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Photoswitchable and long-lived seven-membered cyclic singlet diradicals for the bioorthogonal photoclick reaction†

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Annularly 1,3-localized singlet diradicals are energetic and homolytic intermediates, but commonly too short-lived for widespread utilization. Herein, we describe a direct observation of a long-lived and seven-membered singlet diradical, oxepine-3,6-dione-2,7-diyl (OXPID), *via* spectroscopic experiments and also theoretical evidence from computational studies, which is generated *via* photo-induced ring-expansion of 2,3-diaryl-1,4-naphthoquinone epoxide (DNQO). The photo-generated OXPID reverts to the thermally stable σ -bonded DNQO with $t_{1/2}$ in the μ s level, thus constituting a novel class of T-type molecular photoswitches with high light-energy conversion efficiency ($\eta = 7.8\text{--}33\%$). Meanwhile, the OXPID is equilibrated to a seven-membered cyclic 1,3-dipole as an electronic tautomer that can be captured by ring-strained dipolarophiles with an ultrafast cycloaddition rate (k_{2CA} up to $10^9 \text{ M}^{-1} \text{ s}^{-1}$). The T-type photoswitchable DNQO is then exploited to be a highly selective and recyclable photoclick reagent, enabling spatiotemporal-resolved bioorthogonal ligation on living cell membranes *via* a tailored DNQO-Cy3 probe.

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Introduction

The exploitation of photochemical transformation¹ to establish covalent conjugations in the fashion of click chemistry^{2–5} has emerged as a cutting-edge research field, named photoclick chemistry.^{6–9} Photoclick reactions integrate the innate spatio-temporal resolving merits of light rays for non-invasive manipulation over the construction of organic linking frameworks under mild, catalyst-free conditions in a complex environment.¹⁰ Therefore, photoclick reactions are elaborate chemical tools for the fabrication or functionalization of materials in 3D space,^{11–15} tracking of biomolecules in their native context,^{16–19} and creating hybrid biopolymers *in situ*.^{20,21}

Photoclick reactions employing two inert and exogenous functionalities prior to photoactivated ligation are amenable to single-cell or subcellular bioorthogonal labeling strategies.^{22–28} During the past decade, unremitting efforts have been made to

advance this type of photoclick reaction to meet the strict criteria for utilization in living systems.^{7,25} The cutting-edge approaches encompass light-triggered copper(i)-catalyzed azide-alkyne cycloaddition;²⁹ photo-uncaged cyclooctyne for cycloaddition with azido-reporters;³⁰ the cycloadditions of nitrile imine (NI) produced by photolysis of tetrazole³¹ or sydnone³² toward alkene/yne or cyclic azobenzene; photo-induced hetero-Diels-Alder (HDA) reactions, including photo-oxidative inverse-electron-demand Diels-Alder reactions³³ and light-triggered phenanthrenequinone-alkene cycloadditions;³⁴ light-promoted thiol-ene and -yne ligations.³⁵ Since a prolonged exposure of UV light causes photodamage to cells, photoclick reactions triggered by visible or near-IR irradiation with high quantum efficiency are highly advantageous for *in vivo* applications. The photo-sensitizing approaches can be a good solution for achieving visible or near-IR photoclick reactions.³³

The prerequisites for achieving superior spatial resolution and temporal manipulation in photoclick labeling depend on the chemoselectivity, lifetime, and transition pathway of photoexcited intermediates.^{8,36} Concerning the behaviours of the intermediates, photoclick reactions could be categorized into irreversible and reversible intermediate types,⁷ among which the reversible type is exemplified by the reversible photo-dehydration of 3-(hydroxymethyl)-2-naphthol (HMNP) in aqueous solution³⁷ and the P-type photoswitchable oxidopyrylium ylide (PY).³⁸ The reversible intermediates can be fully restored to the inert precursor to circumvent undesired conjugation towards native biomolecules and to compensate for

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Despite these breakthroughs, dienophiles/dipolarophiles capable of capturing photoswitchable SDRs to compete against the ultrafast σ -self-coupling are yet to be adequately explored.

Herein, a class of ring-expansion/contraction T-type photo-switches between 2,3-diaryl-1,4-naphthoquinone epoxide (DNQO) and oxepine-3,6-dione-2,7-diyl (OXPID, Scheme 1c)^{61,62} was revealed by nanosecond transient absorption spectroscopic (NTAS) analyses. The photochemical homolysis of the carbon-carbon bond in the epoxy moiety of DNQO and the characters of the 1,3-diaryl seven-membered cyclic SDRs (OXPID) were detailed for the first time *via* NTAS, electron paramagnetic resonance (EPR), triplet radical capturing, ³O₂ quenching and the solvent effect in harmony with computational studies. Engrossingly, we found that the photo-energy harvested SDRs (conversion efficiency: η up to 0.33) is seamlessly tautomerized to 3,6-dioxoxepinium-7-ylide (DOXEPEY) as an energetic 1,3-dipole, which can undergo an ultrafast [3 + 2] cycloaddition (k_{2CA} up to $1.01 \times 10^9 \text{ M}^{-1} \text{ s}^{-1}$) with *endo*-bicyclo[6.1.0]nonyn-9-yl-methanol (*endo*-BCN-OH). The light-driven dynamic cycle of the T-type photoswitching was intercepted by the competitive cycloaddition. Therefore, a novel photoclick reaction was proposed and explored, in which the T-type photoswitchable OXPIDs serve as a recyclable intermediate to minimize unwanted side-reactions (towards, *e.g.*, thiol additions) for bioorthogonal photolabeling with better spatiotemporal resolution.

Results and discussion

Absorbance spectroscopic characterization of DNQO

Back in 1974–1980, photo-triggered cycloadditions between DNQO and alkenes were initially explored by Maruyama, Tezuka, and Osuaka *et al.*,⁶³ in which the interconversion among DNQO \rightleftharpoons DR \rightleftharpoons DOXEPEY had been hypothesized. In contrast to 2,3-nondiaryl-NQO, 2,3-diaryl-DNQO prefers to photo-react with dipolarophiles and shows resistance to nucleophilic additions and oxygenation,⁶⁴ which inspire us to conceive bioorthogonality. Therefore, we commenced to establish a library of DNQOs with various heteroaryls or electron donors (eD) and electron acceptors (eA) on the 2,3-diaryl terminals (**1b**–**24b**, Fig. 1a and Table S1, ESI[†]). The UV-Vis spectroscopic characters of the representative DNQOs were summarized with two absorption bands observed ($311 \pm 20 \text{ nm}$ and $340 \pm 40 \text{ nm}$) and two extinction coefficients (ϵ_{311} and ϵ_{365}) displayed in Fig. 1b, respectively (also in Table S2, ESI[†]). TD-DFT calculation and natural transition orbital (NTO) analysis⁶⁵ indicated that the two absorption bands around 311 nm and 340 nm belong to the $S_0 \rightarrow S_4$ and $S_0 \rightarrow S_2$ transitions of **1b** (Fig. 1c, for computational details, Fig. S22, ESI[†]), respectively, which are π - π^* and n - π^* type transitions with the oscillator strength, $f = 0.211$ and 0.009.

