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Recent advances in DNA-based electrogenerated chemiluminescence biosensors

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Electrogenerated chemiluminescence (also called electrochemiluminescence and abbreviated as ECL) has witnessed significant development in fields ranging from biological analysis to clinical diagnosis due to its outstanding advantages such as high sensitivity, wide detection range, and low background signal. DNA, a class of biopolymers with programmable self-assembly and molecular recognition functions, is a significant tool in the field of ECL bioanalysis. Benefiting from the unique structures and excellent properties of DNA, DNA-based ECL biosensors have aroused increasing attention. In this review, we summarize and classify the signal output mode of DNA-based ECL biosensors and introduce the different methods for the immobilization of DNA probes on electrodes. In addition, we discuss the recent progress and applications of DNA-based ECL biosensors, including the detection of nucleic acids, proteins, and small biomolecules. Finally, we present a brief summary and prospects in this field.

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1. Introduction

In biosensing, the issue of selectivity, especially for low-abundant targets and in the presence of interfering substances is critical.^{1–3} Therefore, the development of biosensors with high selectivity and ease of operation can overcome this issue during sensing. In the 1960s, Clark and Lyons invented the first biosensor for the electrochemical measurement of glucose in biological samples and defined the basic concept of biosensors.⁴ Since then, biosensors have been rapidly applied in many fields, including analytical chemistry, medical diagnosis, environment monitoring, and food safety inspection.^{5–7} A biosensor is “a set of stand-alone integrated devices that provide specific quantitative and semi-quantitative analytical information by placing biometric elements in direct contact with transducers”.^{8,9} It is mainly composed of two parts, as follows: (i) bioactive materials, also known as molecular recognition elements, which determine the selectivity of the biosensor, and (ii) transducer, which can be physical or chemical and converts physical or chemical changes produced by various active substances into electrical signals.^{10–12} Thus, by matching the molecular recognition elements with appropriate transducers, the developed

biosensors can show good analytical performances. With the assistance of the transducer, the relevant changes caused by the target are output as the related electrical signal (including capacitance, resistance, potential or current, *etc.*).^{13,14} The higher the concentration of the target, the higher/lower the electrical signal. Based on this principle, the quantitative analysis of the target can be realized. Biosensors combine the specificity and sensitivity of biological systems and the computational power of microprocessors, which have the characteristics of high specificity, fast speed, simple operation, and small sample preprocessing.^{15–17}

ECL biosensors are analytical technology that take biomolecules such as proteins, DNA/RNA, small molecules, and ions as targets and converts the signals generated during their specific recognition into optical signals for the quantitative detection of the targets.¹⁸ Compared with other biosensors, ECL biosensors have outstanding advantages, such as high sensitivity, low background signal, and high controllability.^{19–21} Thus, they are reliable platforms for detecting important disease markers with low abundance, which can be used for early disease diagnosis and prognostic monitoring.^{22–24} Since the 21st century, ECL biosensors have successfully entered the limelight and developed quickly. To build superior ECL biosensing systems, numerous types of materials have been intensively developed and applied.²⁵

As one of the most attractive biological polymers, DNA has great potential in the field of biological analysis. In addition to being the carrier of genetic information and responsible for the conversion of genetic code into proteins, it can also be used as a functional material for bioanalysis, DNA nanotechnology, and nanomedicine.^{26–28} In recent years,

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DNA-based ECL biosensors have attracted significant attention for the ultra-sensitive detection of biomolecules.^{29–31} This type of biosensor typically uses DNA as a molecular recognition element to capture the target in the sample.³² By modifying single-stranded DNA (ssDNA) or DNA self-assembled nanostructured probe on the surface of the electrode, the DNA probe molecules can selectively hybridize with the target under the appropriate conditions and convert the generated biological signals into detectable chemical signals, thus achieving the purpose of target detection.³³ With the advancement of nanotechnology, ingenious signal amplification strategies and nano-functional materials have been gradually developed and various DNA-based ECL biosensors manufactured with improved sensitivity and selectivity.

Herein, we review the latest research on DNA-based ECL biosensors in the past few years, focusing on their main signal output modes and the methods for the immobilization of DNA probes during the construction of ECL biosensors. In addition, the recent progress on DNA-based ECL biosensors in the field of biological analysis is highlighted according to the classification of the targets (Scheme 1). Due to the proliferation of related literature in recent years, this review only briefly introduces some of these studies as references as needed.

2. Signal output mode of DNA-based ECL biosensors

With the rapid development of DNA and the ECL technique, researchers have realized a variety of signal output modes for ECL biosensors, including single signal output mode and multi-signal switching mode.³⁴ Generally, most ECL systems are based on a single signal output mode. The single signal output mode typically includes the “signal on” and “signal off” modes. In the “signal on” mode, the ECL signal usually increases with an increase in the analyte concentration, while the “signal off” mode is the opposite. The single signal output mode has certain disadvantages in ECL analysis, for example, it is not conducive to the stability, accuracy and efficiency. Alternatively, multi-signal mode ECL systems can avoid most of the defects of the single signal output mode, making ECL analysis more flexible and accurate. Generally,

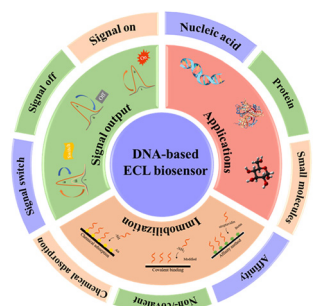
the multi-signal switching mode is used for the analysis of more than two analytes, and the ECL signal changes with the analytes concentration in real time, resulting in more complex signal changes. In this section, the different signal output modes of DNA-based ECL biosensors are categorized and summarized, and their applications in the field of bioanalysis are discussed and explained.

2.1. “Signal on” mode

In DNA-based ECL biosensors, the “signal on” mode is a signal process from zero or weak to strong and is a widely researched ECL signal output mode, which has become an effective method in the field of biological analysis, disease diagnosis, *etc.*^{35–37} There are various conventional means to achieve the “signal on” strategy, including the introduction of luminescent substances, co-reaction reagents or co-reaction promoters and the application of ECL resonance energy transfer (ECL-RET) systems.

Common luminescent materials such as ruthenium (Ru) complexes, luminol and its derivatives, and quantum dots have been used in biosensors. Recently, Peng *et al.*³⁸ modified a luminous Ru complex on the electrode surface as an emitter for signal generation and achieved precise modulation of signal quenching to signal enhancement in a chain substitution reaction-based system by introducing DNA nanoscissors as a control structure (Fig. 1A). In the presence of the target miRNA, a strong ECL emission was obtained. Liu *et al.*³⁹ successfully synthesized sulfur quantum dots (SQDs) as a signal emitter *via* the hydrothermal method. In combination with a three-stranded DNA (tsDNA) nanostructure and duplex-specific nuclease (DSN)-assisted target cycle amplification signal strategy, the ultra-sensitive detection of miRNA-21 was achieved (Fig. 1B). Compared with the quantum dots containing heavy metal elements, pure elemental quantum dot materials such as SQDs are suitable for the construction of ECL biosensors due to their advantages of low toxicity, good water solubility, high biocompatibility and outstanding ECL properties. This work extends the application of pure elemental quantum dots in ECL luminescent materials and provides a new direction for the construction of novel quantum dot-based ECL biosensors.

In the co-reactant ECL system, the co-reactant undergoes electrochemical oxidation or reduction to form a reducing or oxidizing intermediate, which can react with an oxidized or reduced ECL probe to generate excited states. The greater the generation of the intermediate of the co-reactant, the higher the ECL efficiency, thus producing a higher signal. The classical co-reactant systems such as Ru complex-TPrA and luminol–hydrogen peroxide have been widely used in the field of ECL analysis. Nowadays, new ECL luminescent materials and their co-reactant reagents have been developed with the deepening of research. Gold nanoclusters (AuNCs) are a class of ECL clusters, which exhibit the quantum size effect and excellent fluorescence properties, and thus



Scheme 1 Schematic overview of DNA-based ECL biosensors.



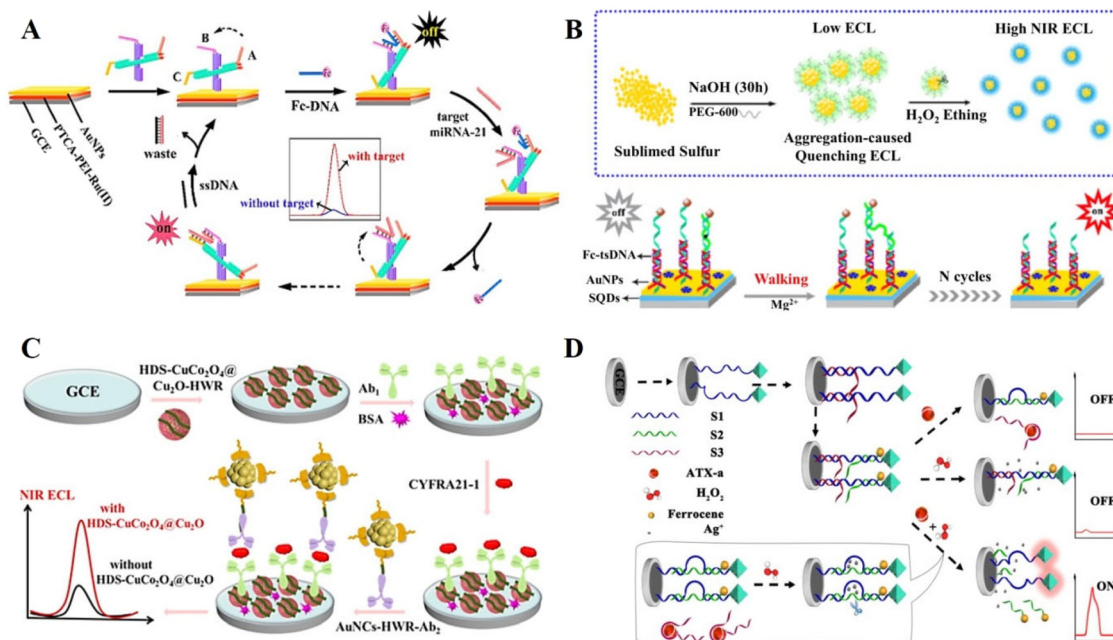
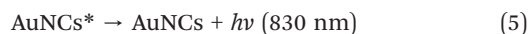
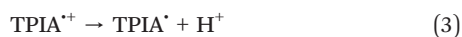
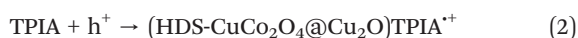
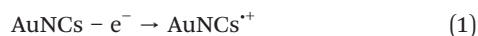


Fig. 1 (A) Schematic illustration of a DNA nanoscissor-based ECL biosensor. Reprinted with permission from ref. 38. Copyright 2019, the American Chemical Society. (B) Schematic illustration of a novel ECL biosensor based on SQUIDs for the detection of miRNA-21. Reprinted with permission from ref. 39. Copyright 2020, the American Chemical Society. (C) Schematic illustration of an ECL biosensor based on HDS-CuCo₂O₄@Cu₂O co-reactant accelerator. Reprinted with permission from ref. 40. Copyright 2022, the American Chemical Society. (D) Schematic illustration of a visual ECL biosensor for the detection of Pb²⁺. Reprinted with permission from ref. 41. Copyright 2018, the American Chemical Society.

attracted extensive attention from researchers in recent years. Jia *et al.*⁴⁰ first prepared a hollow double-shell CuCo₂O₄@Cu₂O (HDS-CuCo₂O₄@Cu₂O) heterostructure, which can be used as a co-reactant accelerator to significantly enhance the luminescence of AuNCs in the ECL reaction (Fig. 1C). HDS-CuCo₂O₄@Cu₂O is involved in the ECL signal generation process mainly by obtaining more electrons from TPIA through holes, which promotes the formation of Triisopropanolamine (TPIA)⁺. The possible mechanism of signal amplification is as follows:



Benefiting from the unique heterostructure of HDS-CuCo₂O₄@Cu₂O, the electron transfer efficiency can be effectively improved in the system, resulting in an increase of nearly 3-fold in the ECL response of the AuNCs.

In addition, the precise and effective introduction of the ECL-RET effect is also an effective strategy for the development of “signal on” mode ECL biosensors. The ECL-RET effect usually occurs when the emission spectrum of a

donor molecule overlaps with the absorption spectrum of another acceptor molecule. The excitation of the donor molecule transfers energy to the acceptor molecule within a certain distance, inducing the absorption of energy by the acceptor molecule to emit light, while the intensity of the donor molecule itself decays. The conventional ECL-RET system usually faces the problem of low effective contact frequency between the donor and acceptor in the solution, resulting in a low quenching efficiency. Xia *et al.*⁴¹ designed a DNA-regulated ECL-RET probe, which can achieve dual quenching and dual stimulation response, with ultra-low background signal and highly strong signal output, to achieve the ultra-sensitive detection of anatoxin-a (Fig. 1D).

