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Facile synthesis of substituted 2-arylbenzo[*b*]thiophen-3-ols to form novel triazole hybrids using click chemistry†

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An efficient one-pot method is proposed for the synthesis of 2-arylbenzo[*b*]thiophen-3-ols from 2-mercaptobenzoic acid and various substituted aryl bromomethyl ketones in the presence of triethylamine. The reaction is likely to proceed through S_N2-type nucleophilic attack of the sulfhydryl group in thiosalicylic acid on bromomethyl ketone in the presence of a base to afford sulfanylbenzoic acid, which undergoes an intramolecular cyclization *in situ* to furnish 2-arylbenzo[*b*]thiophen-3-ol in high yield. To investigate the utility of the synthesized benzothiophene scaffold, an alkyne moiety was introduced at the 3-hydroxy position and subsequently subjected to a click reaction to form novel benzothiophene-triazole hybrids in good yields. A simple and straightforward approach to synthesizing 2-arylbenzo[*b*]thiophen-3-ols can open new avenues for discovering novel biological and pharmaceutical compounds.

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1 Introduction

Heterocycles serve as fundamental components in numerous biologically relevant natural and synthetic compounds. This notion is supported by the fact that a large number of the USA Food and Drug Administration (FDA)-approved drugs contain one or several heterocyclic rings in their structures.^{1–3} Therefore, access to a chemical library of small molecules containing one or more “privileged” heterocyclic rings is generally considered a prerequisite for effective drug discovery endeavors.^{4,5} Among them, sulfur-containing heterocycles are gaining attention in organic synthesis due to their applications in materials sciences, pharmaceutical industries, and food chemistry.^{6,7} Benzo[*b*]thiophene is a versatile sulfur-containing heterocyclic scaffold, which frequently appears in pharmaceutically interesting compounds with a diverse range of biological activities.^{8–12} For instance, various drugs including raloxifene (1), arzoxifene (2), rintodestrant (3), and LSZ102 (4) feature the benzo[*b*]thiophene core and are currently clinically utilized for the treatment of estrogen-receptor-positive breast cancer (Fig. 1). Termed selective estrogen receptor modulators

(SERMs), both 1 and 2 are used for the prevention and treatment of breast cancer and osteoporosis in postmenopausal women.^{13–20} On the other hand, 3 and 4 act as selective estrogen receptor degraders (SERDs), undergoing evaluation for the elimination of estrogen receptor- α from cancer cells in patients diagnosed with aggressive breast cancer.^{21–23} Along with SERM/SERD activities, several benzothiophene-based compounds show anticancer activities by targeting various cellular proteins such as tubulin, topoisomerase, and histone deacetylase.^{24–28}

To exhibit specificity, substituents at the C-2 and the C-3 positions play an important role in target binding and eliciting diverse biological activities associated with benzo[*b*]thiophenes. Upon a cursory inspection of the structures given in Fig. 1, it seems plausible to discern that the 2-aryl and/or 2-aryl moiety at the C-2/C-3 position of the benzo[*b*]thiophene core may be an essential ingredient for the SERM/SERD-like activities of 1–4. We envision that the 2-arylbenzo[*b*]thiophen-3-ol core could provide access to diverse benzo[*b*]thiophene analogs for discovering new SERMs/SERDs. To extend our efforts on the discovery of early-phase inhibitor leads for developing antiviral and anticancer drugs,^{29–32} we herein report a simple, one-pot synthetic method for the preparation of substituted 2-aryl-benzo[*b*]thiophen-3-ol derivatives (5a-s) by reacting 2-mercaptobenzoic acid (6) with substituted aryl bromomethyl ketones (8a-s) in the presence of triethylamine in DMF. To further extend the scope of a 2-aryl-benzo[*b*]thiophen-3-ol substrate (5), we introduced an alkyne moiety at the 3-hydroxy position, conducted a click reaction, and prepared novel benzothiophene-triazole hybrids. We believe that this robust method for the direct synthesis of 5 starting from 2-

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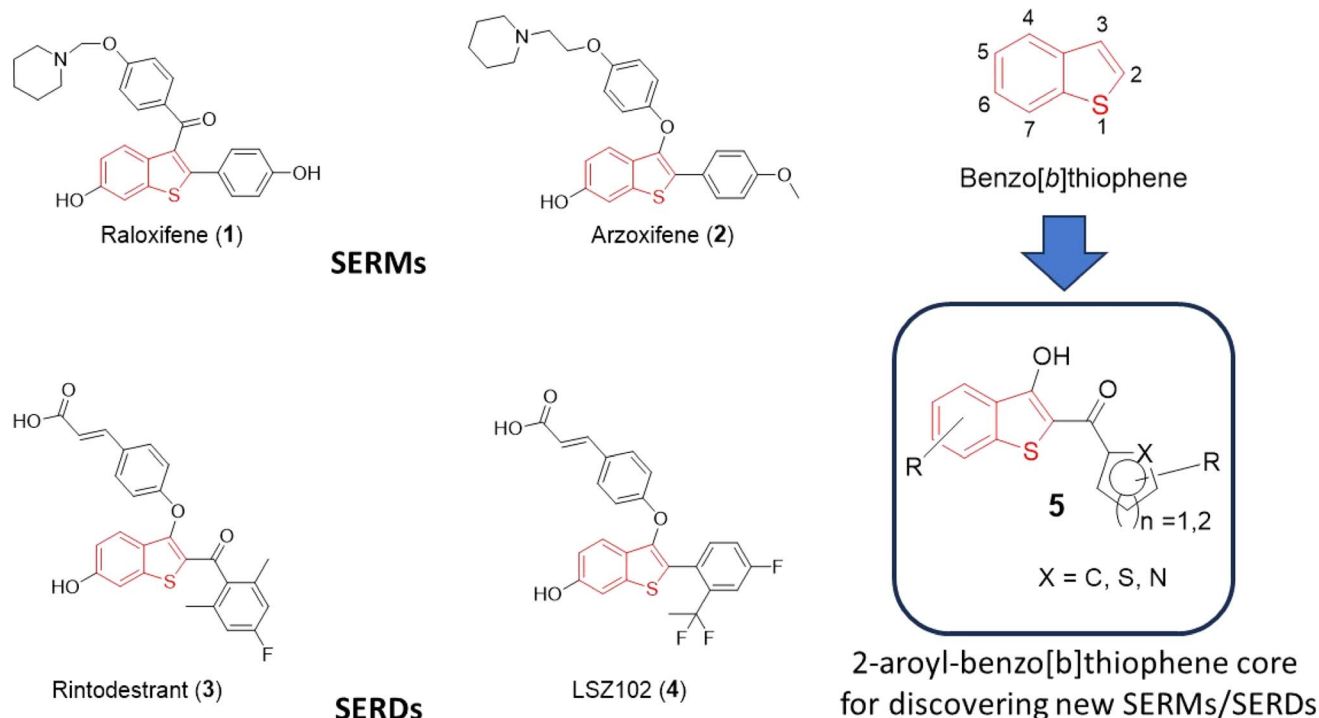
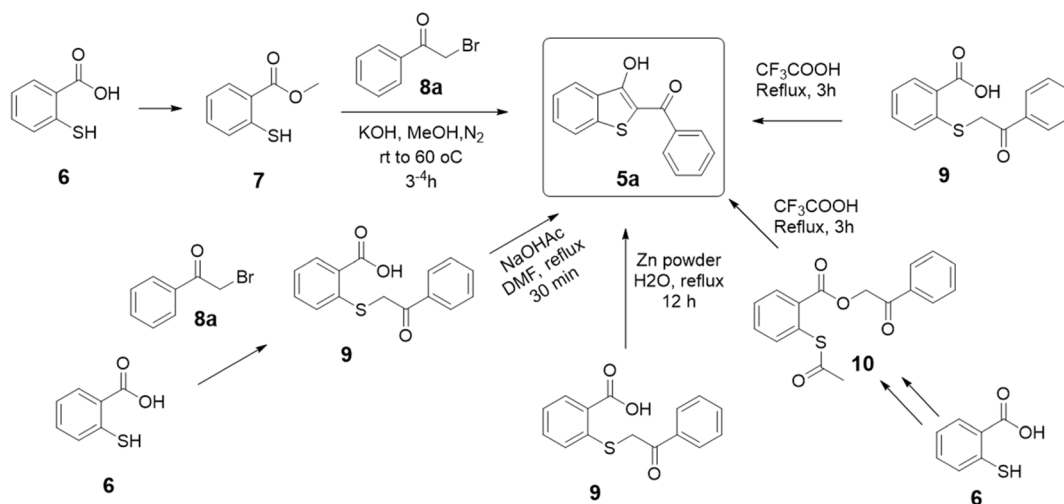


Fig. 1 Structures of selective estrogen receptor modulators (SERMs) and selective estrogen receptor down-regulators (SERDs) containing the benzo[b]thiophene scaffold and identification of the motif 2-aryl-benzo[b]thiophen-3-ol (**5**) for synthesizing new SERM/SERD hits.

