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## Stimuli-responsive synthetic helical polymers

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Synthetic dynamic helical polymers (supramolecular and covalent) and foldamers share the helix as a structural motif. Although the materials are different, these systems also share many structural properties, such as helix induction or conformational communication mechanisms. The introduction of stimuli responsive building blocks or monomer repeating units in these materials triggers conformational or structural changes, due to the presence/absence of the external stimulus, which are transmitted to the helix resulting in different effects, such as asymmetry amplification, helix inversion or even changes in the helical scaffold (elongation, J/H helical aggregates). In this review, we show through selected examples how different stimuli (e.g., temperature, solvents, cations, anions, redox, chiral additives, pH or light) can alter the helical structures of dynamic helical polymers (covalent and supramolecular) and foldamers acting on the conformational composition or molecular structure of their components, which is also transmitted to the macromolecular helical structure.

## 1. Introduction

Non-natural helical polymers including foldamers,<sup>1–17</sup> supramolecular<sup>5,18–38</sup> and covalent helical polymers<sup>5,39–60</sup> have been extensively studied during the last decades due to the possibility of creating a large variability of helical scaffolds with different functionalities by rational design. These materials emerged as consequence of the structure/function relationship found in biomacromolecules such as DNA, peptides or polysaccharides, whose helical scaffolds are directly related to their important biological functions such as signalling, storage, duplication, amplification, and processing of information.<sup>61–63</sup> However, while the building blocks used by nature are limited, chemistry offers a wide range of chemical structures with different functionalities limited mainly by the creativity of the scientist.<sup>64–70</sup> In this line, synthetic helical polymers with stimuli-responsive properties have been extensively studied due to the possibility of using an external stimulus (input) to produce a structural change, such as helix inversion (output), in the polymer. In this review we will focus our attention on those synthetic helical structures (covalent polymers, supramolecular polymers and foldamers) whose helical scaffold (sense and elongation) can be altered if an external stimulus triggers a rational mechanism of information transmission through changes in molecular conformations.<sup>71</sup>

To create a synthetic stimuli-responsive helix by design, the starting material must meet certain requirements, some of which are discussed below.

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### 1.1 Foldamers

In helical foldamers<sup>1–17</sup> (Fig. 1a), the building blocks must preferably adopt a twisted conformation that propagates towards the formation of a helical structure when an oligomer is formed.<sup>70–74</sup> A problem found in the preparation of stimuli-responsive foldamers is that if the conformation adopted by the building block is too stable, the foldamer will behave as quasi-static, making it difficult to tune its helical sense. This effect is the most abundant when a foldamer is formed by chiral residues, such as biotic foldamers by  $\beta$ - and/or  $\gamma$ -amino acids. In these systems, a positive or negative torsion of the chiral residue is favoured due to its intrinsic chirality, being difficult to reorient it in the opposite direction. To avoid this problem, the foldamer must be constituted mainly by achiral residues,<sup>75–79</sup> such as aminoisobutyric acid (Aib) (Fig. 1b), employed in the preparation of stimuli-responsive biotic foldamers where the two *P* (plus) and *M* (minus) twisted conformations are equally favoured. To induce a helical sense in the foldamer, two approaches are usually followed, such as supramolecular or covalent interactions with a chiral fragment at one end of the foldamer helix described by the groups of Clayden and Inai (Fig. 1c–e).<sup>80–85</sup> In these foldamers, the conformational preference adopted by the chiral residue is transmitted to the adjacent achiral residue by adoption of a preferred *P* or *M* twisted conformer and propagated to the other achiral residues through a screw sense induction mechanism. Variations in the conformational composition of the chiral residue, or the replacement of the chiral residue by its enantiomer leads to the induction of the opposite helical sense, generating a dynamic helical foldamers.

In other cases, the chiral residue can be introduced in the middle of the foldamer sequence<sup>86</sup> or encapsulated by the



helical scaffold,<sup>13,87</sup> although the mechanism of screw sense induction is produced mostly by fixation of a conformation in one of the residues of the foldamer backbone which is then transmitted to the rest of the oligomer that adopts a preferred *P* or *M* screw sense excess.

## 1.2 Dynamic helical polymers

In stimuli responsive covalent helical polymers, helix induction mechanisms are different from those observed in foldamers. In general, in these polymers, the main chain or polymer backbone is achiral, *e.g.*, polyacetylenes and their derivatives,<sup>39–57</sup> polyisocyanides,<sup>58,59</sup> polysilanes<sup>88–90</sup> or poly(quinoxaline-2,3-diy)ls (PQXs)<sup>53,91–97</sup> (Fig. 2a), which adopt a preferred helical sense due to the conformational composition adopted by the pendant group. Thus, if the pendant is chiral, *e.g.*, poly-5, a poly(phenylacetylene) (PPA) that bears the benzamide of (*S*)-phenylglycine methyl ester,<sup>98</sup> the presence of a preferred conformer at the chiral centre results in an induced helical sense in the polymer main chain.

Variations in the conformational composition of the pendant group by the presence of external stimuli, such as solvent

polarity or the addition of metal ions, can result in either helical sense enhancement or helical inversion effect (Fig. 2b). On the other hand, if the helical polymer is achiral, *e.g.*, a PPA bearing a benzoic group as pendant (poly-6),<sup>99,100</sup> a phenomenon of asymmetry amplification (helical induction) arises from interactions between the polymer and external chiral molecules, such as chiral amines used as external stimuli.

The most frequently helix induction mechanism that governs the screw sense excess adopted by chiral and achiral polymers is tele-induction.<sup>101</sup> In this mechanism, changes in the spatial distribution of substituents in the chiral centres or in the species associated with the pendant groups result in stimuli being transmitted through space to the polymer backbones, which adopt specific conformations that generate *P* or *M* helical structures (Fig. 2b and c).

Interestingly, other helix induction mechanisms operating from the pendants to the main chains of polymers have been explored in recent years.

### 1.2.1 Helix induction mechanisms in dynamic helical polymers

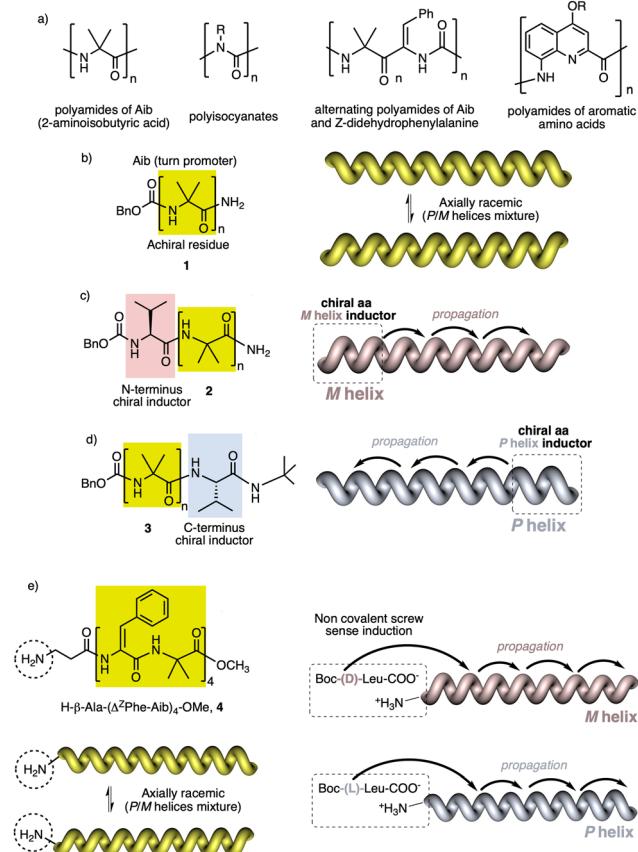
1.2.1.1 *Chiral harvesting.* It is based on the introduction of a chiral source at a remote position separated from the backbone



From left to right: María Lago, Rafael Rodríguez, Manuel Fernández-Míguez, Félix Freire, Emilio Quiñoá

PhD at the University of Santiago de Compostela/CiQUS (Spain) working on the structural elucidation and applications of stimuli responsive helical polymers—awarded as best thesis in science in 2018—under the supervision of Prof. Félix Freire and Prof. Emilio Quiñoá. He performed two postdoctoral stays at Kanazawa University (Japan) and the Institut des Sciences Chimiques de Rennes (University of Rennes 1, France) in the Research groups of Prof. Katsuhiko Maeda and Dr Jeanne Crassous to work in chiral materials with CPL activity. In 2021, he was awarded with a Juan de la Cierva Incorporación fellowship to move back to University of Santiago de Compostela/CiQUS (Spain) to develop chiral materials with switchable CPL activity. Recently, he was awarded with a prestigious Ramón y Cajal contract. Emilio Quiñoá obtained his PhD in Chemistry from the University of Santiago de Compostela (USC) on the topic of bioactive marine natural products. After postdoctoral research at the University of California, Santa Cruz (UCSC), he became an Associate Professor (1989) and later a Full Professor of Organic Chemistry (1998) at USC. His research then addressed the development of methods for the assignment of absolute configurations by NMR spectroscopy among other interests. He has coauthored more than 160 scientific publications, several textbooks and patent applications and has received several research awards. In 2010 he joined the Center for Research in Biological Chemistry and Molecular Materials (CiQUS) at USC. His current research interests focus on the study of dynamic helical polymers and their applications in nanotechnology. Félix Freire obtained his BSc (2000), MSc (2002), and PhD (2005) in Chemistry from the University of Santiago de Compostela (USC). In 2005 he did a postdoctoral stay in the group of Prof. Jesús Jiménez Barbero at CSIC-Madrid, where he worked on NMR studies of biomolecules. During 2006–2008 he was a Fulbright Postdoctoral Fellow in the group of Samuel H. Gellman, studying the folding of parallel  $\beta$ -sheets in water. Since 2009 he has worked at the University of Santiago, first as a Ramón y Cajal Researcher and now as Full Professor of Organic Chemistry (2023). His research interests include stimuli-responsive polymers, molecular self-assembly, and chiral polymer particles.



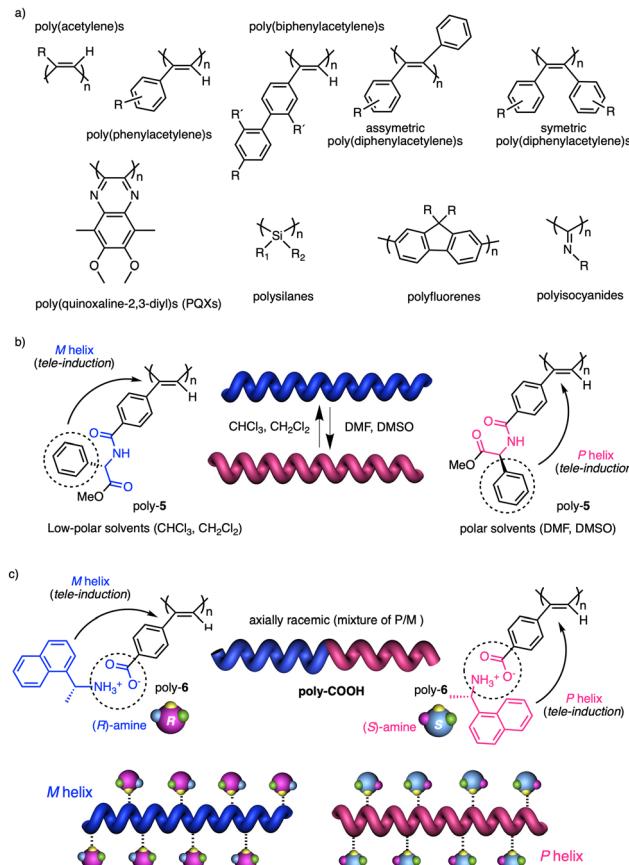


**Fig. 1** (a) Some achiral oligomer families forming axially racemic helices. Schematic illustration of (b) an axially racemic oligoamide of Aib and its (c) *M* or (d) *P* screw sense induction by introducing a chiral (L)-amino acid at the N- and C-terminus respectively. (e) Schematic illustration of the non-covalent screw sense induction of the axially racemic foldamer H-β-Ala-(Δ<sup>2</sup>Phe-Aib)<sub>4</sub>-OMe by interaction with chiral acids.

by achiral spacers. In such a case, the information transmission mechanism can follow two different paths. The induction of: (i) a conformational change in the achiral spacer,<sup>102</sup> or (ii) a tilting degree in the supramolecular arrangement of the achiral spacers along the helix.<sup>103</sup> Both mechanisms are triggered by changes in the conformational composition of a chiral group placed in a remote position, which are transmitted to the achiral spacer and further harvested by the polymer backbone that adopts a *P* or an *M* screw sense excess.

Two illustrative examples are poly-7 and poly-8, PPAs that bear an achiral and flexible Aib residue<sup>102</sup> and a planar and rigid oligo(phenylene-ethynylene) (OPE)<sup>103</sup> as spacers (Fig. 3a and b, respectively).

**1.2.1.2 Chiral overpass.** Usually, when the pendant of a helical polymer is composed by two or more chiral centres, the one closest to the backbone of the polymer is more likely to control its handedness, while the other centre has low-to-null impact on the structural features of the helical structure.<sup>104</sup> Interestingly, the possibility of playing with the conformational composition of these multi-chiral side chains makes possible to activate a chiral-overpass effect.<sup>105,106</sup> For example, in poly-9,



**Fig. 2** (a) Some families of dynamic helical polymers. (b) Helix inversion of a chiral poly(phenylacetylene) bearing 4-benzamide of (S)-phenylglycine methyl ester as pendant group by tuning its conformational composition using solvent polarity as external stimulus. (c) Chiral amplification of an achiral poly(phenylacetylene) bearing 4-benzoic acid as pendant using chiral amines as external stimuli.

which bears the anilide of (S)-alanine-(R)-methoxyphenylacetate—(S)-Ala-(R)-MPA dimer—as pendant,<sup>106</sup> two different conformations, extended and bent, can occur (Fig. 4a). Thus, if the dimer adopts an extended conformation, (S)-alanine, located closer to the poly(phenylacetylene) main chain, commands a preferred macromolecular structure (screw sense and elongation) in the PPA (*M* helix, Fig. 4a). On the other hand, by fixing a bent structure at the dimeric pendant, it is possible to place the second chiral residue close to the backbone, which is now responsible for the screw sense preference adopted by the macromolecular helical polymer<sup>105,106</sup> (*P* helix, Fig. 4a).

However, if the (S)-Ala-(S)-MPA pendant is used, *M* helices are obtained in both extended and bent conformations (Fig. 4a).

**1.2.1.3 Substituent priority overpass.** This helix induction mechanism is found in dynamic helical polymers bearing conformationally locked chiral pendants such as chiral allenes.<sup>107</sup> In these systems, the relative spatial orientation of the substituents is always the same, and it is not possible to modulate it by altering the conformational composition of the chiral moiety. In such cases, to invert the helical sense of the polymer it is necessary to modify the relative volume of one of the substituents.

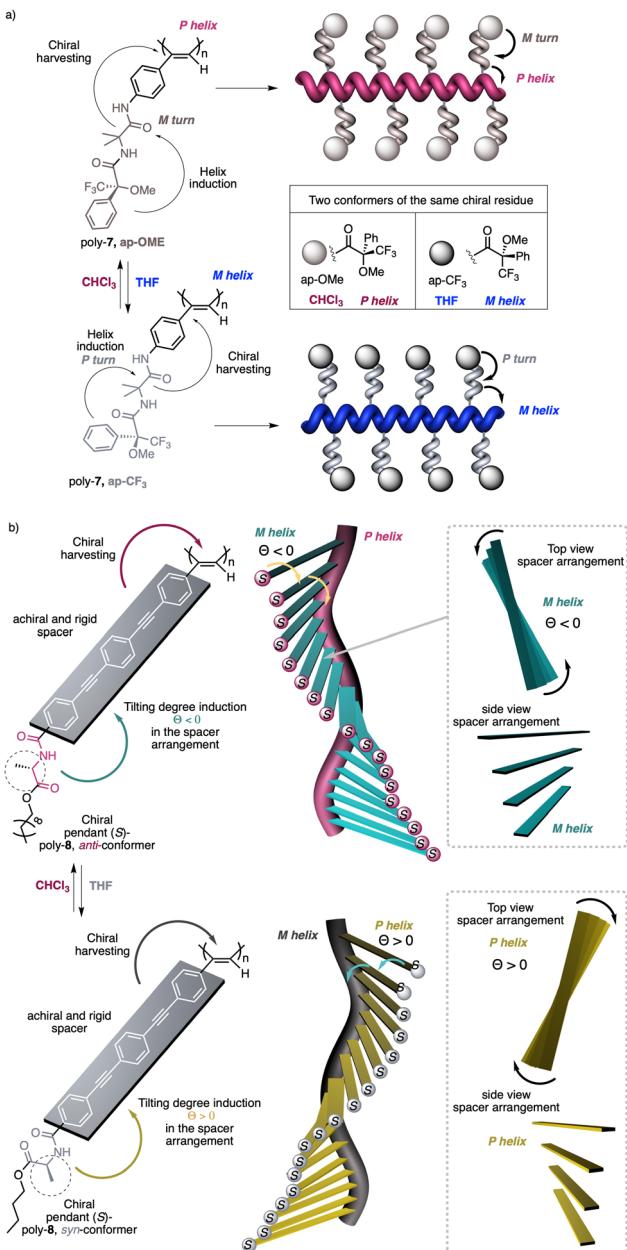


Fig. 3 Screw sense induction of helical polymers through chiral harvesting using (a) flexible and (b) rigid achiral spacers.

As a result of this volume change, a new substituent located in a different position will be responsible for producing variations in the macromolecular structure and inducing the screw sense preference.

To show this effect, poly-10 was chosen as illustrative example.<sup>107</sup> This polymer bears an allene pendant (Fig. 4b) in which its two consecutive double bonds place their four substituents in two perpendicular planes. If these substituents are different, the allene becomes chiral, as in poly-10, which shows *P* axial chirality. In poly-10, the screw sense is governed by the *tert*-butyl group located closest to the backbone (substituent a). However, if a secondary or tertiary amine are added to a solution containing poly-10, a supramolecular interaction is

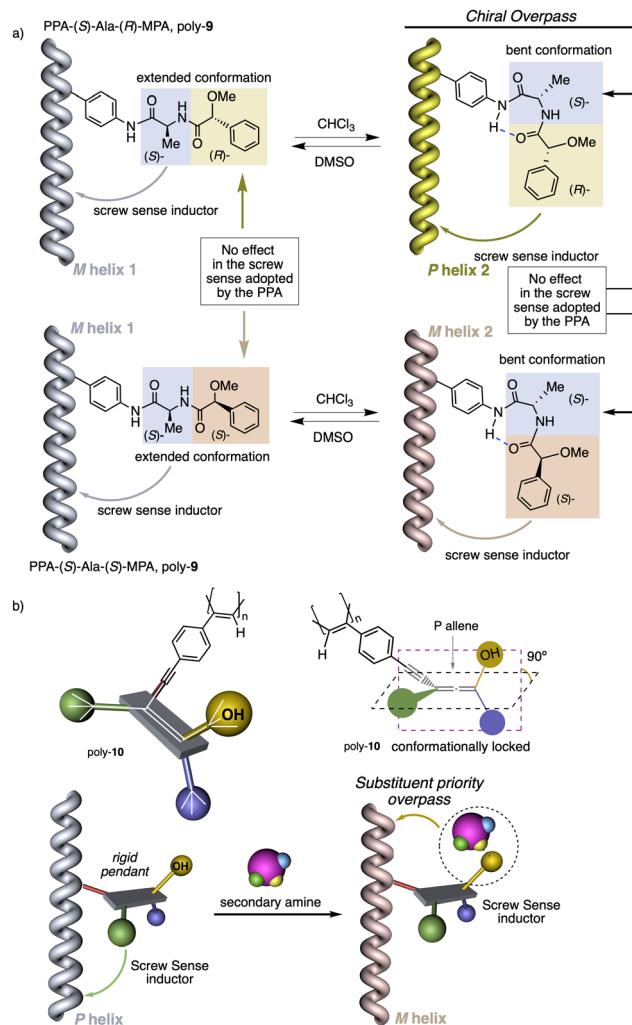


Fig. 4 (a) Schematic illustration of the chiral overpass effect in poly(phenylacetylenes). (b) Chiral amplification of a poly(phenylacetylene) bearing a chiral allene as pendant using chiral amines as external stimuli.

established between the alcohol group (substituent b) of the allene and the amine, increasing the size of this substituent. As a result, substituent b, placed in a different spatial orientation than substituent a, now controls the helical sense that poly-10 adopts, resulting in a helix inversion process.

**1.2.2 Dynamic helical copolymers.** Until now, we have described the most abundant helix induction mechanisms found in dynamic helical polymers constituted by a single monomer. However, these helix induction mechanisms can also occur in dynamic helical copolymers made by two different monomers. In this case, in addition to the helical sense induction effect produced by the pendants in the polymer backbone, it is necessary to consider other communication mechanisms between the different components of the copolymer, which can be either chiral/achiral or chiral/chiral. In general, in these systems, one of the monomers must adopt a preferred conformation that activates a specific conformation in the other comonomer (chiral or achiral). Thus, depending on the chiral composition of the comonomers, different effects can be triggered, such as:



**1.2.2.1 Sergeants and Soldiers effect<sup>108–111</sup>** (*Sergeant: chiral (R or S), minor component; Soldier: achiral, major component*). The Sergeant orders the Soldier to adopt a conformation that promotes the same helical scaffold (sense and elongation) induced by the Sergeant.

**1.2.2.2 Abnormal Sergeants and Soldiers effect<sup>94</sup>** (*Sergeant: chiral, minor component (R or S); Soldier: achiral, major component*). The Sergeant activates a conformation in the Soldier that promotes a different helical scaffold than the one adopted by the Sergeant (sense and/or elongation).

**1.2.2.3 Chiral to chiral Sergeants and Soldiers effect<sup>112</sup>** (*Sergeant: chiral, minor component (R or S); Soldier: chiral (R or S), major component*). The Sergeant commands a specific conformation on the Soldier, which differs depending on the absolute configuration of the chiral Soldier (Fig. 5a and b). Thus, the two enantiomeric configurations of the Soldier can adapt to the scaffold induced by the Sergeant. For example, a PPA bearing the 4-anilide of (R or S)-methoxyphenylacetic acid (poly-11) behaves axially racemic (ECD = 0, mixture of *P* and *M* helices) even though the pendant is chiral.<sup>112</sup> This fact is due to the presence of two conformers in equilibrium at the pendant, *ap* and *sp*, which place the substituents of the chiral moiety in different spatial locations, inducing opposite helical senses in the PPA (Fig. 5a). Interestingly, replacement of the phenyl group by an anthryl ring (poly-12) shifts the conformational composition of the pendant towards the *ap* conformer (Fig. 5a).<sup>111</sup> When the monomers of poly-12 (Sergeant) and poly-11 (Soldier) are copolymerized to create poly[(S)-12<sub>(Sergeant)</sub>-co-(S)-11<sub>Soldier(1-r)</sub>] and poly[(R)-12<sub>(Sergeant)</sub>-co-(S)-11<sub>Soldier(1-r)</sub>] copolymer series (Fig. 5b), it was observed that both series adopt opposite helical senses, *P* and *M*, commanded by the (S)- or (R)-chirality of the Sergeant respectively. The (S)-Soldier fits into the *P* and *M* helical scaffolds adopting the *ap* or *sp* conformations respectively.<sup>112</sup>

**1.2.2.4 Chiral coalition or abnormal chiral to chiral Sergeants and Soldiers effect<sup>113</sup>** (*Sergeant: chiral, minor component; Soldier: chiral, major component*). In this case, the role of the Sergeant is to behave like a chiral dopant, triggering a specific conformation in the Soldier that is different depending on the chirality of the soldier. As a result, poly[(S)-Sergeant,-co-(S)-Soldier<sub>(1-r)</sub>] and poly[(R)-Sergeant,-co-(S)-Soldier<sub>(1-r)</sub>] adopt the same helical structure, which is commanded by the chirality of the Soldier, *e.g.*, poly[(S)-5,-co-(S)-11<sub>(1-r)</sub>] and poly[(R)-5,-co-(S)-11<sub>(1-r)</sub>] (Fig. 5c).<sup>113</sup> However, the Sergeant is necessary to trigger a specific conformation in the Soldier, otherwise, without its presence, the Soldier homopolymer behaves like an axially racemic helix.

**1.2.2.5 Majority rules<sup>114</sup>** [*Sergeant: chiral; Soldier: chiral (Sergeant's enantiomer)*]. In this case, the screw sense preferences of copolymer series made by two enantiomeric monomers (identical chemical structure, mirror image relationship) are biased towards *P* or *M* helices, commanded by a slight enantiomeric excess of one of the enantiomers. For instance, in poly[(R)-Sergeant<sub>0.56</sub>-co-(S)-Soldier<sub>0.44</sub>], made by enantiomeric monomers

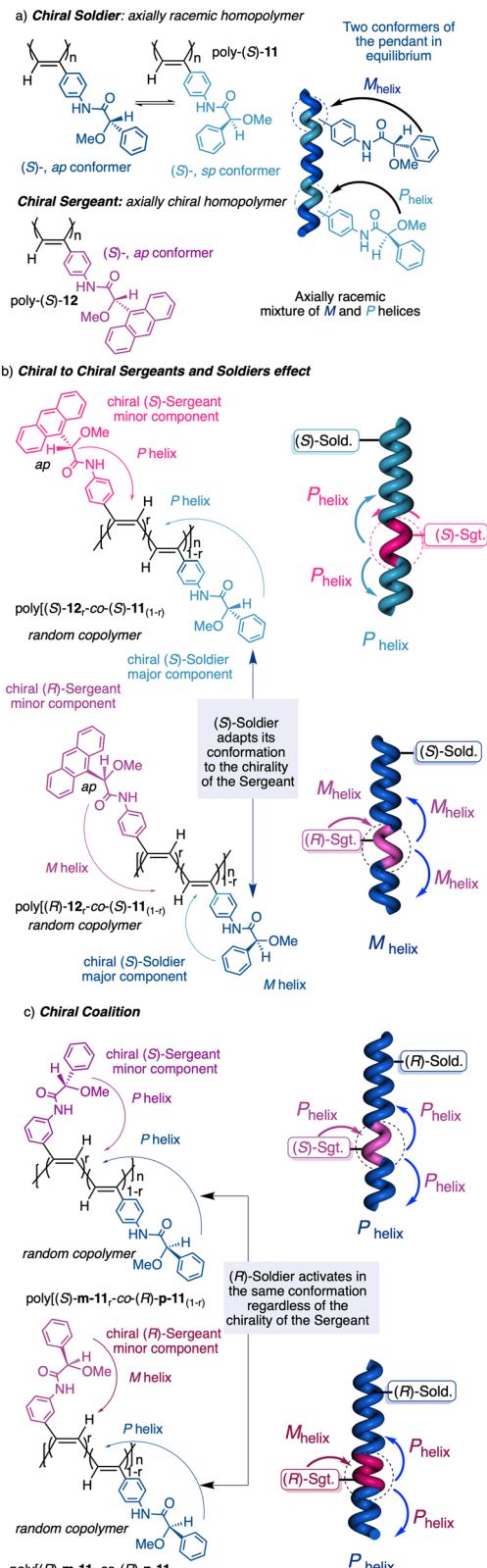


Fig. 5 (a) Chiral Soldier. (b) Chiral to chiral Sergeants and Soldiers effect. (c) Chiral coalition.



of 2,6-dimethylheptyl isocyanate (56% (*R*)-enantiomer/44% (*S*)-enantiomer; ee 12%), the helix adopted by the copolymer is identical to that adopted by the (*R*)-homopolymer.<sup>114</sup>

**1.2.2.6 Chiral accord and chiral conflict**<sup>115–117</sup>. Other interesting phenomena can occur in chiral dynamic helical copolymers although there is not amplification of asymmetry due to the lack of communication between the two chiral components. Thus, if the two comonomers of the copolymer induce independently the same helical sense, a chiral accord<sup>116</sup> emerges, whereas if both comonomers induce opposite helical senses, a chiral conflict<sup>116,117</sup> is produced.

In short, the presence or absence of communication between co-monomers in dynamic helical polymers can produce interesting stimuli-responsive properties in the materials due to the dual or independent stimuli-responsiveness of the comonomers towards a specific stimulus.

### 1.3. Supramolecular helical polymers

In supramolecular helical polymers,<sup>5,18–38</sup> helices are achieved by the presence of tilting degrees generated between building blocks during the supramolecular polymerizations. Positive or negative values of these tilting degrees are induced by the absolute configurations of the chiral groups introduced in the building blocks. Thus, depending on their absolute configurations, *P* or *M* supramolecular helices will be formed.

The stimuli-responsive properties of those helices can produce the disassembly of the helical structures, helix inversion or the formation of other aggregates.

In this review, we will only focus on helical structural changes that result in a helix with opposite helical sense or the formation of another type of helical aggregate, such as J-type or H-type aggregates. To go from one supramolecular helical structure to another, two different mechanisms can be followed, a competitive pathway<sup>118</sup> or a consecutive one.<sup>119,120</sup>

In a competitive pathway, a kinetic supramolecular aggregate disassembles to provide the molecularly dissolved monomer that further aggregates (thermodynamic aggregate) into a different helical structure (sense and/or scaffold). The aggregation of an asymmetric oligo(phenyleneethynylene) (OPE) bearing the anilide of (*S*)-MPA [(*S*)-13] was chosen as illustrative example.<sup>118</sup> In this case, the kinetic aggregate (AggI, *M* helix) was kinetically trapped in MCH/Tol/DCM solvent mixture at room temperature, while the thermodynamic aggregate (AggII, *P* helix) was obtained in MCH/DCM solvent mixture after thermal treatment (Fig. 6a).

In the consecutive pathway, one helical structure is transformed into a different one, without going through the monomeric state. For example, the *N*-annulated perylanenetracarboxamide (*S*)-14<sup>120</sup> aggregates to form fibrillar structures that evolve to form thicker helical assemblies (Fig. 6b).

Supramolecular and helical polymers share many properties, such as the asymmetry amplifying effects found in their copolymers. Thus, Sergeants and Soldiers or Majority Rules can be found in both systems,<sup>121–123</sup> although their information transmission mechanisms to induce screw sense excess are different.

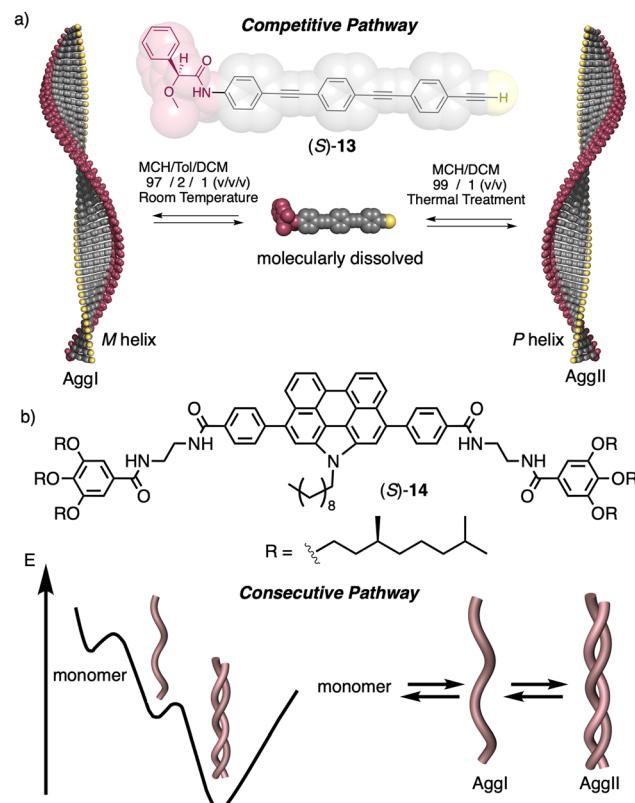


Fig. 6 Supramolecular aggregation following a (a) competitive or a (b) consecutive pathway.

## 2. Stimuli-responsive helical polymers

In this section we will use different examples found in the literature to show how helical structures adopted by foldamers, supramolecular and covalent helical polymers can be modified (helix inversion, amplification of asymmetry or variations in the helical scaffold) through interaction with different stimuli.<sup>124</sup> We will focus our attention on those systems where the structural changes are clearly identified, although we also will go through other interesting examples where the structural changes associated to the presence of an external stimulus are not clear.

### 2.1 External stimulus: temperature

Temperature is involved in the activation/deactivation of supramolecular forces and in the variation of the conformational composition of molecules. Hence, thermal control of screw sense in foldamers and polymers can be achieved by taking these parameters into account during macromolecule design.

**2.1.1 Foldamers.** It is the family of macromolecular helices that presents fewer thermo-responsive examples in the literature. The main reason is the importance of supramolecular forces to keep the structure folded in a specific screw sense excess. Playing with temperature generally affects the stability of the helical oligomer structure, resulting in an equilibrium between a folded structure (low-temperature) and a random coil macromolecular structure (high-temperature).

Inai designed a dynamic helical foldamer consisting of a nonapeptide made mostly by achiral residues such as Aib,  $\beta$ -Ala ( $\beta$ -alanine) and  $\Delta^Z$ Ph ( $Z$ - $\alpha$ , $\beta$ -dehydropheylalanine) and a single chiral residue,<sup>125</sup> L-Phe ( $L$ -phenylalanine), placed in an internal position [H- $\beta$ -Ala- $\Delta^Z$ Ph-Aib-( $L$ )-Phe-( $\Delta^Z$ Ph-Aib)<sub>2</sub>-OMe, 15] (Fig. 7a). The chiral residue is necessary to command a *P* helix in the foldamer, which is protected at the C-terminus with a methyl ester and has a free NH at the N-terminus, necessary to establish supramolecular interactions with chiral acids. Interestingly, when foldamer 15 interacts with Boc-*D*-amino acids such as Boc-*D*-Pro-OH or Boc-*D*-Leu-OH, a *P* to *M* helix inversion occurs in the foldamer. Activation/deactivation of the supramolecular amino/carboxylic acid interaction between the foldamer and the Boc-*D*-amino acids results in a reversible helix inversion process between a *P* helix (high temperature, supramolecular interaction OFF) and an *M* helix (low temperature, supramolecular interaction ON).<sup>125</sup>

In another work, Formaggio and De Zotti<sup>126</sup> found that by replacing the amino alcohol leucenol (Lol) by leucine (Leu-OMe or Leu-OH) at the C-terminus of the natural occurring trichogin (a peptaibol, *n*Oct-Aib-Gly-Leu-Aib-Gly-Gly-Leu-Aib-Gly-Ile-Lol, 16), the new peptides 16-OMe and 16-OH become thermo-responsive transitioning from a *P* to an *M* helix with increasing temperature (Fig. 7b). Again, these peptide sequences contain many flexible and achiral residues, such as Gly and Aib, that can be accommodated in the *P* and *M* helical senses.

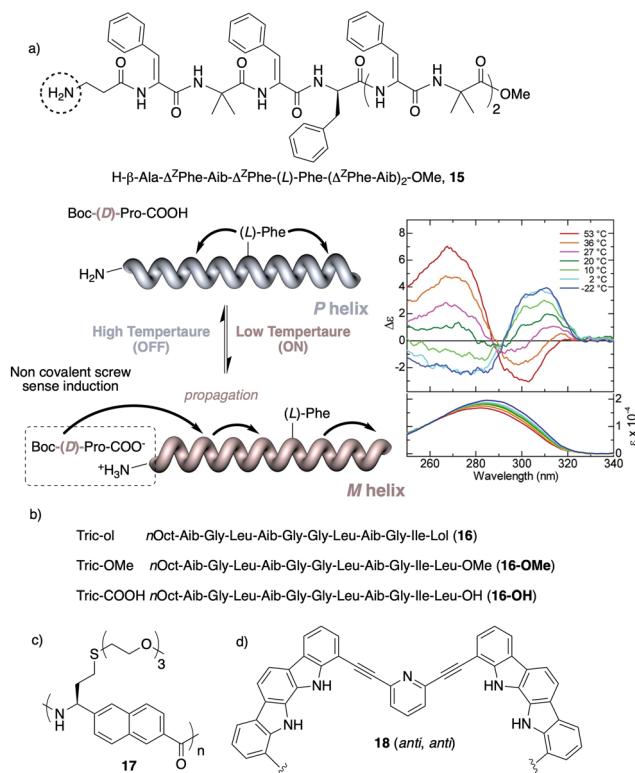


Fig. 7 Temperature responsive foldamers: Helix inversion (a) 15 (reproduced from ref. 125 with permission from the American Chemical Society, copyright 2007) and (b) 16; elongation (c) 17 and (d) 18.

In another example, Onitsuka and Kanbayashi<sup>127</sup> used temperature to change the elongation of an arylopeptide that contains the axially unsymmetrical 2,6-naphthylene as spacer between the amino and the acid group (17) (Fig. 7c). This amino acid has a long oligoether group in the side chain, a thermo-responsive substituent that exhibits lower critical solution temperature (LCST) behavior due to the transition in the hydration of the side chains. At 25 °C, foldamer 17 is soluble in water and adopts a 3<sub>1</sub>-helix, while when heated this helical scaffold stretches and transforms into a 4<sub>1</sub>-helix. In the field of aromatic helical foldamers, Jeong<sup>128</sup> prepared one that contains two indolocarbazole and two pyridine moieties (18) (Fig. 7d). The addition of Pd(CH<sub>3</sub>CN)<sub>2</sub>Cl<sub>2</sub> generated a double-stranded helicate, which at low temperatures favors a more stretched helix (*syn-anti* orientation of the internal pyridine) and a more compact one at high temperatures (*syn-syn* orientation of the internal pyridine).

**2.1.2 Dynamic helical polymers.** In literature there are several examples dealing with temperature responsive helical polymers, being poly(phenylacetylene)s the family with more examples. For instance, Tabata<sup>129,130</sup> studied the irreversible helix rearrangement from *cis-cisoid* (compressed) to *cis-transoid* (stretched) scaffolds in the solid state of PPAs bearing achiral pendant groups such as *n*-octyl or *n*-hexyloxy alkyl chains (19, 20) (Fig. 8a). In both cases, changes in the elongation of the polymer are accompanied by changes in the color of the solid sample. Interestingly, they also found that these structural changes can also occur in solution but in a reversible way. By using a PPA that bears the (*S*)-2-octyl propiolate (21) (Fig. 8a) as pendant group, they found that three helices, with the same helical sense but different elongation degree, coexist within a polymer chain. Variable temperature experiments allowed the equilibrium to be shifted from one helical structure to another in an accordion-like helix oscillation (HELIOS) due to variations in conformational composition in the chiral ester group.<sup>131</sup> In polyacetylenes, Masuda also found a helical polymer bearing a chiral ester group (22) (Fig. 8a and b) that shows helix inversion by thermal treatment.<sup>132</sup>

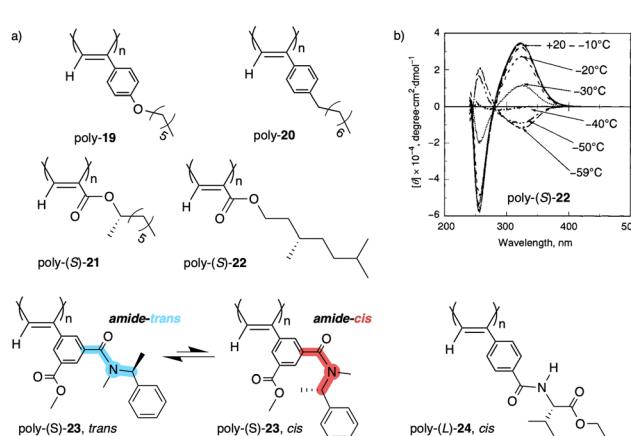


Fig. 8 (a) Chemical structures of polymers poly-(19–24). (b) ECD spectra of poly-(S)-22 at different temperatures (ECD thermal switch) (reproduced from ref. 132 with permission from the American Chemical Society, copyright 2001).

Another interesting example dealing with a temperature driven conformational change at the pendant was described by Wan<sup>133</sup> for a 3,5-disubstituted PPA, poly{[−)-3-methoxycarbonyl-5-[N-methyl-N-(S)-(1-phenylethyl)carbamoyl]-phenylacetylene} (23) (Fig. 8a). In this case, the authors play with the *cis/trans* conformational composition of the tertiary amide using temperature.

Thus, while the *trans* isomer is favored at low temperatures, the *cis* ratio increases when the temperature is raised. As a result, a reversible helix inversion is produced generating a thermal helical switch. Zhang and Liu reported another PPA that works as thermal helical switch by using the benzamide of (L)-valine-OEt (24) as pendant (Fig. 8a).<sup>134</sup> In this case, the macromolecular helical structure adopted by the polymer in chloroform at room temperature can be reversed by cooling the solution to 15 °C or lower. Following in the line of macromolecular helical changes induced by conformational changes driven by temperature in the pendant groups, an interesting example is the PPA designed by Bergueiro, Freire and Calderón,<sup>135</sup> which includes elastin as pendant (poly-25) (Fig. 9a). Elastin is a thermo-responsive peptide, which undergoes conformational changes when the temperature of the solution varies.

In this case, the thermal behavior of elastin with increasing temperature changes from a lower critical solution temperature (LCST) to an upper critical solution temperature (UCST) once polymerized. Below the cloud point temperature, the elastin in the pendants adopts a hydrophobic state ( $\beta$ -spiral conformation), which results in the adoption of an elongated structure. Above the cloud point temperature, the pendant adopts a more hydrophilic state (extended conformation), which induces a more compressed helical scaffold. Yashima demonstrated, through a beautiful example, that a thermal switch can be created from a supramolecular PPA/chiral amine complex.<sup>136</sup> Here, the PPA is achiral and bears a bulky phenyl phosphonate group on the pendant (26) (Fig. 9b).

The resulting polymer is achiral, and a mixture of *P* and *M* helical senses are obtained. However, by adding chiral amines such as (R)-1-(1-naphthyl)ethylamine, a chiral complex is formed in DMSO and a *P* screw sense excess is induced in poly-26 at 25 °C. In this complex, the helical sense is commanded by the phosphonate group, which became chiral after complexation with the chiral amine as inferred from VCD studies. Interestingly, when the DMSO solution of poly-26/(R)-1-(1-naphthyl)ethylamine complex heats up to 65 °C, a helix inversion from *P* to *M* is observed in the PPA, which is accompanied by a planarity of the phosphonate VCD signal. In this case, the authors indicate that phosphonate may racemize or exist as the achiral form due to the resonance effect of the P—OH and P=O group, and therefore the helix induction comes from another moiety of the PPA/amino complex, *i.e.*, the chiral amino group. Yashima created another thermal switch based on supramolecular complexes between an achiral PPA that bears a *N,N*-diisopropylaminomethyl group as pendant (27) (Fig. 9c) and chiral substrates such as chiral acids.<sup>137</sup> In this example, no changes in the helical sense of the poly-27/chiral acid complex were observed by temperature changes. However, they found that a colorimetric thermal switch is obtained by variations in the polymer elongation. Thus, at low temperatures (0–20 °C), a

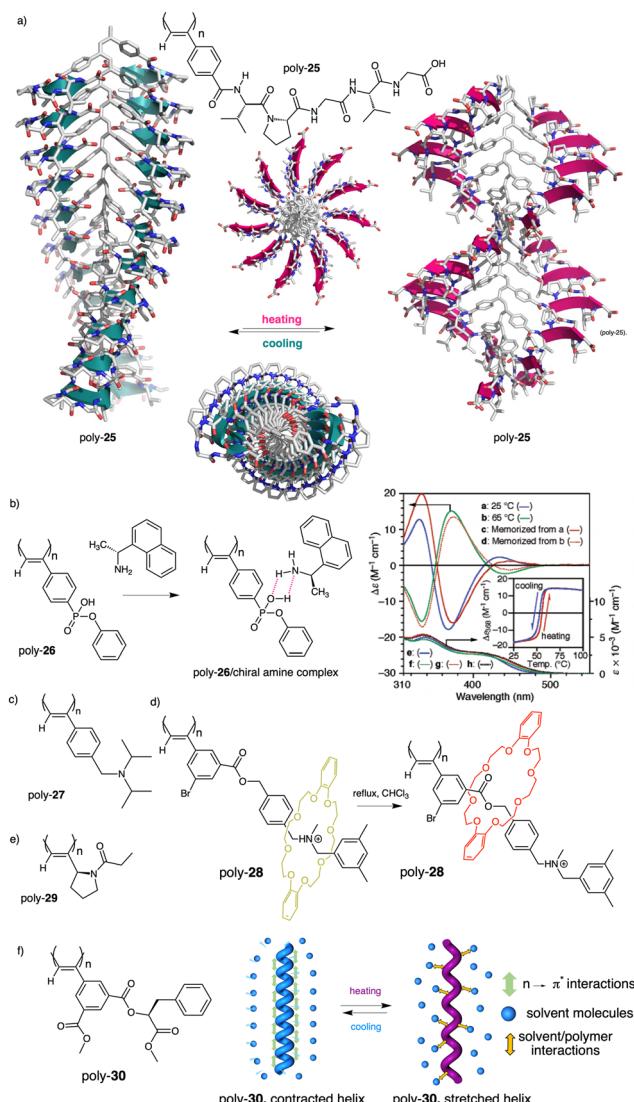


Fig. 9 (a) Schematic representation of the thermal behaviour of a PPA bearing an elastin derivative as pendant group (poly-25). (b) Chemical structure of poly-26 and thermoresponsive behaviour of poly-26/(R)-1-(1-naphthyl)ethylamine complex (reproduced from ref. 136 with permission from the American Chemical Society, copyright 2005). Chemical structures of (c) poly-27, (d) poly-28 and (e) poly-29. (f) Structure of poly-30 and its thermoresponsive behaviour.

compressed helical structure is induced in poly-27/chiral acid complex (yellowish solution) while at higher temperatures (25–40 °C), a stretched helix is obtained (orangeish solution). In relation to the development of PPA thermal switches based on changes in polymer elongation, Takata reported a PPA that has a thermo-responsive rotaxane as pendant (poly-28) (Fig. 9d).<sup>138</sup>

Thus, depending on the position of the crown ether in the pendant, the polymer will adopt different degrees of stretching. At high temperatures, the ring of the rotaxane will sit close to the polyene backbone, resulting in a highly stretched helix. Conversely, at lower temperatures, the crown ether ring will be situated far from the backbone, adopting the PPA a more compressed helical structure. This structural change is accompanied by a



color change of the solution, from red (high temperatures) to yellow (low temperatures) that also occurs in the solid state.

Wan reported a polyacetylene, poly[(S)-2-ethynyl-N-propionylpyrrolidine] (poly-29) (Fig. 9e),<sup>139</sup> derived from proline, that shows a LCST behavior in water accompanied by elongation changes in the chiral polyacetylene helix. Thus, below 32 °C, the polymer adopts a compressed *P* helix that produces a clear solution, whereas at temperatures higher than 32 °C, a more stretched helix is obtained that generates a cloudy solution due to the formation of aggregates. Zhang and Wan reported a 3,5-dibenzoate PPA (poly-30) (Fig. 9f)<sup>140</sup> that also can vary its structure between compressed and stretched helical scaffolds in chloroform by deactivating, at high temperatures, the n- $\pi^*$  interactions between the oxygen of one carbonyl group and the carbon of another carbonyl in nearby upper or lower position within the helix. These n- $\pi^*$  interactions stabilize compressed *cis-cisoidal* polyene structures. At high temperatures, new interactions between solvent and pendants stabilize stretched *cis-transoidal* helices. Percec, Meijer and Zhang studied how temperature produces structural changes (elongation, helix sense) in dendronized PPAs. Thus, Meijer<sup>141</sup> designed a PPA where the dendron grows directly from the phenylacetylene backbone. To do that, they did a 3,4,5-trisubstitution at the aryl ring with a chiral ether—poly[3,4,5-tris((S)-3,7-dimethyloctyloxy)phenylacetylene], poly-31 (Fig. 10a). The resulting polymer shows a helix inversion from *M* to *P* in a very narrow temperature range (*M* helix at 20 °C, *P* helix at 30 °C), which is accompanied by a stretching of the helical scaffold. Percec designed a dendronized PPA<sup>142,143</sup> where the dendron and the PPA backbone are separated by a chiral ether group (poly-32) (Fig. 10b). This polymer shows a reversible *cis-cisoidal* to *cis-transoidal* (contraction/extension) thermoresponsive isomerization of the backbone that can act as a nanomechanical actuators. When extruded as fibers, they have been shown capable of work by displacing objects up to 250-times their mass (Fig. 10b). Li and Zhang<sup>144,145</sup> designed water soluble PPAs bearing dendritic oligo(ethylene glycol) (OEG) as pendants using chiral alanine as spacer between the dendron and the PPA backbone. Different linkages were used to connect the amino acid to the PPA, such as anilide (poly-33) (Fig. 10c) or benzamide (poly-34) (Fig. 10d), which provide the polymer with different thermoresponsive properties such as reversible helix inversion (poly-33) or reversible contraction/extension (poly-34). This fact indicates that the amide linkage, benzamide/anilide, plays an important role in the thermal behavior of dendronized PPAs. Interestingly, the cloud point temperature ( $T_{cp}$ ) of poly-33 can be tuned by salt-in and salt-out anions. Thus, while  $\text{PF}_6^-$  and  $\text{SCN}^-$  (salt in anions) largely increased the  $T_{cp}$ ,  $\text{Cl}^-$  or  $\text{SO}_4^{2-}$  (salt out anions) decreased the  $T_{cp}$ . In the case of adding salt in anions, the helix inversion occurs at the  $T_{cp}$  without structural changes in the PPA, whereas by adding salt out anions, temperature changes produce variation in the polymer elongation. This fact indicates that structural changes are produced by competitive interactions between OEG moieties, water, and anions (Fig. 10e).

