



Cite this: *Sens. Diagn.*, 2023, 2, 1437

Integrated microfluidic devices for point-of-care detection of bio-analytes and disease

Prateechee Padma Behera,^{†a} Natish Kumar,^{†b} Monika Kumari,^b Sumit Kumar,^{ib c} Pranab Kumar Mondal^a and Ravi Kumar Arun ^{ib *b}

Micro- and nanofluidics have enabled the possibility of miniaturized analytical systems, which are commonly known as lab-on-a-chip (LOC) devices. One of the clear benefits of these microfluidic devices is the 'sample-in-answer-out' system, in which the input is a whole/minimally processed biological sample, and the output is a qualitative or quantitative diagnosis of biomarkers of disease or bio-analyte of interest. All the clinical protocols could be integrated into a well-designed LOC device, such as sample pre-treatment, sample/reagent manipulation, separation, reaction, and detection. Microfluidics and nanofluidics interactions revolutionize lab-on-a-chip diagnostics, offering cost-effective alternatives to clinical setups. This article looks at bioanalysis that uses fully integrated microfluidic devices and optical and electrochemical methods like amperometry, potentiometry, cyclic voltammetry, and electrical impedance spectroscopy. These technologies diagnose autoimmune diseases (rheumatoid arthritis, multiple sclerosis, type 1 diabetes, and Addison's disease), acute nutritional deficiencies (iron, vitamin A, D, B₁₂, C, and K), and pharmacological response biomarkers. The purpose of this review is to examine the utility of microfluidics in the detection of biomarkers. Our analysis highlights the potential of microfluidic technologies to transform diagnostics and personalized healthcare by addressing existing issues and proposing new solutions. These systems address sensitive, rapid, and sample volume limits with their sophisticated design and accuracy in managing micro fluid volumes. The present review thoroughly summarizes the qualitative and quantitative characteristics of the diagnostic assay, enabling the biomarker to be found at the appropriate sensitivity levels with a modest analysis time and cost. The study presents an interdisciplinary perspective on modern diagnostics, which could revolutionize healthcare by enabling precise disease identification and management.

Received 3rd July 2023,
Accepted 10th September 2023

DOI: 10.1039/d3sd00170a

rsc.li/sensors

1. Introduction

The integration of microfluidics and diagnostics in recent years has led to an era of innovation in healthcare, transforming bioanalysis and disease detection.¹ Microfluidics has been widely recognized as the main driving force for the paradigm shift in four key domains: molecular analysis, biodefense, molecular biology, and microelectronics. The incorporation of microfluidic elements onto a singular chip has resulted in the emergence of lab-on-a-chip (LOC), micro total analysis systems (μ TAS), and point-of-care (POC) diagnostic devices.² Integrated microfluidic devices are

powerful tools for rapid, sensitive, and portable point-of-care (POC) detection of bio-analytes and disease biomarkers.^{3,4}

The chemical and biological processes occur on a much larger scale in conventional laboratories. To scale down the equipment to the length of the reaction was a wise thing to do. On such a small scale, flow physics works differently. This needs to scale down the processes, thus paving the way for microfluidics and nanofluidics.^{5,6} Recent technology has achieved analytical/antigen detection, drug delivery, biosynthesis, food safety, *etc.* The property of paper to wick fluids has been exploited to detect various biomarkers of diseases and nutrient deficiencies.^{7,8} The introduction of POC testing into the healthcare system has helped in the early detection of diseases, regular monitoring, and blocking many redundant practices. POC devices help analyze minimal volumes of bodily fluids, like whole blood, saliva, and urine.⁹ The smaller size aids in proper disposal of the device after use in cases of contagious diseases. With the help of electrochemical methods, the sensor's sensitivity can be improved to the range of pg mL⁻¹.^{10–12} Further addition of

^a Department of Mechanical Engineering, Indian Institute of Technology, Kamrup, Guwahati, Assam, 783019, India

^b Department of Chemical Engineering, Indian Institute of Technology, Jagti, NH-44, Jammu-181221, India. E-mail: ravi.arun@iitjammu.ac.in

^c Faculty of Mechanical Engineering, Technion-Israel Institute of Technology, Haifa-3200003, Israel

[†] These authors contributed equally.



nanotubes, nanowires, nanofluids, and nanorods enhances the quantification of the signal from POC devices. Most POC devices have the potential to meet the clinical requirements at the global level, but the challenge of standardization remains a problem.^{13–15}

Analytes like proteins, DNA/RNA, antibodies, antigens, or other metabolic compounds in the body could be examined as biomarkers to provide information about diseases or the physiological status of the patient. Monitoring, diagnosis, and risk assessments of diseases are made possible by recognizing and measuring these biomarkers.^{4,15} In research, detecting biomarkers is the gold standard for identifying various diseases, from acute nutritional deficiencies to pathogenic infections, including cancer. Commonly used methods include polymerase chain reaction (PCR), western blotting, immunofluorescence, and enzyme-linked immunosorbent assay (ELISA).^{16,17} However, such analyses are tedious, time-consuming, need large amounts of samples and reagents, and depend on costly, heavy equipment.¹⁸ One major issue that earlier researchers pointed out is the need for more research in multiplexing or the simultaneous detection of multiple biomarkers. Multiple biomarkers can be analyzed simultaneously to provide a deeper understanding of diseases and enable more precise medical diagnosis.¹⁹ These limitations restricted affordable healthcare approaches, particularly in rural and developing regions. Therefore, there is an urgent need for simple, cost-effective, portable diagnostic solutions, especially microfluidics-based point-of-care (POC) devices, which have the potential to detect and monitor disease biomarkers, even in limited resource settings.^{17,20,21}

This multidisciplinary microfluidics approach has resulted in groundbreaking findings with the potential to transform conventional diagnostic techniques and enhance patient care on a worldwide scale.^{15,22} This advancement has opened the way for the design and manufacture of highly efficient miniaturized systems capable of carrying out complex analytical tasks with small sample quantities, including pretreatment, dilution, mixing, chemical reactions, separation, detection, and product extraction of samples, while also significantly cutting down on the time required compared to traditional laboratory techniques.^{23–25} These capabilities are promising for point-of-care applications and lab-on-a-chip technologies, especially in the quick and sensitive identification of disease-specific biomarkers.^{6,21}

In recent years, Song *et al.* presented a microfluidic approach to investigate the effect of macrophages on tumor cell invasion into the surrounding stroma in the presence of interstitial flow. This device has four parallel connected microchannels and could be flexibly used to build a more biomimetic breast cancer microenvironment, containing tumor aggregates, monocytes, macrophages and endothelial cells cultured in a 3D collagen matrix.²⁶ Further, a group of researchers has developed a high-throughput nucleic acid isothermal amplification system that can detect 19 common

respiratory viruses (including SARS-CoV-2) from 16 clinical samples in 90 min.²⁷

Using biomarkers for early identification of diseases and monitoring progression is difficult due to low biomarker concentration that affect downstream analysis. To enhance the extraction of biomarkers from biological matrices, novel techniques need to be developed. Liquid biopsies, which utilize physiological fluids like blood, saliva, and urine and are less invasive than solid tissue samples, are currently used in clinical procedures for early disease identification.¹⁵ However, the complex composition and limited presence of these biomarkers in samples can lead to analysis errors, requiring enrichment protocols to improve the sensing capabilities of analytical techniques.^{28,29} Microfluidic devices enable on-chip point-of-care diagnosis and real-time monitoring, detecting biomarkers for conditions like autoimmune disorders and acute nutrient deficiencies.³⁰

This study provides an in-depth review of integrated microfluidic devices for bioanalysis, emphasizing their potential uses in point-of-care (POC) diagnostics. The paper contributes significantly to the ongoing development of healthcare diagnostics, paving the way for better patient care and outcomes by thoroughly investigating optical and electrochemical methods, amperometry, potentiometry, cyclic voltammetry, and electrical impedance spectroscopy analysis. Microfluidic devices for POC diagnostics enhance understanding and promote translational research. This study focuses on integrated point-of-care microfluidics platforms to diagnose autoimmune diseases (including rheumatoid arthritis, multiple sclerosis, type 1 diabetes, and Addison's disease), acute nutritional deficiencies (like iron, vitamins A, D, B₁₂, C, and K), and pharmacological response biomarkers. Additionally, this paper summarizes about the recent advancement in microfluidic devices for POC diagnostics to drive future research. Integrated microfluidics improves patient outcomes by detecting diseases at the early stage. Health benefits are visible when biomarkers are identified and diagnosed at the point of care so that quick decisions can be made and personalized treatments can be administered.

2. Microfluidics and nanofluidics: next-gen point-of-care lab-on-a-chip devices

2.1. Microfluidics

1950s saw the rise in miniaturization with integrated circuits and photolithography (yes, it is way older than microfluidics). Jay Andrus patented the technique of photoengraving to create finer details that allowed for semiconductor devices.³¹ Further, this process was widely used and known as photolithography. Inkjet printing has a significant influence on the microfluidics industry. In 1879, Lord Rayleigh explained how a falling, continuous fluid stream breaks into droplets to minimize surface area.³² Later, many researchers used the phenomena to fabricate an array of inkjet nozzles over an individual silicon wafer. As time progressed, researchers turned their attention to



molecular analysis. The late 20th century saw the rise of polydimethylsiloxane (PDMS) as the base material for the fabrication of microchannels. A wide range of microfluidic platforms gave rise to many new functionalities – droplet microfluidics,³³ paper analytical devices,³⁴ organ-on-a-chip,³⁵ and open microfluidics.²⁰ Technology has scaled to the microscale to facilitate molecular biology, molecular analysis, and microelectronics.³⁶ Microfluidics helps in achieving a high surface area-to-volume ratio. This makes the microchannel feasible to handle and process only a small volume of liquid *i.e.*, a few microliters. Using microscale, fluid can be relatively easily manipulated using many kinds of external fields (electric, magnetic, pressure, capillary, and so on). Several microfluidic channels can be integrated into a single chip, and parallel observations and analyses can be performed simultaneously. These channels are made of PDMS, acrylic, glass, *etc.*, and can be easily visualized under a microscope owing to the transparency of the materials.³⁷ This technology has gained enormous attention on account of rapid developments in biotechnology and medicine as detection and manipulation at a cellular length scale with minimum reagent volume is advantageous. Advancements in bio-analyte and disease biomarker identification are explained in Fig. 1. The ease of going micro helps in the fundamental studies of physical, chemical, and biological processes and makes the system

portable and inexpensive.^{38,39} Fluids, especially liquids, can be considered continuous in tiny devices and geometries and well-established continuum approaches can be used for flow analysis. To obtain the required governing equations – the fluid flow, heat transfer, and other related physical phenomena in any system are modeled by applying conservation laws of mass, momentum, energy, and species.⁴⁰ These equations, usually in the form of partial differential equations, define the physics of the problem. The detailed approach has been mentioned in various fluid mechanics textbooks.^{41,42} Dimensionless numbers are often used to characterize fluid regimes in fluidics. Reynolds number (Re) is the ratio of inertial forces to viscous forces, $Re = \frac{\rho v D}{\mu}$, where v is the velocity in the microchannel ($m\ s^{-1}$), D is the characteristic dimension of the microchannel (m) and ρ is the density of the fluid ($kg\ m^{-3}$). In microfluidics, $Re < 2300$ and is typically laminar.³⁷ The low Re flow regime of microsystems is called creeping flow or Stokes flow.

This term describes the behavior of fluids in microsystems, where viscosity plays a more significant role than inertia. In macroscopic fluids, mixing occurs convectively, where inertia dominates over viscosity. However, in microsystems, when two fluid streams meet in a microchannel, they flow in parallel without any eddies or turbulence. In such a scenario, mixing happens due to the

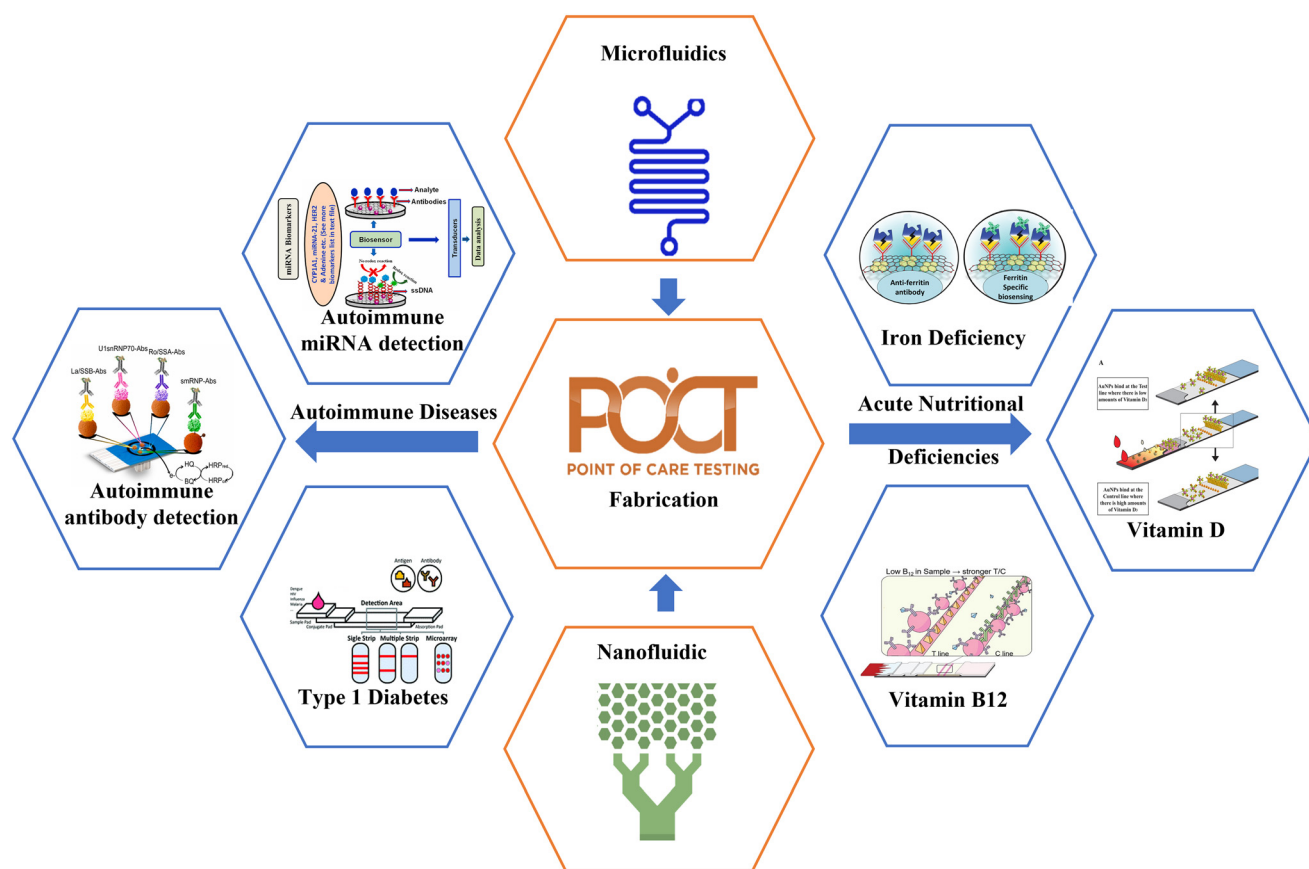


Fig. 1 Schematic illustration of quantitative and qualitative identification of bio-analytes and disease biomarkers; these innovative devices revolutionize diagnostic capabilities, enhancing accessibility and efficiency in healthcare settings.



diffusion of molecules across the interface between the two fluids. This phenomenon plays a crucial role in microfluidics, where precise control over mixing is essential for various applications. By understanding the behavior of fluids in microsystems, researchers and engineers can design and optimize microfluidic devices for various applications. Microscale systems require special machines or attachments to mix two fluids. In micro/nano-fluidics, the surface area-associated quantities are of utmost importance. Specifically, pressure and shear forces become dominant due to the small scale of the system.⁴³ Additionally, the capillary effect, a surface effect, is particularly prominent in microchannels with a bore diameter equal to or less than 50 μm . This effect is a result of the smaller size of the channels and the high surface-to-volume ratio of the components. As a result of this ratio, heat transfer occurs rapidly, facilitated by the low thermal mass of the system.⁴⁴

2.2. Exploring nanofluidics and its contrast from microfluidics

Nanofluidics deals with nanometric scale fluid flow behavior where one characteristic length is at least 1–800 nm.¹⁵ This is the most miniature possible length scale in which fluid flow can be manipulated and visualized owing to the state-of-the-art technologies.⁴⁵ New and unusual physical phenomena and mechanisms are observed at the nanoscale which is different from micro-and macroscale physics. The extra-large surface area to volume ratio (compared to microfluidics) causes a challenge as the wall's proximity and surface characteristics pose a problem in describing accurately the fluid flow and particle dynamics. While most gaseous and liquid microflows are described by continuum mechanics theory (or quasi-continuum, adding velocity slip and temperature jump models), nanofluidics exhibits anomalous behavior.⁴⁶ As a result, the surface characteristics are amplified, and fluid properties such as density and viscosity may fluctuate over interatomic distances. Although microscale material properties are the same as macro scale, the same nanomaterial may exhibit different thermal and electric conduction, optical behavior, and surface reactions, often due to rearranged lattice atoms or molecules.⁴³ Like microfluidics, nanofluidics systems also include supply reservoirs, flow conduits, passive or active pumps, actuators, valves, sensors, heaters, *etc.*⁴⁷

Microfluidics and nanofluidics are interconnected fields focused on manipulating fluids at small scales. Microfluidics operates at the micrometer level and is associated with lab-on-a-chip devices, enabling precise control and rapid assays for applications like point-of-care diagnostics (Fig. 2).^{48–50} Nanofluidics takes this further, working at the nanometer scale (less than 100 nm) to manipulate molecules and explore molecular interactions.⁵¹ Despite their scale differences, both fields share foundational principles and techniques. Microfluidics serves as a basis for nanofluidics, allowing for the development of intricate nanostructures and the

exploration of complex fluidic phenomena.⁵² The relationship between microfluidics and nanofluidics is characterized by their shared focus on fluid behavior at micro/nano scales, the increasing significance of surface effects, and their applications in various fields.⁵³ The relationship leads to innovative applications, particularly in point-of-care diagnosis, where microfluidic and nanofluidic lab-on-a-chip devices are advancing healthcare and diagnostics.

