RSC Advances



View Article Online

View Journal | View Issue

PAPER

Check for updates

Cite this: RSC Adv., 2024, 14, 20061

Mechanisms of photoisomerization of the prenylated flavin mononucleotide cofactor: a theoretical study[†]

Pannipa Panajapo, 🕩 Phorntep Promma and Kritsana Sagarik 🕩 *

The enzymatic decarboxylation of α , β -unsaturated acids using the ferulic acid decarboxylase (Fdc1) enzyme and prenylated flavin mononucleotide (prFMN) cofactor is a potential, environmentally friendly reaction for the biosynthesis of styrene and its derivatives. However, experiments showed that the enzyme activity of Fdc1 depends on the ring structure of prFMN, namely, the iminium and ketimine forms, and the loss of enzyme activity results from $prFMN^{im} \rightarrow prFMN^{ket}$ photoisomerization. To obtain insight into this photochemical process and to improve the enzyme efficiency of Fdc1, two proposed photoisomerization mechanisms with different proton sources for the acid-base reaction were studied herein using theoretical methods. The potential energy surfaces calculated using the density functional theory method with the Becke, 3-parameter, and Lee-Yang-Parr hybrid functionals and DZP basis set (DFT/B3LYP/DZP) and TD-DFT/B3LYP/DZP methods confirmed that the light-dependent reaction occurs in the rate-determining proton transfer process and that the mechanism involving intermolecular proton transfer between prFMN^{im} and Glu282 (external base) is energetically more favorable than that involving intramolecular proton transfer in prFMN^{im} (internal base). The thermodynamic results obtained from the transition state theory method suggested that the exothermic relaxation energy in the photo-to-thermal process can promote the spontaneous formation of a high-energy-barrier transition state, and an effective enzymatic decarboxylation could be achieved by slowing down the formation of the undesirable thermodynamically favorable product (prFMN^{ket}). Because the rate constant for formation of the high-energy-barrier transition state varies exponentially over the temperature range of 273–298 K, and experimental results have shown that incubating Fdc1 on ice results in a complete loss of enzyme activity, it is recommended to perform the decarboxylation reaction at 285 K to strike a balance between minimizing enzyme stability loss at 273 K and mitigating the effects of UV irradiation. The computational strategy and fundamental insights obtained in this study could serve as guidelines for future theoretical and experimental investigations on the same and similar photochemical systems.

Received 17th March 2024 Accepted 31st May 2024

DOI: 10.1039/d4ra02035a

rsc.li/rsc-advances

Introduction

The enzymatic decarboxylation of α , β -unsaturated acids using the ferulic acid decarboxylase (**Fdc1**) enzyme is envisioned as a potential environmentally friendly process for synthesizing styrene and its derivatives from natural resources.¹⁻³ This enzymatic reaction can be effectively accomplished through 1,3dipolar cycloaddition between substrates and enzyme cofactors,⁴ among which the prenylated flavin mononucleotide (**prFMN**) has been proven to be effective for the biosynthesis of styrene.^{1,3-5,7} Payne *et al.*⁴ proposed a mechanism for the enzymatic decarboxylation of α , β -unsaturated acids using the **prFMN** cofactor. The mechanism comprising four consecutive elementary reactions has been widely accepted and further studied using various theoretical and experimental methods. They are 1,3-dipolar cycloaddition, Grob-type decarboxylation, protonation, and retro 1,3-dipolar cycloaddition,⁴ among which the enzyme-catalyzed 1,3-dipolar cycloaddition of **prFMN** was suggested by a mechanism-based inhibitor experiment to play the most important role.²

Ferguson *et al.*¹ studied the enzyme efficiency of **Fdc1** in styrene production from cinnamic acid by monitoring substrate consumption using spectroscopic and kinetic isotope effect methods, from which the cycloelimination was suggested to represent the rate-determining step. In ref. 1, two forms of **prFMN** with different ring structures were considered, namely, the iminium and ketimine forms, abbreviated **prFMN**^{im} and **prFMN**^{ket}, respectively. The experimental results indicated that with **prFMN**^{im}, the decarboxylation reaction proceeded *via* 1,3-dipolar cycloaddition, whereas the reaction with **prFMN**^{ket}

School of Chemistry, Institute of Science, Suranaree University of Technology, Nakhon Ratchasima, 30000, Thailand. E-mail: kritsana@sut.ac.th; Fax: +66-44-224635; Tel: +66-44-224635

[†] Electronic supplementary information (ESI) available. See DOI: https://doi.org/10.1039/d4ra02035a

occurred *via* Michael addition, and the enzyme activity was confirmed to be higher using **prFMN**^{im}.

To study the effect of the **prFMN**^{im} and **prFMN**^{ket} cofactors on the enzyme activity, the enzymatic decarboxylation using **Fdc1** to generate styrene from cinnamic acid was further studied.⁵ Highresolution crystal structures and mass spectrometric and kinetic experiments revealed that the **prFMN**^{im} \rightarrow **prFMN**^{ket} isomerization could occur through an irreversible photochemical reaction, which is independent of the **Glu277–Arg173–Glu282** residue network. This photoisomerization reaction was suggested to be the key factor for the loss of **Fdc1** enzyme activity.

Two photoisomerization mechanisms with different proton sources for the acid–base reaction are proposed in Fig. 1.⁵ Type (1) mechanism involves four consecutive elementary reactions and begins with the intramolecular proton transfer in **prFMN**^{im}, whereas for Type (2) mechanism, the intermolecular proton transfer between **prFMN**^{im} and **Glu282** represents the initial process with only three consecutive elementary reactions. Although the elementary reactions in Type (1) and (2) mechanisms were not known in detail, the light-dependent reaction was anticipated to be in the cyclization process, in which the exposure to UV light at $\lambda^{abs} = 365$ nm for 5 min led to a complete loss of enzyme activity and change in the absorption spectrum.⁵ This light-dependent reaction is similar to maleimide [5 + 2] photocycloaddition reported in ref. 6.

In our previous study,⁷ the elementary reactions proposed in ref. 4 were examined in low and high local dielectric environments ($\varepsilon = 1$ and 78) using the density functional theory (DFT) method with the Becke, 3-parameter, and Lee–Yang–Parr hybrid functionals and DZP basis set (DFT/B3LYP/DZP) and the transition state theory (TST) method. The active site models included the α -methylcinnamate (Cin) substrate, **prFMN**^{im} cofactor, and all relevant **Fdc1** residues. The results confirmed

that the **Fdc1** backbone does not play an important role in the decarboxylation reaction and that indirect cycloelimination in a low local dielectric environment is the rate-determining step.

Literature review showed that in the past decade (2014–2024), there were 44 published research articles reporting the design and/or improvement of enzymatic decarboxylation in organic syntheses, among which 30 studies focused on the decarboxylation of α,β -unsaturated acids, and 14 studies used the **Fdc1** enzyme in the biosynthesis of styrene and its derivatives. Literature review also revealed that only 5 studies investigated the photoisomerization of the enzyme cofactors under UV exposure, among which only one experimental study⁵ examined in detail the loss of the enzyme activity through the **prFMN^{im}** \rightarrow **prFMN^{ket}** photoisomerization.

