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Fluorogenic cell surface glycan labelling with fluorescence molecular rotor dyes and nucleic acid stains†

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We show that covalent labelling of sialic acids on live cell surfaces or mucin increases the fluorescence of the fluorescence molecular rotors (FMRs) CCVJ, Cy3 and thiazole orange, enabling wash-free imaging of cell surfaces. Dual labelling with an FMR and an environmentally insensitive dye allows detection of changes that occur, for example, when cross-linking is altered.

Fluorescence molecular rotor (FMR) dyes are a class of dyes which are exceptionally responsive to the environment.¹ Their responsiveness is based on twisting motions which non-radiatively deplete the excited state. Environments that hinder twisting can induce strong enhancements of fluorescence. This property has been used to measure nanoviscosity^{2–4} and interactions of FMR-labelled ligands with proteins⁵ and nucleic acids.^{6,7} We envisioned that FMR dyes should also be able to show enhanced fluorescence upon labelling of cell surface glycans. The glycocalyx and mucus layers on the cell surface are dense materials, which should provide crowded environments imposing friction on FMR dyes. The resulting fluorogenic labelling is advantageous, because wash-steps typically used for the removal of non-bound probes are no longer required, facilitating the analysis of fast processes. We furthermore envisaged that FMRs, when attached to cell surface glycans, could respond to changes occurring in disease and therapeutic intervention.

For covalent labelling of cell surface glycans, we selected the FMRs CCVJ and Cy3 (Fig. 1A), which show increased fluorescence when high viscosity or the crowded environment of a binding partner hinders twisting motions.^{3,8} So far, binding induced enhancements of CCVJ or Cy3 emission have been demonstrated for interactions with proteins^{9–11} and nucleic acids¹² but to the best of our knowledge increased fluorescence upon attachment to glycans has not been reported. In addition, we explored the use of thiazole orange (TO) previously not considered as a cell surface stain. TO is virtually non-fluorescent in free form but experiences dramatic fluorescence enhancements upon DNA intercalation because rotations around a central methine bridge are hindered.¹³ TO dyes are therefore frequently used as nucleic stains.^{14,15} In addition, TO has been used to sense the vicinity of proteins that hinder twisting.^{16,17} Assuming that glycan labelling and cell surface viscosity would induce a similar effect, we repurposed TO to enable staining of the cell surface. For this purpose, TO as well as CCVJ and Cy3 were attached to a tripartate peptide hydrazide (FMR-3D-Ahx-N₂H₃, Fig. 1B). The negative charges hinder cell membrane permeation of the hydrophobic dyes (Fig. S8, ESI†). The hydrazide group allowed conjugation with aldehyde functions installed at cell surface sialic acid residues upon mild periodate treatment (Fig. 1B and Fig. S8, ESI†).^{18,19} This method has frequently been used to stain cell surfaces. However, fluorogenic labelling with FMR dyes has not been reported. Published labelling protocols involve a total of 0.5–2 h incubation of cells at 0–4 °C, and wash steps to remove excess labelling reagents prior to fluorescence microscopic imaging. This can induce cold stress potentially altering cellular surfaces (Fig. S6, ESI†). We added 5-methoxyanthranilic acid (5MA) as an organocatalyst²⁰ and performed both the periodate treatment and the hydrazone ligation with the acyl hydrazides at 25 °C for 5 minutes only (Fig. S7 and S8, ESI†). This way, delicate cells such as A549 cells were labelled with little, if any, blebbing.

Labelling with TO-3D-Ahx-N₂H₃ allowed clear imaging of cell surfaces (Fig. 1C). Control experiments showed that periodate treatment and the presence of 5MA was required for cell surface staining (Fig. S9, ESI†). Considering that TO in free form is

