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Microfluidic nanobubbles: observations of a sudden contraction of microbubbles into nanobubbles†

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Microfluidic devices are often utilized to generate uniform-size microbubbles. In most microfluidic bubble generation experiments, once the bubbles are formed the gas inside the bubbles begin to dissolve into the surrounding aqueous environment. The bubbles shrink until they attain an equilibrium size dictated by the concentration and type of amphiphilic molecules stabilizing the gas–liquid interface. Here, we exploit this shrinkage mechanism, and control the solution lipid concentration and microfluidic geometry, to make monodisperse bulk nanobubbles. Interestingly, we make the surprising observation of a critical microbubble diameter above and below which the scale of bubble shrinkage dramatically changes. Namely, microbubbles generated with an initial diameter larger than the critical diameter shrinks to a stable diameter that is consistent with previous literature. However, microbubbles that are initially smaller than the critical diameter experience a sudden contraction into nanobubbles whose size is at least an order-of-magnitude below expectations. We apply electron microscopy and resonance mass measurement methods to quantify the size and uniformity of the nanobubbles, and probe the dependence of the critical bubble diameter on the lipid concentration. We anticipate that further analysis of this unexpected microbubble sudden contraction regime can lead to more robust technologies for making monodisperse nanobubbles.

1. Introduction

Nanobubbles (NBs) are nanometer-size bubbles characterized into two categories: bulk and surface NBs.¹ Bulk nanobubbles (BNBs) are gaseous domains surrounded by liquid with a diameter smaller than 1000 nm. Surface NBs are gaseous domains in the shape of a hemispherical cap that originate on solid surfaces. The height of the cap is typically 10–100 nm, and the radius is 50–500 nm.^{1,2} The focus of this paper is on the microfluidic generation of BNBs.

There are emerging applications of BNBs in various fields, including wastewater treatment, aquaculture, medicine, surface cleaning, flotation, and other areas.^{1–6} Different mechanical and chemical production methods have been used to produce BNBs.

In a recent review paper, we discussed each of the existing fabrication methods for making BNBs, their strengths and weaknesses, and how microfluidics may help to address the current challenges in making BNBs.¹

Although many papers report the observation of long-lived BNBs, there are still many debates about the existence and long-term stability of BNBs. The pressure difference between the inside and outside of a bubble is typically dependent on the gas–liquid surface tension and the bubble's radius, as described by the Young–Laplace equation, $\Delta P_{\text{Laplace}} = P_{\text{inside}} - P_{\text{outside}} = 2\gamma/r$, where P_{inside} and P_{outside} are the pressures inside and outside of the bubble, respectively.⁷ Here, γ is the gas–liquid surface tension, and r is the bubble radius.⁷ Therefore, a bubble that has a radius in the nanometer range should have a large internal pressure. According to Henry's law, the excessive pressure difference between the inside and outside of the bubble should lead to a change in the capacity of the liquid surrounding the bubble to absorb the gas.⁸ This should result in one of two destabilizing scenarios: (1) the bubble size increases as additional gas molecules diffuse into the bubble if the liquid is already saturated with the gas, or (2) the bubble shrinks, if the surrounding liquid is not at saturation, until the bubbles completely dissolve from the solution, in a vicious shrinkage cycle.⁸

Why and how stable BNBs are observed experimentally is still being debated. However, one hypothesis for the apparent stability of surfactant stabilized BNBs is the following. After a microbubble

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(MB) forms, the gas inside the bubble diffuses into the surrounding liquid due to the outward driving force caused by Laplace pressure. As a result, the MB's diameter decreases, and if the level of shrinkage is sufficiently high, the MBs shrink to BNBs.^{9,10} Some papers report that during the shrinkage process the surfactants remain on the surface of the bubbles, owing to a combination of van der Waals and hydrophobic interactions between the surfactants that maintains the cohesiveness of the surfactant layer.^{11,12} The presence of the surfactant layer, or shell, lessens the gas-liquid surface tension, thereby reducing the Laplace pressure, and resulting in stable BNBs.¹³ Some papers speculate that the surfactants forming the BNB shell create an electrostatic pressure that balances the Laplace pressure, and prevents the bubble from perishing.^{13,14}

Over the past decade, microfluidics has emerged as a viable tool to produce BNBs. Peyman *et al.* utilized a technique called microfluidic atomization to create BNBs.¹⁵ In the atomization technique, the combination of shear stress and pressure at the microfluidic orifice create an atomization-like phenomenon, forming a mixed population of MBs and BNBs.¹⁵ In another study, Abou-Saleh *et al.* optimized the atomization method,^{15,16} demonstrating a new micro-spray regime by increasing the height of the microchannel downstream of the orifice. In this regime, the sudden pressure drop downstream of the orifice leads to conditions that enable BNB generation.¹⁶ However, a main drawback of the atomization method is that the resulting bubble population includes both MBs and BNBs, making the technique challenging for applications that exclusively require BNBs.^{1,15,16}

In this paper, we utilize a microfluidic MB shrinkage method our group recently developed,¹⁷ and extend the scope of the technology to produce BNBs. We previously showed that the degree of MB shrinkage in a microfluidic setting can be controlled by the solution's lipid concentration, and the MBs can shrink in diameter by as much as tenfold. With this knowledge, here we develop a new microfluidic geometry with a smaller flow-focusing orifice so that we can generate sub-20 μm diameter MBs that may shrink to become BNBs. Surprisingly, we discover that even MBs of larger sizes can suddenly reduce to become stable BNBs. We find that when the initial diameter of MBs is less than a critical threshold, the MBs unexpectedly shrink to approximately the same nanoscale final diameter, regardless of their initial diameter. In contrast, when the MB initial diameter is greater than the critical threshold, the MBs shrink by a factor of ten and do not transform into BNBs. To the best of our knowledge, this is the first experimental report of a critical MB diameter that delineates between MBs shrinking by the expected amount, and MBs shrinking dramatically to become stable and relatively monodisperse BNBs.

2. Materials and methods

2.1 Microfluidic device fabrication

We fabricate our microfluidic chip using the standard photolithography processes. We utilize a MicroWriter ML3 Pro (Durham Magneto Optics Ltd, England) as the photolithography machine. We design our microfluidic design with AutoCAD (AutoCAD 2018, Autodesk, Inc., Dan Rafael, CA) software, and

convert the output file using CleWin software to a format readable by the MicroWriter ML3 Pro machine. We first spin coat 20 μm of SU-8 2010 photoresist onto the surface of a silicon wafer. Then, to prebake the photoresist, we put the silicon wafer on a hot plate for 3 min at 65 $^{\circ}\text{C}$, and then for 5 min at 95 $^{\circ}\text{C}$. We impose our microfluidic design into the prebaked silicon wafer using MicroWriter ML3 Pro. This machine uses a 385 nm semiconductor light source. After this step, we post bake the wafer at 65 $^{\circ}\text{C}$ for 3 minutes and at 95 $^{\circ}\text{C}$ for 5 minutes to stabilize the microfluidic device structure on the silicon wafer. Finally, we remove the uncured SU-8 using a photoresist developer solution. We pour and stir a mixture of polydimethylsiloxane resin (PDMS, Sylgard 184, Dow Corning, USA) and curing agent with a ratio of 10:1 into a disposable plastic bowl.