Because there is no theoretical study on this photoisomerization reaction, to bridge the gap between the experimental and theoretical knowledge, Type (1) and Type (2) mechanisms were studied in detail using the DFT/B3LYP/DZP, TD-DFT/B3LYP/DZP, and TST methods. To obtain insight into the $prFMN^{im} \rightarrow prFMN^{ket}$ photoisomerization, this mechanistic study focused on the molecular processes (scenarios) in the lowest singlet excited (S_1) state and on the kinetics and thermodynamics of nonradiative reaction pathways. The theoretical investigation began with calculations of the equilibrium structures and spectroscopic properties of the selected active site models, from which the potential energy surfaces (PESs) for **prFMN**^{im} \rightarrow **prFMN**^{ket} were optimized in the S₁ and S₀ states. The kinetic and thermodynamic properties of the PESs were computed using the TST method. To suppress/delay the $prFMN^{im} \rightarrow prFMN^{ket}$ photoisomerization process, appropriate thermodynamic conditions were suggested in order to improve the enzymatic decarboxylation of α , β -unsaturated acids using Fdc1.



Fig. 1 The proposed $prFMN^{im} \rightarrow prFMN^{ket}$ photoisomerization mechanisms obtained based on high-resolution crystal structures, mass spectrometric and kinetic experiments.⁵ All the symbols are explained in the text. Type (1) = mechanism involving intramolecular proton transfer in the $C_{1'}^{prFMN} - H \rightarrow O^{prFMN^{im}}$ H-bond in $prFMN^{im}$ with four consecutive elementary reactions; Type (2) = mechanism involving intermolecular proton transfer in the $C_{1'}^{prFMN} - H \rightarrow O^{Glu282}$ H-bond between $prFMN^{im}$ and Glu282 with three consecutive elementary reactions. Int = internal base; Ext = external base.

Computational methods

Active site models

Because the analysis of the equilibrium structures and PESs in our previous study⁷ showed that the **Fdc1** backbone (Fig. 2a) does not play an important role in the decarboxylation reaction, the active site was modeled in this work by substituting the carbon atoms of the **Fdc1** backbone that connect the residues with CH₃ groups (Fig. 2b). For example, C_R^{Gln190} is the carbon atom of the CH₃ group that substitutes the carbon atom of the **Fdc1** backbone that connects the **Gln190** residue. Although the experiment suggested that **prFMN**^{im} \rightarrow **prFMN**^{ket} photoisomerization is independent of the Glu277-Arg173-Glu282 residue network,⁵ to obtain a realistic picture, the active site models used in this study consisted of Glu277, Arg173, Glu282, and Gln190 and prFMN^{im} (Fig. 2b), regarded as the "active site cluster."

Equilibrium structures

The computational strategy and methods used in this study are shown in Fig. 3. The equilibrium structures of **prFMN**^{im}, **prFMN**^{ket}, **Glu277**, **Arg173**, **Glu282**, and **Gln190** (Table S1†) were optimized primarily in the S₀ state (step (a) in Fig. 3) using the DFT/B3LYP/DZP method and the S₀ \rightarrow S₁ energies (ΔE^{Ex}) were



Fig. 2 (a) The structures of the active site, precursor, cofactor and residues involved in the enzymatic decarboxylation of α,β -unsaturated acid using Fdc1 enzyme and prFMN^{im} cofactor obtained from DFT/B3LYP/DZP geometry optimizations.⁷ (b) An example of the model active site cluster (St0) used in the study of the prFMN^{im} \rightarrow prFMN^{ket} photoisomerization. Dash lines show the residue network distances. Int = intramolecular proton transfer in the C^{prFMN}₁ – H \rightarrow O^{prFMN^{im}} H-bond of prFMN^{im}; Ext = intermolecular proton transfer in the C^{prFMN}₁ – H \rightarrow O^{Glu282} Hbond between prFMN^{im} and Glu282; C^{Arg173}_R, C^{Glu277}_R and C^{Glu282}_R = carbon atoms of the CH₃ groups that substitute the carbon atoms of the Fdc1 backbone.



Fig. 3 The computational strategy and methods used to study the $prFMN^{im} \rightarrow prFMN^{ket}$ photoisomerization. DFT/B3LYP/DZP = density functional theory with the B3LYP functional and DZP basis set; TD-DFT/B3LYP/DZP = time-dependent density functional theory with the B3LYP functional and DZP basis set; TST = transition state theory; (S_0/S_1) = intersection of the S_0 and S_1 states; react, Int/ prod = reactant, intermediate and product, respectively.

calculated using the TD-DFT/B3LYP/DZP method. These computational methods were chosen based on our good experience and benchmark calculations in several mechanistic studies. For example, for the photodissociation and formation of glycine,^{8,9} our benchmark calculations against the complete active space multiconfigurational second-order perturbation theory (CASPT2) method showed that the characteristic structures and energies on the S₀ and S₁ PESs (*e.g.*, the transition structures and structures at the S₀/S₁ intersection) obtained from the DFT/B3LYP and TD-DFT/ B3LYP methods were comparable with those obtained from the CASPT2 method, and the DFT/B3LYP/DZP and DFT/B3LYP/TZP methods yielded approximately the same equilibrium structures, relative interaction energies and energy barriers on the PESs for bifunctional proton transfers in the hydrogen-bonds (Hbonds) in poly(benzimidazole) (PBI) systems.¹⁰

In this work, although our model active site clusters contain aromatic compounds, the dispersion correction was not included in the DFT and TD-DFT calculations, because the predominant intermolecular interactions are the H-bond and electrostatic interactions, and there is no π - π interaction directly involved on the hypothesized reaction pathways. Because the model active site clusters used in this work are large in view of quantum chemical methods, based on which the energy gradients on the S₀ and S₁ PESs had to be repeatedly calculated in the reaction pathway optimizations, to compromise between the numerical accuracy and computational resources, we decided to use the DFT/B3LYP/DZP and TD-DFT/ B3LYP/DZP methods without the dispersion correction. Remarks should be made on the application of the TD-DFT method with the B3LYP hybrid functional. The TD-DFT method has been a predominant method for excited state calculations on large molecules. The TD-DFT method circumvents explicit state calculations by focusing only on excitation energy and transition dipole moment.¹¹ However, benchmark TD-DFT calculations revealed that pure density functionals tend to underestimate transition energies, whereas hybrid functionals such as B3LYP yield better results especially for low-lying singlet transition states.¹²

Practically, the presence of nearly degenerate HOMO and LUMO energies, particularly at the intersections of the S_0 and S_1 states, introduces spin instability, which is associated with holes positioned below the Fermi level. The theoretical results reported in ref. 13 demonstrated that for the photochemical ring opening in oxirane, this problem can be avoided by employing the Tamm–Dancoff approximation (TDA) in conjunction with the TD-DFT method. Because the TD-DFT/TDA method is a simplification of the full TD-DFT method, in which the full TD-DFT equations are truncated to mitigate the algorithmic failure due to spin instability, the computational efficiency can be significantly increased.¹⁴ Therefore, the TD-DFT/B3LYP method with TDA was applied in the present and all of our previous studies on large molecular systems.^{15,16}

Analysis of the elementary reactions in Type (1) and (2) mechanisms⁵ resulted in seven active site clusters. In the initial active site cluster St0 (Fig. 1), **prFMN**^{im} is connected to **Gln190** *via* the N^{Gln190}-H···O^{prFMNim,-} H-bond (Fig. 2b). The **Glu277**-

Paper

Arg173–Glu282 network is associated *via* a carboxylate–guanidinium–carboxylate (COO⁻…Gdm⁺…COO⁻) salt-bridge-type interaction (Fig. 2b), which partly stabilizes the molecules in the active site clusters. The equilibrium structures and ΔE^{Ex} of the active site clusters were calculated using the DFT/B3LYP/ DZP and TD-DFT/B3LYP/DZP methods, respectively (Table S2†). All quantum chemical calculations were performed using the TURBOMOLE 7.50 software package.¹⁷ The absorption spectra of the active site clusters were computed from 500 Wigner sampled structures using the NEWTON-X software package¹⁸ interfaced with TURBOMOLE 7.50.

