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A simple approach to determining the efficacy of antiperspirants using paper-based colorimetric paper sensors: SweatSENSE†

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Antiperspirants are commonly used daily, or even multiple times a day, by a large percentage of the global population, in order to address the problems associated with sweating. However, there has been no simple way to test and evaluate the effectiveness of antiperspirants in real time. To address this, we have developed a polydiacetylene based chemosensor that undergoes a blue to red colour change when in contact with acids and alcohols found in sweat, but not when exposed to water. The sensor is prepared via inkjet-printing an imidazolium derivative of a diacetylene monomer onto normal paper to give a transparent square followed by UV curing to produce the navy blue 5 × 5 cm “SweatSENSE” patch. A sensor is applied to the underarm for 5 s and the colour changes to red in the presence of sweat and the intensity of the colour change is recorded. From an *in vivo* trial involving 52 panellists, the sensor was shown to demonstrate a reduction in total sweat area following the application of an antiperspirant, when compared with a deodorant only control. This sensor will allow consumers to test the effectiveness of their antiperspirants during daily life, in real time, without the need for specialised equipment.

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Introduction

In humans, the axilla is a unique body site, containing a high density of hair follicles and sebaceous glands, as well as two types of sweat glands: eccrine and apocrine.^{1,2} These sweat glands differ in their physiology, function and distribution.^{3,4} Eccrine sweat glands are simple, coiled structures that are present throughout the body, although their density varies with body size. They form part of the body's thermoregulatory system, secreting a sterile, dilute electrolyte solution which is predominantly responsible for wetness (perspiration/sweat). Apocrine sweat glands are unique to the axillae and genital areas and secrete an oily fluid containing proteins, lipids, and steroids. Apocrine sweat is typically produced in response to emotional and/or physical stress. At the skin surface of the axilla, microorganisms thrive on these natural secretions, transforming them into volatile organic compounds.⁵

A consensus has emerged that short and medium chain volatile fatty acids (VFAs), along with thioalcohols, are the primary molecules associated with axillary malodour,⁵ with a minor contribution from 16-androstene steroids.⁶ These are by-products of the microbial metabolism of the odourless natural secretions from the eccrine, apocrine and sebaceous glands. In many societies, the detection of underarm wetness and odour brings about feelings of anxiety that affect all aspects of a person's life, from career choices and recreational activities to relationships, emotional well-being, and self-confidence.⁷ People are conscious of wet patches and staining of clothing in the underarm as well as the potentially negative perception of their body odour.

Antiperspirants and deodorants are a large global business estimated to be worth >\$75 billion U.S Dollars in 2019. Many people assume that antiperspirants and deodorants are the same, however, they have different primary functions. The primary aim of a deodorant is to control the concentration of volatile malodorous compounds that are generated as the skin bacteria work on the sweat in the underarm. They contain fragrances and may contain antimicrobial ingredients. Conversely, the first line of defence against underarm wetness is antiperspirants. Antiperspirants are non-invasive, topically applied products worn to reduce the volume of sweat in the

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axilla. An individual product can have one or both of these functionalities.

Numerous methods exist to test the efficacy of an antiperspirant.⁸ The most established test is a gravimetric method, whereby the detection and quantification of collected sweat is used to compare the sweat weight reduction (SWR) of an antiperspirant technology *versus* a control product. Panellists are required to have products applied to the axillae and then sit at elevated temperature in a hot room (for example, at 40 °C and 40% relative humidity) to acclimatise, prior to sweat being collected for a further period on gauze pads applied to the underarms. The pads are weighed before and after application to the underarms which allows the amount of sweat collected to be calculated. This technique provides a means to quantify sweating behaviour under thermal stress conditions but is limited, in that quantification of sweating and measure of sweat control efficacy provided by an antiperspirant is not possible during normal daily activities.

Sensor technology has been driven by advances in materials chemistry combined with the emergence of digital communication technologies and wireless sensor networks. Devices, often quite complex, have been developed for a diverse range of applications, such as forensics, human health, sporting performance, environment and national security.^{9–14} These advances have led to rapid progress in wearable sensor technologies which offer an opportunity to measure sweat flow rate “*in-life*”. A number of recent developments employ electrochemical sensors to record sweat volume or the concentration of specific analytes in sweat.^{15–17} These technologies are more convenient for users than the classic gravimetric methods, however, there can be issues with the response and calibration of such sensors and their use at scale can be cost-prohibitive. Colorimetric sensors use smart materials to induce a response to a specific analyte and can be a simpler, more cost-effective alternative to an electrochemical device.^{18–20}

