



 Cite this: *RSC Adv.*, 2024, 14, 4065

Novel spectrofluorometric approach for assessing vilazodone by blocking photoinduced electron transfer: analytical performance, and greenness–blueness evaluation†

 Sayed M. Derayea,^a Hadeer A. Elhamdy,^b ^{*b} Khalid M. Badr El-Din^a and Mohamed Oraby^b

In this paper, vilazodone (VLD), a serotonin modulator prescribed for major depressive disorder, was investigated using a rapid, highly sensitive, and eco-friendly spectrofluorometric approach. The native fluorescence of VLD, originating from its indole moiety, exhibited an emission peak at 486 nm upon excitation at 241 nm. However, the presence of a piperazinyl nitrogen atom in the VLD structure, acting as an electron donor, significantly diminished the fluorescence intensity through photoinduced electron transfer (PET) to the indole ring. However, by protonating this nitrogen atom using 0.02 M Teorell–Stenhagen buffer (pH 3.5), inhibition of the PET process effectively blocked electron transfer, restoring the fluorescent properties of the drug. Further, an enhancement in the fluorescence was achieved by employing methanol as the solvent, resulting in a 1.5-fold increase. The combined use of PET blockage and methanol enabled the detection of VLD at levels as low as 0.78 ng mL⁻¹. Calibration analysis demonstrated linearity within the range 5–400 ng mL⁻¹, exhibiting a correlation coefficient of 0.9998 and a limit of quantification of 2.37 ng mL⁻¹. The method obeyed the requirements of International Council on Harmonization (ICH). The proposed approach was applied for the accurate measurement of VLD in pharmaceutical tablets, content uniformity testing based on USP requirements, and determining VLD concentration in spiked human plasma. Moreover, the environmental impact, in addition to practical effectiveness, of the proposed approach was evaluated using different metrics.

 Received 23rd November 2023
 Accepted 8th January 2024

DOI: 10.1039/d3ra08034j

rsc.li/rsc-advances

1. Introduction

Vilazodone (VLD) is a potent medication that is widely used in the treatment of major depressive disorder. As a selective serotonin reuptake inhibitor (SSRI) and a partial agonist of the serotonin 5-HT_{1A} receptor, VLD offers a unique pharmacological profile for managing depressive symptoms.¹ The chemical structure of VLD is shown in Fig. 1, and its formula can be given as 5-[4-[4-(5-cyano-1*H*-indol-3-yl)butyl]piperazin-1-yl]-1benzofuran-2-carboxamide. VLD is a piperazine-indole derivative that has properties as an agonist with partial activity at the 5-HT_{1a} receptor and as a selective serotonin reuptake inhibitor (SSRI),² which have seen its use in therapy for major depression.³

Accurate and sensitive analytical methods are essential for the characterization and quantification of VLD in pharmaceutical formulations and biological samples. Few articles have been

published reporting methods for testing VLD, which included spectrophotometry,^{4–7} spectrofluorimetry,⁸ HPTLC,^{7,9} HPLC,^{8,10–13} LC/MS,^{14,15} and UPLC.^{16,17} Although chromatographic methods^{8,10–13}

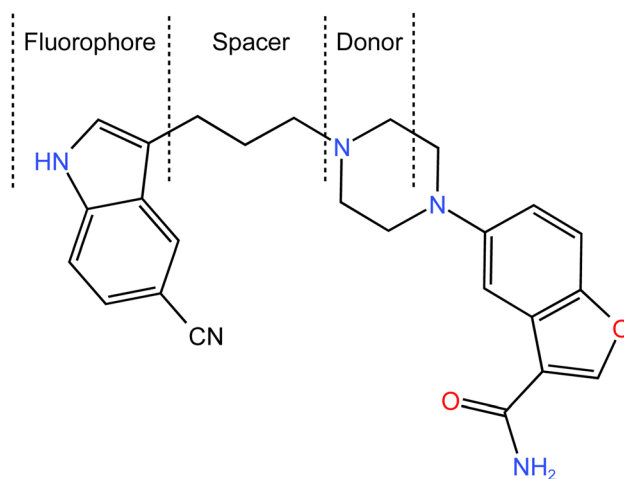


Fig. 1 Chemical structure of VLD, showing the components involved in the photoinduced electron-transfer process (fluorophore, spacer, and donor (receptor)).

^aDepartment of Pharmaceutical Analytical Chemistry, Faculty of Pharmacy, Minia University, Minia 61519, Egypt. E-mail: adelhadeer33@gmail.com

^bDepartment of Pharmaceutical Analytical Chemistry, Faculty of Pharmacy, Sohag University, Sohag 82524, Egypt

† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d3ra08034j>



have the merits of sensitivity and can be applied to separate different compounds in combinations and in the pharmacokinetic study of drugs and studies of degradation products, they utilize expensive equipment, high-purity reagents, and well-trained personnel.^{14,16,17} Alternatively, spectrofluorimetry, a powerful analytical technique, has gained considerable attention in pharmaceutical analysis owing to its high sensitivity and selectivity. In addition, it is simple, rapid, and utilizes less expensive solvents.

VLD has native fluorescence derived from its indole moiety. However, the presence of a piperazinyl nitrogen atom attached to the aliphatic bridge in the VLD structure introduces a significant challenge. This nitrogen atom acts as an electron donor and initiates photoinduced electron-transfer (PET), leading to a substantial reduction in the fluorescent signal.

To overcome this limitation, innovative strategies are required to effectively block the photoinduced electron transfer and restore the fluorescence of VLD. In this study, a highly sensitive spectrofluorimetric approach was proposed that employs PET blocking to enhance the detection and quantification of VLD. By protonating the piperazinyl nitrogen atom using the Teorell–Stenhagen buffer system (pH 3.5), the PET process could be efficiently inhibited, allowing the re-emergence of the drug's fluorescence.

