


 Cite this: *RSC Adv.*, 2024, 14, 16327

Construction of a coumarin-based fluorescent probe for accurately visualizing hydrogen sulfide in live cells and zebrafish†

 Xiao Wei,^{ab} Long Mi,^b Shenglong Dong,^b Hui Yang ^{*b} and Shiyuan Xu^{*a}

Hydrogen sulfide (H₂S), an important gas signaling molecule, is a regulator of many physiological processes, and its abnormal levels are closely related to the onset and progression of disease. It is vital to develop methods for specific tracking of H₂S in clinical diagnosis and treatment. In this study, we designed an ultrasensitive and highly stable coumarin-based fluorescent probe Cou-H₂S. Through the H₂S-initiated tandem reaction, Cou-H₂S successfully achieved highly selective and super-fast detection of H₂S. Cou-H₂S was successfully applied to the monitoring of endogenous and exogenous H₂S at the cellular level and verified the validity of the detection of H₂S in the LPS-induced zebrafish model. Therefore, Cou-H₂S might provide new insights into the study of H₂S-related diseases.

Received 26th January 2024

Accepted 13th May 2024

DOI: 10.1039/d4ra00668b

rsc.li/rsc-advances

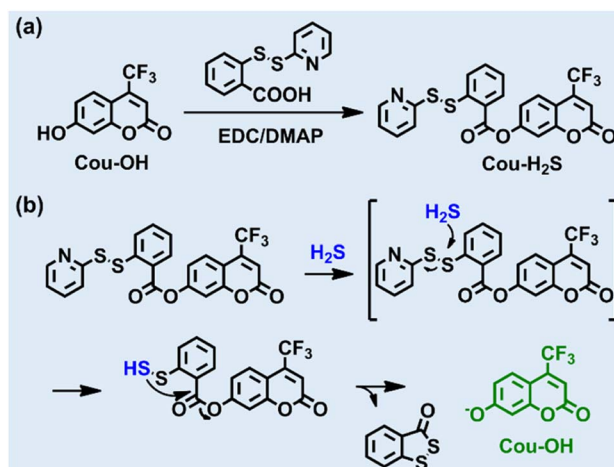
Introduction

Reactive sulfur species (RSS) play a pivotal role as signaling molecules in redox regulation, including hydrogen sulfide (H₂S), glutathione (GSH), cysteine (Cys), and sulfane sulfur.^{1–3} Among them, H₂S stands out as the simplest biothiol involved in the regulation of a range of complex pathophysiological processes in living organisms.^{4–6} These processes include regulation of vascular tone, cytoprotection, and neurotransmission, and so on.^{7–9} For example, H₂S induces vascular relaxation, lowers blood pressure, and plays an important protective role in cardiovascular diseases. It effectively resists oxidative stress and protects cells and tissues.^{10,11} Endogenous H₂S is catalytically produced by intracellular sulfate-reducing enzymes or thio redoxin proteins (cystathionine β-synthase, cystathionine γ-lyase, and 3-mercaptopyruvate sulfur transferase).^{12,13} Abnormal H₂S concentrations in human serum can induce several diseases such as cardiovascular, neurological, and cancer.¹⁴ Therefore, the development of visual detection tools to monitor intracellular H₂S levels is crucial for understanding disease mechanisms and enabling early diagnosis.

Common detection methods for H₂S include chromatography, electrochemistry, and mass spectrometry. Although the above methods can detect the concentration of H₂S *in vitro*, they can cause damage to biological samples.^{15,16} Fluorescence

imaging, with its high sensitivity, high accuracy, and non-invasiveness, enables real-time, non-invasive monitoring of H₂S in organisms. Based on chemical properties such as reducibility, nucleophilicity and metal coordination, researchers have developed a series of H₂S fluorescent probes.^{17–25} However, some of the reported probes have the disadvantages of environmental sensitivity, long reaction time, high detection limit, and the need for large amounts of organic solvents as co-solvents. It is still necessary to develop new small-molecule fluorescent probes for H₂S that can overcome these limitations.

Trifluoromethyl coumarin, with the advantages of high chemical stability, wide emission wavelength range, and high



Scheme 1 (a) Synthesis of Cou-H₂S. (b) Reaction mechanism of Cou-H₂S toward H₂S.

