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Weak effects of prebiotically plausible peptides on self-triphosphorylation ribozyme function†

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Catalytic RNAs (ribozymes) were central to early stages of life on earth. The first ribozymes probably emerged in the presence of prebiotically generated peptides because amino acids can be generated under abiotic conditions, and amino acids can oligomerize into peptides under prebiotically plausible conditions. Here we tested whether the presence of prebiotically plausible peptides could have aided the emergence of ribozymes, by an *in vitro* selection of self-triphosphorylation ribozymes from random sequence in the presence of ten different octapeptides. These peptides were composed of ten different, prebiotically plausible amino acids, each as mixture of *D*- and *L*-stereoisomers. After five rounds of selection and high throughput sequencing analysis, ten ribozymes that appeared most promising for peptide benefits were tested biochemically for possible benefits from each of the ten peptides. The strongest peptide benefit enhanced ribozyme activity by 2.6-fold, similar to the effect from an increase in the pH by one-half unit. Four arbitrarily chosen ribozymes from a previous selection without peptides showed no significant change in their activity in the presence of the ten peptides. Therefore, the used prebiotically plausible peptides – peptides without evolutionarily optimized sequence, without cationic or aromatic side chains – did not provide a strong benefit for the emergence of ribozyme activity. This finding stands in contrast to previously identified polycationic peptides, conjugates between peptides and polyaromatic hydrocarbons, and modern mRNA encoded proteins, all of which can strongly increase ribozyme function. The results are discussed in the context of origins of life.

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Introduction

Early stages of life on earth likely relied on catalytic RNAs to promote self-replication and evolution.^{1–4} This idea is supported by the finding that many of today's enzyme cofactors are nucleotides or variants thereof,^{5,6} and by the idea that RNA's ability to store information as well as catalyze reactions could have been sufficient for an early RNA-dominated life form that was later overtaken by the contemporary, interdependent DNA–RNA–protein systems.^{1–4} The strongest support for this hypothesis is the ribosome, which is a highly evolved ribozyme and synthesizes all genetically encoded proteins in today's life forms.^{7,8} This means that the last universal common ancestor (LUCA) – which contained a complex ribosome⁹ – was preceded by an evolving molecular system with sophisticated catalytic RNAs. However, it is unclear how a system based on

sophisticated RNAs emerged in a prebiotic environment:¹⁰ based on current prebiotic model reactions for nucleotide synthesis and non-enzymatic RNA polymerization^{11–13} and highly controlled experiments that select ribozymes from random sequence,^{14,15} it appears unlikely that sophisticated ribozymes emerged *de novo* in a prebiotic environment. One approach to address this challenge is to investigate the possible help of molecules that likely existed on prebiotic earth but don't have that function in today's biochemistry.¹⁶ An example for this approach lies in α -hydroxy acids, which can catalyze peptide bond formation in wet-dry cycling conditions.¹⁷ Analogously, we explore here whether prebiotically plausible peptides containing five biological and five non-biological amino acids, as mixtures of *D*- and *L*-isomers, could have aided the function of catalytic RNAs. Prebiotically plausible peptides are fundamentally different from ribosome-synthesized peptides and proteins because they contain a set of prebiotically plausible amino acids that only partially overlap with biological amino acids, they contain both (biological) *L* and (non-biological) *D* stereoisomers, and their amino acid sequence was not encoded and therefore not optimized by Darwinian evolution.

Amino acids and peptides can be generated under prebiotically plausible conditions, much easier than the building

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blocks of RNA (compare ref. 17–20 to ref. 12 and 21–23). Since amino acids appear as side products in reactions that may have given rise to nucleotides,²⁴ it is reasonable to assume that the first catalytic RNAs emerged in the presence of amino acids and peptides. Such peptides might have benefitted early ribozymes: peptides can increase the thermodynamic stability of duplex RNA²⁵ and improve ribozyme interactions with their RNA substrate either by direct interactions²⁶ or by co-localizing the ribozyme and its RNA substrate within RNA/peptide coacervates.²⁷ Most of these previous studies used L-amino acids with a high content of arginine and lysine. The exception is a study that also explored the effects of an ornithine decapeptide on the function of a polymerase ribozyme, and found the same beneficial effect as a lysine decapeptide.²⁶ For both decapeptides, the D,L-peptides resulted in the same benefits as the all-L peptides, showing that at least in this case the amino acid chirality is not critical. While ornithine is prebiotically plausible, it is not one of the more frequent amino acids.¹⁹ Therefore, a prebiotic function of ornithine decapeptides would likely have required enrichment steps such as coacervate formation with polycationic RNAs. Most previous studies on peptide benefits for ribozymes tested the effect of peptides on individual, existing ribozymes that were selected in the absence of peptides. The exception is a recent *in vitro* selection study of new ribozymes in the presence of octapeptides composed of the 20 biological L-amino acids, which found that conjugates of octapeptides with polyaromatic hydrocarbons can lead to ~900-fold enhancement of ribozyme activity.²⁸ However, these peptides consisted exclusively of L-amino acids, and included amino acids that would probably not have existed before a macromolecule-catalyzed metabolism. To directly address the question whether prebiotically plausible peptides could have aided the emergence of the first ribozymes, it is necessary to probe RNA sequence space for the existence of ribozymes – for example by *in vitro* selection from random sequence – in the presence of prebiotically plausible peptides that consist of prebiotically plausible amino acids, and include the chiral amino acids as D- and L-stereoisomers.

The model system used in this study to probe RNA sequence space for the existence of ribozymes is an *in vitro* selection for self-triphosphorylation ribozymes.²⁹ This *in vitro* selection system previously yielded more than 300 different ribozyme clusters from 1.7×10^{14} different sequences,³⁰ and has performed robustly in five different selection contexts.^{28,29,31–33} Self-triphosphorylation ribozymes are relevant for prebiotic scenarios because they catalyze the reaction between their 5'-hydroxyl group and the prebiotically plausible energy source ‘cyclic trimetaphosphate’ (cTmp)^{34,35} to generate 5'-triphosphates. The analogous nucleoside 5'-triphosphates (NTPs) are used today as energy currency in all biological organisms, and were thus likely at the core of life forms even before LUCA. Together, the robust performance and prebiotic relevance make this *in vitro* selection setup an ideal model system to study what molecules can increase the frequency of ribozymes in RNA sequence space, in different chemical environments.

