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## Ferrocene- and ruthenium arene-containing glycomimetics as selective inhibitors of human galectin-1 and -3<sup>†</sup>

Vojtěch Hamala,<sup>a,b</sup> Martin Kurfírt,<sup>a,b</sup> Lucie Červenková Šťastná,<sup>a</sup> Hedvika Hujerová,<sup>a</sup> Jana Bernášková,<sup>a</sup> Kamil Parkan,<sup>c,d</sup> Jakub Kaminský,<sup>d</sup> Nina Habanová,<sup>d</sup> Jaroslav Kozák,<sup>d</sup> Alžběta Magdolenová,<sup>d</sup> Martin Zavřel,<sup>d</sup> Tatiana Staroňová,<sup>e,f</sup> Veronika Ostatná,<sup>f</sup> Lucie Žaloudková,<sup>e,f</sup> Aleš Daňhel,<sup>f</sup> Jitka Holčáková,<sup>g</sup> Petr Voňka,<sup>g</sup> Roman Hrstka<sup>g</sup> and Jindřich Karban<sup>a</sup>

Galectins are a family of  $\beta$ -galactoside-binding proteins with an evolutionarily conserved carbohydrate recognition domain. Their dysregulation has been implicated in physiological and pathological processes, including fibrotic disorders, inflammation, and cancer. For example, elevated levels of galectin-1 contribute to tumor cell migration and immune evasion, whereas overexpression of galectin-3 is associated with increased invasiveness and the formation of metastasis. Pharmacological inhibition of these galectins is a promising therapeutic strategy to counteract their oncogenic effects. In this study, we synthesized a novel series of galectin inhibitors with ferrocene and ruthenium arene motifs attached to lactose, *N*-acetyllactosamine, or thiogalactoside scaffolds. We determined their binding affinity toward human galectin-1 (*hgal-1*) and the CRD domain of human galectin-3 (*hgal-3-CRD*) using fluorescence polarization, intrinsic fluorescence of galectin tryptophan residues, and isothermal titration calorimetry. The ferrocene analogs exhibited superior affinity for both *hgal-1* and *hgal-3-CRD* compared with ruthenium arenes. In particular, a symmetrical diferrocene thiogalactoside complex exhibited low nanomolar affinity for *hgal-1* and selectivity over *hgal-3-CRD*. Asymmetrical monoferrocene thiogalactoside complexes exhibited nanomolar affinity and good selectivity for *hgal-3-CRD*. Chronopotentiometric stripping analysis demonstrated that the inhibitors stabilized *hgal-1* against destabilization by electric field effects. <sup>19</sup>F{<sup>1</sup>H} NMR experiments and molecular dynamics simulations suggested that the incorporation of the ferrocene motif limited the accessible binding modes to *hgal-3-CRD* whereas binding to *hgal-1* remained unrestricted, resulting in attenuated binding affinities to *hgal-3-CRD* and selectivity for *hgal-1*. These results open new possibilities for the design and optimization of therapeutic organometallic galectin inhibitors.

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<sup>a</sup>Institute of Chemical Process Fundamentals of the CAS, v. v. i., Rozvojová 1/135, 165 00 Praha, Czech Republic. E-mail: karban@icpf.cas.cz

<sup>b</sup>Department of Organic Chemistry, University of Chemistry and Technology, Technická 5, 166 28 Praha, Czech Republic

<sup>c</sup>Department of Chemistry of Natural Compounds, University of Chemistry and Technology, Prague, Technická 5, 166 28 Prague, Czech Republic

<sup>d</sup>Institute of Organic Chemistry and Biochemistry of the CAS, v. v. i., Flemingovo nám. 542, 160 00 Praha, Czech Republic

<sup>e</sup>Department of Biochemistry, Faculty of Science, Masaryk University, Kotlářská 2, 602 00 Brno, Czech Republic

<sup>f</sup>Institute of Biophysics of the CAS, v. v. i., Královopolská 135, 612 00 Brno, Czech Republic

<sup>g</sup>Research Centre for Applied Molecular Oncology, Masaryk Memorial Cancer Institute, Žlutý kopec 7, 656 53 Brno, Czech Republic. E-mail: hrstka@mou.cz

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## Introduction

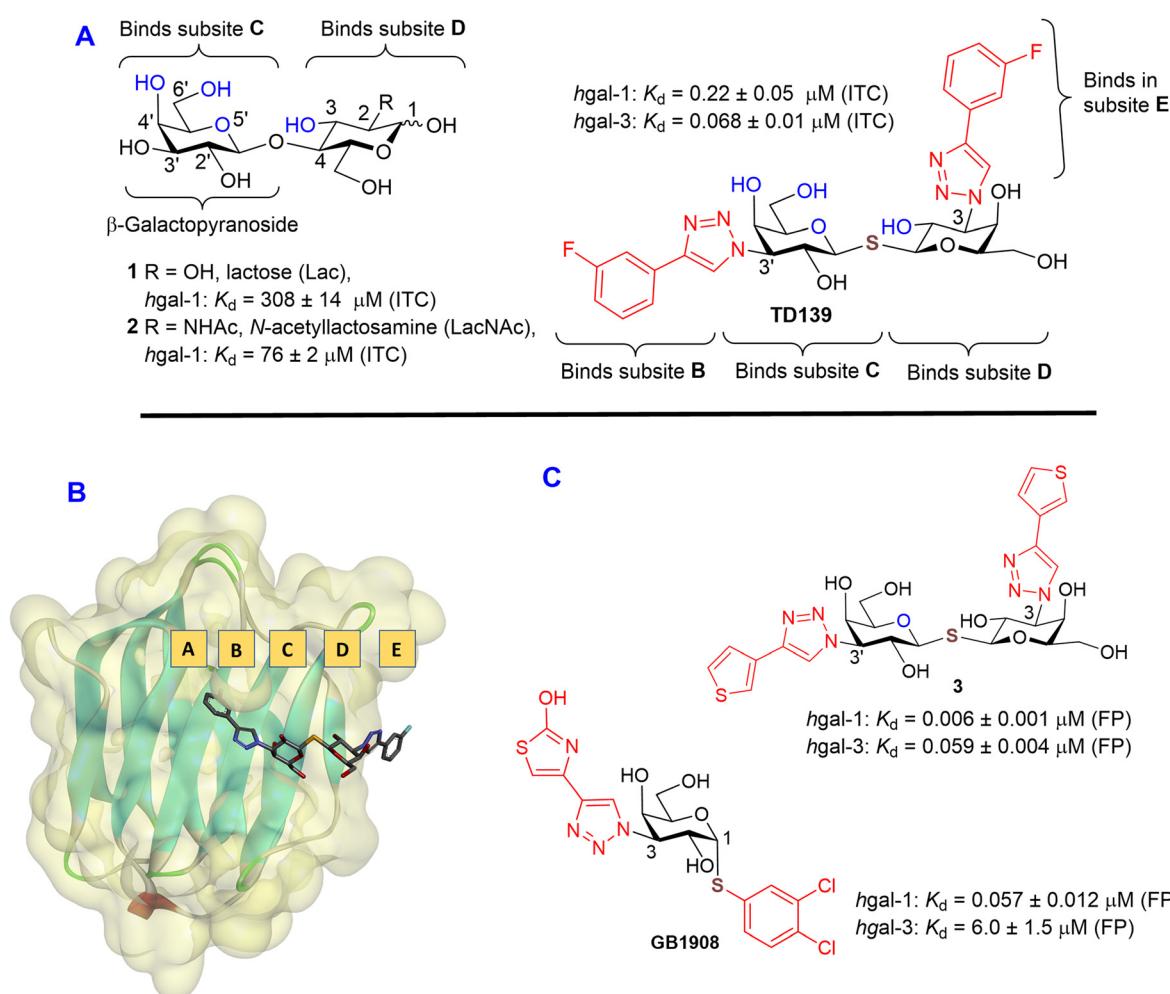
Lectins are glycan-binding proteins that are abundant in all kingdoms of life.<sup>1</sup> In humans, binding of glycans to lectins is essential for many physiological and pathological processes including host-pathogen interactions, inflammation, autoimmune disorders, and cancer progression.<sup>2</sup> Although the key roles of lectins in pathologies and their widespread occurrence make them an important target for drug development,<sup>2b</sup> their pharmacological inhibition by small molecule inhibitors remains underexplored because of the perceived poor druggability of lectins.<sup>3</sup> Ligand binding to the shallow water-exposed binding cavity of lectins is associated with a high desolvation penalty, moderate binding affinities of monovalent sugars, and little ability of an inhibitor to form additional productive



interactions within the binding site.<sup>4</sup> Moreover, carbohydrates have poor pharmacokinetic properties,<sup>4d</sup> and because lectins form homologous families that bind the same structural motif, it is difficult to achieve selectivity in lectin inhibition. Despite these extraordinary challenges, developments in glycochemistry over the past 20 years have produced potent lectin inhibitors.<sup>4b</sup> The most successful approach in the design of lectin inhibitors relies on synthetically modified endogenous carbohydrate ligands (glycomimetics).<sup>4b</sup> Several glycomimetic lectin inhibitors have entered advanced phases of clinical trials,<sup>4b,5</sup> but with the exception of GalNAc-siRNA conjugates, none others have yet been marketed.<sup>4b</sup> Thus, the need for innovative approaches to expand the available chemical space diversity in the design of lectin inhibitors is undeniable.

Galectins are a family of animal lectins that non-covalently bind a  $\beta$ -galactopyranoside motif in membrane-bound and extracellular glycans through an evolutionarily conserved

binding site located within a carbohydrate recognition domain (CRD).<sup>4c,6</sup> Natural disaccharide galectin ligands, such as lactose (Lac) 1 and *N*-acetyllactosamine (LacNAc) 2 (Fig. 1A), bind the highly conserved region of the CRD termed subsites C and D, with each subsite accommodating one hexopyranose moiety (Fig. 1A).<sup>7</sup> The  $\beta$ -galactopyranoside moiety in these disaccharides is recognized by subsite C. Two additional monosaccharides, which can be attached to the 3'-position of the galactopyranoside moiety in endogenous ligands, are recognized by the less conserved extension of the binding groove termed subsites A and B (Fig. 1A and B).<sup>7a,b</sup> The loosely defined subsite E can interact with the substituents attached to the reducing end of the monosaccharide occupying subsite D (Fig. 1B).<sup>7d,8</sup> Galectins are implicated in several pathologies including fibrotic diseases, inflammation, and cancer.<sup>9</sup> Among sixteen mammalian galectins,<sup>7b</sup> human galectin-1 (*hgal-1*) and -3 (*hgal-3*) are probably the most studied to date due to their



**Fig. 1** (A) Structures, numbering, and dissociation constants  $K_d$  of natural disaccharide galectin ligands 1 and 2, and the galectin inhibitor TD139 determined by isothermal titration calorimetry (ITC).<sup>12a</sup> Oxygen functionalities critical for binding are shown in blue, 3-fluorophenyl-triazole moiety of TD139 is shown in red. (B) Binding subsites A–E are shown for the complex of TD139 with *hgal-3*-CRD (PDB: 5H9P). (C) Structures of thiophene-derived *hgal-1* inhibitor 3, thiazole-derived *hgal-1* inhibitor GB1908 and their dissociation constants  $K_d$  determined by competitive fluorescence polarization (FP).<sup>13</sup> (hetero)aromatic moieties shown in red.

expression in many tissues and their prominent role in diseases.<sup>4c,10</sup> The involvement of galectins in pathologies is complex because galectins are multifunctional proteins whose functions depend on their cellular and tissue localization.<sup>11</sup> Moreover, different galectins can play opposite roles in disease progression.<sup>10</sup> Therefore, the availability of galectin inhibitors that are highly selective for individual galectins is essential for both fundamental research and drug development.

