



Cite this: *Mater. Adv.*, 2024,
5, 1480

Received 24th October 2023,
Accepted 22nd January 2024

DOI: 10.1039/d3ma00899a

rsc.li/materials-advances

3D bioprinting cowpea mosaic virus as an immunotherapy depot for ovarian cancer prevention in a preclinical mouse model†

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Implantable polymeric hydrogels loaded with immunostimulatory cowpea mosaic virus (CPMV) were fabricated using digital light processing (DLP) printing technology. The CPMV-laden hydrogels were surgically implanted into the peritoneal cavity to serve as depots for cancer slow-release immunotherapy. Sustained release of CPMV within the intraperitoneal space alleviates the need for repeated dosing and we demonstrated efficacy against ovarian cancer in a metastatic mouse model.

Serous ovarian cancer is a deadly malignancy for women, and about 80% of patients are diagnosed at advanced stages with metastatic disease in the peritoneal space.^{1–3} Standard care for women with ovarian cancer is surgical debulking followed by platinum-based chemotherapy. Although these treatments can provide a short period of remission, up to 70% of patients

experience recurrence.^{3,4} Despite the advances of immunotherapy for cancer treatment, there is no established immunotherapy for ovarian cancer. There is a dire need for therapeutic innovation and development of treatment strategies that prolong survival and prevent recurrence. Immunotherapy is an undeveloped therapeutic option for this disease.

Cancer immunotherapy is fundamentally different from and has multiple advantages over cytotoxic chemotherapy. Immunotherapy empowers the patient's own immune system to recognize and eliminate tumors and metastasis and when anti-tumor immune memory is established, treatment responses can be durable preventing recurrence. Using mouse models of aggressive metastatic ovarian cancer, we have previously reported that the plant virus, cowpea mosaic virus (CPMV), is a potent cancer immunotherapy that generates durable anti-tumor immune responses against ovarian cancer. Weekly intraperitoneal (i.p.) dosing of immunomodulatory CPMV nanoparticles effectively reprograms the ovarian tumor microenvironment (TME) within the i.p. space, leading to survival benefit with inhibited tumor growth, extended survival, and protection demonstrated by re-challenge experiments.^{5,6} The potent efficacy of CPMV has been established in various solid tumor types including in canine patients with spontaneous tumors.^{5–10} Mechanism studies demonstrate that CPMV is a toll-like receptor agonist of TLR2, 4, and 7 and this initially stimulates and recruits immune stimulatory innate immune cells to the tumor sites. Tumor cell killing is mediated by these innate immune cells, which then become antigen-presenting cells to establish adaptive and systemic anti-tumor immunity.^{11,12} To achieve this potency and effectively overcome the immunosuppressed TME, repeated i.p. injections are required. However, this bears a translational barrier because i.p. administration in humans is a complex procedure and requires hospitalization that increases cost and decreases quality of life;¹³ repeated i.p. dosing will be challenging to implement in any clinical treatment plan. To overcome this technological barrier, we turned toward the formulation of CPMV hydrogels to form an intraperitoneal depot

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† Electronic supplementary information (ESI) available: Materials and methods, characterization of CPMV and CPMV-Cy5, recorded mice body weight in the first efficacy study, and averaged circumference and survival rate for the second study. See DOI: <https://doi.org/10.1039/d3ma00899a>

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that mediates sustained release, thus alleviating the need for repeated dosing. Given that women with ovarian cancer almost always require surgeries to remove tumor tissues, such a CPMV hydrogel biomedical device could be readily implanted post-surgical debulking along with CPMV priming. Therefore, no future surgical operation will be needed.

Various slow-release formulations have been developed, such as hydrogels for vaccine or chemotherapy delivery,^{14,15} as well as microparticle depots or microneedle patches for immunotherapy/antibody delivery.^{16,17} In a previous study we turned toward a CPMV-dendrimer self-assembly strategy to form i.p. CPMV depots and showed that the CPMV-dendrimer co-assemblies served as slow-release CPMV formulation and achieved improved efficacy against ovarian cancer after a single treatment.¹⁸ While efficacious in the tumor mouse model, translation may be hampered due to toxicity of the dendrimer formulations;^{19,20} another hurdle is that the formulation relied on self-assembly primed by electrostatic interactions leading to nanoscale aggregates with ill-defined size control.