Here we describe the *in vitro* selection of self-triphosphorylation ribozymes in the presence of ten octapeptides, each composed

of D/L mixtures of ten different, prebiotically plausible amino acids, and with the N-terminal amino group as their only positive charge. After high throughput sequencing analysis of the selected RNA sequences, ten promising ribozymes were tested biochemically with each of the ten peptides individually. The strongest peptide benefit to a ribozyme was a 2.6-fold increase in activity, less than the benefit from increasing the pH by one unit. Similarly, four ribozymes that were previously selected in the absence of peptides were not strongly affected by the same ten peptides. The results are discussed in the context of RNA-centered early life forms.

Results

To test whether prebiotically plausible peptides could help the emergence of ribozymes, we performed an *in vitro* selection of self-triphosphorylating ribozymes in the presence of peptide octamers containing ten different prebiotically plausible amino acids, covering five biological and five non-biological amino acids (Fig. 1). The set of prebiotically available amino acids has been studied in prebiotic model reactions,^{18–20,36–38} and several approaches comparing the amino acid's chemical complexity and the comparisons of their codons.³⁹ Because the current study is focused on the earliest self-replicating systems long before LUCA, the results from prebiotic model reactions were prioritized. The frequency of the ten amino acids in the ten peptides used in this study was based on their abundance in a representative prebiotic model reaction that was analyzed by modern LC-MS, and should be seen as one of the many, similar plausible distributions for prebiotic scenarios:^{18,19} 22 × glycine, 14 × alanine, 9 × aspartate, 8 × b-alanine, 6 × a-amino butyric acid, 6 × b-amino butyric acid, 5 × valine, 5 × a-amino isobutyric acid, 3 × g-amino butyric acid, 2 × serine. All amino acids with a chiral center were represented as mixtures of D- and L-isomers. The presence of α-, β-, and γ-amino acids generated backbones with different spacing between peptide bonds. The sequences of the peptides were designed manually, with an effort to evenly distribute the ten amino acids over the ten peptides, and the eight positions. Each of the peptides contained between three and six chiral amino acids both as D- and L-isomers, with a total of 232 stereoisomers in the ten peptides. The length of eight amino acids per peptide was chosen because this length is accessible *via* wet-dry cycling reactions in the presence of α-hydroxy acids.^{16,17} All ten octapeptides were well soluble at a concentration of at least 20 mM in water.

The *in vitro* selection was conducted as described earlier^{29,33} (Fig. 1), with an effective complexity of 4.2×10^{14} sequences in the first selection round. The RNAs were challenged to catalyze the nucleophilic attack of their 5'-hydroxyl group to the phosphorus of cyclic trimetaphosphate (cTmp), thereby generating a 5'-triphosphate. The incubation with cTmp was done in the presence of the described ten peptides, each at 1 mM concentration. Depending on the number of stereoisomers for a given peptide, the concentration for each stereoisomer was lower. For example, peptides with 3 chiral positions and



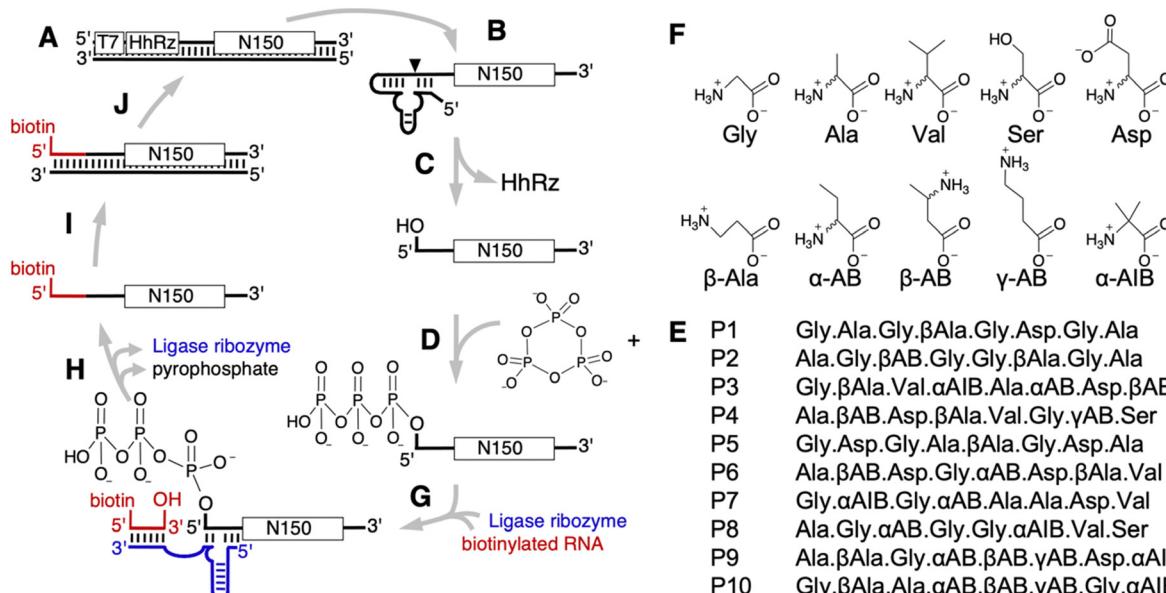


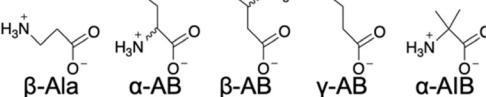
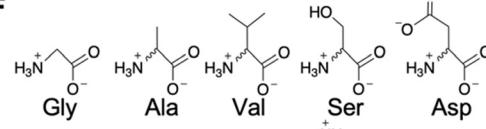
Fig. 1 Schematic of the *in vitro* selection procedure for ribozymes that triphosphorylate their 5'-hydroxyl group in the presence of prebiotically plausible octapeptides. (A) The DNA library contained a promoter for transcription by T7 RNA polymerase (T7), the sequence encoding a hammerhead ribozyme (HhRz), and 150 nucleotides of randomized sequence (N150) flanked by constant regions (bold lines). (B) Transcription into RNA generated a 5'-terminal hammerhead ribozyme, which (C) self-cleaved co-transcriptionally and thereby generated a 5'-hydroxyl terminus at the RNA library. (D) The RNA library was incubated in the presence of 50 mM Tris/HCl pH 8.3, 100 mM MgCl₂, 50 mM Na₃cTmP, and each 1 mM of (E) ten different octapeptides that were composed of (F) ten different, prebiotically plausible amino acids, each as mixtures of their D- and L-isomers. (G) Library RNAs that triphosphorylated their 5'-hydroxyl group were ligated to a 5'-biotinylated oligonucleotide (red) using the R3C ligase ribozyme (blue). This allowed their (H) capture and isolation via streptavidin coated magnetic beads. (I) Reverse transcription to DNA and (J) PCR amplification re-generated the DNA library, now enriched for sequences that were able to self-triphosphorylate in the presence of the ten peptides.