^aDepartment of Anesthesiology, Zhujiang Hospital of Southern Medical University, Guangzhou, 510282, China

^bCentral South University Xiangya School of Medicine Affiliated Haikou Hospital, The First Affiliated Hospital of Hainan Medical University, Haikou, 570102, China. E-mail: 15298989046@163.com; xsy998@smu.edu.cn

† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d4ra00668b>



fluorescence quantum yield, has been widely used in bioimaging and fluorescent probes.^{26–31} In this work, we designed and synthesized a novel coumarin-based fluorescent probe Cou-H₂S using trifluoromethyl coumarin as the molecular scaffold and 2-pyridyl disulfide as the H₂S recognition group (Scheme 1). The reaction of Cou-H₂S with H₂S released a trifluoromethyl coumarin dye that emitted intense fluorescence at 498 nm. Cou-H₂S presented excellent selectivity and sensitivity to H₂S, with a low detection limit of 25 nM. Cou-H₂S could also be employed for fluorescence imaging of endogenous and exogenous H₂S in cells. Furthermore, the upregulation of H₂S levels in LPS-stimulated zebrafish was visualized.

Experimental section

Synthesis of 2-oxo-4-(trifluoromethyl)-2H-chromen-7-yl 2-(pyridin-2-yl)disulfanylbenzoate (Cou-H₂S)

Cou-OH (115 mg, 0.5 mmol) and 2-(pyridin-2-yl)disulfanyl benzoic acid (158 mg, 1.2 mmol) were dissolved in dry dichloromethane (20 mL). Subsequently, EDC (383 mg, 2.0 mmol) and DMAP (25 mg) were added. The reaction mixture was stirred at room temperature for 10 h. After completion of the reaction, the solvent was evaporated *in vacuo*. The resulting product was purified by silica gel column chromatography (eluent: 20% ethyl acetate/80% petroleum ether) to afford the desired compound (76 mg, 32% yield). ¹H NMR (400 MHz, CDCl₃): δ 8.49–8.47 (m, 1H), 8.30 (dd, *J* = 8.0, 1.5 Hz, 1H), 8.01 (dd, *J* = 8.2, 1.1 Hz, 1H), 7.82 (dd, *J* = 8.8, 1.8 Hz, 1H), 7.61–7.53 (m, 3H), 7.41–7.32 (m, 3H), 7.13–7.10 (m, 1H), 6.82 (s, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 163.83, 158.57, 158.44, 155.09, 153.87, 149.72, 142.14, 137.35, 134.34, 132.14, 126.40, 126.35, 125.97, 125.32, 121.14, 119.97, 119.78, 119.23, 115.55, 115.49, 111.59, 111.32; HRMS *m/z*: C₂₂H₁₂F₃NO₄S₂ [M + H]⁺ calcd for 476.0238 found 476.0282.

Results and discussion

Design and synthesis of Cou-H₂S

The chemical structure of Cou-H₂S was well characterized by NMR and HRMS, as shown in Fig. S1–S4.† 2-Pyridyl disulfide masked the hydroxyl group of the fluorescent dye, quenching the fluorescence of Cou-OH. The addition of H₂S induced the release of Cou-OH from Cou-H₂S. The general mechanism was as follows: H₂S reacted with Cou-H₂S *via* a nucleophilic substitution reaction to give Cou-SSH intermediate, followed by an intramolecular esterification reaction to release Cou-OH and 3H-benzo[*c*][1,2]dithiol-3-one. To elucidate the reaction mechanism, we conducted mass spectrometry analysis of the products before and after the reaction of Cou-H₂S with H₂S. As seen in Fig. S5,† Cou-H₂S exhibited a main peak at *m/z* = 476.0282, corresponding to Cou-H₂S. After the excess H₂S reacted with Cou-H₂S, the solution showed two main peaks at *m/z* = 167.9695 and *m/z* = 253.0014, attributed to 3H-benzo[*c*][1,2]dithiol-3-one ([M]⁺) and Cou-OH ([M + Na]⁺).