As a typical single-signal output mode, the signal of the ECL biosensor in the “signal on” mode is gradually enhanced with an increase in the analyte concentration. This is the most commonly used signal output mode with a wide detection range and low background signal. In future research, achieving a stable and continuous signal output, avoiding rapid decay of the ECL signal, and improving the detection sensitivity are worthy of further study.

2.2. “Signal off” mode

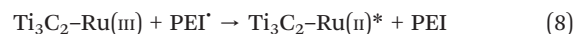
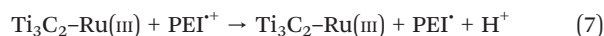
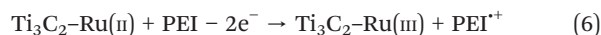
The ECL emission is directly quenched by the ECL quencher, and thus the ECL signal decreases with an increase in the analyte concentration, which is the basic principle of “signal off” mode ECL biosensors.^{42–44}



By cleverly designing and amplifying the ECL response of the initial state, the “signal off” effect of the sensor can be effectively improved.⁴⁵ This provides a new idea for the further development of ultra-sensitive “signal off” mode DNA-based ECL biosensors. Recently, Wang *et al.*⁴⁶ constructed an “on-off” paper-based ECL sensing platform for the highly sensitive detection of miRNA-141. The “signal on” response of this biosensor is based on a dual signal amplification strategy (Fig. 2A). Firstly, 3D graphene with a large specific surface area was introduced on the surface of a paper-based platform, and a large amount of AuPd nanoparticles (NPs) was loaded to build an excellent conductive substrate, which can participate in the ECL reaction system as co-reactants to enhance the ECL emission. Secondly, a DNA walker based on Nt.BsmAI nucleic acid endonuclease was used to further amplify the ECL signal by hydrolyzing DNA through an enzymatic reaction to release the walker probe and excite the 3D DNA walker. The above-mentioned two enhancements caused the sensor to have a strong initial ECL signal. The quenching effect of the ECL biosensor was mediated by clustered regularly interspaced short palindromic repeats (CRISPR)/cas12a-mediated trans-cleavage, and the CRISPR/Cas12a (LbCpf1) protein could indiscriminately digest any non-target ssDNA (called trans-cleavage), resulting in the weakest “signal off” response.

To obtain higher sensitivity and specificity, various new nanomaterials have been introduced in ECL biosensors. Yao *et al.*⁴⁷ successfully prepared an Au@Ti₃C₂@PEI-Ru(dcbpy)₃²⁺

nanocomposite, in which Ti₃C₂ as a class of two-dimensional transition metal compound nanomaterials with the advantages of high loading rate, good electrical conductivity, and polyethylenimine (PEI) was used as a co-reactant to enhance the signal (Fig. 2B). A DNA walker-based amplification strategy was employed to shear hairpin DNA under the action of nucleic acid endonucleases, and the designed model DNA-AgNCs (model DNA-AgNCs) were used as quenchers to quench the ECL. During the ECL reaction, Ti₃C₂-Ru(II) and PEI are firstly oxidized to Ti₃C₂-Ru(III) and PEI⁺. Then, PEI⁺ reacts with PEI' to reduce Ti₃C₂-Ru(III) to the excited-state Ti₃C₂-Ru(II)*. Finally, the excited Ti₃C₂-Ru(II)* radiates to the ground-state Ti₃C₂-Ru(II) and releases photons. The possible luminescence mechanism is as follows:



Consequently, the sensitive detection of SARS-CoV-2 gene was achieved using the “signal off” output mode ECL biosensor. This ECL biosensor has great application potential for clinical detection.

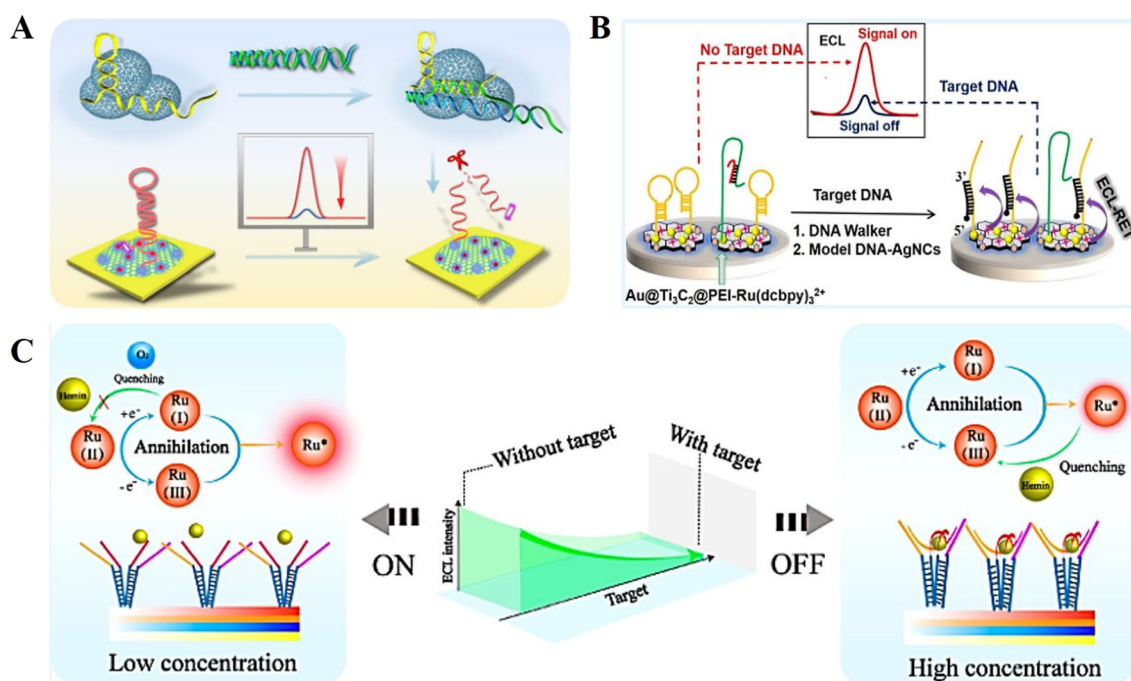


Fig. 2 (A) Schematic illustration of an “on-off” paper-based ECL sensing platform for the highly sensitive detection of miRNA-141. Reprinted with permission from ref. 46. Copyright 2021, the American Chemical Society. (B) Schematic illustration of a “signal off” output mode ECL biosensor for the sensitive detection of SARS-CoV-2. Reprinted with permission from ref. 47. Copyright 2021, the American Chemical Society. (C) Schematic illustration of a “signal off” ECL biosensor based on DNA nanotweezer. Reprinted with permission from ref. 51. Copyright 2019, the American Chemical Society.



The rapid development of DNA nanotechnology has also laid a good foundation for the construction of novel ECL biosensors. The above-mentioned articles both involve the construction of a “signal off” output ECL biosensor with a DNA walker as the main component. In addition, DNA robots, DNA tetrahedrons, DNA dendrimers and other DNA self-assembled structures have been widely used in the design of ECL biosensors.^{48–50} Recently, Bian *et al.*⁵¹ designed a DNA nanotweezer-based ECL biosensor that enables intelligent control of hemin concentration for the ultra-sensitive detection of the early diagnostic biomarker fusion gene PML/RAR α in leukemia (Fig. 2C). Depending on the principle of complementary base pairing, the switching status of the DNA nanotweezer is closely related to the presence or absence of the target. In the absence of the target, the DNA nanotweezer is open, which cannot capture the free hemin in solution, and the ECL response is in the “on” state. In the presence of the target, the DNA nanotweezer changes its configuration to capture hemin, while the high concentration of hemin on the electrode surface quenches the ECL signal and the biosensor is in the “off” state. Based on this highly intelligent and controlled DNA structure design, the proposed method exhibits excellent research value and application potential in bioanalysis.

As an effective signal output mode, the “signal-off” mode has been of increasing interest to researchers due to its high sensitivity, ease of operation, and reagent-free process. In the ongoing development of novel and efficient ECL quenchers, the design of effective steric hindrance strategy or nucleic

acid-based cutting strategy will pave the way for the future “signal-off” mode ECL biosensors.

2.3. “Multiple signal switching” mode

Despite the advantages of “single on” or “single off” mode ECL biosensors, they suffer from the shortcoming of false-positive signals, which limits the application of single-switching mode ECL biosensors in practical sample detection. In recent years, a more complex output mode named “multiple signal switching” mode has attracted widespread attention. Generally, the “multiple signal switching” mode includes the “signal off-on-off” mode, “signal on-off-on” mode, and ratiometric mode.²¹

Hu *et al.*⁵² proposed a “signal off-on-off” mode biosensing strategy for the detection of the antiviral drug amantadine (AMD) (Fig. 3A). In the construction of the biosensor, cyclic multiple changes in the ECL signal were achieved mainly based on the host-guest interaction among AMD, ferrocene (Fc), and cucurbit[7]uril (Q[7]). In the early stage, Fc quenches the Ru(bpy)₃²⁺ luminescence process on the electrode surface, which is in the “off” state with a low signal. Later, by introducing the intermediate Q[7], it will wrap Fc into its cavity, leading to the “on” state with a high signal. Finally, when AMD is added, Fc will be replaced by AMD and cause an obvious signal decrease due to the stronger binding ability between AMD and Q[7], thus achieving an “off-on-off” multiple signal response. Likewise, “signal on-off-on” mode ECL biosensors have been

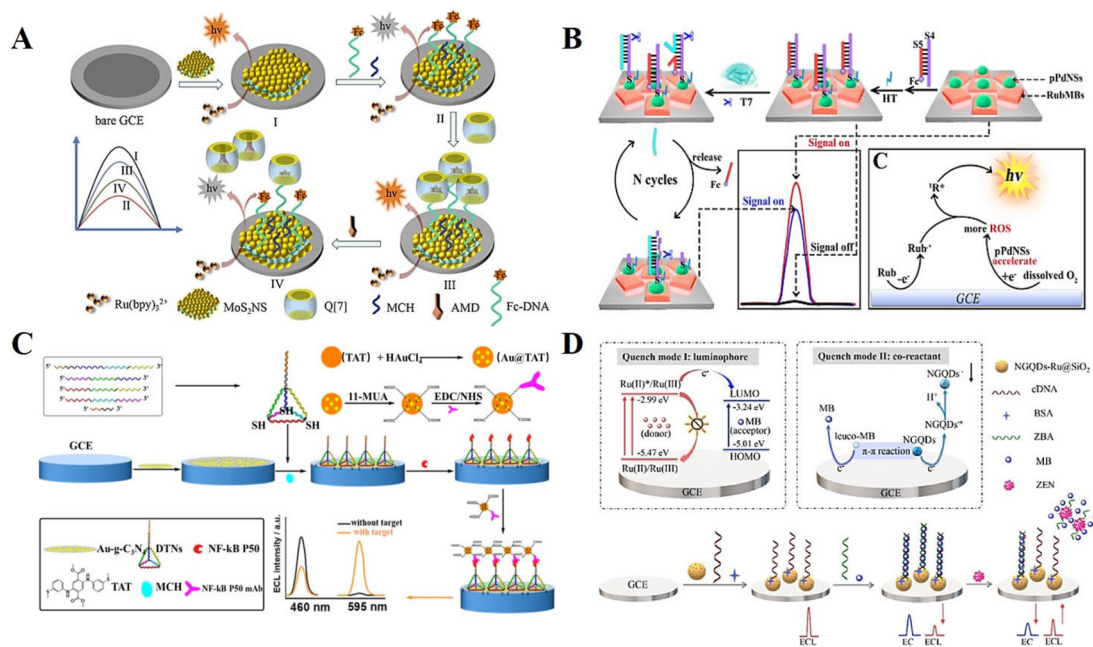
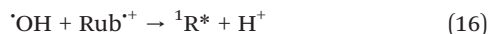
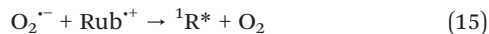
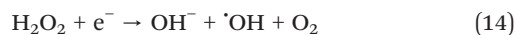
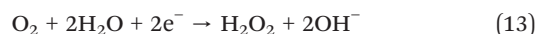


Fig. 3 (A) Schematic illustration of a “signal off-on-off” mode ECL biosensor. Reprinted with permission from ref. 52. Copyright 2021, Elsevier. (B) Schematic illustration of a “signal on-off-on” mode ECL biosensor. Reprinted with permission from ref. 53. Copyright 2018, the American Chemical Society. (C) Schematic illustration of a dual-wavelength emission ECL-RET-based biosensor. Reprinted with permission from ref. 54. Copyright 2020, the American Chemical Society. (D) Schematic illustration of an ECL-EC ratiometric biosensor. Reprinted with permission from ref. 55. Copyright 2023, Elsevier.



developed and used to identify targets with high specificity. Yang *et al.*⁵³ researched a ternary ECL biosensing platform that achieved an initial high intensity ECL signal through material optimization (Fig. 3B). Subsequently, the Fc-labelled nucleic acid probe decreases the electron transfer rate at the electrode surface to obtain a low ECL response. Eventually, an enzyme-mediated strand displacement reaction is triggered in the presence of the target, resulting in signal enhancement and partial restoration of the “signal on” state. The possible ECL mechanism as described in the following eqn (10)–(17). In the solid-phase ternary ECL system, porous palladium nanosphere (pPdNS) with excellent catalytic activity was applied as the co-reaction accelerator to promote the ECL reaction of rubrene microblocks (RubMBs) as luminophores and dissolved O₂ as an endogenous co-reactant.