Apart from PPAs, there are other polyacetylene derivatives, such as poly(biphenylacetylene)s or PDPAs, that show

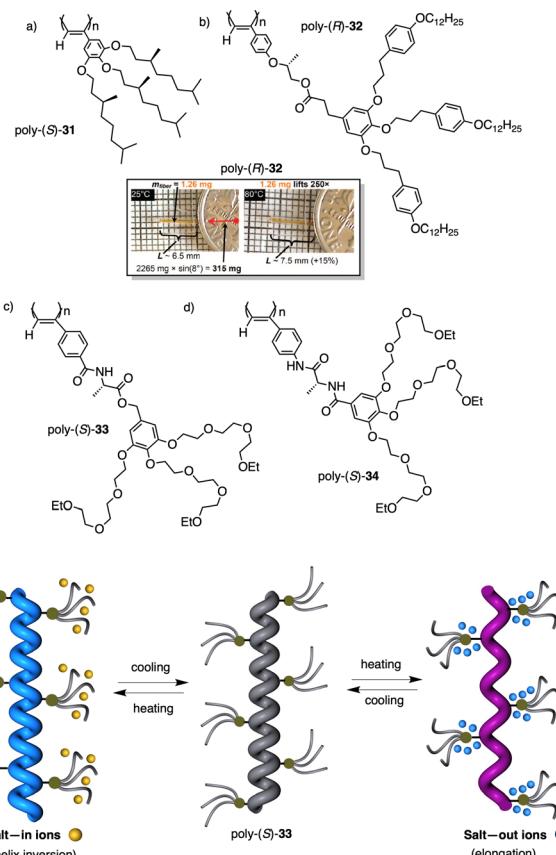


Fig. 10 Structures of (a) poly-31, (b) poly-32 (reproduced from ref. 143 with permission from the American Chemical Society, copyright 2008), (c) poly-33 and (d) poly-34. Schematic representation of the thermal behaviour of poly-33.

thermoresponsiveness. In both systems, the large steric hindrance in the backbone results in a kinetically trapped helical structure with a certain helical sense excess that can evolve towards the thermodynamic helicity by temperature changes. For instance, Maeda<sup>146</sup> designed a PPA that bears 4-biphenyl pendants with methoxymethyl (MOM) groups (poly-35) (Fig. 11a) at the 2,2'-position. This pendant is achiral, and therefore, the resulting polymer behaves as axially racemic.

A screw sense preference can be induced in poly-35 (Fig. 11a) by adding a chiral alcohol as external stimulus. Interestingly, depending on the temperature used to form the complex, the screw sense induced in poly-35/chiral alcohol will be opposite, e.g., poly-35/(S)-1-phenylethan-1-ol complex shows a *P* helix at -10 °C and an *M* helix at 50°. In the field of PDPAs, Freire designed an asymmetric PDPA that bears the benzamide of (L)-alanine methyl ester as pendant (poly-36) (Fig. 11b).<sup>147</sup> This polymer can adopt either *P* or *M* helical sense in different solvents at high temperatures (DMSO, *M* helix;  $\text{CHCl}_3$ , *P* helix). This fact is due to a different conformational composition in the pendant. Interestingly, after solvent removal, the induced helical sense can be kinetically trapped after adding a solvent that induces the opposite helical sense. The evolution to the thermodynamic helix (opposite helical sense) can be controlled by temperature, allowing to create a long-lasting memory effect

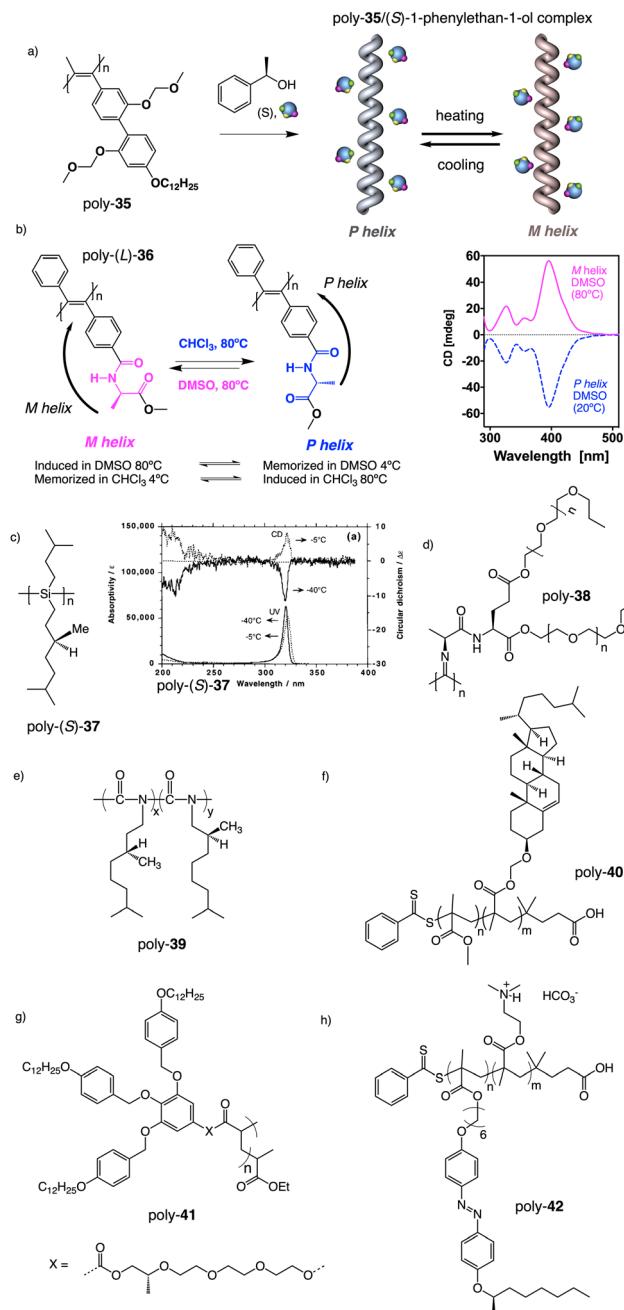


Fig. 11 Thermal driven helix inversion of (a) PBPA poly-35, (b) PDPA poly-36 and (c) polysilylane, poly-37 (reproduced from ref. 149 with permission from the American Chemical Society, copyright 2000). Chemical structures of (d) polyisocyanide poly-38, (e) co-polyisocyanate poly-39, (f) PMMA poly-40, (g) polyacrylate poly-41, (h) PMMA poly-42.

at low temperatures. Fujiki found that poly(silylene)s<sup>89,148–152</sup> containing chiral side chains may exhibit temperature driven helical sense inversion, such as poly{(S)-3,7-dimethyloctyl-3-methylbutylsilylene} (poly-37) (Fig. 11c). Remarkably, in most of these polymers, the macromolecular helix inversion to form the diastereomeric helical structure is accompanied by a bathochromic or hypsochromic shift. Thus, when a depletion and shifting of the ECD signal occurs, in poly(silane)s partial helical inversion takes place. Li and Zhang,<sup>153</sup> studied the thermal

behavior of a polyisocyanide (poly-38) (Fig. 11d) that carries as pendants dipeptides derivatized with oligoethylene glycols (OEGs). In this polymer, the thermal behavior is provided by the OEG chains, while the secondary structure of the polymer remains unaffected. Thus, the OEG chains collapse at high temperatures on the polymer backbone, increasing the turbidity of the solution, while at low temperatures, those chains adopt an extended and solvated conformation.

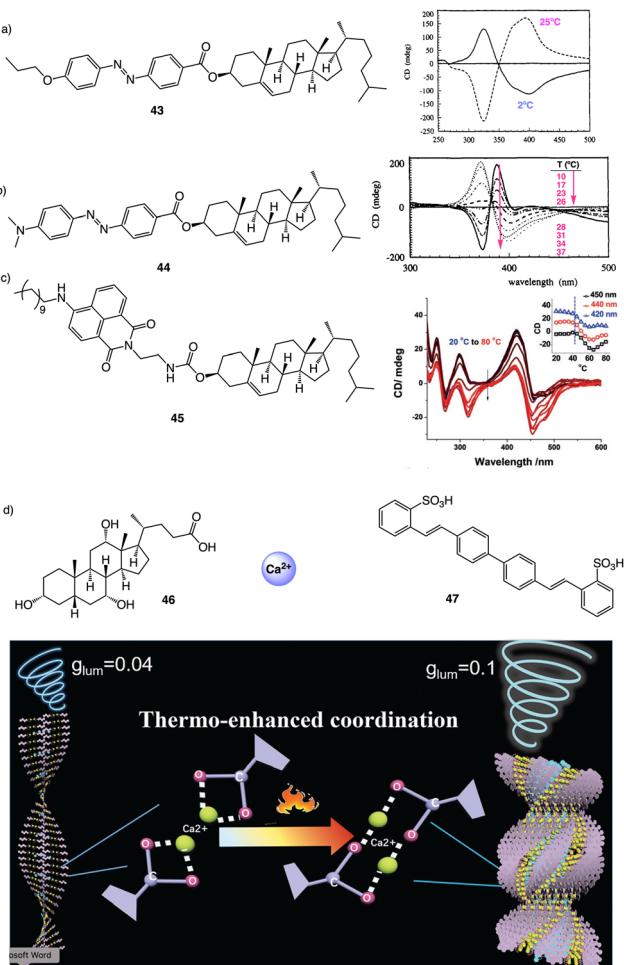
Green and Selinger<sup>154</sup> demonstrated in poly(isocyanate)s, e.g., poly-39 (Fig. 11e), how the absence of chiral-to-chiral communication in helical copolymers made by two chiral comonomers can produce a material with interesting properties such as a thermal behavior. Thus, when a chiral conflict emerges, the two components of a copolymer induce opposite helical senses, and therefore, at a certain comonomer ratio, the copolymer is axially racemic. Interestingly, playing with the temperature, it is possible to modulate the conformational composition of the two components, and therefore the extension of the chiral conflict, which can result in the adoption of a *P* or *M* helical sense depending on the temperature used. Also, in the field of helical copolymers, Mehl, Zhang and Liu<sup>155</sup> found that the thermal behavior of thin film block copolymers (BCPs), comprising poly(methylmethacrylate) (PMMA) and poly(cholesteryl-oxyhexylmethacrylate) (PChMA) blocks, depends on the relative lengths of the blocks (poly-40) (Fig. 11f).

Thus, when the PMMA block is short, a thermal helix inversion is produced in the BCP, while no structural changes are observed for those containing longer PMMA blocks.

They attribute this different behavior in the two-copolymer series to the number of methyl substituents in the backbone, which affect the copolymer packing. In polyacrylates and PMMAs, Percec<sup>156</sup> and Zhang<sup>157</sup> demonstrated through beautiful designs how, regardless of the tacticity or the helical structure adopted by the backbones, the pendants can self-assemble into *P* or *M* axial arrays by inserting into them a flexible chain used as a spacer that is connected to a planar and rigid fragment involved in  $\pi$ - $\pi$  supramolecular interactions [aromatic dendrons, poly-41 (Percec) (Fig. 11g); azo derivatives, poly-42 (Zhang) (Fig. 11h)]. Thus, while Percec introduced the chiral center in the flexible spacer,<sup>156</sup> Zhang introduced it as an external substituent of the azo group at the periphery of the helix.<sup>157</sup>

**2.1.3 Supramolecular helical polymers.** In general, supramolecular helical polymers are sensitive to temperature changes due to the intrinsic nature of supramolecular interactions, weak forces that can be activated/deactivated by temperature. In this review we will focus our attention on those supramolecular aggregates whose axial arrangements change from *P* to *M* or *vice versa* by tuning the temperature. An interesting example was described by Shinkai for cholesterol derivatives containing azobenzene moieties (43–44) (Fig. 12a and b).<sup>158</sup> In these systems, the supramolecular chirality of the aggregate is directly related to the cooling rate used to form a gel. Thus, 43 forms a gel with a positive exciton coupling when a methanolic solution at 60 °C was slowly cooled in air. On the other hand, a negative exciton was obtained when the temperature was rapidly cooled in an ice-water bath at 2 °C. Interestingly, a consecutive





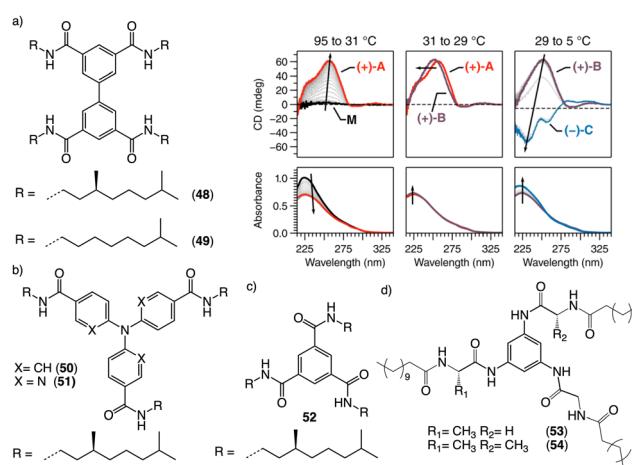
**Fig. 12** Thermal driven helix inversion of supramolecular aggregates using (a) and (b) azo (**43–44**) (reproduced from ref. 157 with permission from the American Chemical Society, copyright 1994) and (c) naphthalimide cholesterol derivatives (**45**) (reproduced from ref. 158 with permission from Wiley-VCH, copyright 2018) as building blocks. (d) CPL enhancement of **47** aligned in a thermoresponsive chiral metallosupramolecular polymer (**46** +  $\text{Ca}^{2+}$ ) (reproduced from ref. 159 with permission from Wiley-VCH, copyright 2018).

*P* to *M* conversion was obtained by heating a cyclohexane gel of compound **44**, where the ECD spectrum gradually inverts from *P* to *M*, with a critical inversion temperature of 27 °C, different from the sol-gel transition temperature that takes place at 45 °C. In a similar design, Zhao prepared cholesterol derivatives containing naphthalimide conjugates (**45**) (Fig. 12c) to create temperature sensitive chiral vesicles by dispersing a small portion of THF stock solution in water.<sup>159</sup>

In the aggregate, the chirality of the cholesterol moiety was transferred to the naphthalimide group producing an *M* chiral aggregate, which progressively evolves towards an aggregate with opposite chirality by heating. This chiral inversion is attributed to the ability of the vesicles to entrap water, which can be released once the system is heated above 45 °C. Cholic acid (**46**) (Fig. 12d) was also used by Yan to create thermo-responsive metallo-supramolecular hydrogels.<sup>160</sup> Cholic acid self-assembles in the presence of  $\text{Ca}^{2+}$  to form fibrillar

structures that promote the formation of a chiral gel in water. The coordination mode between the carbonate groups of cholic acid and the  $\text{Ca}^{2+}$  ions released by  $\text{Ca}(\text{NO}_3)_2$  changes from the metastable bidentate chelating mode to the thermally stable bridging mode with increasing temperature. As consequence, individual nanohelices bunch together to form large scale helical bundles. This effect was used to align a light emitting fluorescent dye within the fibers and create a circularly polarized light emitting system (CPL). The CPL intensity of the hydrogel doped with the fluorescent dye (**47**) (Fig. 12d) increases approximately 4-fold and the  $g_{\text{em}}$  emission dissymmetry factor increases from 0.04 to 0.1 as the temperature goes from 25 to 50 °C.

Water can play an important role in the formation of supramolecular aggregates even when present as a trace in an organic solvent. Through elegant examples, Meijer showed that chiral supramolecular aggregates based on tetracarboxiamides can adopt different helical structures due to the incorporation of water as structural comonomer. This effect is found not only in homochiral aggregates (**48**) (Fig. 13a),<sup>161</sup> but also in copolymer series that follow the Sergeants and Soldiers effect<sup>121–123</sup> [**48** (chiral, Sergeant) + **49** (achiral, Soldier)].<sup>162</sup> Therefore, these systems depend on water and temperature variations led to different interactions between supramolecular aggregates and water molecules, producing different helical structures with diverse scaffolds and/or senses. This effect was also observed in the self-assembly of triarylamines (**50**, **51**) (Fig. 13b) and chiral benzene tricarboxiamide (BTA) (**52**) (Fig. 13c).<sup>161,162</sup> Choi and Jung developed trianilides with different *D*-alanine and/or glycine units (**53**, **54**) (Fig. 13d).<sup>163</sup> Interestingly, those containing one (**53**) or two (**54**) alanine segments can self-assemble into *M* or *P* supramolecular helical structures in toluene by playing with temperature. Thus, while an *M* supramolecular helix is obtained at low temperature (15 °C), an evolution to a supramolecular helix with opposite *P* chirality is observed by heating, reaching the



**Fig. 13** Water effects in the thermal driven helix inversion of supramolecular aggregates using (a) tetracarboxiamides derivatives (**48**, **49**) (reproduced from ref. 162 with permission from the American Chemical Society, copyright 2020), (b) triarylamines (**50**, **51**) and (c) chiral benzene tricarboxiamide (**52**). (d) Chemical structures of thermoresponsive chiral 1,3,5-trianilides (**53**, **54**).

maximum ECD spectra at *ca.* 45 °C. This helix inversion is attributed to conformational changes of the amide groups at the glycine residues attached to the trianilide core.

Meijer and George reported another interesting example showing how the solvent–aggregate interaction affects the axial *P/M* chirality of the helical aggregate.<sup>164</sup> They designed a coronene bisimide that has a 3,5-dialkoxy substitution in the imide phenyl groups (**55**) (Fig. 14a). This molecule self-assembles in methylcyclohexane (MCH), generating “molecular pockets” in the assembly. Solvent molecules can intercalate or form clathrates within the molecular pockets of the aggregate at low temperature (−10 °C), inducing an *M* helical structure. Interestingly, this weak solvent/aggregate interaction can be disrupted at 10 °C, resulting in *M* to *P* helix inversion.

Likewise, Percec showed that dendron-based C<sub>3</sub>-symmetric self-assembling dendrimers connected at their apex *via* trisesters and trisamides of 1,3,5-benzenetricarboxylic show thermal helical inversion due to the change of the lattice symmetry.<sup>165</sup>

Temperature can be used to discriminate between kinetic and thermodynamic aggregates that possess opposite chirality. Thus, Freire found that an asymmetric chiral OPE (**13**, Fig. 6a)<sup>118</sup> can self-assemble into *P* or *M* supramolecular aggregates through a competitive aggregation pathway controlled by thermal treatment. Thus, while a kinetic *P* helix is obtained at room temperature in a

DCM/MCH mixture, the thermodynamic *M* helix is formed at 0 °C after a heating/cooling cycle. Interestingly, the morphologies of these aggregates are different, thus while the kinetic *P* helix is formed by short oligomers that produced twisted sheets, the thermodynamic *M* helix forms columnar helical aggregates.

Dou and Feng found that a similar effect is obtained during the co-assembly of the two components of the building block.<sup>166</sup> Thus, C<sub>2</sub> phenylalanine-based aggregates (**L-PF**, **56**) (Fig. 14b) can be transformed into kinetically trapped architectures with opposite helicity through a kinetic co-assembly pathway. In this case, a *P* helix was formed by heating/cooling or anti-solvent assembly of the L-phenylalanine derivative. Interestingly, the addition of an achiral pyridine derivative (**57**) (Fig. 14b), which can establish supramolecular interactions with the L-PF derivative, generates achiral aggregates after thermal treatment. For its part, *M* chiral aggregates are obtained using the anti-solvent assembly protocol. The morphology of the aggregates depends also on the pathway followed by the self-assembly. In this sense, Carretero found that a quinquethiophene–rhodanine derivative (**58**) (Fig. 14c) self-assembles through hydrogen bonding interactions into opposite chiral helical structures before and after a heating/cooling cycle in pure toluene or in a 20/1 toluene/chloroform mixture.<sup>167</sup>

Supramolecular chemistry can also be used to induce chirality in an achiral polymer by interacting with a chiral molecule. Thus, Chen, Quan and Zheng created helical nanofibers by the co-assembly of achiral pyrenyl based polymer (**59**) and chiral binaphthyl based inducers (**60**) (Fig. 14d) through π–π interactions. The co-assemblies show chiroptical properties due to the formation of helical fibers which can be tuned by thermal annealing towards *P* or *M* helical senses. The emissive properties of the pyrenyl moiety allows to create a thermal CPL switch.<sup>168</sup>

## 2.2 External stimuli: anions

Negatively charged ions can be used as external stimuli to promote helix inversion of foldamers, dynamic covalent helical polymers and supramolecular polymers. Anions can establish supramolecular interactions with different functional groups, disrupting preexisting interactions or generating new ones that result in an inversion of the helical structure.

**2.2.1 Anion-responsive foldamers.** In this section, we highlight recent advances that have been made in anion-responsive foldamers whose morphology and function are modified in the presence of such stimuli. Due to their relevance, several examples of anion foldamers can be found in the literature, so the reader can find excellent reviews focused solely on these systems.<sup>15,169</sup> However, we will focus this section on the foldamer/anion binding that leads to a modification in the elongation or sense of a preformed helical scaffold. In this sense, non-covalent interactions are the primary contact with the anion and therefore, the solvent used plays an important role, favouring supramolecular foldamer/anion interactions in non-polar environments over polar ones. Jeong reported anion-induced folding of an oligoindole–ethynylene skeleton (**60**) (Fig. 15a) through indole–NH hydrogen bonds with Cl<sup>−</sup> delivered as TBACl (tetrabutylammonium chloride).<sup>170</sup> This group developed also chiral oligoindole-based

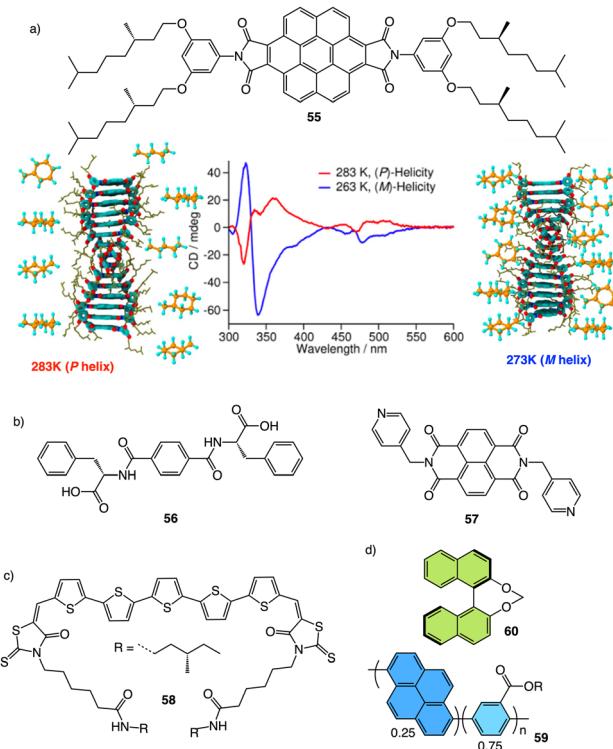


Fig. 14 (a) “Clathrate effect” in the thermal behaviour of a coronene bisimide derivative (**55**) (reproduced from ref. 164 with permission from the American Chemical Society, copyright 2017). Chemical structures of (b) a two-component building block (**56**, **57**) and (c) a quinquethiophene–rhodanine derivative (**58**) that generates chiral thermoresponsive aggregates. (d) Chiroptical thermal co-assembly based on achiral pyrenyl based polymer (**59**) and chiral binaphthyl based inducers (**60**).

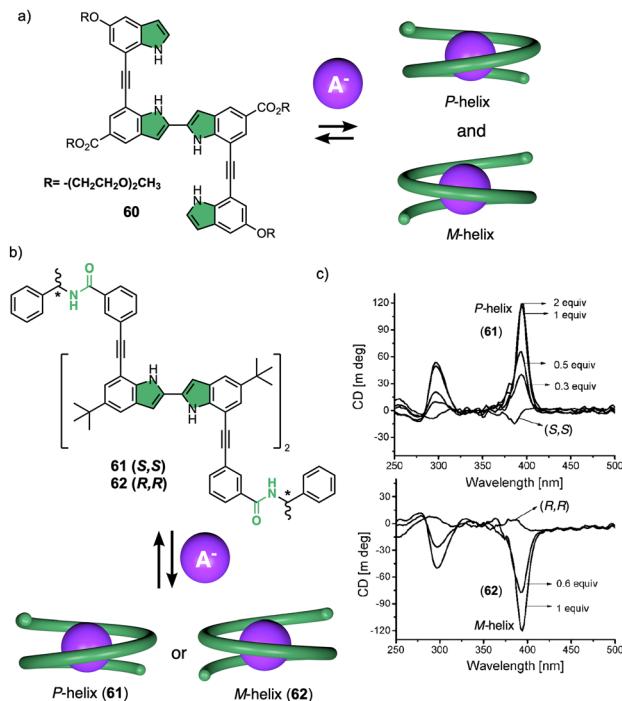


Fig. 15 (a) Chemical structure of an oligoindole–ethynylene foldamer (**60**) and schematic illustration of its anion folding. (b) Chemical structure and (c) ECD spectra of an oligoindole based foldamer bearing (S)- (**61**) and (R)-phenylethylamido (**62**) groups as chiral termini (reproduced from ref. 171 with permission from the American Chemical Society, copyright 2008).

foldamers that bear (1*S*)- and (1*R*)-phenylethylamido groups as chiral termini (**61**, **62**) (Fig. 15b). These foldamers show a screw sense excess in solution that depends on the chirality of the side chain and that can be enhanced by interaction with Cl<sup>-</sup> anions (Fig. 15c).<sup>171</sup> Thus, the chirality has been effectively transferred in the folding process giving rise to a specific helical sense.

However, in these works the helical scaffolds cannot be modulated from *P* to *M* or *vice versa*. So, the researchers took a step forward and reported the first example of a switchable anion-responsive foldamer based on a chiral indocarbazole dimer that folds into a helical conformation by intramolecular hydrogen bonding (**63**) (Fig. 16a).<sup>172</sup> The helical sense of this dimer can be reversibly switched by binding it to a sulphate anion in CH<sub>2</sub>Cl<sub>2</sub>. Thus, the anion disrupts the intramolecular hydrogen bonds of the foldamer that stabilize the helical structure commanded by the chiral groups at the terminal ends. This leads to a different helical scaffold with opposite screw sense due to the formation of a new cavity with the NHs oriented towards the interior where the anion is located (Fig. 16b). In another example, they used a three-indolocarbazole-ethynylene trimer bearing a terminal amide-linked (S)-arylethylamido group at both ends (**64**).<sup>173</sup> In this system, they found that a helix inversion can be achieved by using anions of appropriate size (Cl<sup>-</sup>, Br<sup>-</sup>, or CH<sub>3</sub>COO<sup>-</sup>) that fit in the foldamer cavity.

Recently, Talukdar and coworkers reported a selective chloride anion responsive foldamer in which a double helical structure suffers a conformational change (**65**) (Fig. 17a).<sup>174</sup> The foldamer-anion interaction causes the unwinding of the double helical

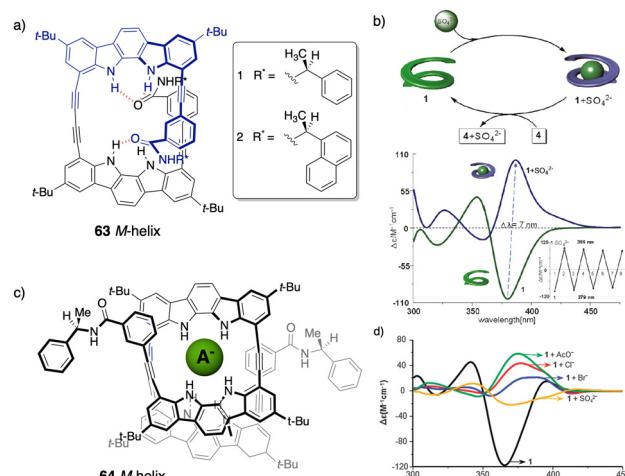


Fig. 16 (a) Chemical structure of the foldamer (**63**). (b) Schematic representation and ECD spectra of the reversible helix inversion of **63** upon addition and removal of sulphate ions (reproduced from ref. 172 with permission from the American Chemical Society, copyright 2011). (c) Molecular structure of a complex between **64** and different anions. (d) ECD spectra of **64** in the absence (black) and in the presence of different tetrabutylammonium (TBA) anion salts (reproduced from ref. 173 with permission from the Royal Society of Chemistry, copyright 2011).

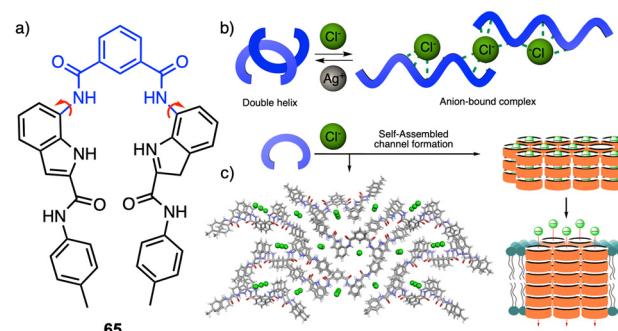


Fig. 17 (a) Chemical structure of **65** that folds into a double helix. (b) Schematic illustration of conformational changes from double helix to anion-coordinated supramolecular assembly. (c) Anion-induced supramolecular channel (reproduced from ref. 174 with permission from Springer Nature, copyright 2022).

structure to form an anion-coordinated supramolecular polymeric channel, and where the double helical structure can be recovered by adding Ag<sup>+</sup> salts (Fig. 17b), demonstrating the reversibility of the process. Moreover, the formation of the anion-induced supramolecular ion channel results in efficient ion transport across lipid bilayer membranes with excellent chloride selectivity (Fig. 17c). This work demonstrates that anion–cation-assisted stimulus responsive unwinding and rewinding of artificial double-helix systems paves the way for developing smart materials with potential biomedical applications.

In general, most of the reported anion-responsive foldamers are not selective for a single anion. They usually bind to different families of anions, *e.g.*, halides (Cl<sup>-</sup>, Br<sup>-</sup>, and I<sup>-</sup>) and oxoanions (NO<sub>2</sub><sup>-</sup>, H<sub>2</sub>PO<sub>4</sub><sup>-</sup>, HSO<sub>4</sub><sup>-</sup> and CH<sub>3</sub>COO<sup>-</sup>).<sup>175,176</sup>

However, selectivity is improving in recent years as the reversal mechanisms become clearer. In this regard, a selective oxoanion foldamer-switch was recently reported by Clayden,<sup>177</sup> who developed axially racemic oligourea foldamers [66-(a-g)] (Fig. 18a) made from mesocyclohexane-1,2-diamine residues that can adopt a preferred screw-sense, differentiating their termini groups by the different characteristics of their hydrogen bonds.

Thus, a dynamic equilibrium between two alternative screw sense conformers is set up whose relative population is determined by the competing hydrogen-bonding properties of the terminal groups, dictating the global hydrogen-bond directionality of the foldamer. In this case, these preferences are relatively insensitive to solvent effects and to neutral hydrogen-bond donors or acceptors. However, addition of a geometrically compatible anionic binding partner (acetate or phosphate) can induce a global refolding of the oligomer. This foldamer/anion interaction causes the exposure of previously embedded hydrogen-bond donors, resulting in the inversion of the conformational preference. For example, upon addition of achiral anion phosphate delivered from a tetrabutylammonium salt to a solution containing foldamer 66-b, an inversion of the CD spectra is observed. This suggests an inversion of the foldamer screw-sense (Fig. 18b) induced by anion binding to the thiourea-terminated foldamer that leads to a conformational change creating a switch in the hydrogen-bond directionality to favour the bind between the thiourea and the phosphate.

Helix inversion can also be achieved from metallo-foldamers when they interact with anions. Thus, Miyake created a peptide

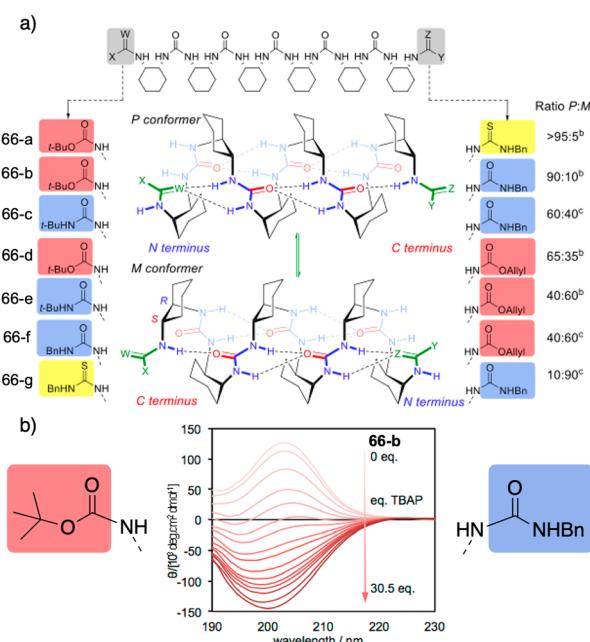


Fig. 18 (a) 2.5<sub>12/14</sub>-helical oligourea foldamers as torsion balances for hydrogen-bonding strength. (b) Screw-sense reversal and hydrogen-bond-directionality reversal upon addition of tetrabutylammonium phosphate (TBAP) (0.2 mM in MeCN). The end groups of the foldamer used in the titration are displayed on either side of the CD spectra. Reproduced from ref. 177 with permission from the American Chemical Society, copyright 2018.

foldamer based on Inai's design<sup>82-85</sup> that contains, in the middle of the sequence, a chiral *N,N'*-ethylenebis[N-methyl-(S)-alanine] connected to the N terminus of two H-(Aib- $\Delta$ Phe)<sub>2</sub>-Aib-OCH<sub>3</sub> pentamers through a peptide bond (67).<sup>178</sup> Addition of a hexa-coordinated metal center [Co(II), Zn(II), or Ni(II), delivered as M(ClO<sub>4</sub>)<sub>2</sub> salts] to a foldamer solution, forms a  $\Lambda$ - $\Delta$  complex with the *N,N'*-ethylenebis[N-methyl-(S)-alanine] of the foldamer that induces a *P* helix. Additional addition of NO<sub>3</sub><sup>-</sup> anions (from TBANO<sub>3</sub>) produces a helix inversion from *P* to *M* due to a  $\Lambda$ - to  $\Delta$ -form change in the metal complex, resulting in a chiral transfer from the metal center to the pentapeptide chains (Fig. 19a). Interestingly, analogous *P* to *M* helix inversion process was observed for Zn(II), Co(II) and Ni(II)-based complexes, although with a different time scale, which is based on the substitution lability of the metal center. Thus, the present metallo-peptide complex offers time-tunable peptide helix inversion that has a time scale from milliseconds to hours, as revealed by the stopped flow CD measurements (Fig. 19b).

**2.2.2 Anion-responsive supramolecular polymers.** Anions-responsive supramolecular helical polymers that result in a structural change in the supramolecular helix (sense, elongation) are a family of supramolecular helices with few examples in the literature. This fact is probably due to the ability of anions to disrupt the supramolecular forces that keep monomers stacked, such as hydrogen bonds, leading to a molecularly dissolved state. One of these examples was designed by Lee who reported an anion-directed self-assembly of a coordination polymer whose secondary structure is tunable.<sup>179</sup> To do that, a

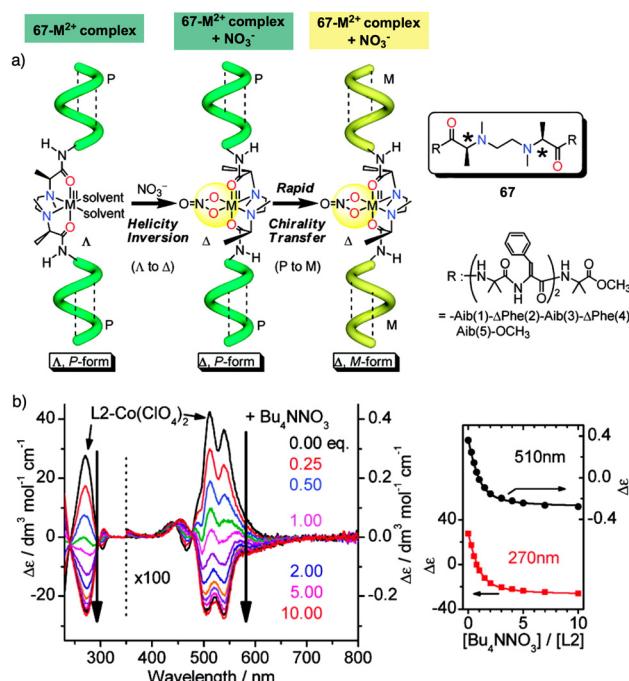


Fig. 19 (a) Anion induced helix inversion of metallo-foldamer 67-M<sup>2+</sup> (M<sup>2+</sup> = Co, Zn, Ni). (b) CD spectra changes of the 67-Co(ClO<sub>4</sub>)<sub>2</sub> complex upon addition of Bu<sub>4</sub>NNO<sub>3</sub> and titration profiles of the CD wavelength at 270 and 510 nm. Reproduced from ref. 178 with permission from the American Chemical Society, copyright 2008.

bent-shaped bipyridine ligand containing a dendritic aliphatic side chain (**68**) was synthesized and complexed with silver ions through a self-assembling process (Fig. 20a). The resulting complexes showed a self-assemble into ordered structures that differ as a function of the counterion size in the solid state.

For instance, fibers obtained by combining **68** with silver nitrate ( $\text{AgNO}_3$ ) or silver tetrafluoroborate ( $\text{AgBF}_4^-$ )—small anions—self-assemble into helical chains that are organized in a 2D hexagonal lattice (Fig. 20b). However, when a silver triflate salt is used [ $\text{Ag}(\text{CF}_3\text{SO}_3^-)$ ], dimeric cycles are formed that are stacked on top of each other, giving rise to columns that are assembled laterally in a hexagonal fashion (Fig. 20b). This assembly behavior changes dramatically when silver salts containing large anions are used. For instance, metallo-supramolecular polymer chains based on silver salts as heptafluorobutyrate ( $\text{CF}_3\text{CF}_2\text{CF}_2\text{CO}_2^-$ ) are organized in a lamellar structure (Fig. 20b). Therefore, these results demonstrate how a variation in the size of the counteranion can regulate the secondary structure of the coordination polymer chain, from helical, cyclic, to unfolded linear chain conformations in the solid state. This unique self-assembly behavior can be explained by considering the size of the counteranion and consequent chain conformation to maximize the electrostatic interactions.

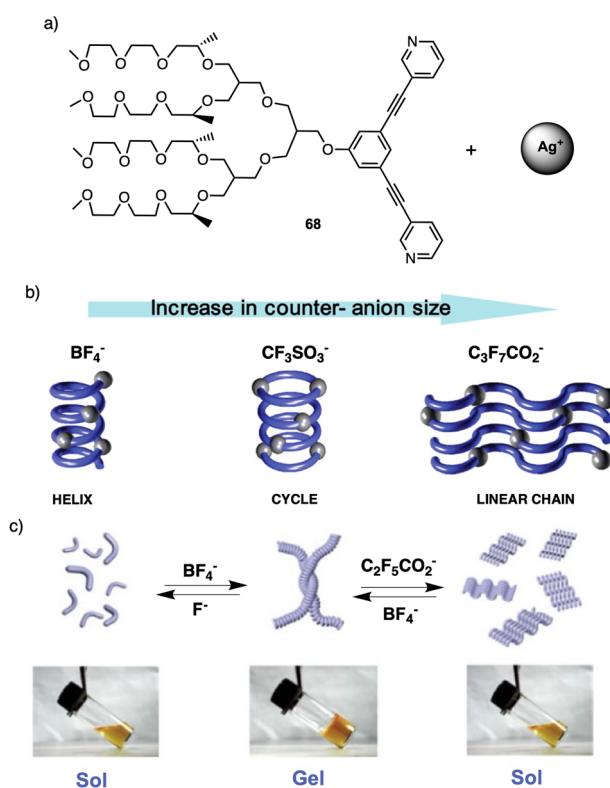


Fig. 20 (a) Chemical structure of **68**. (b) Schematic representation of the different morphologies adopted by **68**– $\text{Ag}^+$  coordination polymer induced by the anion size. Reproduced from ref. 179 with permission from the American Chemical Society, copyright 2004. (c) Schematic representation of the reversible conversion between folded and unfolded conformations of the **68**– $\text{Ag}^+$  coordination chain upon counteranion exchange Reproduced from ref. 180 with permission from John Wiley and Sons, copyright 2018.

This transition was attributed to the strong attraction between fluoride ( $\text{F}^-$ ) and silver ions leading to depolymerization of the helical polymer into individual molecules. In the case of  $\text{C}_2\text{F}_5\text{CO}_2^-$  Later, Lee found that anion exchange in the coordination polymer **68**– $\text{Ag}^+$  led to a reversible sol-gel transition in water.<sup>179</sup> This sol-gel effect occurs due to a change in the secondary structure of the coordination polymer, from a folded helical conformation to an unfolded zigzag conformation. Thus, in the presence of  $\text{BF}_4^-$  anions, **68**– $\text{Ag}^+$  results in a gel that is liquified when fluoride ( $\text{F}^-$ ) or pentafluoropropionate ( $\text{C}_2\text{F}_5\text{CO}_2^-$ ) are present (Fig. 20c), the gel liquefaction occurs due to the large size of this counterion compared to  $\text{BF}_4^-$  (Fig. 20c).<sup>180</sup>

More recently, Muranaka reported an anion-responsive  $\pi$ -conjugated supramolecular assembly based on the chiral anion receptor **69** (Fig. 21a).<sup>181</sup> A tripropyl-substituted triazatriangulenium planar cation (TATA $^+$ ) (Fig. 21a) forms an electrostatic interaction with anion receptor **69**, which traps the  $\text{Cl}^-$  anion delivered from the TATA-Cl salt. This electrostatic interaction results in a layer-by-layer chiral aggregate consisting of planar units with opposite charges (Fig. 21b). Aggregation studies of **69** in the absence and presence of TATA-Cl produce ECD and CPL spectra with opposite signs (Fig. 21c and d). Since the assembly modes depend on the ion-free and ion-pairing states of emissive  $\pi$ -conjugated molecules, the chiroptical properties change accordingly and thus result in readily tuneable CPL-active materials.

**2.2.3 Anion-responsive dynamic covalent polymers.** Dynamic helical polymers, such as PPAs, have been used to sense anions due to their stimuli-responsive properties. Thus, Kakuchi designed different achiral pendant groups that bear functional groups able to interact with anions such as amides,<sup>182</sup> ureas,<sup>183,184</sup> or

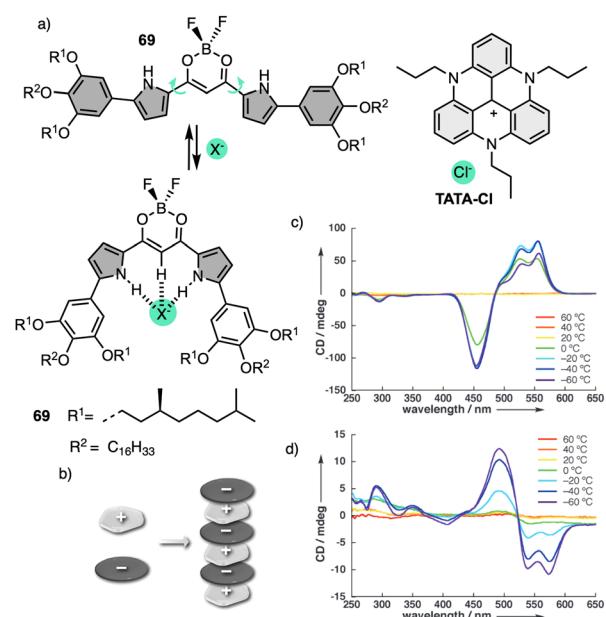


Fig. 21 (a) Chemical structure and anion-binding modes of anion receptors **69**. (b) Schematic model of charge-by-charge assembly. VT CD spectra of **69** in (c) absence  $\text{Cl}^-$  and (d) presence of  $\text{Cl}^-$  added as a TATA salt (1 equiv.). Reproduced from ref. 181 with permission from John Wiley and Sons, copyright 2013.

sulfonamides.<sup>43,185</sup> These functional groups show high affinity towards anions due to the acidity of their protons. The PPA/anion interaction produces an elongation of the polymer due to the size and/or basicity of the anion, which is usually accompanied by a yellow to red colour change. The accommodation of the anions within the macromolecular helical structure results in an elongation of the helical scaffold.

In elongated PPA structures, the conjugation between alternating double bonds increases producing a red shift of the polyene band in the UV-vis spectra that is accompanied by a change in colour of the solution. Hua and Tian also reported another PPA that bears a naphthalimide as pendant connected to the PPA through an amide bond.<sup>186</sup> This polymer works in a similar way to the previous ones, acting as a colorimetric and fluorescent chemosensor for the fluoride anion.

To create a selective anion detector based on a dynamic helical polymer, such as a PPA, the design of the monomer is crucial, as with many other stimuli. For instance, Kakuchi designed a chiral PPA that bears, as a pendant, an amino acid connected to the backbone through a urea group —L-leucine, L-glutamic acid, L-aspartic acid, L-phenylalanine, L-isoleucine, and L-alanine; PPA-Leu, PPA-Glu, PPA-Asp, PPA-Phe, PPA-Ile, and PPA-Ala, respectively.<sup>183,187</sup> Thus, while the chiral amino acid is used to induce a specific helical sense in the PPA, the acidic urea groups are used to interact with anions. The anion/chiral pendant interaction causes a conformational change in the monomer repeating unit, which is harvested by the polyene backbone producing an elongation and/or a helical sense change. All these polymers showed modulable helicities by external stimuli. Interestingly, the addition of various anions as ammonium salts, such as tetra-*n*-butylammonium acetate (TBAA), benzoate (TBAB), nitrate (TBAN), azide (TBAN3), fluoride (TBAF), chloride (TBACl) and bromide (TBABr), to solutions containing the different polymers showed different ECD responses depending on polymer and anion (Fig. 22a for PPA-Leu). Therefore, the anionic signalling property of these polymers depends on both the size of the anion and the amino acid selected as the pendant group. For instance, while the addition of  $\text{CH}_3\text{COO}^-$  to a solution of PPA-Leu, PPA-Ala, and PPA-Ile induces a *P* helix in the polymer ( $\text{ECD}_{500} > 0$ ), an *M* helix is induced in PPA-Phe, PPA-Asp, and PPA-Glu ( $\text{ECD}_{500} < 0$ ) (Fig. 22b). Similar effects were observed with other anions, such as  $\text{C}_6\text{H}_5\text{COO}^-$ ,  $\text{F}^-$ ,  $\text{Cl}^-$ , and  $\text{Br}^-$ , indicating that these polymers can be used for distinguishing anionic hosts due to the resulting unique patterns. However, the mechanisms of helix modulation have not been fully described.

Working in the same direction, Kakuchi subsequently reported analogous studies but using a different group for anionic binding, *i.e.*, sulfonamide group.<sup>188</sup> During the PPA/anion interaction studies, the behavior of sulfonamide-PPAs is coincident with that of urea-PPAs described above, confirming that the amino acid side chain plays a relevant role in the anion-PPA interaction.

On the other hand, PPAs can be used as chiroptical sensors to detect anions. Freire reported a PPA (poly-(*R*)-11) that bears an acidic anilide group as pendant (Fig. 23).<sup>189</sup> This polymer behaves as axially racemic (mixture of *P* and *M* helices) due to

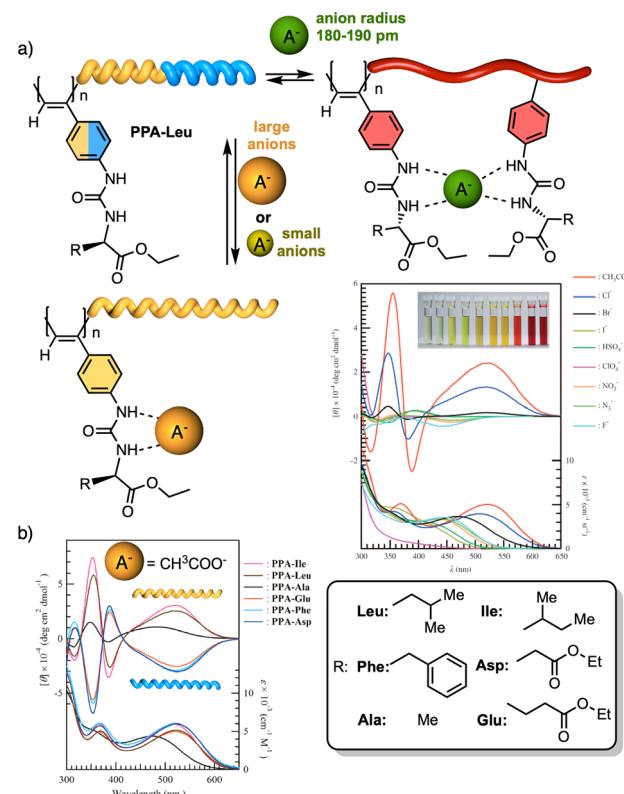


Fig. 22 (a) Schematic representation of size-selective detection for anions based on a PPA. ECD (upper) and UV/Vis absorption (lower) spectra of poly-PPA-Leu upon addition of several anions (1 equiv. of anion to a  $3.3 \text{ mmol L}^{-1}$  THF solution of poly-PPA-Leu) (reproduced from ref. 183 with permission from John Wiley and Sons, copyright 2008). (b) CD (upper) and UV-vis (lower) spectra of urea-functionalized polymers with  $\text{CH}_3\text{COO}^-$ . Reproduced from ref. 187 with permission from the American Chemical Society, copyright 2004.

the presence, in the pendant, of two conformers in equilibrium that have the carbonyl and methoxy groups  $[(\text{O}-\text{C})-(\text{C}=\text{O})]$  in *antiperiplanar* (*ap*) or *synperiplanar* (*sp*) orientations. However, upon interaction with different TBAA salts, the equilibrium can shift toward the *M* or *P* helix due to a supramolecular interaction between the anilide group and the anion. Thus, while  $\text{AcO}^-$ ,  $\text{BzO}^-$  and  $\text{N}_3^-$  stabilize the *sp* conformer in the pendant group of poly-(*R*)-11 and induce an *M*-helix,  $\text{F}^-$  and  $\text{CN}^-$  stabilize the *ap* conformer inducing a *P*-helix in the PPA (Fig. 23).

Interestingly, upon adding larger amounts of TBAA salts, a colorimetric shift from yellow to red is observed in the polymer solution (Fig. 23). This colour change is produced by the deprotonation of the anilide group, and the induced elongated *P* helix depends only on the accommodation of the deprotonated anilide(-)/TBA(+) ion pair within the helix, a phenomenon independent of the anion used to deprotonate the anilide group.

