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Transient self-assembly of metal–organic complexes†

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Implementing transient processes in networks of dynamic molecules holds great promise for developing new functional behaviours. Here we report that trichloroacetic acid can be used to temporarily rearrange networks of dynamic imine-based metal complexes towards new equilibrium states, forcing them to express complexes otherwise unfavourable in their initial equilibrium states. Basic design principles were determined for the creation of such networks. Where a complex distribution of products was obtained in the initial equilibrium state of the system, the transient rearrangement temporarily yielded a simplified output, forcing a more structured distribution of products. Where a single complex was obtained in the initial equilibrium state of the system, the transient rearrangement temporarily modified the properties of this complex. By doing so, the mechanical properties of an helical macrocyclic complex could be temporarily altered by rearranging it into a [2]catenane.

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Introduction

To rival the capabilities of biological systems, temporal control over chemical reactivity must be achieved in synthetic systems. Most synthetic self-assembly processes are designed to generate well-ordered structures with high thermodynamic or kinetic stabilities—these structures being at the global minimum of the energy landscape or trapped in local minima.¹ By modifying the energy landscape using external stimuli (such as pH, light, or the addition of a chemical species) to create a new minimum, these structures could be forced to rearrange into new ones, yielding stimuli-responsive self-assembly processes.² While this approach produces highly functional systems,³ it requires the repeated intervention of an operator to administer opposing stimuli at appropriate times to switch the system back and forth between its different functional states.

To overcome this limitation and inspired by biological systems,^{1b,4} chemists have coupled self-assembly and energy-dissipating processes so that self-assembly processes could transiently express different structures *via* an influx of energy in the form of light, heat or chemicals.^{1b,5} These so-called “transient self-assemblies” require a constant input of energy to persist in time. If the energy supply is stopped, these structures dismantle, their components being recaptured by the initial

state of the system (*i.e.*, its state prior to the energy influx). Interest in chemically-induced transient self-assembly is rapidly growing encouraged by examples of their unique capabilities, including transient gel formation,⁶ operation of molecular machines,⁷ temporal control over host–guest systems,⁸ transient formation of supramolecular assemblies⁹ and non-equilibrium self-replication.¹⁰ In most current reports, however, chemically-induced transient processes are mostly used to modulate the behaviour of single component self-assemblies. Few examples exist in which the consumption of a chemical reactant regulates the organization of networks of dynamic molecules.^{10c,11} Using classical stimuli-responsive self-assembly, networks of dynamic molecules have enabled the emergence of complex functions inaccessible with simpler systems.^{3a,d,3g,12,13} Combining the capabilities of such networks with energy-dissipating processes would enable the development of new adaptive behaviours, furthering our understanding of complex systems (including biological ones).

To date, libraries of dynamic metal–organic complexes consisting of amine and 2-formylpyridine components reversibly condensed into imine ligands around labile metal-ions have yielded some of the most advanced artificial networks of dynamic molecules.^{3a,d,3g,12g,12k,13c–i}

Here we demonstrate that a ‘transient acid’^{5h,7b–g,7i,8c,11f,12j} can force libraries of imine-based metal complexes to temporarily rearrange towards a new equilibrium state, generating complexes otherwise unfavourable in the absence of the acid. The study provides basic design principles for creating networks of dynamic complexes with temporarily modifiable functions, showing that the properties of metal–organic complexes can be altered by their transient rearrangement.

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Results and discussion

Rational and main design

Due to the differences of basicity between arylamine and alkylamine, we envisaged that trichloroacetic acid—an acid capable of temporarily acidifying a medium^{5h,7c,7d,i,11f,12j}—could be used to induce the transient rearrangement of imine-based metal complexes.

Here we will use the notation (**a**,**b**) to refer to the imine-based ligand obtained by the condensation of aldehyde **a** with amine **b**. The more basic ditopic amine **2** (Fig. 1A and B) should yield the most thermodynamically stable complex with Cu(I) and 2-formylpyridine **3** under neutral conditions, based on previous work.^{11f,13f,14} Complex $[\text{Cu}(\mathbf{3}_2, \mathbf{2})]^+$ should therefore be the unique complex observed when Cu(I) is added to an equimolar mixture of **1**, **2** and **3**. Upon addition of trichloroacetic acid, the initial acidification of the medium should displace **2** from the complex by protonation, allowing the incorporation of the less basic arylamine **1** (Fig. 2A and B). A new transient equilibrium state should quickly be achieved, expressing $[\text{Cu}(\mathbf{3}_1, \mathbf{1})_2]^+$ as its most thermodynamically stable complex. The slow decarboxylation of trichloroacetate^{5h,7c,7d,i,11f,12j} should then incrementally return the system of the metal complexes to its initial equilibrium state (*i.e.*, prior to the addition of acid) *via* a quasi-static process continually maintaining the metal complexes at equilibrium (the rearrangement of the metal complexes being faster than the decarboxylation of trichloroacetate). The decarboxylation of trichloroacetate generates a strong base—trichloromethyl anions—that should restore neutral alkylamine **2**, regenerating $[\text{Cu}(\mathbf{3}_2, \mathbf{2})]^+$ with chloroform and carbon dioxide as the only side products.