The “multiple signal switching” mode described above is mainly based on a single signal intensity change. During the detection process, false positive/negative errors may still occur due to instrument errors or environmental changes, such as degradation and dissociation of reagents, co-reactant concentrations, and pH value. The emergence of ratiometric ECL biosensors effectively avoids these interferences. The quantification principle of ratiometric ECL biosensors relies on the change in the ratio of two emission intensities rather than the absolute value, which is normalized by self-calibration, improving the accuracy of the detection. Fan *et al.*⁵⁴ constructed a dual-wavelength emission ECL-RET-based biosensor, which achieved the ultrasensitive detection of nuclear factor-kappa B (NF-κB) based on the ratio change by comparing the signal emission intensity at 595 and 460 nm (Fig. 3C). Mutually independent, non-interfering dual-coupled ECL-electrochemical (EC) signal detection is also an effective method for constructing ratiometric ECL biosensors. Luo *et al.*⁵⁵ firstly discovered the double quenching mechanism of methylene blue (MB) in nanocomposites (nitrogen-doped graphene quantum dots on Ru(bpy)₃²⁺-doped silica nanoparticles, NGQDs-Ru@SiO₂) (Fig. 3D). Relying on this mechanism, an ECL-EC ratiometric biosensor was

cleverly designed for zearalenone (ZEN) detection. Due to the MB-mediated quenching effect, the ECL biosensor initially possessed a high EC signal and a low ECL signal. After recognition binding with ZEN, the ECL signal was recovered and the EC signal was reduced, thus realizing the proportional detection.

Compared with the single signal output mode, the “multiple signal switching” mode undoubtedly has a more sophisticated design concept and corresponding application prospects. This intelligent signal output mode not only has the characteristics of low background signal and high sensitivity, but also can effectively eliminate false negative or positive signals and improve the accuracy, thus promoting the application of ECL biosensors in real sample analysis. In the future, it is necessary to design more “multiple signal switching” strategies to meet different application requirements, and the controllability, reproducibility and stability of the “multiple signal switching” mode in the ECL biosensors also require more attention.

3. Immobilization methods of DNA on electrodes

In the construction of DNA-based ECL biosensors, the immobilization of DNA molecules on the electrode surface is a key step.⁵⁶ To ensure that the DNA probes on the electrode surface possess high activity, stability and directionality, it is necessary to properly regulate the immobilization process of DNA probes and the coverage of the electrode surface.⁵⁷ Whether as a signal or capture probe, the target recognition response in typical DNA-based ECL biosensors typically occurs at the interface.³¹ Therefore, new immobilization strategies need to be continuously developed to improve the accessibility, sensitivity, and specificity of DNA probes and the low thermodynamic and kinetic response at the biosensing interface. The method of immobilizing the probe usually depends on the electrode material and the substrate that modifies the electrode surface.⁵⁸ The commonly used probe immobilization strategies generally include four types including chemical adsorption, physical adsorption (non-covalent bonding), covalent bonding and affinity method. In this section, we briefly introduce these four common types of DNA probe immobilization and illustrate their role in the construction of DNA-based ECL biosensors.

3.1. Chemical adsorption method

The most common chemical adsorption method is the self-assembly adsorption of DNA probes labelled with sulfhydryl (or sulfhydryl derivatives) on the surface of gold (silver, platinum, palladium, *etc.*) electrodes through gold-sulfur bonds (Au-S).⁵⁹ Given that there are no stable oxides under general environmental conditions, sulfhydryl groups can be spontaneously adsorbed on the gold surface through Au-S bonds, forming a dense and highly ordered self-assembled monolayer.⁶⁰ Considering the simplicity and feasibility of this



immobilization method, many reports in the literature fixed sulfur-containing probe DNA on the surface of gold electrodes for the construction of DNA-based ECL biosensors. However, this method of DNA immobilization method faces the problem of non-specific adsorption of non-sulfhydryl-functionalized DNA sequences. DNA probes tend to adsorb on metal surfaces such as gold, silver, platinum, and copper through base pairs non-specifically, making the hybridization efficiency based on pairs at the interface low.⁶¹ Fortunately, some interfacial engineering techniques have been used to position DNA probes vertically on the electrode surface, as well as using short-chain alkane thiols such as 6-mercaptohexanol (MCH) to replace and seal non-specific adsorption at the interface, or the direct adoption of sulfhydryl-functionalized DNA probes to form mixed monolayers for co-assembly, effectively reducing the non-specific adsorption of sulfhydryl-functionalized DNA.^{62–64} Thus, chemisorption allows the single-stranded DNA probe to be firmly and gently fixed to the gold electrode surface, and also preserves the DNA conformation.

Recently, a wide variety of DNA-based ECL biosensors have been developed for clinical diagnostics, chemical research, pharmaceuticals and bioassays based on chemisorption-based DNA probe immobilization. Zhang *et al.*⁶⁵ proposed an ECL strategy for miRNA detection. Hairpin DNA was immobilized on the electrode surface through Au–S bonds, and hybridized with target cyclic amplification products ssDNA, and thus the hairpin DNA opened and acted as a capture probe to achieve the capture of DNA nanoclaws loaded with ECL signal labels (Fig. 4A). The designed ECL biosensor could achieve the highly selective detection of

miRNA-21 under the condition of complex biological samples. Similarly, Zhu *et al.*⁶⁶ used Fe₃O₄@AuNP nanocomplexes with hairpin DNA fixed on their surface as the capture substrate. The target miRNA hybridized with the hairpin DNA to cause conformational changes, thus enabling the detection of miRNAs (Fig. 4B). Besides, Li *et al.*⁶⁷ successfully achieved the capture hybridization of target DNA using sulfhydrylated capture DNA attached to the surface of a glassy carbon electrode (GCE) modified with AuNPs (Fig. 4C). Moreover, Ling *et al.*⁶⁸ achieved early screening of group B streptococci (GBS), an important pathogen at the perinatal stage, based on enzyme-free nucleic acid amplification by assembling thiol-modified capture probes on the surface of gold-modified electrodes through gold thiol bonding (Fig. 4D).

In recent years, DNA immobilization by chemical adsorption, represented by the Au–S bond, has become the most commonly used method for probe immobilization because of its advantages of high structural order and surface binding stability. However, due to the high purity requirements of sulfhydryl DNA, the separation and purification operation are complicated, which may lead to an increase in the experimental cost. Generally, there are two methods for the chemisorption immobilization of sulfhydryl (or sulfhydryl derivatives) DNA probes on electrodes. One is to immobilize the modified probe directly on the surface of the gold electrode. The other is to immobilize the modified probe on gold nanoparticles (AuNPs) at the electrode, which can greatly increase the binding sites, thus improving the ECL response signal and reducing the detection limit.

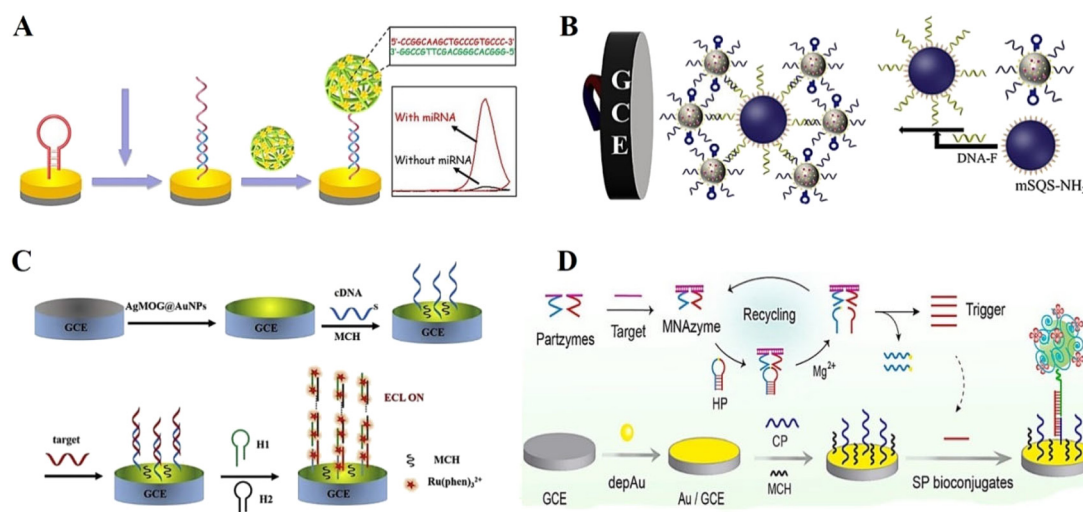


Fig. 4 (A) Schematic illustration of an ECL biosensor in which hairpin DNA was immobilized on the electrode surface through Au–S bonds. Reprinted with permission from ref. 65. Copyright 2020, Elsevier. (B) Schematic illustration of an ECL biosensor based on Fe₃O₄@AuNP nanocomplexes with hairpin DNA fixed on the surface as the capture substrate by Au–S bonding. Reprinted with permission from ref. 66. Copyright 2019, Elsevier. (C) Schematic illustration of an ECL biosensor in which sulfhydrylated capture DNA probes are attached to the electrode (GCE) via Au–S bonds. Reprinted with permission from ref. 67. Copyright 2019, Elsevier. (D) Schematic illustration of an ECL biosensor based on enzyme-free nucleic acid amplification, in which thiol-modified capture probes are assembled on the electrode through gold thiol bonding. Reprinted with permission from ref. 68. Copyright 2019, Elsevier.



3.2. Physical adsorption method (non-covalent bonding)

Physical adsorption (non-covalent bonding) is also a classical immobilization method in DNA-based ECL biosensors. Physical adsorption of DNA monolayers usually depends on the utilization and combination of certain features and factors, including the charge on the electrode surface, the negatively charged phosphate skeleton, the structure of the DNA, and the electrostatic/hydrophobic interactions of the bases.⁶⁹

Graphene, as a new material with sp^2 -hybridized connected carbon atoms tightly stacked into a monolayer two-dimensional honeycomb lattice structure, has excellent optical, electrical, and mechanical properties. It has important application prospects in materials science, micro and nano processing, energy, biomedicine, and drug delivery, and is considered a revolutionary material of the future.⁷⁰ Nowadays, the majority of DNA-based ECL biosensor designs are limited to two-dimensional (2D) graphene, whereas 3D graphene has a larger specific surface area and higher catalytic activity, which can play a more significant role in ECL detection.⁷¹ Wu *et al.*⁷² developed a functionalized ECL sensing platform based on 3D graphene, where $Ru(bpy)_3^{2+}$ -modified DNA probes can self-assemble on the surface of the platform by non-covalent attraction and act as hybridization probes emitting strong ECL signals (Fig. 5A). When the target is present, spontaneous hybridization and complementary pairing will occur to reduce the signal, thus enabling the detection of the target DNA. Song *et al.*⁷³ employed a silver orthophosphate and zirconium-based metal-organic framework (Ag_3PO_4 -UiO-66) as an ECL donor and graphene nanosheets as an ECL acceptor for the fabrication of an ECL-RET biosensor for the highly sensitive detection of diethylstilbestrol (DES) (Fig. 5B). In the sensing platform

construction, aptamer and graphene nanosheets are fixedly attached by electrostatic adsorption, which is used to control the distance between the ECL donor and acceptor to obtain the desired ECL-RET effect.

As one of the simplest methods of DNA immobilization, physical adsorption mainly depends on electrostatic interactions. Given that electrostatic adsorption does not require any modification, the effect on the DNA molecule is weaker. However, the direct non-covalent fixation of DNA makes the DNA skeleton close to the electrode or material surface, causing damage to the DNA probe and the target molecule. This is also a pressing issue for DNA probe immobilization by physical adsorption in the construction of DNA-based ECL biosensors.

