


 Cite this: *RSC Adv.*, 2024, 14, 3359

Functional electrospun nanofibers: fabrication, properties, and applications in wound-healing process

 Qianlan Zheng,^{ID}^a Yuewei Xi^{ID}^{*ab} and Yunxuan Weng^{*ab}

Electrostatic spinning as a technique for producing nanoscale fibers has recently attracted increasing attention due to its simplicity, versatility, and loadability. Nanofibers prepared by electrostatic spinning have been widely studied, especially in biomedical applications, because of their high specific surface area, high porosity, easy size control, and easy surface functionalization. Wound healing is a highly complex and dynamic process that is a crucial step in the body's healing process to recover from tissue injury or other forms of damage. Single-component nanofibers are more or less limited in terms of structural properties and do not fully satisfy various needs of the materials. This review aims to provide an in-depth analysis of the literature on the use of electrostatically spun nanofibers to promote wound healing, to overview the infinite possibilities for researchers to tap into their biomedical applications through functional composite modification of nanofibers for advanced and multifunctional materials, and to propose directions and perspectives for future research.

 Received 17th October 2023
 Accepted 28th December 2023

DOI: 10.1039/d3ra07075a

rsc.li/rsc-advances

1. Introduction

In recent years, there has been a rapid development of research on nanomaterials. Nanofibers are prepared *via* stretching, deposition, self-assembly, phase separation, *etc.*, but there are problems such as complicated processes, low efficiency, and high environmental requirements, which limit the application of nanofibers.¹ Electrostatic spinning technology has emerged to keep pace with the times.²

Electrostatic spinning was initially used to produce nanofibers for textiles³ and filtration applications.⁴ With the continuous development of science and technology, electrostatic spinning has important applications in many fields. For instance, noteworthy applications exist in the realm of food packaging pertaining to the preservation of food through antimicrobial and antioxidant properties,^{5–7} enhancement in the encapsulation efficiency (EE) of actives,⁸ enzyme immobilization,^{9,10} edible food coatings,¹¹ and smart packaging,^{6,12,13} and thermal management composites for phase-change materials also have significant applications. For example, they are used in water filtration treatment,¹⁴ photocatalytic treatment of water pollution,¹⁵ electrocatalysis,¹⁶ and filtration in the field of environmental protection. According to reports, there are already dozens of polymers that have been made into nanofiber

structures through electrospinning technology,^{17,18} such as poly-L-lactic acid (PLA),^{19,20} polyvinyl alcohol (PVA),^{21,22} polycaprolactone (PCL),^{22–25} polyurethane (PU),²⁶ polyvinylidene fluoride (PVDF),²⁷ and polyhydroxybutyric acid ester hydroxy valerate (PHBV),^{28,29} which are also used in the development of electrospinning materials.

Functional electrostatically spun nanofibers are a kind of fibrous material prepared by the electrostatic spinning technology with multiple functions. Due to their simplicity, multifunctionality, and loadability; their high specific surface area and porosity; biocompatibility and biodegradability; easy control over size; and easy surface functionalization (*e.g.*, surface coating and surface modification), they have recently attracted increasing interest, and they have a promising future for a wide range of applications in the field of biomedicine, especially in the field of wound healing.³⁰ In the field of biomedicine as well as tissue engineering,³¹ Hu *et al.*³² prepared composite nanofibers with polydopamine (PDA)-modified hydroxyapatite nanoparticles and PLA by the ES method, which exhibit better hydrophilicity, mechanical properties, and cellular activity, and can be used for tissue engineering scaffolds. Hwang *et al.*³³ coated the surface of PLA fibers with tantalum (Ta), which has high biocompatibility, to improve the bone conduction of PLA. This can be used as a guiding membrane for bone regeneration. Liu *et al.*³⁴ encapsulated cinnamaldehyde (CA) in cyclodextrin (CD) and introduced CA/CD into PLA composite fibers by electrospinning, thereby improving the mechanical properties, hydrophilicity, and antibacterial activity, as well as lowering the cytotoxicity, of the nanofibers. They can be used as wound dressings. Electrospinning has certain advantages in the preparation of fiber

^aCollege of Light Industry Science and Engineering, Beijing Technology and Business University, Beijing 100048, China. E-mail: zheng.qianlan@163.com; xiyuewei@btbu.edu.cn; wyxuan@th.btbu.edu.cn

^bBeijing Key Laboratory of Quality Evaluation Technology for Hygiene and Safety of Plastics Beijing Technology and Business University, Beijing 100048, China



membranes. For example, electrospun fibers with randomly oriented network structures can simulate the three-dimensional structure of the extracellular matrix (ECM), with high porosity and permeability. They are nontoxic, do not cause rejection when implanted in the body, and can be completely metabolized and excreted from the body.^{25,35}

Currently, numerous studies have reported the application of functional electrospun nanofibers in promoting wound healing of various types. For instance, they can be used in acute skin wounds,^{36,37} chronic wound healing,^{38–40} and other aspects. At the same time, there are many studies devoted to the development of new functional electrospun nanofiber materials and exploring different application areas and mechanisms⁴¹ (Fig. 1).

The popularization of electrostatically spun nanotechnology has expanded the use of nanomaterials in biomedical applications. This paper reviews the applications of electrostatically spun nanofibers in the field of promoting wound healing. An introduction to the ES technology is given, followed by a collation of the structure and healing mechanisms of skin and wound healing. In this review, we focus on the preparation and properties of functionalized nanofiber dressings such as antimicrobial, anti-inflammatory, hemostatic, antioxidant, controlled drug release, responsiveness, exudate management, and wound monitoring, as well as their applications in the wound-healing process. Recent advances in research on the healing of chronic wounds, such as burns and diabetes-related wounds, are also briefly described. Functional electrostatically spun nanofiber dressings provide a suitable microenvironment at the wound site and provide a powerful aid to wound healing and consequently better addressing the critical needs in biomedicine.

1.1. ES

The history of ES dates back as far as 1897, when it was first observed by Riley. In 1934, a patent was filed by Formhals. At that time, L. Bocquillon, an American chemist, reported – for the first time – a method of converting polymer materials into fine filaments using electrostatic force. This technology was then called “electrospinning” and was applied to the manufacture of chemical fiber products. In the following decades, ES technology was continuously improved and developed. Taylor’s work with electric jets laid the groundwork for ES, and in 1969, DuPont introduced Kevlar, a polyamide fiber based on ES technology, for the first time.^{42,43}

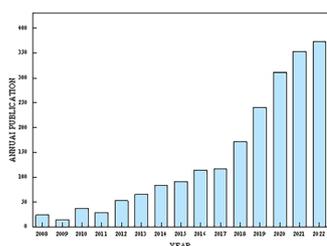


Fig. 1 Annual issues (2008–2022); search the science web for terms “electrospinning” and “wound healing”.

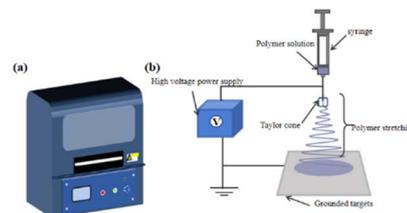


Fig. 2 (a) Schematic of the appearance of the electrospinning machine (b) schematic of the electrospinning process.

1.1.1. Basic composition. The ES machine mainly consists of a high-voltage power supply, a propulsion pump, a syringe, a needle, and a collector (Fig. 2).

1.1.2. Working principle of ES. Electrospinning operates on the principle that exposing a polymer solution to high-voltage electrostatics of thousands to tens of thousands of volts and applying a high voltage to the tip of a syringe containing the spinning solution³⁴ creates repulsion between the charges in the spinning solution and attraction between the grounded collection devices. This creates an electric-field force that overcomes the surface tension of the solution, resulting in the atomization effect of the electrospun liquid, splitting it into tiny jets when the electric-field force is sufficiently large.⁴⁴ A polymer droplet, charged and under the influence of an electric field, accelerates toward the tip of a Taylor cone. The droplet undergoes a transition from a spherical to a conical shape at the needle tip, thereby forming the “Taylor cone”.^{45–47} Subsequently, fiber filaments are produced by extending from the cone tip, undergoing stretching, solvent evaporation, and curing, ultimately yielding nanoscale polymer fibers.⁴⁸

1.1.3. Classification of electrospinning. The production process of ES can be divided into dry spinning, wet spinning,⁴⁹ coaxial ES,^{5,8,22,50,51} and bubble spinning (Fig. 3).

Table 1 shows a comparison of the classification as well as advantages and disadvantages of dry, wet, coaxial, and bubble ES.

Depending on the number of needles, the main types of ES techniques for the preparation of nanofibers are needleless ES³¹ (Fig. 4(a)), single-needle ES⁵² (Fig. 4(b)), coaxial ES^{53,54} (Fig. 4(c)), and multineedle ES^{55,56} (Fig. 4(d)).

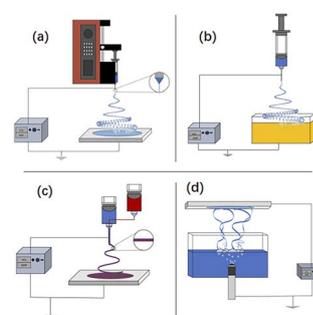


Fig. 3 (a) Dry electrospinning (b) wet electrospinning (c) coaxial electrospinning and (d) bubble electrospinning, illustrated from the literature.²²



Table 1 Classification and comparison of dry, wet, coaxial and bubble ES

Classification	Fiber diameter	Fiber morphology	Advantages	Disadvantages
Dry electrospinning	Tens of microns to over a millimeter	Single root	Fast and efficient manufacturing processes; mild process conditions; does not contain irritating solvents	Poor mechanical strength
Wet electrospinning	Tens of microns to over a millimeter	Single root	Rapid and efficient manufacturing process; high porosity and large pore size	Limited substance encapsulation induced by longer chemical exposure times; poor mechanical strength
Coaxial electrospinning	Micron to submicron/nano	Single fiber with porous, robust, hollow, core-shell structure; aligned/random pattern	Multiple polymer compatibility; multiple system solution spinning; ability to easily fabricate nanofibers; high surface area-to-volume ratio; easy handling; excellent material handling; scalable production	High voltage required; the solvent needs to be removed; selection of nuclear sheath solution
Bubble electrospinning	Nanoscale	Multi-jet twisting	Spinning is done without static electricity, avoiding static pollution and fire explosion caused by high voltage static electricity	The size of the bubbles is difficult to control, and the bubbles affect each other

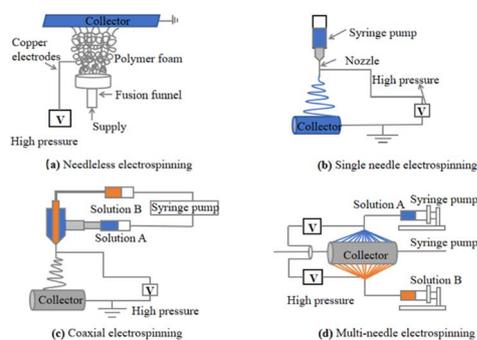


Fig. 4 (a) Needleless electrospinning^{57–59} (b) single-needle electrospinning⁶⁰ (c) coaxial electrospinning^{61–63} and (d) multi-needle electrospinning.⁶⁴

Table 2 lists the classification comparison as well as the advantages and disadvantages of needleless, single-needle, coaxial, and multi-needle ES.

1.1.4. Influence of parameters on fiber morphology. The morphology of nanofibers and the spinning effect are affected by many parameters,⁴⁴ including solution parameters, process parameters, and environmental parameters. The solution parameters include polymer molecular weight, solution concentration, solution viscosity, spinning solution conductivity, liquid surface tension, solvent volatility, and dielectric constant. The process parameters include working voltage, injection flow rate, collection distance and height, needle inner diameter, and so on. Further, the morphology is affected by humidity and temperature, which comprise the environmental parameters (Table 3 lists the parameters of electrospinning and their influence on fiber morphology, and Table 4 lists the

Table 2 Comparison of needleless electrospinning, single-needle electrospinning, coaxial electrospinning, and multi-needle electrospinning

Classification	Advantages	Disadvantages	References
Needleless electrospinning	Strong self-cleaning ability, stable multi-jet ejection, and improved productivity	High production environment requirements; requires a large amount of organic solvents	43 and 50
Single-needle electrospinning	High fidelity, high purity, high productivity	Easy to tangle, easy to form beaded fibers	51 and 52
Coaxial electrospinning	Long-term stable operation, also easy to handle during the lamination process	Low productivity	49 and 53
Multi-needle electrospinning	The release rate can be adjusted by adjusting parameters such as fiber diameter and porosity	The selection range of materials is narrow	54–56
	Containing a variety of active substances; enhancing productivity	Needle interactions are not conducive to stable spraying; film uniformity is poor; nozzles clog easily	57
	Long-term stretching maintains stability, good repeatability and stabilization; sensing performance and capacitance changes are unaffected by stretching rate	The production environment for manufacturing smart wearable electronics requires high standards	49



Table 3 Parameters of electrospinning and their effects on fiber morphology

Classification	Impact parameters	Effect on fiber morphology	References	
Solution parameters	Relative molecular mass	Fiber diameters obtained from polymer solutions with too high a relative molecular mass are generally larger	43	
	Concentration	Fiber diameter increases with concentration, while strength and modulus decrease The higher the concentration, the smaller the chance of forming microspheres, the higher the concentration, the easier it is to clog the nozzle, the lower the concentration, the lower the splash	56 65 and 66	
	Viscosity	As viscosity increases, the number of droplets and stringers decreases and fiber diameter increases	67	
	Conductivity	High conductivity forms thick and continuous nanofibers, low viscosity forms fine and shorter nanofibers	68	
	Conductivity	Fiber diameter decreases with increasing conductivity; high conductivity leads to thinner nanofibers and less chance of bead formation; high conductivity affects coaxial fiber uniformity	69	
	Polarity	The greater the polarity of the solvent, the smaller the diameter of the fiber	5	
	Surface tension	High surface tension leads to instability of the jet, usually low surface tension helps the solution to be electrostatically spun at a low electric field	46	
	Volatile	Higher volatility of solvents is associated with higher porosity and increased surface area	50	
Process parameters	Voltage	Too low a voltage, a high number of fiber beads and larger diameter; too high a voltage cannot form a stable jet, fiber breakage and non-uniformity The higher the voltage, the smaller the diameter of the fiber, but will also increase the corresponding fiber jet speed and fiber scattering problems Within a certain range, the higher the voltage, the smaller the fiber diameter	26 70 65	
	Feed speed	Fiber diameter decreases with decreasing feed rate, and when the flow rate exceeds a critical value and continues to increase, the fiber diameter increases and the number of beaded structures increases Lower transport velocities increase the retention time of the solution between the electrodes, which facilitates the generation and alignment of nanofibers; an increase in flow rate is associated with an increase in fiber diameter	42 5	
	Needle diameter	Needle diameter will directly affect the fiber diameter, large diameter is easy to form beaded fibers	71	
	Work distance	Both excessive proximity and excessive distance can result in the formation of bead-like structures in fibers	47	
	Collector	Smooth fibers can be obtained from a metal collector, while flat collection yields randomly oriented fibers, and rotary collection can generate fibers with a certain degree of orientation, demonstrating anisotropy	72	
	Environmental parameters	Humidity	In high humidity, fibers tend to form beads and pores	58 and 73
		Temperature	The formation of fibers is influenced by both environmental and fluid temperatures, typically resulting in uniform nanofiber diameters at higher temperatures	22

electrospinning parameters and their influence on fiber morphology).