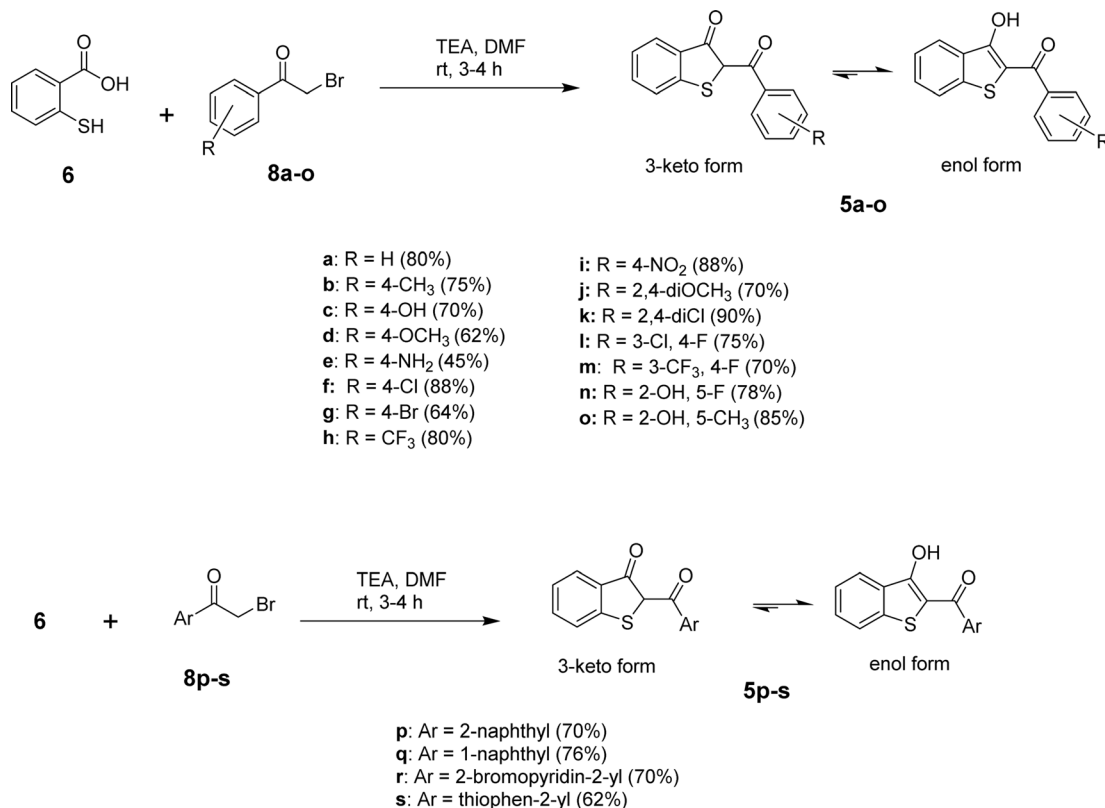
mercaptobenzoic acid (**6**) under basic conditions will complement the previously reported method under acidic conditions.³³

The core 2-aryl-benzo[b]thiophen-3-ol (**5**) structure has been synthesized previously from 2-mercaptobenzoic acid (**6**) using a 2 or 3-step-long reaction sequence and/or using harsh conditions as shown in Scheme 1. For instance, P. Guglielmi *et al.*¹¹ reported the synthesis of **5** by converting **6** to its methyl ester **7**, followed by a reaction with phenacyl bromide (**8a**) in methanol and potassium hydroxide as a base. Several other methods have been reported where a prior conversion of **6** to 2-

((2-oxo-2-phenylethyl)thio)benzoic acid (**9**) is required for the subsequent intramolecular cyclization reaction to **5** in an additional step, such as by using sodium acetate,¹² trifluoroacetic acid,³³ or zinc powder.²² In another report, **6** was first converted to thio-aspirin, then to phenacyl ester **10**, and finally to **5** by using trifluoroacetic acid under reflux.³³ Taking insight from these reports, we anticipated that **5** could be synthesized in a one-pot reaction using basic conditions. The reaction of the sulfhydryl (–SH) group in **6** with **8a** under basic conditions is a well-known S_N2-type reaction to afford sulfanyl benzoic acid



Scheme 1 Various pathways for converting 2-mercaptobenzoic acid (**6**) into benzo[b]thiophene (**5a**).

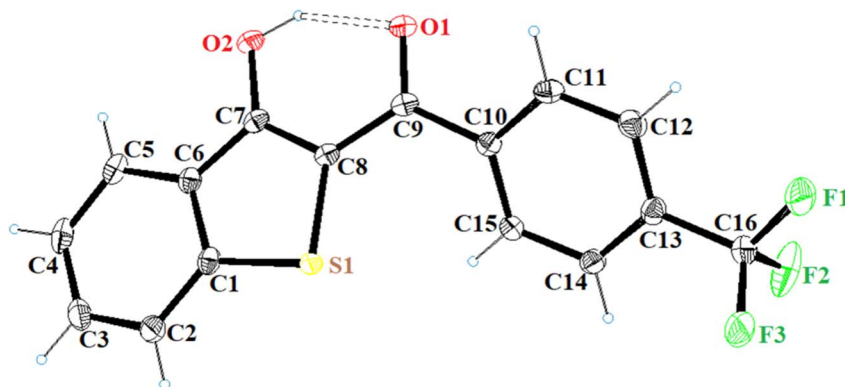
Scheme 2 Synthesis of 2-arylbenzo[*b*]thiophen-3-ols using triethylamine in DMF.

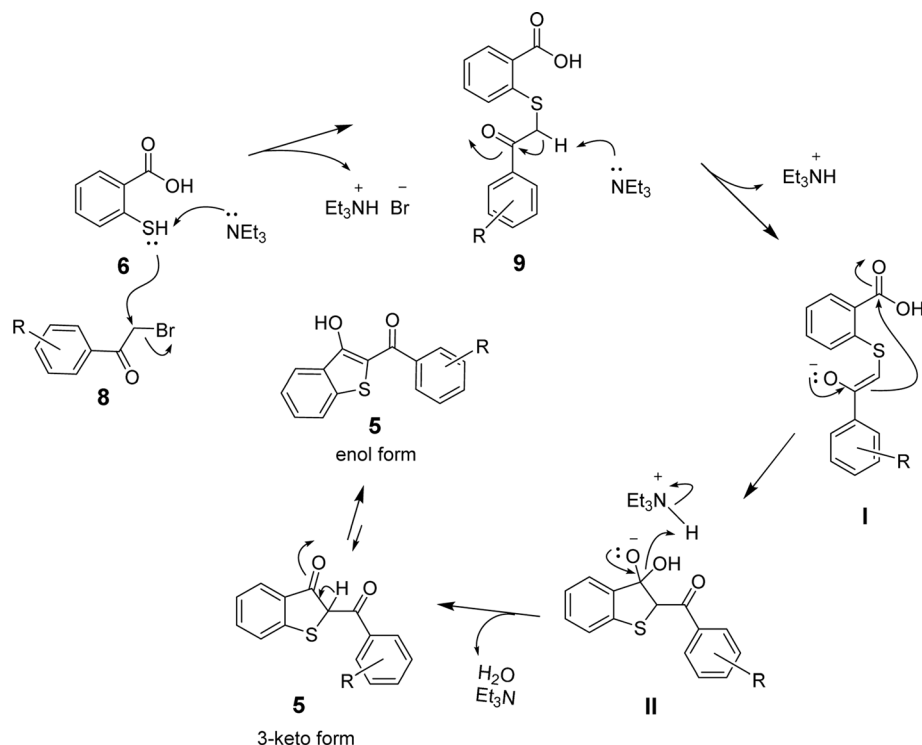
temperature for 3 hours yielded the maximum amounts of the desired product **5a**. Finally, the formation of **5a** was tested in DMSO using TEA as a base (entries 25 and 26). However, the observed yield of the target benzo[*b*]thiophene was lower than what we obtained using the TEA/DMF system.