Typical glycomimetic small molecule galectin inhibitors comprise a galactose-containing mono- or disaccharide scaffold capable of forming the critical hydrogen bonds found in natural agonists **1** and **2** (Fig. 1A).<sup>8</sup> The scaffold is typically decorated with aromatic non-polar substituents that introduce additional interactions within the binding groove.<sup>8</sup>

The glycomimetic inhibitor **TD139** (later renamed **GB0139**, Fig. 1A) targets four subsites and illustrates this approach.<sup>14</sup> The hydroxyl groups at the 3-, 4'- and 6'-positions of natural ligands **1** and **2** (including their stereochemistry), the pyranose oxygen O5', and the hydrophobic  $\alpha$ -side of the galactoside residue are reproduced in the thiogalactoside scaffold of **TD139**, and direct the inhibitor into subsites C and D (Fig. 1A) *via* key interactions with amino acid residues in these subsites. The fluorophenyl-triazole moiety forms guanidinium-arene cation- $\pi$  interactions with the arginine residues Arg144 and Arg186 in subsites B and E of *hgal-3*, respectively, while fluorine is involved in orthogonal multipolar interactions<sup>15</sup> with the polypeptide backbone, collectively leading to nanomolar affinities toward *hgal-3* and approximately 3-fold weaker binding to *hgal-1* according to ITC (Fig. 1A).<sup>12b</sup> Modifications of **TD139** and other inhibitors resulted in a series of excellent *hgal-3* selective disaccharide<sup>8,16</sup> and monosaccharide<sup>17</sup> inhibitors. **TD139** entered the Phase IIb of clinical trials for the treatment of idiopathic pulmonary fibrosis (IPF), but unfortunately failed due to its low efficacy and its development for IPF treatment was discontinued.<sup>18</sup>

Although the development of *hgal-3* inhibitors has advanced considerably, the development of potent *hgal-1* selective inhibitors remains challenging despite their therapeutic potential.<sup>19</sup> To the best of our knowledge, the only reported small molecule, single-digit nanomolar inhibitors of *hgal-1* are the thien-3-yl derivative **3**<sup>13a,14</sup> (Fig. 1C) and its thiazol-2-yl analog.<sup>13a</sup> Both had an approximately 10-fold selectivity for *hgal-1* over *hgal-3* as determined by a competitive fluorescence polarization (FP) assay. However, no further bioactivity of compound **3** has been reported, and it appears that this inhibitor is not being further developed. Very recently, a thiogalactoside-based inhibitor named **GB1908** with a two orders of magnitude *hgal-1* selectivity has been reported (Fig. 1C).<sup>13b</sup>

Recent X-ray diffraction and <sup>19</sup>F NMR studies have suggested differences between *hgal-1* and *hgal-3* in the binding mode of **TD139** for subsites B and E.<sup>12b</sup> Inspired by these findings, we hypothesize that lipophilic barrel-shaped organometallic complexes such as ferrocene or cyclopentadienyl-ruthenium arenes may exploit these differences and interact with the binding subsites of galectins more efficiently or selectively than the planar fluorophenyl moiety, leading to higher

binding affinity or selectivity. Organotransition metal complexes, due to their three-dimensional coordination sphere, offer structural diversity in protein-ligand interactions that is inaccessible to purely organic compounds.<sup>20</sup> For example, improved affinity or selectivity after substituting a ferrocene moiety for a planar arene has been reported for the anticancer drugs paclitaxel,<sup>21</sup> tamoxifen,<sup>20c</sup> the drug candidate plinabulin,<sup>22</sup> and the dopamine D<sub>3</sub> receptor antagonist BP 897.<sup>23</sup> In spite of their advantages, organotransition metal complexes have not been tested as antagonists in lectin inhibition, although multivalent lactose-ferrocene conjugates have been evaluated as electrochemical probes for the detection of galectin-3.<sup>24</sup>

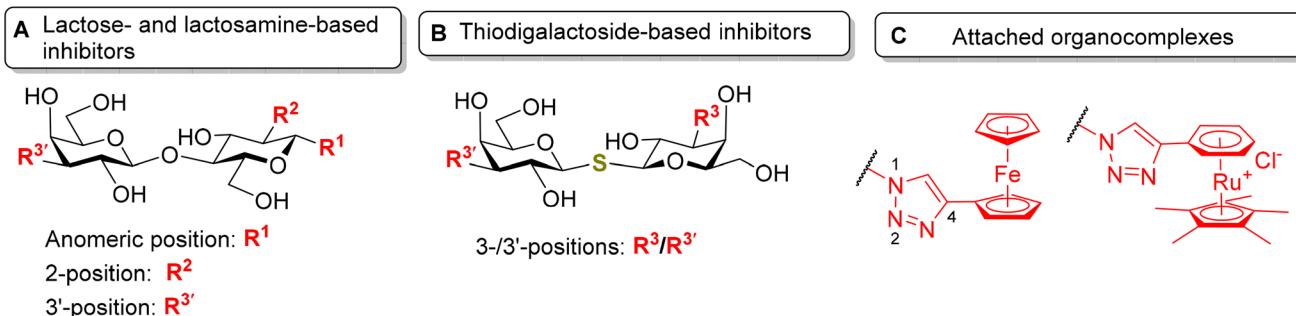
We prepared a series of lactose, *N*-acetyllactosamine- and thiogalactoside-based inhibitors bearing ferrocene or pentamethylcyclopentadienyl ruthenium arene complexes and determined their affinities to *hgal-1* and *hgal-3* using the FP assay, intrinsic fluorescence of galectin tryptophan residues, and ITC. Here, we report that introducing two ferrocene-triazole moieties into the thiogalactoside scaffold resulted in an *hgal-1* inhibitor with potency comparable to **TD139** but with up to 50-fold higher selectivity for *hgal-1* over *hgal-3*, highlighting the potential of organometallic fragments in the design of lectin inhibitors. Subsequent <sup>19</sup>F{<sup>1</sup>H} NMR monitoring of the 3-fluorophenyl-triazole moiety and molecular dynamics simulations were used to explain the differences in the binding mode between *hgal-1* and *hgal-3*. In addition, the electrochemical activity of selected ferrocene derivatives was studied by cyclic voltammetry (CV), and the ability of the synthesized inhibitors to stabilize *hgal-1* against electric current was evaluated using chronopotentiometric stripping analysis.

## Results

### Synthesis of galectin antagonists

The ferrocenyl and ruthenium arene moieties were attached to the selected disaccharide scaffolds *via* a 1,2,3-triazole linker substituted at the 4-position with an organocomplex (Fig. 2C). The linker was formed by a copper(I)-catalyzed cycloaddition reaction between ferrocenyl- or phenylacetylene and an azido group at the selected positions of the disaccharides (Fig. 2, substituents **R**<sup>1</sup>, **R**<sup>2</sup>, **R**<sup>3</sup>, **R**<sup>3'</sup>). These positions were chosen because their modifications generally do not interfere with recognition by galectins and organometallic substituents at these positions enabled us to explore possible attractive interactions with subsites A/B and E flanking the most conserved region of subsites C and D.<sup>7b,8,25</sup> For Lac- and LacNAc-derived galectin ligands, we decided to attach the organometallic substituent at the reducing end anomeric position (Fig. 2A, substituent **R**<sup>1</sup>) to exploit possible interactions with subsite E. Similar reasoning applies to the 3'-OH hydroxyl of *N*-acetyllactosamine **2** (Fig. 2A, substituent **R**<sup>3'</sup>), which points toward subsites A and B. In addition, the 2-position of the lactosamine scaffold (Fig. 2A, substituent **R**<sup>2</sup>) was also modified because this position tolerates modifications with bulky sub-





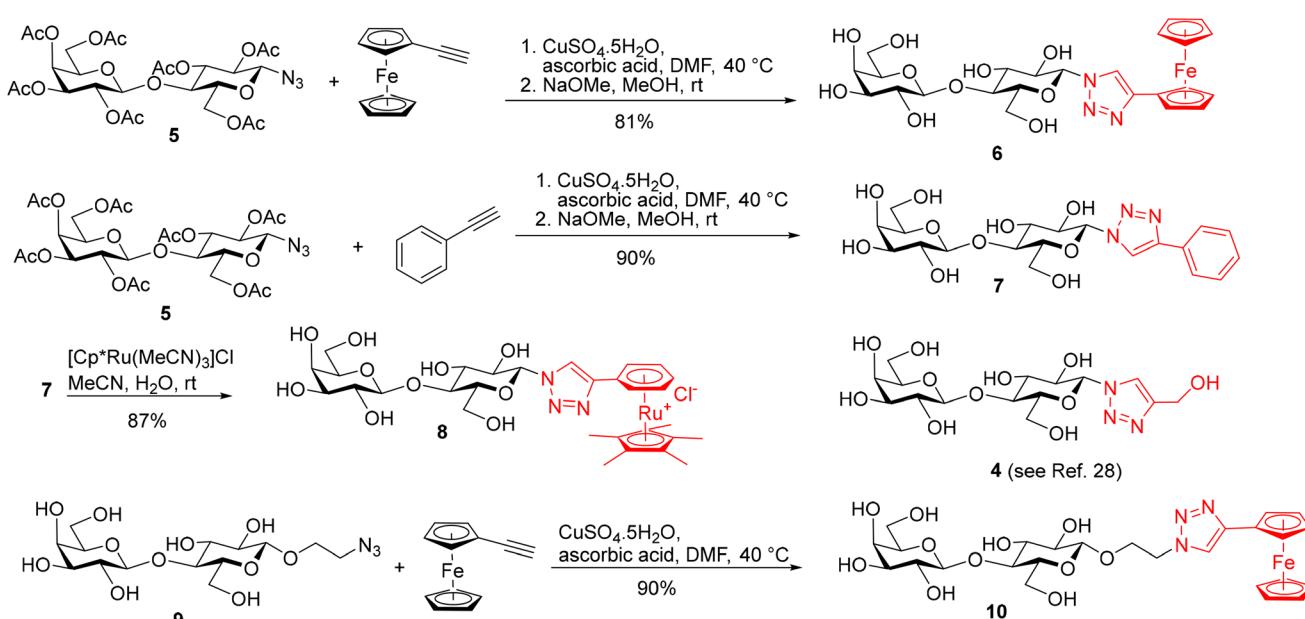
**Fig. 2** The positions selected for the attachment of modifying substituents to (A) lactose ( $R^2 = OH$ ) or lactosamine ( $R^2 = NH_2$ ), (B) thiodigalactoside. (C) Organocomplexes attached to carbohydrate scaffolds and numbering of the 1,2,3-triazole linking moiety.

stituents.<sup>7b</sup> For the symmetrical thiogalactoside scaffold, we modified the 3/3'-position (Fig. 2B, substituents  $R^3/R^3'$ ). Modification at this position with an arene group has previously produced potent galectin inhibitors.<sup>8,26</sup> The ferrocene moiety was introduced directly into the disaccharide scaffold by azide–alkyne cycloaddition of the corresponding azido-disaccharides and ethynylferrocene. The (pentamethylcyclopentadienyl)ruthenium arene moiety was introduced by the complexation reaction of a phenyltriazole moiety with the  $[Cp^*Ru(MeCN)_3]Cl$  complex generated *in situ*.<sup>27</sup> Non-metallic compounds bearing a hydroxymethyl, phenyl, or 3-fluorophenyl substituent attached to the disaccharide scaffold *via* a triazole linker were prepared for comparison (Scheme 1, compounds 4 and 7; Scheme 3, compounds 26, 28 and 29; Scheme 4, compound 37).

The synthesis of Lac-derived galectin ligands 4, 6–8, and 10 modified at the anomeric position is summarized in Scheme 1. Hydroxymethyl-triazole-substituted lactose 4 was