The orbital distributions also suggest that both the π - π^* and the n - π^* transitions were mainly involved by the oxygen atoms of the epoxide together with the two carbonyl groups, while the 2,3-diaryl terminals have negligible contribution. Therefore, it rationalizes the experimental observation that variation on the diaryl moiety has a trivial effect on the absorption wavelength.

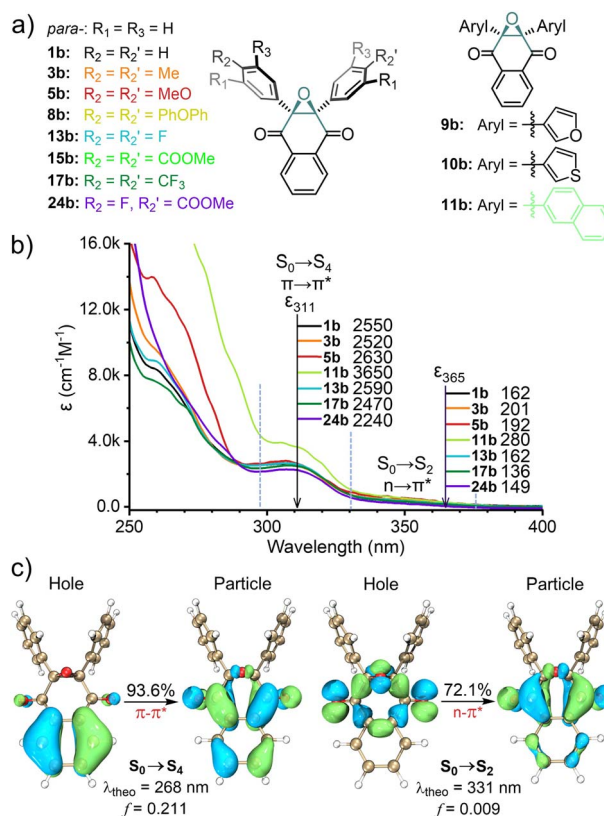


Fig. 1 (a) The structure of the representative DNQOs. (b) Absorption spectra of 100 μM DNQOs in $\text{CH}_3\text{CN} : \text{H}_2\text{O} = 1 : 1$ mixed solvent at 298 K, and the molar absorption coefficients (ϵ) at the designated wavelengths. (c) Theoretical absorption wavelength (λ_{theo}) and the NTOs of the $S_0 \rightarrow S_4$ and $S_0 \rightarrow S_2$ transitions for **1b**, calculated at the TD-M06-2X/6-31G** (SMD, ACN : $\text{H}_2\text{O} = 1 : 1$) level of theory. The oscillator strength (f) and eigenvalue of each NTO pair (in percentage) were shown, and the isovalue of NTO plots was 0.04.

Screening and optimization on the photoclick reactions

To identify a robust reactivity, DNQO (100 μM) was subjected to photo-activation in the presence of typical alkenes/alkynes (500 μM) *via* irradiation with various light sources (*e.g.*, a 311 nm hand-held mercury lamp and 365 nm or 373 nm LED arrays) for 60 s in a mixed aqueous solvent and comparing with conditions of keeping in the dark for 48 h, respectively (Fig. 2 and Tables S3–S5, ESI[†]). The alkenes with ring-strain (**1c**), electron-deficiency (**2c**–**3c**), and electron-donors (**4c**) *versus* the alkynes with ring-strain (**6c** and **7c**), alkyl substituent (**8c**), and electron-deficiency (**9c**) were compared to examine both the electron effect and the ring-strain promotion on the cycloaddition. Based on quantitative HPLC-MS screening (Fig. 2a–c), the results can be summarized as follows: (i) the dipolarophiles without both ring-strain and sufficient electron-deficiency (**3c**, **4c** and **8c**) barely reacted with all the DNQOs. Delightedly, the ring-strained dipolarophiles (**1c**, **6c** and **7c**) could photo-react efficiently with the majority of the DNQOs, while the highly electron-deficient ones (**2c** and **9c**) were able to afford cycloadducts sluggishly; (ii) in the presence of the ring-strained dipolarophiles, DNQOs possessing electron withdrawing groups (EWGs) or fluorine



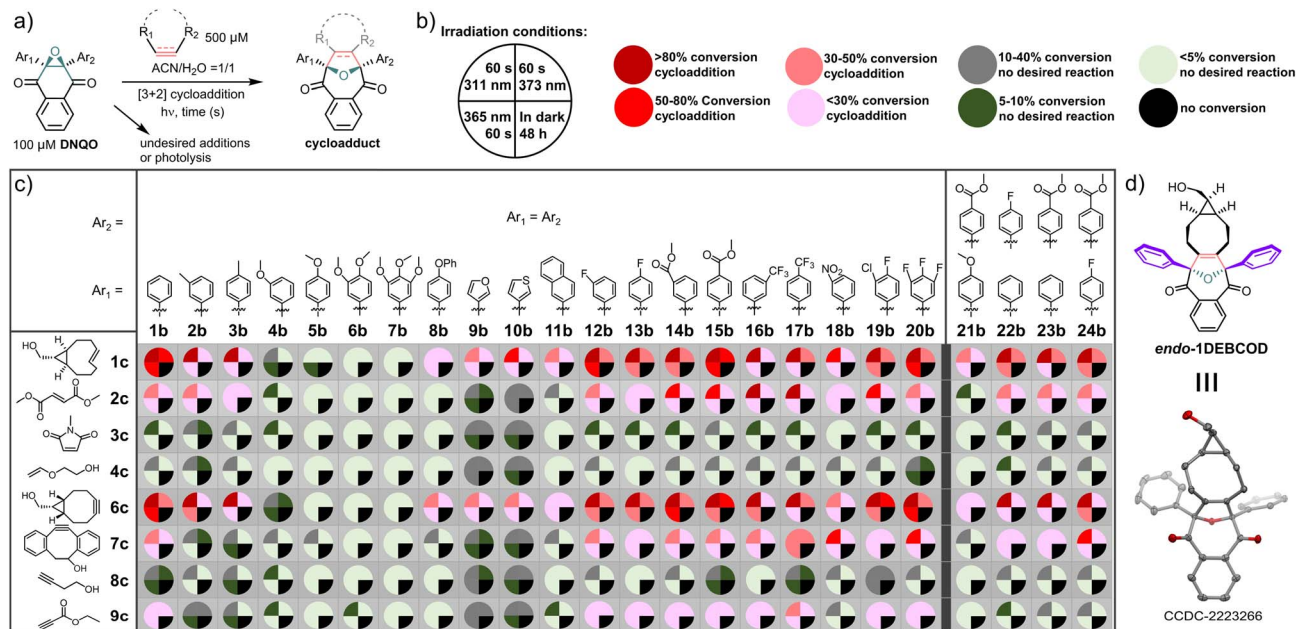


Fig. 2 Screening on DNQO/dipolarophile combinations for the photo-initiated or thermal cycloaddition reactions. Experiments were carried out at 298 K in ACN : H₂O = 1 : 1, irradiated with the indicated light sources. [DNQO] = 100 μM and [dipolarophile] = 500 μM. (a) Reaction schemes. (b) The four irradiation conditions for screening, and the colour codes for the characteristics of each photo-induced [3 + 2] cycloaddition analysed via HPLC-MS. (c) Colour-coded screening results. (d) X-Ray structure of the *endo*-cycloadduct between **1b** and **6c**.

