



Cite this: *Nanoscale*, 2024, **16**, 8533

Membrane fluidity properties of lipid-coated polylactic acid nanoparticles†

Yuanqing Gu ^a and Björn M. Reinhard *^b

Lipid coating is considered a versatile strategy to equip nanoparticles (NPs) with a biomimetic surface coating, but the membrane properties of these nanoassemblies remain in many cases insufficiently understood. In this work, we apply C-Laurdan generalized polarization (GP) measurements to probe the temperature-dependent polarity of hybrid membranes consisting of a lipid monolayer adsorbed onto a polylactic acid (PLA) polymer core as function of lipid composition and compare the behavior of the lipid coated NPs (LNPs) with that of liposomes assembled from identical lipid mixtures. The LNPs were generated by nanoprecipitation of the polymer in aqueous solutions containing two types of lipid mixtures: (i) cholesterol, dipalmitoylphosphatidylcholine (DPPC), and the ganglioside GM3, as well as (ii) dioleoylphosphatidylcholine (DOPC), DPPC and GM3. LNPs were found to exhibit more distinct and narrower phase transitions than corresponding liposomes and to retain detectable phase transitions even for cholesterol or DOPC concentrations that yielded no detectable transitions in liposomes. These findings together with higher GP values in the case of the LNPs for temperatures above the phase transition temperature indicate a stabilization of the membrane through the polymer core. LNP binding studies to GM3-recognizing cells indicate that differences in the membrane fluidity affect binding avidity in the investigated model system.

Received 18th December 2023,
Accepted 31st March 2024

DOI: 10.1039/d3nr06464f

rsc.li/nanoscale

Introduction

Lipid-coated nanoparticles (LNPs) contain a lipid membrane assembled around a central nanoparticle (NP) core. LNPs are versatile hybrid materials whose core can either be an inorganic “hard” NP, or a polymeric “soft” NP.^{1–3} LNPs with an inorganic core have found diverse applications, for instance as drug carriers (*e.g.* silica NP core^{4–6}), in disease diagnosis and therapy (*e.g.* gold^{7–9} or iron oxide NP core^{10–12}), or as bio-sensors (*e.g.* silver NPs¹³). Lipid-coated gold NPs have also been used as mimics to study the lipid-mediated cellular interactions of enveloped viruses¹⁴ and as nanoreactors¹⁵ that localize photoreactive species in the lipid coating around the core where they experience strong electrical (E-) field enhancements. Of particular interest for biomedical applications are, however, LNPs with a biodegradable polymeric core.^{16–19} The core can be loaded with pharmaceutical compounds or con-

trast agents, and the LNPs can serve as effective delivery agents or imaging probes. LNPs mimic enveloped viruses,¹⁴ and since their surface properties derive from the lipid coating, they can have fortuitous stealth properties. Most phospholipids, for instance, are zwitterionic which facilitates the assembly of LNPs with low affinity for non-specific biomolecule adhesion.^{20,21} Furthermore, bio-active lipids can also be utilized to provide LNPs with specific targeting functionalities. Ganglioside GM3 functionalized LNPs (GM3-LNPs), for instance, were shown to facilitate the selective targeting of CD169-myeloid cells and were developed into nanocarriers for antiretrovirals (ARVs) that target viral niches in macrophages and dendritic cells.^{22,23}

Lipid-coated polymeric nanoparticles derive important surface properties from the lipid membrane, but the physico-chemical properties of this membrane still pose important questions and may also depend on details of the fabrication. Two main strategies are applied to generate LNPs with a polymer core: the single step and the two-step assembly.^{18,19} In the two-step strategy, polymeric nanoparticles are generated first and then combined with pre-formed liposomes to form LNPs in which the polymeric core is encapsulated by a lipid bilayer membrane through direct hydration, sonication, or extrusion.^{17–19,24,25} In the single-step strategy LNPs are formed by nanoprecipitation of hydrophobic polymers in a lipid-containing aqueous solution or by emulsification–solvent–evaporation (ESE) in the presence of lipids as surfactants.^{17–19,26} Both

^aDepartment of Chemistry and The Photonics Center, Boston University, Boston, MA 02215, USA. E-mail: yqgu@bu.edu

^bDepartment of Chemistry and The Photonics Center, Boston University, Boston, MA 02215, USA. E-mail: bmr@bu.edu

†Electronic supplementary information (ESI) available: Additional results including cholesterol assay (Fig. S1), darkfield/fluorescence colocalization (Fig. S2), DSC thermograms (Fig. S3), comparison of GP measurements of LNPs and PLA NP (Fig. S4), GP measurements with and without background correction (Fig. S5), and representative tangent constructions for T_{on} and T_{off} determination (Fig. S6). See DOI: <https://doi.org/10.1039/d3nr06464f>



single step strategies have in common that lipids form around the nascent core due to hydrophobic interactions between the hydrocarbon chains of the lipids and the polymer. Consequently, the lipid membrane formed under these conditions is expected to contain a monolayer of lipids that point with their hydrophobic tails towards the polymer core and with their hydrophilic head towards the aqueous phase.^{7,8,13} This membrane is best described as a hybrid membrane, and its structure and key properties, such as lipid packing and membrane fluidity require further investigation. Characterizing key properties of the hybrid membrane of LNPs is the focus of this study.

In the case of liposomes recent studies have shown that membrane fluidity is a factor that affects binding to²⁷ and uptake²⁸ by target cells. The potential effect of lipid order and membrane fluidity of the hybrid membrane on cell binding and uptake of LNPs is less clear. However, especially for membrane-embedded ligands that are weak monovalent binders, as in the case of GM3, and thus require multivalent interactions to increase the avidity,^{23,29,30} it stands to reason that the physical properties of the membrane affect the binding properties, for instance, through changes in the lateral mobility of the gangliosides. Structure and fluidity of membranes are frequently studied with polarity sensitive fluorescent probes, such as Laurdan or the structurally closely related but photo-physically more stable C-Laurdan^{31,32} integrated into the membrane of interest. C-Laurdan is an amphiphilic molecule whose fluorescent emission spectrum is sensitive to the lipid packing in a membrane. The dye monitors changes in the polarity in the lipid membrane at the depth of the glycerol backbone due to changes in the packing of the lipids that modulate the water content in the membrane.^{33,34} The

C-Laurdan emission peaks at 440 nm when the membrane is in a gel-phase with a relatively low concentration of water penetrating the membrane, and it shifts to 490 nm when the membrane is in a disordered fluid phase with a higher water content.^{31,32,35} Consequently, the generalized polarization (GP) defined by the following equation provides a measure for membrane polarity that correlates with membrane order and fluidity:

$$GP = \frac{I_{440} - I_{490}}{I_{440} + I_{490}} \quad (1)$$

GP values, which are independent of the local probe concentration,³⁶ range from -1 to 1 , where high GP values indicate low polarity (*i.e.* high level of membrane order) and low GP values indicate high polarity (*i.e.* low level of membrane order). GP measurements have emerged as an effective experimental strategy to characterize membrane phase transitions in cellular membranes,³⁷ liposomes^{33,38,39} and other hybrid vesicles.⁴⁰ For liposomes GP values measured with the related Laurdan dye were found to distinguish between distinct phase states. Gel-phase, fluid-phase, and intermediate phase were assigned GP value ranges of $GP > 0.55$ (gel phase), $GP < -0.05$ (fluid phase), and $-0.05 < GP < 0.55$ (intermediate liquid-ordered and -disordered phases).³⁶

In this work, we apply C-Laurdan GP measurements to probe the polarity of hybrid membranes of different compositions self-assembled around a polylactic acid (PLA) polymer core. The hybrid membranes were assembled from mixtures of **M1**: saturated dipalmitoylphosphatidylcholine (DPPC), cholesterol, and small amounts of GM3, or **M2**: unsaturated dioleoylphosphatidylcholine (DOPC), DPPC, and small amounts of GM3 (Fig. 1a and b). These lipid compositions were chosen

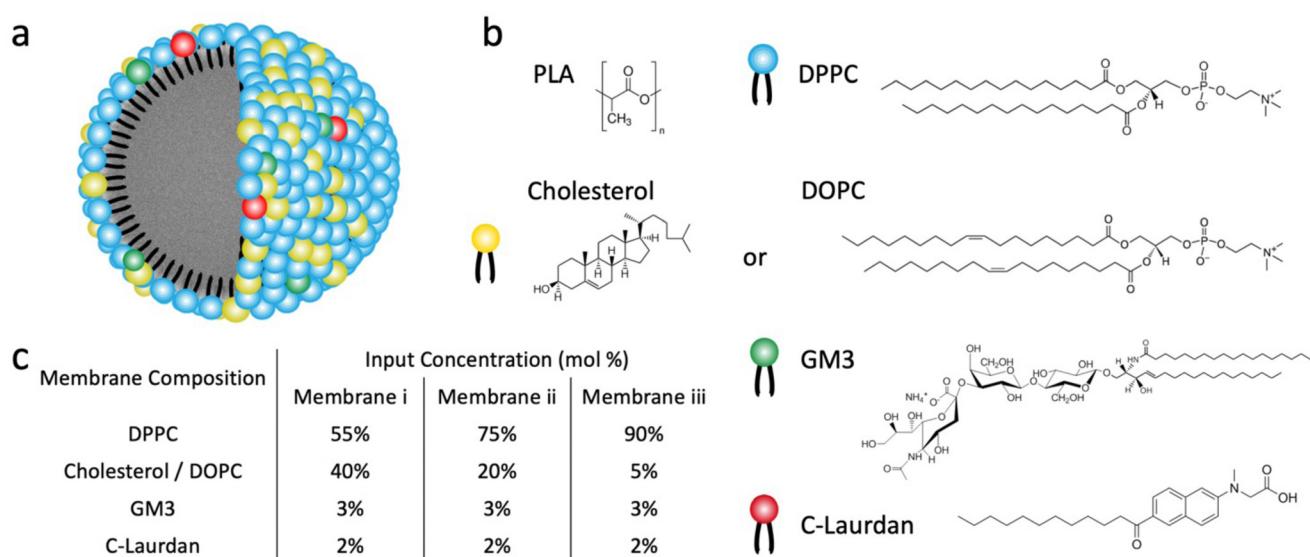


Fig. 1 Structure and composition of GM3-LNPs and liposomes. (a) Schematic drawing of the GM3-LNP structure. (b) Chemical structure of the polymer core (PLA NP) and of membrane components (DPPC, Cholesterol/DOPC, GM3, C-Laurdan). (c) The input mol% of different lipid components for three different (i–iii) membranes of type **M1** (DPPC, cholesterol, GM3, C-Laurdan) and type **M2** (DOPC, DPPC, GM3, C-Laurdan) investigated in this work.