Further, the choice of solvent plays a crucial role in enhancing the fluorescence signal. By carefully selecting the appropriate solvent, further fluorescence enhancement could be achieved, thus further improving the sensitivity and accuracy of VLD quantification.

In this research article, a comprehensive investigation of the spectrofluorimetric analysis of VLD is presented utilizing PET blocking and using methanol as a solvent. The developed approach allows the rapid, highly sensitive, and eco-friendly detection of VLD in pure form, and in pharmaceutical tablets. The great sensitivity of the suggested method could expand its application to examine VLD in biological samples. Furthermore, the ultimate simplicity of the current approach enabled its use in the content uniformity testing of commercial VLD tablets. The method's performance characteristics, including its limit of detection, linearity, and precision, were thoroughly evaluated, and validated according to regulatory guidelines. Moreover, the environmental impact of the proposed method was assessed using established indices, including the National Environmental Method Index (NEMI), eco-scale, GAPI, and AGREE, all highlighting its greenness, applicability, and sustainable attributes. In addition, the practical usefulness of the proposed method was evaluated using the Blue Applicability Grade Index (BAGI).

Overall, this study aimed to provide an innovative and reliable spectrofluorimetric approach for the assessment of VLD, contributing to the advancement of analytical methods in pharmaceutical analysis and facilitating the quality control of VLD-containing formulations.

2. Experimental

2.1. Instrumentation

Spectrofluorometric measurements were conducted using a Jasco FP-8350 spectrofluorometer (Hachioji, Tokyo, Japan).

The instrument features a 150 W Xe-arc lamp and a PMT tuned to 400 V. The emission and excitation monochromator slits were both 5 nm wide, and the scanning speed was 1000 nm min⁻¹. Aquatron Water Still A4000D system producing double-distilled water (Cole-Parmer, Staffordshire, UK), Jenway 3510 pH meter (Staffordshire, UK), and Mettler Toledo 5-digit balance (Greifensee, Switzerland) were also used in the study.

For measurement of the absorbance for the reported method, a T80 double beam UV-VIS spectrophotometer (PG Instruments, Leicestershire, UK) was used. Two matched 1 cm quartz cells were utilized in all the measurements.

2.2. Materials and reagents

VLD was kindly obtained from Rameda Co. for Pharmaceutical Industries & Diagnostic Reagents (6th of October City, Egypt). Vilaphoria® tablets (20 mg VLD) were bought *via* the local market. Analytical grade acetonitrile, methanol, ethanol, β -cyclodextrin (β -CD), Tween 80, and spectroscopic-grade perchloric acid were obtained from Merck (Darmstadt, Germany). Analytical grade sodium hydroxide was supplied by Fischer Scientific (Loughborough, United Kingdom). Sodium dodecyl sulfate (SDS) of analytical grade, cetyltrimethylammonium bromide (CTAB), acetone, sulfuric, phosphoric, acetic, and hydrochloric acid were purchased from El Nasr Pharmaceutical & Chemical Co. (Cairo, Egypt).

2.3. Stock solutions

To make the solution of VLD (1000 $\mu\text{g mL}^{-1}$), 25 mg of VLD was dispersed in methanol in an amount equal 25 mL. Next, 10 mL of that solution (1000 $\mu\text{g mL}^{-1}$) was placed in a volumetric flask measuring 100 mL and methanol was properly added to achieve a 100 $\mu\text{g mL}^{-1}$ solution of VLD. Various concentrations of standard VLD were made utilizing a similar solvent. These solutions were preserved in the fridge for use in the subsequent work.

2.4. General assay procedures

Standard VLD solutions at concentrations from 5–400 ng mL⁻¹ were put in 5 mL volumetric flasks, and 0.5 mL of Teorell–Stenhagen buffer (0.02 M, pH 3.5) was incorporated and then the flask was filled to the mark using methanol. The contents were then thoroughly mixed. The fluorescence intensity of the solutions was determined at 486 nm after excitation at 241 nm. The blank experiment was prepared in a similar way but without the addition of VLD solution. The resulting fluorescence *vs.* VLD concentrations were plotted to obtain a graph and the linear regression equation was obtained.

2.5. Procedures for method validation

In the evaluation of the accuracy, precision, and robustness of the proposed method, different standard solutions of the studied drug under the specified conditions were analyzed using the general procedure. The values of the measured fluorescence intensities were utilized to calculate the drug concentrations by applying a linear regression equation for the



calibration curve. The % recovery was calculated by dividing the found drug concentration by the claimed concentration in each standard solution, and then multiplying this by 100. The analysis was carried out in triplicate and the standard deviation was estimated.

2.6. Procedures for assessing VLD tablets

Ten Vilaphoria® tablets (20 mg per tablet) were weighed and then pulverized using a mortar and pestle. A weight of the finely powdered tablets corresponding to 20 mg VLD was added in to a 50 mL volumetric flask, and then 30 mL of methanol was introduced, and the solution was sonicated for 30 min and filled to the mark with the same the solvent. The contents of the flask were subjected to filtering, and the initial portion of the filtrate was discarded. Aliquots of this stock solution were diluted to provide concentrations that fell inside the linear range. The general assay procedure was performed as mentioned previously. The nominal concentration in the test sample solution was computed using the previously estimated regression equation.