Before pouring the polymer solution onto the structured silicon wafer, we degas the polymer mixture employing a desiccator to remove air bubbles. Next, we bake the mixture in an oven at 70 $^{\circ}\text{C}$ for 1 h. For each experiment, we cut and peel off a device from the PDMS slab. We use a 1 mm diameter biopsy punch (Integra Miltex, Inc., Rietheim-Weilheim, Germany) to create inlets and outlets. Finally, we utilize oxygen plasma bonding (Harrick Plasma, Ithaca, NY) to bond the PDMS device to a glass microscope slide (Corning microscope slides, 75 mm \times 25 mm, Sigma-Aldrich, St. Louis, MI). We connect Teflon tubing (PEEK, Upchurch) to the inlets to supply gas and liquid, and to the channel outlet to collect the bubbles.

2.2 Lipid formulation

We synthesize a total of 100 mg of lipid solution comprising 6:1:2:1 mass ratio of 1,2-dibehenoyl-*sn*-glycero-3-phosphocholine (DBPC), 1,2-dipalmitoyl-*sn*-glycero-3-phosphate (DPPA), 1,2-dipalmitoyl-*sn*-glycero-3-phosphoethanolamine (DPPE), and 1,2-distearoyl-*sn*-glycero-3-phosphoethanolamine-*N*-[carboxy(polyethylene glycol)-2000] (DSPE-mPEG-2000) (Avanti Polar Lipids, Alabaster, AL).^{17,18} To make a lipid solution with the concentration of 0.50 mg mL⁻¹, we heat 200 mL of phosphate-buffered saline (PBS; Thermo Fisher Scientific, Inc., Ontario, Canada) and 2 mL of glycerol (MilliporeSigma Canada Co., Ontario, Canada). Then, we mix the solution and add the solution to the lipids.

To ensure all lipids are entirely dissolved, we heat the lipid solution for 2 h at 80 $^{\circ}\text{C}$, and sonicate for 20 min. Before each experiment, we add 200 μL of Pluronic F-68 (Thermo Fisher Scientific, Inc., Ontario, Canada) to every 4 mL of lipid solution. Then, we filter the lipid solution through a 2 μm filtration membrane (Acrodisc Syringe Filters, VWR, Ontario, Canada) to remove any unwanted particles.

2.3 Blended gas

In all experiments, we use a mixed gas composing 0.17 wt% octafluoropropane (C₃F₈) in nitrogen (N₂) (Messer Canada, Inc., Mississauga, Canada). We use a microfluidic pressure pump (FLOW EZ, Fluigent, Paris, France) to accurately deliver the gas into the microfluidic device.

2.4 BNB generation

We generate BNBs in a flow-focusing chip with an orifice width of 4 μm connected to an outlet microchannel with a width of

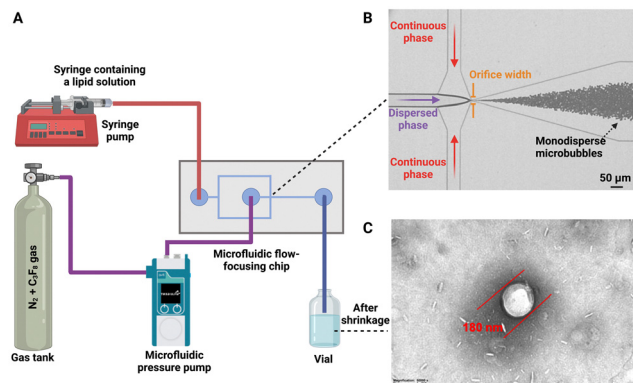


Fig. 1 (A) Schematic of the flow-focusing microfluidic design to generate BNBs. The width of the orifice is 4 μm , and the height of the microchannels is 20 μm . (B) Formation of monodisperse MBs at the orifice. (C) TEM image of a BNB after shrinkage with a diameter of 180 nm. The BNB's initial diameter before shrinkage was 10.0 μm . We use a 0.17 wt% C_3F_8 gas and a lipid solution with the concentration of 0.50 mg mL^{-1} .

502 μm . The width of the dispersed and continuous phase channels is 90 μm , and we keep the microchannel height at 20 μm everywhere in the device. As shown in Fig. 1A, we utilize a microfluidic pressure-controlled pump to have real-time control of the pressure of the gas entering the microfluidic chip. We deliver the aqueous lipid solution through the liquid inlet of the microfluidic chip employing a constant flow rate syringe pump (Harvard Apparatus, PHD 2000, Holliston, MA). The lipid and blended gas phases are pushed toward the flow-focusing orifice. The lipid solution enters the orifice from lateral microchannels and squeezes the gas at the orifice (Fig. 1B). Consequently, MBs are created at the orifice and travel downstream into the outlet microchannel. The outlet microchannel is connected, through a Teflon tubing, to a plastic vial that is open to the atmosphere.

2.5 MB and BNB analysis

We capture images of MBs at the microfluidic orifice using an inverted microscope (AX10, Carl Zeiss AG, Oberkochen, Germany) equipped with a high-speed camera (Phantom M110, Vision Research, Wayne, NJ). We use ImageJ (National Institutes of Health, Bethesda, MD) software calibrated using 5.9 μm diameter standard polystyrene particles (Thermo Fisher Scientific, Inc., Ontario, Canada) to measure the diameter of MBs.

We utilize several characterization methods to analyze the BNBs. First, we experiment with the Tyndall effect to confirm the existence of BNBs in the collected sample.^{3,19,20} To conduct this test, we first generate bubbles, and collect the bubbles in a plastic vial open to the atmosphere. We keep the bubbles in the vial for 1 day to allow gas exchange between the bubbles and the liquid environment. In the test, we illuminate the solution containing BNBs with a red laser beam. BNBs in the sample scatter the light and brighten the solution inside the vial, confirming BNBs in the solution.