After determining the optimized conditions for the formation of product **5a**, the substrate scope of the reaction was explored. Using diverse aryl bromomethyl ketones (**8a-s**), we were able to obtain the target benzo[*b*]thiophen-3-ols (**5a-s**) or their keto tautomers in 45–87% yields as depicted in Scheme 2. The electronic nature of the substituent on the aromatic ring of the arylbromomethyl ketone seems to control the formation of **5** in the keto or enol form. In the case of electron-withdrawing

substituents, the predominant product was obtained as the enol tautomer, while the electron-donating substituents favored the formation of the product in the 3-keto form. Further experiments to delineate the electronic factors of the reaction are under investigation in our laboratory.

All the synthesized benzo[*b*]thiophen-3-ol derivatives were characterized using spectroscopic techniques. We were able to obtain decent crystal of **5h** and, therefore, the compound was subjected to X-ray crystallography for the corroboration of the assigned structure. Fig. 2 shows the ORTEP diagram of **5h** and Table S1† reports the XRD parameters. The detailed analysis of the single-crystal XRD data will be published elsewhere. The ¹H NMR spectra exhibited a characteristic downfield signal around

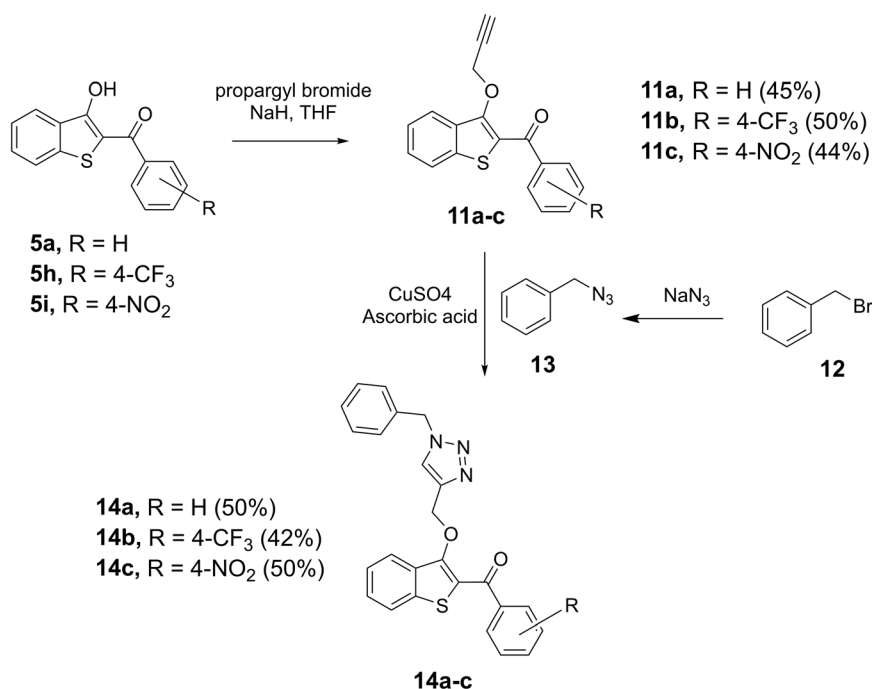
Fig. 2 ORTEP diagram of **5h** (CCDC 2297186).



Scheme 3 The proposed mechanism for the formation of 2-aryl-benzo[*b*]-thiophen-3-ol (5) from 2-mercaptobenzoic acid (6) and aryl-bromomethyl ketone (8).

13 ppm for the 3-OH moiety, indicating the formation of the enolic tautomer. The absence of this peak and the appearance of an up-field singlet around 4.5–5.5 ppm, assigned to the H-2, revealed the formation of the keto tautomer. Thus, out of the 19

benzo[*b*]thiophen-3-ols, most of the synthesized compounds were confirmed as enol analogs, whereas **5b**, **5d**, **5f** and **g**, and **5j** and **k** were obtained as the respective keto tautomers. The chemical shifts for aromatic protons were observed in the



Scheme 4 Synthesis of benzo[*b*]thiophene-triazole hybrids.



expected range from 6.00–8.50 ppm, whereas ^{13}C NMR spectra exhibited the peak for the carbonyl moieties in the 187–200 ppm range.

Based on the above results and the supporting literature,³⁵ we propose a plausible reaction mechanism involving the initial nucleophilic attack of the sulfhydryl group of **6** on the α -carbon of the arylbromomethyl ketone (**8**) to eliminate the bromide ion. The $\text{S}_{\text{N}}2$ -type mechanism is facilitated by triethylamine, which abstracts a proton from S-atom to drive the formation of sulfanyl benzoic acid (**9**) and triethylammonium bromide ($\text{Et}_3\text{NH}^+\text{Br}^-$). The abstraction of another proton from the α -carbon of **9** by triethylamine then leads to enolate **I**, which undergoes intramolecular cyclization *via* the attack on the carboxylic group in the presence of the triethylammonium ion to afford the intermediate **II**. Finally, eliminating the hydroxyl group in the form of H_2O leads to the formation of benzo[*b*]thiophene **5** in the keto-form. The keto–enol tautomerism in the presence of acid (Et_3NH^+) or base (Et_3N) yields 2-aryl-benzo[*b*]thiophen-3-ol as the desired product, as shown in Scheme 3.

After the successful construction of the 2-aryl-benzo[*b*]thiophen-3-ol scaffold, the strategic utilization of the 3-hydroxyl group was examined for the introduction of an alkyne moiety. Various bases were attempted, and we found sodium hydride (NaH) to be a suitable base for converting selected benzothiophenes, **5a**, **5h**, and **5i** to the corresponding benzothiophen-3-olates, which were individually reacted with propargyl bromide to afford the alkyne derivatives **11a**, **11b**, and **11c**, respectively in 45–50% yields. Alkynes **11a–c** were then reacted with benzyl azide **13**, prepared in parallel by reacting benzyl bromide with sodium azide (NaN_3) in the same flask, to accomplish the copper-catalyzed azide–alkyne cycloaddition (CUAAC) reaction in the presence of copper sulfate (CuSO_4) and ascorbic acid. Thus, the target triazols **14a–c**, novel hybrid pharmacophores containing benzo[*b*]thiophene and triazole together, were prepared in 50%, 42%, and 50% yields, respectively as shown in Scheme 4.

3 Experimental

3.1. Materials

All the chemicals and reagents were purchased from different vendors (Merck, Sigma-Aldrich & TCI) and were used without further purification. Solvents for syntheses and chromatographic work were distilled and properly dried before use (if required). The progress of the reactions was monitored by TLC plates (aluminium sheets pre-coated with silica gel) by visualization under a UV lamp at 254 nm wavelength. The reaction mixtures were purified using silica gel column chromatography (pore size 60 Å, 230–400 mesh size, 40–63 μm). FTIR spectra were recorded using a Bruker Alpha Platinum AT by applying the ATR sampling technique through neat samples. NMR spectra were recorded in deuterated solvent using a Bruker 600 MHz NMR instrument (Avance Neo Bruker). GCMS data were obtained using a 1300 GC-MS (Thermo Scientific). Melting points were measured using the STUART SMP3 apparatus and have been reported uncorrected.