1.2. Introduction of skin and wound healing

1.2.1. Introduction to the skin. The skin – the largest organ in the human body – possesses physiological functions such as regulating body temperature and preventing dehydration. It can also protect the body from adverse effects caused by external pressures, such as mechanical damage, radiation, chemical

substances, bacteria, and viruses.^{74,75} According to a survey conducted by QYResearch,⁷⁶ the global wound care management market was approximately 188.2 billion yuan (RMB) in 2021, and is expected to reach 243.4 billion yuan by 2028, with a compound annual growth rate (CAGR) of 3.7% during the period of 2022–2028. In the United States, it has been reported that over 6.5 million wound-related patients spend \$25 billion on the healthcare system annually, which accounts for 2–3% of the medical budget. The loss of skin tissue due to physical



Table 4 Physiological structure of the skin, from the literature⁷⁹

Skin structure	Connect with the outside world	Formation	Functionality
Epidermis	Direct contact	Cuticle and hair growth layer	Blocking tissue fluid outflow, friction reduction, anti-infection
Dermis	Not in direct contact	Papillary and reticular layers	Provides mechanical strength to the skin

trauma or disease can result in disability or even death, causing immense suffering for millions of people each year. Skin-related diseases pose a global public health issue. According to statistics, chronic nonhealing wounds affect approximately 80 000 medical insurance beneficiaries, and the estimated medical insurance cost for all such wounds is projected to increase from \$220 million to \$8.1 billion. This not only requires more medical treatment resource, but also imposes endless economic burdens on society and families.

Before discussing the process of wound healing, we first understand the structure of normal skin. Under normal human conditions, the body's autoimmunity can heal skin wounds, which itself has dynamic and complex responses. The physiological structure of the skin is usually divided into the epidermis and dermis,^{77,78} as shown in Table 4.

1.2.2. Wound healing. Wound healing is divided into four phases, namely, hemostasis, inflammation, proliferation, and remodeling⁷⁵ (Fig. 5), carried out by immune cells, endothelial cells, fibroblasts,⁸⁰ and keratin-forming cells, which are interspersed and intertwined with one another to form a highly complex healing process. Normally, almost within minutes after trauma, the body initiates a series of immune and inflammatory responses to control and reduce bleeding as it removes necrotic tissue and bacteria. This process may cause symptoms such as pain, swelling, and redness. Platelet aggregation and fibrin clot formation then produce hemostasis, and the wound-healing process rapidly shifts to the inflammatory phase. During this phase, the ECM interacts with fibrin-like proteins in the blood and forms a clot, creating a coagulation barrier that protects vascular access and prevents infection. Inflammatory cells such as neutrophils (a type of leukocyte) and monocytes are pooled at the wound site and the resulting inflammatory response not only removes foreign bodies, bacteria, and damaged

endogenous tissues but also secretes chemokines and growth factors (GFs) to further attract cells. Neutrophil counts usually peak 24–48 h after injury and significantly decline after 3 days. As the leukocytes leave, macrophages continue to remove debris and secrete GFs and proteins that attract immune system cells to the wound to promote tissue repair, thereby directing the healing process into the proliferative phase. During the proliferative phase (within 2–3 days after the creation of the wound), cells multiply and differentiate to reestablish the tissue structure, whereas toxins in the wound need to be dealt with to prevent further damage to the body from the flora surrounding the wound. During this stage, inflammatory cells are activated, as well as stem cells begin to migrate to other adjacent tissues; at the same time, other cells (*e.g.*, fibroblasts, dermatocytes, and so on) get involved, and the traumatic fibroblasts and keratinogen-forming cells produce some light-pink tissues to fill in the defects, repairing and protecting the skin and other tissues that have been damaged, and are called granulation tissues due to the shape of granulation (similar to that of a sarcomere). The granulation tissue mainly consists of infiltrating inflammatory cells, fibroblasts, and some new capillaries. Important factors in this phase include cytokines, GFs, and mesenchymal components that promote cell growth and multiplication. Because multiple cells and tissues are involved at this stage, there may be multiple factors that contribute to slow or poor wound healing. As granulation tissue forms, cells begin to move toward the wound and divide to form new tissue. At the same time, collagen and elastin fibers begin to rearrange and synthesize, and basal cells surrounding the defective tissue continue to proliferate and migrate into the wound to form new epithelial cells. When the entire wound is covered by epithelial cells, it marks the completion of reepithelialization, whereas the completion of reepithelialization after the granulation tissue fills the defect results in the formation of a scar epithelium. The cells also need to grow and differentiate further so that new cells can differentiate at the wound and form new blood vessels of the tissue interconnecting with the surrounding tissue to support the circulation of blood flow and nutrients. During the early stages, cells fill wounds by growing at intervals and later form interconnected structures between cells, which eventually grow and reconfigure the entire tissue through multiple growths and proliferations.^{36,37}

The process of wound healing is an intricate and sophisticated one, with each stage playing a crucial role in achieving favorable healing outcomes. It involves the participation of various cell types and cytokines, including platelets, lymphocytes, and fibroblasts.^{81,82} This paper mainly concerns the following aspects, reviewing researchers' advanced multi-functionalization of materials by functional composite

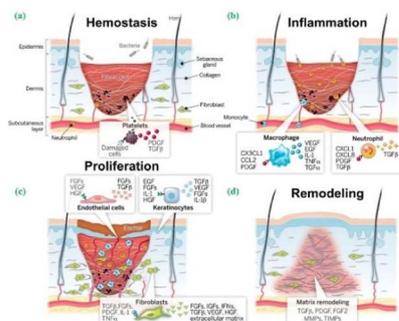


Fig. 5 Four stages of wound healing: (a) hemostasis (b) inflammation (c) proliferation and (d) remodeling. The aforementioned references serve as the foundation for this information. The citation is derived from ref. 81.



modification of nanofibers to promote the process of wound healing. Future research directions and perspectives are proposed for the scientific study of ES in wound healing.

2. Functionality of electrostatically spun nanofiber materials

The original use of electrospun nanofiber wound dressings was to simply physically isolate the outside environment.^{14,15} However, with the advancement of basic research and the increasing demand for wound healing in clinical settings, there has been a growing requirement for enhanced fiber material properties. Consequently, there has been an emergence of electrospun nanofiber dressings with increasingly diverse dressings.

2.1. Antibacterial and antiinflammatory electrostatically spun fiber wound dressing

In recent years, antimicrobial and antiinflammatory fiber dressings with excellent mechanical properties⁸³ and superior biocompatibility prepared using nanotechnology have received increasing attention.^{84,85} Not only do these materials exhibit antibacterial and antiinflammatory properties (effectively alleviating wound infection and pain as well as promoting wound healing) but they can also activate the repair mechanisms of damaged tissues, playing a crucial role in the wound-healing process.^{86,87}

Bacterial infection poses the most common and inevitable challenge to wound healing. Inflammation – the second phase of wound healing^{88,89} – primarily serves to eradicate bacteria and clear debris. Excessive generation of free radicals at the wound site leads to tissue damage, enzyme inactivation, and lipid peroxidation, as well as significantly delays the wound-healing process.⁹⁰ In the case of wound infection, bacteria may induce persistent inflammatory responses at the infection site, further prolonging the healing process of the inflammatory stage.

Severe inflammation often results in poor wound healing and can even lead to complications, including bacteremia, sepsis, and septicemia.

Commonly used antimicrobial additives include antimicrobial metal nanoparticles, such as zinc oxide nanoparticles (ZnONPs),^{91,92} silver nanoparticles (AgNPs),⁹³ and silver oxide (Ag₂O). A study found that the use of PLA nanofibers containing nanosilver significantly promoted the healing of skin wounds in mice,^{94,95} which was attributed to the fact that nanosilver could kill bacteria in the wounds, and at the same time, the high specific surface area of nanofibers could increase the contact area between the cells and the ECM, which could promote the regeneration of cells and healing.⁹⁶ Furthermore, it has been reported that the long-term use of excessive Ag-containing content can considerably increase the prevalence of “Argyria”.⁹⁷

ZnO nanoparticles are also widely used in wound dressings due to their excellent antimicrobial activity, low cytotoxicity, and ability to promote the proliferation of fibroblasts. Zhou *et al.*⁹⁸ investigated a bilayer electrostatically spun fibrous membrane with an outer layer of randomly oriented ZnO/PCL

fibers and an inner layer of neatly aligned nucleosheath structure of PCL@CS. The results showed that the outer layer could confer strong antibacterial activity on the bilayer membrane and inhibit bacterial growth. The inner layer could provide antiinflammatory and effective functions of guiding cellular arrangement. The bilayer CS/PCL electrostatically spun membrane loaded with 1.2 wt% ZnO showed enhanced tensile strength, significant inhibition of *Escherichia coli* and *Staphylococcus aureus*, and exhibited no cytotoxic behavior toward fibroblasts. In addition, the bilayer membrane was able to maintain the high bioavailability of ZnO nanoparticles and synchronized with the aligned structural features of CS fibers, thereby reducing inflammation and stimulating cell migration and reepithelialization *in vivo*.

Sun *et al.*⁹⁹ prepared tunable core–sheath-structured polyacrylonitrile (PAN)/HBPE nanofibers using hyperbranched polyester (HBPE) loaded with ZnONPs by phase separation of the two components during centrifugal spinning and further introduced ZnONPs into the sheath layer. The results showed that the ZnONPs were dispersed by HBPE encapsulation. The PAN/HBPE/ZnONP nanofibers doped with 17 wt% ZnONPs successfully delivered 86.6 wt% of Zn to the surface of the nanofibers, which is a 90% improvement compared with nanofibers without HBPE. PAN/HBPE/ZnONP nanofibers exhibited excellent breathability and good biocompatibility, as well as antibacterial rates as high as 96%. This facile processing strategy cleverly combines material and structural advantages and demonstrates an efficient and stable method to prepare core–sheath-structured nanofibers with functional nanoparticles. The method is also expected to be used for loading small molecules, showing a variety of potential applications such as organophosphorus pesticide detection¹⁰⁰ and food-packaging materials.¹⁰¹

To date, antimicrobial drugs are still the preferred strategy for the clinical treatment of infected wounds, and a large number of drugs have been reported to be encapsulated in electrostatically spun nanofibers for the preparation of antimicrobial wound dressings, including penicillins (*e.g.*, penicillin, amoxicillin, and so on), macrolides (such as erythromycin and clarithromycin), aminoglycosides (*e.g.*, gentamicin, kanamycin,¹⁰² streptomycin eticlopramide, and amikacin), tetracyclines (*e.g.*, doxycycline and tetracycline), sulfonamides (*e.g.*, metribenzylidene and complex sulfonamides), fluoroquinolone antibiotics, and ciprofloxacin hydrochloride.¹⁰³ Alotaibi *et al.*¹⁰⁴ prepared composite electrostatically spun nanofibers loaded with neomycin (Ne) carrageenan, polyacrylonitrile and pullulan polysaccharides^{20,88,105–110} (CG/PAN/PU) by ES for cutaneous wound healing, which was especially effective for burn wounds. In rabbit model experiments, wounds healed in a short period even with drug-free CG/PAN/PU/Ne nanofibers compared with conventional surgical gauze; therefore, drug-loaded CG/PAN/PU/Ne nanofibers have greater healing potential. Peng *et al.*¹¹¹ successfully prepared hybrid poly(lactic glycolic acid) (PLGA)/silkfibroin (SF) membranes loaded with artemisinin (ART) as an antiinflammatory agent onto fiber membranes by the ES technique. The experimental results revealed that the cumulative ART release could reach 69% after three weeks, and the



prepared membrane had a good slow-release effect. Meanwhile, the PLGA/SF/ART fiber membrane shortened the inflammatory period of the wound and facilitated skin regeneration by promoting the expression of inflammatory factors.

Although antibiotics provide excellent clinical control of infections, the problem of bacterial resistance is becoming increasingly serious. Therefore, the search for better antimicrobial strategies has become a topic of considerable interest.¹¹²

To address the challenges of long acute wound-healing time and post-healing scars, Jatoi *et al.*¹¹³ developed a three-phase sustained antimicrobial nanofiber of PVA/carbon nanotubes-AgNPs. Dai and Qin *et al.*¹¹⁴ presented a new method for the preparation of multifunctional nanomaterials with antimicrobial, antioxidant, and thermoregulatory functions. This material was prepared by the coaxial ES technique for core-sheath-structured fibers - PLA/Cur/*n*-octadecane (OD) phase-change thermoregulatory fibers were prepared by selecting OD as the core material. By optimizing the manufacturing conditions of the nanofibers, the researchers successfully fabricated this composite nanomaterial containing linear copolymers, natural antimicrobial substances, and nanoparticles of zinc oxide (ZnO), as well as successfully applied it to functional materials for antimicrobials, which have a wide range of applications in many fields, such as medicine, food, and pharmaceuticals.