Recently, Gao designed an optically active helical polyacetylene carrying the amino acid (*L*)-alanine as pendant group, connected to the PA backbone through the C-terminus and functionalized at the N-terminus with a naphthalene diimide (NDIS) derivative (poly-70) (Fig. 24).<sup>190</sup> In this polymer, the PA/anion interaction is used to regulate the stacking modes



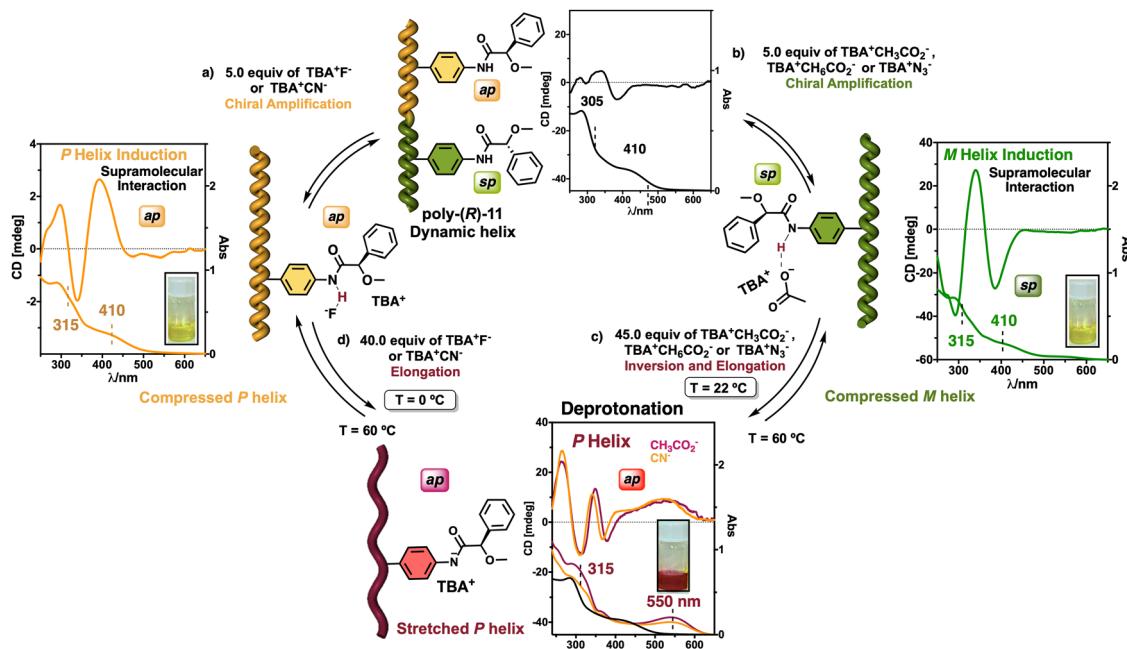


Fig. 23 (a) *P* and (b) *M* helix induction in poly-(*R*)-11 by addition of 5.0 equiv. of TBAA (A = fluoride or cyanide) or (A = acetate, benzoate or azide) respectively. Elongation and *P* helix induction in poly-(*R*)-11 by addition of (c) 45.0 equiv. of TBAA (A = acetate, benzoate or azide) at rt or (d) 40.0 equiv. of TBAA (A = fluoride or cyanide) at 0 °C. Reproduced from ref. 189 with permission from Elsevier, copyright 2021.

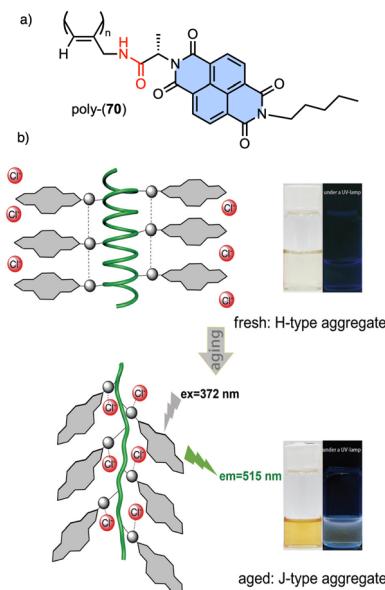


Fig. 24 (a) Chemical structure of poly-70. (b) Schematic representation of time-dependent H to J structural changes in poly-70 by addition of Cl<sup>-</sup> and the corresponding fluorescence effects. Reproduced from ref. 190 with permission from the Royal Society of Chemistry, copyright 2021.

of  $\pi$ -conjugated NDIs building blocks. Thus, anions such as Cl<sup>-</sup> induce a dynamic reorganization of the molecular arrangement of NDIs within the helical scaffold, causing a transition from H-type to J-type aggregates, which activates the fluorescence emission at 515 nm and disrupts the induced helical structure of the polymer backbone.

Xie designed an asymmetrically 2,4-disubstituted PPA bearing a benzamide of (L)-alanine ethyl ester and a methoxycarbonyl group as chiral and achiral substituents respectively (poly-71) (Fig. 25). This polymer adopts a contracted helix stabilized by intramolecular hydrogen bonding networks among neighbouring pendants amides.<sup>191</sup> This helical scaffold possesses

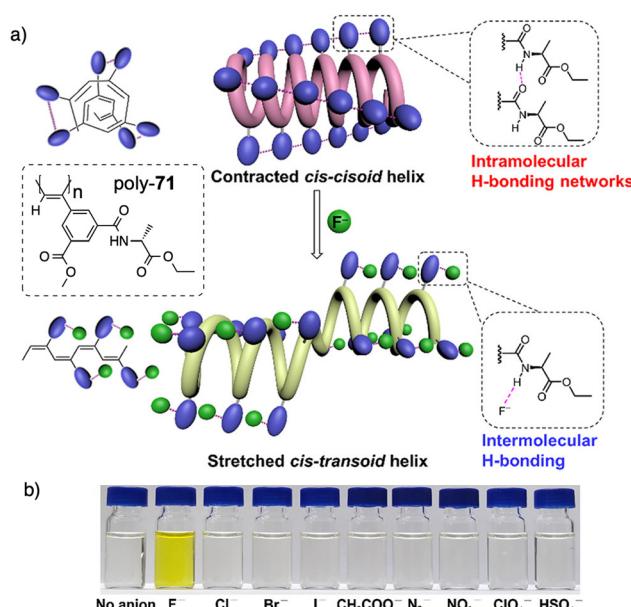


Fig. 25 (a) Schematic mechanism of specific and (b) colorimetric fluoride ion recognition by poly-71. Reproduced from ref. 191 with permission from the American Chemical Society, copyright 2022.

small pockets suitable for accommodating the smallest anion,  $\text{F}^-$ , which has a high basicity, excluding other anions. During the poly-71/ $\text{F}^-$  interaction, the contracted *cis-cisoidal* backbone of poly-71 transforms into a stretched *cis-transoidal* exhibiting a nonpreferred-hand-sense helix, facilitating subsequent binding of  $\text{F}^-$  anions. Such unzipping effect gives rise to an ultralow detection limit for  $\text{F}^-$  ( $2.85 \times 10^{-8}$  M or 0.54 ppb). Meanwhile, the colour of the solution undergoes a change from colourless to yellow due to the conjugated *cis-transoidal* conformation. As a result, the study achieves a remarkable naked-eye recognition of fluoride ions ( $\text{F}^-$ ) with high specificity and sensitivity.<sup>192</sup>

### 2.3 External stimuli: cations

Metal ions can coordinate to many functional groups inducing conformational changes, folding or aggregation of molecules or macromolecules. This ability, in combination with the toxicity properties of metals, makes helical polymers responsive to metal ion stimuli a very attractive field.

**2.3.1 Cation-responsive foldamers.** The coordination of metal ions with molecules or macromolecules can lead to the formation of artificial helical molecules, such as metallofoldamers or helicates.<sup>193–195</sup> Akine and Nabeshima designed a helical lanthanide metal complex<sup>196</sup> equipped with a transducer mechanism based on the (*S,S*)-*trans*-1,2-disubstituted ethylenediamine unit (Fig. 26a), whose helical handedness can be inverted by taking advantage of the host-guest interaction between the diammonium guests and 18-crown-6 substituents (72, Fig. 26b).<sup>197</sup> Thus, short alkanediammonium guests— $\text{H}_3\text{N}^+(\text{CH}_2)_n\text{NH}_3^+$  ( $n = 4, 6, 8$ )—induce a *P* screw sense in the foldamer, while longer guest such as  $\text{H}_3\text{N}^+(\text{CH}_2)_{12}\text{NH}_3^+$  promote *M* (Fig. 26c).

In another example, the authors use metal exchange of helical complexes to induce a helix inversion.<sup>198,199</sup> In this case, the ligand **H<sub>6</sub>73** (Fig. 26d) underwent a four-step conversion (**H<sub>6</sub>73**  $\rightarrow$  73Zn<sub>3</sub>  $\rightarrow$  73Zn<sub>5</sub>  $\rightarrow$  73Zn<sub>3</sub>Ba  $\rightarrow$  73Zn<sub>3</sub>La) upon sequential metal addition ( $\text{Zn}^{2+}$ ,  $\text{Ba}^{2+}$ , then  $\text{La}^{3+}$ ). Associated with the conversion, three-step helicity inversion took place (73Zn<sub>3</sub>, *P* helix  $\rightarrow$  73Zn<sub>5</sub>, *M* helix  $\rightarrow$  73Zn<sub>3</sub>Ba, *P* helix  $\rightarrow$  73Zn<sub>3</sub>La, *M* helix) (Fig. 26e).

Tanaka prepare a discrete chiral metallofoldamer composed of homochiral metallosalen complexes  $[(\text{74}_{RR}\text{-Ni})_2\text{Pd}]$  (Fig. 27a).<sup>200</sup> Connecting chiral Ni-salen moieties to the side chains with a transition metal center like Pd(II), at proper coordination bond angles, would lead to a helical organization of the columnar assembly. The metallosalen exhibited thermotropic columnar liquid crystalline properties and formed hexagonal columnar phases, which exhibited high thermal stability and showed orientation along the extrusion direction (Fig. 27b).

Another interesting example is the foldamer based on triazole-linked phenanthroline ligands (75–76) (Fig. 27c) designed by Zhu and Liu.<sup>201</sup> In the extended solid-state, these ligands formed oligomeric channels through  $\pi$ – $\pi$  interactions, that can trap small guest molecules (Fig. 27d). Upon complexation with copper ions (Fig. 27e), the foldamers intertwine to form a double-stranded helix consisting of two foldamers coordinated with four Cu<sup>+</sup> ions, resulting in a metallofoldamers (Fig. 27e and f). This structural change is reversible, and it is possible to interconvert the hollow channels observed in the extended solid-state

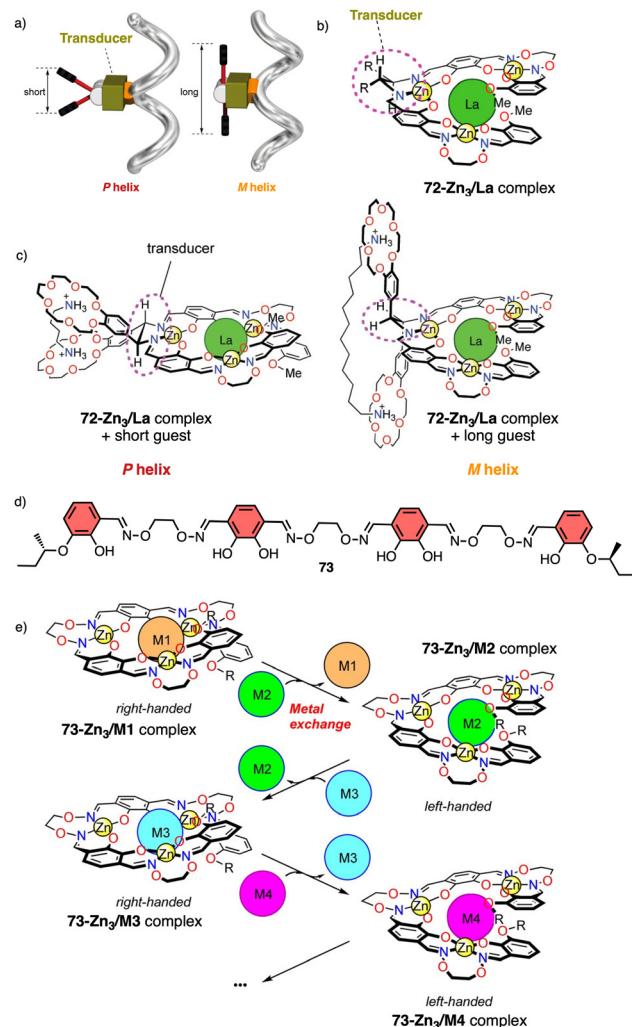


Fig. 26 (a) Schematic representation of helix inversion based on a transducer mechanism. (b) chemical structure for 72-Zn<sub>3</sub>/La complex. (c) Helix induction of 72-Zn<sub>3</sub>/La complex by interaction with diamines (reproduced from ref. 197 with permission from the American Chemical Society, copyright 2011). (d) Chemical structure of ligand 73. (e) Schematic representation of helix inversion via metal exchange of helical complexes (reproduced from ref. 198 with permission from the American Chemical Society, copyright 2013).

structure of the ligands into the “closed” structure of the copper complex (Fig. 27f) and *vice versa* ( $\text{Cs}^+$ ,  $\text{Rb}^+$ ,  $\text{K}^+$ ,  $\text{Na}^+$ ,  $\text{Li}^+$ ). By adding copper ions, ion transport is inhibited due to the structural change in the foldamer metal complex, which can be recovered once the Cu<sup>+</sup> ions are removed (Fig. 27g).

Maayan demonstrated the positive allosteric cooperativity ability of a peptoid that incorporates four 8-hydroxyquinoline (HQ) ligands at fixed positions to create two distinct metal-binding sites (77) (Fig. 28).<sup>202</sup> Peptoid 77 can bind a copper ion (Cu<sup>2+</sup>) in one site that leads a conformational change that in turn facilitates the coordination of a metal ion—Zn<sup>2+</sup>, Co<sup>2+</sup>—to the second metal binding site, demonstrating a positive allosteric cooperativity in peptidomimetics (Fig. 28). This finding was confirmed by competition experiments using a foldamer (78) that did not exhibit an increase in order upon complexation with Cu<sup>2+</sup> (78Cu) (Fig. 28).



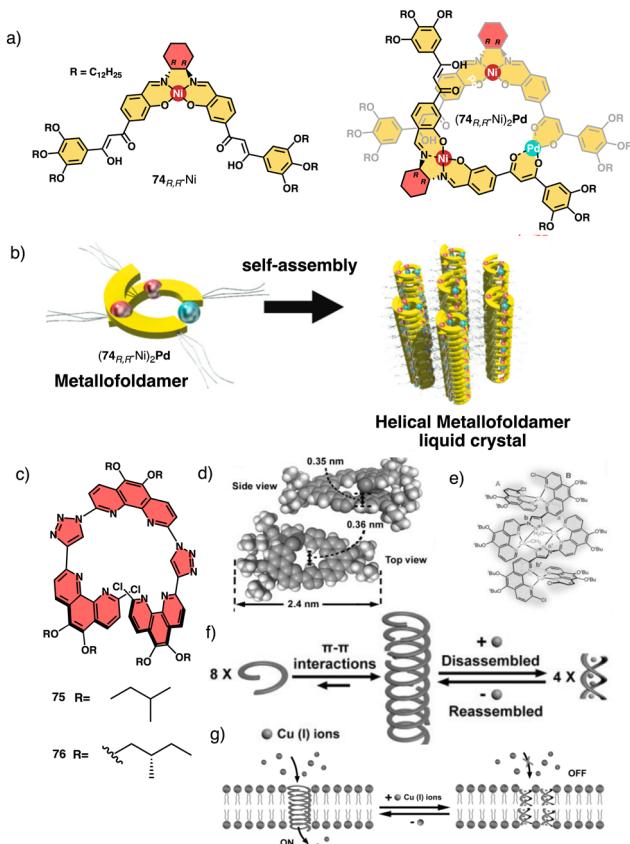


Fig. 27 (a) Chemical structure for  $74_{RR}$ -Ni and  $(74_{RR}-Ni)_2Pd$ . (b) Helically assembled LC metallofoldamer  $(74_{RR}-Ni)_2Pd$  (reproduced from ref. 200 with permission from the Royal Society of Chemistry, copyright 2022). (c) Molecular structures of **75** and **76**. (d) Crystal structure of **75**. (e) Molecular structure of  $75_2$ - $[\text{Cu}(\text{CH}_3\text{CN})_4\text{PF}_6]_4$ . (f) schematic representation of the tentative formation, disassembly and reassembly of the 1D hollow helical tubes from **75**. (g) Schematic representation of the reversible ligand-gated ion channel formed by **75** triggered by  $\text{Cu}^+$  and  $\text{NH}_3\text{-H}_2\text{O}$  reproduced from ref. 201 with permission from John Wiley and Sons, copyright 2020.

Jeong demonstrated the interplay between temperature and metal ions as stimuli in a double-stranded dinuclear helicate.<sup>128</sup> Through self-assembly, the study combined an aromatic helical foldamer **18** with dichloropalladium(II) (Fig. 7), yielding a helicate that undergoes *syn-anti* and *syn-syn* conformational switching in response to temperature changes.<sup>128</sup> In the field of aromatic oligoamides foldamers, Huc introduced, within a foldamer sequence, a central diacid residue known as pyridazine-pyridine-pyridazine (pyz-pyr-pyz) (Fig. 29a).<sup>203,204</sup> The resulting aromatic helical foldamers **79** and **80** have a central residue that can effectively trap metal ions within their helix-shaped cavities (Fig. 29a). In the absence of metal ions, the central pyz-pyr-pyz residue is not folded. However, by addition of alkali or alkaline earth metal ions, the foldamer folds forming a helical structure. Interestingly, the metal ion coordination sphere remains accessible for guest recognition, where cooperative interactions with the helix host strengthen the binding of the guests (see Fig. 29b). Moreover, depending on the metal ion used, the metal-foldamer binding may involve first or second coordination spheres of the metal hydrates (Fig. 29c).

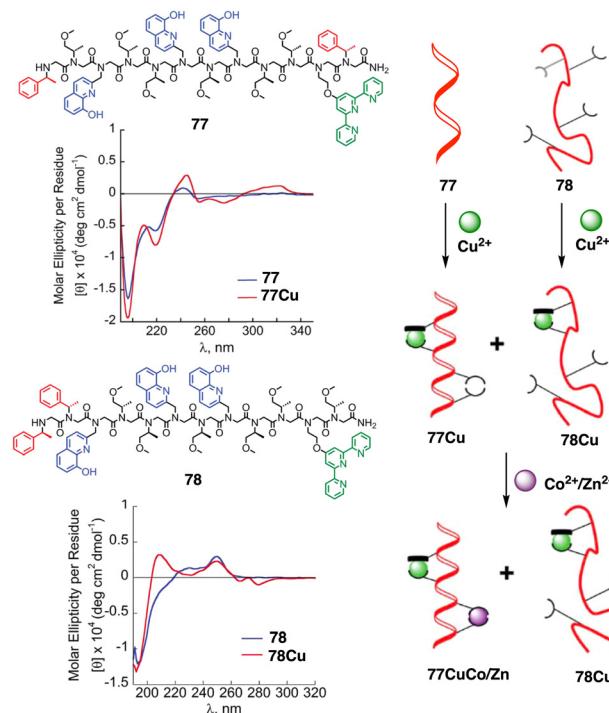


Fig. 28 Chemical sequences of peptoid oligomers **77** and **78**. CD spectra of peptoid **77** with **77Cu** complex and **78** with **78Cu**. A cartoon shows the positive allosteric cooperative binding of **77Cu** and **78Cu**. Reproduced from ref. 202 with permission from the Royal Society of Chemistry, copyright 2019.

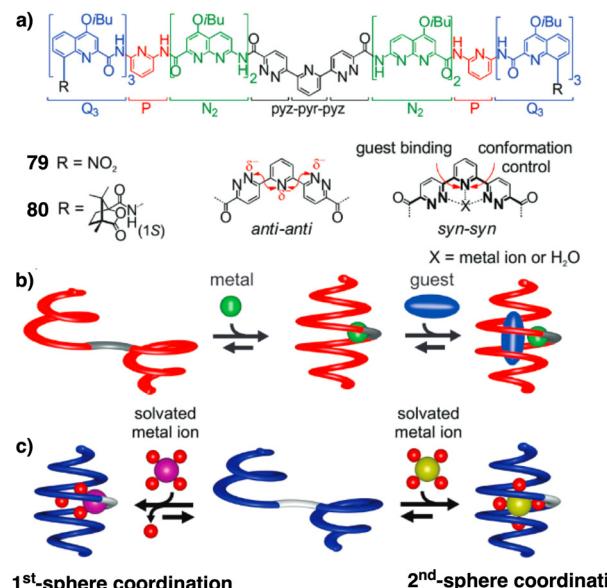


Fig. 29 (a) Chemical of aromatic oligoamide sequences **79** and **80** containing the central diacid residue pyz-pyr-pyz and its preferred conformations in the absence and in the presence of a guest. (b) Depiction of helical-capsule folding upon metal coordination and guest binding in the capsule cavity assisted by the metal ion (reproduced from ref. 203 with permission from John Wiley and Sons, copyright 2017). (c) Schematic representation of metal-induced folding modes of a helical-capsule: first-sphere coordination (left) and second-sphere coordination (right). Reproduced from ref. 204 with permission from the Royal Society of Chemistry, copyright 2017.



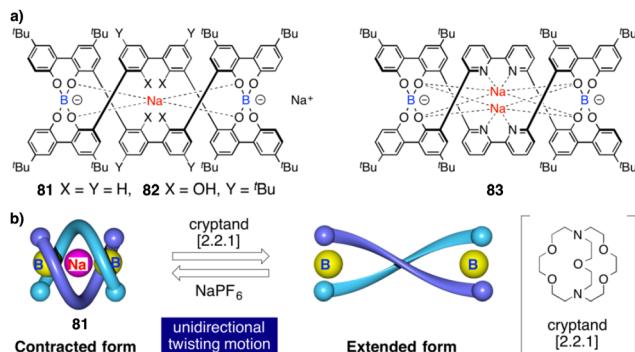


Fig. 30 (a) Chemical structures of double-stranded spiroborate helicates **81**, **82**, and **83**. (b) Schematic representation of the unidirectional spring-like motion upon  $Na^+$ -ion release and binding in **81**. Reproduced from ref. 206 with permission from the Royal Society of Chemistry, copyright 2019.

Contraction motions of double-stranded spiroborate helicates have been studied, in response to the binding and release of protons and/or metal ions, by Yashima.<sup>205,206</sup> Thus, novel helicates composed of two tetraphenol strands bridged by two spiroborate groups (81, Fig. 30a) exhibited a unique unidirectional spring-like motion triggered by the presence of sodium cations. The contracted double-stranded helicate with embedded sodium ions undergoes a conformational change upon ion release with cryptand [2.2.1] that results in the extension of the helicate (Fig. 30b). However, when *ortho*-linked biphenol 82 or 6,6'-linked 2,2'-bipyridine (bpy) 83 are used as central residues (Fig. 30a), the extended form cannot be induced after addition of cryptand [2.2.1] due to their strong interaction with the sodium ion.

**2.3.2 Cation-responsive supramolecular polymers.** Addition of metal ions can affect supramolecular self-assembly of building blocks used in supramolecular chemistry. For instance, Bhattacharya showed that the addition of  $Ca^{2+}$  ions to a pyrene-appended oligopeptide sequence VPGKP (PyP) reinforces the mechanical strength and reduces the critical gelator concentration of the native gel through coordination with the free  $-COO^-$  group of the gelator.<sup>207</sup>

In another example, Yam found that supramolecular helical metallofoldamers that conduct to a gel can be obtained by combining an oligomeric *m*-phenyleneethynylene backbone with  $Pt^{(II)}$  ions. This metallofoldamer self-assembles through Pt-Pt and  $\pi-\pi$  interactions to create a gel.<sup>208</sup> G. Feng reported also how, depending on the metal used, a ditopic ligand composed of terpyridine and acetylene segments can self-assemble into a helical ( $Cu^{2+}$ ) or non-helical ( $Zn^{2+}$ ) aggregate.<sup>209</sup> C.-L. Feng reported the self-assembly of (L)-phenylalanine derivatives in presence of different metal ions that determine the helical sense induction of the assemblies.<sup>210</sup>

Continuing with metal–metal and  $\pi-\pi$  interactions, Akine contributed to the field with the study of metal complexes that exhibit stacking behaviour consisting of acyclic pyridine-phenol ligands and  $Ni^{2+}$  or  $Pd^{2+}$  ions.<sup>211</sup> However, in these systems, the sense or morphology of the chiral aggregate cannot be controlled by the metal ion. In this line, Liu found during the coassembly of *para*-pyridine imine-linked cholesterol conjugate

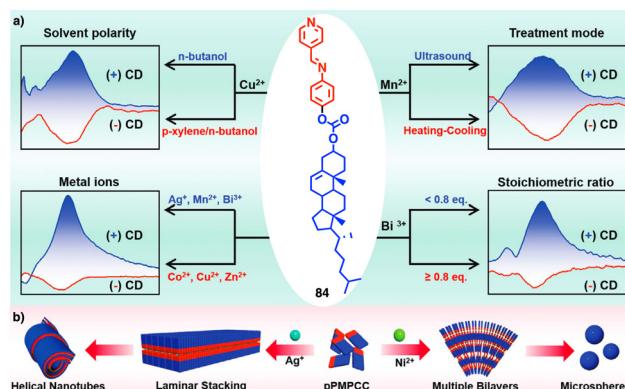


Fig. 31 (a) Quadruple modulated dynamic SMCI of MOSP-based aggregation modulated by the stoichiometric ratio, type of metal ions, solvent polarity, and the treatment of heating–cooling and ultrasound. (b) Proposed self-assembly mechanism and aggregation evolution of **84** +  $AgNO_3$  and **84** +  $NiCl_2$  complex-based coassemblies. Reproduced from ref. 212 with permission from John Wiley and Sons, copyright 2023.

(pPMPCC, 84, Fig. 31a) with different metal salts that a precise control over the chirality of the resulting metal–organic supramolecular polymer (MOSP) aggregates can be obtained by choosing the correct metal ion (Fig. 31b).<sup>212</sup> For instance, metallosupramolecular polymer obtained from 84 and  $Co^{2+}$ ,  $Cu^{2+}$  and  $Zn^{2+}$  produced chiral aggregates with opposite chirality to those obtained by combination of 84 and  $Ag^+$ ,  $Mn^{2+}$  and  $Bi^{3+}$ . Interestingly, in the case of the 84/ $Bi^{3+}$  complex a transition between opposite chiral aggregates can be obtained by varying the 84/ $Bi^{3+}$  ratio (Fig. 31a).

Yashima and co-workers investigated helical foldamer 85, which has a flexible metal-coordination moiety of 2,6-pyridinebis(acylhydrazone) (Fig. 32a).<sup>213</sup> This foldamer can adopt different helical structures in the presence or absence of silver metal ions. Moreover, foldamer and metallofoldamer self-assemble to form helical nanofibers with different shapes, which can undergo reversible transformations by adding or removing the silver ions. This macromolecular conformational change is driven by the metal ions and occurs in a cooperative and positive allosteric manner, meaning that the binding of one ion influences the binding of others. Thus, the metal-coordination-driven conformational change of 2,6-pyridinebis(acylhydrazone) linkers from a W-form to a U-form facilitates the formation of another helical foldamer composed of U-shaped linkers (Fig. 32b).

Lin studied the self-assembly and stimuli-responsive properties of a tripodal quinolinamido-based supramolecular organogel (86, TBT-gel) (Fig. 33).<sup>214</sup> Through experimental and theoretical studies, it is demonstrated how the TBT-gel assembles into a one-dimensional helical supramolecular polymer through strong hydrogen bonds and  $\pi-\pi$  interactions. The addition of  $Fe^{3+}$  ions leads to the formation of a metallogel (86-Fe-gel) through coordination interactions. Interestingly, the TBT-gel selectively responds fluorescently to  $Fe^{3+}$  and  $F^-$  ions *via* a competitive coordination mechanism, which is attributed to an intramolecular charge transfer process (Fig. 33).

Feng explored the control of circularly polarized luminescence (CPL) in supramolecular gels obtained by co-assembly of



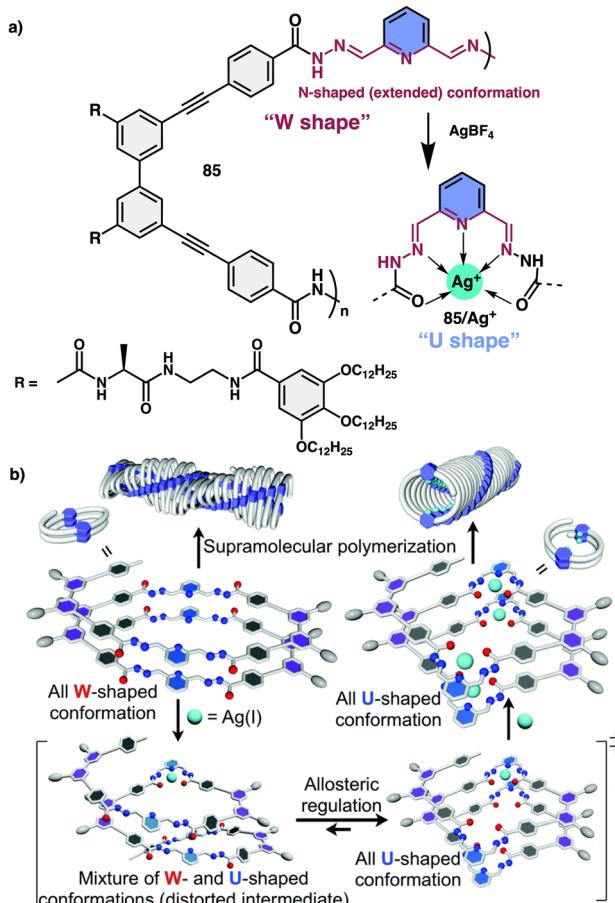


Fig. 32 (a) Chemical structure of **85**. (b) Schematic representation of  $\text{Ag}(\text{I})$ -triggered W-to-U-shaped conformational changes of 2,6-pyridinebis(acylhydrazone) linker units. Reproduced from ref. 213 with permission from the Royal Society of Chemistry, copyright 2018.

a chiral phenylalanine-derived hydrogelator (**87**) (Fig. 34) and achiral coumarin derivatives triggered by addition of metal ions.<sup>215</sup> The study demonstrates that the handedness of CPL in these hydrogels can be efficiently inverted by the incorporation of coumarin derivatives through non-covalent interactions, and further tuned by the addition of metal ions. The key factors influencing CPL inversion are hydrogen bonds, coordination interactions, and steric hindrance.

The co-assembled hydrogels exhibit chiral inversion and the formation of different nanostructures (nanotwists, nanohelices, and nanotubes) depending on the combination of components (Fig. 34).

Zhao investigated the self-assembly of a cholesterol-azopyridine conjugate known as PAzPCC (**88**) and the influence of light and metal ions in the supramolecular chirality of the aggregates (Fig. 35a).<sup>216</sup> Thus, PAzPCC forms organogels that undergo a dimensional transition from 2D microbelts to 1D nanotubes and finally to 0D nanoparticles when exposed to UV light, which is driven by *E/Z* photoisomerization of the azopyridine unit. Furthermore, the self-assembled structures show helicity inversion mediated by metal ions through metal coordination. Interestingly, variations in the solvent while keeping the metal ion unaltered can

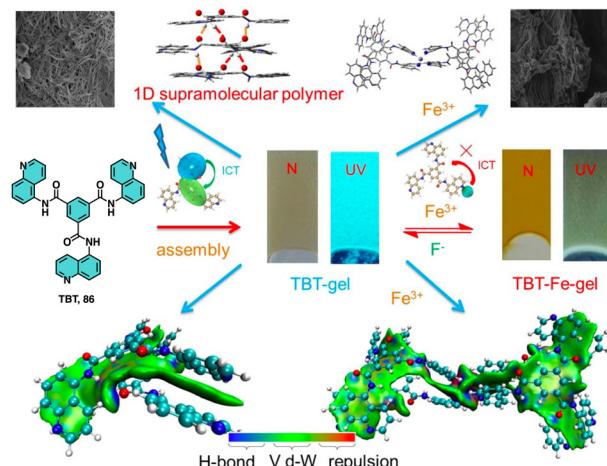


Fig. 33 Structure of **86**. Proposed assembly and ion response mechanisms of **86** with  $\text{Fe}^{3+}$  and  $\text{F}^-$ . Reproduced from ref. 214 with permission from John Wiley and Sons, copyright 2021.

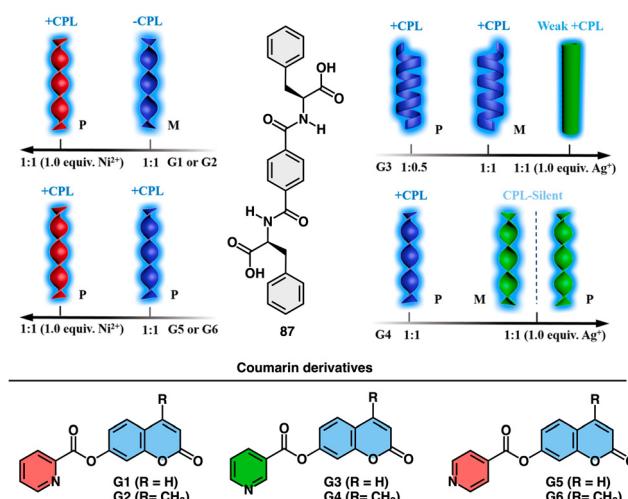


Fig. 34 Schematic representation of the co-assembly of chiral LPF (**87**) and achiral fluorescent coumarin derivatives (**G1-G6**) and their response to metal ions ( $\text{Ni}^{2+}$  and  $\text{Ag}^+$ ). Reproduced from ref. 215 with permission from the American Chemical Society, copyright 2019.

lead to the formation of structures with opposite or racemic helical sense (Fig. 35b and c).

Jaworsky and Jung described a *P/M* reversible helical switch based on the formation of a chiral supramolecular gel using terpyridine-based ligands (**89**) and  $\text{Co}^{2+}$  ions at different metal/ligand ratios (Fig. 36a).<sup>217</sup> Thus, at low  $\text{Co}^{2+}$  concentrations, octahedral complexes that promote the formation of *M* aggregates are formed, while above 0.6 equivalents, a co-existing square pyramidal complex that promotes the formation of *P* aggregates emerges (Fig. 36a). The helical inversion was fully reversible and controlled. Being enantiomeric ligands, **89** and **90** exhibited analogous  $\text{Co}^{2+}$  concentration-dependent switching mechanism (Fig. 36b).

**2.3.3 Cation-responsive dynamic covalent polymers.** Maeda and Freire designed an achiral poly(phenylisocyanate) that

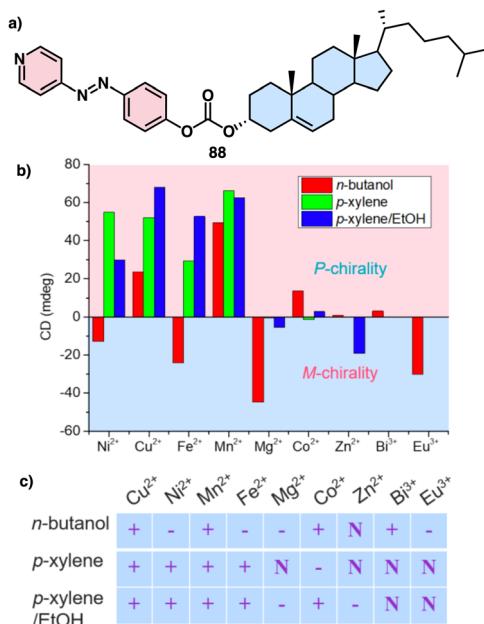


Fig. 35 (a) Chemical structure of **88** (b) relationship between the ECD absorption at 450 nm and corresponding chirality. (c) Supramolecular chirality of nanostructures based on PAzPCC and metal ions regulated by metal ion and solvent effects. "+" represents positive Cotton effect, "-" represents negative Cotton effect, and "N" stands for negligible supramolecular chirality. Reproduced from ref. 216 with permission from the American Chemical Society, copyright 2018.

bears a single chiral MPA derivative at the polymer terminus ( $\alpha$ -end) (poly-**91**) (Fig. 37a).<sup>218</sup> This chiral residue has a switchable conformation that allows, through a conformational communication mechanism, to control the *P*/*M* helical sense of the polymer backbone when different external stimuli such as solvents or metal ions are used (Fig. 37b and c). The helix enhancement is produced *via* a domino effect, whose helical sense was certainly assigned by a combination of VCD spectroscopy and DFT calculations (Fig. 37d). Freire and Riguera used the same chiral MPA group as pendant to control the helical sense of a PPA by using monovalent or divalent metal ions (Fig. 38a and b).<sup>219</sup> Thus, while in poly-(*R*)-**11** an *M* helix is induced by addition of monovalent metal ions, which stabilize an *ap* conformer at the pendant, addition of divalent metal ions induces a *P* helix by fixing a *sp* conformer.

The cross-linking ability of metal ions was employed to prepare nanospheres with tuneable size and chirality, which were achieved by controlling the metal-polymer ratio (Fig. 38c).<sup>220–223</sup> Interestingly, in the case of poly-(*R*)-**11**/*M*<sup>+</sup> helical polymer–metal complexes, a dual *P* or *M* helical sense can be induced by adding a cosolvent that switches the coordination mode between the monovalent ion and the pendant group, allowing tuning of the *sp* or *ap* conformer.<sup>224</sup> The ability of poly-(*R*)-**11** to adopt three different chiral states in the absence (axially racemic) and the presence of monovalent (*M* helix) and divalent (*P* helix) metal ions was used by Maeda and Freire to create a three-state switchable chiral stationary phase that allowed inverting the elution time of enantiomers by changing the helical sense of the polymer (Fig. 38d).<sup>225,226</sup>

The capacity of the MPA pendant group to interact selectively with monovalent and divalent metal ions was used by Freire and Riguera to explore the activation of the Sergeants and Soldiers effect in an axially racemic copolymer<sup>227</sup> series—poly-**11**<sub>r</sub>–co–**92**<sub>1–r</sub>—made by a dormant Sergeant (**11**) and an achiral Soldier, that bears the anilide of diphenyl acetic acid (**92**) (Fig. 39). By tuning the conformational composition of the chiral MPA pendant with monovalent and divalent metal ions in the poly-[**11**<sub>r</sub>–co–**92**<sub>1–r</sub>] copolymer series, it was possible to selectively activate the Sergeants and Soldiers effect, adopting the copolymer a *P* or an *M*

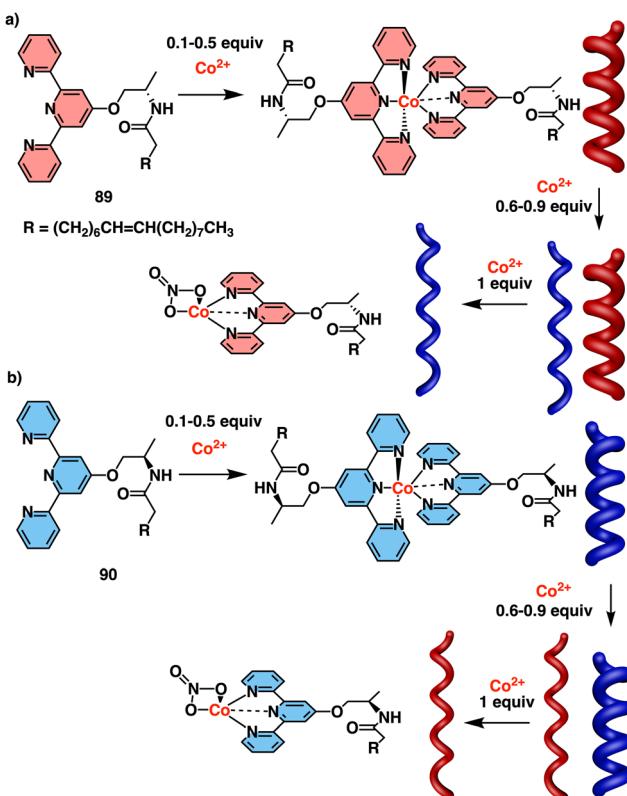


Fig. 36 Structures of (a) **89** and (b) **90** and their coordination with different equivalents of Co<sup>2+</sup>. Reproduced from ref. 217 with permission from the Royal Society of Chemistry, copyright 2014.

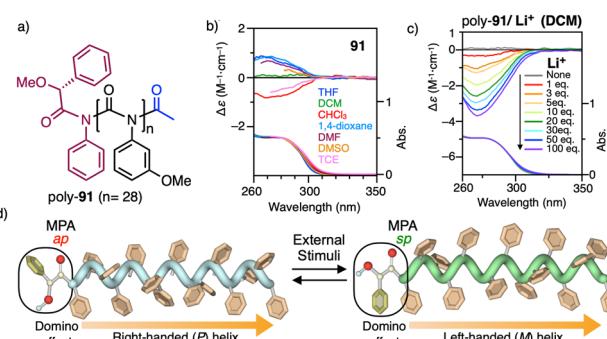


Fig. 37 (a) Chemical structure for poly-**91**. (b) ECD spectra of poly-**91** in different solvents and (c) in dichloromethane after addition of different amounts of LiClO<sub>4</sub>. (d) Schematic illustration of the switchable domino effect along a polymer chain. Reproduced from ref. 218 with permission from the Royal Society of Chemistry, copyright 2019.



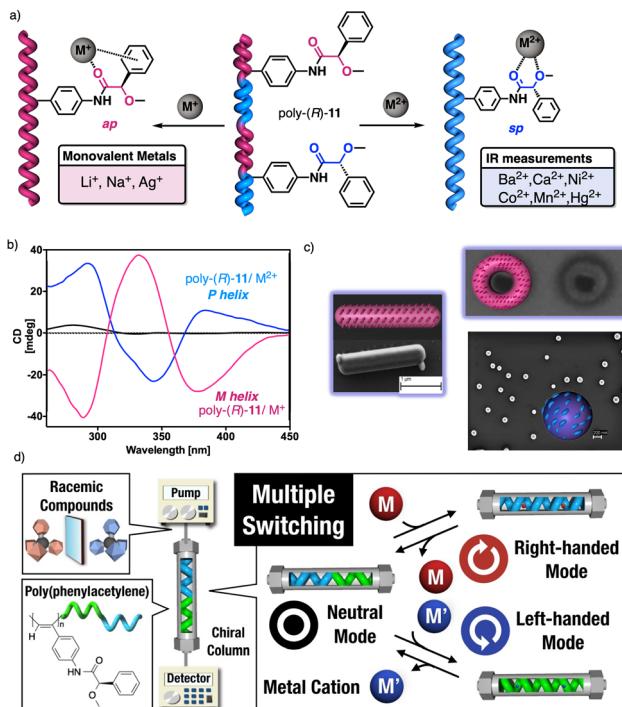


Fig. 38 (a) Schematic illustration and (b) ECD spectra of the helical sense induction in poly-(R)-11 by monovalent and divalent metal ions. (c) Different nanostructures obtained for poly-(R)-11/M<sup>+</sup>. Reproduced from ref. 222 with permission from John Wiley and Sons, copyright 2014. (d) Application of poly-(R)-11/M<sup>+</sup> as multiple-switch chiral stationary phase. Reproduced from ref. 225 with permission from the American Chemical Society, copyright 2019.

helical sense commanded by the Sergeant (Fig. 39). Additionally, it was also found that both *P* and *M* helical senses can be induced in poly-[11<sub>1</sub>-co-92<sub>1-n</sub>] copolymer series by using monovalent metal ions and a cosolvent.<sup>228</sup>

Working on the mechanisms of conformational communication along fully chiral copolymer chains, selective MPA/M<sup>n+</sup> ( $n = 1, 2$ ) interactions were employed to describe the helix induction mechanisms in copolymer series that show chiral coalition,<sup>113,116</sup> chiral to chiral Sergeants and Soldiers effect,<sup>112</sup> chiral accord and chiral conflict.<sup>117</sup>

Riguera, Seco and Freire explored the coordination of metal ions with PPAs containing amino acids as pendant groups to change the conformational composition in the amino acids and induce a helical change (sense/elongation) in the PPAs. Thus, in seminal work, Riguera and Seco discovered that an *anti-syn* conformational change can be triggered in a PPA bearing the benzamides of (L)-phenylglycine methyl ester (PGME, poly-5) as pendant when dissolved in low polar solvents.<sup>98</sup> Therefore, if a metal ion is added to the two carbonyl groups of the pendant when they are oriented *antiperiplanar* (*anti*) in low polarity solvents, a chelate is produced between the metal ion and the amino acid that orients the two carbonyls in a *synperiplanar* (*syn*) manner.

This conformational change is accompanied by helical inversion of the PPA (Fig. 2b). Subsequently, Riguera and Freire demonstrated that PPAs containing aromatic amino acids as

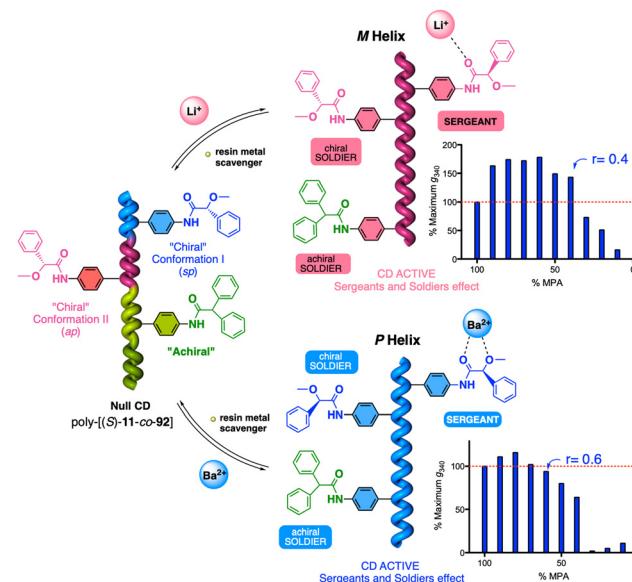


Fig. 39 On/Off Sergeants and Soldiers effect in poly-[11<sub>1</sub>-co-92<sub>1-n</sub>] copolymer series by metal ions (on) and resin metal scavengers (off). Reproduced from ref. 227 with permission from the Royal Society of Chemistry, copyright 2014.

pendants, such as the benzamide of (L)-phenylalanine methyl ester (poly-93), or the benzamide of (L)-phenylglycine methyl ester (poly-5), can adopt up to four different helical scaffolds, in chloroform *via* dynamic coordination chemistry, when Li<sup>+</sup> and a cosolvent as methanol are used as external stimuli (Fig. 40a).<sup>229</sup> Thus, while the Li<sup>+</sup> ion can create a tripodal coordination with the carbonyls and the aryl ring of the pendant, the cosolvent (MeOH) has the ability to change the coordination mode of the Li<sup>+</sup> due to the selective disruption of Li<sup>+</sup>-O=C and Li- $\pi$  interactions (Fig. 40a).

If other amino acids that lack an aromatic ring in the side chain are used as pendants, three different helices, instead of four, can be obtained from this dynamic metal-pendant coordination.

The crosslinking ability of the metal ions was used by Freire and Riguera to create *P* and *M* macroscopically chiral nanospheres from a single PPA-metal complex (Fig. 40b).<sup>230</sup>

## 2.4 External stimulus: redox

Metal complexes with a chiral response to redox stimuli<sup>231-234</sup> have attracted the attention of the scientific community, as well as the chiral response of rotaxanes that exhibit redox responsiveness.<sup>235</sup> Currently, there is growing interest in harnessing redox processes to control the properties and functions of larger macromolecules, such as polymers and foldamers (Fig. 41a), enabling the design of stimuli-responsive systems as well as the investigation of their properties.

**2.4.1 Redox-responsive foldamers.** Tetrathiafulvalene (TTF) is a very popular electro-donating molecule with a rich redox chemistry found in numerous research articles due to its efficient radical formation. Therefore, combining TTF with other fragments with self-assembly properties can generate interesting materials. For instance, Sallé designed a tetrathiafulvalene redox

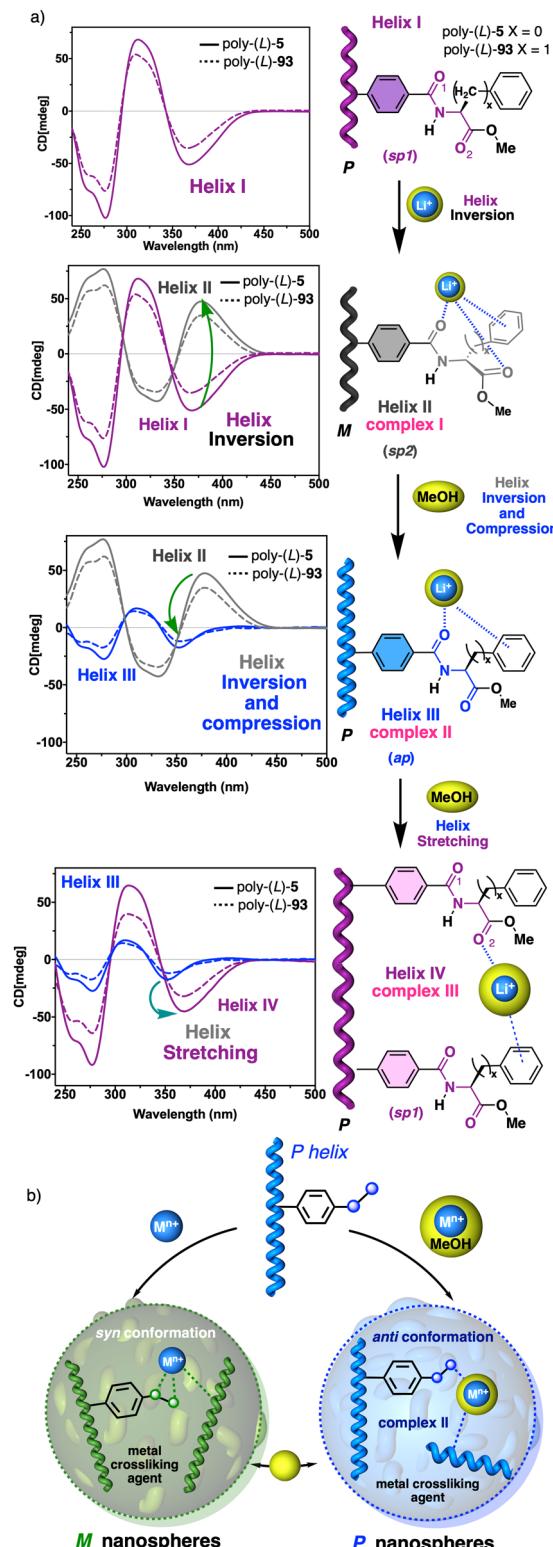


Fig. 40 (a) Elongation and helical sense control via dynamic coordination chemistry in poly-(L)-5 and poly-(L)-93/Li<sup>+</sup>/MeOH complexes. (b) Illustration of nanostructuration via dynamic coordination chemistry. Reproduced from ref. 229 with permission from the Royal Society of Chemistry, copyright 2017.

unit combined with an oligopyridine-dicarboxamide foldamer (**94**)<sup>236</sup> (Fig. 41b) that folds, after oxidation, into stable radical-cation dimers. A nonlinear increase in difference of UV-Vis absorption with the concentration before and after the electrolysis confirmed the intermolecular interactions of **94** (Fig. 41c).