For analyzing the commercial tablets with the reported method,⁵ different standard solutions of the drug (1.0–6.0 $\mu\text{g mL}^{-1}$) were prepared in methanol. The absorbance of each solution was measured at 241 nm against blank methanol. The calibration curve for the reported method was constructed by plotting the drug concentration *versus* the absorbance. After that, the powdered commercial tablet was extracted with methanol as described above. A portion of the filtrate was diluted with methanol to give 3.0 $\mu\text{g mL}^{-1}$ as a final concentration and its absorbance was measured at 241 nm. The obtained absorbance was utilized to determine the drug concentration in the analyzed sample solution. The obtained concentration was divided by the claimed concentration (3.0 $\mu\text{g mL}^{-1}$) in the sample solution and then multiplied by 100 to get the recovery %. All measurements were carried out in triplicate.

2.7. Procedure for assessing the content uniformity

The content uniformity testing of VLD in tablet formulation was performed following US guidelines (Chap. 905).¹⁸ Here, 10 Vilaphoria® 20 mg tablets were individually examined to affirm that their contents were homogeneous using the formerly mentioned procedures under the analysis of VLD tablets.

2.8. Procedures for evaluating VLD in human plasma spiked with VLD

Blood was collected from the forearm vein of a healthy volunteer and placed in a series of EDTA tubes. The procedures for collecting plasma from healthy volunteers were carried out according to the Recommendations of the Declaration of Helsinki.¹⁹ All experiments that included the use of human plasma were performed in accordance with the Guidelines Applied to Research on Human Subjects of Minia University and approved by the “Commission on the Ethics of Scientific Research”, Faculty of Pharmacy, Minia University. In all cases, informed written consent was obtained from each participant before donating the blood samples. A volume of 5 mL of blood was

separated into plasma by 20 min centrifuging at 4000 rpm. The obtained plasma was stored at $-20\text{ }^{\circ}\text{C}$. Next, 1.0 mL of plasma was transferred into a clean tube and mixed with 1.0 mL of VLD standard solution ($5\text{--}400\text{ }\mu\text{g mL}^{-1}$) and 2.0 mL acetonitrile was added to precipitate plasma protein. The tube was vortex-mixed for 1 min before centrifuging for 10 min at 4000 rpm. After transferring the transparent supernatant onto a 5 mL volumetric flask, the general assay method was carried out. Experiments were also performed for the blank prepared using drug-free plasma.

3. Results and discussion

The molecular structure of VLD contains two moieties that may be responsible for its native fluorescence: indole and benzofuran (Fig. 1). Actually, the indole moiety is the fluorophore since it possesses a conjugated system of double bonds with a rigid aromatic structure. This conjugation allows for the absorption of ultraviolet (UV) or visible light, leading to the excitation of electrons to higher energy levels. Upon relaxation, these excited electrons emit light, resulting in fluorescence. On the other hand, the benzofuran moiety does not possess the same features as the indole ring. Consequently, the benzofuran moiety is not typically responsible for fluorescence in this compound. The piperazinyl moiety in VLD contains two nitrogen atoms. The distal nitrogen atom from the benzofuran, which is attached to the aliphatic chain, can act as a potential electron donor. In its protonated form (when the pH is slightly acidic, between 2.0 and 6.0), this piperazinyl nitrogen atom becomes positively charged. This protonation alters the electronic properties of the molecule and disrupts the electron-transfer process. However, in its neutral unprotonated form (at pH higher than 6.0), the piperazinyl nitrogen is not protonated and can readily donate electrons to the indole moiety. In this state, the excited state of the indole can undergo the PET process, which involves the transfer of electrons from the donor (piperazinyl nitrogen) to the electron acceptor (indole). The structural properties of the drug make it susceptible to the PET process as in the case of amino(ethanesulfonate) anthracene indicator²⁰ and others.^{21,22} VLD contains the indole ring as a fluorophore, butyl bridge as a spacer, and the proximal piperazine nitrogen as an electron donor (Fig. 1). The PET process effectively quenches the fluorescence emission of the drug. When the pH of the medium is 6.0 or lower, the piperazinyl moiety becomes protonated, which hinders its ability to donate electrons. As a result, the inhibition of PET takes place, allowing the excited state of the molecule to persist for a longer duration and emit fluorescence. Therefore, the high fluorescence exhibited by VLD at pH between 2.0 and 6.0 can be attributed to the protonation of the piperazinyl moiety, which inhibits the photoinduced electron-transfer process and allows for the augmentation of the fluorescence emission.

VLD has an emission wavelength of 486 nm after an excitation wavelength of 241 nm (Fig. 2) and displayed an enhancement in VLD fluorescence upon the addition of Teorell–Stenhagen buffer (pH 3.5). In addition, an additional



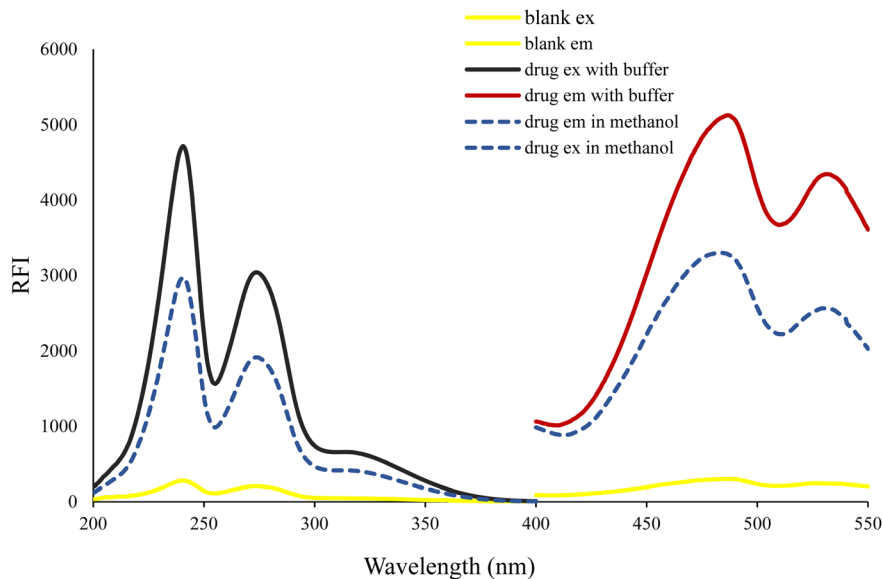


Fig. 2 Excitation and emission spectra of 200 ng mL⁻¹ VLD and the excitation and emission spectra of the blank in methanol.

enhancement of the fluorescence intensity of VLD was observed in the presence of methanol as a diluting solvent.