$prFMN^{im} \rightarrow prFMN^{ket}$ photoisomerization pathways

To simplify the discussion, the symbols shown in Fig. 1 are used and the proposed photoisomerization pathways⁵ are explained in detail. Type (1) mechanism comprises four elementary reactions: (1) $St0 \rightarrow St1^{Int} = C_{1'}^{prFMN} - H \rightarrow O^{prFMN}$ intramolecular proton transfer; (2) $St1^{Int} \rightarrow St2^{Int} =$ covalent bond dissociation; (3) $St2^{Int} \rightarrow St3^{Int} =$ cyclization; and (4) $St3^{Int} \rightarrow St4 =$ reverse protonation. For the Type (2) mechanism, only three elementary reactions were proposed: (1) $St0 \rightarrow St1^{Ext} = C_{1'}^{prFMN} - H \rightarrow O^{Glu282}$ intermolecular proton transfer; (2) $St1^{Ext} \rightarrow St2^{Ext} =$ covalent bond dissociation; and (3) $St2^{Ext} \rightarrow St3^{Ext} \rightarrow St4 =$ concerted cyclization and reverse protonation.

To characterize the structures and energetics of the active site clusters on the PESs, additional symbols are used; $[...]^{Int}$ and $[...]^{Int,*}$ denote structures on the S_0 and S_1 PESs of the Type (1) mechanism, respectively; $[...]^{Ext,\ddagger}$ and $[...]^{Ext,\$}$ denote transition structure (‡) and structure at the S_0/S_1 intersection (§) on the PESs of the Type (2) mechanism, respectively; $\Delta E^{Ex} = S_0 \rightarrow$

 S_1 energy; $\Delta E^{\$} =$ energy at the S_0/S_1 intersection (§); and $\Delta E^{\ddagger} =$ energy barrier. The equilibrium structures of the active site clusters obtained from the DFT/B3LYP/DZP geometry optimization (Table S2†) were used in the elementary reaction pathway optimization in the S_1 state (step (b) in Fig. 3). The elementary reaction pathway optimization was aimed at finding the possibility of the $S_1 \rightarrow S_0$ nonradiative relaxation of the $S_0 \rightarrow S_1$ excited active site clusters.

Because the light-induced reaction was not clearly identified in ref. 5, the $S_0 \rightarrow S_1$ excited precursors for photoisomerization were hypothesized based on the radiation wavelength used in the experiment ($\lambda^{abs} = 365 \text{ nm}/\Delta E^{Ex} = 3.40 \text{ eV}$), ΔE^{Ex} obtained from TD-DFT/B3LYP/DZP calculations, and ΔE^{\ddagger} for the formation of the precursors in the S_0 state. Herein, 7–15 structures connecting the precursor, transition state, and intermediate/ product were optimized using a constrained optimization algorithm¹⁹ included in TURBOMOLE 7.50. If the S_0/S_1 intersection was found, the pathway being optimized was refined using the S_0/S_1 structure as the intermediate/product (step (c) in Fig. 3).

Kinetics and thermodynamics of photoisomerization

Characteristic structures of the model active site clusters on the S_0 and S_1 PESs were used for the calculation of the rate constants using the TST method (step (d) in Fig. 3).^{20,21} Herein, the quantized-vibrational rate constants (k^{Q-vib}) were computed over the temperature range of 273–353 K using eqn (1):²²

$$k^{\text{Q-vib}}(T) = \frac{k_{\text{B}}T}{h} \frac{Q_{\text{ZPE}}^{\ddagger}}{Q_{\text{ZPE}}^{\text{R}}} e^{-\Delta E_{\text{ZPE}}^{\ddagger}/k_{\text{B}}T}$$
(1)



Reaction path

Fig. 4 Schematic diagram showing calculations of Gibbs free energies on the proposed "photo-to-thermal pathway" using the TST method. All the symbol used are explained in the kinetics and thermodynamics of photoisomerization subsection. (I)* \rightarrow (II)** = relaxation of the S₀ \rightarrow S₁ vertically excited structure (I)* to structure (II)** at the S₀/S₁ intersection; (II)* \rightarrow (III) \rightleftharpoons (IV)* \rightarrow (V) = relaxation of structure (II)* at the S₀/S₁ intersection; (II)* \rightarrow (III) \rightleftharpoons (V) = relaxation of structure (II)* at the S₀/S₁ intersection; (II)* \rightarrow (III) \rightleftharpoons (V) = relaxation of structure (II)* at the S₀/S₁ intersection; (II)* \rightarrow (III) \rightleftharpoons (IV)* \rightarrow (V) = relaxation of structure (II)* at the S₀/S₁ intersection; (II)* \rightarrow (III) \rightleftharpoons (IV)* \rightarrow (V) = relaxation of structure (II)* at the S₀/S₁ intersection; (II)* \rightarrow (III) \rightleftharpoons (IV)* \rightarrow (V) = relaxation of structure (II)* at the S₀/S₁ intersection; (II)* \rightarrow (III) \rightleftharpoons (IV)* \rightarrow (V) = relaxation of structure (II)* at the S₀/S₁ intersection; (II)* \rightarrow (III) \rightleftharpoons (IV)* \rightarrow (V) = relaxation of structure (II)* at the S₀/S₁ intersection; (II)* \rightarrow (III) \rightleftharpoons (IV)* \rightarrow (V) = relaxation of structure (II)* at the S₀ state.

In eqn (1), the energy barrier $(\Delta E_{ZPE}^{\ddagger})$ was obtained with the zero-point vibrational energy correction. Q_{ZPE}^{R} and Q_{ZPE}^{\ddagger} are the partition functions of the precursor and transition structures, respectively. $k_{\rm B}$ and h are the Boltzmann and Planck constants, respectively.

The thermodynamic properties of interest were the activation free energies (ΔG^{\ddagger}) and enthalpies (ΔH^{\ddagger}). They were derived from $k^{\text{Q-vib}}(T)$ using eqn (2) and (3), respectively:

$$\Delta G^{\ddagger} = \left[\ln\left(\frac{k_{\rm B}T}{h}\right) - \ln k^{\rm Q-vib}(T) \right] RT \tag{2}$$

$$\ln k^{\text{Q-vib}}(T) = \ln A + \frac{\Delta S^{\ddagger}}{R} - \frac{\Delta H^{\ddagger}}{RT}$$
(3)

 ΔS^{\ddagger} in eqn (3) is the activation entropy and *R* is the gas constant. The value of ΔH^{\ddagger} is obtained from the slope of the linear relationship between $\ln k^{Q-\text{vib}}(T)$ and 1000/T. To calculate the thermodynamic properties of the nonradiative photo-isomerization pathway, the $S_0 \rightarrow S_1$ excited active site clusters were assumed to barrierlessly relax to the S_0/S_1 intersection and to the structures in the S_0 state.