Because millifluidics, microfluidics, and nanofluidics are distinguished in different length scales, the dominating interactions in millifluidics and microfluidics include viscous drag, inertial force, interfacial tension, and capillary force, whereas electrostatic force, van der Waals attraction, hydrogen bonding, and steric repulsion become dominant in nanofluidics.⁴⁸

DNA sequencing using nanofabrication, superhydrophobic surfaces, and carbon nanotubes are some of the revolutionary applications of nanofluidics. The application can be seen in a much more comprehensive range of nanometric objects, *e.g.*, nanoparticles, DNAs, viruses, exosomes, lipid vesicles, and polymers.^{54,55} The advent of nanofluids and the suspension of nano-sized particles in a base fluid led to an increased reach of nanofluidics. These days, nanorods, nanofibers, nanowires, *etc.* can be seen as important constituents in various devices such as those used in water purification, biomedical applications, electronics, photonics, mechanics, *etc.*^{56,57} However, nanofluidics is in its infancy compared to microfluidics; nanochannels are ultras-small confined spaces and most standard tools used in microfluidics are challenging to apply to the field of nanofluidics. This is anticipated as the phenomenon experienced at the nanoscale has yet to be understood entirely. To evaluate the underlying

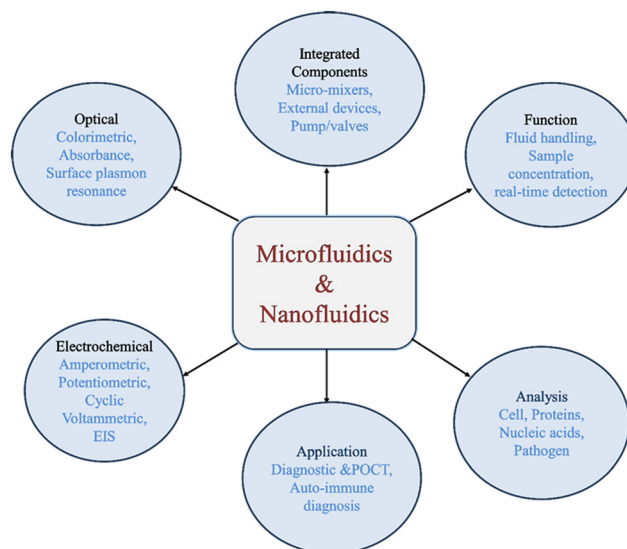


Fig. 2 The amalgamation of nanofluidics and microfluidics has ushered in a new era of possibilities in healthcare. Through their innovative capabilities, these technologies drive advancements that promise enhanced precision, accessibility, and efficacy in diagnostics, treatments, and overall patient care.



physics at the nanoscale, there should be a proper understanding of device dimensions, surface properties, ionic strength, *etc.*^{58,59}

2.3. Point-of-care diagnosis

Advancements in microfluidics, nanotechnology, microelectromechanical systems (MEMS) technology, data analytics, and 3D printing have opened the doors for significant developments in diagnostics in resource-limited places. In such low-resource settings, testing the mass for diseases or infections can be achieved by emerging POC technologies. Strengthening the healthcare system is the best way to keep the population healthy and productive for a country's best interest.⁶⁰ A large portion of the people in developing countries have no or very little access to appropriate healthcare. Tests that are performed at the decentralized level at the point of need or near the patient are called POC diagnoses. These require minimal training of the health care workers and are affordable and rapid in generating results. Although central clinical labs provide more accurate and sensitive results, they are labor-intensive, expensive, require specialized equipment, and have long patient wait times.⁶¹ With the help of POC test kits, preliminary screening can be done by a caregiver or the frontline nursing staff, reducing the burden on hospitals, and giving more time to focus on treatment. Rapid diagnosis will help in specific and effective treatment and contain the spread of infectious diseases.^{34,62} According to the World Health Organization (WHO), POC tests that address infectious disease control needs, especially for developing countries, should follow "ASSURED" criteria: 1) affordable, 2) sensitive, 3) specific, 4) user-friendly, 5) rapid and robust, 6) equipment-free and 7) deliverable to end-users.⁶³ POC device settings include 1) a sample collection kit, 2) portable instrumentation with ready-to-use reagents to process the sample collected, 3) a reader to quantify the results, for example – a smartphone, and 4) minimally trained personnel.⁶⁴ Lab-on-a-chip/disc (LOC/LOD) devices,

lateral flow devices, microfluidic paper-based analytical devices (μ PADS), *etc.*, are the many types of POC devices.^{65,66} This sector has contributed to blood glucose level detection and cardiological analysis. Blood glucose monitoring POC devices have proved efficient in diabetes control and management among the vast population.^{67,68} Other high-end POC devices include blood pressure and oxygen measuring devices. Fig. 3 shows the current trends of POC.

Gou *et al.* developed a compact and affordable smartphone-integrated dPCR (digital polymerase chain reaction) device, coupled with a self-priming dPCR chip, which offers precise DNA quantification down to 10 copies and single-molecule detection, showing promise for POC applications.⁶⁹ Cao *et al.* established a portable digital loop-mediated isothermal amplification (dLAMP) platform that integrates microscale hydrogel arrays for simplified DNA sample partitioning, enabling cost-effective and user-friendly absolute DNA quantification for potential point-of-care nucleic acid testing.⁷⁰ Advances in miniaturized nucleic acid testing (NAT) are propelling the development of fully integrated, simplified, and high-sensitivity point-of-care NAT devices with digital readouts, showing potential for MARCHED (miniaturized, automatic, reagent-preloaded, commercialized, high-throughput, environment-independent, and disposable) applications.²³ Recently, Gupta *et al.* introduced a novel spiral-shaped microfluidic device with a reduced graphene oxide-modified nanohybrid membrane for efficient urea filtration, mimicking the kidney's Glomerulus and showcasing the potential for organ-on-a-chip applications.⁷¹ POC devices that detect biological analytes offer rapid diagnostics even in resource-limited or remote healthcare settings. The seamless integration of POC devices with advanced technologies like fifth-generation wireless communications (5G) and the internet-of-things (IoT) can enable efficient healthcare solutions. The next-generation healthcare approach could become more patient-friendly and cost-effective with fast, low-latency internet connectivity, high bandwidth, cloud-based storage, and real-time computing capabilities. By combining microfluidic POC devices with mobile connectivity and cloud computing, remote disease monitoring and improved doctor-patient communication can be achieved.⁷²

2.4. Lab-on-a-chip (LOC) devices

LOC is a small, integrated analytical platform that can perform multiple tasks in a single platform of only a few square millimeters up to centimeters in size. Combining fluidic channels and chambers with these platforms can achieve the miniaturization, integration, automation, and parallelization of a wide range of biochemical processes. Researchers have explored various enzyme-based biosensor technologies for biomolecule detection, motivated by advancements in analytical methodologies and the need

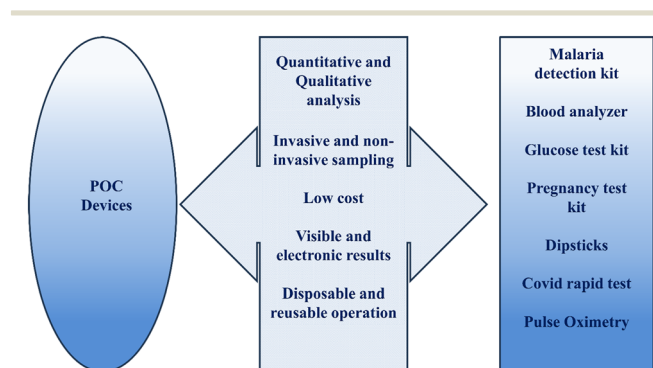


Fig. 3 Current status of point-of-care diagnostic devices in the healthcare system with an update on the COVID-19 pandemic. POC devices are allocated according to the need and usage in different fields.⁶⁸



for efficient, accurate, and economically viable alternatives.⁷³ This means that multiple tests can be performed simultaneously, with each test providing reliable and accurate results. In addition, the LOC platform is designed to function as its full-scale counterpart, making it a highly efficient and cost-effective solution for various laboratory needs.^{74,75} Lateral flow assays (LFA) are an excellent modern-day example of lab-on-a-chip technology that brings sample preparation and analysis to a single platform. This technology combines the convenience of an analytical test strip with the versatility of a microfluidic device. The test strip is either enclosed in a cassette or can be made as a dipstick tube. On-chip storage of dry reagents is well-developed on an LFA. The signal detection of an LFA is either optically interpreted (HIV test kit, pregnancy test kit) or read out by a reader.⁷⁶ These devices typically comprise a membrane, a sample pad, an absorbent pad, a conjugate pad, and a wicking pad. The sample is applied to the sample pad and then spread onto the absorbent pad by capillary action. When the sample meets the conjugate pad, a reaction takes place between the analyte and the antibody labeled with a dye or a marker. The reaction mixture then moves forward to the wicking pad, and the presence or absence of the analyte is detected by a visual signal. LFA technology has revolutionized the diagnostic field by providing precise, rapid, and low-cost results.^{76,77}

3. Microfluidics: from clinical setups to lab-on-a-chip

3.1. Scaling down for qualitative and quantitative analysis

Scaling down clinical tests and processes is a breakthrough in diagnostics. Qualitative and quantitative analysis at the point-of-care setting can cut long hospital waiting times and lets professionals focus on serious patients first.⁷⁸ The major limitation of laboratory tests is the long turn-around time. The patient must travel to the lab/hospital to have the blood drawn. The specimen is sent to specialized labs for testing and after that, the results are provided to the doctor. In such cases, the patient has to wait for a day or more for the results.⁷⁹ In addition to this limitation, high cost, low test speed, complicated assay procedures, and requirement of skilled personnel prove that the lab-based test is not suitable for point-of-care settings or limited-resource settings.⁸⁰

The POCT technology can be divided into two categories: a) a small hand-held biosensor or an LFA (lateral flow assay) for the qualitative or quantitative determination of several analytes.⁸¹ For example – glucose biosensor strips⁸² and lateral flow strips to determine cardiac markers.⁸² b) Bench-top devices, which are essentially the instruments that are used in labs but are reduced in size and complexity. For example – the use of PCR to provide infectious disease testing in a sufficiently small device at the POC.^{83,84}

3.2. Micro- and nanofluidic technology for a cost-effective accessible platform

Biosensors and point-of-care testing (POCT) technologies have become increasingly important in medical diagnostics. Fluidic systems are widely found across various scientific domains owing to their benefits in scaling down, creating distinctive instruments for disease detection, and isolating biomolecules.⁸⁵ One of the key factors in achieving these objectives is using robust reagents and consumables in storage and usage. This can be accomplished through micro- and nanofluidics, which provide efficient throughput processing and enhance control of transport. By handling minimal volumes of reagents, biosensors and POCT technologies can be made more reliable and user-friendly. The ease of integration of detection on the same stage has also led to multiplexing on-site detection of different analytes from a single sample.⁸⁶ The design and fabrication of a microfluidic platform are carefully addressed in the literature, taking its dimension, material, and method of fabrication into consideration which will, in turn, improve the biocompatibility of the device. In the early stages of a disease, biomarkers are found in lower concentrations, making it difficult to diagnose using conventional techniques. Here, using the present technology, ultra-sensitive modules can be fabricated.⁶⁴ Movement of fluid on a micron-scale or less has a wide range of applications such as biomedicine, cell biology, bio-sensing, and chemical synthesis. Previously, only valves and pumps were used to drive working fluid into the microchannel, categorized as an enclosed microfluidic system. Later, they were categorized as continuous flow and droplet flow systems. The emerging digital microfluidics could achieve discrete droplet movement under electrical potential and a magnetic field could be achieved. Photolithographic fabrication of channels in a cleanroom increases the overall fabrication cost and contradicts the promise of lower experimental prices.⁴⁰ To control the cost and bulky techniques, paper-based microfluidics involving open channels and capillary forces for liquid flow is gaining popularity. For mass detection in resource-limited settings, the target application for paper-based devices has been an inexpensive POCT technology.⁸⁷ Martin and Synge used paper as a diagnostic platform to access chromatographic techniques which was later recognized for Nobel Prize in 1952.^{88,89}

4. Bioanalysis by a fully integrated microfluidic device

4.1. Optical analysis

Optical analysis offers unique advantages to traditional analytical techniques in providing real-time detection and non-labeling of biological and chemical substances in a more sensitive, precise, and less expensive way. Optical biosensors are based on detecting visual signals produced by biological recognition by an analyte or biocatalytic or bio-affinitive



reactions. These visual aids can be used to absorb light, appearance, light, Raman scattering (RS), and refractive index (RI) changes in the eye area that are known to occur due to specific biomolecular interactions.⁹⁰ Optical biosensors are used in health care, environmental analysis, and biotechnology. These applications benefit from a wide range of applications, including low concentration and the required number of analyses, faster analysis termination, and reusable sensor chips.⁹¹ In general, two diagnostic procedures can be used for optical biosensing: fluorescence-based detection and label-free detection. In fluorescence-based detection, targeted molecules or biorecognition molecules are labeled with fluorescent markers, such as dyes; fluorescence intensity indicates the presence of targeted molecules and the potential for interaction between targeted molecules and biorecognition elements.⁹² There are a few studies done in the field of medicine and the environment. In one of the studies, gold nanoparticles (AuNPs) were attached to the tetanus antigen and taken from an experimental line-up with the structure of an AuNP-labelled tetanus antigen, tetanus antibody, and tetanus antigen.⁹³

This study employed an AuNP-based testing platform to detect tetanus antibody values. The test and control lines were prepared with human tetanus and antibodies for routine human tetanus. Then, samples containing tetanus antibodies were added to the sample pad, and subsequently, a red band with a mark can be seen in the test site in Fig. 4 due to the accumulation of AuNPs. The red band can also be seen in the control area, indicating the feasibility of the test line.⁹³

4.2. Electrochemical analysis

For electrochemical analysis, the biosensor has an electrochemical transducer (Fig. 5A). The electrochemical

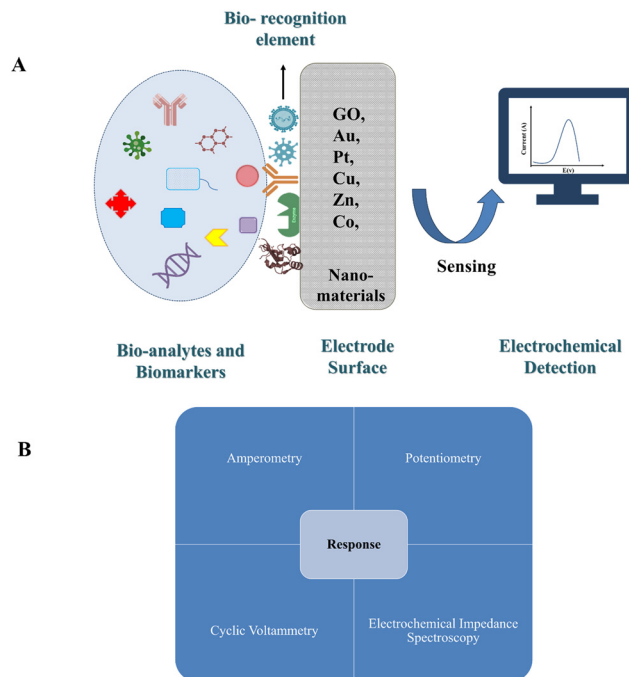


Fig. 5 Schematic representation of (A) electrochemical biosensors and their components – bio-analytes and biomarkers, electrode surface which is modified with various nanomaterials and the sensing platform. (B) Type of electrochemical response: the various responses help in quantitative analysis by processing the response signals.

transducer is classified as a chemical-modified-electrode (CME)^{93,94} as an electronic conducting, ionic conducting, or semi-conducting material treated with a bio-recognition element.