3D bioprinting is an additive manufacturing strategy for the fabrication of bio-constructs. To date, extrusion-, inkjet-, and light-based 3D bioprinting strategies have been developed to print constructs with live cells, biomaterials, or biological molecules, which constitute bioinks. Extrusion-based and inkjet-based bioprinting feature the precise deposition of the bioink with defined placement and pattern.^{21,22} Light-based printing, featuring digital light processing (DLP) printing, uses light to selectively initiate bioink photopolymerization in a vat, enabling layer-by-layer construction with fine control over the geometry of the structure.²³ Compared to extrusion- and inkjet-based bioprinting, DLP printing exhibits significant advantages in printing speed, making it promising for large-scale manufacturing.²⁴ DLP printing is also less dependent than extrusion printing or inkjet printing on specific rheological properties of the bioinks.^{21,22} Additionally, DLP printing offers excellent printing resolution, achieving details up to micron-scale.²⁵ Overall, the DLP bioprinting technology will not only enable us to formulate biodegradable and biosafe 3D depot but also offer us more engineering space to incorporate various antigens and adjuvants in our future cancer immunotherapy development.

In this work we sought to explore the feasibility of using DLP^{26,27} printing technology to fabricate biocompatible polymeric CPMV-laden hydrogels of defined size and geometry (Fig. 1A), to establish this platform for future development of more complex systems. First, CPMV was propagated in black eyed peas and purified as previously described.^{28,29} Then, CPMV was mixed with a defined concentration of gelatin methacrylate (GelMA, 4% w/v) and Polyethylene glycol diacrylate (PEGDA, 0.1% w/v) to prepare a bioink, which was then exposed to 405 nm light for 30 s (light intensity = 79.4 mW cm⁻²) to achieve polymerization and hydrogel formation. Hydrogels of 9.5 mm × 6 mm × 3.2 mm in a slab geometry were bioprinted containing 900 µg CPMV. The polymerization process and blending of CPMV were characterized by Fourier-transform infrared spectroscopy (FTIR). Acrylate

polymerization was effectively initiated for both CPMV laden-hydrogel and sham hydrogel with no CPMV during the DLP printing process, as validated by the pronounced drop in transmittance at 1640 cm⁻¹ (Fig. 1B). Upon the incorporation of CPMV, a group of subtle absorbance signals were noted around ~3000 cm⁻¹. However, the spectrum did not display any characteristic absorbance indicative of covalent or non-covalent bonding. This suggests that CPMV was incorporated into the hydrogel through simple blending, which led to a diffusion-based release mechanism. The microarchitecture was examined by scanning electron microscopic (SEM) imaging of the lyophilized samples. A porous hydrogel architecture stemming from, and characteristic for, the polymeric network of GelMA and PEGDA was observed (Fig. 1C). A consistent pore size distribution was observed in both samples; data suggest that CPMV did not disrupt the polymerization. Mechanical testing further corroborated this, as sham and CPMV-laden hydrogels exhibited comparable Young's moduli (Sham gel: 6531 ± 628.0 Pa, CPMV gel: 6117 ± 2670 Pa, *p* = 0.8065) (Fig. 1D). *In vitro* release of the bioprinted CPMV in PBS was evaluated by ELISA.²⁶ ~30% of CPMV was released by day 1, and ~50% of CPMV was released by day 3. The release then continued slowly to day 14 when it reached a plateau of ~60%, which followed the pattern of Fick's diffusion³⁰ (Fig. 1E).

To evaluate the sustained release of CPMV within the i.p. space, we generated hydrogels incorporating Cy5-labeled CPMV (CPMV-Cy5). Each CPMV particle offers 300 surface-exposed lysine residues suitable for bioconjugation using N-hydroxy-succinimide (NHS) chemistry.³¹ To synthesize CPMV-Cy5, CPMV was reacted with sulfo-Cy5-NHS ester using previously established protocol.^{29,32} As demonstrated by the denaturing gel electrophoresis (NuPAGE), Cy5 signal co-migrated with the CPMV coat proteins, demonstrating the successful conjugation of Cy5 to CPMV (Fig. S1A, ESI†). UV-Vis spectrophotometry analysis determined that ~78 Cy5 molecules were conjugated per CPMV (Fig. S1A, ESI†). Transmission electron microscopy confirmed that CPMV-Cy5 remained intact and structural sound similar to unconjugated CPMV (Fig. S1C, ESI†). Subsequently, we loaded 600 µg of CPMV-Cy5 into two distinct hydrogel formulations: one comprising 5% GelMA + 0.2% PEGDA and the other containing 4% GelMA + 0.1% PEGDA. This allowed us to investigate whether the *in vivo* release of CPMV could be modulated by the formulation; 600 µg soluble CPMV-Cy5 was i.p. injected as a control. To validate that the CPMV-laden hydrogels formed a depot for sustained CPMV release *in vivo*, CPMV-Cy5-loaded hydrogels were surgically implanted into the i.p. space of BALB/c mice, and Cy5 fluorescence was continuously monitored and recorded for a duration of 30 days. The two formulations are designed to provide different kinetics of release – a higher GelMA and PEGDA concentration created a denser polymeric network to sustain the diffusion of CPMV and lead to a slower release.³³ PEGDA increases the stiffness of the implant and therefore improves its handleability. The formulation with 0.1% PEGDA is softer and therefore more compatible to the mechanical environment of the peritoneal although it required more skillful handling for