This led to the consecutive "photo-to-thermal pathway" in Fig. 4, the (I)^{*} \rightarrow (II)^{*,§} relaxation in the S₁ state followed by (II)[§] \rightarrow (III) \rightleftharpoons (IV)[‡] \rightarrow (V) in the S₀ state, in which (III) \rightleftharpoons (IV)[‡] \rightarrow (V) was assumed to be in quasi-equilibrium. Based on this hypothesized consecutive reaction pathway, the total Gibbs free energy ($\Delta G^{\circ, \text{Tot}}$) was computed using ΔG^{\ddagger} obtained from the TST method. For (I)^{*} \rightarrow (II)^{*}, $\Delta G^{\circ,(1)^* \rightarrow (II)^*,\$} = -\Delta G_r^{\ddagger,(1)^* \leftarrow (II)^*,\$}$, whereas $\Delta G^{\circ,(1)^* \rightarrow (III)} = -(\Delta G_r^{\ddagger,(1)^* \leftarrow (II)^*,\$} + \Delta G_r^{\ddagger,(II)^{\$} \leftarrow (III)})$ for (I)^{*} $\rightarrow (\mathrm{II})^{*,\$}/(\mathrm{II})^{\$} \rightarrow (\mathrm{III}). \text{ For } (\mathrm{III}) \rightleftharpoons (\mathrm{IV})^{\ddagger} \rightarrow (\mathrm{V}), \ \Delta G^{\mathrm{o},(\mathrm{III}) \rightarrow (\mathrm{V})} =$ $\Delta G_{f}^{\ddagger,(\mathrm{III})\to(\mathrm{IV})\ddagger} - \Delta G_{r}^{\ddagger,(\mathrm{IV})\ddagger\leftarrow(\mathrm{V})}$. Therefore, $\Delta G^{\circ,\mathrm{Tot}} = \Delta G^{\circ,(\mathrm{I})^*\to(\mathrm{III})}$ + $\Delta G^{\circ,(\mathrm{III}) \to (\mathrm{V})}$. Likewise, the Gibbs free energy for the formation of the transition structure $(IV)^{\ddagger}$ in the S₀ state $\Delta G^{0,(I)^* \to (IV)\ddagger} = \Delta G$ $_{O},(I)^* \rightarrow (III) + \Delta G_f^{\ddagger,(III) \rightarrow (IV)\ddagger}$. The same method was used to calculate the entropy changes $(\Delta S^{o,Rx})$ of the isomerization reaction (system). All the kinetic and thermodynamic properties were calculated using the DL-FIND program²³ included in the ChemShell software package.24

Results and discussion

Equilibrium structures of the active site clusters

The DFT/B3LYP/DZP results show that starting from the seven hypothesized active site clusters shown in Fig. 1, the optimized structures are slightly changed (Table S2a[†]), except for St2^{Int} and St2^{Ext}. For St2^{Int}, DFT/B3LYP/DZP geometry optimization yielded the precursor St0 (**prFMN^{im}** in the active site cluster), whereas starting from St2^{Ext} resulted in the product St4 (**prFMN^{ket}** in the active site cluster). These results suggest that only five active site clusters are stationary points (intermediates/ products) on the S₀ PESs, and St2^{Int} and St2^{Ext} could only be the transition structures on the reaction pathways.

Analysis of the equilibrium structures of the five active site clusters revealed that in the S₀ state, the residue-to-residue (R-to-R) distances in Fig. 2b are not significantly different, characterized by standard deviations less than ± 0.15 Å (Table S2b†); the average R-

to-R distances were calculated using the distances between the carbon atoms of the CH₃ groups that substituted the carbon Fdc1 backbone. atom of the For example, $R_{C_{p}^{Glu277}-C_{p}^{Glu282}} = 13.26 \pm 0.09, R_{C_{p}^{Glu190}-C_{p}^{Glu282}} = 17.11 \pm 0.06$ Å, and $R_{C_{c}^{Gln190}-C_{c}^{Arg173}} = 12.52 \pm 0.15$ Å. Because our previous study⁷ also revealed that the residue-to-residue distances (active site volume) do not change significantly during the enzymatic reaction, the lock-and-key model could explain the substrate specificity of the Fdc1 enzyme, and the catalytic efficiency of this enzymatic decarboxylation reaction is partly connected to the efficiency of the proton exchange between the substrate and Glu282, which is a part of the conserved Glu277-Arg173-Glu282 residue network. These results further suggested that the model active site clusters without the Fdc1 backbone are reasonable and can be used for further studies.

DFT/B3LYP/DZP geometry optimization further suggested that St4 possesses the lowest total energy in the S₀ state, and St0 is 24 kJ mol⁻¹ less stable. The TD-DFT/B3LYP/DZP method showed that the $S_0 \rightarrow S_1$ energies of **prFMN**^{im} and **prFMN**^{ket} are $\Delta E^{\text{Ex}} = 3.15 \text{ and } 2.25 \text{ eV} (\lambda^{S_0 \rightarrow S_1} = 394 \text{ and } 551 \text{ nm})$, respectively. Comparison of ΔE^{Ex} in Tables S1 and S2a[†] indicated that ΔE^{Ex} of St0 is red-shifted from that of **prFMN**^{im}; ΔE^{Ex} of St0 is 0.55 eV (~83 nm red-shifted) lower than that of **prFMN**^{im}, whereas ΔE^{Ex} of St4 and that of **prFMN^{ket}** are almost the same. The red-shift is hypothesized to result from strong H-bond interaction between prFMN^{im} and residues in the S₁ state; based on TD-DFT calculations on the camphorsulfonic acid doped polyaniline,²⁵ the electronic spectral red- and blue-shifts could be induced by the excited state H-bond dynamics, for which strong H-bond interaction in the excited states leads to an increase in the oscillator strength and red-shift.

Analysis of the HOMOs and LUMOs in Fig. 5a suggested that while the electron density distributions for **prFMN**^{ket} in the S₀ and S₁ states are not substantially different, those for **prFMN**^{im} are significantly different, especially for the HOMOs, which are characterized by a lower π -character in the phenyl ring. The Hbond formation between the cofactor and residues results in nearly the same electron density distributions in the S₀ and S₁ states for St0 and St4 (Fig. 5b); in the S₀ state, electron density distributions for St0 and St4 are extensive in the **Glu277– Arg173–Glu282** network, whereas in the S₁ state, the electron densities at the cofactor are the highest, resulting in an increase in the π -character at the cofactors. This could explain why ΔE^{Ex} for St0 and St4 are not significantly different (Table S2a†), ΔE^{Ex} = 2.60 and 2.69 eV, respectively.

Remarks should be made on the HOMOs of the **Glu277– Arg173–Glu282** network in Fig. 5b. The extensive charge (electron density) distributions in the S₀ state within the **Glu277– Arg173–Glu282** network reflect the predominant electrostatic interactions in the COO⁻...Gdm⁺...COO⁻ salt-bridge; Gdm⁺ stabilizes the two carboxylate anions. Because similar COO⁻... Gdm⁺...COO⁻ salt-bridges were found in many protein structures, and theoretical studies have shown that the Gdm⁺... COO⁻ attractive interaction affects the ligand recognition and binding, as well as enzyme folding and activity, the COO⁻... Gdm⁺...COO⁻ salt-bridge with specific electrostatic interaction could be synthesized and applied, for example, in drug design;



Fig. 5 The HOMOs and LUMOs of the prFMN^{im} and prFMN^{ket} cofactors (a), and model active site clusters (b), obtained from DFT/B3LYP/DZP geometry optimizations.