Electrochemical analysis is the chemical sensory phase, where the electrode is used as the transducer element in which the analyte is located. The current generated from the

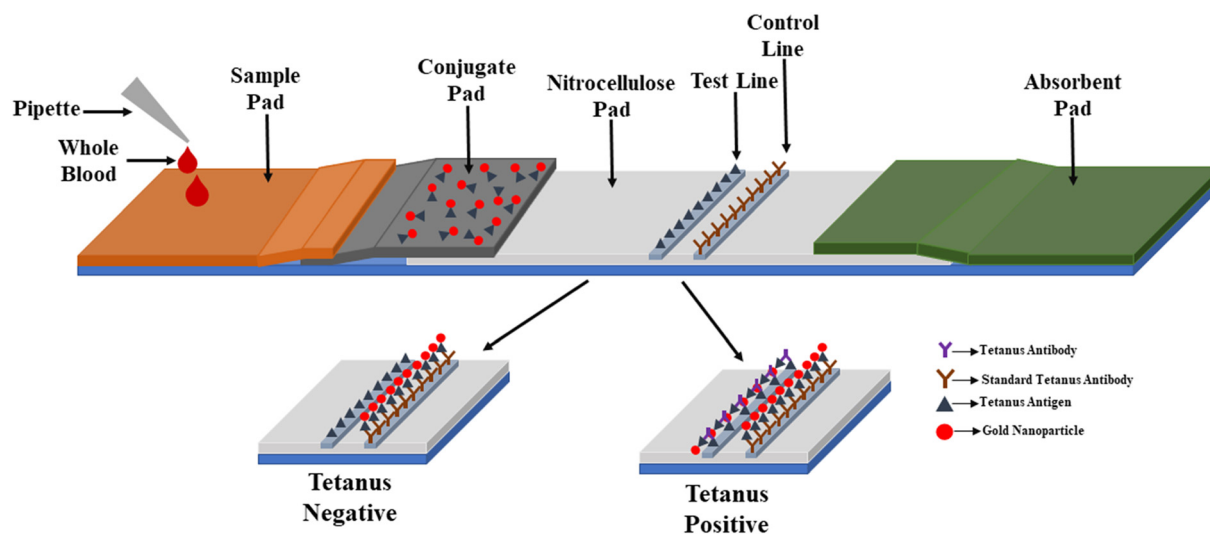


Fig. 4 Schematic of AuNP-based lateral flow assay to detect tetanus antibodies. The illustration shows the migration of the AuNP-tetanus-tetanus antibody complex towards the test zone for colorimetric detection of tetanus antibody.⁹³



response is monitored and used to calculate important data such as the concentration of the analyte. Electrochemical techniques have great potential for easy miniaturized use of portable devices in various applications such as medical diagnostics and environmental monitoring. In the electrochemical biosensor due to the direct transformation of the biological event to the electronic signal, this provides an attractive way to analyze the biological sample.⁹⁴ The sensory pathway of electrochemical sensors is categorized in Fig. 5B according to the method used.⁹⁵

4.2.1. Amperometry. An amperometric transducer studies charge transfer between phase interactions, for example,

between two electrodes separated by an electrolyte. An electrochemical cell is often used to describe a system of phases and boundary parameters. One half-cell reaction within an electrochemical cell is carefully controlled to study changes in charge transfer at the interaction of another cell part reaction, commonly called a working electrode.⁹⁶ Amperometric types are based on the current concentration associated with oxidation or reduction of the electroactive element type involved in the recognition process. By controlling the fixed or differential energy throughout the electrochemical cell, higher energy can be created, which is the difference between the applied energy and the

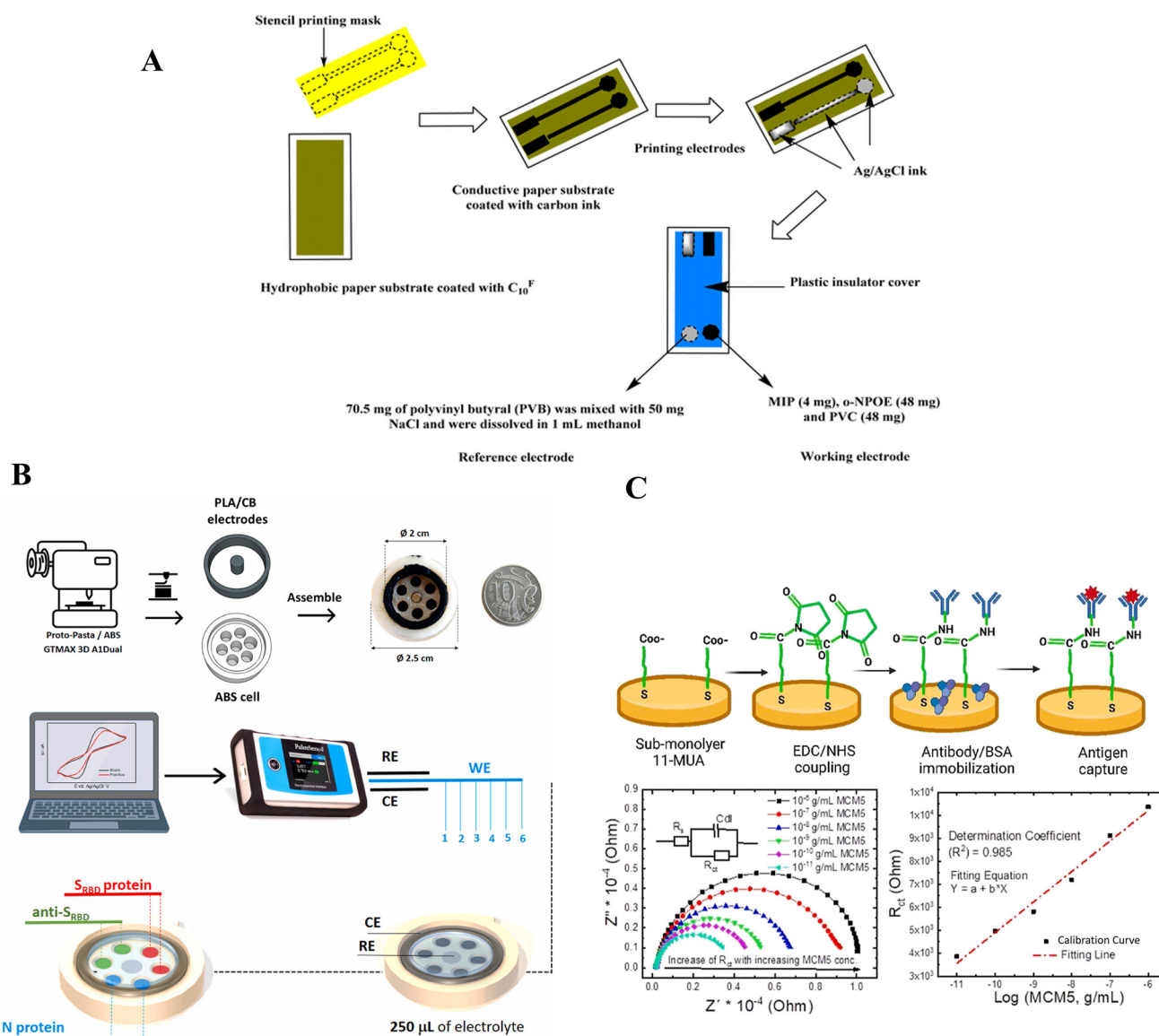


Fig. 6 Schematic illustration showing (A) development of a paper-based analytical biosensor for potentiometric sensing of myoglobin, a cardiac vascular disease biomarker. A hydrophobic paper substrate that has been coated with fluorinated alkyl silane, specifically (CF₃ (CF₂)₇CH₂CH₂SiCl₃, CF₁₀) for potentiometric sensing. (B) 3D-printed cell and electrochemical set-up for detection of three Covid biomarkers. The combination of polylactic acid and carbon black in fabricating the working electrodes ensures reliable and sensitive detection of N protein, SRBD, and anti-SRBD. (C) Electrochemical immunosensor for cervical cancer biomarker MCM5 on a gold disk electrode. MCM5 involves the formation of sub-monolayer capping layers of 11-MUA, activation of the terminal -COOH groups, coupling of MCM5 antibodies and subsequent blocking with BSA, MCM5 antigen-binding at various concentrations, and electrochemical readout for quantification.



equilibrium force. In the overpotential formulation, electron transfer becomes thermodynamically strong and will be followed by oxidative or reducing reactions. These processes are called faradaic processes as they comply with Faraday's law; the amperometric biosensor may be more attractive due to its high sensitivity and wide linearity.⁹⁷ The recent development of an amperometric biosensor offers promising results in quantifying urinary disease biomarkers. Specifically, this biosensor is designed to detect the biomarker p-HPA using allosteric modulation of the bacterial reductase C1. This reductase is embedded and electrically wired to the electrode surface in a redox hydrogel, allowing for reliable and sensitive biomarker detection. The modulation of the sensor current observed at a constant NADH supply facilitates the quantification of the biomarker, making this biosensor a valuable tool for diagnostic purposes. In summary, the C1-*hpah*/redox polymer biosensors have shown great potential in the detection and quantification of p-HPA, paving the way for future developments in the field of urinary disease diagnosis.⁹⁸

4.2.2. Potentiometric. In potentiometric analysis, we determine the accumulation of charge power at the working electrode relative to the reference electrode in the electrochemical cell where zero or no current flow is significant between them. In addition, potentiometry brings information related to ion activity when electrochemical reactions occur. In potentiometric measurements, the relationship between concentration and energy is controlled by Nernst's equations (eqn (1)) where E_{cell} represents the observed cell potential at zero current.⁹⁹ This is sometimes also referred to as the electromotive force or EMF. $E_{\text{o cell}}$ is a constant potential contribution to the cell, R is the universal gas constant, T is the absolute temperature in degree Kelvin, n is the charge number of the electrode reaction, F is the Faraday constant and Q is the ratio of ion concentration at the anode to ion concentration at the cathode.

$$E_{\text{cell}} = E_{\text{o cell}} - RT/nF \ln Q \quad (1)$$

Potentiometric sensors are suitable for measuring low concentrations in small sample volumes, as they offer the advantage of not exposing the sample to chemicals. Varieties of ions, which may not be able to achieve low limits, are currently limited and do not have significant solvents such as nickel, manganese, mercury, and arsenate ions.¹⁰⁰

Almehizia *et al.* presented a potentiometric PAD coupled with an myoglobin (Mb) imprinting polymer/MWCNTs composite for Mb detection (Fig. 6A). In this method, the appropriate MIP particles were added to a polymeric membrane to act as an Mb receptor. The presented sensors showed a linear range of 5.0×10^{-8} to 1.0×10^{-4} M with a potentiometric slope of -57.1 ± 0.3 mV per decade ($R^2 = 0.9998$) and a detection limit of 28 nM at pH 4. This approach offers a reliable and sensitive method for Mb detection, which can be used in various applications. The Mb imprinting

polymer/MWCNTs composite used in this study provides an effective way to detect Mb in biological samples. Therefore, this method can be used as a potential tool for clinical diagnosis and therapeutic monitoring of various diseases.¹⁰¹ Moreover, Akl *et al.*'s research paper discusses the production of a nano-Cu-complex (N1) through a Schiff base ligand (A1) reaction with copper chloride. The nano Cu complex (N1) has been utilized to develop a potentiometric membrane biosensor specifically designed for the early detection of prostate-specific antigen (PSA). The results of the study indicate that the electrode exhibits a stable Nernstian response of 29.26 ± 0.87 mV per decade for PSA within a linear dynamic range of 5.0 pg mL⁻¹ to 10.0 ng mL⁻¹. The pH range for this response is between 6.5 and 9.2, with a rapid response time of 25 ± 5 s. The lifespan of the sensor is yet to be determined, but the results from the study provide a promising foundation for further research into the development of a reliable and efficient PSA detection method.¹⁰²

4.2.3. Cyclic voltammetry. Cyclic voltammetry (CV) is a type of electrochemical technique that is used to investigate the electrochemical characteristic related to electroactive surfaces. CV is the handiest electrochemical process that permits the understanding of the redox reaction mechanism, electron transfer kinetics, and reaction reversibility of an electroactive molecule in a sample. CV facilitates rapid information on the kinetic electron-transfer reactions of heterogeneous reactions, on the adsorption process or coupled chemical reactions, and on the thermodynamic redox procedures. CV is the primary electrochemical experiment achieved to characterize an electrode material for several types of applications.¹⁰³ A cyclic voltammetry analysis incorporates a three-electrode system, which includes the relative potential to the RE scanned at a working electrode, however, the resulting current flowing through a counter electrode is observed in the analyte solution.¹⁰⁴

Various studies have been conducted using CV, and a novel 3D-printed cell has been applied for multiplex voltammetric detection of three main COVID-19 biomarkers: N protein, SRBD protein, and anti-SRBD in saliva and serum samples (see Fig. 6B). The multiple sensors demonstrated a wide linear range, with detection limits of 5, 1, and 1 pg mL⁻¹ for N protein, SRBD-protein, and anti-SRBD, respectively. Similarly, Bakhsh *et al.* presented a newly developed electrochemical sensor that has several attractive features. The sensor is capable of simultaneous determination of vitamin C and vitamin B6 with high sensitivity, selectivity, and rapidity. The SnO₂/GCE-based modified electrode has been found to exhibit remarkable electrochemical properties for the oxidation of both vitamin C and vitamin B6. After optimizing the parameters, including the amplitude, pulse width, and pH level at 0.05 V, 0.045 s, and 0.1 M PBS pH 7, respectively, the modified electrode demonstrated enhanced sensitivity over a broad linear dynamic range from 10 to 140 μM. The limit of detection for vitamin C



was measured at 0.067 μM , while for vitamin B6 it was found to be 0.048 μM . These results suggest that the SnO_2/GCE -based modified electrode is a promising tool for the detection and quantification of both vitamins in various samples.¹⁰⁵

4.2.4. Electrical impedance spectroscopy. Electrical impedance spectroscopy (EIS) analysis is where sinusoidal voltage or test current is applied to a biological sample or other samples, and its effect is determined in the appropriate frequency range. EIS is a powerful means of studying several types of electrical properties of building materials. Functionally, the impedance spectra measured by EIS, usually incorporated into a compatible electric model, represent a sample fingerprint that provides insight into its properties and behavior. EIS is used in various applications as a fast and easy automated way to display solid, liquid, balanced, natural, and inanimate materials.^{106,107}

An immunosensor for label-free electrochemical detection of Mini Chromosome Maintenance Protein 5 (MCM5) has been developed in Fig. 6C. MCM5 is a protein that is overexpressed in cervical cancer, which makes it a promising biomarker for early detection of the disease. The electrode surface was modified by creating a sub-monolayer of 11-mercaptopundecanoic acid (11-MUA) as a capture layer. The next step involved activating the electrode with *N*-(3-dimethylaminopropyl)-*N*-ethylcarbodiimide hydrochloride (EDC) and *N*-hydroxysuccinimide (NHS) to immobilize the capture antibody. During the formation of the 11-MUA layers, the change in the electrode surface properties, specifically its wettability, was determined using the static water contact angle method. The binding of MCM5 antigens to the capture antibody can be effectively monitored using cyclic voltammetry (CV) and electrochemical impedance spectroscopy (EIS) techniques. The results of exposing the antibody-functionalized surface to MCM5 antigens show a linear increase in reticulocyte count test (R_{ct}) with the logarithmic value of MCM5 antigen concentration. The linear dynamic range of this increase is between 10^{-6} and 10^{-11} g mL^{-1} , with a high correlation coefficient of 0.99 and the detection limit for this assay was 2.9 pM (10^{-11} g mL^{-1}).¹⁰⁸ Similarly Aydin *et al.*'s study focuses on the fabrication of an electrochemical impedance spectroscopy (EIS)-based immunosensor that enables the quantification of calreticulin (CALR), a new biomarker of breast carcinoma. The immunosensor is fabricated using a reduced graphene oxide/ amino substituted polypyrrole polymer (rGO-PPyNH₂) nanocomposite modified disposable electrode. The development of a highly sensitive electrochemical biosensor for the detection of calreticulin (CALR) antigens has been achieved using anti-CALR antibodies attached to a rGO-PPyNH₂ nanocomposite-coated electrode. The resulting immunocomplex induces a considerably larger interfacial electron transport resistance (R_{ct}) which increases the sensitivity and accuracy of the biosensor. The R_{ct} variation has been found to exhibit a linear relationship with CALR levels in the detection range of 0.025 to 75 pg mL^{-1} . This

relationship has been established using a suggested biosensor highly selective to CALR with a detection limit of 10.4 fg mL^{-1} . The biosensor has also demonstrated good storage stability and reproducibility.¹⁰⁹