Polydiacetylenes (PDAs) are a family of conjugated polymers arising from the 1,4 topochemical polymerization of diacetylene monomers.²¹ The polymerization typically occurs under UV irradiation in the absence of solvents and catalysts, yielding blue π -conjugated polymer networks due to visible light absorption. PDAs exhibit a well-known colorimetric response towards various analytes which is a consequence of polarity changes that disrupt their conjugation. It has been proposed that such disruption causes conformational transitions in the PDA backbone (from all *trans* to *gauche*) that eventually lead to colorimetric changes from blue to red due to an increase in the energy required for electronic transitions in the visible region.^{22,23}

It has been reported that PDAs have been used in flexible patch-type hydrochromic sensors for human sweat pore mapping on the hands and fingertips.^{24,25} PDAs have been formulated into an inkjet-printable micro-emulsion system and applied to paper substrates.²⁶ This simple, low cost approach provides a chemosensor that detects minute amounts of sweat excreted from the sweat glands through a permanent and quantifiable colour change. However, the main issue with this

technique for quantifying underarm sweating is that the axilla is a unique and complex body site, hard to access topographically, and with typically high humidity levels (60–99%). In this case, a sensitive hydrochromic sensor is saturated by the water vapour in the underarm area, causing a total colour change of the patch and inhibiting the quantification of sweat.

The work presented herein aims to develop a simple yet effective low cost, chemosensing system that can be used to both demonstrate and quantify antiperspirant efficacy. A “*sweatochromic*” sensor has been developed that responds to the weak organic acids present in sweat but does not respond to water. The sensor reaction time is tuned to allow complete detection within a 5 seconds skin contact time. This is beneficial as it allows for careful placement and removal of the sensors in the difficult to access axillary vault and enables detection of sweat generated at a single point in time during normal daily life.

Experimental

Materials

10,12-Pentacosadiynoic acid ($\geq 97\%$), *N*-hydroxysuccinimide (98%), 1-propanol ($\geq 99.5\%$, ACS reagent), dichloromethane (anhydrous), propylene glycol methyl ether (PGME), and 3-bromopropionitrile (99%) were purchased from Sigma-Aldrich. Sodium hydroxide (analytical reagent grade), sodium carbonate anhydrous (laboratory reagent grade), potassium hydroxide (analytical reagent grade), potassium carbonate anhydrous (laboratory reagent grade), magnesium sulfate dried (laboratory reagent grade), 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (EDC, 98+%) were supplied by Fisher Scientific. Propionic acid (for synthesis) and triethylamine (99%) were sourced from VWR International. All materials were used as received.

PCDA-imidazole (PCDA-I[†]H)

10,12-Pentacosadiynoic acid (PCDA) (5 g, 13.4 mmol) was suspended in anhydrous CH_2Cl_2 (100 mL) under a N_2 atmosphere and cooled to 0 °C in an ice bath with constant stirring, to which was added *N*-(3-dimethylaminopropyl)-*N'*-ethylcarbodiimide hydrochloride (3.838 g, 20.0 mmol, 1.5 equiv.) and *N*-hydroxysuccinimide (0.153 g, 1.33 mmol, 0.1 equiv.). Following this, 1-(3-aminopropyl)imidazole (2.39 mL, 20.0 mmol, 1.5 equiv.) and *N,N* diisopropylethylamine (2.56 mL, 14.7 mmol, 1.1 equiv.) were added to the reaction mixture. The solution was removed from the ice bath and stirred at 25 °C for 18 h. The CH_2Cl_2 was removed *in vacuo* and the oily residue resuspended in ethyl acetate (75 mL) and washed with water (3×75 mL). The organic layer was dried over anhydrous magnesium sulfate, filtered, and the solvent removed *in vacuo* to give PCDA-I as an oily white solid. ¹H NMR (CDCl_3 , 400 MHz) δ (ppm): 7.50 (s, 1H), 7.05 (s, 1H), 6.95 (s, 1H), 5.55 (s, 1H), 4.0 (t, 2H), 3.30 (s, 2H), 2.25 (t, 3H), 2.15 (t, 2H), 2.00 (t, 2H), 1.44 (m, 34H), 1.24 (t, 3H).



