


 Cite this: *RSC Adv.*, 2024, 14, 16318

Ecological assessment and development of analytical methods for concurrent quantification of valsartan and sacubitril: whiteness comparative study based on relative scoring †

 Ahmed K. Kammoun,^a Mostafa A. Afify,^{ID} ^b Rasha M. Youssef,^{ID} ^b Sara A. El-Nahass^c and Sameh E. Younis^{*d}

Sustainable analytical chemistry is gaining great interest in global environmental pollution control. In addition, valsartan (VAS) and sacubitril (SAB) have been recently approved by the FDA as a fixed-dose combination "LCZ696". It showed efficacy and safety enough to extend its application from heart failure to hypertension control. VAS/SAB dual therapy is considered expensive; however, its prescription has increased significantly worldwide. This prescription increased the demand for developing sustainable analytical methods that simultaneously analyze VAS and SAB. Highly sensitive and selective spectrofluorimetric methods have been developed for this purpose. A synchronous spectrofluorimetric technique was applied. In one method, it was followed by spectral derivatization at the first-order level. The signals were recorded at 230 and 211 nm for VAS and SAB, respectively. Synchronous spectrofluorimetry was coupled to a dual-wavelength mathematical approach in the second method. Signals were derived by subtracting synchronous responses at 241 nm, 226 nm, and 239 nm from the response at 208 nm for VAS and SAB, respectively. Method validation was carried out following ICH guidelines. VAS showed linear calibration curves spanning the range of 60–200 and 80–600 ng mL⁻¹ for the derivative and dual wavelength-assisted approaches, respectively. SAB achieved linear responses in the range of 17–190 and 30–350 ng mL⁻¹ for the first and second methods, respectively. The green profile of the proposed methods was confirmed using the analytical eco-scale (AES), green analytical procedure index (GAPI), and analytical greenness metric (AGREE) tools. The proposed hybrid methods proved highly sustainable through the whiteness RGB 12 algorithm evaluation approach. Whiteness was comparatively assessed for the proposed and reported methods based on relative scoring depending on the parameters of each method. Despite this scoring approach being accurate as a relative score for comparative purposes, it gave rise to underestimated absolute scores. Therefore, to obtain a proper conclusion from the comparative whiteness study, all the methods were ranked according to their whiteness score, illustrating the excellent whiteness ranks of the proposed methods. Upon complete comparison with the reported methods, the suggested ones showed several advantages concerning analytical performance and the greenness level. The proven affordability and simplicity encourage their wide industrial application in developing countries.

 Received 15th March 2024
 Accepted 6th May 2024

DOI: 10.1039/d4ra01997k

rsc.li/rsc-advances
^aDepartment of Pharmaceutical Chemistry, Faculty of Pharmacy, King Abdulaziz University, P.O. Box 80260, Jeddah 21589, Saudi Arabia

^bDepartment of Pharmaceutical Analytical Chemistry, Faculty of Pharmacy, Alexandria University, Alexandria, Egypt

^cHealthy Care Clinics, Alexandria/INTRAWOOD, International Trading of Wood, Alexandria, Egypt

^dDepartment of Pharmaceutical Chemistry, Faculty of Pharmacy, Pharos University in Alexandria, Alexandria, Egypt. E-mail: smhyones@yahoo.com; sameh.yones@pua.edu.eg

 † Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d4ra01997k>

1. Introduction

Heart failure (HF) is a disease in which the mortality rate increases by six to nine times more than the normal population.¹ This mortality rate makes ongoing efforts to develop new HF therapies a necessity; one of these therapies is LCZ696. However, it was used as a late-line treatment owing to its high commercial price. However, its role was highly considerable to improve the HF therapy and avoid the toxic "digoxin" use or even heart replacement.² LCZ696 is a valsartan (VAS)/sacubitril (SAB) complex; Fig. 1.³ Thus, it has a dual pharmacological action. VAS blocks the angiotensin II receptor, and SAB prevents



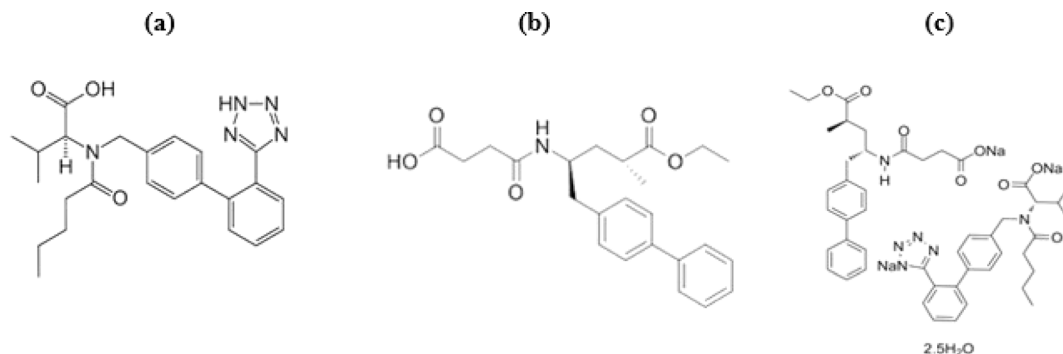


Fig. 1 Chemical structures of VAS (a), SAB (b) and LCZ696 (c).

the degradation of vasoactive peptides *via* neprilysin inhibition. SAB did not succeed as a single HF therapy. On the contrary, the use of the VAS/SAB complex presented a significant improvement for HF patients, even more than that attained by VAS alone.⁴

LCZ696 was rapidly introduced into all HF guidelines and became essential in the treatment protocols for developed and developing countries.^{5,6} This required establishing reliable analytical methods to analyze this combined therapy. Two spectrophotometric^{7,8} and three HPTLC^{9–11} methods have been reported for this purpose. In addition, LCZ696 was assayed by HPLC using many approaches.^{9,10,12–24} In addition, VAS and SAB were simultaneously analyzed *via* spectrofluorometric technique.^{25–27} Very recently, a MEKC method was developed for the same purpose.²⁸

Although synchronous spectrofluorimetry (SF) provides better selectivity and sensitivity than the traditional approach, SF needs further coupling with another analytical method for complete spectral resolution in some cases. Derivative-mathematical and dual-wavelength treatments are the most widely coupled techniques with SF.²⁹ On the other hand, green chemistry has become crucial in developing analytical procedures. There are various qualitative and quantitative metric systems. However, using them together is preferred for the complete picture of the method of greenness.

