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# Acetohydroxamic acid salts: mild, simple and effective degradation reagents to counter Novichok nerve agents†

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Novichoks is the latest known class of organophosphorus nerve agents to be developed. These highly lethal persistent agents, which exert their toxicity mainly through dermal exposure, pose new major challenges in mitigating their effect, mainly in respect to decontamination and medical countermeasures. Herein we report on the effective degradation of Novichok agents (A-230, A-232 and A-234) by hydroxamic acid salts. This class of  $\alpha$ -nucleophiles, with emphasis on the FDA approved drug acetohydroxamic acid, were found to promote rapid hydrolysis of these extremely toxic agents. Using  $^{31}\text{P}$  NMR the Novichoks degradation rates were determined to be in time scale of minutes with the following order of reactivity A-230>A-232>A-234. The degradation efficiency was found to be dependent on the nucleophiles, their counter-cations and the specific solvent mixture used. Hence, these scavengers can serve as efficient and mild decontaminants in various scenarios including surfaces, dermal decontamination (as an alternative to active lotions such as the RSDL® kit) and also as a medical countermeasure in the form of "catch-up therapy".

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## 1 Introduction

Novichok chemical warfare agents (CWAs), also named the A-series, were developed by the Soviet Union during the cold war (1970s–1990s) and are considered as the 4th generation of highly toxic organophosphorus compounds (OPs).<sup>1–3</sup> General public attention towards these agents was obtained, however, only during early 2018, after a Novichok agent was used in a poisoning incident in England.<sup>4</sup> This event, prompted a revision of the OPCWs' convention on the prohibition of the development, production, stockpiling, use and destruction of chemical weapons. Specifically, by 2020 these agents were explicitly added to schedule 1 of the annex of chemicals.<sup>5</sup> This fact, combined with the knowledge gained from the recent poisoning events regarding the extreme toxicity and stability of these agents, led to the realization that new countermeasures must be developed to effectively tackle Novichoks.<sup>6,7</sup> These include analytical detection,<sup>8–16</sup> decontamination (*vide infra*) methodologies and medical treatments. Structurally, Novichoks

resemble the G series more than the V-series of nerve agents, since both Novichoks and G agents possess a fluoride leaving group (Fig. 1), which is replaced by the hydroxyl of serine at the acetyl choline esterase (AChE) active site during the initial step of intoxication.<sup>17</sup> However, although they share a similar biochemical mechanism, as was published also in a recent case report,<sup>18</sup> G-agents' and Novichoks' hydrolytic properties were found to markedly differ, the latter being more similar to V-agents, in this respect.<sup>19,20</sup> Specifically, while G-agents rapidly

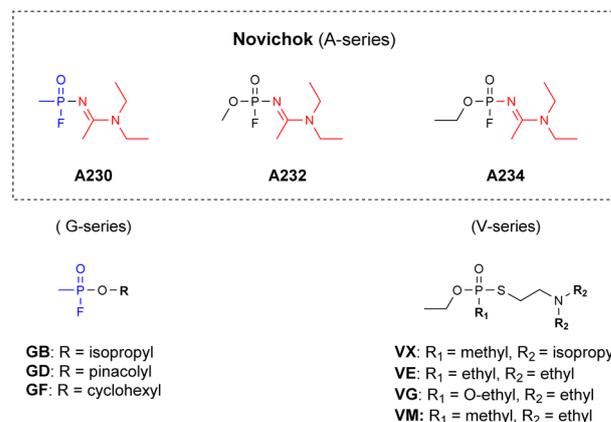


Fig. 1 The structure of the Novichok agents as compared to known CWAs (the unique amidine moiety is marked in red).