5. Diseases, acute nutritional deficiencies, biomarkers

Diseases are frequently defined as medical disorders characterized by distinct signs and symptoms. External causes such as infections or internal dysfunctions can both cause illness. Internal immune system malfunction, for example, can result in various conditions such as immunodeficiency, hypersensitivity, allergies, and autoimmune disorders. A disease is a specific aberrant state that negatively impacts the structure or function of all or part of an organism and is not caused by external harm.¹¹⁰ Nutrient deficiency illnesses are developed when critical nutrients for growth and health are deficient. These illnesses are caused by a lack of food, which results in malnutrition or hunger. A structural or biological imbalance in the individual's metabolic system might also cause a deficient condition, nutritional shortage, or illness, whether currently treated or untreated. Protein energy malnutrition (PEM), scurvy, rickets, beriberi, hypocalcaemia, osteomalacia, vitamin K deficiency, pellagra, xerophthalmia, and iron deficiency are a few examples.¹¹⁰ Biomarkers are molecular traits that offer information on biological states, whether regular, diseased, or medically changed. They can help in diagnosis and prognosis, monitor illness, and measure medication success. While several biomarkers are widely used in therapeutic settings, they represent just a small fraction of all biomarkers found. Various variables contribute to the sluggish adoption of these novel biomarkers, including difficulties encountered along the biomarker development process.^{111,112} Microfluidics provides the field of biomarkers with two critical opportunities: first, it can address some of these developmental challenges, and second, it can provide the precise and complex platform required to bridge the gap between biomarker research and the biomarker-based analytical device market (see Fig. 7C). Indeed, microfluidics has opened up a new path for progress, boosting the clinical use of biomarkers and associated analytical platforms.⁴

5.1. Autoimmune diseases

Autoimmune diseases occur when the immune system defects and misidentifies self-tissues as foreign. Autoimmune illnesses are a group of chronic diseases that are either organ-specific (*e.g.*, thyroid diseases, Graves' disease, type 1 diabetes, and primary biliary cirrhosis) or systemic (*e.g.*, antiphospholipid syndrome, rheumatoid arthritis, and systemic lupus erythematosus).^{117,118} These illnesses are typically caused by a lack of normal immunological self-tolerance, which results in the development of high-affinity autoantibodies that erroneously target the body's cells,



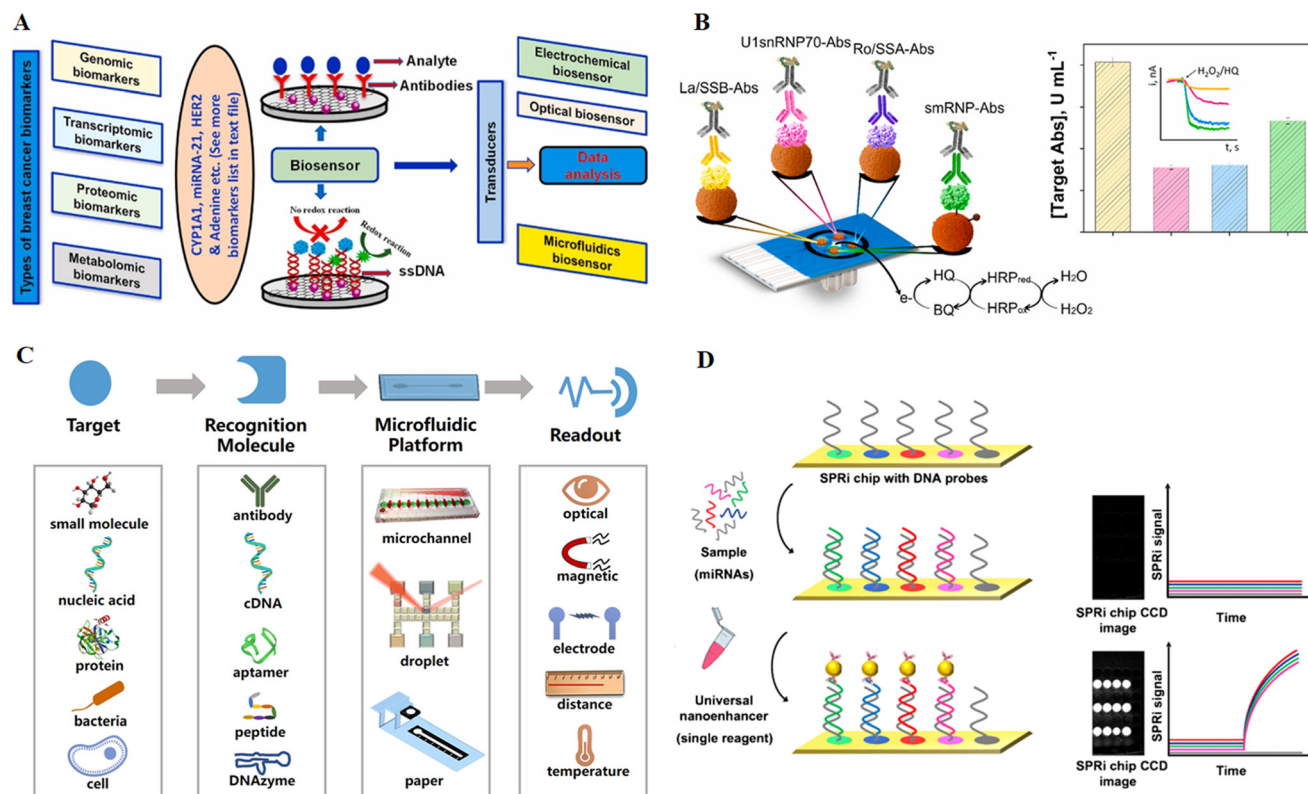


Fig. 7 Innovative approaches for autoimmune disorder detection. (A) Designing and implementing point-of-care biosensors for detecting autoimmune disorders by analysing genetic, gene expression, protein, and metabolic biomarkers. Reproduced from ref. 113 with permission from Elsevier, copyright 2020. (B) Multiplexed electrochemical platforms for the early and precise diagnosis of autoimmune conditions. Reproduced from ref. 114 with permission from Elsevier, copyright 2023. (C) An illustration of a microfluidics-based biosensor with a biological detection element, a microfluidic device, and a signal reading device. Reproduced from ref. 115 with permission from American Chemical Society, copyright 2018. (D) Strategy for simultaneously detecting multiple microRNAs in MS patients using a single nanostructured enhancer of surface plasmon resonance imaging (SPRI). Reproduced from ref. 116 with permission from Springer Nature, copyright 2019.

tissues, and organs. Only around 8% of the population may be affected by autoimmune illnesses. They are, nevertheless, one of the leading causes of severe disability, organ failure, and fatality, resulting in substantial medical costs. As a result, early detection of these illnesses is critical for enhancing patients' quality of life. Autoantibodies (AABs) in clinical samples are the greatest diagnostic biomarkers for predicting autoimmune illnesses' beginning, progression, and severity because they may be discovered years before clinical signs and irreparable damage appear.¹¹⁹ It is essential to identify biomarkers associated with these disorders. Recent advances in biosensor-based diagnostic techniques have enabled the identification (shown in Fig. 7A) of various genomic, transcriptomic, proteomic, and metabolomic biomarkers to diagnose autoimmune diseases. These biosensors can detect biomarkers in patient samples and provide accurate diagnoses. Furthermore, these biosensors detect multiple biomarkers associated with different autoimmune disorders, allowing for a more comprehensive diagnosis. Understanding the various biomarkers associated with autoimmune diseases makes it possible to develop more targeted treatments and therapies. This will enable better outcomes for patients with

autoimmune disorders, and improve their quality of life.¹²⁰ Arévalo, Beatriz *et al.* have presented pioneering research on the simultaneous detection of autoantibodies associated with autoimmune diseases. Their innovative approach utilizes modified magnetic microparticles (MBs) specific to the antigens. These MBs employ His-tag or carbodiimide/hydroxysuccinimide (EDC/NHS) chemistries to capture the corresponding autoantibodies (IgGs). The captured autoantibodies are then enzymatically labeled with HRP-conjugated secondary antibodies. The detection is achieved through amperometric transduction using the H₂O₂/HQ system on disposable platforms, enabling quadruple detection, Fig. 7B. The performance of this platform is suitable for clinical applications. It demonstrates the potential to differentiate between healthy individuals and patients diagnosed with autoimmune diseases. This differentiation is accomplished by determining the levels of the four autoantibodies in serum within a quick and reliable 45-minute timeframe.¹¹⁴

Furthermore, due to their specificity and durability during diseases, autoantibodies are beneficial and essential indicators. Currently, the most prevalent conventional techniques for AABs detection in autoimmune illnesses



include enzyme-linked immunosorbent assay (ELISA), indirect immunofluorescence (IIF), and western blotting. However, these approaches are costly, time-consuming, and inconvenient since they require complex tools. Because of their potential to detect disease-related biomarkers in real time, biosensors are among the most promising technologies for detecting disease-related biomarkers.¹²¹ A biosensor is a device with an integrated receptor – a transducer that may be used for quantitative or semi-quantitative analytical detection based on the signal transducer used. The biosensors are classified as optical, electrochemical, or piezoelectric devices. Nanomaterials (NMs) are prospective candidates for biosensor manufacture to boost sensitivity and decrease detection limits (LOD). Aside from AAbs, several proteins, chemokines, cytokines, cells, and ions have been identified as prospective biomarkers for diagnosing or monitoring autoimmune diseases.¹²² Current biomarker finding efforts are centered on three types of proteins that represent the autoimmune disease process: (1) degradation products resulting from tissue damage, (2) enzymes involved in tissue degradation, and (3) cytokines and other proteins related to immune activation.¹²⁰

5.1.1. Graves' disease. Graves' Disease (GD) is an autoimmune illness in which thyroid-stimulating antibodies (TRAB) replicate the effect of thyroid-stimulating hormone, resulting in hyperthyroidism. 5% of GD patients develop inflammatory Graves' orbitopathy (GO), characterized by proptosis and visual problems. According to UNO, Graves' illness affects females 5–10 times more frequently than males. The disease most commonly manifests in middle age, with a peak prevalence of 40–60%, although it can also afflict children, adolescents, and the elderly. Graves' illness may be found practically anywhere on the planet. Graves' illness is thought to afflict 2% to 3% of the general population. The most prevalent cause of hyperthyroidism is Graves' disease. Circulating proteins and microRNA (miRNA) are the two most often used blood biomarkers in clinical research due to miRNA's (Fig. 7D) growing mechanistic role in specific illnesses such as metabolic and cardiovascular disorders such as cancer.¹¹⁷ Serum diiodotyrosine DIT levels might be used as a novel biomarker to differentiate between destructive thyroiditis and Graves' disease, with values of 359.9 pg ml⁻¹ preferring Graves' disease and 359.9 pg ml⁻¹ favoring destructive thyroiditis.¹²³ Foster, Hayley, and colleagues showed that a microfluidic device can preserve human Graves' disease tissue, allowing for the extraction and characterization of Graves' disease-specific exosomes. The exosome markers CD63 and CD81 were found in 2/5 of the exosomes from Graves' disease tissue, while CD63 was also found in exosomes from a single non-Graves' sample. MiRNA-146a and miRNA-155 were found in all the samples, with no differences between tissue cohorts. Treatment did not affect miRNA expression in exosomes extracted from Graves' disease tissue. Although miRNA-146a and miRNA-155 were enhanced after dexamethasone and methimazole therapy of non-Graves' disease tissue, the rise was modest.

The results of this work show that incubating tissue on a microfluidic device allows for the identification, separation, and characterization of extracellular vesicles from human tissue biopsies.⁵⁴

5.1.2. Rheumatoid arthritis. Rheumatoid arthritis (RA) and osteoarthritis (OA) are the two most frequent types of degenerative joint disorders, and they share several symptoms while having different pathogenesis. The importance of early diagnosis and therapy in arthritis management is to prevent additional joint deterioration and maximize functional capacity. Rheumatoid arthritis (RA) is thought to affect 0.24 to 1% of the population and is twice as frequent in women as in males.¹²⁴ Biomarkers might be employed to understand pathobiological processes in the joint and differentiate the degree of articular cartilage deterioration. It is vital to detect existing biosensor technologies, sensing modalities, and the strengths and drawbacks of these techniques in resolving gaps in business attitudes. This detection mechanism is quicker than traditional methods, and no additional signal amplification is required.^{121,125} This raises the prospect of rapid, dependable, cost-effective screening for RA and OA. Biosensor research and commercialization can be critical in diagnosing and managing bone disorders and arthritis due to their features and benefits.¹²⁵ Around 20% of patients with diagnosed RA will not have an abnormal rheumatoid factor (RF) RF test, while 5% of those without RA will have an abnormal RF test. The typical RF range is 0–20 IU ml⁻¹. RF levels exceeding 20 IU ml⁻¹ are not regarded as sufficient to diagnose RA since the RF level might be raised for various causes.¹²⁶ Other autoimmune disorders, some chronic infections, diabetes, bacterial endocarditis, cancer, natural aging, immunizations, and transfusions are examples of conditions and medical treatments that might boost RF levels. OA is a common illness that requires early detection to avoid joint harm. However, OA clinical diagnosis is challenging, particularly in the early stages of the disease, and the absence of effective therapy is most likely due to late detection. The most frequently used techniques for OA diagnosis include radiographic imaging, which exposes patients to radiation. As a result, it is critical in creating novel selective and simple biosensors capable of detecting OA biomarkers at an early stage.¹²⁷

5.1.3. Multiple sclerosis (MS). MS is a neurodegenerative autoimmune condition characterized by nerve demyelination and axon destruction. The immune system begins perceiving myelin proteins as alien components, leading to the breakdown of myelin sheath. MS had an overall incidence rate of 3.6 per 100 000 person-years in women (95 percent CI 3.0, 4.2) and 2.0 (95 percent CI 1.5, 2.4) in males.¹²⁸ Apoptosis of oligodendrocytes, the cells responsible for the formation and maintenance of myelin, causes additional neuronal disintegration and function loss. The Food and Drug Administration (FDA) has authorized over a dozen medications with distinct mechanisms of action to treat MS attacks (*e.g.*, dimethyl fumarate, interferon (IFN), fingolimod,



teriflunomide, ocrelizumab, *etc.*).¹²⁹ At the moment, known biomarkers can aid with MS diagnosis and prognosis, as well as monitoring therapy response and assessing the risk of adverse effects. These include oligoclonal bands and the IgG index, anti-AQP-4 antibodies, IFN- and natalizumab neutralizing antibodies, and anti-JCV and anti-VZV antibodies. Furthermore, promising biomarkers such as NFL and CHI3L must be validated in additional studies.¹³⁰ Given the significance of demyelination in MS, Kerman *et al.* used stem cell biology and microfluidic technology to convert mouse embryonic stem cells into myelinating oligodendrocytes, then assessed and quantified myelin development over several days. According to live imaging and confocal analyses, the oligodendrocytes anchor to the naked axons before wrapping them and producing the myelin sheets. This model is an excellent tool for better understanding the myelination process and developing innovative therapies for demyelinating illnesses like MS.¹³¹

5.1.4. Type 1 diabetes. Type 1 diabetes is caused by an autoimmune reaction (the body mistakenly fights itself) that prevents your body from producing insulin. Type 1 diabetes affects around 5–10% of all people with diabetes. Type 1 diabetes symptoms can appear fast. It is most diagnosed in children, teenagers, and young adults. To survive, if you have type 1 diabetes, you must take insulin daily. Now, no one knows how to avoid type 1 diabetes. The meta-analysis findings revealed that the global incidence of type 1 diabetes was 15 per 100 000 persons, with a prevalence of 9.5 percent (95 percent CI: 0.07 to 0.12), which was statistically significant.¹³² The technique employs a microchip and sensor to detect blood indicators that can pinpoint the early loss of beta cells, which persons with type 1 diabetes cannot create. Researchers have already produced a proof-of-concept sensor coated with unique nanoparticles. The sensor can detect the presence of specific biomarkers in the blood, changing color if the molecule is present. Electrochemical measurements are reported more frequently in the literature than other approaches. As a result, they are the primary focus of our investigation. Biosensors remain the primary focus of most teams due to their excellent selectivity for glucose measurement.¹³³