Tang *et al.*¹¹ fabricated edible gelatin fibers by incorporating peppermint oil (PO) and chamomile oil (CO). The experimental results showed that all the gelatin nanofibers containing PO, CO, or PO/CO exhibited concentration-dependent antimicrobial properties against *E. coli* and *S. aureus*, as well as some antioxidant properties. In gelatin nanofibers, the combined action of PO and CO exhibited better bioactivity than either PO or CO alone. Finally, the absence of cytotoxicity of PO/CO-composite

gelatin fibers was revealed by the tetramethyl azole salt colorimetric assay. The developed gelatin/PO/CO nanofibers were shown to be biologically safe wound-healing dressings.

Dalgic *et al.*¹¹⁵ prepared a bilayer electrospun scaffold by the dry ES of hydrophilic PUL fiber membranes onto hydrophobic PHBV3D fiber mats that underwent wet electrospinning in the presence of the crosslinking agent glutaraldehyde (GTA), using pullulan polysaccharide and PHBV¹¹⁶ as the raw materials. The results showed that the PHBV layer of the scaffold was developed as a micron-sized ($5.68 \pm 2.12 \mu\text{m}$) regenerated fiber layer (regeneration layer) with a large pore size, which promotes cell survival, proliferation, and migration, as well as O₂ and nutrient delivery. Meanwhile, the PUL layer of the scaffold was produced as a nanosized ($264.1 \pm 38.8 \text{ nm}$) barrier fiber layer (protective membrane) with a smaller pore size, and the nanosized pores are expected to provide the required barrier properties to effectively block bacterial transmission while achieving optimal O₂ and water vapor transport. In *in vivo* enzymatic degradation experiments, the PUL/PHBV scaffolds lost approximately 20% of their body weight after incubation in a lysozyme solution at 37 °C for 14 days,²⁹ and the degradation of the PUL/PHBV scaffolds is more suitable for wound-healing-promoting applications; therefore, this new naturally sourced PUL/PHBV-type bilayered scaffolds are a promising material for modified wound repair.

In addition, Wang *et al.*¹¹⁷ prepared poly(γ -benzyl-L-glutamic acid)/poly(lactic acid) (PBLG/PLA) nanofibrous membranes as a three-dimensional culture matrix for melanoma cells *in vitro* and found that compared with PLA nanofibrous membranes, PBLG/PLA nanofibrous membranes could better support cell survival and proliferation, showing that PBLG/PLA nanofibrous membranes promote a tumor-like structure. Bi *et al.*²¹ prepared PLA and PLA/PVA/sodium alginate (SA) nanofibrous

Table 5 Brief summary of antimicrobial and antiinflammatory electrospun-fiber-based wound dressings

Polymer matrix	Active ingredient	Results	References
PCL/CS	ZnO	The double membrane has strong antimicrobial activity, the inner layer is anti-inflammatory, reduces inflammation, stimulates cell migration and re-epithelialization in the body	97
HBPE	ZnO	PAN/HBPE/ZnONP nanofibers exhibit excellent air permeability and good biocompatibility, with a high antibacterial rate of 96%	204
CG/PAN/PU	Ne	It has a significant effect on burn wound healing. Drug-loaded CG/PAN/PU/NE nanofibers heal in a short time	103
PLGA/SF	ART	PLGA/SF/ART fiber membrane shortens the inflammatory phase of wounds by promoting the expression of inflammatory factors, which is conducive to skin regeneration	110
PLA	Cur	The composite film exhibits excellent bactericidal and antioxidant properties	55
Gelatin	PO and CO	Gelatin nanofibers containing PO, CO or PO/CO all exhibited antimicrobial properties with concentration and had certain antioxidant properties. PO has good antibacterial activity, while CO has good antioxidant activity	11
Pullulan and PHBV		Fiber promotes cell survival, proliferation, and migration, while promoting the transmission of O ₂ and nutrients. Nanoscale pores provide barrier properties that effectively stop the spread of bacteria while enabling optimal O ₂ oxygen and water vapor transport	28
PLA/PVA	SA	PLA/PVA/SA nanofiber membranes promote cell adhesion and proliferation, and can reduce inflammatory responses during early wound healing	21
PLA	PBLG	PBLG/PLA nanofiber membranes can better support the survival and proliferation of cells	116



membranes by ES and carried out a comparative study and found that PLA/PVA/SA nanofibrous membranes showed slightly better adhesion and proliferation of rat fibroblasts than PLA fibrous membranes and reduced inflammatory response during early wound healing⁷¹ (Table 5 summarizes the main features of this subsection).

2.2. Electrostatically spun nanofiber wound dressing with hemostatic, adhesive, and antioxidant properties

Hemostasis occurs at the earliest stage of wound healing. Hemostatic wound dressings include not only hydrogel dry-frozen “hemostatic sponges”^{18,118,119} and peptide-based hemostatic collagen scaffolds¹⁰⁵ but also electrostatically spun nanofiber dressings with hemostatic properties that have been shown to have a positive impact on promoting wound healing. Studies have shown that hemostasis, as an essential function, is usually used in combination with other functions.¹²⁰ For example, combining the adhesive properties of electrostatically spun nanofibers not only seals the wound to provide hemostasis¹²¹ but also seamlessly adheres to the wound site for a long period, avoiding the potential risk of infection caused by contact between the wound and the external environment.^{122,123}

Jiang *et al.*¹²⁴ first used different concentrations of sodium hypochlorite (NaClO) to isolate SF to obtain multiscale silk fibers (MSFs). These MSFs are then implanted into the PLA fiber membrane by electrospinning to improve hydrophilicity. The results showed that the MSFs obtained by splitting SF by NaClO not only improved the hydrophilicity of PLA but also improved the cytocompatibility, promoted cell proliferation, and did not cause inflammation. Yao *et al.*²⁴ introduced vascular endothelial growth factor (VEGF) on the surface of electrostatically spun fibrous scaffolds of polycaprolactone (ϵ -DA) polymers by UV photoinitiation using thiol-ene chemistry. Experimental studies showed that the tensile strength and modulus of elasticity of the 30% PCL-DA fibrous scaffolds were significantly increased compared with unfunctionalized scaffolds. Coupling with the VEGF peptide increased the surface-water wettability of the scaffolds. Experimental studies *in vitro* and in chicken chorio-allantoic membrane (CAM) showed that scaffolds functionalized with the VEGF peptide induced the phosphorylation of VEGF receptor and promoted the survival, proliferation, and adhesion of vascular endothelial cells. It suggests that VEGF-

peptide-functionalized PCL nanofiber scaffolds promote angiogenesis *in vivo*.

Investigations have shown that wound dressings doped with antioxidant-rich ingredients can scavenge free radicals from the wound site and increase the rate of healing.^{22,79,125} Antioxidants are substances that help trap and neutralize free radicals, thereby eliminating the damage caused by free radicals to the body.¹²⁶ Therefore, electrostatically spun nanofibers with antioxidant properties can significantly promote wound healing. Common antioxidants include vitamin C, vitamin E, and glutathione. Polyphenolic^{8,74,110,127} antioxidants are one of the most widely used natural antioxidants with good stability and storage resistance. Common natural polyphenols such as tea polyphenols, resveratrol, ferulic acid,^{128,129} curcumin,^{88,130} and anthocyanins, as well as some flavonoids, have been loaded into electrostatically spun nanofibers to play an antioxidant role in promoting wound healing.^{131,132}

Polyvinylpyrrolidone (PVP) is a nonionic, water-soluble, high-molecular-weight compound formed by the polymerization of *N*-vinylpyrrolidone monomers. It exhibits excellent physiological inertness; does not participate in human metabolism; and demonstrates remarkable biocompatibility and not irritating to the skin, mucous membranes, or eyes.¹³³ Pusporini *et al.*¹³⁴ demonstrated that blending PVP with green tea extract (GTE) and subsequent spinning can produce PVP/GTE nanofibers with antioxidant activity. Polyethylene oxide (PEO), a nontoxic and biologically safe substance, has been widely utilized in the development of composite functional materials and nanocomposites. Locicento *et al.*¹³⁵ utilized the electrospinning technique to produce nanofiber membranes composed of PLA and PEO, with the addition of grape seed extract (GSE). These membranes exhibited antioxidant properties and demonstrated effective control over the release of GSE. Furthermore, the PLA/PEO nanofiber membranes loaded with GSE displayed excellent hydrophilicity and demonstrated good biocompatibility with cells (Table 6 summarizes the main features of this subsection).

2.3. Electrostatically spun nanofiber wound dressing for drug delivery and controlled release

The porous structure of electrospun nanofibers makes them naturally suitable for loading various substances and slowly

Table 6 Brief summary of wound dressings for hemostasis, adhesion, and antioxidant electrospun fibers

Polymer matrix	Active ingredient	Results	References
PLA	SF	Increased hydrophilicity for PLA, improved cytocompatibility, promoted cell adhesion and proliferation, and did not cause inflammation	123
PCL	VEGF	PCL scaffolds functionalized with VEGF peptides were able to induce phosphorylation of VEGF receptors and promote angiogenesis <i>in vivo</i>	24
PVP	GTE	Antioxidant active PVP/GTE nanofibers promote wound healing	133
PLA and PEO	GSE	GSE with antioxidant properties controlled release, and GSE-loaded PLA/PEO nanofiber membranes have good hydrophilicity and cytocompatibility	134



releasing them at specific locations. Electrospun nanofibers are widely applied due to their high efficiency in sustained drug release¹³⁶ and inherent antibacterial properties.¹³⁷ However, as mentioned earlier, most of the drugs related to antibacterial and antiinflammatory delivery are antibiotics. Wound healing also requires cellular interactions between various cell types, including keratinocytes, fibroblasts, endothelial cells, neutrophils, and macrophages, which are regulated by the endogenous release of cytokines and chemokines at the site of the wound. Therefore, the local delivery of exogenous cells or cytokines has shown significant advantages in promoting wound healing.^{36,37} In general, biomedical materials should have the following basic requirements when used in clinical medicine: they should be nontoxic, noncarcinogenic, and nonteratogenic, and should show no sudden reaction to human cells and tissue cells; good compatibility with human tissues and show no poisoning, hemolysis, coagulation, fever, and allergic reactions; stable chemical properties and resistance to the action of antibodies, blood, and enzymes; possess physical and mechanical characteristics that are compatible with natural tissues; and specific functions for different purposes. For example, they can have applications in biomedical nanomaterials such as drug release materials and drug carriers.^{138,139} It is crucial to ensure the safety of the materials to the environment and the human body, and simultaneously improve the encapsulation efficiency (EE) of the contents or loaded components as well as protecting their activity and effectiveness.

Fig. 6 is a simple schematic of a device with controlled drug release for ES.¹⁴⁰ The device mainly consists of a high-voltage power supply, a nozzle, a drug liquid supply system, and a collector. In operation, the drug solution is fed into the nozzle through the drug supply system, whereas the high-voltage power supply provides a high voltage to eject the drug solution through the nozzle, and is subjected to electrostatic action as it is being ejected to form nanoscale fibers. These nanofibers have a large specific surface area and high porosity, which can increase the solubility and release rate of the drug, thereby achieving the controlled release of the drug (Fig. 7).

2.3.1. Classification of ES technology for drug delivery. ES in drug delivery mainly includes sequential ES,¹⁴¹ synchronized ES,¹⁴² coaxial ES,¹⁴³ and side-by-side ES¹⁴⁴ (Fig. 8 and Table 7).

Sequential ES is the ES of different material solutions or melts one at a time through a single spinneret on a case-by-case basis, which can be regarded as multiple replications of the ES

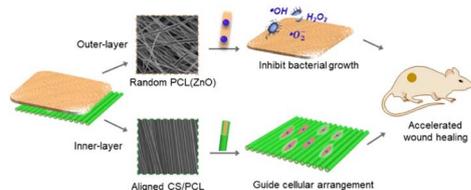


Fig. 6 Graphical abstract from the article titled "Electrospun ZnO-loaded chitosan/PCL bilayer membranes with spatially designed structure for accelerated wound healing" is paraphrased here.

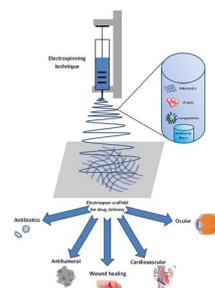


Fig. 7 Schematic of the controlled release of electrospinning drugs, from the literature.¹⁴⁰

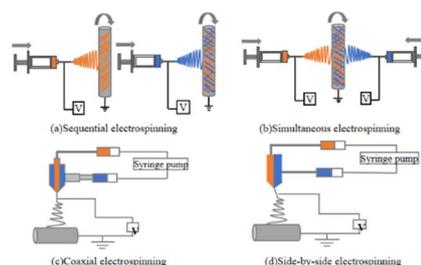


Fig. 8 Schematic of (a) sequential ES (b) synchronized ES (c) coaxial ES and (d) side-by-side ES.

of a single material.¹⁴⁵ Therefore, sequential ES can be easily realized from a variety of materials using the same experimental setup and process parameters as conventional ES.¹⁴¹ A major disadvantage of sequential ES is poor interfacial bonding between the layers of different materials. The controlled release of drugs is used to regulate the release of single drugs, as well as the co-delivery of multiple drugs.

Simultaneous ES,⁷² sometimes referred to as co-ES, refers to the simultaneous ES of nanofibers of different materials stacked on top of each other using two or more spinnerets on a single collector to achieve an integrated fiber structure.¹⁴⁶ When multiple jets are simultaneously ejected from different spinnerets, the interaction of external electric fields and Coulomb repulsion between the jets usually results in unstable and irregular paths of the jets, which makes it difficult to achieve the desired fiber structure.¹⁴¹ Synchronized ES is suitable for more selective material systems. Thus, these well-mixed fiber structures produce more biomimetic scaffolds by mimicking multiple features of the natural ECM and integrating multiple drug-carrying fibers for co-delivery.