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hydrolyse in aqueous media even at a mildly basic pH (Sarin  $t_{1/2}$  = 5.5 min at pH 10,<sup>21</sup> Soman  $t_{1/2}$  = 12 min at pH 10),<sup>22,23</sup> Novichok agents are extremely stable at this pH (we measured the  $t_{1/2}$  to be  $\gg 110$  h at pH 11). In fact, their stability at neutral pH is even greater than that of VX and VR.<sup>19</sup>

As part of our ongoing research to develop CWAs decontamination methodologies, we were interested in developing effective detoxification agents that would be universal. Namely, effective towards all types of OP nerve agents including Novichoks. Specifically, we were interested in the decontamination of delicate surfaces such as electronic devices and most importantly human skin for which highly corrosive solutions, such as bleach or the organic decontamination solution DS2 cannot be implied.<sup>23–28</sup>

To the best of our knowledge, to date, only three reports studied Novichoks' stability towards hydrolysis and possible methods for their degradation (enzymatic, using MOFs or standard decontaminants) using the actual agents.<sup>19,20,29</sup> All reports were published after the Salisbury incident. Thus, there is still much to be done to thoroughly examine their effective decontamination using existing protocols, as well as to develop effective medical countermeasures against them.

In light of the above-mentioned stability of Novichoks, we envisioned that only strong nucleophiles in an appropriate solvent mixture would effectively react with the Novichok agents to provide efficient degradation. Recently, following a thorough investigation, we found that  $\alpha$ -nucleophiles like hydroxamic acid salts degrade VX effectively on time scale of a few minutes.<sup>30</sup> Moreover, we also demonstrated the therapeutic value and dermal and systemic safety of one of our lead lotions as a “catch-up therapy” against dermal intoxication by VX in live pigs.<sup>31</sup> Decades ago, these types of nucleophiles were already shown to effectively hydrolyze G-agents like sarin.<sup>32</sup> Herein we report our findings regarding the ability of hydroxamate salts to degrade Novichoks in a relevant time frame, which is applicable for both surface, dermal and transdermal detoxification. The study was conducted using <sup>31</sup>P NMR spectroscopy, which allowed an accurate time-dependent monitoring of the reaction progress in respect to the agents and their degradation products, thus enabling the extraction of the reactions' kinetic parameters.

## 2 Experimental section

Caution! OP's are extremely toxic compounds, with special emphasis on Novichok agents. Experiments with these compounds should only be performed by trained personal using applicable safety procedures.

### 2.1 NMR spectroscopy

<sup>31</sup>P{1H} spectra were obtained at 202 MHz at room temperature on a 11.7T (500 MHz) Bruker spectrometer (Avance III HD). Chemical shifts were calibrated to trimethyl phosphate as 0 ppm. The spectra were recorded using standard parameters of the TopSpin software (version 3.5). Each data point was obtained from 32 scans with a spectral width of 200 ppm and 4 s

recycle delay. All spectra were taken under identical conditions and integration was done automatically using TopSpin 3.5.2 software and the built in AU multi\_integ3.

### 2.2 Degradation kinetics

Hydroxamic acids (26 equiv.) and the appropriate base (KOH, NaOH, amine) were added to water, mPEG, DMSO or PG solution (1 mL) and stirred for 1 min until completely dissolved. Novichok agents (A230, A232 or A234) (2  $\mu$ L, 1 equiv.) were carefully added, and the solution was transferred to an NMR tube for kinetic analysis. Kinetic NMR acquisition started as soon as possible (without lock and shimming) until full degradation or no further change in reaction conversion were observed. Analysis of the results was performed by the GraphPad Prism 5 software to determine half-life times.