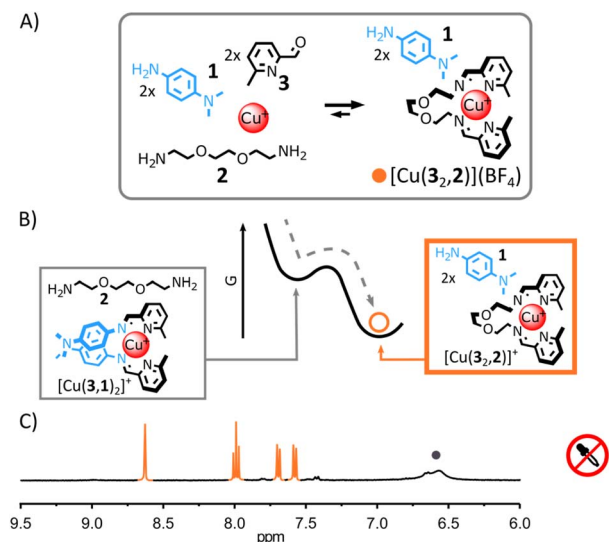


Fig. 1 (A) Self-assembly of $[\text{Cu}(\mathbf{3}_2, \mathbf{2})]^+$ under thermodynamic control. Reaction conditions: **1** : **2** : **3** : $\text{Cu}(\text{BF}_4)$ (**2** : **1** : **2** : **1**), CD_3CN , 60 °C, 18 h. (B) Schematic representation of the Gibbs free energy landscape of the reaction; $[\text{Cu}(\mathbf{3}_1, \mathbf{1})_2]^+$ only forms as a metastable intermediate during the self-assembly process. (C) Partial ^1H NMR spectrum (400 MHz, CD_3CN , 297 K) of the crude reaction mixture after 12 h at 60 °C; the diagnostic signals of $[\text{Cu}(\mathbf{3}_2, \mathbf{2})]^+$ are coloured in orange and two of the diagnostic signals of **1** are highlighted by a grey circle.