therefore 8 stereoisomers contained only 1/8 of that concentration – 125 μM – for each stereoisomer. Importantly, there was no selection pressure to utilize these peptides: the procedure only selected for self-triphosphorylation catalysis. Therefore, if a large fraction of the selected ribozymes would depend on peptides, it would show that evolving RNAs adapt to the presence of peptides, and suggest that prebiotic peptides may increase the frequency of ribozymes in sequence space. RNAs that successfully catalyzed the reaction during the incubation with cTmP and the ten peptides, and therefore carried a 5'-triphosphate, were isolated *via* ligation to a biotinylated oligonucleotide and streptavidin-coated magnetic beads, reverse transcribed, and PCR amplified to complete one cycle of selection. After four rounds of selection, the RNA library was dominated by catalytically active sequences as judged by the decrease of PCR cycles required after reverse transcription (Fig. S1, ESI†).

High throughput sequencing (HTS) analysis showed that more than 200 sequence clusters were enriched in the selection protocol (Fig. 2A). To identify whether individual RNAs were benefitting from the presence of peptides, a fifth selection round was appended, in 22 different variants that differed only in the presence of peptides. The first variant was in the presence of all ten peptides, each at 1 mM concentration. The second variant was done in the absence of peptides. The remaining 20 variants were done in the presence of each one of the ten peptides, at either 1 mM or 10 mM concentration. After

HTS analysis of these 22 sub-libraries and the four preceding selection rounds, the enrichment of sequence clusters in the presence of individual peptides was determined (Fig. S2, ESI†). To identify ribozymes that may benefit from peptides in their activity, the increase of reads in the presence of individual peptides was determined. No cluster with at least 20 reads showed a stronger peptide-mediated read enhancement than 4-fold, suggesting that none of the ribozyme clusters benefitted more than 4-fold from peptides (the 9.4-fold peptide benefit for cluster 408 could be a result of statistical error because this cluster contained only 16 reads). To test the outcome with the ribozymes that appeared most likely to have a peptide benefit, 30 sequences with a high enrichment in the presence of peptides, and a number of reads without peptides that was high enough for statistical purposes were chosen for further analysis (colored symbols in Fig. 2B; sequences in Fig. S3, ESI†).

To identify peptide/ribozyme pairs where the peptide may increase ribozyme activity, the 30 chosen ribozyme sequences were tested for catalytic activity under selection conditions, in the absence and presence of the specific peptides that were suggested as beneficial by HTS analysis (Fig. S4A, ESI†). The strongest peptide benefit for ribozyme activity was 1.65-fold, consistent with the idea that under selection conditions, these peptides mediated little benefit to the ribozymes. To test whether peptides could benefit selected ribozymes under sub-optimal reaction conditions, the pH in the assay was decreased to pH 7.0, which facilitated about 2-fold peptide benefit for



E

P1	Gly.Ala.Gly. β Ala.Gly.Asp.Gly.Ala
P2	Ala.Gly. β AB.Gly.Gly. β Ala.Gly.Ala
P3	Gly. β Ala.Val. α AIB.Ala. α AB.Asp. β AB
P4	Ala. β AB.Asp. β Ala.Val.Gly. γ AB.Ser
P5	Gly.Asp.Gly.Ala. β Ala.Gly.Asp.Ala
P6	Ala. β AB.Asp.Gly. α AB.Asp. β Ala.Val
P7	Gly. α AIB.Gly. α AB.Ala.Ala.Asp.Val
P8	Ala.Gly. α AB.Gly.Gly. α AIB.Val.Ser
P9	Ala. β Ala.Gly. α AB. β AB. γ AB.Asp. α AIB
P10	Gly. β Ala.Ala. α AB. β AB. γ AB.Gly. α AIB