Coaxial ES technology is the most widely studied. Coaxial ES employs a coaxial nozzle, consisting of two or more concentrically nested needles supplied by solutions or melts of different materials,¹⁴⁷ to synthesize multiple fluids into one-dimensional nanostructures with different morphologies. It is capable of forcing materials without filament-forming properties into one-dimensional structures or producing high-quality nanofibers from poorly spinnable polymers. The main disadvantage of coaxial ES is the limited choice of material pairs that can be used to prepare high-quality core-shell fibers. Fine-tuning of



Table 7 Comparison of sequential electrostatic spinning, synchronized electrostatic spinning, coaxial electrostatic spinning and side-by-side electrostatic spinning

Categorization	Sequential electrostatic spinning	Synchronized electrostatic spinning	Coaxial electrostatic spinning	Side-by-side electrostatic spinning
Schematic diagram of fiber morphology				
Advantages	Builds multi-layer fiber structures; adapts to a variety of materials; no equipment modifications required	Construction of hybrid fiber structures; independent of compatibility with different materials	Construction of fibers with different morphologies; making fibrous structures out of non-electrospinnable materials	Construction of Janus fibers; making fiber structures out of non-electrospinnable materials
Disadvantages	Poor contact surface bonding	The collector needs to avoid interactions	Small range of material options	Janus structures are of low quality
Applications	Multifunctional scaffolds; modulation of single drug release profiles	Multiple characterization scaffolds; multidrug delivery	Multiple characterization scaffolds; single drug release; multi-drug co-delivery	Amphiphilic applications
References	115	116	117	118

the drug release profiles through shells that act as buffers,¹⁴⁸ as well as the co-delivery of multiple drugs with independent controlled release behavior, offers greater potential for tissue engineering and drug delivery applications.⁵¹

In side-by-side ES,¹⁴⁴ Janus fibers are fabricated by using a spinneret having two or more parallel-needle arrangements in which each side of the structure can be individually designed on the basis of the chemical composition and function. Side-by-side ES also allows for the treatment of nonspinnable fluids with ES fluids to build Janus-structured nanocomposites. However, ES solutions can easily mix, resulting in the inability to produce well-defined Janus fibers.^{70,149} To date, Janus fibers have been widely used in adjustable single-drug-release profiles or controlled multidrug delivery by doping drug active ingredients into both sides of the Janus structure.¹⁵⁰

Alves *et al.*¹⁵¹ successfully prepared drug-loaded PLA fibers using the electrospinning technique, with acetic acid dexamethasone (DEX) or betamethasone 17-valerate (BET) as the corticosteroids. PLA/BET and PLA/DEX electrospun films exhibited promising drug release properties for sustained release in local applications, thus providing a potential drug delivery system.

Glycyrrhizoidic acid (GA),¹³⁸ a natural product of the genus *Glycyrrhizae*, is a novel mitochondrion that targets ligands and improves mitochondrial permeability and enhances mitochondrial drug uptake. GA-functionalized nanocarriers have also shown low toxicity, suggesting that it could be a useful tool for drug delivery. Cacciotti *et al.*¹⁵² prepared 18-beta-glycyrrhetic acid (GA)-nanoparticle-loaded chitosan and poly(lactic acid-glycolic acid copolymer)-based nanoparticles (GA-NPs) as well as poly(lactic acid) fibers (GA-FBs) to compare the release behaviors of free and encapsulated GA in two different biopolymer delivery systems to ensure the controlled release of GA *in vitro*. MTT assays demonstrated that no cytotoxic effects on regular human gingival fibroblasts (HGFs) were observed.

On the other hand, GA-NPs and GA-FBs increased the toxic effects on human oral squamous cell carcinoma (PE/CA-PJ15) cells *in vitro*. The specific effect of GA on PE/CA-PJ15 cells was mainly due to the different sensitivity of cancer cells to reactive oxygen species (ROS) overproduction; GA-NP and GA-FB preparations increased the toxic effects on oral cancer cells *in vitro*. Therefore, FBs can be considered for the treatment of local manifestations of systemic diseases (*i.e.*, blistering diseases, oral lichen planus, and aphthous ulcers), as there is no need for the drug to penetrate the oral mucosa.

Locilento *et al.*¹⁵³ prepared nanofibrous membranes based on PLA and PEO with GSE by the ES technique with antioxidant properties and well-controlled release of GSE, and PLA/PEO nanofibrous membranes loaded with GSE had good hydrophilicity and good cytocompatibility. Goreninskii *et al.*¹⁵⁴ used ES to form PCL/PVP scaffolds with hexafluoroisopropanol as a solvent, which had extreme hydrophilicity and high cellular activity. The PVP-containing materials exhibited enhanced initial release. Because of the good solubility of proteins, drugs and other bioactive compounds in this solvent, the fabrication of PCL/PVP scaffolds using HFIP expands their application in drug delivery systems.

Yang *et al.*¹⁵⁵ developed a detachable triaxial spinneret to direct three ES fluids for the preparation of a series of nucleosheath nanostructures. The three fluids used for ES consisted of a common solvent (outer fluid), an electrospinnable dilute cellulose acetate (CA)¹⁴³ solution (intermediate fluid), and an electrospinnable ibuprofen-methanolysin solution (inner fluid), which led to the formation of an ibuprofen/methanolysin amorphous solid dispersion coated with a thin layer of CA, and the thickness of the coating could be adjusted by precisely adjusting the CA concentration of the intermediate solution. Experimental results show that single ibuprofen/myristolol protein fibers produce an initial burst release, but this release is eliminated in systems with CA coatings, and this tunable way



of precisely manipulating drug release provides a new approach for the development of advanced functional nanomaterials, which can be controlled by the process nanostructure in triaxial ES property relationship to control their performance. Kyzioł *et al.*¹⁰³ developed a novel method for the fabrication of SA nanofibers loaded with ciprofloxacin hydrochloride (CpHCl) under the presence of PEO and Pluronic F-127 surfactant. The experimental findings indicate that approximately 24% of CpHCl is released from alginate fibers within the first 20 h, exhibiting an overall loading efficiency of 51% for the investigated antibiotic.

2.3.2. Multiple drug delivery. During the healing process of wounds, the heterogeneity of the wound site and the development of resistance after long-term use may render monotherapy delivery systems insufficient for clinical practice.¹⁵⁶ In order to overcome chemical resistance, the combined use of multiple drugs with different modes of action is necessary to achieve a synergistic effect. Additionally, a localized drug delivery system is required to inhibit multidrug resistance and mitigate more severe toxicity.¹⁵⁷ Nanofibers encapsulating drugs present an optimal carrier for the delivery of dual drugs,^{52,131,155} without the occurrence of uncontrolled release. Furthermore, they surpass the limitations of traditional drug-polymer combinations by allowing for the placement of multiple drugs within one or two layers possessing distinct polymer, chemical and physical characteristics, thereby achieving the desired release behavior.⁷²

Dual drug-carrying drug delivery scaffolds¹⁴⁷ and multiaxial ES technology adds new dynamics by fabricating multilayered nano- and microsized fibers.¹⁵⁸ These techniques offer the possibility of forming fibers with features such as co-bonding, reinforced cores, and porous and hollow structures. Unique fabrication processes can be used to tailor the mechanical energy, biological, and release of various factors that may be useful in various controlled drug delivery applications. Utilizing these advantages, various polymers and their combinations have been explored in many drug delivery and controlled release applications.

Ma *et al.*¹⁵⁹ successfully prepared PLA/SF nucleosheath fibers containing diclofenac sodium as a steroidal antiinflammatory drug and ZnONPs by coaxial ES. Their results showed that the

initial release concentration was reduced, and the cumulative drug release rates of the nucleosheath fibers were 48.8% and 72.6% at 100 and 600 min, respectively, which showed significant slow-release antimicrobial effects and could be used for the prevention of wound infections (Table 8 summarizes the main features of this subsection).

2.4. Self-healing and stimulus-responsive electrostatically spun nanofiber wound dressing

2.4.1. Self-healing fiber membrane. A self-healing fibrous membrane (SFM) exhibits a porous structure and self-repair ability in a particular material.¹⁶⁰ Unlike nonporous self-healing materials, SFM – with its more complex repair mechanism and microstructure – can introduce self-healing characteristics into porous fibrous membrane materials.⁸³

Zhu *et al.*¹⁶¹ developed intrinsic self-healing thermochromic fiber membranes (STFMs) with biomimetic-constrained protective structures. The self-healing fibers mainly consist of synthesized polydimethylsiloxane-based polyurea (PDMS-PUa), known as the original STFMs. A “closed protective layer” composed of polyacrylic acid (PAA) and branched polyethyleneimine (bPEI) is self-assembled onto the shell of the original SFM. This protective layer prevents supramolecular interactions between PDMS-PUa and ensures the stability of the inherent morphology of the STFMs. By introducing thermochromic microcapsules into the system, the resulting fiber membrane exhibits the thermochromic functionality. Furthermore, without the use of external adhesives, the surface chemical structure of the fiber membrane can be further adjusted by utilizing hydrogen bonds formed at the interfaces between different functional layers to assemble electronic skin sensors. The artificial electronic skin demonstrates self-healing capabilities without external stimuli, offering new directions for the field of biomimetic and advanced flexible electronic skin.¹⁶²

2.4.2. Stimuli-responsive wound dressing. Stimuli-responsive polymers (SRPs) have attracted a great deal of attention from the scientific community because of their unique ability to change their properties in response to external stimuli.⁴¹ Stimulated nanofibers are materials that can release bioactive substances at the wound site to promote wound healing. This material can be used to achieve a variety of

Table 8 Comprehensive compilation of electrospun nanofiber wound dressings for drug delivery and controlled release

Polymer matrix	Active ingredient	Results	References
PLA	DEX and BET	PLA/BET and PLA/DEX electrospun membranes provide sustained drug release for topical applications	144
CS	GA	GA-NPs and GA-FBs have toxic effects on PE/CA-PJ15 cells	145
CA	Ibuprofen-gliadin solution	Monoibuprofen/gliadin fibers with CA coating completely eliminate the initial burst release, precisely manipulating the drug release	148
PEO	CpHCl	CpHCl releases the studied antibiotic from alginate fibers within the first 20 hours with a final loading efficiency of 51%	102
PLA/SF	ZnO NPs	PLA/SF fibers, with reduced initial release concentration, have a sustained release effect, improved antibacterial properties, and good cytocompatibility, which can be used to prevent wound infection	151



therapeutic effects such as promoting skin regeneration, reducing pain, and inhibiting infection by controlling the rate and manner of release. Currently, many stimulating nanofibers are used for wound therapy. Some of the most common stimuli are light,^{163,164} temperature,^{165,166} and pH.¹⁶⁷ If an inflammatory reaction occurs during the wound-healing process, the pathological manifestations are redness, swelling, heat, pain, and so on, in which heat is an indication of increased temperature; therefore, the body temperature at the wound will be higher than other centers. On the basis of the relatively common normal physiological temperature of 37 °C of the human body, heat-sensitive materials used in electrostatically spun nanofiber wound dressings generally exhibit a low critical dissolution temperature (LCST) near the physiological temperature.¹⁶⁸

As an alternative antimicrobial method, photodynamic therapy (PDT),¹⁶⁹ which has recently attracted considerable attention, utilizes photosensitizers (PSs) to produce highly bactericidal ROS in the presence of light and oxygen.

γ -PGA,¹⁷⁰ also known as poly- γ -glutamic acid, is a peptide molecule synthesized through the amide bond polymerization of the monomers of L-glutamic acid and D-glutamic acid.¹⁷¹ Its main chain consists of numerous peptide bonds and carboxyl groups. γ -PGA is a natural anionic biopolymer that exhibits excellent moisturizing and water-absorbing properties.¹⁷² Due to its outstanding biocompatibility and biodegradability, it has found wide applications in the fields of drug delivery, wound dressings, and tissue engineering. Sun *et al.*¹⁷³ prepared an electrospun nanofiber scaffold composed of γ -PGA and a cationic photosensitizer, namely, 5,10,15,20-tetra(1-methylpyridin-4-yl)porphyrin tetra(*p*-toluenesulfonate), which was chemically crosslinked for stability. This scaffold demonstrates excellent cytocompatibility and antibacterial properties, as it utilizes the PS to generate highly bactericidal ROS under oxygen presence and light irradiation.

Tao *et al.*⁸⁸ conducted a study and observed that the abundance of aromatic rings in mesoporous polydopamine nanoparticles (PDANPs) enables the loading of a substantial amount of drugs through π - π stacking and/or hydrogen-bonding interactions. These loaded drugs can be released upon exposure to near-infrared laser irradiation.

Han *et al.*¹⁷⁴ developed the first self-incinerating polymer (SIP) nanofiber membrane in which self-incineration could be triggered by an external stimulus. Electrostatically spun SIP/PAN fibers provided depolymerization 25 times faster and were more responsive (*i.e.*, incineration) than cast membranes under triggering conditions. The depolymerization of SIP in the SIP/PAN co-blended fibrous membranes resulted in a shift in surface properties from hydrophobic ($\sim 110^\circ$) to hygroscopic ($\sim 0^\circ$). The triggered release of encapsulated functional molecules was demonstrated using coaxial electrostatically spun fiber membranes made of SIP/PAN co-blended sheaths and PVP/dye cores. The coaxial fiber with a SIP/PAN sheath provided a minimal release of the encapsulated material in a non-triggered solution, whereas it immediately released the encapsulated material when the triggering condition was met. The versatility is enhanced compared to non-SIP coaxial fibers, which do not trigger a response due to external stimuli (Table 9 summarizes the main features of this subsection).