The different spectroscopic data align with the dimerization of **94**, while the possibility of supramolecular macrocycle formation is ruled out based on the high experimental value of  $K_{\text{dim}}$  (Fig. 41d and e), which cannot be achieved through  $\pi$ -dimer interactions alone, as it happens in the macrocycle structure.

They propose that the oligopyridine-dicarboxamide has stronger dimerization tendency in the oxidized foldamer compared to its neutral counterpart [ $K_{\text{dim}}(\mathbf{94}^{2(\bullet^+)})) = 10^2 \times K_{\text{dim}}(\mathbf{94})$ ]. Moreover, it was found that the length of the oligopyridine-dicarboxamide foldamer plays an important role in the supramolecular polymerization of **94**. For instance, an oligomer containing the terminal

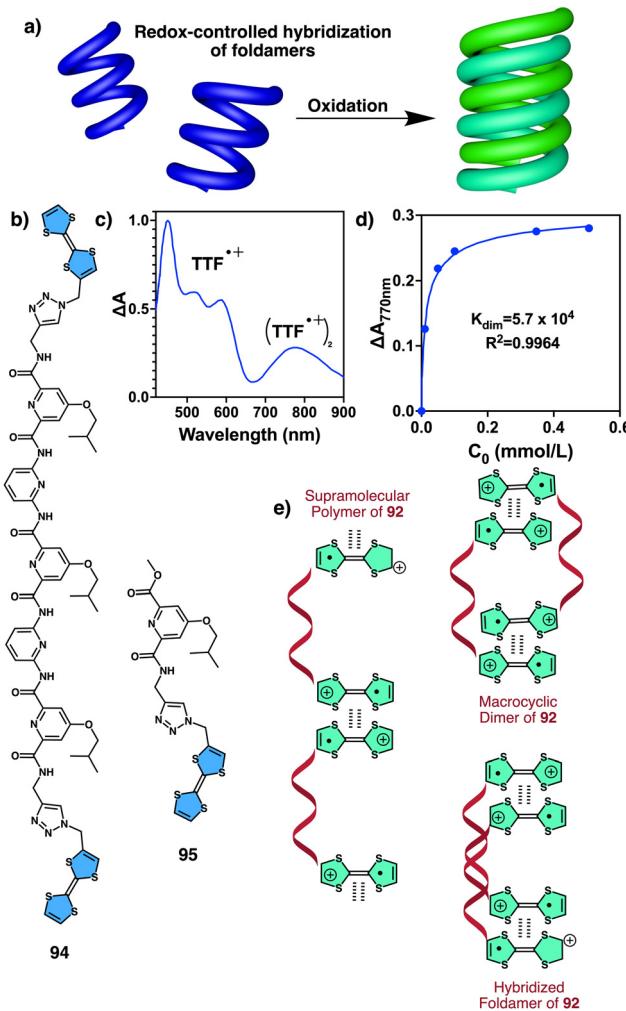


Fig. 41 (a) Schematic representation of foldamer hybridization after oxidation. (b) Chemical structure of foldamer **94** and molecule **95**. (c) Typical normalized spectrum obtained through spectroelectrochemical measurement upon oxidation of **94**. (d) Evolution of  $\Delta A_{770}$  upon oxidation of **94** (e) possible supramolecular arrangements accounting for intermolecular radical-cation dimerization of **94**. Reproduced from ref. 236 with permission from the Royal Society of Chemistry, copyright 2019.



TTF and a pyridine carboxiamide residue (95) (Fig. 41b) needs to increase the solution concentration up to 250-fold to force its aggregation. This result also indicates that the terminal TTF groups do not have propensity to form 1D polymers, and therefore excluding as aggregation products from 94, the formation of 1D polymers.

Stoddart studied the design, folding behaviour, and potential applications of redox-responsive foldamers constructed using oligobipyrinidinium chains.<sup>237</sup> Varying the number of BIPY<sup>2+</sup> units in oligoviologens 96-(2V<sup>4+</sup>), 96-(3V<sup>6+</sup>), 96-(4V<sup>8+</sup>), 96-(5V<sup>10+</sup>) and 96-(12V<sup>24+</sup>) (Fig. 42a), and utilizing short, flexible p-xylylene linkers, the researchers achieved extended conformations through Coulombic repulsion in the fully oxidized state. Upon reduction, the foldamers folded into stable conformations due to interactions between BIPY<sup>•+</sup> radical cations.

This study shows how the folding behaviour of 96 was influenced by intra- and intermolecular radical–radical interactions (Fig. 42b), which depended on the chain length of the oligoviologens. Thus, longer oligoviologens, *i.e.*, 96-(12V<sup>24+</sup>), showed greater radical stabilization, resulting in lower energy levels in their reduced states and the formation of intramolecular trisradical complexes. In contrast, the shorter oligoviologens, 96-(2-5V<sup>n+</sup>), displayed reduced radical stabilization, rendering them unable to form trisradical complexes (Fig. 42c). Consequently, foldamers with a reduced number of

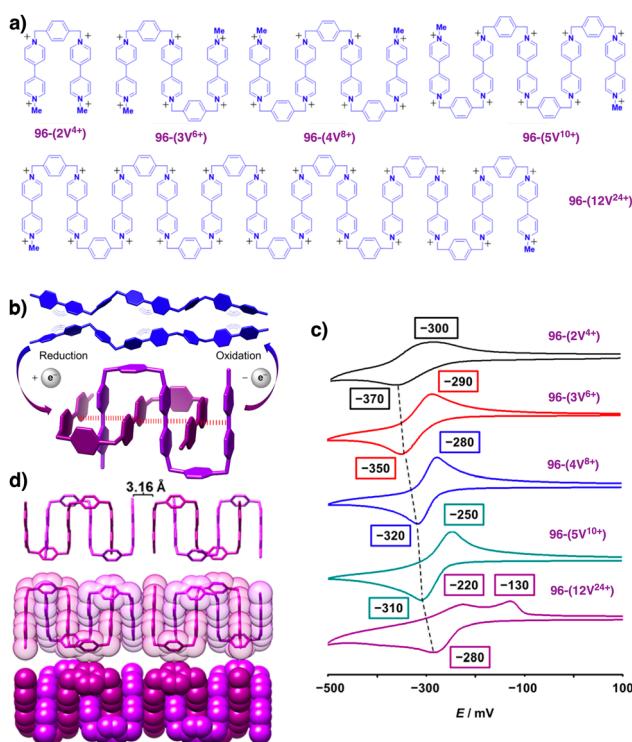


Fig. 42 (a) Structural formulas of 96-(2V<sup>4+</sup>), 96-(3V<sup>6+</sup>), 96-(4V<sup>8+</sup>), 96-(5V<sup>10+</sup>), and 96-(12V<sup>24+</sup>). PF<sub>6</sub><sup>−</sup> counterions are omitted. (b) Schematic illustration of the switching process under redox stimulus. (c) Cyclic voltammograms of five oligoviologens. (d) Representations along the *a*-axis of the long-range packing order of the radical cationic species 3V<sup>3•+</sup>. Reproduced from ref. 237 with permission from the American Chemical Society, copyright 2015.

viologens units form radical–radical dimers and infinite packing due to intermolecular radical–radical interactions (Fig. 42d).

**2.4.2 Redox-responsive supramolecular polymers.** Redox phenomena play an important role in the chirality and morphology of supramolecular helical aggregates. Thus, Feng found that the chirality of a supramolecular gel consisting of a derivative of (L)-phenylalanine diethylene glycol (LPFEG) (97) (Fig. 43a) can be tuned by the oxidation state of graphene oxide (LPFEG–GO) added to the system.<sup>238</sup> It was found that reduction of graphene oxide (GO) to reduced graphene oxide (RGO) disrupts the hydrogen bonding and hydrophobic interactions between GO and the chiral molecules (97), leading to a chiral inversion in the LPFEG–GO gel (Fig. 43b and c). The redox phenomena and reduction of GO to RGO significantly influenced the supramolecular chirality and drove the reversible chiral inversion observed in the LPFEG–GO gel system.

The redox phenomenon also plays a significant role in the sol–gel transition of various supramolecular systems containing cholesteric groups on their outer surface. A noteworthy study by Fang and co-workers showed the redox sensitivity of a chiral cholesteric ferrocene gel to different oxidants.<sup>239</sup> Thus, the building block 98 (Fig. 44a) was designed introducing a redox sensitive ferrocene group derivatized, in each cyclopentadiene ring, with a glycine residue that has a cholesterol unit anchored at the C terminus. This molecule self-assembles to form a chiral gel, which can be oxidized using Ce<sup>4+</sup> [(NH<sub>4</sub>)<sub>2</sub>Ce(No<sub>3</sub>)<sub>6</sub>] and producing a colour change from yellow to green. This colour switch is attributed to the different oxidation states of the ferrocenyl residues within the gel, which are transformed into ferrocenium cations. The presence of positive charges at the core of the supramolecular aggregate disrupts the intermolecular hydrogen bonding network and the interaction of cholesteric residues resulting in a gel to sol process.

Conversely, although the addition of hydrazine for the reduction of ferrocenium cations to ferrocenyl produces the desired yellow colour, the gel state could not be recovered (Fig. 44a). Other examples of supramolecular assemblies involving ferrocene derivatized with peptide moieties have been used to demonstrate sol–gel

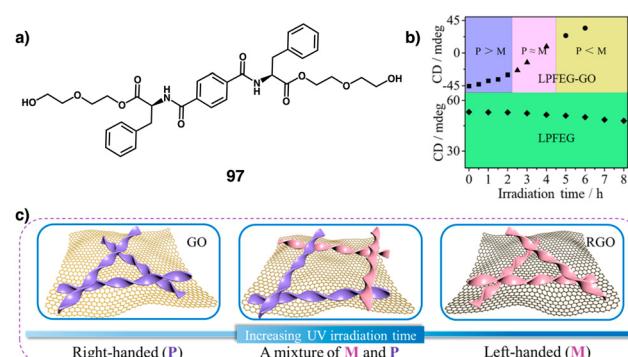


Fig. 43 (a) Structure of monomer 97. (b) Plot of CD intensity of LPFEG and LPFEG–GO hydrogels at around 270 nm versus UV irradiation time. (c) The representative transition stages from right-handed nanofibers to left-handed twists with increasing UV irradiation time. Reproduced from ref. 238 with permission from the American Chemical Society, copyright 2020.

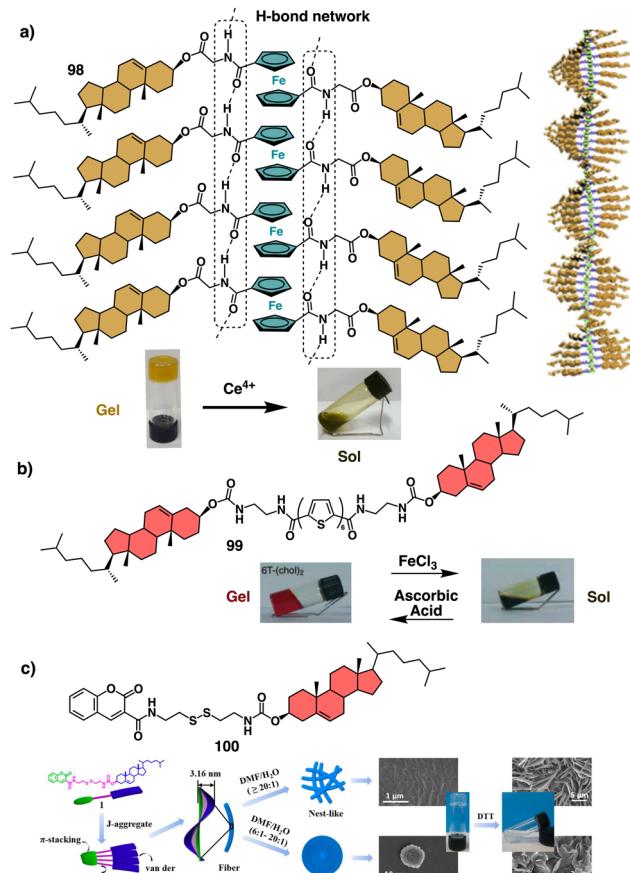


Fig. 44 (a) Self-assembly structure of **98** and gel–sol transition after oxidation with  $(\text{NH}_4)_2\text{Ce}(\text{NO}_3)_6$ . Reproduced from ref. 239 with permission from Elsevier, copyright 2015. (b) Structure of monomer **99** and the reversible gel–sol transition after oxidation and reduction. Reproduced from ref. 241 with permission from John Wiley and Sons, copyright 2005. (c) Structure of monomer **100**. Formation of a gel made of J aggregates and gel–sol transition after oxidation with DTT. Reproduced from ref. 242 with permission from the Royal Society of Chemistry, copyright 2014.

transitions induced by a redox process, as observed in the work conducted by Kraatz and Adhikari on the supramolecular transformation of FeCO-Val-Phe-Phe-OMe from a nanofibrous gel to a spherical micelle.<sup>240</sup> Shinkai<sup>241</sup> designed an oligothiophene-based gelator bearing cholesteryl groups at both ends of the thiophene sequences (**99**) to study its redox properties. In this case, a reversible redox sol–gel transition was found by adding  $\text{FeCl}_3$  as an oxidant and ascorbic acid as a reductant (Fig. 44b). Ju described another interesting redox cholesteric gel using a disulfide bond as redox stimuli responsive group (**100**).<sup>242</sup> Thus, compound **100** forms a gel through a combination of H-bonds and  $\pi$ – $\pi$  interactions when the disulfide bond is present. However, addition of a reductant agent such as 1,4-dithiothreitol (DTT) destroys the fibres, obtaining a solution that can be monitored by SEM (Fig. 44c).

**2.4.3 Redox-responsive dynamic covalent polymers.** Dynamic covalent helical polymers also play an important role in the development of redox responsive macromolecular helical structures. Thus, Wu<sup>243</sup> prepared a polyisocyanide with a thioether pendant (poly-**101**) that can be oxidized to a sulfoxide group

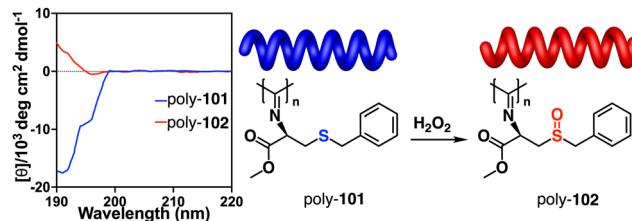


Fig. 45 Chemical structures of poly-**101** and poly-**102**. ECD spectra showing the helix inversion after oxidation of poly-**101** to poly-**102**. Reproduced from ref. 243 with permission from De Gruyter, copyright 2019.

(poly-**102**) using hydrogen peroxide (Fig. 45). Upon oxidation, the specific optical rotations and Cotton effects of poly-**101** are reversed, suggesting helix inversion of the polyisocyanide macromolecular structure (Fig. 45).

In a different example, also using polyisocyanides as dynamic helical polymers, Wu studied the use of organoiodine-functionalized helical block copolymers as catalysts for asymmetric oxidations (poly-**103**) (Fig. 46).<sup>244</sup> By immobilizing the organocatalyst on the helical polymers, the researchers not only facilitated catalyst recycling from homogeneous reactions, but also observed a significant boost in enantioselectivity. This immobilization strategy allowed the creation of an organoiodine-functionalized single left- and right-handed helical polyisocyanides block copolymerized with chiral monomers. These helical polyisocyanides served as catalysts for three asymmetric oxidations, delivering the desired products with high yields and excellent enantioselectivity (Fig. 46).

Amabilino explored the potential of electroactive organic polymers as chiral redox-polymer systems for applications in molecular electronics.<sup>245</sup> Thus, a chiral polyisocyanide that incorporates a tetrathiafulvalene (TTF) derivatized with stereogenic centres (poly-**104**) was designed and prepared (Fig. 47). This polymer shows a reversible interconversion between three univalent and two mixed-valence redox states, each of which displays distinct chiroptical properties associated with the

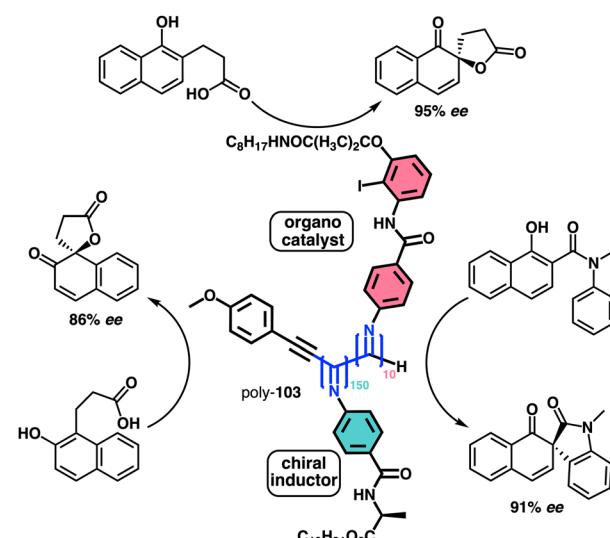


Fig. 46 Structure of poly-**103** and three asymmetric catalysed reactions.



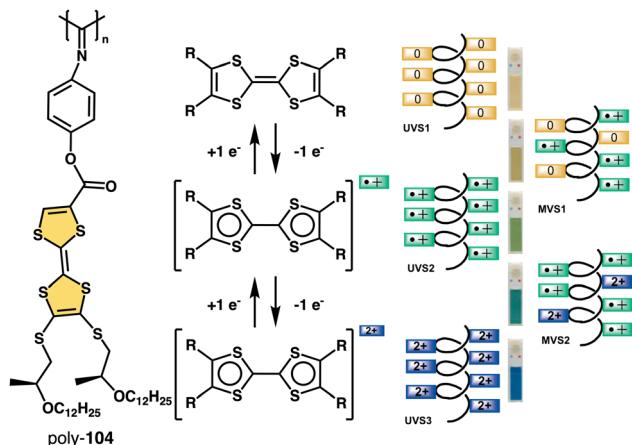


Fig. 47 Cartoon representation of the redox states of poly-104 with TTF units in the side chains that can be interconverted through redox reactions and the colours shown by the solutions of these species. Reproduced from ref. 245 with permission from John Wiley and Sons, copyright 2005.

presence of helical structures, as deduced from ECD studies (Fig. 47).

Freire found that during the synthesis of hybrid materials from dynamic helical polymers and metal nanoparticles,<sup>246,247</sup> adding HAuCl<sub>4</sub> to an anilide-PPA [poly-(R)-11], oxidation of the polyene backbone occurs, which leads to the loss of the helical structure due to a delocalization of the radical along the polyene chain (Fig. 48a–c). To overcome this problem, TOAB (tetraoctylammonium bromide) was added to the polymer solution in order to trap the Au<sup>3+</sup> cations and prevent the oxidation of the polyene (Fig. 48d).

In this new protocol, gold interact with the polymer once it is reduced to Au<sup>0</sup>, which is obtained after the addition of NaBH<sub>4</sub> as reducing agent. In this poly-11-AuNPs nanocomposite, the AuNPs are stabilized by supramolecular anilide–AuNPs interactions (Fig. 48d).

This protocol was also used to successfully prepare a poly-11-AgNPs nanocomposite.<sup>248</sup> In a different example, Freire explored the helix inversion of a PPA during a redox metal translocation process, a crucial step during the synthesis of helical poly(phenylacetylene)-metal nanocomposites.<sup>249</sup> A PPA copolymer made by two comonomers that contain the benzamides of methionine methyl ester [(L)-105] and (L)-alanine methyl ester [(L)-106] as pendants in 0.2 (L)-105/0.8 (L)-106 ratio—poly[(L)-105<sub>0.2</sub>-co-(L)-106<sub>0.8</sub>]—was prepared. In a low-polarity solvent environment such as CHCl<sub>3</sub>, both pendant groups adopt an *anti*-conformation that induces an *M* helix into the copolymer (Fig. 49). However, upon complexation with gold (Fig. 49a–d) or silver (Fig. 49e–h) cations, the copolymer undergoes helix inversion from *M* to *P* due to the adoption of *syn* conformations in the pendants through chelation with metal ions. Further reduction of metal ions with NaBH<sub>4</sub> produces translocation of the metal towards the tioether group, which is accompanied by a conformational change from *syn* to *anti* in the pendants accompanied by a *P* to *M* helix inversion of the polymer (Fig. 49).

Yashima studied helical poly(phenylacetylene)s (PPAs) that bear riboflavin as pendants by cyclic voltammetry (poly-107)

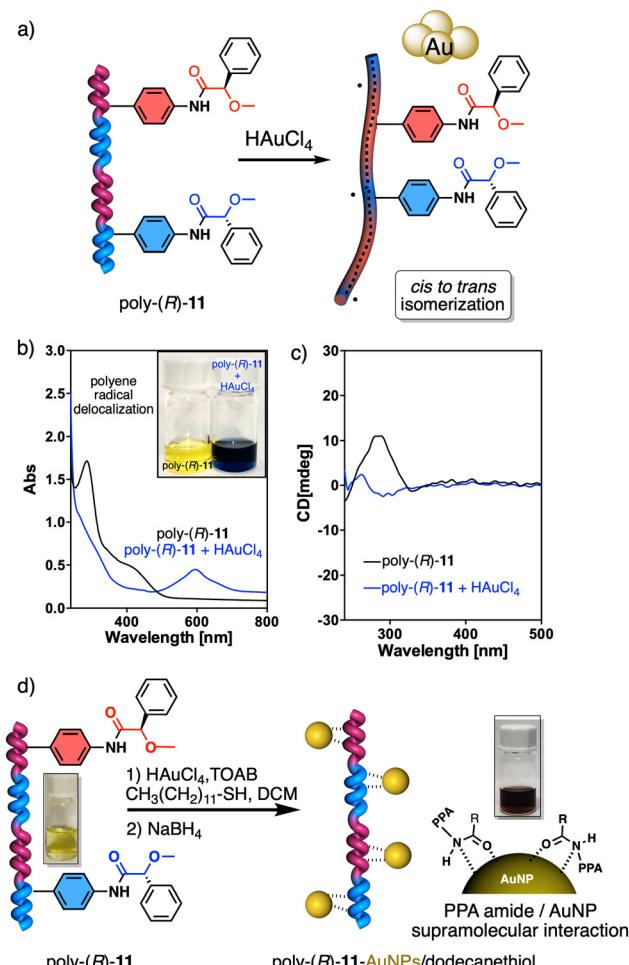


Fig. 48 (a) Schematic representation for the formation of radicals in poly-(R)-11 in presence of HAuCl<sub>4</sub>. (b) UV-vis and (c) CD spectra of poly-(R)-11 and poly-(R)-11 in presence of HAuCl<sub>4</sub>. (d) Schematic representation for the formation of poly-(R)-11-AuNPs nanocomposites by a variation of Brust–Schiffman method. Reproduced from ref. 247 with permission from John Wiley and Sons, copyright 2021.

(Fig. 50a).<sup>250</sup> The study aimed to understand the electron transfer capabilities of riboflavin pendants in these polymers. The values obtained were similar to those of the monomer, indicating that the redox behaviour was mainly governed by the riboflavin units. On the other hand, when using another connection point between riboflavin and PPA (poly-108) (Fig. 50a), a reduction curve was found at a more positive potential, indicating a relationship between the connector used to link riboflavin and PPA and its redox potential. These redox processes reversibly induced conformational changes in the flavin skeleton that are transmitted to the helical main chain (Fig. 50b and c).

Aida and Fukushima introduced an *o*-phenylene oligomer (109) (Fig. 51b) that exhibits dynamic motion triggered by redox reactions.<sup>251</sup> Although the oligomer helices demonstrate rapid inversion in a solution environment, a remarkable phenomenon of chiral symmetry-breaking is observed during crystallization. Each crystal contains exclusively the right-handed or left-handed enantiomeric form, resulting in a mixture of

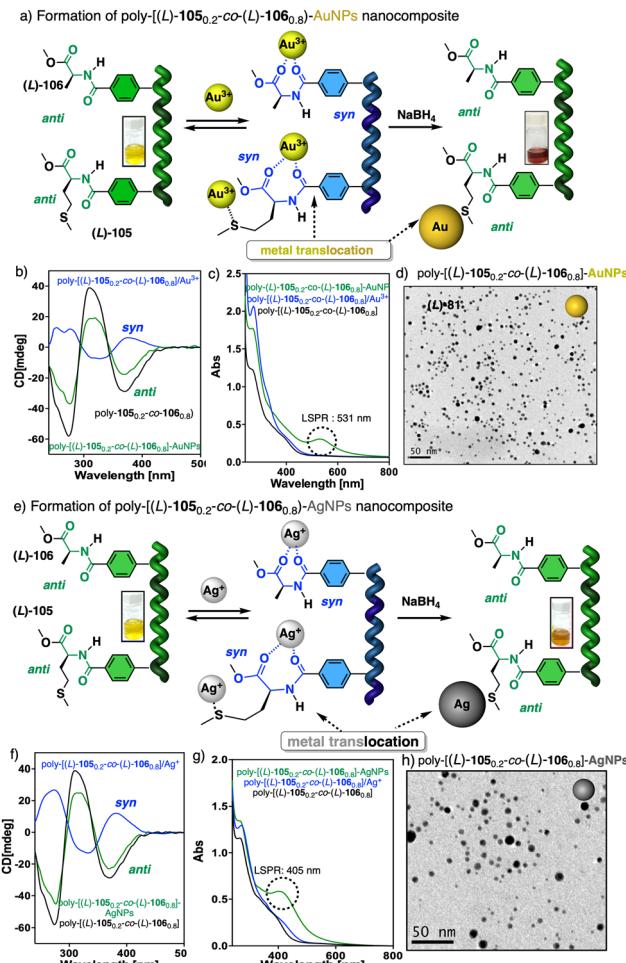


Fig. 49 (a) Schematic illustration for the preparation of poly- $[(L)-105_{0.2}-co-(L)-106_{0.8}]$ -AuNPs and its chiroptical responses associated to a redox driven translocation of a gold metal centre. (b) CD and (c) UV-vis spectra for poly- $[(L)-105_{0.2}-co-(L)-106_{0.8}]$ , poly- $[(L)-105_{0.2}-co-(L)-106_{0.8}]Au^{3+}$  and poly- $[(L)-105_{0.2}-co-(L)-106_{0.8}]$ -AuNPs. (d) TEM image for poly- $[(L)-105_{0.2}-co-(L)-106_{0.8}]$ -AuNPs nanocomposites. (e)–(h) Idem for silver. Reproduced from ref. 249 with permission from the Royal Society of Chemistry, copyright 2022.

non-racemic crystals (Fig. 51a). Furthermore, the dynamic motion of the helical oligomer in solution is considerably suppressed upon one-electron oxidation (Fig. 51b). Using X-ray crystallography, analysis of the neutral and oxidized forms reveals the generation of a hole after oxidation, which is shared by the repeating *o*-phenylene units. This shared hole facilitates locking of the helix conformation, leading to sustained chiroptical memory. These findings offer valuable insights into the behaviour of redox-active helical polymers and their potential applications in various domains.

The synthesis and properties of a series of  $\pi$ -conjugated polymers, formed by repeating units of tetrathiafulvalene vinylogue (TTFV) and phenylacetylene, were investigated as supramolecular hosts for non-covalent functionalization of single-walled carbon nanotubes (SWNTs). These polymers adopt a folding conformation in the neutral state, allowing them to interact effectively with individual SWNTs and form supramolecular complexes with good solubility in organic solvents. The polymers can be

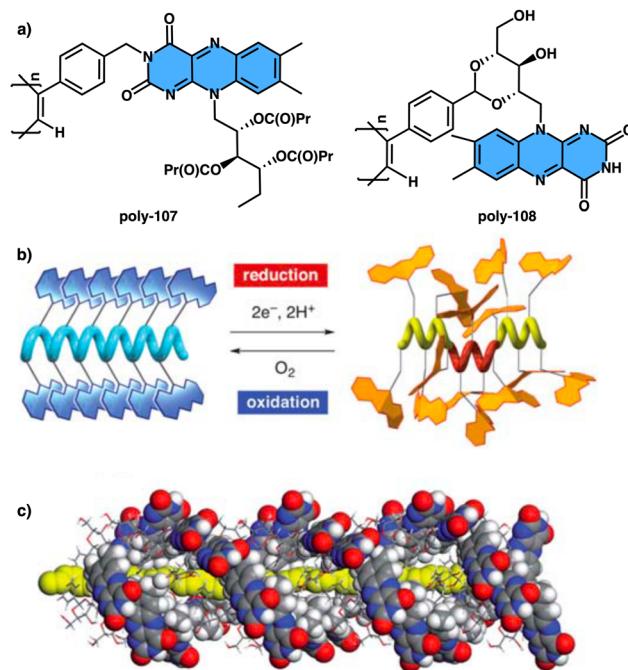


Fig. 50 (a) Chemical structures of poly-107 and poly-108. (b) Schematic illustration of the redox-triggered switching of the helical structure of poly-108. (c) Possible helical structure of poly-108 (30-mer). Reproduced from ref. 250 with permission from the Royal Society of Chemistry, copyright 2010.

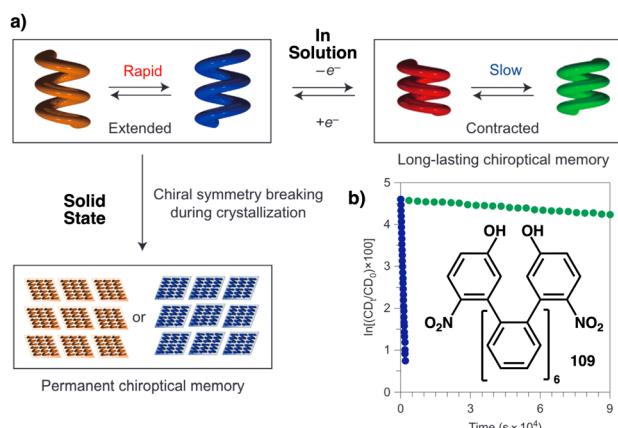


Fig. 51 (a) Schematic representation of the helical inversion dynamics of oligomer 109 in solution and in solid state. (b) chemical structures of 109 and decay profiles of the CD intensities in spectra of the oligomer dissolved in MeCN at 265 nm and the oligomer dissolved in MeCN in the presence of the one-electron oxidant  $(4-BrC_6H_4)_3N^+ [SbCl_6]^-$  at 273 nm (green). Reproduced from ref. 251 with permission from Springer Nature, copyright 2010.

activated by redox stimuli to change their molecular shapes to linear structures in the oxidized state. It results in reversible dissociation of polymers from SWNTs and efficient release of pristine SWNTs from polymer dispersants.<sup>252,253</sup> There are also examples of sol-gel transition compounds in which the disulphide bond can be cleaved with reducing agents such as dithiothreitol (DTT).<sup>254</sup>



## 2.5 External stimulus: light

The development of light-sensitive molecules has attracted the attention of numerous scientist due to the possibility of modulating the structures and functions of molecules after light irradiation. The introduction of these stimuli-responsive molecules or functional groups as components of macromolecules allows the generation of materials with unique light-responsive behavior. As a result, there are many examples in the literature of various types of well-established photo-responsive molecules, such as azobencene,<sup>255</sup> diarylethenes,<sup>256–260</sup> stilbenes,<sup>261,262</sup> molecular motors,<sup>263–266</sup> or acylhydrazones<sup>267,268</sup> among others. In this section we will focus our attention on the photomodulation of the structure, properties, and functions of macromolecular helical structures.<sup>269</sup>

**2.5.1 Covalent helical polymers.** Feringa and coworkers reported the preparation of a polyisocyanate (PIC) with photo-switchable screw sense (poly-110) (Fig. 52a). To this end, a single chiral molecular motor was introduced as the initiator of the PIC. Photoisomerization of the initiator triggers the chiral

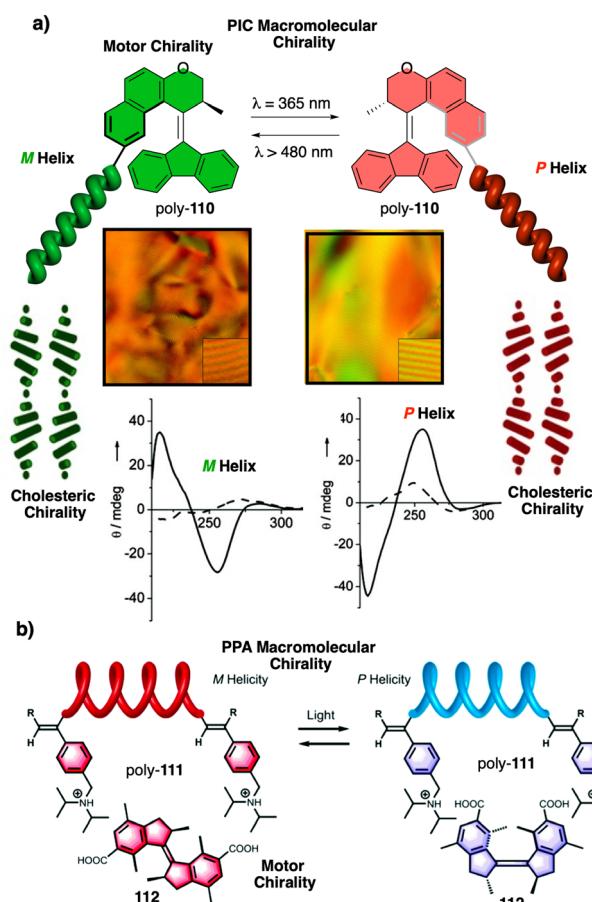


Fig. 52 (a) Helix inversion process of poly-110 bearing a chiral photo-active molecular motor and subsequent hierarchical chiral transmission from molecular motor to helical PIC cholesteric phase. Reproduced from ref. 271 with permission from the American Chemical Society, copyright 2008. (b) Helix inversion process of PPA poly-111 via salt-bridge formation with chiral molecular motor 112. Reproduced from ref. 272 with permission from the Royal Society of Chemistry, copyright 2017.

inversion of the macromolecular PIC helix through a conformational communication mechanism, which is further transferred, in a hierarchical way, to the cholesteric liquid crystalline phases formed by these polymers (Fig. 52a).<sup>270,271</sup>

The same group also reported the photoswitchable screw sense of an achiral PPA by establishing a supramolecular interaction between the pendant and a chiral photoswitchable molecular motor. They prepared a PPA that bears a trialkyl amine as pendant (poly-111), which was combined with a chiral photoswitchable molecular motor functionalized with two carboxylic acids (Fig. 52b). From this system they observed that photomodulation of the motor chirality triggers the helical inversion of the PPA main chain, as inferred by ECD spectroscopy (Fig. 52b).<sup>272</sup>

Zentel reported several examples of light-driven helical inversion of PIC copolymers consisting of achiral monomers bearing an alkyl chain and chiral monomers bearing a photo-switchable azo group (poly-113 and poly-114) (Fig. 53).<sup>273–277</sup> Depending on the chemical structure of the chiral azo monomer, the *E/Z* photoisomerization process produced either helical inversion (poly-113) or screw sense excess variation (poly-114) (Fig. 53a and b). Due to the good photostability and reversibility of the isomerization process, the PIC screw sense of poly-113 and poly-114 can be switched back and forth several times, opening venues for the preparation of chiroptical switches.

**2.5.2 Supramolecular helical polymers.** Light has been applied as stimulus to modulate the properties of supramolecular polymers. The introduction of photoresponsive groups in the monomer building blocks made it possible to modify the optical, chiroptical and morphological properties of different types of these macromolecular entities.<sup>278</sup> Meijer's group has recently employed light as an orthogonal external stimulus to modulate the assembly of BTAs through chemical modification

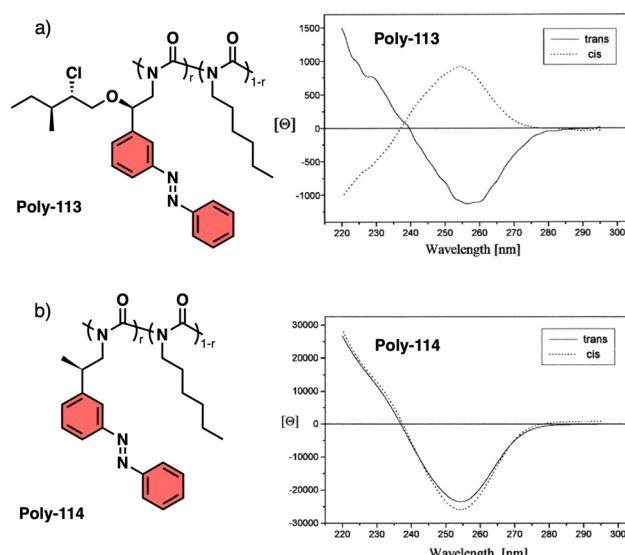


Fig. 53 Structures of PICs (a) poly-113 and (b) poly-114 copolymers and CD monitoring of the helical variations in the *E/Z* photoisomerization processes. Reproduced from ref. 276 with permission from the American Chemical Society, copyright 1998.

and photoisomerization of the side chains. Specifically, they employed light to covalently modify the side chain of a cysteine-based BTA (**115**) after a thiol-ene reaction. As the photoreaction proceeds, the hydrogen bonding pattern of the newly functionalized BTA (**116**) changes from one-dimensional self-assembled stacks to homo and heterodimers, as proven by several spectroscopic techniques (Fig. 54a and b).<sup>279</sup> In a different example, a BTA derivative bearing an acyl hydrazone as side chain (**117**) effectively co-assembles as an intercalator with a classical C<sub>3</sub>-symmetrical BTA **52** (Fig. 54c). Photoirradiation triggers *E/Z* isomerization of the acyl hydrazone moiety and, consequently, switches its behavior from intercalator to chain capper in a reversible way.

This process modulates the polymer chain length and triggers a macroscopic sol-gel transition (Fig. 54c and d).<sup>280</sup>

Diarylethenes (DAEs) represent a family of photoswitches whose structure can be tuned from an open to a close conformation by light irradiation. This chemical transformation not only changes the optical properties of the molecule, but also its rigidity. This fact makes DAEs very interesting molecules to

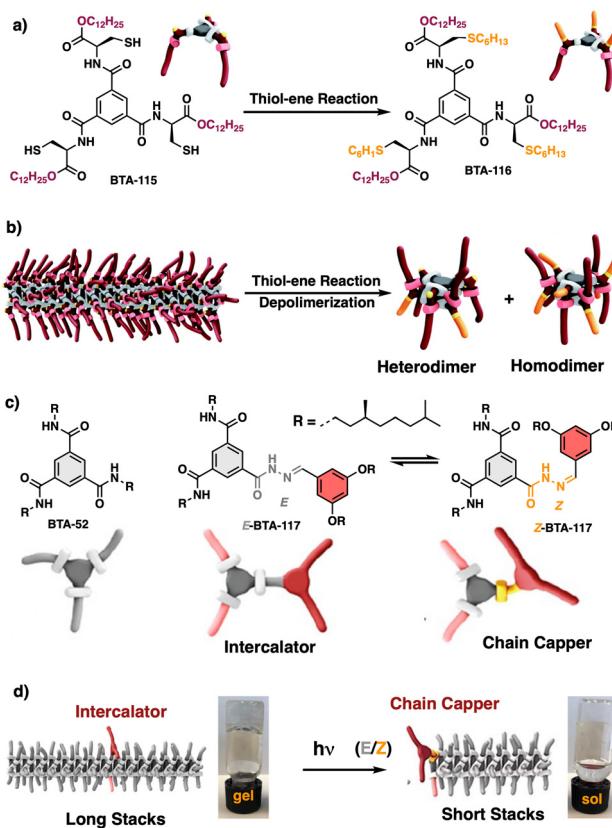


Fig. 54 (a) Structure of BTAs **115** and **116**, bearing free and alkylated cysteine moieties respectively. (b) Schematic illustration of the photo-depolymerization process from one dimensional stack to homo/heterodimers as the alkylation of the cysteine moieties takes place. Reproduced from ref. 279 with permission from the Royal Society of Chemistry, copyright 2021. (c) Structure of BTAs **52** and **117**. (d) Schematic illustration of the photo-triggered *E/Z* isomerization process of BTA **117** that triggers a change of function from intercalator to chain capper that is accompanied by a gel–sol state. Reproduced from ref. 280 with permission from the American Chemical Society, copyright 2020.

create photoactive functional supramolecular aggregates. For instance, Zhu reported the self-assembly of the DAE **118**, which is functionalized with naphthylamide units carrying a phenylalanine-OH functional group (Fig. 55a). The supramolecular polymerization of the system is assisted by the formation of carboxylate bridges that generate macromolecular helical structures in CCl<sub>4</sub>. The handedness of the systems depends on the absolute configuration of the amino acid, while the morphology of the aggregate depends on the open/closed state of the DAE group. Photoisomerization of the DAE moiety triggers a reversible morphological transformation from helical structures to nanosheets related to the change in the DAE scaffold (Fig. 55a–d).

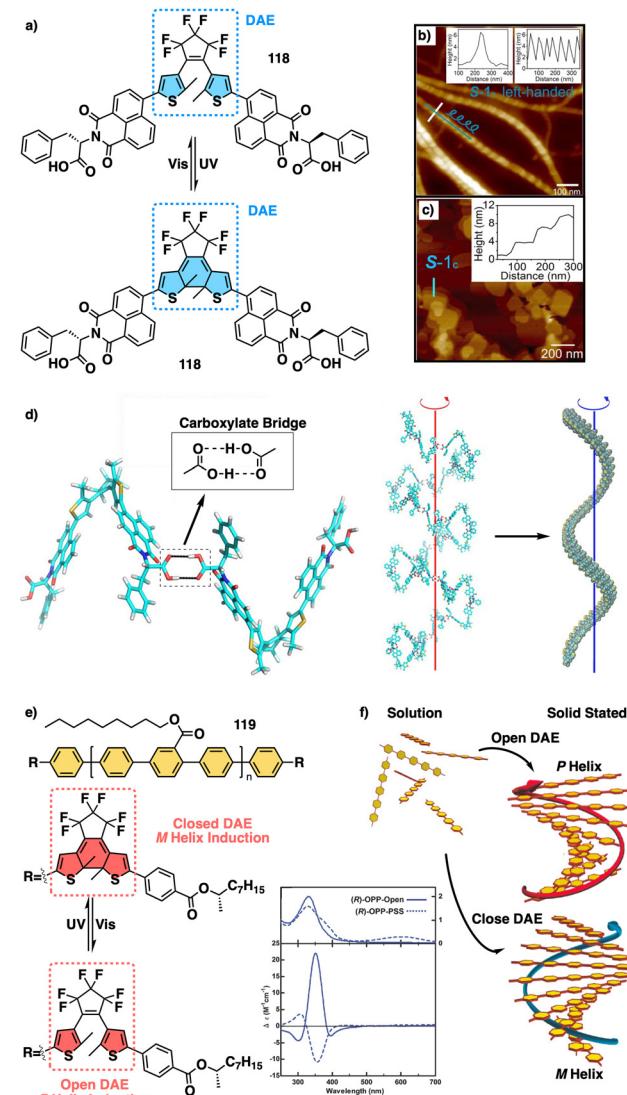


Fig. 55 (a) Structures of DAE-naphthylamide-Phe derivative **118** in open and closed states. Morphological transformation from (b) helical structures to (c) nanosheets upon light irradiation. (d) Schematic illustration of carboxylate bridge-triggered self-assembly generating helical structures. Reproduced from ref. 281 with permission from the American Chemical Society, copyright 2016. (e) Structure of molecule **119**. (f) Schematic representation of helix induction in open–closed states of DAE-OPP derivatives. Reproduced from ref. 282 with permission from John Wiley and Sons, copyright 2014.



Akagi reported the self-assembly of oligo(*para*-phenylenes) **119** functionalized with DAE bearing chiral esters. The macromolecular chirality of the system is reversible, showing inversion after irradiation with UV-vis light. This helical transformation is related to the open/closed form of the DAE moiety that promotes opposite handedness in the different states. Noteworthy, this process works both in solution and in solid state (Fig. 55e and f).<sup>282</sup> Feringa introduced a photoswitchable molecular motor in a supramolecular helical polymer building block endowed with a urea side chain (**120**) for the construction of photo-responsive supramolecular chiral aggregates (Fig. 56a).<sup>283–287</sup> In this system, the chirality of the molecular motor (*P,P*)-**120** is hierarchically transferred to supramolecular structures generating preferred-handed helical fibers.

Photoirradiation triggers rotation-induced morphological changes (*e.g.*, chiral, and geometric) that dramatically influence

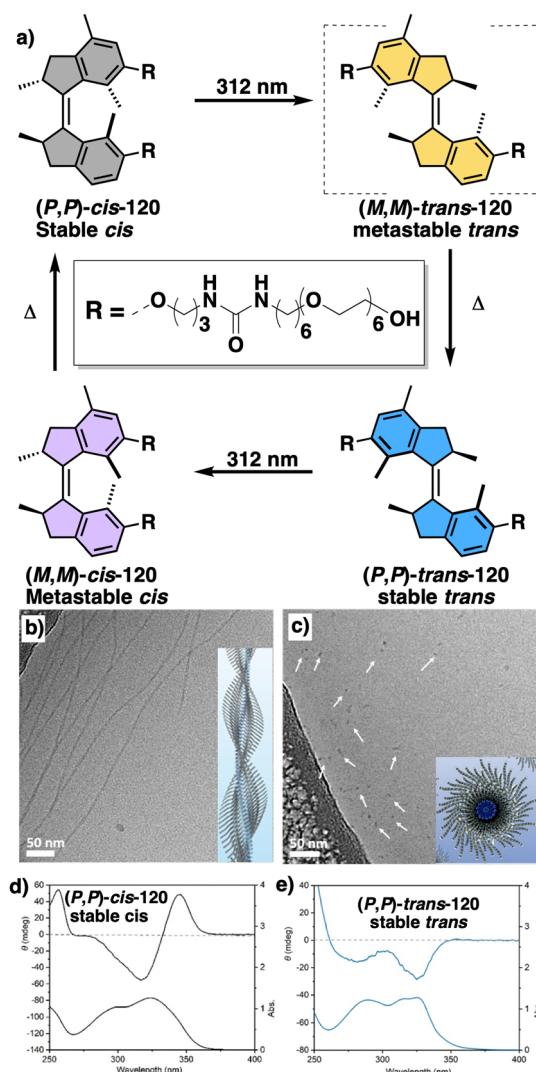


Fig. 56 (a) Structures of photoactive chiral molecular motor **120** and illustration of multistate rotation. Cryo-TEM image images of (b) *(P,P)-cis-120* and (c) after irradiation with 312 nm light to reach *(P,P)-trans-120*. CD and UV-vis spectra of (d) *(P,P)-cis-120* and (e) *(P,P)-trans-120*. Reproduced from ref. 290 with permission from the American Chemical Society, copyright 2022.

the morphology and chirality of the supramolecular polymer (Fig. 56a–e). Through this process they achieved dynamic control of the supramolecular polymer in multiple self-assembled states. Therefore, a cascade of morphological transformations from initial helical fibers to micelles, worm-like micelles, to finally recovering helical fibers, is possible simply playing with light irradiation.<sup>288</sup> Yagai designed a cone shape alkoxyazobenzene functionalized with chiral amides that self-assemble to generate uniform toroidal nanostructures that can hierarchically organize into nanotubes or axially chiral nanofibrils under the control by light, temperature, or concentration.<sup>289,290</sup>

**2.5.3 Foldamers.** Feringa introduced oligo bipyridine moieties as side chains of a photoswitchable molecular motor (**121**) (Fig. 57a). In the presence of Cu(i), a metallofoldamer is formed that folds into a double helix [**121**-Cu(i)]. Photochemical isomerization of the molecular motor triggers not only the chiral

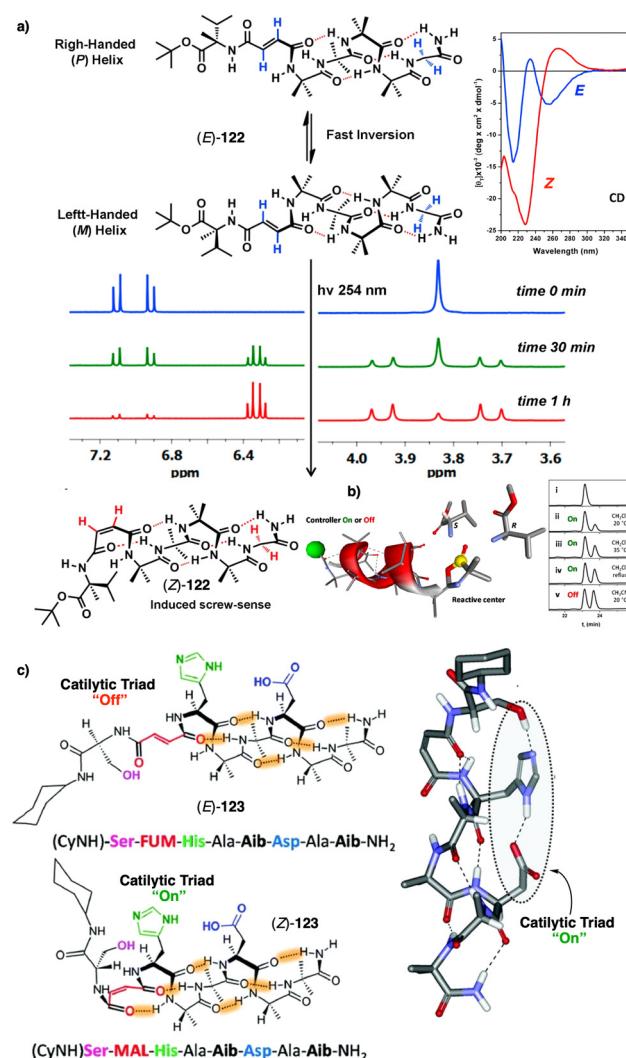


Fig. 57 (a) Structure of molecular motors endowed with bipyridine units. (b) Optimized molecular structures of *(P,P)-cis-121-Cu2* and *(M,M)-cis-121-Cu2*. (c) CD spectra showing the helical inversion process of these complexes. Reproduced from ref. 291 with permission from Springer Nature, copyright 2016.

inversion of the system but also the orientation of the coordinating bipyridine units (Fig. 57a).