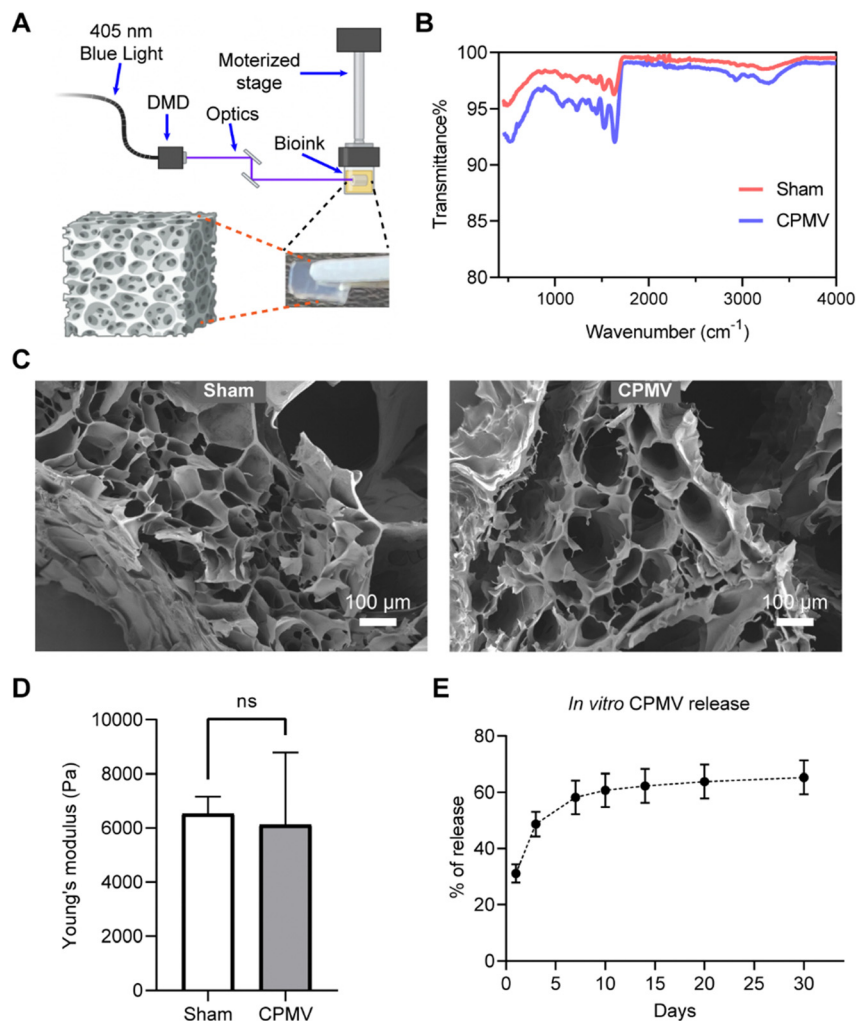


Fig. 1 Bioprinting and characterization of CPMV implants. (A) Schematic diagram of the digital light processing (DLP) bioprinting approach. Characterization of empty gel (Sham) and CPMV loaded implants using (B) FTIR spectroscopy, (C) SEM, and (D) Young's modulus (statistical analysis was calculated by unpaired T test) (D). (E) *In vitro* releasing profile of printed CPMV.