Optical response of Cou-H₂S to H₂S

The response time serves as a crucial metric for assessing the probe's performance, providing insight into the kinetic process of its binding to target analytes. Upon synthesizing Cou-H₂S, we opted for an initial concentration of 10 μM and recorded the time course of Cou-H₂S in the absence or presence of 20 μM Na₂S (an inorganic H₂S donor substance) in the PBS solution containing 100 μM hexadecyl trimethyl ammonium bromide (CTAB). The fluorescence intensity at 498 nm of Cou-H₂S remained constant for 300 s (Fig. 1a). When 20 μM Na₂S was added, the fluorescence intensity at 498 nm reached the maximum at 100 s and remained stable. The changes in fluorescence intensity before and after the reaction of Cou-H₂S with Na₂S across various pH values were then explored. As seen in

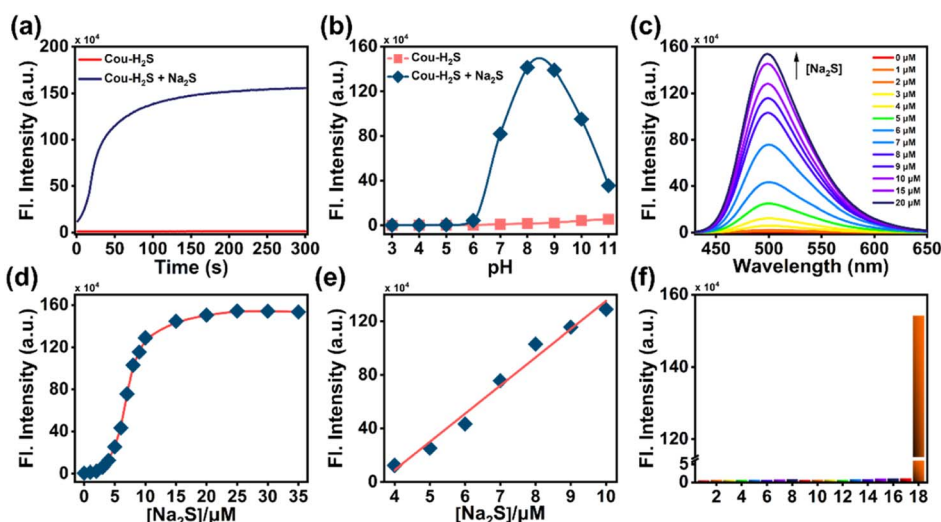


Fig. 1 (a) Fluorescence intensities at 498 nm of Cou-H₂S (10 μM) in the presence or absence of Na₂S (20 μM). (b) pH-dependent fluorescence intensities of Cou-H₂S (10 μM) in the absence or presence of Na₂S (20 μM). (c) Fluorescence response of Cou-H₂S (10 μM) to varying concentrations of Na₂S (0–20 μM). (d) Plot of fluorescence intensities of Cou-H₂S (10 μM) at 498 nm versus Na₂S concentrations (0–35 μM). (e) Linear relationship between fluorescence intensities of Cou-H₂S (10 μM) at 498 nm and Na₂S concentrations (4–10 μM). (f) Fluorescence response of Cou-H₂S (10 μM) at 498 nm to common interfering analytes (100 μM, respectively). (1) Blank, (2) Met, (3) Gly, (4) Arg, (5) Pro, (6) S₂O₃²⁻, (7) SO₃²⁻, (8) SO₄²⁻, (9) Cl⁻, (10) Mg²⁺, (11) Al³⁺, (12) Ca²⁺, (13) Fe³⁺, (14) ClO⁻, (15) Cys, (16) Hcy, (17) GSH, (18) H₂S. λ_{ex} = 405 nm.