Current work aims to develop green, reliable, and affordable analytical approaches for the simultaneous analysis of VAS and SAB in their raw materials and LCZ696 tablets. They couple SF with first-derivative and dual-wavelength manipulations. No tedious pretreatment steps were needed. All ICH validation parameters were fulfilled. The green profile was confirmed *via* the eco-scale penalty points, GAPI, and AGREE tools to avoid any hazardous and risky exposure to the analyst or environment while applying the proposed methods.

Moreover, the AGREE assessment was employed to compare the greenness level between the suggested and reported methods. In addition, the methods' whiteness was comparatively investigated *via* the RGB 12 model proposed by Nowak *et al.* that was applied in recent publications for comparative purposes.^{30,31} A complete comparative study of sustainability was performed for the proposed and reported methods based on relative scoring depending on the parameters of each method.

2. Experimental

2.1. Materials, reagents and pharmaceutical formulations

VAS raw material was obtained free of charge from the Egyptian International Pharmaceutical Industries Company (Egypt). SAB authentic powder was purchased from Beijing Mesochem Technology Co. Ltd, China. Lobachemie Pvt. Ltd (India) and El-Nasr Chemical Company (Egypt) were the suppliers of analytical-grade methanol and sulfuric acid. The distilled water was filtered through 0.45 μm disposable filters before use. A commercial product (Entresto 50[®], batch number TDW73) was studied.

2.2. Instrumentation

The spectrofluorometer used was the G9800A Cary Eclipse fluorescence spectrophotometer (Agilent Technologies, USA). Its light source was a long-life Xenon flash lamp. Samples were placed in a 1 cm quartz cell. The grating monochromators were adjusted at 5 nm slit width for both excitation and emission. The operation voltage was 600 volts.

2.3. Standard solutions

Individual VAS and SAB stock solutions were prepared in methanol at a concentration of 100 $\mu\text{g mL}^{-1}$. They were appropriately diluted with methanol to 5 $\mu\text{g mL}^{-1}$ working solutions.

2.4. Construction of calibration graphs

Various volumes from VAS and SAB working standard solutions were poured into separate 10 mL volumetric flasks. Each flask was completed to the mark with 0.1 M sulfuric acid to obtain the concentration ranges of 60–200 and 17–190 ng mL^{-1} for VAS and SAB, respectively, in the SF-D¹ coupled approach. The concentration ranges of 80–200 and 30–350 ng mL^{-1} for VAS and SAB were also prepared for the SF- $\Delta\lambda$ coupled approach. Spectrofluorimetric measurements were carried out in triplicates, using synchronous mode at constant wavelength $\Delta\lambda$ of 130 nm between excitation and emission wavelengths. The zero-order synchronous spectrofluorimetric spectra were recorded and corrected by subtracting the blank spectrum.



2.4.1. SF-D¹ method. The first-order derivative spectra were derived at $\Delta\lambda$ of 5 nm. SF-D¹ amplitudes were measured for VAS at 230 nm (zero-crossing of SAB) and for SAB at 211 nm (zero-crossing of VAS).

2.4.2. SF- $\Delta\lambda$ method. Difference signals in the corrected synchronous values at (226–241) nm were determined for VAS where SAB contribution is zero. For SAB, signals were derived at (208–239) nm where VAS contribution is zero.

The measured amplitudes in each method were correlated to the corresponding concentrations, and regression equations for VAS and SAB were acquired.

2.5. Procedure for tablet analysis

VAS and SAB were determined in their combined commercial tablets (Entresto 50®, batch number TDW73). A part of the powdered ground tablets containing 8.5 and 8.0 mg VAS and SAB, respectively, were used. It was placed in a 100 mL volumetric flask. The flask was filled with methanol up to half its volume and placed in the sonicator for 30 minutes. The solution was completed with methanol till it reached the volume mark. One hundred μL of this filtrate was appropriately diluted with 0.1 M sulfuric acid to 100 mL volume. Finally, a clear tablet extract containing 85 and 80 ng mL^{-1} VAS and SAB was obtained. Then, the steps in the “Generation of regression equations” were followed for both hybrid approaches.

3. Results and discussions

3.1. Method optimization

In SF, the excitation and emission wavelengths are scanned at the same time. A wavelength difference ($\Delta\lambda$) is determined and set between λ_{em} and λ_{ex} throughout all measurements, resulting in a significantly narrower band. Consequently, a simpler spectrum is obtained, and the resolution and selectivity are enhanced. Thus, SF represents a sensitive approach for concurrently analyzing multicomponent samples. However, SF may not achieve the required resolution for all components in a mixture. In such cases, coupling it with various mathematical treatments will be beneficial to obtain enhanced performance. As a coupled technique, the derivative approach presents a synergistic role in enhancing SF signals concerning resolution and sensitivity. In addition, dual-wavelength presents another coupled technique that is coadjutant to SF. Its signal arises from the difference in SF amplitude at two wavelengths, and the correct choice for the measured analyte is the most important step in this technique to reach a difference signal for the co-existing analyte at a non-significant level.

Regarding the proposed methods, VAS has a proven native fluorescence.^{32,33} For SF spectra generation, $\Delta\lambda$ ($\lambda_{\text{em}} - \lambda_{\text{ex}}$) of 130 nm was chosen as the best compromise between peak separation, peak integrity, and signal intensities. As a diluting solvent, 0.1 M sulfuric was chosen for the highest synchronous response for both drugs (Fig. 2). Also, the synchronous spectra of VAS and SAB were not well separated. Based on these results, we employed two hybrid approaches to resolve VAS and SAB completely, resulting in high assay reliability.