3.3. Covalent bonding method

Covalent bonding is another commonly used method to immobilize probes in DNA-based ECL biosensors, which is based on the formation of covalent bonds between chemically modified DNA probe ends and activated substrate surfaces.^{74–76} Schiff base reactions have been widely used for the construction of this type of sensor.⁷⁷ For example, indium tin oxide (ITO) electrodes were silanized by 3-aminopropyl triethoxy silanes (APTS), and acetaldehyde-modified trapped DNA was subjected to a Schiff base reaction to obtain electrodes that immobilized the capture probe. The silanized ITO electrode was cross-linked with a glutaraldehyde cross-linker, and could also be used to immobilize amino-modified DNA or aptamer.⁷⁸ Another common covalent bonding method is carbodiimide bonding, which is formed by immobilizing chemically modified DNA on the surface of an activated sensor with oxygen groups.⁷⁹ For instance, DNA attached to an amino group can be bound to the modified electrode with carboxyl group through the classical amide reaction. Besides, click reaction also provides a promising means for DNA immobilization.⁸⁰ However, compared with chemical adsorption and physical adsorption, the covalent bonding reaction conditions are more complex and demanding, and some covalent binding reaction paths will even damage the base and the recognition ability of the DNA probe. Therefore, once covalent binding is selected, the conditions under which bonding occurs are limited, and it is difficult to ensure that the reaction occurs under mild conditions without affecting the subsequent hybridization reactions. However, a significant advantage of covalent binding is that the bond force is very strong, which not only ensures the fixed amount of DNA probe, but also allows easy removal of small molecules with weak binding force and non-specific adsorption.⁸¹

Feng *et al.*⁸² constructed an ECL sensing platform based on a DNA tetrahedral structure (TDN), which acts as a capture probe with a strand extending from the top and three vertices at the bottom modified with amino functional groups, thus immobilizing multilayer silica nanoparticles on the electrode surface (Fig. 6A). The constructed ECL

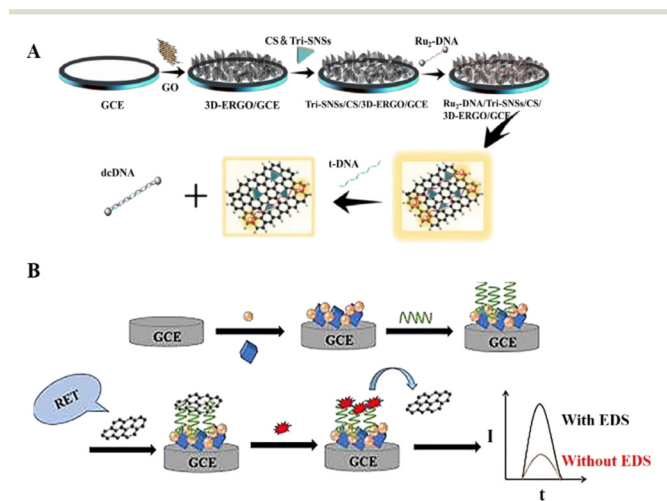


Fig. 5 (A) Schematic illustration of a functionalized ECL biosensor based on 3D graphene in which DNA probes can self-assemble on the surface of the platform by non-covalent attraction. Reprinted with permission from ref. 72. Copyright 2018, Elsevier. (B) Schematic illustration of an ECL-RET-based biosensor relying on physical adsorption binding. Reprinted with permission from ref. 73. Copyright 2023, Elsevier.



biosensor achieved the accurate determination of glucose oxidase (GOD) with a detection limit as low as 40 aM. Liu *et al.*⁸³ investigated the effect of the distance between MoS₂ nanosheets and sulfur-doped boron nitrogen QDs (S-BN QDs) on the enhancement of the MoS₂ surface plasmon response and developed a surface plasmon coupling (SPC) ECL biosensor based on this (Fig. 6B). Hairpin DNA was covalently coupled to S-BN QDs *via* amino and carboxyl groups and served to modulate the DNA strand distance, thus exploring the signal effects of different lengths of DNA strands on plasmonic nanostructures. As a proof of concept, the constructed ECL sensor could achieve the effective detection of the hepatitis C virus (HCV) gene. Xu *et al.*⁸⁴ developed a Zn²⁺-driven DNA rolling machine by borrowing the design concept of DNA walker, which was successfully used in the ultrasensitive detection of miRNA (Fig. 6C). During the construction of the sensing platform, the amino-modified ssDNA is covalently bound to the carboxyl-modified CdS:Mn QDs incubated on the electrode surface, and the ssDNA serves as a track path, which enables the DNA rolling machine to walk, and finally generate the ECL signal. Liu *et al.*⁸⁵ developed an efficient ECL biosensor for miRNA detection based on the signal amplification strategy of nicking enzymes Nb.BbvCI (Fig. 6D). The amino-functionalized hairpin DNA and QDs with carboxyl groups form covalent coupling through amide bonds. The

combination of hairpin probes endowed the biosensor with excellent selectivity (Fig. 6).

3.4. Affinity method

The affinity method represents another effective immobilization method, which has been widely used in DNA-based ECL biosensors. Although there are a variety of affinity methods, the biotin–streptavidin system is one of the most commonly used.⁸⁶ Because streptavidin has a tetramer conformation, it can bind four biotin molecules with high affinity and selectivity. This diversity makes it possible to amplify weak signals and improve the detection sensitivity. As a promising immobilization method, the assembly of DNA probes by the affinity method will be further applied in future biosensing applications.

Cui *et al.*⁸⁷ proposed a dual-signal amplification ECL sensing strategy based on isothermal strand-displacement polymerase reaction (ISDPR) and bridge DNA-AuNP nanocomposites for the highly sensitive detection of miRNA-21, in which a streptavidin-modified Ru(bpy)₃²⁺ complex (SA-Ru) can be immobilized with bridge DNA-AuNPs by the specific binding between biotin and streptavidin, resulting in an ECL emission (Fig. 7A). Li *et al.*⁸⁸ developed a highly specific ECL biosensor based on catalytic hairpin assembly and nonmetallic surface plasmon resonance (SPR) effect,

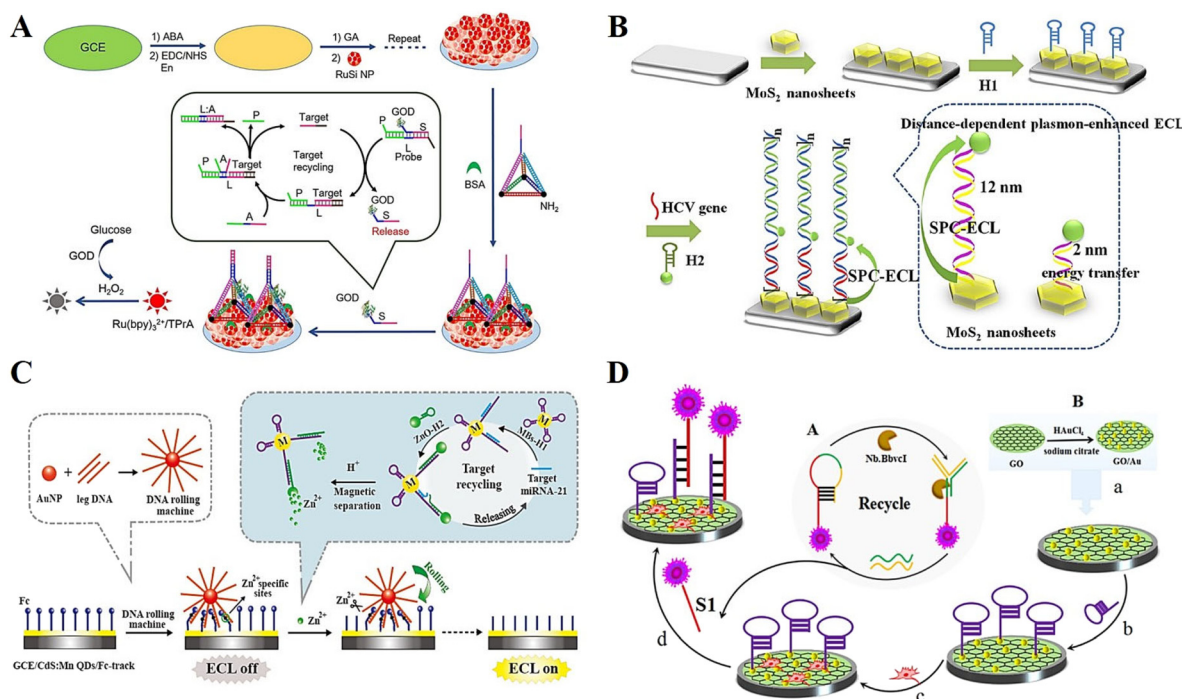


Fig. 6 (A) Schematic illustration of an ECL biosensor based on DNA tetrahedral structure. Reprinted with permission from ref. 82. Copyright 2017, Elsevier. (B) Schematic illustration of an ECL sensor platform, showing the distance relation between MoS₂ nanosheet plasmonic effects and the ECL luminescence of S-BN QDs. Reprinted with permission from ref. 83. Copyright 2020, Elsevier. (C) Schematic illustration of a Zn²⁺-driven DNA rolling machine-based biosensor. Reprinted with permission from ref. 84. Copyright 2019, the American Chemical Society. (D) Schematic illustration of an efficient ECL biosensor for miRNA detection based on the signal amplification strategy of nicking enzymes Nb.BbvCI. Reprinted with permission from ref. 85. Copyright 2017, Elsevier.



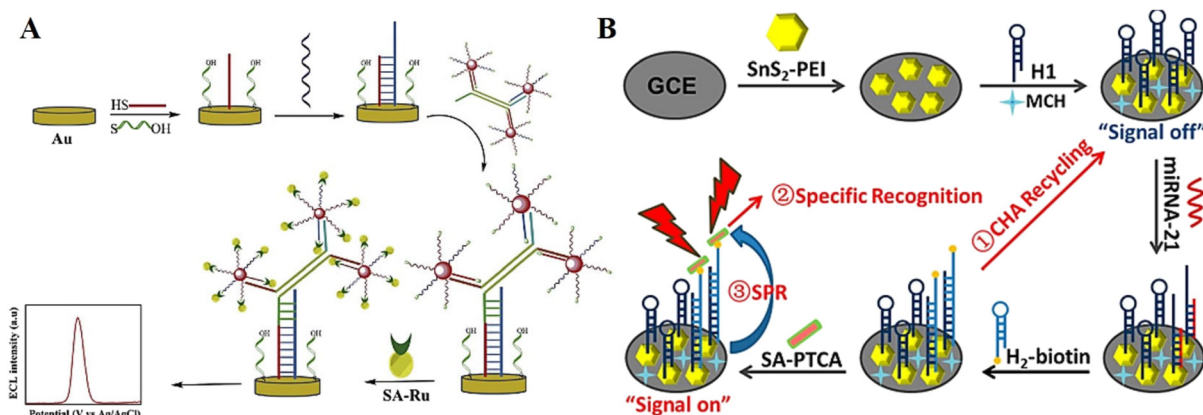


Fig. 7 (A) Schematic illustration of a dual-signal amplification ECL sensing platform. Reprinted with permission from ref. 87. Copyright 2019, Elsevier. (B) Schematic illustration of a highly specific ECL biosensor based on the catalytic hairpin assembly cascade nonmetallic SPR effect. Reprinted with permission from ref. 88. Copyright 2022, Elsevier.

which can be used for the highly sensitive detection of miRNA-21. Hairpin DNA is bound to the surface of SnS₂ nanoplates *via* the specific interactions between biotin and streptavidin, enabling the immobilization of the biosensor signal probe (Fig. 7B).

The biotin–streptavidin binding method is fast, specific and has high affinity and can withstand the extreme effects of pH, temperature, organic solvents and denaturants. Considering the unique advantages of this affinity method, it can realize powerful streamlined bioassay applications. Moreover, owing to the versatility of the biotin–streptavidin system, streptavidin can be coupled to a variety of molecular markers and participate in a variety of immunoassay processes. Therefore, this affinity method is gradually gaining popularity among researchers.

4. Applications of DNA-based ECL biosensors

Nucleic acids, proteins, and small biological molecules in organisms play an essential role in many physiological activities.^{89,90} Their expression levels are also closely related to the occurrence of various human diseases (*e.g.*, malignant tumors, cardiovascular and cerebrovascular diseases), and thus they are widely used as relevant markers for disease diagnosis and prediction.^{91,92} However, due to the low expression level of these disease markers at the early stage, monitoring abnormal changes at their low abundance levels, and thus judging the occurrence of diseases, have become a huge challenge for early diagnosis.