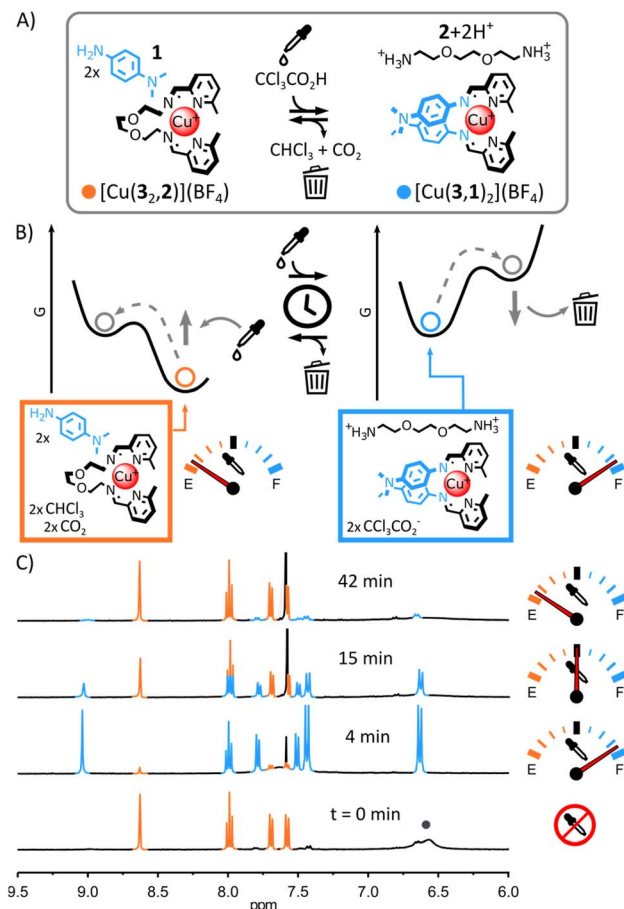


Fig. 2 (A) Transient rearrangement of $[\text{Cu}(\mathbf{3}_2, \mathbf{2})]^+$ into $[\text{Cu}(\mathbf{3}_1, \mathbf{1})_2]^+$. Conditions: CCl_3COOH (2.5 eq.), CD_3CN , r.t. (B) Schematic representation of the Gibbs free energy landscape of the rearrangement; $[\text{Cu}(\mathbf{3}_1, \mathbf{1})_2]^+$ forms under thermodynamic control as part of a quasi-static process and $[\text{Cu}(\mathbf{3}_2, \mathbf{2})]^+$ forms under thermodynamic control in the default equilibrium state of the system. (C) Partial ^1H NMR spectra (400 MHz, CD_3CN , 297 K) showing the evolution of the crude reaction mixture upon addition of CCl_3COOH ; the diagnostic signals of $[\text{Cu}(\mathbf{3}_2, \mathbf{2})]^+$ and $[\text{Cu}(\mathbf{3}_1, \mathbf{1})_2]^+$ are coloured in orange and blue, respectively, two of the diagnostic signals of **1** are highlighted by a grey circle.

This rational was tested. Combining alkylamine **2**, 2-formylpyridine **3** and Cu(I) in a **2** : **2** : **1** ratio in CD_3CN led to the quantitative formation of $[\text{Cu}(\mathbf{3}_2, \mathbf{2})]^+$ after 12 h at 60 °C (see ESI† for details and full characterization data of new compounds). When arylamine **1** (2 eq.) was added to the solution, a new equilibrium state was quickly achieved.¹⁵ $[\text{Cu}(\mathbf{3}_2, \mathbf{2})]^+$ remained the only complex observable by ^1H NMR spectroscopy (Fig. 1C and S17, ESI†) and electrospray ionization mass spectrometry (ESI-MS, ESI, Fig. S33†), hinting at its greater stability compared to $[\text{Cu}(\mathbf{3}_1, \mathbf{1})_2]^+$. Addition of trichloroacetic acid (2.5 eq.) to this stable mixture of $[\text{Cu}(\mathbf{3}_2, \mathbf{2})]^+$ and **1** immediately caused the rearrangement of the system, yielding $[\text{Cu}(\mathbf{3}_1, \mathbf{1})_2]^+$ and $[2+2\text{H}^+]$ as unique products observable by ^1H NMR spectroscopy (Fig. 2C and S24–S26, ESI†). The formation of $[\text{Cu}(\mathbf{3}_1, \mathbf{1})_2]^+$ was also confirmed by ESI-MS (ESI, Fig. S34†). After a few minutes, the signals of $[\text{Cu}(\mathbf{3}_1, \mathbf{1})_2]^+$ and $[2+2\text{H}^+]$ started to diminish giving place to those of $[\text{Cu}(\mathbf{3}_2, \mathbf{2})]^+$ and **1**—the decarboxylation of



trichloroacetate slowly returning the system to its initial state (ESI, Fig. S24–S26†). After *ca.* 40 min, the ^1H NMR spectra of the reaction mixture closely matched that prior to the addition of acid, differing only by the emergence of a strong signal at 7.58 ppm attributed to the side product CHCl_3 (Fig. 2C).

The robustness of the system was tested by subjecting the same sample to five consecutive rearrangement cycles (ESI, Fig. S26–31†). The evolution of the system was monitored by ^1H NMR spectroscopy, invariably showing behaviours identical to those observed during the first rearrangement cycle (ESI, Fig. S26–31†) and showing only minimal deterioration of the system even after five consecutive cycles (ESI, Fig. S32†). Having established that trichloroacetic acid could induce the transient rearrangement of a library of metal complexes—temporarily forming a complex unfavourable in the initial equilibrium state of the system—we sought to probe the tolerance of the system to modifications.

Structural tolerance of the design

The structural tolerance of the system shown in Fig. 2 was studied by systematic modification of the nature of (i) the alkylamine, (ii) the tetrahedral metal ion and (iii) the arylamine.

