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Surface engineering of carbon microspheres with nanoceria wrapped on MWCNTs: a dual electrocatalyst for simultaneous monitoring of molnupiravir and paracetamol[†]

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In the present study, nanoceria-decorated MWCNTs (CeNPs@MWCNTs) were synthesized using a simple and inexpensive process. Molnupiravir (MPV) has gained considerable attention in recent years due to the infection of severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2). Since some people infected with COVID-19 experience fever and headaches, paracetamol (PCM) has been prescribed to relieve these symptoms. Therefore, there is an urgent need to monitor and detect these drugs simultaneously in pharmaceutical and biological samples. In this regard, we developed a novel sensor based on nanoceria-loaded MWCNTs (CeNPs@MWCNTs) for simultaneous monitoring of MPV and PCM. The incorporation of CeNPs@MWCNTs electrocatalyst into a glassy carbon microsphere fluorolube oil paste electrode (GCMFE) creates more active sites, which increase the surface area, electrocatalytic ability, and electron transfer efficiency. Interestingly, CeNPs@MWCNTs modified GCMFE demonstrated excellent detection limits (6.0 nM, 8.6 nM), linear ranges (5.0–5120 nM, 8.0–4162 nM), and sensitivities (78.6, 94.3 $\mu\text{A }\mu\text{M}^{-1}\text{ cm}^{-2}$) for simultaneous detection of MPV and PCM. The developed CeNPs@MWCNTs electrocatalyst modified GCMFE exhibited good repeatability, anti-interference capability, stability, and real-time analysis with good recovery results, which clearly indicates that it can be used for real-time industrial applications.

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Introduction

Monitoring of drugs is a significant aspect in pharmaceuticals, healthcare, and biological samples for the controlled analysis of drug dosages. Antiviral drugs have gained considerable attention in recent years due to the severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) infection. Molnupiravir (MPV), an oral ribonucleoside analog with broad-spectrum antiviral activity, is a prodrug of the synthetic nucleoside derivative N4-hydroxycytidine (NHC). Clinical trials have shown that treatment with MPV reduces deaths and hospitalizations in patients with COVID-19 infections.^{1,2} Paracetamol (PCM), a well-known antipyretic and analgesic compound, is extensively used for the treatment of fever, cough, cold, and pain including muscular ache, chronic pain, headache, backache and

toothache.^{3–5} Moreover, PCM has been frequently used to reduce fever during the COVID-19 pandemic.^{6–8} Abnormal MPV and PCM levels in the body can affect the immune system, which may lead to some diseases. Therefore, there is an urgent need to simultaneously monitor and detect these two drugs in human biological samples.

Although a wide range of techniques are available, the detection technique with factors, including simple, easier handling/operation, and quick and sensitive detection, is the most preferable one.^{9–11} The electrochemical sensor covers all these features; hence, it is utilized to simultaneously monitor and measure MPV and PCM, particularly in human biological fluids and pharmaceutical products. Many methods are described in the literature that can measure MPV or PCM using HPLC,^{12–14} UV spectrophotometry,^{15,16} and electrochemical methods.^{17–28} However, except for electrochemical methods, all other analytical methods are complicated, tedious, time-consuming, and expensive. Electrochemical methods are time-saving, simple, real-time, user-friendly, and economically worthwhile.

The implementation of an electrochemical sensor for the simultaneous assay of MPV and PCM requires electrode modification. The beneficial factors of the modified electrodes include improved sensitivity, higher transfer of electrons,

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enhanced conductivity, and surface area. A huge variety of materials, such as metal oxide nanoparticles and carbon-based composites, are available.^{29–32} In recent years, nanoparticle-based metal oxide electrochemical sensors have garnered a great deal of interest in electrocatalysis.^{33–35} Due to its unique properties, nano-CeO₂ (CeNPs) has been applied in various fields, including supercapacitors, solar cells, photocatalysis, and sensors.^{36–38} Nevertheless, it has been observed that CeNPs tend to aggregate, leading to a decrease in the electrocatalytic activity. Furthermore, the electrocatalytic efficacy of CeNPs is limited by their comparatively low conductivity. Multi-walled carbon nanotubes (MWCNTs) exhibit high electron transfer efficiency and mechanical stability and are therefore suitable conductive substrates for flexible sensors.³⁹ Therefore, the combination of MWCNTs and CeNPs is an effective approach for increasing the conductivity and promoting the electrocatalytic activity of CeNPs.

Carbon, in many respects, is an ideal electrode substrate owing to its wide anodic potential range, low residual current, chemical inertness, and low cost. Moreover, carbon electrodes exhibit a fast response time and can be easily fabricated in different configurations and sizes. Among the family of carbon, glassy carbon microspheres (GCMs) are the most popular electrode material, offering attractive electrochemical reactivity, negligible porosity, and good mechanical rigidity.^{29,34} Glassy carbon paste electrodes (GCPES) combine the attractive properties of composite electrodes and glassy carbon because the preparation of these electrodes includes the mixing of GCMs with organic-pasting liquid. GCPES offer high electrochemical reactivity, a wide accessible potential window; low background current; are easy to prepare, modify, and renew; and are inexpensive.^{40,41}

In this work, for the first time, we developed a glassy carbon microsphere fluorolube oil paste electrode (GCMFE) modified

with CeNPs@MWCNTs electro-catalyst for simultaneous determination of MPV and PCM. To the best of our knowledge, simultaneous determination of the anti-coronavirus MPV and PCM has not been carried out. The CeNPs@MWCNTs electrocatalyst-modified GCMFE exhibits excellent electrochemical sensing capability for simultaneous detection of MPV and PCM in human biological fluids and pharmaceutical samples, which reveals the potential practical applications of the CeNPs@MWCNTs electrocatalyst.

