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RuC nanosheet as a promising biosensing material for detecting the aromatic amino acids: a DFT study†

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Density functional theory (DFT) calculations were performed to examine the potential of the RuC nanosheet as a biosensor towards the aromatic amino acids (AAA; tryptophan (TRP), histidine (HIS), tyrosine (TYR), and phenylalanine (PHE)). The AAA molecules were placed vertically and horizontally with respect to the RuC surface and then subjected to geometrical relaxation. According to the geometry relaxation results, it was found that all AAA molecules preferred to be adsorbed on the RuC surface in a horizontal configuration rather than a vertical one, except the HIS molecule, which desired to be vertically adsorbed on the RuC nanosheet. From the energy manifestations, the adsorption process within the TRP···RuC complexes had the greatest desired negative adsorption energy (E_{ads}), followed by HIS···, TYR···, and then PHE···RuC complexes ($E_{\text{ads}} = -40.22$, -36.54 , -23.95 , and -16.62 kcal mol⁻¹, respectively). As indicated by the FMO data, changes in the E_{HOMO} , E_{LUMO} , and E_{gap} values of the RuC nanosheet following the adsorption process demonstrated the capacity of the RuC nanosheet to adsorb the AAA molecules. The outcomes of Bader charge transfer revealed that the RuC nanosheet had the ability to donate electrons to the AAA molecules during the adsorption process, supported by the positive Q_t values. Consistent with the E_{ads} conclusions, the TRP···RuC complexes had the largest Q_t values, indicating the potential affinity of the RuC nanosheet to adsorb the TRP molecule. Following the adsorption of AAA molecules on the RuC nanosheet, new peaks and bands were discovered based on the DOS and the band structure plots, respectively, revealing the validity of the adsorption process. Additionally, the current adsorption findings on the RuC nanosheet were compared to those on the graphene (GN) nanosheet. The outcomes of the comparison demonstrated the outperformance of the RuC nanosheet over the GN nanosheet in adsorbing the AAA molecules. These outcomes provide a solid foundation for further research on the RuC nanosheets to detect small biomolecules.

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1. Introduction

The efficient exfoliation of the graphene (GN) nanosheet has paved an essential path toward the creation of low-dimensional nanomaterials.^{1–3} Motivated by the potential of GN in a wide range of practical applications, several free-standing 2D structures, planar and non-planar, have been constructed from their bulk equivalents.^{4,5} Eventually, this resulted in the formation of layered double hydroxides,^{6,7} MXenes,^{8,9} vertical and lateral van der Waals heterostructures,¹⁰ transition metal dichalcogenides (TMDs),¹¹ borophene,^{12–15} and several families of 2D materials.^{3,16} The distinct characteristics of the 2D materials have prompted additional research into the possibility of producing novel 2D materials, including transition metals and groups III and IV elements.^{9,17} Among these, carbides based on transition metals have been attracting much interest due to their promising characteristics, including energy storage and



superconductivity.¹⁸ The ruthenium carbide (RuC), a novel structure of 2D transition metal carbide formed by combining carbon and ruthenium atoms in a planar surface, was computationally hypothesized by Gorkan *et al.* in 2020.¹⁹ The RuC nanosheet has outstanding structural, mechanical, and electrical stabilities and is a very stable type of 2D transition metal carbide.¹⁹ The history of RuC goes back to the 1960s when the bulk ruthenium carbides were reported as superhard materials that could be synthesized at high temperatures and ambient pressure.²⁰ When using a laser-heated diamond anvil process at high temperature and pressure, Kumar *et al.* announced later in 2012 that they had synthesized Ru₂C stoichiometry in Fe₂N type hexagonal structure with the *P3m1* space group.²¹ Using a simple hydrothermal synthesis approach, the most recent study by Cored *et al.* examined the processes for fabricating and stabilizing surface ruthenium carbide species on a metallic Ru core.²² Moreover, no additional experimental research on ruthenium carbides has been completed. Computational analysis of bulk ruthenium carbides has been defined, involving studies of different stoichiometries such as rhombohedral RuC^{23–25} and hexagonal RuC, Ru₂C, and Ru₃C.^{23,26}

Utilization of the RuC nanosheet as a detector towards small molecules, such as NH₃, NO, and NO₂, was examined using density functional theory (DFT) computations.²⁷ The adsorption energy (E_{ads}) outcomes found that the NO adsorption over the RuC nanosheet had the most favorable negative E_{ads} of -1.718 eV.²⁷ According to our knowledge, no further studies have been carried out to investigate the potential of RuC nanosheets to adsorb biomolecules.

The way that biological molecules interact with nanomaterials is of great interest to researchers studying biomedical and biosensing applications.^{28–30} Among the biological molecules, the adsorption of nucleic acid molecules and proteins has garnered significant interest.^{31,32} Amino acids (AAs) are essential for all living things due to their important function as the building blocks of enzymes and proteins.^{31–33} These AAs also serve as important metabolic intermediates, such as in the synthesis of vitamins, hormones, and neurotransmitters.^{34,35} Among the AAs, the biogenic aromatic α -amino acids (AAA), including histidine (HIS), tyrosine (TYR), phenylalanine (PHE), and tryptophan (TRP), are crucial residues containing aromatic systems that determine how protein residues interact with the environment.^{36–38}

The application of 2D nanosheets as a biosensor for AAA molecules has garnered significant attention. Therefore, the current work was dedicated to evaluating the adsorption features of the AAA molecules (AAA; tryptophan (TRP), histidine (HIS), tyrosine (TYR), and phenylalanine (PHE)) on the RuC nanosheet by means of DFT calculations (Fig. 1). To determine the minimal structures, relax computations were first applied to the geometric structures of AAA···RuC complexes. Subsequently, the adsorption energies were determined for each complex under investigation. To clarify how the adsorption process affects the properties of the studied RuC nanosheet, computations of Bader charge transfer, density of state (DOS), frontier molecular orbital (FMO), and band structures were carried out. In addition, a comparison with the GN nanosheet