3.2. General procedure for the synthesis of 2-aryl-benzo[*b*]thiophen-3-ols

To a stirred solution of 2-mercaptobenzoic acid (1 mmol) and arylbromomethyl ketone (1.2 mmol) in 5 mL of DMF, triethylamine (1 mL) was added, and the reaction mixture was stirred for 2–4 h at room temperature. The progress of the reaction was monitored through TLC analysis. After completion of the reaction, the mixture was poured into 30 mL of ice-cold water and the pH was adjusted to approximately 7 using a 2 N solution of hydrochloric acid. The precipitate of benzo[*b*]thiophen-3-ol was filtered *via* a Buchner funnel and washed with cold water. The ppt was recrystallized using hot methanol to afford the target.

(3-Hydroxybenzo[*b*]thiophen-2-yl)(phenyl)methanone **5a**. Yellow powder (205 mg, 80%), m.p. = 103–107 °C. ^1H NMR (600 MHz, chloroform-*d*): δ (ppm) = 13.45 (s, 1H, OH), 8.08–8.05 (m, 3H, C_6H_4), 7.74 (d, J = 8.2 Hz, 1H, C_6H_4), 7.62 (t, J = 7.4 Hz, 1H, C_6H_5), 7.57–7.54 (m, 3H, C_6H_5), 7.44 (t, J = 7.6 Hz, 1H, C_6H_5). ^{13}C NMR (150 MHz, chloroform-*d*) δ (ppm) = 191.9(CO), 165.5 (C-3), 140.9, 138.3, 132.7, 130.3, 130.2, 128.8, 128.5, 124.9, 124.1, 123.1 (C_6H_5 and C_6H_4) 109.7 (C-2). IR (cm^{-1}): 3473 (broad OH); 1682 (C=O); 1590 (C=C). EI-MS (m s^{-1}): Calcd. for $\text{C}_{15}\text{H}_{10}\text{O}_2\text{S}$ [M^+]: 254.30. GC-MS (EI) m/z (%relative intensity) 254.04 (M^+ , 100%), 197.05 (10%), 176.04 (95%).

(3-Hydroxybenzo[*b*]thiophen-2-yl)(*p*-tolyl)methanone **5b**. Yellow amorphous solid (134 mg, 50%), m.p. = 99–101 °C. ^1H NMR (600 MHz, DMSO-*d*₆): δ (ppm) = 8.05–8.03 (dd, J = 7.62 Hz, 1H), 7.92 (d, J = 8.16 Hz, 2H), 7.70 (m, 1H), 7.66 (m, 1H), 7.63 (m, 1H), 7.40 (d, J = 7.98 Hz, 2H), 5.69 (s, 1H), 2.42 (s, 3H). ^{13}C NMR (150 MHz, DMSO-*d*₆) δ (ppm) = 193.15, 191.39, 165.63, 144.95, 136.57, 132.27, 131.50, 129.60, 129.41, 127.94, 66.60, 21.79. EI-MS: Calcd. for $\text{C}_{16}\text{H}_{12}\text{O}_2\text{S}$ [M^+]: 268.33. GC-MS (EI) m/z (%relative intensity) 268.06 (M^+ , 70%), 254.01(15%), 176.04 (100%).

(3-Hydroxybenzo[*b*]thiophen-2-yl)(4-hydroxyphenyl)methanone **5c**. Light yellow amorphous solid (189 mg, 70%). ^1H NMR (600 MHz, DMSO-*d*₆): δ (ppm) = 10.47 (bs, 1H), 7.96 (d, J = 8.4 Hz, 2H), 7.89–7.88 (d, J = 7.8 Hz, J = 1.2 Hz, 1H), 7.49–7.47 (m, 1H), 7.44–7.43 (m, 1H), 7.21 (t, J = 7.8 Hz, 1H), 6.88 (d, J = 9.0 Hz, 2H). ^{13}C NMR (150 MHz, DMSO-*d*₆) δ (ppm) = 193.23, 168.00, 162.96, 140.89, 132.75, 131.73, 131.36, 127.53, 126.46, 124.51, 115.77. EI-MS: Calcd. for $\text{C}_{15}\text{H}_{10}\text{O}_3\text{S}$ [M^+]: 270.30. GC-MS (EI) m/z (%relative intensity) 270.04 (M^+ , 100%), 254.04 (40%), 176.04 (90%).

2-(4-Methoxybenzoyl)benzo[*b*]thiophen-3(2H)-one **5d**. Yellow amorphous solid (176 mg, 62%), m.p. = 182–183 °C. ^1H NMR (600 MHz, DMSO-*d*₆): δ (ppm) = 8.05–7.99 (m, 4H), 7.71–7.62 (m, 2H), 7.16–7.10 (d, J = 7.8 Hz, 2H), 5.66 (s, 1H), 3.87 (s, 3H). ^{13}C NMR (150 MHz, DMSO-*d*₆) δ (ppm) = 193.09, 191.23, 165.64, 164.21, 137.25, 134.21, 132.94, 131.34, 130.77, 130.31, 128.29, 127.17, 123.96, 114.66, 67.53, 56.12. EI-MS: Calcd. for $\text{C}_{16}\text{H}_{12}\text{O}_3\text{S}$ [M^+]: 284.33. GC-MS (EI) m/z (%relative intensity) 284.05 (M^+ , 70%), 254.04 (30%), 176.04 (100%).

(4-Aminophenyl)(3-hydroxybenzo[*b*]thiophen-2-yl)methanone **5e**. Light Yellow powder (40 mg, 45%); mp 164–166 °C. ^1H NMR (600 MHz, DMSO): δ (ppm) = 11.10 (brs, 1H), 7.90 (d, J = 7.2 Hz, 2H), 7.47 (d, J = 8.4 Hz, 2H), 7.21 (t, J =

7.2 Hz, 2H), 7.09 (t, $J = 7.2$ Hz, 2H), 4.40 (brs, 2H-NH₂). ¹³C NMR (150 MHz, DMSO-*d*₆): δ (ppm) = 189.08, 164.56, 139.58, 130.05, 129.99, 129.20, 123.76, 123.03, 121.89, 114.92, 114.77. EI-MS: Calcd. for C₁₅H₁₁NO₂S [M]⁺: 269.32 GC-MS (EI) m/z (%relative intensity) 269.05 (M⁺, 30%), 254.04 (70%), 176.04 (100%).

(4-Chlorophenyl)(3-hydroxybenzo[*b*]thiophen-2-yl) methanone **5f**. Yellow powder (240 mg, 88%), m.p. = 164–166 °C. ¹H NMR (600 MHz, chloroform-*d*) δ (ppm) = 8.17 (d, $J = 7.8$ Hz, 1H), 7.85 (d, $J = 7.8$ Hz, 2H), 7.58 (m, 2H), 7.51 (m, 1H), 7.30 (d, $J = 7.8$ Hz, 2H), 5.53 (s, 1H). ¹³C NMR (150 MHz, chloroform-*d*) δ (ppm) = 198.28, 191.52, 165.75, 145.07, 136.70, 132.40, 131.62, 129.75, 129.72, 129.53, 128.10, 128.06. EI-MS: Calcd. for C₁₅H₉ClO₂S [M + H]⁺: 288.75. GC-MS (EI) m/z (% relative intensity) 288.0 (M⁺, 70%), 176.04 (100%), 112.01 (40%).

(4-Bromophenyl)(3-hydroxybenzo[*b*]thiophen-2-yl) methanone **5g**. Yellow amorphous solid (180 mg, 64%), m.p. = 168–170 °C. ¹H NMR (600 MHz, chloroform-*d*): δ (ppm) = 8.15 (m, 2H), 7.87 (m, 2H), 7.40 (m, 4H), 7.20 (t, $J = 7.3$ Hz, 1H), 4.46 (s, 1H). ¹³C NMR (150 MHz, chloroform-*d*): δ (ppm) = 193.91, 168.10, 166.50, 164.88, 140.19, 132.78, 132.50, 132.18, 132.12, 131.31, 126.44, 124.64, 116.34, 116.20. EI-MS: Calcd. for C₁₅H₉BrO₂S [M]⁺: 333.20. GC-MS (EI) m/z (%relative intensity) 331.95 (M + 1, 97.8%), 333.95 (M⁺, 100%), 176.04 (70%).