Fig. 1b, the fluorescence intensity of Cou-H₂S showed minimal variation over a wide pH range from 3.0 to 10.0, highlighting its insensitivity to the pH of the detection system. In the presence of Na₂S, the fluorescence intensity closely resembled free Cou-H₂S within the pH range of 3.0–6.0. A notable increase in fluorescence intensity was observed beyond pH 6.0, reaching its peak at pH 8.0–9.0. The absorption spectra of Cou-H₂S changed significantly before and after the addition of Na₂S (Fig. S6†). The relationship between fluorescence intensity changes of Cou-H₂S and Na₂S concentration was also investigated (Fig. 1c). Under the excitation light of 405 nm, the fluorescence of the free Cou-H₂S was weak. Upon sequentially adding different concentrations of Na₂S, the fluorescence emission peak of Cou-H₂S at 498 nm gradually enhanced (Fig. 1d). In the Na₂S concentration range of 0–20 μM, the fluorescence of Cou-H₂S at 498 nm increased by 340-fold. This substantial change indicated a high signal-to-noise ratio for the *in vitro* detection of H₂S. In addition, the fluorescence intensities of Cou-H₂S at 498 nm displayed an excellent linear relationship with the Na₂S concentration (4–10 μM), represented by the linear equation $y/10^4 = 21.07x - 75.49$ (where y was the recorded fluorescence intensity at 498 nm, and x was the Na₂S concentration), yielding a linear correlation coefficient of 0.99 (Fig. 1e). Utilizing the detection limit formula $3\sigma/k$ (σ represented the standard deviation of the recorded fluorescence intensity of free Cou-H₂S at 498 nm 12 times, and k represented the slope of the linear equation), it could be concluded that the detection limit was 25 nM, lower than that of several reported fluorescent probes for H₂S (Table S1†). To evaluate the selectivity of Cou-H₂S toward H₂S, the performance of Cou-H₂S was tested in response to common interfering analytes, including amino acids (Met, Gly, Arg, Pro), anions (S₂O₃²⁻, SO₃²⁻, SO₄²⁻, Cl⁻), metal ions (Mg²⁺, Al³⁺, Ca²⁺, Fe³⁺), reactive oxygen species (ClO⁻), and biothiols (Cys, Hcy, GSH). As depicted in Fig. 1f, there was no significant variation in the fluorescence intensity at 498 nm for amino acids, anions, metal ions, and reactive oxygen species, and only Na₂S triggered an enhancement of fluorescence intensity. These results underscore the excellent sensitivity and selectivity of Cou-H₂S for H₂S, suggesting its potential use in complex biological environments.

H₂S imaging in live cells

Evaluating the cytotoxicity of Cou-H₂S would provide a basis for its potential biological applications. Fig. S7† showed that incubation with different concentrations ranging from 0 to 30 μM of Cou-H₂S for 24 h resulted in more than 90% cell survival, confirming the commendable biocompatibility of Cou-H₂S in cellular and *in vivo* imaging experiments. As illustrated in Fig. 2, co-incubating Cou-H₂S with HeLa cells yielded weak green fluorescence. When the cells were pre-incubated with Cys, followed by incubation with Cou-H₂S, a moderate green fluorescence emerged due to the conversion of Cys to H₂S catalyzed by the thiosulfate transferase enzyme. Notably, the cells treated with Na₂S induced robust green fluorescence, whereas treatment with Na₂S and ZnCl₂ (an H₂S scavenger) in turn led to reduced green fluorescence. The results established the

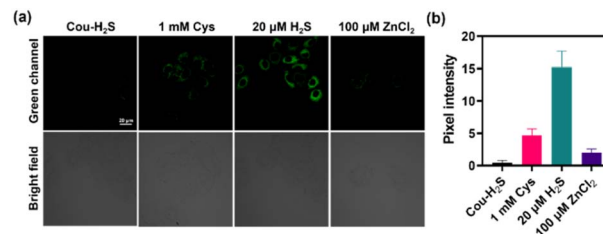


Fig. 2 Confocal laser fluorescence imaging of HeLa cells under different pretreatments. (a) The cells were treated with Cys (1 mM, 30 min), Na₂S (20 μM, 30 min), ZnCl₂ (20 μM Na₂S + 100 μM ZnCl₂, 30 min) at 37 °C respectively, then incubated with Cou-H₂S (10 μM) and CTAB (50 μM) for 30 min before imaging. (b) Pixel intensities of the cells in plane a. $\lambda_{\text{ex}} = 405 \text{ nm}$, $\lambda_{\text{em}} = 450\text{--}550 \text{ nm}$. Scale bar: 20 μm.