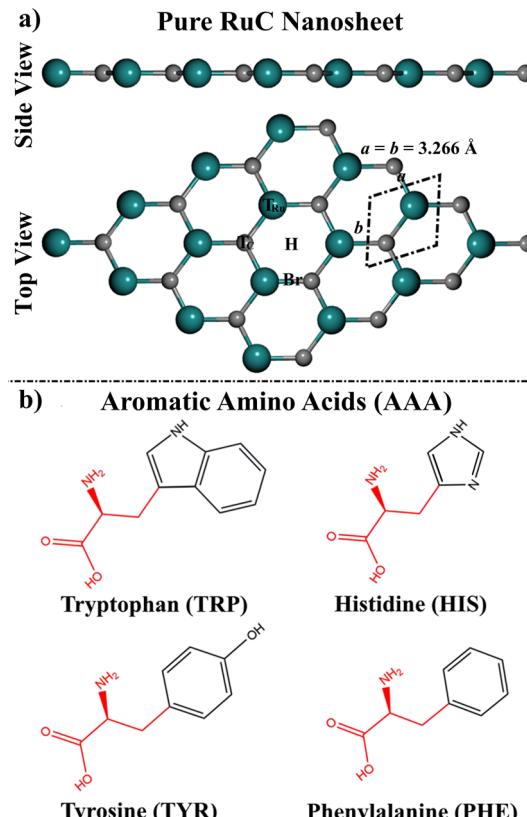


Fig. 1 (a) Side and top perspectives of a $4 \times 4 \times 1$ supercell of the RuC nanosheet. The dashed rhombus represents the unit cell of the RuC. The adsorption sites of the top, bridge, and hollow are represented as T, H, and Br, respectively. (b) 2D chemical structures of the studied aromatic amino acids (AAA; tryptophan (TRP), histidine (HIS), tyrosine (TYR), and phenylalanine (PHE)). In our models, the red fragments were replaced by a hydrogen atom.

provided more insight into the efficiency of the RuC nanosheet in adsorbing the AAA molecules. Thus, the AAA···GN complexes were relaxed, and their corresponding E_{ads} were evaluated. The outcomes of this investigation will provide insightful information on the potential applications of the RuC nanosheet in sensing biological molecules in the future.

2. Computational methodology

All computations were carried out using Quantum ESPRESSO 7.1 package^{39,40} utilizing Density Functional Theory (DFT) methods.^{41,42} DFT calculations were employed by using the projector augmented wave (PAW).⁴³ Implementing the Perdew–Burke–Ernzerhof (PBE) exchange–correlation function, Generalized Gradient Approximation (GGA) was utilized.⁴⁴ All calculations were supported with the Grimme-D3 correction to adjust the van der Waals interactions.^{45,46} The charge density and energy cutoffs were determined at 400 Ry and 50 Ry, respectively. The force and energy convergence criteria for all computations were 10^{-4} eV Å⁻¹ and 10^{-5} eV, respectively. For structure relaxation and density of state computation, the Brillouin zone was sampled using Monkhorst–Pack grids of $5 \times 5 \times 1$ and $15 \times 15 \times 1$ k -points, respectively. The Marzari–



Vanderbilt smearing technique was implemented in relaxation computations.⁴⁷ For all calculations, a 20 Å vacuum was placed along the vertical direction of the RuC nanosheet to prevent interactions between the periodic images.

For the purpose of investigating the adsorption of AAA molecules over the RuC nanosheet, the optimized RuC unit cell was widened to a 4 × 4 × 1 supercell with 16 Ru and 16 C atoms (Fig. 1). The AAA molecules were then placed vertically and horizontally with respect to the RuC surface to explore all possible adsorption configurations. Adsorption energies (E_{ads}) were then assessed as follows:

$$E_{\text{ads}} = E_{\text{AAA} \cdots \text{RuC}} - (E_{\text{AAA}} + E_{\text{RuC}}) \quad (1)$$

where $E_{\text{AAA} \cdots \text{RuC}}$, E_{AAA} , E_{RuC} are the energies of the AAA···RuC complex, AAA molecule, and RuC nanosheet, respectively. Bader charge analysis^{48,49} was carried out to assess the charge transfer to/from the RuC nanosheet according to the following equation:

$$Q_t = Q_{(\text{combined RuC})} - Q_{(\text{isolated RuC})} \quad (2)$$

where the $Q_{(\text{combined RuC})}$ and $Q_{(\text{isolated RuC})}$ are the charge of the RuC nanosheet after and prior to the adsorption process of AAA molecules, respectively. Maps representing the charge density difference ($\Delta\rho$) were created relating to the subsequent equation:

$$\Delta\rho = \rho_{\text{AAA} \cdots \text{RuC}} - \rho_{\text{AAA}} - \rho_{\text{RuC}} \quad (3)$$

where $\rho_{\text{AAA} \cdots \text{RuC}}$, ρ_{AAA} , and ρ_{RuC} demonstrate the charge density of complex, AAA molecule, and the RuC nanosheet, respectively. Employing the VESTA visualization software, the charge density maps were generated.⁵⁰ Frontier molecular orbital (FMO) computations were executed to get additional insight into the interaction between adsorbates and an adsorbent.⁵¹ For the relaxed AAA···RuC complexes, the energies of the lowest unoccupied molecular orbital (E_{LUMO}) and highest occupied molecular orbital (E_{HOMO}) were calculated throughout the FMO studies, and the energy gap (E_{gap}) was also estimated as follows:

$$E_{\text{gap}} = E_{\text{LUMO}} - E_{\text{HOMO}} \quad (4)$$

Band structure and the density of states (DOS) calculations were also investigated to comprehend the electronic properties of the RuC nanosheet. The desorption of the AAA molecules from the RuC nanosheet was assessed by computing the recovery time (τ) for the AAA···RuC complexes as follows:

$$\tau = \nu^{-1} \exp(-E_{\text{ads}}/KT) \quad (5)$$

where the value of attempt frequency (ν) is 10^{18} s⁻¹, and Boltzmann constant (K) is 0.00199 kcal mol⁻¹ K⁻¹. T represents root temperature (298.15 K) and human body temperature (310.15 K).