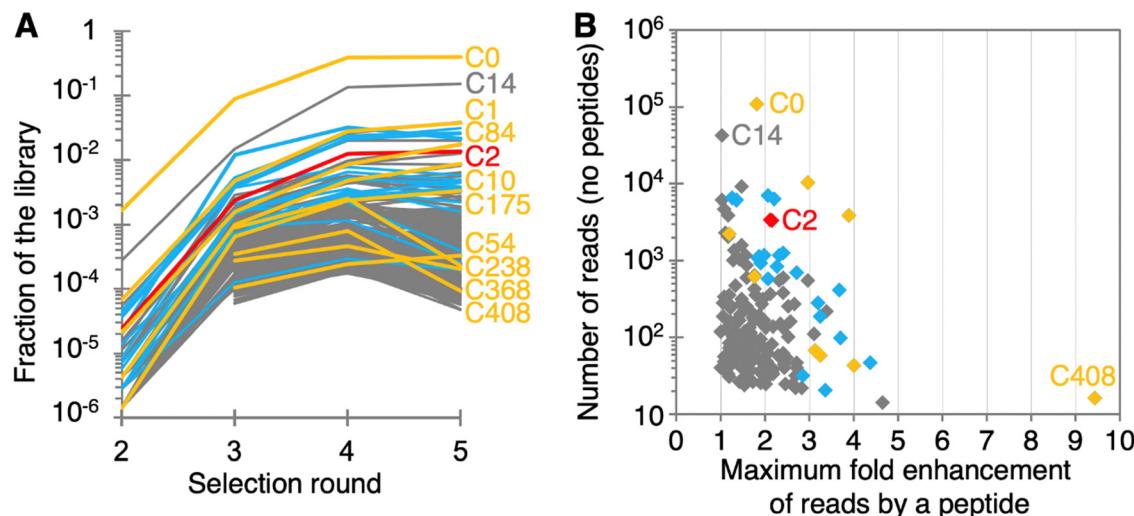


Fig. 2 High throughput sequencing analysis of selection rounds 2–5. (A) Enrichment of sequence clusters over multiple rounds of selection. Each line represents one sequence cluster. The number of sequence reads is shown for each of the 212 most abundant clusters with at least 500 reads in the 25 sub-libraries of selection round 5. Lines in blue denote clusters that appeared promising for peptide benefit and were screened in a biochemical assay. Lines in yellow denote ten clusters that were then tested with attention to the individual peptide's influence on reaction pH. The red line denotes cluster C2, which was analyzed in more detail. (B) Predicted peptide benefits for the same sequence clusters as in (A). Each diamond represents one sequence cluster, using the same color coding as in (A), with four clusters labeled to help correlation with (A). The vertical axis shows the number of reads in the selection sub-round 5 without peptides, while the horizontal axis shows the fold enhancement by the most beneficial peptide. This plot was used to choose sequences for biochemical analysis.

ribozyme S2 (Fig. S4B, ESI†). For additional insight into ribozyme S2, it was truncated at its 3'-terminus, reducing its length from 201 nucleotides in the original isolate to 180 nucleotides while increasing ribozyme activity (Fig. S5, ESI†), and its secondary structure was probed using SHAPE (Fig. S6, ESI†).

Because the HTS data appeared unreliable to identify small peptide benefits, the biochemical assays were expanded to a larger number of ribozyme/peptide combinations. The activity of ten different ribozymes was measured in the presence of the ten individual peptides, in 100 ribozyme/peptide combinations (Fig. 3A). The effect of the ten peptide preparations on the activity of the ten ribozymes was largely described by their influence on the reaction pH. This pH influence was mostly due to an unintended carryover of ammonium hydrogen carbonate from the purification procedure: NH₄HCO₃ leads to a pH increase in solution because CO₂ has a higher vapor pressure than NH₃ in aqueous solution. For nine of the ten ribozymes, the logarithm of ribozyme activity was well-correlated with the reaction pH with a slope of 1.0. This slope is indicative of a single deprotonation event as the rate limiting step of catalysis, and was observed previously for a self-triphosphorylation ribozyme.²⁹ This deprotonation is likely the deprotonation of the 5'-hydroxyl group, which is rate limiting for its nucleophilic attack on a phosphorus of cyclic trimetaphosphate. However, ribozyme S2 showed significant deviations from this correlation (yellow symbols in Fig. 3A), with increases of activity in the presence of peptides P4 and P2. Therefore, ribozyme S2 was studied in more detail.

To measure the peptide influence on the activity of ribozyme S2 without the confounding influence of ammonium hydrogen carbonate (Fig. 3B), all peptide preparations were subjected to

an additional, 12-hour long lyophilization procedure. The remaining influence of peptide preparations on the reaction pH was therefore caused by the influence of the peptide's amino and carboxy groups, reflected in the peptide's overall charge (the carboxy terminus was used as amide and therefore uncharged). When ribozyme S2 was then incubated with cTmp in the presence or absence of the re-purified peptides, the three peptides with an overall positive charge (P2, P8, P10) resulted in the expected 2- to 3-fold increase in ribozyme activity. Peptides 5 and 6 carried an overall negative charge and did not increase ribozyme activity. In contrast, peptide 4 mediated a 2.6-fold increase in ribozyme activity. Peptides P1, P3, P7 and P9 also did not carry an overall charge and also appeared to increase ribozyme activity by an average of 1.6 ± 0.2-fold. Together, this means that these uncharged peptides mediate a less than 2-fold, nonspecific benefit for ribozyme activity, and that peptide 4 increased the activity of ribozyme S2 slightly more, due to specific interactions of peptide 4 with the ribozyme.

All ribozymes tested above had been selected in the presence of the ten studied peptides, and might therefore have adapted to the presence of these peptides. To test whether ribozymes that were selected in the absence of peptides would be influenced by the peptides, four ribozymes that were selected in the presence of only 50 mM MgCl₂, 100 mM Na₃cTmp, and 50 mM Tris/HCl pH 8.3^{29,30} were tested in the same buffer but in the presence of the peptides used in this study (Fig. 4). None of the four ribozymes showed a peptide-mediated increase or decrease in activity. Together, these data show that ten octapeptides consisting of D/L mixtures from prebiotically plausible amino acids did not strongly increase, or decrease the activity of self-triphosphorylation ribozymes.