This different orientation of the coordinating units promotes the formation of polymeric or discrete helical structures (*trans*- and *cis*-conformations, respectively). In addition to modifying the *cis*-/*trans*-equilibrium of the molecular motor, the chirality of the system is also modulable with light, so the (*P,P*)-**121** and the (*M,M*)-**121** conformations induce opposite handedness in the discrete double helical structures (Fig. 57b and c).<sup>291</sup>

Moretto and Clayden reported a series of AIB-based foldamers containing photoswitchable fumaramide/maleamide linkers to tune the *P/M* screw sense excess of the foldamers by light. These moieties bearing carbonyl groups present different compatibility with the hydrogen-bonding pattern of the foldamers, with the *Z* maleamide being compatible with the HB network of the foldamer while the *E* fumaramide configuration is not. Photoswitching from the insulating fumaramide to the conducting maleamide allowed activation of the stereoselective reactivity of **122**, as proven by chiral HPLC (Fig. 58a and b).<sup>292</sup>

Following this work, they have also reported the photoactivation of a catalytic triad of reactive side chains – Ser, His and Asp – present in foldamer **123**. Light modulation of the *E/Z* forms allows control of the reactivity of the system, as it only works in the maleamide form, where the reactive groups are placed in the proper orientation to catalyze the corresponding

reaction (Fig. 58c).<sup>292</sup> Clayden and coworkers also reported an azobenzene containing AIB foldamer **124** inserted into an artificial lipid membrane whose screw-sense was controlled by a configurational change in the azobenzene unit. The *E/Z* conformers stabilize different hydrogen bond patterns within the AIB foldamers, biasing the rate of the helix inversion process (Fig. 58d).<sup>293</sup> The introduction of fluorine atoms into the foldamer allowed direct monitoring of this process by <sup>19</sup>F NMR experiments (Fig. 58).<sup>291</sup>

Huc reported an aromatic oligoamide (**125**) bearing a DAE central linker (**S**) (Fig. 59a and b). Binding of fructose results in the induction of opposite handedness depending on the open/close state of the DAE unit, as demonstrated by CD (Fig. 59c and d). This fact indicates that the DAE/fructose binding occurs in a different way. Interestingly, subsequent irradiation of the host-guest complex allows switching between *P/M* helical scaffolds (Fig. 59e).<sup>294</sup> To date, many foldamer containing azobenzene groups has been reported, both as side chains and as part of the macromolecular main chain. The introduction and photomodulation of these azobenzene moieties allows control of the folding of the foldamer, and consequently, its properties.<sup>295</sup> For instance, the *E/Z* structural variation of the azobenzene group in the side chains of the foldamer creates variations in the supramolecular interaction that governs the folding of the systems.<sup>296–301</sup>

Pieroni and Ciardelli<sup>296</sup> reported examples on this topic in poly(glutamic) **126** derivatives and Ueno and Osa in poly(aspartic)<sup>300</sup> **127** derivatives (Fig. 60a and b). This strategy was also employed by Ciardelli and coworkers using spiropyrans **128** as photoresponsive side chains instead of azo derivatives (Fig. 60c).<sup>302</sup>

Short helical foldamers containing photoresponsive units in the main chain or in the side chains can also be found in the literature.<sup>295</sup> In these systems, the photoisomerization of the

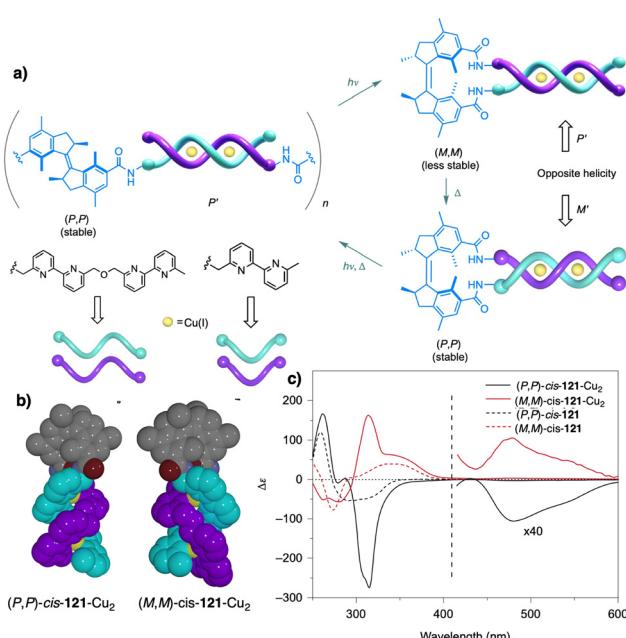


Fig. 58 (a) Screw sense induction process by light irradiation in AIB foldamer **122** demonstrated by CD and NMR. (b) Schematic representation of the diastereoselective chainextension reactions followed by chiral HPLC. (c) Light-driven turn on/off catalytic activity of **123** bearing a catalytic triad esterase. Reproduced from ref. 292 with permission from the American Chemical Society, copyright 2016. (d) Switching the terminal azobenzene chromophore between *E* and *Z* configurations changes the population distribution between right- and left-handed conformations of foldamers **124**. Reproduced from ref. 293 with permission from the Royal Society of Chemistry, copyright 2021.

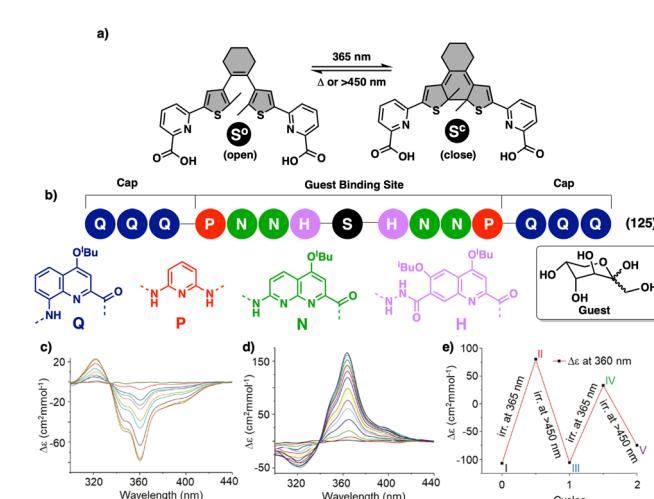


Fig. 59 (a) Structure of DAE core and (b) oligoamide sequence of **125**. CD spectra showing an increase of chiroptical activity after irradiation with different wavelengths: (c)  $\lambda = 365$  nm and (d)  $\lambda = 450$  nm. (e) CD switching upon subsequent irradiation cycles. Reproduced from ref. 294 with permission from the Royal Society of Chemistry, copyright 2021.



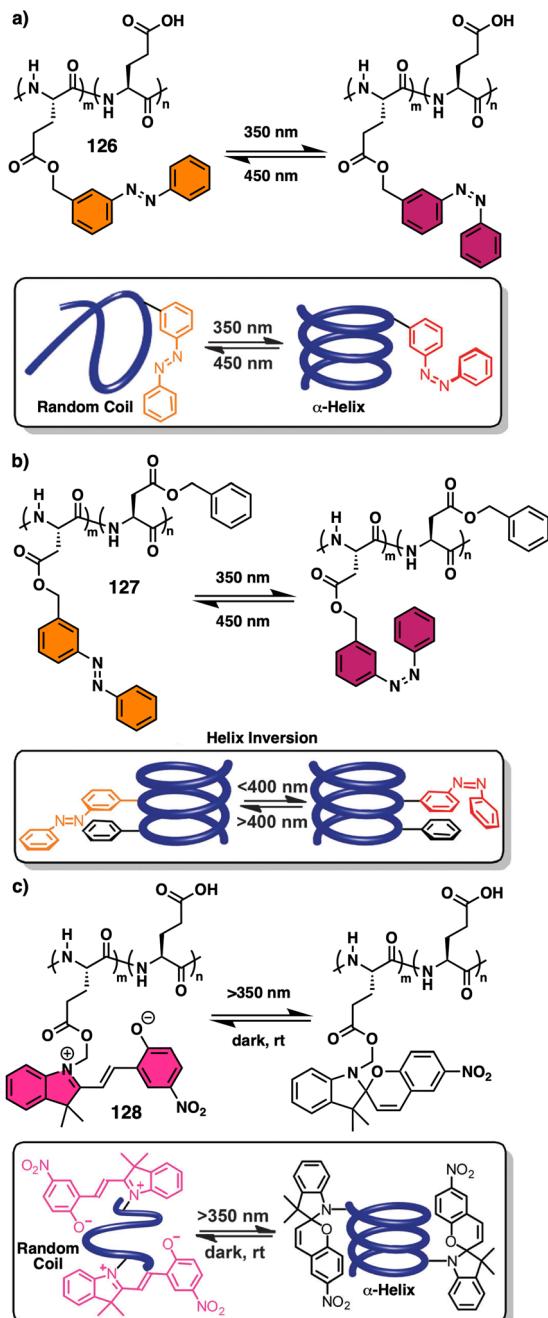


Fig. 60 Structure and behavior with light of  $\alpha$ -helical foldamers composed by (a) glutamic acid-azobenzene derivatives (126) and (b) aspartic acid-azobenzene derivatives (127). (c) Structure of glutamic acid-spiropyran derivative (128). A schematic illustration of the photoisomerization impact on the folding of the molecules is depicted below each section. Reproduced from ref. 295 with permission from the Royal Society of Chemistry, copyright 2016.

foldamer can trigger the unfolding of the system, a dimerization process, or affect their guest-binding ability.<sup>302–305</sup> As an illustration, Jiang designed a small foldamer containing an azobenzene and two phenyl-1,2,3-triazole moieties (129) that, through photoisomerization of the foldamer backbone, changes the binding affinity for anions (Fig. 61a).<sup>306</sup> Flood showed that

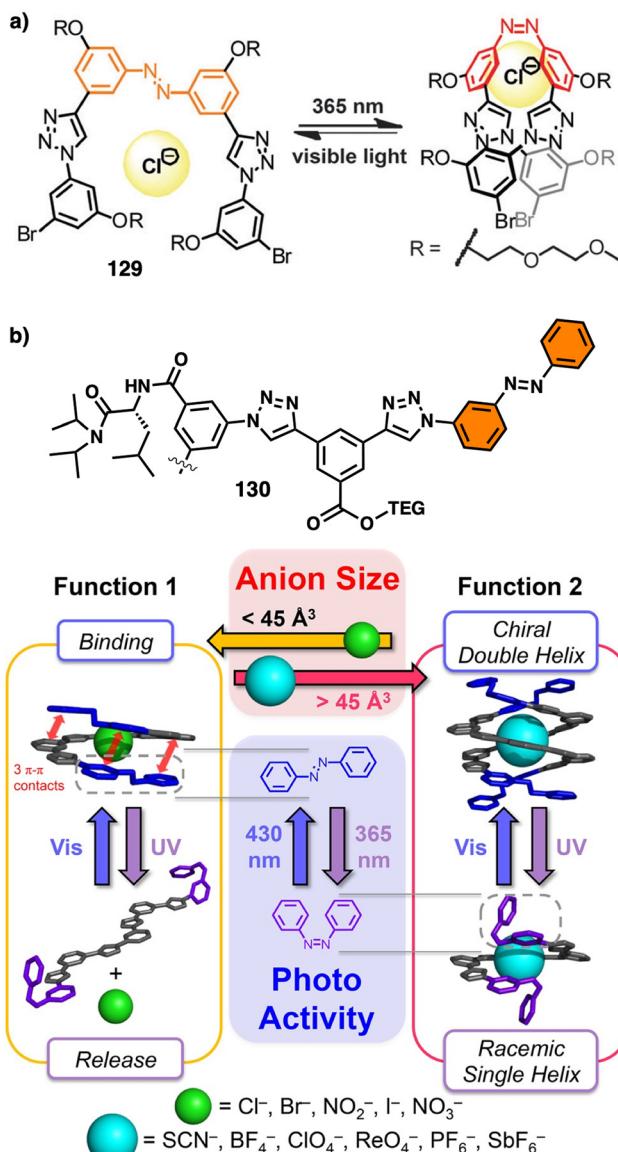


Fig. 61 (a) Structure of foldamer 129, binding to Cl<sup>-</sup> and light-driven conformational change. Reproduced from ref. 303 with permission from the American Chemical Society, copyright 2010. (b) Structure of foldamer 130 and schematic illustration of the cascade of structural transformations mediated by photoirradiation and anion binding. Reproduced from ref. 307 with permission from the American Chemical Society, copyright 2018.

foldamer 130, which contains a central benzene endowed with an amino acid (Leu) along with aryl triazoles-azobenzene units, can be used to bind or release small anions such as Cl<sup>-</sup>, depending on the *E* or *Z* configuration of the azo group.<sup>307</sup>

In case of using large anions such as BF<sub>4</sub><sup>-</sup> to interact with 130, the *Z/E* isomerization of the azo group leads to the formation of single or double helices.<sup>307</sup>

Foldamers based on oligo(*meta*-phenyleneethynylene)s (OmPES) provided inspiration for the preparation of photoresponsive materials by including of photoresponsive azobenzene groups in the side chains or, mainly, within the sequence.<sup>308–314</sup> Based on these designs, Hecht reported several examples of photoresponsive

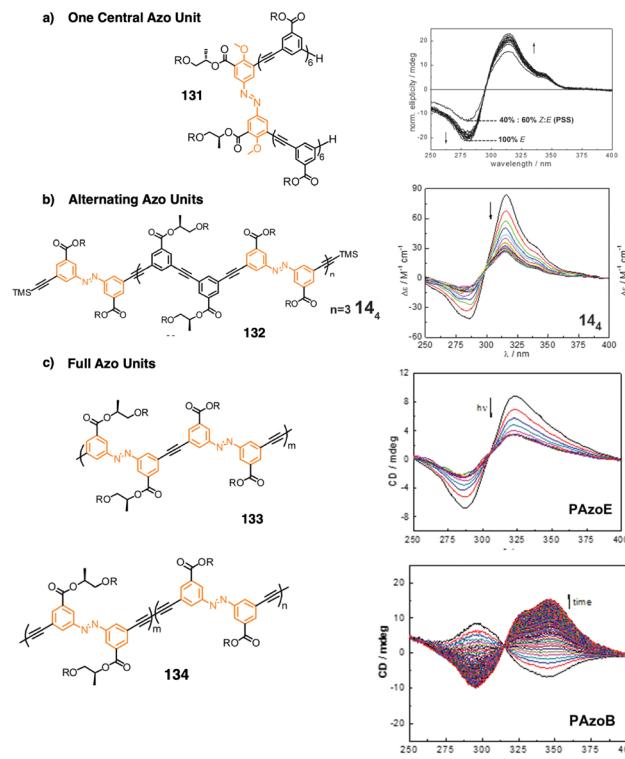


Fig. 62 Structures and CD spectra showing variations in the helical structures of foldamers containing (a) one central azo group (**131**) (reproduced from ref. 312 with permission from John Wiley and Sons, copyright 2006). (b) Alternating azo units (**132**) Reproduced from ref. 313 with permission from the Royal Society of Chemistry, copyright 2013 and (c) composed by azo units with alkynyl connectors of different lengths (**133** and **134**) Reproduced from ref. 314 with permission from John Wiley and Sons, copyright 2015. Otherwise, stated variations in the ECD spectra correlate with irradiation time.

OmPEs foldamers bearing, for instance, only the central azo unit (**131**) (Fig. 62a),<sup>315–319</sup> different alternating orientations of azobenzenes within the OmPEs-azo sequence (**132**) (Fig. 62b),<sup>313</sup> or composed entirely of azobenzene units (**133**) (Fig. 62c).<sup>313,316</sup> Most of the reported examples show variations in their folding degree upon photoirradiation. Generally, a decrease in the intensity of the CD signal is observed once the corresponding photostationary state is reached.<sup>295</sup> Representative examples of the structures and their chiroptical properties are depicted in Fig. 62. Hecht also reported that slight variations in the connectivity of the units that make up the foldamers have a large impact on their chiroptical responses.

Replacing the alkynyl connector in **133** by a bisalkynyl unit resulted in self-assembly of foldamer **134**, leading to a CD inversion. The assembly/disassembly process is easily controllable by simply irradiating the sample (Fig. 62c).<sup>314</sup>

## 2.6 External stimulus: circular polarized luminescence

Circular polarized luminescence (CPL) is an intriguing property of certain types of chiral luminophores that emit light with preferred handedness.<sup>317–322</sup> In recent years, this type of materials at both the molecular,<sup>323–325</sup> macromolecular and supramolecular

levels<sup>326–329</sup> have found applications in the development of CP-OLEDs,<sup>330–332</sup> security encryption systems,<sup>333</sup> tags for biology<sup>334</sup> and asymmetric synthesis,<sup>335</sup> in among many others. Although CPL emission is a well-established research topic whose importance is constantly growing year by year, the application of CP-light as an asymmetry inducer is still in its infancy. Therefore, a small number of examples on this topic can be found in the literature.<sup>336</sup> Here we summarize some of the most relevant examples of asymmetry induction in covalent and supramolecular helical polymers.

**2.6.1 Covalent polymers.** The first example of asymmetry induction by CP-light in helical polymers was reported by Selinger, Green and coworkers.<sup>337</sup> They prepared a series of co-poly(isocyanate)s (PIC)—(poly-X<sub>n</sub>-co-Y<sub>1-n</sub>) copolymer series **135**, **136**, **137** depicted in Fig. 63a—composed by a monomer bearing an achiral alkyl group and another monomer possessing a photoresolvable ketone pendant group (Fig. 63b). As expected, for a racemic helical polymer, both helical senses are equally populated and do not show a circular dichroism spectrum. However, after irradiation with CP-light, the ketone pendant (Fig. 63c) exhibits partial photoresolution. Thus, following the Majority Rules effect,<sup>114</sup> an efficient chirality transfer from the enantio-enriched pendant to the polymer backbone results in an amplified photoresolution.

The *ortho*-, *meta*- or *para*-linkage of the photoactive moiety to the polymeric backbone is also a key factor in the transfer of asymmetry from the pendant to the main chain, losing effectiveness when we go from *ortho*, to *meta* and *para*. It is important to highlight that the helical sense of the polymer can be switched reversibly by alternating irradiation with (+)- or (-)-CP-light and

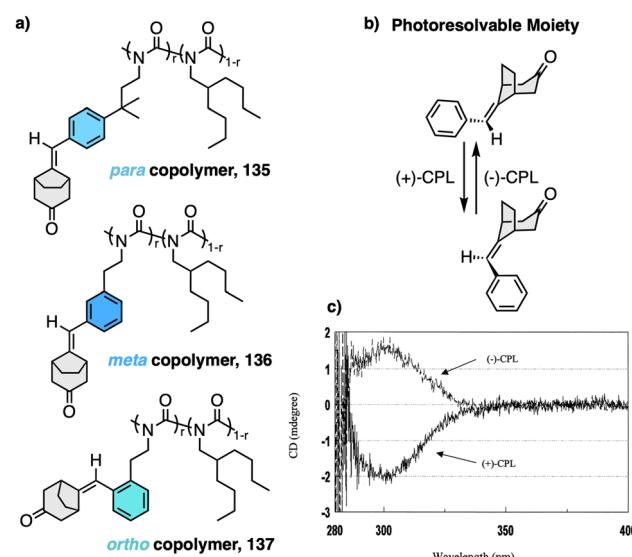


Fig. 63 (a) Structure of the copolymers employed in this work **135**, **136** and **137**. (b) Schematic illustration of the photo-resolution of the chiral moiety by irradiation with CP-light. (c) ECD spectra of **136** showing the asymmetry induction after irradiation with (+) or (-)-CP-light. Reproduced from ref. 337 with permission from the American Chemical Society, copyright 2000.



return to the axially racemic state by irradiation with non-polarized light.<sup>21</sup>

Nakano and coworkers induced a preferred handedness in polyfluorene derivatives **138–140** by irradiation with either (+)- or (−)-CP-light (Fig. 64a).<sup>60</sup> They found that the helix induction mechanism is based on a photoderacemization process in the photostationary state. This is conceivable through a chirality amplification process that takes place through the deactivation of a coplanar conformation in polyfluorene leading to right- and left-handed twisted dyads.

Asymmetry induction could be achieved for polymers with longer alkyl chains [*i.e.*, poly(DOF) **138**, octyl chain, and poly(DDF) **139**, dodecyl chain], while it was not feasible for the one carrying a shorter chain [hexyl chain, poly(DHF) **140**], which required an auxiliary molecule to aid the induction of chirality. The chirality-induced samples in the film state remained stable for years if stored protected from light irradiation and at room temperature (Fig. 64b and c).<sup>60</sup> Natansohn reported asymmetry induction in polymethylmethacrylate **141** endowed with photoresponsive azobenzene units as side chains (Fig. 65a). The generated liquid crystalline phase takes advantage of the photoisomerization of all azobenzene groups in the film plane, inducing a preferred arrangement of the chromophores that changes from a random to a chiral environment (Fig. 65b). The chirality of the films can be exchanged after subsequent irradiation cycles with either (+) or (−)-CP-light, giving rise to interesting chiroptical switches in the condensed state (Fig. 65c and d).<sup>338</sup> Similarly, Li, Yang and coworkers reported the impact of the linker between the azobenzene unit that now holds a stereogenic center and the PMA backbone of **142(x)** ( $x = 0–6$ ) (Fig. 65e) in the light-driven asymmetry induction.

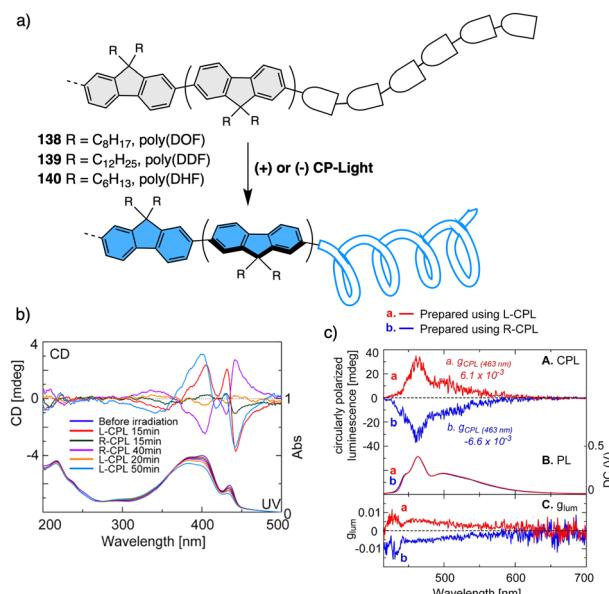


Fig. 64 (a) Schematic illustration of the asymmetry induction by CP-light irradiation and structures of polymers **138–140**. (b) ECD monitoring of asymmetry induction after exposure to CP-light of poly(DOF) film (**138**). (c) CPL spectra of photo-induced films of poly(DOF) (**138**). Reproduced from ref. 50 with permission from the American Chemical Society, copyright 2018.

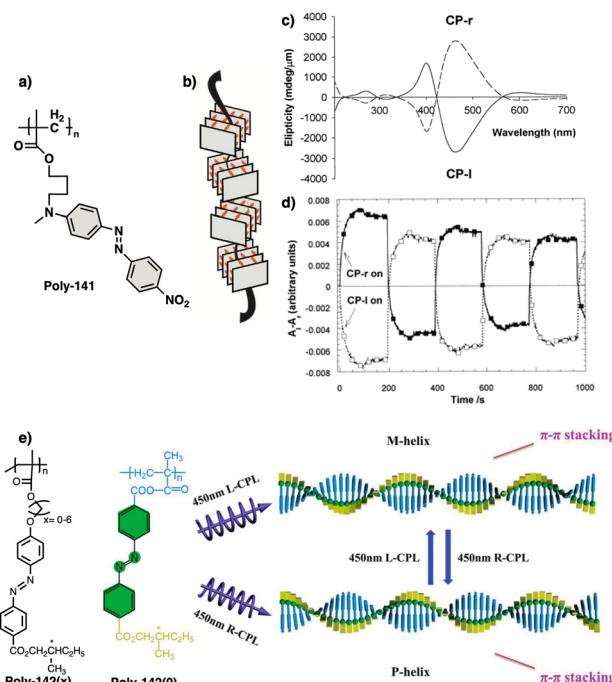


Fig. 65 (a) Structure of poly-141. (b) Schematic illustration of the cholesteric phase generated upon CP irradiation. Reproduced from ref. 338 with permission from the American Chemical Society, copyright 2000. (c) ECD spectra of thin films showing the helical orientation of the molecules in the crystalline phase after irradiation with (L) or (R)-CP-light. (d) Several cycles of chirality switching in the crystalline phase by subsequent irradiation with CP-light of different chirality. (e) Structures of Azo polymers and schematic illustration of the supramolecular chiral structures of poly-142(0) induced by 450 nm CP-light. Reproduced from ref. 339 with permission from John Wiley and Sons, copyright 2022.

They found that shorter linkers did not produce efficient chirality harvesting after thermal annealing, while the other derivatives, bearing longer linkers (from Azo-PMA-142(**1**) to Azo-PMA-142(**6**)), exhibited effective asymmetry transfer after thermal annealing. Remarkably, all derivatives show asymmetry induction after irradiation with (+) or (−)-CP-light, except **142(6)** which can only be folded/unfolded by UV light irradiation.<sup>339</sup> In the field of azobenzene polymer derivatives, Meijer, Vantomme and Meskers reported alternating copolymers formed by azobenzene and fluorene units (poly-**143**) that form cholesteric phases with strong optical activity upon thermal annealing (Fig. 66a). Taking advantage of the *trans/cis/trans* isomerization of the azobenzene units, the macromolecular helicity and organization of the polymer chains could be modulated by CP-light (Fig. 66b–e). Remarkably, due to the diastereomeric nature of the polymer chains and the light, there is asymmetry in the interconversion pathway.<sup>340</sup> The same team also reported an azobenzene–oligo-siloxane alternating copolymer that shows a similar behavior, the formation of a liquid crystalline chirality that can be controlled by CP-light irradiation.<sup>341</sup>

**2.6.2 Supramolecular polymers.** Circular polarized luminescence has also emerged as an effective tool to induce asymmetry in supramolecular polymers.

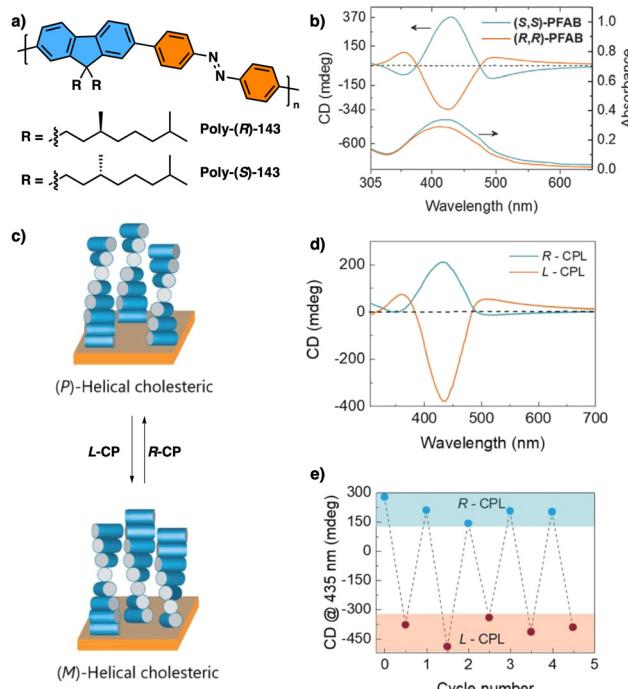


Fig. 66 (a) Structure of alternating poly-**143**. (b) ECD spectra showing the helical orientation of the molecules in the crystalline phase after thermal annealing. (c) Schematic illustration of the cholesteric helical inversion after irradiation with (L)- or (R)-CP-light. (d) ECD spectra showing the helical inversion after irradiation with (L) or (R)-CP light. (e) Rewritable chirality of the crystalline phase by subsequent irradiation with CP-light of different chirality. Reproduced from ref. 340 with permission from John Wiley and Sons, copyright 2021.

Kim, Seo and coworkers reported the asymmetry induction by CP-irradiation of triphenylamine<sup>342–346</sup> derivative **144a** bearing diacetylene units (Fig. 67a).

Irradiation with CP-light produced a chiral bias in the arrangement of the triphenylamine core that was subsequently amplified throughout the supramolecular polymerization process. Further irradiation with light triggered the photopolymerization of the diacetylene units, generating the corresponding covalent polymer with memorized chirality (Fig. 67a–c).<sup>343</sup> They also showed that supramolecular chirality can emerge from a transiently chiral molecular building block by transmitting chiral information inherited from the CP-light irradiation.<sup>344</sup> Consequently, a chiral radical cation was generated that aggregated into preferred-handed helical stacks (Fig. 67d). The nonlinear screw sense excess as a function of CPL irradiation time is reminiscent of the chiral induction expressed by the Sergeants-and-Soldier principle, where the co-assembly of a transiently chiral Soldier with a permanently chiral Sergeant takes place. Moreover, matching the handedness of the CPL to the Sergeant's direction produced an abnormally enhanced chiral response. However, in the mismatched scenario (CPL vs. mismatched Sergeant's direction) (Fig. 67e), a critical molar fraction of the Sergeant was found to bifurcate the bias: above a critical point, the helicity determined by the Sergeant persisted. Below the critical point, the CPL can override the molecular

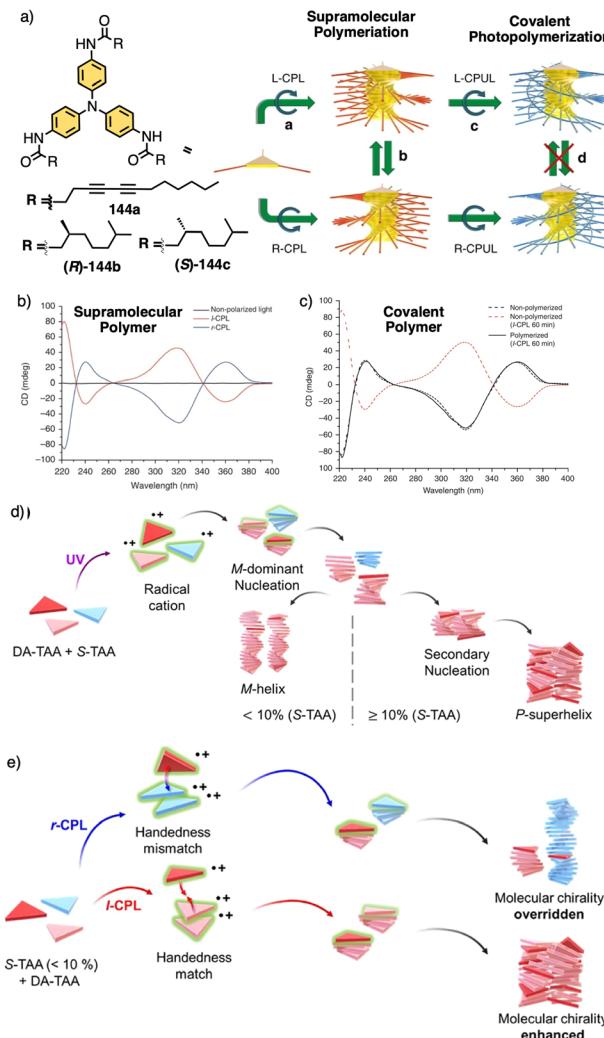


Fig. 67 (a) Structures of derivatives **144a–c** and schematic illustration of the asymmetry induction and photopolymerization of **144a**. ECD spectra showing the mirror images generated after (b) CP-irradiation and (c) photopolymerization. Reproduced from ref. 343 with permission from Springer Nature, copyright 2015. (d) Sergeants and Soldiers coassembly of **144a** with (S)-**144c** as a molecular Sergeant: Schematic depiction of the copolymerization to *M*-helix and *P*-superhelix formation. (e) Amplifying and overriding molecular chiral information in the supramolecular state by CPL: Schematic depiction of **144a**/(S)-**144c** copolymerization upon l- and r-CPL exposure. Reproduced from ref. 344 with permission from the American Chemical Society, copyright 2022.

chirality with increasing irradiation time. Therefore, the authors demonstrated that supramolecular chirality can be developed following the handedness of the external stimulus and overturn—in some extent—the one imposed *via* molecular constraints (Fig. 67e).<sup>344</sup> Recently, Raynal and coworkers demonstrated how the presence of radical species in triarylamine triamides affects to its self-assembly.<sup>345</sup> Zou reported a similar CP-light asymmetry induction in porphyrins **145** endowed with diacetylene moieties (Fig. 68a).

Irradiation of these molecules with (+) or (–)-CP-light generated photo-induced protonation of the basic internal rim of the macrocycle, producing *P*- or *M*-helical stacks, respectively



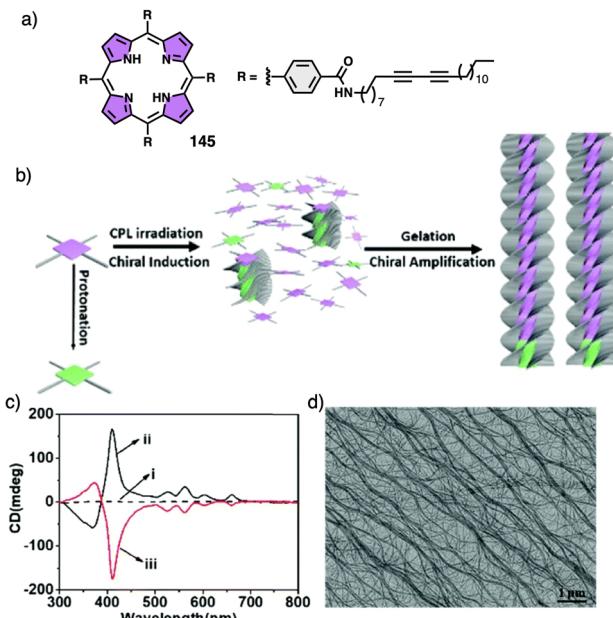


Fig. 68 (a) Structure of the porphyrin **145** used in this work. (b) Schematic illustration of the photoinduced asymmetry induction. (c) ECD showing the induction of mirror image helical structures upon irradiation with (+)- or (−)-CPL. (d) TEM image of the supramolecular fibbers generated upon self-assembly of **145**. Reproduced from ref. 346 with permission from the Royal Society of Chemistry, copyright 2019.

(Fig. 68b–d). Further irradiation with non-polarized light triggered the polymerization of the diacetylene moieties yielding the corresponding covalent polymer with memorized handedness.<sup>346</sup> Sánchez reported asymmetry induction in the supramolecular polymerization of achiral *N*-annulated perylene derivative **146** (Fig. 69a). Irradiation with (+) or (−)-CP-light during the elongation process results in the adoption of preferred *P* or *M* helical assemblies as determined by ECD (Fig. 69b and c). Additionally, after removing CP irradiation, these systems exhibit a “memory” that shows, after 2 hours, an ECD trace virtually identical to that initially prepared.<sup>120</sup>

Lee and co-workers reported the co-assembly of amidoamine **147** with monoacid **148** bearing diacetylene units that self-assemble to generate supramolecular ribbons in both gel and film states

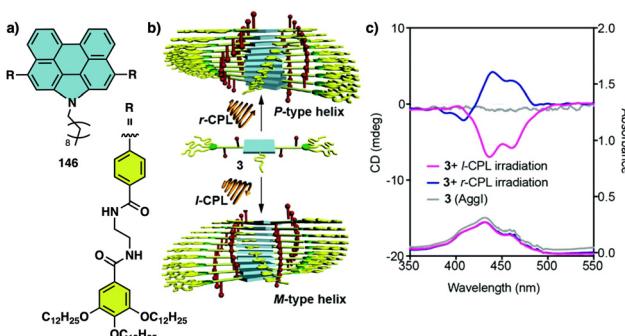


Fig. 69 (a) Structure of **146**. (b) Schematic illustration of the asymmetry induction by CP-light irradiation. (c) ECD Spectra of the opposite handed helical structures generated by irradiation with CP-light. Reproduced from ref. 120 with permission from the Royal Society of Chemistry, copyright 2020.

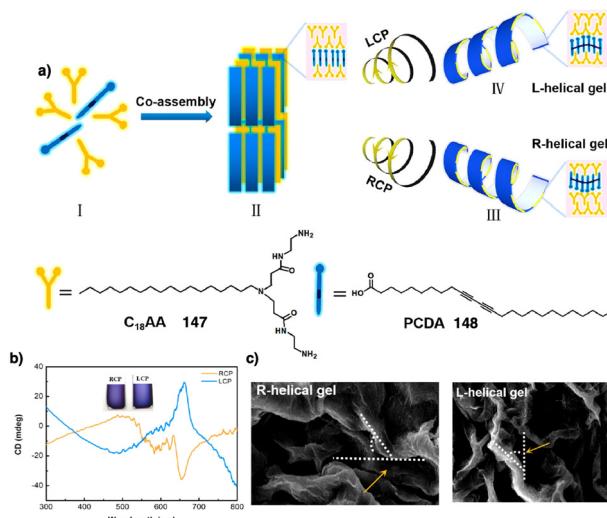


Fig. 70 (a) Structures **147** and **148** and schematic illustration of the asymmetry induction by CP-light irradiation of **147**–**148** co-assembly. (b) ECD spectra showing opposite chirality after irradiation with *L*/*R* CP-light. (c) SEM images of the gels with different handedness. Reproduced from ref. 347 with permission from the American Chemical Society, copyright 2022.

(Fig. 70). After irradiation with (+) or (−) CP-light, these ribbons fold into preferred left- or right-handed structures, as determined by ECD and SEM visualization.<sup>347</sup>

## 2.7 External stimuli: solvents

The main endoergic contribution to solute–solvent interactions is the formation of a cavity in the solvent to accommodate the solute. This is a chemical property of the solvent, which depends on the association of its molecules in a liquid state.<sup>348</sup> The interactions between solvents and solutes are of paramount significance in the field of organic chemistry. They play a central role in governing various aspects, such as solubility, reactivity, and structural characteristics of the compounds involved.

**2.7.1 Solvent-responsive foldamers.** Helical folding in foldamers is a complex process influenced by both internal and external factors.<sup>14,349</sup> The shape and rigidity of monomers, as well as intermolecular interactions, such as attraction and repulsion, rotational restrictions, solvophobic effects or aggregation, play a role in the folding behaviour.

An important parameter that affects the folding of the foldamers is the nature of the solvent, which greatly affects their secondary structure. For instance, oligophenylethylenes are also called “solvophobic foldamers” due to their sensitivity to solvents.<sup>350–352</sup> On the other hand, the aromatic amide foldamers developed by Huc and co-workers showed remarkable solvent folding stability in a wide range of solvents,<sup>353</sup> which is quite unique compared to other foldamers. However, this apparent and atypical lack of solvent dependence led them to carry out more in-depth studies on the subject. Thus, a helical aromatic oligoamide foldamer (**149**) bearing triethyleneglycol (Tg) side chains (Fig. 71a) was prepared to study its folding in a wide range of solvents.<sup>354</sup>

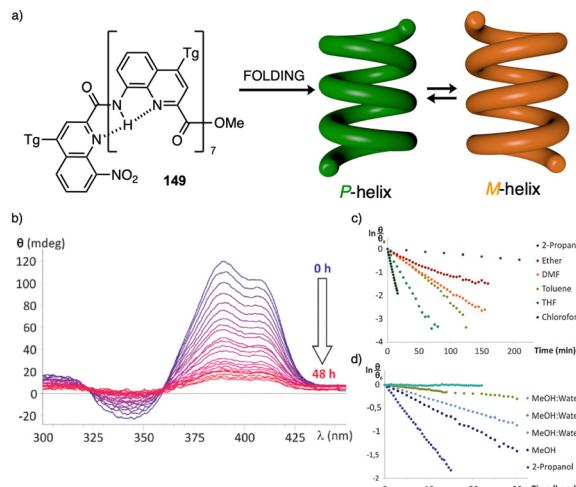


Fig. 71 (a) Chemical structure of foldamer **149** and its folding into a mixture of *P* and *M* helices. Time dependent ECD racemization studies for *P*-**149** as a function of solvent in (b) MeOH, (c) different solvents and (d) MeOH:Water mixtures. Reproduced from ref. 354 with permission from the Royal Society of Chemistry, copyright 2012.

Although the polymer is achiral, the two *P* and *M* helical structures of **149** were isolated by chiral HPLC when folded into an axially racemic mixture, allowing their exceptional stability to be determined (Fig. 71a). The helices did not racemize during the chromatography studies. By performing time dependent ECD studies in different solvents (Fig. 71b-d), they found that in chloroform, which was believed to provide highly stable conformations, turned out to be one of the least favourable solvents for these foldamers (Fig. 71c).

On the other hand, protic solvents,<sup>355</sup> such as methanol–water mixtures, demonstrated a remarkable ability to stabilize helical conformations (Fig. 71d).

Another example of solvent-dependent aromatic foldamers was reported by Li who prepared hybrid oligomers containing one aromatic amide trimer or pentamer and one or two aromatic 1,2,3-triazole tetramers (**150**) (Fig. 72a).<sup>356</sup> In this case, the researchers explored a conformational communication mechanism to induce a helical sense in the 1,2,3-triazole tetramer from the aromatic amide region. It was found that while in polar solvents the conformational communication from the aromatic amide to the triazole tetramer stops, in low polar solvents the conformational communication flows between the two fragments (Fig. 72b).

Flood and Liu described a more complex system where two stimuli, solvent, and anions, were combined to produce a structural change from single to double helices.<sup>357</sup> Thus, they studied a series of *C*<sub>2</sub>-symmetric aryl-triazole foldamers (**151**) (Fig. 73a) in which the interconversion between chloride-stabilized single and double helices can be understood and controlled by sequence modification strategies. Interestingly, a 1:1 single to 2:2 duplex (Fig. 73c) helical switch could be obtained at room temperature by increasing the solvent polarity (Fig. 73b and d), indicating that solvophobically driven  $\pi$ -stacking plays a critical role in stabilization of the 2:2 double

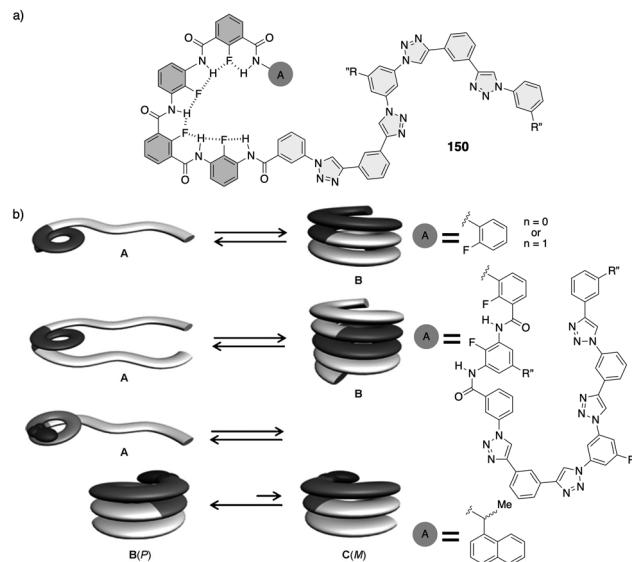


Fig. 72 (a) General foldamer structures **150** used in this work. (b) Schematic representations of the solvent-dependent (A polar; B low-polar) folding-induced processes for the different **150** derivatives. Reproduced from ref. 356 with permission from John Wiley and Sons, copyright 2014.

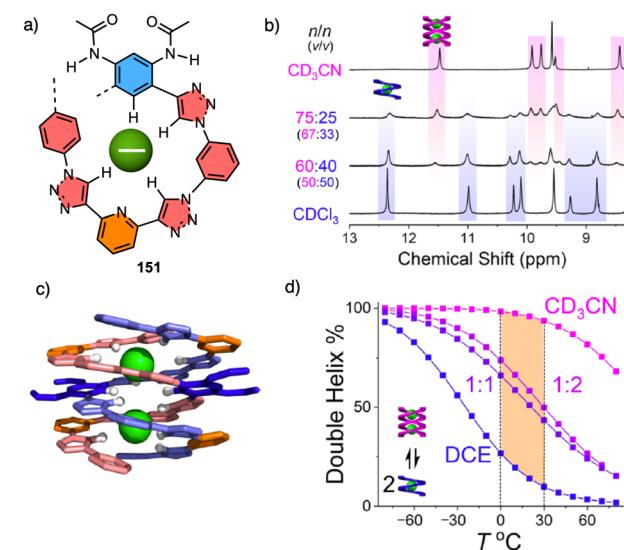


Fig. 73 (a) Chemical structure of anion foldamer host **151**. (b) Double helix formation of **151**-Cl<sup>-</sup> in response to solvent polarity (5 mM, 15 equiv. of TEACl). (c) DFT-optimized geometry of the 2:2 double helix formed by **151**-Cl<sup>-</sup>. (d) Two factors combined showing maximal solvent-induced swings in double helix population around ambient temperatures (orange shade). Reproduced from ref. 357 with permission from the American Chemical Society, copyright 2018.

helix compared to the single helix. For instance, there was a notable increase in the population of the double helix as the proportion of acetonitrile in chloroform or DCE increased (Fig. 73b). This indicates that the enhanced stability of double helices in polar solvents is associated with greater burial of  $\pi$  surfaces within the double helix structure.



Another family of foldamers whose helical structure is also solvent dependent are the AIB containing foldamers described by Clayden.<sup>1</sup> These foldamers tend to form a  $3_{10}$ -helix that shows a solvent dependent decay of screw-sense excess induced by chiral residues introduced in both ends (**152**) or at one end (**153**) of the foldamer chain.<sup>358,359</sup> The observed signal decay was attributed to the occurrence of 'tendril perversions', which are characterized by random breaks in hydrogen bonds along the helix (Fig. 74a). These breaks cause a helical reversal in the specific location where they occur, producing the observed

decay in the signal. After exhaustive study using a variety of analytical and computational techniques, the intrusion of 'tendril perversions' was shown to have an effect analogous to a  $\gamma$ -turn, a less prevalent turn that involves the formation of a hydrogen bond between residues  $i$  and  $i + 2$  (Fig. 74b). Furthermore,  $\gamma$ -turn conformations were also found to be more frequent in polar solvents, as tendril perversion increases from 0.5% per residue in THF to 6% per residue in methanol (Fig. 74c).

In supramolecular helical polymers, monomeric repeating units are held together by non-covalent interactions. The dynamic nature of these interactions makes that the nature of the solvent can affect supramolecular polymerization pathways.<sup>360</sup> Consequently, investigating interactions between supramolecular polymers and solvents remains challenging as solvent molecules will participate in the molecular stacking arrays of various ways. So, in recent years detailed solvent effects on supramolecular polymers can be found in several excellent works, focusing on stability, kinetics, pathway selection, and structure.<sup>28,361–364</sup> This review will focus mainly on supramolecular polymers whose supramolecular helix evolves into a different one due to the action of solvents.

**2.7.2 Solvent-responsive supramolecular polymers.** For instance, Meijer demonstrated how the morphology of bipyridine-decorated BTA aggregates (**154**) can be tuned by the properties of the solvent mixture itself (Fig. 75).<sup>365</sup> In this work, the correlation between the formation of single or triple helical superstructures of bipyridine-decorated BTAs (**154**) and the water content in water-isopropanol mixtures was found to be significant. Therefore, amphiphilic discotic molecules self-assemble into supramolecular fibres formed by one or three discotic molecules depending on the low (single) or high (triple) water content of the water-isopropanol mixtures (Fig. 75).

Meijer and George reported a coronene bisimide bearing a 3,5-dialkoxy substitution on the imide phenyl groups (**55**) (Fig. 14) that shows solvent-dependent aggregation behavior.<sup>164</sup>

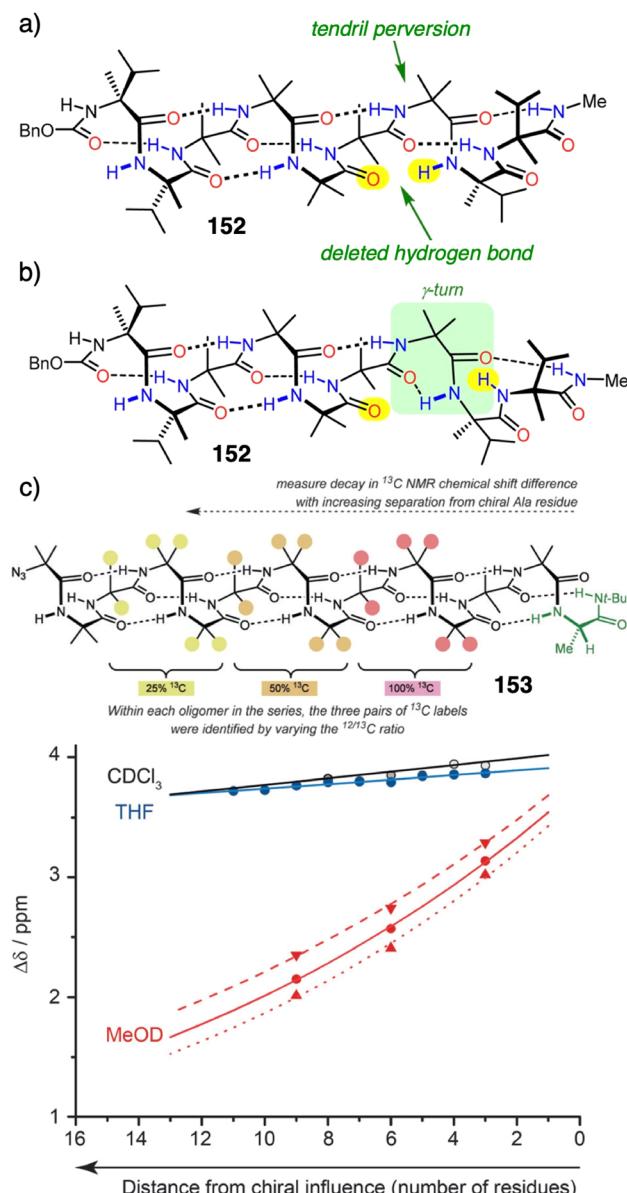


Fig. 74 (a) Tendril perversions and (b)  $\gamma$ -turn in AIB foldamer **152**. (c) Chemical shift differences ( $\Delta\delta$ ) measured in a series of three 'frame-shifted' oligomers, each containing three pairs of abundance-labelled (100%, 50% or 25%) <sup>13</sup>C labels. Screw-sense preference exhibited by Aib oligomer **153** decays as a terminal chiral influence becomes more distant. Reproduced from ref. 358 with permission from the Royal Society of Chemistry, copyright 2016.