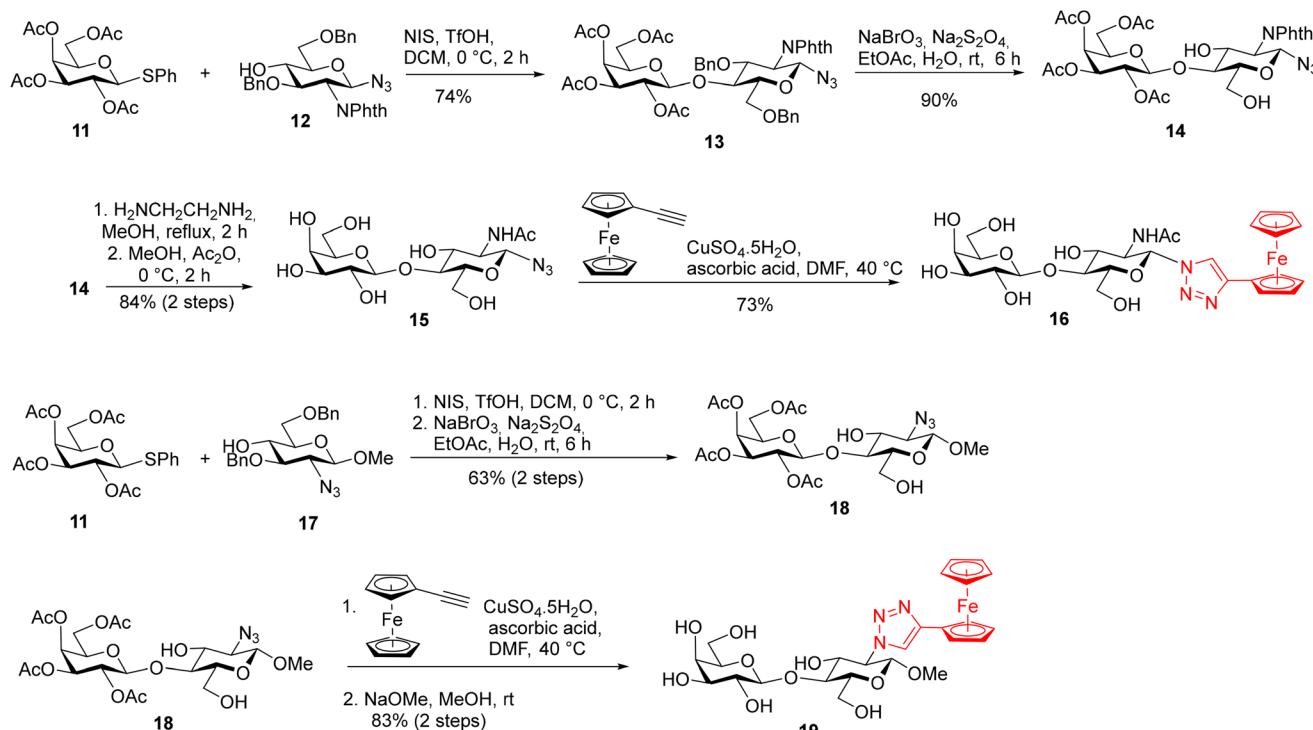
prepared for comparison as reported.<sup>28</sup> Starting azido disaccharides 5<sup>29</sup> and 9<sup>30</sup> were prepared from lactose 1 as reported and converted to the target ferrocene analogs 6 and 10 using cycloaddition with ethynylferrocene as the key reaction. The phenyl triazole 7 was obtained from azide 5 by reaction with phenyl acetylene followed by Zemplén deacetylation. Phenyl analog 7 was subjected to the complexation reaction with the  $[Cp^*Ru(MeCN)_3]Cl$  complex generated *in situ* from the tetramer  $[Cp^*RuCl]_4$  as described previously.<sup>27</sup> This complexation reaction afforded the corresponding ruthenium arene 8 (Scheme 1).

The synthesis of LacNAc-derived galectin ligands 16 and 19 modified at the 1- and 2-positions, respectively, is summarized in Scheme 2. Azides 15 and 18 for the copper-catalyzed cycloaddition were prepared from monosaccharide building blocks by chemical glycosylation. *N*-Acetylglucosaminyl azide 15 was obtained by NIS/TfOH-promoted glycosylation<sup>31</sup> of known 2-phthalimido- $\beta$ -glucosyl-azide 12<sup>32</sup> with thiogalactoside 11<sup>33</sup>



**Scheme 1** Synthesis of galectin ligands 4, 6–8, and 10 derived from lactose modified at the anomeric position.





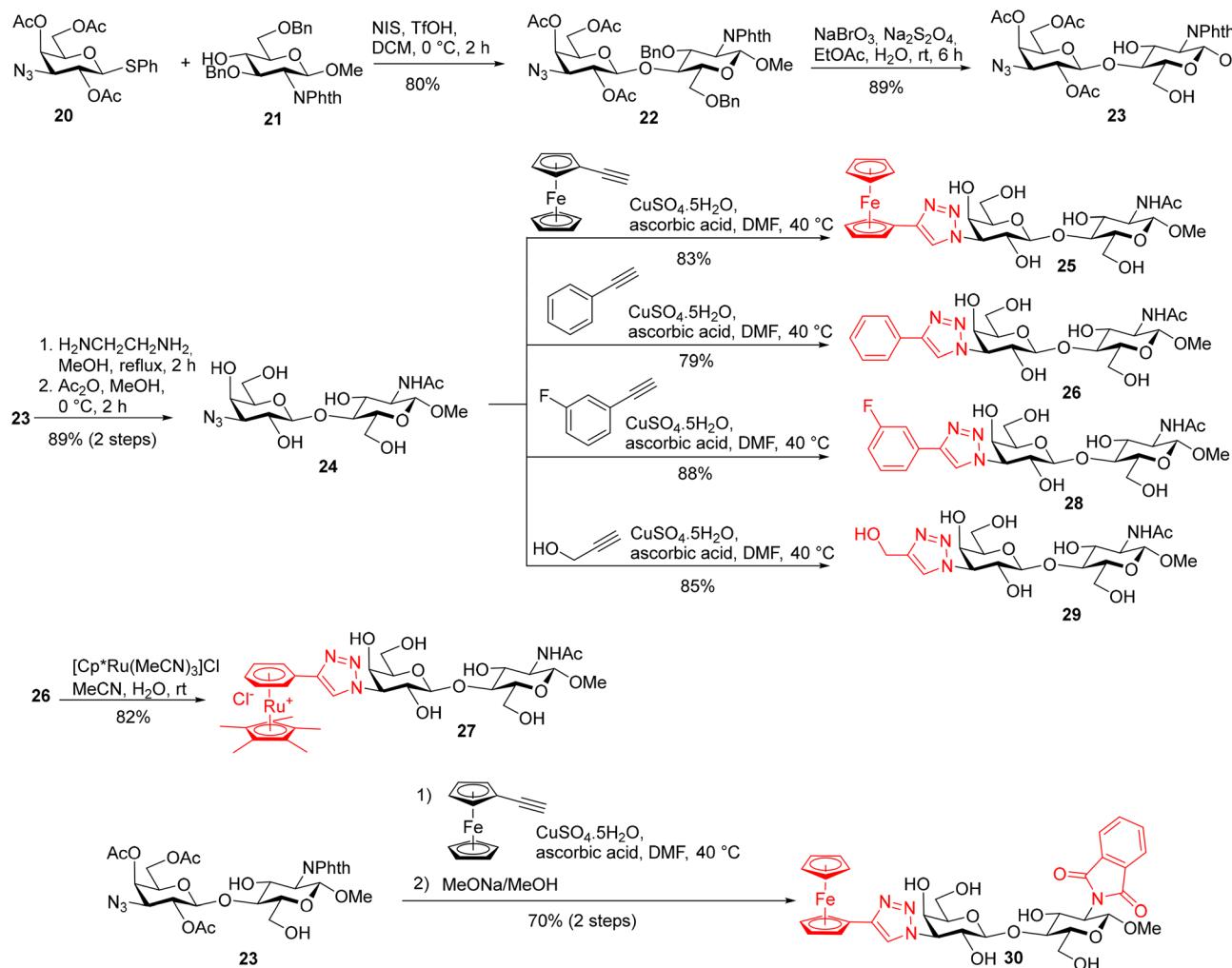
**Scheme 2** Synthesis of galectin ligands **16**, and **19** derived from LacNac modified at the 1- and 2-positions.

followed by sequential deprotection, using first oxidative de-*O*-benzylation with the NaBrO<sub>3</sub>/Na<sub>2</sub>S<sub>2</sub>O<sub>4</sub> system<sup>34</sup> to give compound **14**, then treatment with ethylenediamine in refluxing methanol to liberate the 2-amino group,<sup>35</sup> and finally *N*-acetylation (Ac<sub>2</sub>O, MeOH). Methyl 2-azido- $\beta$ -lactoside **18** was obtained by NIS/TfOH-promoted glycosylation of methyl 2-azido- $\beta$ -glucoside **17**<sup>36</sup> with thiogalactoside **11** followed by oxidative de-*O*-benzylation. Copper(I)-catalyzed cycloaddition between azido disaccharides **15**, and **18** and ethynylferrocene, followed by Zemplén deacetylation in the case of acetylated disaccharide **18**, afforded the target ferrocenyl-triazole-substituted disaccharides **16** and **19** (Scheme 2).

The synthesis of LacNac analogs modified at the 3'-position is described in Scheme 3. The methyl  $\beta$ -glycosides **23** and **24** of 3'-azido-LacNac required for azide–alkyne cycloaddition were synthesized by glycosylation of methyl 2-phthalimido- $\beta$ -glucoside **21**<sup>37</sup> with 3'-azido-thiogalactoside **20**.<sup>38</sup> The resulting disaccharide **22** was oxidatively de-*O*-benzylated to yield diol **23** that was subjected to phthalimide deprotection (ethylenediamine in refluxing MeOH) and *N*-acetylation to afford the 3'-azido-LacNac-OMe **24**. Copper(I)-catalyzed cycloaddition between azido disaccharides **23**, and **24** and ethynylferrocene, followed by Zemplén deacetylation in the case of acetylated disaccharide **23**, afforded the target ferrocenyl-triazole-substituted disaccharides **30** and **25**, respectively. The bulky phthalimido group (Phth) was retained at the 2-position of disaccharide **30** because a large aromatic substituent at this position may influence the selectivity in interactions with galectins. For comparison, 3-fluorophenyl-triazole-substituted LacNac **28**

and hydroxymethyl-triazole-substituted LacNac **29** were prepared from disaccharide **24** by cycloaddition with 3-fluorophenylacetylene and propargyl alcohol, respectively (Scheme 3). The cycloaddition of azide **24** with phenylacetylene afforded disaccharide **26** carrying a phenyl-triazole ligand suitable for the complexation reaction with the [Cp\*Ru(MeCN)<sub>3</sub>]Cl complex, which afforded the corresponding ruthenium arene **27** (Scheme 3). The parent disaccharides **26** along with disaccharide **7** (Scheme 1), both modified with the phenyl-triazole moiety, were also used as the reference compounds to determine how the substitution of the ruthenium arene or ferrocene for a planar phenyl substituent influenced the affinity. All Lac- and LacNac-based ligands were prepared as  $\beta$ -anomers at the reducing end anomeric position. When this anomeric position did not carry a modifying organometallic substituent, it was protected as a methyl  $\beta$ -glycoside (compounds **19**, **25**–**30**) to prevent the formation of a mixture of anomers.

For the thiogalactoside-based inhibitors, we attached the modifying substituent at the 3- and 3'-positions of the thiogalactoside scaffold (Fig. 2B), as previous work has documented that binding to galectins benefits from the attachment of arene-triazole substituents at these positions.<sup>14</sup> This produced compounds **34**, **36**–**38**, **41**, and **42** (Scheme 4). Starting 3-azido-thiogalactoside **33** was obtained by the tetrabutylammonium fluoride-promoted reaction of 3-azido-galactosyl bromide **31**<sup>39</sup> with thiogalactoside **32**<sup>40</sup> in acetonitrile following the published protocol (Scheme 4).<sup>40,41</sup> 3,3'-Diazido-thiogalactoside **35**,<sup>26c,39</sup> 3,4,5-trifluorophenyl-analog **39**,<sup>16</sup> and 3-fluorophenyl