In recent years, ECL biosensing has been developing rapidly. By combining DNA nanotechnology, electronics and optical imaging technology, it has attracted much attention in the fields of food safety and environmental monitoring. Accordingly, achieving efficient and sensitive detection has become one of the research hotspots in the field of ECL bioanalysis. It is capable of target conversion and signal amplification by recognizing targets through a target

conversion strategy that converts a small number of targets into a large number of analog targets through chemical reactions or intermolecular specific recognition interactions.^{93,94} With the help of enzyme-free nucleic acid chain reaction, catalytic hairpin assembly (CHA), hybridization chain reaction (HCR), rolling circle amplification (RCA), DNzyme-induced chain reaction and other signal amplification strategies, the emergence of DNA-based ECL biosensors provides highly sensitive detection tools for disease marker analysis, which is mainly reflected in the detection of nucleic acids, proteins, small molecules, and other types of targets.^{95–97} In this section, we focus on the developments and practical applications of DNA-based ECL biosensors in recent years.

4.1. Nucleic acid detection

Nucleic acid is an essential constituent of all known life forms, which formed by the polymerization of many nucleotide monomers. Depending on their chemical composition, nucleic acids can be mainly classified as deoxyribonucleic acid (DNA) and ribonucleic acid (RNA).^{98,99} The detection of specific nucleic acid sequence targets, including DNA, RNA, miRNA and siRNA, is important for the early diagnosis and treatment of genetic diseases and medical analysis.^{100,101} Currently, the main approach for the detection of nucleic acids is based on the principle of complementary base pairing. In the presence of a target nucleic acid, it undergoes base recognition with the corresponding nucleic acid to induce nucleic acid amplification or DNA self-assembly, after which a small number of target nucleic acid molecules is converted into a mock target associated with an ECL signal, thus enabling detection. In the past few years, many high-performance DNA-based ECL biosensors have been developed for the sensitive detection of specific nucleic acid sequences, and significant progress has been made in the development of strategies for the construction of biosensors.



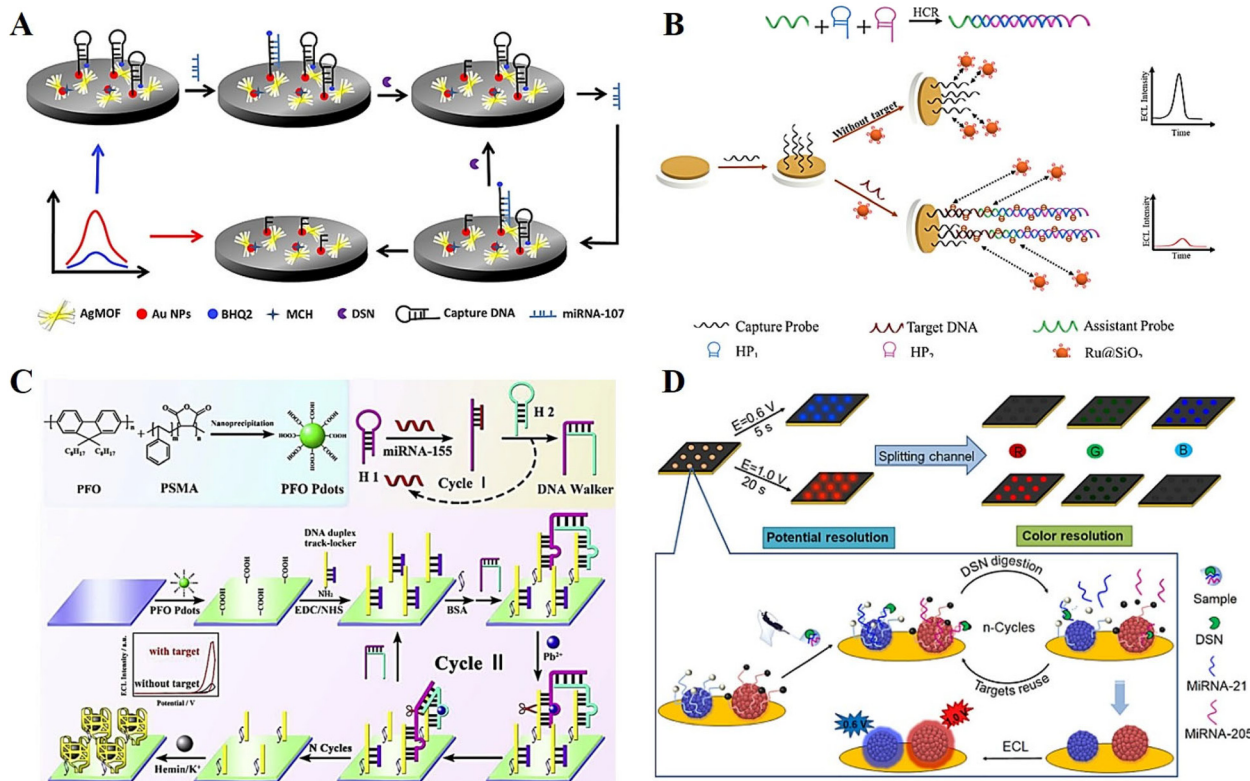


Fig. 8 (A) Schematic illustration of a novel ECL biosensor based on silver metal organic framework for the detection of miRNA-107. Reprinted with permission from ref. 103. Copyright 2022, the American Chemical Society. (B) Schematic illustration of a novel homogeneous ECL biosensor based on Ru@SiO₂ NPs for the sensitive detection of HPV16. Reprinted with permission from ref. 104. Copyright 2022, the American Chemical Society. (C) Schematic illustration of a dual-wavelength emission ECL-RET-based biosensor. Reprinted with permission from ref. 107. Copyright 2020, Elsevier. (D) Schematic illustration of an ECL-EC ratiometric biosensor. Reprinted with permission from ref. 108. Copyright 2021, the American Chemical Society.

To further improve the ECL efficiency, various materials have been developed to enhance the signal. Among them, silver-based materials can be used as a co-reactant of peroxydisulfate to promote the production of radicals and excited states during electrochemical reactions, which is an effective ECL-enhanced emitter.¹⁰² Xiao *et al.*¹⁰³ developed a novel silver metal organic framework (AgMOF) that can catalyze the production of $\text{SO}_4^{\cdot-}$ in the ECL reaction system, enabling self-enhanced ECL emission for the detection of miRNA-107 (Fig. 8A). In addition, the proposed biosensor exhibited a good ECL performance with a linear range of 20 to 120 fM and a detection limit as low as 4.33 fM by combining double-specific nuclease (DSN) for cyclic amplification of the target and resonance energy transfer effects between the donor and acceptor.

Most conventional DNA-based ECL biosensors require the DNA probe and the ECL reporter to be immobilized on the electrode surface, which is usually a tedious and low reproducible process. In contrast, homogeneous ECL biosensors based on sequence length differences are simple and reproducible by using the electrostatic interaction between the DNA strand length variation and the electrode surface to achieve ECL signal changes. Lu *et al.*¹⁰⁴ constructed a novel homogeneous ECL biosensor based on

tris(2,2'-bipyridyl) ruthenium(II) chloride hexahydrate-doped SiO₂ nanoparticles (Ru@SiO₂ NPs) for the highly sensitive detection of human papillomavirus 16 (HPV16) (Fig. 8B). The strength of the ECL signal was varied by the electrostatic interaction between the negative charge carried by the Ru@SiO₂ NPs and the negatively charged electrode surface, which enabled the detection of the target HPV concentration by using long and short strand DNA.

miRNA is a type of important endogenous RNA in the human body. It plays an important regulatory role in the immune response and cellular activity, and can be used as an important marker in the diagnosis and treatment of cancer and other major diseases.¹⁰⁵ Currently, polymer dots (Pdts), as an emerging nanomaterial with excellent luminescence and carrier mobility, have attracted wide interest in the fields of optoelectronic materials research and electrochemical analysis.¹⁰⁶ Furthermore, novel ECL sensing platforms based on Pdts have become an emerging research hotspot in miRNA detection. Herein, Liu *et al.*¹⁰⁷ developed an ECL sensor based on poly(9,9-di-*n*-octylfluorenyl-2,7-diyl) polymer dots (PFO Pdts) and CHA-triggered DNA walker for the ultrasensitive detection of miRNA-155 (Fig. 8C). At +1.25 V, PFO Pdts have strong ECL emission, which can be inhibited and quenched by adding H₂O₂. The DNA walker introduced



by the target-triggered CHA reaction can consume the H_2O_2 in the solution, allowing the recovery of the ECL signal, and thus enabling the accurate determination of intracellular miRNAs.

In addition to single-component detection, multi-component detection is also a hot research topic in the field of ECL biosensors. In this case, Wang *et al.*¹⁰⁸ synthesized luminol-doped Pdots (L-Pdots) and diethylamine-coupled Pdots (N-Pdots) and successfully implemented a potential and color dual-localization ECL sensing strategy for the simultaneous detection of multi-pathway miRNAs (Fig. 8D). L-Pdots and N-Pdots emit the strongest blue and red ECL emission at +0.6 V and +1.0 V, respectively, without interfering with each other. This provides a high-throughput sensing means for dual-potential and dual-color identification, further expanding the application of ECL bioanalysis and clinical diagnosis.

4.2. Protein detection.

Protein, a polypeptide chain composed of amino acids, is coiled and folded into a spatially structured substance during the “dehydration condensation” process, which is the material basis of life.¹⁰⁹ Thus, the detection of proteins plays

a key role in early clinical diagnosis and disease prevention. Currently, the strategies employed for protein detection are mainly specific biological recognition (including antigen-antibody reactions and nucleic acid aptamer binding, *etc.*) and chemical reactions between substances.^{110–112}

The selection of suitable high-performing ECL luminophores for efficient emission is one of the important issues in the field of ECL biosensors. Nanomaterials have been used to improve the sensitivity and selectivity of DNA-based ECL biosensors for protein detection due to their better biocompatibility, easy modification of aptamers and bioenzyme activity. Chen *et al.*¹¹³ developed a polymeric nanomaterial luminescence cluster induced by β -cyclodextrin (β -CD)-mediated enrichment of co-reactants (Fig. 9A). Due to the good stability and excellent ECL performance of the nanocomposite material, the SARS-CoV-2 nucleocapsid protein (ncovNP) could be sensitively detected by a sandwich-type immune system with detection limits down to the femtomolar level.

Metal organic frameworks (MOFs), as a type of new nanomaterials with high porosity, low density and large specific surface area, have attracted wide attention in the field of electrochemical analysis in recent years.¹¹⁴ Among them, zeolitic imidazolate frameworks (ZIFs) are often used

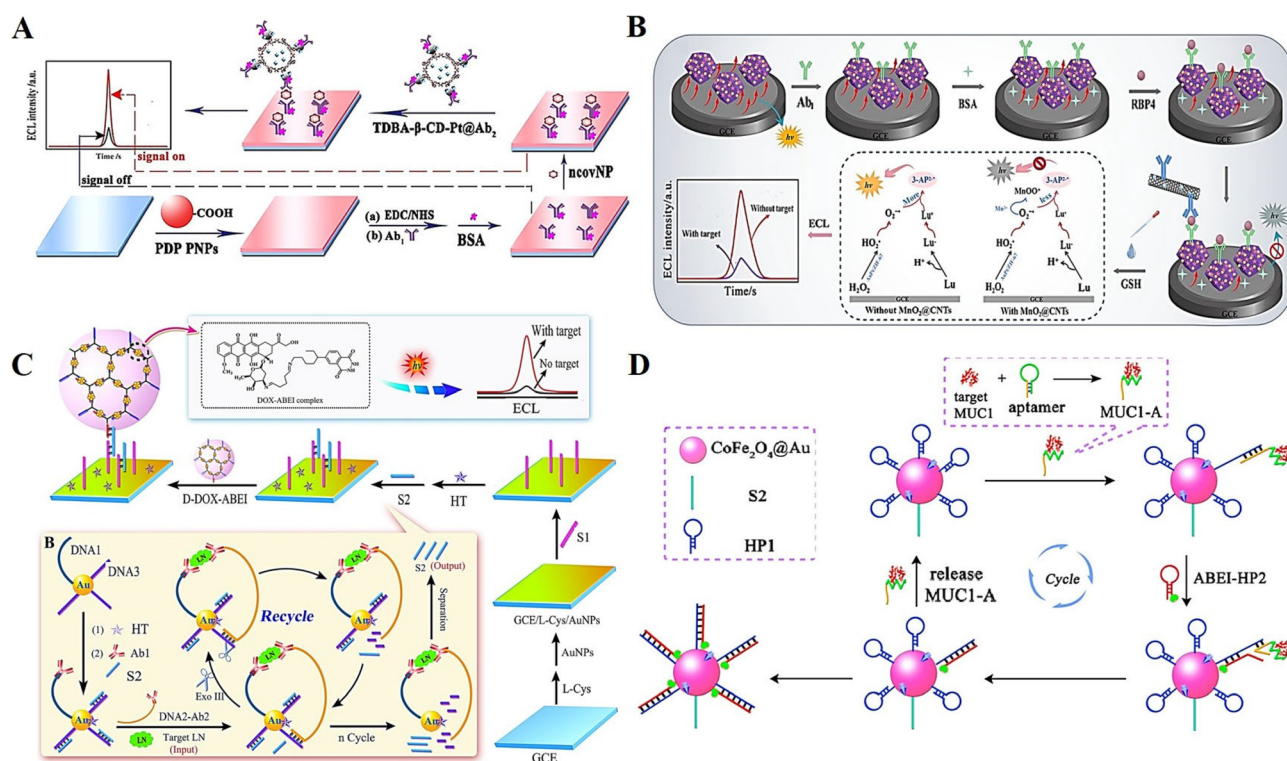


Fig. 9 (A) Schematic illustration of an ECL biosensor based on polymeric nanomaterial luminescence cluster induced by β -CD-mediated enrichment of co-reactants for ncovNP determination. Reprinted with permission from ref. 113. Copyright 2022, the American Chemical Society. (B) Schematic illustration of a double-quenched ECL-RET immunosensor for the detection of typical diabetic proteins. Reprinted with permission from ref. 116. Copyright 2021, Springer Nature. (C) Schematic illustration of an ECL biosensor based on dendritic macromolecular structure for LN detection. Reprinted with permission from ref. 117. Copyright 2018, Elsevier. (D) Schematic illustration of an ECL biosensor relying on an aptamer-binding-initiated DNA nanomachine for ultra-sensitive detection of MUC1. Reprinted with permission from ref. 120. Copyright 2017, the American Chemical Society.



to fabricate composites with good ECL performances through nano-doping due to their excellent catalytic and loading performance.¹¹⁵ Gong *et al.*¹¹⁶ firstly synthesized an Au and Pt bimetallic-anchored and luminol-loaded ZIF material and used it to construct a double-quenched ECL-RET immunosensor for the detection of typical diabetic proteins (Fig. 9B).