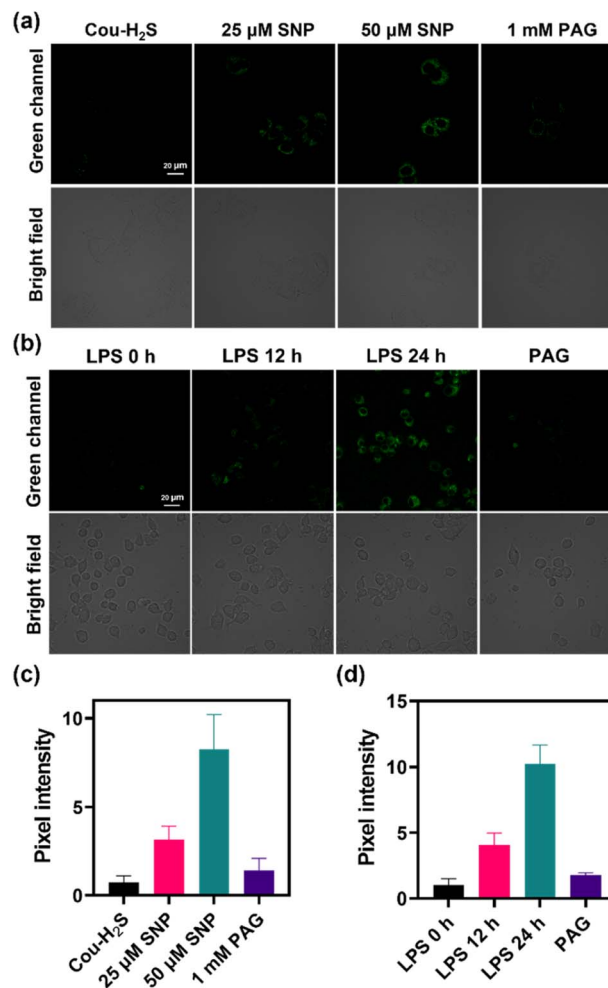


Fig. 3 (a) HeLa cells were treated with SNP (25, 50 μM, 30 min), SNP (50 μM, 30 min) + PAG (1 mM, 30 min) at 37 °C respectively, then incubated with Cou-H₂S (10 μM) and CTAB (50 μM) for 30 min before imaging. (b) RAW264.7 cell were stimulated with LPS (1 μg mL⁻¹, 0, 12, 24 h), LPS (1 μg mL⁻¹, 24 h) + PAG (1 mM, 30 min) at 37 °C respectively, then incubated with Cou-H₂S (10 μM) and CTAB (50 μM) for 30 min before imaging. (c) Pixel intensities of the cells in plane a. (d) Pixel intensities of the cells in plane c. $\lambda_{\text{ex}} = 405 \text{ nm}$, $\lambda_{\text{em}} = 450\text{--}550 \text{ nm}$. Scale bar: 20 μm.



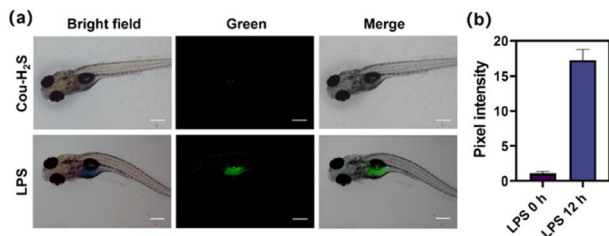


Fig. 4 (a) The zebrafish were stimulated with LPS ($2 \mu\text{g mL}^{-1}$) for 12 h at 37°C , then incubated with Cou- H_2S ($10 \mu\text{M}$) for 30 min before imaging. (d) Pixel intensities of the cells in plane a. $\lambda_{\text{ex}} = 405 \text{ nm}$, $\lambda_{\text{em}} = 450\text{--}550 \text{ nm}$. Scale bar: $200 \mu\text{m}$.

capability of Cou- H_2S to image exogenous H_2S , positioning it for biological applications.

Sodium nitroprusside (SNP) was a commonly used nitric oxide (NO) donor that induced hydrogen sulfide production.³² Fig. 3a revealed a faint green fluorescence upon incubating the cells with Cou- H_2S , with varying concentrations of SNP leading to distinct enhancements in green fluorescence. Treatment of $50 \mu\text{M}$ SNP followed by propargylglycine (PAG, an H_2S scavenger)³³ resulted in a rapid decline in green fluorescence, demonstrating the ability of Cou- H_2S to monitor SNP-induced endogenous H_2S (Fig. 3c).