**Scheme 3** Synthesis of galectin ligands 25–30 derived from LacNAc modified at the 3'-position.

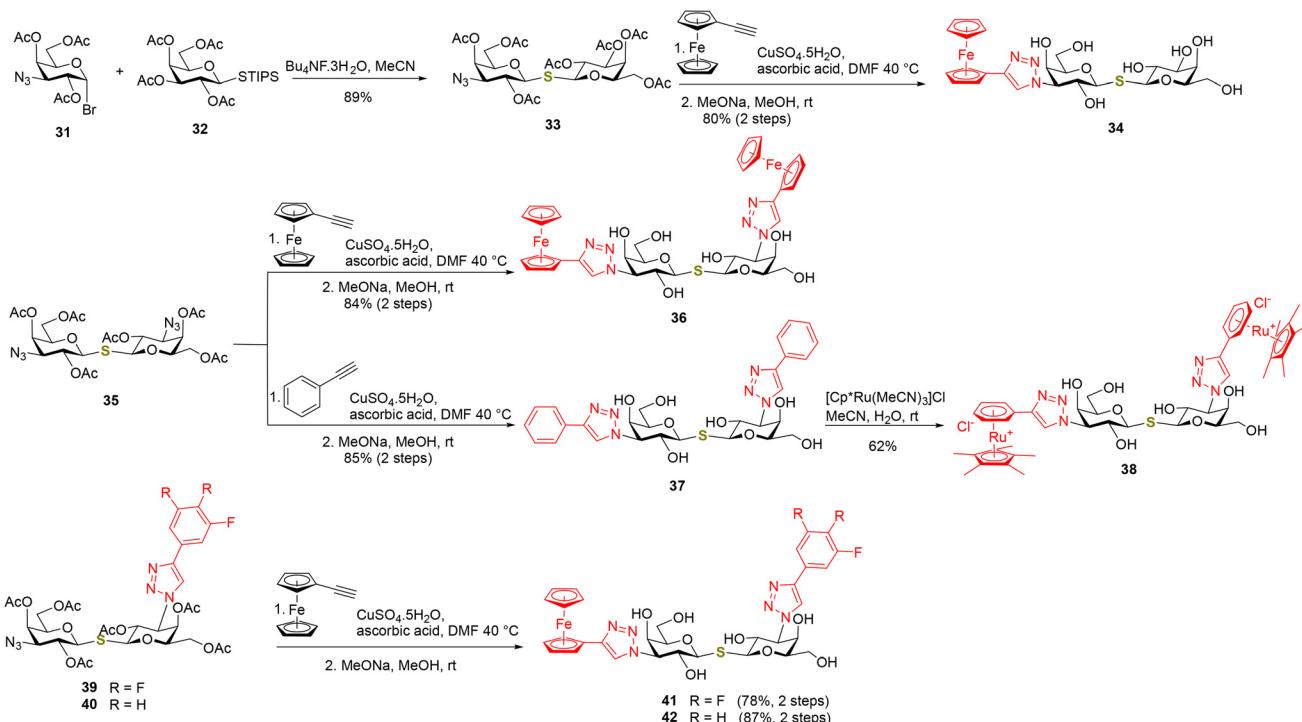
analog **40**<sup>16</sup> were prepared as reported. Cycloaddition of azides **33**, **35**, **39**, and **40** with ethynylferrocene followed by Zemplén deacetylation gave ferrocenyl-triazole-substituted analogs **34**, **36**, **41**, and **42**, respectively, whereas azide–alkyne cycloaddition of diazide **35** with phenylacetylene afforded the phenyl-triazole ligand **37** (Scheme 4). The complexation reaction of compound **37** with  $[\text{Cp}^*\text{Ru}(\text{MeCN})_3]\text{Cl}$  generated *in situ* was expected to afford the bisruthenium arene disaccharide **38**. The product produced the expected molecular ion mass in high resolution mass spectra; however, the <sup>1</sup>H NMR spectrum showed two signals for the triazole proton and four signals for the methyl protons of the pentamethylcyclopentadienyl ring. We speculate that compound **38** was prepared as a mixture of four inseparable stereoisomers due to hindered rotation at the bond between the triazole and the phenyl ligand. All prepared ferrocenes were obtained as either crystalline or amorphous solids; most of them with limited solubility in water (see ESI for details, section B.2.†), whereas the ruthenium arenes were obtained as white amorphous solids with good water solubility. Both types of compounds exhibited stability in air and in

aqueous or DMSO solutions, as confirmed over a period of three weeks.

### Compound characterization

**NMR and HRMS spectroscopy.** The structure of the prepared compounds was confirmed by NMR and HRMS spectroscopy. The 1,2-*trans* (or  $\beta$ -) configuration of the newly formed glycosidic linkage in disaccharides **13**, **18**, **22**, and **33**, was evidenced by the magnitude of the coupling constant  $^3J_{(\text{H}-1',\text{H}-2')}$  = 8.0–9.8 Hz. Ferrocenyl- or phenyltriazole-substituted disaccharides **6**, **7**, and **16**, obtained from glycosyl azides **5** and **15** by cycloaddition, retained the  $\beta$ -configuration at the 1-position and the <sup>4</sup>C<sub>1</sub> chair conformation of both pyranose rings, as evidenced by the values of the vicinal coupling constants  $^3J_{\text{H,H}}$  (ESI, Table S1†). The galactoside ring also retained its <sup>4</sup>C<sub>1</sub> chair conformation after the attachment of the modifying substituent at the 3/3'-position (compounds **25–30**, **34**, **36–38**, **41** and **42**). When signal overlap in <sup>1</sup>H NMR spectra precluded direct determination of the diagnostic  $^3J_{\text{H,H}}$  couplings, 2D <sup>1</sup>H–<sup>13</sup>C gHSQC NMR spectra were used for their estimation.





**Scheme 4** Synthesis of galectin ligands **34**, **36–38**, **41** and **42** derived from thiogalactoside modified at the 3 and 3'-positions.

The formation of 1,2,3-triazole was also confirmed by NMR spectroscopy. The proton of the triazole CH group resonated at 8.01–8.90 ppm in <sup>1</sup>H NMR spectra, while the corresponding carbon resonated at 119.8–124.7 ppm in <sup>13</sup>C NMR spectra, which is characteristic of 1,4-substituted 1,2,3-triazoles.<sup>42</sup> The quaternary carbon of the 1,2,3-triazole ring showed a chemical shift in the range of 141.8–148.4 ppm. Ruthenium complexation led to shielding of the corresponding phenyl ring,<sup>27</sup> resulting in the decreased values of the proton and carbon chemical shifts ( $\delta_H = 5.95$ –6.64 ppm/ $\delta_C = 84.98$ –99.3 ppm for the phenyl ring in the ruthenium complexes,  $\delta_H = 7.30$ –7.88 ppm/ $\delta_C = 125.0$ –131.9 ppm for the phenyl rings in the parent phenyl-substituted disaccharides).

**Cyclic voltammetry.** Ferrocene is a widely used electrochemical redox probe, particularly for biological applications in conjugation with biomolecules.<sup>43</sup> In this study, we established the electrochemical activity of three inhibitors with the ferrocenyl-triazole substituent (**6**, **25**, and **36**) in comparison with ferrocenecarboxylic acid (**FeCOOH**)—a reference electrochemical redox probe—using cyclic voltammetry (CV) at a basal-plane pyrolytic graphite electrode (bPGE).<sup>44</sup> Unmodified disaccharides are electrochemically inactive substances. Because of the presence of readily oxidizable ferrocene tag(s), compounds **6**, **25**, and **36** offered reversible double peak with  $E_{1/2}$  varying between 300 mV (**25**) and 323 mV (**36**), similar to the probe **FeCOOH** with  $E_{1/2} = 320$  mV (Fig. S1 and Table S2 in the ESI†).  $\Delta E_p = 84$  mV of **FeCOOH** was approximately 6 mV (**6**) to 14 mV (**25**) higher than  $\Delta E_p$  of the corresponding ferrocenyl-triazole-modified disaccharides and approximately 25 mV

higher than the theoretical value of 59 mV for an ideally reversible system. Together with higher  $J_p^{a3}/J_p^{c}$  ratios ranging from 1.67 to 2.14, with an ideal value of 1.0, it is possible to attribute the ferrocene redox signals observed at bPGE to quasi-reversible processes,<sup>45</sup> which are not significantly affected by the disaccharide structures. For more details, see ESI, pages S24–S28.†

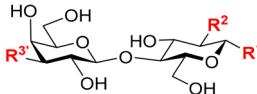
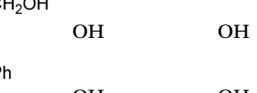
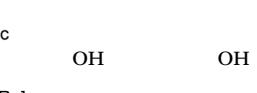
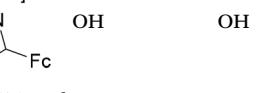
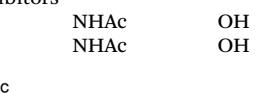
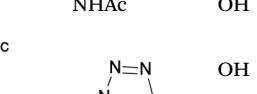
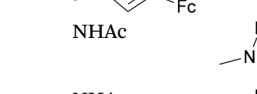
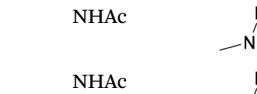
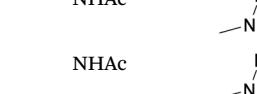
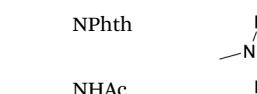
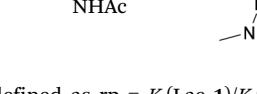
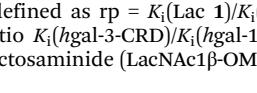
#### Binding to *hgal-1* and *hgal-3*

**Competitive fluorescence polarization (anisotropy) assay.** We screened the affinity of all prepared organometallic complexes and the corresponding nonmetallic reference compounds to *hgal-1* and human galectin-3 CRD domain (*hgal-3-CRD*) using a competitive FP assay.<sup>46</sup> This assay allows a rapid determination of the apparent inhibition constant  $K_i$  with minimal consumption of the inhibitor (Tables 1 and 2, see the ESI† for the details of the calculation).<sup>46</sup> The *hgal-3-CRD* was employed instead of *hgal-3* to facilitate comparison with the literature data and the ITC measurements (*vide infra*).<sup>47</sup> We prepared and used methyl  $\beta$ -glycoside of LacNAc, designated 2 $\beta$ -OMe in his study, as the parent compound for the comparison with the LacNAc-derived ligands in FP and ITC assays.<sup>36</sup>

The attachment of an organometallic moiety to positions 1 and 2 of disaccharides Lac and LacNAc was aimed at the formation of additional interactions with the galectin subsite E.<sup>48</sup> Both galectins contain an arginine residue in this subsite (Arg74 in *hgal-1* and Arg186 in *hgal-3*) capable of forming cation– $\pi$  interactions with arene moieties of the inhibitor.<sup>7a,49</sup> On the whole, modifications at these positions mostly resulted in a modest improvement in binding affinity (Table 1, entries



**Table 1** Apparent inhibition constant  $K_i$ , relative potency rp<sup>a</sup> and selectivity values obtained from competitive fluorescence polarization for inhibitors derived from Lac and LacNAc

Entry	R <sup>1</sup>	R <sup>2</sup>	R <sup>3'</sup>	Compound	$K_i$ (hgαl-1) (μM)	rp (hgαl-1)	$K_i$ (hgαl-3-CRD) (μM)	rp (hgαl-3-CRD)	Selectivity to hgαl-1 <sup>b</sup>
					Compound	$K_i$ (hgαl-1) (μM)	rp (hgαl-3-CRD)	Selectivity to hgαl-1 <sup>b</sup>	
Lac-derived inhibitors <sup>c</sup>									
1	OH	OH	OH	<b>1</b> (lactose)	334 ± 28	1.0	94 ± 7	1.0	0.28
2		OH	OH	<b>4</b>	157 ± 31	2.1	55 ± 4	1.7	0.35
3		OH	OH	<b>7</b>	241 ± 36	1.4	67 ± 7	1.4	0.28
4		OH	OH	<b>6</b>	105 ± 20	3.2	35 ± 1	2.7	0.33
5		OH	OH	<b>8</b>	262 ± 51	1.3	132 ± 15	0.7	0.50
6		OH	OH	<b>10</b>	141 ± 6	2.4	30 ± 1	3.1	0.21
LacNAc-derived inhibitors <sup>c</sup>									
7	OMe	NHAc	OH	2β-OMe <sup>d</sup>	182 ± 12	1.0	49 ± 3	1.0	0.27
8		NHAc	OH	<b>16</b>	73 ± 19	2.5	15 ± 1	3.3	0.20
9	OMe		OH	<b>19</b>	54 ± 2	3.4	3.9 ± 0.1	13	0.07
10	OMe	NHAc		<b>29</b>	43 ± 8	4.2	22 ± 5	2.2	0.51
11	OMe	NHAc		<b>26</b>	5.2 ± 0.8	35	2.7 ± 0.7	18.1	0.52
12	OMe	NHAc		<b>28</b>	1.7 ± 0.1	107	0.26 ± 0.01	188	0.15
13	OMe	NHAc		<b>25</b>	3.7 ± 1.2	49	5.0 ± 0.2	10	1.35
14	OMe	NPhth		<b>30</b>	3.4 ± 0.5	54	1.3 ± 0.1	38	0.38
15	OMe	NHAc		<b>27</b>	78 ± 11	2.3	14.8 ± 3.9	3.3	0.19

<sup>a</sup> Relative potency defined as rp =  $K_i$ (Lac 1)/ $K_i$ (inhibitor) for Lac-derived inhibitors, rp =  $K_i$ (2β-OMe)/ $K_i$ (inhibitor) for LacNAc-based inhibitors.

<sup>b</sup> Defined as the ratio  $K_i$ (hgαl-3-CRD)/ $K_i$ (hgαl-1). <sup>c</sup> Fc = ferrocenyl, [Ru] = ruthenium arene moiety, 3F-Ph = 3-fluorophenyl. <sup>d</sup> 2β-OMe denotes methyl N-acetyl-β-lactosaminide (LacNAc1β-OMe).<sup>36</sup>

1–9). The selectivity was always in favor of hgαl-3-CRD. The best improvement was obtained for compound **19** resulting from the attachment of ferrocenyltriazole to the 2-position of LacNAc. It bound hgαl-3-CRD 13-fold more strongly than the parent methyl N-acetyl-β-lactosaminide<sup>36</sup> 2β-OMe (entry 9). Complex **19** had affinity and selectivity comparable with that obtained for 5,6,7,8-tetrahydronaphthoate **43** (Fig. 3), which was among the most selective hgαl-3 binders in a series of previously reported lactose 2-esters.<sup>49</sup> The remaining complexes **6**, **8**, **10**, and **16** had at most a two- to threefold improved affinity for both galectins relative to the parent disaccharides Lac (entries 4–6 vs. entry 1) and 2β-OMe (entry 8 vs.

entry 7). Ferrocenes **6** and **10** had only marginally better affinity for both hgαl-1 and hgαl-3-CRD than the non-metallic phenyl and hydroxymethyl compounds **7** and **4** (entries 4 and 6 vs. entries 3 and 2), respectively. Ruthenium arene **8** was a slightly weaker binder than the corresponding phenyl analog **7** (entries 5 and 3).

