

## RESEARCH ARTICLE

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## Novel octacationic-resorcin[4]arenes featuring quaternary ammonium groups as multivalent biocides†

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Novel resorcin[4]arene-based octacationic quaternary ammonium compounds (**ResQAC<sup>R</sup>**) were obtained by linking tetraalkylammonium groups on the macrocycle wider rim to give multivalent multi-QACs. Their structures were elucidated through 1D and 2D NMR experiments, HR-MS analysis, and crystallographic studies. Additionally, the conformational dynamics of these octacationic-resorcinarenes were investigated using <sup>1</sup>H variable-temperature (VT) NMR experiments. The antimicrobial properties of **ResQAC<sup>R</sup>** derivatives were studied by *in vitro* biological investigations. We identified that the **ResQAC<sup>butyl</sup>** derivative shows an impressive bacteriostatic activity against *S. aureus*. A remarkable multivalent effect was observed for this bacteriostatic activity. Interestingly, cytotoxic studies indicate that **ResQAC<sup>butyl</sup>** showed no adverse impact on cell viability of a human cell line, even at concentrations 30 times greater than the MIC for *S. aureus* and approximately 3 times higher than the MIC for *E. coli*. **ResQAC<sup>undecyl</sup>** exhibits greater bacteriostatic activity against *E. coli* than against *S. aureus*, but it is cytotoxic at lower concentrations (IC<sub>50</sub> of 12.1 μM) than its MIC. **ResQAC<sup>phenyl</sup>** shows scarce bacteriostatic activity against both *S. aureus* and *E. coli*, while no multivalent effect was observed. This is likely attributed to the conformational rigidity of the boat C<sub>2v</sub> conformation of **ResQAC<sup>phenyl</sup>**, which hinders optimal matching between the cationic chains and the negatively charged bacterial surface.

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## Introduction

*Staphylococcus aureus* and *Escherichia coli* represent two of the eight bacteria considered significant for public health.<sup>1</sup> These pathogenic microorganisms are responsible for causing a considerable number of diseases.<sup>1</sup> It is crucial to control the environmental transmission of these organisms to safeguard public health. In recent decades, genetic recombination, widespread antibiotic usage, and other factors have led to bacterial resistance, raising the possibility that common infections will once again become potentially fatal. Very recently, analyses have been conducted<sup>1</sup> to study the correlations between antimicrobial consumption (AMC) and antimicrobial resistance (AMR) for specific combinations of bacteria and antibiotics. In detail, statistically significant decreasing trends in AMC and

AMR were observed in food-producing animals and humans from 2014 to 2021.<sup>1</sup> Nevertheless, these measures necessitate reinforcement to maintain reductions in AMC. This emphasizes the importance of initiatives promoting human and animal health, such as vaccination and improved hygiene practices, alongside the use of disinfectants.

Among disinfectants, quaternary ammonium compounds (Fig. 1, QACs)<sup>2</sup> are a well-known class of cationic biocides, boasting a wide spectrum of antimicrobial activity.<sup>3,4</sup> QACs are commonly present in cleaning products, hand sanitizers, personal care products, various types of wipes, and diverse pesticidal products. They are frequently integrated into polymeric materials (poly-QAC in Fig. 1). Interestingly, QACs serve as antimicrobial treatments for biomedical instruments and high-touch surfaces in public areas.<sup>2–4</sup> In addition, the use of QACs as antimicrobial agents has experienced a notable increase in response to the COVID-19 pandemic.<sup>5</sup>

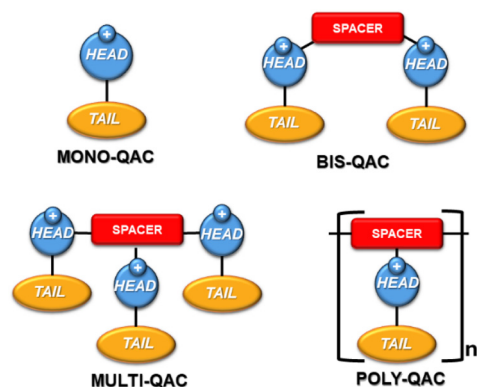
Over recent decades, numerous studies have highlighted the role of macrocycles as useful scaffolds for obtaining antibacterial derivatives (spacer in Fig. 2).<sup>6–11</sup> Interestingly, the synthetic versatility of pillararenes and calixarenes enabled the incorporation of multiple functionalities onto their rims.<sup>6–15</sup>