The PCDA-I product (5 g, 10.4 mmol) was then suspended in acetonitrile (100 mL), to which was added 3-bromopropionitrile (1.1 equiv.) and for 18 h under reflux at 90 °C. The reaction mixture was then allowed to cool before concentration *in vacuo* to yield a pale yellow solid of PCDA-I⁺H. ¹H NMR (CDCl₃, 400 MHz, Fig. S3†) δ (ppm): 14.24 (s, 1H), 9.10 (s, 1H), 7.96 (s, 1H), 7.83 (s, 1H), 7.72 (s, 1H), 4.20 (t, 2H), 3.38 (s, H₂O), 3.04 (t, 2H), 2.51 (s, DMSO), 2.27 (t, 2H), 2.08 (t, 2H), 1.94 (t, 2H), 1.44 (m, 6H), 1.24 (m, 26H). For storage, due to the light sensitivity of PCDA-I⁺H, the solid product was then resuspended at a concentration of 1 M in 1-propanol and stored in the dark.

Ink formulation and ink-jet printing

The ink formulation comprised of PCDA-I⁺H at 350 mM in 1:1 H₂O/1-propanol, with 10% propylene glycol methyl ether (PGME). It was added to the “black” ink tank of an Epson EcoTank inkjet printer, which was then covered with aluminium foil to shield from exposure to light. Images of black rectangles (5 × 5 cm) were then printed on 80 gsm A4 paper, using standard print settings, the sheets were then allowed to dry at room temperature (*ca.* 5 minutes) before the patches were cured for 12 s at 500 000 $\mu\text{J cm}^{-2}$ in a box UV curer designed for sterilisation ($\lambda = 254 \text{ nm}$).

Sweat study

The performance of the sensors was assessed with a “hot room study” involving 52 subjects, where half were given a topologically applied antiperspirant and the other half a control deodorant product. Subjects applied the products for three consecutive days and on the fourth day they sat in a Hot Room under controlled conditions of temperature and humidity (40 °C/40% relative humidity) for 40 minutes. After this time, the SweatSense™ patches were applied to the subjects' underarms for 5 seconds using a purpose-built applicator. This applicator allowed for a uniform and consistent force to be applied to each sensor when used in the underarm. Sensors were applied to the centre of the axillary vault. The area of coverage of sweat on the sensor is calculated to determine sweating behaviour at a point in time, *i.e.*, one hour after being in the hot room. This study was performed under approval of the local Unilever Safety and Environmental Assurance Centre (SEAC) following their guidelines.

Results and discussion

A common strategy to introduce sensing properties to PDA systems is by covalently attaching polar moieties to diacetylene acid monomers thus enabling polarity changes and colour transitions after exposure to certain analytes.²⁷ A study by Kim *et al.* demonstrated PCDA bromoacetonitrile imidazolium salts able to sense water by undergoing such transitions.²⁵ However, it was found that smaller chain-length PCDAs showed no response towards water indicating that minor structural changes can affect polarity alternations. Since the goal of this study was to develop PDA sensors with minimal response to

water molecules, a comparable approach was envisioned wherein the modification focused on the polar moiety rather than altering the alkyl chain of PCDA.

More specifically, a similar imidazolium derivative of PCDA was desired but this time reacting with bromopropionitrile in the final step. The PCDA monomer salt was synthesized in a simple two-step procedure, Fig. 1A. In the first step, a one-pot EDC/NHS coupling was used to generate the neutral imidazolium derivative. The product was then heated under reflux for 18 hours with 1.1 equivalents of bromopropionitrile and was isolated as a pale yellow oily solid.

This was quickly redispersed in 1-propanol and protected from light to ensure no polymerisation occurred prior to printing. Surprisingly, the attained final product was not the expected PCDA bromopropionitrile imidazolium salt but instead the protonated PCDA imidazolium salt derivative (PCDA-I⁺H). This was confirmed by both ¹H-NMR and high-resolution mass spectrometry (Fig. S2 and S3†) showing that molecule 1 is the major product of this reaction at the used experimental conditions with the characteristic protonated nitrogen peak (–NH) appearing at $\delta = 9.20 \text{ ppm}$ and an $m/z = 496.42 \text{ g mol}^{-1}$. The basic character of the lone pair electrons on nitrogen at position 3 of the imidazole ring likely showed preference to abstract an acidic proton (–CH₂–Br) from bromopropionitrile resulting in protonation.