Experimental

Detailed information regarding the chemicals, reagents, instruments, and preparation of the real samples can be found in the ESI.†

Synthesis of CeNPs and CeNPs@MWCNTs electrocatalyst

Detailed information regarding the synthesis procedure of CeNPs can be found in the ESI.† CeNPs@MWCNTs were prepared by mixing CeNPs and MWCNTs (1 : 1, 1 : 2, 2 : 1 w/w) in ethanol. These mixtures were subjected to sonication for 1 h and they were then transferred to a glass Petri dish and heated (70 °C) for 24 h to evaporate the ethanol. Finally, the dry black solid CeNPs@MWCNTs electrocatalyst was obtained.

Preparation of the electrodes

A glassy carbon microsphere paraffin oil paste electrode (GCMPE) was fabricated by blending paraffin oil and GCM (80 : 20 w/w). A glassy carbon microsphere fluorolube oil paste electrode (GCMFE) was then developed using the same procedure, however, with fluorolube oil rather than paraffin oil. The produced paste was placed into a Teflon tube ($A_G = 0.07 \text{ cm}^2$). The CeNPs@MWCNTs modified GCMFEs



Scheme 1 Schematic diagram of preparation process of CeNPs@MWCNTs modified GCMFE.



(CeNPs@MWCNTs/GCMFE) were fabricated by blending (5, 10, 15, and 20 mg) of CeNPs@MWCNTs with GCM and 20% fluorolube oil. Similarly, CeNPs/GCMFE and MWCNTs/GCMFE were prepared (Scheme 1).

Ethical statement

All experiments were performed in compliance with the relevant laws and Assiut University's guidelines. These studies were approved by the ethics committees of Assiut Medical University – Joint Institutional Review Board. All of the subjects signed an informed consent form before the examination.

Results and discussion

Morphological studies

The crystal structures of MWCNTs, CeO₂ nanoparticles (CeNPs), and CeNPs@MWCNTs were studied using XRD analysis, as shown in Fig. S1A.† The XRD pattern of MWCNTs demonstrated an intense diffraction peak around $2\theta = 25.32^\circ$ and a low intense diffraction peak around 41.67° , which are ascribed to (002) and (100), respectively. This result shows that the MWCNTs are well graphitized (Fig. S1A,† curve a). As expected, CeNPs display sharp diffraction peaks at (111), (200), (220), (311), (222), and (400) for the face-centered cubic CeO₂. The XRD patterns agreed well with the standard CeO₂ crystal lattice (JCPDS card No. 04-013-4459) (Fig. S1A,† curve b). Fig. S1A† (curve c) shows the XRD profiles of CeNPs@MWCNTs with sharp diffraction peaks at (111), (200), (220), (311), (222), and (400) for the face-centered cubic CeO₂. Notably, the XRD peaks

of MWCNTs disappeared from the CeNPs@MWCNTs nanocomposite. This may be attributed to the high degree of CeNPs wrapped on MWCNTs. Furthermore, MWCNTs are used to support metal oxides (CeNPs) to enhance the catalytic properties of MWCNT composites through the strong interactions between them.⁴² Fig. S1B† shows the TEM image of the CeNPs material, indicating that it consists of a large number of particles. The magnified HR-TEM image shown in Fig. S1C† shows the lattice spacing of about 0.34 nm, hkl (111), and 0.29 nm, hkl (200), which are typical for cubic CeO₂ structures. The histogram of Fig. S1D† reveals the particle size distribution with an average diameter of 12 nm.

The surface morphology of the CeNPs/MWCNTs electrocatalyst (2 : 1 w/w) was examined by SEM and TEM techniques (Fig. 1A and B). As can be seen, most of the MWCNTs are decorated with CeNPs, which provides a large active surface area. The surface morphology of GCMFE and CeNPs@MWCNTs electrocatalyst-modified GCMFE was studied by SEM. Fig. 1C shows highly packed non-porous microspheres and no topographic features characterize the surface of the GCM-fluorolube oil electrode (GCMFE); however, Fig. 1D displays how the CeNPs@MWCNTs electrocatalyst helps to wire each GCM into an electrical contact and to bind and stabilize the microspheres within the film structure.

Electrochemical analysis

The electrochemical impedance spectroscopy (EIS) performance of the bare GCMFE and fabricated electrodes were