based on prior work with conventional liposomes^{38,40-42} and are anticipated to generate hybrid membranes with different levels of lipid packing and fluidity. Small amounts of GM3 were included in the lipid mix to characterize GM3-mediated binding to CD169-expressing cells as function of GP. In all experiments, liposomes of identical composition and similar size as the LNPs were included as benchmarks to facilitate a comparison of the LNP hybrid membrane properties with those of a conventional lipid bilayer. We found that LNPs have more distinct and narrower phase transitions than corresponding liposomes. Intriguingly, LNPs were found to exhibit detectable phase transitions even for cholesterol or DOPC concentrations that yielded no detectable transitions in liposomes. Furthermore, the GP values in the high temperature limit of all investigated GM3-LNPs were higher than those of liposomes with the same composition. Together these observations evidence that the presence of the polymer core decreases the polarity in the lipid layer surrounding the NPs when compared to conventional liposomes, indicating a higher packing density of hydrophobic chains, potentially induced by the interdigitation of polymer chain fragments and lipid tails in the hybrid membrane.

Results and discussion

Fabrication and characterization of GM3-LNPs and corresponding liposomes

All LNPs used in this work were generated through dropwise addition of an organic solution of poly(lactic acid) (PLA) (poly(D, L-lactide)), Resomer R207S, into an aqueous phase containing both lipids and C-Laurdan under stirring, followed by subsequent sonication. Liposomes were fabricated following established procedures²³ through sonication of a re-hydrated lipid film that also contained C-Laurdan and subsequent extrusion. LNPs and liposomes generated with lipid mix M1 are in the following referred to as LNP_x^{CHOL} and $Liposome_x^{CHOL}$ respectively, in which x stands for the input mol% of cholesterol in the lipid mix M1. Similarly, LNPs and liposomes generated with lipid mix M2 are referred to as LNP_x^{DOPC} and $Liposome_x^{DOPC}$, in which x means the input mol% of DOPC in the lipid mix M2. In all preparations the input concentration of GM3 was maintained at 3 mol%, and C-Laurdan was kept at 2 mol%. The input concentrations of cholesterol or DOPC were 5, 20, 40 mol%, respectively, and the rest of the lipid mix was made up by DPPC (90, 75, 55 mol%) (Fig. 1c). A total of six kinds of LNPs, namely LNP_5^{CHOL} , LNP_{20}^{CHOL} , LNP_{40}^{CHOL} , LNP_5^{DOPC} , LNP_{20}^{DOPC} , LNP_{40}^{DOPC} , and corresponding six kinds of liposomes, namely $Liposome_5^{CHOL}$, $Liposome_{20}^{CHOL}$, $Liposome_{40}^{CHOL}$, $Liposome_5^{DOPC}$, $Liposome_{20}^{DOPC}$, $Liposome_{40}^{DOPC}$ were investigated in this work. Given the complexity of the assembly process, it is unclear how the input concentration relates to the actual lipid ratios in the assembled membranes. Therefore, LC-MS was applied to measure the relative lipid concentrations in the assembled liposomes and LNPs (Fig. 2a). Overall, the measured lipid concentrations followed the input concen-

trations, but the measured ratios showed some deviations from the input ratios. Notably, the difference in cholesterol concentration between LNP_{40}^{CHOL} and LNP_{20}^{CHOL} was smaller than expected based on the input concentrations. This discrepancy was independently verified by a cholesterol fluorescence quantification assay (Fig. S1†). Furthermore, the GM3 concentrations in LNP_x^{DOPC} were systematically higher than those in LNP_x^{CHOL} . Despite these deviations, the LC-MS data confirms the successful assembly of LNPs with systematic differences in cholesterol and DOPC content.

The size and ζ -potentials for LNP_x^{CHOL} and $Liposome_x^{CHOL}$ are summarized in Fig. 2b, the corresponding data for LNP_x^{DOPC} and $Liposome_x^{DOPC}$ are presented in Fig. 2c. For both LNP_x^{CHOL} and LNP_x^{DOPC} , the average hydrodynamic diameters show no significant dependence on the lipid composition. The average hydrodynamic diameters are 136 ± 1 nm for LNP_x^{CHOL} and 136 ± 2 nm for LNP_x^{DOPC} , with an average polydispersity index < 0.28 . While LNP_x^{CHOL} and LNP_x^{DOPC} have essentially identically hydrodynamic diameters, the liposomes are smaller with average hydrodynamic diameters between 97 ± 7 and 111 ± 30 nm for $Liposome_x^{CHOL}$ and between 68 ± 14 and 96 ± 20 nm for $Liposome_x^{DOPC}$ with an average polydispersity index < 0.28 . Due to its sialic acid GM3 carries a negative charge, and the fabricated LNPs and liposomes are negative with ζ -potentials in the range between -21 ± 3 mV to -12 ± 1 mV. Transmission electron microscopy (TEM) images of LNP_{40}^{CHOL} , LNP_{40}^{DOPC} and PLA NPs (without lipid membrane) treated with sodium phosphotungstate $Na_3P(W_3O_10)_4$ are shown in Fig. 2d-f. To further verify the successful assembly of a lipid membrane around the polymer NP core, we colocalized LNPs and fluorescently labelled lipid membrane with an average colocalization percentage of 73% (Fig. S2†), which represents a conservative estimate of the fraction of lipid-coated NPs due to the rapid bleaching of the fluorescence. Consistent with the lipid-wrapped polymer NP design, the DSC thermograms (Fig. S3†) of LNP_{20}^{CHOL} show two endothermic peaks, which correspond to the glass transition temperature of the polymer core and the phase transition temperature of the lipid membrane respectively.

C-Laurdan GP measurement of LNP hybrid membranes and liposomes

In a first experiment we established baseline GP values for PLA NP controls generated by nanoprecipitation in a C-Laurdan containing solution in the absence of lipids. Fig. 3 contains the GP values of PLA NPs (no C-Laurdan) and PLA NPs with C-Laurdan in the temperature range between 25 °C to 55 °C. The GP values of C-Laurdan in PLA NPs are only slightly higher than for PLA NP background. Consistent with previous reports,^{43,44} the fluorescence signal of C-Laurdan in PLA NPs shows only a weak temperature dependence. In fact, the GP(T) curves for PLA NPs with C-Laurdan have nearly the same slope as the PLA controls without C-Laurdan and show a negligible decrease ($\Delta GP < 0.04$) when compared with LNPs (Fig. S4†).

Next, we investigated the GP(T) relationships for LNP_x^{CHOL} and $Liposome_x^{CHOL}$. Fig. 4a contains GP(T) plots for $Liposome_5^{CHOL}$, $Liposome_{20}^{CHOL}$, $Liposome_{40}^{CHOL}$ over the temp-



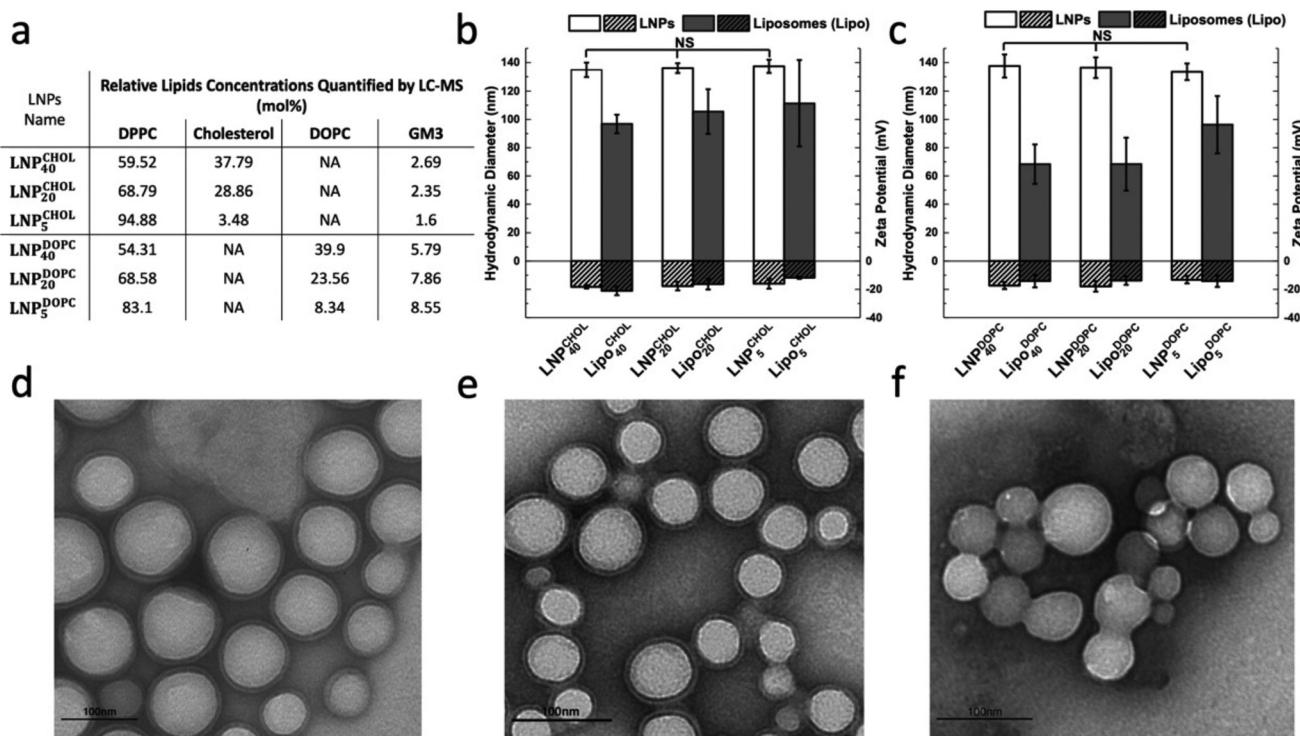


Fig. 2 Characterization of GM3-LNPs and liposomes. (a) Relative lipids concentrations in GM3-LNPs quantified by LC-MS. (b and c) Hydrodynamic diameter and zeta potential of LNP_x^{CHOL} and corresponding Liposome_x^{CHOL} (b), and of LNP_x^{DOPC} and corresponding Liposome_x^{DOPC} (c). Error bars represent standard deviation. (d–f) TEM images of LNP₄₀^{CHOL} (d), LNP₄₀^{DOPC} (e), and PLA NPs (without lipid membrane) (f). Samples were treated with sodium phosphotungstate (1% w/v in water).