## 3 Results and discussion

This current study commenced by examining the Novichok decontamination activity of four selected hydroxamic acid derivatives, which were previously shown to effectively degrade VX and its simulant.<sup>30</sup> A-232, was chosen as the first agent to be examined. The hydroxamic acids, *i.e.*, acetohydroxamic, salicylhydroxamic, caprylhydroxamic and bufexamac (Table 1) were converted to the various salts using different inorganic and organic bases. Notably, all these compounds are FDA-approved for topical skin applications. In addition, the solvent mixtures tested were chosen based on common solvents used in pharmaceutical products. Specifically, the degradation reactions in this study were performed by dissolving 1 eq. of the Novichok agent in the selected solvent mixtures and adding it to a solution containing 26 eq. of the hydroxamate salt, in order to obtain pseudo-first order conditions. It should be noted that a much higher ratio is expected to be used in real-life scenarios. For example in our “catch-up therapy” protocol an excess of  $\sim 10^3$  of the AHA salt is used,<sup>31</sup> similar to the skin decontamination protocol using RSDL® (Reactive Skin Decontamination Lotion) where a  $\sim 1 : 10^3$  ratio is also desirable.<sup>33</sup> Such a high ratio should obviously further accelerate the reactions (see the kinetic equations below). Directly after mixing, the samples were consecutively examined using <sup>31</sup>P NMR spectroscopy to study the reaction's progress and the resulting kinetic data was analysed *via* integration using the TOPSPIN software and GraphPad Prism 5 (Fig. 2 and S1–S48†), to provide half-life times ( $t_{1/2}$ ) of these reactions, which are summarized in Table 1. In previous work we showed that adding up to 20% DMSO to the lotion dramatically enhances the skin penetration capability of the hydroxamic acid salts and also increases the solubility of the more lipophilic hydroxamate derivatives in the aqueous media, while still maintaining reasonable degradation rates of the persistent nerve agent VX.<sup>30</sup> Thus, lotions holding H<sub>2</sub>O/DMSO mixtures should be preferred for skin (including in-skin) decontamination.

As may be seen in Table 1, among the tested hydroxamic acid derivatives, salts of acetohydroxamic acid 1 and salicylhydroxamic acid 2 in DMSO, H<sub>2</sub>O and their mixtures were found to



**Table 1** Half-life time of A-232 degradation by the hydroxamate salts and solvent mixtures used in this study (a cut off of <10 min. was arbitrary chosen for the initial screening of the nucleophiles)

#	Compound name	Counter cation	Solvent	$t_{1/2}$ (min)
1	Acetohydroxamate (AHA)	Na <sup>+</sup> ( <b>1a</b> )	DMSO	30.4
		Na <sup>+</sup> ( <b>1a</b> )	DMSO/H <sub>2</sub> O 1 : 1	7.9
		Na <sup>+</sup> ( <b>1a</b> )	PG/H <sub>2</sub> O 1 : 1	28.1
		Na <sup>+</sup> ( <b>1a</b> )	mPEG/H <sub>2</sub> O 9 : 1	>> 300
		K <sup>+</sup> ( <b>1b</b> )	H <sub>2</sub> O	<2
		K <sup>+</sup> ( <b>1b</b> )	DMSO/H <sub>2</sub> O 1 : 1	7.4
		Diethylamine ( <b>1c</b> )	PG/H <sub>2</sub> O 1 : 1	36.4
		Diethylamine ( <b>1c</b> )	DMSO/H <sub>2</sub> O 1 : 1	41.7
		Olamine ( <b>1d</b> )	DMSO/H <sub>2</sub> O 1 : 1	128.0
		Epolamine ( <b>1e</b> )	DMSO/H <sub>2</sub> O 1 : 1	161.9
		Piperazine ( <b>1f</b> )	DMSO/H <sub>2</sub> O 1 : 1	124.3
2	Salicylhydroxamate (SHA)	2Na <sup>+</sup> ( <b>2a</b> )	DMSO	<3
		2Na <sup>+</sup> ( <b>2a</b> )	PG/H <sub>2</sub> O 1 : 1	16.4
		2Diethylamine ( <b>2c</b> )	DMSO/H <sub>2</sub> O 1 : 1	111.2
		2Olamine ( <b>2d</b> )	DMSO/H <sub>2</sub> O 1 : 1	N.R. <sup>c</sup>
		2Piperazine ( <b>2f</b> )	DMSO/H <sub>2</sub> O 1 : 1	N.R.
3	Caprylhydroxamate (CHA) <sup>a</sup>	Na <sup>+</sup> ( <b>3a</b> )	PG/H <sub>2</sub> O 1 : 1	~20 <sup>b</sup>
		Diethylamine ( <b>3e</b> )	DMSO/H <sub>2</sub> O 1 : 1	69.1
4	Bufexamac (BUF) <sup>a</sup>	Na <sup>+</sup> ( <b>4a</b> )	PG/H <sub>2</sub> O 1 : 1	~20 <sup>b</sup>