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‡ Deceased



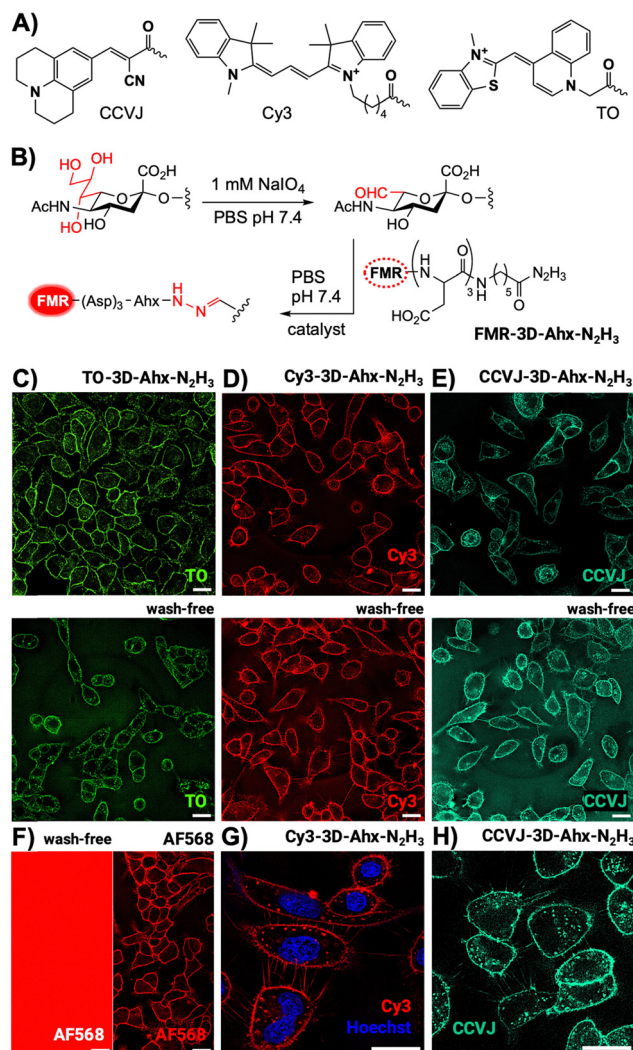


Fig. 1 (A) FMR dyes used for (B) cell surface labelling via mild and rapid oxidation with NaIO_4 at room temperature followed by organocatalyzed hydrazine ligation with FMR-functionalized peptide hydrazide probes FMR-3D-Ahx- N_2H_3 . Fluorescence microscopy images of A549 cells labelled with (C) TO-3D-Ahx- N_2H_3 , (D) Cy3-3D-Ahx- N_2H_3 , (E) CCVJ-3D-Ahx- N_2H_3 , (F) AF568- N_2H_3 , (G) and (H) All probes stain filopodia. Conditions: *i*. 1 mM NaIO_4 in PBS pH 7.4, 5 min, 25 °C, *ii*. 10 μM (with wash) or 1 μM (no wash) hydrazide probe, 10 mM 5MA in PBS pH 7.4, 5 min, 25 °C. Scale bar: 20 μm .

essentially non-fluorescent in aqueous solution,²¹ we attribute the signals provided by the glycan-bound TO probe to properties of the cell surface matrix (*vide infra*). As anticipated, washing is not required after treatment with the TO probe (Fig. 1C, low). Similarly, A549 cells were treated, after rapid periodate treatment, with peptide hydrazides containing Cy3 and CCVJ. Again, a short (5 min) 5MA-catalyzed labelling with Cy3-3D-Ahx- N_2H_3 and CCVJ-3D-Ahx- N_2H_3 was sufficient to stain the surface of living A549 cells (Fig. 1D and E). Even filopodial protrusions were clearly visible (Fig. 1G and H). Again, no washing was required indicating that the fluorescence of non-reacted Cy3 and CCVJ probes is lower than the fluorescence of probes on the cell surface. In a control experiment, we labelled

cells with AF568 hydrazide (Fig. 1F). While clear cell surface staining was achieved after removal of non-reacted probe by washing, the no-wash protocol failed to provide contrast. Regardless of the imaging probe used, staining by direct labelling of the surface glycans was never exclusive for the surface of A549 cells. The punctuate signals inside cells are likely due to rapid internalization as a result of constitutively active endocytotic processes. Of note, rapid room temperature labelling with the FMR-3D-Ahx- N_2H_3 probes (FMR = Cy3, CCVJ and TO) also succeeded with CEM cells as an example of a suspension cell line (Fig. S12, ESI[†]).

In studies of porcine mucin, we investigated the factors responsible for enhancing the fluorescence of covalently bound FMR on the cell surface. Porcine mucin in water is viscous and can be labelled with FMR hydrazides due to its approx. 1.2% sialic acid content. The fluorescence of the CCVJ-3D-Ahx- N_2H_3 probe increased substantially when allowed to react with periodate-treated mucin (Fig. 2A). For comparison, the reactive CCVJ-3D-Ahx- N_2H_3 probe was added to mucin without prior periodate activation. As a further control, fluorescence of an unreactive CCVJ-OH was measured in PBS and after addition of the periodate-activated mucin (Fig. S16, ESI[†]). Fluorescence remained low in both cases. This suggests that the close proximity to glycans induced upon formation of covalent bonds is very efficient in hindering CCVJ torsional motions, demonstrating, for the first time, that CCVJ can respond to interactions with glycans. This raises the question of whether mucin-conjugated