(i) Transient self-assembly of $[\text{Cu}(\mathbf{3},\mathbf{1})_2](\text{BF}_4)$ from a mixture of $\text{Cu}(\text{BF}_4)$, **4**, **3** and **1**. The substitution of ditopic alkylamine **1** for monotopic alkylamine **4** resulted in a distribution of homoleptic and heteroleptic complexes in the initial equilibrium state of the system (Fig. 3); however, the acid-induced rearrangement of the system remained unchanged. Upon addition of arylamine **1** (2 eq.) to $[\text{Cu}(\mathbf{3},\mathbf{4})_2]^+$ in CD_3CN , ^1H NMR signals corresponding to $[\text{Cu}(\mathbf{3},\mathbf{1})_2]^+$ and to the heteroleptic complex $[\text{Cu}(\mathbf{3},\mathbf{1})(\mathbf{3},\mathbf{4})]^+$ appeared besides those of $[\text{Cu}(\mathbf{3},\mathbf{4})_2]^+$ (Fig. 3B and S18 and S42, ESI†). The lack of the self-sorting in the initial equilibrium state of the system¹⁵ could be attributed to the reduced entropic driving force for the formation of $[\text{Cu}(\mathbf{3},\mathbf{4})_2]^+$ compared to the formation of $[\text{Cu}(\mathbf{3},\mathbf{2})_2]^+$ (loss of the chelate effect).^{14b,16} Adding trichloroacetic acid (2.5 eq.) to the system immediately simplified the reaction mixture, with only $[\text{Cu}(\mathbf{3},\mathbf{1})_2]^+$ and $[\mathbf{4}+2\text{H}^+]$ being observable by ^1H NMR spectroscopy (ESI, Fig. S35 and S36;† the formation of $[\text{Cu}(\mathbf{3},\mathbf{1})_2]^+$ was also confirmed by ESI-MS, ESI, Fig. S43†). This ‘simplified state’ persisted for a few minutes before slowly returning to the initial distribution of products (after *ca.* 40 min, ESI, Fig. S37†). This transient ‘simplified state’ could be recalled three times with little signs of fatigue (ESI, Fig. S27–41†). This system exemplifies how a network of dynamic complexes can be forced to temporarily express fewer products—a simplified output—by populating a different more structured equilibrium state *via* an energy-dissipating process.

(ii) Transient self-assembly of $[\text{Ag}(\mathbf{3},\mathbf{1})_2](\text{SbF}_6)$ from a mixture of $\text{Ag}(\text{SbF}_6)$, **3**, **2** and **1**. Upon addition of **1** to $[\text{Ag}(\mathbf{3},\mathbf{2})_2]^+$ and equilibration at 60 °C for 18 h, the increased plasticity of the coordination geometry of $\text{Ag}(\text{I})$ ¹⁷ yielded a distribution of complexes in the initial equilibrium state of the system (ESI, Fig. S19–21 and S49†), that immediately simplified upon addition of trichloroacetic acid. Following the addition of acid (2.5 eq.), $[\text{Ag}(\mathbf{3},\mathbf{1})_2]^+$ and $[\mathbf{2}+2\text{H}^+]$ were the sole

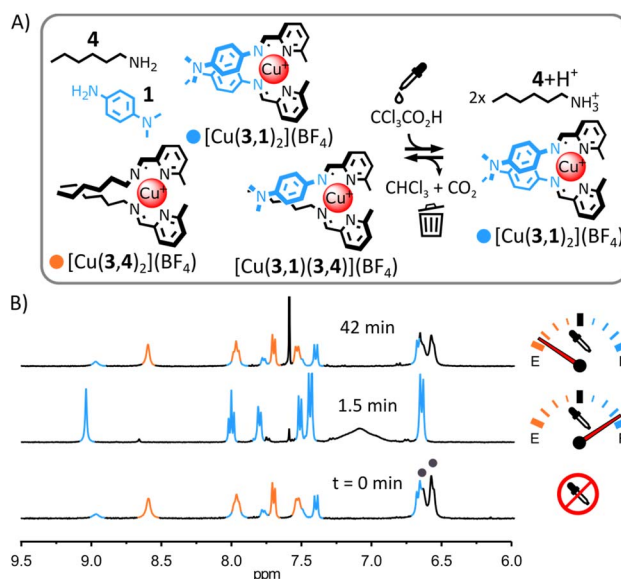


Fig. 3 (A) Transient ‘simplification’ of a mixture of complexes into $[\text{Cu}(\mathbf{3},\mathbf{1})_2]^+$. Conditions: CCl_3COOH (2.5 eq.), CD_3CN , r. t. (B) Partial ^1H NMR spectra (400 MHz, CD_3CN , 297 K) showing the evolution of the crude reaction mixture upon addition of CCl_3COOH ; the diagnostic signals of $[\text{Cu}(\mathbf{3},\mathbf{4})_2]^+$ and $[\text{Cu}(\mathbf{3},\mathbf{1})_2]^+$ are coloured in orange and blue, respectively, two of the diagnostic signals of **1** are highlighted by a grey circle, the diagnostic signals of $[\text{Cu}(\mathbf{3},\mathbf{1})(\mathbf{3},\mathbf{4})]^+$ overlap with those of $[\text{Cu}(\mathbf{3},\mathbf{1})_2]^+$ and $[\text{Cu}(\mathbf{3},\mathbf{4})_2]^+$.