5.1.5. Addison's disease. AD (autoimmune Addison disease) is one of the rarest autoimmune endocrinopathies. Most instances of AAD are caused by an abnormal immunological response aimed, at least in part, against the steroidogenic enzymes of the adrenal cortex (particularly 21-hydroxylase). Poudyal *et al.* created a simple, low-cost, microfluidic, field-deployable biosensor using a screen-printed carbon electrode (SPCE) for sweat cortisol detection in point-of-care applications. The detection of cortisol in artificial sweat is a valuable screening technique for identifying and monitoring a variety of health disorders such as Addison's disease, stress disorder, and Cushing's syndrome.¹³⁴ On the SPCE electrode, a self-assembled monolayer of graphene oxide (GO) is functionalized, onto

which cortisol antibodies are immobilized for cortisol detection. The equally dispersed GO increased the surface area available for antibody immobilization. Cortisol was identified in a linear range of 0.1 to 150 ng mL⁻¹, with the amplitude of the current decreasing with increasing cortisol concentration due to a decrease in the number of free electrons. The created microfluidic cortisol detection biosensor served as the foundation for a sweat cortisol sensor with POC applications, and it may also be employed in customized health diagnosis or monitoring.¹³⁴ AAD can advance slowly, with an insidious deterioration into adrenal insufficiency over months or years. AAD has a significant hereditary component. However, due to its rarity, the recurrence probability in family members is just 2%. Several disease susceptibility alleles have been found, including three loci linked to organ-specific autoimmunity (MHC, CTLA4, and PTPN22) and others that encode proteins implicated in innate immune responses (Mitchell, Anna L., and Simon HS Pearce).¹¹⁹

5.2. Acute nutritional deficiencies

5.2.1. Vitamin A deficiency. Vitamin A (VA) is a liposoluble vital micronutrient. It is obtained primarily from diet alone as the human body does not produce vitamin A. The vitamin can be drawn from animal products as VA (retinol and its close derivatives) or as provitamin A (carotenoids) from vegetables.⁸⁰ Vitamin A deficiency leads to night blindness (this causes trouble seeing at night), impaired vision, and increased morbidity and mortality in children and mothers in poor populations. Every year >250 000 children are born blind. The deficiency in vitamin A will deliver stunted growth, difficulty in reproduction, and difficulty in fighting infections.¹³⁵ Progress is made globally in controlling child deaths and blindness through effective oral supplementation in early childhood but it is also population dependent. Due to ambiguous findings, the WHO continues to recommend high supplement doses for children and infants; frequency and effective doses have not yet been determined. A close monitoring is highly required to determine the supplementation needs and when to stop it. For VAD diagnostics, different RBP cut-offs are proposed – 14.5 µg mL⁻¹ (0.69 µmol L⁻¹), 14.7 µg mL⁻¹ (0.70 µmol L⁻¹), and 17.4 µg mL⁻¹ (0.83 µmol L⁻¹) as the molar ratio between retinol and RBP depends on a lot of factors.¹³⁶

Keratomalacia and Bitot's spots are among the non-invasive indicators of clinical vitamin A deficiency (VAD). Though better biochemical indicators are always explored for better evaluation of VAD with minimal invasion. Vitamin A status can be monitored by determining the serum retinol concentration of an individual.¹³⁷ Retinol binding protein (RBP) is present in 1:1:1 M concentration with retinol and transthyretin. As serum RBP is present in a 1:1 M complex with retinol, it can be chosen as a surrogate biomarker for serum retinol



concentration.¹³⁸ It is easier to access serum RBP than serum retinol. Serum RBP is more stable with reference to light and temperature. Also, analysis of serum RBP requires finger prick blood *i.e.*, 10–20 μL .¹³⁹ However, acute infection and inflammation in the human body affect the serum retinol and serum RBP levels. So, it is always wise to measure C-reactive protein (CRP, a biomarker for infection) along with serum RBP.¹³⁷

A third of the population is affected by VAD and it is the need of the hour to carry out mass testing for close monitoring and administration of doses. Though clinical results are accurate, it takes at least one day, and the patient must travel to the laboratory for blood tests. So, it is challenging to do mass testing at the clinical level in resource-limited settings.^{140,141} Lu *et al.* have developed a test strip to quantitate ferritin (a biomarker for iron deficiency), RBP, and CRP. It is a multiplexed test strip with a reusable

stand-alone reader. To read the concentration of the markers in the sample, fluorescence tags are bound on the test line. The intensity is measured with the help of fluorescence imaging mode and the concentration of the marker is read out.¹³⁶

5.2.2. Iron deficiency (ID). Iron is an essential component of human blood production. 70% of iron is present as hemoglobin in the erythrocytes, which helps in oxygen transfer from the lungs to the tissues. It is also an important component of enzyme systems, which is helpful in oxidative metabolism. Iron is stored as ferritin in the liver, spleen, muscle tissue, and bone marrow and is transferred in the body by transferrin (a blood protein that binds to iron). Iron deficiency is a widespread nutritional disorder worldwide and is most prevalent among pregnant women, infants, school children, and non-pregnant women.¹⁴² Iron deficiency anaemia (occurs when the body does not have enough iron

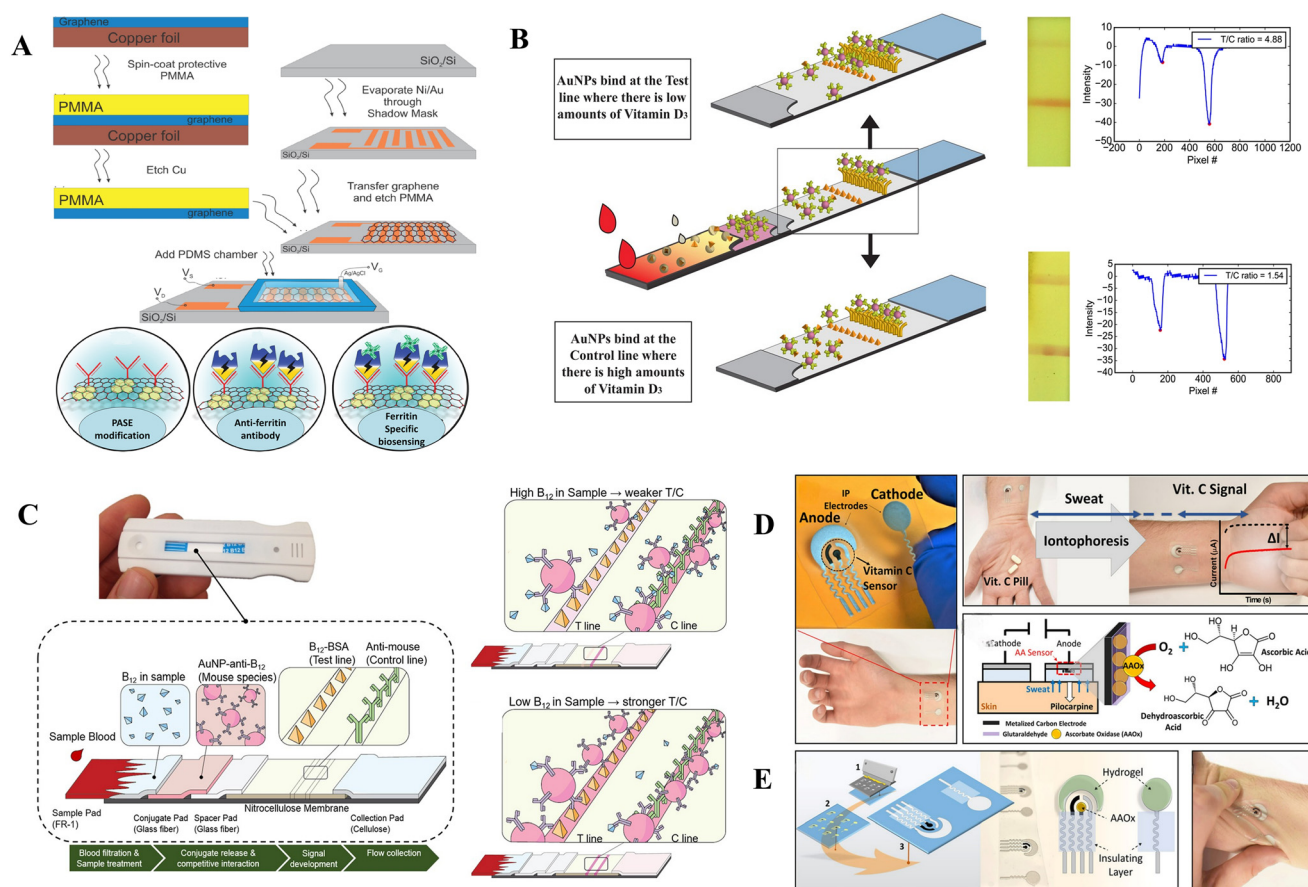


Fig. 8 Advancing point-of-care techniques. (A) Developing a complete strategy for detecting ferritin involves a combination of modern methods. This includes using GFET technology to improve graphene-based biosensors, a sequential fabrication process, encapsulating the sensor with PDMS, and a multi-step functionalization strategy. Reproduced from ref. 145 with permission from MDPI, copyright 2020. (B) Comprehensive visualization of the vitamin D₃ lateral flow assay includes the strip design and a participant's intensity profile indicating low vitamin D₃ and elevated T/C ratio. Reproduced from ref. 146 with permission from Nature, copyright 2017. (C) Visual depiction of the vitamin B₁₂ lateral flow assay, showcasing the specialized test strip design, competitive interactions at the test line, and signal intensity variations indicating high and low B₁₂ levels. Reproduced from ref. 147 with permission from Nature, copyright 2016. (D) Detection of vitamin C in sweat through a three-electrode system, involving simultaneous stimulation and detection, a biosensing protocol based on sweat stimulation before and after vitamin C intake, and a sweat-based enzymatic reaction for accurate vitamin C measurement. (E) Development of a vitamin C biosensor featuring hydrogel and enzyme layers, and visualization of the epidermal sensor's mechanical strain response (twisting). Reproduced from ref. 148 with permission from American Chemical Society, copyright 2020.



levels) has high-frequency occurrence in the developing world and it has substantial health and economic costs. Anaemic children are more susceptible to upper respiratory tract infections than healthy children.¹⁴³ ID increases the risk of preterm labour in the first two trimesters of pregnancy, low birth weight, and infant mortality.¹⁴⁴

Estimating iron deficiency anemia based on hemoglobin concentration alone fails to account for nutritional deficiencies, infectious disorders, *etc.* A combination of indicators best assesses iron status. However, ferritin is primarily accepted as the iron-storage molecule and an increase in iron supplies in cells increases the production of serum ferritin. So, serum ferritin is a significant biomarker for iron status as it correlates with relative total body iron reserve. Nevertheless, the concentration of ferritin elevates in the presence of infection or inflammation. The normal range of ferritin in serum is between 15 ng mL⁻¹ (32 pmol L⁻¹) and 150 ng mL⁻¹ (316 pmol L⁻¹) for females or 200 ng mL⁻¹ (421 pmol L⁻¹) for males, while the cut-off for depleted iron storage in children less than 5 years old is <12 ng mL⁻¹ (26 pmol L⁻¹) and <30 ng mL⁻¹ (63 pmol L⁻¹) if inflammation is present.¹³⁶ In Fig. 8 graphene-based field-effect transistor (GFET) biosensors are developed in a step-by-step process as a ferritin specific biosensing platform. To start, a GFET device is fabricated using standard lithography and etching techniques. Then, a polydimethylsiloxane (PDMS) well is placed on top of the GFET device to secure an electrolyte. After that, the graphene layer is functionalized with pyrene-butanoic acid, succinimidyl ester (PASE) (Fig. 8A). This enables the anti-ferritin antibody to attach to the graphene, which is the next step in the process. Finally, the GFET-based biosensor is capable of ferritin-specific biosensing after the completion of these steps.¹⁴⁵

Lu *et al.*¹³⁶ have reported a rapid diagnostic test for quantifying ferritin, RBP, and CRP simultaneously. It detects ferritin concentrations as low as 15 ng mL⁻¹. Oshin *et al.*¹⁴⁵ demonstrated a graphene-based biosensor with a ferritin detection range of 5.3 ng L⁻¹ to 0.5 µg L⁻¹.

5.2.3. Vitamin D deficiency. Vitamin D belongs to a group of fat-soluble secosteroids responsible for the absorption of calcium, phosphate, and zinc in the body. Vitamin D is best known for preventing bone disease, rickets in growing children, and osteomalacia (soft bone) in adults. The clinical level for vitamin D deficiency is less than 50 nmol L⁻¹, a value between 50 and 75 nmol L⁻¹ is insufficient and a vitamin D level greater than 75 nmol L⁻¹ is considered healthy.¹⁴⁶ Vitamin D levels have been linked to depression, Parkinson and Alzheimer's disease.^{149,150} Also, deficient vitamin D levels in pregnant women lead to pre-eclampsia, gestational diabetes, and adverse pregnancy outcomes.

25-Hydroxyvitamin D or 25(OH)D is a pro-hormone produced in the liver and is a biologically inert form of vitamin D that is produced in the skin. 25(OH)D is a major circulating form of vitamin D, and it can be considered as the best indicator of vitamin D status.¹⁴⁶ Total 25(OH)D is a measure of 25(OH)D₂ (ergocalciferol) and 25(OH)D₃

(cholecalciferol). However, studies suggest that 25(OH)D₃ supplementation alone can lead to an overall increase in vitamin D status in the body.¹⁵¹

Vemulapati, S. *et al.* have created a fast diagnostic test designed for precise and quantitative evaluation of vitamin D levels. The test specifically targets the measurement of 25(OH)D₃, which is the primary component of circulating 25(OH)D and is particularly significant in the deficient range (<50 nmol L⁻¹), Fig. 8C. This range is essential as it represents the area with the highest demand for accurate assessment.

Peter *et al.*¹⁵² developed a fully automated microfluidic assay along with a readout for measuring vitamin D levels. The lab-on-a-chip used a microarray-based competitive immunoassay to detect 25(OH)D₃. Lee *et al.*¹⁴⁷ presented the vitaAID – vitamin AuNP-based immunoassay device – a system for rapid quantitation of vitamin D levels on a smartphone. The device compares the differences in brightness between the detection area and the reference area and estimates the concentration of 25(OH)D independent of the exposure setting. A rapid diagnostic test integrated as a lateral flow assay was developed by Vemulapati *et al.*¹⁴⁶ incorporating competitive type architecture capable of processing blood on the strip and quantifying 25(OH)D₃ in the deficient range.

5.2.4. Vitamin B₁₂ deficiency. Vitamin B₁₂, also known as cobalamin, is an essential factor in DNA synthesis and neurologic functions. This vitamin is only synthesized by microorganisms and not by plants. Thus the only source of vitamin B₁₂ in our diet is the flesh of other animals. Vitamin B₁₂ is bound to food enzymes and released by gastric enzymes and acid and again bound to a protein that is taken up in the distal ileum. Pernicious anemia, a human autoimmune disease, is caused by loss of secretion of the protein which causes fatal vitamin B₁₂ deficiency. Low vitamin B₁₂ status also leads to several acute and chronic conditions such as anemia, paraesthesia, and cognitive impairments.^{153,154} Diagnosis of vitamin B₁₂ deficiency is correlated to low serum B₁₂ levels. The clinical level of deficiency is <100 pg mL⁻¹ (<73.8 pmol L⁻¹).¹⁵⁵ Along with serum vitamin B₁₂, methylmalonic acid, and total homocysteine values are also compared with vitamin B₁₂ levels in patients and are often elevated prior to any clinical abnormalities.

Lee *et al.*¹⁴⁷ developed a Nutriphone which consists of a smartphone accessory and a lateral flow test strip (Fig. 8C) that quantifies deficiency of vitamin B₁₂. Silver enhancement of the initial colorimetric signal and quantification with the app and smartphone detected the lower range of clinical serum vitamin B₁₂ levels.

5.2.5. Vitamin C deficiency. Vitamin C, scientifically known as L-ascorbic acid (AA), is an essential water-soluble nutrient crucial for the human body's proper growth and functioning. This vitamin plays a vital role in numerous bodily functions.¹⁵⁶ It is involved in synthesizing collagen, which aids in wound healing and promotes healthy skin.