3.1 Improvement of the experimental parameters

The influence of various experimental parameters on the native fluorescence intensity of VLD were examined to get the optimal parameters to maximize the fluorescence intensity.

3.1.1 Effect of the buffer and pH modifier. To improve the intrinsic fluorescence of VLD, different solutions with varying pH values were examined. First, 1.0 mL aliquots of Teorell-Stenhagen buffer solution covering the pH ranges of 2.5–8.0 were investigated, as shown in Fig. S1†. In addition, the fluorescence intensity of VLD was also examined in both highly acidic and highly alkaline media (see ESI†). The examined acids were sulfuric, perchloric, acetic, and hydrochloric acids, while 1.0 mL of 1 M NaOH was also assessed to examine the performance in a highly alkaline pH medium (Fig. S2†). It was noticed that the highest fluorescence intensity was attained using acidic buffer solutions in the pH range of 2.0–6.0. The use of buffer solutions having a pH higher than 6.0 or the addition of 1.0 M NaOH dramatically diminished the fluorescence. This could be explained on the basis of the PET process. VLD contains an indole moiety, which acts as a fluorophore. This indole is linked to the benzofuran ring through an aliphatic bridge containing the piperazinyl moiety. Upon increasing the pH to higher than 6.0, the nitrogen atom is no longer protonated and thus its lone pair of electrons can be donated to the indole through the PET process, resulting in a severe decrease in the VLD fluorescence. This was also true in the case of using 1.0 mL of 1 M NaOH, which also reduced the native fluorescence of VLD.

To further confirm the protonation of the studied drug at pH below 6.0, the effect of the pH on the absorption spectrum of the drug was studied. The results are presented in the ESI (S1†)

Although the change was not high, it was clear that increasing the pH led to a slight but distinct decrease in absorbance at 241 nm. Furthermore, a very slight red shift in the λ_{max} at 241 nm was also observed upon increasing the pH above 6.0. Both changes in the position and absorbance confirmed the deprotonation of the drug by increasing the pH beyond 6.0.

At low pH values, below the pK_{a} of the relevant ionizable group, the nitrogen of the indole ring is likely to be protonated ($>\text{NH}_2^+$). This protonation can disrupt the electronic structure and conjugation within the indole ring, leading to a decrease in fluorescence efficiency. Protonation of the indole nitrogen can also increase the internal conversion or nonradiative decay processes, reducing the population of the excited state and hence decreasing the fluorescence intensity. Furthermore, in highly acidic medium (below pH 2.0), the cyano group attached to the indole moiety can be converted into the carboxylic functional group. The conversion of the cyano group ($-\text{CN}$) into carboxylic group ($-\text{COOH}$) can have a significant impact on the fluorescence properties of a molecule, often resulting in fluorescence quenching. This quenching effect is due to the higher electron-withdrawing nature of the carboxylic group compared to the cyano group, which can decrease the electron density within the indole fluorophore. The reduced electron density can lead to a less favorable excited state and an increased likelihood of nonradiative decay pathways, resulting in a decreased fluorescence intensity or shortened fluorescence lifetime. Furthermore, the fluorescence strength of VLD at pH 2.5–4.5 was increased by 1.5-fold compared with methanol alone. Hence, 0.02 M Teorell-Stenhagen solution of buffer at pH 3.5 was chosen for the following work (Fig. S1†).

3.1.2 Effect of the volume of the buffer. The impact of the Teorell-Stenhagen buffer volume (0.02 M, pH 3.5) on the intensity of VLD fluorescence was investigated utilizing various volumes (0.1–1 mL) of 0.02 M Teorell-Stenhagen buffer (pH



3.5). It was revealed that increasing the buffer volume led to an increase in the fluorescence intensity until reaching a steady state at 0.3–0.6 mL and then a decrease in the relative fluorescence intensity (RFI) was observed with further increasing the volume of the buffer. Therefore, the optimum volume for the maximum fluorescence intensity of VLD was 0.5 mL (Fig. 4).

3.1.3 Effects of various organized media. The study made use of a number of organized media to augment the fluorescence of the methanolic VLD solution. Surfactants with anionic properties (SDS, 0.5% w/v), cationic (CTAB), and Tween 80 0.5% v/v, and macromolecules (β -CD, 0.5% w/v) were examined (Fig. S3†). From the tested organized media, Tween 80 produced the highest enhancement in the fluorescence of VLD. In contrast, the intensity of VLD's fluorescence was dramatically decreased by β -CD. However, the augmentation effect produced by the organic solvents (methanol, ethanol) was much greater than that induced by Tween 80. The combined use of both Tween 80 and the organic solvents together, especially alcohols, to produce a synergistic effect on the fluorescence was not feasible owing to the destructive effect of these solvents on the micellar system of the surfactant. It was concluded that the use of these chemicals was not suitable for the optimal augmentation of the fluorescence of VLD. Therefore, none of these media were employed.