As Fig. 1C shows, we secondly utilize transmission electron microscopy (TEM, Hitachi HT7800, Japan) to capture images of

the BNBs after shrinkage.²¹ The bubbles are kept inside a plastic vial for 1 hour before TEM imaging. To start the TEM imaging, first we plasma clean the TEM grid before each imaging session. We pipet 25 μL of BNB solution and three droplets of DI water, as a dilution reservoir, onto a parafilm sheet. We put the TEM grid on the BNBs droplets and leave it for 3 minutes. Then, we dilute the BNBs into the grid by placing the grid on DI water droplets and leaving it there for 1 minute. After dilution, we apply 20 μL of 2% uranyl acetate on the grid as a stain. We remove the excess stain and air-dry the grid before imaging.

Finally, to measure the diameter of BNBs, we use the Resonant Mass Measurement (RMM) technique (Archimedes, Malvern Instruments). An accurate BNB characterization method is essential to ensure the particles under investigation are indeed NBs, not solid nanoparticles. Among the most common characterization techniques, RMM is the only method which can differentiate bubbles from non-gaseous particles, such as contaminants and surfactants.^{6,22–24}

We use 1 mL of BNBs solution to quantitatively characterize BNBs with RMM. We utilize a 10 nm resolution RMM nanosensor in all experiments with a measurement limitation of 100 nm to 2.0 μm . Before obtaining the data, we calibrate the sensor employing NIST traceable 500 nm polystyrene beads (MilliporeSigma Canada Ltd 2149, Oakville ON, Canada) dispersed in water solution. Before and after each experiment, we run three cycles of nanosensor cleaning through filtered Milli-Q water to ensure the microchannels inside the sensor are not blocked and are free of any unwanted particles. We dilute each sample filtered PBS with a factor of 10 to keep the coincidence percentage less than 5% and decrease the number of “coincidence bumps” in the obtained result. We let the instrument automatically adjust the detection limit in each experiment. We use the density of 0.008 mg cc^{-1} for positively buoyant particles. We calculate this number through the particle density measurement guideline recommended by Archimedes user manual.

3. Results and discussion

In a recently published paper, we found a linear relationship between the initial diameter, D_i , of bubbles and their post-shrinkage final diameter, D_f .¹⁷ We also found that when the continuous lipid phase concentration is decreased, bubbles experience increased levels of shrinkage.¹⁷ We suggest that this happens because when bubbles form in a solution of higher lipid concentration, more lipid molecules are initially adsorbed onto the gas-liquid interface in comparison with bubbles produced using a lower lipid concentration solution. Consequently, in solutions with higher lipid concentration, lipids at bubble interfaces jam earlier, and result in lower levels of shrinkage.¹⁷ For example, when we utilize a lipid concentration of 1.10 mg mL^{-1} , and make MBs with $D_i \approx 25.0 \mu\text{m}$, the post-shrinkage final mean diameter $D_f \approx 3.7 \mu\text{m}$. This means D_f is approximately 6.7 times smaller than D_i . Therefore, in this paper, we expand the previous research¹⁷ by hypothesizing that

decreasing the initial diameter D_i of MBs and the concentration of the lipid solution would further increase the degree of shrinkage and lead to the formation of BNBs. We also hypothesize that the size of the resulting BNBs can be tuned by adjusting the initial diameter D_i of MBs, and the lipid solution concentration.

3.1 Unexpected MBs shrinkage to BNBs

In a set of representative experiments, we generate MBs with a mean initial diameter $D_i < 20.0\ \mu\text{m}$ in a solution with a lipid concentration of $0.50\ \text{mg mL}^{-1}$. Fig. 2a–d show bright-field images of MBs at the orifice with $D_i = 7.0, 12.0, 15.0$, and $20.0\ \mu\text{m}$, respectively. Fig. 2e–h show corresponding TEM images of the resulting bubbles after they have been collected in a plastic vial. We observe that bubbles with $D_i = 15.0$ and $20.0\ \mu\text{m}$ have final diameters, $D_f \approx 1.4$ and $2.0\ \mu\text{m}$, respectively, confirming that the degree of shrinkage, $D_i/D_f \approx 10.0$. This observation is consistent with the results published in our recent paper, where there is a linear relationship between MB D_i and D_f .¹⁷

Surprisingly, TEM images from MBs with $D_i = 7.0$ and $12.0\ \mu\text{m}$ show that the final bubble diameter $D_f \approx 189$ and $195\ \text{nm}$, respectively, with $D_i/D_f \gg 10.0$. This finding means that 7.0 and $12.0\ \mu\text{m}$ diameter MBs deviate from the linear trend between bubble D_i and D_f that is found for MBs that have just slightly larger D_i . Another unexpected finding is that both $D_i = 7.0$ and $12.0\ \mu\text{m}$ bubbles shrink to approximately the same diameter BNBs.

3.2 Confirmation of BNBs using the Tyndall effect

We utilize the Tyndall effect to confirm the presence of BNBs in samples we collect from our microfluidic experiments. Namely, we make MBs with an initial mean diameter $D_i = 7.0\ \mu\text{m}$, a lipid solution concentration of $0.50\ \text{mg mL}^{-1}$, and a blended gas of $0.17\ \text{wt\% C}_3\text{F}_8$ in N_2 . After 1 day, we investigate the presence of BNBs in the collected sample by running the Tyndall test. For this test, we irradiate a laser beam, from right to left, towards

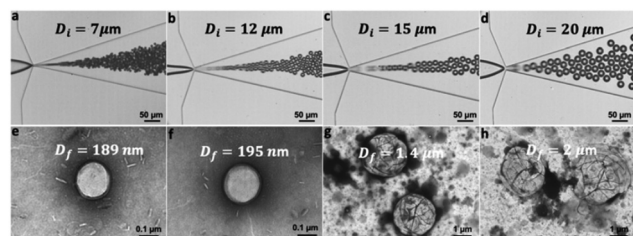


Fig. 2 (a–d) Bright field, and (e–h) TEM images of the bubbles before and after shrinkage, respectively. The lipid concentration in all of these experiments is $0.5\ \text{mg mL}^{-1}$, and the gas is composed of $0.17\ \text{wt\% C}_3\text{F}_8$ in pure N_2 . In (a–d) the MB initial diameter $D_i = 7.0, 12.0, 15.0$ and $20.0\ \mu\text{m}$, respectively. (e–h) The corresponding final mean diameters $D_f = 189\ \text{nm}, 195\ \text{nm}, 1.4\ \mu\text{m}$, and $2.0\ \mu\text{m}$, measured using TEM. When we reduce D_i from 20.0 to $7.0\ \mu\text{m}$, the final bubble diameter D_f goes from $2\ \mu\text{m}$ to $189\ \text{nm}$. There appears to be a threshold $D_i = 13.0$ – $14.0\ \mu\text{m}$, that separates bubbles that shrink to MBs and those that shrink to BNBs. Bright field and TEM images are taken 1 hour after bubbles are generated. We vary the gas phase pressure from 14.0 to $18.3\ \text{psi}$ and keep the aqueous flow rate fixed at $45\ \mu\text{L min}^{-1}$. The width of the orifice in (a–d) are $4, 7, 10$, and $10\ \mu\text{m}$.