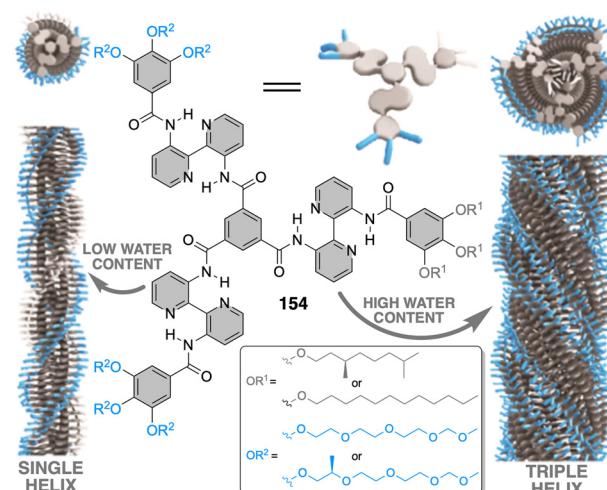


Fig. 75 Chemical structure of amphiphilic BiPy-BTA (**154**) and its hierarchical self-assembly from supramolecular fibre to triple helical bundle depending on the water content of water-isopropanol mixtures. Reproduced from ref. 365 with permission from the American Chemical Society, copyright 2014.

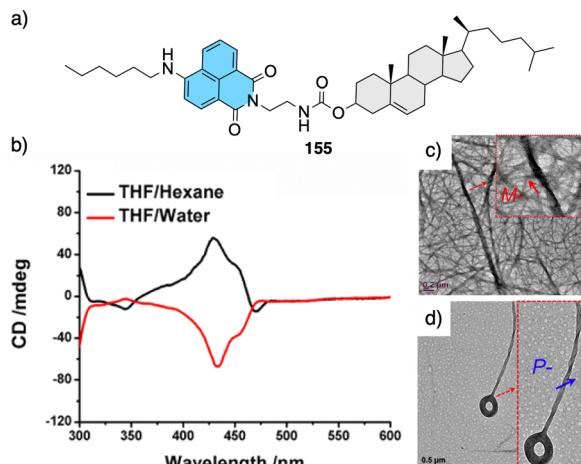


Fig. 76 (a) Chemical structure of **155**. (b) Normalized CD spectra of a vesicle sample (mixture of THF/water) and a gel sample (mixture of THF/hexane). TEM images of (c) *M*- and (d) *P*-type twist nanofibers observed in hexane and water, respectively. Reproduced from ref. 366 with permission from the American Chemical Society, copyright 2016.

Thus, helix inversion is observed when cyclic solvents are used to form the chiral aggregate, *e.g.*, *cis/trans*-decalin mixture or cyclohexane. Zhao reported another example of aggregation solvent dependence using, as building block, a molecule that contains naphthalimide and cholesteryl groups (**155**) (Fig. 76a).<sup>366,367</sup> This molecule self-assembles in low-polarity solvents (THF/hexane) into right-handed supramolecular helices ( $ECD > 0$ ) leading to *M*-oriented fibres observed in transmission electron microscopy studies (TEM) (Fig. 76b and c). In solvents with increased polarity (THF/water), this molecule self-assembles into left-handed supramolecular helices ( $ECD < 0$ ), that promote the formation of toroidal structures formed by *P* fibres (Fig. 76b and d).

Ajayaghosh reported a chiral OPE (**156**) that assembles, in *n*-decane, into a helical supramolecular polymer that in turn, with increasing building block concentration, assembles into superhelices of opposite handedness (Fig. 77a). Interestingly, the addition of chloroform recoils the superhelices to get the initial supramolecular helices.<sup>33</sup>

Liu reported another example in which the solvent dominates the helical structure over the rest of the stimuli.<sup>368</sup> The co-assembly of L-glutamide derivatives (**157** = PULG) with achiral anionic dyes (**158**, **159**) (Fig. 77b) promotes the formation of *P* or *M* helical structures in presence or absence of water. Thus, while co-assembly of monomers in ethanol results in the formation of *M*-oriented nanoribbon-like structures (Fig. 77c and d), addition of water to the ethanol solution led to a helical inversion into a *P*-helix (Fig. 77c and e). The amplification of asymmetry was mainly driven by electrostatic interaction between the gelator (**157**) and the achiral dyes (**158**, **159**). Remarkably, this interaction caused the transformation of the nanofibrous structures of the gelator into uniform left-handed helices stabilized by hydrogen bonds of **157**, while addition of water produced the formation of right-hand helices stabilized by  $\pi$ - $\pi$  stacking of dyes **158** or **159**. Ortí and Sanchez designed a chiral *N*-annulated perylene bisimide that self-assembles into supramolecular

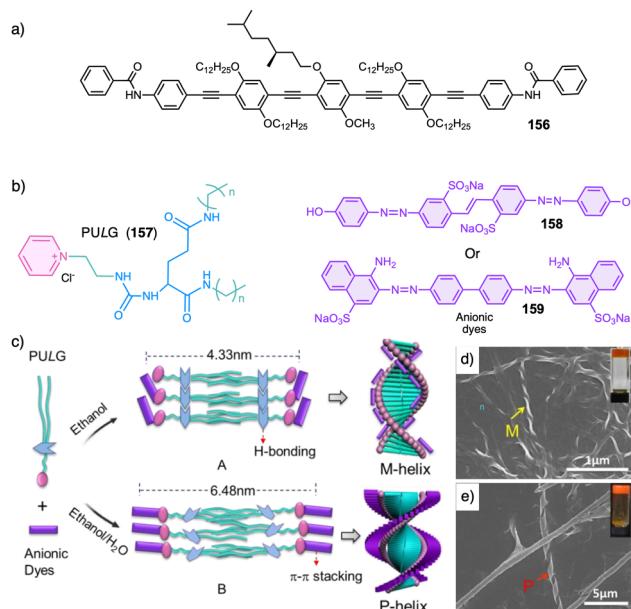


Fig. 77 (a) Chemical structure of OPE **156**. (b) Chemical structures of L-glutamide derivatives (**157**) and achiral dyes **158** and **159**. (c) Schematic representation of different helical co-assemblies of **157** with **158** or **159** induced by solvent conditions. (d) SEM images of aggregates showing an *M* helix obtained from ethanol. (e) SEM images of aggregates showing a *P* helix obtained from ethanol/water. Reproduced from ref. 368 with permission from the Royal Society of Chemistry, copyright 2019.

helical polymers with different handedness depending on the solvent (*methylcyclohexane* or *toluene*).<sup>369</sup>

**2.7.3 Solvent-responsive covalent polymers.** In chiral covalent polymers, solvents are frequently employed as external stimuli to induce helix inversion due to their effects on the conformational flexibility of the chiral pendant groups. Therefore, the use of organic and inorganic solvents, each of which has unique physical and chemical properties, has enabled screw sense control of dynamic covalent helical polymers.

Therefore, PPAs carrying chiral amino acid derivatives as pendants usually show helix inversion in different solvents due to conformational changes in the pendant group (Fig. 2b),<sup>98</sup> which places the substituents of the chiral pendant in different spatial orientations. In this way, a mechanism is produced that allows opposite helical senses to be induced.

For instance, Tang and Yashima described PPAs bearing L-leucine<sup>370</sup> and D/L-alanine<sup>371</sup> derivatives as pendants that showed solvent-induced helix inversion. Moreover, Yashima, using a PPA with a chiral alanine as pendant (poly-**160**), was able to visualize, through high-resolution AFM images, the two helical senses adopted by poly-**160** in solvents with different polarity, *e.g.*, THF and benzene (Fig. 78).<sup>372,373</sup> Helix inversion also occurs in the solid state by placing a 2D crystal in vapors of a solvent that induces the opposite chirality (Fig. 78).

This dynamic behavior of PPAs in the solid state was also employed by Tsuchihara to produce a colour change in a PPA film under solvent exposure.<sup>374</sup> Thus, a chiral *para*-substituted ether-PPA bearing an alcohol group and a long alkyl chain (poly-**161**) (Fig. 79a) shows a colour change in the film state



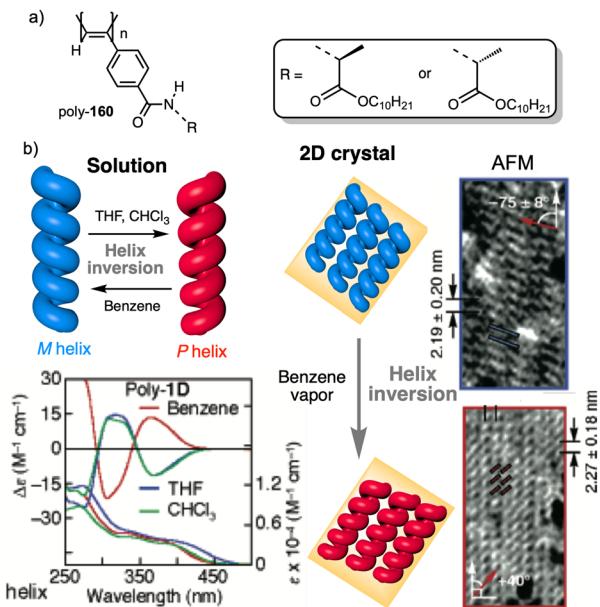


Fig. 78 (a) Chemical structure of poly-L/D-160. (b) Schematic representation of poly-D-158 and its macromolecular helicity inversion in solution and in 2D crystal state. Reproduced from ref. 372 with permission from the American Chemical Society, copyright 2006.

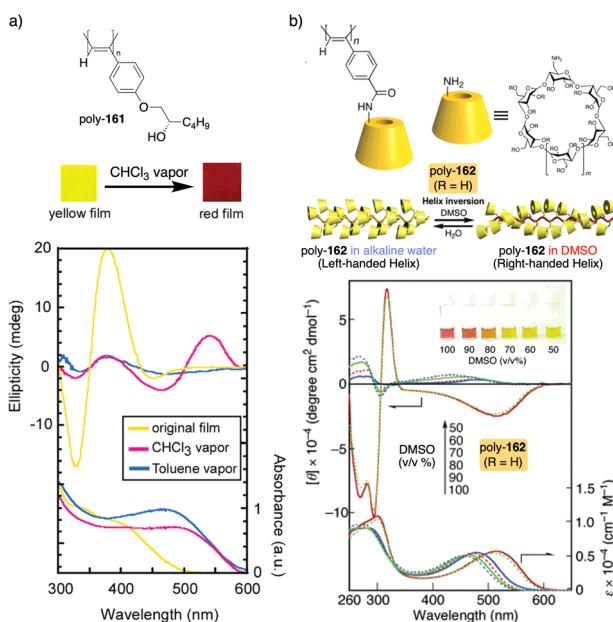


Fig. 79 (a) Chemical structure of poly-161 (top), colour change of its film under vapor exposure (middle), and ECD and UV spectra of its films in different solvents (bottom). (b) Chemical structure of poly-162 (top), schematic representation of its helical inversion (middle), and CD and UV-vis spectra in DMSO/water at varied ratios. Reproduced from ref. 374 and 383 with permission from the American Chemical Society, copyright 2006.

prepared in diethylamine upon exposure of the film to solvent vapors such as acetone, methanol, THF, and  $\text{CHCl}_3$ . The yellow to red colour change of the film is accompanied by a bathochromic UV shift from 400 to 500 nm, and by a new ECD

pattern with a positive Cotton band at 550 nm, which indicates the presence of a highly stretched helix (Fig. 79a).

Another interesting examples of helix inversion under different solvent vapor exposures can be found in literature of PPAs<sup>375,376</sup> and PDPAs.<sup>377</sup> In some cases, solvent can produce changes in both the elongation and the helical sense of dynamic helical polymers. Yashima described PPA poly-162 containing optically active cyclodextrin (CyD) as pendant (Fig. 79b),<sup>378</sup> whose elongation and screw sense depend on the solvent used to dissolve the polymer. Thus, while poly-162 adopts a more compressed helix in alkaline water, when the DMSO content increases in the solvent mixture, a bathochromic shift is observed in the polyene band, which is accompanied by a colour change of the solution from yellow to red and a helix inversion, as inferred from ECD studies (Fig. 79b). These structural changes are due to a different array of the CyD units along the polymer chains in both solvents.

Over the last few years, several examples have been reported involving solvent-induced helical structural changes in dynamic helical polymers (sense and/or elongation) induced by solvents that affect intra-pendant interactions within a helix.<sup>378–383</sup>

Recently, Wan<sup>384</sup> reported a PPA bearing achiral alkoxy carbonyl and chiral alkylamide substituents at *meta*-positions of its phenyl ring (poly-163) (Fig. 80a). In this case, a variation in the elongation of the polymer between a *cis-transoid* (stretched) and *cis-cisoid* (compressed) can be produced by playing with the polarity of the solvent (Fig. 80b–d).

Another family of dynamic helical polymers, whose helical scaffold shows solvent dependence, are polyquinoxalines (PQXs) developed by Sugino and coworkers. In a first example, they

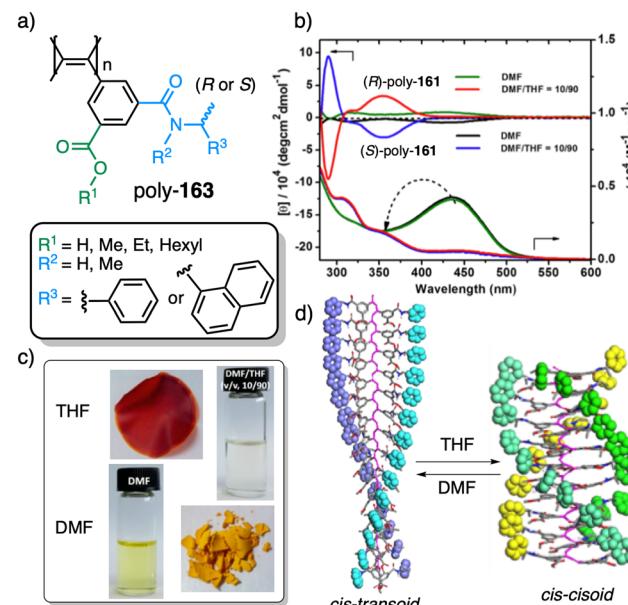


Fig. 80 (a) Chemical structure of poly-163. (b) CD and UV-Vis spectra of (R)/(S)-poly-163 in DMF and DMF/THF mixture (v/v, 10/90). (c) Photographs of poly-163 in solid and solution states in THF and in DMF. Reproduced from ref. 384 with permission from the American Chemical Society, copyright 2017.

reported a poly(quinoxaline-2,3-diyl)s that bears chiral *(R)*-2-butoxy groups as side chains (poly-**164**).<sup>385</sup> This polymer adopts a *P* helix in  $\text{CHCl}_3$ , whereas an *M* helix is adopted in 1,1,2-trichloroethane (1,1,2-TCE) (Fig. 81a), a helix inversion triggered by a conformational change in the pendant. In a different example, a polyquinoxanoline based phosphine (PQXphos) copolymer, consisting of a chiral comonomer carrying *(R)*-2-butoxymethyl groups and an achiral comonomer with a bulky phosphorous group in a 950/50 ratio, also shows a helix inversion in chloroform and in 1,1,2-trichloroethane/toluene (2/1) mixture.<sup>386</sup>

Next, the same group explored how solvents can affect the Sergeants and Soldiers effect in poly(quinoxaline-2,3-diyl) copolymers. Thus, different copolymer series were prepared using, as chiral Sergeants, the monomeric repeating units carrying *(R)*-2-butoxymethyl (**165a**), *(S)*-3-methylpentyl (**165b**), *(R)*-(pentan-2-yloxy)methyl (**165c**) and *(S)*-(2-methylbutoxy)methyl (**165d**) side chains at positions 6 and 7 of the quinoxaline rings. Each Sergeant was copolymerized with an achiral Soldier that bears an achiral propoxymethyl chain also at positions 6 and 7 of the quinoxaline rings (Fig. 81b).<sup>387</sup> In all cases, an effective *P* or *M* solvent-dependent Sergeants and Soldier effect was observed. The helical sense adopted by the copolymer is that dictated by the chiral comonomer (Fig. 81b). The ability of these polymers to adopt a single-handed helix, even when copolymerized with an achiral quinoxanoline, was employed by the same group to prepare a PQXphos acting as a catalysts, whose helical structure can be transferred to an enantiospecific hydrosilylation reaction.<sup>388</sup> These PQXs were also used to create a solvent-induced CPL switch.<sup>389</sup> Thus, after annealing in  $\text{CHCl}_3$ , a PQX film displayed reflection of right-handed CPL, whereas chiral reversion of reflected CPL was observed after annealing in 1,2-DCE vapor.

Although most examples deal with the polarity of the solvents, there is another relevant parameter that solvents

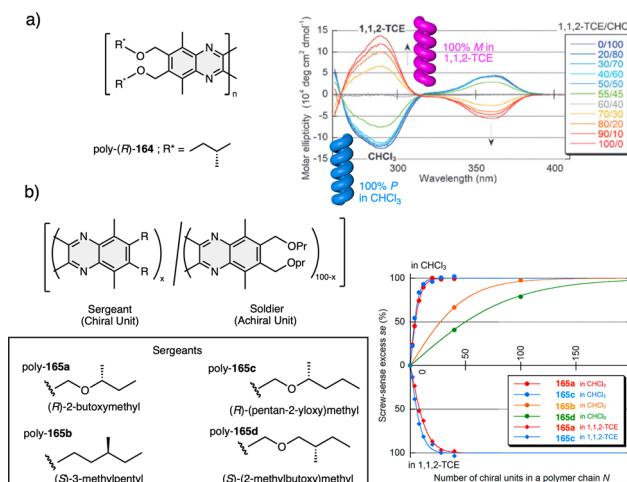


Fig. 81 (a) Chemical structure of poly-(*R*)-**164** and its CD spectra in 1,1,2-TCE/CHCl<sub>3</sub> with varied ratios. Reproduced from ref. 385 with permission from the Royal Society of Chemistry, copyright 2010. (b) General chemical structures of copolymers series **165a-d** and their screw sense excess in different solvents. Reproduced from ref. 387 with permission from the American Chemical Society, copyright 2013.

possess and that must be considered, the donor character, which is the ability to solvate cations and Lewis's acids and is related to the presence of atoms with free electron pairs. Freire and Riguera developed a PPA that was able to differentiate solvents based on their polarity and donor number, due to the presence of two independent tuneable bonds in the pendant. To do that, a PPA that bears *(R)*- $\alpha$ -methoxy- $\alpha$ -(trifluoromethyl)-phenyl acetic acid (MTPA) as pendant [poly-(*R*)-**166**] was prepared (Fig. 82).<sup>390</sup> In this polymer, the amide bond (H-N-C(=O)) is sensitive to the donor ability of the solvent—*trans*, non-donor solvents (CHCl<sub>3</sub>); *cis*, donor solvents (THF), while the (O=)C-C(-O) bond responds to the polarity of the solvent—*sp*, polar solvents (DMSO); *ap*, low-polar solvents (DCM). The *sp/ap* conformational change (O=)C-C(-O) produces a helical inversion in the polymer due to a different spatial location of the substituents, while the *cis/trans* conformational change in the amide (H-N-C(=O)) produces changes in the elongation of the polymer. As a result, four different helical scaffolds are obtained for poly-(*R*)-**166** taken into account the donor character (compressed/stretched) and the polarity (*P/M*) of the solvent (Fig. 82a). Moreover, this solvent behaviour for poly-(*R*)-**167** is observed in both solution and film state. The authors used the ability of this polymer to form *cis* amide bonds in THF to create a stereocomplex, where poly-(*R*)-**166** interacts with its enantiomeric and complementary helix to form fibre-like aggregates that evolve into a gel at high concentrations.<sup>391</sup>

Freire performed also solvent-dependent studies on PDPAs functionalized with chiral amino acids as pendants,<sup>147,392,393</sup> obtaining *syn/anti* conformational equilibria similar to those

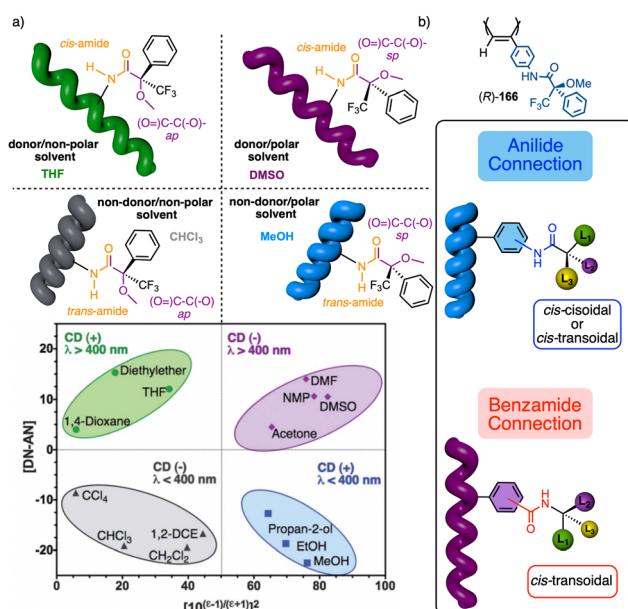


Fig. 82 (a) Correspondence between the four different structural states of poly-(*R*)-**166** and the polar/low-polar and donor/non-donor properties of the solvents. (b) Structure of poly-(*R*)-**166** and PPA with *ortho*-, *meta*- and *para*-aromatic substitution and anilide and benzamide connectors. Reproduced from ref. 391 with permission from the Royal Society of Chemistry, copyright 2015.

obtained for PPAs, although the activation energy to promote helix inversion is very high providing the polymers a memory effect. Also, the same group studied how the aromatic substitution in chiral PPAs (*i.e.*, *ortho*, *meta*, *para*) and/or the connector used to link pendant and backbone (*i.e.*, anilide, benzamide) affects the dynamic behaviour of PPAs and their solvent response (Fig. 82b).<sup>394,395</sup>

The conformational control of the pendants by solvents was also employed by Freire and Riguera to study conformational communication mechanisms between chiral comonomers along the polymer chain. Thus, they were able to prepare several series of fully chiral copolymers in which the different conformations adopted by the chiral Sergeants in various solvents are transmitted to the chiral Soldiers through Sergeants and Soldiers<sup>112,115</sup> or Chiral Coalition effects.<sup>113</sup>

Most of the solvent response behavior of dynamic covalent polymers found in the literature is based on a teleinduction phenomenon, where the chiral information flows through the different spatial orientation of substituents at the chiral centre towards the polyene backbone. Lately, Freire's group reported two different chiral harvesting mechanisms to induce a helical sense in the polymer (Fig. 3),<sup>102,103,396,397</sup> where the conformational information in the chiral pendant placed at a remote position is transferred to an achiral spacer and then harvested by the polyene backbone. Another helix induction mechanism that can be activated/deactivated by solvents is the Chiral Overpass effect<sup>105,106</sup> (Fig. 4a), where different chiral residues of a multichiral pendant group can control the helical sense of the helical polymer depending on the conformation adopted by the pendant.

## 2.8 External stimuli: chiral molecules

Chiral molecules have been widely used as a stimulus to induce the transfer of asymmetry from the molecular to the macromolecular level. Different strategies have been employed, from simple chiral solvation to the development of sophisticated receptors for specific targets. In this section we will show different examples on the topic.

**2.8.1 Covalent polymers.** Chiral molecules represent the archetypical stimuli for inducing asymmetry in covalent polymers. Therefore, over the last decades, Okamoto, Yashima, Maeda and others have developed a set of conjugated macromolecules that carry different types of receptors such as carboxylic acids (*e.g.*, poly-6),<sup>100,398–401</sup> crown ethers,<sup>402–405</sup> boronates,<sup>406</sup> phosphorylated derivatives (*e.g.*, poly-26),<sup>136,407–409</sup> sulphonic acids,<sup>410</sup> amines (*e.g.*, poly-27),<sup>141,411,412</sup> peptide sequences,<sup>413,414</sup> cyclodextrines,<sup>382,415</sup> and binaphthyls (*e.g.*, poly-35),<sup>146,416</sup> among many others. Upon formation of the corresponding supramolecular interaction between the polymer-bound receptor and the corresponding chiral molecule, an efficient asymmetry transfer occurs towards the macromolecule inducing preferred handedness. Remarkably, for some of the systems mentioned above, subsequent removal of the chiral unit and its replacement with a non-chiral counterpart gave rise to the memorized helical scaffold.<sup>100</sup> Further examples in other helical systems demonstrated an efficient memory effect without the presence of a non-chiral

chaperon if there is a sufficient energy barrier between the *P/M* helices.<sup>146</sup> The seminal example on this topic was reported by Okamoto, Yashima and coworkers who worked with a polycarboxylic polymer (poly-6) (Fig. 2, 83). The interaction of this polymer with a chiral amine (*e.g.*, (*R*)- or (*S*)-naphthylethylamine) gives rise to the salt pair that promotes the adoption of a preferred handedness. Replacement of the chiral amine with an achiral amine or amino alcohol results in memorization of the macromolecular screw sense.<sup>100</sup> This phenomenon was denoted as Dynamic Helix Memory effect (Fig. 83a). In a second generation of polymers, Maeda and Yashima managed to remove the chiral inductor without adding other stimuli to stabilize the helical structure, this phenomenon was called Static Helix Memory (Fig. 83b).

This protocol has been successfully applied by Yashima's group working not only with PPAs with the aforementioned receptors but also with other types of polymers such as poly(isocyanide)s (PICs),<sup>416,417</sup> poly(phospazanes),<sup>418,419</sup> or PDPAs<sup>420</sup> among others, giving rise to different types of memory effects. Similar effects occur in double-helical polymers stabilized by carboxylate bridges through the intercalation of chiral amines.<sup>421</sup> A detailed explanation of each receptor and the memory effects related to the nature of the polymer backbone can be found in specific reviews.<sup>422</sup>

Yashima also reported efficient asymmetry transfer during the formation of inclusion complexes in the helical cavity of PMMAs. Syndiotactic PMA (*st*-PPMA, poly-167) is known to encapsulate fullerenes within its helical cavity.<sup>423</sup> Therefore, they prepared a fullerene functionalized with a *P* or an *M* helical 3<sub>10</sub> peptide (168) (Fig. 84a). Upon encapsulation of the corresponding fullerene, poly-167 folds into a preferred helical conformation, as demonstrated by VCD. Similarly, they found that

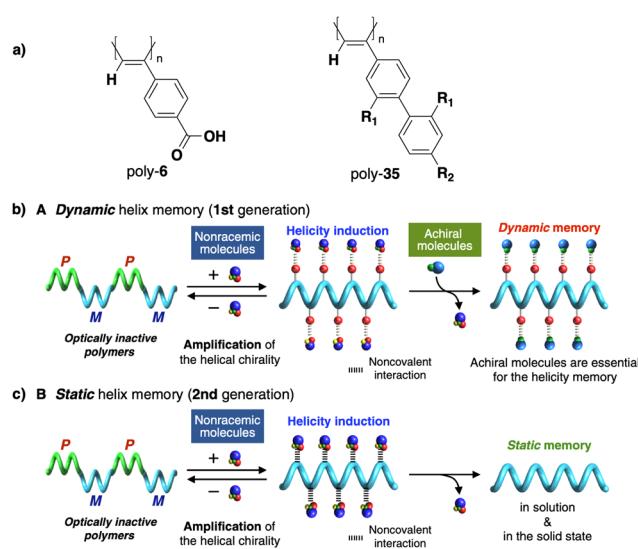
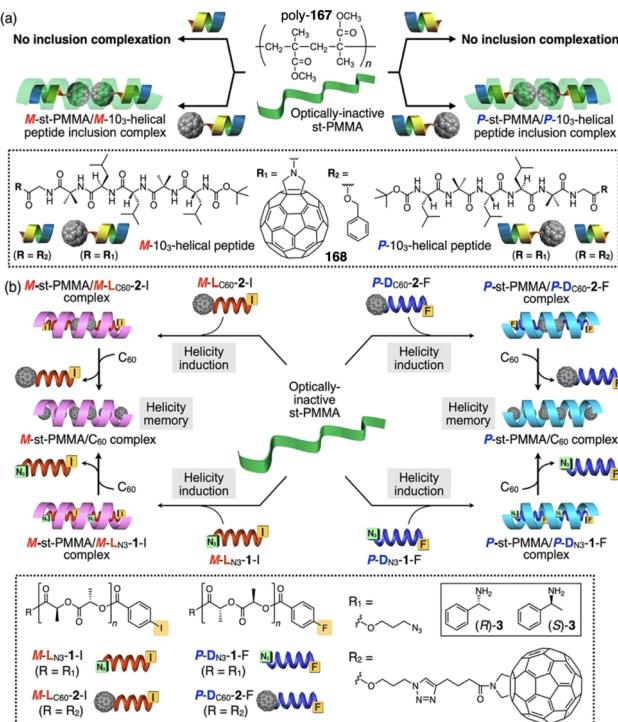


Fig. 83 (a) Structures of polymers poly-6 and poly-35. Schematic illustration of asymmetry induction and memory effect after (b) replacing the chiral molecules by achiral ones (first generation, dynamic memory) and (c) removal of the chiral molecules (second generation, static memory). Reproduced from ref. 422 with permission from the Chemical Society of Japan, copyright 2021.





encapsulation of fullerene bearing *P* or *M* polylactic acid also resulted in the folding of poly-167 into a preferred handedness (Fig. 84b).<sup>424</sup>

Apart from directed supramolecular interactions between a designed receptor and a target chiral molecule, in the literature we can find a large number of examples of efficient asymmetry transfer from solvent to solute (*i.e.*, helical macromolecule). Therefore, the use of chiral solvents as a source of chirality has been applied for the helical induction in different macromolecules. For instance, Kwak reported the asymmetry induction of a series of PDPAs containing trialkylsilyl moieties as pendant groups (poly-169) by thermal annealing in (*R*)- or (*S*)-limonene (Fig. 85a). During the annealing process, the macromolecule concomitantly adopts a preferred helical sense that depends on the absolute configuration of the solvent, as demonstrated by ECD and CPL spectroscopies (Fig. 85b).<sup>425</sup>

In another example, Zhang employed the chirality of limonene as an efficient asymmetry inductor in achiral PMMA-azo (poly-170) (Fig. 85c and d)<sup>426</sup> or crosslinked films of achiral poly-fluorene derivatives<sup>427</sup> with chiral storage and self-recovering properties. Thus, the chiroptical activity of poly-170 after induction with limonene can be easily analyzed by CD spectroscopy due to the presence of a strong bisignate band in the azo region (Fig. 85e).<sup>428</sup>

Interestingly, in the case of chiral polyfluorene derivatives [poly-(*S*)-171, Fig. 85f], Zhang induced opposite handedness in

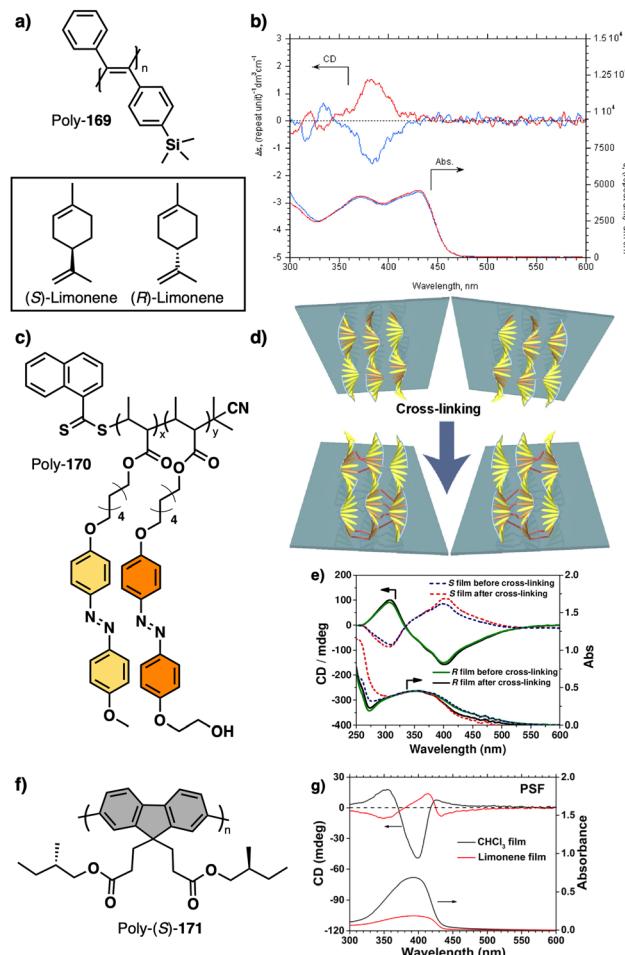


Fig. 85 (a) Structure of poly-169 and (b) ECD and UV spectra in (*S*)- and (*R*)-limonene (red and blue respectively). (c) Structure of poly-170, (d) Schematic illustration of the solvent drive symmetry breaking and further crosslinking upon self-assembly. (e) CD and UV spectra of chiral poly-170 films before and after cross-linking. Reproduced from ref. 425 with permission from the American Chemical Society, copyright 2012. (f) Structure of poly-(*S*)-171. (g) CD and UV-Vis spectra of poly-(*S*)-171 showing the helix inversion of the films from chloroform solution and from assemblies in limonene. Reproduced from ref. 428 with permission from John Wiley and Sons, copyright 2017.

films generated from  $\text{CHCl}_3$  or limonene solutions, because the chirality came from a combination of the polymer side chains and the solvents (Fig. 85g).<sup>428</sup> Following this protocol, this group has also prepared efficient multicolor CPL sources.<sup>429</sup>

Suginome reported different examples of PQXs endowed with solubilizing achiral lateral chains that adopt a preferred-handed helical structure upon dissolving in chiral solvents, such as poly-172 (Fig. 86a) that adopts a *P* helix in the presence of (*R*)-limonene (Fig. 86b). This induction of asymmetry later occurred in asymmetric catalysts for Suzuki, asymmetric silaborative C–C cleavage and hydrosilylation reactions (Fig. 86c).<sup>430</sup>

Nitschke and coworkers reported chiral induction, assisted by chiral solvents, in double-helical metallocopolymers 173 prepared *via* subcomponent self-assembly (Fig. 87a). During these studies, it was found that the metallocopolymer/chiral solvent



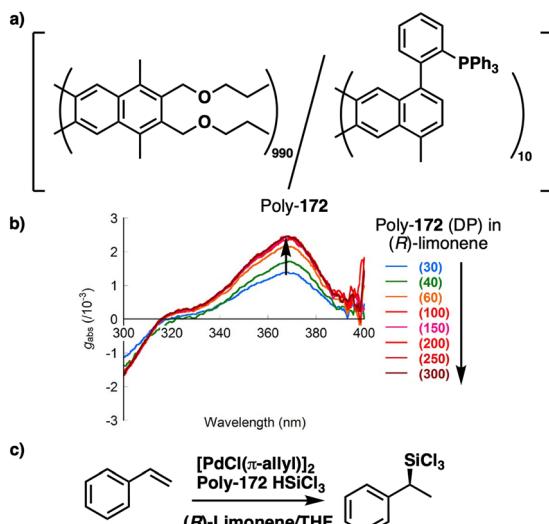


Fig. 86 (a) Structure of poly-172. (b) CD spectra of poly-172 upon helical induction with (R)-limonene. (c) Enantioselective reaction catalysed by poly-172 and induced by (R)-limonene. Reproduced from ref. 430 with permission from the American Chemical Society, copyright 2019.

interaction (*i.e.*, (S)-ethyl lactate) induces the adoption of *P* double helices with a degree of asymmetry similar to the use of a chiral initiator (Fig. 87b). Remarkably, asymmetry induction works only during the self-assembly process, since the addition of the chiral solvent once the polymer was generated did not trigger any screw sense excess, as demonstrated by CD spectroscopy (Fig. 87b).<sup>431</sup>

**2.8.2 Supramolecular polymers.** Chiral solvents represent an effective strategy to induce asymmetry in supramolecular polymers.<sup>432</sup> Meijer and co-workers studied the impact of a chiral solvent on the self-assembly of a chiral BTA-BIPY discotic molecule [(S)-174] (Fig. 88a). Aggregation in dodecane shows an active CD trace indicative of the formation of preferred-handed supramolecular helices, induced by the intrinsic chirality of BTA. Nonetheless, if aggregation occurs in a chiral alkane such as (S)-(-)-2,6-dimethyloctane, the CD spectra show a slight decrease in the CD trace, indicating a decrease in the asymmetry of the aggregate (Fig. 88a).<sup>122</sup> This group has also reported the use of chiral (S)- or (R)-1-chloro-3,7-dimethyloctane to induce asymmetry in triphenylene-2,6,10-tricarboxamides (TTAs)<sup>433,434</sup> bearing non-chiral solubilizing chains (175 and 176) (Fig. 88b).<sup>434</sup> Surprisingly, only 20% ee of the chiral solvent is needed to fully induce a preferred *P/M*-handedness in one-dimensional stack, following a supramolecular majority rules effect. Taking advantage of the induced handedness, these helical fibers were employed as efficient spin-filters attaining spin-polarization values of up to 40%.<sup>434</sup>

This group also reported the impact of chiral solvents on the self-assembly of TTAs containing chiral side chains [(S)-177 and (R)-177]. They discovered that depending on the temperature, the degree of solvation of the supramolecular polymer changes and, consequently, the point chirality of the TTAs side chains or the chirality of the solvents govern the screw sense of the supramolecular polymer. Therefore, at low temperatures, when the polymer is more solvated, chiral solvents govern the helicity of the systems, while at high temperatures, when the polymer is less solvated, point chirality is what controls the handedness of the polymers (Fig. 88b).<sup>433</sup> Moving from discotic to linear molecules, the same group reported the self-assembly of three different achiral oligo(*p*-phenylenevinylene)s (OPVs) with diaminotriazine (A-OPVTs) (178) or ureidotriazine (A-OPVUTs) (179, 180) units devoid of chiral side chains (Fig. 88c). As expected, aggregation in non-polar solvents gave rise to a racemic mixture of helical aggregates, while the inclusion of a small percentage of a chiral alcohol [*i.e.*, (R)- or (S)-citronellol] produced a chiral bias towards *M*- or *P*-helical stacks, respectively, as inferred by ECD spectroscopy (Fig. 88d). Intriguingly, they found that chiral alkanes were not able to efficiently transfer asymmetry to the aggregate, thus indicating the presence of precise supramolecular forces between solvent and solute (*e.g.*, hydrogen bond between the alcohol and the triazine unit) as a key factor in biasing the handedness of the aggregates.<sup>435</sup> Bouteiller and coworkers reported on the asymmetry induction in the formation of supramolecular nanotubes composed by benzene bisureas. In this system, self-assembly of the achiral derivatives in the presence of (R)- or (S)-limonene provided 33% of the maximum net helicity achieved for analogous building blocks containing chiral side chains.<sup>436</sup>

The chiral solvent-mediated asymmetry induction found in supramolecular polymers in the solution state also operates at the solid–liquid interface. Thus, De Feyter reported the multi-component assembly of coronene, isophthalic acid and dehydrobenzo[12]annulene at the solid–liquid interface, where

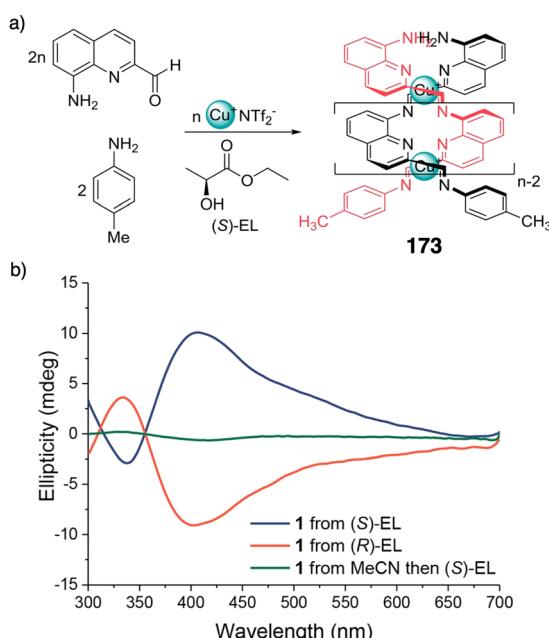


Fig. 87 (a) Subcomponent self-assembly polymerization of double helical metallopolymer 173 in the presence of a chiral solvent. (b) CD spectra showing the formation of enantiomeric helical structures as function of the enantiomeric form of the chiral solvent. Reproduced from ref. 431 with permission from the American Chemical Society, copyright 2018.

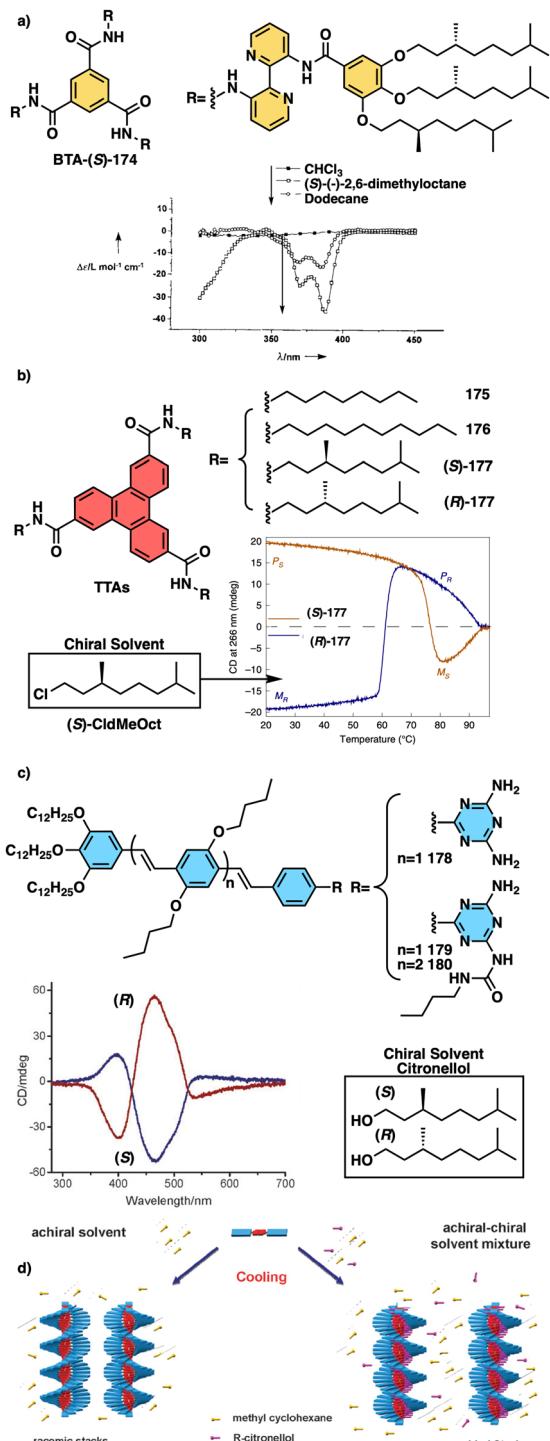


Fig. 88 (a) Structure of BTA-(S)-174 and CD spectra showing the impact of chiral solvent on the screw sense excess of the supramolecular polymer. Reproduced from ref. 121 with permission from John Wiley and Sons, copyright 1997. (b) Structures of TTAs 175 and 176. Structures of TTAs (S)-177 and (R)-177 and CD spectra showing their temperature dependent stereomutation in (S)-CldMeOct. Reproduced from ref. 433 with permission from Springer Nature, copyright 2021. (c) Structures of the OPVs 178-180 and CD spectra showing the adoption of opposite handed helices depending on absolute configuration of the chiral solvent. (d) Schematic illustration of the solvent drive symmetry breaking during self-assembly. Reproduced from ref. 435 with permission from the Royal Society of Chemistry, copyright 2011.

the chirality of the system depended solely on the chiral solvent used.<sup>437</sup> Another strategy to study the transfer of asymmetry from a chiral molecule to supramolecular polymers is based on the inclusion of a receptor at the periphery of the assembling molecule. Following this idea, George and coworkers reported on the formation of supramolecular polymers based on naphthalene diimide that carried zinc-complexes as substituents (NDPA, 181). Opposite handedness is induced in the supramolecular polymer that binds adenophosphates to these receptors as a function of the number of phosphates anchored to the chiral adenosine (*i.e.*, ATP, *P* helix; AMP and ADP, *M* helix). Additionally, the enzymatic transformation from ATP to ADP mediated by alkaline phosphatase produces helix inversion (Fig. 89).<sup>438,439</sup>

**2.8.3 Foldamers.** As explained in the introductory section, Inai and coworkers reported the so-called Domino effect in foldamers composed by non-natural achiral amino acids alternating, in the sequence, *Z*-dehydrophenylalanine and Aib units. The interaction of either the C or N terminus (COOH or NH, respectively) of the foldamer with chiral amines or carboxylic acids respectively, triggered the adoption of an excess of preferred screw sense in the foldamer as depicted in Fig. 1c.<sup>82-85,439-441</sup>

This asymmetry induction mechanism was exploited by Clayden and coworkers to develop stimuli-responsive foldamers based on 3<sub>10</sub>-helical Aib peptides (Fig. 1)<sup>358</sup> and oligourreas 66 (Fig. 18).<sup>177</sup> Different receptors such as amino groups, ureas, boronic esters, pyridines, or metal complexes that upon interaction with the corresponding guests induced changes in the helical scaffold were introduced into the N or C terminus of the foldamer. These interactions produced variations in the spectroscopic data from NMR, CD or photoluminescence among other techniques employed.<sup>60,70,72-74,78-81,442-444</sup> As an example, the authors reported the preparation of an Aib foldamer carrying a 2-(aminomethyl)phenylboronic acid derivative (182)—known to interact with geminal diols—placed at the N-terminal position (Fig. 90a). Coordination of the diol 183 with the boron atom results in the adoption of a preferred screw sense in the Aib foldamer. The helical bias is monitored by the presence of evident chemical shifts and diastereotopic

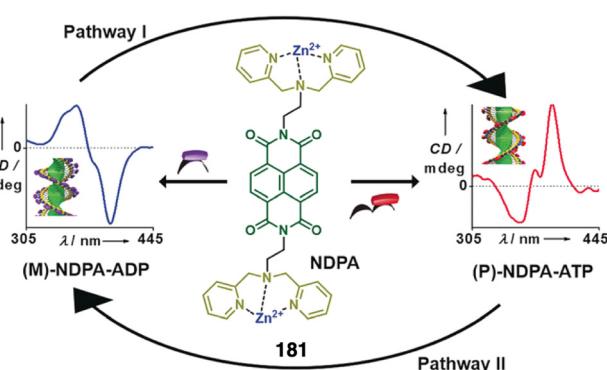


Fig. 89 Chemical structure of naphthalene diimide 181, bearing as substituents zinc-complexes, that promotes *P* or *M* helical coassemblies by interaction with ATP or ADP respectively. Reproduced from ref. 438 with permission from John Wiley and Sons, copyright 2017.



NMR signals in  $^{11}\text{B}$  and  $^1\text{H}$  experiments, respectively (Fig. 90b).<sup>445</sup> A very interesting asymmetry induction found in foldamers is based on the encapsulation of achiral/chiral guests within the helical cavity.<sup>13,445–449</sup> Huc and Ferrand reported different oligoamide-based foldamers (e.g., **184**, Fig. 91a) and metallofoldamers that encapsulate chiral guests such as tartaric acid<sup>450</sup> or different saccharides.<sup>13</sup> Encapsulation of the corresponding guest induced the adoption of a preferred-handed helical structures (Fig. 91b and c). Remarkably, modifications in certain regions of the foldamer with light<sup>291</sup> (Fig. 59) or addition of metals induced additional helix inversion processes (Fig. 29b and c).<sup>203,204</sup> Li studied other examples of foldamers that encapsulate saccharides, and designed oligohydrazide-type foldamers capable of encapsulating glucose through hydrogen bonds. Changes in foldamer length allowed the binding of different saccharides to be modified, while the inclusion of chiral handles, such as proline residues, allowed the binding events to be monitored using CD spectroscopy.<sup>451–453</sup>

*meta*-Phenylene ethynlenes (mPEs) represent another interesting family of foldamers that adopt helical structures induced by solvophobic effects.<sup>454–458</sup> For instance, Moore reported that mPE **185** (Fig. 92a), with a triethyleneglycol side chain, folds into a helical structure with no screw sense preference in MeCN/H<sub>2</sub>O mixtures. Upon addition of chiral terpenes, such as pinene, a CD signal arises for both compounds indicating

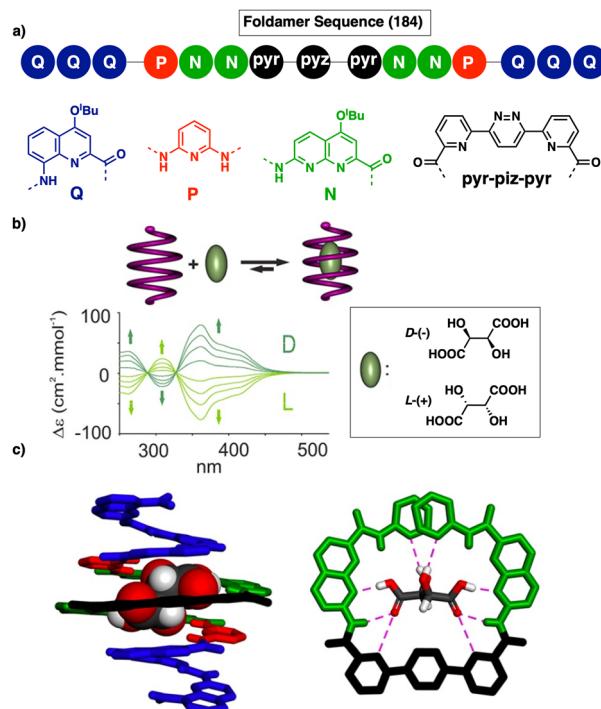


Fig. 91 (a) Structure of foldamer **184**. (b) Schematic illustration of the host–guest formation monitored by CD and structures of the guests. (c) Host–guest complex formed by foldamer **184** and D-tartaric acid. Reproduced from ref. 450 with permission from the American Chemical Society, copyright 2010.