Further comparative analysis of the adsorption efficiency of the RuC and GN nanosheets toward AAA molecules was carried out. Therefore, geometrical relaxation, E_{ads} , and Bader charge calculations were conducted for the AAA···GN complexes.

3. Results and discussion

3.1. Geometric structures

Prior to the adsorption process of AAA molecules, the 4 × 4 × 1 supercell of the RuC nanosheet was modeled and relaxed. The optimal structure of the RuC nanosheet is presented in Fig. 1a.

From the optimized RuC unit cell, it was found that the Ru–C bond length and lattice constant were 1.906 Å and $a = b = 3.266$ Å, respectively. The aforementioned findings were in line with previous investigations, in which the lattice parameter was $a = 3.25$ and 3.24 Å.^{19,27} According to the relaxed RuC nanosheet, four adsorption sites can be observed: two top (T_{Ru} and T_{C}), bridge (Br), and hollow (H) sites (Fig. 1).

3.2. Adsorption energy

The adsorption of AAA molecules over the RuC nanosheet was investigated at all possible adsorption sites (Fig. S1†). The adsorption energies (E_{ads}) and associated equilibrium adsorption distances for the relaxed AAA···RuC complexes were computed (Fig. S1†). The structures and the atomic coordinates of the most stable relaxed AAA···RuC complexes are given in Fig. 2 and Table S1,† respectively. All AAA molecules preferred to be adsorbed on the RuC nanosheet through their aromatic rings except the HIS molecule, which showed more desirability to be adsorbed through the lone pair electrons on its nitrogen atom.

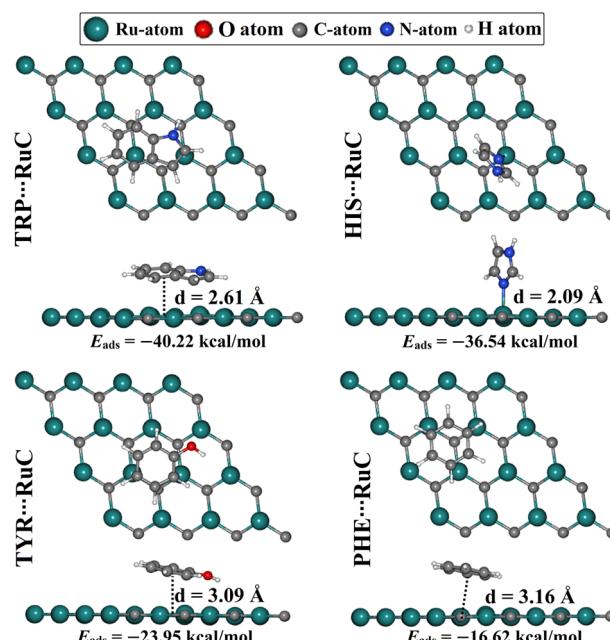


Fig. 2 Side and top representations of the relaxed AAA···RuC complexes (AAA; tryptophan (TRP), histidine (HIS), tyrosine (TYR), and phenylalanine (PHE)). The adsorption energies (E_{ads}) and the equilibrium adsorption distances (d) between the AAA molecules and the RuC nanosheet are in kcal mol⁻¹ and Å, respectively. The d values were measured between the centroid of the AAA rings and the RuC surface, except for the HIS···RuC, d was measured between the N atom of the HIS molecule and the RuC surface.



The considerable capability of the RuC nanosheet to adsorb AAA molecules was confirmed by the negative values of E_{ads} for all AAA···RuC complexes under investigation. Among the adsorption processes of the AAA molecules over the RuC nanosheet, the most significant negative E_{ads} value of $-40.22 \text{ kcal mol}^{-1}$ was denoted for the TRP···RuC complex. The superior adsorption behavior of TRP, compared to the other AAA molecules, aligns with the results of an earlier study on the adsorption of AAA molecules on a graphene nanosheet.⁵² The size of the π -system of the studied AAA molecules might be the reason for their favorability of being adsorbed on the RuC nanosheet.⁵³ In this regard, the RuC nanosheet was more suitable for adsorbing molecules containing two aromatic rings (*i.e.*, TRP) than those containing a single aromatic ring (*i.e.*, HIS, TYR, and PHE). Turning to the second potent AAA molecule, the HIS adsorption over the RuC nanosheet showed a negative E_{ads} value of $-36.54 \text{ kcal mol}^{-1}$ and an equilibrium adsorption distance of 2.09 \AA . For TYR, its oxygen atom enhanced the adsorption process on the RuC nanosheet, demonstrating the third promising negative E_{ads} value of $-23.95 \text{ kcal mol}^{-1}$. Finally, the PHE···RuC complex had the smallest negative E_{ads} value of $-16.62 \text{ kcal mol}^{-1}$ since the PHE molecule contains only carbon and hydrogen as heteroatoms. Remarkably, it was observed that the adsorption distance within the AAA···RuC complexes decreased as the negative E_{ads} increased. Numerically, the equilibrium adsorption distances were 3.16 , 3.09 , and 2.61 \AA within PHE···, TYR···, and TRP···RuC complexes, with E_{ads} values of -16.62 , -23.95 , and $-40.22 \text{ kcal mol}^{-1}$, respectively (Fig. 2). For the latter complexes, the correlation coefficient (R^2) value between the E_{ads} values and the adsorption distances was 0.93 . However, the HIS···RuC complex showed the smallest adsorption distance of 2.09 \AA , due to the adsorption of HIS *via* the lone pair of electrons on its nitrogen atom over the RuC nanosheet (Fig. 2).