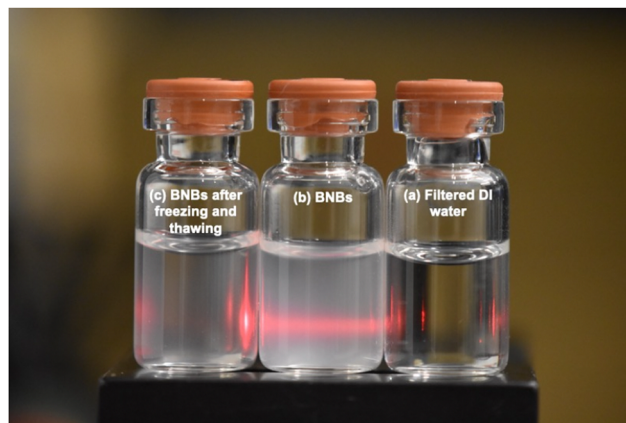


Fig. 3 Tyndall effect test. This image is taken 1 day after making BNBs. We shoot a red laser beam from the right side of the image to the left. (a) The vial containing DI water does not scatter the light as there are no particles inside the sample. (b) The sample containing the BNB solution scatters the laser light due to the presence of BNBs suspended in the vial. (c) This image shows a BNB sample after freezing for $24\ \text{h}$ and thawing. The degree of scattered light in the thawed BNB solution is considerably reduced compared to the BNB vial without freezing–thawing. This is likely due to the rupture of BNBs during the freezing–thawing process.¹⁴ Both BNB suspensions in (b) and (c) are made by first generating MBs with $D_i = 7.0\ \mu\text{m}$, with $0.17\ \text{wt\% C}_3\text{F}_8$ gas, and a lipid solution concentration of $0.50\ \text{mg mL}^{-1}$.

three vials (see Fig. 3). We observe that the control vial of filtered deionized (DI) water does not scatter the laser light (Fig. 3a). However, the vial containing BNBs scatters the laser light (Fig. 3b). We hypothesize that the observed Tyndall phenomenon is due to the presence of BNBs suspended in the solution that scatter the laser beam.

It is also important to confirm that the nanoparticles in the solution are actual gas-filled BNBs. To do this, we utilize the technique suggested by Alheshibri and Craig on distinguishing gas-filled BNBs from solid nanoparticles.²⁵ In this method, we keep a vial of BNBs in a freezer at the temperature of $-20\ ^\circ\text{C}$ for $24\ \text{h}$. After $24\ \text{h}$, we leave the vial at room temperature to let the liquid inside the vial thaw. As Nirmalkar *et al.* have shown, most BNBs burst as a result of freezing and thawing; however, contaminants and surfactants remain.¹⁴ After thawing the vial, we repeat the Tyndall test. As Fig. 3c shows, the laser beam passes through the vial, suggesting that the BNBs have also burst in the sample. In the next step, we utilize the RMM characterization method to further analyze the BNBs.

3.3 MB critical initial diameter that enables shrinkage to BNBs

We further investigate the unexpected phenomenon of the sudden and dramatic shrinkage of MBs to BNBs that we observe in some experiments. Here, we generate MBs with different initial diameters D_i . These experiments aim to see how the final diameter D_f changes with different values of initial diameters D_i . We also attempt to understand whether there is a relationship between D_i and D_f when bubbles are generated below the threshold diameter. Moreover, we investigate the effect of altering the lipid solution on the observed phenomenon.

To better examine the behaviour of bubbles when shrinking, we make MBs with a mean initial diameter $D_i \approx 4.0\text{--}24.0\ \mu\text{m}$, varying the lipid concentration from $0.30\text{--}0.75\ \text{mg mL}^{-1}$. We use RMM to measure the resulting bubbles when the final diameter $D_f < 1.0\ \mu\text{m}$. When $D_f > 1.0\ \mu\text{m}$, bubble diameter is measured *via* brightfield microscopy. We also define a critical initial diameter, $D_{i,c}$, that is the average of the two closest values of D_i , where the larger value of D_i results in the expected amount of shrinkage and the smaller value of D_i results in the unexpected, drastic shrinking of BNBs.

Fig. 4 shows the relationship between the initial mean diameter D_i of the MBs and their final mean diameter D_f after shrinkage. We use a mixed gas composed of $0.17\ \text{wt\% C}_3\text{F}_8$ and pure N_2 in all experiments. We observe that $D_f \propto D_i$ at larger values of D_i , as expected, in all experiments.¹⁷ However, at lower values of D_i , all bubbles converge to a final diameter $D_f \approx 150\text{--}250\ \text{nm}$, with a critical initial diameter $D_{i,c}$ that appears to depend on the solution lipid concentration. We define the “critical region” as the region in the plot that delineates between the two regimes.

In Fig. 4, black, red, and blue symbols represent data points from experiments with lipid concentrations of 0.75 , 0.50 , and $0.30\ \text{mg mL}^{-1}$, respectively. We observe the same trend in the data from all three lipid concentrations. However, as the inset shows, we also see a shift in the $D_{i,c}$ with lipid concentration. Namely, $D_{i,c}$ declines from approximately 15.0 to $12.0\ \mu\text{m}$ when lipid concentration is increased from 0.30 to $0.75\ \text{mg mL}^{-1}$.