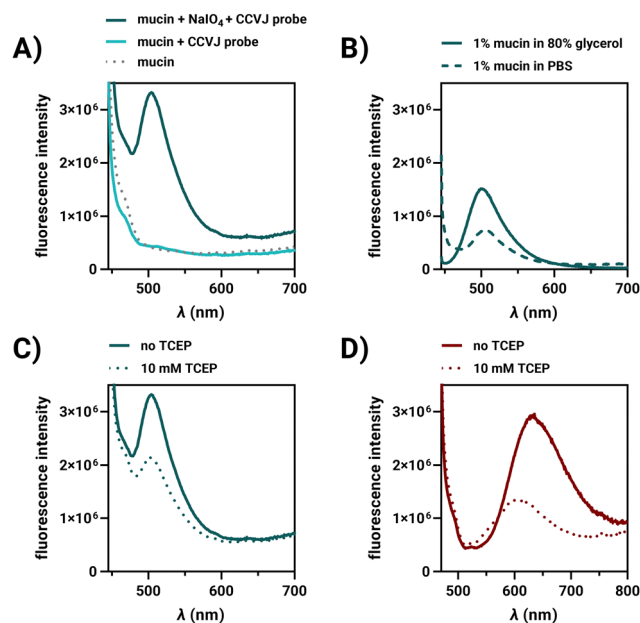


Fig. 2 Fluorescence spectra of (A) CCVJ-3D-Ahx- N_2H_3 in presence of porcine stomach mucin (8% w/v) with or without periodate activation and blank mucin alone, (B) CCVJ-3D-Ahx- N_2H_3 -labelled mucin (1% w/v) in PBS or 80% glycerol, (C) CCVJ-3D-Ahx- N_2H_3 -labelled or (D) TO-3D-Ahx- N_2H_3 -labelled porcine mucin before and after addition of 10 mM TCEP. Conditions: 10 μM CCVJ- or TO-3D-Ahx- N_2H_3 , in treated (1 mM NaIO_4 , PBS, pH 7.4, 37 °C, 30 min, washing) or untreated mucin in PBS or 80% glycerol (v/v) or 10 mM TCEP, PBS, pH 7.4, 37 °C, 30 min. CCVJ: λ_{ex} = 435 nm; TO: λ_{ex} = 460 nm.



CCVJ would still be able to sense changes in viscosity. Glycerol (80% v/v) was included in the buffer, and an increase in CCVJ emission was observed (Fig. 2B). In a second experiment, TCEP was added, which was reported to decrease viscosity of mucin by cleavage of disulfide crosslinks.²² Intensity of CCVJ emission decreased upon addition of TCEP (Fig. 2C). Rheological measurements showed that TCEP treatment affected both the storage modulus G' and the loss modulus G'' (Fig. S19, ESI†). The decrease in fluorescence might therefore be caused by both decreases of molecular crowding around CCVJ and reduced viscosity. Control measurements involving addition of TCEP to CCVJ in glycerol (a matrix which lacks cleavable disulfides), showed that chemical quenching contributed only little to the decrease of dye emission (Fig. S18A, ESI†). We concluded that CCVJ on the glycans of porcine mucin remains responsive to changes of the environment that alter friction.

Labelling of porcine mucin was also performed with TO-3D-Ahx-N₂H₃. The emission from TO on mucin was red-shifted with a maximum at 635 nm (Fig. 2D), indicative of TO aggregation.¹⁶ As observed for CCVJ, treatment of the TO-labelled mucin with TCEP led to a decrease of fluorescence, which did not occur when TCEP was added to TO in glycerol (Fig. S18B, ESI†). For Cy3-3D-Ahx-N₂H₃ we also observed a fluorescence increase upon covalent labelling and a viscosity responsiveness (Fig. S17, ESI†), both of which may contribute to the fluorogenicity of cell surface glycan labelling.