<sup>a</sup> CHA and BUF salts have a very poor solubility in DMSO and its mixtures with water, therefore the reaction with A232 couldn't be performed in these solvents. <sup>b</sup> Due to low solubility of the degradation products in the reaction mixture the reaction did not proceed to completion. <sup>c</sup> N. R. – no reaction.

be the most effective, rapidly degrading A-232 with a  $t_{1/2}$  of a few minutes (**1a-b** and **2a**). The degradation of A-232 by organic salts of **1**, namely, diethylamine, olamine, epolamine and piperazine (**1c-f**) respectively, was however much slower, with half-life values ranging between 36 and 162 min, while organic salts of **2**, olamine (**2d**) and piperazine (**2f**) didn't affect A-232 degradation at all. Among the organic salts examined in a DMSO/H<sub>2</sub>O mixture, diethylammonium salt of AHA (**1c**) showed the best activity. Surprisingly, the degradation of A-232 using the AHA Na salt (**1a**) in a mPEG/H<sub>2</sub>O 9 : 1 mixture, a solution used in the RSDL lotion, proceeded extremely slowly with  $t_{1/2}$  of more than 300 min, while under the same conditions VX was degraded much faster, with a  $t_{1/2}$  value of 7.45 min.<sup>30</sup>

The more hydrophobic hydroxamic acid salts, Caprylhydroxamate **3a** and Bufexamac **4a**, were found to have poor solubility in DMSO and its mixtures with water, therefore the reaction of these derivatives with A-232 was performed in a PG/H<sub>2</sub>O 1 : 1 mixture to show that they are less active than inorganic salts of **1** and **2**, degrading A-232 with a  $t_{1/2}$  of ~20 min.

Next, we proceeded to determine the effectiveness of the best hydroxamic acid derivative, identified, to degrade other members of the Novichok family *i.e.*, A-230 and A-234. In this respect, the AHA salts (Na<sup>+</sup>, K<sup>+</sup> and DEA<sup>+</sup>) were found to be more suitable than the SHA (2Na<sup>+</sup> salt), although showing similar activity. This is because the latter acid derivative holds two labile protons, which need to be deprotonated in order to react efficiently with A-232, as ascertained from the salt partial stability in aqueous mixtures. Water or DMSO/H<sub>2</sub>O mixtures were selected as the solvents of choice as they provided the best kinetic results for A-232 degradation among the solvents and solvent mixtures evaluated.

A summary of the Novichok agents' degradation kinetics using AHA salts is presented in Table 2. As can be seen, in addition to three different AHA counter ions, namely, sodium, potassium and diethylamine, each experiment was also performed in three solvent conditions, *i.e.*, water and two DMSO/H<sub>2</sub>O mixtures at room temperature.