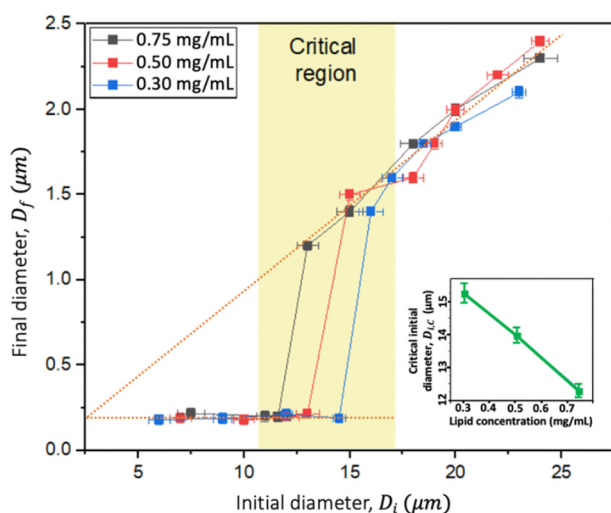


Fig. 4 Plot of the post-shrinkage final mean diameter D_f versus the initial mean diameter D_i of the MBs. Different colours represent data associated with different lipid concentrations, as shown in the legend. We use three lipid concentrations in our experiments: $0.30\ \text{mg mL}^{-1}$ (blue squares), $0.50\ \text{mg mL}^{-1}$ (red squares), and $0.75\ \text{mg mL}^{-1}$ (black squares). We use a mixed gas of $0.17\ \text{wt\% C}_3\text{F}_8$ and pure N_2 in all experiments. When the $D_i > D_{i,c}$, the final diameter, $D_f \propto D_i$, and the shrinkage degree $D_i/D_f \approx 10.6$, 10.0 , and 9.4 , when the lipid concentration is $0.30\ \text{mg mL}^{-1}$, $0.50\ \text{mg mL}^{-1}$, and $0.75\ \text{mg mL}^{-1}$, respectively. However, when $D_i < D_{i,c}$, the final and initial diameters, D_f and D_i , become non-monotonically related. In this regime, the final diameter $D_f = 150\text{--}250\ \text{nm}$ for all BNBs, regardless of their initial diameter D_i . The inset shows the critical initial diameter $D_{i,c}$ versus the solution lipid concentration. As we increase the lipid concentration from 0.30 to $0.75\ \text{mg mL}^{-1}$, $D_{i,c}$ declines from approximately 15.0 to $12.0\ \mu\text{m}$.

We suppose the reason that a higher lipid concentration results in a lower $D_{i,c}$ is due to a phenomenon called “lipid jamming”.^{17,26–28} Here, as bubbles form at the orifice, lipid molecules adsorb on the gas–liquid interface *via* self-assembly. As gas inside the MBs dissolve into the surrounding aqueous phase, the MBs shrink. The shrinkage process stops when the lipid molecules on the gas–liquid interface mechanically compress together, forming a packed interface with a surface tension that approaches zero.^{17,29} MBs that are generated in a high lipid concentration solution begin with a higher density of lipid molecules absorbed onto the gas–liquid interface. Consequently, the interfacial lipids jam earlier, arresting MB shrinkage, and reducing the degree of bubble shrinkage.¹⁷

In our experiments with lipid concentration between 0.30 and $0.75\ \text{mg mL}^{-1}$, bubbles that are formed with $D_i > D_{i,c}$ have a shrinkage degree $D_i/D_f \approx 10.0$. This is consistent with findings in our recent paper, in which we showed the impact of lipid concentration on MB shrinkage.¹⁷ We found that the shrinkage degree $D_i/D_f \approx 2.0\text{--}6.7$ when the lipid concentration is from 10.0 to $1.1\ \text{mg mL}^{-1}$.¹⁷

Table 1 shows the final diameter D_f of bubbles generated with lipid concentrations of 0.30 , 0.50 , and $0.75\ \text{mg mL}^{-1}$, when $D_i < D_{i,c}$. Notably, in this regime we do not observe any correlation between D_i and D_f . Moreover, it appears that the final diameter is consistently $D_f = 150\text{--}250\ \text{nm}$.

Fig. 5a–d show distribution plots of the final diameter D_f of BNBs, obtained using RMM. The BNBs are produced using a lipid concentration of $0.50\ \text{mg mL}^{-1}$ and $0.17\ \text{wt\% C}_3\text{F}_8$ blended gas. Here, the BNBs in a–d have initial diameters $D_i = 7.0$, 10.0 , 12.0 , and $13.0\ \mu\text{m}$. Fig. 5e shows a TEM image of BNBs, with the red arrows showing BNBs with $D_f \approx 180\ \text{nm}$. $D_f = 150\text{--}250\ \text{nm}$ for all BNBs observed, and there is good agreement between RMM and TEM imaging results. We also monitor the stability of BNBs with $D_f \approx 189\ \text{nm}$, starting at 1 hour post-generation for a period of 48 hours. Over this time, the average diameter of BNBs increases from $189 \pm 18\ \text{nm}$ to $220 \pm 14\ \text{nm}$.

We compare the shrinkage process between two single bubbles, both made with a lipid concentration of $0.50\ \text{mg mL}^{-1}$. We generate the two MBs with initial diameters $D_i = 12.0$ and

Table 1 Final vs. initial diameter of BNBs formed with the lipid concentrations of $0.30\text{--}0.75\ \text{mg mL}^{-1}$ when $D_i < D_{i,c}$

Lipid concentration (mg mL^{-1})	Initial diameter, D_i (μm)	Final diameter, D_f (nm)
0.30	14.5 ± 0.3	189 ± 30
0.30	12.0 ± 0.2	210 ± 31
0.30	9.0 ± 0.4	185 ± 33
0.30	6.0 ± 0.5	179 ± 25
0.50	13.0 ± 0.5	215 ± 21
0.50	12.0 ± 0.6	195 ± 24
0.50	10.0 ± 0.4	181 ± 28
0.50	7.0 ± 0.5	189 ± 29
0.75	11.6 ± 0.6	195 ± 30
0.75	11.0 ± 0.4	200 ± 32
0.75	7.5 ± 0.6	215 ± 30

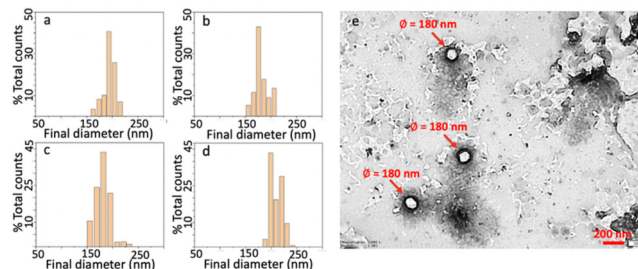


Fig. 5 RMM and TEM characterization of BNBs. (a–d) Representative RMM measurements of NB diameter generated with 0.5 mg mL^{-1} lipid concentration and $0.17 \text{ wt\% C}_3\text{F}_8$ gas. The initial diameter D_i of bubbles in (a–d) are 7.0 , 10.0 , 12.0 , and $13.0 \mu\text{m}$, respectively. The results show that the all BNBs in our experiments have $D_f = 150\text{--}250 \text{ nm}$. (e) TEM image of NBs. Red arrows show NBs with $D_f \approx 180 \text{ nm}$.