Lipopolysaccharide (LPS) induced a cellular immune response leading to upregulation of H_2S expression in RAW264.7 cells.³⁴ As displayed in Fig. 3b, faint green fluorescence was observed in RAW264.7 cells incubated with Cou- H_2S . When the cells were stimulated with LPS for 12 h or 24 h, and then incubated with Cou- H_2S , green fluorescence showed a gradual enhancement compared to that of the control group (Fig. 3d). Conversely, treatment with PAG post-LPS exposure led to a reduction in green fluorescence, highlighting Cou- H_2S 's ability to track the LPS-induced upregulation of H_2S levels in this cellular inflammation model.

H_2S imaging in zebrafish

LPS induced up-regulation of the expression and activity of H_2S synthase in zebrafish, thereby promoting H_2S production. As depicted in Fig. 4, Cou- H_2S was incubated with zebrafish, only weak green fluorescence was observed, likely due to the release of trifluoromethyl coumarin dye from the reaction between the low concentration of endogenous H_2S and Cou- H_2S . The green fluorescence of zebrafish treated with LPS for 12 h increased rapidly compared to free Cou- H_2S . This enhancement was attributed to the significant increase in the endogenous H_2S concentration in zebrafish stimulated with LPS. The findings suggested that Cou- H_2S was capable of tracking changes in H_2S concentration levels *in vivo*.

Conclusions

In summary, we have designed and synthesized a novel fluorescence probe, namely Cou- H_2S , for the detection of H_2S based on trifluoromethyl coumarin dye. Cou- H_2S showed high specificity for H_2S , which overcame the interference of common

biological analytes, such as metal ions, anions, amino acids, reactive oxygen species, and biothiols. Cou- H_2S responded rapidly to H_2S with response times as low as 100 s. In addition, the negligible cytotoxicity of Cou- H_2S made it suitable for imaging exogenous and endogenous H_2S in live cells. With the aid of Cou- H_2S , LPS-induced upregulation of endogenous H_2S levels in zebrafish was verified. This work developed a powerful chemical tool for dynamic monitoring of H_2S levels in cells and zebrafish, underpinning the study of H_2S -related diseases.

Conflicts of interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

This research was funded by the Scientific Research Project of Health and Family Planning Industry in Hainan Province (21A200120) and Hainan Province Clinical Medical Center (2021).

References

- 1 T. V. Mishanina, M. Libiad and R. Banerjee, *Nat. Chem. Biol.*, 2015, **11**, 457–464.
- 2 M. Iciek, A. Bilska-Wilkosz, M. Kozdrowicki and M. Górný, *Biosci. Rep.*, 2022, **42**, BSR20221006.
- 3 M. Iciek, A. B. Wilkosz, M. Kozdrowicki and M. Górný, *Antioxid. Redox Signaling*, 2023, **39**, 1000–1023.
- 4 P. Rose, P. K. Moore and Y. Z. Zhu, *Cell. Mol. Life Sci.*, 2017, **74**, 1391–1412.
- 5 R. Wang, *Antioxid. Redox Signaling*, 2010, **12**, 1061–1064.
- 6 L. Alvarez, C. L. Bianco, J. P. Toscano, J. Lin, T. Akaike and J. M. Fukuto, *Antioxid. Redox Signaling*, 2017, **27**, 622–633.
- 7 L. Zhang, Y. Wang, Y. Li, L. Li, S. Xu, X. Feng and S. Liu, *Front. Pharmacol.*, 2018, **9**, 1066.
- 8 S. J. Tripathi, S. Chakraborty, E. Miller, A. A. Pieper and B. D. Paul, *Br. J. Pharmacol.*, 2023, 1–18.
- 9 J. W. Calvert, W. A. Coetzee and D. J. Lefer, *Antioxid. Redox Signaling*, 2010, **12**, 1203–1217.
- 10 Z. Zhao, W. Guo, C. Xu, Q. Wang, C. Mao and M. Wan, *Chem. Eng. J.*, 2023, **452**, 139089.
- 11 J. L. Wallace and R. Wang, *Nat. Rev. Drug Discovery*, 2015, **14**, 329–345.
- 12 H. Liu, M. N. Radford, C. T. Yang, W. Chen and M. Xian, *Br. J. Pharmacol.*, 2019, **176**, 616–627.
- 13 P. Kamoun, *Amino Acids*, 2004, **26**, 243–254.
- 14 X. Cao, Z. Z. Xie, Y. Yang, M. Whiteman, P. K. Moore and J. S. Bian, *Antioxid. Redox Signaling*, 2019, **31**, 1–38.
- 15 H. M. Smith and M. D. Pluth, *JACS Au*, 2023, **3**, 2677–2691.
- 16 H. Ibrahim, A. Serag and M. A. Farag, *J. Adv. Res.*, 2021, **27**, 137–153.
- 17 K. Shimamoto and K. Hanaoka, *Nitric Oxide*, 2015, **46**, 72–79.
- 18 T. T. Jia, Y. Zhang, J.-T. Hou, H. Niu and S. Wang, *Front. Chem.*, 2023, **11**, 1126309.