3.3. Frontier molecular orbital (FMO) calculations

Frontier molecular orbitals (FMO) theory was utilized to better understand how the adsorption process of the AAA molecules impacted the electronic features of the RuC nanosheet. The energies of the highest occupied molecular orbitals (E_{HOMO}), the lowest unoccupied molecular orbitals (E_{LUMO}), and energy gap (E_{gap}) values were determined (Table 1). Furthermore, the HOMO and LUMO distributions prior to and following the adsorption process were produced to comprehend the electron transfer regioselectivity for the systems under investigation (Fig. 3).

According to the findings in Table 1, the E_{HOMO} , E_{LUMO} , and E_{gap} values of the studied systems showed significant changes before and following the adsorption process. As an example, the E_{HOMO} value of the pure RuC nanosheet was -3.412 eV , while the TRP···, HIS···, TYR···, and PHE···RuC complexes had E_{HOMO} values of -2.534 , -2.488 , -2.873 , and -2.899 eV , respectively (Table 1). Further, the E_{gap} values of the AAA molecules and RuC nanosheet altered following the adsorption process, revealing the validity of the adsorption process. In this regard, the E_{gap} of the pure RuC nanosheet had a value of

Table 1 E_{HOMO} , E_{LUMO} , and E_{gap} of both the isolated adsorbent/adsorbate and the relaxed AAA···RuC complexes (AAA: tryptophan (TRP), histidine (HIS), tyrosine (TYR), and phenylalanine (PHE))

System	E_{HOMO} (eV)	E_{LUMO} (eV)	E_{gap} (eV)
Isolated systems			
Pure RuC	-3.412	-3.341	0.071
Tryptophan (TRP)	-4.794	-1.032	3.762
Histidine (HIS)	-5.582	-0.705	4.876
Tyrosine (TYR)	-5.342	-1.071	4.271
Phenylalanine (PHE)	-6.139	-1.039	5.100
Combined systems^a			
TRP···RuC	-2.534	-2.479	0.056
HIS···RuC	-2.488	-2.470	0.016
TYR···RuC	-2.873	-2.866	0.007
PHE···RuC	-2.899	-2.790	0.109

^a The relaxed AAA···RuC complexes are shown in Fig. 2.

0.071 eV that decreased following the adsorption of TRP to 0.056 eV (Table 1). Interestingly, the E_{gap} was indicated with modest values, implying that the charge might be transferred within the complex.

The propensity of the molecule to donate electrons is shown by a high E_{HOMO} value, whereas a low E_{LUMO} value reveals the tendency of the molecule to accept electrons. Besides, a small E_{gap} value demonstrates a considerable chemical reactivity of the molecule. According to data listed in Table 1, TRP showed the highest chemical reactivity, followed by TYR, HIS, and then PHE, with E_{gap} values of 3.762 , 4.271 , 4.876 , and 5.100 eV , respectively.

From Fig. 3a, the LUMO distributions were located on the C–N, N–H, C–H, and C–O bonds of the AAA molecules, suggesting that these atoms served as locations for accepting electrons during the adsorption process over the RuC nanosheet. The amount of the LUMO distribution on the TRP molecule was more than that of the other AAA molecules, indicating its significant electron-accepting property. For the relaxed AAA···RuC complexes, the LUMO orbitals distribution can be observed on the Ru atoms of the nanosheet, demonstrating their significant role in the adsorption process (Fig. 3b).

3.4. Charge transfer calculation

Bader charge analysis is regarded as a useful technique for figuring out how charges are transferred during the adsorption process.^{48,49} Consequently, the charge transfer (Q_t) was determined for all relaxed AAA···RuC complexes (Fig. 4).

According to the charge transfer calculations, all estimated Q_t values were positive, indicating that the charge was moved from the RuC nanosheet to the adsorbed AAA molecules. In line with the E_{ads} results, the Q_t values increased in order: PHE··· < TYR··· < HIS··· < TRP···RuC complexes. In terms of values, the Q_t values of the PHE···, TYR···, HIS···, and TRP···RuC complexes were 0.1055 , 0.1461 , 0.1926 , and $0.2820e$, respectively (Fig. 4). Overall, the RuC nanosheet within the AAA···RuC complexes showed to have an electron-donating nature by the positive Q_t values. To examine the distribution of charge for the



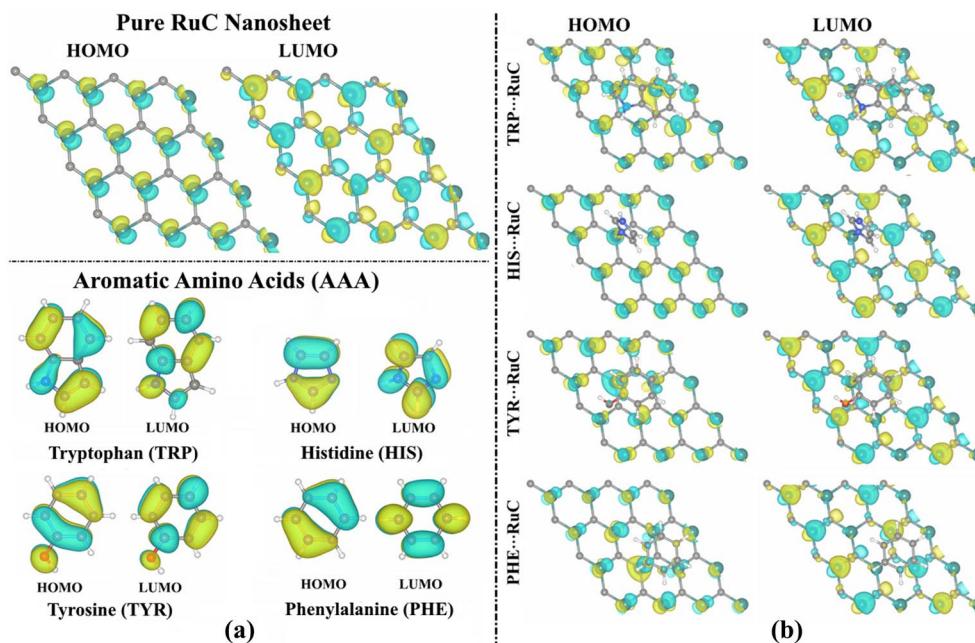


Fig. 3 HOMO and LUMO distributions of (a) pure RuC nanosheet and isolated AAA molecules, and (b) the relaxed AAA...RuC complexes (AAA; tryptophan (TRP), histidine (HIS), tyrosine (TYR), and phenylalanine (PHE)).