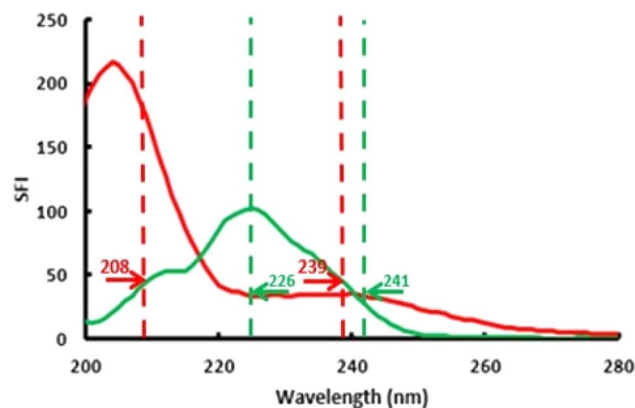


Fig. 2 Dual wavelength points on the synchronous fluorescence spectra ($\Delta\lambda = 130$ nm) of 200 ng mL^{-1} VAS (green line) and 190 ng mL^{-1} SAB (red line) in $0.1 \text{ M H}_2\text{SO}_4$.

In the first approach, spectrofluorimetric synchronous spectra were derivatized. Derivatization at the first order was enough to achieve optimum resolution between VAS and SAB. Higher-order derivatives did not provide better performance. Derivative wavelength increment ($\Delta\lambda$) is critical to control spectral resolution, sensitivity, and background noise.^{34–36} $\Delta\lambda$ of 5 nm was selected as it achieved the most acceptable compromise between these items. To eliminate cross-interference, SF-D¹ amplitudes of each drug were measured at the zero-crossing of the other component. Thus, VAS and SAB were measured at 230 and 211 nm, respectively (Fig. 3).

Regarding the second approach, the simple dual wavelength method was coupled.³⁷ For VAS determination, the interference of SAB was eliminated by recording the difference in VAS synchronous values at wavelengths where SAB has the same contribution (226 and 241 nm). For SAB quantification, the interference of VAS was eliminated by subtracting synchronous values at 239 nm from the response at 208 nm where VAS has the same signal intensity (Fig. 2).

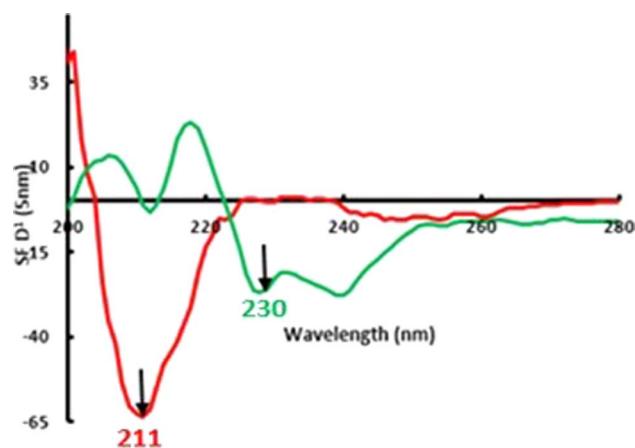
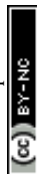


Fig. 3 Zero-crossing points on the first derivative synchronous spectra ($\Delta\lambda = 5$ nm) of 200 ng mL^{-1} VAS (green line) and 190 ng mL^{-1} SAB (red line) in $0.1 \text{ M H}_2\text{SO}_4$.



3.2. Validation of the proposed methods

ICH guidelines were followed to validate the suggested SF-D¹ and SF-Δλ methods.³⁸

3.2.1. Selectivity. Solutions of different VAS/SAB concentrations (200/100, 200/190, and 100/190 ng mL⁻¹) were prepared in the laboratory. The triplicate approach was used to analyze them by the suggested two hybrid methods for VAS and SAB contents. The resulting mean % recoveries did not exceed ±2% for VAS and SAB (Table 1). These acceptable values proved the absence of cross-interference between the two analytes.

3.2.2. Accuracy and precision. Ground powdered tablets of the VAS/SAB supramolecular complex were employed. According to the labeled claim, an amount of it was weighed to contain 8.5 and 8.0 mg VAS and SAB, respectively. Then, as described in section (2.5), the steps were followed to prepare the tablet extract solution at 85 and 80 ng mL⁻¹ of VAS and SAB, respectively. Spiking with authentic VAS and SAB powders was carried out at 80, 100, and 120% of the label claim. The above-mentioned prepared concentrations allowed the acquisition of VAS and SAB final concentration levels to be analyzed after standard addition within the linearity scales of Table 1. Then, the assay was performed in triplicates for each method. Er% values were low enough (<±1%) to ensure the accuracy of each method, Table 1.

The previous laboratory-prepared VAS/SAB mixtures were analyzed in triplicates. Analysis was performed on the same day or three days to evaluate the intra- or inter-day precision approaches, respectively. RSD% values (<2%) were within the limits that fulfill the precision parameter, Table 1.