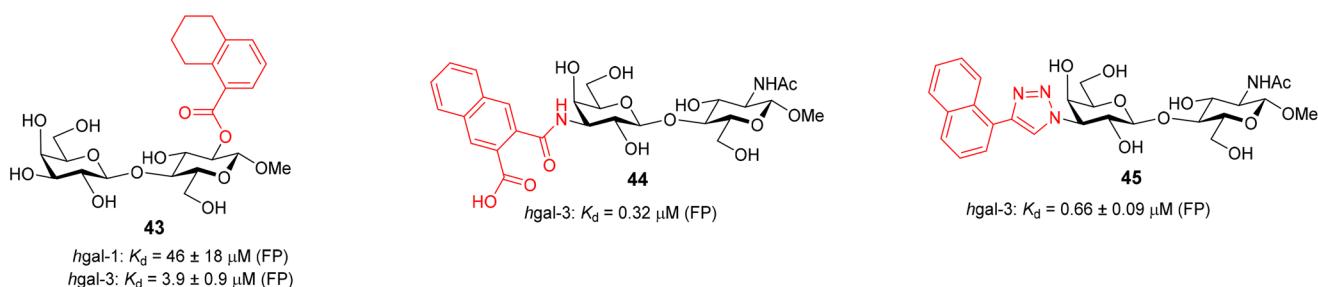
Next, we modified the 3'-position of the N-acetyllactosamine scaffold (compounds **25**–**30**, Table 1, entries 10–15), with the aim of introducing cation-π interactions between Arg144 in subsite B of the hgαl-3 CRD and the ferrocene/ruthenium arene complex attached to the 3'-position.<sup>12b,26a,50</sup> A similar approach has been used previously in the preparation of non-



**Table 2** Apparent inhibition constant  $K_i$ , relative potency rp<sup>a</sup> and selectivity values obtained from competitive fluorescence polarization for thiogalactoside analogs<sup>b</sup>

Entry	$R^3$	$R^{3'}$	Compound	$K_i$	$rp$	$K_i$	$rp$	Selectivity to
				( <i>hgal-1</i> ) ( $\mu M$ )	( <i>hgal-1</i> )	( <i>hgal-3-CRD</i> ) ( $\mu M$ )	( <i>hgal-3-CRD</i> )	<i>hgal-1</i> <sup>c</sup>
1			TD139	0.12 ± 0.01	1.00	0.008 ± 0.001	1.00	0.07
2	OH		34	1.9 ± 0.2	0.06	1.7 ± 0.4	<0.01	0.89
3			37	0.25 ± 0.04	0.48	0.033 ± 0.002	0.24	0.13
4			36	0.035 ± 0.017	3.43	0.26 ± 0.066	0.03	7.43
5			38	0.82 ± 0.08	0.15	0.72 ± 0.16	0.01	0.88
6			41	0.15 ± 0.03	0.80	0.013 ± 0.004	0.62	0.09
7			42	0.16 ± 0.01	0.75	0.026 ± 0.002	0.31	0.16

<sup>a</sup> Relative potency with respect to TD139, one of the most potent inhibitor, defined as  $rp = K_i(\text{TD139})/K_i(\text{inhibitor})$ . <sup>b</sup> Fc = ferrocenyl, [Ru] = ruthenium arene moiety. <sup>c</sup> defined as the ratio  $K_i(\text{hgal-3-CRD})/K_i(\text{hgal-1})$ .



**Fig. 3** Structures of previously reported galectin inhibitors 43–45 and their affinities to *hgal-1* and *hgal-3* from FP.<sup>26a,49,50</sup> The affinity of 44 and 45 to *hgal-1* was not reported.

organometallic *hgal-3* inhibitors<sup>7d</sup> and resulted in affinities up to the high nanomolar range (see *N*-acetyllactosaminides 44 and 45 for examples, Fig. 3). Interestingly, the affinity for *hgal-1* has not been previously determined for 3'-modified lactosamines. We found that 3-fluorophenyltriazole 28 (entry 12) and ruthenium arene 27 (entry 15) bound *hgal-3-CRD* approximately 7- and 5-fold more strongly than *hgal-1*, respectively, whereas the remaining 3'-modified lactosamine analogs displayed comparable or only 2- to 3-fold higher affinities for *hgal-3-CRD* than for *hgal-1* despite the absence of an arginine residue in subsite B of *hgal-1*.

The 3'-hydroxymethyltriazole 29 (entry 10) bound both galectins better than the unmodified 2 $\beta$ -OMe due to the interaction of the triazole ring with the galectins.<sup>7d</sup> Substituting the hydroxymethyl group with phenyl in 26 significantly improved the affinity to the single-digit micromolar range. In agreement with the previous observations made for thiogalactosides,<sup>14</sup> substituting fluorine for 3-hydrogen in the phenyl moiety of the analog 28 (entry 12) further increased the affinity, especially for *hgal-3-CRD*, which reached nanomolar values. Replacing the phenyl with ferrocene (26 → 25, entry 13) resulted in an affinity comparable to that of the phenyl ana-



logue **26**, but for the first time in our series, the selectivity was very slightly tilted in favor of *hgal-1*. Substituting the bulky phthalimide in **30** (entry 14) for the acetamide in **25** (entry 13) further improved binding to *hgal-3-CRD*, producing the best inhibitor among our LacNAc- and Lac-based metal organocomplexes, probably owing to simultaneous interactions with subsites B and E. Disappointingly, the ruthenium arene complex **27** was a weaker inhibitor of both galectins compared with its ferrocene and phenyl counterparts **25** and **26**, respectively (entries 15 vs. 13 and 11).

Some thiogalactoside-based inhibitors, such as compound **TD139**, are among the most potent *hgal-1* and -3 inhibitors.<sup>14</sup> Also in this study, inhibitors resulting from the attachment of an organometallic substituent to the thiogalactoside scaffold outperformed (Table 2) Lac- and LacNAc-derived inhibitors. Inhibitor **TD139** (entry 1) was more selective for *hgal-3-CRD* in our hands than originally reported. While the affinity for *hgal-3-CRD* ( $K_i = 0.008 \pm 0.001 \mu\text{M}$ ) was close to the reported value ( $K_i = 0.014 \pm 0.003 \mu\text{M}$ ),<sup>14</sup> the affinity for *hgal-1* was one order of magnitude lower ( $K_i = 0.12 \pm 0.01 \mu\text{M}$ ) than originally reported ( $K_i = 0.012 \pm 0.003 \mu\text{M}$ ).<sup>14</sup> However, a value very close to ours was reported by Zetterberg *et al.* this year ( $K_i = 0.109 \pm 0.022 \mu\text{M}$  as determined by FP).<sup>13b</sup> Symmetrically substituted phenyltriazolyl-thiogalactoside **37** was a selective *hgal-3-CRD* inhibitor, although less potent than **TD139** because it lacked the fluorine-galectin interactions contributing to the high binding affinity of **TD139** (Table 2, entries 1 and 3).<sup>14</sup> Substituting both planar phenyls in **37** with ferrocene to give **36** resulted in a significant drop in affinity toward *hgal-3-CRD* (almost by an order of magnitude, entries 3 and 4) and an improvement in affinity toward *hgal-1*. As a result, diferrocene **36** was the best and most selective organometallic inhibitor of *hgal-1* in the series, because it bound *hgal-1* 7-fold more strongly than *hgal-3-CRD*, surpassing not only the diphenyl analog **37** (entries 3 and 4, 7-fold increase in affinity for *hgal-1*) but also the inhibitor **TD139** (entries 1 and 4, 3-fold increase), despite the absence of a fluorine substituent. The asymmetric monoferrocene complex **34** exhibited only micromolar affinity for both galectins (entry 2). The asymmetric ferrocene-phenyltriazole complexes **41** and **42** showed a good *hgal-3-CRD* selectivity (entries 6 and 7), although **42** bound *hgal-3-CRD* 3-fold more weakly than **TD139**. Ruthenium arene complex **38** (entry 5) displayed a significantly lower affinity for both galectins compared with its phenyl and ferrocenyl counterparts **37** and **36**, respectively, and was not further investigated.

**Intrinsic tryptophan fluorescence.** We determined the binding affinity expressed as a dissociation constant ( $K_d$ , Table 3) for selected ligands **6**, **19**, **25**, **27**, **28**, **36**, **37**, **41**, and **42** by an orthogonal method that utilizes ligand-induced changes in the intrinsic fluorescence of tryptophan residues.<sup>51</sup> Inhibitors **6**, **19**, and **25** (entries 3–5) represent the Lac and LacNAc scaffolds modified at the 1-, 2-, and 3'-positions, respectively. Inhibitor **36** was selected because it is the most potent and selective *hgal-1* inhibitor in the whole series, whereas disaccharides **27**, **28**, **37**, **41**, and **42** were chosen because of their good selectivity in inhibition of *hgal-3-CRD* as

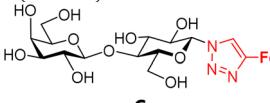
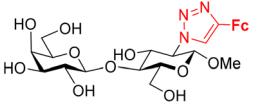
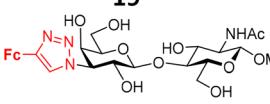
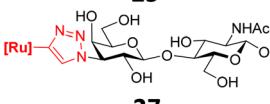
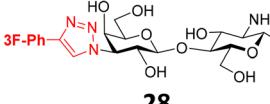
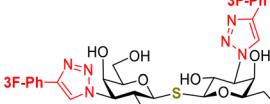
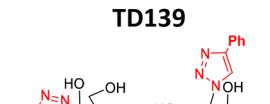
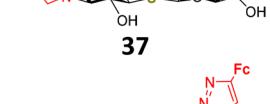
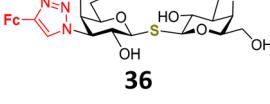
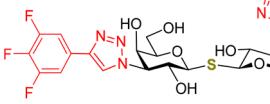
determined by fluorescence anisotropy. For comparison, the affinity of the endogenous ligands Lac and LacNAc and the known inhibitor **TD139** were also measured (Table 3, entries 1, 2, and 8). Unlike the competitive FP assay and ITC, which employed *hgal-3-CRD* to avoid *hgal-3* oligomerization, this method has been also successfully applied to full-length *hgal-3*.<sup>51a</sup>

In all cases, the measured  $K_d$  to *hgal-3-CRD* was slightly lower than that obtained for full-length *hgal-3*; however, the difference did not appear significant in most cases, and the affinity trends were similar for both *hgal-3* and *hgal-3-CRD*. Also, the values for disaccharides determined by the intrinsic tryptophan fluorescence assay and by the FP assay were comparable and did not differ by more than a factor of three in most cases. A more significant difference between the two method was found for the potent *hgal-1* inhibitor in the series **36** (Table 3, entry 10). The intrinsic tryptophan fluorescence assay indicated a higher affinity and selectivity of diferrocene **36** for *hgal-1* than FP: Complex **36** bound *hgal-1* 50-fold more strongly than *hgal-3*, and 40-fold more strongly than *hgal-3-CRD*, whereas the selectivity using the FP assay was lower ( $K_i(\text{hgal-3-CRD})/K_i(\text{hgal-1}) = 7.4$ ). The intrinsic tryptophan fluorescence assay also confirmed the selectivity of disaccharides **19**, **27**, **28**, **37**, **41**, **42** and **TD139** towards *hgal-3*. In contrast to the FP and ITC assays (see Table 4 for the ITC data), there was practically no difference between disaccharides **41** and **42** in binding to *hgal-3*, as both had low nanomolar affinity (Table 3, entries 11 and 12). This resulted in the 3-fluorophenyl analog **42** being a more selective *hgal-3* inhibitor than the trifluorophenyl analog **41**, again in contrast to the FP results (Table 2, entries 6 and 7). Since these potent *hgal-3* inhibitors **41** and **42** and *hgal-1* inhibitor **36** carry an electroactive ferrocene moiety, we envision their application in electrochemical sensing of galectin inhibition. The commercial inhibitor **TD139** showed 4-fold higher *hgal-3* affinity and 2-fold higher *hgal-3* selectivity in the tryptophan fluorescence assay (Table 3, entry 8) than in the FP assay (Table 2, entry 1). Discrepancies between the two assays can be attributed to the different bioanalytical principles on which the assays are based. Importantly, both assays confirmed that the affinity and selectivity for *hgal-1* were significantly improved by insertion of two ferrocene-triazole motifs into the thiogalactoside scaffold to yield a potent *hgal-1* inhibitor **36**.