Gdm⁺ derivatives have already been used to treat various deceases related to muscle weakness, cancer and diabetes.²⁶

Potential energy surfaces for $prFMN^{im} \rightarrow prFMN^{ket}$ photoisomerization

The structures and energies of the active site clusters on the S_0 and S_1 PESs obtained from the DFT/B3LYP/DZP and TD-DFT/ B3LYP/DZP reaction path optimization (Fig. S1–S6†) are analyzed in detail. To study the possibility of St2^{Int} and St2^{Ext} being the $S_0 \rightarrow S_1$ photoexcited precursors in the [5 + 2] photocycloaddition, as proposed in ref. 6, the PESs for St0 \rightarrow St1^{Int} \rightarrow St2^{Int} and St0 \rightarrow St1^{Ext} \rightarrow St2^{Ext} in the S_0 state were primarily calculated; the intra- and intermolecular proton transfers and covalent bond dissociation occurring in the S_0 state serve as prerequisites for the light-dependent cycloaddition in Type (1) and Type (2) mechanisms,⁵ respectively.

The results showed that both St0 \rightarrow St1^{Int} \rightarrow St2^{Int} and St0 \rightarrow St1^{Ext} \rightarrow St2^{Ext} involve extraordinarily high energy barriers in the S₀ state, especially for the covalent bond dissociation, ΔE^{\ddagger} = 456.4 and 435.9 kJ mol⁻¹ (Fig. S1b and S2b,† respectively). Based on these findings and the observations that St2^{Int} and St2^{Ext} are not stationary points with S₀ \rightarrow S₁ energies significantly different from the experimental radiation wavelength ($\lambda^{abs} = 365 \text{ nm}/\Delta E^{Ex} = 3.40 \text{ eV}$),⁵ these two active site clusters were ruled out from further study; TD-DFT/B3LYP/DZP single point calculations suggested that for St2^{Int} and St2^{Ext}, $\Delta E^{Ex} =$ 2.96 and 1.88 eV, respectively (Table S2a†).

Similarly, because the intra- and intermolecular proton transfers (St0 \rightarrow St1^{Int} and St0 \rightarrow St1^{Ext}) possess high energy barriers in the S₀ state, $\Delta E^{\ddagger} = 262.6$ (Fig. S1a[†]) and 125.7 kJ mol⁻¹ (Fig. S2a[†]), respectively, it is unlikely that St1^{Int} and St1^{Ext} serve as the photoexcited precursors for S₀ \rightarrow S₁. To study the possibility of St0 being the S₀ \rightarrow S₁ photoexcited precursor, UV-visible absorption spectra were calculated at 277 and 300 K using 500 Wigner sampled initial conditions. The

results in Fig. 6 show two outstanding peaks, *e.g.*, at 300 K, $\lambda^{abs} = 384$ and 474 nm. Because the structured peak at $\lambda^{abs} = 384$ nm is close to the radiation wavelength used in an experiment,⁵ St0 was chosen as the S₀ \rightarrow S₁ photoexcited precursor for Type (1) and Type (2) mechanisms.

The PESs obtained from reaction path optimization show that the $S_0 \rightarrow S_1$ vertically excited structure barrierlessly relaxes to the structure at the S_0/S_1 intersection (St0^{*} \rightarrow St0^{*,§} in Fig. S3a[†]). The S_0 PES (Fig. S4[†]) illustrates that after the $S_1 \rightarrow S_0$ nonradiative relaxation, the intramolecular proton transfer (St0[§] \rightarrow St0-[1]^{Int} \rightarrow St1^{Int}) in the Type (1) mechanism possesses a rather high energy barrier, $\Delta E^{\ddagger} = 209.5$ kJ mol⁻¹, whereas $\Delta E^{\ddagger} = 86.9$ kJ mol⁻¹ is for the intermolecular proton transfer (St0[§] \rightarrow St0-[1]^{Ext} \rightarrow St1^{Ext}) in the Type (2) mechanism



Fig. 6 The UV-visible spectra of the model active site cluster St0 (prFMN^{im} in the residue network) obtained based on 500 Wigner sample structures at 277 and 300 K.





Fig. 7 (a) and (b) Type (3) mechanism and potential energy profiles involving the $S_0 \rightarrow S_1$ excitation of St0, intermolecular proton transfer in $C_{1'}^{\text{prfMN}} - H \rightarrow O^{Glu282}$ H-bond between **prFMN**^{im} and **Glu282**, ring expansion and reverse protonation, respectively. (c) The H-bond distances ($R_{C_{1}^{\text{prfMN}} - H \dots O^{Glu282}}$ and $R_{O}^{Glu282} - H \dots O^{\text{prfMN}}$) and local valence charge densities on the high-energy barrier intermolecular proton transfer process (St0-[1]^{Ext} \rightarrow St0-[1]^{Ext.‡} \rightarrow St1^{Ext}). (d) and (e) Type (4)

(Fig. S5a[†]). Hence, one can conclude that the external base pathway is energetically more favorable than the internal one, and only the Type (2) mechanism is further discussed in detail.

To test the hypothesis that the red-shift in ΔE^{Ex} for St0 compared to that of **prFMN**^{im} results from strong H-bond interaction between the **prFMN**^{im} cofactor and residues in the S₁ state, the variation of the N^{Gln190}-H···O^{prFMN,-} H-bond distance in the S₁ state was analyzed as an example. The results revealed that on the S₁ PES (Fig. S3a†), while the $R_{\text{N}^{\text{Gln190}}-\text{H}\cdots\text{O}^{\text{prFMN},-}}$ H-bond distance decreases, the $R_{\text{N}^{\text{Gln190}}-\text{H}}$ distance increases (Fig. S3b†), reflecting an increase in the N^{Gln190}-H···O^{prFMN,-} H-bond strength (red-shift) in the S₁ state towards the S₀/S₁ intersection.

Because St2^{Ext} had already been excluded from the photoisomerization pathway, an attempt was made to bypass this active site cluster. The results showed that after the $S_1 \rightarrow S_0$ nonradiative relaxation at the S_0/S_1 intersection (Fig. S3a[†]) and $C_{1'}^{prFMN} - H \rightarrow O^{Glu_{282}}$ intermolecular proton transfer from **prFMN**^{im} to **Glu282** (Fig. S5a[†]), the S₀ PES shows a low energy barrier for St1^{Ext} \rightarrow St3^{Ext}, $\Delta E^{\ddagger} = 11.2$ kJ mol⁻¹ (Fig. S5b[†]), represented by concerted N_{5a}^{prFMN} - C_{5a}^{prFMN} and $C_{4'}^{prFMN}$ - C_{5a}^{prFMN} covalent bond dissociation and formation in Fig. 7a, respectively. This concerted process can be collectively regarded as "ring expansion." To complete the formation of the St4 product $(prFMN^{ket} in the active site cluster)$, the S₀ PES for the reverse protonation from **Glu282** to **prFMN**^{im} (St3^{Ext} \rightarrow St4) has ΔE^{\ddagger} = 63.9 kJ mol⁻¹ (Fig. S5c[†]). The potential energy profile, which bypasses St2^{Ext}, is regarded as the Type (3) mechanism shown in Fig. 7b.