3.1.4 Effect of the solvent. Water, methanol, ethanol, acetone, and acetonitrile were considered to dilute VLD (Fig. 3). The intensity of fluorescence was severely decreased in acetone and water. It was noted that an increase in the fluorescence intensity was obtained in ethanol, acetonitrile, and methanol. In methanol, the highest intensity of fluorescence was observed and therefore this was used for the subsequent work.

The presence of an electron-transfer state in tryptophan to a nearby amide in the protein backbone, which is available in polar environments, may result in a quenching of the fluorescence.²³ Thus, the amide group attached to the benzofuran ring in VLD promoted fluorescence quenching, arising from a loss of energy owing to an electron transfer from the aromatic indole ring to this amide. Since the electron-transfer state is expected to be high in energy, normally this would not be a possible outcome, but because of its large dipole, such a state should be

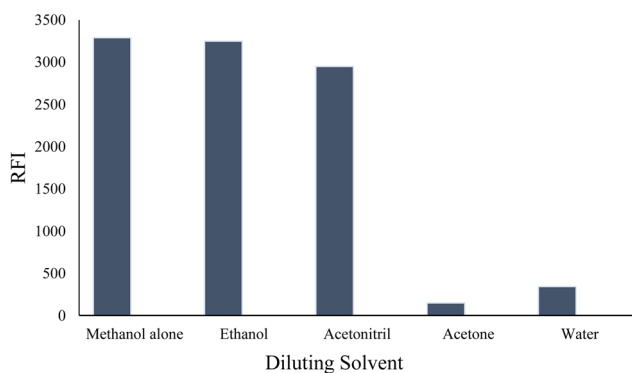


Fig. 3 Effect of different diluting solvents on the relative fluorescence intensity (RFI) of 200 ng per mL VLD.

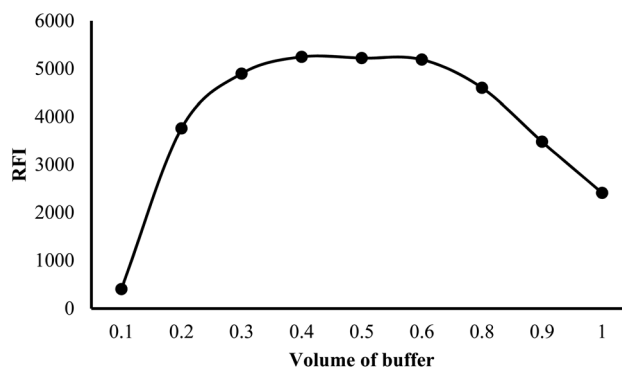


Fig. 4 Effect of different volumes of buffer on the relative fluorescence intensity (RFI) of 200 ng per mL VLD.

more accessible in polar solvents, such as water, which is a polar protic solvent with a relatively high dielectric constant.

3.2 Validation of the method

The validation of the analytical process was carried out in accordance with the International Council on Harmonization (ICH) guidelines.²⁴ The study included assessing the linearity, range, limit of detection (LOD), limit of quantification (LOQ), accuracy, precision, and robustness.

3.2.1 Linearity and range. Under optimal conditions, the calibration curve of VLD was obtained by plotting different concentrations of VLD against the fluorescence intensity. The curve that was used for calibration was linear in the 5–400 ng mL⁻¹ range and had a high correlation coefficient (r) value of 0.9998 and determination coefficient (r^2) value of 0.9996. The statistical parameters for the proposed spectrofluorometric method are summarized in Table 1.

3.2.2 Quantification (LOQ) and detection (LOD) limits. The LOD and LOQ values were calculated depending on the slope and intercept standard deviation of the calibration graph. The LOD and LOQ values were discovered to be 0.78 and 2.37 ng mL⁻¹, respectively, indicating a high sensitivity of the proposed method for the analysis of VLD (Table 1). The ICH equations $LOD = 3.3SD/b$ and $LOQ = 10SD/b$ (b = slope and SD = standard deviation of the intercept) were utilized to calculate the LOD and LOQ.

Table 1 Regression equation and validation parameters for the determination of VLD with the proposed spectrofluorometric method

Parameters	Value
Linear range (ng mL ⁻¹)	5–400
Slope (b)	23.72
Standard deviation of slope (S_b)	0.20
Intercept (a)	131.54
Standard deviation of intercept (S_a)	5.63
Correlation coefficient (r)	0.9998
Determination coefficient (r^2)	0.9996
Number of determinations	5
Limit of detection (LOD ng mL ⁻¹)	0.78
Limit of quantification (LOQ ng mL ⁻¹)	2.37



Table 2 Evaluation of the accuracy of the analytical spectrofluorometric method for VLD determination

Amount taken (ng mL ⁻¹)	Amount found (ng mL ⁻¹)	% Recovery ^a ± SD
50	50.67	101.34 ± 1.23
100	98.61	98.61 ± 0.72
200	201.02	100.51 ± 0.60
300	302.49	100.83 ± 1.83
400	400.16	100.04 ± 0.73

^a Mean of three determinations, SD, standard deviation.

3.2.3 Accuracy. Based on the estimation of the standard deviation (SD) and % recovery, the accuracy of the suggested method's accuracy was estimated for five distinct concentrations of VLD standard solution (50, 100, 200, 300, and 400 ng mL⁻¹). Both standard deviations and the % recovery were determined in triplicate for each concentration. The % recoveries were found to be in the range of 98.61–101.34%, indicating the good accuracy of the suggested method due to the closeness of the calculated results to the true values. Table 2 summarizes the findings.