The experiments described reflect a significant difference in the stability of the three Novichok agents tested, under the conditions examined. The phosphonate derivative A-230 is degraded in less than a few minutes ( $t_{1/2} < 2$  min) regardless of the solvent or counter-cation used. The degradation of A-232 was also rather quick, albeit slower than that of A-230, specifically in the case of the AHA DEA salt and a mixture of DMSO/water 1 : 1. A-234 was found to be the most stable, with half-life values ranging between 8 and 140 min, depending on the counter-cation and the percent of DMSO in the solution. Interestingly, the same tendency of Novichok agents' susceptibility to hydrolysis was previously observed also at pH 7.2, where the hydrolysis rate of A-230 agent was the fastest between the Novichok agents and the hydrolysis rate of A-234 was 50 times slower.<sup>19</sup> Importantly, the degradation rates of all three agents, using a small and simple scavenger, such as the AHA salt, do not fall short of the rates obtained with complicated structures as Zr based MOFs (reported by de Koning *et al.*).<sup>20</sup> The nucleophile ionization state was found to have a significant impact on the effectiveness of the degradation reactions, as was nicely demonstrated in the A-234 experiments, and to a lesser extent in the A-232 experiments, Table 2. Specifically, as the percentage of DMSO in the solution mixture was increased the concentration of the ionized nucleophile decreased, leading to slower reaction rates. Importantly, this trend was observed only when



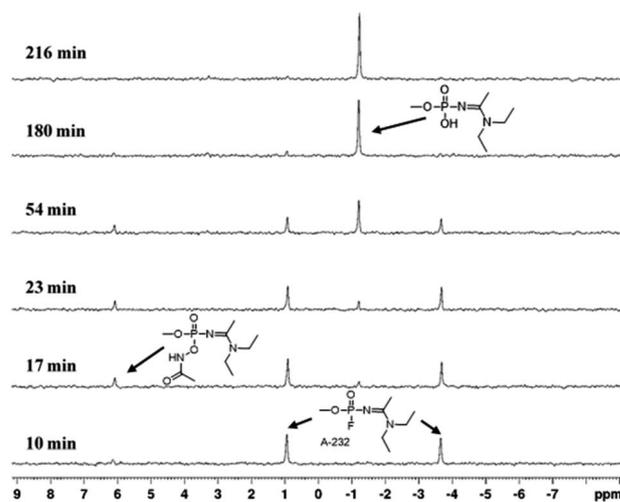
**Table 2** Half-life time of Novichok agents' degradation by acetohydroxamate salts using different cations and various solvent mixtures

Agent	AHA salt	% DMSO <sup>a</sup>	<i>k</i> <sub>obs</sub> (min <sup>-1</sup> )	<i>t</i> <sub>1/2</sub> (min)
A-230	AHA Na	0	— <sup>b</sup>	<2 <sup>b</sup>
	AHA Na	20	—	<2
	AHA Na	50	—	<2
	AHA K	0	—	<2
	AHA K	20	—	<2
	AHA K	50	—	<2
	AHA DEA	0	—	<2
	AHA DEA	20	—	<2
	AHA DEA	50	—	<2
A-232	AHA Na	0	0.1924	3.6
	AHA Na	20	—	<3.5
	AHA Na	50	0.0872	7.9
	AHA K	0	—	<2
	AHA K	20	—	<3
	AHA K	50	0.0938	7.4
	AHA DEA	0	0.1474	4.7
	AHA DEA	20	0.1440	4.8
	AHA DEA	50	0.0166	41.7
A-234	AHA Na	0	0.07825	8.86
	AHA Na	20	0.03905	17.75
	AHA Na	50	0.02068	33.51
	AHA K	0	0.08161	8.49
	AHA K	20	0.04566	15.18
	AHA K	50	0.02007	34.53
	AHA DEA	0	0.06982	9.93
	AHA DEA	20	0.03003	23.08
	AHA DEA	50	0.00489	141.88

<sup>a</sup> Percent of DMSO in a solution of DMSO/H<sub>2</sub>O (v/v). <sup>b</sup> Due to NMR time scale limitations, more accurate values of *t*<sub>1/2</sub> and *k*<sub>obs</sub> couldn't be determined.

degradation in water alone was not immediate, as inferred from the experiments with A-230.