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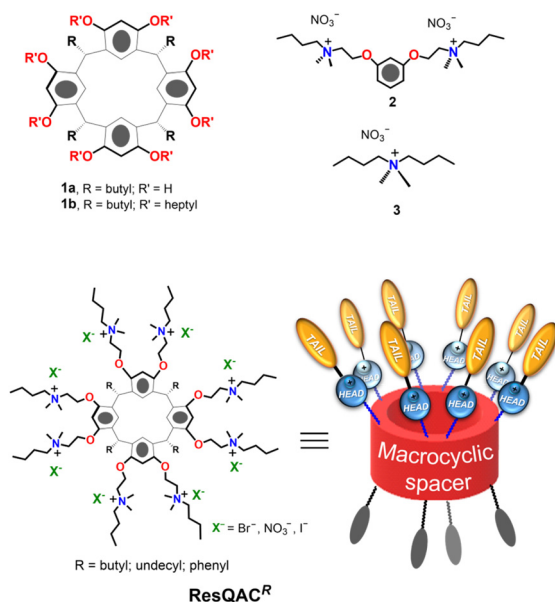
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**Fig. 1** Schematic representation of mono-QAC, bis-QAC, multi-QAC, and poly-QAC.<sup>3</sup> A mono-QAC is constituted by a single tetraalkylammonium group. In the case of bis-QAC, multi-QAC, and poly-QAC, the segment connecting the charged nitrogen atoms ("head") is named "spacer". The alkyl chains extending from the head are called "tails".



**Fig. 2** Resorcin[4]arene-based multi-QACs (ResQAC<sup>R</sup>) investigated in this work.

Thus, in 2016, Cohen and coworkers<sup>12</sup> were able to obtain pillar[*n*]arenes decorated with 10 quaternary ammonium groups (five ammonium groups on each side of the macrocycle) which inhibited the formation of biofilms by two significant Gram-positive pathogens, *Staphylococcus aureus* and *Enterococcus faecalis*.

Consoli and colleagues<sup>13,14</sup> focused on calixarene scaffolds, designing and synthesizing novel water-soluble polycationic calix[4]arenes. Subsequently, the authors demonstrated the ability of these novel calix[4]arene-multi-QACs to work as antibacterial agents against Gram-positive and Gram-negative bacteria, both independently and in combination with appropriate antibiotics. The same research group reported a calixarene

featuring long alkyl chains at the lower rim, which exhibited micellar behavior.<sup>14</sup> These micelles inhibited the formation of biofilms by Gram-negative bacteria.

In the literature, a few rare examples have been reported where macrocycle-based antibacterial agents exhibited increased antibacterial efficacy by exploiting the multivalent effect.<sup>15,16</sup> Multivalency<sup>17–19</sup> refers to the phenomenon where multiple receptor/ligand interactions occur simultaneously, leading to enhanced overall affinity and selectivity of binding. Natural systems extensively utilize multivalency, such as in immunoglobulin function, virus particle adhesion to target cells, and regulation of cell–cell interactions. This concept has acquired increasing attention in the literature,<sup>17–19</sup> where multivalency was exploited in designing macrocyclic ligands<sup>20–22</sup> targeting pathologically relevant enzymes.

Resorcin[4]arene<sup>23,24</sup> (**1a**, Fig. 2) is a macrocycle bearing four resorcinol rings, and is obtained from resorcinol/aldehyde acid-catalysed condensation.<sup>23,24</sup> This macrocycle possesses eight functionalizable OH groups, all positioned on one side of the macrocycle (wider rim, see Fig. 2). Thus, the resorcin[4]arene scaffold can be considered as an ideal candidate to design multivalent quaternary ammonium compounds (multi-QACs, Fig. 1 and 2), where the valence indicates the number of ammonium units (heads in Fig. 1 and 2) linked to the macrocycle. To the best of our knowledge, no examples of polycationic ammonium resorcinarene compounds have been reported in the literature to date.

Inspired by these considerations, this study focused on the synthesis of novel multi-QAC resorcin[4]arenes (denoted as ResQAC<sup>R</sup> in Fig. 2), featuring eight cationic ammonium groups on the wider rim (Fig. 2). In addition, conformational dynamics of these novel octacationic resorcinarenes will be investigated.