3.2.4 Precision. The repeatability (intra-day) and reproducibility (inter-day) precisions of the suggested approach were checked by evaluating three distinct concentrations of VLD (50, 200, and 400 ng mL⁻¹) in triplicate using the general assay procedures. The analysis of the three concentrations was carried out within one day for the intra-day precision and three successive days for the inter-day precision. The intra-day precision relative standard deviation (RSD) was found to be 1.23% (50 ng mL⁻¹), 0.6% (200 ng mL⁻¹), and 0.73% (400 ng mL⁻¹). The inter-day precision (RSD) was found to be 1.71% (50 ng mL⁻¹), 1.73% (200 ng mL⁻¹), and 1.27% (400 ng mL⁻¹). The findings shown in Table 3 demonstrate that the RSD was less than 2%, showing that the suggested approach had a high degree of precision.

3.2.5 Robustness. The study of the robustness involved studying the effect of minor changes in method variables, including pH (3.5 ± 0.1) and the buffer solution volume (0.5 ± 0.1), on the method's sensitivity and suitability. The outcomes in Table 4 show that the effect of these variables on the method's sensitivity was not significant. Therefore, the suggested method was considered robust.

3.3 Applications

3.3.1 Analysis of real pharmaceutical preparations. Ten Vilaphoria tablets (20 mg VLD) were subjected to VLD analysis

Table 4 Robustness of the proposed method for the determination of VLD (200 ng mL⁻¹) in pure form

Parameter		% Recovery ± SD ^a
pH of buffer	3.4	98.58 ± 0.48
	3.6	99.29 ± 0.32
Volume of buffer (mL)	0.4	98.91 ± 1.28
	0.6	98.07 ± 1.51

^a Mean of three determination, SD = standard deviation.

using the suggested method. There was no interference from the tablet additives as confirmed by the good %recoveries obtained with the proposed method. The estimated % recovery was 99.31 ± 1.64% for the spectrofluorimetric method. The proposed approach's outcomes were compared with those of a method that had already been published⁵ using *t*- and *F*-tests at a 95% confidence interval (CI). Since the *F*- and *t*-tests findings fell below those given with a 95% CI, no appreciable differences were discovered between the recommended and published procedures (Table 5). The calculated *t*- and *F*-tests values decreased from the tabulated values, confirming that there was no noticeable alteration between the proposed and published methods in terms of precision and accuracy. These results affirmed the ability of the suggested method to analyze VLD precisely and accurately in tablet dosage form.

3.3.2 Content uniformity (CU) testing. Ten Vilaphoria 20 mg tablets were tested for content uniformity testing to confirm that each tablet's composition matched US standards,¹⁸ which require that if the ratio of a drug in tablet dosage form is less than or no more than 25% of the tablet's entire weight, content uniformity (CU) testing of the tablets should be performed.¹⁸ The suggested method was used for the first time to examine the CU of VLD in tablet formulation. Here, if the calculated results of the acceptance value (AV) are less than or equal to the maximum allowed acceptance value (L1), the amount of active ingredient in the tablets under study could be presented as uniform. Following US standards,¹⁸ the equation below could be utilized for this determination:

$$AV; AV = KS + |M - \bar{X}|$$

where *M* is the reference value, *K* is the acceptability constant (equal to 2.4 in the case of 10 tablets), *S* is the standard deviation of the sample, and \bar{X} is the mean of the % recovery of the

Table 3 Evaluation of intra-day and inter-day assay precisions for VLD determination using the proposed method

VLD conc. (ng mL ⁻¹)	Intra-day precision % recovery ± SD ^a	Inter-day precision % recovery ± SD ^a
50	101.34 ± 1.23	99.66 ± 1.71
200	100.51 ± 0.60	99.68 ± 1.73
400	100.04 ± 0.73	98.68 ± 1.27

^a Mean of three determination, SD = standard deviation.



Table 5 Analysis of VLD in tablet dosage form by the proposed spectrofluorometric and reported methods⁵

Dosage form	Proposed method	Reported method	<i>t</i> -Test ^b value	<i>F</i> -Test ^b value
	% Recovery ± SD ^a	% Recovery ± SD ^a		
Vilaphoria® 20 mg per tablets	99.31 ± 1.64	99.59 ± 1.22	0.30	1.82

^a Mean of five measurements, SD = standard deviation. ^b Tabulated value at 95% confidence limit, *F* = 6.388 and *t* = 2.364.

Table 6 Application of the proposed spectrofluorometric method for the content uniformity testing of Vilaphoria tablets

Tablet number	Proposed method
1	102.08
2	98.35
3	98.10
4	99.49
5	97.23
6	98.12
7	101.85
8	101.62
9	98.33
10	97.07
Mean (<i>X</i>)	99.22
Standard deviation (<i>S</i>)	1.93
Acceptance value (<i>AV</i>)	5.12
Maximum allowed acceptance value (<i>L1</i>)	15

individual content. The AV must be smaller than the maximum permitted AV (*L1* = 15). The aforementioned equation changes depending on the value of \bar{X} .

— If $98.5\% \leq \bar{X} \leq 101.5\%$, then $M = \bar{X}$ ($AV = KS$).

— If $\bar{X} < 98.5\%$, then $M = 98.5\%$ ($AV = 98.5 - \bar{X} + KS$).

— If $\bar{X} > 101.5\%$, then $M = 101.5\%$ ($AV = \bar{X} - 101.5 + KS$).

The obtained AV value using the suggested spectrofluorometric method for the analysis of the 10 Vilaphoria 20 mg tablets was lower than the *L1* value, thus affirming the uniformity of the studied tablet formulations (Table 6).

3.3.3 Spiked human plasma application. The relative bioavailability of VLD is 72% when administered with meals. The average highest concentration in the blood (C_{\max}) at steady state was reported to be 156 ng mL⁻¹ following daily VLD 40 mg dosage under fed circumstances.³ Because of the elevated sensitivity of the proposed method, VLD levels in human plasma that were spiked with various VLD concentrations could be determined. The concentration of VLD was calculated utilizing the regression equation $Y = 23.72x + 131.54$, and the produced mean recovery values were (96.36–104.54%) ± (0.30–2.23%) for human plasma. The high calculated mean recoveries shown in Table S1† confirm the convenience of the suggested method for the examination of VLD in human plasma.