3.2.3. Linearity and range. Five VAS and SAB solutions of suitable concentrations were analyzed for each hybrid method. All linearity parameters (including *r*, *r*², *S*_a, *S*_b, *S*_{y/x}, *S*_b%, *F* and

Table 2 Assay of VAS and SAB in their combined tablets using the proposed hybrid methods (*n* = 5)

| Entresto ^{®a} | Mean% recovery ± SD | <i>t</i> ^c | <i>F</i> ^c |
|------------------------------|---------------------|-----------------------|-----------------------|
| VAS | | | |
| SF-D ¹ | 100.56 ± 0.98 | 1.06 | 1.43 |
| SF-Δλ | 101.10 ± 1.05 | 1.62 | 1.23 |
| Reported method ^b | 99.62 ± 1.17 | | |
| SAB | | | |
| SF-D ¹ | 99.89 ± 0.72 | 2.18 | 1.78 |
| SF-Δλ | 100.63 ± 0.59 | 1.01 | 2.60 |
| Reported method | 101.21 ± 0.96 | | |

^a Labeled to contain 25.70 mg VAS and 24.30 mg SAB per tablet (batch number TDW73). ^b First derivative spectrofluorimetry in 0.1 M H₂SO₄ at 448 and 300 nm for VAS and SAB, respectively. ^c Theoretical values of *t* and *F* at *p* = 0.05 are 2.31 and 6.39, respectively.

significance *F*) were calculated. The correlation coefficient (*r*) should be ≥ 0.999 as a measure of correlation between concentration and signal. R-squared (*r*²) should be ≥ 0.99 as a better linearity indicator. Linearity was further evaluated by the relative standard deviation of slope (*S*_b%); it should be less than 2%. The experimental points become closer to the straight line as the standard deviation of the regression (*S*_{y/x}) decreases. The standard deviation of intercept (*S*_a) and slope (*S*_b) are important parameters, and their low values indicate low uncertainties of intercept and slope. As such, they have implications for method sensitivity, LOD, and LOQ. In addition, increased *F*-values (decreased significance *F*-*F*-values) indicate a high and low mean of squares due to regression and residuals, respectively. As shown in Table 1, values of linearity statistical parameters were good enough to conclude that the responses

Table 1 Validation parameters of the proposed hybrid methods

| Parameters | SF-D ¹ | | SF-Δλ | |
|--|-------------------------|-------------------------|-------------------------|-------------------------|
| | VAS | SAB | VAS | SAB |
| Selectivity ^a | 100.88 | 101.13 | 101.52 | 99.04 |
| Accuracy ^b | 101.58 | 99.60 | 99.81 | 100.74 |
| Intra-day precision ^c | 1.10 | 0.81 | 1.37 | 0.53 |
| Inter-day precision ^c | 1.34 | 1.07 | 0.94 | 0.79 |
| Linearity range ^d | 60–200 | 17–190 | 80–600 | 30–350 |
| LOD ^d | 7.03 | 5.06 | 14.39 | 8.91 |
| LOQ ^d | 23.44 | 16.87 | 47.96 | 29.72 |
| Intercept (<i>a</i>) | 2.31 | −0.81 | −0.99 | 7.28 |
| Slope (<i>b</i>) | 3.47 × 10 ⁻² | 0.41 | 5.95 × 10 ⁻² | 1.17 |
| Correlation coefficient (<i>r</i>) | 0.9995 | 0.9984 | 0.9996 | 0.9993 |
| <i>r</i> ² | 0.9991 | 0.9968 | 0.9992 | 0.9990 |
| <i>S</i> _a ^e | 8.13 × 10 ⁻² | 0.69 | 0.29 | 3.46 |
| <i>S</i> _b ^e | 6.14 × 10 ⁻⁴ | 8.23 × 10 ⁻³ | 9.71 × 10 ⁻⁴ | 2.21 × 10 ⁻² |
| <i>S</i> _{y/x} ^(b,e) | 6.36 × 10 ⁻² | 1.56 | 0.42 | 6.17 |
| <i>S</i> _b % | 1.77 | 2.01 | 1.63 | 1.89 |
| <i>F</i> | 3189.04 | 946.39 | 3745.824 | 2145.17 |
| Significance <i>F</i> | 1.22 × 10 ⁻⁵ | 7.55 × 10 ⁻⁵ | 9.61 10 ⁻⁶ | 2.22 × 10 ⁻⁵ |

^a Calculated as mean % recovery from synthetic mixtures of three different ratios (*n* = 9). ^b Calculated as (mean % recovery ± RSD%) from standard addition method synthetic mixtures at three concentration levels (*n* = 9). ^c Calculated as mean RSD% from synthetic mixtures of three different ratios (*n* = 9). ^d Concentration in ng mL⁻¹. ^e *S*_a is the standard deviation of intercept, *S*_b is the standard deviation of the slope, and *S*_{y/x} is the standard deviation of residuals.



Table 3 Analytical eco-scale (a) and GAPI (b) for the greenness evaluation of the proposed hybrid methods

| (a) Analytical eco-scale | | |
|--|-------------------------|----------------------|
| Eco-scale parameter | Subtotal penalty points | Total Penalty points |
| 0.1 M H ₂ SO ₄ amount (<10 mL) | 1 | 2 ^b |
| 0.1 M H ₂ SO ₄ hazard | 2 ^a | |
| Methanol amount (<1 mL) | 1 | 6 ^b |
| Methanol hazard | 6 ^a | |
| Spectrofluorimeter (≤0.1 kW h/sample) | | 0 |
| Sonicator (≤0.1 kW h/sample) | | 0 |
| Waste volume (10 mL) | 3 | 6 ^c |
| Waste treatment (none) | 3 | |
| Overall penalty points | | 14 |
| Total eco-scale score | | 86 |

| (b) GAPI | |
|--|------------------------------|
| GAPI parameter | Status |
| Sample preparation | |
| (1) Collection | At-line |
| (2) Preservation | None |
| (3) Transport | None |
| (4) Storage | Under none conditions |
| (5) Type of method (direct or indirect) | Filtration |
| (6) Scale of extraction | None |
| (7) Solvents/reagents used | None |
| (8) Additional treatment | None |
| Reagents and solvents used (9–11) | |
| (9) Amount | < 10 mL |
| (10) Health hazard | NFPA health hazard score = 3 |
| (11) Safety hazard | NFPA fire hazard score = 0 |
| Instrumentation | |
| (12) Energy | ≤ 0.1 kW h/sample |
| (13) Occupational hazard | — |
| (14) Waste | 1–10 mL |
| (15) Waste treatment | No treatment |
| Quantification | Yes |

^a Calculated as (GSH hazard pictograms × degree of hazard). ^b Calculated as (amount PP × hazard PP). ^c Calculated as (waste volume PP + waste treatment PP).

were linearly correlated to the corresponding concentrations for both VAS and SAB.³⁹

3.2.4. Limit of detection and limit of quantitation. [The standard error of the *y*-intercept (*S_a*)/slope of the calibration graph (*b*)] was multiplied by 3 and 10 to calculate LOD and LOQ, respectively. Their values could confirm the excellent sensitivity of the proposed methods (Table 1).³⁹

3.2.5. Robustness. The previously, laboratory-prepared VAS/SAB mixtures were analyzed. In the triplicate set, sulfuric acid concentration was liable to show ± 0.02 M intended variation. Similarly, the detection wavelength and Synchronous Δλ were varied by ± 1 nm. The resulting RSD% did not exceed 2%, fulfilling the robustness requirement.