Ink-jet printing is a convenient method for depositing PCDA-I⁺H onto paper, in a scalable and reproducible manner. However, careful consideration must be given to the ink formulation, which needs to contain appropriate solvents to avoid both particulates forming in the ink and fast evaporation, both of which lead to deposition of solids at the nozzle leading to the printer blocking. This results in streaks forming in the product, Fig. S4.† Ink was produced by dissolving 1 in a 50:50 mixture of water and 1-propanol to form a clear, slightly yellow solution, with added PGME to lower the volatility, Fig. 1B. The ink was transferred into an Epson EcoTank inkjet printer and printed at standard quality on standard 80 gsm A4 paper. The ink was then allowed to fully dry on the paper surface before being irradiated with $\lambda = 254 \text{ nm}$ light at 5000 $\mu\text{J cm}^{-2}$ for 12 seconds to produce a uniform blue square. Optimizing the time and energy of curing was important to give the highest quality sensors. Three different curing times with two predetermined energy outputs were examined and their performance was assessed *in vivo* after hot room studies (Fig. S5–S8 and Table S1†). Results indicated that longer irradiation times reduced the sensitivity and quality of the sensor while a 12 seconds curing using 5000 $\mu\text{J cm}^{-2}$ was optimal, giving uniform patches with good colour contrast, high droplet clarity and minimal droplet spreading.

The response of the printed patches to various stimuli was first tested in the laboratory. For the intended application, it is important that the sensor only responds to the acids and alcohols in sweat, and not to water. Fig. 2 demonstrates the colorimetric response to a range of acids, alcohols and tap water. The Raman spectra of a cured PDA patch (blue phase) and the same region after exposure to a drop of propionic acid (red



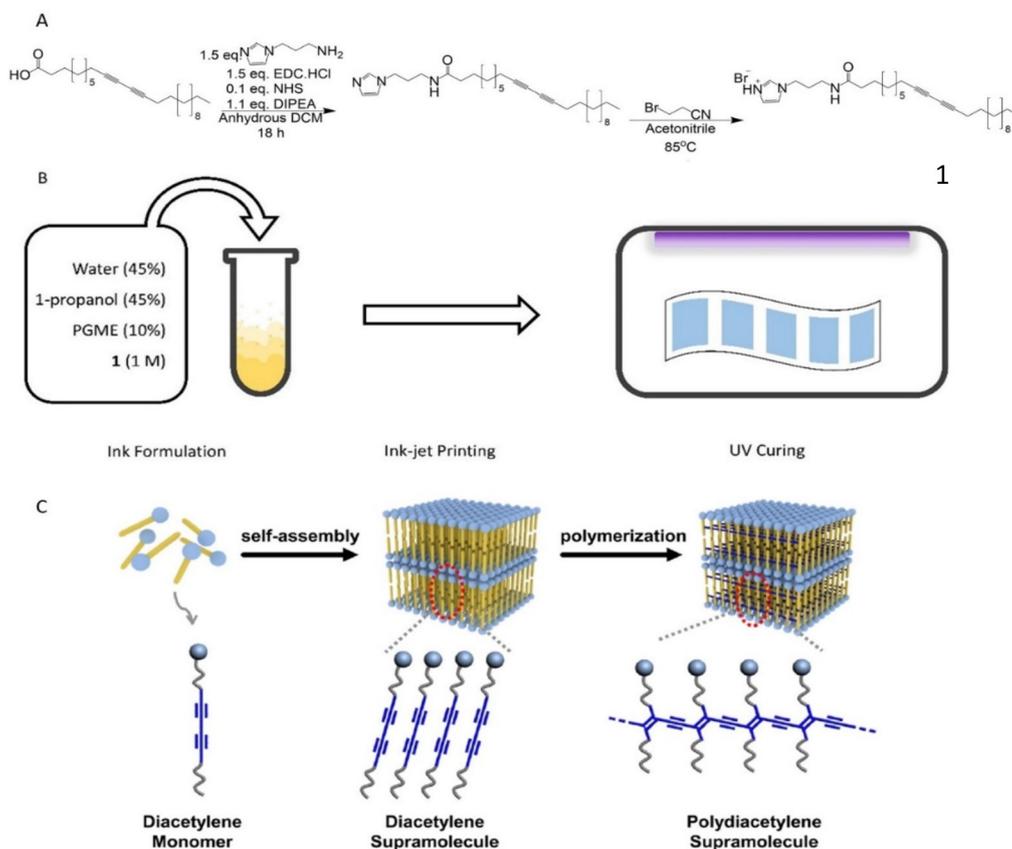


Fig. 1 A: Reaction scheme for the synthesis of PCDA-I⁺H B: schematic illustration of sensor preparation C: cartoon showing ordering and polymerisation adapted from Lee *et al.* under licence CC BY-NC-ND 3.0.²⁴