Based on the results of the mucin labelling experiments, we hypothesized that conjugation of FMR dyes with cell surface glycans might allow detection of changes in the glycocalyx environment, such as might be caused by modifications/remodelling of cell surface components. However, in cell measurements, low fluorescence intensity does not necessarily have to be due to such changes, but can also be caused by low occupancy, *i.e.*, a lower amount of dye. We, therefore, investigated simultaneous labelling of A549, HeLa and CEM cells with two dyes (Fig. 3). The FMR dye would serve as friction sensor, while Alexa Fluor 568 hydrazide, would function as an environmentally insensitive concentration reporter. A mixture of both dyes was used for labelling and the ratio of staining intensities in the FMR and the AF568 channels was determined for each cell surface. In control experiments, we verified that AF568 fluorescence is not affected by viscosity (Fig. S15, ESI†). The analysis was focused on cell surface regions (Fig. 3A–D and Fig. S14, ESI†). In one set of experiments, A549 cells were treated with TCEP and washed prior to labelling. As shown in Fig. 2C and D, TCEP treatment reduced the degree of molecular crowding/viscosity of mucus, and we reasoned that it might also act on the glycocalyx. Remarkably, cells which were incubated for 60 min with 100 μ M TCEP before labelling showed significantly reduced CCVJ/AF568 (compare Fig. 3A, B and E) and TO/AF568 (compare Fig. 3C, D and F) fluorescence ratio values compared to control cells. Quantitative analysis of a total of 400–600 cells (Fig. 3E and F) showed that labelling of TCEP-treated cells resulted in mean FMR/AF568 ratios that were 29% (FMR = CCVJ) or 35% (FMR = TO) lower than the ratio determined for untreated cells. TCEP is a reducing agent,

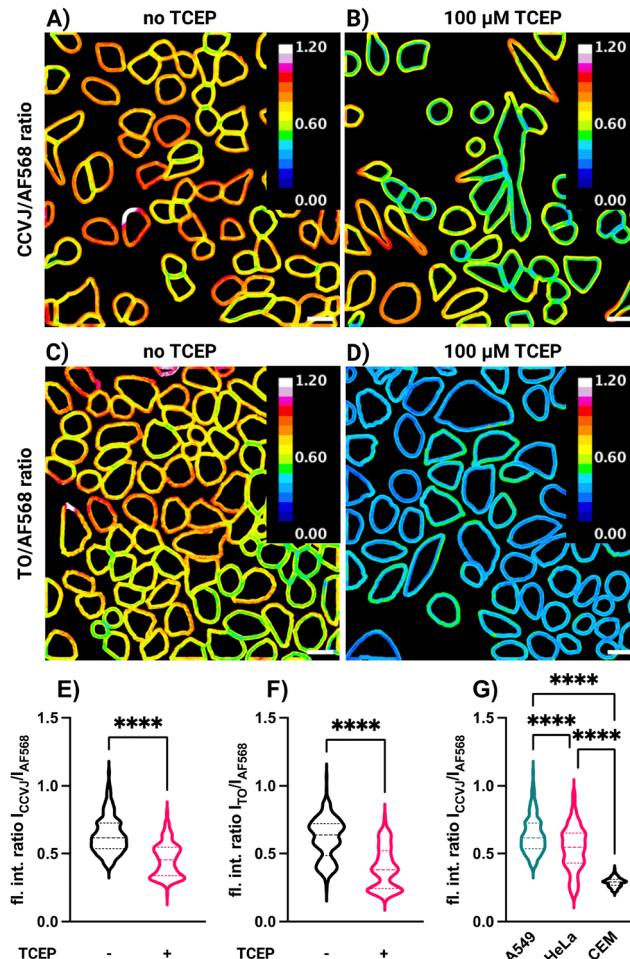


Fig. 3 Ratio imaging for A549 cells labelled with a mixture of either CCVJ-3D-Ahx-N₂H₃ or TO-3D-Ahx-N₂H₃ with AF568-N₂H₃. Cell surface fluorescence intensity ratios of (A) and (B) $I_{\text{CCVJ}}/I_{\text{AF568}}$ and (C) and (D) $I_{\text{TO}}/I_{\text{AF568}}$ for (A) and (C) untreated A549 cells and (B) and (D) cells treated with the reducing agent TCEP (100 μ M) prior to labelling. Violin plots for fluorescence intensity ratios (E) $I_{\text{CCVJ}}/I_{\text{AF568}}$ and (F) $I_{\text{TO}}/I_{\text{AF568}}$ for surfaces of untreated A549 cells and cells treated with TCEP in DMEM and 5% CO₂ for 1 h at 37 °C prior to labelling. (G) Fluorescence intensity ratios $I_{\text{CCVJ}}/I_{\text{AF568}}$ for A549, HeLa and CEM cells labelled with a mixture of AF568- and CCVJ-3D-Ahx-N₂H₃. Conditions: *l.* 1 mM NaIO₄, PBS, pH 7.4, 5 min, 25 °C, *ll.* 1 μ M AF568-N₂H₃, 10 μ M TO- or CCVJ-3D-Ahx-N₂H₃, 10 mM 5MA, PBS, pH 7.4, 5 min, 25 °C. Data in (E), (F) and (G) is from $N = 2$ replicates and analysis of 100–150 cells per replicate. Statistical analysis: unpaired, two-tailed *t* test. ns: not significant, $p > 0.05$; **** $p < 0.0001$. Scale bar = 20 μ m.