substituents (e.g., **12b–17b** and **19b–20b**) tend to show excellent conversions (>80%), compared with very poor conversion for those with multiple electron-donating substituents (e.g., **4b–7b**); (iii) without appropriate dipolarophiles, most of the DNQOs appear to be inert under continuous irradiation, especially under a 365 or a 373 nm LED. But severe photolyses were observed for most of the DNQOs under 311 nm light, which induces mainly the S₀ → S₄ transition of DNQO. These phenomena reflect the reversible and the short-lived nature of the photogenerated intermediates in the cycloaddition reaction; (iv) there was no background reactions detected in the dark even after incubation for 48 h, implying a high stability of DNQOs; (v) for the ring-strain dipolarophiles, the reaction of **7c** towards DNQOs was found to be slower than **1c** and **6c**, probably due to the steric effects. Therefore, DNQOs **1b**, **3b**, **5b**, **13b**, **17b**, and **24b** with representative substituents were preferred candidates for in-depth evaluation of the performance of the photoclick reaction toward the ring-strained BCN-OH. 365 nm LED was chosen as an optimal light source, considering the high phototoxicity, the photolysis of DNQOs under 311 nm light as well as the poor photo-conversion under 373 nm light (Table S2, ESI†). After a synthetic scale preparation (0.48 mmol scale and 71% yield) and crystallization, the structure of the *endo*-cycloadduct (**1DEBCOD**, Fig. 2d) from **1b** and BCN-OH was identified via single crystal XRD (Table S7, ESI†).

Proofs for the presence of SDRs and the substituent effects

Given the relatively short-lived intermediates (SDRs, Fig. 3a) from the photochemical conversion of DNQO **1b**, we adopted NTAS measurement ($\lambda_{\text{ex.}} = 355$ nm, 1 ns pulse duration, 7 μJ, and 1 kHz) to unravel the absorption spectrum and the decaying

of the intermediates in CH₃CN : H₂O = 1 : 1. Intriguingly, a strong absorption band was observed in the visible region ($\lambda_{\text{max}} = 550$ nm, Fig. 3b) with an exponential decay to the original state ($t_{1/2} = 18$ μs, Fig. 3c) under an air atmosphere. This transient species was then discovered to be a seven-membered cyclic singlet diradical, **1SDR**, based on the following experimental evidence:^{55,59} (i) this species absorbing at 550 nm was EPR-silent in a methanol matrix at 140 K (Fig. S6, ESI†); (ii) the lifetime of this transient species was scarcely altered in the presence of saturated ³O₂ or by typical triplet quenchers (Fig. S4, ESI†); (iii) spin trapping of this transient intermediates (5.0 mM **1b** in ACN/H₂O at 298 K) by radical scavengers,⁶⁶ e.g., 5,5-dimethyl-1-pyrroline *N*-oxide (DMPO), under continuous 365 nm irradiation resulted in failures in identifying the corresponding spin adducts based on HPLC-MS analysis (Fig. S7, ESI†).

As theoretical proofs, the optimized geometry of **1SDR** was obtained by the broken-symmetry wavefunction method (for computational details, see the ESI†).^{67,68} Spin density analysis (by using Multiwfn 3.8dev⁶⁹ and visualization by using VMD 1.9.3⁷⁰) support the existence of single electrons with opposite spin on the two bond-broken carbon atoms (Fig. 3d). It is also calculated that the Gibbs free energy of **1SDR** is lower than that of the triplet diradical **1TDR**, exhibiting a prominent singlet-triplet energy gap, $\Delta E_{\text{S-T}} = -3.5$ kcal mol⁻¹, and $S^2 = 0.32$ at the M06-2X/6-31G** level of theory (by using Shermo 2.3,⁷¹ Fig. 3d and Tables S9 and S14, ESI†). The predicted λ_{max} for the absorption of **1SDR** in the visible region ($\lambda_{\text{max}}^{\text{calc.}} = 517$ nm and $f = 0.355$) at the TD-UM06-2X/6-31G** level of theory⁷² also matches well with the experimental value, $\lambda_{\text{max}}^{\text{exp.}} = 550$ nm (Fig. 3g, Table S11, ESI†). According to the most stable electronic configuration