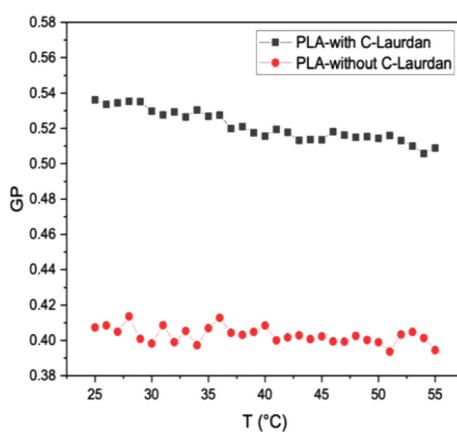


Fig. 3 C-Laurdan GP measurements of PLA NPs (without lipid membrane). C-Laurdan GP(T) plots for PLA NPs without C-Laurdan (red circles), and PLA NPs with C-Laurdan (black squares).

eratures range between 25 °C to 65 °C. Fig. 4b shows equivalent data for LNP_x^{CHOL} whose lipid coating was assembled from the same lipid mixture as liposomes. For both liposomes and LNPs the GP(T) curves show a much stronger decrease as function of temperature for all three investigated (hybrid) membrane compositions than observed for the PLA core. The decreasing sigmoidal GP(T) curves that are obvious for several

experimental conditions indicate discrete phase transitions in the lipid membranes of liposomes and LNPs. All data were fitted to a Boltzmann sigmoidal⁴⁵ (solid lines in Fig. 4a and b):

$$GP(T) = GP_2 + \frac{(GP_1 - GP_2)}{1 + e^{\frac{T - T_m}{p}}} \quad (2)$$

T_m represents an apparent thermal transition temperature of the membrane and corresponds to the temperature at which the GP value has dropped to 50% of its initial value.³⁷ GP₁ and GP₂ are the low and high temperature asymptotic values of the GP(T) fit, and p accounts for the slope of the curve, which determines the width of the transition and is a measure of the cooperativity during the membrane phase transition. The absolute value of the first derivative of the decreasing sigmoidal function ($|d(GP)/dT|$) is a bell-shaped curve which allows for an easy determination of the onset (T_{on}) and offset (T_{off}) temperatures (Fig. 4c).⁴⁰ T_m , T_{on} , T_{off} , ΔT between T_{on} and T_{off} , GP values at 25 °C and 65 °C, ΔGP between 25 °C and 65 °C, as well as p values for Liposome_x^{CHOL} and LNP_x^{CHOL} are summarized in Table 1.

The Liposome₅^{CHOL} GP(T) curves indicates a distinct phase transition from gel ($GP > 0.55$) to liquid-crystalline phase ($GP < -0.05$) (Fig. 4a). For Liposome₂₀^{CHOL} this transition is substantially broadened, and for Liposome₄₀^{CHOL} the GP(T) has no longer a detectable sigmoidal character. The phase transition



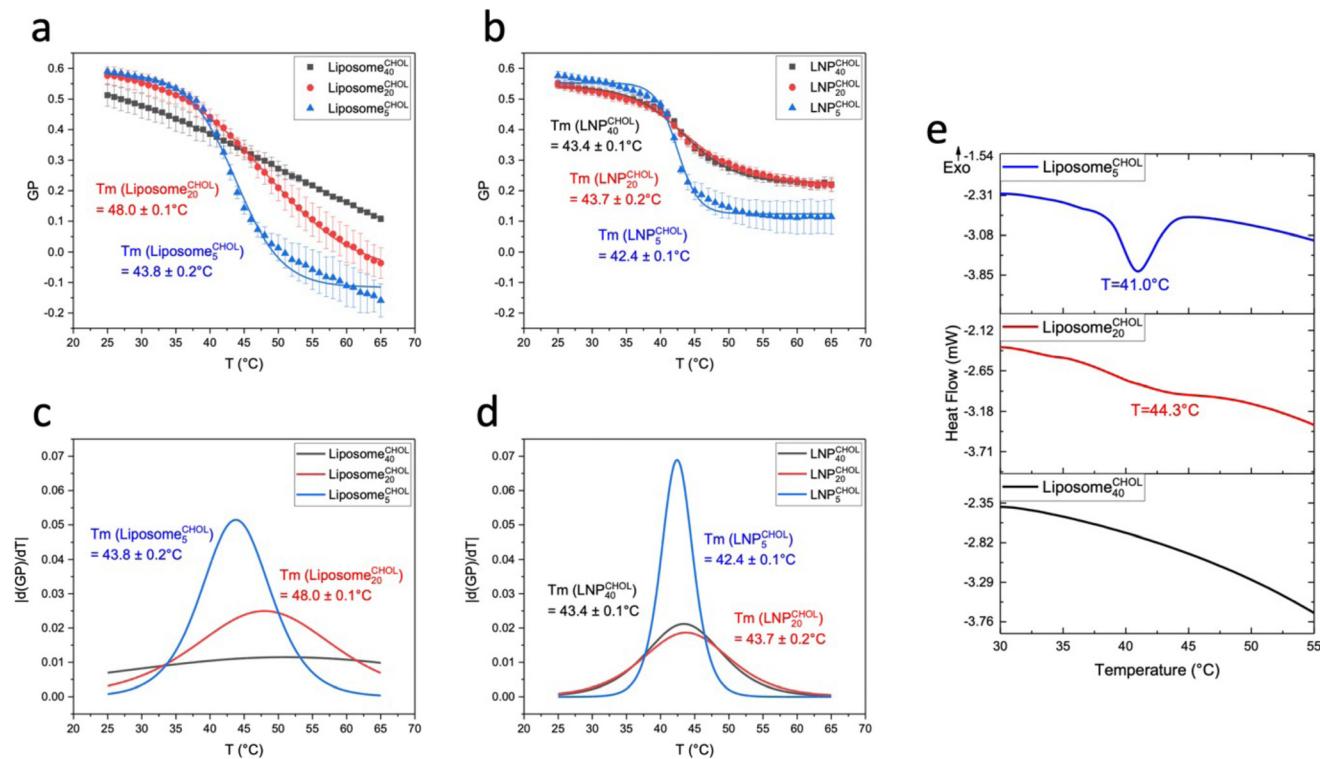


Fig. 4 C-Laurdan GP(T) and DSC measurements of Liposome_x^{CHOL} and LNP_x^{CHOL}. (a) GP(T) plots for Liposome_x^{CHOL} ($n = 3$). (b) GP(T) plots for LNP_x^{CHOL} ($n = 4$). Error bars in (a and b) represent standard deviation. (c) $|d(GP)/dT|$ vs. T plots for Liposome_x^{CHOL}. (d) $|d(GP)/dT|$ vs. T plots for LNP_x^{CHOL}. (e) DSC thermograms of Liposome_x^{CHOL}. T gives the phase transition temperature of the respective liposomes.

Table 1 The apparent thermal transition temperature (T_m), GP values at 25 °C and 65 °C, ΔGP between 25 °C and 65 °C, p values, onset (T_{on}) and offset (T_{off}) temperatures of the apparent phase transition, ΔT between T_{on} and T_{off} for Liposome_x^{CHOL} and LNP_x^{CHOL}

	T_m (°C)	GP (25 °C)	GP (65 °C)	$\Delta GP(GP_{T=25\text{ }^{\circ}\text{C}} - GP_{T=65\text{ }^{\circ}\text{C}})$	p	T_{on} (°C)	T_{off} (°C)	$\Delta T(T_{off} - T_{on})$ (°C)
Liposome ₄₀ ^{CHOL}	NA	0.51 ± 0.04	0.11 ± 0.01	0.40 ± 0.05	17.59 ± 1.62	<25.0	>65.0	>40.0
Liposome ₂₀ ^{CHOL}	48.0 ± 0.1	0.57 ± 0.03	-0.04 ± 0.05	0.61 ± 0.08	6.76 ± 0.12	27.6	>65.0	>38.0
Liposome ₅ ^{CHOL}	43.8 ± 0.2	0.59 ± 0.01	-0.16 ± 0.05	0.75 ± 0.06	3.40 ± 0.11	31.0	57.0	26.0
LNP ₄₀ ^{CHOL}	43.4 ± 0.1	0.55 ± 0.01	0.22 ± 0.02	0.33 ± 0.03	3.73 ± 0.17	29.8	57.5	27.7
LNP ₂₀ ^{CHOL}	43.7 ± 0.2	0.55 ± 0.01	0.22 ± 0.02	0.33 ± 0.03	4.26 ± 0.19	29.1	59.3	30.2
LNP ₅ ^{CHOL}	42.4 ± 0.1	0.57 ± 0.01	0.11 ± 0.06	0.46 ± 0.07	1.55 ± 0.12	35.9	49.0	13.1

has been “washed out” and is no longer detectable. The changes in the GP(T) curves observed with GM3-containing liposomes for increasing cholesterol concentrations is consistent with differential scanning calorimeter (DSC) measurements performed with the same liposomes (Fig. 4e) as well as with previous Laurdan studies of the DPPC/cholesterol system and has been rationalized by the formation of a liquid-ordered phase throughout the investigated temperature range in the presence of medium to high concentrations of cholesterol.^{38,41,46,47} The observations that in the low temperature limit the GP values for Liposome₄₀^{CHOL} lie lower than those of Liposome₅^{CHOL}, Liposome₂₀^{CHOL}, and that this order reverses at high temperatures reflects the well-known temperature dependence of the cholesterol effect on membrane fluidity, which means cholesterol decreases fluidity at high temperatures and increases fluidity at low temperatures.^{48,49}

Unlike in the case of liposomes, the GP(T) curves for LNPs indicate discernible phase transitions for all investigated conditions (Fig. 4b). Even at high input concentrations of cholesterol, LNP₄₀^{CHOL} still show a decreasing sigmoidal GP(T) behaviour, albeit with a smaller total change in GP. For all cholesterol concentrations, the fits yield p values whose absolute values are smaller than those of liposomes with identical composition, which is consistent with narrower transitions (*i.e.* a small temperature differences (ΔT) between T_{on} and T_{off}) (Fig. 4c, d and Table 1) due to a higher degree of cooperativity. The recorded GP values of LNP₅^{CHOL} indicate transitions from a gel phase ($GP = 0.57 \pm 0.01$) to liquid-disordered intermediate phase ($GP = 0.11 \pm 0.06$). For LNP₂₀^{CHOL} and LNP₄₀^{CHOL}, GP values of 0.22 ± 0.02 are measured in the high temperature limit, suggesting a state between liquid-ordered and liquid-disordered phase. The measured phase transition temperatures

T_m for LNPs with different cholesterol content are very similar with a maximum difference of 1.3 °C. For Liposome_x^{CHOL} an increase in T_m of 4.2 °C is recorded between Liposome₅^{CHOL} and Liposome₂₀^{CHOL}, indicating a stronger cholesterol dependence of T_m in liposomes than in LNPs.