$15.0 \mu\text{m}$ (Fig. 6). We use the same blended gas, $0.17 \text{ wt\% C}_3\text{F}_8$ in pure N_2 , in both experiments. After generating the MBs, we collect them in a plastic vial that is open to the atmosphere. We transfer a drop of the bubble suspension from the microfluidic chip outlet to a flow cell (IBIDI μ -slide). The inlet and outlet of the flow cell is open to the atmosphere. We dilute the bubble population inside the flow cell with water (Milli-Q, Millipore) to enable visualization with bright-field microscope imaging.

Fig. 6a–e show the shrinkage of an MB with $D_i = 12.0 \mu\text{m}$ at different times. The total time for the bubble to shrink to the nanoscale is approximately 33 s . We describe the total shrinkage time as the duration from bubbles entering the flow cell until they complete shrinkage and reach a stable diameter. In our experiments, $t = 0$ is when bubbles first enter the flow cell. Here, we measure the size of the collected bubble sample *via* the RMM method and find the final diameter $D_f = 195 \text{ nm}$. Fig. 6k shows that the instantaneous bubble diameter D , at early times, decreases approximately linearly with time t during the shrinkage process. The bubble shrinkage accelerates towards the end of the shrinkage process, as also predicted using the classical Epstein and Plesset theory.³⁰ The difference observed here is that the bubble does not completely vanish but stabilizes to a BNB. Fig. 6f–j shows the shrinkage of an MB with $D_i = 15.0 \mu\text{m}$. In this case, the post-shrinkage final diameter $D_f = 1.6 \mu\text{m}$. The bubble reaches its final size after around 15 s .

3.4 Shrinkage of MBs to BNBs with similar final diameters

Interestingly, MBs bigger and smaller than the critical initial diameter, $D_{i,c}$, follow two different shrinkage patterns. The former experiences a linear shrinkage trend, with D_f dependant on the value of D_i , while the latter shrinks into a diameter that is always between $150\text{--}250 \text{ nm}$, independent of D_i .

We attempt to address the question of why bubbles that have $D_i < D_{i,c}$ appear to shrink to a characteristic diameter $D_f = 150\text{--}250 \text{ nm}$. We approach this investigation with the assumption that the gas–liquid interface experiences “jamming” due to the close packing of lipids. Namely, when gas dissolution occurs, bubbles start to shrink and the lipids form a monolayer with an increasing packing density until the interfacial tension approaches zero.^{29,31} The decrease in bubble volume also

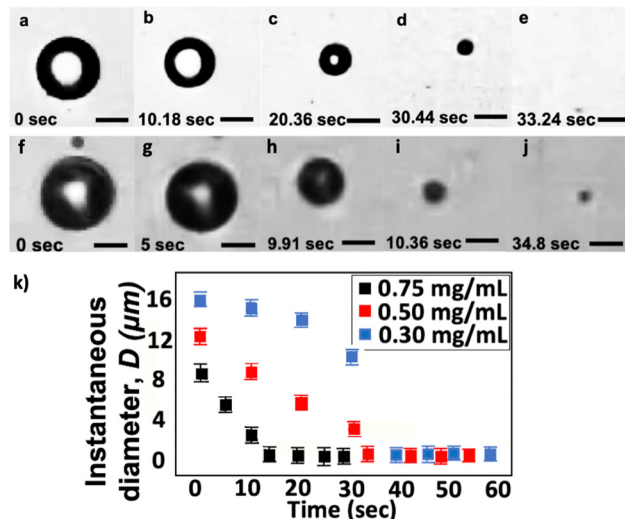


Fig. 6 (a–e) Brightfield images of an MB shrinking into a BNB. (a) The bubble has an initial diameter, $D_i = 12.0 \mu\text{m}$ at $t = 0 \text{ s}$. We define $t = 0 \text{ s}$ as the moment when bubbles enter the flow cell and begin recording their shrinkage time. (b–d) High-speed brightfield images of the MB shrinking at $t = 10$, 20 , and 30 s , respectively. (e) At $t = 33 \text{ s}$, the bubble disappears from the screen of the microscope. RMM size characterization after the experiment shows the corresponding final diameter $D_f = 195 \text{ nm}$, for the initial diameter $D_i = 12.0 \mu\text{m}$. (f–j) Time-lapse images of an MB with $D_i = 15.0 \mu\text{m}$ shrinking to a smaller MB with $D_f = 1.6 \mu\text{m}$. The shrinkage stops after 15 s , and the bubble maintains its final diameter $D_f = 1.6 \mu\text{m}$. (k) This graph shows the instantaneous bubble diameter D , versus time t , with lipid concentrations of 0.75 mg mL^{-1} , 0.50 mg mL^{-1} , and 0.30 mg mL^{-1} . Here, instantaneous microscale bubble diameters D and microscale final diameters D_f are measured utilizing high-speed brightfield microscopy and ImageJ software. Nanoscale final diameters D_f are measured *via* RMM. Bubbles in (a and f) are formed using a lipid solution concentration of 0.50 mg mL^{-1} and a gas of $0.17 \text{ wt\% C}_3\text{F}_8$. Scale bars represent $5.0 \mu\text{m}$.

decreases the average area per insoluble surfactant molecule at the interface, which decreases surface tension until the packing of lipids prevents the nanobubble from further dissolution.^{29,31} This is also described in molecular dynamics simulations, by Zhang *et al.*, and other researchers, showing that when lipids at the gas–liquid interface pack together, the interfacial tension can approach zero.^{29,32–34}

When the lipids are packed together on the gas–liquid interface, the interfacial free energy of BNBs is dominated by the free energy of the surface curvature.^{29,31} The Helfrich model can be used to describe the free energy of curvature.^{35–37} According to this model, when surface tension is zero, bending free energy is $F_s = \int \kappa/2 (C_1 - C_2)^2 dA$. In this equation, κ represents the bending rigidity or bending modulus, C_1 and C_2 are bubble surface curvature and the spontaneous lipid monolayer curvature, respectively. Different studies show that when short-chain molecules (*e.g.* alcohols) are present in the continuous lipid solution, the bending rigidity κ is as small as $k_B T$, where k_B and T are Boltzmann constant and temperature, respectively.^{29,31,38–41} On a bubble's shell, short chains sit between the long chains and space them apart so that their ends swing freely.⁴² Moreover, short-chains may also move across in the gas–liquid interface and disrupt the packing of lipid hydrocarbon chains on the bubble's

shell. All these factors may contribute to a considerable reduction in the bending stiffness κ of the bubble's shell.³⁹

Closely examining the lipid solution used in our study reveals that glycerol comprises a large amount of the lipid solution compared to the lipids present. Glycerol, also referred to as trihydroxy alcohol, is classified as a short-chain alcohol.^{43,44} Our lipid solution has a high glycerol-to-lipid mass ratio of 120.