DNA nanomaterials with non-toxicity and high biocompatibility, are an excellent and efficient nanoluminescence carriers with potential. Recently, Li *et al.*¹¹⁷ constructed an ECL biosensor based on a DNA dendrimer-containing luminophore system for the ultrasensitive detection of laminin. The DNA dendrimer with abundant double-stranded DNA was synthesized *via* hybridization, which could be loaded with a large amount of ECL luminophore (Fig. 9C). A DNA nanomachine was also involved, which could achieve the ultra-sensitive detection of target proteins by an enzyme-induced cyclic amplification system. This ECL biosensor may hold promising potential in biological analysis and clinical application.

Additionally, the strategy for protein detection of aptamer-based target conversion is the mainstream way of building DNA-based ECL biosensors. In most reported biosensors, a common way to achieve the detection of the target protein is to convert the protein into output ssDNA through transformation by enzymatic cleavage, which is then used to participate in the fabrication of a biosensor.^{118,119} However,

this approach suffers from a series of drawbacks such as complicated operation, time-consuming process and high cost. Fortunately, with the rapid development of DNA nanotechnology, a variety of DNA nanoprobe, nanostructures and nanomachines has been developed, thus enabling direct protein recovery assays under laboratory conditions. As a typical design, Jiang *et al.*¹²⁰ developed an ECL biosensor based on a protein-aptamer binding-initiated DNA nanomachine and successfully used it for the ultra-sensitive detection of mucin 1 (MUC1) (Fig. 9D). Due to its excellent sensitivity, this strategy may provide an efficient method for clinical application, especially in trace protein determination.

4.3. Small biomolecule detection

In the fields of molecular biology and pharmacology, small biomolecules refer to organic compounds with a low molecular weight (usually less than 1000 Daltons), which are directly involved in biological reactions as substrates or products, such as glucose, amino acids and cholesterol, as well as secondary metabolites such as lipids, glycosides, alkaloids and natural phenols.¹²¹ Small biomolecules can combine with specific large biomolecules and act as effectors to change the target activity or function.¹²² Similarly, given that small biomolecules cannot be detected directly by the principle of complementary base pairing, indirect binding to

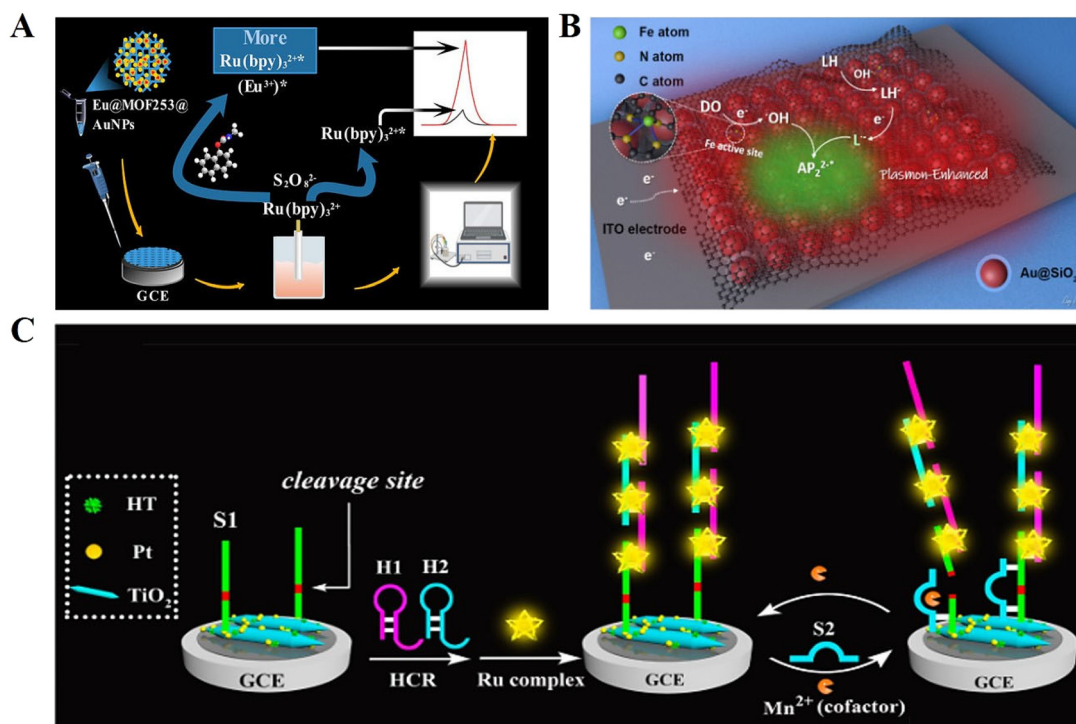


Fig. 10 (A) Schematic illustration of an ECL biosensor based on the dual identification of Eu@MOF253 for carbaryl detection. Reprinted with permission from ref. 124. Copyright 2022, the American Chemical Society. (B) Schematic illustration of a novel ECL biosensor based on two-dimensional layered Au@SiO₂ nanomaterial for the detection of dopamine. Reprinted with permission from ref. 125. Copyright 2021, the American Chemical Society. (C) Schematic illustration of a sensitive ECL biosensor for GSH detection. Reprinted with permission from ref. 127. Copyright 2019, the American Chemical Society.



DNA through intercalation or electrostatic interaction, and the ability to bind small molecules to aptamers also provide new ideas for small molecule detection.

Small biomolecules such as food additives, pesticide residues, drugs and other chemicals are closely related to daily life. Carbaryl is a broad-spectrum insecticide that can protect trees and crops from pests.¹²³ However, due to its high stability at room temperature, it may be residual and accumulate in agricultural crops and related foods. Therefore, it is very important to establish a fast and sensitive method to detect carbaryl residues in food. Liu *et al.*¹²⁴ developed an ECL biosensor based on the dual identification of Eu@MOF253, and successfully achieved highly sensitive and selective carbaryl detection in actual samples (Fig. 10A).

In addition, some biomolecules (*e.g.*, the neurotransmitter dopamine and antioxidant glutathione) are also important markers indicating life activities. Bushira *et al.*¹²⁵ synthesised a 2D Au@SiO₂ nanomaterial coupled with Fe-SACs and further employed it to boost luminol-DO ECL (Fig. 10B). The developed ECL biosensor successfully achieved the rapid and sensitive detection of dopamine and broadened the application of plasmon effect-based ECL signal enhancement strategy in the field of biosensors. Glutathione (GSH) is widely found in animals and plants, and its expression in living organisms plays an important role in maintaining normal immune system function.¹²⁶ Unfortunately, the aptamer of glutathione has not been successfully screened thus far, and it is not feasible to achieve the detection of

glutathione using the target-aptamer target transformation strategy. Zhang *et al.*¹²⁷ proposed a new GSH detection strategy by using the properties of glutathione to reduce MnO₂ nanosheets to Mn²⁺, and then by detecting Mn²⁺ to achieve the conversion of the target (Fig. 10C). TiO₂ acts as a co-reactant in the ECL ternary reaction system to enhance the ECL luminescence, and the helical space groove in the middle of the DNA phosphate backbone provides an insertion site for Ru complexes. SsDNA is modified on the electrode surface, which initially has a high ECL signal and suppresses the signal generation when Mn²⁺ is present, thus achieving the highly sensitive detection of GSH.

4.4. Others

In addition to the above-mentioned biosensors, DNA-based ECL biosensors are also widely used for cellular analysis, metal ions, *etc.* Considering the important role of cells in life sciences and human health, cell-related bioassays have become a popular research topic in recent years. By introducing different types of functional nanomaterials and aptamers, novel ECL cell sensors are being developed rapidly.¹²⁸

Caspase-3 plays an irreplaceable role in apoptosis, and thus is an important biomarker for monitoring apoptosis.¹²⁹ Liang *et al.*¹³⁰ prepared a high-efficiency ECL emitter by doping carbon dots (CDots) and AuNPs in ZIF-8. CDots@ZIF-8/AuNPs not only acted as a carrier for the ECL luminophore, but also a co-reactant to enhance the signal. As a proof of

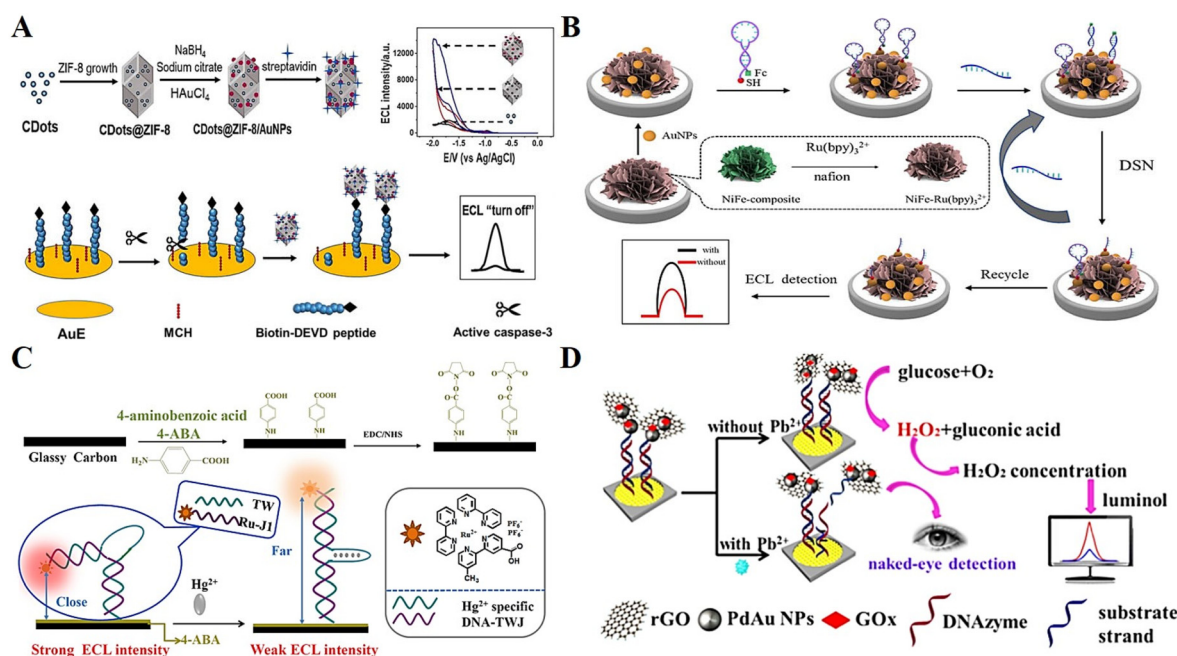


Fig. 11 (A) Schematic illustration of a ZIF-based ECL sensing platform for detection of caspase-3. Reprinted with permission from ref. 130. Copyright 2021, Elsevier. (B) Schematic illustration of an ultrasensitive ECL biosensor based on multiple signal amplification for the detection of cellular exosome. Reprinted with permission from ref. 132. Copyright 2022, Elsevier. (C) Schematic illustration of a highly selective and stable ECL biosensor for the detection of Hg²⁺. Reprinted with permission from ref. 133. Copyright 2019, Elsevier. (D) Schematic illustration of a visual ECL biosensor for the detection of Pb²⁺. Reprinted with permission from ref. 134. Copyright 2018, the American Chemical Society.



concept, CDots@ZIF-8/AuNPs were successfully applied in the detection of caspase-3 activity during cell apoptosis (Fig. 11A). Exosomes are a class of small membrane vesicles containing complex RNA and proteins, which can be used as effective immune markers for the diagnosis and treatment of tumor-like diseases.¹³¹ Zhang *et al.*¹³² proposed a multiplex ECL signal amplification strategy based on NiFe–Ru(bpy)₃²⁺ nanocomposites and DSN-mediated target cycle for cellular exosome detection, exemplifying the great potential of DNA-based ECL analysis technology in the early diagnosis of diseases (Fig. 11B).