products observed in solution by ^1H NMR spectroscopy (ESI, Fig. S44 and S45;† the formation of $[\text{Ag}(\mathbf{3},\mathbf{1})_2]^+$ was also confirmed by ESI-MS, ESI, Fig. S50†). The system gradually returned to its initial distribution of products over *ca.* 40 min (ESI, Fig. S46†). This process could be repeated with no visible deterioration (ESI, Fig. S46–48†), indicating the compatibility of $\text{Ag}(\text{I})$ with our transient rearrangement process. Our transient rearrangement process could also be operated from a non-equilibrated mixture of $[\text{Ag}(\mathbf{3},\mathbf{2})_2](\text{SbF}_6)$ and **1** (ESI, Fig. S51–S59†).

(iii) Transient self-assembly of $[\text{Cu}(\mathbf{3},\mathbf{5})_2](\text{BF}_4)$ from a mixture of $\text{Cu}(\text{BF}_4)$, **5**, **3** and **2**. The use of the less electron-rich anisidine **5** in place of 4-(dimethylamino)aniline **1** was uninfluential on the initial self-assembly of the system (ESI, Fig. S22 and S64†) or its transient rearrangement (Fig. 4 and S60 and S61, ESI†). From ^1H NMR spectra, $[\text{Cu}(\mathbf{3},\mathbf{2})_2]^+$ and **5** were the only species present initially and $[\text{Cu}(\mathbf{3},\mathbf{5})_2]^+$ and $[\mathbf{2}+2\text{H}^+]$ were the only species present upon addition of trichloroacetic acid (Fig. 4B and S62, ESI;† the formation of $[\text{Cu}(\mathbf{3},\mathbf{5})_2]^+$ was also confirmed by ESI-MS, ESI, Fig. S65†). However, in subsequent rearrangement cycles (Fig. 4B, top spectrum and ESI, Fig. S63†), **5** led to the rapid deterioration of the system *via* the trichloromethylation of imine (**3,5**) (ESI, Fig. S66†).¹⁸ This result indicates that in our original system (Fig. 2) the strong electron-donating *N,N*-dimethyl group of **1** must reduce the reactivity of imine (**3,1**) towards nucleophilic attack by the trichloromethyl anions generated by the decarboxylation of trichloroacetate.

Having established basic design principles, we envisaged that the transient rearrangement of metal–organic complexes



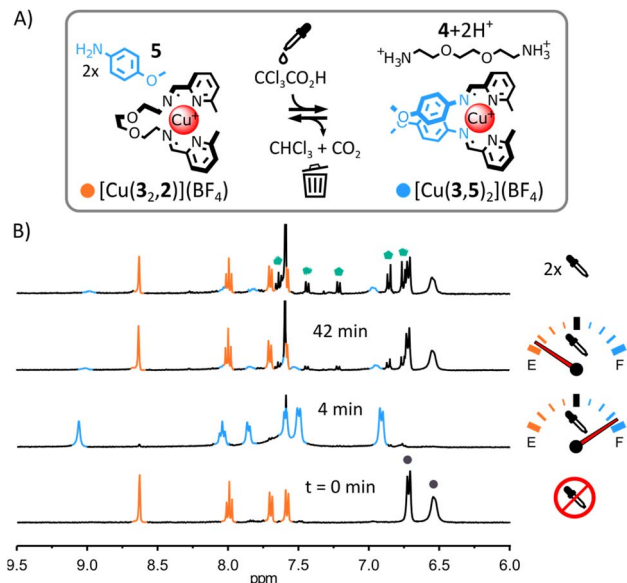


Fig. 4 (A) Transient rearrangement of $[\text{Cu}(\mathbf{3},\mathbf{2})]^+$ into $[\text{Cu}(\mathbf{3},\mathbf{5})]^+$. Conditions: CCl_3COOH (2.5 eq.), CD_3CN , r. t. (B) Partial ^1H NMR spectra (400 MHz, CD_3CN , 297 K) showing the evolution of the crude reaction mixture upon addition of CCl_3COOH and after two rearrangement cycles; the diagnostic signals of $[\text{Cu}(\mathbf{3},\mathbf{2})]^+$ and $[\text{Cu}(\mathbf{3},\mathbf{5})]^+$ are coloured in orange and blue, respectively, two of the diagnostic signals of $\mathbf{5}$ are highlighted by a grey circles, the diagnostic signals of trichloromethylated imine ($\mathbf{3},\mathbf{5}$) are highlighted by green pentagons.