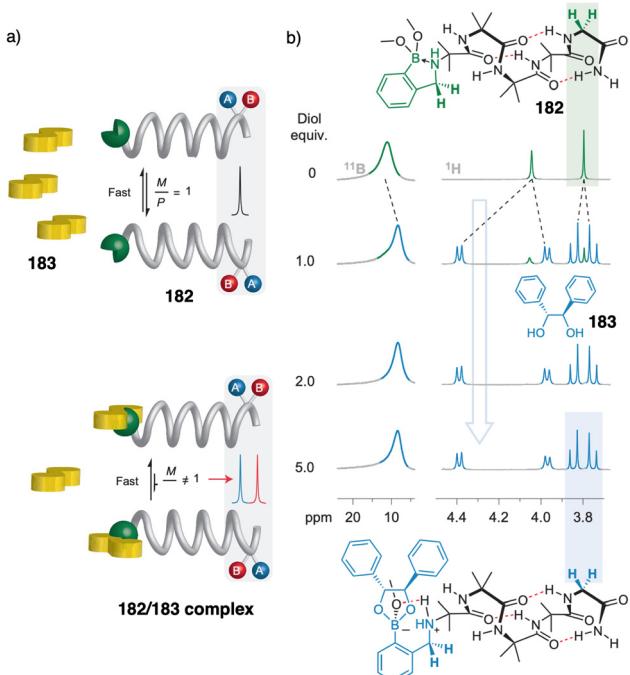


Fig. 90 (a) Schematic illustration of the helical bias upon binding of a substrate in the receptor of the Aib foldamer **182** and the corresponding NMR spectroscopic output upon binding event. (b) Structure of the Aib based foldamer **182** containing the boronate receptor for diols (top) and  $^{11}\text{B}$  and  $^1\text{H}$  NMR experiments showing the diastereotopic signals of **182** upon adoption of preferred screw-sense excess when interacts with the chiral diol **183**. Reproduced from ref. 445 with permission from Springer Nature, copyright 2013.

the adoption of preferred handed helices. The asymmetry induction comes from the formation of inclusion complexes, where pinene is encapsulated within the helical cavity of the mPE foldamer. Moore also prepared longer derivatives of up to 28 mPE units and also mPEs with different core linkers, which were able to form inclusion complexes with chiral aromatic pillars such as **186**, **187** (Fig. 92b–d).<sup>454–458</sup>

Inoue developed an *m*-pyridine ethynylene foldamer that, upon binding to  $\beta$ -glucosides, folds into a helical structure with preferred handedness in MeOH/H<sub>2</sub>O mixtures. Protonation of the pyridine units also resulted in direct folding into helical structures. Curiously, in this state, the chirality of the foldamer (*i.e.*, *P* and *M* helical conformations) depends on the mutarotation of glucose. Additional modifications to the sequence of the foldamer allowed the preparation of systems that self-assemble into double-helical structures, which can be disassembled upon binding the corresponding guests.<sup>459,460</sup>

## 2.9 External stimulus: pH

The influence of pH on stimuli-responsive helical polymers has attracted significant attention from scientists working in this field due to the possibility of controlling the helical sense through the presence of positive or negative charges in the chiral material.

**2.9.1 pH-Responsive foldamers.** In Chemical Biology, pH control is crucial because many biological processes are affected by pH fluctuations. Clayden and coworkers prepared

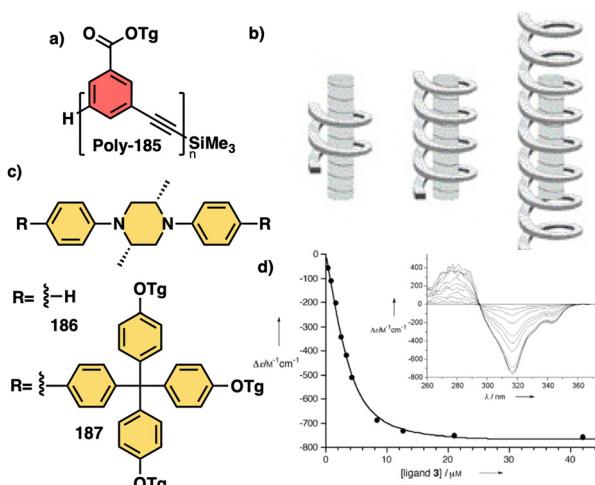


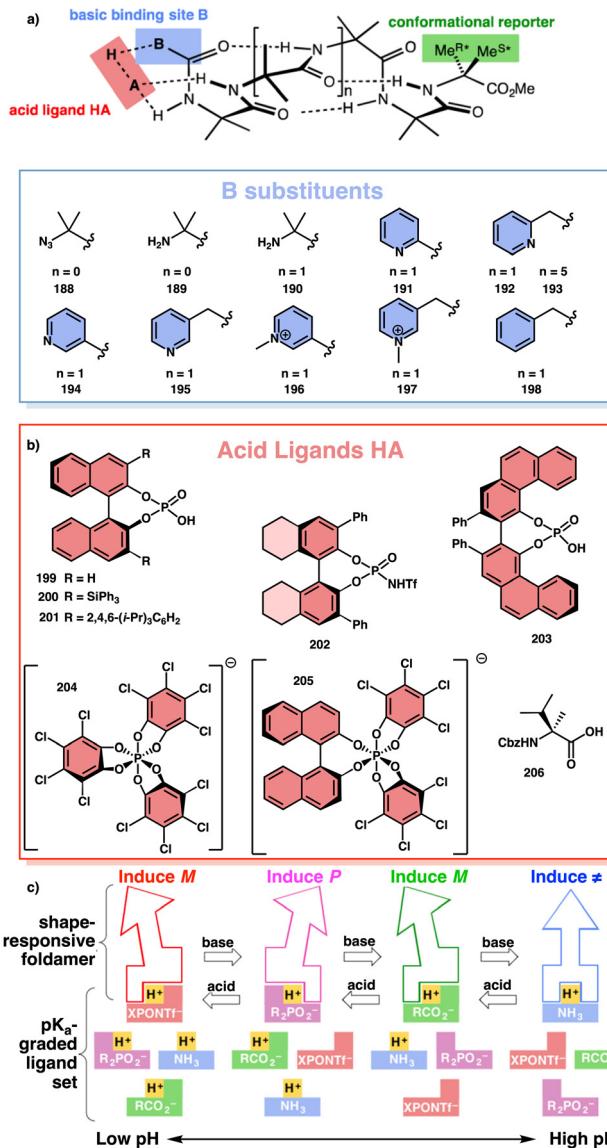
Fig. 92 (a) Structure of foldamer 185. (b) Schematic illustration of the host-guest formation with (c) chiral pillars 186 and 187 (reproduced from ref. 457 with permission from the American Chemical Society, copyright 2001). ECD spectra upon the addition of a chiral molecule to foldamer 185. Reproduced from ref. 458 with permission from John Wiley and Sons, copyright 2002.

a series of Aib foldamers that respond to pH changes (188–198) (Fig. 93a).<sup>444</sup> They demonstrated that these achiral helical foldamers selectively interact with chiral acidic ligands (199–206) (Fig. 93b), inducing conformational preferences—screw sense induction—detectable by  $^{13}\text{C}$ -NMR. By adding bases or acids to the foldamer-ligand mixture, the conformational preference of the foldamer chain can be switched between left- and right-handed conformations (Fig. 93c). This pH-dependent conformational response allows the foldamer-ligand system to function as a “proton-counting” molecular device, providing a tuneable and reversible response to changes in its environment.

In a project led by Huc, a helix-rod was designed with host-guest functionality.<sup>461</sup> This innovative design involved the folding, binding, and random sliding motions of helical foldamers (specifically, aromatic oligoamides such as 207) along rod-like guests (Fig. 94a). To facilitate this process, two distinct binding stations were created along the rod-like guest 208: one with a heptyl segment and the other with a diethylamine segment.

The host-guest complex demonstrated negligible bias between the two binding sites. However, during the experimental work, an interesting observation was made. By adding an acid, the researchers were able to selectively trap the host-guest complex at a single binding site. Conversely, the addition of a base, which served to deprotonate, successfully reactivated the dynamic motion of the system (Fig. 94b).

**2.9.2 pH-Responsive supramolecular polymers.** pH-Responsive chiral supramolecular aggregates have been employed to primarily investigate mostly the pH-dependent chirality of co-assembled hydrogels. In a specific study conducted by Feng and coworkers,<sup>462</sup> a phenylalanine-derived gelator and an achiral cationic polyacrylamide were co-assembled to form a chiral hydrogel. The chirality could be controlled by pH adjustments,



so at pH 7, the hydrogels showed right-handed chirality, while at pH 7 and above, the chirality switched to a left-handed configuration. This observation suggests that at lower pH levels, hydrogen bonds between carboxylic acids and amide groups maintain the alignment of the nanostructures with a certain twisting degree. In contrast, at higher pH values, the ionization of carboxylic acid groups results in their interaction with cationic quaternary ammonium ions through electrostatic interactions, resulting in the rearrangement of packing patterns and the formation of opposite chiral nanostructures.

Similarly, in another system investigated by Bhosale and coworkers using NDI-L-Glu monomers (209), chiral supramolecular helices were formed (Fig. 95a).<sup>463</sup> In this case, the pH response was as observed in the previous example. At acidic pH,



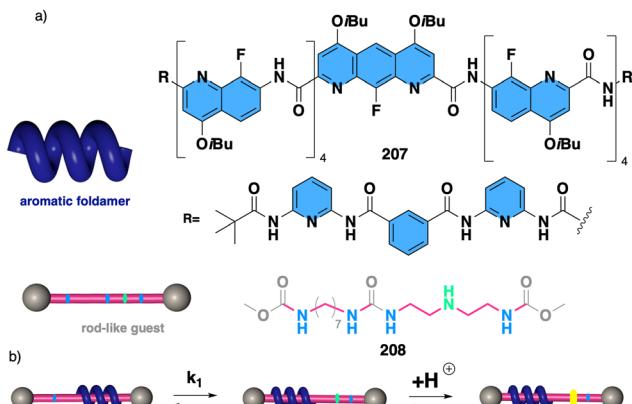


Fig. 94 (a) Molecular structure of aromatic oligoamide **207** and the rod-like guest **208**. (b) Scheme of helix sliding along a nondegenerate guest possessing a station that can be blocked or unblocked upon protonation or deprotonation, respectively. The green dot symbolizes the amine function, which can be included in the helical cavity. The yellow dot is the corresponding ammonium, which is not included in the helical cavity.

the monomers interacted through hydrogen bonding between the amides and carboxylic acids, while at high pH the carboxylic acids were deprotonated, and electrostatic repulsion stabilized the opposite helical sense (Fig. 95b). Additionally, in this system, it was found that the *P/M* helical sense of the supramolecular aggregate could be switched and interconverted after several pH cycles (Fig. 95c).

The pH-responsive self-assembly of a histidine-functionalized perylene diimide (PHP, **210**) was studied by Govindarajan (Fig. 96a). This molecule exhibited a pH reversible chiroptical supramolecular aggregate switch with tunable 1D nanostructures

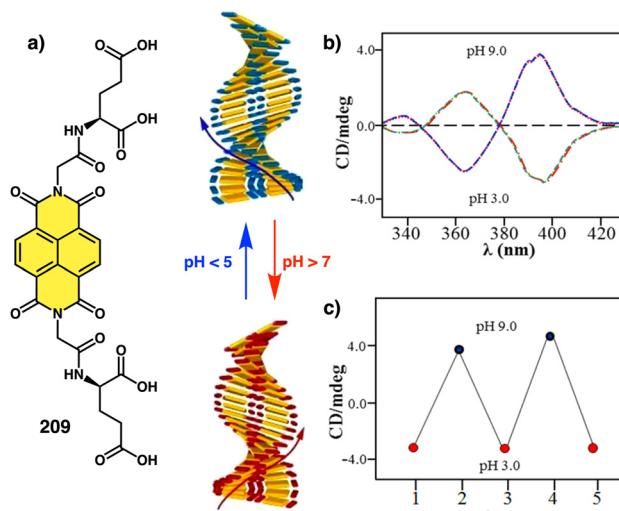


Fig. 95 (a) Molecular structure of **209** and graphical illustration of *P* or *M* helical senses of the supramolecular aggregate tuneable by the action of pH. (b) CD spectra of **209** at pH 3 and 9. (c) pH chiroptical switch of a supramolecular aggregate of **209** after different cycles at pH = 3 and pH = 9. Reproduced from ref. 463 with permission from Springer Nature, copyright 2018.

between thin or thick fibrils and belts.<sup>464</sup> The pH-induced self-assembly of **210** was confirmed by CD measurements (Fig. 96b and c).

Besenius also explored pH-responsive supramolecular aggregates using oppositely charged phenylalanine-based dendritic peptide amphiphiles (**211** and **212**, Fig. 97a), which enabled selective homopolymerization by pH modulation and charge switching into comonomers.<sup>465</sup> The strategy involves attractive Coulomb interactions in oppositely charged dendritic peptide amphiphiles that reinforce weak non-covalent interactions, resulting in the formation of alternating supramolecular copolymers. By altering the pH and switching the peptidic monomers, attractive electrostatic interactions are disrupted, leading to selective polymerization and the formation of homopolymers **211** of the basic comonomer at high pH and homopolymers **212** of the acidic comonomer at low pH. The transition between copolymers and homopolymers is reversible (Fig. 97b).

**2.9.3 pH-Responsive dynamic covalent polymers.** Yu explored the synthesis of water-soluble, double-helical polymers that exhibit responsiveness to multiple stimuli, including pH.<sup>466</sup> The authors achieved this goal by employing Passerini multicomponent polymerization-induced assembly (PMPIA), which resulted in the

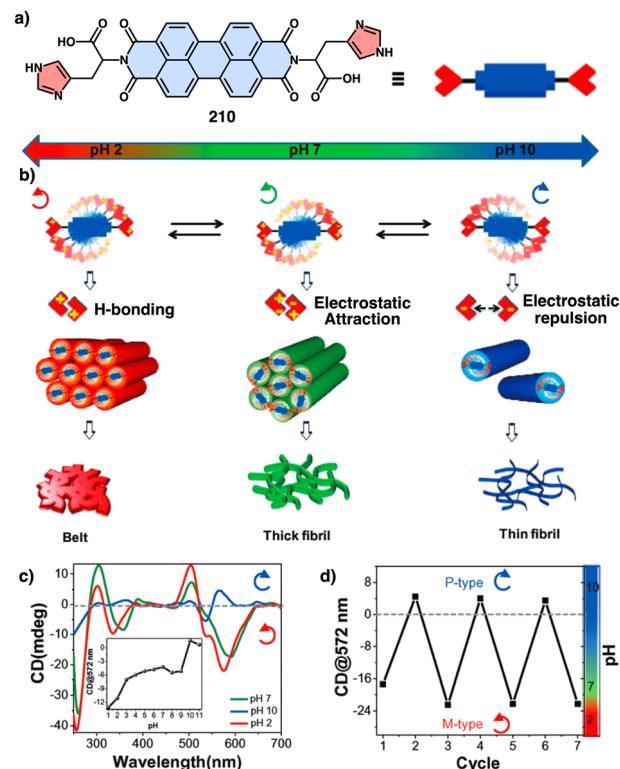


Fig. 96 (a) Molecular structure and schematic representation of **210**. (b) Schematic illustration of the pH-responsive reversible chiroptical aggregate switch and its tuneable 1D morphology induced by protonation-deprotonation. Curly arrows indicate the handedness of the molecular assembly of **210**. (c) CD spectra of **210** at different pH. Inset: Plot of CD intensity at 572 nm versus pH. (d) Reversible chiroptical switching at 572 nm as a function of pH cycles. Curly arrows indicate the handedness of the molecular assembly of **210**. Reproduced from ref. 464 with permission from the Royal Society of Chemistry, copyright 2016.



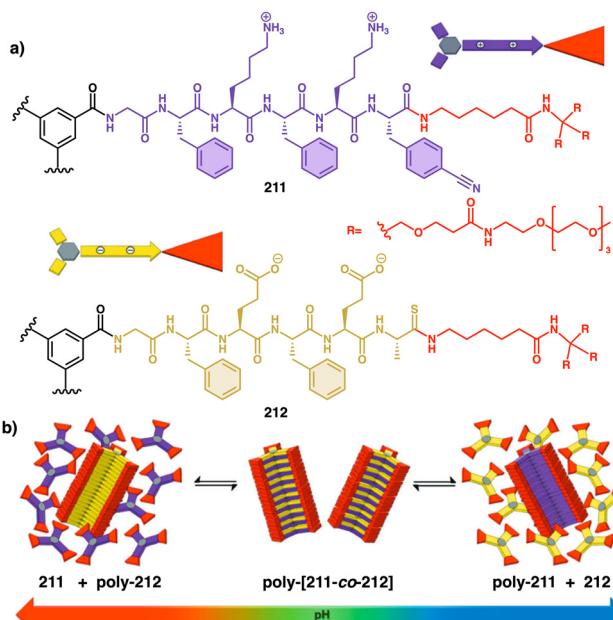


Fig. 97 (a) Chemical structures of the C<sub>3</sub>-symmetric dendritic peptide comonomers **211** and **212** and (b) their pH-regulated supramolecular polymerization into homopolymers of **211** and **212**, at high and low pH respectively, and **211-212** copolymers at neutral pH. Reproduced from ref. 465 with permission from John Wiley and Sons, copyright 2015.

formation of reversible polymers that adopt a helical structure and possess an aggregation-induced emission (AIE) effect. The assembly of polymers was driven by the polymerization-induced generation of supramolecular interactions in chiral  $\alpha$ -acyloxyamides (poly-**213**) (Fig. 98). Under neutral pH conditions, the polymers self-assembled into a double helix structure, which was stabilized by hydrogen bonds between the pyridines and the protons of the amides. This helical arrangement exhibited circularly polarized luminescence (CPL) and fluorescence due to  $\pi$ -stacking interactions of the pyridines. As the pH decreased, the pyridines in the  $\alpha$ -acyloxyamides became protonated. Consequently, supramolecular interactions were simultaneously disrupted (Fig. 98). Increasing pH causes neutralization of hydrogen atoms in amide bonds, leading to breakdown of hydrogen bond interactions.

Freire successfully synthesized water-soluble helical amine-PPAs with pH-responsive behaviour, such as poly-**214**.<sup>467</sup> Importantly, the location of the amino group within the polymer determines the mechanism and direction of the helical response. At acidic pH, the preferred helical conformation is stabilized by a hydrogen bond between the ammonium and carbonyl groups, while at higher pH values, when the amino group is deprotonated, an antiperiplanar orientation is preferred between both groups, placing the substituents of the chiral centre in different locations and therefore, promoting opposite helical senses. Helix inversion could be observed in MeOH, but not in water, due to solubility problems of the neutral form of poly-**214** (Fig. 99).

Masuda reported poly(phenyleneethynylene-pyromellitic diimide)s,<sup>468</sup> which exhibited stable helical conformations that could be disrupted by the addition of NaOH and subsequently recovered by the addition of HCl. Zheng and Bao studied the

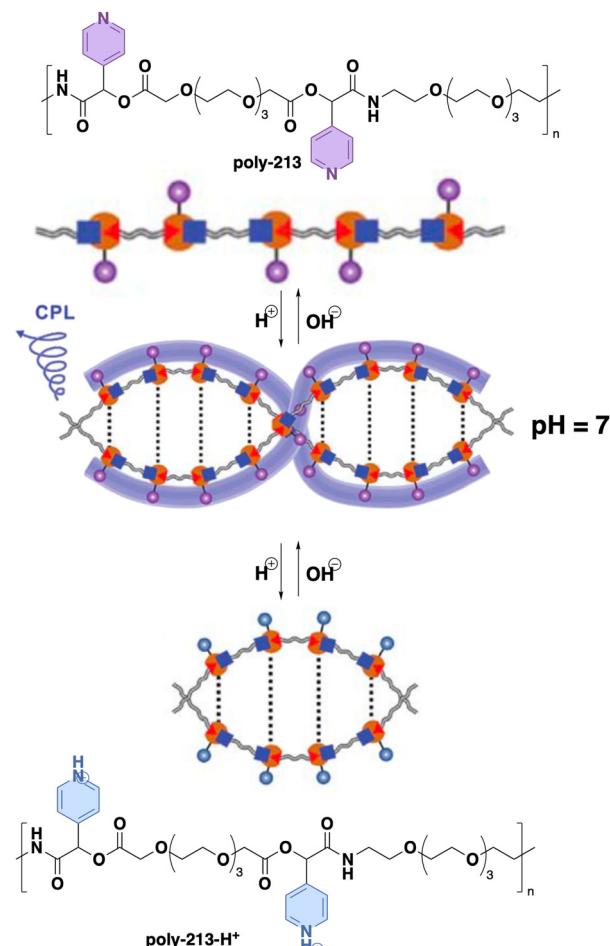
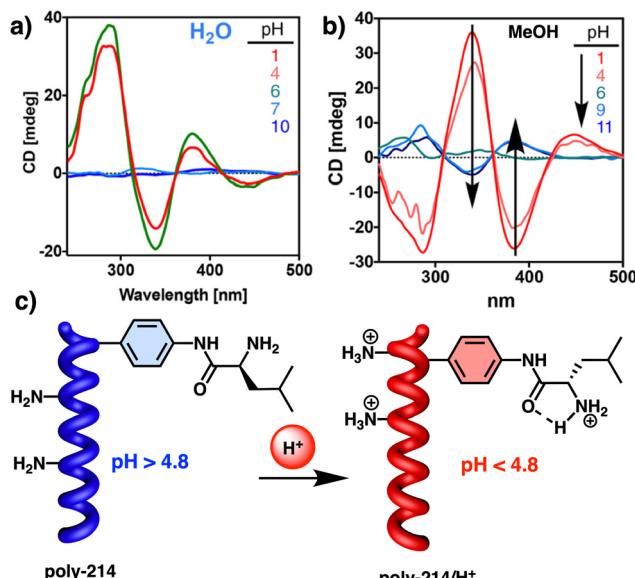


Fig. 98 Schematic illustration of double-helical AIE-active polymers **213** with multiple responsiveness via the PMPA. Reproduced from ref. 466 with permission from the Royal Society of Chemistry, copyright 2020.

self-assembly of 1D gadolinium(III) phosphonate coordination polymers (CPs) using different metal sources and ligands. They achieved the formation of helical superstructures with controllable twist directions by manipulating interchain interactions and pH. The specific counter-anions present in the system and the pH determined the chemical composition and morphology of the CPs.<sup>469</sup> Other examples dealing with a helix to random coil transition triggered by pH are found in the literature for unnatural polypeptides<sup>470</sup> and graft copolymers,<sup>471</sup> however, the main goal of this review is to describe systems that form helical structures whose scaffolds can be altered by the presence of external stimuli.

### 3 Conclusions and outlook

Dynamic helical polymers (covalent and supramolecular) and foldamers share the helix as structural motif, which can be altered (sense and elongation) by the presence of external stimuli. In this review, we highlight the most important helix induction and conformational communication mechanisms triggered by the presence of different external stimuli, which



**Fig. 99** CD spectra for poly-214 at different pH in (a) water and (b) MeOH. (c) Conformational composition of amino-PPAs in basic and acidic media. [amino-PPA] = 0.5 mg mL<sup>-1</sup>. Reproduced from ref. 467 with permission from the American Chemical Society, copyright 2018.

provoke conformational or structural changes in the monomer repeating units or building blocks and which are shared by these different materials.

Thus, although helical polymers (covalent and supramolecular) and foldamers belong to different research areas, the knowledge and advances achieved in the different materials can also be applied to others. Therefore, through a rational design of the components of helical polymers, it is possible to create dynamic helices with stimuli-responsive properties that can be used in different applications such as sensing, molecular recognition, asymmetric synthesis, signalling, information storage, and son on.

In summary, this review shows the main stimuli-responsive properties of dynamic chiral helical polymers, and the mechanisms involved in the structural changes induced in macromolecular and supramolecular helices. However, although the helices are different and each one has its own intrinsic properties, the stimuli-response capacity of the building blocks or monomer repeating units, and the mechanisms involved in the structural changes of the helices are similar, making it possible to extract information from a specific system and employ it in another different material. Thus, by combining information from different systems, it is possible to search for novel helical polymers with new or improved functionalities.

## Author contributions

All authors have contributed to the preparation of the manuscript and agreed with the final version.

## Conflicts of interest

There are no conflicts to declare.

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## Notes and references

- D. T. J. Morris and J. Clayden, *Chem. Soc. Rev.*, 2023, **52**, 2480–2496.
- T. Seedorf, A. Kirschning and D. Solga, *Chem. – Eur. J.*, 2021, **27**, 7321–7339.
- R. Aksakal, C. Mertens, M. Soete, N. Badi and F. Du Prez, *Adv. Sci.*, 2021, **8**, 20004038.
- H.-W. Schmidt and F. Würthner, *Angew. Chem., Int. Ed.*, 2020, **59**, 8766–8775.
- E. Yashima, N. Ousaka, D. Taura, K. Shimomura, T. Ikai and K. Maeda, *Chem. Rev.*, 2016, **116**, 13752–13990.
- A. Das and S. Ghosh, *Chem. Commun.*, 2016, **52**, 6860–6872.
- M. Shigeno, Y. Kushida and M. Yamaguchi, *Chem. Commun.*, 2016, **52**, 4955–4970.
- D.-W. Zhang, X. Zhao, J.-L. Hou and Z.-T. Li, *Chem. Rev.*, 2012, **112**, 5271–5316.
- Z. C. Girvin and S. H. Gellman, *J. Am. Chem. Soc.*, 2020, **142**(41), 17211–17223.
- P. Rivera-Fuentes and F. Diederich, *Angew. Chem., Int. Ed.*, 2012, **51**, 2818–2828.
- G. Maayan, *Eur. J. Org. Chem.*, 2009, 5699–5710.
- A. Das and S. Ghosh, *Angew. Chem., Int. Ed.*, 2014, **53**, 2038–2054.
- Y. Ferrand and I. Huc, *Acc. Chem. Res.*, 2018, **51**, 970–977.
- I. Huc and S. Hecht, *Foldamers: Structure, Properties, and Applications*, John Wiley & Sons, Weinheim, 2007.
- E. A. John, C. J. Massena and O. B. Berryman, *Chem. Rev.*, 2020, **120**, 2759–2782.
- D. J. Hill, M. J. Mio, R. B. Prince, T. S. Hughes and J. S. Moore, *Chem. Rev.*, 2001, **101**, 3893–4012.
- S. H. Gellman, *Acc. Chem. Res.*, 1998, **31**, 173–180.
- O. J. G. M. Goor, S. I. S. Hendrikse, P. Y. W. Dankers and E. W. Meijer, *Chem. Soc. Rev.*, 2017, **46**, 6621–6637.
- M. H. Bakker, C. C. Lee, E. W. Meijer, P. Y. W. Dankers and L. Albertazzi, *ACS Nano*, 2016, **10**, 1845–1852.
- T. Aida, E. W. Meijer and S. I. Stupp, *Science*, 2012, **335**, 813–817.
- S. Biswas, K. Kinbara, N. Oya, N. Ishii, H. Taguchi and T. Aida, *J. Am. Chem. Soc.*, 2009, **131**, 7556–7557.
- O. Dumele, J. Chen, J. V. Passarelli and S. I. Stupp, *Adv. Mater.*, 2020, 1907247.
- Q. Zhang, R. Toyoda, L. Pfeifer and B. L. Feringa, *J. Am. Chem. Soc.*, 2023, **145**, 6976–6985.
- Y. Yamaguchi, Y. Ochi, S. Miyamura, T. Tanaka, S. Kobayashi, T. Wakamiya, Y. Matsubara and Z. Yoshida, *J. Am. Chem. Soc.*, 2006, **128**, 4504–4505.

25 A. P. H. J. Schenning and E. W. Meijer, *Chem. Commun.*, 2005, 3245–3258.

26 M. Raynal, F. Portier, P. W. N. M. van Leeuwen and L. Bouteiller, *J. Am. Chem. Soc.*, 2013, **135**, 17687–17690.

27 C. Kulkarni, E. W. Meijer and A. R. A. Palmans, *Acc. Chem. Res.*, 2017, **50**, 1928–1936.

28 T. F. A. de Greef, M. M. J. Smulders, M. Wolffs, A. P. H. J. Schenning, R. P. Sijbesma and E. W. Meijer, *Chem. Rev.*, 2009, **109**, 5687–5754.

29 Y. Dorca, J. Matern, G. Fernández and L. Sánchez, *Isr. J. Chem.*, 2019, **59**, 869–880.

30 A. Sorrenti, J. Leira-Iglesias, A. J. Markvoort, T. F. A. de Greef and T. M. Hermans, *Chem. Soc. Rev.*, 2017, **46**, 5476–5490.

31 E. E. Greciano, J. Calbo, E. Ortí and L. Sánchez, *Angew. Chem., Int. Ed.*, 2020, **132**, 17670–17677.

32 F. Würthner, C. R. S. Möller, B. Fimmel, S. Ogi, P. Leowanawat and D. Schmidt, *Chem. Rev.*, 2016, **116**, 962–1052.

33 M. Hifsudheen, R. K. Mishra, B. Vedhanarayanan, V. K. Praveen and A. Ajayaghosh, *Angew. Chem., Int. Ed.*, 2017, **56**, 12634–12638.

34 Y. Dorca, E. E. Greciano, J. S. Valera, R. Gómez and L. Sánchez, *Chem. – Eur. J.*, 2019, **25**, 5848–5864.

35 Y. Yang, Y. Zhang and Z. Wei, *Adv. Mater.*, 2013, **42**, 6039–6049.

36 J.-M. Lehn, *Angew. Chem., Int. Ed. Engl.*, 1990, **11**, 1304–1319.

37 M. Liu, L. Zhang and T. Wang, *Chem. Rev.*, 2015, **115**, 7304–7397.

38 F. García, R. Gómez and L. Sánchez, *Chem. Soc. Rev.*, 2023, **52**, 7524–7548.

39 E. Yashima, K. Maeda and Y. Furusho, *Acc. Chem. Res.*, 2008, **41**, 1166–1180.

40 E. Yashima, K. Maeda, H. Iida and Y. Furusho, *Chem. Rev.*, 2009, **109**, 6102–6211.

41 J. Liu, J. W. Y. Lam and B. Z. Tang, *Chem. Rev.*, 2009, **109**, 5799–5867.

42 T. Nakano and Y. Okamoto, *Chem. Rev.*, 2001, **101**, 4013–4038.

43 R. Sakai, E. B. Barasa, N. Sakai, S. I. Sato, T. Satoh and T. Kakuchi, *Macromolecules*, 2012, **45**, 8221–8227.

44 E. Yashima and K. Maeda, *Macromolecules*, 2008, **41**, 3–12.

45 K. Maeda and E. Yashima, *Top. Curr. Chem.*, 2006, **265**, 47–88.

46 C. Zhang, Y. Qiu, S. Bo, F. Wang, Y. Wang, L. Liu, Y. Zhou, H. Niu, H. Dong and T. Satoh, *J. Polym. Sci., Part A: Polym. Chem.*, 2019, **57**, 1024–1031.

47 J. Shen and Y. Okamoto, *Chem. Rev.*, 2016, **116**, 1094–1138.

48 C. I. Simionescu and V. Percec, *Prog. Polym. Sci.*, 1982, **8**, 133–214.

49 M. Milton, R. Deng, A. Mann, C. Wang, D. Tang and M. Weck, *Acc. Chem. Res.*, 2018, **54**, 2397–2408.

50 J. G. Rudick and V. Percec, *Acc. Chem. Res.*, 2008, **41**, 1641–1652.

51 N. Liu, L. Zhou and Z. Q. Wu, *Acc. Chem. Res.*, 2021, **54**, 3953–3967.

52 R. P. Megens and G. Roelfes, *Chem. – Eur. J.*, 2011, **17**, 8514–8523.

53 Y. Nagata, R. Takeda and M. Sugimoto, *ACS Cent. Sci.*, 2019, **5**, 1235–1240.

54 K. Maeda, D. Hirose, M. Nozaki, Y. Shimizu, T. Mori, K. Yamanaka, K. Ogino, T. Nishimura, T. Taniguchi, M. Moro and E. Yashima, *Sci. Adv.*, 2021, **7**, eabg5381.

55 B. A. San Jose, S. Matsushita and K. Akagi, *J. Am. Chem. Soc.*, 2012, **134**, 19795–19807.

56 S. Wang, D. Hu, X. Guan, S. Cai, G. Shi, Z. Shuai, J. Zhang, Q. Peng and X. Wan, *Angew. Chem., Int. Ed.*, 2021, **60**, 21918–21926.

57 F. Freire, E. Quiñoá and R. Riguera, *Chem. Rev.*, 2016, **116**, 1242–1271.

58 J. J. L. M. Cornelissen, A. E. Rowan, R. J. M. Nolte and N. A. J. M. Sommerdijk, *Chem. Rev.*, 2001, **101**, 4039–4070.

59 E. Schwartz, M. Koepf, H. J. Kito, R. J. M. Nolte and A. E. Rowan, *Polym. Chem.*, 2011, **2**, 33–47.

60 Y. Wang, T. Harada, L. Q. Phuong, Y. Kanemitsu and T. Nakano, *Macromolecules*, 2018, **51**, 6865–6877.

61 E. Kervio, A. Hochgesand, U. E. Steiner and C. Richert, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 12074–12079.

62 A. Brewer and A. P. Davis, *Nat. Chem.*, 2014, **6**, 569–574.

63 D. M. Lovinger, *Alcohol Res. Health*, 2008, **31**, 196–214.

64 B. Liu, C. G. Pappas, J. Ottelé, G. Schaeffer, C. Jurissek, P. F. Pieters, M. Altay, I. Maric, M. C. A. Stuart and S. Otto, *J. Am. Chem. Soc.*, 2020, **142**, 4184–4192.

65 T. Kosikova, N. I. Hassan, D. B. Cordes, A. M. Z. Slawin and D. Philp, *J. Am. Chem. Soc.*, 2015, **137**, 16074–16083.

66 G. Clixby and L. Twyman, *Org. Biomol. Chem.*, 2016, **14**, 4170–4184.

67 G. De Bo, M. A. Y. Gall, S. Kuschel, J. DeWinter, P. Gerbaux and D. A. Leigh, *Nat. Nanotechnol.*, 2018, **13**, 381–385.

68 R. K. O'Reilly, A. J. Turberfield and T. R. Wilks, *Acc. Chem. Res.*, 2017, **50**, 2496–2509.

69 F. G. A. Lister, B. A. F. Le Bailly, S. J. Webb and J. Clayden, *Nat. Chem.*, 2017, **9**, 420–425.

70 M. de Poli, W. Zawodry, O. Quinonero, M. Lorch, S. J. Webb and J. Clayden, *Science*, 2016, **352**, 575–580.

71 J. Zhang, D. Luo, C. Ma and Q. Gan, *Nat. Commun.*, 2021, **12**, 2659.

72 S. M. Wales, D. T. J. Morris and J. Clayden, *J. Am. Chem. Soc.*, 2022, **144**, 2841–2846.

73 D. T. J. Morris, S. M. Wales, D. Tilly, E. H. E. Farrar, M. N. Grayson, J. W. Ward and J. Clayden, *Chem.*, 2021, **7**, 2460–2472.

74 J. E. Jones, V. Diemer, C. Adam, J. Raftery, R. E. Ruscoe, J. Sengel, M. I. Wallace, A. Bader, S. L. Cockcroft, J. Clayden and S. J. Webb, *J. Am. Chem. Soc.*, 2016, **132**, 688–695.

75 Y. Zhong, T. A. Sobiech, B. Kauffmann, B. Song, X. Li, Y. Ferrand, I. Huc and B. Gong, *Chem. Sci.*, 2023, **14**, 4759–4768.

76 J. Wang, B. Wicher, A. Méndez-Ardoy, X. Li, G. Pecastaings, T. Buffeteau, D. M. Bassani, V. Maurizot and I. Huc, *Angew. Chem., Int. Ed.*, 2021, **60**, 18461–18466.

77 M. De Zotti and J. Clayden, *Org. Lett.*, 2019, **21**, 2209–2212.



78 D. P. Tilly, J. P. Heeb, S. J. Webb and J. Clayden, *Nat. Commun.*, 2023, **14**, 2647–2653.

79 F. G. A. Lister, N. Eccles, S. J. Pike, R. A. Brown, G. F. S. Whitehead, J. Raftery, S. J. Webb and J. Clayden, *Chem. Sci.*, 2018, **9**, 6860–6870.

80 J. Solá, S. P. Fletcher, A. Castellanos and J. Clayden, *Angew. Chem., Int. Ed.*, 2010, **49**, 6836–6839.

81 J. Solá, M. Helliwell and J. Clayden, *J. Am. Chem. Soc.*, 2010, **132**, 4548–4549.

82 Y. Inai, N. Ousaka and T. Okabe, *J. Am. Chem. Soc.*, 2003, **125**, 8151–8162.

83 Y. Inai, K. Tagawa, A. Takasu, T. Hirabayashi, T. Oshikawa and M. Yamashita, *J. Am. Chem. Soc.*, 2000, **122**, 11731–11732.

84 Y. Inai, Y. Ishida, K. Tagawa, A. Takasu and T. Hirabayashi, *J. Am. Chem. Soc.*, 2002, **124**(11), 2466–2473.

85 Y. Inai and H. Komori, *Biomacromolecules*, 2004, **5**, 1231–1240.

86 N. Ousaka, Y. Takeyama, H. Iida and E. Yashima, *Nat. Chem.*, 2011, **3**, 856–861.

87 G. Lautrette, B. Wicher, B. Kauffmann, Y. Ferrand and I. Huc, *J. Am. Chem. Soc.*, 2016, **138**, 10314–10322.

88 M. Fujiki, *J. Am. Chem. Soc.*, 1994, **116**, 6017–6018.

89 A. Ohira, M. Kunitake, M. Fujiki, M. Naioto and A. Saxena, *Chem. Mater.*, 2004, **16**, 3919–3923.

90 M. Fujiki, *Chem. Rec.*, 2009, **9**, 271–298.

91 M. Sugino, *Molecular Technology*, John Wiley & Sons, Weinheim, 2018.

92 Y. Nagata, T. Nishikawa, M. Sugino, S. Sato, M. Sugiyama, L. Porcar, A. Martel, R. Inoue and N. Sato, *J. Am. Chem. Soc.*, 2018, **140**, 2722–2726.

93 Y. Nagata, T. Kuroda, K. Takagi and M. Sugino, *Chem. Sci.*, 2014, **5**, 4953–4956.

94 Y. Nagata, T. Nishikawa and M. Sugino, *Chem. Commun.*, 2018, **54**, 6867–6870.

95 Y. Nagata, T. Nishikawa, K. Terao, H. Hasegawa and M. Sugino, *J. Polym. Sci., Part A: Polym. Chem.*, 2019, **57**, 260–263.

96 T. Nishikawa, Y. Nagata and M. Sugino, *ACS Macro Lett.*, 2017, **6**, 431–435.

97 T. Yamamoto, R. Murakami, S. Komatsu and M. Sugino, *J. Am. Chem. Soc.*, 2018, **140**, 3867–3870.

98 I. Louzao, J. M. Seco, E. Quiñoá and R. Riguera, *Angew. Chem., Int. Ed.*, 2010, **49**, 1430–1433.

99 K. Maeda, K. Morino, Y. Okamoto, T. Sato and E. Yashima, *J. Am. Chem. Soc.*, 2004, **126**(13), 4329–4342.

100 E. Yashima, K. Maeda and Y. Okamoto, *Nature*, 1999, **399**, 449–451.

101 Y.-Z. Ke, Y. Nagata, T. Yamada and M. Sugino, *Angew. Chem., Int. Ed.*, 2015, **54**, 9333–9337.

102 R. Rodríguez, E. Suárez-Picado, E. Quiñoá, R. Riguera and F. Freire, *Angew. Chem., Int. Ed.*, 2020, **59**, 8616–8622.

103 Z. Fernández, B. Fernández, E. Quiñoá, R. Riguera and F. Freire, *Chem. Sci.*, 2020, **11**, 7182–7187.

104 J. J. L. M. Cornelissen, J. J. J. M. Donners, R. De Gelder, W. S. Graswinckel, G. A. Metselaar, A. E. Rowan, N. A. J. M. Sommerdijk and R. J. M. Nolte, *Science*, 2001, **293**, 676–680.

105 E. Suárez-Picado, E. Quiñoá, R. Riguera and F. Freire, *Angew. Chem., Int. Ed.*, 2020, **59**, 4537–4543.

106 R. Rodríguez, E. Rivadulla-Cendal, M. Fernández-Míguez, B. Fernández, E. Quiñoá and F. Freire, *Angew. Chem., Int. Ed.*, 2022, **61**, e202209953.

107 M. Lago-Silva, M. M. Cid, E. Quiñoá and F. Freire, *Angew. Chem., Int. Ed.*, 2023, **62**, e202303329.

108 M. M. Green, M. P. Reidy, R. D. Johnson, G. Darling, D. J. O'Leary and G. Willson, *J. Am. Chem. Soc.*, 1989, **111**, 6452–6454.

109 M. M. Green, N. C. Peterson, T. Sato, A. Teramoto, R. Cook and S. Lifson, *Science*, 1995, **268**, 1860–1866.

110 H. Gu, Y. Nakamura, T. Sato, A. Teramoto, M. M. Green, S. K. Jha, C. Andreola and M. P. Reidy, *Macromolecules*, 1998, **31**, 6362–6368.

111 M. M. Green, J. W. Park, T. Sato, A. Teramoto, S. Lifson, R. L. Selinger and J. V. Selinger, *Angew. Chem., Int. Ed.*, 1999, **38**, 3138–3154.

112 K. Cobos, E. Quiñoá, R. Riguera and F. Freire, *J. Am. Chem. Soc.*, 2018, **140**, 12239–12246.

113 S. Arias, R. Rodríguez, E. Quiñoá, R. Riguera and F. Freire, *J. Am. Chem. Soc.*, 2018, **140**, 667–674.

114 M. M. Green, B. A. Garetz, B. Munoz, H. Chang, S. Hoke and R. G. Cooks, *J. Am. Chem. Soc.*, 1995, **117**(14), 4181–4182.

115 K. Cobos, R. Rodríguez, E. Quiñoá, R. Riguera and F. Freire, *Angew. Chem., Int. Ed.*, 2020, **59**, 23724–23730.

116 V. Jain, K.-S. Cheon, K. Tang, S. Jha and M. M. Green, *Isr. J. Chem.*, 2011, **51**, 1067–1074.

117 M. Alzubi, S. Arias, R. Rodríguez, E. Quiñoá, R. Riguera and F. Freire, *Angew. Chem., Int. Ed.*, 2019, **58**, 13365–13369.

118 Z. Fernández, B. Fernández, E. Quiñoá and F. Freire, *Angew. Chem., Int. Ed.*, 2021, **60**, 9919–9924.

119 M. A. Martínez, E. E. Greciano and L. Sánchez, *Chem. – Eur. J.*, 2019, **25**, 16012–16016.

120 E. E. Greciano, R. Rodríguez, K. Maeda and L. Sánchez, *Chem. Commun.*, 2020, **56**, 2244–2247.

121 A. R. A. Palmans and E. W. Meijer, *Angew. Chem., Int. Ed.*, 2007, **46**, 8948–8968.

122 A. R. A. Palmans, J. A. J. M. Vekemans, E. E. Havinga and E. W. Meijer, *Angew. Chem., Int. Ed. Engl.*, 1997, **36**, 2648–2651.

123 M. M. J. Smulders, P. J. M. Stals, T. Mes, T. F. E. Paffen, A. P. H. J. Schenning, A. R. A. Palmans and E. W. Meijer, *J. Am. Chem. Soc.*, 2010, **132**(2), 620–626.

124 G. Liu, M. G. Humphrey, C. Zhang and Y. Zhao, *Chem. Soc. Rev.*, 2023, **52**, 4443–4487.

125 H. Komori and Y. Inai, *J. Org. Chem.*, 2007, **72**, 4012–4022.

126 M. De Zotti, V. N. Syryamina, R. Hussain, E. Longo, G. Siligardi, S. A. Dzuba, L. Stella and F. Formaggio, *ChemBioChem*, 2019, **20**, 2125–2132.

127 Y. Ishido, N. Kanbayashi, T. Okamura and K. Onitsuka, *Macromol. Rapid Commun.*, 2021, **42**, 2100250.

128 H. G. Jeon, H. K. Lee, S. Lee and K. S. Jeong, *Chem. Commun.*, 2018, **54**, 5740–5743.

129 A. Motoshige, Y. Mawatari, Y. Yoshida, R. Motoshige and M. Tabata, *Poly. Chem.*, 2014, **5**, 971–978.

130 A. Motoshige, Y. Mawatari, Y. Yoshida, C. Seki, H. Matsuyama and M. Tabata, *J. Polym. Sci., Part A: Polym. Chem.*, 2012, **50**, 3008–3015.

131 Y. Yoshida, Y. Matawari, A. Motoshige, R. Motoshige, T. Hiraoki, M. Wagner, K. Mullen and M. Tabata, *J. Am. Chem. Soc.*, 2013, **135**, 4110–4116.

132 H. Nakako, R. Nomura and T. Masuda, *Macromolecules*, 2001, **34**, 1496–1502.

133 X. Guan, S. Wang, G. Shi, J. Zhang and X. Wan, *Macromolecules*, 2021, **54**, 4592–4600.

134 Y. Zhou, C. Zhang, Y. Qiu, L. Liu, T. Yang, H. Dong, T. Satoh and Y. Okamoto, *Molecules*, 2016, **21**, 1583.

135 S. Arias, F. Freire, M. Calderón and J. Bergueiro, *Angew. Chem., Int. Ed.*, 2017, **56**, 11420–11425.

136 T. Miyagawa, A. Furuko, K. Maeda, H. Katagiri, Y. Furusho and E. Yashima, *J. Am. Chem. Soc.*, 2005, **127**, 5018–5019.

137 K. Nagai, K. Maeda, Y. Takeyama, T. Sato and E. Yashima, *Chem. – Asian J.*, 2007, **2**, 1314–1321.

138 N. Zhu, K. Nakazono and T. Takata, *Chem. Commun.*, 2016, **52**, 3647–3649.

139 G. Shi, S. Wang, X. Guan, J. Zhang and X. Wan, *Chem. Commun.*, 2018, **54**, 12081–12084.

140 S. Wang, S. Xie, H. Du, H. Zeng, J. Zhang and X. Wan, *Sci. China: Chem.*, 2023, **66**, 887–895.

141 A. P. H. J. Schenning, M. Fransen and E. W. Meijer, *Macromol. Rapid Commun.*, 2002, **23**, 265–270.

142 V. Percec, M. Peterca, J. G. Rudick, E. Aqad, M. R. Imam and P. A. Heiney, *Chem. – Eur. J.*, 2007, **13**, 9572–9581.

143 V. Percec, J. G. Rudick, M. Peterca and P. A. Heiney, *J. Am. Chem. Soc.*, 2008, **130**, 7503–7508.

144 F. Wang, C. Zhou, K. Liu, J. Yan, W. Li, T. Masuda and A. Zhang, *Macromolecules*, 2019, **52**, 8631–8642.

145 Y. Cao, L. Ren, Y. Zhang, X. Lu, X. Zhan, J. Yan, W. Li, T. Masuda and A. Zhang, *Macromolecules*, 2021, **54**, 7621–7631.

146 K. Maeda, K. Shimomura, T. Ikai, S. Kanoh and E. Yashima, *Macromolecules*, 2017, **50**, 7801–7806.

147 J. J. Tarrío, R. Rodríguez, J. Crassous, E. Quiñoá and F. Freire, *Angew. Chem., Int. Ed.*, 2023, **62**, e202307059.

148 J. K. Koe, M. Fujiki, M. Motonaga and H. Nakashima, *Chem. Commun.*, 2000, 389–390.

149 M. Fujiki, *J. Am. Chem. Soc.*, 2000, **122**, 3336–3343.

150 M. Fujiki, J. R. Koe, M. Motonaga, H. Nakashima, K. Terao and A. Teramoto, *J. Am. Chem. Soc.*, 2001, **123**, 6253–6261.

151 M. Fujiki, *Macromol. Rapid Commun.*, 2001, **22**, 539–563.

152 H. Nakashima, M. Fujiki, J. R. Koe and M. Motonaga, *J. Am. Chem. Soc.*, 2001, **123**, 1963–1969.

153 G. Hu, W. Li, Y. Hu, A. Xu, J. Yan, L. Liu, X. Zhang, K. Liu and A. Zhang, *Macromolecules*, 2013, **46**, 1124–1132.

154 K. Tang, M. M. Green, K. S. Cheon, J. V. Selinger and B. A. Garetz, *J. Am. Chem. Soc.*, 2003, **125**, 7313–7323.

155 V. Krishnasamy, W. Qu, C. Chen, H. Huo, K. Ramanagul, V. Gothandapani, G. H. Mehl, Q. Zhang and F. Liu, *Macromolecules*, 2020, **53**, 4193–4203.

156 K. A. Andreopoulou, M. Peterca, D. A. Wilson, B. E. Partridge, P. A. Heiney and V. Percec, *Macromolecules*, 2017, **50**, 5271–5284.

157 Y.-J. Sun, X.-X. Cheng, T.-F. Miao, H.-T. Ma, W. Zhang and X.-L. Zhu, *Chin. J. Polym. Sci.*, 2022, **40**, 56–66.

158 K. Murata, M. Aoki, T. Suzuki, T. Harada, H. Kawabata, T. Komori, F. Ohseto, K. Ueda and S. Shinkai, *J. Am. Chem. Soc.*, 1994, **116**, 6664–6676.

159 P. Xing, Y. Li, Y. Wang, P.-Z. Li, H. Chen, S. Z. F. Phua and Y. Zhao, *Angew. Chem., Int. Ed.*, 2018, **57**, 7774–7779.

160 W. Qi, C. Ma, P. Liao, H. Li, T. Gu, T. Wu, J. Huang and Y. Yan, *Adv. Optical Mater.*, 2023, **11**, 2201229.

161 N. J. V. Zee, B. Adelizzi, M. F. J. Mabesoone, X. Meng, A. Aloi, H. Zha, M. Lutz, I. A. W. Filot, A. R. A. Palmans and E. W. Meijer, *Nature*, 2018, **558**, 100–103.

162 N. J. V. Zee, M. F. J. Mabesoone, B. Adelizzi, A. R. A. Palmans and E. W. Meijer, *J. Am. Chem. Soc.*, 2020, **142**, 20191–20200.