the implantation. Upon surgical administration of the CPMV-laden hydrogels and i.p. injection of CPMV-Cy5, the Cy5 fluorescence signal within the i.p. space was monitored by IVIS

imaging (Fig. 2A). Quantitative analysis showed an initial decrease followed by a gradual increase in fluorescence intensity for both hydrogel formulations – this fluctuation can be

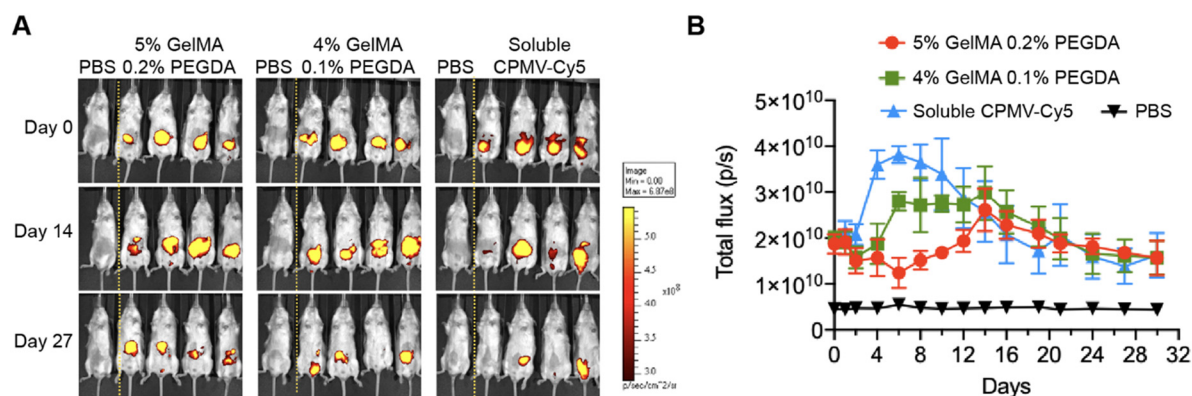


Fig. 2 Release of CPMV in the intraperitoneal space. (A) Fluorescence imaging of BALB/c mice post-surgical implantation of CPMV-Cy5 hydrogels on days 0, 14, and 27. (B) Quantified fluorescent intensity in the intraperitoneal space over time.



explained by initial fluorescence quenching.³⁴ At day 15 post administration, maximum fluorescence intensity was achieved for both hydrogels, then the fluorescence signals declined gradually for 30 days post administration, which indicated the sustained release of CPMV in the i.p. space (Fig. 2B). In comparison, the soluble CPMV-Cy5 showed early increasing of Cy5 fluorescence with peaking around day 7 and faster decay over time. Overall, no major differences in the release profiles were noted comparing the two hydrogel formulations; therefore, we proceeded with 4% GelMA + 0.1% PEGDA for efficacy testing.

Next, we assessed the efficacy of the CPMV slow-release hydrogel using a mouse model of ovarian cancer, ID8-Defb29/Vegf-a-Luc ovarian tumors, disseminated in the peritoneal cavity of C57BL/6J mice.^{35,36} To mimic the remission stage and application of the implant post-surgery, we first surgically implanted hydrogels containing 900 μ g of CPMV into the i.p. space on day -3 prior to tumor challenge. In addition, we administered a single dose of soluble CPMV (100 μ g in 200 μ L of PBS, i.p.). The addition of soluble CPMV is to kick-start the immunostimulation and avoid delayed responses due to slow-release of CPMV from the hydrogel ($n = 9$). The slow-release

hydrogel group was compared to: (1) soluble 1 mg dose of CPMV (i.p. administration, $n = 9$), (2) weekly administration of 100 μ g-doses of CPMV (in 200 μ L of PBS, weekly for 6 weeks, $n = 10$). PBS ($n = 8$) and sham gel ($n = 9$) were included as controls (Fig. 3A). On day 0, mice were challenged with 5×10^6 ID8-Defb29/Vegf-a-Luc cells administered i.p. Tumor progression was monitored closely by tracking changes in body weight (Fig. S2A and B, ESI†) and increases in body circumference (Fig. 3B and C) due to tumor burden and ascites. On day 50, all groups received one additional dose of 100 μ g soluble CPMV *via* i.p. injection to further improve treatment efficacy. The study concluded on day 80 when all remaining mice were free of tumors. Mice were euthanized if ascites and tumor growth caused body weight to exceed 35 g or their abdominal circumference exceeded 9 cm.⁶

