the variance ratio F-test. These results were then compared with the results of a previously published approach, as shown in Table 5.[61]

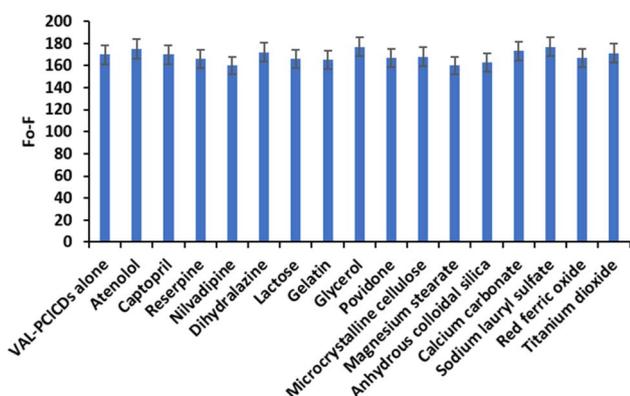The present spectrofluorometric method for VAL determination is superior or comparable to previously reported analytical methods (Table 6), such as spectrophotometric, fluorescence, electrochemical, and chromatographic methods. It should be noted that the developed probes are based on the use of simple and available instruments, nontoxic materials, and mild synthetic conditions.

3.5.4. Detection mechanism. The primary quenching process in sensing is the interaction of a quencher molecule with a fluorophore. This occurs as a result of either dynamic or static collisions between the quencher and the fluorophore. Static-based quenching inhibits the fluorophore from absorbing light and from transferring an electron to the excited state by forming a complex between the quencher and the fluorophore in the ground state (before excitation). On the other hand, when an excited fluorophore and a quenching molecule collide, dynamic quenching occurs, causing the fluorescent molecule to return to its ground state. To gain a deeper understanding of the fluorescence quenching mechanism, the fluorescence quenching data were examined using the Stern–Volmer (S–V) equation, which is shown as follows (2):

$$F_0/F = 1 + K_{SV}\ [Q] \tag{2}$$

where F_0 and F are the FL intensities of the CDs in the absence and presence of VAL, respectively. [Q] is the quencher (VAL) concentration, and K_{sv} is the Stern–Volmer quenching constant of the CDs.

Table 5 Application of the proposed and comparison methods for the determination of VAL in its pharmaceutical formulation (Diovan® 160 mg VAL per tablet) in the presence of PClCDs

	Taken ($\mu g\ mL^{-1}$)	%Recovery
Statistical analysis	4	97.50
	20	99.10
	40	98.58
	60	99.67
	80	99.38
	100	99.40
Mean ± SD		98.94 ± 0.80
n		6
Variance		0.63
%SE[a]		0.32
%RSD[b]		0.80
t-Test		2.06 (2.228)[c]
F-Test		3.76 (5.05)[c]
Reported		99.69 ± 0.41
Method[61]		(n = 6)

[a] %SE = SD/$\sqrt{n}$. [b] %RSD = (SD/mean) × 100. [c] The figures between parentheses are the tabulated t- and F values at $p < 0.05$.

Fig. 13 Selectivity of the PClCDs-VAL system for different species at a concentration of 10 μg per mL VAL.

Table 6 A comparative study between the results obtained from the determination of VAL utilizing spectrofluorometry in the presence of PClCDs and previously published analytical techniques