2.5. Electrospun nanofiber dressings for exudate management and wound-monitoring functions

2.5.1. Electrostatically spun nanofiber dressing with exudate management function.

Excessive wound exudate in clinical treatment can impede the wound-healing process.¹⁴⁹ The healing of wounds may be influenced by the pH of the wound or the pH of the wound exudate present in the wound. Healthy skin typically has a pH within the acidic range between ~ 4.0 and 5.7 .¹⁵⁶ In damaged wounds, particularly in cases of chronic wounds or infected wounds, alkaline pH can often be observed in the wound or the wound exudate. The pH of the wound exudate can be measured, for instance, through *in vitro* measurements.¹⁶⁷ The transition from acidic to alkaline pH can be caused, for example, by bacterial proliferation or the development of necrosis.¹⁷⁵

When damaged skin is in the inflammatory phase, histamine increases the capillary permeability, leading to excessive exudate production from the wound.¹⁷⁶ Excessive wound exudate tends to lead to severe infection, which, in turn, hinders the wound-healing process.¹⁷⁷ Although the widely used hydrophilic wet dressings have some wound exudate absorption function, when

Table 9 Main summary of self-healing and stimulus-responsive electrospun nanofiber wound dressings

Polymer matrix	Active ingredient	Results	References
γ -PGA	TMPyP	γ -PGA-TMPyP nanofiber mats show good cytocompatibility and antimicrobial properties using photosensitizers to produce highly bactericidal reactive oxygen species (ROS) in the presence of oxygen in the presence of light	131
PEGDA	Cur	Under NIR laser irradiation, the concentration of released Cur increased dramatically, showing a concentration gradient-dependent drug diffusion	78
PAN	SIP	Fiber membranes containing SIPs in the triggering solution achieved a triggered release of encapsulated functional molecules. The SIP-containing fiber membrane in non-triggering solution provides minimal release of the encapsulated material. And when the trigger condition is met, it releases the encapsulated material immediately	132



the wound exudate is excessive or is not removed in time, the wet dressings are prone to become a source of contamination, resulting in the hydration and infection of the wound tissue.¹⁵⁰ For this reason, electrostatically spun nanofiber dressings with the function of absorbing or pumping the wound exudate have emerged. Yang *et al.*¹⁷⁸ added ϵ -polylysine (EPL) as a low-cost and safe natural antimicrobial agent to hyaluronic acid (HA) nanofiber dressings, and the prepared EPL/HA composite nanofiber mats were extremely hydrophilic, absorbing 26.3 times its own mass of wound exudate. However, the hydrophilic-absorbent dressing made it difficult to retain the exudate, especially under pressure, and was prone to rewet the wound.

Researchers electrospun hydrophobic substances onto a hydrophilic fiber network to construct a self-pumping dressing with a biofluid pump that unidirectionally drains excess biofluid from the hydrophobic side to the hydrophilic side, preventing the biofluid from wetting the wound, which is referred to as a “self-pumping dressing”,¹⁵⁷ and has promising potential in wound exudate management.^{149,150} In contrast to hydrophilic dressings, self-pumping dressings prevent rewetting of the wound while managing the wound exudate. Currently, the construction of hydrophilic/hydrophobic asymmetric (Janus) structures⁷² is an important method for the preparation of self-pumping exudate management dressings.¹⁷⁹ The Janus structure confers different properties to each side of the material, such as hydrophilicity on one side and hydrophobicity on the other. This asymmetric wettability allows the transport of liquid from the hydrophobic layer to the hydrophilic layer and prevents exudate from flowing back from the hydrophilic layer to the hydrophobic layer.¹⁸⁰

Shi *et al.*¹⁸¹ who electrostatically spun a hydrophobic PU nanofibrous membrane on the surface of cotton medical gauze found that PU/cotton dressings could exclude excessive wound exudate from the wound by a self-pumping action and showed faster healing than conventional dressings. In addition to the micro-/nanofiber-composite-based Janus self-pumping dressing, researchers also developed a pure nanofiber Janus self-pumping dressing with antimicrobial properties.

Qi *et al.*¹⁸² embedded the macromolecular antimicrobial agent poly(hexamethylene guanidine hydrochloride) (PHGC) into multilayered nanofiber membranes (polyurethane/polyacrylonitrile/sodium polyacrylate PU/PAN/SPA) with a hydrophobic/hydrophilic gradient structure and self-pumping effect by ES to prepare unidirectional water-transporting antimicrobial wound dressings. It was found that when the mass fraction of PHGC was 0.06%, the dressing could achieve nearly 100% antimicrobial activity against *E. coli* and *S. aureus*. Yang *et al.*¹⁸³ fabricated Janus membranes based on hydrophilic PVP and hydrophobic ethylcellulose (EC) polymers by side-by-side ES and prepared antimicrobial wound exudate management dressings by loading ciprofloxacin (CIP) and AgNPs on the PVP side and the EC side, respectively. It was found that when the PVP side was dissolved, the CIP was rapidly released to kill the bacteria, and at the same time, due to the insolubility of EC and AgNPs, the EC/AgNP side could play an antimicrobial role when the Janus fibers were in contact with the wound

exudate. The development of Janus electrostatically spun nanofiber dressings that combine wound exudate management with the antimicrobial function will be an effective way to address wound overhydration and bacterial infection.

2.5.2. Electrostatically spun nanofiber wound dressing for wound monitoring. The healing of wounds is a complex process, and various parameters in the vicinity of the wound are constantly changing.⁴⁰ This necessitates the implementation of additional strategies for wound management and monitoring. Wound monitoring using electrospun nanofibers typically involves three components: sensors, drug delivery, and support structures.^{23,103} When applied to wound monitoring, sensors can detect parameters such as wound temperature, humidity, and pH, and transmit these data to external devices for analysis and processing.^{184,185} When applied to wound treatment, nanofibers can release drugs, promoting wound healing and preventing infection.^{186,187}

Currently, the effectiveness of wound care primarily relies on (semi-)blind assessment methods,⁴⁰ which involve visually observing the wound to judge its healing status. However, this approach is unable to provide an accurate assessment of wound healing, and the frequent removal of dressings can lead to secondary damage and cause pain to patients.¹⁸⁸ Therefore, researchers have increasingly paid attention to the incorporation of sensors or stimuli-responsive materials into dressings, which provide real-time monitoring and diagnostic capabilities with respect to wounds.¹⁸⁵ A variety of physical, chemical, or biological markers have been loaded into nanofiber dressings that can be used to monitor indicators such as bacteria, pH, temperature, ROS, and blood glucose at the wound site.^{189,190} The information provided by the detection metrics can enable clinicians to monitor the wound status in real time and provide patients with optimal wound-care protocols. Singh *et al.*¹⁸⁴ designed a medical dressing (TH-WD) for the simultaneous monitoring and treatment of bacterial infections. TH-WD consists of a blend of PU, the antimicrobial agent ciprofloxacin precursor (Pro-Cip), and hemicyanin reactive dyes (H-Cy), and is prepared *via* the ES technology. When encountering a bacterial infection, TH-WD releases H-Cy and hydrolyzes Pro-Cip to ciprofloxacin, and the wound undergoes a color change from yellow to green to red, enabling rapid wound monitoring and care. Furthermore, Gong *et al.*¹⁹¹ utilized electrospinning technology to fabricate a nanofiber membrane composed of conductive patterns and a load of moxifloxacin hydrochloride (MOX) thermosensitive polymer, known as the Smart Conductive-Polymer Nanofiber Hydrogel Membrane (SC-PNHM). This membrane enables the real-time monitoring of wound temperature: an infection leads to an increase in the wound temperature. Once the temperature rises, the thermal-responsive fibers are triggered to release MOX, effectively eliminating the bacterial infection. Combining the monitoring sensor with electrospun nanofibers, the development of nanofiber dressings with wound-monitoring capabilities has emerged as a new trend in the field of medical materials (Table 10 summarizes the main features of this subsection).



Table 10 Key summary of electrostatically spun nanofiber dressings with exudate management and wound monitoring features

Polymer matrix	Active ingredient	Results	References
HA	EPL	EPL/HA composite nanofiber pads are extremely hydrophilic and can absorb 26.3 times their own mass of wound exudate	136
PU	Cotton medical gauze	Absorbs excess wound exudate from the wound through a self-pumping action and demonstrates faster healing than conventional dressings	139
LA	IBU	Photothermal antimicrobial and synergistic antimicrobial properties of NIR irradiation. Janus nanofiber dressings have asymmetric wettability for targeted water transport to drain excess wound exudate	142
PU/PAN/SPA	PHGC	Unidirectional water delivery properties that drive the spontaneous flow of wound exudate from the inside out. Powerful antimicrobial capabilities	140
PVP and EC	CIP and AgNPs	When the PVP side is dissolved, CIP is rapidly released to kill bacteria, while the EC/AgNPs side can still exert antimicrobial effects when Janus fibers come into contact with wound exudates due to the insolubility of EC and AgNPs	141
PU	Pro-Cip	When bacterial infection is encountered, TH-WD releases H-Cy and hydrolyzes Pro-Cip to ciprofloxacin, and the wound undergoes a color change from yellow to green to red, enabling rapid wound monitoring and care	150
Heat-responsive polymers	MOX	Enables real-time monitoring of wound temperature, and infection causes an increase in wound temperature, which triggers the release of MOX from thermo-responsive fibers to eliminate bacterial infection	151

2.6. Refractory wound dressing

Difficult-to-heal wounds, also known as chronic wounds, refer to the condition where wounds are unable to heal or heal slowly due to various reasons. This is considered to be a significant public health issue that severely affects both health and quality of life.⁸² Acute wounds typically heal within 2 to 4 weeks, whereas chronic wounds fail to fully heal even after subjected to standard care for 4 weeks.¹⁷⁶

Common chronic wounds include burns, lower extremity venous ulcers (VLUs),^{192,193} pressure ulcers (PUs),¹⁹⁴ diabetic foot ulcers (DFUs),¹⁹⁵ and ischemic ulcers (IUs),¹⁹⁶ which continue to present a daunting challenge to clinicians and researchers alike.¹⁹⁷

2.6.1. Burns. Burn injury refers to tissue damage caused by external stimuli such as high temperatures, electric currents, strong radiation, or chemical substances.¹⁹⁸ It is a catastrophic trauma that can result in long-term residual effects and functional impairments even after healing. Consequently, the healing process of burn wounds is challenging and requires the implementation of effective measures to minimize the impact of injury as well as promote healing.¹⁹⁹

The utilization of electrospun nanomaterials to facilitate the healing process of burn wounds is a subject of significant interest. Electrospun nanomaterials can be fabricated into various forms according to specific application requirements, allowing for a tailored selection based on factors such as burn-related wound size and severity.²⁰⁰ Furthermore, these nanomaterials possess diverse properties that can be harnessed to achieve distinct functionalities, such as promoting cell growth, enhancing cell adhesion, or facilitating cell proliferation. Of

utmost importance, these nanomaterials exhibit excellent biocompatibility, ensuring their safe implementation.²⁰¹

Pandey *et al.*²⁰² developed a novel composite electrostatically spun nanofiber antimicrobial dressing (PVP–Ce–CurNF) consisting of PVP, cerium nitrate hexahydrate ($\text{Ce}(\text{NO}_3)_3 \cdot 6\text{H}_2\text{O}$), and curcumin for addressing burn-related wound infection and scar formation. In the experiment, a PVP–Ce–CurNF dressing was directly applied to a full circular excision wound in rats. It showed complete healing and reepithelialization within 20 days without any scarring.

Hadisi *et al.*⁹¹ investigated the preparation of a novel wound dressing made of HA–filiprotein/ZnO nucleosheath structure nanofibers for the treatment of burns. ZnO, the nuclear fluid antimicrobial agent in the core–sheath structure, can be continuously released and also maintain its biological activity. The results of *in vivo* studies showed that loading 3 wt% of ZnO in a wound dressing significantly improved the wound-healing process and significantly reduced the inflammatory response at the wound site.

2.6.2. Diabetes. Diabetes is a chronic disease characterized by elevated blood sugar levels due to insufficient insulin secretion or decreased response to insulin.²⁰³ As the population ages, the excessive intake of various sugars in daily life leads to an increase in the obese population, and the prevalence of diabetes continues to rise each year.²⁰⁴ For instance, approximately one-quarter of diabetes patients suffer from foot ulcers, with 10–20% of such cases resulting in lower-limb amputation. The treatment of diabetes trauma has become one of the major challenges of the global medical system.^{194,197}

In diabetic trauma, macrophage polarization from proinflammatory (M1) to antiinflammatory (M2) types becomes very



Table 11 A major summary of refractory wound dressings

Polymer matrix	Active ingredient	Results	References
PVP	CurNF	Addresses infection and scar formation in burn wounds	193
HA-SF	ZnO	ZnO can continuously release and maintain its bioactivity, 3 wt% ZnO wound dressing significantly improved the wound healing process and reduced the inflammatory response at the wound site for the treatment of burns	90
PLLA	QCS and BP	Using BP to convert NIR radiation into heat and stimulate oxygen release from Hb. QCS is a hemostatic and broad-spectrum antimicrobial material. Photothermal therapy increases bacterial susceptibility to QCS and has great potential for diabetic wound healing	201
miR-92a		Release of anti-miR in diabetic ulcers has efficacy in promoting endothelial cell angiogenesis and overall healing	202
PEO/CS/PVP	Vancomycin/ imipenem/cilastatin	Nanofiber dressings can be used as a suitable drug delivery device for diabetic foot ulcer infections in clinical care, as well as for other chronic wounds	71

difficult due to the specific high-glucose environment that interferes with the physiological process of wound healing.²⁰⁵ Large numbers of M1 macrophages accumulate at the trauma site, releasing proinflammatory cytokines such as tumor necrosis factor (TNF- α)¹¹¹ and producing ROS.²⁰⁶ Diabetes mellitus – because of the specificity of the condition – considerably increases the rate of infection in chronic wounds, leading to a prolonged inflammatory phase and vasculopathy, which ultimately leads to difficult-to-heal wounds and increasing the risk of infection: therefore, it creates a vicious circle. It is very probable to cause a variety of complications, including cardiovascular and cerebrovascular diseases,²⁰⁷ renal diseases,²⁰⁴ and retinopathy,²⁰⁸ as well as affect the function of various tissues and organs throughout the body.

Diabetic skin injury (DSI)²⁰⁹ usually occurs on the legs and feet, referred to as the “diabetic foot.” The wound-healing process in DSI can last for more than 12 weeks; often, this process can be impeded by a harsh hypoxic microenvironment (HME), which consists of a range of endogenous and exogenous factors, such as localized hemorrhage, bacterial infection, and microangiopathy-hypoxia feedback loops.

Zhao *et al.*²¹⁰ used charged quaternized chitosan (QCS) and HA to self-assemble black phosphorus (BP) nanosheets and hemoglobin (Hb) in a layer-by-layer manner onto electrostatically spun poly(L-propylene-alpha-lipoic acid ester) (PLLA) nanofibers. Hb is a natural oxygen carrier with excellent thermoresponsiveness. On the basis of NIR-assisted oxygen delivery combined with biopolymer bioactive properties, BP was utilized to convert NIR radiation into heat and stimulate oxygen release from Hb. QCS is a hemostatic and broad-spectrum antimicrobial material. Photothermal therapy increases the bacterial susceptibility to QCS and has great potential for diabetes-related wound healing.