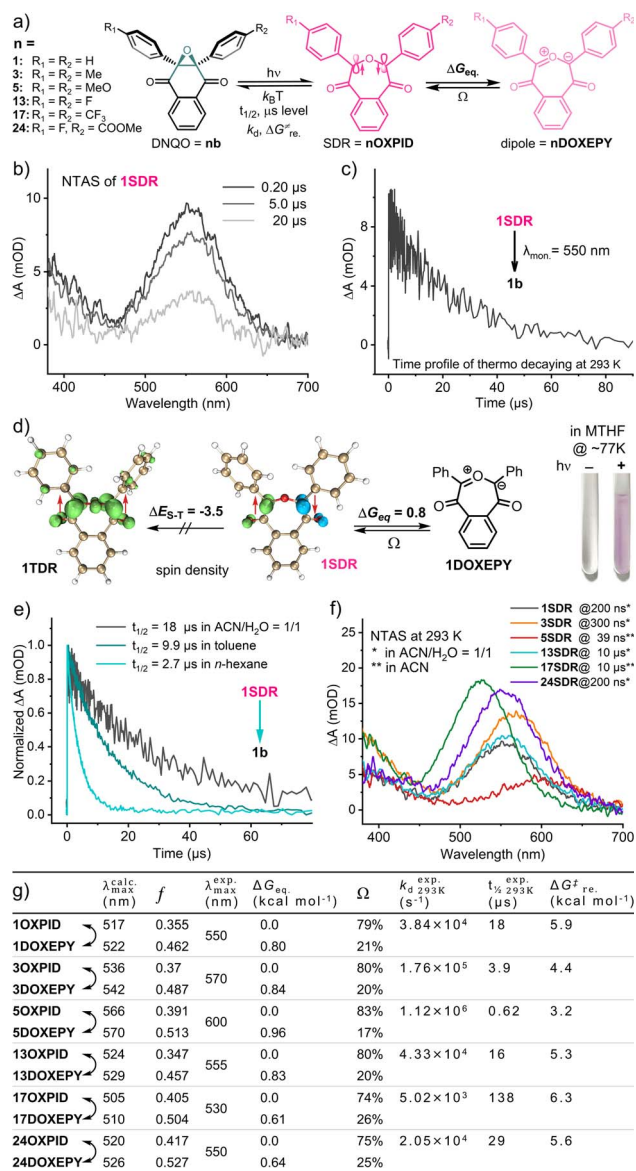


Fig. 3 (a) The scheme for the T-type photoswitching of DNQOs and the substitution effect on the tautomerization of SDRs \rightleftharpoons DOXEPIs in $\text{CH}_3\text{CN}:\text{H}_2\text{O} = 1:1$ at 293 K. (b) Transient absorption spectrum of **1SDR** at various delay times, generated from the flash photolysis of **1b** ($\lambda_{\text{ex.}} = 355$ nm). (c) Time-dependent decaying profile of **1SDR** ($\lambda_{\text{mon.}} = 550$ nm). (d) Visualization of the spin density analysis (isovalue = 0.01), the relative energies for **1TDR** \rightleftharpoons **1SDR** \rightleftharpoons **1DOXEPI**, and the photograph of *in situ* formed **1SDR** in an NMR tube. (e) Transient decaying traces of **1SDR** in various solvents and corresponding half-life. (f) Transient absorption spectra of the selected SDRs at 293 K. (g) The tabular parameters for the spectral characters of the transient species and their thermodynamic properties.

in the ground-singlet-state, SDRs can be classified into two types. Type-I has the symmetric orbital ψ_S as the highest occupied molecular orbital (HOMO) and the antisymmetric orbital ψ_A as the lowest unoccupied molecular orbital (LUMO).⁵⁹ In contrast, type-II adopts ψ_A as the HOMO and ψ_S as the LUMO. CASSCF calculations at the CAS(2,2)/6-31G** (SMD, ACN : H₂O = 1 : 1) level of theory indicated that **1SDR** is a type-II SDR with the HOMO of antisymmetric ψ_A (Fig. 4a). The coefficients (*c*) of

electronic configurations I and III were 0.92 and -0.38 , respectively. Therefore, the numbers of occupied electrons on the ψ_A and ψ_S orbitals were $1.69 (= 2 \times 0.92^2)$ and $0.29 [= 2 \times (-0.38)^2]$, respectively (Fig. 4a). And the diradical character y of **1SDR** was determined to be 0.29.

Taking the seven-membered cyclic dipole (**1DOXEPI**, Fig. 3a) into account, we subsequently reassessed the absorption spectrum and decaying of the transient **1SDR** in various solvents because the predicted absorbance maximum of **1DOXEPI** ($\lambda_{\text{max}}^{\text{calc.}} = 522$ nm, $f = 0.462$, Fig. 3g) is also in good agreement with the experimental value. As a result of the solvent effect,⁶⁶ the lifetime of **1SDR** was notably prolonged in more polar solvent (Fig. 3e) implying that the radical-radical σ -coupling energy barrier ($\Delta G_{\text{re.}}^{\ddagger}$) increased, but the $\lambda_{\text{max}}^{\text{exp.}}$ was almost unchanged (Fig. S5a, ESI[†]). The higher $\Delta G_{\text{re.}}^{\ddagger}$ supports the presence of **1DOXEPI**, which is calculated to be in equilibrium *via* electronic tautomerism with **1SDR** (Fig. 3d) and stabilized by polar solvents (Fig. S5b, ESI[†]).⁵⁹ This absorbance peak ($\lambda_{\text{max}}^{\text{exp.}} = 550$ nm, Fig. 3b) was assigned to the characteristic $\pi-\pi^*$ (HOMO-LUMO) electronic transition of **1SDR** and its electronic tautomer **1DOXEPI** after comparing with their TD-DFT results. The major electronic transitions were located at 517 nm ($f = 0.355$) and 522 nm ($f = 0.462$, Table S11, ESI[†]), respectively. Based on their free energy difference ($\Delta G_{\text{eq.}} = 0.80$ kcal mol⁻¹, Fig. 3d), the Boltzmann distribution between **1SDR** and **1DOXEPI** (Ω , Fig. 3g and Table S11, ESI[†]) at 298 K was obtained, indicating a component of 79% **1SDR** *versus* 21% **1DOXEPI**. And, this distribution is constant during the thermal decay of **1SDR** \rightarrow **1b** ($k_d = 3.8 \times 10^4$ s⁻¹) because the electronic tautomerism is an ultrafast equilibrium with no transition states⁷³ and only the reconstitution in the electronic structure. At about 77 K, we were able to photograph the pink-coloured **1SDR** and its thermo-relaxation process in a MTHF matrix, clarifying the T-type photoswitching nature (Fig. 3d).

In view of the apparent substitution effect on the cycloaddition yield (Fig. 2), we further investigated the properties of the selected SDRs (Fig. 3f). With strong electron-donors (eDs), the absorption maximum of **5SDR** was red-shifted to 600 nm in contrast to **17SDR** with strong electron-acceptors (eAs) showing a hypochromic shift to 530 nm. And these experimental spectra were consistent with the predicted spectra obtained *via* TD-DFT calculation ($\lambda_{\text{max}}^{\text{exp.}}$ vs. $\lambda_{\text{max}}^{\text{calc.}}$, Fig. 3g). Accordingly, the thermal ring-contraction rate (k_d) of **3SDR** and **5SDR** with eDs accelerated to 1.73×10^5 s⁻¹ (4.6-fold) and 1.12×10^6 s⁻¹ (29-fold), respectively, while that of **17SDR** with eAs decelerated to 5.02×10^3 s⁻¹ (0.13-fold, Fig. 3g and S3, ESI[†]) by using **1SDR** as a benchmark. As the transition states (TS) searching explicated (Table S11, ESI[†]), the thermodynamic barrier ($\Delta G_{\text{re.}}^{\ddagger}$) for the relaxation of SDR \rightarrow DNQO descended from 6.3 to 3.2 kcal mol⁻¹ along with a substituent order from eAs (**17SDR** > **1SDR** > **13SDR**) to eDs (**3SDR** > **5SDR**), which was also in conformity to the experimental $t_{1/2}^{\text{exp.}}$ (Fig. 3g). By comparing the $\Delta G_{\text{eq.}}$ with the $\Delta G_{\text{re.}}^{\ddagger}$ in Fig. 3g, we speculate that the thermal decaying observed in the NTAS was presumably rate-determined by the radical-radical σ -coupling process of SDR \rightarrow DNQO rather than the faster tautomerization between SDR \rightleftharpoons DOXEPI. Since there is only small variation in the $\Delta G_{\text{eq.}}$ of the equilibrium of SDR \rightleftharpoons DOXEPI among various