Fig. 5a–c compares GP(T) curves of Liposome_x^{CHOL} and LNP_x^{CHOL} with identical membrane composition. In the low temperature limit, the GP values for liposomes and LNPs are nearly identical except Liposome₄₀^{CHOL} and LNP₄₀^{CHOL}, for which the GP values of the LNP₄₀^{CHOL} lie slightly higher than those of Liposome₄₀^{CHOL}. In the high temperature limit, the GP values of LNPs lie overall higher than those of liposomes. However, since the GP values of liposomes also show a more pronounced increase as function of increasing cholesterol input, the gap between the high temperature GP limits of liposomes and LNPs narrows with increasing cholesterol concentration in the high temperature limit. The increase in GP with increasing cholesterol content in liposomes for $T > T_m$ indicates an increasing membrane order that can be understood in terms of the established cholesterol condensing effect that results in a closer packing of lipids.^{48,49} It is also interesting that the high temperature GP values for LNP₅^{CHOL} are similar to those of Liposome₄₀^{CHOL}. This observation implies that the polymer core of the LNPs has a similar effect on the polarity of the lipid layer as cholesterol in liposomes in the high temperature range. We attribute this behaviour to an interdigitation of hydrophobic polymer chains into the surrounding lipid layer (Fig. 6). These additional polymer segments increase the density of hydrophobic components in the resulting hybrid membrane, and it is conceivable that hydrophobic interactions between polymer segments and lipid tails favour a parallel alignment that result in an increased order and tighter packing. A similar interpretation was previously presented for the rigidification of a lipid monolayer around cross-linked polysaccharide NPs functionalized with long chain (C16) fatty acids by Peyrot *et al.*⁵⁰ Stabilizing hydrophobic interactions between lipid tails and surface polymer chains could also explain the weak cholesterol dependence of T_m observed for LNPs.

After characterizing GP(T) curves for Liposome_x^{CHOL} and LNP_x^{CHOL}, we will now turn our attention to GP(T) curves for liposomes and LNPs with DOPC input concentrations of 5, 20,

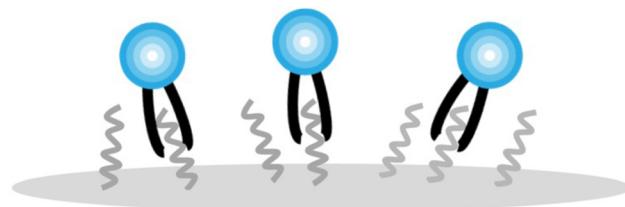


Fig. 6 Schematic representation of the interdigitation between hydrophobic polymer chains and surrounding lipid tails.

and 40 mol% (Fig. 7a and b). Liposomes and LNPs exhibited decreasing sigmoidal GP(T) curves that were well described by eqn (2) for all DOPC concentrations. Fig. 7c and d contains plots of the first derivative of GP versus the temperature ($|d(GP)/dT|$). T_m , T_{on} , T_{off} , ΔT between T_{on} and T_{off} , GP values at 25 °C and 65 °C, ΔGP between 25 °C and 65 °C, as well as p values are summarized in Table 2. LNPs and liposomes with identical DOPC content show comparable transition temperatures, T_m , which systematically decrease with increasing DOPC concentration due to increasing membrane disorder.

The measured GP values of Liposome₅^{DOPC} indicate a transition from gel phase (GP = 0.55 ± 0.02) to fluid phase (GP = −0.27). The GP values of liposomes in the low temperature limit decrease with increasing DOPC concentration. The GP ($T = 25$ °C) values for Liposome₂₀^{DOPC} and Liposome₄₀^{DOPC} are 0.49 ± 0.03 and 0.38 ± 0.05, respectively. These data evidence an increase in average membrane polarity in the low temperature limit due to a decrease of the average lipid packing density with increasing concentration of unsaturated lipid. Previous studies have shown that a binary DOPC/DPPC mix exhibits a gel – liquid crystalline coexistence region over a broad DOPC concentration range at 25 °C.⁴² The decrease in GP observed for increasing DOPC concentration is consistent with a gradual increase in the contribution from the liquid crystalline phase in the coexistence region. In the high temperature limit, the differences in the GP values for liposomes with different DOPC concentrations are less distinct and the GP(T) curves converge.

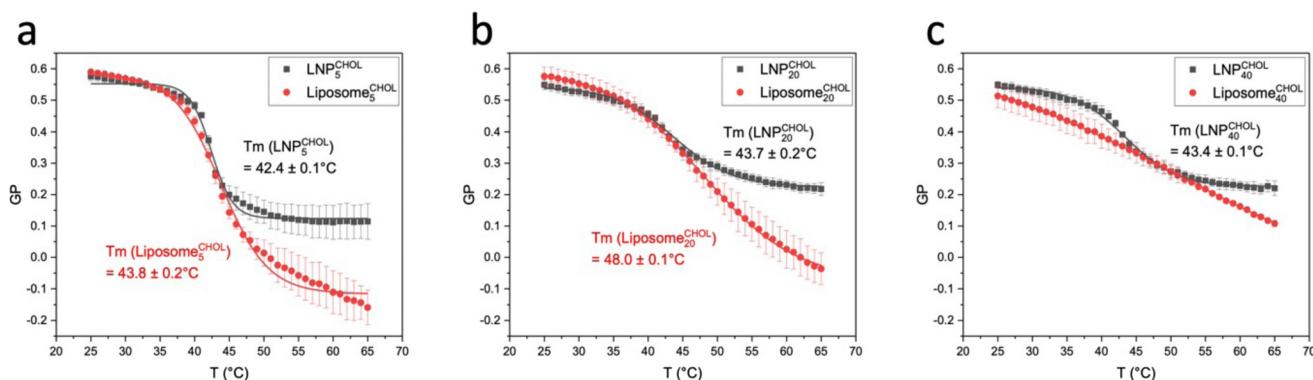


Fig. 5 Comparison of GP(T) curves of LNP_x^{CHOL} and Liposome_x^{CHOL} with the same cholesterol content. GP(T) curves of LNP_x^{CHOL} and Liposome_x^{CHOL} with $x = 5$ (a), $x = 20$ (b), and $x = 40$ (c). Error bars represent standard deviation.



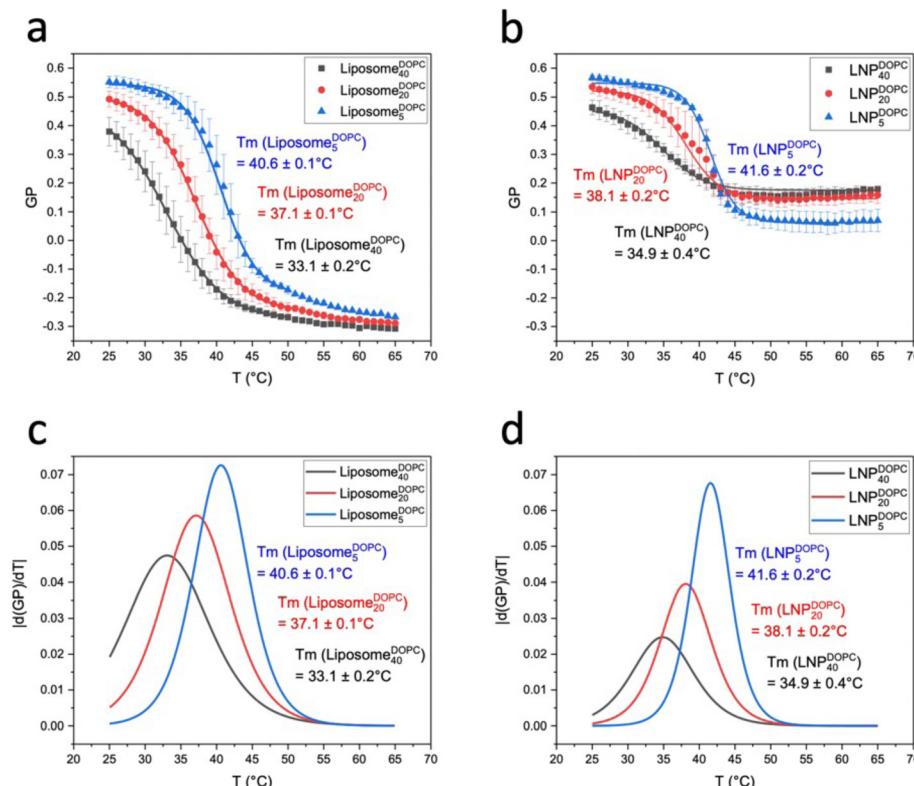


Fig. 7 C-Laurdan GP(T) measurements of Liposome_x^{DOPC} and LNP_x^{DOPC}. (a) C-Laurdan GP(T) plots for Liposome_x^{DOPC} ($n = 3$). (b) C-Laurdan GP(T) plots for LNP_x^{DOPC} ($n = 4$). Error bars in (a and b) represent standard deviation. (c) $|d(GP)/dT|$ vs. T plots for Liposome_x^{DOPC}. (d) $|d(GP)/dT|$ vs. T plots for LNP_x^{DOPC}.