Berger *et al.*²¹¹ assembled nanofiber coatings containing anti-miR by ES to package and control the release of anti-miR-92a (92ai). Experimental tests showed that the dressing of 92ai enhanced healing and the release of anti-miR from the dressing exhibited efficacy in promoting endothelial cell angiogenesis and overall healing in a relevant model of diabetic

ulcers. Davani *et al.*³⁹ adopted ES to prepare a coating consisting of PEO, chitosan, and vancomycin shells and PVP, gelatin, and imipenem/cilastatin of novel core-shell double-drug delivery nanofibers. The results showed that the nanofiber dressing exhibited significant antimicrobial activity against *S. aureus*, methicillin-resistant *S. aureus* (MRSA), and *Pseudomonas aeruginosa* (Gram-negative bacteria) without cytotoxicity. It can be used as a suitable drug delivery device for DFU infections in clinical practice, as well as for other chronic wounds¹⁶⁵ (Table 11 summarizes the main features of this subsection).

3. Conclusions and prospects

At present, with the demand for a healthy and comfortable life, the research of ES as wound dressing has made great progress in ES technology and material design, but in order to meet the needs of specific biomedical needs, multifunctionally coordinated and facilitated wound dressings with certain advantages, both multifunctional and precisely prepared as mentioned before, such as antimicrobial, antiinflammatory, antioxidant, controlled drug release, impulse response, wound exudate management, wound monitoring, and other multifunctional nanofiber dressings will be the main research direction in the future. Research on burn-related wound healing and diabetes-related wound healing in recent years is also presented. Functional electrostatically spun nanofiber dressings provide a suitable microenvironment for wound healing.

Although electrostatically spun nanofibers show good potential for wound therapy, more practical studies and experimental proof are still needed. Electrostatically spun nanofibrous membranes have structural advantages, but electrostatically spun nanofibers still need to consider a series of problems such as the interference of nozzle and recycling-agent evaporation during spinning, solvent toxicity, and fiber deposition uniformity. Therefore, there are still significant challenges in the selection of suitable polymers and solvents and the optimization of existing spinning technologies to achieve stable, safe, and large-scale greening of nanofiber dressings for preparation and application. The further development of



efficient electrostatically spun multifunctional dressings still needs to be urgently addressed.

In the future, with the continuous development and improvement of nanotechnology, it is believed that functional nanofibers and electrospun nanofibers will be more widely used in the field of biomedicine.

Author contributions

Conceptualization, Y. X.; writing – original draft preparation, Q. Z.; data collection, Q. Z. and Y. X.; writing – review and editing, Y. X. and Y. W.; project administration and funding acquisition, Y. W. All authors have read and agreed to the published version of the manuscript.

Conflicts of interest

There are no conflicts to declare.

References

- C. Xue and L. D. Wilson, *Carbohydr. Polym.*, 2022, **275**, 118751.
- A. M. Al-Dhahebi, J. Ling, S. G. Krishnan, M. Yousefzadeh, N. K. Elumalai, M. S. M. Saheed, S. Ramakrishna and R. Jose, *Appl. Phys. Rev.*, 2022, **9**, 011319.
- D. Hu, *J. Aust. Ceram. Soc.*, 2022, **58**, 1509–1517.
- A. I. Kamil and M. M. Munir, *Powder Technol.*, 2023, **430**, 118978.
- C. Drosou, M. Krokida and C. G. Biliaderis, *Food Hydrocolloids*, 2022, **133**, 107949.
- F. Topuz and T. Uyar, *Food Res. Int.*, 2020, **130**, 108927.
- S. R. Falsafi, H. Rostamabadi, K. Nishinari, R. Amani and S. M. Jafari, *Food Chem.*, 2022, **374**, 131826.
- B. S. Isik, F. Altay and E. Capanoglu, *Food Chem.*, 2018, **265**, 260–273.
- C. Shi, A. Zhou, D. Fang, T. Lu, J. Wang, Y. Song, L. Lyu, W. Wu, C. Huang and W. Li, *Chem. Eng. J.*, 2022, **445**, 136746.
- T. Min, L. Zhou, X. Sun, H. Du, Z. Zhu and Y. Wen, *Food Chem.*, 2022, **391**, 133239.
- Y. Tang, Y. Zhou, X. Lan, D. Huang, T. Luo, J. Ji, Z. Mafang, X. Miao, H. Wang and W. Wang, *J. Agric. Food Chem.*, 2019, **67**, 2227–2234.
- T. S. M. Kumar, K. S. Kumar, N. Rajini, S. Siengchin, N. Ayrilmis and A. V. Rajulu, *Composites, Part B*, 2019, **175**, 107074.
- P. Müller and M. Schmid, *Foods*, 2018, **7**(10), 170.
- A. H. Behroozi, M. Al-Shaeli and V. Vatanpour, *Desalination*, 2023, **558**, 116638.
- H. Lv, Y. Liu, Y. Bai, H. Shi, W. Zhou, Y. Chen, Y. Liu and D.-G. Yu, *Catalysts*, 2023, **13**(4), 758.
- L. Luan, X. Ji, B. Guo, J. Cai, W. Dong, Y. Huang and S. Zhang, *Biotechnol. Adv.*, 2023, **63**, 108098.
- N. Yuan, K. Shao, S. Huang and C. Chen, *Int. J. Biol. Macromol.*, 2023, **240**, 124321.
- W. Zheng, Z. Zhang, Y. Li, L. Wang, F. Fu, H. Diao and X. Liu, *Chem. Eng. J.*, 2022, **447**, 137482.
- I. Armentano, N. Bitinis, E. Fortunati, S. Mattioli, N. Rescignano, R. Verdejo, M. A. Lopez-Manchado and J. M. Kenny, *Prog. Polym. Sci.*, 2013, **38**, 1720–1747.
- W. Zheng, Z. Zhang, Y. Li, L. Wang, F. Fu, H. Diao and X. Liu, *Chem. Eng. J.*, 2022, **447**, 137482.
- H. Bi, T. Feng, B. Li and Y. Han, *Polymers*, 2020, **12**(4), 839.
- B. Kong, R. Liu, J. Guo, L. Lu, Q. Zhou and Y. Zhao, *Bioact. Mater.*, 2023, **19**, 328–347.
- V. Shadman-Manesh, A. Gholipour-Kanani, N. Najmoddin and S. Rabbani, *Sci. Rep.*, 2023, **13**, 919.
- T. Yao, H. Chen, R. Wang, R. Rivero, F. Wang, L. Kessels, S. M. Agten, T. M. Hackeng, T. G. A. M. Wolfs, D. Fan, M. B. Baker and L. Moroni, *Bioact. Mater.*, 2023, **20**, 306–317.
- J. Fernández-Pérez, K. E. Kador, A. P. Lynch and M. Ahearne, *Mater. Sci. Eng., C*, 2020, **108**, 110415.
- M. M. Demir, I. Yilgor, E. Yilgor and B. Erman, *Polymer*, 2002, **43**, 3303–3309.
- Z. Feng, Z. Zhao, Y. Liu, Y. Liu, X. Cao, D.-G. Yu and K. Wang, *Adv. Mater. Technol.*, 2023, **8**, 2300021.
- A. D. Dalgic, E. Koman, A. Karatas, A. Tezcaner and D. Keskin, *Biomater. Adv.*, 2022, **134**, 112554.
- M. Kouhi, M. P. Prabhakaran, M. Shamanian, M. Fathi, M. Morshed and S. Ramakrishna, *Compos. Sci. Technol.*, 2015, **121**, 115–122.
- R. S. Bhattarai, R. D. Bachu, S. H. S. Boddu and S. Bhaduri, *Pharmaceutics*, 2019, **11**(1), 5.
- D. M. Dos Santos, D. S. Correa, E. S. Medeiros, J. E. Oliveira and L. H. C. Mattoso, *ACS Appl. Mater. Interfaces*, 2020, **12**, 45673–45701.
- S. Hu, J. Wu, Z. Cui, J. Si, Q. Wang and X. Peng, *J. Appl. Polym. Sci.*, 2020, **137**, 49077.
- C. Hwang, S. Park, I.-G. Kang, H.-E. Kim and C.-M. Han, *Mater. Sci. Eng., C*, 2020, **115**, 111112.
- Y. Liu, X. Liang, R. Zhang, W. Lan and W. Qin, *Polymers*, 2017, **9**(10), 464.
- G. Patel, K.-S. Na, H. J. Lee and W.-G. Koh, *Curr. Ophthalmol. Rep.*, 2021, **9**, 146–157.
- S. V. Langwald, A. Ehrmann and L. Sabantina, *Membranes*, 2023, **13**(5), 488.
- K. C. Castro, M. G. N. Campos and L. H. I. Mei, *Int. J. Biol. Macromol.*, 2021, **173**, 251–266.
- P. Zhao, W. Chen, Z. Feng, Y. Liu, P. Liu, Y. Xie and D.-G. Yu, *Int. J. Nanomed.*, 2022, **17**, 4137–4162.
- F. Davani, M. Alishahi, M. Sabzi, M. Khorram, A. Arastehfar and K. Zomorodian, *Mater. Sci. Eng., C*, 2021, **123**, 111975.
- E. R. Ghomi, M. Niazi and S. Ramakrishna, *Polym. Adv. Technol.*, 2023, **34**(2), 520–530.
- I. Cacciotti, M. Ciocci, E. Di Giovanni, F. Nanni and S. Melino, *Int. J. Mol. Sci.*, 2018, **19**(8), 2368.
- T. J. Sill and H. A. von Recum, *Biomaterials*, 2008, **29**, 1989–2006.
- N. Bhardwaj and S. C. Kundu, *Biotechnol. Adv.*, 2010, **28**, 325–347.



- 44 B. Pant, M. Park and A. A. Kim, *Pharmaceutics*, 2023, **15**(5), 1347.
- 45 Z. Zhao, J. Li, X. Yuan, X. Li, Y. Zhang and J. Sheng, *J. Appl. Polym. Sci.*, 2005, **97**, 466–474.
- 46 A. K. Haghi and M. Akbari, *Phys. Status Solidi A*, 2007, **204**, 1830–1834.
- 47 C. S. Ki, D. H. Baek, K. D. Gang, K. H. Lee, I. C. Um and Y. H. Park, *Polymer*, 2005, **46**, 5094–5102.
- 48 Q. Wang, Y. Li, X. Zhou, T. Wang, L. Qiu, Y. Gu and J. Chang, *Polymers*, 2019, **11**(9), 1413.
- 49 R. Tonndorf, D. Aibibu and C. Cherif, *Mater. Sci. Eng., C*, 2020, **106**, 110105.
- 50 B. Pant, M. Park and S.-J. Park, *Pharmaceutics*, 2019, **11**(7), 305.
- 51 J. Li, Y. Liu and H. E. Abdelhakim, *Molecules*, 2022, **27**(6), 1803.
- 52 R. Boda, I. Lázár, A. Keczánné-Üveges, J. Bakó, F. Tóth, G. Trencsényi, I. Kálmán-Szabó, M. Béresová, Z. Sajtos, E. D. Tóth, Á. Deák, A. Tóth, D. Horváth, B. Gaál, L. Daróczi, B. Dezső, L. Ducza and C. Hegedűs, *Int. J. Mol. Sci.*, 2023, **24**(8), 7562.
- 53 S. S. Ganesh, R. Anushikaa, V. S. Swetha Victoria, K. Lavanya, A. Shanmugavadivu and N. Selvamurugan, *J. Funct. Biomater.*, 2023, **14**(5), 288.
- 54 N. Angel, S. Li, F. Yan and L. Kong, *Trends Food Sci. Technol.*, 2022, **120**, 308–324.
- 55 Y. Yang, W. Chen, M. Wang, J. Shen, Z. Tang, Y. Qin and D.-G. Yu, *Polymers*, 2023, **15**(10), 2237.
- 56 G. Xu, X. Chen, Z. Zhu, P. Wu, H. Wang, X. Chen, W. Gao and Z. Liu, *Adv. Compos. Hybrid Mater.*, 2020, **3**, 98–113.
- 57 G. Zheng, J. Jiang, X. Wang, W. Li, W. Zhong and S. Guo, *Appl. Phys. A*, 2018, **124**, 473.
- 58 A. G. Thite, R. D. Kale, P. K. Panda and D. M. More, *Cellulose*, 2023, **30**, 4873–4888.
- 59 M. Hwang, M. O. Karenson and Y. A. Elabd, *ACS Appl. Polym. Mater.*, 2019, **1**, 2731–2740.
- 60 M. P. Falaschetti, F. Rondina, E. Maccaferri, L. Mazzocchetti, L. Donati, A. Zucchelli and L. Giorgini, *Compos. Struct.*, 2023, **311**, 116845.
- 61 T. Dai, Z. Qin, S. Wang, L. Wang, J. Yao, G. Zhu, B. Guo, J. Militky, M. Venkataraman and M. Zhang, *Polym. Adv. Technol.*, 2022, **33**, 4062–4071.
- 62 Y. Zhao, C. Tian, Y. Liu, Z. Liu, J. Li, Z. Wang and X. Han, *Biomaterials*, 2023, **295**, 122029.
- 63 F. Topuz and T. Uyar, *Food Res. Int.*, 2020, **130**, 108927.
- 64 N. Zhan, Y. Li, C. Zhang, Y. Song, H. Wang, L. Sun, Q. Yang and X. Hong, *J. Colloid Interface Sci.*, 2010, **345**, 491–495.
- 65 N. S. A. Halim, M. D. H. Wirzal, S. M. Hizam, M. R. Bilad, N. A. H. M. Nordin, N. S. Sambudi, Z. A. Putra and A. R. M. Yusoff, *J. Environ. Chem. Eng.*, 2021, **9**, 104613.
- 66 S. Noamani, S. Niroomand, M. Rastgar and M. Sadrzadeh, *npj Clean Water*, 2019, **2**, 20.
- 67 X. Han, P. Huo, Z. Ding, P. Kumar and B. Liu, *Pharmaceutics*, 2019, **11**(9), 449.
- 68 L. Shahreen and G. G. Chase, *J. Eng. Fibers Fabr.*, 2015, **10**, 155892501501000308.
- 69 A. Koski, K. Yim and S. Shivkumar, *Mater. Lett.*, 2004, **58**, 493–497.
- 70 X. Jiao, J. Xie, H. Du, X. Bian, C. Wang, L. Zhou and Y. Wen, *Food Packag. Shelf Life*, 2023, **37**, 101066.
- 71 S. Zhao, X. Wu, L. Wang and Y. Huang, *J. Appl. Polym. Sci.*, 2004, **91**, 242–246.
- 72 J. Xing, M. Zhang, X. Liu, C. Wang, N. Xu and D. Xing, *Mater. Today Bio*, 2023, 100710.
- 73 A. Refate, Y. Mohamed, M. Mohamed, M. Sobhy, K. Samhy, O. Khaled, K. Eidaroos, H. Batikh, E. El-Kashif, S. El-Khatib and S. Mehanny, *Heliyon*, 2023, **9**, e17051.
- 74 P. Kewlani, L. Singh, B. Singh and I. D. Bhatt, *Sustainable Chem. Pharm.*, 2022, **29**, 100791.
- 75 L. Barnum, M. Samandari, T. A. Schmidt and A. Tamayol, *Expert Opin. Drug Delivery*, 2020, **17**, 1767–1780.
- 76 S. Jacob, A. B. Nair, J. Shah, N. Sreeharsha, S. Gupta and P. Shinu, *Pharmaceutics*, 2021, **13**(3), 357.
- 77 A. Zimmerman, L. Bai and D. D. Ginty, *Science*, 2014, **346**, 950–954.
- 78 B. S. Kim, G. Gao, J. Y. Kim and D.-W. Cho, *Adv. Healthcare Mater.*, 2019, **8**, 1801019.
- 79 Y. Liang, J. He and B. Guo, *ACS Nano*, 2021, **15**, 12687–12722.
- 80 M. B. Buechler, R. N. Pradhan, A. T. Krishnamurthy, C. Cox, A. K. Calviello, A. W. Wang, Y. A. Yang, L. Tam, R. Caothien, M. Roose-Girma, Z. Modrusan, J. R. Arron, R. Bourgon, S. Müller and S. J. Turley, *Nature*, 2021, **593**, 575–579.
- 81 J. Koehler, F. P. Brandl and A. M. Goepferich, *Eur. Polym. J.*, 2018, **100**, 1–11.
- 82 M. Sadeghi-Aghbash, M. Rahimnejad, H. Adeli and F. Feizi, *Curr. Pharm. Biotechnol.*, 2023, **24**, 1079–1093.
- 83 G. Selvakumar and S. Lonchin, *Int. J. Biol. Macromol.*, 2020, **164**, 1592–1599.
- 84 X. Zhang, Y. Wang, Z. Gao, X. Mao, J. Cheng, L. Huang and J. Tang, *J. Appl. Polym. Sci.*, 2024, **141**(1), e54746.
- 85 A. Memic, T. Abudula, H. S. Mohammed, K. J. Navare, T. Colombani and S. A. Bencherif, *ACS Appl. Bio Mater.*, 2019, **2**, 952–969.
- 86 Z. Norouzi and M. Abdouss, *Int. J. Biol. Macromol.*, 2023, **233**, 123518.
- 87 L. Bardoňová, A. Kotzianová, K. Skuhrovcová, O. Židek, H. Vágnerová, J. Kulhánek, T. Hanová, M. Knor, J. Starigazdová, K. M. Kutlákova and V. Velebný, *Int. J. Biol. Macromol.*, 2022, **194**, 726–735.
- 88 B. Tao, C. Lin, Z. Yuan, Y. He, M. Chen, K. Li, J. Hu, Y. Yang, Z. Xia and K. Cai, *Chem. Eng. J.*, 2021, **403**, 126182.
- 89 F. Zhou, X. Jia, Y. Yang, Q. Yang, C. Gao, Y. Zhao, Y. Fan and X. Yuan, *Mater. Sci. Eng., C*, 2016, **68**, 623–631.
- 90 H.-S. Jung, M. H. Kim, J. Y. Shin, S. R. Park, J.-Y. Jung and W. H. Park, *Carbohydr. Polym.*, 2018, **193**, 205–211.
- 91 Z. Hadisi, M. Farokhi, H. R. Bakhsheshi-Rad, M. Jahanshahi, S. Hasanpour, E. Pagan, A. Dolatshahi-Pirouz, Y. S. Zhang, S. C. Kundu and M. Akbari, *Macromol. Biosci.*, 2020, **20**, 1900328.
- 92 C. T. Chasapis, P.-S. A. Ntoupa, C. A. Spiliopoulou and M. E. Stefanidou, *Arch. Toxicol.*, 2020, **94**, 1443–1460.