(3-Hydroxybenzo[*b*]thiophen-2-yl)[4-(trifluoromethyl)phenyl] methanone **5h**. Yellow amorphous solid (258 mg, 80%), m.p. = 150–152 °C. ¹H NMR (600 MHz, DMSO-*d*₆) δ (ppm) = 11.96 (s, 1H), 8.08 (d, $J = 8.0$ Hz, 1H), 8.04 (d, $J = 8.0$ Hz, 2H), 7.98 (d, $J = 8.16$ Hz, 1H), 7.93 (d, $J = 8.1$ Hz, 2H), 7.62 (t, $J = 7.3$ Hz, 1H), 7.49 (t, $J = 7.3$ Hz, 1H). ¹³C NMR (150 MHz, DMSO-*d*₆) δ (ppm) = 206.95, 158.04, 142.89, 139.93, 131.85, 130.20 (q, ² $J_{CF} = 32.07$ Hz), 129.50, 125.80, 125.78 (q, ³ $J_{CF} = 3.85$ Hz), 125.38, 124.11 (q, ¹ $J_{CF} = 272.98$ Hz), 124.07, 123.466, 114.01. EI-MS: Calcd. for C₁₆H₉F₃O₂S [M]⁺: 322.30. GC-MS (EI) m/z (%relative intensity) 322.03 (M⁺, 80%), 176.04 (100%), 146.03 (20%).

(3-Hydroxybenzo[*b*]thiophen-2-yl)(4-nitrophenyl)methanone **5i**. Orange powder (230 mg, 88%), m.p. = 202–204 °C. ¹H NMR (600 MHz, DMSO-*d*₆): δ (ppm) = 12.01 (s, 1H), 8.37 (d, $J = 8.7$ Hz, 2H), 8.36 (m, 3H), 7.98 (d, $J = 8.4$ Hz, 1H), 7.62 (t, $J = 1$ Hz), 7.49 (t, 1H). ¹³C NMR (150 MHz, DMSO-*d*₆) δ (ppm) = 193.09, 164.22, 137.25, 134.21, 132.95, 131.34, 131.16, 130.75, 130.71, 130.31, 128.30, 127.18, 123.96. EI-MS: Calcd. for C₁₅H₉NO₄S [M]⁺: 299.30. GC-MS (EI) m/z (%relative intensity) 299.03 (M⁺, 70%), 176.04 (100%), 123.03 (20%).

2-(2,4-Dimethoxybenzoyl)benzo[*b*]thiophen-3(2H)-one **5j**. Light yellow amorphous solid (220 mg, 70%) m.p. = 200–204 °C. ¹H NMR (600 MHz, DMSO-*d*₆) δ (ppm) = 8.19–8.18 (dd, $J = 7.8$ Hz, $J = 1.2$ Hz, 1H), 8.00 (d, $J = 8.8$ Hz, 1H), 7.59 (m, 1H), 7.55 (m, 1H), 7.50 (m, 1H), 6.58 (dd, $J = 8.4$ Hz, $J = 1.4$ Hz, 1H), 6.47 (d, $J = 2.4$ Hz, 1H), 5.41 (s, 1H), 3.94 (s, 3H), 3.88 (s, 3H). ¹³C NMR (150 MHz, DMSO-*d*₆) δ (ppm) = 193.35, 190.73, 165.87, 165.49, 161.52, 136.48, 133.79, 133.24, 132.01, 131.48, 129.35, 128.79, 117.20, 105.92, 98.05, 55.66, 55.60. EI-MS: Calcd. for C₁₇H₁₄O₄S [M]⁺: 314.36. GC-MS (EI) m/z (%relative intensity) 314.06 (M⁺, 80%), 254.04 (70%), 176.04 (100%).

(2,4-Dichlorophenyl)(3-hydroxybenzo[*b*]thiophen-2-yl) methanone **5k**. Yellow amorphous solid (290 mg, 90%), m.p. = 149–152 °C. ¹H NMR (600 MHz, chloroform-*d*) δ (ppm) = 8.08

(dd, $J = 7.8$ Hz, 1H), 7.50 (t, $J = 8.4$ Hz, 1H), 7.44 (d, $J = 2.4$ Hz, 1H), 7.37 (m, 3H), 7.28 (m, 1H), 7.25 (m, 1H), 4.37 (s, 1H). ¹³C NMR (150 MHz, chloroform-*d*) δ (ppm) = 196.78, 170.51, 140.40, 138.21, 135.73, 133.51, 132.55, 132.18, 131.03, 130.44, 127.52, 126.96, 126.73, 125.26. EI-MS: Calcd. for C₁₅H₈Cl₂O₂S [M]⁺: 323.19. GC-MS (EI) m/z (%relative intensity) 321.96 (M⁺, 80%), 176.04 (100%), 145.97 (30%).

(3-Chloro-4-fluorophenyl)(3-hydroxybenzo[*b*]thiophen-2-yl) methanone **5l**. Yellow amorphous solid (230 mg, 75%) m.p. = 167–169 °C. ¹H NMR (600 MHz, chloroform-*d*): δ (ppm) = 11.94 (s, 1H), 8.08 (m, 2H), 7.98 (d, $J = 8.16$ Hz, 1H), 7.90 (m, 1H), 7.62 (m, 2H), 7.48 (m, 1H). ¹³C NMR (150 MHz, chloroform-*d*) δ (ppm) = 195.18, 167.93, 140.56, 136.01, 134.11, 132.81 (d, ¹ $J_{CF} = 245.26$ Hz), 131.41, 129.25 (d, ³ $J_{CF} = 8.74$ Hz), 129.06, 126.50 (d, ² $J_{CF} = 23.15$ Hz), 124.63. EI-MS: Calcd. for C₁₅H₈ClFO₂S [M]⁺: 306.74. GC-MS (EI) m/z (%relative intensity) 307.99 (M + 1, 36.6%), 305.99 (M⁺, 100%), 176.04 (90%).

(4-Fluoro-3-(trifluoromethyl)phenyl)(3-hydroxybenzo[*b*]thiophen-2-yl)methanone **5m**. Yellow amorphous solid (238 mg, 70%), m.p. = 160–162 °C. ¹H NMR (600 MHz, DMSO-*d*₆) δ (ppm) = 11.91 (s, 1H), 8.22 (m, 2H), 7.99 (d, $J = 7.8$ Hz, 1H), 7.88 (d, $J = 7.8$ Hz, 1H), 7.65 (t, $J = 8.8$ Hz, 1H), 7.55 (t, $J = 7.2$ Hz, 1H), 7.40 (t, $J = 7.2$ Hz, 1H). ¹³C NMR (150 MHz, DMSO-*d*₆) δ (ppm) = 190.00, 162.00, 140.19 (d, ¹ $J_{CF} = 245.26$ Hz), 136.53, 136.26, 129.84 (m, ² $J_{CF} = 32.93$ Hz), 128.36, 124.79 (m, ³ $J_{CF} = 3.98$ Hz), 124.31 (q, ¹ $J_{CF} = 273.49$ Hz), 123.90 (d, ³ $J_{CF} = 8.74$ Hz), 117.56 (d, ² $J_{CF} = 23.15$ Hz), 117.42. EI-MS: Calcd. for C₁₆H₈F₄O₂S [M]⁺: 340.29. GC-MS (EI) m/z (%relative intensity) 341.02 (M⁺, 18.3%), [M]⁺: 340.02 (100%), 176.04 (100%).

(5-Fluoro-2-hydroxyphenyl)(3-hydroxybenzo[*b*]thiophen-2-yl) methanone **5n**. White amorphous solid (224 mg, 78%) m.p. = 166–169 °C. ¹H NMR (600 MHz, DMSO-*d*₆) δ (ppm) = 13.48 (bs, 1H), 8.04–8.02 (dd, $J = 7.8$ Hz, $J = 1.8$ Hz, 1H), 7.93–7.91 (dd, $J = 7.8$ Hz, $J = 1.2$ Hz, 1H), 7.62 (m, 1H), 7.58–7.55 (m, 1H), 7.52–7.51 (d, $J = 8.4$ Hz, 1H), 7.38–7.34 (m, 1H), 7.19 (t, $J = 7.8$ Hz, 1H), 3.37 (bs, 1H). ¹³C NMR (150 MHz, DMSO-*d*₆) δ (ppm) = 200.46, 168.09, 139.40, 133.76, 132.83, 132.06 (d, ¹ $J_{CF} = 245.26$ Hz), 131.83, 131.44, 128.54 (d, ³ $J_{CF} = 8.66$ Hz), 126.98, 126.47 (d, ² $J_{CF} = 24.11$ Hz), 125.47, 124.90. EI-MS: Calcd. for C₁₅H₉FO₃S [M]⁺: 288.29. GC-MS (EI) m/z (%relative intensity) 289.03 (M + 1, 17.3%), 288.03 (M⁺, 100%), 176.04 (70%).