A plausible degradation mechanism of Novichok agents by AHA salts is implied from the <sup>31</sup>P NMR spectra of the reaction progress. Specifically, in all experiments, the Novichok agent first appears as a distinct doublet stemming from the phosphorus–fluoride (P–F) coupling. As the reaction with the AHA nucleophile progresses, the doublet intensity decreases while two new signals arising from the degradation products appear. This pattern is nicely seen in Fig. 2, which presents <sup>31</sup>P NMR spectra of the reaction of A-232 with AHA DEA in DMSO/H<sub>2</sub>O 1 : 1, where A-232 signals appear at 0.97, –3.75 ppm, and the products signals at 6.2 and –1 ppm, while at the end of the reaction only the signal, at –1 ppm, remains. Initially, the AHA nucleophile attacks the phosphorous center to obtain an acetamidophosphoate intermediate, which is observed as an emerging signal at 6.2 ppm. Then, at a second step, this intermediate converts *via* Lossen rearrangement to the final stable product, phosphate acid, observed at –1 ppm, Scheme 1. The fact that the acetamidophosphoate intermediate is observed indicates that the Lossen rearrangement step is slower than the initial step of the nucleophilic attack. Although it is not clear which specific step of the Lossen reaction is decelerated, the deprotonation or the concerted dephosphorylation-alkyl migration, one may speculate on the former because the solution is



**Fig. 2** Representative <sup>31</sup>P NMR spectra presenting A-232 degradation by AHA DEA in DMSO/H<sub>2</sub>O 1 : 1, emphasizing the parent agent as a doublet which decomposes over time, giving rise to the degradation product.

only mildly basic (pH = 11). Importantly, this suggested degradation pathway of Novichoks, promoted by hydroxamic acid salts was previously reported for other organophosphorus nerve agents such as the persistent VX, Tabun and Sarin.<sup>30,34</sup> Notably, although toxicological data regarding Novichok degradation products has not been published, it is safe to assume that their toxicity would be drastically reduced compared to the parent agent, exactly as was found for other nerve agents (*i.e.*, ~10<sup>3</sup> decrease in lethality).<sup>35</sup>

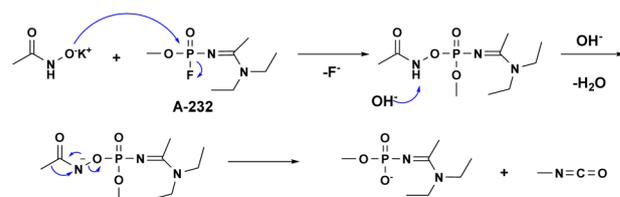
According to the suggested mechanism the reaction kinetics may be described by the following equation:

$$\text{Rate} = k_{\text{obs}} [\text{A-agent}] \quad (1)$$

where

$$k_{\text{obs}} = k_0 + k_{\text{OH}} [\text{OH}^-] + k_2 [\text{AHA}^-] \quad (2)$$

*k*<sub>0</sub> is the rate constant of the un-catalysed reaction in water, *k*<sub>OH</sub> is the second-order reaction rate between OH<sup>-</sup> and the A-agent, and *k*<sub>2</sub> is the enhancement resulting from the addition of the AHA nucleophile to the system.



**Scheme 1** Proposed mechanism for A-232 degradation by potassium acetohydroxamate through nucleophilic attack on the agent followed by Lossen rearrangement to form a non-toxic phosphoric acid derivative.



As the hydroxamate ion is a very strong nucleophile the agent's hydrolysis by the hydroxide ions present in the water basic solutions may be essentially neglected. Indeed, in a buffer of pH 11, *i.e.* a similar value to that of the AHA solutions, A-232 and A-234 were found highly stable with half-life exceeding 4 days (data not shown). Therefore, the rate constant,  $k_{\text{obs}}$ , can be expressed as:

$$k_{\text{obs}} = k_2 [\text{AHA}^-] \quad (3)$$

Moreover, it is assumed that the reaction kinetics is highly dependent on the ionization state of the active nucleophile in the solution, which is present in large excess and kept at a constant pH ( $\sim 11$ ). The addition of DMSO, an organic solvent, to the aqueous reaction mixture leads to an increase in the  $\text{p}K_{\text{a}}$  value of the acetohydroxamic acid (from 9.55 in case of 0% DMSO to 13.3 in case of 50% DMSO, as was previously shown).<sup>30</sup> This leads to a decrease in the concentration of the ionized nucleophile, and accordingly to a decrease in the reaction rate. Therefore, eqn (3) can actually be expressed by:

$$k_{\text{obs}} = k_2 [\text{AHA}]_t \times \alpha \text{AHA}^- \quad (4)$$

( $t$  – total concentration,  $\alpha$  – ionized fraction).