- 93 Y. Zhu, J. Li, J. Kim, S. Li, Y. Zhao, J. Bahari, P. Eliahoo, G. Li, S. Kawakita, R. Haghniaz, X. Gao, N. Falcone, M. Ermis, H. Kang, H. Liu, H. Kim, T. Tabish, H. Yu, B. Li, M. Akbari, S. Emaminejad and A. Khademhosseini, *Biomaterials*, 2023, **296**, 122075.
- 94 K. Shameli, M. Bin Ahmad, W. M. Z. W. Yunus, N. A. Ibrahim, R. A. Rahman, M. Jokar and M. Darroudi, *Int. J. Nanomed.*, 2010, **5**, 573–579.
- 95 L. Song, Y. Guo, J. Fan, X. Fan, Y. Xie, Z. Xiao, H. Wang, D. Liang and Y. Wang, *Composites, Part A*, 2022, **163**, 107231.
- 96 M. Cui, S. Li, X. Ma, J. Wang, X. Wang, N. E. Stott, J. Chen, J. Zhu and J. Chen, *Int. J. Biol. Macromol.*, 2023, 128088.
- 97 N. Chow, K. Fain, J. Truitt and C. Stetson, *Proc. - Bayl. Univ. Med. Cent.*, 2022, **35**, 382–384.
- 98 F. Zhou, C. Cui, S. Sun, S. Wu, S. Chen, J. Ma and C. M. Li, *Carbohydr. Polym.*, 2022, **282**, 119131.
- 99 M. Sun, T. Hou, J. Zhou, L. Zhou, Z. Zhang, X. Li and B. Yang, *ACS Appl. Nano Mater.*, 2023, **6**, 9707–9717.
- 100 I. Cacciotti, F. Pallotto, V. Scognamiglio, D. Moscone and F. Arduini, *Mater. Sci. Eng., C*, 2020, **111**, 110744.
- 101 I. Cacciotti, E. Fortunati, D. Puglia, J. M. Kenny and F. Nanni, *Carbohydr. Polym.*, 2014, **103**, 22–31.
- 102 J. J. Ahire, D. D. Robertson, A. J. van Reenen and L. M. T. Dicks, *Biomed. Pharmacother.*, 2017, **86**, 143–148.
- 103 A. Kyzioł, J. Michna, I. Moreno, E. Gamez and S. Irusta, *Eur. Polym. J.*, 2017, **96**, 350–360.
- 104 B. S. Alotaibi, M. Shoukat, M. Buabeid, A. K. Khan and G. Murtaza, *J. Drug Delivery Sci. Technol.*, 2022, **74**, 103502.
- 105 G. Selvakumar and S. Lonchin, *Biomater. Adv.*, 2022, **140**, 213078.
- 106 I. Esparza, N. Jimenez-Moreno, F. Bimbela, C. Ancin-Azpilicueta and L. Gandia, *J. Environ. Manage.*, 2020, **265**, 110510.
- 107 Z. Feng, S. Chen, A. Ahmad, L. Chen and W. Bai, *Carbohydr. Polym.*, 2022, **295**, 119836.
- 108 B. Bernier, *Can. J. Microbiol.*, 1958, **4**, 195–204.
- 109 H. Bender, J. Lehmann and K. Wallenfels, *Biochim. Biophys. Acta*, 1959, **36**, 309–316.
- 110 F. Zhou, Z. Gu, Z. Zeng, X. Tang, C. Li, Z. Fang, B. Hu, H. Chen, C. Wang, S. Chen, H. Wu, W. Wu and Y. Liu, *Food Biosci.*, 2022, **49**, 101898.
- 111 Y. Peng, Y. Ma, Y. Bao, Z. Liu, L. Chen, F. Dai and Z. Li, *Int. J. Biol. Macromol.*, 2021, **183**, 68–78.
- 112 Z. Zeng, S. Deng, Y. Liu, C. Li, Z. Fang, B. Hu, H. Chen, C. Wang, S. Chen, W. Wu and Y. Liu, *Food Hydrocolloids*, 2023, **142**, 108781.
- 113 A. W. Jatoi, H. Ogasawara, I. S. Kim and Q.-Q. Ni, *Mater. Lett.*, 2019, **241**, 168–171.
- 114 T. Dai, Z. Qin, S. Wang, L. Wang, J. Yao, G. Zhu, B. Guo, J. Militky, M. Venkataraman and M. Zhang, *Polym. Adv. Technol.*, 2022, **33**, 4062–4071.
- 115 A. D. Dalgic, E. Koman, A. Karatas, A. Tezcaner and D. Keskin, *Biomater. Adv.*, 2022, **134**, 112554.
- 116 P. M. Tyubaeva, I. A. Varyan, E. D. Nikolskaya, M. R. Mollaeva, N. G. Yabbarov, M. B. Sokol, M. V. Chirkina and A. A. Popov, *Nanomaterials*, 2023, **13**(2), 236.
- 117 Y. Wang, J. Qian, T. Liu, W. Xu, N. Zhao and A. Suo, *Mater. Sci. Eng., C*, 2017, **76**, 313–318.
- 118 J. He, G. Ye, H. Ma, S. Jia, J. Ma, J. Lv, D. Jia, Y. Song, F. Liu, P. Li, J. Wang, K. Gyal, K. Gou, M. La and R. Zeng, *Int. J. Biol. Macromol.*, 2023, **240**, 124487.
- 119 H.-J. Shu, C.-X. Wu, K. Yang, T.-W. Liu, C. Li and C.-L. Cao, *J. Mater. Eng.*, 2019, **47**, 124–129.
- 120 W.-L. Luo, X. Qiu, J. Zhang, P.-Y. Hu, X.-F. Liu, J.-J. Liu, M. Yu, S. Ramakrishna and Y.-Z. Long, *Mater. Sci. Eng., C*, 2019, **101**, 380–386.
- 121 R. Chen, Z. Liu, T. Cui, X. Zhang, C.-F. Wang, G.-X. Li, G. Wang and S. Chen, *ACS Appl. Mater. Interfaces*, 2023, **15**, 46322–46332.
- 122 V. Deineka, O. Sulaieva, N. Pernakov, J. Radwan-Pragłowska, L. Janus, V. Korniienko, Y. Husak, A. Yanovska, I. Liubchak, A. Yusupova, M. Piatkowski, A. Zlatska and M. Pogorielov, *Mater. Sci. Eng., C*, 2021, **120**, 111740.
- 123 W. Liang, Q. Lu, F. Yu, J. Zhang, C. Xiao, X. Dou, Y. Zhou, X. Mo, J. Li and M. Lang, *Biomater. Sci.*, 2021, **9**, 7124–7133.
- 124 F. Jiang, D. Yan, J. Lin, H. Kong and Q. Yao, *Mater. Today Chem.*, 2021, **21**, 100494.
- 125 Y. Xi, J. Ge, M. Wang, M. Chen, W. Niu, W. Cheng, Y. Xue, C. Lin and B. Lei, *ACS Nano*, 2020, **14**, 2904–2916.
- 126 I. Solaberrieta, A. Jimenez, I. Cacciotti and M. Carmen Garrigos, *Polymers*, 2020, **12**(6), 1323.
- 127 P. Kewlani, L. Singh, B. Singh and I. D. Bhatt, *Sustainable Chem. Pharm.*, 2022, **29**, 100791.
- 128 F. Wahid, L.-H. Huang, X.-Q. Zhao, W.-C. Li, Y.-Y. Wang, S.-R. Jia and C. Zhong, *Biotechnol. Adv.*, 2021, **53**, 107856.
- 129 Z. Ding, X. Chang, X. Fu, H. Kong, Y. Yu, H. Xu, Y. Shan and S. Ding, *Int. J. Biol. Macromol.*, 2022, **219**, 121–137.
- 130 E. Shekarforoush, A. C. Mendes, V. Baj, S. R. Beeren and I. S. Chronakis, *Molecules*, 2017, **22**(10), 1708.
- 131 E. Subroto, R. Andoyo and R. Indiarito, *Antioxidants*, 2023, **12**, 633.
- 132 J. L. C. de Queiroz, I. Medeiros, A. C. Trajano, G. Piuvezam, A. C. F. Nunes, T. S. Passos and A. H. A. Morais, *Food Chem.*, 2022, **385**, 132593.
- 133 R. K. Deshmukh, P. Kumar, R. Tanwar and K. K. Gaikwad, *Food Bioprocess Technol.*, 2022, **15**, 2839–2853.
- 134 P. Pusporini, D. Edikresnha, I. Sriyanti, T. Suciati, M. M. Munir and K. Khairurrijal, *Mater. Res. Express*, 2018, **5**, 5.
- 135 D. A. Locilento, L. A. Mercante, R. S. Andre, L. H. C. Mattoso, G. L. F. Luna, P. Brassolatti, F. F. Anibal and D. S. Correa, *J. Nanomater.*, 2019, **2019**, 2472964.
- 136 W.-F. Lai, O. S. Reddy, D. Zhang, H. Wu and W.-T. Wong, *Sci. Technol. Adv. Mater.*, 2023, **24**, 2167466.
- 137 O. S. Fenton, K. N. Olafson, P. S. Pillai, M. J. Mitchell and R. Langer, *Adv. Mater.*, 2018, **30**, 1705328.
- 138 C. Zhang, Z. Liu, Y. Zheng, Y. Geng, C. Han, Y. Shi, H. Sun, C. Zhang, Y. Chen, L. Zhang, Q. Guo, L. Yang, X. Zhou and L. Kong, *Small*, 2018, **14**, 1703306.