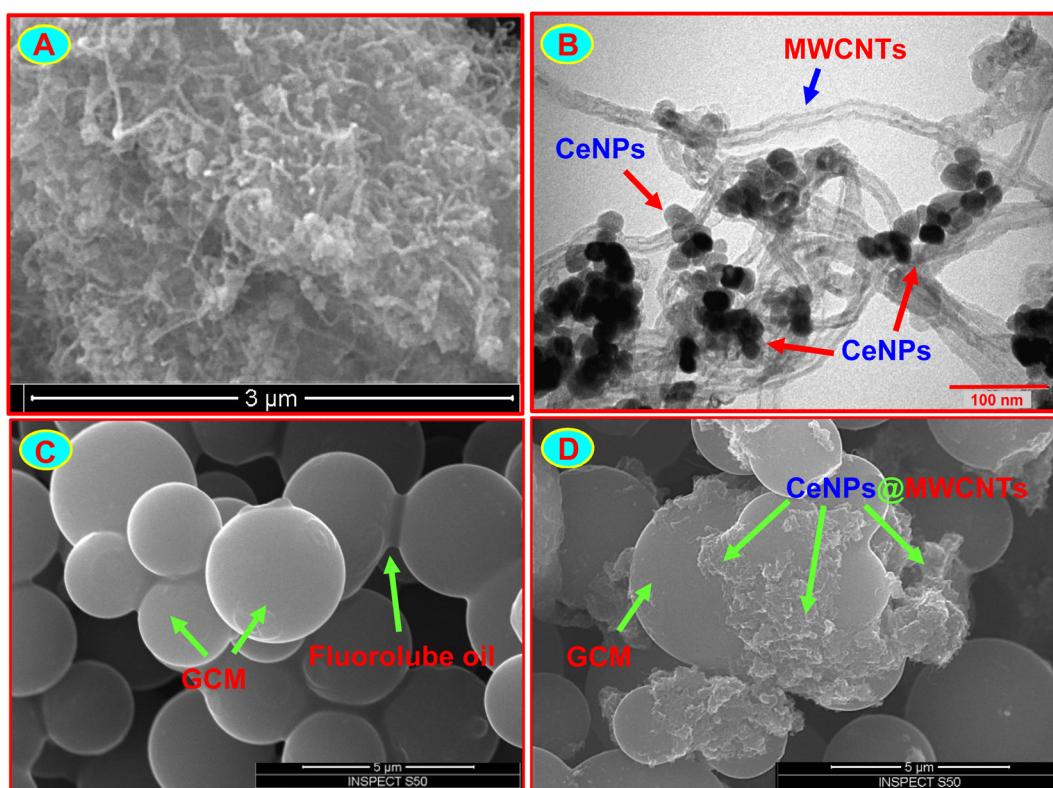


Fig. 1 SEM (A) and TEM (B) images of CeNPs@MWCNTs. SEM images of GCMFE (C) and CeNPs@MWCNTs/GCMFE (D).



evaluated in a three-electrode system containing 5 mM $\text{K}_3\text{Fe}(\text{CN})_6/\text{K}_4\text{Fe}(\text{CN})_6$ and 0.1 M KCl (Fig. 2A). It was observed that GCMFE presented a charge transfer resistance ($R_{\text{ct}} = 2225 \pm 46 \Omega$) lower than GCMPE ($R_{\text{ct}} = 3025 \pm 53 \Omega$), which may be related to the presence of fluorolube oil, accelerating the electronic transfer. When modified by CeNPs, R_{ct} considerably decreased to $1438 \pm 38 \Omega$. After the application of the MWCNTs nanomaterial to the electrode surface, the resistance decreased to $963 \pm 24 \Omega$, which indicates that the presence of MWCNTs may facilitate the transfer of charge and improve the conductivity of the electrode. Among the electrode materials, CeNPs@MWCNTs modified GCMFE exhibits the smallest R_{ct} value ($225 \pm 13 \Omega$). This result is mainly due to the benefits of the synergistic effect of CeNPs with large specific surface areas and MWCNTs with high electrical conductivity.

Characterization of the modified electrodes was performed by cyclic voltammetry (CV) and the obtained data are shown in Fig. 2B. After the GCMFE was incorporated into CeNPs, MWCNTs, and CeNPs@MWCNTs nanomaterials, the potential difference (ΔE_p) between the redox peaks of the $\text{K}_3\text{Fe}(\text{CN})_6/\text{K}_4\text{Fe}(\text{CN})_6$ decreased to 223, 198 and 161 mV, respectively, accompanied by the peak current increment. This indicates that the CeNPs@MWCNTs/GCMFE have higher catalytic ability than bare GCMPE and CeNPs, MWCNTs modified GCMFEs. The EIS data agreed with the CV and demonstrated the successful construction of the electrochemical sensor.

The values of the charge transfer rate constant (k_{et}) for the modified electrodes were calculated from R_{ct} via eqn (1):⁴³

$$k_{\text{et}} = RT/F^2 R_{\text{ct}} A C \quad (1)$$

The k_{et} values for GCMPE, GCMFE, CeNPs/GCMFE, MWCNTs/GCMFE and CeNPs@MWCNTs/GCMFE were measured as 2.5×10^{-4} , 3.4×10^{-4} , 6.0×10^{-4} , 8.0×10^{-4} , and $33.8 \times 10^{-4} \text{ cm s}^{-1}$, respectively. The larger k_{et} value of the CeNPs@MWCNT-modified GCMFE compared with those of the other electrodes showed highly efficient electron transport at the electrolyte/electrode interface of CeNPs@MWCNTs/GCMFE.⁴⁴ Moreover, to estimate the electrocatalytic ability, the exchange current (I_0) is frequently measured using eqn (2):⁴⁵

$$I_0 = RT/nFR_{\text{ct}} \quad (2)$$

The I_0 values calculated for GCMPE, GCMFE, CeNPs/GCMFE, MWCNTs/GCMFE, and CeNPs@MWCNTs/GCMFE were 8.5 , 11.5 , 17.8 , 26.7 , and $114.0 \mu\text{A cm}^{-2}$, respectively, which confirmed the higher electrocatalytic activity of the CeNPs@MWCNTs electrocatalyst. Effective surface area (Fig. S2† and 2C) was calculated using Randles–Sevcik eqn (3).^{46,47}

$$I_{\text{pa}} = (2.69 \times 10^5) n^{3/2} A_{\text{eff}} D^{1/2} v^{1/2} C \quad (3)$$