**Chronopotentiometric stripping analysis.** We tested the formation of *hgal-1* complex with selected ligands using chronopotentiometric stripping analysis (CPS),<sup>52</sup> in which proteins yield peak H. The structural transition between native and denatured forms of protein can be observed in dependence of peak H on stripping current  $I_{\text{STR}}$  (Fig. 4). The obtained result provides information whether the individual substances stabilize or destabilize the protein structure, which is manifested by a structural transition shift to lower or higher stripping current values.<sup>52</sup> *Hgal-1* yielded a structural transition between  $I_{\text{STR}}$  of  $-17.5 \mu\text{A}$  and  $-30 \mu\text{A}$  with an  $I_{\text{STR}1/2}$  value<sup>53,54</sup> about  $-25 \mu\text{A}$ . The structural transition was accompanied by the splitting of a single peak H into two peaks characteristic of the



Table 3 Dissociation constant  $K_d$  values obtained using intrinsic tryptophan fluorescence

Entry	Compounds	$K_d$ ( <i>hgal-1</i> ) ( $\mu\text{M}$ )	$K_d$ ( <i>hgal-3</i> ) ( $\mu\text{M}$ )	$K_d$ ( <i>hgal-3-CRD</i> ) ( $\mu\text{M}$ )	Selectivity to <i>hgal-1</i> <sup>a</sup>
1	<b>1</b> (Lac)	$244 \pm 29$	$297 \pm 28$	$142 \pm 23$	1.2
2	<b>2</b> (LacNAc)	$84 \pm 13$	$80 \pm 15$	$45 \pm 12$	0.9
3		$193 \pm 21$	$49 \pm 8$	$30 \pm 2$	0.5
4		$52 \pm 4$	$5 \pm 1$	$2.9 \pm 0.3$	0.1
5		$1.8 \pm 0.5$	$13 \pm 2$	$8 \pm 1$	7.2
6		$84 \pm 12$	$10 \pm 1$	$5.2 \pm 0.5$	0.12
7		$2.2 \pm 0.2$	$0.15 \pm 0.03$	$0.08 \pm 0.02$	0.07
8		$0.08 \pm 0.01$	$0.0023 \pm 0.0004$	$0.0021 \pm 0.0004$	0.03
9		$0.12 \pm 0.02$	$0.031 \pm 0.004$	$0.012 \pm 0.001$	0.26
10		<b><math>0.010 \pm 0.001</math></b>	<b><math>0.50 \pm 0.04</math></b>	<b><math>0.40 \pm 0.10</math></b>	50
11		$0.035 \pm 0.002$	$0.0067 \pm 0.0006$	$0.0040 \pm 0.0005$	0.19
12		$0.11 \pm 0.01$	$0.0044 \pm 0.0008$	$0.0038 \pm 0.0006$	0.04

<sup>a</sup> Defined as the ratio  $K_d(\text{hgal-3})/K_d(\text{hgal-1})$ .

Table 4 Thermodynamic parameters for binding to *hgal-1* and *hgal-3*-CRD obtained by ITC at  $T = 298$  K

Entry	Compound	$K_d$ ( $\mu\text{M}$ )	$\Delta G$ (kcal mol $^{-1}$ )	$\Delta H$ (kcal mol $^{-1}$ )	$-T\Delta S$ (kcal mol $^{-1}$ )	$n$	Selectivity to <i>hgal-1</i> <sup>a</sup>
							<i>hgal-1</i>
1	TD139 <sup>b</sup>	$0.220 \pm 0.05$	$-9.1 \pm 0.2$	$-16.7 \pm 0.4$	$7.6 \pm 0.4$	$0.96 \pm 0.04$	0.31
2	TD139	$0.279 \pm 0.001$	$-8.9 \pm 0.1$	$-16.9 \pm 0.4$	$8.0 \pm 0.4$	$1.03 \pm 0.04$	0.07
3	37	$0.288 \pm 0.006$	$-8.9 \pm 0.1$	$-9.6 \pm 0.4$	$0.7 \pm 0.4$	$1.04 \pm 0.02$	0.33
4	36	$0.258 \pm 0.004$	$-9.0 \pm 0.1$	$-12.8 \pm 1.4$	$3.8 \pm 1.5$	$1.06 \pm 0.07$	3.6
5	41	$0.340 \pm 0.019$	$-8.9 \pm 0.1$	$-10.2 \pm 0.1$	$1.3 \pm 0.1$	$1.03 \pm 0.00$	0.07
6	42	$0.366 \pm 0.031$	$-8.8 \pm 0.1$	$-9.5 \pm 0.1$	$0.6 \pm 0.1$	$1.03 \pm 0.02$	0.18
7	TD139 <sup>b</sup>	$0.068 \pm 0.01$	$-9.9 \pm 0.4$	$-18.2 \pm 0.8$	$8.3 \pm 0.9$	$0.96 \pm 0.01$	—
8	TD139	$0.019 \pm 0.003$	$-10.6 \pm 0.1$	$-16.4 \pm 0.4$	$5.8 \pm 0.3$	$1.08 \pm 0.01$	—
9	37	$0.096 \pm 0.005$	$-9.6 \pm 0.1$	$-11.2 \pm 0.3$	$1.7 \pm 0.2$	$1.03 \pm 0.03$	—
10	36	$0.923 \pm 0.040$	$-8.3 \pm 0.1$	$-12.7 \pm 0.2$	$4.4 \pm 0.1$	$1.08 \pm 0.02$	—
11	41	$0.023 \pm 0.006$	$-10.5 \pm 0.2$	$-14.0 \pm 1.3$	$3.6 \pm 1.5$	$1.03 \pm 0.04$	—
12	42	$0.066 \pm 0.010$	$-9.8 \pm 0.1$	$-12.0 \pm 0.2$	$2.3 \pm 0.4$	$1.01 \pm 0.00$	—

<sup>a</sup> Defined as the ratio  $K_d(hgal-3)/K_d(hgal-1)$ . <sup>b</sup> Taken from ref. 12b.

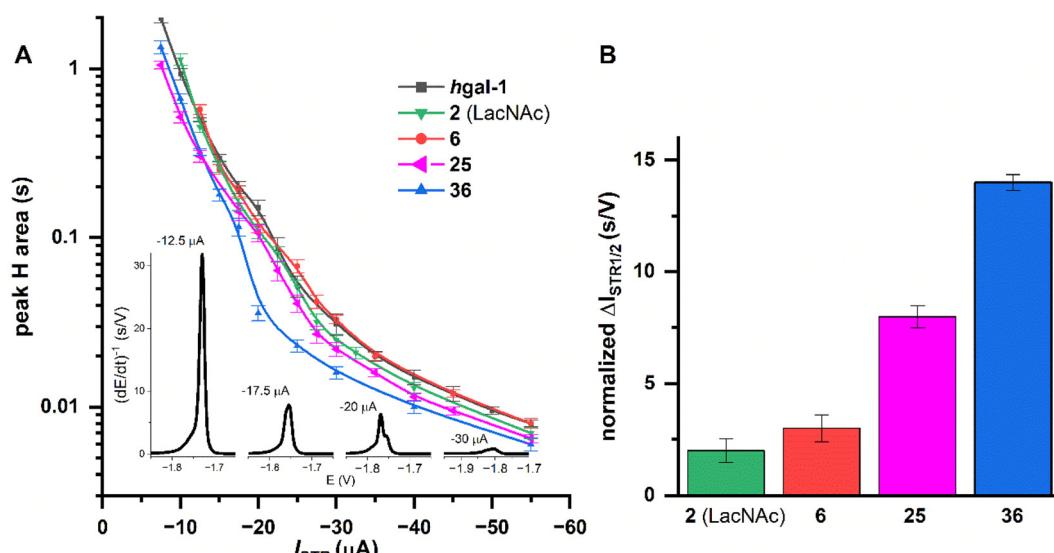


Fig. 4 (A) The dependence of peak H area on stripping current  $I_{STR}$  for free *hgal-1* (grey) and that in complex with compounds 2 (green), 6 (red), 25 (magenta), and 36 (blue). Inset: peak H height of free *hgal-1* measured using different stripping currents. A double peak of structural transition can be seen at  $-20 \mu\text{A}$ . (B)  $\Delta I_{STR1/2}$  values of *hgal-1* complexes normalized<sup>52b</sup> to free *hgal-1*.

native and denatured *hgal-1* (Fig. 4A, inset). Interaction with LacNAc (2) and compounds 6, 25, and 36 shifted the structural transition characterized by  $I_{STR1/2}$  to lower values, indicating the stabilization of *hgal-1* molecules against the effect of the electric field. The average values of this structural transition shift for *hgal-1* in complex with disaccharides compared to those of free *hgal-1* are in good agreement with the results

(binding affinities) obtained by other methods, giving the lowest  $\Delta I_{STR1/2}$  value for the strongest binder 36 (Fig. 4B). However, the determination of  $I_{STR1/2}$  for *hgal-1* interaction with compounds 2 and 6 is within the experimental limits of the CPS method.

**Isothermal titration microcalorimetry (ITC).** Ferrocene-modified thiogalactosides emerged as the most potent

organometallic inhibitors tested in this study. To assess how the substitution of ferrocene for the phenyl and fluorophenyl groups affects the thermodynamics of binding in the thiogalactoside series, we determined the thermodynamic binding parameters (Table 4 and Fig. 5) of five TDG analogs, three symmetrically substituted thiogalactoside-based inhibitors, namely bis(3-fluorophenyl) inhibitor **TD139**, and its diphenyl and diferrocene analogs **37** and **36**, and two asymmetrically substituted thiogalactosides, trifluorophenyl-ferrocenyl inhibitor **41** and 3-fluorophenyl-ferrocenyl analog **42**. We used *hgal*-3-CRD instead of complete *hgal*-3 to minimize complications resulting from oligomerization of *hgal*-3.<sup>16</sup> The ITC measurement yielded lower affinities than FP and intrinsic tryptophan fluorescence, but this phenomenon has been reported previously for galectin inhibitors including **TD139**.<sup>12b</sup> The data for binding of **TD139** to *hgal*-1 were essentially identical to those recently reported (Table 4, entries 1 and 2),<sup>12b</sup> whereas the binding affinity for *hgal*-3-CRD was approximately 3.6-fold higher in our hands than that reported<sup>12b</sup> but still comparable (entries 7 and 8). The ITC assay confirmed the selectivity of the diferrocene inhibitor **36** towards *hgal*-1, albeit with a lower selectivity value of 3.6 (entries 4 and 10), than that determined by FP (7.4) and intrinsic tryptophan fluorescence (40). ITC also confirmed the *hgal*-3 selectivity of the asymmetrically substituted thiogalactosides **41** and **42**, which exhibited selectivity values comparable to those obtained by FP (entries 5, 6, 11, and 12).

The ITC data confirmed that the binding affinity for *hgal*-3-CRD decreased when moving from the bis(fluorophenyl) analog **TD139** to the diphenyl analog **37** and the diferrocene analog **36** (Table 4, entries 8–10). This trend was consistent with the FP assay. Compared to diphenyl analog **37** (entry 3), bis(fluorophenyl) inhibitor **TD139** (entry 2) displayed a higher

binding enthalpy to *hgal*-1 (−16.9 vs. −9.6 kcal mol<sup>−1</sup>) accompanied by a significant increase in the entropic penalty (8.0 vs. 0.7 kcal mol<sup>−1</sup>), leading to a perfect enthalpy-entropy compensation with a negligible change in affinity for *hgal*-1. Binding of **TD139** to *hgal*-3-CRD also showed a gain in enthalpy relative to diphenyl analog **37** (entries 8 and 9, −16.4 vs. −11.2 kcal mol<sup>−1</sup>). However, the increased entropic penalty (5.8 vs. 1.7 kcal mol<sup>−1</sup>) did not fully compensate for this gain, resulting in a higher overall affinity of **TD139** to *hgal*-3-CRD. Thus, the ITC data suggests that while the fluorine substituent in **TD139** interacts with the protein backbone in both galectins,<sup>12b</sup> it produces a significant gain in affinity only in the case of *hgal*-3-CRD. This is probably due to better protein preorganization.