An excess or lack of metal ions in the human body will cause an imbalance, leading to serious diseases. Ma *et al.*¹³³ assembled a unique structure of DNA three-way junction (DNA-TWJ) and applied it for the preparation of a highly selective and stable ECL biosensor (Fig. 11C). Finally, they successfully realized the sensitive detection of the metal ion Hg²⁺ with a detection limit as low as 0.04 pM. Xu *et al.*¹³⁴ designed a visual ECL biosensor based on a cross-like all-in-one lab-on-paper analytical device to realize the on-site visible monitoring of Pb²⁺ in tap water and river water, providing a new scheme for the detection of heavy metal ions in the technical field of environmental monitoring (Fig. 11D).

5. Summary and prospect

In the last few years, DNA-based ECL biosensors have been flourishing based on different signal output modes and immobilization methods, combined with various types of signal amplification strategies. Has accordingly, they have become an excellent analytical tool and widely used in many fields such as medical diagnostics, biopharmaceuticals, food safety, and environmental monitoring. Currently, there are two main paths for the development and innovation of DNA-based ECL biosensors, as follows: (i) novel materials. Continuously developing new materials with high luminescence efficiency, high stability and electrocatalytic properties or co-reactant materials with the ability to accelerate electron transfer and improve the reaction rate of ECL systems. In this regard, benefiting from the rapid development of nanotechnology, various nanomaterials, such as carbon nanomaterials, noble metal nanomaterials, semiconductor nanomaterials and conductive polymer nanomaterials in ECL biosensing are also increasing. (ii) Signal amplification strategy. Efficient ECL emission can be achieved by combining multiple signal amplification reactions through clever strategy design. In addition, the in-depth research on DNA nanotechnology has overcome the limitation of using DNA as genetic material, and thus DNA is no longer limited to being a capture probe or signal hybridization probe for DNA-based ECL biosensors. Through precise structural design based on the principle of complementary base pairing, it is used for the construction of biosensors in various forms and means (*e.g.*, DNA molecular motors, DNA walkers, and DNA robots). This bringing a new perspective to the field of biosensors.

Looking ahead, we believe that there are still many problems that need to be overcome and improved. For example, the current ECL biosensors are still mainly focused on single-component detection, and most of the existing studies for two-component detection are based on two luminescent reagents to achieve common detection, with cross-reaction. A few sensing devices that implement one luminescent reagent require the building of complex mathematical models, which are laborious and time-consuming. Therefore, one of the problems to be solved in the future is the crossing of the direct influence between two luminescent reagents in ECL two-component detection and realizing the purpose of using one luminescent reagent to achieve the two-component detection. In addition, building a universal ECL biosensing platform, solving the problem that traditional biosensors are difficult to regenerate, minimizing the regeneration steps, and reducing experimental costs should also be the focus of future research. Furthermore, achieving the miniaturization of ECL biosensing technology will be the focus of future application in commercial products. By combining microelectrode array technology and microfluidic platforms, the migration from laboratory biosensing detection tools to commercial biological medical devices will receive increasing attention.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

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Notes and references

- 1 M. Oliverio, S. Perotto, G. C. Messina, L. Lovato and F. De Angelis, *ACS Appl. Mater. Interfaces*, 2017, **9**, 29394–29411.
- 2 S. Lakard, I.-A. Pavel and B. Lakard, *Biosensors*, 2021, **11**, 179.
- 3 E. A. Songa and J. O. Okonkwo, *Talanta*, 2016, **155**, 289–304.
- 4 L. C. Clark Jr and C. Lyons, *Ann. N. Y. Acad. Sci.*, 1962, **102**, 29–45.
- 5 M. Soltani, B. R. Davis, H. Ford, J. A. D. Nelson and B. C. Bundy, *Biochem. Eng. J.*, 2018, **138**, 165–171.
- 6 A. Salari and M. Thompson, *Sens. Actuators, B*, 2018, **255**, 3601–3615.
- 7 P. Mehrotra, *J. Oral. Biol. Craniofac. Res.*, 2016, **6**, 153–159.
- 8 D. R. Thévenot, K. Toth, R. A. Durst and G. S. Wilson, *Anal. Lett.*, 2001, **34**, 635–659.
- 9 D. R. Thevenot, K. Toth, R. A. Durst and G. S. Wilson, *Pure Appl. Chem.*, 1999, **71**, 2333–2348.
- 10 S. A. Lim and M. U. Ahmed, *RSC Adv.*, 2016, **6**, 24995–25014.



- 11 R. R. Breaker, *Curr. Opin. Biotechnol.*, 2002, **13**, 31–39.
- 12 F. S. Ligler, K. E. Sapsford, J. P. Golden, L. C. Shriver-Lake, C. R. Taitt, M. A. Dyer, S. Barone and C. J. Myatt, *Anal. Sci.*, 2007, **23**, 5–10.
- 13 R. S. Sethi, *Biosens. Bioelectron.*, 1994, **9**, 243–264.
- 14 S. P. Mohanty and E. Kougiyanos, *IEEE Potentials*, 2006, **25**, 35–40.
- 15 C. Ziegler and W. Göpel, *Curr. Opin. Chem. Biol.*, 1998, **2**, 585–591.
- 16 M. N. Velasco-Garcia and T. Mottram, *Biosyst. Eng.*, 2003, **84**, 1–12.
- 17 S. Vigneshvar, C. Sudhakumari, B. Senthilkumaran and H. Prakash, *Front. Bioeng. Biotechnol.*, 2016, **4**, 11.
- 18 V. Gaudin, in *Electrochemical Biosensors*, Elsevier, 2019, pp. 307–365.
- 19 H. Qi and C. Zhang, *Anal. Chem.*, 2019, **92**, 524–534.
- 20 Z. Xu, L. Liao, Y. Chai, H. Wang and R. Yuan, *Anal. Chem.*, 2017, **89**, 8282–8287.
- 21 Y. Zhuo, H.-J. Wang, Y.-M. Lei, P. Zhang, J.-L. Liu, Y.-Q. Chai and R. Yuan, *Analyst*, 2018, **143**, 3230–3248.
- 22 J. E. Ardill and T. M. O'Dorisio, *Endocrinol. Metab. Clin.*, 2010, **39**, 777–790.
- 23 P. Steinacker, A. Huss, B. Mayer, T. Grehl, J. Grosskreutz, G. Borck, J. Kuhle, D. Lulé, T. Meyer and P. Oeckl, *Amyotrophic Lateral Scler. Frontotemporal Degener.*, 2017, **18**, 112–119.
- 24 M. Khalil, C. E. Teunissen, M. Otto, F. Piehl, M. P. Sormani, T. Gattringer, C. Barro, L. Kappos, M. Comabella and F. Fazekas, *Nat. Rev. Neurol.*, 2018, **14**, 577–589.
- 25 M. A. Arugula, Y. Zhang and A. L. Simonian, *Anal. Chem.*, 2014, **86**, 119–129.
- 26 J. Kim and C. J. Easley, *Bioanalysis*, 2011, **3**, 227–239.
- 27 N. C. Seeman and H. F. Sleiman, *Nat. Rev. Mater.*, 2017, **3**, 1–23.
- 28 R. Chhabra, J. Sharma, Y. Liu, S. Rinker and H. Yan, *Adv. Drug Delivery Rev.*, 2010, **62**, 617–625.
- 29 J. Zhang, H. Qi, Y. Li, J. Yang, Q. Gao and C. Zhang, *Anal. Chem.*, 2008, **80**, 2888–2894.
- 30 M.-L. Zhao, W.-J. Zeng, Y.-Q. Chai, R. Yuan and Y. Zhuo, *Anal. Chem.*, 2020, **92**, 11044–11052.
- 31 L. D. Zhao, X. Yang, X. Zhong and Y. Zhuo, *ChemPlusChem*, 2022, **87**, e202200070.
- 32 Z. Ning, M. Chen, G. Wu, Y. Zhang and Y. Shen, *Biosens. Bioelectron.*, 2021, **191**, 113462.
- 33 D. Li, S. Song and C. Fan, *Acc. Chem. Res.*, 2010, **43**, 631–641.
- 34 H. Jin, R. Gui, J. Yu, W. Lv and Z. Wang, *Biosens. Bioelectron.*, 2017, **91**, 523–537.
- 35 J. Zhang, P. Chen, X. Wu, J. Chen, L. Xu, G. Chen and F. Fu, *Biosens. Bioelectron.*, 2011, **26**, 2645–2650.
- 36 A. Cui, J. Zhang, W. Bai, H. Sun, L. Bao, F. Ma and Y. Li, *Biosens. Bioelectron.*, 2019, **144**, 111664.
- 37 X. Ou, X. Tan, X. Liu, Q. Lu, S. Chen and S. Wei, *Biosens. Bioelectron.*, 2015, **70**, 89–97.
- 38 L. Peng, Y. Yuan, X. Fu, A. Fu, P. Zhang, Y. Chai, X. Gan and R. Yuan, *Anal. Chem.*, 2019, **91**, 3239–3245.
- 39 L. Liu, Y. Zhang, R. Yuan and H. Wang, *Anal. Chem.*, 2020, **92**, 15112–15119.
- 40 H. Jia, J. Li, L. Yang, D. Fan, X. Kuang, X. Sun, Q. Wei and H. Ju, *Anal. Chem.*, 2022, **94**, 7132–7139.
- 41 M. Xia, F. Zhou, X. Feng, J. Sun, L. Wang, N. Li, X. Wang and G. Wang, *Anal. Chem.*, 2021, **93**, 11284–11290.
- 42 L. Liu, Q. Ma, Y. Li, Z. Liu and X. Su, *Biosens. Bioelectron.*, 2015, **63**, 519–524.
- 43 W. Bai, A. Cui, M. Liu, X. Qiao, Y. Li and T. Wang, *Anal. Chem.*, 2019, **91**, 11840–11847.
- 44 Y. Zhang, Y. Li, Y. Wei, H. Sun and H. Wang, *Talanta*, 2017, **170**, 546–551.
- 45 M. H. Jiang, S. K. Li, X. Zhong, W. B. Liang, Y. Q. Chai, Y. Zhuo and R. Yuan, *Anal. Chem.*, 2019, **91**, 3710–3716.
- 46 Q. Wang, Y. Liu, J. Yan, Y. Liu, C. Gao, S. Ge and J. Yu, *Anal. Chem.*, 2021, **93**, 13373–13381.
- 47 B. Yao, J. Zhang, Z. Fan, Y. Ding, B. Zhou, R. Yang, J. Zhao and K. Zhang, *ACS Appl. Mater. Interfaces*, 2021, **13**, 19816–19824.
- 48 K. Zhang, Z. Fan, Y. Huang, Y. Ding and M. Xie, *Talanta*, 2022, **236**, 122868.
- 49 K. Zhang, Z. Fan, Y. Ding and M. Xie, *Chem. Eng. J.*, 2022, **429**, 132472.
- 50 L. Li, C. Niu, T. Li, Y. Wan, Y. Zhou, H. Wang, R. Yuan and P. Liao, *Biosens. Bioelectron.*, 2018, **101**, 206–212.
- 51 X. Bian, B. Guo, M. Zhao, D. Han, W. Cheng, F. Song and S. Ding, *ACS Appl. Mater. Interfaces*, 2019, **11**, 3715–3721.
- 52 L. Hu, M. Lei, J. Zhang and Y. Dong, *Sens. Actuators, B*, 2021, **344**, 130328.
- 53 S. S. Yang, M. H. Jiang, Y. Q. Chai, R. Yuan and Y. Zhuo, *ACS Appl. Mater. Interfaces*, 2018, **10**, 38648–38655.
- 54 Z. Fan, Z. Lin, Z. Wang, J. Wang, M. Xie, J. Zhao, K. Zhang and W. Huang, *ACS Appl. Mater. Interfaces*, 2020, **12**, 11409–11418.
- 55 L. Luo, X. Liu, X. Bi, L. Li and T. You, *Biosens. Bioelectron.*, 2023, **222**, 114991.
- 56 X.-m. Chen, B.-y. Su, X.-h. Song, Q.-a. Chen, X. Chen and X.-R. Wang, *TrAC, Trends Anal. Chem.*, 2011, **30**, 665–676.
- 57 E. Sobhanie, F. Salehnia, G. Xu, Y. Hamidi, S. Arshian, A. Firoozbakhtian, M. Hosseini, M. R. Ganjali and S. Hanif, *TrAC, Trends Anal. Chem.*, 2022, 116727.
- 58 C. Fang, H. Li, J. Yan, H. Guo and T. Yifeng, *ChemElectroChem*, 2017, **4**, 1587–1593.
- 59 W.-W. Zhao, J.-J. Xu and H.-Y. Chen, *Chem. Rev.*, 2014, **114**, 7421–7441.
- 60 C. Vericat, M. Vela, G. Benitez, P. Carro and R. Salvarezza, *Chem. Soc. Rev.*, 2010, **39**, 1805–1834.
- 61 P. Kingshott and H. J. Griesser, *Curr. Opin. Solid State Mater. Sci.*, 1999, **4**, 403–412.
- 62 K. Thapa, W. Liu and R. Wang, *Wiley Interdiscip. Rev.: Nanomed. Nanobiotechnol.*, 2022, **14**, e1765.
- 63 J. J. Gooding, *Electroanalysis*, 2002, **14**, 1149–1156.
- 64 S. Campuzano, F. Kuralay and J. Wang, *Electroanalysis*, 2012, **24**, 483–493.
- 65 Y. Zhang, G. Xu, G. Lian, F. Luo, Q. Xie, Z. Lin and G. Chen, *Biosens. Bioelectron.*, 2020, **147**, 111789.
- 66 H.-Y. Zhu and S.-N. Ding, *Biosens. Bioelectron.*, 2019, **134**, 109–116.