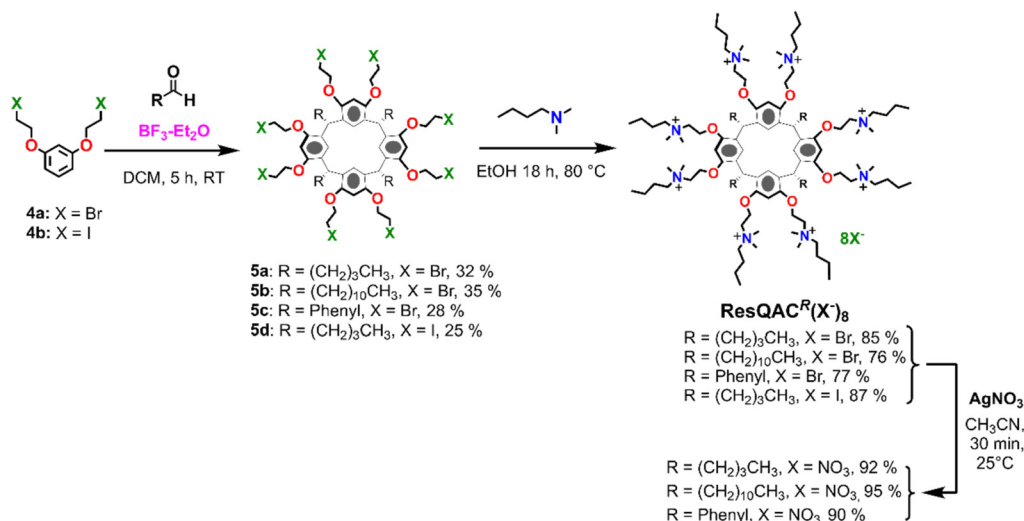
Finally, the antimicrobial effects of ResQAC<sup>R</sup> against *Staphylococcus aureus* and *Escherichia coli* will be evaluated, alongside an assessment of their cytotoxic properties.

## Results and discussion

### Synthesis of ResQACs<sup>R</sup>

Octacationic ResQACs<sup>R</sup> (Fig. 2) were easily synthesized drawing inspiration from a method previously reported by Huang in 2011 for the preparation of cationic pillararenes.<sup>25</sup> Thus, resorcinarenes **5a–d**, bearing bromoalkane groups, were initially synthesized as suitable precursors of ResQAC<sup>R</sup> derivatives (Scheme 1). To obtain resorcinarenes **5a–d**, we tested the direct macrocyclization of monomers **4a–b** with appropriate aldehydes in the presence of BF<sub>3</sub>·Et<sub>2</sub>O in CH<sub>2</sub>Cl<sub>2</sub> as solvent (Scheme 1). Precursors **5a–d** were obtained in reasonable yields (25–35%). It is noteworthy that this represents the first example of direct macrocyclization of resorcinol monomers bearing alkyl bromide chains (**4a–b**) to yield the corresponding resorcin[4]arenes. Successively, **5a–d** were treated with an excess of dimethylbutyl amine (30 equiv.) in ethanol solvent (Scheme 1). The mixture was stirred for 18 hours at 80 °C, and





**Scheme 1** Direct macrocyclization of resorcinols **4a,b** bearing bromoalkane groups with aldehydes to obtain resorcin[4]arenes **5a–d** as precursors for the synthesis of cationic resorcinarenes **ResQAC<sup>R</sup>**.

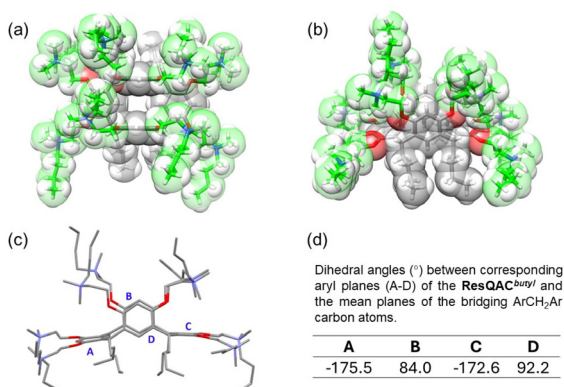
after usual work-up, **ResQAC<sup>R</sup>** derivatives were obtained as bromide or iodide salts. Finally, halide to nitrate exchange was carried out by treating **ResQAC<sup>R</sup>(X<sup>-</sup>)<sub>8</sub>** (X<sup>-</sup> = Br<sup>-</sup> or I<sup>-</sup>) with AgNO<sub>3</sub>, leading to **ResQAC<sup>R</sup>(NO<sub>3</sub><sup>-</sup>)<sub>8</sub>** in 90–95% yields (Scheme 1). The structures of **ResQAC<sup>R</sup>** were confirmed using HR FT ICR mass spectra, 1D–2D NMR spectra, and X-ray studies (see the ESI†).