For the diferrocene complex **36**, the higher enthalpy of binding to *hgal*-1 compared to its diphenyl analog **37** (−12.8 vs. −9.6 kcal mol<sup>−1</sup>) was completely compensated for by the increased entropic penalty of **36** (3.8 vs. 0.7 kcal mol<sup>−1</sup>) leading to virtually identical binding affinities of compounds **36** and **37** for *hgal*-1 (entries 3 and 4). In contrast, the entropic penalty of diferrocene **36** relative to its diphenyl counterpart **37** (4.4 vs. 1.7 kcal mol<sup>−1</sup>) exceeded the enthalpy gain (−12.7 vs. −11.2 kcal mol<sup>−1</sup>) in binding to *hgal*-3-CRD, resulting in a significantly lower affinity of **36** for this galectin (entries 9 and 10), and hence selectivity for *hgal*-1 over *hgal*-3. We interpret these ITC data in terms of an entropically unfavorable structural transition that *hgal*-3-CRD has to undergo to accommodate the bulky ferrocene moiety, which outweighs any attractive interaction. Asymmetrically substituted mono-ferrocene analogs **41** and **42** showed binding behavior similar to **TD139**, with significantly higher affinity for *hgal*-3-CRD than for *hgal*-1 (Table 4, entries 5, 6, 11, and 12), suggesting that *hgal*-3-CRD accommodates one ferrocene group in inhibitors **41** or **42**.

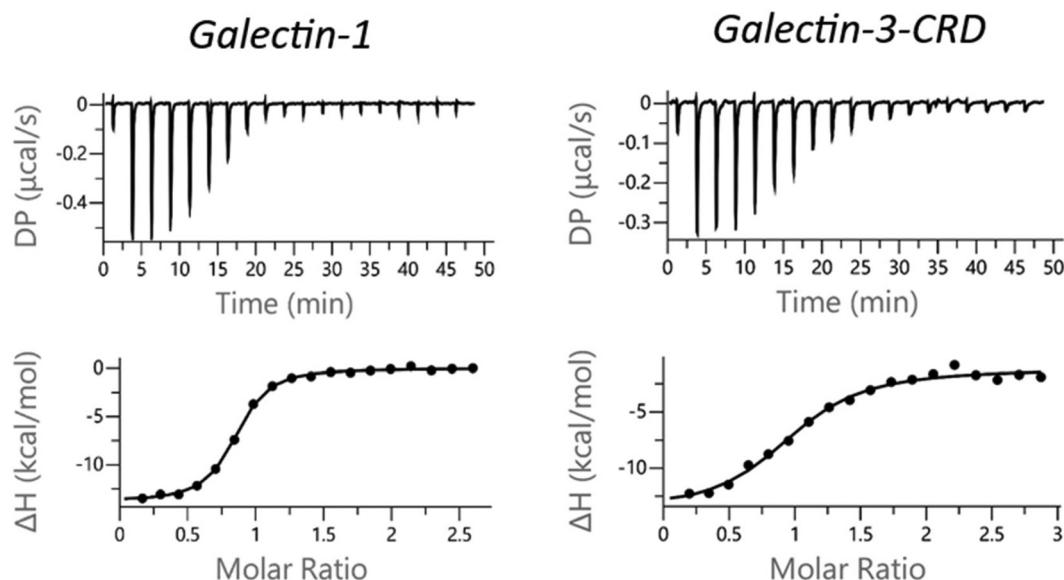


Fig. 5 Thermograms and binding isotherms obtained by isothermal titration calorimetry for the interaction of **36** with *hgal*-1 and *hgal*-3-CRD.

more effectively than two ferrocenes in compound 36. This was further investigated by computational modeling and  $^{19}\text{F}$  NMR spectroscopy.

**Computational modelling of ligand-galectin interactions.** The asymmetric monoferrocene complex 34 exhibited only a micromolar and essentially identical affinity for both galectins (Table 2, entry 2). Because the symmetrical difterrocene complex 36 exhibited a 54-fold increase in affinity toward *hgal*-1 but only a 7-fold increase toward *hgal*-3-CRD compared to 34, it seems likely that the symmetrically placed ferrocene moieties in inhibitor 36 can interact more efficiently with subsites A/B and E in *hgal*-1 than in *hgal*-3-CRD. To evaluate possible interactions between the ferrocene group and these subsites and to compare them with those of the planar fluorophenyl group, we attempted to estimate the binding energies of mono- and difterrocene analogs 34 and 36, respectively, and fluorophenyl ferrocene analog 42 for *hgal*-1 and *hgal*-3-CRD using molecular modeling tools (Table 5). For the calculation, we used a simple model based on a combination of molecular dynamics of the ligand-galectin complex and subsequent estimation of the ligand binding enthalpy for a set of sampled MD geometries. MD simulations were performed in Desmond,<sup>55,56</sup> and binding enthalpies were calculated at the semiempirical level in MOPAC<sup>57</sup> (see ESI† for a detailed method description). Table 5 summarizes the computationally estimated binding enthalpies for disaccharides 34, 36, and 42. Because of the asymmetric character of compounds 34 and 42, two binding modes were considered. In the first binding mode, the ferrocene moiety is positioned in subsites A and B

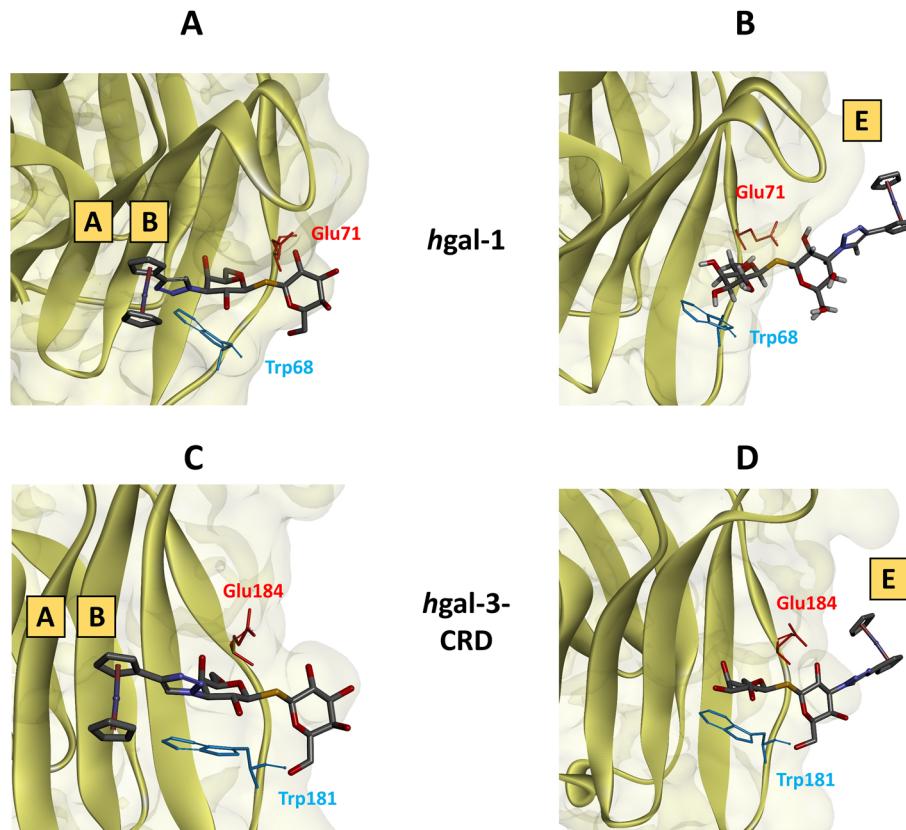
of a galectin. In the second mode, it resides in the E subsite (Fig. 6 and Fig. S80–S83 in ESI†). Only one binding mode was considered for difterrocene 36 (Fig. S78–S79 in ESI†) because of its symmetry. It is obvious from the comparison of the calculated  $\Delta H_{\text{bind}}$  (Table 5, entries 3–5) and experimentally determined  $\Delta H$  for difterrocene 36 (Table 4, entries 4 and 10) and mono-ferrocene 42 (entries 6 and 12) that the calculation overestimates binding enthalpies (see discussion in ESI, pp. S81–S82†).

Notwithstanding the aforementioned inaccuracies and the absence of the entropic penalty from the calculations, the determined binding enthalpies correlated consistently with the experimentally determined affinities. Accordingly, complex 36 demonstrated the highest binding enthalpy of all calculated disaccharides in binding to *hgal*-1 ( $-34.1\text{ kcal mol}^{-1}$ ), along with relatively strong binding to *hgal*-3-CRD ( $-30.4\text{ kcal mol}^{-1}$ ). These results are consistent with the ITC data, where complex 36 displayed a similar enthalpy change in binding to both galectins, whereas the decrease in affinity to *hgal*-3-CRD stemmed from a higher entropic penalty (Table 4, entries 4 and 10). Notably, complexes 34 and 42 exhibited significantly higher binding enthalpies in binding to *hgal*-1 when the ferrocene moiety occupied subsites A and B. Conversely, the preferred binding mode of these complexes to *hgal*-3-CRD positioned the ferrocene moiety in subsite E. Thus, the MD simulation suggests that *hgal*-1 accommodates the bulky ferrocene moiety in subsites A and B more efficiently than *hgal*-3-CRD.

**Monitoring of the  $^{19}\text{F}$  [ $^1\text{H}$ ] resonances of the 3-fluorophenyl moiety.** The presence of the fluorine atom in compounds 28,

**Table 5** Binding enthalpy calculated from MD simulation

Entry	Compounds	Binding mode (subsites in which the ferrocene moiety is positioned)	$\Delta H_{\text{bind}}$ (kcal mol $^{-1}$ ) <i>hgal</i> -1	$\Delta H_{\text{bind}}$ (kcal mol $^{-1}$ ), <i>hgal</i> -3-CRD
1				
2		A and B (Fig. 6A and C) E (Fig. 6B and D)	-16.9 -8.4	-16.3 -25.8
3		A and B, and E (Fig. S78–S79 in ESI†)	-34.1	-30.4
4		A and B (Fig. S80 and S82 in ESI†) E (Fig. S81 and S83 in ESI†)	-32.3 -27.3	-30.4 -32.9
5				



**Fig. 6** Geometries of compound 34 bound to *hgal-1* or -3-CRD derived from molecular dynamics calculations showing characteristic amino acid residues (Glu71 and Trp68 for *hgal-1*, Glu184 and Trp181 for *hgal-3*-CRD) interacting with the hexopyranose rings. (A) Ferrocene moiety in subsites A and B of *hgal-1*. (B) Ferrocene moiety is in subsite E of *hgal-1*. (C) Ferrocene moiety in subsites A and B of *hgal-3*-CRD. (D) Ferrocene moiety in subsite E of *hgal-3*-CRD.

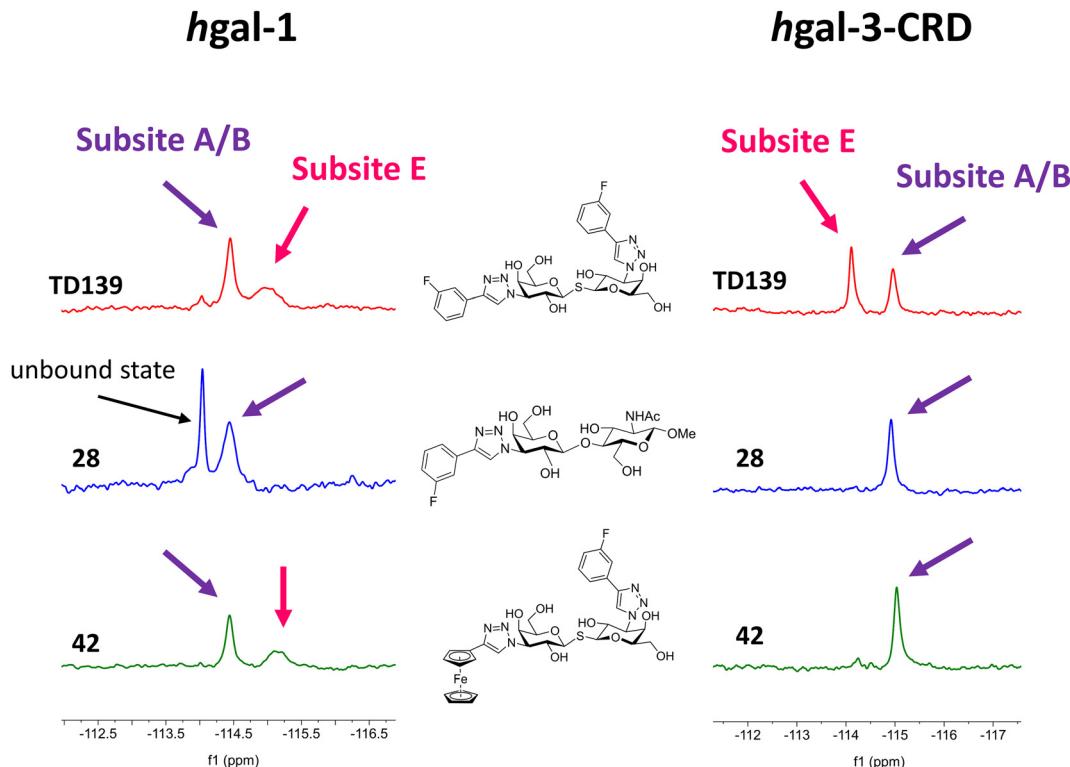
42, and TD139 enabled us to use sensitive  $^{19}\text{F}\{^1\text{H}\}$  NMR experiments to map the position of the 3-fluorophenyl-triazole moiety in both galectins. All investigated compounds TD139, 28, and 42 showed a single sharp  $^{19}\text{F}$  resonance close to  $-114$  ppm in the unbound state (Fig. S84 in ESI $\dagger$ ). According to a previous report, the interaction of TD139 with either *hgal-1* or *hgal-3*-CRD resulted in similar  $^{19}\text{F}$  NMR spectra revealing multiple resonances of the galectin-bound states in slow chemical exchange. $^{12b}$  In our hands, measuring the solution of TD139 with the excess of either *hgal-1* or *hgal-3*-CRD relative to the ligand produced two bound state  $^{19}\text{F}$ -NMR resonances (Fig. 7, red spectra). Repetition of the experiment with an excess of TD139 relative to the galectins showed an additional resonance of unbound TD139, distinct from the bound state signals (Fig. S85 and S88 in ESI $\dagger$ ).