where A_{eff} represents the effective surface area, I_{pa} represents the anodic peak current, n represents the number of electrons transferred in the redox reaction ($n = 1$), C represents the concentration of $[\text{Fe}(\text{CN})_6]^{3-/-4-}$ (mol cm^{-3}), and D represents the diffusion coefficient of the ferricyanide solution ($7.6 \times 10^{-6} \text{ cm}^2 \text{ s}^{-1}$). The obtained A_{eff} values increased in the following order: CeNPs@MWCNTs/GCMFE ($0.86 \pm 0.022 \text{ cm}^2$) > MWCNTs/GCMFE ($0.43 \pm 0.017 \text{ cm}^2$) > CeNPs/GCMFE ($0.33 \pm 0.014 \text{ cm}^2$) > GCMFE ($0.25 \pm 0.010 \text{ cm}^2$) > GCMPE ($0.19 \pm 0.006 \text{ cm}^2$). Finally, CeNPs@MWCNTs/GCMFE possessed the most A_{eff} among the electrodes, indicating that the CeNPs@MWCNTs electrocatalyst-modified GCMFE exhibited improved electrocatalytic performance. Furthermore, the surface roughness factor ($R_f = A_{\text{eff}}/A_G$) assessed for CeNPs@MWCNTs/GCMFE ($R_f = 12.6$) was larger than that of GCMPE ($R_f = 2.7$). The results show that the CeNPs@MWCNTs electrocatalyst has a higher surface area and therefore provides more catalytic reaction sites, thereby accelerating the electron mass transfer efficiency.

Electro-catalytic oxidation of MPV and PCM

CVs of the unmodified and differently modified working electrodes toward MPV and PCM were examined in the Britton–Robinson buffer (BRB). The oxidation peak of MPV at 440 mV was relatively weak and broad at bare GCMPE, indicating the lower electrocatalytic activity of the unmodified electrode (Fig. 3A). Compared with the bare GCMPE, the oxidative peak negatively shifted to 338 mV when CeNPs@MWCNTs/GCMFE was used. Importantly, the peak current observed at the CeNPs@MWCNTs electrocatalyst-modified GCMFE was almost 5 times that of the unmodified GCMPE, which can be attributed to the electrocatalytic performance of CeNPs@MWCNTs/GCMFE on the electro-oxidation of MPV. Then, the electrochemical performances

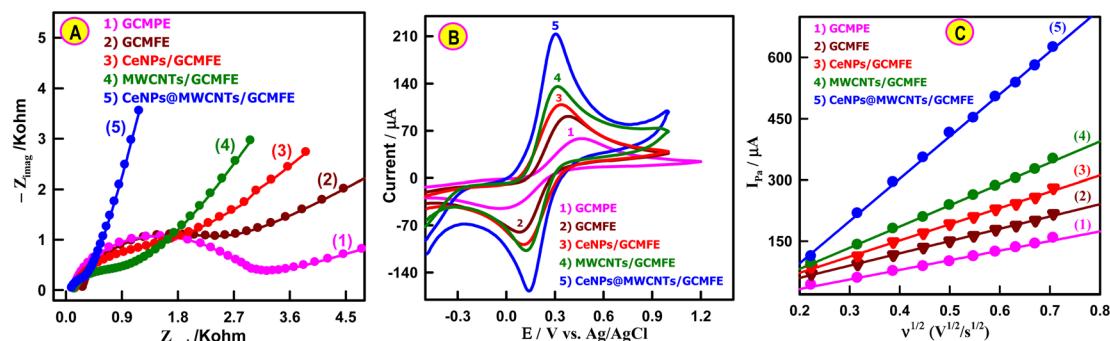


Fig. 2 (A) Nyquist plots and (B) CVs of (1) GCMPE, (2) GCMFE, (3) CeNPs/GCMFE, (4) MWCNTs/GCMFE and (5) CeNPs@MWCNTs/GCMFE in 5 mM $[\text{Fe}(\text{CN})_6]^{3-/-4-}$ (1 : 1 mixture) solution containing 0.1 M KCl at a scan rate of 100 mV s^{-1} . (C) Plot of I_{pa} vs. v .



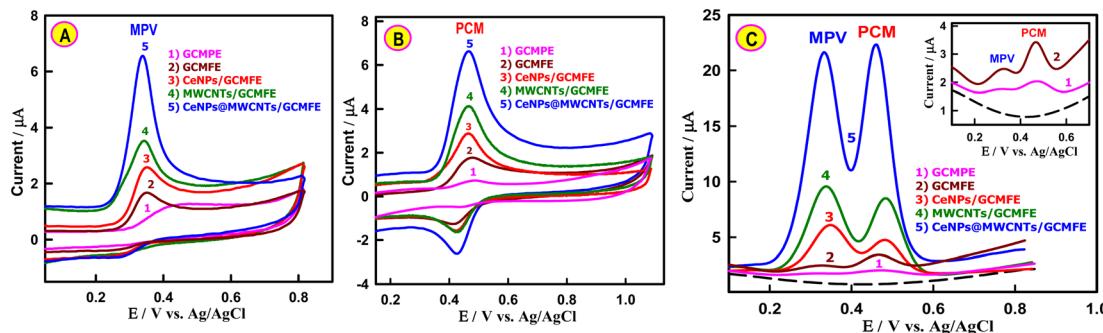


Fig. 3 CVs of (A) $1.5 \mu\text{M}$ MPV and of $1.2 \mu\text{M}$ PCM (B) in BRB solution of pH 6.0 obtained at (1) GCMPE, (2) GCMFE, (3) CeNPs/GCMFE, (4) MWCNTs/GCMFE and (5) CeNPs@MWCNTs/GCMFE. (C) Ads-SW voltammograms at (1) GCMPE, (2) GCMFE, (3) CeNPs/GCMFE, (4) MWCNTs/GCMFE, and (5) CeNPs@MWCNTs/GCMFE in BRB solution (pH 6.0) containing $2.6 \mu\text{M}$ MPV and $2.0 \mu\text{M}$ PCM.