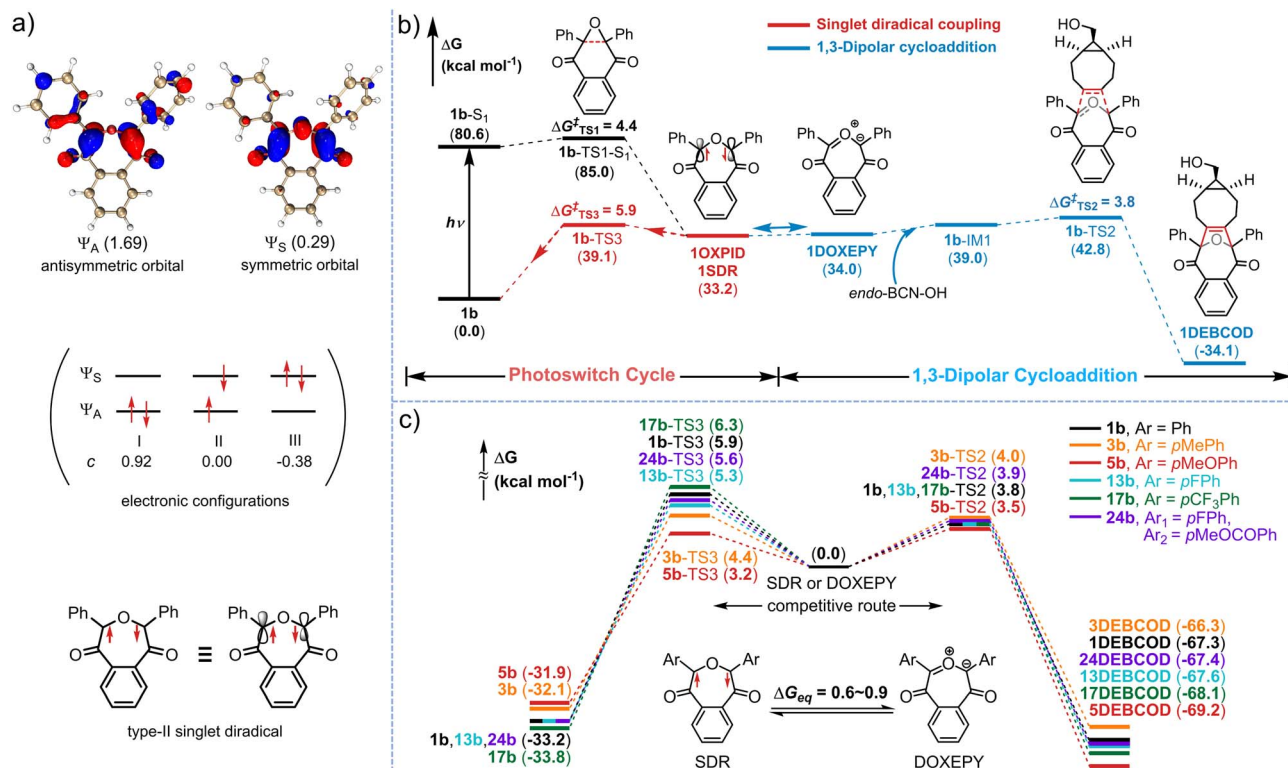


Fig. 4 (a) The molecular orbitals of the two active spaces (ψ_A and ψ_S) of **1SDR** are shown (isovalue = 0.05) with the values of the occupation numbers of the two electrons displayed in parentheses. The three electronic configurations (I–III) with the corresponding coefficients “c” are also given. (b) Gibbs free energy profile for the TS and intermediates in the photoswitching and the cycloaddition reaction pathways. The red pathway is the radical σ -coupling of **1SDR**, and the blue pathway is the cycloaddition between BCN-OH and **1DOXEPEY**. (c) Free energy barrier of the σ -coupling of SDRs (ΔG_{TS3}^\ddagger , left) vs. the 1,3-dipolar cycloaddition between BCN-OH and DOXEPEYs (ΔG_{TS2}^\ddagger , right) with various substituents.

substitutions, the Boltzmann distribution was always dominated by the SDRs (a content of 74–83%, Fig. 3g).

DFT calculation on the competitive mechanism of the photoswitch cycles versus the cycloaddition

To decipher the mechanism of the photoclick process mediated via the photoswitchable **1SDR**, further studies on the photochemical pathways were performed via (TD-)DFT (Fig. 4b). Once **1b** was excited to the first singlet state (**1b-S₁**) by 365 nm light (both $S_0 \rightarrow S_1$ and $S_0 \rightarrow S_2$ transitions), **1SDR** could be obtained as a converged structure via a homolytic cleavage of the C–C bond in the epoxy via **1b-TS_{1-S₁}** with a free energy barrier (ΔG_{TS1}^\ddagger) of 4.4 kcal mol⁻¹ (Fig. 4b). Without BCN-OH, **1SDR** would primarily undergo a thermodynamic radical–radical coupling, returning to **1b** via transition state **1b-TS₃** ($\Delta G_{TS3}^\ddagger = \Delta G_{re.\ddagger} = 5.9$ kcal mol⁻¹) to complete an ultrafast photoswitch cycle. In the presence of BCN-OH, a competitive pathway leading to a 1,3-dipolar cycloaddition between BCN-OH and 1,3-dipole (**1DOXEPEY**, as the electronic tautomer of **1SDR**) was involved, which affords the final cycloadduct **1DEBCOD** via transition state **1b-TS₂**. Notably, the ΔG_{TS2}^\ddagger of the cycloaddition step was substantially lower than that of the radical coupling process (3.8 vs. 5.9 kcal mol⁻¹), suggesting that BCN-OH is capable of capturing the transient **1DOXEPEY** if [BCN-OH] is given at 0.5 mM (theoretical $k_d/k_{2CA} = 58$). Considering the

bioorthogonality of these reactive intermediates, we have further searched for the nucleophilic addition pathways of the 1,3-dipole (**1DOXEPEY**) with respect to thiols, e.g., glutathione (GSH, Fig. S23 and Table S9, ESI[†]). As a result, the energy barrier in the nucleophilic addition by GSH was incredibly higher than that of the ring-strain promoted cycloaddition with BCN-OH (17.6 vs. 3.8 kcal mol⁻¹) and therefore implies a superior chemoselectivity toward the cycloaddition.