Table 2 The apparent thermal transition temperature (T_m), GP values at 25 °C and 65 °C, ΔGP between 25 °C and 65 °C, p values, onset (T_{on}) and offset (T_{off}) temperatures of the apparent phase transition, ΔT between T_{on} and T_{off} for Liposome_x^{DOPC} and LNP_x^{DOPC}

	T_m (°C)	GP (25 °C)	GP (65 °C)	$\Delta GP(GP_{T=25\text{ }^{\circ}\text{C}} - GP_{T=65\text{ }^{\circ}\text{C}})$	p	T_{on} (°C)	T_{off} (°C)	$\Delta T(T_{off} - T_{on})$ (°C)
Liposome ₄₀ ^{DOPC}	33.1 ± 0.2	0.38 ± 0.05	-0.31 ± 0.00	0.69 ± 0.05	3.93 ± 0.09	<25.0	48.9	>23.9
Liposome ₂₀ ^{DOPC}	37.1 ± 0.1	0.49 ± 0.03	-0.29 ± 0.00	0.78 ± 0.03	3.21 ± 0.07	<25.0	50.3	>25.3
Liposome ₅ ^{DOPC}	40.6 ± 0.1	0.55 ± 0.02	-0.27 ± 0.00	0.82 ± 0.02	2.53 ± 0.09	30.6	51.4	20.8
LNP ₄₀ ^{DOPC}	34.9 ± 0.4	0.46 ± 0.03	0.18 ± 0.00	0.28 ± 0.03	2.88 ± 0.32	<25.0	46.6	>21.6
LNP ₂₀ ^{DOPC}	38.1 ± 0.2	0.54 ± 0.02	0.16 ± 0.03	0.38 ± 0.05	2.39 ± 0.13	29.0	47.9	18.9
LNP ₅ ^{DOPC}	41.6 ± 0.2	0.57 ± 0.01	0.07 ± 0.04	0.50 ± 0.05	1.80 ± 0.12	34.1	49.3	15.2

LNP_x^{DOPC} also show a decrease in GP values with increasing DOPC content in the low temperature limit, but the GP values for $T = 25$ °C do not drop below GP = 0.45 for LNP₄₀^{DOPC}. In the high temperature limit the GP(T) curves for LNPs lie systematically higher than for liposomes (Fig. 8). The GP ($T = 65$ °C) values are negative for liposomes but positive for LNPs. There are also smaller differences between LNPs with different DOPC concentrations in the high temperature limit; the GP values for LNP₅^{DOPC} lie systematically lower than those of LNP₂₀^{DOPC} or LNP₄₀^{DOPC}, but the difference is small (~0.1 unit). Overall, the lower polarity of the hybrid membranes in the high temperature limit as indicated by higher GP values for LNPs suggests that the phase transition in the hybrid membrane of DOPC-containing LNPs is associated with a less distinct structural change than in DOPC liposomes. We again attribute this

difference to an interdigitation of polymer chain fragments and self-assembled lipid monolayer in the case of the LNPs. Lateral hydrophobic interactions between polymer chain fragments and both saturated and unsaturated lipids can stabilize the hybrid membrane and decrease the change in membrane polarity associated with the phase transition. The polymer chain fragments represent a framework that impedes lateral lipid motion, thus limiting the fluidity of the hybrid membrane at elevated temperatures.

GM3-CD169-mediated binding of GM3-LNPs to Raji B CD169⁺ cells

GM3-LNPs are interesting drug delivery platforms as they combine GM3-mediated targeting of CD169-expressing cells with the programmable release properties of the PLA core.²²

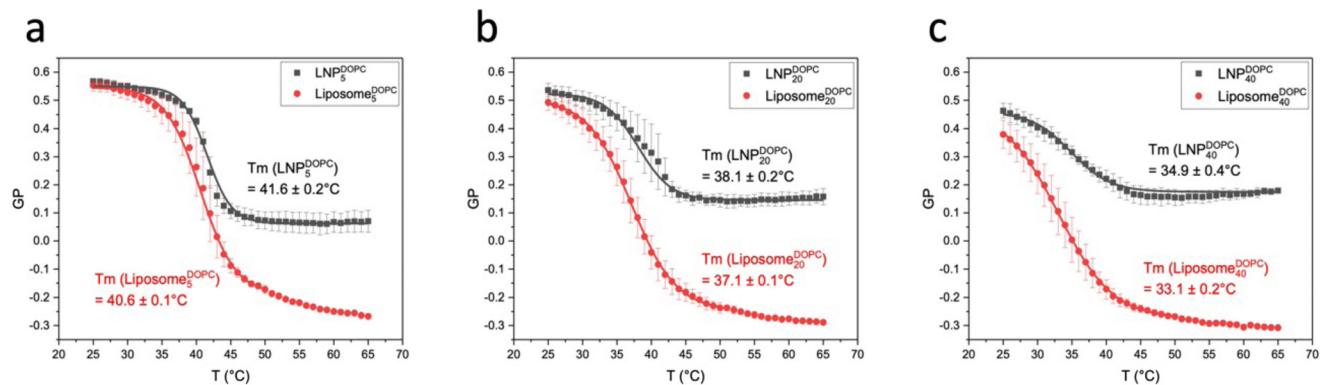


Fig. 8 Comparison of $GP(T)$ curves of LNP_x^{DOPC} and $\text{Liposome}_x^{\text{DOPC}}$ with the same DOPC content. $GP(T)$ curves of LNP_x^{DOPC} and $\text{Liposome}_x^{\text{DOPC}}$ with $x = 5$ (a), $x = 20$ (b), and $x = 40$ (c). Error bars represent standard deviation.

Individual GM3-CD169 contacts are relatively weak with a dissociation constant K_D in the mM range, but multivalent presentation can result in an increase in avidity.^{51,52} Differences in the mobility of GM3 in the membrane of liposomes or LNPs could affect collective binding interactions and thus avidity.²⁹ In this context it is therefore relevant that the measured GP values for DOPC liposomes and LNPs indicate differences in lipid packing and fluidity at 37 °C. These differences may affect GM3-mediated binding to CD169-expressing cell surfaces. Fig. 9 compares the specific GM3 binding of LNP_5^{CHOL} and LNP_{40}^{CHOL} , as well as LNP_5^{DOPC} and LNP_{40}^{DOPC} to $CD169^+$ Raji B cells as determined by flow cytometry. In all experiments, the LNP concentration was monitored by UV-Vis measure-

ments and kept constant, and LNPs without GM3 (the DPPC content was increased to account for the lack of GM3) were included as controls. All GM3-containing LNPs showed much more binding than the LNP controls, confirming that binding is dominated by GM3-CD169 interactions. Consistent with this interpretation, GM3-LNPs showed negligible binding to $CD169^-$ Raji B cells (Raji B cells which did not express CD169). Intriguingly, LNP_{40}^{DOPC} which have a lower GP and thus more fluid membrane than all other investigated GM3-LNPs, show significantly more binding to $CD169^+$ Raji B cells than any other experimental condition. This is especially remarkable considering that – according to our LC-MS measurements in Fig. 2a – the GM3 content of LNP_{40}^{DOPC} was lower than for LNP_5^{DOPC} or LNP_{20}^{DOPC} . The observed increase in binding for GM3-LNPs with the lowest GP value is indicative of a gain in binding avidity and corroborates the hypothesis that increased membrane fluidity benefits GM3-CD169-mediated cell binding.

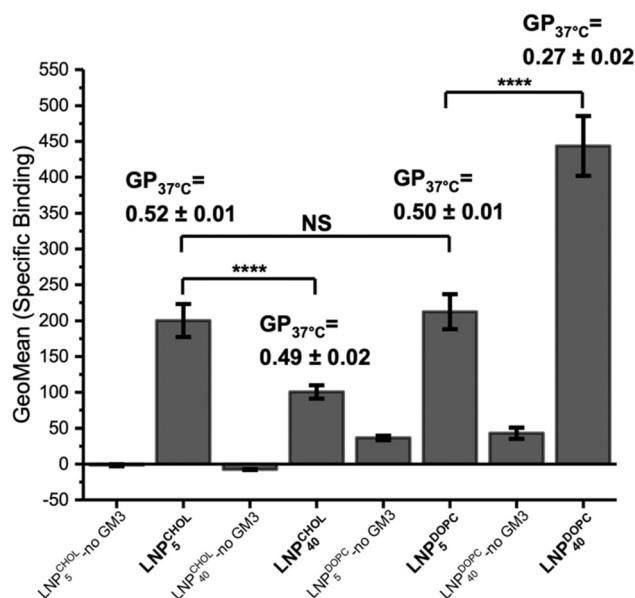


Fig. 9 Binding of different GM3-LNPs to $CD169^+$ Raji B cells. From left to right: LNP_5^{CHOL} without GM3, LNP_5^{CHOL} , LNP_{40}^{CHOL} without GM3, LNP_{40}^{CHOL} , LNP_5^{DOPC} without GM3, LNP_5^{DOPC} , LNP_{40}^{DOPC} without GM3, LNP_{40}^{DOPC} . The GP ($T = 37$ °C) are included for LNP_5^{CHOL} , LNP_{40}^{CHOL} , LNP_5^{DOPC} , and LNP_{40}^{DOPC} ($n = 10$).

Conclusions

This work has investigated the temperature-dependent polarity of hybrid membranes around a PLA core using the fluorescent C-Laurdan probe. In the low temperature limit (*i.e.* at room temperature) LNP_x^{CHOL} and LNP_x^{DOPC} were found to have GP values that are comparable to those of liposomes with identical membrane composition. However, consistently higher GP values for LNPs than for liposomes at elevated temperatures evidence that LNPs contain the C-Laurdan probe located in an environment that is less polar than in the corresponding liposomes. One model to account for this gain in polarity for the LNP hybrid membrane is a structure that contains an outer lipid layer at least partially interdigitating into an inner layer of polymer chain segments that define the surface of the PLA NP. The polymer segments of the PLA core structure the lipid layer through lateral hydrophobic interactions and reduce the fluidity of the lipid membrane. Due to this structure-stabilizing effect, which was observed for both saturated and unsatu-



rated lipids, LNPs retain detectable phase transitions even for membrane compositions that lack detectable phase transitions in liposomes. The fluidity of the hybrid membrane of GM3-LNPs was found to increase GM3-CD169-mediated cell binding under otherwise identical conditions, underlining the potential for controlling lipid-mediated binding through rationally designed LNPs.