3.3. Assay of pharmaceutical tablets

The VAS/SAB solution was prepared by extraction from their commercial combined tablets. Mean% recovery and RSD% were

obtained from 5-replicate analysis by the suggested SF-D¹ and SF-Δλ methods. Their values were acceptable, so interference from tablet excipients was predicted to be insignificant. The

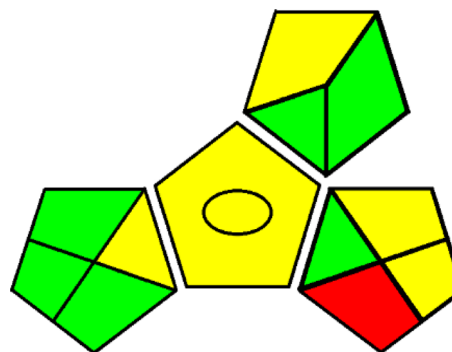


Fig. 4 GAPI pictogram of the proposed hybrid methods.



suggested method was statistically compared with the reported one.²⁵ Tht- and F-test outputs were within the acceptable theoretical limits (Table 2). Thus, the methods proved highly reliable with respect to the VAS/SAB fixed-dose combination assay.

3.4. Evaluation of the greenness of the method

Green analytical chemistry (GAC) aims to encourage chemists to consider the effect of their analytical procedures on the environment, personal safety, and health. GAC necessitates implementing dedicated approaches to outline the green profiles of the analytical methods. The analytical eco-scale tool was employed as a preliminary judgment on the greenness of the proposed methods.^{40–43} It quantifies a method's green

parameters. It is based on penalty points (PPs) to reach a total numerical score as an accurate description of greenness. This score is calculated by subtracting penalty points of reagent toxicity, instrument energy consumption, and waste generation from a base of 100. The result is determined by the number of residual points: >75 represents excellent green analysis, >50 indicates acceptable green analysis, and <50 means inadequate green analysis. Table 3 presents the detailed penalty points for each reagent and instrument employed in the suggested hybrid methods. For both methods, the overall penalty points were 14 only. Thus, the total eco-scale score was 86; much higher than 75. This proved the excellent green profile of the proposed hybrid methods.