described in Fig. 2 could be used to temporarily modify the properties of supramolecular architectures, enabling a macrocyclic complex to become a [2]catenane—two mechanically interlocked macrocycles.

Transient rearrangement of a macrocyclic complex into a [2]catenane

Dialdehyde **6** (Fig. 5A) is known to self-assemble with *p*-anisidine **5** and tetrahedral metal ions to produce dinuclear double-helical complexes (e.g., $[\text{Cu}_2(\mathbf{6},\mathbf{5})_2]^{2+}$).^{13b} Based on previous work by Nitschke,^{14b} we anticipated that both a macrocyclic complex and a [2]catenane could be generated from this helical motif if appropriate diamines were used in place of **5** (Fig. 5A).

In the presence of $\text{Ag}(\text{I})$ and aided by the gauche effect,¹⁹ dialkylamine **2** should have the right length and flexibility, to preferentially bridge two distinctive dialdehydes **6** while being incorporated in the helical complex, generating the dinuclear helical macrocycle $[\text{Ag}_2(\mathbf{6},\mathbf{2})_2]^{2+}$ (Fig. 5A). In contrast, the longer length and increased rigidity of diarylamine **7** should favour its condensation with the same dialdehyde **6** in the helical complex, yielding the dinuclear helical [2]catenane $[\text{Ag}_2(\mathbf{6},\mathbf{7})_2]^{2+}$ (Fig. 5A).

To confirm their architectures, both complexes were prepared by heating an equimolar mixture of dialdehyde **6**, diamine **2** or **7** and $\text{Ag}(\text{SbF}_6)$ in $\text{CD}_3\text{CN} : \text{CDCl}_3$ 3 : 1 at 60 °C for 12 h. In both cases, a single product was obtained with NMR spectra and ESI-MS analyses consistent with the structures shown in Fig. 5 (ESI, Section 2.4.2 and 2.4.3[†]). X-ray-quality crystals of both architectures were grown by liquid–liquid

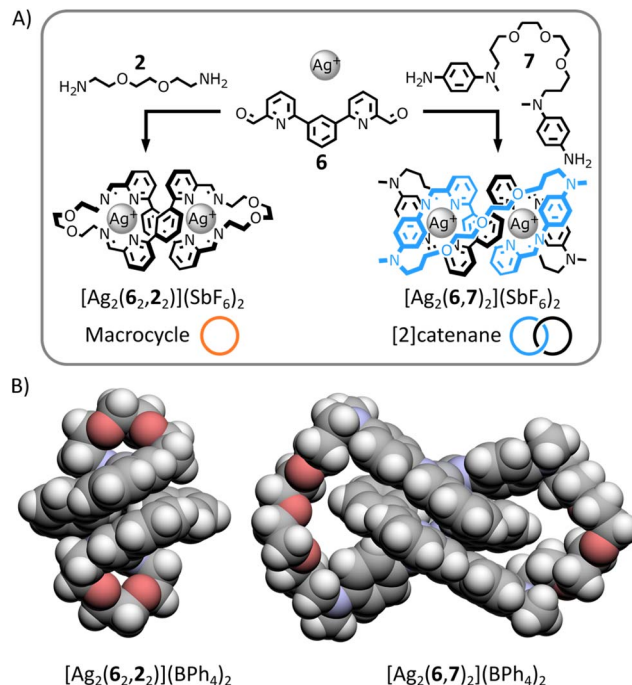


Fig. 5 (A) Self-assembly of macrocycle $[\text{Ag}_2(\mathbf{6},\mathbf{2})_2]^{2+}$ and [2]catenane $[\text{Ag}_2(\mathbf{6},\mathbf{7})_2]^{2+}$ under thermodynamic control. Reaction conditions: **6** : **2** : $\text{Ag}(\text{SbF}_6)$ (1 : 1 : 1) or **6** : **7** : $\text{Ag}(\text{SbF}_6)$ (1 : 1 : 1), $\text{CD}_3\text{CN} : \text{CDCl}_3$ 3 : 1, 60 °C, 12 h. (B) Single crystal X-ray structures of macrocycle $[\text{Ag}_2(\mathbf{6},\mathbf{2})_2](\text{BPh}_4)_2$ and of [2]catenane $[\text{Ag}_2(\mathbf{6},\mathbf{7})_2](\text{BPh}_4)_2$; solvent molecules and counterions have been omitted for clarity.

diffusion (ESI, Section 5.1 and 5.2[†]). The solid-state structures of both complexes unambiguously confirmed their topologies (Fig. 5B and ESI, Section 5.1 and 5.2[†]).