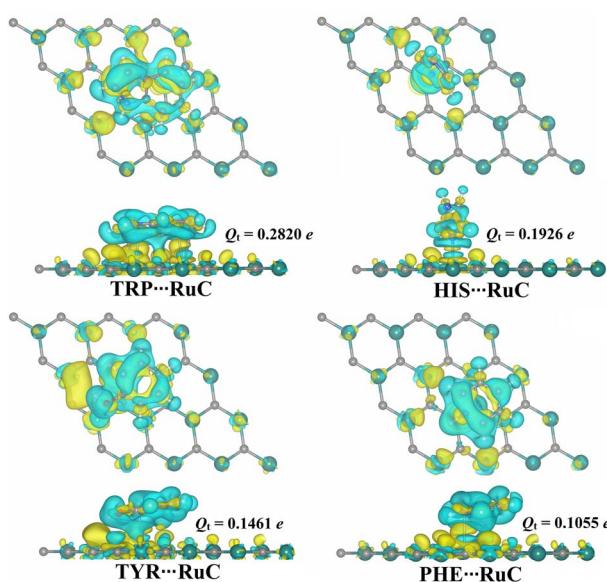


Fig. 4 Top and side perspectives of the charge density difference ($\Delta\rho$) maps for the relaxed AAA...RuC complexes (AAA; tryptophan (TRP), histidine (HIS), tyrosine (TYR), and phenylalanine (PHE)). Sites of electron depletion and accumulation are shown by cyan and yellow-colored regions, respectively. Charge transfer difference (Q_t) values for the RuC nanosheet before and after the adsorption of AAA molecules are in e.

relaxed AAA...RuC complexes, maps of the charge density difference ($\Delta\rho$) were plotted and shown in Fig. 4.

In Fig. 4, positive (yellow) and negative (cyan) regions represent the regions of electron accumulation and depletion, respectively. The most remarkable electron accumulation

region was observed in the TRP...RuC complex, demonstrating the superior ability of the TRP molecule to be adsorbed on the studied RuC nanosheet compared to the other AAA molecules. In all investigated AAA...RuC complexes, the electron depletion region was observed above the surface of the RuC nanosheet, demonstrating its electron-donating property (Fig. 4).

3.5. Recovery time

Recovery time (τ) computations can also be carried out to assess the desorption of AAA molecules from the RuC nanosheet. As a result, the τ was estimated for the relaxed AAA...RuC complexes at two different temperatures, and the findings are listed in Table 2.

Based on eqn (5), the τ results in Table 2 demonstrated the direct correlation between the τ and the E_{ads} outcomes; τ increased as negative E_{ads} values increased. For instance, the TRP...RuC complex, which had the most significant negative E_{ads} with a value of $-40.22\text{ kcal mol}^{-1}$, exhibited the longest τ with a value of 2.75×10^{11} and $2.00 \times 10^{10}\text{ s}$ at the room and

Table 2 Recovery time (τ , in s) for the relaxed AAA...RuC complexes at room temperature (298.15 K) and human body temperature (310.15 K)

Complexes ^a	Recovery time (s)	
	$T = 298.15\text{ K}$	$T = 310.15\text{ K}$
TRP...RuC	2.75×10^{11}	2.00×10^{10}
HIS...RuC	5.58×10^8	5.15×10^7
TYR...RuC	3.39×10^{-6}	7.12×10^{-2}
PHE...RuC	1.46×10^{-6}	4.95×10^{-7}

^a The relaxed AAA...RuC complexes are presented in Fig. 2.



human body temperatures, respectively. As observed in Table 2, the τ values decreased as the temperature increased. For example, the τ of the PHE \cdots RuC complex was 1.46×10^{-6} s at room temperature (*i.e.*, 298.15 K) and became 4.95×10^{-7} s at human body temperature (*i.e.*, 310.15 K). The τ results validated the affinity of the RuC nanosheet to adsorb AAA molecules effectively.

3.6. Band structure

Band structures were carried out before and after the adsorption of the AAA molecules over the RuC nanosheet to find out how the adsorbates changed the electronic characteristics of the adsorbent (Fig. 5).

For all AAA \cdots RuC complexes, the density of valence bands near the Fermi level increased as the adsorption energy increased following the order PHE \cdots < TYR \cdots < HIS \cdots < TRP \cdots RuC complexes (Fig. 5b). Upon the adsorption of the TRP molecule over the RuC surface, additional valence and

conduction bands were observed in its band structure. For instance, additional bands were noted in the valence region at around -1.60 and -1.80 eV (Fig. 5b). A significant shift in valence bands (from -0.03 to 0.00 eV) can also be observed for the TRP \cdots RuC complex (Fig. 5b).

New valence bands appeared in the band structure of the HIS \cdots RuC complex, reflecting the interaction of the HIS molecule with the pure RuC nanosheet. Additional valence bands were observed at around -1.65 eV and between -0.25 to -0.80 eV, whereas new conduction bands were identified at approximately 1.75 and 1.07 eV (Fig. 5b). The valence and conduction bands of the HIS \cdots RuC complex were moved nearer the Fermi level, indicating the considerable of the HIS adsorption over the RuC nanosheet. As noted in the band structures depicted in Fig. 5, the electronic characteristics of the RuC nanosheet seemed to be impacted by the adsorption of TYR and PHE molecules, exhibiting an increase in valence and conduction bands. The E_{gap} findings supported the observed band