To understand the electronic effects on the efficiency of the cycloaddition, five exemplary DNQOs (**3b**, **5b**, **13b**, **17b** and **24b**, Fig. 4c) were subjected to mechanism interpretation at the same level of theory. The computational results illustrated that the introduction of various substituents (either eD or eA) displayed comparable energy barriers during the ring-strain promoted cycloaddition step (ΔG_{TS2}^\ddagger , 3.5–4.0 kcal mol⁻¹, Table S12, ESI[†]). However, the energy barrier associated with the radical–radical coupling processes (ΔG_{TS3}^\ddagger) became differentiated. The eA groups (such as trifluoromethyl in **17SDR**) dramatically elevated the $\Delta G_{TS3}^\ddagger = 6.3$ kcal mol⁻¹, prolonging the half-life of the SDR and facilitating the cycloaddition with BCN-OH ($\Delta G_{TS2}^\ddagger = 3.8$ kcal mol⁻¹, Fig. 4c). In contrast, the eD group, e.g., fluorine atoms in **13SDR**, methyl groups in **3SDR** and methoxyl groups in **5SDR**, lowered the ΔG_{TS3}^\ddagger in the radical-coupling by 0.6, 1.5 and 2.7 kcal mol⁻¹, respectively, shortening the half-life of the SDRs. Especially for **5SDR** with the shortest half-life ($t_{1/2} = 620$ ns, Fig. 3g), the ΔG_{TS3}^\ddagger of 3.2 kcal mol⁻¹ was even slightly lower



than that in the cycloaddition step ($\Delta G_{\text{TS2}}^\ddagger = 3.5 \text{ kcal mol}^{-1}$), leaving few chances for trapping the SDR *via* the cycloaddition (theoretical $k_d/k_{2\text{CA}} = 3320$ when $[\text{BCN-OH}] = 0.5 \text{ mM}$). The theoretical calculation was consistent with the experimental observations that DNQOs with eD groups showed very poor photo-conversion (**4b–7b**, Fig. 2c). Upon further calculation, there was no significant alteration found in both the orbital characteristics and the electron occupied numbers by the various substituents in the five SDRs (Fig. S24, S25 and Table S13, ESI†). The relatively long-lived SDRs⁵⁵ were likely stabilized by both the carbonyl and the diaryl moieties, thus enabling the cycloaddition with BCN-OH to establish an ultrafast photoclick reaction.

Key parameters for the photoswitching and the photoclick reaction

As a proof-of-concept for the photoclick strategy utilizing T-type photoswitchable SDRs, the quantum yield (Φ_{max}) and the light-to-chemical-energy conversion efficiency (η) for the photochemical transformation from DNQO to SDRs as well as the cycloaddition rate ($k_{2\text{CA}}$) are essential parameters. Since the apparent quantum yield (Φ) for a photoswitchable intermediate is dependent on the concentration of $[\text{BCN-OH}]$ under given irradiation conditions, we were able to derive both the Φ_{max} and the $k_d/k_{2\text{CA}}$ (based on experimental studies) by varying $[\text{BCN-OH}]$, according to reported methods (Fig. S8 and S9, ESI†).⁷⁴ Herein, the k_d is directly obtained by monitoring the decay of SDRs from NTAS measurements. Then, the $k_{2\text{CA}}$ is calculated based on the competition reactions between the thermal decay and the cycloaddition of DOXEPEY. As an energetic SDR molecule, the η is the ratio of the potential energy difference ($\Delta G_{\text{SDR-DNQO}}$) between DNQO and the “metastable” SDR harvested during the photochemical conversion with a quantum yield (Φ_{max}) *versus* the incident photon energy ($E_{\text{light}} = h\nu N_A$, Table S15, ESI†).⁷⁵ These parameters for the photoclick reaction between the five DNQOs and BCN-OH are summarized in Table 1, except **5b** due to a very poor reactivity. Firstly, we found that the [3 + 2] cycloaddition rates between DOXEPEYs and the ring-strained BCN-OH are ultrafast, ranging from $4.86 \times 10^7 \text{ M}^{-1} \text{ s}^{-1}$ for unsymmetric **24b** to $1.01 \times 10^9 \text{ M}^{-1} \text{ s}^{-1}$ for **13b** with *p*-F-phenyl. The cycloaddition rates at this level imply promising kinetics to ligate to extremely diluted BCN reporters on biomolecules. Regardless of the substitutions on the diaryl, all the SDRs hold a high and

consistent $\Delta G_{\text{SDR-DNQO}}$ energy, about $32.1\text{--}33.8 \text{ kcal mol}^{-1}$. Therefore, a higher η depends mainly on a better Φ_{max} . Symmetric *p*-FPh (**13b**), *p*-CF₃Ph (**17b**) on 1,3-positions of DNQO and unsymmetric *p*-FPh combined with *p*-COOMePh (**24b**) terminals were favourable to a high Φ_{max} ($= 0.56, 0.58$ and 0.76) in the photo-ring-opening step, and accordingly the value of η can be gained as 24%, 25% and 33%, respectively. Thus, the light irradiation not only manages the process of the photoclick reaction, but its energy is also efficiently converted to the chemical energy of the long-lived SDR and DOXEPEY during the T-type photoswitching, powering an ultrafast cycloaddition with BCN-OH.

Evaluation of the chemoselectivity and the stability of DNQOs in a biological medium

In a complex living context, photolysis or quenching of the photo-generated intermediates (type-I, Fig. 5a) by the abundant

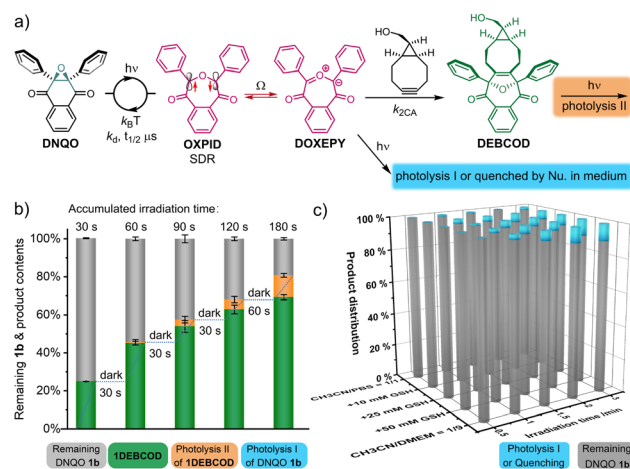


Fig. 5 Studies on the chemoselectivity of the photoclick reaction between **1b** with BCN-OH upon 365 nm LED irradiation at 298 K under biocompatible conditions. (a) The schematic illustration. (b) Percentage distribution of each component after the photoclick reactions between **1b** ($40 \mu\text{M}$) + BCN-OH ($500 \mu\text{M}$) with GSH (5.0 mM) in $\text{CH}_3\text{CN}/\text{PBS} = 1/1$. (c) The remaining **1b** and photolysis products after the irradiation in the absence of BCN-OH with various concentrations of GSH in $\text{CH}_3\text{CN}/\text{PBS} = 1/1$ or without GSH in $\text{CH}_3\text{CN}/\text{DMEM} = 1/9$ with 10% FBS + 1% antibiotics. DMEM = Dulbecco's modified Eagle's medium.