Materials and methods

Materials

All chemicals were used as received. 1,2-dipalmitoyl-*sn*-glycero-3-phosphocholine (DPPC), 1,2-dioleoyl-*sn*-glycero-3-phosphocholine (DOPC), cholesterol, GM3 ganglioside (Milk, Bovine-Ammonium Salt) and 1,2-dipalmitoyl-*sn*-glycero-3-phosphoethanolamine-*N*-(lissamine rhodamine B sulfonyl) (ammonium salt) (16:0 Liss Rhod PE) were purchased from Avanti Polar Lipids Inc. *N*-Methyl-*N*-[6-(1-oxododecyl)-2-naphthalenyl]glycine (C-Laurdan) was purchased from R&D Systems Inc. Chloroform, methanol, acetonitrile (all HPLC grade), sodium phosphotungstate hydrate, and ester terminated polylactic acid (PLA) (poly(D,L-lactide)), Resomer R207S,⁵³ with a mass-average molecular weight (M_w) of 262 000 g mol⁻¹ and polydispersity of 1.6 were purchased from Sigma-Aldrich Inc.

Methods

Preparation of lipid-wrapped polymeric nanoparticles. Lipid-coated polymeric nanoparticles (LNPs) were prepared through one-step nanoprecipitation synthesis, as described previously.^{23,54} A total lipid amount of 0.25 μ mole,⁵⁴ an aqueous/organic solution volume ratio of 10:1⁵⁵ and a lipid/polymer weight ratio of 15%²⁶ was chosen for the synthesis. A lipid mixture containing DPPC (25 mg mL⁻¹ in chloroform), cholesterol (25 mg mL⁻¹ in chloroform), GM3 (2 mM in methanol/chloroform (1:1)), and C-Laurdan (1 mM in chloroform) (M1) or DPPC (25 mg mL⁻¹), DOPC (25 mg mL⁻¹ in chloroform), GM3 (2 mM), and C-Laurdan (1 mM) (M2) with mol% as-specified in the text was added to around 4 mL of Milli-Q water. Next, around 0.4 mL of high molecular weight PLA solution in acetonitrile (2.5 mg mL⁻¹) was pipetted dropwise to the above aqueous solution under stirring. The final solution was vortexed for 10 s and then sonicated in a bath sonicator (Branson Ultrasonics, No. 5510, Danbury, CT) for 6 min. Finally, NPs were washed 3 times (4500 g, 15 min) using 10 kDa Amicon Ultra-4 centrifugal filter (Millipore Sigma, Burlington, MA) to remove organic solvent and free lipid molecules. Polymer NPs without membrane (PLA NPs) were obtained following the same procedure in the absence of lipids but with C-Laurdan. DLS was applied to measure the diameter of the samples. 2 mol% of fluorescent lipid (16:0 Liss Rhod PE) instead of C-Laurdan was incorporated to all LNPs for labelling when LNPs were subjected to flow cytometry. The different LNP preparations used in the cell binding experiments had essentially identical diameters. Assuming

identical dye concentrations in the lipid coating, UV-Vis absorbance measurements were used to ensure identical concentrations for the different LNP preparations in the binding studies.

Fabrication of liposomes. Liposomes were formulated according to the Bangham method⁵⁶ with certain adjustments. A lipid mixture containing DPPC (10 mM), cholesterol (10 mM), GM3 (2 mM), C-Laurdan (1 mM) (M1) or DPPC (10 mM), DOPC (10 mM), GM3 (2 mM), C-Laurdan (1 mM) (M2) in chloroform with mol% as-specified in the text was added to a 25 mL round-bottom flask. The total lipid amount was maintained at 1 μ mole. Then, the solvent was removed by rotary evaporation (34 °C, 10 min) to obtain a homogeneous and thin lipid film and the samples were dried overnight under vacuum. Multilamellar vesicles (MLVs) were obtained by adding 1 mL of Milli-Q water to this lipid dry film and then sonicating for 5 min using a probe sonicator (120 Sonic Dismembrator, Fisher Scientific, Waltham, MA). Finally, a dispersion of small unilamellar vesicles (SUVs) was obtained by extruding the obtained MLV dispersion 6 times through a calibrated polycarbonate membrane with a pore diameter of 100 nm using the Avanti® mini extruder (Avanti Polar Lipids Inc).

Dynamic light scattering (DLS) and zeta-potential measurements. Hydrodynamic size and zeta-potential of LNPs and liposomes were measured by Zetasizer Nano ZS90 (Malvern, Worcestershire, UK) at room temperature. For hydrodynamic size measurements, NPs were diluted with Milli-Q water. The zeta potential was determined in 10 mM NaCl solution.

Transmission electron microscopy (TEM) characterization. To prepare samples for inspection in TEM, lipid-coated polymeric nanoparticles (LNPs) and polymer NPs without membrane (PLA NPs) NPs were drop-cast onto carbon-coated TEM grids and incubated for 20 min before removing the excess solution by a clean filter paper. Next, samples were stained with 1% sodium phosphotungstate Na₃P(W₃O₁₀)₄ in water (w/v) for 10 s and excess stain solution was removed. Samples were dried and stored under vacuum before imaging using a Hitachi 7800 TEM (HT7800) with 100 kV acceleration voltage.

Quantification of relative lipids concentrations. Lipids were extracted from lipid-coated polymer nanoparticles (LNPs) through a mixture of chloroform–methanol (1:1, v/v), and a single-phase solution was obtained after stirring overnight. LC-MS analysis was performed by the Harvard Center for Mass Spectrometry. All samples were run on a Thermo Orbitrap QE+/Ultimate 3000 LC (ThermoFisher Qexactive Plus) with source HESI+, full MS 70k resolution, 3×10^6 target AGC, m/z 300–1500, max IT 100 ms. A Dikma biobond C4 (50 \times 4.6 mm) column was used with column temperature at 45 °C, sample temperature at 4 °C, and injection volume of 5 μ L.

C-Laurdan generalized polarization (GP) measurements. Steady-state fluorescence measurements with C-Laurdan were performed with a spectrofluorometer (Horiba, Piscataway, NJ) equipped with temperature control, using 1 cm path length quartz cuvettes. C-Laurdan emission spectra (420–510 nm) of LNPs and liposomes were collected from 25 °C to 65 °C at an



interval of 1 °C with an equilibration time of 0.5 min at each temperature upon excitation at 405 nm. C-Laurdan GP values at different temperature were calculated using the difference in emission intensities at 440 and 490 nm according to eqn (1) and GP(T) plots were obtained. Note that background intensities of LNPs and liposomes without C-Laurdan were minimal and negligible (Fig. S5†), so only the fluorescence intensities of the samples containing C-Laurdan without background subtraction were recorded for GP measurements. The decreasing sigmoidal GP(T) curve was fitted to a Boltzmann sigmoidal function (eqn (2)) to derive the apparent thermal transition temperature (T_m) of the membrane, low (GP_1) and high (GP_2) temperature asymptotic values, and p values which show the cooperativity during the membrane phase transition. To assess the onset (T_{on}) and offset (T_{off}) temperatures, the absolute value of the first derivative with respect to temperature of the decreasing GP(T) sigmoidal function was further evaluated, which showed a bell-shaped curve with baseline values corresponding to the small slopes of the GP(T) curves. The points at which the slope begins to deviate sharply from and returns to its baseline values (*i.e.* the start and the end of the peak) correspond to T_{on} and T_{off} . To determine these points, tangent lines were constructed along the peaks and representative tangent constructions are shown in the Fig. S6.† T_{on} and T_{off} were taken to be the averages of the intersection of two left-side tangents and two right-side tangents with the baseline of each peak respectively, analogous to the procedure followed to find transition onset and completion temperatures using differential scanning calorimetry (DSC).⁴⁰

Differential scanning calorimetry (DSC) measurements. To characterize thermal behaviour of LNPs and liposomes, 20 μ L of concentrated solution was transferred into an aluminum pan (Thermal Support, Hayesville, NC) and sealed. Thermograms from 30 °C to 55 °C, at a heating rate of 5 °C min⁻¹ were recorded by using a Mettler Toledo Polymer DSC R (Mettler-Toledo, Columbus, OH). The DSC software integrated into the instrument was used to determine the phase transition temperatures.

Determination of NP concentrations through UV-Vis. The absorption spectra of LNPs in Milli-Q water were acquired using a Spectronic 200 UV-Vis spectrometer (Fisher Scientific, Waltham, MA) and Milli-Q water was used for baseline correction. Beer's law was used to calculate the concentration of LNPs using the absorption value of fluorescently labelled lipid (16 : 0 Liss Rhod PE) at the wavelength of 570 nm (Lambda max), and the molar absorptivity of $\epsilon = 73\,000\,M^{-1}\,cm^{-1}$.²³ The final volume of samples was adjusted to 100 μ L with a typical concentration of 10^{12} NPs mL⁻¹.

Cell culture. Raji B cells were cultured in complete RPMI-1640 medium (Gibco, Thermo Fisher Scientific, No. 11875093) containing 10% FBS (Gibco, Thermo Fisher Scientific, No. 16000044) and 1% penicillin-streptomycin (Gibco, Thermo Fisher Scientific, No. 15070063). Raji B cells were transduced with VSV-G pseudotyped LNC-CD169 mutant retroviral vectors, followed by 1 mg mL⁻¹ G418 (Gibco, Thermo Fisher Scientific) selection, as described previously.⁵⁷ CD169⁺

Raji B cells were cultured with addition of antibiotic G418, while CD169⁻ Raji B cells were cultured in the same medium without G418. All cells were cultured at 37 °C in a humidified atmosphere containing 5% CO₂.