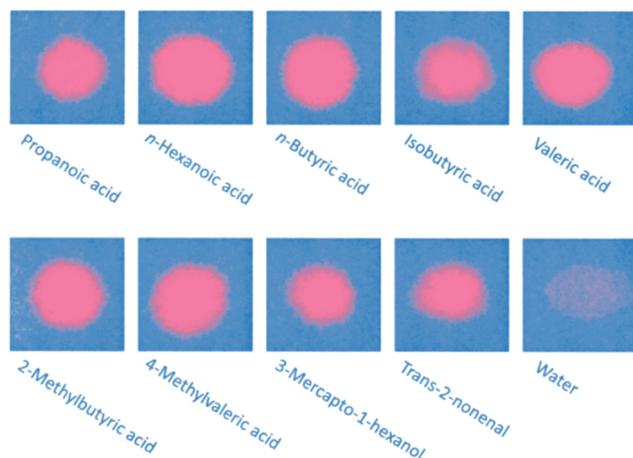


Fig. 2 Chromic response of poly(PCDA) printed on paper to a range of odorous sweat components and a tap water control.

phase) shows indicative changes, with the disappearance of a sharp peak at 1513 cm^{-1} moving to 1454 cm^{-1} and 2118 cm^{-1} moving to 2081 cm^{-1} , Fig. 3A. It is apparent that the addition of the acid increased the frequency of both the carbon-to-carbon double and triple bond vibrations, indicating that their bond lengths decrease. Upon the addition of water,

no visual change was detected (Fig. S9[†]). This potentially indicated that water molecules did not cause enough polar changes to disrupt the conjugated PDA network thus leading to minor conformational transitions of the backbone.

Heat can also be used as a stimulus to induce a colour change in these PDAs.²⁸ For underarm sweat analysis, it is necessary that the PDA patches are stable at body temperature. To test this, we taped a sheet of polyethylene film (PE) to paper and printed and cured a PDA patch on to the surface. The blue region on the PE was then clamped in a quartz demountable cuvette and its UV-vis spectra were measured over a temperature range from 25–95 °C (Fig. 3B). With increasing temperature, the absorbance band of the PDA undergoes a significant blue shift, from 664 nm to 553 nm. The increase in the energy of the π^* transitions shows the disruption of the large conjugated system of the blue phase PDA and the formation of the red phase, which begins at around 45 °C. The dashed line in Fig. 3B shows that at body temperature the PDA patch is still in the blue phase.

Following the benchtop tests, we proceeded to test the effectiveness of the PDA sensors for detection of sweat from human underarms (Fig. 4A). For this we setup an experiment to test the differences between a deodorant and an antiperspirant product. Antiperspirants typically contain active ingredients that block pores and reduce sweat volume and so we should



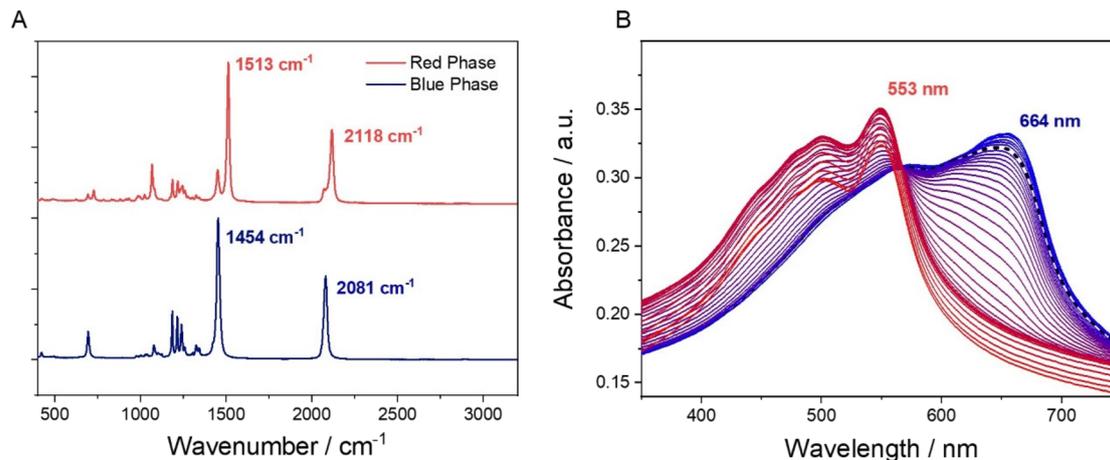


Fig. 3 A: Raman spectra of blue phase PDA (bottom) and red phase PDA (top) after treatment with propionic acid. B: UV spectra of PDA printed on PE film with temperature increasing at 2 °C increments from 25–95 °C. The spectrum at 37 °C is given as a dotted black line.