3.4 Method greenness evaluation

Green processes are those having no or minimal usage of dangerous chemicals, the elimination of waste, and a decrease in energy consumption.

The method was evaluated in accordance with the National Environmental Method Index (NEMI),²⁵ which aims to avoid the use of permanent, bio-accumulative, and toxic solvents (PBTs). Methanol used in the developed spectrofluorometric method is not a PBT solvent. Also, the Teorell–Stenhagen buffer pH of the proposed method was 3.5, which is not judged as corrosive. Furthermore, the volume of the waste was not greater than 50 mg or 50 mL. According to these results, the proposed fluorometric method conserved solvents while producing low amounts of waste. As a result of these factors, it achieved each of the four quadrants of the greenness profile and is regarded as an eco-friendly green approach, as shown in Table 7.

The eco-scale is a straightforward method that can be used in the practice of quality control laboratories. The subsequent equation (analytical eco-scale score = 100 – total penalty)²⁶ is employed to determine the penalty point value for each of the procedure's defined parameters, such as the quantity of chemicals used, dangers to employees, waste production, and energy consumption. If the score is greater than 75, the analytical method is considered green. The eco-scale score of the developed fluorometric method was found to be 91 (Table 7), which is regarded as environmentally friendly.

Another trend, called GAPI,²⁷ can be used to assess how environmentally friendly an analytical process is, from sample collection to final analysis. The greenness of each stage in the process of analysis is evaluated using a pictogram, which has three color levels: green, yellow, and red, in the GAPI tool. The suggested technique revealed 5 yellow, 7 green, and 3 red areas when evaluated using the GAPI metric. These regions are related to the toxicity and solvent volume used. As a result, the suggested approach had minimal environmental impact, see Fig. 5a.

The most recent metric is AGREE.²⁸ The submitted criteria for the AGREE metric are flexible and can be weighted a different way, drawing design ideas from the 12 significance principles. Twelve input variables are graded from 0 to 1. The total of each principle's assessment results is the final evaluation result. The result is presented as a graph that resembles a clock, with the overall score and a color representation in the middle (Fig. 5b). The evaluation can be carried out with special software, which also generates a report in addition to an auto-generated graph. The AGREE evaluation depends on



Table 7 Evaluation of the greenness of the proposed method using the Eco-score scale tool^b

Parameters		Penalty points	NEMI pictogram
Reagents	None	0	
Instrument	Spectrofluorometer	0	
Solvent	Methanol	6	
Teorell–Stenhagen buffer (0.02 M)		0	
Energy consumption	(Less than 0.1 kW h per sample)	0	
Occupational hazard	(Analytical process hermitization)	0	
Waste	(10 mL)	3	
Total penalty points		9	
Analytical eco-scale total score ^a		91	

^a If the score is greater than 75, it represents excellent green analysis. If the score is greater than 50, it represents acceptable green analysis. If the score is less than 50, it represents inadequate green analysis. ^b NEMI = National Environmental Method Index; PBT = persistent, bio-accumulative, and toxic solvents.

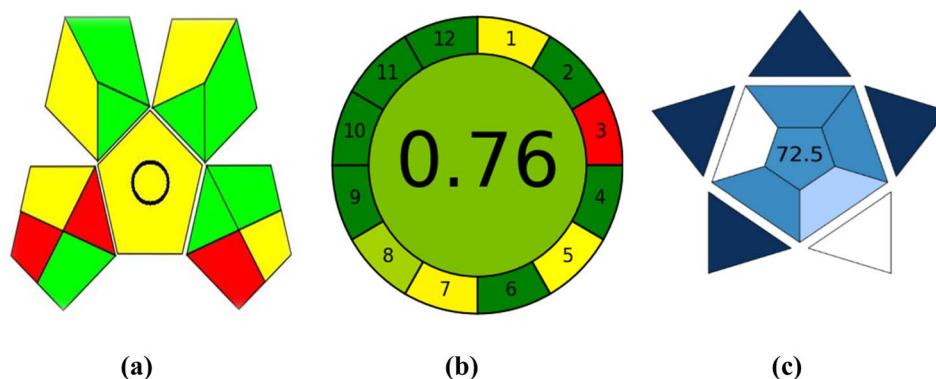


Fig. 5 Evaluation of the greenness using GAPI (a) and AGREE (b) metrics and the blueness using BAGI (c) of the proposed spectrofluorometric method.

numerous variables, including the solvent type, volume used, quantity of chemicals throughout each run, and solvent toxicity to individuals and the surrounding environment. According to the AGREE evaluation metric, the proposed method received an overall score of 0.76.

3.5 Blueness evaluation

The Blue Applicability Grade Index (BAGI)²⁹ complements the established green analytical chemistry metrics. It is a new tool that can be used to assess an analytical method's practical considerations. In order to create a pictogram and a score that illustrate the usefulness and effectiveness of an analytical approach, BAGI takes into account 10 factors (Table S2†). The BAGI metric tool yields two different sets of results: an asteroid-shaped pictogram as a graphic illustration and a score in number at the center of the pictogram. It is advised that the final score should be greater than 60 in order for the analytical procedure to be deemed "practical". The asteroid-shaped pictogram, which is made up of several shades of blue to represent varying degrees of compliance (dark blue for high, blue for moderate, light blue for low, and white for non-compliance), serves as a visual representation of the

assessment result. The proposed approach achieved an overall score of 72.5 in the final rating, as shown in the pictogram's center in Fig. 5c.