Characterization of GM3-CD169-mediated binding of GM3-LNPs to CD169⁺ Raji B Cells. 5×10^5 CD169⁺ Raji B and CD169⁻ Raji B cells were pelleted after centrifugation (270g, 5 min). 2 mol% of fluorescent lipid (16 : 0 Liss Rhod PE) was incorporated into all LNPs for labelling, and the absorbance of LNPs was controlled to be the same. 30 μ L LNPs with and without GM3 (in 0.1 mL of 10% FBS RPMI-1640) were added to the cell pellets and the mixture was incubated at 37 °C, 5% CO₂ for 5 min. Unbound LNPs were washed twice with PBS by centrifugation (270g, 5 min), and cells were subsequently fixed with 4% paraformaldehyde (PFA) (Sigma-Aldrich, St Louis, MO) for 10 min at room temperature. Following the last washing step, the fluorescence intensity of each sample was measured by flow cytometry using a FACSCalibur cytometer (BD Biosciences, San Jose, CA) and the data was analyzed through Flowing Software 2. All fluorescence intensities were background corrected (cells without treatment were used as background), and the calculated difference was used for the analysis. GeoMean intensities for specific binding (Fig. 9) were determined by subtracting non-specific binding (binding of LNPs to CD169⁻ Raji B cells) from total binding (binding of LNPs to CD169⁺ Raji B cells).

Statistical analysis. All data are presented as mean \pm standard deviation (SD). Data presentation and sample size for statistical analysis of individual experiments are specified in figure captions. Statistical significance of data was determined using two-sample Student's *t*-test as implemented in Origin. One asterisk (*) indicates significant differences at $p \leq 0.05$, two asterisks (**) for $p \leq 0.01$, three asterisks (***) for $p \leq 0.001$, and four asterisks (****) for $p \leq 0.0001$. NS was used to demonstrate nonsignificant differences.

Author contributions

BMR and YG conceived and designed the project. YG performed experiments, analysed the data, and generated figures. BMR and YG wrote the manuscript. All authors assisted with the interpretation of the results and contributed to the final manuscript.

Conflicts of interest

BMR holds patents for GM3-functionalized nanoparticles.

Acknowledgements

BMR acknowledges support from the National Institutes of Health through grants 1R01AI175068 and R01CA138509. YG thanks Madison Tang for rendering the scheme of the lipid-coated polymeric nanoparticles in Fig. 1a.



References

- 1 K. M. Vargas and Y.-S. Shon, Hybrid Lipid–Nanoparticle Complexes for Biomedical Applications, *J. Mater. Chem. B*, 2019, **7**(5), 695–708.
- 2 A. Luchini and G. Vitiello, Understanding the Nano-Bio Interfaces: Lipid-Coatings for Inorganic Nanoparticles as Promising Strategy for Biomedical Applications, *Front. Chem.*, 2019, **7**, 343.
- 3 M. Mirahadi, S. Ghanbarzadeh, M. Ghorbani, A. Gholizadeh and H. Hamishehkar, A Review on the Role of Lipid-Based Nanoparticles in Medical Diagnosis and Imaging, *Ther. Delivery*, 2018, **9**(8), 557–569.
- 4 I.-A. Pavel, M. Girardon, S. E. Hajj, S. Parant, F. Amadei, S. Kaufmann, M. Tanaka, V. Fierro, A. Celzard, N. Canilho and A. Pasc, Lipid-Coated Mesoporous Silica Microparticles for the Controlled Delivery of β -Galactosidase into Intestines, *J. Mater. Chem. B*, 2018, **6**(35), 5633–5639.
- 5 T. Sun, Y. Sun and H. Zhang, Phospholipid-Coated Mesoporous Silica Nanoparticles Acting as Lubricating Drug Nanocarriers, *Polymers*, 2018, **10**(5), 513.
- 6 J. Tu, J. Bussmann, G. Du, Y. Gao, J. A. Bouwstra and A. Kros, Lipid Bilayer-Coated Mesoporous Silica Nanoparticles Carrying Bovine Hemoglobin towards an Erythrocyte Mimic, *Int. J. Pharm.*, 2018, **543**(1), 169–178.
- 7 B. Du, L. Tian, X. Gu, D. Li, E. Wang and J. Wang, Anionic Lipid, pH-Sensitive Liposome–Gold Nanoparticle Hybrids for Gene Delivery – Quantitative Research of the Mechanism, *Small*, 2015, **11**(19), 2333–2340.
- 8 B. Du, X. Gu, X. Han, G. Ding, Y. Wang, D. Li, E. Wang and J. Wang, Lipid-Coated Gold Nanoparticles Functionalized by Folic Acid as Gene Vectors for Targeted Gene Delivery in Vitro and in Vivo, *ChemMedChem*, 2017, **12**(21), 1768–1775.
- 9 J. H. Kang and Y. T. Ko, Lipid-Coated Gold Nanocomposites for Enhanced Cancer Therapy, *Int. J. Nanomed.*, 2015, **10**(Spec Iss), 33–45.
- 10 J. Liang, X. Zhang, Y. Miao, J. Li and Y. Gan, Lipid-Coated Iron Oxide Nanoparticles for Dual-Modal Imaging of Hepatocellular Carcinoma, *Int. J. Nanomed.*, 2017, **12**, 2033–2044.
- 11 A. Luchini, C. Irace, R. Santamaria, D. Montesarchio, R. K. Heenan, N. Szekely, A. Flori, L. Menichetti and L. Paduano, Phosphocholine-Decorated Superparamagnetic Iron Oxide Nanoparticles: Defining the Structure and Probing in Vivo Applications, *Nanoscale*, 2016, **8**(19), 10078–10086.
- 12 A. A. Allam, M. E. Sadat, S. J. Potter, D. B. Mast, D. F. Mohamed, F. S. Habib and G. M. Pauletti, Stability and Magnetically Induced Heating Behavior of Lipid-Coated Fe₃O₄ Nanoparticles, *Nanoscale Res. Lett.*, 2013, **8**(1), 426.
- 13 D. Bhowmik, K. R. Mote, C. M. MacLaughlin, N. Biswas, B. Chandra, J. K. Basu, G. C. Walker, P. K. Madhu and S. Maiti, Cell-Membrane-Mimicking Lipid-Coated Nanoparticles Confer Raman Enhancement to Membrane Proteins and Reveal Membrane-Attached Amyloid- β Conformation, *ACS Nano*, 2015, **9**(9), 9070–9077.
- 14 X. Yu, A. Feizpour, N.-G. P. Ramirez, L. Wu, H. Akiyama, F. Xu, S. Gummuluru and B. M. Reinhard, Glycosphingolipid-Functionalized Nanoparticles Recapitulate CD169-Dependent HIV-1 Uptake and Trafficking in Dendritic Cells, *Nat. Commun.*, 2014, **5**(1), 4136.
- 15 X. An, D. Stelter, T. Keyes and B. M. Reinhard, Plasmonic Photocatalysis of Urea Oxidation and Visible-Light Fuel Cells, *Chem*, 2019, **5**, 2228–2242.
- 16 W. T. Al-Jamal and K. Kostarelos, Liposome–Nanoparticle Hybrids for Multimodal Diagnostic and Therapeutic Applications, *Nanomedicine*, 2007, **2**(1), 85–98.
- 17 S. Krishnamurthy, R. Vaiyapuri, L. Zhang and J. M. Chan, Lipid-Coated Polymeric Nanoparticles for Cancer Drug Delivery, *Biomater. Sci.*, 2015, **3**(7), 923–936.
- 18 B. Mandal, H. Bhattacharjee, N. Mittal, H. Sah, P. Balabathula, L. A. Thoma and G. C. Wood, Core-Shell-Type Lipid-Polymer Hybrid Nanoparticles as a Drug Delivery Platform, *Nanomedicine*, 2013, **9**(4), 474–491.
- 19 K. Hadinoto, A. Sundaresan and W. S. Cheow, Lipid-Polymer Hybrid Nanoparticles as a New Generation Therapeutic Delivery Platform: A Review, *Eur. J. Pharm. Biopharm.*, 2013, **85**(3), 427–443.
- 20 K. P. García, K. Zarschler, L. Barbaro, J. A. Barreto, W. O’Malley, L. Spiccia, H. Stephan and B. Graham, Zwitterionic-Coated “Stealth” Nanoparticles for Biomedical Applications: Recent Advances in Countering Biomolecular Corona Formation and Uptake by the Mononuclear Phagocyte System, *Small*, 2014, **10**(13), 2516–2529.
- 21 F. Xu, M. Reiser, X. Yu, S. Gummuluru, L. Wetzler and B. M. Reinhard, Lipid-Mediated Targeting with Membrane Wrapped Nanoparticles in the Presence of Corona Formation, *ACS Nano*, 2016, **10**(1), 1189–1200.
- 22 B. Eshaghi, J. Fofana, S. B. Nodder, S. Gummuluru and B. M. Reinhard, Virus-Mimicking Polymer Nanoparticles Targeting CD169 + Macrophages as Long-Acting Nanocarriers for Combination Antiretrovirals, *ACS Appl. Mater. Interfaces*, 2022, **14**(2), 2488–2500.
- 23 B. Eshaghi, N. Alsharif, X. An, H. Akiyama, K. A. Brown, S. Gummuluru and B. M. Reinhard, Stiffness of HIV-1 Mimicking Polymer Nanoparticles Modulates Ganglioside-Mediated Cellular Uptake and Trafficking, *Adv. Sci.*, 2020, **7**(18), 2000649.
- 24 J. Thevenot, A.-L. Troutier, L. David, T. Delair and C. Ladavière, Steric Stabilization of Lipid/Polymer Particle Assemblies by Poly(Ethylene Glycol)-Lipids, *Biomacromolecules*, 2007, **8**(11), 3651–3660.
- 25 F. Vanneste, A. Faure, M. Varache, M. Menendez-Miranda, V. Dyon-Tafani, S. Dussurgey, E. Errazuriz-Cerda, V. L. Padula, P. Alcouffe, M. Carrière, R. Gref, F. Laurent, J. Josse and C. Ladavière, LipoParticles: A Lipid Membrane Coating onto Polymer Particles to Enhance the Internalization in Osteoblast Cells, *Nanoscale*, 2023, **15**(44), 18015–18032.
- 26 L. Zhang, J. M. Chan, F. X. Gu, J.-W. Rhee, A. Z. Wang, A. F. Radovic-Moreno, F. Alexis, R. Langer and



O. C. Farokhzad, Self-Assembled Lipid–Polymer Hybrid Nanoparticles: A Robust Drug Delivery Platform, *ACS Nano*, 2008, **2**(8), 1696–1702.