and cleavage of accessible disulfide bridges could cause partial shedding of cell surface glycans. This could reduce the amount of sialic acid residues available for reactions with the rotor dyes. Nonetheless, this should affect both, the FMR and AF568 dyes. It is therefore plausible to assume that the decrease in FMR/AF568 intensity ratio is due to a decrease in steric hindrance experienced by the FMR. This decrease could be due to a reduced viscosity or an increase of degrees of freedom when reductive cleavage of the disulfide bridges reduces crowding in the FMR's environment.

Interestingly, simultaneous labelling with AF568-N₂H₃ and CCVJ-3D-Ahx-N₂H₃ showed cell type-specific emission ratios.



Compared to A549 and HeLa cells, CEM cells were characterized by a significantly lower ratio value (Fig. 3G). We speculate that this difference can be caused by different compositions of the cell surfaces. A549 cells originate from the lung epithelium and HeLa from the cervix. These epithelial cell types might form a thicker glycocalyx than CEM cells, which originate from circulating lymphocytes.

The data shown in Fig. 2 and 3 suggest that direct labelling of neuraminic acid with an FMR dye such as CCVJ can detect changes of the cell surface composition. For further validation we assessed the CCVJ-labelled cells by fluorescence lifetime imaging microscopy (FLIM).²³ The mean fluorescence lifetime of 0.37 ns determined for CCVJ-3D-Ahx-N₂H₃ in PBS at pH 7.4 increased after labelling of the cell surface to 1.9 ± 0.2 ns (Table S1 and Fig. S20, ESI†). This confirms that the signals measured by fluorescence intensity microscopy (Fig. 2) are not due to dye enrichment but to interactions of the dye with the cell surface layer. Of note, the lifetime of CCVJ emission on the cell is much longer than the lifetime measured in highly viscous glycerol, which is in the subnanosecond range.²⁴ This might be attributed to the additional contribution of interactions with glycans induced upon covalent attachment. Cells that were treated with TCEP prior to labelling with CCVJ-3D-Ahx-N₂H₃ showed a decrease in the mean fluorescence lifetime to 1.3 ns (Table S1 and Fig. S20B, ESI†). Thus, we conclude that conjugation of CCVJ with cell surface sialic acids allows detection of changes in the glycocalyx layers that occur, for example, when disulfide bridges are cleaved.

In summary, we explored covalent labelling of cell surface sialic acids with the FMRs CCVJ, Cy3 and thiazole orange (TO). TO is a nucleic acid stain, which, to our knowledge, has not been used for staining cell surfaces. Our data shows that sialic acid labelling of cell surfaces or isolated mucin *via* the periodate oxidation/peptidyl hydrazone ligation sequence guides the FMRs into an environment where the free volume available for twisting motions is effectively restricted. This can be caused by viscosity describing the resistance of solvent molecules to displacements and/or molecular crowding/confinement referring to the steric effect of biomolecules. To the best of our knowledge, this is the first example showing that FMR dyes can be used to detect interactions with glycans. Both CCVJ and TO are essentially dark when free in aqueous solution, and their fluorogenic response allows omission of washing after labelling. Co-labelling of cell surfaces with an environmentally insensitive dye and intensity ratio imaging proved that CCVJ and TO fluorescence is affected by alterations induced upon the treatment of cells with the reducing agent TCEP. This suggests that treatment of cells, and also mucin, with TCEP reduced frictional forces acting on the glycan-bound dyes, most likely by decreasing crowding in their environment through disulfide cleavage. In diseases such as mucoviscidosis or chronic obstructive pulmonary disease (COPD), the surface of airway epithelial cells is covered by abnormally thick mucus. Our

feasibility study suggests that it should be possible to monitor changes of cell surface properties using equipment commonly available to cell biology labs. We envision applications, such as in screening campaigns searching for mucolytic agents or to decipher the effects that occur upon bacterial/viral challenge. In this context, it is beneficial that direct labelling of glycans allows studies of wildtype cells.

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Conflicts of interest

There are no conflicts to declare.

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