Additionally, vitamin C facilitates iron absorption, helps treat colds, and strengthens the immune system to defend against viral infections and neurological disorders. Furthermore, it contributes to the well-being of the cardiovascular system, gums and even assists in combating cancer. Accurately measuring vitamin C levels in biological fluids is significant as it helps prevent imbalances and reduces the risks associated with various disorders and diseases.^{148,156–158} An enzyme-based biosensor is one approach to use an enzyme called ascorbate oxidase (AOx), which specifically reacts with vitamin C (Fig. 8D). This enzyme catalyzes the oxidation of ascorbic acid, producing dehydroascorbic acid and hydrogen peroxide. The generated hydrogen peroxide can be quantitatively measured to determine the concentration of vitamin C (Fig. 8D). The biosensor system typically consists of three components: the enzyme (AOx), an electrode surface, and a transducer (Fig. 8E). The enzyme is immobilized onto the electrode surface, and when vitamin C is present in the sample, it reacts with the immobilized AOx. This reaction produces hydrogen peroxide, which can be detected using electrochemical techniques such as amperometry or potentiometry.¹⁴⁸

5.2.6. Vitamin K deficiency. Vitamin K, famously known as a blood coagulation vitamin, functions as a cofactor for a single microsomal enzyme, γ -carboxy glutamyl carboxylase (GGCX). Many vitamin K dependent proteins (Gla proteins) are found in many cell and tissue types. This vitamin is a clinical anticoagulant and rodenticide. Vitamin K deficiency symptoms include spontaneous cutaneous purpura, epistaxis, gastrointestinal, genitourinary, or other bleeding.^{159–161}

Vitamin K is the most lipophilic and least abundant vitamin. Measurement is a challenging task in human samples. Uncarboxylated vitamin K proteins, factor VII, uncarboxylated osteocalcin, and urinary concentration of Gla residues are some of the markers for vitamin K depletion.¹⁶²

Li *et al.* developed an eco-friendly, rapid, and sensitive detection of anticoagulant rodenticides (ARs) in emergency clinics and food samples with a point-of-care assay. The developed QNs were prepared by encapsulating Cd-free InP/ZnS QDs into PS nanospheres. The obtained QNs have good potential in clinical diagnosis and food safety monitoring.¹⁶³

5.3. Novel biomarker as a pharmacological response to diseases

Capturing target genes in microwells with a small sample volume can improve sensitivity. Furthermore, gene quantification is now attainable with the advancement of considerable sample volume minimization employing droplet technology. These cost, automation, usability, and portability advancements have enabled point-of-care applications to decentralize testing platforms from laboratory-based settings to field usage against infections.¹⁶⁴ Biosensors are critical for illness diagnosis at the point of care or bedside. The key to effective illness therapy is early detection mainly in infectious diseases,

cardiovascular disease, neurological disorders, and so on which are as important in this regard as these diseases are responsible for a high incidence of morbidity and death globally.¹⁶⁵ In the case of biosensors electrochemical, optical, piezoelectric, or nucleic acid biosensors have been developed. Most biosensors function as immunosensors, detecting immunological markers in biological samples. These ultrasensitive instruments eliminate the time-consuming and arduous stages of traditional sample processing and analysis. Biosensors provide an accurate diagnosis with high specificity and sensitivity and a better prognosis.¹⁶⁶ Transcriptome, metabolomic, and proteomic (including miRNA and single-cell T cell) profiling yield new biomarkers. To analyze and integrate big datasets and guide data interpretation, advanced computational approaches are necessary. Longitudinal sample analysis will be critical in future studies, first for identification and, more crucially, validation of chosen sets of putative biomarkers.¹⁶⁷ Wu, Tsung-Hsien, *et al.* demonstrated an extensive microfluidic-based dual-target detection method for RF and anti-CCP. This research explores practical parallel processing and prospective uses for point-of-care rheumatoid arthritis diagnosis and monitoring. This study lays the path for improved rapid diagnostic capabilities in the management of rheumatoid arthritis by combining cutting-edge microfluidic technology with sensitive ELISA tests.³ Lin, Xiaodong, *et al.* incorporated a lanthanide-based ratiometric fluorescent probe within a self-designed Fermat spiral microfluidic chip that enables rapid and sensitive automated determination of bacterial endospore biomarker, dipicolinic acid (DPA), emphasizing the potential for streamlined and efficient point-of-care testing applications.¹⁶⁸ Pt-Pd bimetallic nanoclusters are used by Chen, Chaozhan, *et al.* in a novel microfluidic device with mass-producible micropillar array electrodes for enhanced biomarker detection sensitivity, as demonstrated by notable improvements in H_2O_2 detection and high-sensitivity biosensing of glucose, uric acid, and sarcosine in human serum.¹⁶⁹ A novel technique for early clinical sepsis detection was developed by Yin, Binfeng, *et al.*, using commercially available 3M tapes inside a proprietary microfluidic device for automated and sensitive analysis of sepsis-related biomarkers in serum. As a promising platform for accurate clinical and field diagnostics, the tape-based microfluidic chip exhibits high sensitivity and selectivity in detecting C-reactive protein (CRP) with a limit of detection (LOD) of 0.23 g mL^{-1} , procalcitonin (PCT) at 0.14 ng mL^{-1} , and interleukin-6 (IL-6) at 12.53 pg mL^{-1} .¹⁷⁰ The study introduces an innovative microfluidic system capable of automatically detecting alpha defensin HNP-1 and C-reactive protein biomarkers from synovial fluid, demonstrating high specificity and accurate diagnosis potential for post-arthroplasty periprosthetic joint infections.¹⁷¹ This section focuses on improvements in microfluidic-based biosensors for point-of-care diagnosis and monitoring of various diseases. It illustrates cutting-



edge methods like dual-target detection, lanthanide-based fluorescent probes, bimetallic nanoclusters, and commercially available tapes, all of which show enhanced sensitivity and specificity in detecting biomarkers, particularly in sepsis and post-arthroplasty periprosthetic joint infections.

6. Evolution of POCT devices

6.1. How it started

Dr. Kost began researching biosensors and measuring pH changes *in vivo* during cardiac stress and shock in 1972. In innovative visual logistics depicting clinical pathophysiology, he conceived the new area of patient-focused testing.¹⁷² Earlier in 1955, hematological testing instructions in England described on-site patient treatment. The phrase “point-of-care testing” was created by Dr. Kost in the early 1980s while

doing biosensor-based serial measurements of ionized calcium in whole blood at the bedside of patients undergoing liver transplantation at the University of California, Davis Medical Center.¹⁷³ Dr. Kost personally performed a whole-blood analysis at the patient's side in the operating room and instantly conveyed the results to the anesthesia and surgical teams, who acted on them. The pathologic dissociation of ionized calcium from total calcium due to the chelating effects of extensive blood transfusion required to sustain patient vascular volume in the early days of liver transplantation was an important finding at the time.¹⁷⁴

6.2. New platforms

The recently faced global pandemic has made everyone realize the importance of patient-oriented technology which reduces and distributes the burden on hospitals. Care and

Table 1 Biomarkers for various diseases and their detection sensitivities

Disease name	Biomarker	Sensitivity and specificity	LOD	Remark	Ref.
Rheumatoid arthritis	Rheumatoid factor (RF), anti-citrullinated peptide antibody (ACPA), and the newly developed anti-mutated citrullinated vimentin (anti-MCV)	Anti-MCV POCT – 64% and 97% RF POCT – 51% and 95%	—		180
Graves' disease	Hormone thyroxine (T4)/l-thyroxine	—	$3.0 \times 10^{-9} \text{ mol L}^{-1}$		181
Multiple sclerosis	Anti-myelin basic protein (MBP) autoantibody	—	Gelatin MBP biosensor – $0.1528 \text{ ng mL}^{-1}$ Gelatin TiO_2 MBP – $0.1495 \text{ ng mL}^{-1}$		182
Multiple sclerosis	Matrix metalloproteinase 9 (MMP-9)	—	15 ng mL^{-1}		183
Type 1 diabetes	Glucose and lactate	Glucose biosensor – $0.33 \mu\text{A cm}^{-2} \text{ mM}^{-1}$ Lactate biosensor – $0.78 \mu\text{A cm}^{-2} \text{ mM}^{-1}$	Glucose biosensor – 0.18 mM Lactate biosensor – 0.12 mM		118
Type 1 diabetes	Glucose	—	Fluorescence spectroscopy – $0.14 \mu\text{M}$ UV-vis spectroscopy – $3.26 \mu\text{M}$		184
Addison's disease	Cortisol – a glucocorticoid hormone	$0.25 \text{ ohm ng}^{-1} \text{ mL}^{-1}$	1 pg mL^{-1}		185
Addison's disease	Cortisol	—	$0.87 \pm 0.12 \text{ pg mL}^{-1}$	Electrochemical-digital immunosensor – revealed an accuracy of 96.3% in comparison with ELISA	186
Acute nutritional deficiencies					
Vitamin A deficiency	Retinol binding protein	Sensitivity – 100% Specificity – 100%	$2.2 \mu\text{g mL}^{-1}$	Multiplexed fluorescence test strip and mobile enable platform as reader	136
Iron deficiency (ID)	Soluble transferrin receptor (sTfr)	Sensitivity – 100% Specificity – 83.3%	$6 \mu\text{g mL}^{-1}$	An immunochromatographic assay-based POC screening device	187
	Serum ferritin	Sensitivity – 90% Specificity – 100%	$18 \mu\text{g L}^{-1}$		188
Vitamin D deficiency	$25(\text{OH})\text{D}_3$ (vitamin D_3)	Sensitivity – $0.90 \Omega \text{ nM}^{-1} \text{ mm}^{-2}$	0.28 ng mL^{-1}	Response time <1 min	189
Vitamin B ₁₂ deficiency	Label-free immunosensor	—	2.36 ng mL^{-1}		190
	Vitamin B ₁₂	Sensitivity – 87% Specificity – 100%	258 pmol L^{-1}		147
Vitamin K deficiency	Vitamin B ₁₂ Anticoagulant rodenticides (ARs)	—	0.109 nM		191 163



treatment can be given to those who are ill or critically ill.¹⁷⁵ Over the last few decades, point-of-care testing has shown itself in epidemics, disasters, crises, hospital surgical practice, critical care, bedside decision-making, personal health maintenance, and self-testing. POCT technology briefly has two essential parts – one gives a visual conclusion for qualitative data and the other includes a physicochemical component for quantitative inference.¹⁷⁶ Going for only a visual inference such as in LFA gives us a quick result but it cannot be favorable for continuous monitoring in near-patient situations. Conversely, a read-out modality attached to the POCT device gives a better interpretation of the illness and keeps the patient in check. For example, a patient who monitors his/her glucose level at home using a glucose meter can check the blood glucose levels and consult a doctor in case of urgency.¹⁷⁷

Recent developments have also led to multiplexing of analyte testing on a single platform using the same specimen. Point-of-careology results from the evolution of POCT and the POC specialists that do it across the world. In each environment and nation, point-of-careologists will create standards of care, the test reference values, and best practices.¹⁷⁸ The development of point-of-careology reflects improvements in contemporary medical, technology, and social expectations in general, mostly in tandem with the introduction of cell phones and other convenient gadgets that individuals may quickly obtain. There are improvements in the marginal value of testing and time savings in proportion to the severity of the medical crisis. New testing modalities on ambulances, airplanes, ships, and other mobile carriers provide simplicity and speed. Reliable results eliminate specimen time obsolescence when transported or stored.^{178,179}

7. Conclusion

The future of personalized medicine will depend heavily on biomarkers because of their enormous potential to benefit society and the healthcare field. The biomarkers can provide valuable information about the state of our health, aid in early disease detection, guide treatment decisions, and monitor the effectiveness of therapies. Several microfluidic platforms including different sample detection optical and electrochemical techniques have been developed for rapid detection of biomarkers of various diseases (Table 1). Microfluidics technology provides a great ability to execute and analyze complex operations which include detection, drug delivery, synthesis, *etc.* The high surface-to-volume ratio can improve reliability and result in shorter trial times. The use of this state-of-the-art technology has provided the opportunity to perform experiments using small volumes which results in a reduction of reagent volume and low energy consumption. The advent of biomarkers to help detect diseases and deficiencies has led us to a path where we can stop future pandemics. Different response-analyses such as

optical analysis, electrochemical analysis, *etc.* aid the POCT technology for faster and more accurate results.

Although there are many challenges, integrating biomarkers with microfluidics technology has great potential. Encouraging multidisciplinary collaboration among biologists, engineers, and doctors to build and improve microfluidic devices is the main problem. Another challenge is ensuring the durability and robustness of these devices in practical applications, which requires careful testing and validation. For wider acceptance, it is essential to address the problem of the low throughput that microfluidic devices have under the complex channel design. A possible solution, enhancing the capabilities of microfluidics and allowing additional functionalities, is the merging of microfluidics with 3D printing. Additionally, despite significant efforts, only a few biomarkers find therapeutic use, making identifying and using them challenging. The abilities of a suitable, functional POC device must meet the “ASSURED” criteria, which calls for the device to have fast detection, high accuracy, a low detection limit, low cost, good repeatability and stability, and user-friendly characteristics. Microfluidic technology may overcome these difficulties, especially when resolving challenges associated with biomarker specificity, sensitivity, and validation methods. Microfluidics' ability to multiplex allows the technology to increase accuracy by combining several indicators for different biomarkers' analysis. Consequently, even though biomarker-integrated microfluidics has incredible promise, it requires a collaborative effort to overcome its challenges and fully achieve the benefits in real-world applications.

The study investigates a state-of-the-art bioanalytical method that integrates microfluidic devices with advanced optical and electrochemical methods. Precise diagnoses are made possible by this ground-breaking combination of scientific techniques, including electrical impedance spectroscopy, cyclic voltammetry, and amperometry. By correctly identifying autoimmune conditions such as rheumatoid arthritis, multiple sclerosis, type 1 diabetes, and Addison's disease, the approach establishes its importance and enables earlier treatments and better management. Evaluating iron and essential vitamins also plays a crucial part in recognizing acute nutritional shortages and allowing targeted therapies. The method also provides insights into indicators of pharmacological response, increasing personalized treatments and accelerating pharmaceutical research. As we reflect on the evolution of point-of-care testing devices, it is evident that we stand at the threshold of a transformative era in healthcare diagnostics characterized by innovation, accessibility, and improved patient care.

In the future of healthcare, the rapidly emerging science of bioanalysis has enormous potential. The potential for changing diagnoses and treatments has been demonstrated with the development of miniaturized diagnostic tools, wearable microfluidics devices, multi-analyte detection systems, AI integration, improved sensor materials,



smartphone-enabled diagnostics, and standardized platforms.

The growing number of novel applications and advancements has facilitated the integration of enhanced health monitoring capabilities into cell phones. Several smartphones can now conduct sophisticated immunoassays, similar to those often performed in laboratory settings, frequently using additional accessories. The integration of advancements in smartphone technology with microfluidic devices has the potential to significantly revolutionize the healthcare field, particularly in mobile health (mHealth) and disease monitoring. This innovation has the potential to enhance accessibility by enabling smartphone users to access laboratory-like diagnostic capabilities. These developments enable people to take control of their health by providing real-time information, early diagnosis, and personalized therapies. Precision, accessibility, and better medical outcomes will be at the forefront of healthcare in the future as these technologies continue to develop and integrate.

The next phase of microfluidic devices involves novel methodologies for sample collection and preparation, reagent storage, and the development of complete lab-on-a-chip systems. The significance of on-chip sample preparation is often disregarded since most devices available on the market have partial on-chip analysis capabilities. To achieve widespread acceptance among healthcare professionals, it is essential for microfluidic platforms to effectively tackle real-world challenges and substantiate the durability and benefits of microfluidic biochips compared to conventional approaches. Anticipated advancements are foreseen due to the collaborative efforts of the microfluidic lab-on-a-chip community, specialists in biology, and clinical practitioners.

Conflicts of interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence this paper.

Acknowledgements

The CSIR-UGC Junior Research Fellowship greatly assisted NK in his research endeavors, for which we acknowledge and are deeply grateful. We sincerely thank DBT-BIRAC for their invaluable assistance and contributions to MK academic endeavor. PPB is thankful for the fellowship provided by the MHRD and IIT Guwahati.