The analysis of the H-bond distances, local valence charge densities and energetics in the high-energy barrier intermolecular proton transfer process $(\text{St0-[1]}^{\text{Ext}} \rightarrow \text{St0-[1]}^{\text{Ext},\ddagger} \rightarrow \text{St1}^{\text{Ext}})$ in Fig. 7c reveals that although the $C_{1'}^{\text{prFMN}} - \text{H} \cdots \text{O}^{\text{Glu282}}$ H-bond in St0-[1]^{Ext} ($R_{\text{CP}^{\text{PFMN}} - \text{H} \cdots \text{O}^{\text{Glu282}} = 2.18$ Å) is longer than the O^{Glu282} -H $\cdots \text{O}^{\text{prFMN}}$ H-bond in St1^{Ext} ($R_{\text{O}^{\text{Glu282}}-\text{H} \cdots \text{O}^{\text{prFMN}} = 1.52$ Å), due to the strong electrostatic interaction between **Glu282** and **prFMN^{im**}, St0-[1]^{\text{Ext}} is more stable than St1^{Ext}. It also appears that proton transfer to one of the COO⁻ groups in the COO⁻ \cdots Gdm⁺ \cdots COO⁻ salt bridge leads to a strong decrease in the electrostatic interaction between **Glu282** and **prFMN^{im**}, recognized from a reduction of the local valence charge density at the COOH group of St1^{Ext}, and the high-energy barrier in St0-[1]^{Ext,‡} \rightarrow St0-[1]^{Ext,‡} \rightarrow St1^{Ext} could be attributed the $C_{1'}^{\text{prFMN}} - \text{H}$ covalent bond breaking.

Because the Type (1) mechanism⁵ has been established in this work to be energetically not favorable, to search for an alternative internal base pathway, the S₀ PES directly connecting structure St0[§] at the S₀/S₁ intersection and St4 (St0[§] \rightarrow St4) was optimized. It appeared that St0[§] \rightarrow St0-[1]^{direct} \rightarrow St0-[1]^{direct,‡} \rightarrow St4

mechanism and potential energy profiles involving the $S_0 \rightarrow S_1$ excitation of St0 and concerted $C_1^{prFMN} - H \rightarrow N_5^{prFMN}$ intramolecular proton transfer in $prFMN^{im}$ and ring expansion. The characteristic structures are shown in detail in Table S2.† $(\ldots)^{S_0} \rightarrow {}^{S_1} = S_0 \rightarrow S_1$ vertical excitation energy; $(\ldots)^{Rel}$ and $(\ldots)^{\ddagger}$ = relative and transition energies on the PES; $(\ldots)^{S_1S_0} =$ difference between the total energies in the S_0 and S_1 states at the S_0/S_1 intersection.

Paper

(Fig. S6[†]) can occur without proton exchange between the **prFMN**^{im} cofactor and the **Glu282** residue. This internal base pathway (Type (4) mechanism shown in Fig. 7d) involves concerted $C_{1'}^{prFMN} - H \rightarrow N_{1'}^{prFMN}$ intramolecular proton transfer, $N_{5}^{prFMN}-C_{5a}^{prFMN}$ and $C_{1'}^{prFMN} - C_{5a}^{prFMN}$ covalent bond dissociation and formation, respectively, with $\Delta E^{\ddagger} = 31.7$ kJ mol⁻¹ (Fig. 7e).

Thermodynamics of the elementary reactions

To study the kinetic and thermodynamic properties of the two energetically favorable pathways, the characteristic active site clusters in Type (3) and Type (4) mechanisms shown in Fig. 7 were used in TST calculations, and the results are summarized in Tables S3–S5.† The results discussed are included in Tables 1

Table 1 Thermodynamics of the elementary reactions for the external base $prFMN^{ket}$ photoisomerization (Type (3) mechanism), obtained from the DFT/B3LYP/DZP, TD-DFT/B3LYP/DZP and TST methods. ΔG° and ΔH° = Gibbs free energy and enthalpy in kJ mol⁻¹; ΔS° = entropy in kJ mol⁻¹ K⁻¹; $\Delta G^{\circ,Tot}$ = total reaction Gibbs free energy for $prFMN^{ket}$ and; $\Delta S^{\circ,Rx}$ = reaction entropy of all the elementary processes; T = temperature in K



Table 2 Thermodynamics of the elementary reactions for the internal base $prFMN^{im} \rightarrow prFMN^{ket}$ photoisomerization (Type (4) mechanism), obtained from the DFT/B3LYP/DZP, TD-DFT/B3LYP/DZP and TST methods. ΔG° and ΔH° = Gibbs free energy and enthalpy in kJ mol⁻¹; ΔS° = entropy in kJ mol⁻¹ K⁻¹; $\Delta G^{\circ,Tot}$ = total reaction Gibbs free energy for $prFMN^{im} \rightarrow prFMN^{ket}$ and; $\Delta S^{\circ,Rx}$ = reaction entropy of all the elementary processes; T = temperature in K



and 2. Analysis based on the photo-to-thermal pathway in Fig. 4, (I)^{*} \rightarrow (II)^{*,8}/(II)[§] \rightarrow (III), revealed that for the Type (3) mechanism at T = 298 K, although the entropy for the formation of St0-[1]^{Ext,‡} is negative ($\Delta S^{\circ,(I)^* \rightarrow (IV)^{\ddagger}} = -7.4 \times 10^{-2}$ kJ mol⁻¹), the high exothermic relaxation energy for St0^{*} \rightarrow St0[§] \rightarrow St0-[1]^{Ext} ($\Delta H^{\circ,(I)^* \rightarrow (III)} = -178.4$ kJ mol⁻¹ in Table 1a) allows the spontaneous formation of this high energy-barrier transition state with $\Delta G^{\circ,(I)^* \rightarrow (IV)^{\ddagger}} = -77.7$ kJ mol⁻¹.

Based on the same approach, the total Gibbs free energy and reaction entropy for the **prFMN**^{im} \rightarrow **prFMN**^{ket} photoisomerization *via* the Type (3) mechanism were computed to be $\Delta G^{\circ,Tot} = -226.3 \text{ kJ mol}^{-1}$ and $\Delta S^{\circ,Rx} = -7.5 \times 10^{-2} \text{ kJ mol}^{-1}$ K⁻¹ at 298 K (Table 1c). For the Type (4) mechanism, $\Delta H^{\circ,(I)^* \rightarrow (III)} = -221.1$, $\Delta G^{\circ,Tot} = -226.3 \text{ kJ mol}^{-1}$, and $\Delta S^{\circ,Rx} = -7.4 \times 10^{-2} \text{ kJ mol}^{-1}$ K⁻¹, with the spontaneous formation of St0-[1]^{direct,‡}, $\Delta G^{\circ,(I)^* \rightarrow (IV)\ddagger} = -131.6 \text{ kJ mol}^{-1}$ (Table 2). Therefore, based on the values of $\Delta G^{\circ,(I)^* \rightarrow (IV)\ddagger}$ of the rate-determining step, the Type (4) mechanism is thermodynamically more favorable than the Type (3) mechanism. Because $\Delta G^{\circ,Tot}$ and $\Delta S^{\circ,Rx}$ of Type (3) and Type (4) mechanisms are the same (Tables 1 and 2, respectively), the hypothesized photo-to-thermal pathway in



Fig. 8 (a) and (b) Correlations between $\tau_{1/2}^{Q-\text{vib}}$ and *T*, and $\ln k_f^{Q-\text{vib}}$ and 1/T for the rate determining step (St0-[1]^{Ext} \rightarrow St0-[1]^{Ext,‡} in Fig. 7(b) in Type (**3**) mechanism obtained from the TST method, compared with experiment.⁵ $\tau_{1/2}^{\text{loss},T}$ and $k^{\text{loss},T} = \text{half-life}$ and firstorder rate constant for the loss of enzyme stability at 273 K;⁵ $\tau_{1/2}^{\text{loss},\text{UV}}$ and $k^{\text{loss},\text{UV}} = \text{half-life}$ and first-order rate constant for the loss of enzyme activity due to UV radiation at 365 nm;⁵ $\tau_{1/2}^{Q-\text{vib}}$ and $k_f^{Q-\text{vib}} =$ half-life and first-order rate constant for the rate-determining intermolecular proton transfer (St0-[1]^{Ext} \rightarrow St0-[1]^{Ext,‡}) in Fig. 7(b).