Given the impacts of short chains on the bending rigidity κ of the shell, numerous studies have shown that when κ is comparable to $k_B T$, thermal fluctuations determine the radius of the shell's curvature.^{29,31,38–41} In our study, the radius of the shell's curvature corresponds to the radius of a bubble. Assuming that the temperature fluctuations govern the bending energy of the shell, we can demonstrate the relationship between the two using the persistence length of the bubble's shell, $\xi_k = \xi_0 \exp(4\pi\kappa/2k_B T)$.³¹ In this equation, $\xi_0 = 10 \text{ \AA}$, which is approximately equal with the length of the lipid molecules.³¹ With $\kappa = k_B T$ and $\xi_0 = 10 \text{ \AA}$, the model of Chen *et al.* would predict a final BNB diameter of approximately 132 nm,³¹ which has the same order-of-magnitude as the BNB diameters we observe in our experiments. We, therefore, find a reasonable agreement between the experimental results of our study and the theoretical predictions of Chen *et al.*³¹ Some differences between the predicted and experimental BNB diameters may be attributed to the type of lipids used to generate BNBs. Specifically, our lipid solution may not accurately fit the assumption that bending rigidity is entirely dominated by $k_B T$ (*i.e.* the actual bending rigidity may be slightly higher, which would increase the diameter of the final BNB).

4. Conclusions

In this study, we identify a critical initial diameter, $D_{i,c}$ of MBs made with microfluidic flow-focusing, where bubbles larger and smaller than $D_{i,c}$ exhibit distinct shrinking behaviours. When the initial diameter $D_i < D_{i,c}$, an unexpected phenomenon occurs in which all the MBs shrink to the same range of final diameters. In contrast, when $D_i > D_{i,c}$, MBs shrink by the amount predicted by previous literature.¹⁷ Specifically, when MB $D_i < D_{i,c}$, MBs appear to always shrink to a diameter $D_f = 150\text{--}250 \text{ nm}$, using the lipid concentrations in our experiments. We observe that the critical initial diameter, $D_{i,c}$ ranges between $13.0\text{--}15.0 \text{ \mu m}$, for lipid concentrations of $0.30\text{--}0.75 \text{ mg mL}^{-1}$, respectively. To the best of our knowledge, this is the first experimental report of a critical MB diameter that delineates between MBs shrinking by an anticipated amount, and MBs shrinking dramatically to become stable BNBs.

Conflicts of interest

The authors declare no conflict of interest.

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References

- 1 A. A. Paknahad, L. Kerr, D. A. Wong, M. C. Kolios and S. S. H. Tsai, Biomedical nanobubbles and opportunities for microfluidics, *RSC Adv.*, 2021, **11**(52), 32750–32774.
- 2 C. Li and H. Zhang, A review of bulk nanobubbles and their roles in flotation of fine particles, *Powder Technol.*, 2022, **395**, 618–633.
- 3 E. P. Favvas, G. Z. Kyzas, E. K. Efthimiadou and A. C. Mitropoulos, Bulk nanobubbles, generation methods and potential applications, *Curr. Opin. Colloid Interface Sci.*, 2021, **54**, 101455.
- 4 M. Zhang, J. Yang, Z. Kang, X. Wu, L. Tang and Z. Qiang, *et al.*, Removal of micron-scale microplastic particles from different waters with efficient tool of surface-functionalized microbubbles, *J. Hazard. Mater.*, 2021, **404**, 124095.
- 5 R. H. Perera, H. Wu, P. Peiris, C. Hernandez, A. Burke and H. Zhang, *et al.*, Improving performance of nanoscale ultrasound contrast agents using N,N-diethylacrylamide stabilization. Nanomedicine: Nanotechnology, *Biol. Med.*, 2016, 1–9.
- 6 A. A. Exner and M. C. Kolios, Bursting microbubbles: How nanobubble contrast agents can enable the future of medical ultrasound molecular imaging and image-guided therapy, *Curr. Opin. Colloid Interface Sci.*, 2021, **54**, 101463.
- 7 J. L. Parker, P. M. Claesson and P. Attard, Bubbles, cavities, and the long-ranged attraction between hydrophobic surfaces, *J. Phys. Chem. C*, 1994, **98**(34), 8468–8480.
- 8 G. S. Manning, On the thermodynamic stability of bubbles, immiscible droplets, and cavities, *Phys. Chem. Chem. Phys.*, 2020, **22**(31), 17523–17531.
- 9 L. Zhou, S. Wang, L. Zhang and J. Hu, Generation and stability of bulk nanobubbles: A review and perspective, *Curr. Opin. Colloid Interface Sci.*, 2021, **53**, 101439.
- 10 P. A. Satpute and J. C. Earthman, Hydroxyl ion stabilization of bulk nanobubbles resulting from microbubble shrinkage, *J. Colloid Interface Sci.*, 2021, **584**, 449–455.
- 11 D. H. Kim, M. J. Costello, P. B. Duncan and D. Needham, Mechanical properties and microstructure of polycrystalline phospholipid monolayer shells: Novel solid microparticles, *Langmuir*, 2003, **19**(20), 8455–8466.
- 12 S. R. Sirsi and M. A. Borden, Microbubble compositions, properties and biomedical applications, *Bubble Sci., Eng., Technol.*, 2009, **1**(1–2), 3–17.