Table 1 Comparative key parameters for the photoclick reaction between the representative DNQOs and BCN-OH^a

DNQO	Φ_{max}	$k_d/k_{2\text{CA}}$ (mM)	$k_{2\text{CA}}$ ($\text{M}^{-1} \text{ s}^{-1}$)	$t_{1/2}^b$ (μs)	$\Delta G_{\text{SDR-DNQO}}$ (kcal mol^{-1})	η (%)
1b	0.25	0.114	3.38×10^8	4.1	33.2	10.6
3b	0.19	0.278	6.32×10^8	2.2	32.1	7.8
5b	N.A.	N.A.	N.A.	N.A.	31.9	N.A.
13b	0.56	0.043	1.01×10^9	1.4	33.2	23.7
17b	0.58	0.031	1.62×10^8	8.6	33.8	25.0
24b	0.76	0.422	4.86×10^7	29	33.2	33.1

^a $20 \mu\text{M}$ DNQOs upon 365 nm LED irradiation for a specific period of time (3–28 s, Fig. S8, ESI) in $\text{CH}_3\text{CN}:\text{H}_2\text{O} = 1:1$ at 298 K in the presence of $[\text{BCN-OH}] = 0.4 \text{ mM}, 0.6 \text{ mM}, 1.0 \text{ mM}$ or 1.6 mM , respectively. ^b The half-life of DOXEPEY in the pseudo-first-order cycloaddition reaction, when $[\text{BCN-OH}] = 500 \mu\text{M}$. N.A. = not available.



chemicals before coming into contact with desired reporters are a major reason for undesired side-reactions in photoclick chemistry. The reversion of the reactive **1SDR** \rightleftharpoons **1DOXEPEY** back to **1b** *via* the superfast σ -coupling provides an alternative pathway to circumvent these undesired processes, and the SDR is naturally insensitive to $^3\text{O}_2$. To evaluate the biocompatibility (Fig. 5a), we rescreened the DNQO-BCN photoclick reactions by moving to a biological mimicking medium containing both 10.0 mM GSH and nucleophilic natural amino acids (1.0 mM each, Fig. S10, ESI †). Without BCN-OH, the concentration of DNQOs (**1b**, **13b**, **15b** and **24b**) was almost unchanged after incubation in the dark for 24 h or under 365 nm irradiation for 60 s, verifying a superior photostability of the DNQOs against a variety of nucleophiles as well as an excellent recovery through photoswitching cycles. Afterward, temporally controlled cycloadditions were then realized in the presence of BCN-OH by using incremental irradiation with the cycloadduct yield increased along the accumulation of the irradiation time (Fig. 5b and S11,

ESI †). Although the formation of by-products became notable after prolonged irradiation (0–14.3% for 30–180 s, the orange columnar segments, Fig. 5b), they were attributed to the photolysis of the desired cycloadducts (type-II, Fig. 5a) rather than the type-I photolysis or nucleophilic quenching of the SDR or DOXEPEY (Fig. S12, S13, ESI †). To further monitor type-I photolysis, BCN-OH has to be omitted from the mixture. However, by-products produced by type-I photolysis after 180 s exposure in simply $\text{CH}_3\text{CN}/\text{PBS} = 1/1$ mixed solvent remain negligible. Furthermore, a high concentration of GSH (10–50 mM) as well as mammalian cell culture medium were included to reassess the photostability of **1SDR** (Fig. 5c and S14–S18, ESI †). Gratifyingly, the remaining **1b** could be kept up to 80% in the presence of 50.0 mM GSH (almost saturated solution) or even in the Dulbecco's modified Eagle's medium (DMEM with 10% fetal bovine serum + 1% antibiotics) after 3.0 min irradiation of a 365 nm LED. It is worth noting that DNQO **1b** did not deteriorate after storage at 298 K for more than 1 year or under strong