## 4 Conclusions

In conclusion, the experiments performed in this work demonstrate that AHA salts are effective nucleophilic scavengers that lead to fast and full Novichok agents' degradation. Our previous results demonstrated the therapeutic value and dermal and systemic safety of one of our AHA salts as a potential “catch-up therapy” against dermal intoxication by VX in live pigs.<sup>31</sup> Combined, the above suggest that DMSO/H<sub>2</sub>O solutions of AHA salts have a high potential to serve as a generic tool for the decontamination of low-volatility CWAs. Specifically, they may be useful for both skin surface decontamination and within-skin “catch-up therapy” (manuscript in preparation).

## Conflicts of interest

There are no conflicts to declare.

## Notes and references

- 1 V. S. Mirzayanov, *STATE SECRETS: An Insider's Chronicle of the Russian Chemical Weapons Program*, Outskirts Press, 2008.
- 2 M. Kloske and Z. Witkiewicz, *Chemosphere*, 2019, **221**, 672–682.
- 3 G. Bauer, A. Wildauer, G. Provoden, B. Menzi and C. Curty, *Forensic Sci.*, 2023, **3**, 231–244.
- 4 M. Peplow, *Chem. Eng. News*, 2018, **96**, 3–4.
- 5 *Schedule 1*, <https://www.opcw.org/chemical-weapons-convention/annexes/annex-chemicals/schedule-1>.
- 6 T. L. Blom and T. T. Wingelaar, *Mil. Med.*, 2023, usad464.
- 7 M. Hrabínova, J. Pejchal, V. Hepnarova, L. Muckova, L. Junova, J. Opravil, J. Zdarova Karasova, T. Rozsypal, A. Dlabkova, H. Rehulkova, T. Kucera, Z. Vecera, F. Caisberger, M. Schmidt, O. Soukup and D. Jun, *Arch. Toxicol.*, 2024, **98**, 1135–1149.
- 8 M. Otsuka, A. Yamaguchi and H. Miyaguchi, *Forensic Toxicol.*, 2023, **41**, 221–229.
- 9 J. Y. Lee, J. Y. Shin and H. S. Kim, *J. Anal. Toxicol.*, 2023, **47**, 81–88.
- 10 A. Yamaguchi, H. Miyaguchi and M. Tokeshi, *Anal. Chem.*, 2022, **94**, 4658–4665.
- 11 F. Mirbabaei, A. Mohammad-Khah, M. T. Naseri, M. Babri, S. M. Faraz, S. E. Hosseini and D. Ashrafi, *Anal. Bioanal. Chem.*, 2022, **414**, 3429–3442.
- 12 J. H. Lee, W. E. Jang, J. H. Park, H. B. Mohammad, J.-Y. Lee, W.-H. Jeong and M.-S. Kim, *Bull. Korean Chem. Soc.*, 2022, **43**, 444–449.
- 13 H. John, M. Dentzel, M. Siegert and H. Thiermann, *Anal. Chem.*, 2022, **94**, 2048–2055.
- 14 M. Eskandari, S. M. Faraz, S. E. Hosseini, S. Moradi and H. Saeidian, *Int. J. Mass Spectrom.*, 2022, **473**, 116794.
- 15 D. Noort, A. Fidler, D. van der Riet-van Oeveren, R. Busker and M. J. van der Schans, *Chem. Res. Toxicol.*, 2021, **34**, 1926–1932.
- 16 M. de Bruin-Hoegée, L. Lamriti, J. P. Langenberg, R. C. M. Olivier, L. F. Chau, M. J. van der Schans, D. Noort and A. C. van Asten, *Anal. Methods*, 2023, **15**, 142–153.
- 17 S. Costanzi, J.-H. Machado and M. Mitchell, *ACS Chem. Neurosci.*, 2018, **9**, 873–885.
- 18 D. Steindl, W. Boehmerle, R. Koerner, D. Praeger, M. Haug, J. Nee, A. Schreiber, F. Scheibe, K. Demin, P. Jacoby, R. Tauber, S. Hartwig, M. Endres and K.-U. Eckardt, *Lancet*, 2021, **397**, 249–252.
- 19 S. P. Harvey, L. R. McMahon and F. J. Berg, *Heliyon*, 2020, **6**, e03153.
- 20 M. C. de Koning, C. Vieira Soares, M. van Grol, R. P. T. Bross and G. Maurin, *ACS Appl. Mater. Interfaces*, 2022, **14**, 9222–9230.
- 21 R. L. Gustafson and A. E. Martell, *J. Am. Chem. Soc.*, 1962, **84**, 2309–2312.
- 22 J. R. Ward, Y. C. Yang, R. B. Wilson Jr. and W. D. Burrows, *Bioorg. Chem.*, 1988, **16**, 12–16.
- 23 Y.-C. Yang, J. A. Baker and J. R. Ward, *Chem. Rev.*, 1992, **92**, 1729–1743.
- 24 H.-J. Altman and A. Richardt, Decontamination of chemical warfare agents, in *Decontamination of Warfare Agents: Enzymatic Methods for the Removal of B/C Weapons*, ed. A. Richardt and M.-M. Blum, Wiley-VCH Verlag GmbH & Co. KGaA, 2008, ch. 7, pp. 83–115.
- 25 K. Kim, O. G. Tsay, D. A. Atwood and D. G. Churchill, *Chem. Rev.*, 2011, **111**, 5345–5403.
- 26 S. S. Talmage, A. P. Watson, V. Hauschild, N. B. Munro and J. King, *Curr. Org. Chem.*, 2007, **11**, 285–298.
- 27 A. W. Khan, S. Kotta, S. H. Ansari, J. Ali and R. K. Sharma, *Def. Sci. J.*, 2013, **63**, 487–496.
- 28 B. Picard, I. Chataigner, J. Maddaluno and J. Legros, *Org. Biomol. Chem.*, 2019, **17**, 6528–6537.