References

- 1 S. Preetam, B. K. Nahak, S. Patra, D. C. Toncu, S. Park, M. Syväjärvi, G. Orive and A. Tiwari, *Biosens. Bioelectron.*, 2022, **10**, 100–106.
- 2 N. T. Nguyen, M. Hejazian, C. H. Ooi and N. Kashaninejad, *Micromachines*, 2017, **8**, 186.
- 3 T. H. Wu, Y. C. Tsai, F. C. Kuo, M. S. Lee, C. C. Hu and G. Bin Lee, *Sens. Actuators, B*, 2023, **383**, 133587.
- 4 S. Nahavandi, S. Baratchi, R. Soffe, S. Y. Tang, S. Nahavandi, A. Mitchell and K. Khoshmanesh, *Lab Chip*, 2014, **14**, 1496–1514.
- 5 K. Yamamoto, N. Ota and Y. Tanaka, *Anal. Chem.*, 2021, **93**, 332–349.
- 6 J. Hong, J. B. Edel and A. J. deMello, *Drug Discovery Today*, 2009, **14**, 134–146.
- 7 P. Wang, L. Ge, M. Yan, X. Song, S. Ge and J. Yu, *Biosens. Bioelectron.*, 2012, **32**, 238–243.
- 8 N. Kumar, M. Kumari and R. K. Arun, in *Miniaturized Biosensing Devices*, Springer Nature Singapore, Singapore, 2022, pp. 99–122.
- 9 A. Sharma, M. Badea, S. Tiwari and J. L. Marty, *Molecules*, 2021, **26**, 1–32.
- 10 H. Lee, T. K. Choi, Y. B. Lee, H. R. Cho, R. Ghaffari, L. Wang, H. J. Choi, T. D. Chung, N. Lu, T. Hyeon, S. H. Choi and D. H. Kim, *Nat. Nanotechnol.*, 2016, **11**, 566–572.
- 11 N. P. Shetti, S. D. Bukkigar, K. R. Reddy, C. V. Reddy and T. M. Aminabhavi, *Colloids Surf., B*, 2019, **178**, 385–394.
- 12 I. R. Suhito, K. M. Koo and T. H. Kim, *Biomedicines*, 2021, **9**, 1–20.
- 13 Y. Xiang, C. Hu, G. Wu, S. Xu and Y. Li, *TrAC, Trends Anal. Chem.*, 2023, **158**, 116835.
- 14 P. Malik, V. Katyal, V. Malik, A. Asatkar, G. Inwati and T. K. Mukherjee, *ISRN Nanomater.*, 2013, **2013**, 1–9.
- 15 M. Kumari, V. Gupta, N. Kumar and R. Kumar, *Mol. Biotechnol.*, 2023, 1–24.
- 16 X. L. Sanjay, T. Sharma, G. Fu, M. Dou, F. Xu, R. Liu and H. Qi, *Analyst*, 2015, **140**, 7062–7081.
- 17 B. Yin, X. Wan, A. S. M. M. F. Sohan and L. Xiaodong, *Micromachines*, 2022, **13**(8), 12–38.
- 18 A. Ganguli, A. Mostafa, J. Berger, M. Y. Aydin, F. Sun, S. A. Stewart de Ramirez, E. Valera, B. T. Cunningham, W. P. King and R. Bashir, *Proc. Natl. Acad. Sci. U. S. A.*, 2020, **117**, 22727–22735.
- 19 K. R. Mitchell, J. E. Esene and A. T. Woolley, *Anal. Bioanal. Chem.*, 2022, **414**, 167–180.
- 20 J. Berthier, Tech. Proc. 2013 NSTI Nanotechnol. Conf. Expo, NSTI-Nanotech 2013, 2013, 2, pp. 342–345.
- 21 E. K. Sackmann, A. L. Fulton and D. J. Beebe, *Nature*, 2014, **507**, 181–189.
- 22 G. Luka, A. Ahmadi, H. Najjaran, E. Alocilja, M. Derosa, K. Wolthers, A. Malki, H. Aziz, A. Althani and M. Hoorfar, *Sensors*, 2015, **15**, 30011–30031.
- 23 Z. Li, Y. Bai, M. You, J. Hu, C. Yao, L. Cao and F. Xu, *Biosens. Bioelectron.*, 2021, **177**, 112952.
- 24 W. Su, X. Gao, L. Jiang and J. Qin, *J. Chromatogr. A*, 2015, **1377**, 13–26.
- 25 D. Migliozi, B. Pelz, D. G. Dupouy, A. L. Leblond, A. Soltermann and M. A. M. Gijs, *Microsyst. Nanoeng.*, 2019, **5**, 59.
- 26 C. Song, D. Gao, T. Yuan, Y. Chen, L. Liu, X. Chen and Y. Jiang, *Chin. Chem. Lett.*, 2019, **30**, 1038–1042.
- 27 W. Xing, Y. Liu, H. Wang, S. Li, Y. Lin, L. Chen, Y. Zhao, S. Chao, X. Huang, S. Ge, T. Deng, T. Zhao, B. Li, H. Wang, L. Wang, Y. Song, R. Jin, J. He, X. Zhao, P. Liu, W. Li and J. Cheng, *Engineering*, 2020, **6**, 1130–1140.



- 28 P. D. Sinawang, F. Soto, M. O. Ozen, D. Akin and U. Demirci, *Prog. Biomed. Eng.*, 2021, **3**, 043001.
- 29 A. M. Urdea, L. A. Penny, S. S. Olmsted, M. Y. Giovanni, P. Kaspar, A. Shepherd, P. Wilson, C. A. Dahl, S. Buchsbaum, G. Moeller and D. C. H. Burgess, *Nature*, 2006, **444**, 73–79.
- 30 S. Gupta, A. Venkatesh, S. Ray and S. Srivastava, *Biochim. Biophys. Acta, Proteins Proteomics*, 2014, **1844**, 899–908.
- 31 J. Andrus, Fabrication of semiconductor devices, *US Pat.*, 3122817, 1964.
- 32 L. Rayleigh, *Proc. R. Soc. London*, 1879, **29**, 71–97.
- 33 M. G. Pollack, A. D. Shenderov and R. B. Fair, *Lab Chip*, 2002, **2**, 96–101.
- 34 Y. Hou, C. C. Lv, Y. L. Guo, X. H. Ma, W. Liu, Y. Jin, B. X. Li, M. Yang and S. Y. Yao, *J. Anal. Test.*, 2022, **6**, 247–273.
- 35 S. N. Bhatia and D. E. Ingber, *Nat. Biotechnol.*, 2014, **32**, 760–772.
- 36 G. M. Whitesides, *Nature*, 2006, **442**, 368–373.
- 37 J. Cottet and P. Renaud, in *Drug Delivery Devices and Therapeutic Systems*, Elsevier, 2021, pp. 3–17.
- 38 Z. Liao, J. Wang, P. Zhang, Y. Zhang, Y. Miao, S. Gao, Y. Deng and L. Geng, *Biosens. Bioelectron.*, 2018, **121**, 272–280.
- 39 D. Rodrigues, A. I. Barbosa, R. Rebelo, I. K. Kwon, R. L. Reis and V. M. Correlo, *Biosensors*, 2020, **10**, 79.
- 40 Y. Abbas, J. Miwa, R. Zengerle and F. von Stetten, *Micromachines*, 2013, **4**, 80–89.
- 41 H. Bruus, *Theoretical Microfluidics*, 1st edn, 2008.
- 42 R. W. Fox and A. T. McDonald, *Introduction to Fluid Mechanics*, Wiley, New York, 1985.
- 43 C. Kleinstreuer, *Microfluidics and Nanofluidics: Theory and Selected Applications*, Wiley, 2014, vol. 7.
- 44 S. K. Mitra and S. Chakraborty, *Microfluidics and Nanofluidics Handbook-Fabrication, Implementation, and Applications*, 2012.
- 45 L. Lin, *Micro/Nano fluidics Enabled Single-Cell Biochemical Analysis*, 2019.
- 46 E. O. Ige, R. K. Arun, P. Singh, M. Gope, R. Saha, N. Chanda and S. Chakraborty, *Microfluid. Nanofluid.*, 2019, **23**, 1–8.
- 47 C. Carrell, A. Kava, M. Nguyen, R. Menger, Z. Munshi, Z. Call, M. Nussbaum and C. Henry, *Microelectron. Eng.*, 2019, **206**, 45–54.
- 48 L. Chen, C. Yang, Y. Xiao, X. Yan, L. Hu, M. Eggersdorfer, D. Chen, D. A. Weitz and F. Ye, *Mater. Today Nano*, 2021, **16**, 100136.
- 49 L. Chen, C. Yang, Y. Xiao, X. Yan, L. Hu, M. Eggersdorfer, D. Chen, D. A. Weitz and F. Ye, *Mater. Today Nano*, 2021, **16**, 100–136.
- 50 S. Kumar, S. Kumar, M. A. Ali, P. Anand, V. V. Agrawal, R. John, S. Maji and B. D. Malhotra, *Biotechnol. J.*, 2013, **11**, 1267–1279.
- 51 W. Sparreboom, A. Van Den Berg and J. C. T. Eijkel, *Nat. Nanotechnol.*, 2009, **4**, 713–720.
- 52 S. Bayda, M. Adeel, T. Tuccinardi, M. Cordani and F. Rizzolio, *Molecules*, 2019, **25**, 112.
- 53 D. G. Haywood, A. Saha-shah, L. A. Baker and S. C. Jacobson, *Anal. Chem.*, 2014, **87**, 172–187.
- 54 H. Foster, M. Wade, J. England, J. Greenman and V. Green, *Organs-on-a-Chip*, 2021, **3**, 100011.
- 55 Y. Ai, R. Xie, J. Xiong and Q. Liang, *Small*, 2020, **16**, 1–24.
- 56 B. Y. Xia, P. Yang, Y. Sun, Y. Wu, B. Mayers, B. Gates, Y. Yin, F. Kim and H. Yan, *Adv. Mater.*, 2003, **15**, 353–389.
- 57 D. Zhang, S. Wei and Z. Guo, *MRS Online Proc. Libr.*, 2009, **1240**, 1006.
- 58 K. E. Bates and H. Lu, *Biophys. J.*, 2016, **110**, 1684–1697.
- 59 K. Y. Lien, C. J. Liu, Y. C. Lin, P. L. Kuo and G. Bin Lee, *Microfluid. Nanofluid.*, 2009, **6**, 539–555.
- 60 O. M. Ikumapayi, E. T. Akinlabi, A. O. M. Adeoye and S. O. Fatoba, *Mater. Today: Proc.*, 2021, **44**, 1154–1162.
- 61 K. S. Prasad, X. Cao, N. Gao, Q. Jin, S. T. Sanjay, G. Henao-Pabon and X. J. Li, *Sens. Actuators, B*, 2020, **305**, 127516.
- 62 Y. T. Chen, Y. C. Lee, Y. H. Lai, J. C. Lim, N. T. Huang, C. T. Lin and J. J. Huang, *Biosensors*, 2020, **10**, 1–22.
- 63 A. Tay, A. Pavesi, S. R. Yazdi, C. T. Lim and M. E. Warkiani, *Biotechnol. Adv.*, 2016, **34**, 404–421.
- 64 K. Mahato, A. Srivastava and P. Chandra, *Biosens. Bioelectron.*, 2017, **96**, 246–259.
- 65 M. Xiao, F. Tian, X. Liu, Q. Zhou, J. Pan, Z. Luo and M. Yang, *Adv. Sci.*, 2022, **9**, 2105904.
- 66 B. Nasser, N. Soleimani, N. Rabiee, A. Kalbasi, M. Karimi and M. R. Hamblin, *Biosens. Bioelectron.*, 2018, **117**, 112–128.
- 67 R. Rajendran and G. Rayman, *J. Diabetes Sci. Technol.*, 2014, **8**, 1081–1090.
- 68 A. N. Konwar and V. Borse, *Sens. Int.*, 2020, **1**, 100015.
- 69 T. Gou, J. Hu, W. Wu, X. Ding, S. Zhou, W. Fang and Y. Mu, *Biosens. Bioelectron.*, 2018, **120**, 144–152.
- 70 L. Cao, X. Guo, P. Mao, Y. Ren, Z. Li, M. You, J. Hu, M. Tian, C. Yao, F. Li and F. Xu, *ACS Sens.*, 2021, **6**, 3564–3574.
- 71 U. Gupta, N. Kumar, A. Lata, P. Singh and R. K. Arun, *Int. J. Biol. Macromol.*, 2023, **241**, 124614.
- 72 J. R. Mejía-Salazar, K. R. Cruz, E. M. M. Vásquez and O. N. de Oliveira, *Sensors*, 2020, **20**, 1–19.
- 73 U. Gupta, V. Gupta, R. K. Arun and N. Chanda, *Biotechnol. Bioeng.*, 2022, **119**, 3393–3407.
- 74 G. P. Nikoleli, C. G. Siontorou, D. P. Nikolelis, S. Bratakou, S. Karapetis and N. Tzamtzis, *Nanotechnol. Biosens.*, 2018, 375–394.
- 75 R. Augustine, A. Hasan, S. Das, R. Ahmed, Y. Mori, T. Notomi, B. D. Kevadiya and A. S. Thakor, *Biology*, 2020, **9**, 1–17.
- 76 E. B. Bahadir and M. K. Sezgentürk, *Anal. Biochem.*, 2015, **478**, 107–120.
- 77 S. K. Sia, V. Linder, B. A. Parviz, A. Siegel and G. M. Whitesides, *Angew. Chem., Int. Ed.*, 2004, **43**, 498–502.
- 78 A. J. Rodriguez-Morales, J. A. Cardona-Ospina, E. Gutiérrez-Ocampo, R. Villamizar-Peña, Y. Holguin-Rivera, J. P. Escalera-Antezana, L. E. Alvarado-Arnez, D. K. Bonilla-Aldana, C. Franco-Paredes, A. F. Henao-Martinez, A. Paniz-Mondolfi, G. J. Lagos-Grisales, E. Ramírez-Vallejo, J. A. Suárez, L. I. Zambrano, W. E. Villamil-Gómez, G. J. Balbin-Ramon, A. A. Rabaan, H. Harapan, K. Dhama, H. Nishiura, H. Kataoka, T. Ahmad and R. Sah, *Travel Med. Infect. Dis.*, 2020, **34**, 101623.