of PCM at bare GCMPE, GCMFE, CeNPs/GCMFE, MWCNTs/GCMFE, and CeNPs@MWCNTs/GCMFE were investigated (Fig. 3B). Comparing the related CVs, the bare GCMPE demonstrates a pair of weak redox peaks, while CeNPs@MWCNTs/GCMFE possesses a pair of well-defined redox peaks with higher current responses. The CeNPs@MWCNTs/GCMFE sensor is more responsive to PCM compared to the unmodified GCMPE owing to its excellent electroactive area.

In addition, the simultaneous electrocatalytic responses of $2.6 \mu\text{M}$ MPV and $2.0 \mu\text{M}$ PCM were explored by adsorptive stripping square-wave voltammetry (Ads-SWV) in BRB solution of pH 6.0. At the bare GCMPE (inset of Fig. 3C), the MPV and PCM exhibited two small anodic peaks compared to the GCMFE, suggesting low conductivity and slow electron transfer of paraffin oil as a binder. However, after incorporating CeNPs or MWCNTs into GCMFE, the oxidation peak currents of MPV and PCM improved (Fig. 3C, curves 3 and 4). Interestingly, the fabricated CeNPs@MWCNTs/GCMFE showed considerably higher MPV and PCM oxidation currents (12-fold and 11.2-fold, respectively) compared to bare GCMPE (Fig. 3C, curve 5). This significant change was mainly attributed to the large electroactive surface area of the nanostructured CeNPs@MWCNTs that facilitated the contact of the electrolyte with the electrode surface, thus improving the detection of MPV and PCM by strengthening their absorption on the electrode.

Optimization studies

To acquire the strongest signal response and the highest sensitivity for the detection of MPV and PCM, we optimized the experimental conditions using the Ads-SWV method, including the pH of the BRB solution, supporting electrolyte and mass ratio of CeNPs@MWCNTs electrocatalyst to GCM. The effect of the pH range of the buffer, from 3 to 8, on peak currents and potentials of the two analytes in the BRB solution containing $2.6 \mu\text{M}$ MPV and $2.0 \mu\text{M}$ PCM on CeNPs@MWCNTs/GCMFE was investigated. The voltammograms displayed two oxidation peaks at all pH values (Fig. S3A†). When pH was 6.0, the current responses of MPV and PCM had the maximum value; therefore, a BRB solution with a pH of 6.0 was used for the experiments. Furthermore, the peak potential (E_{pa}) gradually moved in a more negative

direction with the increase of pH, which indicates that the proton participates in the redox reaction process. The regression equations are provided as part of the analysis, $E_{\text{Pa,MPV}} (\text{V}) = 0.71 - 0.061 \text{ pH}$, $R^2 = 0.9981$ and $E_{\text{Pa,PCM}} (\text{V}) = 0.9 - 0.064 \text{ pH}$, $R^2 = 0.9986$ (Fig. S3B and C†). The slope values in these equations were 61 and 64 mV pH^{-1} , which were almost consistent with the Nernstian standard value of 59.0 mV pH^{-1} . This result indicated that the electro-oxidation process of MPV and PCM on CeNPs@MWCNTs electrocatalyst-modified GCMFE involved an equal number of electrons and protons. Detailed information regarding the influence of the supporting electrolyte, mass ratio of CeNPs@MWCNTs electrocatalyst to GCM, and Ads-SWV parameters can be found in the ESI.†

Impact of the scan rate

The study of electrode kinetic characteristics can be realized by researching the scanning rate. Fig. S6A and D† show the CV curves of CeNPs@MWCNTs modified GCMFE with the scanning rate increasing from 50 to 400 mV s^{-1} in MPV and PCM, respectively. The illustrations show the linear relationship between I_{pa} and the scan rate (ν) (Fig. S6B and E†). The linear regression equations were: $I_{\text{pa}} (\mu\text{A}) = 2.5 + 0.042\nu (\text{mV s}^{-1})$ ($R^2 = 0.992$) for MPV, $I_{\text{pa}} (\mu\text{A}) = 2.03 + 0.05\nu (\text{mV s}^{-1})$ ($R^2 = 0.996$) for PCM. Moreover, when examining the plot correlating the logarithm of I_{pa} and logarithm of ν (Fig. S6C and F†), linear equations were derived, and the corresponding slopes were 0.75 and 0.8, respectively. The results indicated that MPV and PCM oxidation on the CeNPs@MWCNTs/GCMFE were controlled by the kinetic adsorption process. Moreover, the E_{pa} was shifted towards the anodic side by increasing the scan rate. Plots illustrating the relationship between E_{pa} and the natural logarithm of ν were generated (Fig. S6G and H†). The corresponding linear equations between the peak potential and natural logarithm of the scan rate emerged: $E_{\text{Pa(MPV)}} (\text{V}) = 0.4 + 0.024 \ln \nu$ ($R^2 = 0.991$) and $E_{\text{Pa(PCM)}} (\text{V}) = 0.5 + 0.023 \ln \nu$ ($R^2 = 0.989$). As per the Laviron equation,⁴⁸ the values of n during the oxidation of MPV and PCM were calculated to be 2.14 and 2.2, respectively. Therefore, the results indicated that the electrochemical reactions of MPV and PCM on CeNPs@MWCNTs electrocatalyst-modified GCMFE involved two electrons and two protons.