## Paper

- 29 H. Jung, J. Heo, N. Park, K. C. Lim and H. Jung, *J. Hazard. Mater.*, 2023, **451**, 131150.
- 30 V. Nahum, U. Nili, E. Bloch-Shilderman, B. Smolkin and N. Ashkenazi, *Int. J. Pharm.*, 2021, **603**, 120689.
- 31 E. Bloch-Shilderman, U. Nili, V. Nahum, B. Smolkin and N. Ashkenazi, *Arch. Toxicol.*, 2023, **97**, 2771–2783.
- 32 M. A. Stolberg and W. A. Mosher, *J. Am. Chem. Soc.*, 1956, **79**, 2618–2620.
- 33 *Reactive Skin Decontamination Lotion (RSDL) – Medical Countermeasures Database – CHEMM (and references therein)*, [https://chemm.hhs.gov/countermeasure\\_RSDL.htm](https://chemm.hhs.gov/countermeasure_RSDL.htm), (accessed 30 April 2024).
- 34 A. Bierwisch, M. Koller, F. Worek and S. Kubik, *Eur. J. Org. Chem.*, 2016, 5831–5838.
- 35 N. B. Munro, S. S. Talmage, G. D. Griffin, L. C. Waters, A. P. Watson, J. F. King and V. Hauschild, *Environ. Health Perspect.*, 1999, **107**, 933–974.