The weekly dosing of 100 μ g CPMV is the established 'standard dose' for the ID8-Defb29/Vegf-a-Luc i.p. ovarian cancer model in C57BL/6J mice,^{5,7} consistent with our prior work.⁵⁻⁷ This repeated dosing of CPMV indeed showed potent efficacy controlling tumor growth and leading to survival benefits (Fig. 3B-D). Also, the surgically implanted CPMV hydrogels primed potent efficacy against the ovarian tumors

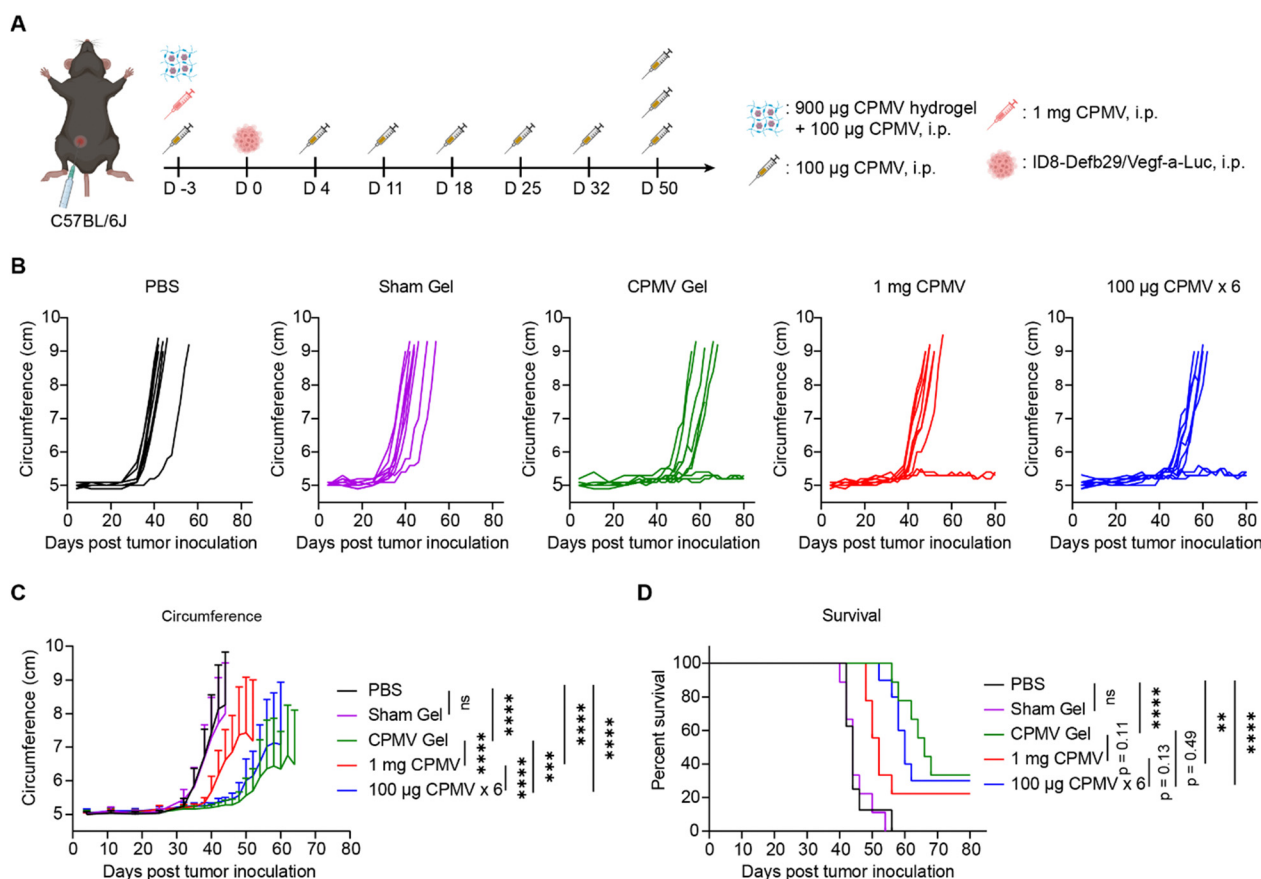


Fig. 3 Treatment efficacy of CPMV hydrogels. (A) Treatment and tumor injection schedule. The groups are PBS ($n = 8$), Sham hydrogel ($n = 9$), CPMV hydrogel (900 μ g + 100 μ g soluble CPMV) ($n = 9$), bolus treatment (1 mg CPMV) ($n = 9$), and weekly CPMV dosing using 100 μ g ($n = 10$). (B) Individual circumferences in each group. (C) Average body circumferences for all treatment groups. Data points are cut off when $n < 5$ for the treatment group. (D) Survival rates for all treatment groups. Statistical significance in (C) was calculated by two-way ANOVA (** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$). Statistical significance in (D) was calculated using the log-rank (Mantel-Cox) test (** $p < 0.01$, **** $p < 0.0001$).