| Analytical techniques	Reagents	Linearity ($\mu g \ mL^{-1}$)	LOD ($\mu g \ mL^{-1}$)	Reference
Spectrophotometry	VAL and nebivolol dissolved in methanol	4–80	1.183	62
Fluorescence	VAL and amlodipine dissolved in methanol and HCl	0.25–4.50	0.04	63
Fluorescence	VAL and sacubitril dissolved in methanol	0.06–0.20	0.0126	64
Fluorescence	VAL, amlodipine and Fe_3O_4 nanoparticles	0.03–0.27	0.004	65
Fluorescence	VAL, metolazone and acetic acid as a solvent	0.01–0.1	0.003	66
Electrochemical	VAL, amlodipine and hydrochlorothiazide by graphene oxide-chitosan sensor	0.4–100.2	0.04	67
Chromatography	VAL and nebivolol in potassium hydrogen phosphate buffer, acetonitrile and methanol	48–112	0.78	68
Chromatography	VAL, acetonitrile and acetate buffer as a mobile phase	0.005–10	0.001	69
Chromatography	VAL, hydrochlorothiazide, irbesartan, acetonitrile and ammonium acetate buffer	0.05–6.02	0.050	70
Proposed method	Spectrofluorometric measurement in the presence of PClCDs	4.0–100.0	1.23	PClCDs-VAL
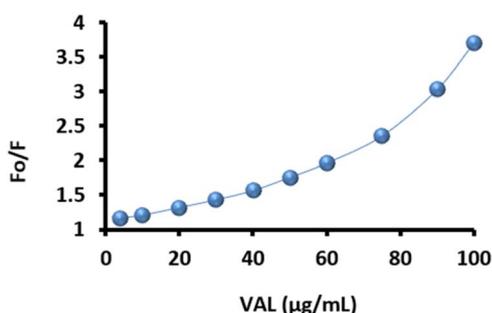
Fig. 14 Nonlinear Stern–Volmer plots for PClCDs-VAL system.

The S–V plots shown in Fig. 14 for the PClCDs-VAL system demonstrate that the FL intensities of the PClCDs gradually decreased (increased F_0/F) with the addition of VAL from 4–100 $\mu g \ mL^{-1}$. These results illustrate that VAL can effectively quench the FL of PClCDs. However, the S–V plots of the PClCDs-VAL system do not fit a conventional linear relationship between F_0/F and the concentration of VAL; thus, both static and dynamic quenching mechanisms were speculated to be responsible for the quenching process of these systems. According to Tanwar *et al.*, the presence of a second-order polynomial curve in the S–V plot indicates that both static and dynamic quenching mechanisms are present in any sensing system.[71] The S–V equation of the polynomial curve is given by (3):

$$F_0/F = (1 + K_{SV}[Q]) (1 + K_S[Q])$$
$$= 1 + (K_{SV} + K_S)[Q] + K_{SV}K_S[Q]^2 \quad (3)$$

where K_{SV} and K_S are the dynamic (S–V) and static quenching constants, respectively.

4. Conclusions

The present work reports a facile and eco-friendly approach for CDs synthesis using lignin extracted from date seeds as a precursor under different conditions and modifications.

Phosphorous and chlorine co-doped CDs derived from lignin are successfully obtained by a hydrothermal treatment in alkaline solution. In addition to being environmentally friendly and availability, low-cost biomass production offers greater benefits than other chemical approaches. The synthesized lignin-based CDs exhibit good fluorescence capabilities and demonstrate excellent solubility in water, along with satisfactory stability. The characterization of the produced CDs indicated the presence of several functional groups with a graphene oxide-like structure. The quantum yield and average size of the synthesized CDs were 37.7% and 5.08 ± 1.60 nm, respectively. The synthesized probe was used for sensitive and selective detection of VAL. The linear range and the detection limit were 4.0–100.0 $\mu g \ mL^{-1}$ and 1.23 $\mu g \ mL^{-1}$, respectively. This research is essential for expanding the exploration and use of fluorescent CDs in the field of drug detection.

Data availability

The data supporting the outcomes of this study have been incorporated within the manuscript.

Author contributions

All the authors contributed to the study conception and design; material preparation, data collection, and analysis were performed by Fatemah Aldakhil, Nawal A. Alarfaj, Salma A. Al-Tamimi, Maha F. El-Tohamy. The first draft of the manuscript was written by Fatemah Aldakhil. All the authors read and approved the final manuscript.

Conflicts of interest

There are no conflicts to declare.

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