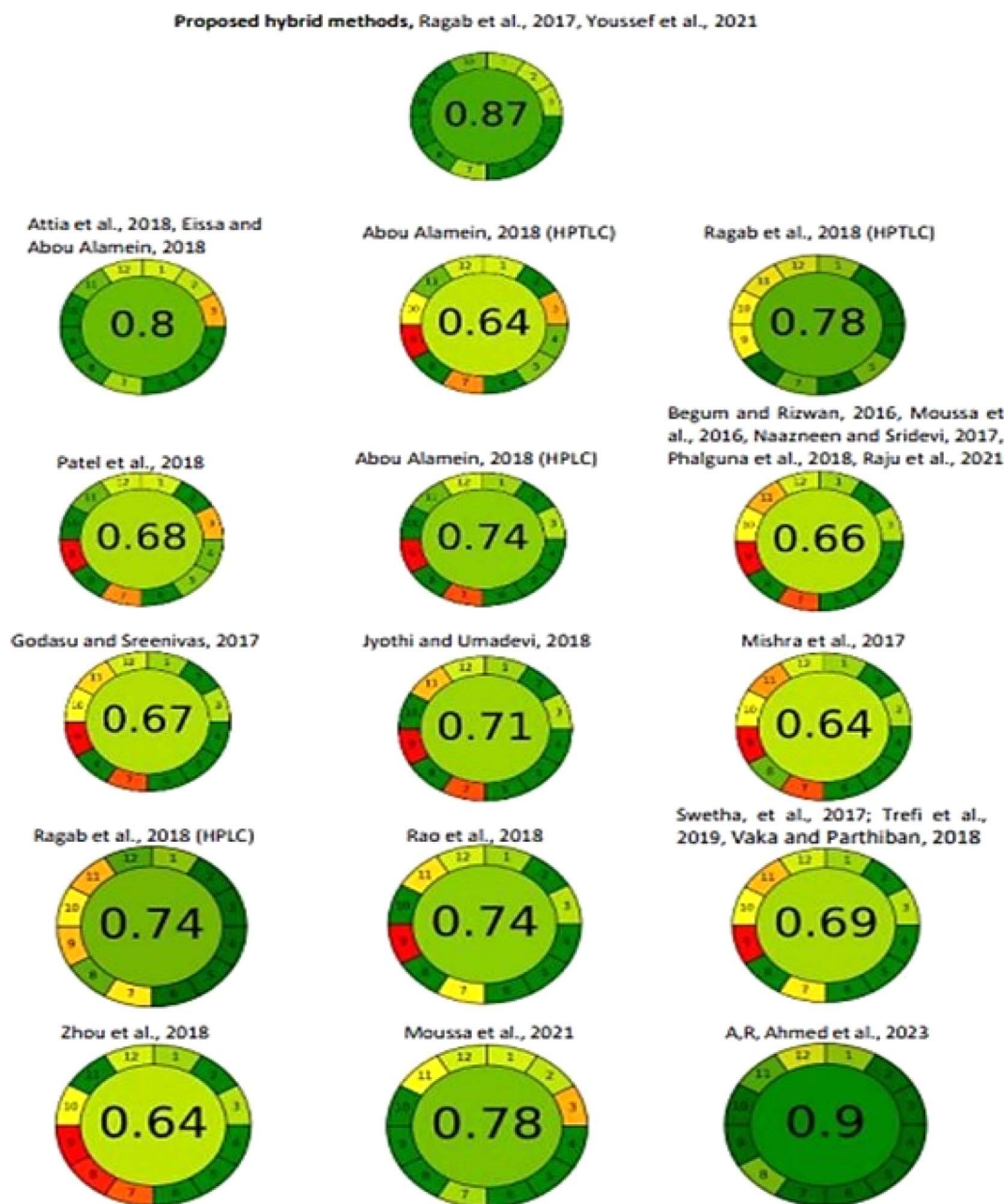


Fig. 5 Comparative AGREE charts of the proposed and reported methods.



A green analytical procedure index (GAPI) metric was applied to attain an inclusive greenness evaluation and rational ranking of the applied methods.^{44–46} This metric offers an ecological assessment of the entire method, from sample preparation to the final determination step. It is represented as a colored pictogram composed of 15 evaluated parameters. These parameters include sample collection, preservation, transportation, solvents and reagents, energy consumed, waste generated, and other instrumental parameters. Each section is colored green, yellow, or red, indicating a low, medium, or high negative environmental impact. After inspection of the developed methods' GAPI pictograms visually, it was found that both proposed hybrid methods are ranked as green since they both obtained six green zones, five yellow zones, and only one red zone (Fig. 4, Table 3).

The recent AGREE tool was implemented to provide a more comprehensive and informative appraisal of the methods' greenness.^{47,48} This downloadable software was used to evaluate the method's greenness based on the 12 GAC principles (SIGNIFICANCE). After inputting the 12 variables of each method, automatically, an output of a clock-like graph was generated. This graph comprises 12 sections, colored from deep green to red, with a total numerical score in the center. The score is a fraction of unity; it ranges from 0 to 1. From the illustrated AGREE chart of the proposed SF-D¹ and SF-Δλ methods (Fig. 5), the AGREE score was found to be 0.87, indicating that the developed hybrid methods are green.

In addition, the AGREE tool was applied to the proposed *versus* all reported methods, in a comparative approach (Fig. 5). The proposed hybrid method revealed an AGREE score much

higher than those of almost all the reported methods. Two reported spectrofluorimetrics had the same AGREE score as the suggested ones. Only the recently reported MEKC method showed a slightly higher AGREE score in a non-significant way.

3.5. Comparative evaluation of the method whiteness

Method whiteness was comparatively evaluated for the proposed method and previously reported methods *via* the RGB 12 approach, as illustrated in ESI 1† and Table 4. The proposed SF-Δλ and SF-D¹ methods showed the second and the fifth rank, respectively.

The scoring of R2, R3, R4, G1, G2, G4, B1, B2, and B3 metrics followed relative ranking. Despite this scoring approach being accurate as a relative score for comparative purposes, it gave rise to underestimated absolute scores. Therefore, to obtain the right conclusion from the comparative whiteness study, a whiteness rank was calculated as shown in Table 4, illustrating excellent whiteness ranks of the proposed methods.

3.6. Comparative evaluation of the method performance

While spectrofluorimetric spectra depend on λ_{ex} and λ_{em} , spectrophotometric ones depend on λ_{max} only.⁴⁹ Hence, the reported spectrophotometric methods^{7,8} have lower spectral resolution than the proposed methods. In addition, applying the synchronous mode improves their selectivity over the reported traditional spectrofluorimetric ones.^{25,26} Moreover, one of them²⁶ employs less green and more expensive solvent (acetonitrile). Compared with the reported synchronous-derivative ratio method,²⁷ coupling with derivative and dual-

Table 4 Summary of the whiteness comparative evaluation featuring the whiteness ranks of the studied methods

| Method number | Method name | R (%) | G (%) | B (%) | Whiteness (%) | Whiteness (rank) | Reference |
|---------------|------------------------------|-------|-------|-------|---------------|------------------|-----------|
| 1 | Proposed (SF-D1) | 59.3 | 60.8 | 72.8 | 64.3 | 5 | |
| 2 | Proposed (SF-Δλ) | 70.7 | 60.8 | 72.8 | 68.1 | 2 | |
| 3 | Ragab et al., 2017 | 58.0 | 73.8 | 57.7 | 63.2 | 9 | 25 |
| 4 | Youssef et al., 2021 | 69.1 | 59.2 | 74.3 | 67.6 | 3 | 27 |
| 5 | Attia et al., 2018 | 63.6 | 71.7 | 47.5 | 60.9 | 11 | 7 |
| 6 | Eissa and Abou Alamein, 2018 | 53.6 | 73.8 | 48.5 | 58.6 | 17 | 8 |
| 7 | Godasu and Sreenivas, 2017 | 61.6 | 81.1 | 47.5 | 63.4 | 6 | 21 |
| 8 | Ragab et al., 2018 (HPLC-FD) | 96.2 | 61.3 | 54.1 | 70.5 | 1 | 10 |
| 9 | Zhou et al., 2018 | 56.5 | 34.7 | 53.6 | 48.3 | 23 | 12 |
| 10 | Abou Alamein, 2018 (HPTLC) | 81.7 | 72.7 | 41.3 | 65.2 | 4 | 9 |
| 11 | Abou Alamein, 2018 (HPLC) | 72.2 | 56.1 | 41.8 | 56.7 | 19 | 9 |
| 12 | Jyothi and Umadevi, 2018 | 79.2 | 58.7 | 44.4 | 60.7 | 12 | 15 |
| 13 | Rao et al., 2018 | 63.9 | 75.9 | 33.4 | 57.7 | 18 | 17 |
| 14 | Moussa et al., 2021 | 45.9 | 82.6 | 50.6 | 59.7 | 14 | 26 |
| 15 | Ragab et al., 2018 (HPTLC) | 71.1 | 64.9 | 53.5 | 63.2 | 8 | 10 |
| 16 | Moussa et al., 2018 | 62.2 | 51.9 | 53.3 | 55.8 | 20 | 22 |
| 17 | Naazneen and Sridevi, 2017 | 71.1 | 57.1 | 47.8 | 58.7 | 16 | 18 |
| 18 | Phalguni et al., 2018 | 63.9 | 66.5 | 47.5 | 59.3 | 15 | 13 |
| 19 | Raju et al., 2021 | 73.7 | 59.7 | 45.9 | 59.8 | 13 | 24 |
| 20 | Mishra et al. 2017 | 73.3 | 72.2 | 44.4 | 63.3 | 7 | 23 |
| 21 | Swetha et al., 2017 | 76.9 | 56.6 | 54.8 | 62.7 | 10 | 19 |
| 22 | Trefi et al., 2019 | 30.3 | 57.1 | 43.9 | 43.8 | 24 | 20 |
| 23 | Vaka and Parthiban, 2018 | 53.2 | 70.1 | 39.2 | 54.2 | 22 | 14 |