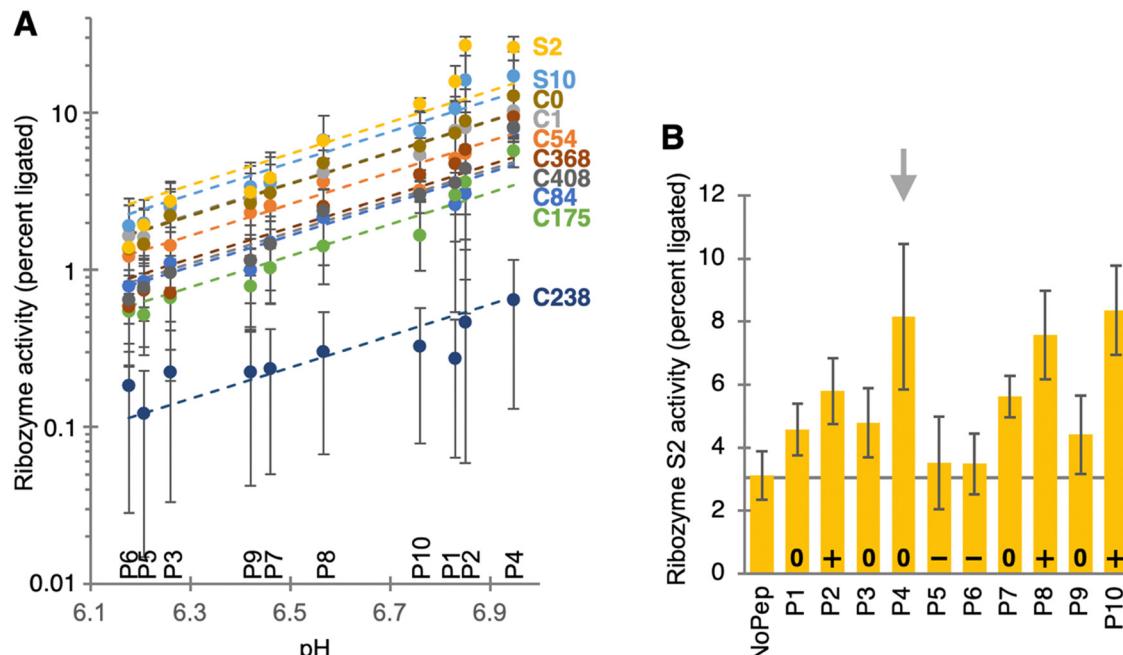


Fig. 3 Biochemical analysis of ribozyme activity dependent on the presence of 10 different peptides. (A) Ribozyme activity plotted on a logarithmic scale as function of the pH shift caused by peptide preparation for each of 10 peptides at 10 mM concentration. The color of each ribozyme's data is annotated with the color of its label on the right. The dashed lines are fitted to each ribozyme's data with a slope of 1.0. Error bars are standard deviations of triplicate experiments. (B) Activity of ribozyme S2 in the presence of 10 mM of each individual peptide, after removing traces of ammonium carbonate from the peptide preparations. The remaining overall charge of each peptide is indicated at the bottom of each column. The peptide without overall charge and the strongest ribozyme benefit (P4) is indicated with a grey arrow. Error bars are standard deviations of triplicate experiments.

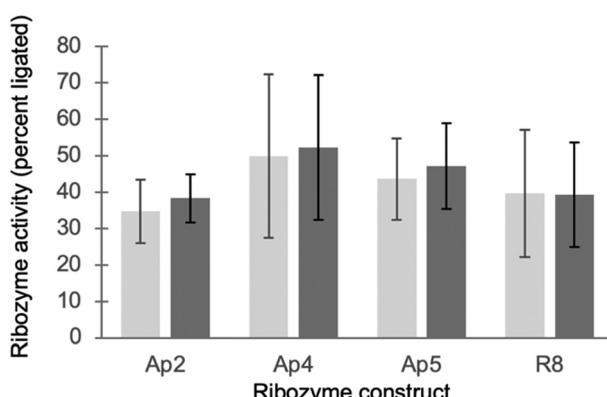


Fig. 4 Biochemical analysis of the effect of prebiotically plausible peptides on four ribozymes that were selected in the absence of peptides. Ribozyme activity is shown as a function of the different ribozyme constructs (Ap2, Ap4, Ap5, R8), in the absence (light grey) or presence (dark grey) of the same peptides as used in the selection of the current study. Error bars are standard deviations of triplicate experiments.

Discussion

To test whether prebiotically plausible peptides can increase the frequency and/or activity of ribozymes in RNA sequence space, an *in vitro* selection for self-triphosphorylation ribozymes was conducted in the presence of ten prebiotically plausible peptides. This selection identified hundreds of ribozyme clusters, and 10 sequences were analyzed biochemically with each individual peptide for possible benefits. Most peptide

effects on ribozyme activity were explained by their effect on reaction pH. A less than 2-fold, non-specific benefit for ribozyme activity was found for four uncharged peptides, while the uncharged peptide P4 mediated a 2.6-fold benefit for ribozyme S2. The data suggest for the specific system used (self-triphosphorylation ribozymes in the presence of ten specific octamers at 1 mM concentration, at 50 mM free Mg²⁺ concentration and pH 8.3) the peptides provided only weak benefits for the frequency of ribozymes in sequence space, and the activity of the selected ribozymes.

The chemical structure of the ten peptides (Fig. 5) allows to speculate what features of the peptides may be responsible for the beneficial effect especially of peptide 4 on ribozyme S2. However, the small differences between the benefits of uncharged peptides could be caused by interactions weaker than a single hydrogen bond, therefore the current data set is insufficient to pin-point the feature(s) of peptide P4 mediating the 2.6-fold benefit for ribozyme S2.

This study explored how prebiotically plausible peptides could have influenced the emergence and function of catalytic RNAs. The interpretation of this mostly negative result with respect to the origin of life – no RNA/peptide couples were identified where peptides increased RNA activity more than 3-fold – needs to be done with care, given that only ten specific octapeptides were studied, that ribozymes were selected only for self-triphosphorylation activity, that all experiments were done with relatively high Mg²⁺ concentration (50 mM free Mg²⁺), and that only peptide concentrations up to 10 mM were