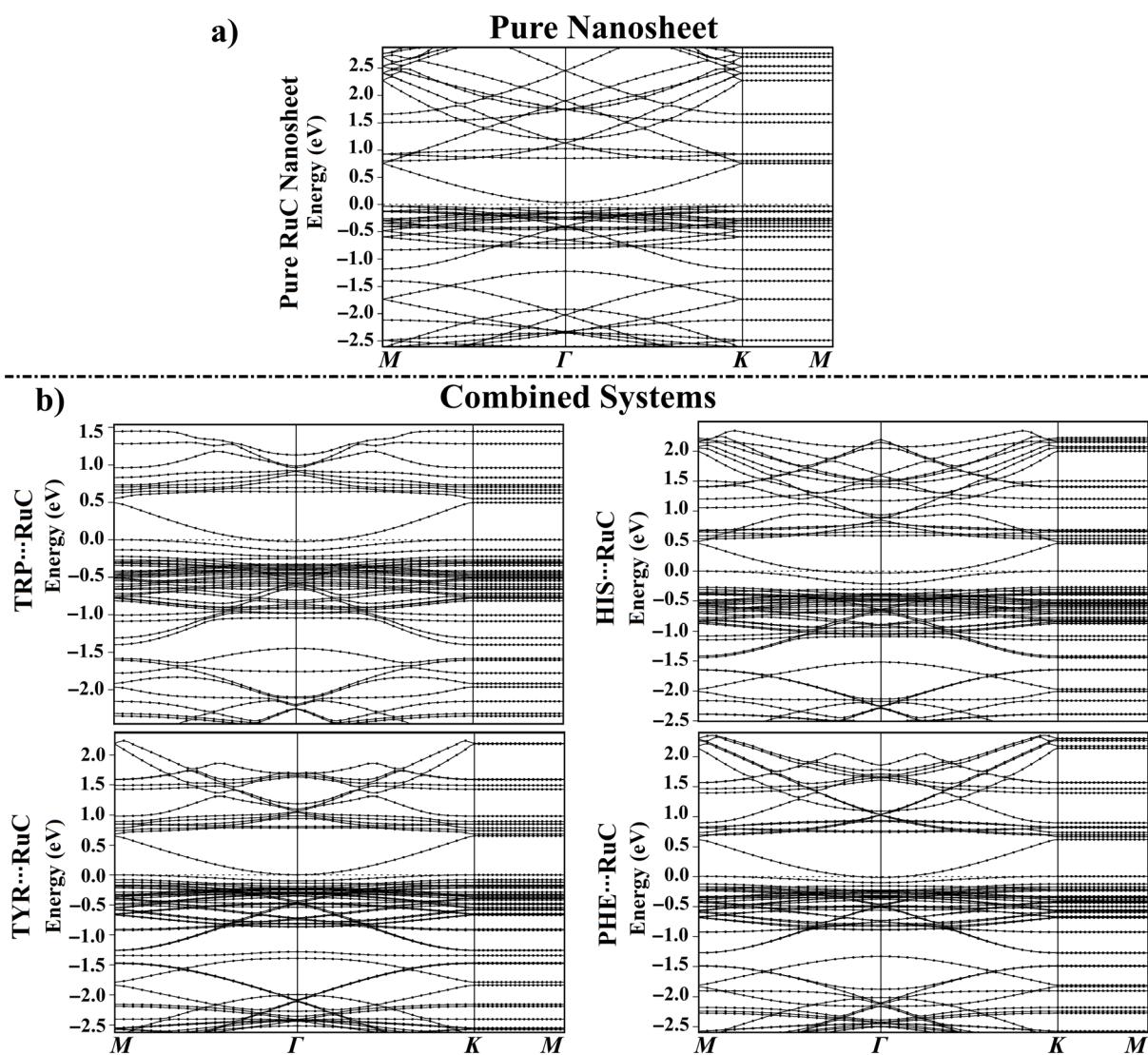


Fig. 5 Electronic band structure along the high symmetry path of the Brillouin zone ($M-\Gamma-K-M$) for (a) the pure RuC nanosheet and (b) the relaxed AAA \cdots RuC complexes (AAA; tryptophan (TRP), histidine (HIS), tyrosine (TYR), and phenylalanine (PHE)). The Fermi level is shown at zero.