27 J. Bompard, A. Rosso, L. Brizuela, S. Mebarek, L. J. Blum, A.-M. Trunfio-Sfarghiu, G. Lollo, T. Granjon, A. Girard-Egrot and O. Maniti, Membrane Fluidity as a New Means to Selectively Target Cancer Cells with Fusogenic Lipid Carriers, *Langmuir*, 2020, **36**(19), 5134–5144.

28 H. Abumanhal-Masarweh, D. Da Silva, M. Poley, A. Zinger, E. Goldman, N. Krinsky, R. Kleiner, G. Shenbach, J. E. Schroeder, J. Shklover, J. Shainsky-Roitman and A. Schroeder, Tailoring the Lipid Composition of Nanoparticles Modulates Their Cellular Uptake and Affects the Viability of Triple Negative Breast Cancer Cells, *J. Controlled Release*, 2019, **307**, 331–341.

29 C. M. Nycholat, C. Rademacher, N. Kawasaki and J. C. Paulson, In Silico-Aided Design of a Glycan Ligand of Sialoadhesin for in Vivo Targeting of Macrophages, *J. Am. Chem. Soc.*, 2012, **134**(38), 15696–15699.

30 F. Xu, A. Bandara, H. Akiyama, B. Eshaghi, D. Stelter, T. Keyes, J. E. Straub, S. Gummuluru and B. M. Reinhard, Membrane-Wrapped Nanoparticles Probe Divergent Roles of GM3 and Phosphatidylserine in Lipid-Mediated Viral Entry Pathways, *Proc. Natl. Acad. Sci. U. S. A.*, 2018, **115**(39), E9041–E9050.

31 H. M. Kim, H.-J. Choo, S.-Y. Jung, Y.-G. Ko, W.-H. Park, S.-J. Jeon, C. H. Kim, T. Joo and B. R. Cho, A Two-Photon Fluorescent Probe for Lipid Raft Imaging: C-Laurdan, *ChemBioChem*, 2007, **8**(5), 553–559.

32 M. M. Dodes Traian, F. L. Gonzalez Flecha and V. Levi, Imaging Lipid Lateral Organization in Membranes with C-Laurdan in a Confocal Microscope, *J. Lipid Res.*, 2012, **53**(3), 609–616.

33 L. A. Bagatolli and E. Gratton, Two Photon Fluorescence Microscopy of Coexisting Lipid Domains in Giant Unilamellar Vesicles of Binary Phospholipid Mixtures, *Bioophys. J.*, 2000, **78**(1), 290–305.

34 J. Barucha-Kraszewska, S. Kraszewski and C. Ramseyer, Will C-Laurdan Dethrone Laurdan in Fluorescent Solvent Relaxation Techniques for Lipid Membrane Studies?, *Langmuir*, 2013, **29**(4), 1174–1182.

35 E. Sezgin, T. Sadowski and K. Simons, Measuring Lipid Packing of Model and Cellular Membranes with Environment Sensitive Probes, *Langmuir*, 2014, **30**(27), 8160–8166.

36 K. Gaus, E. Gratton, E. P. W. Kable, A. S. Jones, I. Gelissen, L. Kritharides and W. Jessup, Visualizing Lipid Structure and Raft Domains in Living Cells with Two-Photon Microscopy, *Proc. Natl. Acad. Sci. U. S. A.*, 2003, **100**(26), 15554–15559.

37 S. Mukherjee and A. Chattopadhyay, Monitoring the Organization and Dynamics of Bovine Hippocampal Membranes Utilizing Laurdan Generalized Polarization, *Biochim. Biophys. Acta, Biomembr.*, 2005, **1714**(1), 43–55.

38 F. M. Harris, K. B. Best and J. D. Bell, Use of Laurdan Fluorescence Intensity and Polarization to Distinguish between Changes in Membrane Fluidity and Phospholipid Order, *Biochim. Biophys. Acta, Biomembr.*, 2002, **1565**(1), 123–128.

39 E. R. Moulton, K. J. Hirsche, M. L. Hobbs, J. M. Schwab, E. G. Bailey and J. D. Bell, Examining the Effects of Cholesterol on Model Membranes at High Temperatures: Laurdan and Patman See It Differently, *BBA, Biochim. Biophys. Acta, Biomembr.*, 2018, **1860**(8), 1571–1579.

40 N. Hamada, S. Gakhar and M. L. Longo, Hybrid Lipid/Block Copolymer Vesicles Display Broad Phase Coexistence Region, *BBA, Biochim. Biophys. Acta, Biomembr.*, 2021, **1863**(4), 183552.

41 H. A. Wilson-Ashworth, Q. Bahm, J. Erickson, A. Shinkle, M. P. Vu, D. Woodbury and J. D. Bell, Differential Detection of Phospholipid Fluidity, Order, and Spacing by Fluorescence Spectroscopy of Bis-Pyrene, Prodan, Nystatin, and Merocyanine 540, *Biophys. J.*, 2006, **91**(11), 4091–4101.

42 M. L. Schmidt, L. Ziani, M. Boudreau and J. H. Davis, Phase Equilibria in DOPC/DPPC: Conversion from Gel to Subgel in Two Component Mixtures, *J. Chem. Phys.*, 2009, **131**(17), 175103.

43 I. R. Calori, W. M. Pazin, K. Brunaldi, D. S. Pellosi, W. Caetano, A. C. Tedesco and N. Hioka, Laurdan as Fluorescent Probe to Determine the Critical Micelle Temperature of Polymers from Pluronic®-Coated Fluid Phase Liposomes, *J. Mol. Liq.*, 2019, **294**, 111562.

44 R. Sachl, M. Stepanek, K. Prochazka, J. Humpolickova and M. Hof, Fluorescence Study of the Solvation of Fluorescent Probes Prodan and Laurdan in Poly(Epsilon-Caprolactone)-Block-Poly(Ethylene Oxide) Vesicles in Aqueous Solutions with Tetrahydrofuran, *Langmuir*, 2008, **24**(1), 288–295.

45 G. Neunert, J. Tomaszecka-Gras, A. Baj, M. Gauza-Włodarczyk, S. Witkowski and K. Polewski, Phase Transitions and Structural Changes in DPPC Liposomes Induced by a 1-Carba-Alpha-Tocopherol Analogue, *Molecules*, 2021, **26**(10), 2851.

46 M. R. Vist and J. H. Davis, Phase Equilibria of Cholesterol/ Dopalmitoylphosphatidylcholine Mixtures: Deuterium Nuclear Magnetic Resonance and Differential Scanning Calorimetry, *Biochemistry*, 1990, **29**(2), 451–464.

47 T. P. W. McMullen and R. N. McElhaney, New Aspects of the Interaction of Cholesterol with Dipalmitoylphosphatidylcholine Bilayers as Revealed by High-Sensitivity Differential Scanning Calorimetry, *BBA, Biochim. Biophys. Acta, Biomembr.*, 1995, **1234**(1), 90–98.

48 S. J. Singer and G. L. Nicolson, The Fluid Mosaic Model of the Structure of Cell Membranes, *Science*, 1972, **175**(4023), 720–731.

49 J. Zhang, Q. Li, Y. Wu, D. Wang, L. Xu, Y. Zhang, S. Wang, T. Wang, F. Liu, M. Y. Zaky, S. Hou, S. Liu, K. Zou, H. Lei, L. Zou, Y. Zhang and H. Liu, Cholesterol Content in Cell Membrane Maintains Surface Levels of ErbB2 and Confers a Therapeutic Vulnerability in ErbB2-Positive Breast Cancer, *Cell Commun. Signaling*, 2019, **17**(1), 15.

50 M. Peyrot, A. M. Sautereau, J. M. Rabanel, F. Nguyen, J. F. Tocanne and D. Samain, Supramolecular Biovects (SMBV): A New Family of Nanoparticulate Drug Delivery



Systems, Synthesis and Structural Characterization, *Int. J. Pharm.*, 1994, **102**(1–3), 25–33.

51 P. R. Crocker, J. C. Paulson and A. Varki, Siglecs and Their Roles in the Immune System, *Nat. Rev. Immunol.*, 2007, **7**(4), 255–266.

52 L. Martinez-Pomares and S. Gordon, CD169+ Macrophages at the Crossroads of Antigen Presentation, *Trends Immunol.*, 2012, **33**(2), 66–70.

53 C. Sungkapreecha, M. J. Beily, J. Kressler, W. W. Focke and R. Androsch, Phase Behavior of the Polymer/Drug System PLA/DEET: Effect of PLA Molar Mass on Subambient Liquid-Liquid Phase Separation, *Thermochim. Acta*, 2018, **660**, 77–81.

54 H. Zang, M. Siddiqui, S. Gummuluru, W. W. Wong and B. M. Reinhard, Ganglioside-Functionalized Nanoparticles for Chimeric Antigen Receptor T-Cell Activation at the Immunological Synapse, *ACS Nano*, 2022, **16**(11), 18408–18420.

55 R. H. Fang, S. Aryal, C.-M. J. Hu and L. Zhang, Quick Synthesis of Lipid-Polymer Hybrid Nanoparticles with Low Polydispersity Using a Single-Step Sonication Method, *Langmuir*, 2010, **26**(22), 16958–16962.

56 A. D. Bangham, M. M. Standish and J. C. Watkins, Diffusion of Univalent Ions across the Lamellae of Swollen Phospholipids, *J. Mol. Biol.*, 1965, **13**(1), 238–252.

57 W. B. Puryear, H. Akiyama, S. D. Geer, N. P. Ramirez, X. Yu, B. M. Reinhard and S. Gummuluru, Interferon-Inducible Mechanism of Dendritic Cell-Mediated HIV-1 Dissemination Is Dependent on Siglec-1/CD169, *PLoS Pathog.*, 2013, **9**(4), e1003291.