Mechanism of detection of MPV and PCM

The electrochemical-sensing mechanisms of both drugs were proposed based on the results of the pH, sweep rate, and EIS measurements. Scheme 2 shows the possible mechanisms of the electrochemical reactions of MPV and PCM at CeNPs@MWCNTs electrocatalyst-modified GCMFE. As is known, MWCNTs are a good electron acceptor and CeNPs are a good electron donor.⁴⁹ The redox properties of CeNPs with dual oxidation states ($\text{Ce}^{4+}/\text{Ce}^{3+}$) co-existing on the surface may act as oxidants for biomolecule electro-oxidation.⁵⁰ They can provide a large number of reactive oxygen species and active sites and improve the electrical conductivity of the nanocatalyst materials.⁵¹ This will increase the electrons capable of transferring from CeNPs to MWCNTs, thereby obtaining special electrochemical properties by promoting the oxidation reactions of MPV and PCM. Equal numbers of electrons and protons participated in the oxidation of both drugs, as confirmed by the pH and scan rate studies. The low R_{ct} ($225 \pm 13 \Omega$) and the increased surface area ($0.86 \pm 0.022 \text{ cm}^2$) of the CeNPs@MWCNTs/GCMFE caused the rapid transfer of electrons to the electrode. Therefore, CeNPs@MWCNTs electrocatalyst-modified GCMFE is more suitable for the simultaneous determination of MPV and PCM in practical applications.

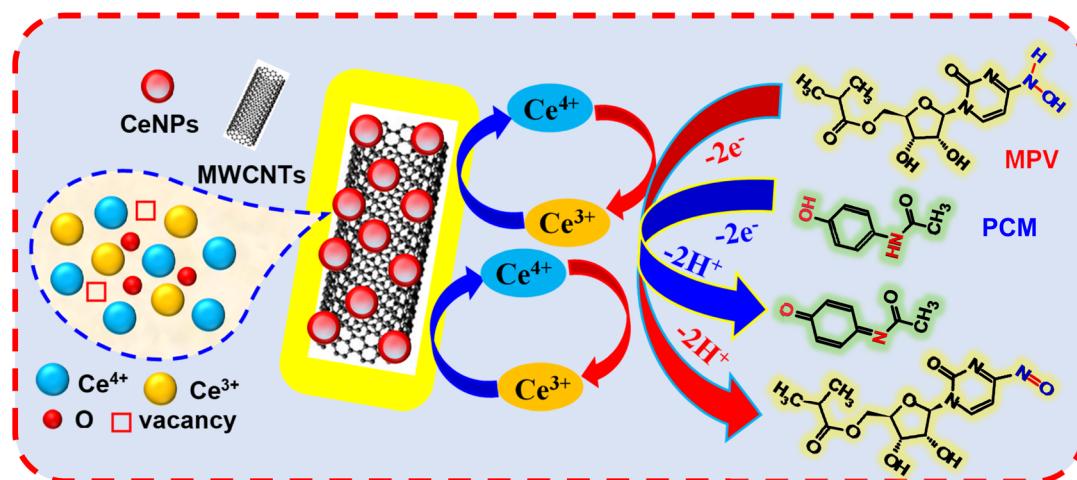
Selective determination of MPV and PCM

The electrochemical performance of CeNPs@MWCNTs electrocatalyst-modified GCMFE towards the selective assays of MPV and PCM was monitored. A controlled AdS-SWV experiment was performed at the CeNPs@MWCNTs/GCMFE sensor, keeping the concentration of PCM (2.8 μM) fixed with a variable concentration of MPV from 8.0 to 3500 nM (Fig. S7A†). The anodic response of MPV increases linearly with the increasing addition of MPV, as can be seen in Fig. S7A† (inset). The LOD, LOQ, and sensitivity of MPV were estimated to be 8.2 nM, 27 nM, and $78.6 \mu\text{A } \mu\text{M}^{-1} \text{ cm}^{-2}$, respectively. Similarly, as the concentration of PCM increased from 10.0 to 2500 nM in the presence of 2.6 μM MPV, the current responses of PCM gradually increased while the peak current of MPV remained almost

unchanged, which revealed that the concentration variation of PCM did not interfere with the peak current of MPV (Fig. S7B†). The sensor presented excellent analytical performance toward PCM detection with high sensitivity ($77.1 \mu\text{A } \mu\text{M}^{-1} \text{ cm}^{-2}$) and a low LOD (10.5 nM). This demonstrated the high-resolution CeNPs@MWCNTs electrocatalyst-modified GCMFE for simultaneous detection of MPV and PCM in their binary mixture without interfering with each other.