(2-Hydroxy-5-methylphenyl)(3-hydroxybenzo[*b*]thiophen-2-yl)methanone **5o**. Yellow solid (227 mg, 85%) m.p. = 130–132 °C. ¹H NMR (600 MHz, DMSO-*d*₆) δ (ppm) = 12.87 (s, 1H), 12.16 (s, 1H), 7.85 (m, 2H), 7.56 (m, 3H), 7.54 (dd, $J = 7.8$ Hz, $J = 1.2$ Hz, 1H), 6.90 (d, $J = 7.8$ Hz, 2H). ¹³C NMR (150 MHz, DMSO-*d*₆) δ (ppm) = 195.18, 167.93, 140.56, 136.01, 134.11, 132.81, 131.41, 129.25, 129.06, 126.50, 124.63, 31.17. EI-MS: Calcd. for C₁₆H₁₂O₃S [M]⁺: 284.33. GC-MS (EI) m/z (%relative intensity) 284.03 (M⁺, 100%), 254.04 (30%), 176.04 (70%).

(3-Hydroxybenzo[*b*]thiophen-2-yl)(naphthalen-2-yl) methanone **5p**. Yellow amorphous solid (210 mg, 70%) m.p. = 126–128 °C. ¹H NMR (600 MHz, DMSO-*d*₆) δ (ppm) = 12.85 (s, 1H), 7.96 (m, 2H), 7.70 (m, 3H), 7.54 (m, 1H), 7.47 (m, 3H), 7.06 (d, $J = 8.4$ Hz, 1H), 6.98 (t, $J = 7.8$ Hz, 1H). ¹³C NMR (150 MHz, DMSO-*d*₆) δ (ppm) = 193.90, 163.61, 145.53, 136.47, 134.60, 130.98, 130.56, 129.69, 129.00, 128.71, 128.44, 120.02, 118.90,



118.79. EI-MS: Calcd. for $C_{19}H_{12}O_2S$ $[M]^+$: 304.36 GC-MS (EI) m/z (%relative intensity) 304.06 (M^+ , 60%), 176.04 (100%), 128.06 (30%).

(3-Hydroxybenzo[*b*]thiophen-2-yl)(naphthalen-1-yl) methanone **5q**. Yellow amorphous solid (231 mg, 76%) m.p. = 162–166 °C. 1H NMR (600 MHz, DMSO- d_6) δ (ppm) = 13.18 (s, 1H), 8.92 (m, 1H), 8.20 (m, 2H), 7.96 (m, 2H), 7.66 (m, 1H), 7.50 (m, 1H), 7.72 (m, 3H), 7.57 (m, 1H). ^{13}C NMR (150 MHz, DMSO- d_6) δ (ppm) = 195.10, 167.91, 140.73, 135.47, 134.60, 130.98, 130.56, 129.69, 129.01, 128.71, 128.44, 124.35, 124.03. EI-MS: Calcd. for $C_{19}H_{12}O_2S$ $[M]^+$: 304.36. GC-MS (EI) m/z (%relative intensity) 304.06 (M^+ , 60%), 176.04 (100%), 128.06 (30%).

(6-Bromopyridin-2-yl)(3-hydroxybenzo[*b*]thiophen-2-yl) methanone **5r**. Light Yellow amorphous solid (233 mg, 70%), m.p. = 138–142 °C. 1H NMR (600 MHz, DMSO- d_6) δ (ppm) = 13.17 (s, 1H), 8.02 (s, 1H), 7.99 (m, 2H), 7.90 (dd, J = 7.8 Hz, J = 1.2 Hz, 1H), 7.50 (t, J = 7.2 Hz, 1H), 7.45 (d, J = 7.8 Hz, 1H), 7.22 (t, J = 7.2 Hz, 1H). ^{13}C NMR (150 MHz, DMSO- d_6) δ (ppm) = 194.10, 167.96, 153.32, 141.52, 141.17, 140.30, 133.08, 132.92, 131.47, 128.46, 126.31, 124.77, 122.14. EI-MS: Calcd. for $C_{14}H_8BrNO_2S$ $[M]^+$: 334.19. GC-MS (EI) m/z (%relative intensity) 332.95 (M + 1, 98.3%), 334.94 (M^+ , 100%), 176.04 (70%).

(3-Hydroxybenzo[*b*]thiophen-2-yl)(thiophen-2-yl)methanone **5s**. Yellow amorphous solid, (161 mg, 62%), m.p. = 182–184 °C. 1H NMR (600 MHz, chloroform- d) δ (ppm) = 13.24 (s, 1H), 8.04 (m, 1H), 7.88 (m, 1H), 7.77 (m, 1H), 7.60 (m, 1H), 7.47 (m, 1H), 7.20 (m, 1H). ^{13}C NMR (150 MHz, chloroform- d) δ (ppm) = 181.13, 166.07, 144.59, 140.22, 134.43, 132.97, 131.79, 130.97, 130.63, 125.29, 124.23, 124.08, 123.51, 123.27, 123.20. EI-MS: Calcd. for $C_{13}H_8O_2S_2$ $[M]^+$: 260.33. GC-MS (EI) m/z (%relative intensity) 260.00 (M^+ , 60%), 176.04 (100%), 84.06 (30%).

3.3. General procedure for the synthesis of phenyl(3-(prop-2-yn-1-yloxy)benzo[*b*]thiophen-2-yl)methanone

In a 50 mL round-bottom flask, 1 mmol of a benzothiophene derivative (compound 5) and 4 mmol of NaH were dissolved in 8 mL of DMF. To this stirred solution, 10 mmol of propargyl bromide was added. The stirring was continued at room temperature for approximately 1 hour. After this time, the reaction progress was assessed using TLC, revealing the presence of some benzothiophene derivative remaining in its protonated form. Consequently, 4 mmol of NaH and 5 mmol of propargyl bromide were added. Following overnight stirring, the reaction mixture was transferred to a separating funnel and dissolved in a solution consisting of 5 mL of ice-cold deionized water, 5 mL of brine, and 5 mL of ethyl acetate. The excess ethyl acetate was removed using a rotary evaporator, and the organic layer was collected in a clean vial. A brownish-yellow semi-liquid was obtained, which was ultimately purified through column chromatography to yield a brownish-yellow solid.

Phenyl(3-(prop-2-yn-1-yloxy)benzo[*b*]thiophen-2-yl) methanone **11a**. Compound **11a** was synthesized from **5a** following the general procedure. Brownish-yellow solid, (45%). 1H NMR (600 MHz, chloroform- d) δ (ppm): 8.050–8.080 (m, 3H, Ar), 7.775–7.761 (m, 1H, benzothiophene), 7.765–7.761 (m, 1H, benzothiophene), 7.510–7.490 (m, 3H, Ar), 7.463–7.445 (m, 1H,

benzothiophene), 4.678–4.567 (m, 2H, CH_2), 2.226–2.222 (m, 1H, CH). ^{13}C NMR (150 MHz, chloroform- d) δ (ppm) = 189.27, 152.17, 138.68, 138.33, 133.98, 132.88, 129.63, 128.26, 128.08, 124.83, 124.73, 123.46, 122.90, 78.03, 76.89, 62.26. EI-MS ($m\ s^{-1}$): Calcd. For $C_{18}H_{12}O_2S$ $[M]^+$: 292.35. GC-MS (EI) m/z (% relative intensity) 260.00 (M^+ , 60%), 188.04 (100%), 84.06 (30%).