The organic ligand of macrocycle $[\text{Ag}_2(\mathbf{6},\mathbf{2})_2]^{2+}$ forms a 42-atom-loop twisted in a double-helical shape by two $\text{Ag}(\text{I})$ ions each coordinated to two iminopyridine domains of the ligand in a flattened tetrahedral geometry. The architecture of [2]catenane $[\text{Ag}_2(\mathbf{6},\mathbf{7})_2]^{2+}$ consists of two interlocked 36-membered rings entwined by an helical core created by two $\text{Ag}(\text{I})$ ions coordinating each to one of the iminopyridine domain of each macrocycles in an irregular four-coordinate geometry.

These two structures exemplify the importance of controlling crossing point connectivity while synthesizing interlocked architectures.²⁰ Both complexes have the same helical core, providing the same initial three crossing points. The difference of chemical topology between macrocycle $[\text{Ag}_2(\mathbf{6},\mathbf{2})_2]^{2+}$ and [2]catenane $[\text{Ag}_2(\mathbf{6},\mathbf{7})_2]^{2+}$ arises from the retention of a different number of these initial crossing points when connecting the end-groups of the helical core with diamine **2** or **7**. By connecting the closest end-groups of the helical core, diamine **2** retains none of the initial crossing points, yielding a topologically trivial macrocycle. By connecting the end-groups of the same strand of the helical core, diamine **7** retains one of the initial crossing points, yielding a [2]catenane.

Having established the nature of both complexes, we attempted the transient rearrangement of $[\text{Ag}_2(\mathbf{6},\mathbf{2})_2]^{2+}$ and **7** into $[\text{Ag}_2(\mathbf{6},\mathbf{7})_2]^{2+}$ and $[\mathbf{2} + 2\text{H}^+]$.



As shown in Fig. 6, when diarylamine **7** (1 eq.) was added to macrocycle $[\text{Ag}_2(\mathbf{6}_2, \mathbf{2}_2)]^{2+}$ in $\text{CD}_3\text{CN}:\text{CDCl}_3$ 3 : 1, ^1H NMR spectroscopy provided no evidence of reaction between these two species. This absence of reactivity combined with the ability of $[\text{Ag}_2(\mathbf{6}_2, \mathbf{2}_2)]^{2+}$ to selectively self-assemble from a mixture of $\text{Ag}(\text{SbF}_6)$, **7**, **6** and **2** (ESI, Fig. S23†) hint at the greater stability of $[\text{Ag}_2(\mathbf{6}_2, \mathbf{2}_2)]^{2+}$ over $[\text{Ag}_2(\mathbf{6}, \mathbf{7})_2]^{2+}$ in the initial equilibrium state of the system.²¹ Addition of trichloroacetic acid (10 eq.) triggered the transient rearrangement of the system: macrocycle $[\text{Ag}_2(\mathbf{6}_2, \mathbf{2}_2)]^{2+}$ was gradually converted into $[\text{Ag}_2(\mathbf{6}, \mathbf{7})_2]^{2+}$ and dialkylamine **2** was trapped in its protonated form (Fig. 6B, S67 and S68, ESI†). The concentration of $[\text{Ag}_2(\mathbf{6}, \mathbf{7})_2]^{2+}$ peaked at ca. 240 min before starting to decrease—slowly at first, but accelerating with depletion of the acid (ESI, Fig. S71 and S73†). After ca. 870 min, macrocycle $[\text{Ag}_2(\mathbf{6}_2, \mathbf{2}_2)]^{2+}$ was again the only complex visible by ^1H NMR spectroscopy.