Simultaneous determination of MPV and PCM

Simultaneous monitoring of MPV and PCM is a crucial necessity because they are co-administered for COVID-19 patients. Under optimized AdS-SWV conditions (Fig. S5†), the simultaneous determination of MPV and PCM at CeNPs@MWCNTs/GCMFE in the BRB solution (pH 6.0) was analyzed. Fig. 4A shows the voltammetric responses of the two analytes at the fabricated sensor. It shows that both the MPV and PCM peaks are well delineated, and the ΔE_{pa} of MPV-PCM was 160 mV. Fig. 4B and C show that the current responses of MPV and PCM linearly increased with their concentrations. The analytical parameters and sensitivity using the AdS-SWV technique for both drugs are presented in Table S2.† To evaluate the performance and sensitivity of the CeNPs@MWCNTs/GCMFE sensor, it was compared with other reported electrodes for the individual-level detection of MPV and PCM (Table 1). This newly fabricated sensor displayed remarkable electrochemical performance for the simultaneous determination of MPV and PCM with low LODs of 6.0 nM and 8.6 nM, respectively, and a wide LDR from 5.0 to 5120 nM for MPV and from 8.0 to 4160 nM for PCM compared to previously modified electrodes. The following factors led to the impressive performance of the sensor that was constructed: (i) the large effective surface area has more active sites and adsorption centers; (ii) the introduction of MWCNTs effectively improves the electrical conductivity of nanoceria; and (iii) MWCNTs and nanoceria play a synergistic role to considerably improve the detection performance and sensitivity of the sensing platform.



Scheme 2 The proposed electrochemical oxidation mechanism of MPV and PCM at CeNPs@MWCNTs/GCMFE.



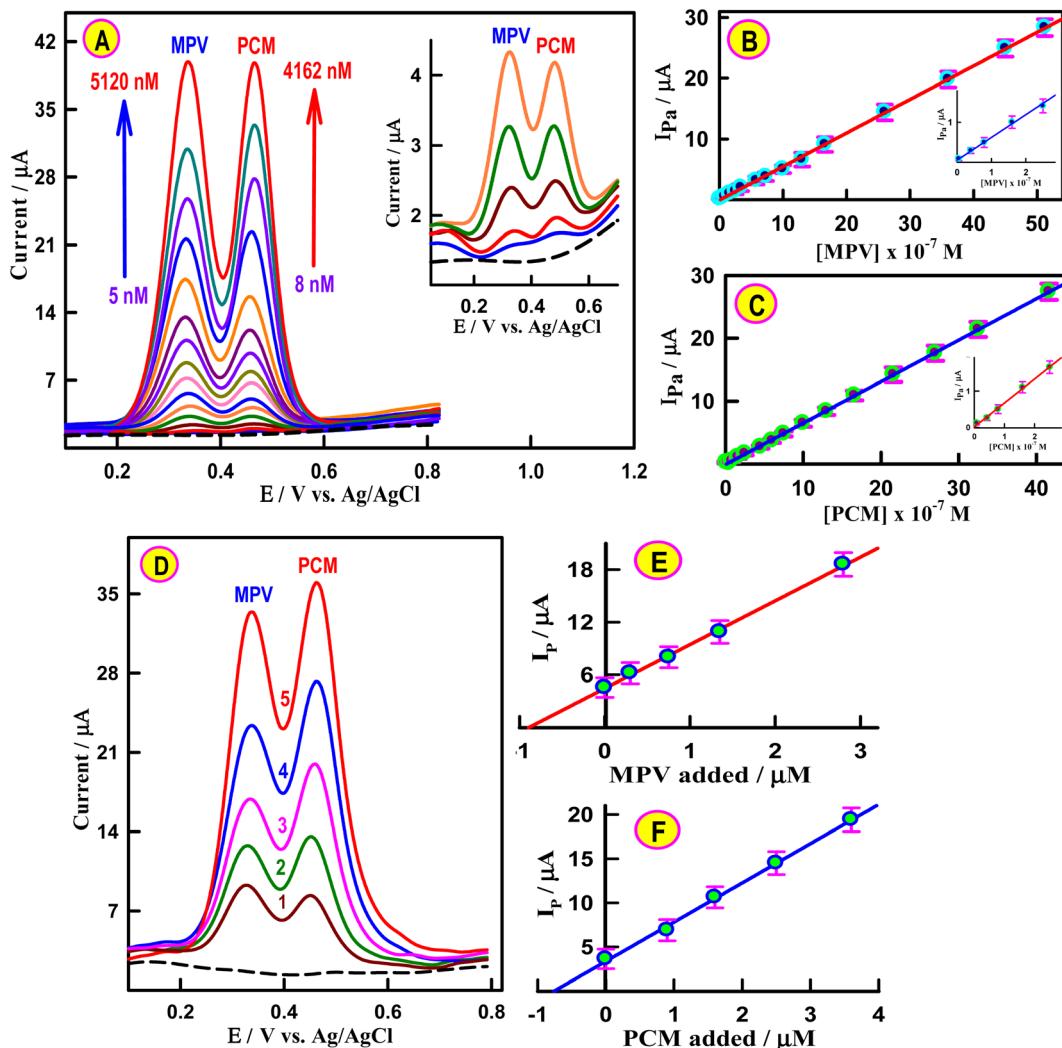


Fig. 4 (A) AdS-SW voltammograms of simultaneous determination of MPV (5.0–5120 nM) and PCM (8.0–4162 nM) CeNPs@MWCNTs/GCMFE in BRB solution (pH 6.0). (B&C) Corresponding calibration plots for MPV and PCM. (D) AdS-SW voltammograms of (1) the diluted samples of Molnupiravir-Rameda and Panadol tablets formulation and after standard additions of (2) 0.3 + 0.9, (3) 0.75 + 1.6, (4) 1.4 + 2.5, and (5) 2.8 + 3.6 μM MPV + PCM in BRB solution (pH 6.0) at CeNPs@MWCNTs/GCMFE sensor. (E and F) Corresponding linear calibration plots of peak current vs. concentration of added MPV and PCM. The error bar represents the standard deviation of triple measurements.