Single crystals of **ResQAC<sup>butyl</sup>** as an iodide salt, suitable for X-ray diffraction, were obtained by slow evaporation of a H<sub>2</sub>O/MeOH solution (Fig. 3). The X-ray structure of resorcinarene **ResQAC<sup>butyl</sup>** (Fig. 3) was determined using synchrotron radiation with cryogenic techniques. The structure of the macrocycle shows a boat conformation with a pseudo C<sub>2v</sub> symmetry (Fig. 3). The two opposite aryl rings, which define the sides of the boat, make almost orthogonal dihedral angles (Fig. 3d)

with respect to the mean plane of the carbon bridging atoms (Fig. 3d). Angles less than 90° indicate that the aryl rings lean inwards towards the twofold axis of the molecule (Fig. 3c and d) and *vice versa*. In particular, one aryl ring of **ResQAC<sup>butyl</sup>** is slightly inward oriented, and the other is slightly outward oriented (Fig. 3c and d), defining two almost parallel inclined sides of the boat (3.8°). This observation of a more open conformation of **ResQAC<sup>butyl</sup>** than other resorcinarenes in the boat conformation<sup>26,27</sup> is consistent with the presence of the electrostatic repulsion between the positively charged quaternary alkylammonium side chains. Close inspection of the <sup>1</sup>H NMR spectrum of **ResQAC<sup>butyl</sup>** in CD<sub>3</sub>CN (Fig. 4a, 400 MHz, 298 K) clearly indicated that the macrocycle adopts, also in solution, a C<sub>2v</sub>-boat conformation, stable on the NMR time scale at 298 K (400 MHz). In fact, four <sup>1</sup>NCH<sub>3</sub> singlets were detected at 3.25, 3.24, 3.04, and 3.01 ppm (“f and g” in Fig. 4), which correlated in the 2D HSQC spectrum with carbon signals between 52.0 and 52.2 ppm. Furthermore, in accordance with the C<sub>2v</sub> symmetry of **ResQAC<sup>butyl</sup>**, the aromatic region of its <sup>1</sup>H NMR spectrum revealed the presence of four singlets at 7.35, 6.99, 6.69, and 6.15 ppm (“a and b” in Fig. 4), which are attributable to the aromatic protons.

In addition, the <sup>1</sup>H NMR spectrum of **ResQAC<sup>butyl</sup>** at 298 K showed the presence of two sets of OCH<sub>2</sub>CH<sub>2</sub>N<sup>+</sup> diastereotopic resonances between 3.0 and 5.0 ppm (c,c’ and d,d’ in Fig. 4). Subsequently, the conformational dynamics of **ResQAC<sup>butyl</sup>** was studied by <sup>1</sup>H VT NMR.<sup>27,28</sup> At room temperature, this macrocycle is frozen in the C<sub>2v</sub> boat conformation (Fig. 4c and d).

Upon heating, a broadening was observed for the aromatic signals “a,b” and the diastereotopic aliphatic “c,d” resonances (Fig. 4c and d), while their coalescence was observed at 373 K. This coalescence (Fig. 4d) clearly indicates conformational mobility of the C<sub>2v</sub> boat conformation of **ResQAC<sup>butyl</sup>** due to a boat(C<sub>2v</sub>)-crown(C<sub>4v</sub>)-boat(C<sub>2v</sub>) conformational interconversion (Fig. 4c),<sup>21,27</sup> which becomes fast on the NMR time scale



**Fig. 3** (a and b) X-ray structure of **ResQAC<sup>butyl</sup>** (stick representation with van der Waals surfaces). (c) Simple stick representation of **ResQAC<sup>butyl</sup>**. Hydrogen atoms are omitted for clarity. (d) Dihedral angles (°) between the corresponding aryl planes (A–D in Fig. 3c) of the **ResQAC<sup>butyl</sup>** structure.