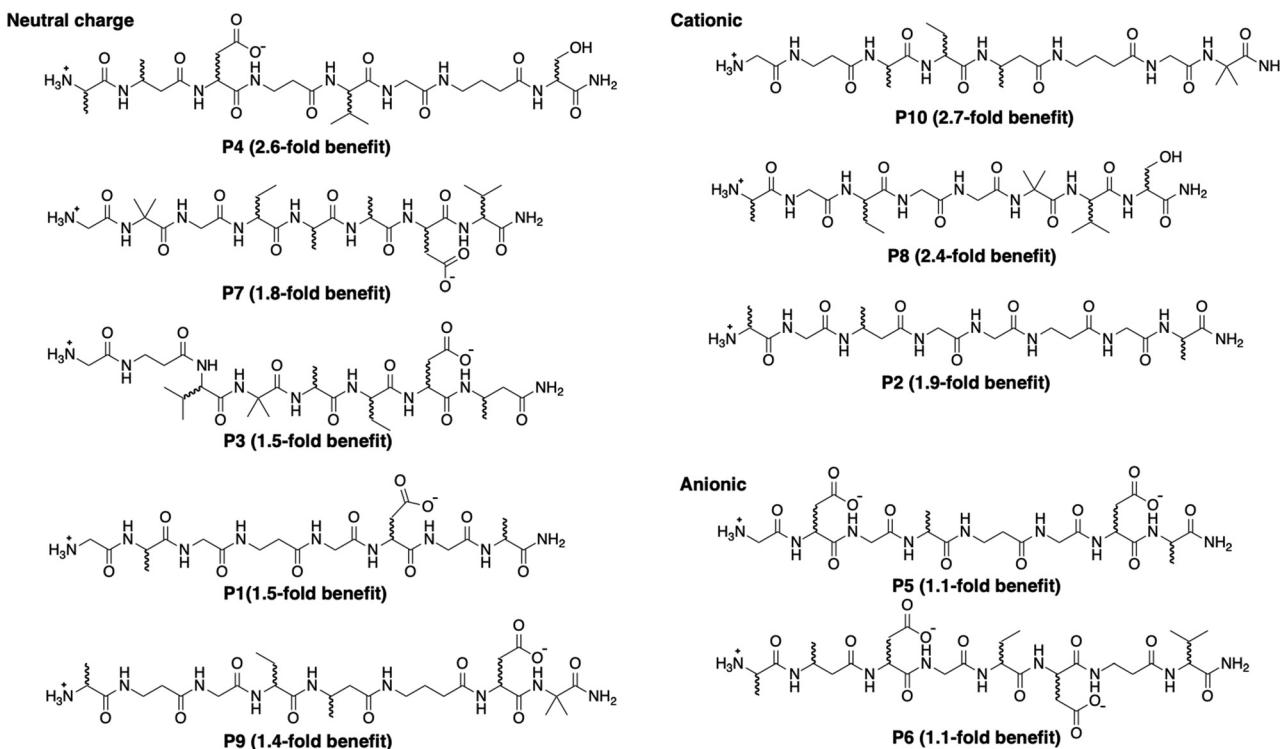


Fig. 5 Chemical structure of the ten peptides used in this study. The peptides are sorted by their overall charge (neutral charge left, cationic top right, anionic bottom right) as well as their effect on ribozyme activity within each group, with the top peptides more beneficial. The fold benefit on ribozyme activity is given next to the peptide name. Note that the effects of cationic and anionic peptides are easiest interpreted by their influence on reaction pH. All chiral centers carry both enantiomers, indicated by squiggly lines.

explored. In addition, it needs to be considered that peptides with encoded sequences were likely not present at the emergence of an RNA world and only generated at the end of an RNA world, when a precursor to the ribosome was able to polymerize amino acids with some sequence fidelity. Given these constraints, it is useful to consider the two extreme hypotheses: the hypothesis that prebiotically plausible peptides were essential for catalytic RNAs is likely incorrect because many ribozymes have been selected, and are active in the absence of peptides. The hypothesis that peptides had no influence on the rise of an RNA world organism is likely also incorrect because peptides likely existed in the same environment in which RNA emerged, and some peptides can support ribozyme function (see introduction and the two paragraphs below). The question is whether peptides in a prebiotic environment could have supported the emergence and function of catalytic RNAs. The narrow interpretation of the results in the present study is that the type of ribozymes studied here (self-triphosphorylation ribozymes) does not benefit more than 3-fold from the ten used octapeptides, under the specific conditions. While only ten octapeptides were studied, it needs to be considered that prebiotically plausible peptides were not synthesized by a ribosome, and did not have an encoded sequence. In the absence of such sequence encoding, the representation of the ten used prebiotically amino acids by the ten used peptides can serve as an 'average view' that suggests that most peptides based on these ten amino acids are unlikely to strongly affect the emergence of self-triphosphorylation

ribozymes, and perhaps many catalytic RNAs. However, it is possible that other sequences of the same amino acids would benefit self-triphosphorylation and other ribozymes. For this to be relevant in a prebiotic scenario, local enrichment by a process of chemical evolution would have to be postulated. For example, polycationic peptides could become enriched in coacervates with RNA⁴⁰ and aromatic amino acids may form amyloids or other aggregates, especially if they are homochiral.⁴¹ This situation changes after the appearance of the first proto-ribosome and amino acid metabolism, when amino acid sequences can be encoded, when homochiral as well as aromatic amino acids become available, and when the benefits seen in today's proteins can complement the function of catalytic RNAs.^{42–44}

If cationic amino acids such as ornithine and 2,4-diamino butyric acid could have been abundant in prebiotic peptides then the corresponding peptides may have changed the outcome of this study. These amino acids were not included in the current study because their abundance in prebiotic model reactions is far below that of the ten used amino acids.¹⁹ However, it is well documented that peptides that are rich in the cationic amino acids arginine or lysine can improve ribozyme activity by increasing the thermodynamic stability of RNA/RNA duplexes,²⁵ stabilizing the sequence non-specific binding of an RNA substrate by a ribozyme,²⁶ or by forming coacervates with the RNA that lead to better RNA/RNA interactions.²⁷ Peptides containing multiple cationic and multiple hydrophobic amino acids can enhance the activity of polymerase ribozymes through the formation of

aggregates.⁴⁵ All of these studies required multiple cationic amino acids to benefit ribozyme function, and the requirement of multiple rare amino acids further reduces the peptide's expected abundance in a prebiotic setting. However, very rare peptides could have become enriched and co-localized with RNA by partitioning of polycationic peptides in coacervates with polyanionic RNA. It remains to be seen whether polycationic peptides can be generated, and enriched in prebiotic model reactions in sufficient abundance to provide their benefit to catalytic nucleic acids.

While aromatic amino acids are rare or absent in prebiotic model reactions,¹⁹ peptides containing multiple aromatic amino acids are able to aggregate in amyloids and then interact sequence specifically with short RNA fragments.⁴¹ Moreover, aromatic moieties have been shown to benefit ribozyme activity: conjugates between polycyclic aromatic hydrocarbons and peptides are able to increase the activity of some ribozymes by 100-fold or more.²⁸ Since insoluble organic matter is the dominant form of carbon in meteorites, and rich in polycyclic aromatic hydrocarbons,^{46,47} it is possible that the prebiotic environment was rich in polycyclic aromatic hydrocarbons, and present in prebiotic peptides. This may have acted as 'prebiotic equivalent' to the aromatic amino acids present in modern proteins. However, it is unclear whether this usefulness of PAHs for ribozymes is a rare phenomenon or whether it may have been wide-spread and important during an RNA world. Future work will show what kind of prebiotically plausible molecules could have aided the emergence and function of an RNA world organism.