Fig. 4 is validated and the applicability of the TST method is confirmed.

Kinetics of elementary reactions

To correlate the theoretical results with the experimental data⁵ and to explore the possibility of improving the enzyme efficiency of **Fdc1**, the rate constants (k^{Q-vib}) for Type (3) and Type (4) mechanisms (Tables S3 and S4,† respectively) were analyzed and discussed in detail. The experimental rate constants revealed that the enzyme activity of **Fdc1** purified in the dark was slightly higher than that under visible light, $k^{cat} = 9.3 \pm 0.1$ and 7.6 \pm 0.2 s⁻¹, respectively, and when **Fdc1** was prepared in the dark, the enzyme activity remained constant for many hours. However, direct exposure of the **Fdc1** enzyme incubated in the dark with UV radiation at $\lambda = 365$ nm for ~5 min led to a complete loss of enzyme activity and a change in the UV-visible spectra.

The experimental results also showed that after purification, **Fdc1** incubated on ice resulted in a loss of enzyme stability with a half-life $\tau_{1/2}^{\log s,T} \approx 30$ min. Based on the assumption that the loss of enzyme activity at 273 K follows the first-order kinetics, $k^{\log s,T} = 0.693/\tau_{1/2}^{\log s,T} = 3.85 \times 10^{-4} \text{ s}^{-1}$. In addition, assuming that the photochemical experiment⁵ was conducted at room temperature, the complete loss of enzyme activity due to UV radiation is approximated to be $\tau^{\log s,UV} = 3.00$ s, with the rate constant $k^{\log s,UV} = 1/\tau^{\log s,UV} = 3.33 \times 10^{-3} \text{ s}^{-1}$ at 298 K. The value of $k^{\log s,UV}$ is compatible with the rate-determining step in the Type (3) mechanism; for the $C_{1'}^{\text{prFMN}} - H \rightarrow O^{\text{Glu282}}$ intermolecular proton transfer from **prFMN**^{im} to **Glu282**, $k_{\rm f}^{\text{Q-vib}} = 3.53 \times 10^{-3} \text{ s}^{-1}$ at 298 K (Table S3†).

Although the internal base pathway in Type (4) mechanism is energetically and thermodynamically more favorable than Type (3) mechanism, the concerted rate-determining process is too fast compared with the experimental rate constant, $k_f^{Q,vib} = 3.26 \times 10^0 \text{ s}^{-1}$ compared with $k^{\text{loss,UV}} = 3.33 \times 10^{-3} \text{ s}^{-1}$ at 298 K. Therefore, Type (3) mechanism, which requires an external base, is likely to represent the **prFMN**^{im} \rightarrow **prFMN**^{ket} photoisomerization mechanism.

Balance between the loss of enzyme stability and the efficiency by UV radiation

Based on the above discussion on the theoretical and experimental data,⁵ to reduce the **prFMN**^{im} \rightarrow **prFMN**^{ket} photoisomerization rate without substantial loss of the **Fdc1** stability, an appropriate enzymatic decarboxylation temperature should be chosen; an effective enzymatic decarboxylation using **Fdc1** could be achieved by slowing down the photo-to-thermal process (formation of the high-energy barrier transition state) in Type (3) mechanism, compared with the enzymatic decarboxylation rate.

To recommend the appropriate temperature, at which the styrene production is kinetically controlled in the presence of UV radiation, the plot of the half-life for the formation of the high-energy barrier transition state ($\tau_{1/2}^{\text{Q-vib}}$) *versus T* and the Arrhenius plot for this photo-to-thermal pathway were constructed over the studied temperature range and are shown in

Fig. 8. Fig. 8a shows that $\tau_{1/2}^{\text{Q-vib}}$ varies exponentially over the temperature range of 273–298 K. Thus, to balance between the loss of the enzyme stability at 273 K and that under UV irradiation, the appropriate temperature should be in this temperature range, $\tau_{1/2}^{\text{loss},T} = 1800 (T = 273 \text{ K})$ and $\tau_{1/2}^{\text{loss},\text{UV}} = 208 \text{ s} (T = 298 \text{ K})$.

As a rule of thumb, the rate of chemical reaction doubles if the temperature increases by 10 K. Therefore, the experimental rate constants to produce styrene in this temperature range are $k^{\text{cat}} = 3.00$ and 7.60 s⁻¹, and the rate constants for the $C_{1'}^{\text{prFMN}} - \text{H} \rightarrow \text{O}^{\text{Glu282}}$ intermolecular proton transfer (rate determining step for **prFMN**^{im} \rightarrow **prFMN**^{ket}) are $k_{\text{f}}^{\text{Q-vib}} = 1.98 \times 10^{-4} \text{ s}^{-1}$ (273 K) and 3.53 $\times 10^{-3} \text{ s}^{-1}$ (298 K).

The Arrhenius plot in Fig. 8b reveals that in this temperature range, $k^{\text{loss},T} = 3.85 \times 10^{-4} \text{ s}^{-1}$ (T = 273 K) and $k^{\text{loss},\text{UV}} = 3.33 \times 10^{-3} \text{ s}^{-1}$ (T = 298 K), and the optimal temperature should be at T = 285 K with $k_{\text{f}}^{\text{Qvib}} = 8.16 \times 10^{-4} \text{ s}^{-1}$ for the photo-to-thermal pathway. Therefore, the enzyme activity of **Fdc1** purified under visible light at 285 K is approximated to be $k^{\text{cat}} \approx 3.8 \text{ s}^{-1}$. This experimental rate constant is ~4600 times larger than the rate-determining step for **prFMN**^{im} \rightarrow **prFMN**^{ket} at the same temperature, and styrene production becomes kinetically controlled in the presence of UV radiation at this temperature.

Conclusions

The enzymatic decarboxylation of α , β -unsaturated acids using the **Fdc1** enzyme and **prFMN** cofactor is a potential, environmentally friendly reaction for the biosynthesis of styrene and its derivatives. However, an experiment indicated that the enzyme activity of **Fdc1** depends on the ring structure of **prFMN**, namely, the iminium and ketimine forms, and the loss of the enzyme activity results from the **prFMN**^{im} \rightarrow **prFMN**^{ket} photo-isomerization with the light-dependent reaction suggested to occur in the cyclization process.