- 79 M. H. Nawaz, A. Hayat, G. Catanante, U. Latif and J. L. Marty, *Anal. Chim. Acta*, 2018, **1026**, 1–7.
- 80 D. N. D'Ambrosio, R. D. Clugston and W. S. Blaner, *Nutrients*, 2011, **3**, 63–103.
- 81 J. Yang, K. Wang, H. Xu, W. Yan, Q. Jin and D. Cui, *Talanta*, 2019, **202**, 96–110.
- 82 Q. Ma, H. Ma, F. Xu, X. Wang and W. Sun, *Microsyst. Nanoeng.*, 2021, **7**, 19.
- 83 A. S. John and C. P. Price, *Clin. Biochem. Rev.*, 2014, **35**, 155–167.
- 84 A. W. Martinez, S. T. Phillips, E. Carrilho, S. W. Thomas, H. Sindi and G. M. Whitesides, *Anal. Chem.*, 2008, **80**, 3699–3707.
- 85 A. Pandya and V. Singh, *Micro/Nanofluidics and Lab-on-Chip Based Emerging Technologies for Biomedical and Translational Research Applications - Part A*, 1st edn, 2022.
- 86 C. Dincer, R. Bruch, A. Kling, P. S. Dittrich and G. A. Urban, *Trends Biotechnol.*, 2017, **35**, 728–742.
- 87 S. Chen and M. H. Shamsi, *J. Micromech. Microeng.*, 2017, **27**, 083001.
- 88 A. J. P. Martin and R. L. M. Synge, *Biochem. J.*, 1941, **35**(12), 1358.
- 89 F. A. Robinson, R. L. M. Synge, K. A. Williams and F. R. Cropper, *Analyst*, 1946, **71**(843), 251b–267.
- 90 G. Alarcón-Angeles, G. Arturo, G. Álvarez-Romero, G. Rosati, C. Parolo and A. Merkoçi, *Reference module in biomedical sciences*, 2022, pp. 772–788.
- 91 P. Damborský and J. Katrlík, *Essays Biochem.*, 2016, **60**(1), 91–100.
- 92 X. Fan, I. M. White, S. I. Shopova, H. Zhu, J. D. Suter and Y. Sun, *Anal. Chim. Acta*, 2008, **620**, 8–26.
- 93 J. Liu, J. Wang, Z. Li, H. Meng, L. Zhang, H. Wang, J. Li and L. Qu, *Microchim. Acta*, 2018, **185**, 110.
- 94 Y. Wang, H. Xu, J. Zhang and G. Li, *Sensors*, 2008, **8**, 2043–2081.
- 95 W. Jin and G. Maduraiveeran, *J. Anal. Sci. Technol.*, 2018, **9**, 18.
- 96 M. Pohanka, *Int. J. Anal. Chem.*, 2021, 1–9.
- 97 J. L. Hammond, N. Formisano, P. Estrela, S. Carrara and J. Tkac, *Essays Biochem.*, 2016, **60**(1), 69–80.
- 98 S. Teanphonkrang, A. Ernst, S. Janke, P. Chaiyen, J. Sucharitakul, W. Suginta, P. Khunkaewla, W. Schuhmann, A. Schulte and A. Ruff, *ACS Sens.*, 2019, **4**, 1270–1278.
- 99 M. Pohanka and P. Skladal, *J. Appl. Biomed.*, 2008, **6**, 57–64.
- 100 D. Grieshaber, R. MacKenzie, J. Vörös and E. Reimhult, *Sensors*, 2008, **8**, 1400–1458.
- 101 A. A. Almezizia, A. M. Naglah, L. S. Alrasheed, M. G. Alanazi, A. E.-G. E. Amr and A. H. Kamel, *RSC Adv.*, 2023, **13**, 15199–15207.
- 102 M. A. Akl, E. S. R. El-Gharkawy, N. A. El-Mahdy, S. M. El-Sheikh and S. M. Sheta, *Dalton Trans.*, 2020, **49**, 15769–15778.
- 103 H. Karimi-Maleh, Y. Orooji, F. Karimi, M. Alizadeh, M. Baghayeri, J. Rouhi, S. Tajik, H. Beitollahi, S. Agarwal, V. K. Gupta, S. Rajendran, A. Ayati, L. Fu, A. L. Sanati, B. Tanhaei, F. Sen, M. Shabani-nooshabadi, P. N. Asrami and A. Al-Othman, *Biosens. Bioelectron.*, 2021, **184**, 113252.
- 104 V. Climent and J. M. Feliu, in *Encyclopedia of Interfacial Chemistry*, Elsevier, 2018, pp. 48–74.
- 105 H. Bakhsh, I. M. Palabiyik, R. Kumar Oad, N. Qambrani, J. A. Buledi, A. R. Solangi, S. Tufail and H. Sherazi, *J. Electroanal. Chem.*, 2022, **9**, 116181.
- 106 P. Singh, S. Mandal, D. Roy and N. Chanda, *ACS Biomater. Sci. Eng.*, 2021, **7**, 3446–3458.
- 107 M. Grossi and B. Riccò, *J. Sens. Sens. Syst.*, 2017, **6**, 303–325.
- 108 A. Barhoum and R. J. Forster, *Anal. Chim. Acta*, 2022, **1225**, 340226.
- 109 E. B. Aydın, M. Aydın and M. K. Sezgintürk, *Macromol. Biosci.*, 2023, **23**, 1–13.
- 110 H. Oz, *Nutrients*, 2017, **9**, 1085.
- 111 S. Prasad, *Nanobiosens. Dis. Diagn.*, 2014, 1.
- 112 Y. Li, D. Li, P. Zhao, K. Nandakumar, L. Wang and Y. Song, *Micromachines*, 2020, **11**, 1–18.
- 113 P. Ranjan, A. Parihar, S. Jain, N. Kumar, C. Dhand, S. Murali, D. Mishra, S. K. Sanghi, J. P. Chaurasia, A. K. Srivastava and R. Khan, *Anal. Biochem.*, 2020, **610**, 113996.
- 114 B. Arévalo, V. Serafín, M. Garranzo-Asensio, R. Barderas, P. Yáñez-Sedeño, S. Campuzano and J. M. Pingarrón, *Biosens. Bioelectron.*, 2023, **13**, 100325.
- 115 Y. Song, B. Lin, T. Tian, X. Xu, W. Wang, Q. Ruan, J. Guo, Z. Zhu and C. Yang, *Anal. Chem.*, 2019, **91**, 388–404.
- 116 A. Sguassero, Á. Artiga, C. Morasso, R. R. Jimenez, R. M. Rapún, R. Mancuso, S. Agostini, A. Hernis, A. Abols, A. Linē, A. Gualerzi, S. Picciolini, M. Bedoni, M. Rovaris, F. Gramatica, J. M. de la Fuente and R. Vanna, *Anal. Bioanal. Chem.*, 2019, **411**, 1873–1885.
- 117 L. Zhang, G. Masetti, G. Colucci, M. Salvi, D. Covelli, A. Eckstein, U. Kaiser, M. S. Draman, I. Muller, M. Ludgate, L. Lucini and F. Biscarini, *Sci. Rep.*, 2018, **8**, 1–14.
- 118 A. Márquez, J. Aymerich, M. Dei, R. Rodríguez-Rodríguez, M. Vázquez-Carrera, J. Pizarro-Delgado, P. Giménez-Gómez, Á. Merlos, L. Terés, F. Serra-Graells, C. Jiménez-Jorquera, C. Domínguez and X. Muñoz-Berbel, *Biosens. Bioelectron.*, 2019, **136**, 38–46.
- 119 A. L. Mitchell and S. H. S. Pearce, *Nat. Rev. Endocrinol.*, 2012, **8**, 306–316.
- 120 H. E. Prince, *Biomarkers*, 2005, **10**(Suppl 1), 44–49.
- 121 F. Ghorbani, H. Abbaszadeh, A. Mehdizadeh, M. Ebrahimi-Warkiani, M. R. Rashidi and M. Yousefi, *Microchim. Acta*, 2019, **186**, 1–11.
- 122 A. T. Giannitsis, *Est. J. Eng.*, 2011, **17**, 109–139.
- 123 R. Villela and R. Correa, *Clin. Thyroidol.*, 2022, **34**, 147–149.
- 124 S. E. Gabriel and C. S. Crowson, Up to Date, 2018, p. 20181.
- 125 A. Mobed, S. Dolati, S. K. Shakouri, B. Eftekharsadat and B. Izadseresht, *Sens. Actuators, A*, 2021, **331**, 112975.
- 126 R. F. Standard, *HEUMATOID FACTOR (RF) ImmunoTurbidimetry 3rd Generation*, <https://www.sdi-fz.com/wp-content/uploads/2021/12/Rheumatoid-Factor-RF-Immunturbidimetry.pdf>, (accessed October 2021).



- 127 U. G. Longo, V. Candela, A. Berton, S. De Salvatore, S. Fioravanti, L. Giannone, A. Marchetti, M. G. De Marinis and V. Denaro, *Biosensors*, 2021, **11**, 31.
- 128 A. Alonso and M. A. Hernán, *Neurology*, 2008, **71**, 129–135.
- 129 M. I. Teixeira, M. H. Amaral, P. C. Costa, C. M. Lopes and D. A. Lamprou, *Pharmaceutics*, 2020, **12**, 1–37.
- 130 T. Ziemssen, K. Akgün and W. Brück, *J. Neuroinflammation*, 2019, **16**, 1–11.
- 131 B. E. Kerman, H. J. Kim, K. Padmanabhan, A. Mei, S. Georges, M. S. Joens, J. A. J. Fitzpatrick, R. Jappelli, K. J. Chandross, P. August and F. H. Gage, *Development*, 2015, **142**, 2213–2225.
- 132 M. Mobasser, M. Shirmohammadi, T. Amiri, N. Vahed, H. H. Fard and M. Ghojaziadeh, *Health Promot. Perspect.*, 2020, **10**, 98–115.
- 133 C. Chen, X. L. Zhao, Z. H. Li, Z. G. Zhu, S. H. Qian and A. J. Flewitt, *Sensors*, 2017, **17**, 1–19.
- 134 S. S. Poudyal, *PhD thesis*, South Dakota State University, 2017, p. 2156.
- 135 S. A. Tanumihardjo, *J. Nutr.*, 2016, **146**, 1816S–1848S.
- 136 Z. Lu, D. O'Dell, B. Srinivasan, E. Rey, R. Wang, S. Vemulapati, S. Mehta, D. Erickson and A. Sommer, *Proc. Natl. Acad. Sci. U. S. A.*, 2017, **114**, 13513–13518.
- 137 J. Hix, C. Martinez, I. Buchanan, J. Morgan, M. Tam and A. Shankar, *Am. J. Clin. Nutr.*, 2004, **79**, 93–98.
- 138 M. V. Gamble, R. Ramakrishnan, N. A. Palafox, K. Briand, L. Berglund and W. S. Blamer, *Am. J. Clin. Nutr.*, 2001, **73**, 594–601.
- 139 S. de Pee and O. Dary, *J. Nutr.*, 2002, **132**, 2895S–2901S.
- 140 N. I. Krinsky, *J. Nutr.*, 2004, **134**, 290–293.
- 141 V. P. Prakashan, G. Gejo, M. S. Sanu, M. S. Sajna, T. Subin, P. R. Biju and J. Cyriac, *Appl. Surf. Sci.*, 2019, **484**, 219–227.
- 142 World Health Organization, *Evaluating the public health significance of micronutrient malnutrition*, WHO, Geneva, Switzerland, 2009.
- 143 M. B. Zimmermann and R. F. Hurrell, *Lancet*, 2007, **370**, 511–520.
- 144 W. P. Goddard, I. Murray, R. G. Long, B. Scott, R. Barton, M. Salman, R. Frewin, D. Provan and A. Henson, *BMJ*, 1997, **314**, 1759.
- 145 O. Oshin, D. Kireev, H. Hlukhova, F. Idachaba, D. Akinwande and A. Atayero, *Sensors*, 2020, **20**, 3688.
- 146 S. Vemulapati, E. Rey, D. O'Dell, S. Mehta and D. Erickson, *Sci. Rep.*, 2017, **7**, 14142.
- 147 S. Lee, D. O'Dell, J. Hohenstein, S. Colt, S. Mehta and D. Erickson, *Sci. Rep.*, 2016, **6**, 1–8.
- 148 J. R. Sempionatto, A. A. Khorshed, A. Ahmed, A. N. De Loyola, E. Silva, A. Barfidokht, L. Yin, K. Y. Goud, M. A. Mohamed, E. Bailey, J. May, C. Aebischer, C. Chatelle and J. Wang, *ACS Sens.*, 2020, **5**, 1804–1813.
- 149 V. Ganji, C. Milone, M. M. Cody, F. McCarty and Y. T. Wang, *Int. Arch. Med.*, 2010, **3**, 29.
- 150 P. Knekt, A. Kilkkinen, H. Rissanen, J. Marniemi, K. Sääksjärvi and M. Heliövaara, *Arch. Neurol.*, 2010, **67**, 808–811.
- 151 L. R. Wilson, L. Tripkovic, K. H. Hart and S. A. Lanham-New, *Proc. Nutr. Soc.*, 2017, **76**, 392–399.
- 152 H. Peter, N. Bistolas, S. Schumacher, C. Laurisch, P. C. Guest, U. Höller and F. F. Bier, *Methods Mol. Biol.*, 2018, **1735**, 477–486.
- 153 F. Wokes, J. Badenoch and H. M. Sinclair, *Am. J. Clin. Nutr.*, 1955, **3**, 375–382.
- 154 E. Moore, A. Mander, D. Ames, R. Carne, K. Sanders and D. Watters, *Int. Psychogeriatr.*, 2012, **24**, 541–556.
- 155 S. P. Stabler and R. H. Allen, *Annu. Rev. Nutr.*, 2004, **24**, 299–326.
- 156 M. A. O. Dawood and S. Koshio, *Rev. Aquac.*, 2018, **10**, 334–350.
- 157 C. M. Nam, K. W. Oh, K. H. Lee, I. Suh, S. Y. Cho, W. H. Shim and S. H. Jee, *J. Am. Coll. Nutr.*, 2003, **22**, 372–378.
- 158 P. C. Calder, A. C. Carr, A. F. Gombart and M. Eggersdorfer, *Nutrients*, 2020, **12**, 1–3.
- 159 I. Cirilli, P. Orlando, S. Silvestri, F. Marcheggiani, P. V. Duda, N. Kaesler and L. Tian, *BioFactors*, 2022, **48**, 1129–1136.
- 160 D. Neutze and J. Roque, *Am. Fam. Physician*, 2016, **93**, 279–286.
- 161 R. E. Taylor and P. M. Blatt, *J. Am. Acad. Dermatol.*, 1981, **4**, 348–368.
- 162 P. Mladěnka, K. Macáková, L. Kujovská Krémová, L. Javorská, K. Mrštná, A. Carazo, M. Protti, F. Remião and L. Nováková, *Nutr. Rev.*, 2022, **80**, 677–698.
- 163 H. Li, B. Dong, L. Dou, W. Yu, X. Yu, K. Wen, Y. Ke, J. Shen and Z. Wang, *Sens. Actuators, B*, 2020, **324**, 128771.
- 164 A. I. Kathrada, S. C. Wei, Y. Xu, L. F. Cheow and C. H. Chen, *Biomechanics*, 2020, **14**, 1–10.
- 165 X. Huang, Y. Zhu and E. Kianfar, *J. Mater. Res. Technol.*, 2021, **12**, 1649–1672.
- 166 A. Parihar, S. Jain, D. S. Parihar, P. Ranjan and R. Khan, *Biosensor Based Advanced Cancer Diagnostics*, Academic Press, 2022, pp. 47–57.
- 167 C. Mathieu, R. Lahesmaa, E. Bonifacio, P. Achenbach and T. Tree, *Diabetologia*, 2018, **61**, 2252–2258.
- 168 X. Lin, H. Wu, S. Zeng, T. Peng, P. Zhang, X. Wan, Y. Lang, B. Zhang, Y. Jia, R. Shen and B. Yin, *Biosens. Bioelectron.*, 2023, **230**, 115283.
- 169 C. Chen, B. Ran, B. Liu, X. Liu, Z. Zhang, Y. Li, H. Li, M. Lan and Y. Zhu, *Anal. Chim. Acta*, 2023, **1272**, 341450.
- 170 B. Yin, C. Qian, X. Wan, A. S. M. Muhtasim Fuad Sohan and X. Lin, *Biosens. Bioelectron.*, 2022, **212**, 114429.
- 171 T. W. Chen, R. Gandotra, H. Y. Chang, M. S. Lee, F. C. Kuo and G. Bin Lee, *Anal. Chem.*, 2023, **95**, 7693–7701.
- 172 G. J. Kost, *Proceedings of the San Diego Biomedical Symposium*, Academic Press, 1977, vol. 16, pp. 25–33.
- 173 K. Hyde, S. M. Lewis, I. J. Mackie, R. M. Rowan, B. J. Bain, E. M. Matutes, M. F. Murphy, K. Shinton, A. Stephens, J. K. Wood, B. T. Colvin, S. J. Machin, T. Baglin, T. W. Barrowcliffe, M. Greaves, C. A. Ludlam, F. E. Preston, P. E. Rose and I. D. Walker, *Clin. Lab. Haematol.*, 1995, **17**, 301–310.



- 174 G. J. Kost, M. A. Jammal, R. E. Ward and A. M. Safwat, *Am. J. Clin. Pathol.*, 1986, **86**, 61–70.
- 175 T. Nguyen, D. Duong Bang and A. Wolff, *Micromachines*, 2019, 1–7.
- 176 N. Rabiee, M. Bagherzadeh, A. Ghasemi and H. Zare, *Int. J. Mol. Sci.*, 2020, **21**, 5126.
- 177 F. Di Nardo, M. Chiarello, S. Cavallera, C. Baggiani and L. Anfossi, *Sensors*, 2021, **21**, 5185.
- 178 X. Liu, X. Zhu, G. J. Kost, J. Liu, J. Huang and X. Liu, *Point Care*, 2019, **18**, 77–84.
- 179 G. J. Kost, *Front. Public Health*, 2019, **7**, 329.
- 180 P. Rojanasantikul, P. Pattrapornpisut, K. Anuruckparadorn and W. Katchamart, *Clin. Rheumatol.*, 2014, **33**, 919–923.
- 181 L. Mardani, M. T. Vardini, M. Es'Haghi and E. Ghorbani Kalhor, *Anal. Methods*, 2020, **12**, 333–344.
- 182 B. Derkus, E. Emregul, C. Yucesan and K. Cebesoy Emregul, *Biosens. Bioelectron.*, 2013, **46**, 53–60.
- 183 A. Biela, M. Watkinson, U. C. Meier, D. Baker, G. Giovannoni, C. R. Becer and S. Krause, *Biosens. Bioelectron.*, 2015, **68**, 660–667.
- 184 T. Liu, S. Zhang, W. Liu, S. Zhao, Z. Lu, Y. Wang, G. Wang, P. Zou, X. Wang, Q. Zhao and H. Rao, *Sens. Actuators, B*, 2020, **305**, 127524.
- 185 H. B. Lee, M. Meeseepong, T. Q. Trung, B. Y. Kim and N. E. Lee, *Biosens. Bioelectron.*, 2020, **156**, 112133.
- 186 M. S. Khan, K. Dighe, Z. Wang, I. Srivastava, A. S. Schwartz-Duval, S. K. Misra and D. Pan, *Analyst*, 2019, **144**, 1448–1457.
- 187 B. Srinivasan, J. L. Finkelstein, D. O'Dell, D. Erickson and S. Mehta, *EBioMedicine*, 2019, **42**, 504–510.
- 188 B. Srinivasan, D. O'Dell, J. L. Finkelstein, S. Lee, D. Erickson and S. Mehta, *Biosens. Bioelectron.*, 2018, **99**, 115–121.
- 189 S. Wadhwa, A. T. John, S. Nagabooshanam, A. Mathur and J. Narang, *Appl. Surf. Sci.*, 2020, **521**, 146427.
- 190 F. Polli, C. D'Agostino, R. Zumpano, V. De Martino, G. Favero, L. Colangelo, S. Minisola and F. Mazzei, *Talanta*, 2023, **251**, 123755.
- 191 M. Bhaiyya, P. K. Pattnaik and S. Goel, *Sens. Actuators, A*, 2021, **331**, 112831.