Materials and methods

Synthesis and purification of peptides

Peptides were synthesized on rink amide PS resin (0.77 mmol g⁻¹, Novabiochem) and Rink Amide ChemMatrix (0.45 mmol g⁻¹, Biotage) following the Fmoc/tBu strategy using a microwave-assisted peptide synthesizer (Alstra, Biotage). Side chain protecting groups for amino acids were as follows: Ser(tBu), His(Trt), Lys(Boc), Asp(OtBu), Glu(OtBu). Unnatural amino acids used for the syntheses of prebiotically plausible peptides were Fmoc-D-Ala-OH, Fmoc-D-Asp-OH (OtBu), Fmoc-D-Val-OH, Fmoc-D-Ser-OH(tBu), Fmoc-β-alanine, Fmoc-α-L-aminobutyric acid, Fmoc-α-D-aminobutyric acid, Fmoc-L-β-aminobutyric acid, Fmoc-D-β-aminobutyric acid, γ-aminobutyric acid, Fmoc-α-amino butyric acid. For the peptides containing prebiotically plausible residues, a racemic mixture (1:1) was used for each amino acid reagents. Typical coupling reactions were performed at 0.100–0.200 mmol scale with 3.0 equiv. of Fmoc-protected amino acids, 2.94 equiv. of HBTU and 6.0 equiv. of NMM for 5 min at 75 °C. All reagents were pre-dissolved in DMF at 0.5 M. Fmoc deprotections were performed with 20% piperidine in DMF for 10 min at rt. Washing was performed after every deprotection and coupling step using DMF. For the syntheses of some sequences, the deprotection and coupling steps described above were either performed twice or with increased reagent equivalence to ensure reaction completion. Cleavage of the peptide sequences from the solid support

with concomitant side chain deprotection was accomplished by placing the resin in fritted SPE tube and treating with TFA cleavage cocktail (~20 mL g⁻¹) containing 90:5:2.5:2.5 TFA:dimethoxybenzene:H₂O:TIS for 2 hour. Cleaved peptides were then precipitated in cold ether, centrifuged, dissolved in methanol and reprecipitated in cold ether (3×).

HPLC characterization and purification of peptides were carried out at rt on analytical (Jupiter C18 5 μm, 300 Å, 150 × 4.6 mm by Phenomenex, Torrance, CA) and semi-preparative columns (Aquasil C18 5 μm, 100 Å, 150 × 10 mm by Keystone Scientific Inc., Waltham, MA) with Prostar 325 Dual Wavelength UV-Vis Detector from Agilent Technologies with Varian pumps (Santa Clara, CA) with detection set at 225 and 406 nm. Peptides were eluted from column following a gradient using mobile phases A: 0.1% TFA in H₂O and B: 0.1% TFA in CH₃CN. MS analyses were obtained on a LTQ ESI-MS spectrometer (San Jose, CA). Solutions were prepared in either methanol or methanol/water (formic acid 1%) with flow rate of 10 μL min⁻¹, spray voltage at 4.50 kV, capillary temperature at 300 °C, capillarity voltage at 7.00 V, tube lens at 135.00 V. Purified peptides were characterized by analytical HPLC (with purity typically greater than 90%) and MS (either by direct injection or LC-MS).

To remove a possible carryover of TFA, peptides were dissolved in a total volume of 2 to 5 mL 100 mM (NH₄)HCO₃, and frozen as a thin film on the inside of a glass bulb cooled in a dry ice/isopropanol bath. After lyophilizing the frozen solution to dryness in oil vacuum (~2 mbar), the process was repeated once with 100 mM (NH₄)HCO₃, and once or twice with water. The desiccated peptide was weighed, and dissolved in water to a stock concentration of 10 mM.

In vitro selection

A 183 nucleotide (nt) long DNA oligomer that included 150 positions with randomized sequence and flanked by 5'- and 3'-constant regions was purchased from integrated DNA technologies (IDT). 5'-GAGACCGCAAGAGAC(N150)CATGGTTC AGACTACAAC-3' A 82 nt long oligomer was used to add the promoter sequence of T7 RNA polymerase and the sequence of the hammerhead ribozyme (underlined) to the 5' end of the 183-mer. 5'-AATTAAATACGACTCACTATAGGGAGCGGTCTCCCTG ACGAGCTAACGAAACTGCGGAAACGCAGTCGAGACCGCAAGA GAC-3'. Amplification with the 67-mer and a short 3' primer generated the double stranded DNA pool. qPCR was used to determine the amount of amplifiable DNA and this number was used to estimate that the starting DNA pool consisted of 4.2×10^{14} sequences. The DNA pool was transcribed into RNA using T7 RNA polymerase under standard conditions. 100 nM of purified RNA sub-pool was incubated with 50 mM Tris/HCl, 3.3 mM NaOH, 100 mM MgCl₂, 50 mM of freshly dissolved trimetaphosphate and 1 mM each of ten prebiotically plausible peptides (10 mM total). This mixture was allowed to incubate for 3 hours at room temperature. The reaction was quenched by ethanol precipitation. The RNA pellet was washed with chilled water and desalting using P30 Tris/HCl spin columns (Bio-Rad). Active ribozymes were ligated to a biotinylated oligomer (biotin-d(GTAACATAATGA)rU)