Table 1 A comparison of electrodes for the individual determination of MPV and PCM

Analyte	Electrode	LDR/ μM	LOD/nM	Ref.
MPV	rGO-GCE	0.09–4.57	30	17
	Fe_3O_4 /CPE	0.25–1500	50	18
	Cu-BTC@EG-PEDOT:PSS/GCE:PSS/GCE	0.1–63.3	71.3	19
	rGO-SPCE	0.152–18.272	48	20
PCM	AuNPs/MWCNT/GCE	0.09–35	30	21
	PEDOT/GO/GCE	10 – 60	570	22
	Pt/NDG	0.05–90	8.0	23
	GCE/Pd-SB	1.0–50	67	24
	γ -Fe ₂ O ₃ /CNTs	2.5–385	46	25
	ZnO/FMWCNT/CPE	0.5–13	23	26
MPV and PCM	CeNPs@MWCNTs/GCMFE	MPV: 0.005–5.12 PCM: 0.008–4.16	6.0 8.6	This work

Table 2 Recovery of MPV and PCM in human serum and urine samples ($n = 5$)^a

Compound	Spiked (nM)	Serum			Urine		
		Found (nM)	Precision RSD (%)	Recovery (%)	Found (nM)	Precision RSD (%)	Recovery (%)
MPV	70	67 ± 0.15	2.1	95.7	68.5 ± 0.1	1.7	97.9
	350	345 ± 0.5	1.8	98.6	347 ± 0.4	2.3	99.1
	500	508 ± 0.3	2.4	101.6	504.5 ± 0.2	1.9	100.9
PCM	40	40.8 ± 0.12	2.2	102	39.5 ± 0.2	1.6	98.7
	160	158 ± 0.24	1.6	98.7	162 ± 0.4	2.1	101.2
	320	323 ± 0.43	1.9	100.9	321.5 ± 0.5	2.4	100.5

^a Found: mean ± standard error.

Simultaneous analysis of MPV and PCM in real samples

Prior to testing the credibility of the proposed sensor, an examination of its repeatability, reproducibility, and stability was conducted (Fig. S8 and S9†). The simultaneous analysis of MPV and PCM in biological fluids such as human blood serum and urine samples demonstrated the practical application of the sensing electrode. Human blood serum and urine samples were obtained from the hospital of the Assiut University and used shortly after the collection. Fig. S10A† shows representative Ads-SW voltammograms for the simultaneous assay of the coexistence of MPV and PCM spiked in human serum samples. As shown in Fig. S10A† the two peaks at 0.33 V and 0.47 V correspond to the oxidation of MPV and PCM, respectively, and are used for quantification of these drugs in serum samples. Potentially interfering compounds that may be present in serum samples do not indicate any electrochemical response where the analytical peaks appear. As can be seen in Fig. S10B and C,† the oxidation peak currents of MPV and PCM are linearly proportional to their concentrations. The LOD values for MPV and PCM in the spiked serum samples were 7.2 and 10.7 nM, respectively.

The constructed sensor was successfully applied for the simultaneous analysis of the two drugs in spiked human urine samples (Fig. S11A†). Linear relationships were obtained between the peak currents and the concentrations of MPV and PCM in the urine sample (Fig. S11B and C†). The LDR, LOD, LOQ, and sensitivity were calculated and are shown in Table S3.† The calculated recovery results of the MPV and PCM concentrations in biological samples were between 97.9% and 101.2%, with RSDs lower than 2.5% (Table 2). Furthermore, the suggested Ads-SWV was utilized for monitoring MPV and PCM in the pharmaceutical products (Molnupiravir-Rameda 200 mg, Panadol 500 mg) using the constructed sensor (Fig. 4D–F). The recovery values listed in Table S4† were in the range of 96.8–102.3%. Based on these results, CeNPs@MWCNTs electrocatalyst-modified GCMFE demonstrates an excellent capability for the simultaneous electrochemical monitoring of MPV and PCM in human biological samples and commercial tablet dosage forms.

Conclusions

Sensitive sensing platforms have become an effective method to analyze biological compounds. Herein, we constructed a novel

sensor based on nanoceria-decorated MWCNTs electrocatalyst (CeNPs@MWCNTs) for the simultaneous detection of antiviral drugs MPV and PCM. Morphological analysis of the modified electrode clearly revealed that the CeNPs@MWCNTs electrocatalyst was incorporated into the glassy carbon microsphere fluorolube oil paste electrode (GCMFE), which enhanced the electrochemical performance toward simultaneous detection of MPV and PCM. The electrochemical analysis studied for MPV and PCM detection experienced a good current response, which was enhanced owing to the higher surface area and improved electron transfer to the surface. Furthermore, the fabricated sensor showed excellent detection limits, 6.0 nM, 8.6 nM, linear ranges, 5.0–5120 nM and 8.0–4162 nM, and sensitivities, 78.6 and 94.3 $\mu\text{A } \mu\text{M}^{-1} \text{ cm}^{-2}$, for simultaneous detection of MPV and PCM. Furthermore, the proposed sensor exhibited good repeatability, reproducibility, storage stability, and anti-interference capability toward MPV and PCM. In addition, it showed a satisfactory recovery rate in human biological fluids and pharmaceutical products. The obtained results suggested that CeNPs@MWCNT electrocatalyst-incorporated GCMs could become a prospective candidate for the construction of both solid and screen-printed electrodes in the future for the determination of MPV and PCM individually and simultaneously.

Conflicts of interest

There are no conflicts to declare.

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