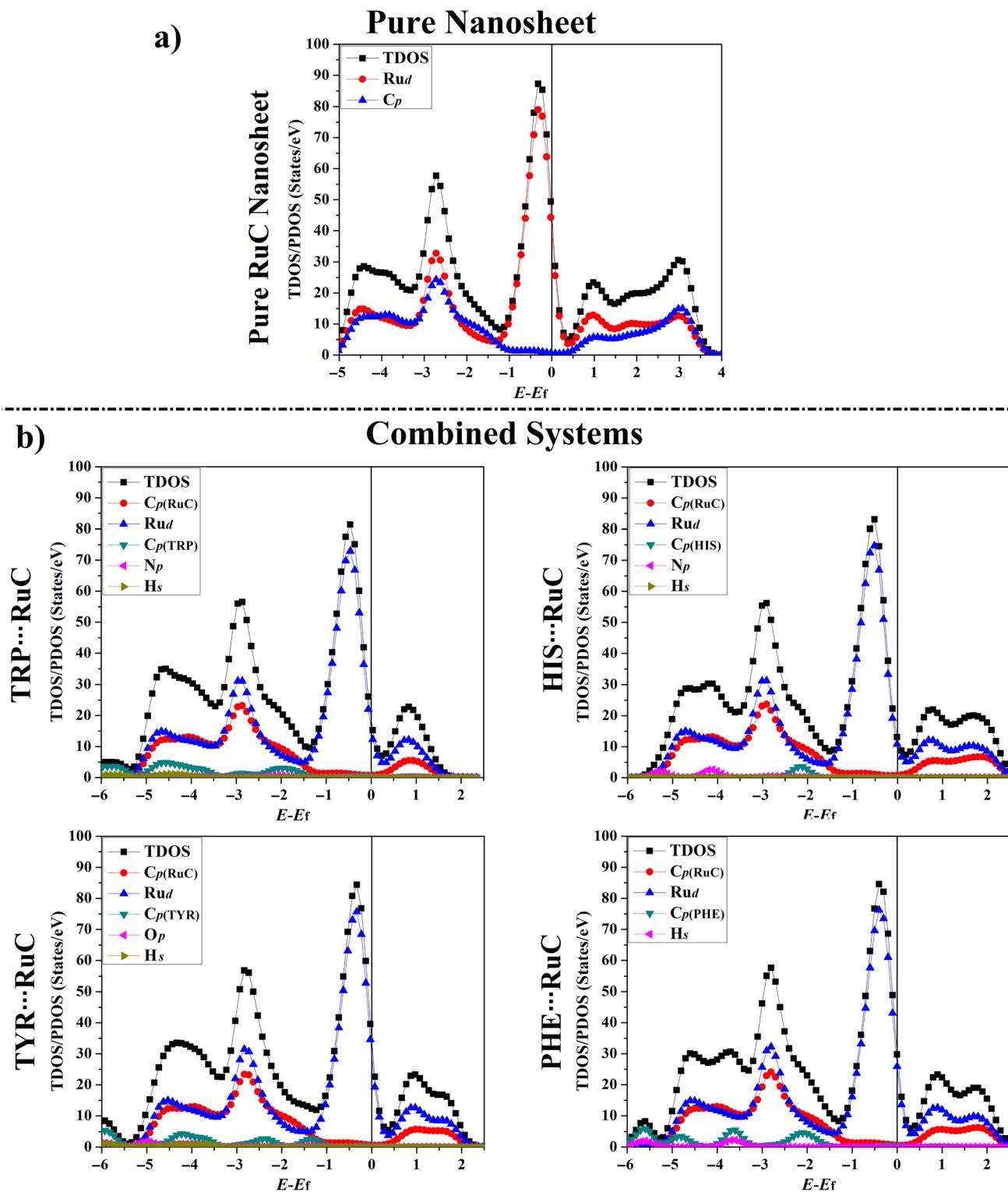


Fig. 6 TDOS and PDOS plots for (a) the pure RuC nanosheet and (b) the relaxed AAA...RuC complexes (AAA: tryptophan (TRP), histidine (HIS), tyrosine (TYR), and phenylalanine (PHE)). The contributions of the d-orbital of ruthenium (Ru_d), p-orbital of carbon (C_p), nitrogen (N_p), oxygen (O_p), and the s-orbital for hydrogen (H_s) in the adsorption process. The Fermi level is located at zero.

shifts; for example, the E_{gap} of the pure RuC nanosheet was 0.071 eV, which shifted to 0.056, 0.016, 0.007, and 0.109 eV within the TRP..., HIS..., TYR..., and PHE...RuC complexes (Table 1).

3.7. Density of state (DOS) calculations

The total density of states (TDOS) and projected density of states (PDOS) plots for the RuC nanosheet prior to and following the

adsorption were generated to determine how the electrical characteristics of the RuC nanosheet were affected during the adsorption of the AAA molecules (Fig. 6).

As seen in Fig. 6a, the PDOS plot of the pure RuC nanosheet revealed that the d-orbitals of the Ru atoms are primarily responsible for the TDOS near the Fermi level. According to Fig. 6b, it can be seen that the Ru_d, N_p, O_p, and C_p had significant contributions to the adsorption process within the relaxed AAA···RuC complexes, and their PDOS peaks appeared between −6.00 and 2.50 eV. As an example, the PDOS plot of the TRP···RuC complex showed that the Ru_d and C_p of the RuC nanosheet were the primary origins of the adsorption process. The C_p of the TRP molecule had the largest contribution to the adsorption process on the RuC nanosheet, appearing in the valence region at energy ranges of −1.50 to −2.50, −3.50 to −5.00, and −5.50 to −7.00 eV (Fig. 6b). The H_s and N_p of the TRP molecule within the TRP···RuC complex showed minor contributions that can be observed in the valence region from −1.50 to −7.00 eV. Obviously, an overlap between the C_p of the RuC nanosheet and the C_p, N_p, and H_s of the TRP molecule was noted from −0.50 to −1.40 eV in the valence region. The latter observation confirmed the strong interaction between the TRP molecule and the RuC nanosheet.

3.8. Adsorption process on graphene (GN) nanosheet

As a point of comparison, the AAA···GN complexes were relaxed at the same configuration as the AAA···RuC complexes, and

then their corresponding E_{ads} values were evaluated (Fig. 7). The atomic coordinates of the relaxed AAA···GN complexes are given in Table S1.†

According to the results, the negative E_{ads} of the AAA···GN complexes increased in the order of HIS··· < PHE··· < TYR··· < TRP···GN complexes with E_{ads} values of −7.63, −9.13, −10.08, and −12.55 kcal mol^{−1}, respectively. The latter results were consistent with the prior study.⁵² In terms of comparison, the E_{ads} values of the AAA···GN complexes in the modeled configurations were less negative than those of the AAA···RuC complexes, demonstrating the superior adsorption efficacy of the RuC nanosheet toward the AAA molecules. For instance, the E_{ads} values of the TRP···RuC and TRP···GN complexes were −40.22 and −12.55 kcal mol^{−1}, respectively.

Upon the relaxed AAA···GN complexes, the Q_t values were computed within the Bader charge method (Fig. 8). Based on the Bader charge results, all AAA···GN complexes showed positive Q_t values, demonstrating that GN nanosheet acted, similarly to RuC nanosheet, as an electron donor during the adsorption process. In this spirit, the Q_t values of the TRP···, TYR···, PHE···, and HIS···GN complexes were 0.0156, 0.0053, 0.0072, and 0.0065 e, respectively (Fig. 8). Additionally, the $\Delta\rho$ maps were produced for the relaxed AAA···GN complexes and are illustrated in Fig. 8. As shown in Fig. 8, cyan regions were observed above the GN nanosheet within all relaxed AAA···GN complexes, demonstrating the electron-donating nature of the GN nanosheet during the adsorption process. Similar to the AAA···RuC complexes, the TRP···GN complex also exhibited the largest amount of the transferred charge supported by the highest Q_t value of 0.0156 e (Fig. 8).