wavelength approaches present much simpler techniques. Thus, the suggested methods do not necessitate the presence of a highly trained operator or highly advanced spectrofluorimeter software.

As spectroscopic methods, both proposed methods are much easier, faster and cheaper than the reported chromatographic ones. Moreover, unlike the reported chromatographic methods that consume organic solvents, the proposed hybrid methods are applied in the aqueous phase. As a consequence, they are more eco-friendly than the reported chromatographic ones.^{9–24,28} Finally, the proposed methods showed more sensitive calibration ranges than most reported methods. Only one reported HPLC-Flu method¹⁰ has a linearity scale at slightly lower concentrations. However, it is limited by its chromatographic nature. In addition, the recently reported chromatographic method²⁸ may be subjected to poor reproducibility due to its MEKC nature. This is referred to as the high sensitivity to minor changes in BGE pH and composition. In conclusion, the highly sensitive linearity ranges, using small amounts of green solvents and ease of operation, and the presented highly sustainable profiles provide experimental evidence for the superiority of the proposed methods over the previously reported ones. In addition, the suggested methods experimentally succeeded in assaying VAS/SAB combined tablets with high reliability concerning accuracy and precision (Section 3.3).

4. Conclusions

The present research provides two reliable hybrid assays for simultaneously analyzing VAS and SAB in their combined tablets. It depends on the synchronous spectrofluorimetric measurement, followed by first-derivative or dual-wavelength mathematical treatments. The proposed methods showed several privileges over the reported ones concerning simplicity, analysis time, and sensitivity. Analytical eco-scale, GAPI, and AGREE metrics were used to assess the greenness of the developed methods. The proposed methods are proven to be white, green, economical, with no need for organic solvents, and consume less time. Their high accuracy and precision recommend the methods used in the routine industrial assay of LCZ696 (VAS/SAB) tablets. The affordable cost potentiates their application in all countries, regardless of their economic state.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

“This research work was funded by Institutional Fund Projects under grant no. (IFPIP: 841-166-1443). The authors gratefully acknowledge technical and financial support provided by the Ministry of Education and King Abdulaziz University, DSR, Jeddah, Saudi Arabia”.

References

- 1 Wellness, Emory Healthcare, *Heart Failure Statistics*, 2018.
- 2 M. B. Andersen, U. Simonsen, M. Wehland, J. Pietsch and D. Grimm, *Basic Clin. Pharmacol. Toxicol.*, 2016, **118**, 14–22.
- 3 Z. Qianqian, C. Qixin and Y. H. Lichun, *China: Worldwide applications; 105622535A*, 2015, p. 15.
- 4 D. G. Waller and A. P. Sampson, in *Medical Pharmacology and Therapeutics*, Elsevier Science Publishing Co Inc, 5th edn, 2018, pp. 131–142.
- 5 P. Ponikowski, A. A. Voors, S. D. Anker, H. Bueno, J. G. F. Cleland, A. J. S. Coats and C. Davies, *Eur. Heart J.*, 2016, **37**, 2129–2200.
- 6 C. Yancy, M. Jessup, B. Bozkurt, J. Butler, D. E. Casey, M. M. Colvin, M. H. Drazner, G. S. Filippatos, G. C. Fonarow, M. M. Givertz, S. M. Hollenberg, J. A. Lindenfeld, F. A. Masoudi, P. E. McBride, P. N. Peterson, L. W. Stevenson and C. Westlake, *Circulation*, 2017, **136**, e137–e161.
- 7 K. Attia, M. Nassar, A. El-Olemy and S. Ramzy, *J. Adv. Res.*, 2018, **2**, 133–141.
- 8 M. S. Eissa and A. M. Abou Al Alamein, *Spectrochim. Acta, Part A*, 2018, **193**, 365–374.
- 9 A. Abou Al Alamein, *J. Appl. Pharm. Sci.*, 2018, **8**, 011–017.
- 10 M. A. A. Ragab, S. M. Galal, M. A. Korany and A. R. Ahmed, *J. Chromatogr. Sci.*, 2018, **56**, 498–509.
- 11 G. Patel, A. Vohra and S. Shah, *International Journal in Pharmaceutical & Nanotechnology*, 2018, **11**, 4208–4218.
- 12 S. Trefi, Y. Bitar and V. Gilard, *Res. J. Pharm. Technol.*, 2019, **12**, 1017–1022.
- 13 S. K. Godasu and S. A. Sreenivas, *Eur. J. Biomed. Pharmaceut. Sci.*, 2017, **4**, 640–645.
- 14 B. A. Moussa, H. M. A. Hashem, M. A. Mahrouse and S. T. Mahmoud, *Chromatographia*, 2018, **81**, 139–156.
- 15 S. Mishra, C. J. Patel and M. M. Patel, *Int. J. Appl. Pharm.*, 2017, **9**, 1–8.
- 16 T. N. Raju, D. R. Kumar and D. Ramachandran, *J. Appl. Chem.*, 2017, **6**(5), 1004–1011.
- 17 L. Zhou, L. Zou, L. Sun, H. Zhang, W. Hui and Q. Zou, *Anal. Methods*, 2018, **10**, 1046–1053.
- 18 Y. Phalgun, N. Jahan, N. Indraja and G. Satheesh, *Asian J. Res. Pharm. Sci. Biotechnol.*, 2018, **8**, 09–16.
- 19 S. Vaka and P. Parthiban, *Int. J. Res.*, 2018, **4**, 17–24.
- 20 U. Jyothi and P. Umadevi, *J. Pharmaceut. Sci. Res.*, 2018, **10**, 2201–2204.
- 21 F. Begum and Sh. Rizwan, *International Journal of Farmacia*, 2016, **2**, 79–87.
- 22 B. S. Rao, M. V. Reddy and B. S. Rao, *World J. Pharm. Pharm. Sci.*, 2018, **7**, 1435–1451.
- 23 S. Naazneen and A. Sridevi, *Int. J. Appl. Pharm.*, 2017, **9**, 9–15.
- 24 V. Swetha, S. V. U. M. Prasad and M. Indupriya, *World Journal of Pharmaceutical and Life Sciences*, 2017, **3**, 185–191.
- 25 M. A. A. Ragab, S. M. Galal, M. A. Korany and A. R. Ahmed, *Luminescence*, 2017, **32**, 1417–1425.