- 139 N. Wang, C. Liu, W. Yao, H. Zhou, S. Yu, H. Chen and W. Qiao, *Colloids Surf., B*, 2021, **205**, 111866.
- 140 A. Luraghi, F. Peri and L. Moroni, *J. Controlled Release*, 2021, **334**, 463–484.
- 141 Y. Montoya, J. Cardenas, J. Bustamante and R. Valencia, *Biomater. Res.*, 2021, **25**, 38.
- 142 H. Luo, J. Hu, Y. Dou, Y. Yang and J. Hou, *Compos. Commun.*, 2020, **22**, 100516.
- 143 Y. Huang, Y. Jin, B. Wang, H. Tian, Y. Weng and S. Men, *J. Polym. Environ.*, 2022, **30**, 1758–1771.
- 144 M.-L. Wang, D.-G. Yu and S. W. A. Bligh, *Appl. Mater. Today*, 2023, **31**, 101766.
- 145 J. Wang, M. Wu, Y. Zhu, Z. Wang, H. Cao, X. Li, Y. Yin, X. Ren, Y. Tian, Z. Guo and X. Zeng, *Tissue Eng., Part A*, 2022, **28**, 958–967.
- 146 G. Yang, L. Zhou, X. Zhang, D. Pan, M. Wang, S. Luo, F. Su, Y. Ji and C. Liu, *Compos. Commun.*, 2023, **37**, 101450.
- 147 J. Li, Y. Liu and H. E. Abdelhakim, *Molecules*, 2022, **27**(6), 1803.
- 148 M. Duan, J. Sun, S. Yu, Z. Zhi, J. Pang and C. Wu, *Int. J. Biol. Macromol.*, 2023, **233**, 123433.
- 149 Z. Xu, J. Fan, W. Tian, X. Ji, Y. Cui, Q. Nan, F. Sun and J. Zhang, *Adv. Funct. Mater.*, 2024, **34**(3), 2307449.
- 150 X. Zhang, R. Lv, L. Chen, R. Sun, Y. Zhang, R. Sheng, T. Du, Y. Li and Y. Qi, *ACS Appl. Mater. Interfaces*, 2022, **14**, 12984–13000.
- 151 P. E. Alves, B. G. Soares, L. C. Lins, S. Livi and E. P. Santos, *Eur. Polym. J.*, 2019, **117**, 1–9.
- 152 I. Cacciotti, L. Chronopoulou, C. Palocci, A. Amalfitano, M. Cantiani, M. Cordaro, C. Lajolo, C. Calla, A. Boninsegna, D. Lucchetti, P. Gallenzi, A. Sgambato, G. Nocca and A. Arcovito, *Nanotechnology*, 2018, **29**(28), 285101.
- 153 D. A. Locilento, L. A. Mercante, R. S. Andre, L. H. C. Mattoso, G. L. F. Luna, P. Brassolatti, F. D. F. Anibal and D. S. Correa, *J. Nanomater.*, 2019, **2019**, 1–11.
- 154 S. Goreninskii, N. Danilenko, E. Bolbasov, A. Evtina, M. Buldakov, N. Cherdynitseva, M. Saqib, N. Beshchasna, J. Opitz, V. Filimonov and S. Tverdokhlebov, *J. Appl. Polym. Sci.*, 2021, **138**, app50535.
- 155 Y. Yang, W. Li, D.-G. Yu, G. Wang, G. R. Williams and Z. Zhang, *Carbohydr. Polym.*, 2019, **203**, 228–237.
- 156 Y. Zhang, X. Gao, X. Tang, L. Peng, H. Zhang, S. Zhang, Q. Hu and J. Li, *Int. J. Biol. Macromol.*, 2023, **253**, 126693.
- 157 Y. Liu, S. Huang, S. Liang, P. Lin, X. Lai, X. Lan, H. Wang, Y. Tang and B. Gao, *ACS Appl. Mater. Interfaces*, 2023, **15**(45), 52244–52261.
- 158 A. Khalf and S. V. Madihally, *Eur. J. Pharm. Biopharm.*, 2017, **112**, 1–17.
- 159 Y. Ma and J. Li, *Mater. Lett.*, 2022, **325**, 132804.
- 160 M. Zhu, J. Yu, Z. Li and B. Ding, *Angew. Chem., Int. Ed.*, 2022, **61**, e202208949.
- 161 M. Zhu, J. Li, J. Yu, Z. Li and B. Ding, *Angew. Chem., Int. Ed.*, 2022, **61**, e202200226.
- 162 H. Zhang, Y. Shao, B. Gao and J. Li, *Eur. Polym. J.*, 2023, **198**, 112429.
- 163 A. K. Brooks, R. G. Ramsey, N. Zhang and V. K. Yadavalli, *ACS Biomater. Sci. Eng.*, 2023, **9**, 5793–5803.
- 164 J. Zhao, Q. Shu, S. H. Jia and J. Tian, *Chin. J. Burns*, 2022, **38**, 870–873.
- 165 Z. Ge, W. Guo, Y. Tao, H. Sun, X. Meng, L. Cao, S. Zhang, W. Liu, M. L. Akhtar, Y. Li and Y. Ren, *Adv. Mater.*, 2023, **35**(47), 2304005.
- 166 T. Wang, B. Ma, G. Hao, Z. Ding, P. Liu, Y. Zhang and J. Liu, *J. Drug Delivery Sci. Technol.*, 2023, **88**, 104961.
- 167 G. R. Nirmal, C.-C. Liao, Z.-C. Lin, A. Alshetali, E. Hwang, S.-C. Yang and J.-Y. Fang, *Drug Delivery*, 2023, **30**(1), 2245169.
- 168 O. A. Oyebode, S. W. Jere and N. N. Houreld, *J. Diabetes Res.*, 2023, **2023**, 1359537.
- 169 B. Wang, M. Wang, A. Mikhailovsky, S. Wang and G. C. Bazan, *Angew. Chem., Int. Ed.*, 2017, **56**(18), 5031–5034.
- 170 Z. Chen, J. Yao, J. Zhao and S. Wang, *Int. J. Biol. Macromol.*, 2023, **225**, 1235–1245.
- 171 J. Tang, W. Yi, J. Yan, Z. Chen, H. Fan, D. Zaldivar-Silva, L. Aguero and S. Wang, *Int. J. Biol. Macromol.*, 2023, **247**, 125754.
- 172 Y. Gong, P. Wang, R. Cao, J. Wu, H. Ji, M. Wang, C. Hu, P. Huang and X. Wang, *ACS Nano*, 2023, **17**(22), 22355–22370.
- 173 L. Sun, L. Song, X. Zhang, R. Zhou, J. Yin and S. Luan, *Mater. Sci. Eng., C*, 2020, **113**, 110936.
- 174 D. Han, X. Yu, Q. Chai, N. Ayres and A. J. Steckl, *ACS Appl. Mater. Interfaces*, 2017, **9**, 11858–11865.
- 175 D.-L. Lam, Y.-T. Cheng and C.-J. Huang, *ACS Appl. Mater. Interfaces*, 2023, **15**, 53297–53309.
- 176 Y. Gao, Z. Qiu, L. Liu, M. Li, B. Xu, D. Yu, D. Qi and J. Wu, *J. Polym. Sci.*, 2022, **60**, 2191–2212.
- 177 Y. Yang and H. Hu, *Polymers*, 2018, **10**(2), 210.
- 178 Q. Yang, Z. Xie, J. Hu and Y. Liu, *Mater. Sci. Eng., C*, 2021, **128**, 112319.
- 179 S. Yang, K. Yin, D. Chu, J. He and J.-A. Duan, *Appl. Phys. Lett.*, 2018, **113**(20), 203701.
- 180 J. Chen, Z.-X. Low, S. Feng, Z. Zhong, W. Xing and H. Wang, *ACS Appl. Mater. Interfaces*, 2021, **13**, 60763–60788.
- 181 L. Shi, X. Liu, W. Wang, L. Jiang and S. Wang, *Adv. Mater.*, 2019, **31**(5), 1804187.
- 182 L. Qi, K. Ou, Y. Hou, P. Yuan, W. Yu, X. Li, B. Wang, J. He, S. Cui and X. Chen, *Mater. Des.*, 2021, **201**, 109461.
- 183 J. Yang, K. Wang, D.-G. Yu, Y. Yang, S. W. A. Bligh and G. R. Williams, *Mater. Sci. Eng., C*, 2020, **111**, 110805.
- 184 H. Singh, W. Li, M. R. Kazemian, R. Yang, C. Yang, S. Logsetty and S. Liu, *ACS Appl. Bio Mater.*, 2019, **2**, 2028–2036.
- 185 B. Mirani, Z. Hadisi, E. Pagan, S. M. H. Dabiri, A. van Rijt, L. Almutairi, I. Noshadi, D. G. Armstrong and M. Akbari, *Adv. Healthcare Mater.*, 2023, **12**(18), 2203233.
- 186 N. Dehghani, F. Haghirsadat, F. Yazdian, F. Sadeghian-Nodoushan, N. Ghasemi, F. Mazaheri, M. Pourmadadi and S. M. Naghib, *Int. J. Biol. Macromol.*, 2023, **238**, 124078.
- 187 C. Li, X. Luo, L. Li, Y. Cai, X. Kang and P. Li, *Int. J. Biol. Macromol.*, 2022, **209**, 344–355.



- 188 J. Xia, H. Zhang, F. Yu, Y. Pei and X. Luo, *ACS Appl. Mater. Interfaces*, 2020, **12**, 24370–24379.
- 189 M. He, F. Ou, Y. Wu, X. Sun, X. Chen, H. Li, D. Sun and L. Zhang, *Mater. Des.*, 2020, **194**, 108913.
- 190 Q. Zeng, X. Qi, G. Shi, M. Zhang and H. Haick, *ACS Nano*, 2022, **16**, 1708–1733.
- 191 M. Gong, P. Wan, D. Ma, M. Zhong, M. Liao, J. Ye, R. Shi and L. Zhang, *Adv. Funct. Mater.*, 2019, **29**(26), 1902127.
- 192 M. Kletter, J. Griffiths, C. Arundel, J. Dumville and On behalf of the VenUS 6 Collaborators, *Trials*, 2023, **24**, 727.
- 193 O. Krizanova, A. Penesova, A. Hokynkova, A. Pokorna, A. Samadian and P. Babula, *Int. Wound J.*, 2023, 1–9.
- 194 E. Cereda, N. Veronese and R. Caccialanza, *Curr. Opin. Clin. Nutr. Metab. Care*, 2024, **27**(1), 3–8.
- 195 M. J. Carter, J. DaVanzo, R. Hought, M. Nusgart, D. Cartwright and C. E. Fife, *J. Med. Econ.*, 2023, **26**, 894–901.
- 196 C. W. Curry, S. M. Sturgeon, B. J. O'Grady, A. Yates, A. Kjar, H. Paige, L. S. Mowery, K. A. Katdare, R. Patel, K. Mlouk, M. R. Stiefbold, S. Vafaie-Partin, A. Kawabata, R. McKee, S. Moore-Lotridge, A. Hawkes, J. Kusunose, K. N. Gibson-Corley, J. Schmeckpeper, J. G. Schoenecker, C. F. Caskey and E. S. Lippmann, *Biomaterials*, 2023, **303**, 122397.
- 197 C. K. Sen, *Adv. Wound Care*, 2023, **12**, 657–670.
- 198 L. Zhou, F. Liu, J. You, B. Zhou, W. Guo, W. Qu, X. Ren and G. Gao, *Adv. Healthcare Mater.*, 2023, e2303460.
- 199 B. Wang, J. Chen, C. Zhang, Q. Zhang, Z. Zhu, L. Qiu, J. Yan, Z. Li, X. Zhu, Y. Zhang and Y. Jiang, *Int. Immunopharmacol.*, 2023, **125**, 111164.
- 200 M. A. M. Ahmed, M. F. Ali, N. M. Mohamed, S. A. L. Bayoumi, A. M. Zahran and K. I. Elsayh, *J. Ethnopharmacol.*, 2024, **319**, 117174.
- 201 O. Ramirez, F. Pomareda, B. Olivares, Y.-L. Huang, G. Zavala, J. Carrasco-Rojas, S. Alvarez, C. Leiva-Sabadini, V. Hidalgo, P. Romo, M. Sanchez, A. Vargas, J. Martinez, S. Aguayo and C. M. A. P. Schuh, *Phytomedicine*, 2024, **122**, 155108.
- 202 V. K. Pandey, G. Ajmal, S. N. Upadhyay and P. K. Mishra, *Int. J. Pharm.*, 2020, **589**, 119858.
- 203 N. Xu, S. Liu, Y. Zhang, Y. Chen, Y. Zuo, X. Tan, B. Liao, P. Li and J. Feng, *Redox Rep.*, 2023, **28**(1), 2246720.
- 204 X. Chen, C. Chen, X. Tian, L. He, E. Zuo, P. Liu, Y. Xue, J. Yang, C. Chen and X. Lv, *Talanta*, 2024, **266**, 125052.
- 205 Y.-J. Fu, Y.-F. Shi, L.-Y. Wang, Y.-F. Zhao, R.-K. Wang, K. Li, S.-T. Zhang, X.-J. Zha, W. Wang, X. Zhao and W. Yang, *Adv. Sci.*, 2023, **10**, 2206771.
- 206 P. Chandra, N. Sachan, N. Saraswat and N. Vyawahare, *Curr. Mol. Pharmacol.*, 2024, **17**(1), e280323215026.
- 207 M. Oishi, K. Tanaka, K. Ishihara, K. Iino, A. Ito and Y. Yokoyama, *J. Matern.-Fetal Neonat. Med.*, 2023, **36**(1), 2183757.
- 208 S. Vujosevic, E. Chew, L. Labriola, S. Sivaprasad and E. Lamoureux, *Ophthalmol. Sci.*, 2024, **4**, 100378.
- 209 S. Zhu, B. Zhao, M. Li, H. Wang, J. Zhu, Q. Li, H. Gao, Q. Feng and X. Cao, *Bioact. Mater.*, 2023, **26**, 306–320.
- 210 Y. Zhao, C. Tian, Y. Liu, Z. Liu, J. Li, Z. Wang and X. Han, *Biomaterials*, 2023, **295**, 122029.
- 211 A. G. Berger, E. Deiss-Yehiely, C. Vo, M. G. McCoy, S. Almofty, M. W. Feinberg and P. T. Hammond, *Biomaterials*, 2023, **300**, 122188.