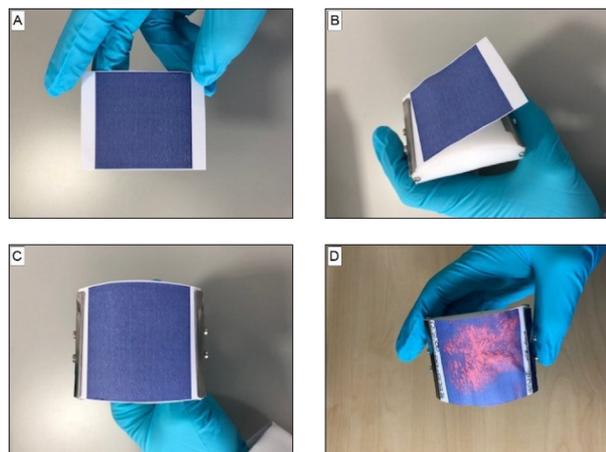


Fig. 4 Application procedure for the PDA patches used in the sweat study. The printed patch (A) is first clipped (B) onto the curved applicator (C) then applied to the underarm. Once removed, the colour change to red (from blue) indicates the presence of sweat (pores) (D).

see a clear difference following the application of these two products. To ensure a controlled environment in the experiment, a hot room study was carried out with 52 panellists. Half were given an antiperspirant to apply for three consecutive days and the other half a deodorant control product. On the fourth day, the panellists sat in a hot room, which was set at 40 °C and 40% relative humidity. After 40 minutes in the hot room, a PDA patch was applied to the underarm of each of the panellists for 5 seconds. For comparison, examples of patches from “untreated” sweat from a hot room study prior to product application are presented in Fig. S10.†

To ensure that an even pressure was applied to the underarms, the PDA patches were secured on an applicator, where the radius of its curvature was designed to maximise surface contact in the underarm (Fig. 4B and C). Following application to the underarm, the area of the patch that was in contact with sweat changed to be red in colour (Fig. 4D).

Images of the patches used in the hot room study were digitized using the Epson EcoTank scanning function. These images were read into MATLAB and processed into binary images with an algorithm based on thresholding of their RGB values. Example images of patches processed from the deodorant control group and the antiperspirant group are given in Fig. 5A, B and C, D respectively. The red area of the patches was determined by calculating the ratio of the number of pixels with a value of 1 to the total number of pixels in the image and finally converting these values to mm².

The processed data from all panellists in the hot room study is given in Table 1. It is apparent that when using the

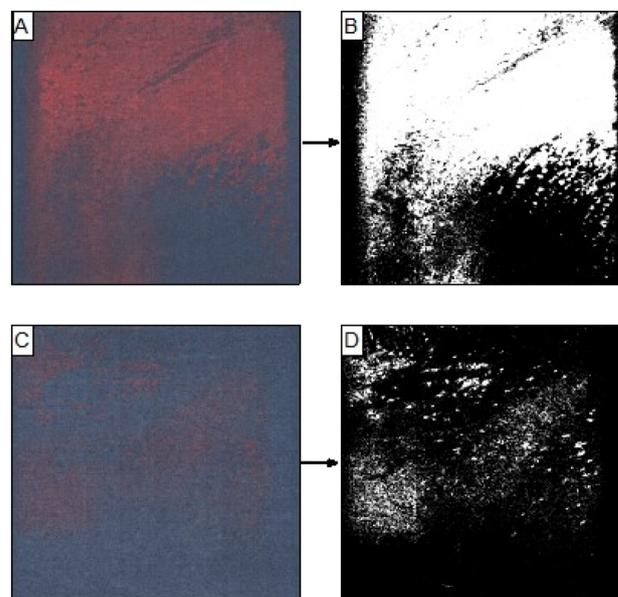


Fig. 5 Scanned images of patches after application on subjects with deodorant (A) and antiperspirant (C) and their corresponding processed images (B and D, respectively).