(3-(Prop-2-yn-1-yloxy)benzo[*b*]thiophen-2-yl)(4-(trifluoromethyl)phenyl)methanone **11b**. Compound **11b** was synthesized from **5h** by following the general procedure. Brown solid, (50%). 1H NMR (600 MHz, chloroform- d) δ (ppm): 8.07 (d, J = 8.0 Hz, 1H), 8.01 (d, J = 8.0 Hz, 2H), 7.98 (d, J = 8.16 Hz, 1H), 7.92 (d, J = 8.1 Hz, 2H), 7.62 (t, J = 7.3 Hz, 1H), 7.53 (t, J = 7.3 Hz, 1H), 4.71 (s, 2H), 3.53 (s, 1H). ^{13}C NMR (150 MHz, DMSO- d_6) δ (ppm) = 206.95, 152.70, 142.06, 138.83, 133.44, 130.26 (q, $^2J_{CF}$ = 32.07 Hz), 129.25, 126.03, 125.87 (q, $^3J_{CF}$ = 3.85 Hz), 125.73, 125.71, 124.14 (q, $^1J_{CF}$ = 272.98 Hz), 123.92, 80.75, 62.64. EI-MS ($m\ s^{-1}$): Calcd. For $C_{19}H_{11}F_3O_2S$ $[M]^+$: 360.35. GC-MS (EI) m/z (% relative intensity) 360.04 (M^+ , 60%), 188.03 (100%), 174.03 (30%).

(4-Nitrophenyl)(3-(prop-2-yn-1-yloxy)benzo[*b*]thiophen-2-yl) methanone **11c**. Compound **11c** was synthesized from **5i** by following the general procedure. Brown solid, (44%). 1H NMR (600 MHz, chloroform- d) δ (ppm): 7.90 (m, 1H), 7.87 (d, J = 8.0 Hz, 1H), 7.72 (m, 2H), 7.55 (d, J = 8.0 Hz, 2H), 7.35 (dd, J = 7.8 Hz, J = 1.2 Hz, 1H), 7.25 (D, 1H), 4.84 (s, 1H), 4.24 (s, 2H). ^{13}C NMR (150 MHz, DMSO- d_6) δ (ppm) = 194.34, 167.79, 144.51, 132.48, 132.44, 131.55, 131.30, 131.10, 130.90, 129.35, 128.87, 128.81, 126.95, 80.95, 68.16, 52.75. EI-MS ($m\ s^{-1}$): Calcd. For $C_{19}H_{11}F_3O_2S$ $[M]^+$: 337.35 GC-MS (EI) m/z (%relative intensity) 337.04 (M^+ , 60%), 188.03 (100%), 151.03 (30%).

3.4. General procedure for the synthesis of 3-((1-benzyl-1H-1,2,3-triazol-4-yl)methoxy)benzo[*b*]thiophen-2-yl(phenyl) methanone

In a 25 mL round-bottom flask, a solution was prepared by dissolving 1.5 mmol of benzyl bromide and 1 mmol of the acetylene derivative. To this stirred solution, 1 mmol of sodium azide was added, along with a 2 mL mixture of *t*BuOH : H₂O (1 mL : 1 mL), and the reaction mixture was stirred at room temperature for approximately 5 minutes. Subsequently, 0.1 mmol of sodium ascorbate and 0.64 mmol of Cu(II) sulfate pentahydrate were added. The reaction mixture was stirred at 60 °C for 2–3 hours. The completion of the reaction was confirmed by TLC, which indicated the consumption of the starting material and the formation of the desired product. After overnight stirring, the reaction mixture was dissolved in 5 mL of ice-cold deionized water, followed by the dropwise addition of 2 mL of aqueous ammonia. The mixture was stirred for 5 minutes. Finally, the reaction mixture was extracted with 12 mL of ethyl acetate. A yellow liquid was obtained, which was ultimately purified through column chromatography to yield a yellowish liquid.

3-((1-Benzyl-1H-1,2,3-triazol-4-yl)methoxy)benzo[*b*]thiophen-2-yl(phenyl)methanone **14a**. Compound **14a** was synthesized from **11a** by following the general procedure. Brown solid (50%). 1H NMR (600 MHz, chloroform- d): δ (ppm) = 7.89



(m, 3H), 7.80 (m, 1H), 7.52 (m, 2H), 7.40 (m, 6H), 7.20 (m, 2H), 6.97 (s, 1H), 5.44 (s, 2H), 5.04 (s, 2H). ^{13}C NMR (150 MHz, chloroform-*d*) δ (ppm) = 189.32, 152.85, 143.36, 138.93, 138.31, 134.31, 133.71, 132.68, 129.58, 129.14, 128.84, 128.17, 128.10, 124.90, 123.38, 123.22, 123.02, 68.56, 54.10. EI-MS (m s^{-1}): Calcd. For $\text{C}_{25}\text{H}_{19}\text{N}_3\text{O}_2\text{S}$ $[\text{M}]^+$: 425.51 GC-MS (EI) m/z (%relative intensity) 425.12 ($\text{M} + 1$, 29%), 426.12 (M^+ , 100%).

(3-((1-Benzyl-1*H*-1,2,3-triazol-4-yl)methoxy)benzo[*b*]thiophen-2-yl)(4-(trifluoromethyl)phenyl)methanone **14b**. Compound **14b** was synthesized from **11b** by following the general procedure. Brownish Yellow solid (42%). ^1H NMR (600 MHz, chloroform-*d*): δ (ppm) = 7.75 (m, 2H), 7.60 (s, 1H), 7.38 (m, 6H), 7.32 (m, 4H), 6.49 (s, 1H), 5.56 (s, 2H), 5.34 (s, 2H). ^{13}C NMR (150 MHz, chloroform-*d*) δ (ppm) = 206.10, 150.00, 144.30, 137.64, 134.40, 132.05 (q, $^2J_{\text{CF}}$ = 32.07 Hz), 129.19, 128.86, 128.13, 125.25 (q, $^3J_{\text{CF}}$ = 3.85 Hz), 123.73 (q, $^1J_{\text{CF}}$ = 272.98 Hz), 122.80, 120.94, 97.51, 63.90, 54.30. EI-MS (m s^{-1}): Calcd. For $\text{C}_{26}\text{H}_{18}\text{F}_3\text{N}_3\text{O}_2\text{S}$ $[\text{M}]^+$: 493.50 GC-MS (EI) m/z (%relative intensity) 494.11 ($\text{M} + 1$, 29%), 493.11 (M^+ , 100%).

(3-((1-Benzyl-1*H*-1,2,3-triazol-4-yl)methoxy)benzo[*b*]thiophen-2-yl)(3-nitrophenyl)methanone **14c**. Compound **14c** was synthesized from **11c** by following the general procedure. Brown solid (50%). ^1H NMR (600 MHz, chloroform-*d*): δ (ppm) = 8.84 (t, J = 7.8 Hz, 1H), 8.40 (m, 1H), 8.37 (m, 1H), 7.65 (m, 3H), 7.38 (m, 5H), 7.33 (m, 3H), 5.56 (s, 2H), 5.50 (s, 2H). ^{13}C NMR (150 MHz, chloroform-*d*) δ (ppm) = 200.01, 164.34, 148.23, 142.52, 135.42, 134.22, 131.48, 129.69, 129.22, 128.95, 128.22, 127.62, 124.76, 124.05, 58.73, 54.34. EI-MS (m s^{-1}): Calcd. for $\text{C}_{25}\text{H}_{18}\text{N}_4\text{O}_4\text{S}$ $[\text{M}]^+$: 470.50. GC-MS (EI) m/z (%relative intensity) 471.11 ($\text{M} + 1$, 29%) $[\text{M}]^+$: 470.10 (M^+ , 100%).

4 Conclusion

In conclusion, an expeditious one-pot method has been developed to synthesize 2-aryl-benzo[*b*]thiophen-3-ols. The method is applicable to a wide range of arylbromomethyl ketones, demonstrating its substrate tolerability. The synthesis of benzothiophene was extended further toward introducing the triazole moiety, exemplifying the efficiency and selectivity inherent in click chemistry methodologies. The outlined approach not only underscores the versatility of click chemistry but also contributes to the expanding repertoire of synthetic methods for constructing complex molecular architectures with potential applications in medicinal and materials science.