- 67 Y. Li, L. He, C. Z. Huang and Y. F. Li, *Biosens. Bioelectron.*, 2019, **134**, 29–35.
- 68 J. Ling, M. Zhao, F. Chen, X. Zhou, X. Li, S. Ding and H. Tang, *Biosens. Bioelectron.*, 2019, **127**, 167–173.
- 69 T.-S. Wong, B. Brough and C.-M. Ho, *MCB Mol. Cell. Biomech.*, 2009, **6**, 1.
- 70 A. K. Geim, *Science*, 2009, **324**, 1530–1534.
- 71 S. Nardecchia, D. Carriazo, M. L. Ferrer, M. C. Gutiérrez and F. del Monte, *Chem. Soc. Rev.*, 2013, **42**, 794–830.
- 72 Y. Wu, J. Yang, Z. Zheng, Z. Li, F. Lu, Y. Chen and W. Gao, *Talanta*, 2018, **188**, 58–65.
- 73 Q. Song, X. Shan, L. Bu, A. Dai, D. Jiang, W. Wang, H. Shiigi and Z. Chen, *J. Electroanal. Chem.*, 2023, **928**, 116987.
- 74 Z. Hu, Z. Suo, W. Liu, B. Zhao, F. Xing, Y. Zhang and L. Feng, *Biosens. Bioelectron.*, 2019, **131**, 237–249.
- 75 A. Sassolas, B. D. Leca-Bouvier and L. J. Blum, *Chem. Rev.*, 2008, **108**, 109–139.
- 76 Y. Du and S. Dong, *Anal. Chem.*, 2017, **89**, 189–215.
- 77 Y. Jia and J. Li, *Chem. Rev.*, 2015, **115**, 1597–1621.
- 78 D. Liu and M. La, *Int. J. Electrochem. Sci.*, 2020, **15**, 11371–11386.
- 79 X.-B. Yin, *TrAC, Trends Anal. Chem.*, 2012, **33**, 81–94.
- 80 M. Tarnowska and T. Krawczyk, *Biosens. Bioelectron.*, 2020, **169**, 112614.
- 81 I. Rahmawati, Y. Einaga, T. A. Ivandini and A. Fiorani, *ChemElectroChem*, 2022, **9**, e202200175.
- 82 Q. M. Feng, Y. H. Guo, J. J. Xu and H. Y. Chen, *ACS Appl. Mater. Interfaces*, 2017, **9**, 17637–17644.
- 83 Y. Liu, Y. Nie, M. Wang, Q. Zhang and Q. Ma, *Biosens. Bioelectron.*, 2020, **148**, 111823.
- 84 Z. Xu, Y. Chang, Y. Chai, H. Wang and R. Yuan, *Anal. Chem.*, 2019, **91**, 4883–4888.
- 85 Q. Liu, C. Ma, X. P. Liu, Y. P. Wei, C. J. Mao and J. J. Zhu, *Biosens. Bioelectron.*, 2017, **92**, 273–279.
- 86 A. Sassolas, L. J. Blum and B. D. Leca-Bouvier, *Biotechnol. Adv.*, 2012, **30**, 489–511.
- 87 A. Cui, J. Zhang, W. Bai, H. Sun, L. Bao, F. Ma and Y. Li, *Biosens. Bioelectron.*, 2019, **144**, 111664.
- 88 J. Li, R. Cai and W. Tan, *Anal. Chem.*, 2022, **94**, 12280–12285.
- 89 Y. Zhao, X. Zuo, Q. Li, F. Chen, Y.-R. Chen, J. Deng, D. Han, C. Hao, F. Huang and Y. Huang, *Sci. China: Chem.*, 2021, **64**, 171–203.
- 90 S. L. Kuan and M. Raabe, *ChemMedChem*, 2021, **16**, 94–104.
- 91 M. Lin, P. Song, G. Zhou, X. Zuo, A. Aldalbahi, X. Lou, J. Shi and C. Fan, *Nat. Protoc.*, 2016, **11**, 1244–1263.
- 92 E. E. Ferapontova, E. M. Olsen and K. V. Gothelf, *J. Am. Chem. Soc.*, 2008, **130**, 4256–4258.
- 93 S. Goggins and C. G. Frost, *Analyst*, 2016, **141**, 3157–3218.
- 94 P. Scrimin and L. J. Prins, *Chem. Soc. Rev.*, 2011, **40**, 4488–4505.
- 95 G. Seelig, D. Soloveichik, D. Y. Zhang and E. Winfree, *Science*, 2006, **314**, 1585–1588.
- 96 P. Yin, H. M. Choi, C. R. Calvert and N. A. Pierce, *Nature*, 2008, **451**, 318–322.
- 97 I. Willner, B. Shlyahovsky, M. Zayats and B. Willner, *Chem. Soc. Rev.*, 2008, **37**, 1153–1165.
- 98 J. D. Watson and F. H. Crick, *Nature*, 1953, **171**, 737–738.
- 99 J. Liu, Z. Cao and Y. Lu, *Chem. Rev.*, 2009, **109**, 1948–1998.
- 100 N. Risch and K. Merikangas, *Science*, 1996, **273**, 1516–1517.
- 101 J. J. McCarthy and R. Hilfiker, *Nat. Biotechnol.*, 2000, **18**, 505–508.
- 102 S. Y. Xiao, Y. Li, S. J. Zhen, C. Z. Huang and Y. F. Li, *Electrochim. Acta*, 2020, **357**, 136842.
- 103 S. Xiao, X. Wang, C. Yang, Y. Jiang, S. Zhen, C. Huang and Y. Li, *Anal. Chem.*, 2022, **94**, 1178–1186.
- 104 Y. Lu, X. Rong, P. Wu, J. Shou, L. Chen, F. Luo, C. Lin, J. Wang, B. Qiu and Z. Lin, *Anal. Chem.*, 2022, **94**, 5823–5829.
- 105 Y. Cai, X. Yu, S. Hu and J. Yu, *Genomics, Proteomics Bioinf.*, 2009, **7**, 147–154.
- 106 C. Wu and D. T. Chiu, *Angew. Chem., Int. Ed.*, 2013, **52**, 3086–3109.
- 107 D. Liu, X. Zhang, J. Zhao, S. Chen and R. Yuan, *Biosens. Bioelectron.*, 2020, **150**, 111872.
- 108 N. Wang, L. Chen, W. Chen and H. Ju, *Anal. Chem.*, 2021, **93**, 5327–5333.
- 109 S. Jones and J. M. Thornton, *Proc. Natl. Acad. Sci. U. S. A.*, 1996, **93**, 13–20.
- 110 X. Cheng, Y. Li, K. Jun, D. Liao, W. Zhang, L. Yin, S. Man and L. Ma, *Biosens. Bioelectron.*, 2022, **215**, 114559.
- 111 H. Xiong, J. Yan, S. Cai, Q. He, D. Peng, Z. Liu and Y. Liu, *Int. J. Biol. Macromol.*, 2019, **132**, 190–202.
- 112 W. Zhao, M. M. Ali, M. A. Brook and Y. Li, *Angew. Chem., Int. Ed.*, 2008, **47**, 6330–6337.
- 113 Y. Chen, Y. He, J. Zhao, J. Zhang, R. Yuan and S. Chen, *Anal. Chem.*, 2022, **94**, 4446–4454.
- 114 J. Zhou, Y. Li, W. Wang, X. Tan, Z. Lu and H. Han, *Biosens. Bioelectron.*, 2020, **164**, 112332.
- 115 J. Zhang, Y. Tan and W.-J. Song, *Microchim. Acta*, 2020, **187**, 1–23.
- 116 W. Gong, S. Yang, F. Zhang, F. Tian, J. Chen, Z. Yin, S. Ding, W. Yang and R. Luo, *J. Nanobiotechnol.*, 2021, **19**, 272.
- 117 L. Li, C. Niu, T. Li, Y. Wan, Y. Zhou, H. Wang, R. Yuan and P. Liao, *Biosens. Bioelectron.*, 2018, **101**, 206–212.
- 118 Z.-H. Yang, Y. Zhuo, R. Yuan and Y.-Q. Chai, *Anal. Chem.*, 2016, **88**, 5189–5196.
- 119 S. Fredriksson, M. Gullberg, J. Jarvius, C. Olsson, K. Pietras, S. M. Gústafsdóttir, A. Östman and U. Landegren, *Nat. Biotechnol.*, 2002, **20**, 473–477.
- 120 X. Jiang, H. Wang, H. Wang, Y. Zhuo, R. Yuan and Y. Chai, *Anal. Chem.*, 2017, **89**, 4280–4286.
- 121 C. Feng, S. Dai and L. Wang, *Biosens. Bioelectron.*, 2014, **59**, 64–74.
- 122 D. D. Ryu and D. H. Nam, *Biotechnol. Prog.*, 2000, **16**, 2–16.
- 123 W. R. Melchert and F. R. Rocha, *Talanta*, 2010, **81**, 327–333.
- 124 C. Liu, L. Cai, Y. Wang, H. Wang, G. Fang and S. Wang, *J. Agric. Food Chem.*, 2022, **70**, 6264–6271.
- 125 F. A. Bushira, S. A. Kitte, C. Xu, H. Li, L. Zheng, P. Wang and Y. Jin, *Anal. Chem.*, 2021, **93**, 9949–9957.
- 126 J. Yin, Y. Kwon, D. Kim, D. Lee, G. Kim, Y. Hu, J.-H. Ryu and J. Yoon, *Nat. Protoc.*, 2015, **10**, 1742–1754.



- 127 R. Zhang, X. Zhong, A. Y. Chen, J. L. Liu, S. K. Li, Y. Q. Chai, Y. Zhuo and R. Yuan, *Anal. Chem.*, 2019, **91**, 3681–3686.
- 128 J. Shen, Y. Li, H. Gu, F. Xia and X. Zuo, *Chem. Rev.*, 2014, **114**, 7631–7677.
- 129 A. G. Porter and R. U. Jänicke, *Cell Death Differ.*, 1999, **6**, 99–104.
- 130 G.-X. Liang, K.-R. Zhao, Y.-S. He, Z.-J. Liu, S.-Y. Ye and L. Wang, *Microchem. J.*, 2021, **171**, 106787.
- 131 P. Li, M. Kaslan, S. H. Lee, J. Yao and Z. Gao, *Theranostics*, 2017, **7**, 789.
- 132 J. Zhang, L. Hao, Z. Zhao, D. Jiang and J. Chao, *Sens. Actuators, B*, 2022, **369**, 132332.
- 133 F. Ma, Y. Chen, Y. Zhu and J. Liu, *Talanta*, 2019, **194**, 114–118.
- 134 J. Xu, Y. Zhang, L. Li, Q. Kong, L. Zhang, S. Ge and J. Yu, *ACS Appl. Mater. Interfaces*, 2018, **10**, 3431–3440.