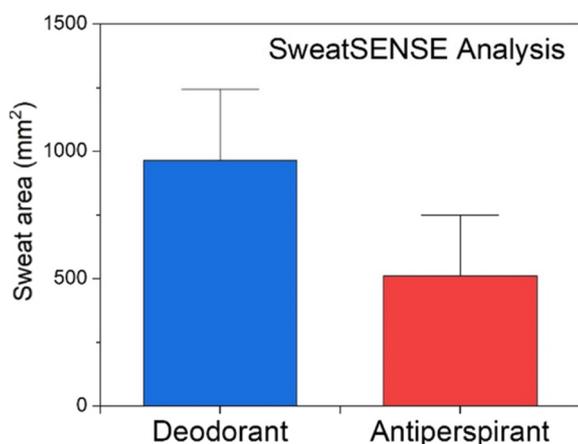
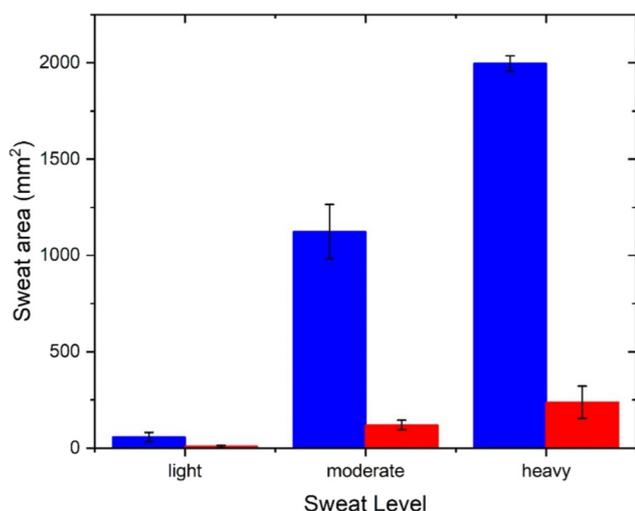


Table 1 Sweat area data from a hot room study where groups of panellists applied either a deodorant or an antiperspirant product

Product	Mean sweat area (mm ²)	Std. dev. (σ)	Std. error mean (SEM)	Wilcoxon signed-rank test (p -value)	Sweat area reduction (SAR) ^a	Number of panellists
Deodorant	963.37	693.26	135.96	0.0002175	46.93%	26
Antiperspirant	511.24	587.37	115.19			26

$$^a \text{SAR} = (1 - \text{mean (Antiperspirant)/mean (Deodorant)}) \times 100.$$

patches, a clear difference in the average area of colour change was detected between the groups where deodorant was applied compared with the antiperspirant group, with the latter group showing an area reduction of 46.93%. The difference between the sets of samples was also tested using the Wilcoxon signed-rank test, which showed a significant difference between the two populations ($p \ll 0.05$), Fig. 6.

**Fig. 6** Comparison of the red area of patches following application on subjects in the antiperspirant and deodorant groups.**Fig. 7** Comparison of the red area of patches following application on subjects in the antiperspirant (red) and deodorant (blue) groups for the data divided into three subsets.

The capability of the PDA sensor was then further examined by analysing the data collected from three subsets of subjects: light, moderate and heavy “sweaters”, again comparing the area of colour change when using an antiperspirant product or a control, Table S2.† Thus, the patches can be used to differentiate between the subsets when wearing or not wearing an antiperspirant product, Fig. 7.

Conclusions

A simple PDA sensor was developed that selectively changes colour from blue to red in response to the acids and alcohols found in sweat, but not to water. Patches were prepared using an inexpensive home/office ink jet printer with refillable ink tanks. The ink, printing method and UV curing time of the sensors were optimised to determine changes in a person's sweat volume in real time and allow for a high data collection rate. This technology is currently being exploited by Unilever.

Author contributions

RH and AW contributed to the investigation, supervision and writing. SE contributed to the investigation and writing. MG, GK contributed to the investigation and methodology. TB contributed to the investigation and methodology and supervision. ECT contributed to the investigation and validation. JF contributed to the conceptualization and project administration. JR and JM contributed to the investigation. AJ contributed to the conceptualization. SB, EK and DH were responsible for conceptualization, supervision and funding acquisition.

Conflicts of interest

There are no conflicts of interest to declare.