Herein, to obtain insight into the $\mathbf{prFMN^{im}} \rightarrow \mathbf{prFMN^{ket}}$ photoisomerization process and to improve the enzyme efficiency of **Fdc1**, two proposed photoisomerization mechanisms with different proton sources for the acid-base reaction were studied using theoretical methods. This mechanistic study focused on the photoisomerization pathways in the S₁ and S₀ states and on the kinetics and thermodynamics of the photo-tothermal process in the active site of **Fdc1**. Analysis of the equilibrium structures of the model active site clusters revealed that because the active site volumes (residue-to-residue distances) obtained from the DFT/B3LYP/DZP geometry optimization were not significantly different, the model active site clusters without the **Fdc1** backbone were proved to be reasonable and can be used in the study of the enzymatic decarboxylation process.

The PESs calculated from the DFT/B3LYP/DZP and TD-DFT/ B3LYP/DZP methods suggested that the light-dependent reaction occurs in the rate-determining proton transfer process (acid-base reaction), and the mechanism involving intermolecular proton transfer between **prFMN**^{im} and **Glu282** (external base) is energetically more favorable than that involving intramolecular proton transfer in **prFMN**^{im} (internal base). The lightdependent reaction was confirmed by the calculated UV-visible spectra to be a high-energy-barrier proton transfer process, for which the $S_0 \rightarrow S_1$ photoexcited precursor is St0 (**prFMN**^{im} in the active site cluster). The thermodynamic results obtained from the TST method suggested that the exothermic relaxation energy in the photo-to-thermal process in the Type (3) mechanism can promote the spontaneous formation of the high-energy-barrier transition state.

Based on the analysis of the theoretical and experimental data, effective enzymatic decarboxylation of the α , β -unsaturated acids using Fdc1 could be achieved by slowing down the formation of the undesirable thermodynamically favorable product (prFMN^{ket}). Because the kinetic results suggested that the rate constant of formation of the high-energy-barrier transition state varies exponentially over the temperature range of 273-298 K, and because the experiment showed that incubating Fdc1 on ice could result in complete loss of enzyme stability, it is recommended to perform the decarboxylation reaction at T =285 K. This temperature allows for kinetically controlled styrene production in the presence of UV radiation, thus addressing the balance between enzyme stability loss at 273 K and under UV radiation. The computational strategy and fundamental insights obtained in this study could serve as guidelines for future theoretical and experimental investigations on the same and similar photochemical systems.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The authors would like to acknowledge the high-performance computer facilities provided by the National e-Science project of the National Electronics and Computer Technology Centre (NECTEC), and the National Science and Technology Development Agency (NSTDA). This work was supported by (I) Suranaree University of Technology (SUT), (II) Thailand Science Research and Innovation (TSRI), and (III) National Science, Research, and Innovation Fund (NSRF). The authors also acknowledge NSTDA Supercomputer Center (ThaiSC) for providing computing resources for this work. This research has received funding support from the NSRF *via* the Program Management Unit for Human Resources & Institutional Development, Research, and Innovation (PMU-B) [grant number B13F660060].

References

- 1 K. L. Ferguson, N. Arunrattanamook and E. N. G. Marsh, *Biochemistry*, 2016, 55, 2857–2863.
- 2 K. L. Ferguson, J. D. Eschweiler, B. T. Ruotolo and E. N. G. Marsh, J. Am. Chem. Soc., 2017, **139**, 10972–10975.
- 3 C.-L. Lan and S.-L. Chen, J. Org. Chem., 2016, 81, 9289-9295.
- 4 K. A. Payne, M. D. White, K. Fisher, B. Khara, S. S. Bailey, D. Parker, N. J. Rattray, D. K. Trivedi, R. Goodacre and R. Beveridge, *Nature*, 2015, **522**, 497–501.

- 5 S. S. Bailey, K. A. P. Payne, K. Fisher, S. A. Marshall, M. J. Cliff, R. Spiess, D. A. Parker, S. E. J. Rigby and D. Leys, *J. Biol. Chem.*, 2018, **293**, 2272–2287.
- 6 D. M. E. Davies, C. Murray, M. Berry, A. J. Orr-Ewing and K. I. Booker-Milburn, *J. Org. Chem.*, 2007, **72**, 1449–1457.
- 7 P. Promma, C. Lao-ngam, R.-Y. Lai and K. Sagarik, *RSC Adv.*, 2022, **12**, 14223–14234.
- 8 J. Nirasok, P. Panajapo, P. Promma, P. Suwannakham and K. Sagarik, *J. Photochem. Photobiol., A*, 2023, **436**, 114354.
- 9 P. Panajapo, P. Suwannakham, P. Promma and K. Sagarik, *R. Soc. Open Sci.*, 2024, **11**, 231957.
- 10 J. Thisuwan, P. Promma and K. Sagarik, *R. Soc. Open Sci.*, 2021, **8**, 211168.
- 11 L. Gonzalez, D. Escudero and L. Serrano-Andres, *ChemPhysChem*, 2012, **13**, 28–51.
- 12 D. Jacquemin, B. Mennucci and C. Adamo, *Phys. Chem. Chem. Phys.*, 2011, 13, 16987–16998.
- 13 E. Tapavicza, I. Tavernelli, U. Rothlisberger, C. Filippi and M. E. Casida, *J. Chem. Phys.*, 2008, **129**, 124108.
- 14 Y.-L. Wang and G.-S. Wu, Int. J. Quantum Chem., 2008, 108, 430-439.
- 15 T. Khrootkaew, S. Wangngae, K. Chansaenpak, K. Rueantong, W. Wattanathana, P. Pinyou, P. Panajapo, V. Promarak, K. Sagarik and A. Kamkaew, *Chem.-Asian J.*, 2023, e202300808.

- 16 S. Siriwibool, N. Kaekratoke, K. Chansaenpak,
 K. Siwawannapong, P. Panajapo, K. Sagarik, P. Noisa,
 R. Y. Lai and A. Kamkaew, *Sci. Rep.*, 2020, **10**, 1283.
- 17 TURBOMOLE V 7.5 2020, A Development of University of Karlsruhe and Forschungszentrum Karlsruhe GmbH, BIOVIA.TURBOMOLE@3ds.com, V7.5 edn, 2019.
- M. Barbatti, M. Bondanza, R. Crespo-Otero, B. Demoulin,
 P. O. Dral, G. Granucci, F. Kossoski, H. Lischka,
 B. Mennucci, S. Mukherjee, M. Pederzoli, M. Persico,
 M. P. Jr, J. Pittner, F. Plasser, E. S. Gil and L. Stojanovic, *J. Chem. Theory Comput.*, 2022, 18, 6851–6865.
- 19 P. Plessow, J. Chem. Theory Comput., 2013, 9, 1305–1310.
- 20 E. Pollak and P. Talkner, *Chaos*, 2005, **15**, 026116.
- 21 P. Hänggi, P. Talkner and M. Borkovec, *Rev. Mod. Phys.*, 1990, **62**, 251–341.
- 22 J. E. House, *Principles of Chemical Kinetics, Kindle Edition*, Academic Press, USA, 2nd edn, 2007.
- 23 J. Kästner, J. M. Carr, T. W. Keal, W. Thiel, A. Wander and P. Sherwood, *J. Phys. Chem. A*, 2009, 113, 11856–11865.
- 24 ChemShell, *A Computational Chemistry Shell*, https://www.chemshell.org/.
- 25 Y. Zhang, Y. Duan and T. Wang, Phys. Chem. Chem. Phys., 2014, 16, 26261.
- 26 A. N. Muttathukattil, S. Srinivasan, A. Halder and G. Reddy, *J. Phys. Chem. B*, 2019, **123**, 9302–9311.