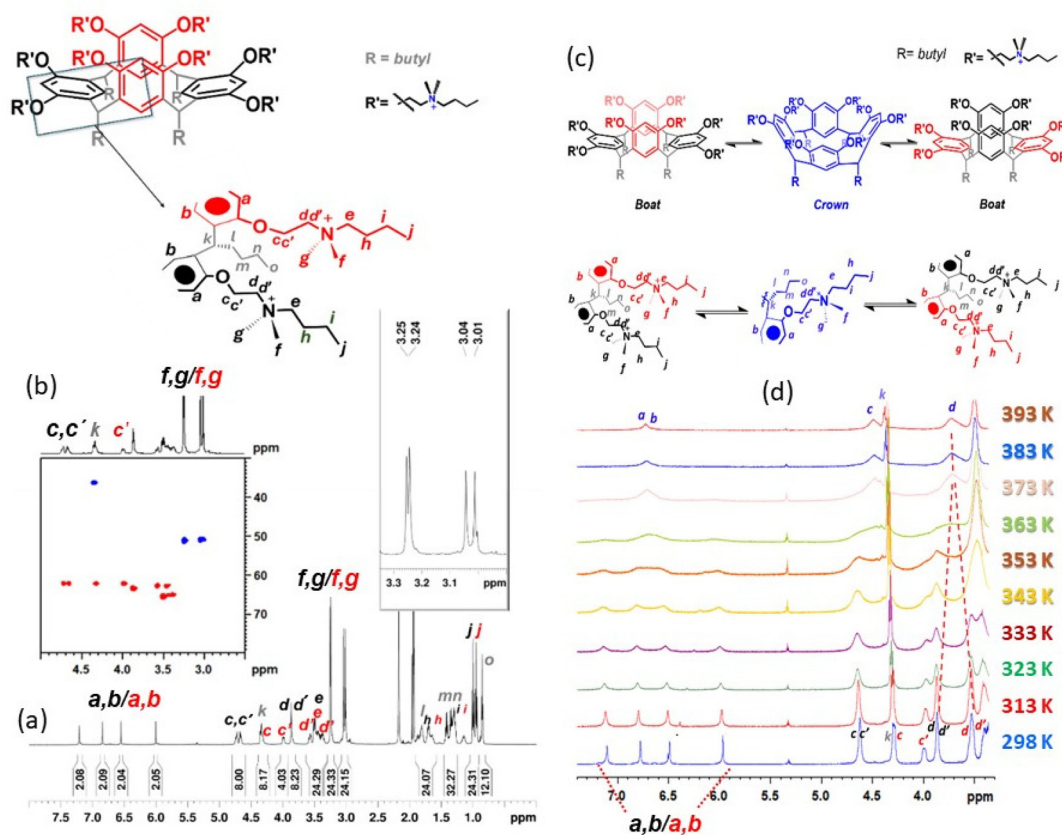


Fig. 4 (a) <sup>1</sup>H NMR spectrum and (b) 2D HSQC spectrum of ResQAC<sup>butyl</sup> (600 MHz, CD<sub>3</sub>CN, 298 K). (c) Chemical drawing of the boat (C<sub>2v</sub>) and crown (C<sub>4v</sub>) conformations of ResQAC<sup>butyl</sup>. (d) <sup>1</sup>H VT NMR experiment (600 MHz) for ResQAC<sup>butyl</sup> in DMSO-*d*<sub>6</sub>.

at 393 K (a  $\Delta G^\ddagger$  value of 17.3 kcal mol<sup>-1</sup> was calculated for this process, see the ESI<sup>†</sup>). In fact, at 393 K, the <sup>1</sup>H NMR spectrum of ResQAC<sup>butyl</sup> becomes simpler (Fig. 4d) and more compatible with an averaged crown-like structure with C<sub>4v</sub> symmetry. In a similar way, the <sup>1</sup>H NMR spectrum at 298 K of ResQAC<sup>undecyl</sup> reveals a stable boat C<sub>2v</sub> conformation (with respect to the NMR time scale), while on heating, a broadening of its NMR signals was observed. Therefore, a boat–crown–boat conformational interconversion (Fig. 4c) occurs within the NMR time scale at higher temperature. A coalescence of NMR signals of ResQAC<sup>undecyl</sup> was observed at 373 K, while at 393 K the <sup>1</sup>H NMR spectrum agreed with an averaged crown C<sub>4v</sub> conformation. From these data, an energy barrier value of 17.2 kcal mol<sup>-1</sup> was calculated for the boat–crown–boat conformational interconversion of ResQAC<sup>undecyl</sup>, a value very similar to that found for ResQAC<sup>butyl</sup>.

With these results in hand, we have investigated the conformational dynamics of ResQAC<sup>phenyl</sup>. In this case, the boat conformation is blocked on the NMR time scale. In fact, on heating, the NMR signals of ResQAC<sup>phenyl</sup> didn't show any evidence of broadening in the temperature range investigated (298–393 K, ESI<sup>†</sup>).

These results clearly indicate that the presence of phenyl groups at the methine bridges of ResQAC<sup>phenyl</sup> increases the

energy barrier to the boat–crown–boat conformational process with respect to the linear aliphatic chains (undecyl and butyl).

#### Antimicrobial activity of novel ResQACs<sup>R</sup>

The antimicrobial activity of compounds ResQAC<sup>butyl</sup>, ResQAC<sup>undecyl</sup>, and ResQAC<sup>phenyl</sup> was evaluated by comparative analysis of both the MIC and the MLD determined for the non-pathogenic Gram-negative strain *Escherichia coli* JM109 and Gram-positive *Staphylococcus aureus* (a pathogenic hospital isolate) (Table 1).