To facilitate the interpretation of the  $^{19}\text{F}\{^1\text{H}\}$  NMR spectra of protein-bound TD139, we performed the same NMR experiments with 3'-fluorophenyl *N*-acetyllactosamine 28. This ligand showed only one  $^{19}\text{F}$  resonance of its bound state upon addition of either *hgal-1* or *hgal-3*-CRD (Fig. 7 blue spectra, Fig. S86 and S89 in ESI $\dagger$ ). Because disaccharide 28 is an unsymmetrical LacNAc analog, the galactopyranoside ring can only occupy subsite C with the 3'-fluorophenyl ring pointing

toward subsites A/B. $^{7b,12a,50,58}$  Therefore, its  $^{19}\text{F}$  resonance was assigned to the fluorophenyl-triazole group located in subsites A/B. The chemical shift of the bound state  $^{19}\text{F}$  resonance in compound 28 closely matched one of the two resonances of bound TD139 in the presence of either *hgal-1* or *hgal-3*-CRD (Fig. 7 red vs. blue spectra). Therefore, this resonance of TD139 was also assigned to the fluorophenyl-triazole group positioned in subsites A/B. Consequently, the remaining broad signal at  $\delta = -115.01$  ppm of TD139 bound to *hgal-1* (Fig. 7, *hgal-1*, pink label) was assigned to the fluorophenyl moiety occupying subsite E. Similarly, the more downfield-shifted  $^{19}\text{F}$  resonance of TD139 bound to the *hgal-3*-CRD (Fig. 7, *hgal-3*-CRD, pink label) was also assigned to the fluorophenyl-triazole moiety in subsite E. It should be noted that this assignment is based on the assumption that the differences between the compounds 28 and TD139 do not affect the interactions of the fluorine atom with the binding subsites A/B or E and thus its chemical shift in these subsites.

In principle, the ferrocene moiety in compound 42 can occupy subsites A/B, or subsite E in both galectins, leading to two distinct binding modes (Fig. 6). Indeed, the  $^{19}\text{F}\{^1\text{H}\}$  NMR spectrum of 42 in the presence of *hgal-1* showed two bound state  $^{19}\text{F}$  NMR signals (Fig. 7, *hgal-1*, green spectrum, Fig. S87





**Fig. 7**  $^{19}\text{F}\{^1\text{H}\}$  NMR spectra of ligands **28**, **42**, and **TD139** in the presence of *hgal-1* or *hgal-3-CRD*, pink arrows indicate resonances of fluorine in subsite E, purple arrows indicate resonances of fluorine in subsites A/B. Except for the combination **28** with *hgal-1*, all these spectra were acquired using an excess of the galectin over the ligand.

in ESI $^{\dagger}$ ), whose chemical shifts matched the two bound state resonances of **TD139** with *hgal-1* (Fig. 7, *hgal-1*, red spectrum). Therefore, the fluorophenyl group in **42** occupies both subsites

A/B and subsite E during recognition by *hgal-1*, and consequently the ferrocene group is likely to occupy these two subsites as well. Conversely, disaccharide **42** exhibited only a

**Table 6**  $IC_{50}$  [ $\mu\text{M}$ ] values obtained by the MTT assay after 72 h treatment of cells with selected compounds $^a$

Entry	Compound	A2780	SK-OV-3	MDA-MB-231	HEK-293
1	<b>6</b> (Lac1 $\beta$ -Fc')	$85 \pm 8$	$>200$	$183 \pm 35$	$>200$
2	<b>8</b> (Lac1 $\beta$ -Ru')	$69 \pm 12$	$>200$	$>200$	$>200$
3	<b>10</b> (Lac1 $\beta$ -OCH <sub>2</sub> CH <sub>2</sub> Fc')	$116 \pm 14$	$>200$	$170 \pm 5$	$198 \pm 35$
4	<b>16</b> (LacNAC1 $\beta$ -Fc')	$>200$	$>200$	$>200$	$>200$
5	<b>19</b> (2-Fc'-Lac1 $\beta$ -OMe)	$159 \pm 54$	$>200$	$>200$	$>200$
6	<b>25</b> (3'-Fc'-LacNAC1 $\beta$ -OMe)	$>200$	$>200$	$>200$	$>200$
7	<b>27</b> (3'-Ru'-LacNAC1 $\beta$ -OMe)	$161 \pm 50$	$>200$	$>200$	$143 \pm 44$
8	<b>30</b> (3'-Fc'-LacNPhth1 $\beta$ -OMe)	$>200$	$>200$	$>200$	$>200$
9	<b>34</b> (3'-Fc'-TDG)	$90 \pm 5$	$>200$	$>200$	$>200$
10	<b>36</b> (3,3'-diFc'-TDG)	$>200$	$>200$	$>200$	$151 \pm 11$
11	<b>38</b> (3,3'-diRu'-TDG)	$>200$	$>200$	$125 \pm 21$	$>200$
12	<b>41</b> (3-F <sub>3</sub> Ph-3'-Fc'-TDG)	$168 \pm 8$	$>200$	$>200$	$152 \pm 6$
13	<b>42</b> (3-3FPh-3'-Fc'-TDG)	$>200$	$>200$	$>200$	$>200$
14	<b>cisPt</b>	$1.7 \pm 0.3$	$5.6 \pm 1.0$	$3.8 \pm 0.5$	$3.7 \pm 0.6$

<sup>a</sup> To facilitate orientation, the structures of compounds are indicated using the abbreviations Lac, LacNAC, and TDG for the disaccharides, and abbreviations Fc', Ru', 3FPh, and F<sub>3</sub>Ph for the triazole-linked moieties as defined below the Table heading.



single bound state  $^{19}\text{F}$  resonance (Fig. 7, *hgal*-3-CRD, green spectrum, Fig. S90 in ESI<sup>†</sup>) in the presence of *hgal*-3-CRD, similar to compound 28 (Fig. 7, *hgal*-3-CRD, blue spectrum), suggesting that the fluorophenyl substituent of 42 interacts only with subsites A/B during recognition by *hgal*-3-CRD. Thus, the bulky ferrocene moiety is unlikely to occupy subsites A/B in *hgal*-3-CRD and compound 42 prefers to bind *hgal*-3-CRD through the binding mode with the ferrocene moiety located in subsite E, which is consistent with computational modeling (Table 5, entries 4 and 5).

### Cytotoxicity

Ferrocene itself is not significantly toxic.<sup>20b</sup> Compounds formed by the insertion of ferrocene into bioactive compounds to enhance a particular bioactivity generally showed low toxicity.<sup>59</sup> However, we can also find examples of highly cytotoxic ferrocene compounds<sup>60</sup> including ferrocene-containing glycomimetics.<sup>61</sup> Therefore, we screened the *in vitro* cytotoxicity of the prepared organometallic hybrid inhibitors against human ovarian cancer cell lines A2780 and SK-OV-3, human triple-negative breast cancer cell line MDA-MB-231, and human embryonic kidney non-malignant cell line HEK-293. A MTT cell viability assay was performed after 72 h of treatment and cell viability was expressed as  $IC_{50}$  values (Table 6). In this initial screening, we did not attempt to correlate the cytotoxicity of inhibitors with galectin expression in the cells, which is reserved for a detailed follow-up study. The tested disaccharides showed minimal or only weak cytotoxicity towards the cancer cell lines SK-OV-3, MDA-MB-231, and the non-malignant HEK-293. Complexes 6, 8 and 34 exhibited moderate cytotoxicity towards A2780 cell line (Table 6, entries 1, 2, and 9). Notably, the potent inhibitors 36, 41 and 42 showed no significant cytotoxicity (entries 10, 12, and 13).

### Conclusion

This study reports a novel approach to the design of galectin inhibitors using disaccharides modified with ferrocenes and ruthenium arenes. We have thoroughly investigated their binding to *hgal*-1 and *hgal*-3-CRD through a series of binding assays and experiments, including fluorescence anisotropy, intrinsic fluorescence of the tryptophan residues, chronopotentiometric stripping analysis, isothermal titration microcalorimetry, molecular dynamics simulation, and the monitoring of  $^{19}\text{F}\{^1\text{H}\}$  NMR resonances. Disaccharides containing a ruthenium arene complex were found to be weaker galectin binders than the corresponding ferrocene analogs. The inhibitors interacting with the galectin subsite E *via* the ferrocene moiety (Lac and LacNAc modified at the 1- or 2-position) exhibited an increase in affinity to both galectins similar to that observed for inhibitors with planar organic arenes at the corresponding disaccharide positions. However, the attachment of ferrocene to positions 3/3' of the galactopyranoside ring significantly enhanced the affinity for both disaccharide scaffolds tested,

*N*-acetyllactosamine and thiogalactoside, and in the case of diferrocenyl-thiogalactoside 36, surpassed the performance of the inhibitors possessing planar arenes, leading to a reversal of the inhibitor selectivity in favor of *hgal*-1 over *hgal*-3.

Notably, the thiogalactoside-based diferrocene inhibitor 36 proved to be a potent and selective *hgal*-1 inhibitor whereas thiogalactoside analogs 41 and 42, which incorporate only a single ferrocene moiety, showed good affinity and selectivity for *hgal*-3 comparable to TD139. A preliminary hypothesis based on molecular dynamics simulations and the measurement of  $^{19}\text{F}\{^1\text{H}\}$  NMR resonances attributed the selectivity of inhibitor 36 to the restrictions imposed by the ferrocene moiety when binding to *hgal*-3-CRD. Further experiments to elucidate the molecular basis of the unusual affinity and selectivity of this inhibitor are underway in our group. We also envisage that the series of ferrocene-containing inhibitors with varying affinities developed in this study can be used to exploit the electrochemical activity of the ferrocene moiety for the development of an electrochemical galectin binding assay, an avenue currently under active investigation in our group.

### Author contributions

Vojtěch Hamala and Martin Kurfiřt contributed equally to this study. Vojtěch Hamala: conceptualization, investigation (synthesis), data curation and writing – original draft. Martin Kurfiřt: investigation (synthesis,  $^{19}\text{F}$  NMR experiments) and data curation. Lucie Červenková Šťastná and Jana Bernášková: investigation (NMR and HRMS analysis). Hedvika Hujerová: investigation (synthesis). Kamil Parkan: investigation (fluorescence probe synthesis). Jakub Kaminský: investigation (molecular modelling). Nina Habanová: investigation ( $^{19}\text{F}$  NMR experiments). Jaroslav Kozák and Alžběta Magdolenová: investigation (fluorescence polarization). Martin Zavřel and Jitka Holčáková: investigation (galectins cloning and purification). Tatiana Staroňová, Veronika Ostatná, Lucie Žaloudková and Aleš Daňhel: investigation (voltammetric analysis, tryptophan fluorescence assay and chronopotentiometric stripping analysis). Petr Voňka: investigation (isothermal titration microcalorimetry). Roman Hrstka: funding acquisition, investigation and supervision. Jindřich Karban: conceptualization, funding acquisition, investigation, project administration, supervision, writing – review & editing.

### Data availability

The data supporting this article have been included as part of the ESI.<sup>†</sup>

### Conflicts of interest

The authors declare no conflict of interest.



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