4. Conclusion

A highly sensitive, simple, accurate, and green spectrofluorimetric was generated to measure VLD in its pure form tablet formulation and in spiked human plasma. The technique depends on boosting VLD's intrinsic fluorescence by the addition of Teorell–Stenhagen buffer (0.02 M, pH 3.5). The method was validated according to the ICH guideline and was found to be linear in the range of 5–400 ng mL⁻¹. The method was also shown to be accurate, precise, and robust. The method's greenness was assessed following the eco-scale, the National Environmental Method Index (NEMI), GABI, and AGREE metrics. In addition, the practical effectiveness of the analytical procedure was assessed using the BAGI metric. Furthermore, the suggested spectrofluorometric method was used to test the content uniformity of VLD in pharmaceutical tablets. The proposed more-sensitive approach made it possible to analyze VLD in spiked human plasma. Since the



proposed method requires low time and effort, and utilizes environmentally friendly solvents, it can be useful in quality control laboratories.

Conflicts of interest

There are no conflicts to declare.

References

- 1 D. Guay, *Consult. Pharm.*, 2012, **27**, 857–867.
- 2 T. de Paulis, *Invest. Drugs*, 2007, **10**, 193–201.
- 3 M. P. Cruz, *Pharmacol. Ther.*, 2012, **37**, 28.
- 4 S. R. Chaudhari, V. K. Salunkhe, H. S. Deore and A. A. Shirkhedkar, *Sustainable Chem. Pharm.*, 2021, **24**, 100534.
- 5 K. R. Vivekkumar, F. C. Chetan, D. P. Pritam and J. S. Sanjay, *J. Pharm. Res.*, 2014, **13**, 20–22.
- 6 C. Mazi, S. Karaderi and F. Arioiz, *Int. J. Pharm. Res. Allied Sci.*, 2018, **7**, 146–152.
- 7 S. S. Panda and R. K. V. Bera, *Indian Drugs*, 2023, **60**, 61–66.
- 8 R. I. El-Bagary, H. Hashem, M. A. Fouad and S. T. Mahmoud, *Int. J. Pharm. Res. Scholars*, 2014, **3**, 113–119.
- 9 K. R. Vivekkumar, F. C. Chetan and J. S. Sanjay, *J. Pharm. Res.*, 2014, **13**, 71–73.
- 10 A. M. El-Kosasy, M. H. Rahman and S. H. Abdelaal, *Int. J. Pharm., Chem. Biol. Sci.*, 2018, **8**, 319–329.
- 11 S. Ghosh, S. Venkatesh and B. V. V. Ravikumar, *Int. J. PharmTech Res.*, 2015, **7**, 204–211.
- 12 A. Petruczynik, K. Wroblewski, M. Szultka-Mlynska, B. Buszewsk, H. Karakula-Juchnowicz, J. Gajewski, J. Morylowska-Topolska and M. Waksmundzka-Hajnosi, *Acta Pol. Pharm.*, 2017, **74**, 765–775.
- 13 M. M. Annapurna, K. Anusha, A. S. Sharief and V. Sindhuri, *Res. J. Pharm. Technol.*, 2017, **10**, 1077–1080.
- 14 W. Sui, X. Yang, W. Yu, Y. Jin, X. Luan, X. Wang and H. Xu, *J. Pharm. Biomed. Anal.*, 2014, **98**, 228–234.
- 15 B. Ghosh, P. Mandal, S. Chakraborty, R. Bera, C. Saha, S. Poddar, S. Barma, J. Khanam, S. Karmakar and T. Pal, *J. Anal. Pharm. Res.*, 2021, **10**, 1–12.
- 16 M. Iqbal, E. Ezzeldin, K. A. Al-Rashood and A. A. Bajrai, *J. Anal. Toxicol.*, 2015, **39**, 106–112.
- 17 R. El-Bagary, H. Hashem, M. Fouad and S. Tarek, *J. Chromatogr. Sci.*, 2016, **54**, 1365–1372.
- 18 M. Rockville, *The United States Pharmacopoeia 30, the National Formulary 25 US Pharmacopeial Convention*, Electronic version, 2007.
- 19 P. Rickham, *Br. Med. J.*, 1964, **2**, 177.
- 20 M. A. Cardona, C. J. Mallia, U. Baisch and D. C. Magri, *RSC Adv.*, 2016, **6**, 3783–3791.
- 21 S. K. Dwivedi, R. C. Gupta, P. Srivastava, P. Singh, B. Koch, B. Maiti and A. Misra, *Anal. Chem.*, 2018, **90**, 10974–10981.
- 22 H. Niu, J. Liu, H. M. O'Connor, T. Gunnlaugsson, T. D. James and H. Zhang, *Chem. Soc. Rev.*, 2023, **52**, 2322–2357.
- 23 A. P. Osysko and P. L. Muiño, *Biophys. J.*, 2013, **104**, 346a.
- 24 *ICH Harmonised Tripartite Guideline*, 2005.
- 25 L. H. Keith, L. U. Gron and J. L. Young, *Chem. Rev.*, 2007, **107**, 2695–2708.
- 26 K. Aken, L. L. Strekowski and L. Patiny, *Beilstein J. Org. Chem.*, 2006, **2**, 3.
- 27 J. Plotka-Wasyłka, *Talanta*, 2018, **181**, 204–209.
- 28 F. Pena-Pereira, W. Wojnowski and M. Tobiszewski, *Anal. Chem.*, 2020, **92**, 10076–10082.
- 29 N. Manousi, W. Wojnowski, J. Plotka-Wasyłka and V. Samanidou, *Green Chem.*, 2023, **25**, 7598–7604.