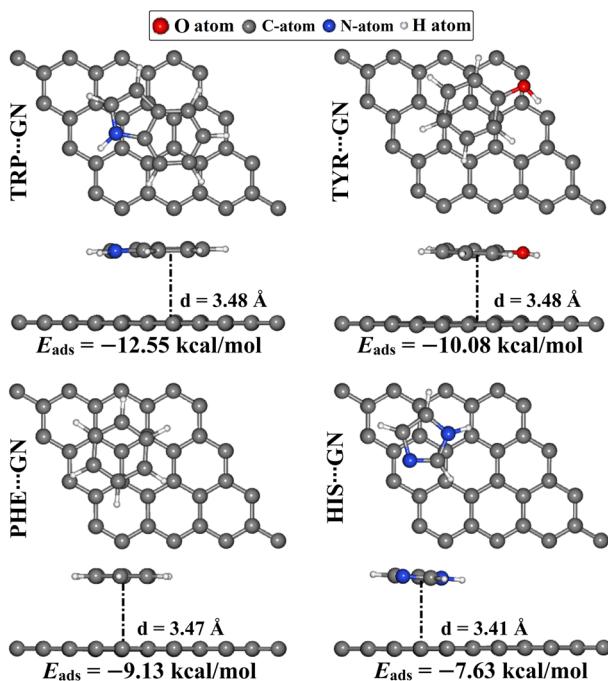


Fig. 7 Top and side representations of the relaxed AAA···GN complexes (AAA: tryptophan (TRP), histidine (HIS), tyrosine (TYR), and phenylalanine (PHE)). The adsorption energies (E_{ads}) and the equilibrium adsorption distances (d) between the AAA molecules and the GN nanosheet are in kcal mol^{−1} and Å, respectively. The d values are measured between the centroid of the AAA rings and the GN surface.

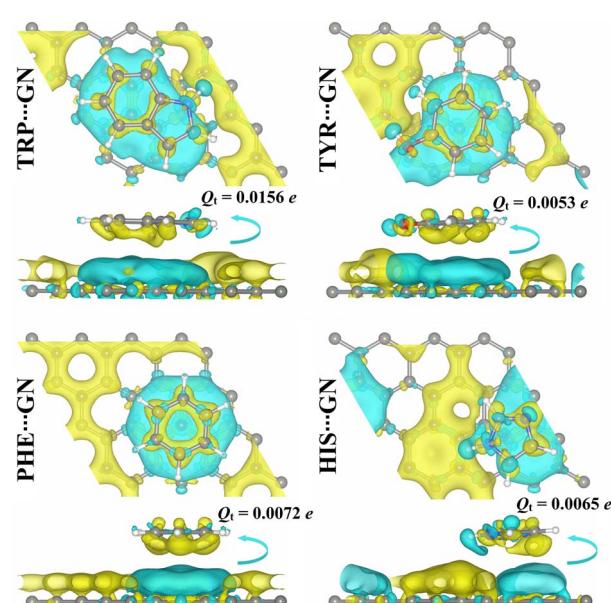


Fig. 8 Top and side perspectives of the charge density difference ($\Delta\rho$) maps for the relaxed AAA···GN complexes (AAA: tryptophan (TRP), histidine (HIS), tyrosine (TYR), and phenylalanine (PHE)). Sites of electron depletion and accumulation are shown by cyan and yellow-colored regions, respectively. Charge transfer difference (Q_t) values for the GN nanosheet prior to and following the adsorption of AAA molecules are in e.



4. Conclusion

AAA molecules are building blocks for synthesizing numerous biologically active compounds necessary to preserve regular biological processes. Based on their significance, the propensity of the RuC nanosheet as a biosensor for different AAA molecules was assessed within both horizontal and vertical configurations employing the density functional theory (DFT). Based on the relaxed structures, it was found that the AAA molecules favored their horizontal configuration within the adsorption process on the RuC nanosheet, while the HIS molecule desired to be adsorbed vertically. As per E_{ads} outcomes, the TRP···RuC complexes showed the greatest negative E_{ads} value of $-40.22\text{ kcal mol}^{-1}$. From the FMO data, notable changes in the E_{HOMO} , E_{LUMO} , and E_{gap} values of the RuC nanosheet following the adsorption process demonstrated the capability of the RuC nanosheet to adsorb the AAA molecules. According to Bader charge transfer analysis, the RuC nanosheet functioned as an electron donor during the adsorption process, supported by positive Q_t values. In line with the E_{ads} values, the TRP···RuC complex had the largest Q_t value of $0.2820e$. The adsorption of AAA molecules on the RuC nanosheet was also confirmed by the band structure and DOS analysis, supported by the appearance of additional bands and peaks in their plots, respectively. The outcomes of a comparative investigation revealed that the RuC nanosheet outperformed the GN nanosheet in adsorbing the AAA molecules. These findings lay a solid basis for further investigations into the potential of the RuC nanosheet as an effective biosensor for small biomolecules.

Data availability

The data supporting this article have been included as ESI.†

Author contributions

Conceptualization, M. A. A. I. and T. S.; methodology, M. A. A. I., A. H. M. M., and M. E. S. S.; software, M. A. A. I.; formal analysis, N. K. M. A. and A. H. M. M.; investigation, N. K. M. A., A. H. M. M., and S. K.; resources, M. A. A. I., M. A. E.-T., A. M. M. A. and T. S.; data curation, N. K. M. A. and A. H. M. M.; visualization, N. K. M. A. and A. H. M. M., and M. E. S. S.; supervision, M. A. A. I.; project administration, M. A. A. I. and A. H. M. M.; writing—original draft, N. K. M. A. and A. H. M. M.; writing—review and editing, M. A. A. I., M. A. E.-T., A. M. M. A., S. K., M. E. S. S. and T. S.; all authors have read and agreed to the published version of the manuscript.

Conflicts of interest

The authors declare no competing interests.

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