Paper

- 19 D. A. Jose, R. Sakla, N. Sharma, S. Gadiyaram, R. Kaushik and A. Ghosh, *ACS Sens.*, 2020, **5**, 3365–3391.
- 20 J. Wang, F. Huo, Y. Yue and C. Yin, *Luminescence*, 2020, **35**, 1156–1173.
- 21 H. Li, Y. Fang, J. Yan, X. Ren, C. Zheng, B. Wu, S. Wang, Z. Li, H. Hua, P. Wang and D. Li, *TrAC, Trends Anal. Chem.*, 2021, **134**, 116117.
- 22 C. Liu, J. Pan, S. Li, Y. Zhao, L. Y. Wu, C. E. Berkman, A. R. Whorton and M. Xian, *Angew. Chem., Int. Ed.*, 2011, **50**, 10327–10329.
- 23 Y. Luo, Y. Zuo, G. Shi, H. Xiang and H. Gu, *Anal. Bioanal. Chem.*, 2022, **414**, 2809–2839.
- 24 Q. Sun, H. Liu, Y. Qiu, J. Chen, F.-S. Wu, X. G. Luo and D. W. Wang, *Spectrochim. Acta A*, 2021, **254**, 119620.
- 25 X. Zhang, W. Qu, H. Liu, Y. Ma, L. Wang, Q. Sun and F. Yu, *Anal. Chim. Acta*, 2020, **1109**, 37–43.
- 26 A. Majhi, K. Venkateswarlu and P. Sasikumar, *J. Fluoresc.*, 2023, DOI: [10.1007/s10895-023-03372-3](https://doi.org/10.1007/s10895-023-03372-3).
- 27 G. Kaur, I. Singh, R. Tandon and N. Tandon, *Inorg. Chem. Commun.*, 2023, **158**, 111480.
- 28 M. López-Corrales, A. Rovira, A. Gandioso, S. Nonell, M. Bosch and V. Marchán, *J. Org. Chem.*, 2023, **88**, 7128–7140.
- 29 A. Rovira, M. Pujals, A. Gandioso, M. López-Corrales, M. Bosch and V. Marchán, *J. Org. Chem.*, 2020, **85**, 6086–6097.
- 30 A. Gandioso, R. Bresolí-Obach, A. Nin-Hill, M. Bosch, M. Palau, A. Galindo, S. Contreras, A. Rovira, C. Rovira, S. Nonell and V. Marchán, *J. Org. Chem.*, 2018, **83**, 1185–1195.
- 31 Y. Fan, Y. Wu, J. Hou, P. Wang, X. Peng and G. Ge, *Coord. Chem. Rev.*, 2023, **480**, 215020.
- 32 A. D. Ivankovich, D. J. Miletich and J. H. Tinker, *Int. Anesthesiol. Clin.*, 1978, **16**, 1–29.
- 33 Q. Sun, R. Collins, S. Huang, L. Holmberg-Schiavone, G. S. Anand, C. H. Tan, S. van-den-Berg, L. W. Deng, P. K. Moore, T. Karlberg and J. Sivaraman, *J. Biol. Chem.*, 2009, **284**, 3076–3085.
- 34 Y. Zheng, N. Luo, D. Mu, P. Jiang, R. Liu, H. Sun, S. Xiong, X. Liu, L. Wang and Y. Chu, *In Vitro Cell. Dev. Biol.: Anim.*, 2013, **49**, 679–688.