All ResQAC<sup>R</sup> compounds were tested as nitrate salts to avoid bromide and iodide counter-anions, thereby enhancing the safety of these compounds for both humans and the environment. In fact, recent reports<sup>29–31</sup> indicate that iodide and bromide salts of QACs typically threaten water sources with pollution. During disinfection, they can produce harmful by-products such as brominated disinfection by-products (Br-DBPs) and iodinated disinfection by-products (I-DBPs), which pose significant health risks.<sup>29–31</sup> All ResQAC<sup>R</sup> compounds showed antimicrobial activity against both microbial strains, but ResQAC<sup>butyl</sup> and ResQAC<sup>undecyl</sup> were more effective, presenting lower MIC and MLD values (Table 1). The toxic activity was variable with respect to the bacterial group. ResQAC<sup>butyl</sup> showed high bacteriostatic and bactericidal activity with very



**Table 1** Antimicrobial activity of the multivalent **ResQAC<sup>R</sup>** compounds and reference compounds **1b**, **2** and **3** towards Gram-positive *Staphylococcus aureus* (a pathogenic hospital isolate) and Gram-negative strain *Escherichia coli* JM109

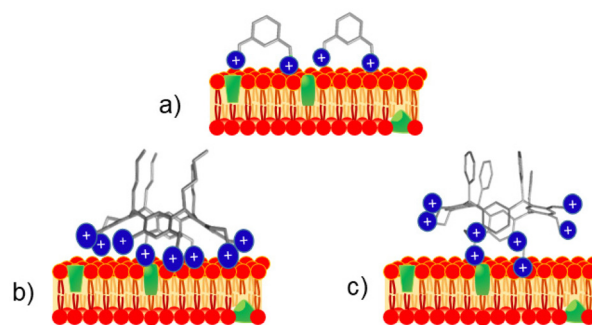
	Valency <sup>a</sup> (n)	<i>Staphylococcus aureus</i>			<i>Escherichia coli</i> JM109		
		MIC <sup>b,c</sup> μM (μg mL <sup>-1</sup> )	r.p.	MLD <sup>d</sup> μM (μg mL <sup>-1</sup> )	MIC <sup>b,c</sup> μM (μg mL <sup>-1</sup> )	r.p.	MLD <sup>d</sup> μM (μg mL <sup>-1</sup> )
<b>ResQAC<sup>butyl</sup></b>	8	0.89 (2)	182	>13 (>30)	≈13 (≈30)	4.6	>125 (>280)
<b>ResQAC<sup>undecyl</sup></b>	8	26 (70)	6.1	≈53 (≈140)	≈6.5 (≈17)	9.4	≈107 (≈280)
<b>ResQAC<sup>phenyl</sup></b>	8	≈43 (≈100)	3.8	>60 (>140)	≈43 (≈100)	1.4	>121 (>280)
<b>1b</b>	0	R <sup>e</sup> > 187 (>280)	—	>187 (>280)	R <sup>e</sup> > 187 (>280)	—	>187 (>280)
<b>2</b>	2	163 (80)	1	≈185 (≈140)	≈61 (≈30)	1	≈143 (≈70)
<b>3</b>	1	R <sup>e</sup> > 954 (>280)	—	>954 (>280)	≈635 (≈140)	—	>954 (>280)

<sup>a</sup> n = number of ammonium units. <sup>b</sup> MIC (minimum inhibitory concentration). <sup>c</sup> r.p. = relative potency = MIC (derivative 2)/MIC (**ResQAC<sup>R</sup>**). <sup>d</sup> MLD (minimum lethal dose). <sup>e</sup> R = resistance to the derivative.

low MIC and MLD (0.89 and >13 μM, respectively) for the Gram-positive strain; on the other hand, **ResQAC<sup>undecyl</sup>** was more bacteriostatic against *E. coli* than against *S. aureus*, but more bactericidal for the latter (lower MLD values). The less efficient **ResQAC<sup>phenyl</sup>** showed similar bacteriostatic activity for the two bacteria (MIC = 43 μM), but a variable bactericidal effect. As reported in Table 1, resorcinarene **1b** (Fig. 2) bearing eight neutral heptyl groups on the wider rim didn't show any bacteriostatic or bactericidal effects for the doses tested. With the aim of quantifying the multivalent effect, we chose the resorcinol diammonium derivative **2** and the dibutyl-dimethylammonium derivative **3** shown in Fig. 2 as the reference divalent and monovalent compounds, respectively. The monovalent derivative **3** exhibited very low activity against both *E. coli* and *S. aureus* (MIC = 635 and >954 μM, respectively, Table 1). The resorcinol diammonium derivative **2**, which mimics a single monomeric unit of the multivalent **ResQAC<sup>R</sup>** macrocycles, showed bacteriostatic activity, MIC = 163 and 61 μM, for *S. aureus* and *E. coli*, respectively (Table 1), which was significantly lower than that found for the **ResQAC<sup>butyl</sup>** compound (0.89 and 13 μM).