and appeared slightly more potent in terms of controlling tumor growth and survival rate (Fig. 3B–D). Lastly, the 1 mg bolus-dose of CPMV also delayed onset of tumor growth, however with lesser efficacy compared to the sustained-release or repeated dosing groups. This underscores the potent anti-tumor efficacy of CPMV. Conversely, the sham hydrogel group or PBS control groups had no discernible effects on tumor growth. While statistical significance was not reached, sustained release CPMV hydrogel treatment trended toward being the most potent formulation as demonstrated by improved controlling of tumor growth (Fig. 3B and C) and enhanced overall survival rates (33.3%, 3/9 mice) in comparison to repetitive dosing (30%, 3/10) and bolus dosing (22.2%, 2/9) (Fig. 3D). Further analysis revealed that CPMV gel implantation extended median survival to 66 days, compared to 60 days with repetitive CPMV dosing and 52 days with a single large CPMV dose. In contrast, both PBS and Sham gel controls had a median survival of 44 days. It was noted that the single bolus CPMV dose of 1 mg, resulted in five mice displaying potential autoimmune symptoms, such as fur loss and white hair growth. From a clinical perspective, CPMV hydrogel implants not only prevent overdosing and excessive immune system overstimulation but also ensure efficacy with a broader remission period for patients to receive additional treatments. This approach can also reduce costs associated with repetitive dosing and hospitalization, ultimately leading to an improved quality of life.

In conclusion, we successfully developed a sustained-release hydrogel formulation of CPMV using state-of-the-art DLP printing technology. Sustained release of the active ingredient, CPMV, was confirmed through *in vitro* and longitudinal imaging studies in live animals. More importantly, through surgical implantation, the CPMV-laden hydrogels proved potent and efficacious against murine ovarian cancer – on par with our standard weekly dosing regimen. From a clinical perspective, CPMV hydrogel implantation offers several advantages over the traditional dosing methods by avoiding repetitive intraperitoneal injection with associated patient stress, potential for problems and increased costs with hospitalization. Less need for repetitive clinical intervention will enhance quality of life for patients, and the slow release potentially would reduce risk of immune-related adverse events. Throughout this study, the hydrogels served as scaffolds for CPMV delivery. In our forthcoming research, we aim to enhance treatment efficacy for ovarian cancer by incorporating additional immunoadjuvants or tumor-specific antigens. Overall, this work lays the groundwork for the future translation of CPMV as a treatment option for human patients in clinical settings.

Author contributions

Z. Z., Y. X., S. F., S. C., and N. F. S. designed and analyzed experiments. Z. Z., Y. X., E. C. K., and S. S. performed experiments. Z. Z., Y. X., S. C., and N. F. S. wrote the manuscript. Z. Z. and Y. X., contributed equally.

Conflicts of interest

The authors declare the following competing financial interest(s): Dr Steinmetz and Dr Fiering are co-founders and have equity and financial interest in Mosaic ImmunoEngineering Inc. Dr Steinmetz is a co-founder of, and serves as manager of Pokometz Scientific LLC, under which she is a paid consultant to Mosaic ImmunoEngineering Inc., Flagship Labs 95 Inc., and Arana Biosciences Inc. The other authors declare no potential COI.

Acknowledgements

This work was supported in part by the NIH (R01-CA253615) to N. F. S., S. C. and S. F., the Shaughnessy Family Fund for Nano-ImmunoEngineering (nanoIE) at the University of California, San Diego (UCSD), and training grant T32EB005970 to E. C. K. We thank the Nano3 core facility at UCSD for transmission electron microscopy imaging. Nano3 hosts San Diego Nanotechnology Infrastructure (SDNI), a member of the National Nanotechnology Coordinated Infrastructure (NNCI), which is supported by the National Science Foundation (Grant ECCS-2025752).

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