163 M. Go, H. Choi, K. Y. Kim, C. J. Moon, Y. Choi, H. Miyake, S. S. Lee, S. H. Jung, M. Y. Choi and J. H. Jung, *Org. Chem. Front.*, 2019, **6**, 1100–1108.

164 C. Kulkarny, P. A. Korevaar, K. K. Bejagam, A. R. A. Palmans, E. W. Meijer and S. J. George, *J. Am. Chem. Soc.*, 2017, **139**, 13867–13875.

165 M. Peterca, M. R. Iman, C.-H. Ahn, V. S. Balagurusamy, D. A. Wilson, B. M. Rosen and V. Percec, *J. Am. Chem. Soc.*, 2011, **133**, 2311–2328.

166 L. Gao, C. Xing, X. Dou, Y. Zou, C. Zhao and C. Feng, *Angew. Chem., Int. Ed.*, 2022, **61**, e202211812.

167 A. M. García and A. Ruiz-Carretero, *Chem. Commun.*, 2022, **58**, 529–532.

168 Y. Zhang, H. Li, Z. Geng, W.-H. Zheng, Y. Quan and Y. Cheng, *ACS Nano*, 2022, **16**, 3173–3181.

169 H. Juwarker and K. S. Jeong, *Chem. Soc. Rev.*, 2010, **39**, 3664–3674.

170 K. J. Chang, B. N. Kang, M. H. Lee and K.-S. Jeong, *J. Am. Chem. Soc.*, 2005, **127**, 12214–12215.

171 M. C. Kim, J. Suk, H. J. Kim, E. Sim, M. Lee, K.-S. Jeong and V. R. Naidu, *Org. Lett.*, 2008, **10**, 5373–5376.

172 J. M. Suk, V. R. Naidu, X. Liu, M. S. Lah and K. S. Jeong, *J. Am. Chem. Soc.*, 2011, **133**, 13938–13941.

173 D. A. Kim, P. Kang, M. G. Choi and K. S. Jeong, *Chem. Commun.*, 2013, **49**, 9743–9745.

174 D. Mondal, M. Ahmad, B. Dey, A. Mondal and P. Talukdar, *Nat. Commun.*, 2022, **13**, 6507.

175 Q. Li, F. Huang, J. Shang, Y. Che, Y. Wang, W. Zhao and H. Jiang, *Tetrahedron Lett.*, 2016, **57**, 1691–1694.

176 J. V. Gavette, C. J. Evoniuk, L. N. Zakharov, M. E. Carnes, M. M. Haley and D. W. Johnson, *Chem. Sci.*, 2014, **5**, 2899–2905.

177 R. Wechsel, M. Źabka, J. W. Ward and J. Clayden, *J. Am. Chem. Soc.*, 2018, **140**, 3528–3531.

178 H. Miyake, H. Kamon, I. Miyahara, H. Sugimoto and H. Tsukube, *J. Am. Chem. Soc.*, 2008, **130**, 792–793.

179 H.-J. Kim, W.-C. Zin and M. Lee, *J. Am. Chem. Soc.*, 2004, **126**, 7009–7014.

180 H.-J. Kim, J.-H. Lee and M. Lee, *Angew. Chem., Int. Ed.*, 2005, **44**, 5810–5814.

181 H. Maeda, W. Hane, Y. Bando, Y. Terashima, Y. Haketa, H. Shibaguchi, T. Kawai, M. Naito, K. Takaishi,



M. Uchiyama and A. Muranaka, *Chem. – Eur. J.*, 2013, **19**, 16263–16271.

182 R. Kakuchi, S. Nagata, Y. Tago, R. Sakai, I. Otsuka, T. Satoh and T. Kakuchi, *Macromolecules*, 2009, **42**, 1476–1481.

183 R. Kakuchi, S. Nagata, R. Sakai, I. Otsuka, H. Nakade, T. Satoh and T. Kakuchi, *Chem. – Eur. J.*, 2008, **14**, 10259–10266.

184 R. Sakai, S. Okade, E. B. Barasa, R. Kakuchi, M. Ziabka, S. Umeda, K. Tsuda, T. Satoh and T. Kakuchi, *Macromolecules*, 2010, **43**, 7406–7411.

185 R. Kakuchi, T. Kodama, R. Shimada, Y. Tago, R. Sakai, T. Satoh and T. Kakuchi, *Macromolecules*, 2009, **42**, 3892–3897.

186 Y. Qu, J. Hua, Y. Jiang and H. Tian, *J. Polym. Sci., Part A: Polym. Chem.*, 2009, **47**, 1544–1552.

187 R. Kakuchi, Y. Tago, R. Sakai, T. Satoh and T. Kakuchi, *Macromolecules*, 2009, **42**, 4430–4435.

188 R. Kakuchi, R. Shimada, Y. Tago, R. Sakai, T. Satoh and T. Kakuchi, *J. Polym. Sci., Part A: Polym. Chem.*, 2010, **48**, 1683–1689.

189 S. Leiras, E. Suárez-Picado, E. Quiñoá, R. Riguera and F. Freire, *Giant*, 2021, **7**, 100068.

190 Y. Gao, T. Gao, L. Wang, X. Ma, R. Jin, C. Kang and L. Gao, *New J. Chem.*, 2021, **45**, 5093.

191 Y. Qiu, Y. Zhang, Q. Jiang, H. Wang, Y. Liao, H. Zhou and X. Xie, *Macromolecules*, 2022, **55**, 9057–9065.

192 M. H.-Y. Chan, S. Y.-L. Leung and V. W.-W. Yam, *J. Am. Chem. Soc.*, 2019, **141**, 12312–12321.

193 J. Liu, C. Sun, W. Ma, Y.-J. Lu, L. Yu, K. Zhang and H. Zeng, *RSC Adv.*, 2014, **4**, 54469–54473.

194 Y.-C. Lin and C.-T. Chen, *Chem. – Eur. J.*, 2012, **19**, 2531–2538.

195 S. Y.-L. Leung, A. Y.-Y. Tam, C.-H. Tao, H. S. Chow and V. W.-W. Yam, *J. Am. Chem. Soc.*, 2011, **134**, 1047–1056.

196 S. Akine, T. Matsumoto and T. Nabeshima, *Chem. Commun.*, 2008, 4604–4606.

197 S. Akine, S. Hotate and T. Nabeshima, *J. Am. Chem. Soc.*, 2011, **133**, 13868–13871.

198 S. Akine, S. Sairenji, T. Taniguchi and T. Nabeshima, *J. Am. Chem. Soc.*, 2013, **135**, 12948–12951.

199 S. Sairenji, S. Akine and T. Nabeshima, *Dalton Trans.*, 2016, **45**, 14902–14906.

200 S. Kawano, K. Narita, Y. Ikemoto, A. Sasaki and K. Tanaka, *Chem. Commun.*, 2022, **58**, 3274–3277.

201 D. Bai, T. Yan, S. Wang, Y. Wang, J. Fu, X. Fang, J. Zhu and J. Liu, *Angew. Chem., Int. Ed.*, 2020, **59**, 13602–13607.

202 M. Baskin, H. Zhu, Z.-W. Qu, J. H. Chill, S. Grimme and G. Maayan, *Chem. Sci.*, 2019, **10**, 620–632.

203 M. Horeau, G. Lautrette, B. Wicher, V. Blot, J. Lebreton, M. Pipelier, D. Dubreuil, Y. Ferrand and I. Huc, *Angew. Chem., Int. Ed.*, 2017, **56**, 6823–6827.

204 P. Mateus, B. Wicher, Y. Ferrand and I. Huc, *Chem. Commun.*, 2017, **53**, 9300–9303.

205 K. Miwa, Y. Furusho and E. Yashima, *Nat. Chem.*, 2010, **2**, 444–449.

206 D. Taura, K. Shimizu, C. Yokota, R. Ikeda, Y. Suzuki, H. Iida, N. Ousaka and E. Yashima, *Chem. Commun.*, 2019, **55**, 12084–12087.

207 B. Maiti, S. Bhattacharjee and S. Bhattacharya, *Nanoscale*, 2019, **11**, 2223–2230.

208 M. H.-Y. Chan, M. Ng, S. Y.-L. Leung, W. H. Lam and V. W.-W. Yam, *J. Am. Chem. Soc.*, 2017, **139**, 8639–8645.

209 X. Yu, Z. Wang, Y. Li, L. Geng, J. Ren and G. Feng, *Inorg. Chem.*, 2017, **56**, 7512–7518.

210 F. Wang and C.-L. Feng, *Angew. Chem., Int. Ed.*, 2018, **57**, 5655–5659.

211 S. Akine, H. Nagumo and T. Nabeshima, *Inorg. Chem.*, 2012, **51**, 5506–5508.

212 C. Liu, K. Fu, S. Zheng, L. Yao, W. Zhou and G. Liu, *Adv. Opt. Mater.*, 2023, **11**, 2300019.

213 S. Kawabata, N. Ousaka and E. Yashima, *Chem. Commun.*, 2018, **54**, 2417–2420.

214 Z. Li, H. Yang, K. M. Adam, H. Yao, T. Wei, Y. Zhang and Q. Lin, *ChemPlusChem*, 2021, **86**, 146–154.

215 F. Wang, W. Ji, P. Yang and C.-L. Feng, *ACS Nano*, 2019, **13**, 7281–7290.

216 G. Liu, J. Sheng, W. L. Teo, G. Yang, H. Wu, Y. Li and Y. Zhao, *J. Am. Chem. Soc.*, 2018, **140**, 16275–16283.

217 S. H. Park, S. H. Jung, J. Ahn, J. H. Lee, K.-Y. Kwon, J. Jeon, H. Kim, J. Jaworski and J. H. Jung, *Chem. Commun.*, 2014, **50**, 13495–13498.

218 M. Fukuda, R. Rodríguez, Z. Fernández, T. Nishimura, D. Hirose, G. Watanabe, E. Quiñoá, F. Freire and K. Maeda, *Chem. Commun.*, 2019, **55**, 7906–7909.

219 F. Freire, J. M. Seco, E. Quiñoá and R. Riguera, *Hierarchical Macromolecular Structures: 60 Years after the Staudinger Nobel Prize II*, 2013, pp. 123–140.

220 F. Freire, J. M. Seco, E. Quiñoá and R. Riguera, *Angew. Chem., Int. Ed.*, 2011, **50**, 11692–11696.

221 F. Freire, J. M. Seco, E. Quiñoá and R. Riguera, *J. Am. Chem. Soc.*, 2012, **134**, 19374–19383.

222 S. Arias, F. Freire, E. Quiñoá and R. Riguera, *Angew. Chem., Int. Ed.*, 2014, **53**, 13720–13724.

223 R. Rodríguez, S. Arias, E. Quiñoá, R. Riguera and F. Freire, *Nanoscale*, 2017, **9**, 17752–17757.

224 S. Arias, F. Freire, E. Quiñoá and R. Riguera, *Polym. Chem.*, 2015, **6**, 4725–4733.

225 D. Hirose, A. Isobe, E. Quiñoá, F. Freire and K. Maeda, *J. Am. Chem. Soc.*, 2019, **141**, 8592–8598.

226 M. Fukuda, T. Nishimura, D. Hirose and K. Maeda, *Chem. Lett.*, 2023, **52**, 136–139.

227 J. Bergueiro, F. Freire, E. P. Wendler, J. M. Seco, E. Quiñoá and R. Riguera, *Chem. Sci.*, 2014, **5**, 2170–2176.

228 S. Arias, J. Bergueiro, F. Freire, E. Quiñoá and R. Riguera, *Small*, 2015, **12**, 238–244.

229 M. Alzubi, S. Arias, I. Louzao, E. Quiñoá, R. Riguera and F. Freire, *Chem. Commun.*, 2017, **53**, 8573–8576.

230 S. Arias, M. Núñez-Martínez, E. Quiñoá, R. Riguera and F. Freire, *Polym. Chem.*, 2017, **8**, 3740–3745.

231 Y.-H. Chiu, O. dos Santos and J. W. Canary, *Tetrahedron*, 1999, **55**, 12069–12078.

232 S. Zahn and J. W. Canary, *Science*, 2000, **288**, 1404–1407.

233 J. Liang and J. W. Canary, *Chirality*, 2010, **23**, 24–33.

234 F. Wendt, C. Näther and F. Tuczek, *J. Biol. Inorg. Chem.*, 2016, **21**, 777–792.

235 M. Gaedke, F. Witte, J. Anhäuser, H. Hupatz, H. V. Schröder, A. Valkonen, K. Rissanen, A. Lützen, B. Paulus and C. A. Schalley, *Chem. Sci.*, 2019, **10**, 10003–10009.

236 L. Faour, C. Adam, C. Gautier, S. Goeb, M. Allain, E. Levillain, D. Canevet and M. Sallé, *Chem. Commun.*, 2019, **55**, 5743–5746.

237 Y. Wang, M. Frasconi, W.-G. Liu, Z. Liu, A. A. Sarjeant, M. S. Nassar, Y. Y. Botros, W. A. Goddard III and J. F. Stoddart, *J. Am. Chem. Soc.*, 2015, **137**, 876–885.

238 Y. Zhang, M. Qin, C. Xing, C. Zhao, X. Dou and C. Feng, *ACS Nano*, 2020, **14**, 17151–17162.

239 J. Yan, J. Liu, H. Lei, Y. Kang, C. Zhao and Y. Fang, *J. Colloid Interface Sci.*, 2015, **448**, 374–379.

240 B. Adhikari and H. B. Kraatz, *Chem. Commun.*, 2014, **50**, 5551–5553.

241 S. Kawano, N. Fujita and S. Shinkai, *Chem. – Eur. J.*, 2005, **11**, 4735–4742.

242 Y. Gao, J. Lu, J. Wu, J. Hu and Y. Ju, *RSC Adv.*, 2014, **4**, 63539–63543.

243 L.-Y. Guo, H. Zhang, S.-T. Huang, M. Di, G. Yao and S. Wu, *e-Polymers*, 2019, **19**, 377–384.

244 Z.-Q. Wu, X. Song, Y.-X. Li, L. Zhou, Y.-Y. Zhu, Z. Chen and N. Liu, *Nat. Commun.*, 2023, **14**, 566–577.

245 E. Gomar-Nadal, J. Veciana, C. Rovira and D. B. Amabilino, *Adv. Mater.*, 2005, **17**, 2095–2098.

246 J. Bergueiro, M. Núñez-Martínez, S. Arias, E. Quiñoá, R. Riguera and F. Freire, *Nanoscale Horiz.*, 2020, **5**, 495–500.

247 M. Núñez-Martínez, S. Arias, J. Bergueiro, E. Quiñoá, R. Riguera and F. Freire, *Macromol. Rapid Commun.*, 2022, **43**, 2100616.

248 M. Núñez-Martínez, S. Arias, E. Quiñoá, R. Riguera and F. Freire, *Chem. Mater.*, 2021, **33**, 4805–4812.

249 M. Núñez-Martínez, E. Quiñoá and F. Freire, *Nanoscale*, 2022, **14**, 13066–13072.

250 H. Iida, T. Mizoguchi, S.-D. Oh and E. Yashima, *Polym. Chem.*, 2010, **1**, 841–848.

251 E. Ohta, H. Sato, S. Ando, A. Kosaka, T. Fukushima, D. Hashizume, M. Yamasaki, K. Hasegawa, A. Muraoka, H. Ushiyama, K. Yamashita and T. Aida, *Nat. Chem.*, 2010, **3**, 68–73.

252 S. Liang, G. Chen and Y. Zhao, *J. Mater. Chem. C*, 2013, **1**, 5477–5490.

253 S. Liang, G. Chen, J. Peddle and Y. Zhao, *Chem. Commun.*, 2012, **48**, 3100–3102.

254 S. Anantharaj and M. Jayakannan, *J. Polym. Sci., Part A: Polym. Chem.*, 2016, **54**, 2864–2875.

255 H. M. D. Bandara and S. C. Burdette, *Chem. Soc. Rev.*, 2012, **41**, 1809–1825.

256 M. Irie, *Chem. Rev.*, 2000, **100**, 1685–1716.

257 M. Irie, T. Fukaminato, K. Matsuda and S. Kobakate, *Chem. Rev.*, 2014, **114**, 12174–12277.

258 K. Matsuda and M. Irie, *J. Photochem. Photobiol. C*, 2004, **5**, 169–182.

259 M. Li and W. H. Zhu, *Acc. Chem. Res.*, 2022, **55**, 3136–3149.

260 G. Likhtenshtein, *Stilbenes: Applications in Chemistry, Life Sciences and Materials Science*, Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim, 2009.

261 T. Dünnebacke, K. K. Kartha, J. M. Wiest, R. Q. Albuquerque and G. Fernández, *Chem. Sci.*, 2020, **11**, 10405–10413.

262 R. Costil, M. Holsheimer, S. Crespi, N. A. Simeth and B. L. Feringa, *Chem. Rev.*, 2021, **121**, 13213–13237.

263 J. C. M. Kistemaker, A. S. Lubbe and B. L. Feringa, *Mater. Chem. Front.*, 2021, **5**, 2900–2906.

264 D. R. S. Pooler, A. S. Lubbe, S. Crespi and B. L. Feringa, *Chem. Sci.*, 2021, **12**, 14964–14986.

265 B. L. Feringa, *J. Org. Chem.*, 2007, **72**, 6635–6652.

266 S. Kassem, T. Van Leeuwen, A. S. Lubbe, M. R. Wilson and B. L. Feringa, *Nat. Rev. Chem.*, 2017, **1**, 0096.

267 X. Su and I. Aprahamian, *Org. Lett.*, 2011, **13**, 30–33.

268 B. Shao and I. Aprahamian, *Chem.*, 2020, **6**, 2162–2173.

269 F. Xu and B. L. Feringa, *Adv. Mater.*, 2023, **35**, 2204413.

270 D. Pijper and B. L. Feringa, *Angew. Chem., Int. Ed.*, 2007, **46**, 3693–3696.

271 D. Pijper, M. G. M. Jongejam, A. Meetsma and B. L. Feringa, *J. Am. Chem. Soc.*, 2008, **130**, 4541–4552.

272 T. Van Leeuwen, G. H. Heideman, D. Zhao, S. J. Wezenberg and B. L. Feringa, *Chem. Commun.*, 2017, **53**, 6393–6396.

273 M. Müllerand and R. Zentel, *Macromolecules*, 1994, **27**, 4404–4406.

274 G. Maxeinand and R. Zentel, *Macromolecules*, 1995, **28**, 8438–8440.

275 M. Müllerand and R. Zentel, *Macromolecules*, 1996, **29**, 1609–1617.

276 S. Mayer, G. Maxeinand and R. Zentel, *Macromolecules*, 1998, **31**, 8522–8525.

277 S. Mayerand and R. Zentel, *Macromol. Chem. Phys.*, 1998, **199**, 1675–1682.

278 X. Yao, T. Li, J. Wang, X. Ma and H. Tian, *Adv. Opt. Mater.*, 2016, **4**, 1322–1349.

279 K. M. Vonk, E. W. Meijer and G. Vantomme, *Chem. Sci.*, 2021, **12**, 13572–13579.

280 E. Weyandt, G. M. Ter Huurne, G. Vantomme, A. J. Markvoort, A. R. A. Palmans and E. W. Meijer, *J. Am. Chem. Soc.*, 2020, **142**, 6295–6303.

281 Y. Cai, Z. Guo, J. Chen, W. Li, L. Zhong, Y. Gao, L. Jiang, L. Chi, H. Tian and W. H. Zhu, *J. Am. Chem. Soc.*, 2016, **138**, 2219–2224.

282 B. A. San Jose, T. Ashibe, N. Tada, S. Yorozuka and K. Akagi, *Adv. Funct. Mater.*, 2014, **24**, 6166–6171.

283 F. K. C. Leung, T. Van Den Enk, T. Kajitani, J. Chen, M. C. A. Stuart, J. Kuipers, T. Fukushima and B. L. Feringa, *J. Am. Chem. Soc.*, 2018, **140**, 17724–17733.

284 S. Chen, L. Yang, F. K. C. Leung, T. Kajitani, M. C. A. Stuart, T. Fukushima, P. Van Rijn and B. L. Feringa, *J. Am. Chem. Soc.*, 2022, **144**, 3543–3553.

285 F. Xu, L. Pfeifer, S. Crespi, F. K. C. Leung, M. C. A. Stuart, S. J. Wezenberg and B. L. Feringa, *J. Am. Chem. Soc.*, 2021, **143**, 5990–5997.

286 T. S. C. MacDonald, B. L. Feringa, W. S. Price, S. J. Wezenberg and J. E. Beves, *J. Am. Chem. Soc.*, 2020, **142**, 20014–20020.



287 S. Chen, S. Chen, F. K. C. Leung, M. C. A. Stuart, C. Wang and B. L. Feringa, *J. Am. Chem. Soc.*, 2020, **142**, 10163–10172.

288 F. Xu, S. Crespi, G. Pacella, Y. Fu, M. C. A. Stuart, Q. Zhang, G. Portale and B. L. Feringa, *J. Am. Chem. Soc.*, 2022, **144**, 6019–6027.

289 S. Yagai, M. Yamauchi, A. Kobayashi, T. Karatsu, A. Kitamura, T. Ohba and Y. Kikkawa, *J. Am. Chem. Soc.*, 2012, **134**, 18205–18208.

290 A. Suzuki, K. Aratsu, S. Datta, N. Shimizu, H. Takagi, R. Haruki, S.-I. Adachi, M. Hollambi, F. Silly and S. Yagai, *J. Am. Chem. Soc.*, 2010, **141**, 13196–13202.

291 D. Zhao, T. Van Leeuwen, J. Cheng and B. L. Feringa, *Nat. Chem.*, 2017, **9**, 250–256.

292 D. Mazzier, M. Crisma, M. De Poli, G. Marafon, C. Peggion, J. Clayden and A. Moretto, *J. Am. Chem. Soc.*, 2016, **138**, 8007–8018.

293 M. Pollastrani, G. Marafon, J. Clayden and A. Moretto, *Chem. Commun.*, 2021, **57**, 2269–2272.

294 S. Pramanik, B. Kauff, S. Hecht, Y. Ferrand and I. Huc, *Chem. Commun.*, 2021, **57**, 93–96.

295 Z. Yu and S. Hecht, *Chem. Commun.*, 2016, **52**, 6639–6653.

296 O. Pieroni, J. L. Houben, A. Fissi, P. Costantino and F. Ciardelli, *J. Am. Chem. Soc.*, 1980, **102**, 5913–5915.

297 J. L. Houben, A. Fissi, D. Bacciola, N. Rosato, O. Pieroni and F. Ciardelli, *Int. J. Biol. Macromol.*, 1983, **5**, 94–100.

298 O. Pieront, D. Fabbri, A. Fissi and F. Ciardelli, *Macromol. Rapid Commun.*, 1988, **9**, 637–640.

299 A. Ueno, J.-i Anzai, T. Osa and Y. Kadoma, *Bull. Chem. Soc. Jpn.*, 1979, **52**, 549–554.

300 A. Ueno, K. Takahashi, J. Anzai and T. Osa, *J. Am. Chem. Soc.*, 1981, **103**, 6410–6415.

301 A. Ueno, K. Adachi, J. Nakamura and T. Osa, *J. Polym. Sci., Part A: Polym. Chem.*, 1990, **28**, 1161–1170.

302 F. Ciardelli, D. Fabbri, O. Pieroni and A. Fissi, *J. Am. Chem. Soc.*, 1989, **111**, 3470–3472.

303 E. D. King, P. Tao, T. T. Sanan, C. M. Hadad and J. R. Parquette, *Org. Lett.*, 2008, **10**, 1671–1674.

304 C. J. Gabriel and J. R. Parquette, *J. Am. Chem. Soc.*, 2006, **128**, 13708–13709.

305 Y. Hua and A. H. Flood, *J. Am. Chem. Soc.*, 2010, **132**, 12838–12840.

306 Y. Wang, F. Bie and H. Jiang, *Org. Lett.*, 2010, **12**, 3630–3633.

307 F. C. Parks, Y. Liu, S. Debnath, R. Stutsman, K. Raghavachari and A. H. Flood, *J. Am. Chem. Soc.*, 2018, **140**, 17711–17723.

308 A. Khan, C. Kaiser and S. Hecht, *Angew. Chem., Int. Ed.*, 2006, **45**, 1878–1881.

309 Z. Yu and S. Hecht, *Angew. Chem., Int. Ed.*, 2011, **50**, 1640–1643.

310 J. Moore and C. Ray, *Adv. Polym. Sci.*, 2005, **177**, 91–149.

311 S. Hecht and A. Khan, *Angew. Chem., Int. Ed.*, 2003, **115**, 6203–6206.

312 A. Khan and S. Hecht, *Chem. – Eur. J.*, 2006, **12**, 4764–4774.

313 Z. Yu, S. Weidner, T. Risse and S. Hecht, *Chem. Sci.*, 2013, **4**, 4156–4167.

314 Z. Yu and S. Hecht, *J. Polym. Sci., Part A: Polym. Chem.*, 2015, **53**, 313–318.

315 R. B. Prince, J. G. Saven, P. G. Wolynes and J. S. Moore, *J. Am. Chem. Soc.*, 1999, **121**, 3114–3121.

316 S. Hecht and J. M. J. Fréchet, *Angew. Chem., Int. Ed.*, 2001, **40**, 74–91.

317 P. D. Kiser, M. Golczakak and K. Palczewski, *Chem. Rev.*, 2014, **114**, 194–232.

318 J. P. Riehl and F. S. Richardson, *Chem. Rev.*, 1986, **86**(1), 1–16.

319 J. P. Riehl, *Chiroptical spectroscopy, emission theory*, Elsevier Ltd., 3rd edn, 2016.

320 Y. Zhang, S. Yu, B. Han, Y. Zhou, X. Zhang, X. Gao and Z. Tang, *Matter*, 2022, **5**, 837–875.

321 G. Longhi, E. Castiglioni, J. Koshoubu, G. Mazzeo and S. Abatte, *Chirality*, 2016, **28**, 696–707.

322 S. Yang, S. Zhang, F. Hu, J. Han and F. Li, *Coord. Chem. Rev.*, 2023, **485**, 215116.

323 E. M. Sánchez-Carretero, A. R. Agarrabeitia, F. Moreno, B. L. Maroto, G. Muller, M. J. Ortiz and S. de la Moya, *Chem. – Eur. J.*, 2015, **21**, 13488–13500.

324 L. Arrico, L. di Bari and F. Zinna, *Chem. – Eur. J.*, 2021, **27**, 2920–2934.

325 S. Y. Li, L. Xu, R. T. Gao, Z. Chen, N. Liu and Z. Q. Wu, *J. Mater. Chem. C*, 2022, **11**, 1242–1250.

326 K. Watanabe and K. Akagi, *Sci. Technol. Adv. Mater.*, 2014, **15**, 044203.

327 X. Liu and R. H. Jin, *Chem. Synth.*, 2022, 1–29.

328 Y. Sang, J. Han, T. Zhao, P. Duan and M. Liu, *Adv. Mater.*, 2020, **32**, 1900110.

329 J. Kumar, T. Nakashima and T. Kawai, *J. Phys. Chem. Lett.*, 2015, **6**, 3445–3452.

330 K. Dhbaibi, L. Abella, S. Meunier-Della-Gatta, T. Roisnel, N. Vanthuyne, B. Jamoussi, G. Pieters, B. Racine, E. Quesnel, J. Autschbach, J. Crassous and L. Favereau, *Chem. Sci.*, 2021, **12**, 5522–5533.

331 J. R. Brandt, X. Wang, Y. Yang, A. J. Campbell and M. J. Füchter, *J. Am. Chem. Soc.*, 2016, **138**, 9743–9746.

332 Y. Yang, R. Correia da Costa, D. M. Smilgies, A. J. Campbell and M. J. Füchter, *Adv. Mater.*, 2013, **25**, 2624–2628.

333 L. E. MacKenzie and R. Pal, *Nat. Rev. Chem.*, 2021, **5**, 109–124.

334 P. Stachelek, L. MacKenzie, D. Parker and R. Pal, *Nat. Commun.*, 2022, **13**, 1–8.

335 C. He and Y. Li, *Chin. Chem. Lett.*, 2022, 108077.

336 J. S. Kang, N. Kim, T. Kim, M. Seo and B. S. Kim, *Macromol. Rapid Commun.*, 2022, **43**, 1–21.

337 J. Li, G. B. Schuster, K. S. Cheon, M. M. Green and J. V. Selinger, *J. Am. Chem. Soc.*, 2000, **122**, 2603–2612.

338 G. Iftime, F. L. Labarthet, A. Natansohn and P. Rochon, *J. Am. Chem. Soc.*, 2000, **122**, 12646–12650.

339 G. Li, M. Xu, S. Zhang, G. Yang and W. Li, *Macromol. Rapid Commun.*, 2022, **43**, 2100904.

340 C. Kulkarni, R. H. N. Curvers, G. Vantomme, D. J. Broer, A. R. A. Palmans, S. C. J. Meskers and E. W. Meijer, *Adv. Mater.*, 2021, **33**, 1–7.

341 H. Sakaino, B. A. G. Lamers, S. C. J. Meskers, E. W. Meijer and G. Vantomme, *J. Polym. Sci.*, 2021, **59**, 1131–1141.

342 E. Moulin, J. J. Armao IV and N. Giuseppone, *Acc. Chem. Res.*, 2019, **52**, 975–983.

343 J. Kim, J. Lee, W. Y. Kim, H. Kim, S. Lee, H. C. Lee, Y. S. Lee, M. Seo and S. Y. Kim, *Nat. Commun.*, 2015, **6**, 6959.

344 J. S. Kang, S. Kang, J. M. Suh, S. M. Park, D. K. Yoon, M. H. Lim, W. Y. Kim and M. Seo, *J. Am. Chem. Soc.*, 2022, **144**, 2657–2666.

345 Q. Sallembien, P. Aoun, S. Blanchard, L. Bouteiller and M. Raynal, *Chem. – Eur. J.*, 2023, **29**, e202203199.

346 J. Hu, Y. Xie, H. Zhang, C. He, Q. Zhang and G. Zou, *Chem. Commun.*, 2019, **55**, 4953–4956.

347 H. Wang, Y. Liu, L. Wang, B. Raktani, T. Yang, K. M. Tran, Y. Luo, J. Yu, H. Lee and H. Lee, *ACS Mater. Lett.*, 2022, **4**, 1954–1961.

348 Y. Marcus, *Chem. Soc. Rev.*, 1993, **22**, 409–416.

349 G. Guichard and I. Huc, *Chem. Commun.*, 2011, **47**, 5933.

350 D. J. Hilland and J. S. Moore, *Proc. Natl. Acad. Sci. U. S. A.*, 2002, **99**, 5053.

351 J.-L. Hou, M.-X. Jia, X.-K. Jiang, Z.-T. Liand and G.-J. Chen, *J. Org. Chem.*, 2004, **69**, 6228.

352 A. J. Zychand and B. L. Iverson, *J. Am. Chem. Soc.*, 2000, **122**, 8898.

353 I. Huc, *Eur. J. Org. Chem.*, 2004, 17–29.

354 T. Qi, V. Maurizot, H. Noguchi, T. Charoenraks, B. Kauffmann, M. Takafuji, H. Iharacg and I. Huc, *Chem. Commun.*, 2012, **48**, 6337–6339.

355 M. Kudo, V. Maurizot, B. Kauffmann, A. Tanatani and I. Huc, *J. Am. Chem. Soc.*, 2013, **135**, 9628–9631.

356 C.-F. Wu, Z.-M. Li, X.-N. Xu, Z.-X. Zhao, X. Zhao, R.-X. Wang and Z.-T. Li, *Chem. – Eur. J.*, 2014, **20**, 1418–1426.

357 Y. Liu, F. C. Parks, W. Zhao and A. H. Flood, *J. Am. Chem. Soc.*, 2018, **140**, 15477–15486.

358 B. A. F. Le Bailly and J. Clayden, *Chem. Commun.*, 2016, **52**, 4852–4863.

359 M. Tomsett, I. Maffucci, B. A. F. Le Bailly, L. Byrne, S. M. Bijvoets, M. G. Lizio, J. Raftery, C. P. Butts, S. J. Webb, A. Contini and J. Clayden, *Chem. Sci.*, 2017, **8**, 3007–3018.

360 D. Zwaag, P. A. Pieters, P. A. Korevaar, A. J. Markvoort, A. J. H. Spiering, T. F. A. de Greef and E. W. Meijer, *J. Am. Chem. Soc.*, 2015, **137**, 12677–12688.

361 L. Brunsved, B. J. Folmer, E. W. Meijer and R. P. Sijbesma, *Chem. Rev.*, 2001, **101**, 4071–4098.

362 S. I. Stupp and L. C. Palmer, *Chem. Mater.*, 2014, **26**, 507–518.

363 J. Matern, Y. Dorca and L. Sánchez, *Angew. Chem., Int. Ed.*, 2019, **58**, 16730–16740.

364 M. F. J. Mabesoone, A. R. A. Palmans and E. W. Meijer, *J. Am. Chem. Soc.*, 2020, **142**, 19781–19798.

365 M. A. J. Gillissen, M. M. E. Koenigs, J. J. H. Spiering, J. A. J. M. Vekemans, A. R. A. Palmans, I. K. Voets and E. W. Meijer, *J. Am. Chem. Soc.*, 2014, **136**, 336–343.

366 P. Xing, H. Chen, L. Bai, A. Hao and Y. Zhao, *ACS Nano*, 2016, **10**, 2716–2727.

367 P. Xing and Y. Zhao, *Acc. Chem. Res.*, 2018, **51**, 2324–2334.

368 C. Liu, D. Yang, L. Zhang and M. Liu, *Soft Matter*, 2019, **15**, 6557–6563.

369 M. A. Martínez, A. Doncel-Giménez, J. Cerdá, J. Calbo, R. Rodríguez, J. Aragó, J. Crassous, E. Ortí and L. Sánchez, *J. Am. Chem. Soc.*, 2021, **143**, 13281–13291.

370 K. K. L. Cheuk, J. W. Y. Lam, J. Chen, L. M. Lai and B. Z. Tang, *Macromolecules*, 2003, **36**, 5947–5959.

371 K. Okoshi, S. Sakurai, S. Ohsawa, J. Kumaki and E. Yashima, *Angew. Chem., Int. Ed.*, 2006, **45**, 8173–8176.

372 S. Sakurai, K. Okoshi, J. Kumaki and E. Yashima, *J. Am. Chem. Soc.*, 2006, **128**, 5650–5651.

373 S. Sakurai, K. Okoshi, J. Kumaki and E. Yashima, *Angew. Chem.*, 2006, **118**, 1267–1270.

374 T. Fukushima, K. Takachi and K. Tsuchihara, *Macromolecules*, 2006, **39**, 3103–3105.

375 T. Fukushima, H. Kimura and K. Tsuchihara, *Macromolecules*, 2009, **42**, 8619–8626.

376 T. Fukushima and K. Tsuchihara, *Macromolecules*, 2009, **42**, 5453–5460.

377 K. K. L. Cheuk, J. W. Y. Lam, B. S. Li, Y. Xie and B. Z. Tang, *Macromolecules*, 2007, **40**, 2633–2642.

378 K. Maeda, H. Mochizuki, M. Watanabe and E. Yashima, *J. Am. Chem. Soc.*, 2006, **128**, 7639–7650.

379 C. Zhang, F. Liu, Q. Geng, S. Zhang, X. Shen, R. Kakuchi, H. Misaka, T. Kakuchi, T. Satoh and R. Sakai, *Eur. Polym. J.*, 2011, **47**, 1923–1930.

380 H. Zhao, F. Sanda and T. Masuda, *Macromol. Chem. Phys.*, 2005, **206**, 1653–1658.

381 H. Zhao, F. Sanda, T. J. Masuda, H. Zhao, F. Sanda and T. J. Masuda, *Polym. Sci., Part A: Polym. Chem.*, 2005, **43**, 5168–5176.

382 F. Sanda, H. Araki and T. Masuda, *Macromolecules*, 2005, **38**, 10605–10608.

383 A. Ikeda, K. Terada, M. Shiotsuki and F. Sanda, *J. Polym. Sci., Part A: Polym. Chem.*, 2011, **49**, 3783–3796.

384 S. Wang, X. Feng, J. Zhang, P. Yu, Z. Guo, Z. Li and X. Wan, *Macromolecules*, 2017, **50**, 3489–3499.

385 T. Yamada, Y. Nagata and M. Suginome, *Chem. Commun.*, 2010, **46**, 4914–4916.

386 T. Yamamoto, T. Yamada, Y. Nagata and M. Suginome, *J. Am. Chem. Soc.*, 2010, **132**, 7899–7901.

387 Y. Nagata, T. Yamada, T. Adachi, Y. Akai, T. Yamamoto and M. Suginome, *J. Am. Chem. Soc.*, 2013, **135**, 10104–10113.

388 Y. Nagata, T. Nishikawa and M. Suginome, *J. Am. Chem. Soc.*, 2015, **137**, 4070–4073.

389 Y. Nagata, K. Takagi and M. Suginome, *J. Am. Chem. Soc.*, 2014, **136**, 9858–9861.

390 S. Leiras, F. Freire, J. M. Seco, E. Quiñoá and R. Riguera, *Chem. Sci.*, 2013, **4**, 2735–2743.

391 S. Leiras, F. Freire, E. Quiñoá and R. Riguera, *Chem. Sci.*, 2015, **6**, 246–253.

392 J. J. Tarrío, R. Rodríguez, B. Fernández, E. Quiñoá and F. Freire, *Angew. Chem., Int. Ed.*, 2022, **61**, e20211507.

393 J. J. Tarrío, B. Fernández, E. Quiñoá and F. Freire, *J. Mater. Chem. C*, 2023, **11**, 8378–8382.

394 R. Rodríguez, E. Quiñoá, R. Riguera and F. Freire, *J. Am. Chem. Soc.*, 2016, **138**, 9620–9628.

395 K. Cobos, R. Rodríguez, O. Domarco, B. Fernández, E. Quiñoá, R. Riguera and F. Freire, *Macromolecules*, 2020, **53**, 3182–3193.

396 R. Rodriguez, E. Quiñoá, R. Riguera and F. Freire, *Small*, 2019, **15**, 1970070.

397 Z. Fernández, B. Fernández, E. Quiñoá and F. Freire, *J. Am. Chem. Soc.*, 2021, **143**, 20962–20969.

398 E. Yashima, T. Matsushima and Y. Okamoto, *J. Am. Chem. Soc.*, 1995, **117**, 11596–11597.

399 K. Morino, N. Watase, K. Maeda and E. Yashima, *Chem. – Eur. J.*, 2004, **10**, 4703–4707.

400 K. Morino, K. Maeda and E. Yashima, *Macromolecules*, 2003, **36**, 1480–1486.

401 E. Yashima, T. Matsushima and Y. Okamoto, *J. Am. Chem. Soc.*, 1997, **119**, 6345–6359.

402 R. Nonokawa and E. Yashima, *J. Am. Chem. Soc.*, 2003, **125**, 1278–1283.

403 R. Nonokawa, M. Oobo and E. Yashima, *Macromolecules*, 2003, **36**, 6599–6606.

404 K. Morino, M. Oobo and E. Yashima, *Macromolecules*, 2005, **38**, 3461–3468.

405 R. Sakai, I. Otsuka, T. Satoh, R. Kakuchi, H. Kaga and T. Kakuchi, *Macromolecules*, 2006, **39**, 4032–4037.

406 E. Yashima, T. Nimura, T. Matsushima and Y. Okamoto, *J. Am. Chem. Soc.*, 1996, **118**, 9800–9801.

407 T. Nishimura, K. Tsuchiya, S. Ohsawa, K. Maeda, E. Yashima, Y. Nakamura and J. Nishimura, *J. Am. Chem. Soc.*, 2004, **126**, 11711–11717.

408 H. Onouchi, T. Miyagawa, A. Furuko, K. Maeda and E. Yashima, *J. Am. Chem. Soc.*, 2005, **127**, 2960–2965.

409 Y. Nishikawa, D. Hirose, S. Sona and K. Maeda, *Chem. Commun.*, 2023, **59**, 8226–8229.

410 H. Onouchi, T. Hasegawa, D. Kashiwagi, H. Ishiguro, K. Maeda and E. Yashima, *Macromolecules*, 2005, **38**, 8625–8633.

411 K. Maeda, Y. Takeyama, K. Sakajiri and E. Yashima, *J. Am. Chem. Soc.*, 2004, **126**, 16284–16285.

412 K. Nagai, K. Maeda, Y. Takeyama, K. Sakajiri and E. Yashima, *Macromolecules*, 2005, **38**, 5444–5451.

413 K. Maeda, H. Tsukui, Y. Matsushita and E. Yashima, *Macromolecules*, 2007, **40**, 7721–7726.

414 K. Maeda, H. Mochizuki, K. Osato and E. Yashima, *Macromolecules*, 2011, **44**, 3217–3226.

415 T. Ikai, M. Ando, M. Ito, R. Ishidate, N. Suzuki, K. Maeda and E. Yashima, *J. Am. Chem. Soc.*, 2021, **143**, 12725–12735.

416 Y. Hase, M. Ishikawa, R. Muraki, K. Maeda and E. Yashima, *Macromolecules*, 2006, **39**, 6003–6008.

417 Y. Hase, K. Nagai, H. Iida, K. Maeda, N. Ochi, K. Sawabe, K. Sakajiri, K. Okoshi and E. Yashima, *J. Am. Chem. Soc.*, 2009, **131**, 10719–10732.

418 E. Yashima, K. Maeda and T. Yamanaka, *J. Am. Chem. Soc.*, 2000, **122**, 7813–7814.

419 K. Maeda, K. Kuroyanagi, S. I. Sakurai, T. Yamanaka and E. Yashima, *Macromolecules*, 2011, **44**, 2457–2464.

420 K. Maeda, M. Nozaki, K. Hashimoto, K. Shimomura, D. Hirose, T. Nishimura, G. Watanabe and E. Yashima, *J. Am. Chem. Soc.*, 2020, **142**, 7668–7682.

421 W. Makiguchi, S. Kobayashi, Y. Furusho and E. Yashima, *Angew. Chem., Int. Ed.*, 2013, **52**, 5275–5279.

422 K. Maeda and E. Yashima, *Bull. Chem. Soc. Jpn.*, 2021, **94**, 2637–2661.

423 N. Ousaka, F. Mamiya, Y. Iwata, K. Nishimura and E. Yashima, *Angew. Chem., Int. Ed.*, 2017, **56**, 791–795.

424 T. Ikai, S. Kawabata, F. Mamiya, D. Taura, N. Ousaka and E. Yashima, *J. Am. Chem. Soc.*, 2020, **142**, 21913–21925.

425 D. Lee, Y. J. Jin, H. Kim, N. Suzuki, M. Fujiki, T. Sakaguchi, S. K. Kim, W. E. Lee and G. Kwak, *Macromolecules*, 2012, **45**, 5379–5386.

426 T. Miao, X. Cheng, H. Ma, Z. He, Z. Zhang, N. Zhou, W. Zhang and X. Zhu, *Angew. Chem., Int. Ed.*, 2021, **60**, 18566–18571.

427 J. Liu, Y. Zhao, H. Chen, Z. Zhang, W. Zhang and X. Zhu, *React. Funct. Polym.*, 2017, **121**, 76–81.

428 Y. Zhao, L. Yin, J. Liu, H. Chen and W. Zhang, *Chirality*, 2017, **29**, 107–114.

429 G. Zhang, Y. Bao, M. Pan, N. Wang, X. Cheng and W. Zhang, *Sci. China: Chem.*, 2023, **66**, 1169–1178.

430 Y. Nagata, R. Takeda and M. Sugino, *ACS Cent. Sci.*, 2019, **5**, 1235–1240.

431 J. L. Greenfield, E. W. Evans, D. Di Nuzzo, M. Di Antonio, R. H. Friend and J. R. Nitschke, *J. Am. Chem. Soc.*, 2018, **140**, 10344–10353.

432 S. Xue, P. Xing, J. Zhang, Y. Zeng and Y. Zhao, *Chem. – Eur. J.*, 2019, **25**, 7426–7437.

433 M. L. Ślęczkowski, M. F. J. Mabesoone, P. Ślęczkowski, A. R. A. Palmans and E. W. Meijer, *Nat. Chem.*, 2021, **13**, 200–207.

434 A. K. Mondal, M. D. Preuss, M. L. Ślęczkowski, T. K. Das, G. Vantomme, E. W. Meijer and R. Naaman, *J. Am. Chem. Soc.*, 2021, **143**, 7189–7195.

435 S. J. George, Ž. Tomović, A. P. H. J. Schenning and E. W. Meijer, *Chem. Commun.*, 2011, **47**, 3451–3453.

436 B. Isare, M. Linares, L. Zargarian, S. Fermandjian, M. Miura, S. Motohashi, N. Vanthuyne, R. Lazzaroni and L. Bouteiller, *Chem. – Eur. J.*, 2010, **16**, 173–177.

437 I. Destoop, E. Ghijssens, K. Katayama, K. Tahara, K. S. Mali, Y. Tobe and S. De Feyter, *J. Am. Chem. Soc.*, 2012, **134**, 19568–19571.

438 S. Dhiman, A. Jain and S. J. George, *Angew. Chem., Int. Ed.*, 2017, **56**, 1329–1333.

439 S. Dhiman, A. Jain, M. Kuma and S. J. George, *J. Am. Chem. Soc.*, 2017, **139**, 16568–16575.

440 N. Ousaka, Y. Inai and R. Kuroda, *J. Am. Chem. Soc.*, 2008, **130**, 12266–12267.

441 N. Ousaka and Y. Inai, *J. Am. Chem. Soc.*, 2006, **128**, 14736–14737.

442 T. Sanji, K. Takase and H. Sukaria, *J. Am. Chem. Soc.*, 2001, **123**, 12690.

443 B. A. F. Le Bailly, L. Byrne and J. Clayden, *Angew. Chem., Int. Ed.*, 2016, **55**, 2132–2136.

444 J. Brioche, S. J. Pike, S. Tshepelevitsh, I. Leito, G. A. Morris, S. J. Webb and J. Clayden, *J. Am. Chem. Soc.*, 2015, **137**, 6680–6691.



445 R. A. Brown, V. Diemer, S. J. Webb and J. Clayden, *Nat. Chem.*, 2013, **5**, 853–860.

446 S. Saha, B. Kauffmann, Y. Ferrand and I. Huc, *Angew. Chem., Int. Ed.*, 2018, **57**, 13542–13546.

447 P. Mateus, B. Wicher, Y. Ferrand and I. Huc, *Chem. Commun.*, 2018, **54**, 5078–5081.

448 M. Vallade, P. Sai Reddy, L. Fischer and I. Huc, *Eur. J. Org. Chem.*, 2018, 5489–5498.

449 P. Mateus, N. Chandramouli, C. D. Mackereth, B. Kauffmann, Y. Ferrand and I. Huc, *Angew. Chem., Int. Ed.*, 2020, **59**, 5797–5805.

450 Y. Ferrand, A. M. Kendhale, B. Kauffmann, A. Grélard, C. Marie, V. Blot, M. Pipelier, D. Dubreuil and I. Huc, *J. Am. Chem. Soc.*, 2010, **132**, 7858–7859.

451 J. L. Hou, X.-B. Shao, G. J. Chen, Y. X. Zhou, X. K. Jiang and Z. T. Li, *J. Am. Chem. Soc.*, 2004, **126**, 12386.

452 C. Li, G. T. Wang, H. P. Yi, X. K. Jiang, Z. T. Li and R. X. Wang, *Org. Lett.*, 2007, **9**, 1797.

453 M. Inouye, M. Waki and H. Abe, *J. Am. Chem. Soc.*, 2004, **126**, 2022–2027.

454 J. C. Nelson, J. G. Saven, J. S. Moore and P. G. Wolynes, *Science*, 1997, **277**, 1793–1796.

455 R. B. Prince, S. A. Barnes and J. S. Moore, *J. Am. Chem. Soc.*, 2000, **122**, 2758–2762.

456 M. T. Stone and J. S. Moore, *Org. Lett.*, 2004, **6**, 469–472.

457 A. Tanatani, M. J. Mio and J. S. Moore, *J. Am. Chem. Soc.*, 2001, **123**, 1792–1793.

458 A. Tanatani, T. S. Hughes and J. S. Moore, *Angew. Chem., Int. Ed.*, 2002, **41**, 325–328.

459 H. Abe, N. Masuda, M. Waki and M. Inouye, *J. Am. Chem. Soc.*, 2005, **127**, 16189–16196.

460 H. Abe, H. Machiguchi, S. Matsumoto and M. Inouye, *J. Org. Chem.*, 2008, **73**, 4650–4661.

461 Q. Gan, Y. Ferrand, C. Bao, B. Kauffmann, A. Grélard, H. Jiang and I. Huc, *Science*, 2011, **331**, 1172–1175.

462 A. Kousar, J. Liu, N. Mehwish, F. Wang, A. Y. Dang-i and C. Feng, *Mater. Today Chem.*, 2019, **11**, 217–224.

463 S. Goskulwad, D. D. La, M. A. Kobaisi, S. V. Bhosale, V. Bansal, A. Vinu, K. Ariga and S. V. Bhosale, *Sci. Rep.*, 2018, **8**, 11220.

464 M. Pandeeswar and T. Govindaraju, *Mol. Syst. Des. Eng.*, 2016, **1**, 202–207.

465 H. Frisch, Y. Nie, S. Raunser and P. Besenius, *Chem. – Eur. J.*, 2015, **21**, 3304–3309.

466 J. Liu, Z. Luo, L. Yu, P. Zhang, H. Wei and Y. Yu, *Chem. Sci.*, 2020, **11**, 8224–8230.

467 E. Suárez-Picado, E. Quiñoá, R. Riguera and F. Freire, *Chem. Mater.*, 2018, **30**, 6908–6914.

468 R. Liu, F. Sanda and T. Masuda, *Macromol. Chem. Phys.*, 2009, **210**, 331–339.

469 L. Wu, Y. Xu, T. Hou, J. Jia, X. Huang, G. Weng, S. Bao and L. Zheng, *Chem. – Eur. J.*, 2021, **27**, 16722–16734.

470 J. Yuan, Y. Zhang, Y. Sun, Z. Cai, L. Yang and H. Lu, *Biomacromolecules*, 2018, **19**, 2089–2097.

471 R. Barman, P. Dey, T. Mondal and S. Ghosh, *Chem. – Asian J.*, 2019, **14**, 4741–4747.