Thus, the relative antimicrobial potencies (r.p. in Table 1) of **ResQAC<sup>R</sup>** derivatives were obtained by calculating the ratio of the MIC values (μM) of the resorcinol diammonium **2** to that of the multivalent **ResQAC<sup>R</sup>**.<sup>32</sup> In detail, a remarkable multivalent effect with an r.p. of 182 was observed for the **ResQAC<sup>butyl</sup>** derivative (MIC = 0.89 μM) against *S. aureus*. According to this finding, each resorcinol diammonium monomer in **ResQAC<sup>butyl</sup>** experiences an antimicrobial potency amplification of 45.5 times compared to the reference **2** (Fig. 5a and b). The data reported in Table 1 clearly show that *S. aureus* is more responsive to tetra-alkylammonium multimerization than *E. coli*. In fact, multivalent **ResQAC<sup>R</sup>** derivatives showed low r.p. values ranging from 1.4 to 9.4 against *E. coli*. Remarkably, the **ResQAC<sup>undecyl</sup>** derivative demonstrated lower bacteriostatic activity against *S. aureus* (MIC = 26 μM) than **ResQAC<sup>butyl</sup>** and exhibited six times greater efficiency than derivative **2** (r. p. = 6.1). Interestingly, **ResQAC<sup>phenyl</sup>** showed a modest bacteriostatic activity towards both *S. aureus* and *E. coli* and very low multivalent effects (r.p. = 1.4 and 3.8). Thus, by close inspection of the data reported in Table 1, we can conclude that:

(a) the antibacterial activity of **ResQAC<sup>R</sup>** derivatives is attributable to the interaction with the bacterial cell wall



**Fig. 5** Cartoon representations depicting the proposed model for the interaction between Gram-positive bacteria and (a) derivative **2**; (b) **ResQAC<sup>butyl</sup>** in the crown conformation; and (c) **ResQAC<sup>phenyl</sup>** in the boat conformation.

(Fig. 5a–c). As is known<sup>33–37</sup> for other quaternary ammonium compounds and polycationic macrocycle derivatives, **ResQAC<sup>R</sup>** also can bind with the negatively charged bacterial surface by electrostatic and hydrophobic interactions (Fig. 5). Differently, no significant biocidal effect of compound **1b** was observed against both *E. coli* and *S. aureus*.

(b) A significant bacteriostatic activity was observed for the **ResQAC<sup>butyl</sup>** derivative (MIC = 0.89 μM) against *S. aureus*. In addition, **ResQAC<sup>butyl</sup>** shows a remarkable multivalent effect against *S. aureus*.

(c) *E. coli* is more responsive to **ResQAC<sup>undecyl</sup>**.

(d) As demonstrated previously by some of the authors,<sup>20,21</sup> the effectiveness of resorcinarene-based multivalent ligands relies on achieving a delicate balance between the preorganization of the macrocycle and its conformational flexibility.<sup>38</sup> In fact, the effectiveness of multi-QACs against bacteria relies on their ability to align all their ammonium groups with the negatively charged bacterial surface (Fig. 5). **ResQAC<sup>butyl</sup>** and **ResQAC<sup>undecyl</sup>** derivatives exhibit conformational adaptability, allowing them to fit and interact optimally with the bacterial surface (Fig. 5b). In contrast, the **ResQAC<sup>phenyl</sup>** derivative features a rigid boat-shaped structure, with four alkylammonium groups positioned away from the surface (Fig. 5c) and four cationic groups facing towards it. Consequently, the decreased activity of **ResQAC<sup>phenyl</sup>** is likely due to its weaker interaction



with the bacterial wall, as it utilizes only four of its available eight cationic groups (Fig. 5c).<sup>38</sup>

### Effect of ResQAC<sup>R</sup> on cell viability of human HaCat cells

To evaluate the ability of ResQAC<sup>R</sup> compounds to affect the cell viability of eukaryotic cells, a model of immortalized human keratinocytes (HaCat cells) was used. A 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay was performed after 24 h of treatment with different concentrations of each compound. The percentage of cell viability measured at 100 μM and the calculated IC<sub>50</sub> values are listed in Table 2. The dose-response curves in the concentration range from 1 to 150 μM for ResQAC<sup>butyl</sup> and compound 2 are presented in Fig. 6. From all these data, we can confirm that ResQAC<sup>butyl</sup> has no negative effect on cell viability at concentrations reaching about 30 times the MIC for *S. aureus* and about 3 times the MIC for *E. coli*. It presents only a moderate toxicity at higher concentrations (IC<sub>50</sub> = 54.3 μM). On the other hand, compound 2 was less cytotoxic

than ResQAC<sup>butyl</sup>, with IC<sub>50</sub> almost twice that of ResQAC<sup>butyl</sup>; however, at 100 μM, *i.e.* a concentration below the MIC for *S. aureus* and at about 1.6 times the MIC for *E. coli*, cell viability was reduced by more than 50%.

Finally, viability assays clearly indicated that ResQAC<sup>undecyl</sup> was cytotoxic at concentrations (IC<sub>50</sub> = 12.1 μM) lower than its MIC for *S. aureus*, while ResQAC<sup>phenyl</sup> had a moderate ability to reduce cell viability (IC<sub>50</sub> = 99.8 μM), whereas compounds **1b** and **3** had no effect on the viability of HaCat cells.