using the R3C ligase ribozyme whose arms were designed to anneal to the 5' constant region of the pool RNA and the biotinylated oligomer. A mixture containing 1.3 μ M of desalted pool RNA, 1 μ M ligase ribozyme, 1.2 μ M biotinylated oligomer, 100 mM KCl, 100 mM Tris/HCl, and 60 pM triphosphorylated RNA was heated to 65 °C for 2 minutes then cooled to 30 °C at a rate of 0.1 °C per second. The triphosphorylated RNA was used to generate a small amount of ligation product and thereby reduce the number of PCR cycles, and prevent artifacts during PCR amplification. After heat renaturation, an equal volume of a mix consisting of 40% (w/v) PEG8000, 4 mM Spermidine, and 50 mM MgCl₂ was added to the ligase reaction mixture. This mixture was incubated at 30 °C for 3 hours. After the incubation step, the reaction was quenched by adding final concentrations of 13.9 mM sodium/EDTA, 50 mM Tris/HCl, 50 mM KCl, 0.012% (w/v) Triton X-100, and 1.19 μ M of a 61 nt oligomer that was complementary to the ligase ribozyme (5'-AGTAACATAATGACT TCAACCCATTCAAAC TGTCTTACGAACAATCGAGCACAAGAGAC-3'). This mixture was heated at 50 °C for 10 minutes. Streptavidin magnetic beads (Promega) were washed thrice with 20 mM HEPES/KOH pH 7.2, 0.01% (w/v) Triton X-100 and 50 mM KCl. Biotinylated RNA was then captured by mixing the RNA with the magnetic beads and rotating end-over-end at room temperature for at least 30 minutes. A magnetic rack was used to focus the beads and the beads were washed twice with a solution containing 0.01% (w/v) Triton X-100 and 20 mM NaOH. Captured RNA was eluted from the beads by incubating the beads in a solution of 25 mM Tris/HCl pH 8.5, 1.56 mM EDTA and 96% formamide at 65 °C for 3 minutes. The beads were removed by immediate centrifugation, and the supernatant was concentrated by ethanol precipitation. 1 μ g of tRNA was used as precipitation carrier. Captured RNA was resuspended in 10 mM Tris/HCl, pH 8.3. The RNA was reverse transcribed using superscript III (Invitrogen) and a reverse transcription primer (5'-GTTGTAGTCTGAACCATG-3') complementary to the 3' constant region. The products were then PCR amplified with 5' and 3' primers that could bind to the sequence of the biotinylated oligomer and the 3' constant region of the pool respectively. A second PCR amplification was performed to add the T7 promoter sequence and the hammerhead ribozyme sequence to the DNA.

HTS analysis

Illumina sequencing primers were added to the ends of rounds 2–5 pools. Pools were sequenced on MiSeq with 10% PhiX. FastQ files were processed using custom Python scripts for rounds 2–5 (including all sub-rounds in round 5). The usearch merge function was used to merge forward and reverse reads. Sequences not containing both constant regions or short randomized regions (<135 nt in length or 10% of 150) were first filtered out. The file was reformatted to Fasta for easier downstream processing. The total counts for each sequence were calculated. The usearch clustering algorithm was used to group similar sequences into clusters. Clusters were defined as sequences that shared at least 75% sequence identity and had at least 100 reads in a given round. Clusters and sequences were tracked through each round. 189 sequences were evaluated further. For each sequence,

the fold enrichment in the round 5 sub-rounds were determined by normalizing to the no peptide sub-round.

Generation of 3' truncations

3' truncations were generated by PCR using Taq DNA polymerase and a series of 3' primers. The 3' primers annealed to different portions of the ribozyme sequence of interest to generate a series of truncations with desired deletions from the 3' end. DNA templates of each truncation were transcribed using T7 RNA polymerase, and purified by denaturing PAGE.

Self-triphosphorylation activity assays

Purified ribozyme (5 μ M) was incubated with 50 mM Tris/HCl, 100 mM MgCl₂, 50 mM of freshly dissolved trimetaphosphate, and either a mixture of peptides, individual peptides or water as a control. This mixture was incubated for 3 hours at room temperature. The products were ligated to a radiolabeled oligomer (γ ³²P-d(GTAACATAATGA)rU) using an R3C ligase ribozyme whose arms were designed to anneal to the 5' constant region of the pool RNA and the oligomer. Following the incubation with buffer, Tmp and peptides, the mixture was diluted 1:10 in a solution that contained a final concentration of 0.5 μ M ligase ribozyme, 0.5 μ M biotinylated oligomer, 100 mM KCl, 100 mM Tris/HCl, and 15 mM sodium EDTA. This mixture was heat renatured at 65 °C for 2 minutes then cooled to 30 °C at a rate of 0.1 °C per second. After heat renaturation, an equal volume of a mix consisting of 40% PEG8000 (w/v), 4 mM Spermidine, and 50 mM MgCl₂ was added to the ligase reaction mixture. This mixture was incubated at 30 °C for 2 hours. After the incubation step, the reaction was quenched by ethanol precipitation. The reaction products were resolved on a denaturing 15% polyacrylamide denaturing gel.

SHAPE probing

Selective 2'hydroxyl acylation analyzed by primer extension (SHAPE) was performed on the 180-nucleotide long ribozyme isolated in this study, with 1-methyl-7-nitro-2H-3,1-benzoxazine-2,4(1H)-dione (1M7) as the chemical probe. 50 pmol of ribozyme were incubated in buffer containing 50 mM MES/NaOH pH 6.0, 100 mM MgCl₂, 3.3 mM NaOH, 50 mM Tmp, and 10 mM peptide 4 at room temperature for 2 minutes. This mixture was incubated at room temperature for 2 minutes or 2 hours. 1M7 was added to this solution such that its final concentration was 2 mM and the concentration of DMSO was 1% of the solution. A negative control was prepared with no 1M7 and 1% DMSO. Both samples were incubated at room temperature for 3 minutes. The reaction was quenched by ethanol precipitation and resuspended in 10 μ L of 5 mM Tris/HCl pH 8.0. The products were reverse transcribed using superscript III reverse transcriptase (Invitrogen) according to manufacturer's instructions and trace amounts of a radiolabeled primers were added. Three reverse transcription primers were used to probe different portions of the ribozyme. The sequences of these primers are (5'-TTAGTCTGAACCATG-3', annealed to pos. 165–180); (5'-CGATGTTGACATTTTC-3', annealed to pos. 112–127); and



(5'-AAACTCAAGTGGGTTT-3', annealed to pos. 53–68). The RNA template in each sample was degraded by alkaline hydrolysis, by incubating for 5 minutes at 80 °C in a solution containing 750 mM NaOH. The reaction was quenched by adding a two-fold stoichiometric excess of acetic acid to generate a NaOAc/HOAc buffer with a final concentration of 300 mM. The products were ethanol precipitated, resuspended in formamide loading buffer and resolved on a denaturing 10% polyacrylamide gel. The RNA secondary structure was predicted with help by the secondary structure prediction tool RNAstructure (ReuterMathews10). To do this, the SHAPE data was mapped onto an initial secondary structure prediction and manually adjusted to better represent the SHAPE data.

Data availability

The data supporting this article have been included as part of the ESI.†

Conflicts of interest

There are no conflicts of interest to declare.

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