Author contributions

Asma Mukhtar: syntheses of benzo[*b*]thiophenes and writing the manuscript. Arif Hussain and Faiza Younus: assistance in the synthesis of benzo[*b*]thiophenes. Sammer Yousuf: X-ray data acquisition and curation, Muhammad Saeed: supervision, experimental planning project administration, writing – review & editing, funding acquisition.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- 1 M. C. Egbujor, P. Tucci, U. C. Onyeije, C. N. Emeruwa and L. Saso, *Molecules*, 2023, **28**, 2751.
- 2 C. Rizzo, S. Amata, I. Pibiri, A. Pace, S. Buscemi and A. Palumbo Piccionello, *Int. J. Mol. Sci.*, 2023, **24**, 7728.
- 3 E. Vitaku, D. T. Smith and J. T. Njardarson, *J. Med. Chem.*, 2014, **57**, 10257–10274.
- 4 M. M. Heravi and V. Zadsirjan, *RSC Adv.*, 2020, **10**, 44247–44311.
- 5 J. Shearer, J. L. Castro, A. D. G. Lawson, M. MacCoss and R. D. Taylor, *J. Med. Chem.*, 2022, **65**, 8699–8712.
- 6 S. Sharma, C. C. Malakar and V. Singh, *Asian J. Org. Chem.*, 2020, **9**, 1857–1868.
- 7 S. Sharma, D. Singh, S. Kumar, Vaishali, R. Jamra, N. Banyal, Deepika, C. C. Malakar and V. Singh, *Beilstein J. Org. Chem.*, 2023, **19**, 231–244.
- 8 R. S. Keri, K. Chand, S. Budagumpi, S. Balappa Somappa, S. A. Patil and B. M. Nagaraja, *Eur. J. Med. Chem.*, 2017, **138**, 1002–1033.
- 9 T. Al-Warhi, N. M. Rashad, H. Almahli, M. M. Abdel-Aziz, Z. M. Elsayed, M. I. Shahin and W. M. Eldehna, *Arch. Pharm.*, 2024, **357**, e2300529.
- 10 T. Barbier, A. Barbry, J. Magand, C. Badiou, F. Davy, A. Baudouin, Y. Queneau, O. Dumitrescu, G. Lina and L. Soulere, *Biomolecules*, 2022, **12**, 131.
- 11 P. Guglielmi, D. Secci, A. Petzer, D. Bagetta, P. Chimenti, G. Rotondi, C. Ferrante, L. Recinella, S. Leone, S. Alcaro, G. Zengin, J. P. Petzer, F. Ortuso and S. Carradori, *J. Enzyme Inhib. Med. Chem.*, 2019, **34**, 1511–1525.
- 12 M. Gerard, L. Caroline and K. Micheline, *French Pat.*, FR2862646, 2005.
- 13 T. W. Burke and C. L. Walker, *Gynecol. Oncol.*, 2003, **90**, S40–S46.
- 14 E. Esposito, A. Iacono, G. M. Raso, M. Pacilio, A. Coppola, R. Di Carlo and R. Meli, *Endocrinology*, 2005, **146**, 3301–3308.
- 15 S. Gizzo, C. Saccardi, T. S. Patrelli, R. Berretta, G. Capobianco, S. Di Gangi, A. Vacilotto, A. Bertocco, M. Noventa, E. Ancona, D. D'Antona and G. B. Nardelli, *Obstet. Gynecol. Surv.*, 2013, **68**, 467–481.
- 16 I. Y. Kim, B. C. Kim, D. H. Seong, D. K. Lee, J. M. Seo, Y. J. Hong, H. T. Kim, R. A. Morton and S. J. Kim, *Cancer Res.*, 2002, **62**, 5365–5369.
- 17 S. S. Ko and V. C. Jordan, *Expert Opin. Pharmacother.*, 2011, **12**, 657–674.
- 18 C. Marchisano-Karpman and M. W. DeGregorio, *IDrugs*, 2003, **6**, 880–885.
- 19 P. N. Munster, *Expert Opin. Invest. Drugs*, 2006, **15**, 317–326.
- 20 M. B. Sporn, *Clin. Cancer Res.*, 2004, **10**, 5313–5315.
- 21 R. Iqbal, M. Yaqub, D. E. Oprea-Lager, Y. Liu, A. M. Luik, A. P. Beelen, R. C. Schuit, A. D. Windhorst, R. Boellaard



- and C. W. Menke-van der Houven van Oordt, *J. Nucl. Med.*, 2022, **63**, 694–699.
- 22 K. Jhaveri, D. Juric, Y. S. Yap, S. Cresta, R. M. Layman, F. P. Duhoux, C. Terret, S. Takahashi, J. Huober, N. Kundamal, Q. Sheng, A. Balbin, Y. Ji, W. He, A. Crystal, S. De Vita and G. Curigliano, *Clin. Cancer Res.*, 2021, **27**, 5760–5770.
 - 23 G. S. Tria, T. Abrams, J. Baird, H. E. Burks, B. Firestone, L. A. Gaither, L. G. Hamann, G. He, C. A. Kirby, S. Kim, F. Lombardo, K. J. Macchi, D. P. McDonnell, Y. Mishina, J. D. Norris, J. Nunez, C. Springer, Y. Sun, N. M. Thomsen, C. Wang, J. Wang, B. Yu, C. L. Tiong-Yip and S. Peukert, *J. Med. Chem.*, 2018, **61**, 2837–2864.
 - 24 J. Ceramella, A. Caruso, M. A. Occhiuzzi, D. Iacopetta, A. Barbarossa, B. Rizzuti, P. Dallemagne, S. Rault, H. El-Kashef, C. Saturnino, F. Grande and M. S. Sinicropi, *Eur. J. Med. Chem.*, 2019, **181**, 111583.
 - 25 N. R. Penthala, V. N. Sonar, J. Horn, M. Leggas, J. S. Yadlapalli and P. A. Crooks, *Medchemcomm*, 2013, **4**, 1073–1078.
 - 26 C. V. Do, A. Faouzi, C. Barette, A. Farce, M. O. Fauvarque, E. Colomb, L. Catry, O. Berthier-Vergnes, M. Haftek, R. Barret and T. Lomberget, *Bioorg. Med. Chem. Lett.*, 2016, **26**, 174–180.
 - 27 R. Romagnoli, D. Preti, E. Hamel, R. Bortolozzi, G. Viola, A. Brancale, S. Ferla, G. Morciano and P. Pinton, *Bioorg. Chem.*, 2021, **112**, 104919.
 - 28 D. J. Witter, S. Belvedere, L. Chen, J. P. Secrist, R. T. Mosley and T. A. Miller, *Bioorg. Med. Chem. Lett.*, 2007, **17**, 4562–4567.
 - 29 F. Batool, E. U. Mughal, K. Zia, A. Sadiq, N. Naeem, A. Javid, Z. Ul-Haq and M. Saeed, *J. Biomol. Struct. Dyn.*, 2022, **40**, 3777–3788.
 - 30 F. Batool, M. Saeed, H. N. Saleem, L. Kirschner and J. Bodem, *Pathogens*, 2021, **10**, 464.
 - 31 H. N. Saleem, S. Kousar, A. H. Jiskani, I. Sohail, A. Faisal and M. Saeed, *Arch. Pharm.*, 2023, **356**, e2300292.
 - 32 H. A. Younus, M. Saeed, A. Mahmood, M. S. K. Jadoon, A. Hameed, A. Asari, H. Mohamad, J. Pelletier, J. Sevigny, J. Iqbal and M. Al-Rashida, *Bioorg. Chem.*, 2023, **134**, 106450.
 - 33 P. Trapani, L. Kvapil, P. Hradil and M. Soural, *Synlett*, 2018, **29**, 810–814.
 - 34 D. W. Plank and J. B. Howard, *J. Biol. Chem.*, 1988, **263**, 8184–8189.
 - 35 G. A.-N. Gohar, *J. Phys. Org. Chem.*, 2012, **25**, 343–350.