- 13 N. Nirmalkar, A. W. Pacek and M. Barigou, Interpreting the interfacial and colloidal stability of bulk nanobubbles, *Soft Matter*, 2018, **14**(47), 9643–9656.
- 14 N. Nirmalkar, A. W. Pacek and M. Barigou, On the existence and stability of bulk nanobubbles, *Langmuir*, 2018, **34**(37), 10964–10973.
- 15 S. A. Peyman, J. R. McLaughlan, R. H. Abou-Saleh, G. Marston, B. R. G. Johnson and S. Freear, *et al.*, On-chip preparation of nanoscale contrast agents towards high-resolution ultrasound imaging, *Lab Chip*, 2016, **16**(4), 679–687.
- 16 R. H. Abou-Saleh, F. J. Armistead, D. V. B. Batchelor, B. R. G. Johnson, S. A. Peyman and S. D. Evans, Horizon: Microfluidic platform for the production of therapeutic microbubbles and nanobubbles., *Rev. Sci. Instrum.*, 2021, **92**(7), 074105.
- 17 I. O. Zalloum, A. A. Paknahad, M. C. Kolios, R. Karshafian and S. S. H. Tsai, Controlled Shrinkage of Microfluidically Generated Microbubbles by Tuning Lipid Concentration, *Langmuir*, 2022, **38**(43), 13021–13029.
- 18 A. De Leon, R. Perera, C. Hernandez, M. Cooley, O. Jung and S. Jeganathan, *et al.*, Contrast enhanced ultrasound imaging by nature-inspired ultrastable echogenic nanobubbles, *Nanoscale*, 2019, **11**(33), 15647–15658.
- 19 Q. Wang, H. Zhao, N. Qi, Y. Qin, X. Zhang and Y. Li, Generation and stability of size-adjustable bulk nanobubbles based on periodic pressure change, *Sci. Rep.*, 2019, **9**(1), 1–9.
- 20 J. Jin, R. Wang, J. Tang, L. Yang, Z. Feng and C. Xu, *et al.*, Dynamic tracking of bulk nanobubbles from microbubbles shrinkage to collapse, *Colloids Surf., A*, 2020, **589**, 124430.
- 21 C. Counil, E. Abenojar, R. Perera and A. A. Exner, Extrusion: A New Method for Rapid Formulation of High-Yield, Mono-disperse Nanobubbles, *Small*, 2022, **18**(24), 1–11.
- 22 L. Zhou, S. Wang, L. Zhang and J. Hu, Generation and stability of bulk nanobubbles: A review and perspective, *Curr. Opin. Colloid Interface Sci.*, 2021, **53**, 101439.
- 23 E. C. Abenojar, I. Bederman, A. C. de Leon, J. Zhu, J. Hadley, M. C. Kolios and A. A. Exner, Theoretical and experimental gas volume quantification of micro-and nanobubble ultrasound contrast agents, *Pharmaceutics*, 2020, **12**(3), 208.
- 24 A. A. Paknahad, L. Kerr, D. A. Wong, M. C. Kolios and S. S. H. Tsai, Biomedical nanobubbles and opportunities for microfluidics, *RSC Adv.*, 2021, **11**, 32750–32774.
- 25 M. Alheshibri and V. S. J. Craig, Differentiating between Nanoparticles and Nanobubbles by Evaluation of the Compressibility and Density of Nanoparticles, *J. Phys. Chem. C*, 2018, **122**(38), 21998–22007.
- 26 R. Shih and A. P. Lee, Post-Formation Shrinkage and Stabilization of Microfluidic Bubbles in Lipid Solution, *Langmuir*, 2016, **32**(8), 1939–1946.
- 27 M. A. Borden and M. L. Longo, Dissolution behavior of lipid monolayer-coated, air-filled microbubbles: Effect of lipid hydrophobic chain length, *Langmuir*, 2002, **18**(24), 9225–9233.
- 28 M. Abkarian, A. B. Subramaniam, S. H. Kim, R. J. Larsen, S. M. Yang and H. A. Stone, Dissolution arrest and stability of particle-covered bubbles, *Phys. Rev. Lett.*, 2007, **99**(18), 1–4.
- 29 H. Zhang, S. Chen, Z. Guo and X. Zhang, The fate of bulk nanobubbles under gas dissolution, *Phys. Chem. Chem. Phys.*, 2022, **24**(16), 9685–9694.
- 30 P. S. Epstein and M. S. Plesset, On the stability of gas bubbles in liquid-gas solutions, *J. Chem. Phys.*, 1951, **19**(2), 256.
- 31 C. Chen, H. Zhang and X. Zhang, Stable bulk nanobubbles can be regarded as gaseous analogues of microemulsions, *ChemRxiv*, 2022, preprint, DOI: [10.26434/chemrxiv-2022-hq85x](https://doi.org/10.26434/chemrxiv-2022-hq85x).
- 32 R. Cantor, Nonionic Diblock Copolymers as Surfactants between Immiscible Solvents, *Macromolecules*, 1981, **14**(5), 1186–1193.
- 33 J. L. Cayias, R. S. Schechter and W. H. Wade, The utilization of petroleum sulfonates for producing low interfacial tensions between hydrocarbons and water, *J. Colloid Interface Sci.*, 1977, **59**(1), 31–38.
- 34 H. Eicke and J. Rehak, On the Formation of Water/Oil-Microemulsions, *Helv. Chim. Acta*, 1976, **59**(306), 2883–2891.
- 35 W. Helfrich, Effect of Thermal Undulations on the Rigidity of Fluid Membranes and Interfaces, *J. Phys.*, 1985, **46**(7), 1263–1268.
- 36 W. Helfrich, Elastic Properties of Lipid Bilayers: Theory and Possible Experiments. Zeitschrift fur Naturforschung-Section C, *J. Biosci.*, 1973, **28**(11–12), 693–703.
- 37 J. Oberdisse and T. Hellweg, Structure, interfacial film properties, and thermal fluctuations of microemulsions as seen by scattering experiments, *Adv. Colloid Interface Sci.*, 2017, **247**, 354–362.
- 38 I. Szleifer, D. Kramer, A. Ben-Shaul, D. Roux and W. M. Gelbart, Curvature elasticity of pure and mixed surfactant films, *Phys. Rev. Lett.*, 1988, **60**(19), 1966–1969.
- 39 B. P. Binks, J. Meunier, O. Abillon and D. Langevin, Measurement of Film Rigidity and Interfacial Tensions in Several Ionic Surfactant-Oil-Water Microemulsion Systems, *Langmuir*, 1989, **5**(2), 415–421.
- 40 J. M. Di Meglio, M. Dvolaitzky and C. Taupin, Determination of the rigidity constant of the amphiphilic film in “birefringent microemulsions”: The role of the cosurfactant, *J. Phys. Chem.*, 1985, **89**(5), 871–874.
- 41 J. Meunier, Measurement of the rigidity coefficient of a surfactant layer and structure of the oil or water microemulsion interface, *J. physique, Lett.*, 1985, **46**(21), 1005–1014.
- 42 C. R. Safinya, D. Roux, G. S. Smith, S. K. Sinha, P. Dimon and N. A. Clark, *et al.*, Steric interactions in a model multi-membrane system: A synchrotron X-ray study, *Phys. Rev. Lett.*, 1986, **57**(21), 2718–2721.
- 43 H. Zhang, F. Feng, J. Li, X. Zhan, H. Wei and H. Li, *et al.*, Formulation of food-grade microemulsions with glycerol monolaurate: Effects of short-chain alcohols, polyols, salts and nonionic surfactants, *Eur. Food Res. Technol.*, 2008, **226**(3), 613–619.
- 44 A. Yaghmur, A. Aserin and N. Garti, Phase behavior of microemulsions based on food-grade nonionic surfactants: Effect of polyols and short-chain alcohols, *Colloids Surf., A*, 2002, **209**(1), 71–81.