- 26 B. A. Moussa, H. M. A. Hashem, M. Alphonse Mahrouse, M. A. Mahrouse and S. T. Mahmoud, *Spectrochim. Acta, Part A*, 2021, **253**, 119613.
- 27 R. M. Youssef, S. A. El-Nahass, S. A. Soliman and S. E. Younis, *Spectrochim. Acta, Part A*, 2021, **256**, 119748.
- 28 A. R. Ahmed, S. M. Galal, M. A. Korany and M. A. A. Ragab, *Sustainable Chem. Pharm.*, 2023, **33**, 101066.
- 29 Y.-Q. Li, X.-Y. Li, A. A. F. Shindi, Z.-X. Zou, Q. Liu, L.-R. Lin and N. Li, Synchronous Fluorescence Spectroscopy and Its Applications in Clinical Analysis and Food Safety, *Evaluation*, 2012, 95–117.
- 30 P. M. Nowak, R. Wietecha-Postuszny and J. Pawliszyn, *Trends Anal. Chem.*, 2021, **138**, 116223.
- 31 M. A. Korany, R. M. Youssef, M. A. A. Ragab and M. A. Afify, *Microchem. J.*, 2024, **196**, 109616.
- 32 R. A. Shaalan and T. S. Bela, *Drug Test. Anal.*, 2010, **2**, 489–493.
- 33 A. M. Zeid, R. Aboshabana and F. A. Ibrahim, *Spectrochim. Acta, Part A*, 2022, **26**, 120591.
- 34 S. E. Younis, S. A. El-Nahass, R. M. Youssef and S. A. Soliman, *Microchem. J.*, 2020, **1**, 156.
- 35 R. M. Youssef, A. M. Abdelhafez, E. M. Hassan and D. A. Gawad, *Spectrochim. Acta, Part A*, 2022, **271**, 120904.
- 36 R. M. Youssef, *Saudi Pharm. J.*, 2010, **18**, 45–49.
- 37 K. A. M. Attia, A. El-Olemy, S. Ramzy, A. H. Abdelazim, M. A. Hasan and R. F. Abdel-Kareem, *Spectrochim. Acta, Part A*, 2021, **248**.
- 38 ICH, *Harmonised Tripartite Guideline.: Text and Methodology Q2(R1)*, Harmon Tripart Guidel, 1995.
- 39 J. Miller and J. Miller, *Statistics and Chemometrics for Analytical Chemistry*, 4th edn, 2000.
- 40 A. F. El-Yazbi, E. F. Khamis, R. M. Youssef, M. A. El-Sayed and F. M. Aboukhalil, *Helyion*, 2020, **6**(9).
- 41 A. F. El-Yazbi, F. M. Aboukhalil, E. F. Khamis, R. M. Youssef and M. A. El-Sayed, *Microchem. J.*, 2021, **163**, 105900.
- 42 M. Tobiszewski, M. Marć, A. Gałuszka and J. Namieśnik, *Molecules*, 2015, **20**, 10928–10946.
- 43 A. Gałuszka, P. Konieczka, Z. M. Migaszewski and J. Namiesnik, *Trends Anal. Chem.*, 2012, **37**, 61–72.
- 44 A. F. El-Yazbi, F. M. Aboukhalil, E. F. Khamis, R. M. Youssef and M. A. El-Sayed, *JPC (J. Planar Chromatogr.) - Mod. TLC*, 2021, **34**, 455–466.
- 45 A. F. El-Yazbi, F. M. Aboukhalil, E. F. Khamis, R. M. Youssef and M. A. El-Sayed, *Beni-Suef University Journal of Basic and Applied Sciences*, 2022, vol. 11, p. 15.
- 46 J. Płotka-Wasyłka, new tool for the evaluation of the analytical procedure: Green Analytical Procedure Index, *Talanta*, 2018.
- 47 A. H. Abo-Gharam and D. S. El-Kafrawy, *Sustainable Chem. Pharm.*, 2022, **29**, 14.
- 48 W. W. Francisco Pena-Pereira and M. Tobiszewski, *Anal. Chem.*, 2020, **92**, 10076–10082.
- 49 A. Nahata, *Pharm. Anal. Acta*, 2011, **02**, 1000107e.