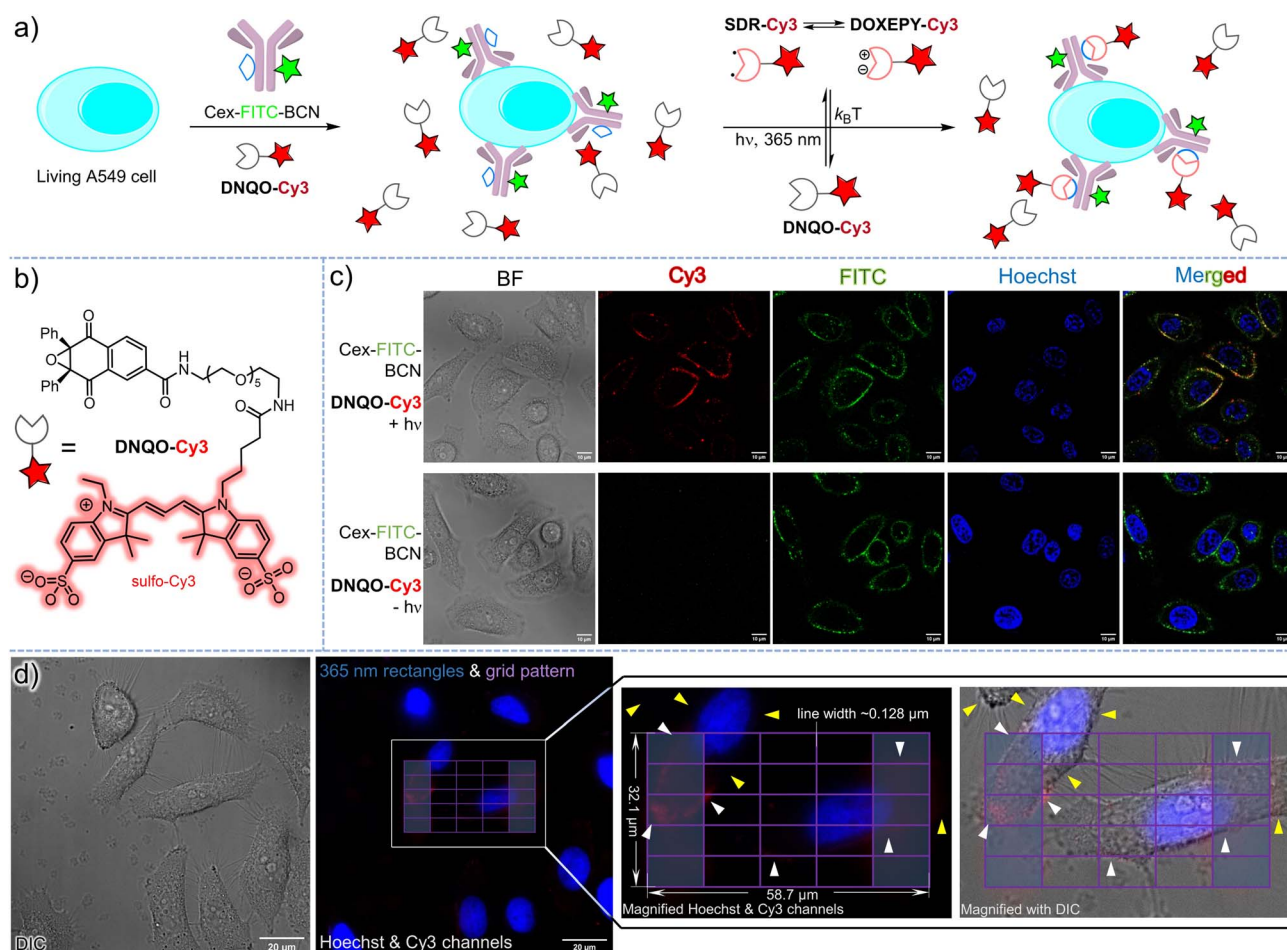


Fig. 6 Spatiotemporally resolved fluorescence labeling on the membrane of living mammalian cells. (a) Schematic illustration for the photoclick labeling procedure on the cell membrane *via* 365 nm light (90 s) to induce the photoswitching of **DNQO-Cy3** \rightarrow **DOXEPEY-Cy3** (total 20 μM); (b) chemical structure of the tailored **DNQO-Cy3** probe; (c) confocal microscopic imaging to identify the colocalization of both Cy3 and FITC signals on cell surface, Hoechst-33342 staining for nucleus of the living cells, BF = bright field, and scale bar = 10 μm ; (d) the merged fluorescence imaging of the cells after exposure to the patterned 365 nm LED stimulation (purple colour) projected through the DMD (*via* Polygon 400, Mightex, 25% 2.0 W LED light power) for 30 s, with the white arrows pointing to the labelled membrane segments and the yellow arrows to the membrane segments outside the stimulation pattern.



acid treatment for 5 h ($\text{CF}_3\text{COOH}:\text{CH}_2\text{Cl}_2 = 1:1$) or even after heating to 100 °C for 10 h in DMF, making it a robust precursor for photoclick ligation in living systems.

Spatiotemporally resolved fluorescence labeling on the membrane of living cells *via* the photoclick reaction

To explore the bioorthogonal decoration on the cell surface *via* the T-type photoswitchable SDR (Fig. 6a), we then tailored a water-soluble fluorophore conjugate (**DNQO-Cy3**, Fig. 6b) in which a sulfo-Cy3 was linked to the core of **1b** *via* oligo ethylene glycol to avoid cell permeability. To incorporate the BCN reporter onto the membrane of a living A549 cancer cell *via* immuno-recognition toward the epidermal growth factor receptor (EGFR+), a chemically modified chimeric monoclonal antibody, cetuximab-FITC-BCN ($20 \mu\text{g mL}^{-1} = 0.13 \mu\text{M}$ Cex-FITC-BCN, Fig. S19, ESI†) was attached *via* incubation for 60 min. After exposure to a 365 nm LED for 90 s with the treatment of $20 \mu\text{M}$ **DNQO-Cy3** in air (Fig. 6c), the concomitant appearance of both Cy3 and FITC fluorescence was observed on the cell surface, colocalizing precisely (Fig. 6c, upper row), suggesting a temporal controllability of this photoclick reaction. Without the exposure, there was no Cy3 signal detected on the cell surface but only the FITC signal was observed due to binding of Cex-FITC-BCN (Fig. 6c, lower row). Without the treatment with Cex-FITC-BCN, both Cy3 and FITC signals were negligible even after identical exposure (for other stringent control conditions, Fig. S20, ESI†).

Encouraged by the ultrafast reaction rates and the high chemoselectivity of the **DNQO-BCN** photoclick reaction, we attempted spatial-resolved fluorescence labeling of cetuximab-BCN on single or even partial cell membranes. To achieve lithographic patterning with sub-micron resolution, we programmed a 30 s continuous 365 nm illumination sequence *via* a digital microarray device (DMD) embedded in an epifluorescence microscope through a $100\times$ objective lens. Meanwhile a lithographic pattern of a grid matrix ($58.7 \times 32.1 \mu\text{m}$, 6×6 purple lines, Fig. 6d) with two rectangle columns at the terminal (light blue regions) filled was drawn as the input shape. As a result, the **DNQO-Cy3** probe in PBS ($20 \mu\text{M}$ and $\text{pH} = 7.4$) was photo-switched on to generate **SDR-Cy3** within the stimulation region, and subsequently conjugated toward the bound cetuximab-BCN (incubated at $120 \mu\text{g mL}^{-1}$ in DMEM for 1 h) in proximity (Fig. 6d, for details, refers to Fig. S21, ESI†). Unreacted **SDR-Cy3** was mostly recovered to **DNQO-Cy3** instantaneously with minimal molecule diffusion because of its μs level half-life. Consequently, most of the cell membrane segments overlaying within the grid-patterned region were covalently marked with the Cy3 fluorophore, resulting in curve-shaped signals (white arrows). But the unexposed portion of the membranes (yellow arrows) scarcely showed any signal, indicating a subcellular spatial-resolving power of the photoclick reaction. However, off-site labeling could also be observed (Fig. S21, ESI†), which might be ascribed to scattering of the light-ray in the specimen or dynamic change of the membrane morphology during the extensive washing procedure or unexpected off-target labeling.

Conclusions

Inspired by the transient SDR intermediate produced from the photoactivation of DNQO, we present a novel T-type photo-switch based on the homolytic ring-expansion in association with the counteracting thermal σ -coupling of the SDR in dynamic cycles. The unique seven-membered cyclic OXPID (SDR) as well as its long-lived character revealed by spectroscopic and computational characterization constitute an efficient photo-energy transducer, which tautomerize into an energetic 1,3-dipole, DOXEPEY. Therefore, the energy harvesting nature of the T-type molecular photoswitch makes the OXPID a highly useful species in a limited repertoire of SDRs that can drive an ultrafast cycloaddition with *endo*-BCN-OH. In this way, we established a type of bioorthogonal photoclick reaction for spatiotemporal labeling on living cells, which is underpinned by the unique features of the SDR, including insensitivity to triplet species and regeneration without capturing by BCN-OH. The facile two-step synthesis of DNQO facilitates an easy access for versatile utilities of the photoswitchable SDR in photoclick chemistry. Currently, the requirement of 365 nm UV light to induce the photoswitching is the limiting factor. Therefore, research on visible-light-triggered bioorthogonal reactions with temporal and spatial control by using DNQO is now underway in our lab.

Data availability

Additional figures as described in the main text, all experimental procedures, synthetic procedures, copies of spectral data for the compounds, data processing, and computational details are available in the ESI.†

Author contributions

F. H. and Z. Y. initiated the idea, designed the experiments, and wrote and revised the manuscript. F. H. synthesized the main compounds and conducted all of the experiments. C. Z., Z. S., and C. H. carried out the theoretical calculations. Z. L., X. X., X. Z., J. F., and B. L. helped with compound synthesis and *in vitro* cell experiments. Y. L. assisted with the NTAS experimental studies. All authors contributed to the manuscript.

Conflicts of interest

There are no conflicts to declare.

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