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References

- 1 A. Watkinson, R. S. Lee, A. E. Moore, S. E. Paterson, P. Pudney and A. V. Rawlings, *Int. J. Cosmet. Sci.*, 2007, **29**(1), 60–60.
- 2 K. Wilke, K. Wick, F. J. Keil, K. P. Wittern, R. Wepf and S. S. Biel, *Exp. Dermatol.*, 2008, **17**(1), 73–81.
- 3 J. Tobin, *Chem. Soc. Rev.*, 2006, **35**(1), 52–67.
- 4 K. Wilke, A. Martin, L. Terstegen and S. S. Biel, *Int. J. Cosmet. Sci.*, 2007, **29**(3), 169–179.
- 5 A. G. James, C. J. Austin, D. S. Cox, D. Taylor and R. Calvert, *FEMS Microbiol. Ecol.*, 2013, **83**(3), 527–540.
- 6 C. Austin and J. Ellis, *J. Steroid Biochem. Mol. Biol.*, 2003, **87**(1), 105–110.
- 7 N. Solish, A. Benohanian and W. K. Jonathan, *Dermatol. Surg.*, 2005, **31**(4), 405–413.
- 8 H. Murota, Old and new approaches for assessing sweating, in *Perspiration Research*, Karger Publishers, 2016, vol. 51, pp. 22–29.
- 9 Y.-R. Kim, S. Jung, H. Ryu, Y.-E. Yoo, S. M. Kim and T.-J. Jeon, *Sensors*, 2012, **12**(7), 9530–9550.
- 10 D. Grieshaber, R. MacKenzie, J. Vörös and E. Reimhult, *Sensors*, 2008, **8**(3), 1400–1458.
- 11 H. Tao, L. R. Chieffo, M. A. Brenckle, S. M. Siebert, M. Liu, A. C. Strikwerda, K. Fan, D. L. Kaplan, X. Zhang, R. D. Averitt and F. G. Omenetto, *Adv. Mater.*, 2011, **23**(28), 3197–3201.
- 12 S. Kim, C. Young, B. Vidakovic, S. G. A. Gabram-Mendola, C. W. Bayer and B. Mizaikoff, *IEEE Sens. J.*, 2010, **10**(1), 145–158.
- 13 A. J. Bandodkar and J. Wang, *Trends Biotechnol.*, 2014, **32**(7), 363–371.
- 14 D. P. Lobo, A. M. Wemyss, D. J. Smith, A. Straube, K. B. Betteridge, A. H. J. Salmon, R. R. Foster, H. E. Elhegni, S. C. Satchell, H. A. Little, R. Pacheco-Gomez, M. J. Simmons, M. R. Hicks, D. O. Bates, A. Rodger, T. R. Dafforn and K. P. Arkill, *Nano Res.*, 2015, **8**(10), 3307–3315.
- 15 Y. Yang, S. Xing, Z. Fang, R. Li, H. Koo and T. Pan, *Lab Chip*, 2017, **17**(5), 926–935.
- 16 P. Salvo, F. D. Francesco, D. Costanzo, C. Ferrari, M. G. Trivella and D. D. Rossi, *IEEE Sens. J.*, 2010, **10**(10), 1557–1558.
- 17 W. Gao, S. Emaminejad, H. Y. Y. Nyein, S. Challa, K. Chen, A. Peck, H. M. Fahad, H. Ota, H. Shiraki, D. Kiriya, D.-H. Lien, G. A. Brooks, R. W. Davis and A. Javey, *Nature*, 2016, **529**, 509.
- 18 A. Koh, D. Kang, Y. Xue, S. Lee, R. M. Pielak, J. Kim, T. Hwang, S. Min, A. Banks, P. Bastien, M. C. Manco, L. Wang, K. R. Ammann, K.-I. Jang, P. Won, S. Han, R. Ghaffari, U. Paik, M. J. Slepian, G. Balooch, Y. Huang and J. A. Rogers, *Sci. Transl. Med.*, 2016, **8**(366), 366ra165.
- 19 X. Huang, Y. Liu, K. Chen, W.-J. Shin, C.-J. Lu, G.-W. Kong, D. Patnaik, S.-H. Lee, J. F. Cortes and J. A. Rogers, *Small*, 2014, **10**(15), 3083–3090.
- 20 A. Soni, R. K. Surana and S. K. Jha, *Sens. Actuators, B*, 2018, **269**, 346–353.
- 21 R. Jelinek and M. Ritenberg, *Polydiacetylenes – recent molecular advances and applications*, 2013.
- 22 J. Kiji, J. Kaiser, G. Wegner and R. C. Schulz, *Polymer*, 1973, **14**(9), 433–439.
- 23 K. Fahsi, J. Deschamps, K. Chougrani, L. Viau, B. Boury, A. Vioux, A. van der Lee and S. G. Dutremez, *CrystEngComm*, 2013, **15**(21), 4261–4279.
- 24 J. Lee, M. Pyo, S.-H. Lee, J. Kim, M. Ra, W.-Y. Kim, B. J. Park, C. W. Lee and J.-M. Kim, *Nat. Commun.*, 2014, **5**, 3736.
- 25 M. Seo, D.-H. Park, B. J. Park and J.-M. Kim, *J. Appl. Polym. Sci.*, 2017, **134**(6), 44419.
- 26 D.-H. Park, W. Jeong, M. Seo, B. J. Park and J.-M. Kim, *Adv. Funct. Mater.*, 2016, **26**(4), 498–506.
- 27 S. Lee, J. Y. Kim, X. Chen and J. Yoon, *Chem. Commun.*, 2016, **52**, 9178–9196.
- 28 B. Yoon, H. Shin, E.-M. Kang, D. W. Cho, K. Shin, H. Chung, C. W. Lee and J.-M. Kim, *ACS Appl. Mater. Interfaces*, 2013, **5**(11), 4527–4535.