## Conclusions

We reported the synthesis of novel resorcin[4]arene multi-QACs (ResQAC<sup>R</sup>) bearing eight tetraalkylammonium groups on the wider rim. This study marks the first example, in the resorcin[4]arene literature, of direct macrocyclization of resorcinol monomers functionalized with bromoalkane groups. In this regard, the information described here could pave the way for the easy synthesis of new resorcin[4]arene hosts.

In the solid state, ResQAC<sup>butyl</sup> adopts a boat conformation with a pseudo C<sub>2v</sub> symmetry (Fig. 3) to minimize the electrostatic repulsion between the positively charged quaternary alkylammonium side chains. For the first time, the conformational dynamics properties of resorcin[4]arenes bearing cationic groups on the wider rim have been investigated. Dynamic NMR investigations indicate that cationic chains do not inhibit the boat-to-crown conformational process, whereas the groups on the CH-bridges play a crucial role in this process. In fact, ResQAC<sup>butyl</sup> and ResQAC<sup>undecyl</sup> derivatives exhibit a boat-crown-boat conformational interconversion. Conversely, for the ResQAC<sup>phenyl</sup> derivative, conformational boat-to-crown interconversion is blocked within the NMR time scale.

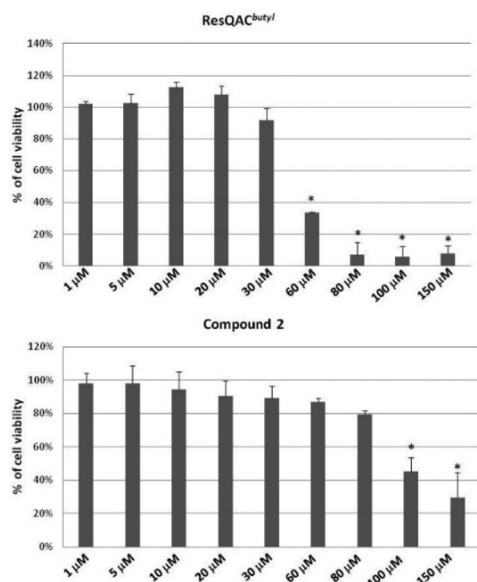
Biological studies unequivocally underscore the antibacterial abilities of ResQAC<sup>butyl</sup> and ResQAC<sup>undecyl</sup> derivatives. ResQAC<sup>butyl</sup> demonstrates a notable bacteriostatic activity against *S. aureus* with a remarkable multivalent effect. Interestingly, studies on the viability of HaCat cells indicate that ResQAC<sup>butyl</sup> exhibits no detrimental effect on cell viability, even at a concentration higher than the MIC for *S. aureus* and *E. coli*. ResQAC<sup>phenyl</sup> show a modest bacteriostatic activity against both *S. aureus* and *E. coli*. Probably, this is due to the conformational rigidity of the resorcinarene framework in ResQAC<sup>phenyl</sup>, which does not facilitate optimal matching between the ammonium groups and the negatively charged bacterial surface. These results corroborate<sup>21</sup> the significance of the conformational flexibility of macrocyclic scaffolds in relation to their interaction with biological targets.<sup>21</sup>

## Author contributions

V. F., V. I. and P. D. S. synthesized and characterized all compounds. L. D. S. and G. V. performed all antimicrobial tests. G. P. and I. C. performed all cell viability tests. S. G. and

**Table 2** Residual cell viability measured in HaCat cells treated for 24 h with each compound at 100 μM. Calculated mean IC<sub>50</sub> values are also reported

	Residual cell viability at 100 (μM)	IC <sub>50</sub> (μM)
ResQAC <sup>butyl</sup>	4.1%	54.3
ResQAC <sup>undecyl</sup>	4.7%	12.1
ResQAC <sup>phenyl</sup>	49.8%	99.8
<b>1b</b>	99.4%	>300
<b>2</b>	45.3%	97.2
<b>3</b>	100%	>1 mM



**Fig. 6** Effect of ResQAC<sup>butyl</sup> and compound 2 on the viability of human keratinocytes (HaCat) after 24 h of treatment with concentrations ranging from 1 μM to 150 μM. Mean values (± standard deviation) of biological triplicate are reported. \**p* < 0.0001 vs. vehicle-treated cells.



N. H. were responsible for the X-ray crystal structure analyses. P. D. S. performed the FT ICR MS analysis. V. I., G. V. and C. T. supervised the project. C. G. secured funding and overall supervised the project. The manuscript was written by C. G., P. N., and C. T. and edited by all co-authors.

## Conflicts of interest

There are no conflicts to declare.

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