Compared to the systems of mononuclear complexes in Fig. 2–4, the rate of rearrangement of this system was considerably lower ($[\text{Ag}_2(\mathbf{6}, \mathbf{7})_2]^{2+}$ peaked after ca. 240 min whereas $[\text{Ag}(\mathbf{3}, \mathbf{1})_2]^{2+}$ peaked after ca. 2 min), reflecting the increased complexity of this rearrangement process. Metal–organic complexes involving dialdehyde and diamine components are known to self-assemble *via* the initial formation of metastable ill-defined oligomers and polymers slowly rearranging into (kinetically or thermodynamically) more stable structures.^{19b,22} A similar scenario is likely when the present system is rearranging towards $[\text{Ag}_2(\mathbf{6}, \mathbf{7})_2]^{2+}$ following the addition of trichloroacetic acid. Indications of the formation of ill-defined oligomeric and polymeric species were found by

monitoring the rearrangement of the system by ^1H NMR spectroscopy. Despite $[\text{Ag}_2(\mathbf{6}_2, \mathbf{2}_2)]^{2+}$ and $[\text{Ag}_2(\mathbf{6}, \mathbf{7})_2]^{2+}$ being the only species visible by ^1H NMR spectroscopy throughout the rearrangement process (Fig. 6B and S71, ESI†), their combined populations remained under ca. 80% of the initial population of $[\text{Ag}_2(\mathbf{6}_2, \mathbf{2}_2)]^{2+}$ (*i.e.*, prior to the addition of trichloroacetic acid), the missing material being presumably trapped in ill-defined oligomers and polymers undetectable by ^1H NMR spectroscopy. After the concentration of $[\text{Ag}_2(\mathbf{6}, \mathbf{7})_2]^{2+}$ peaked, the baseline of the ^1H NMR spectra of the reaction became broad and uneven in the aromatic region (Fig. 6B and S71, ESI†), indicating an increased formation of oligomeric and polymeric species. This observation is consistent with the progressive deprotonation of $[\mathbf{2} + 2\text{H}^+]^{2+}$ during the decomposition process of trichloroacetate into CHCl_3 and CO_2 , making both diamines **2** and **7** available to form oligomeric and polymeric species as the system returns to its initial equilibrium state.

Compared to the systems of mononuclear complexes in Fig. 2–4, a large excess of trichloroacetic acid was required to maximize the formation of $[\text{Ag}_2(\mathbf{6}, \mathbf{7})_2]^{2+}$ (10 eq. of trichloroacetic acid were needed in this case *vs.* 2.5 eq. in the case of the simpler systems). The amount of $[\text{Ag}_2(\mathbf{6}, \mathbf{7})_2]^{2+}$ generated during the transient process and its lifetime were both found to be roughly proportional to the amount of acid added (ESI, Fig. S69–S71 and S73†). When the amount of acid added was increased from 7.5 to 10 eq., the peak population of $[\text{Ag}_2(\mathbf{6}, \mathbf{7})_2]^{2+}$ increased from ca. 35 to 70% of the initial population of $[\text{Ag}_2(\mathbf{6}_2, \mathbf{2}_2)]^{2+}$ and the amount of time required for the system to return to its initial equilibrium state increased from ca. 450 to 810 min. This high requirement in acid could suggest that the rearrangement of the system is slow compared to the rate of decarboxylation of trichloroacetate. An excess of acid would therefore be needed to reach a steady state in the system, maximizing the transient formation of catenane $[\text{Ag}_2(\mathbf{6}, \mathbf{7})_2]^{2+}$ under thermodynamic control.

Higher amounts of acid (up to 12 eq., ESI, Fig. S72 and S73†) did not increase the peak population of $[\text{Ag}_2(\mathbf{6}, \mathbf{7})_2]^{2+}$ further, possibly indicating that some of the oligomeric and polymeric intermediates formed during the rearrangement of the system may be too kinetically stable to rearrange in the time scale of the transient process (in the conditions used).

While all the initial macrocycle $[\text{Ag}_2(\mathbf{6}_2, \mathbf{2}_2)]^{2+}$ could be recovered at the end of the rearrangement using 7.5 eq. of trichloroacetic acid (ESI, Fig. S69 and S73†), the amount of macrocycle recovered at the end of the rearrangement cycle diminished when more acid was used (90 and 70% of the initial macrocycle were recovered when 8.5 or 12 eq. of trichloroacetic acid were used, respectively, ESI, Fig. S70–S73†). Performing a second rearrangement cycle led to significant deterioration of the system (ESI, Fig. S75†).

Conclusions

Trichloroacetic acid has been used to control in time the composition of imine-based metal complexes *via* energy-dissipation. By exploiting the base-promoted decarboxylation of trichloroacetic acid, it was possible to cycle between the

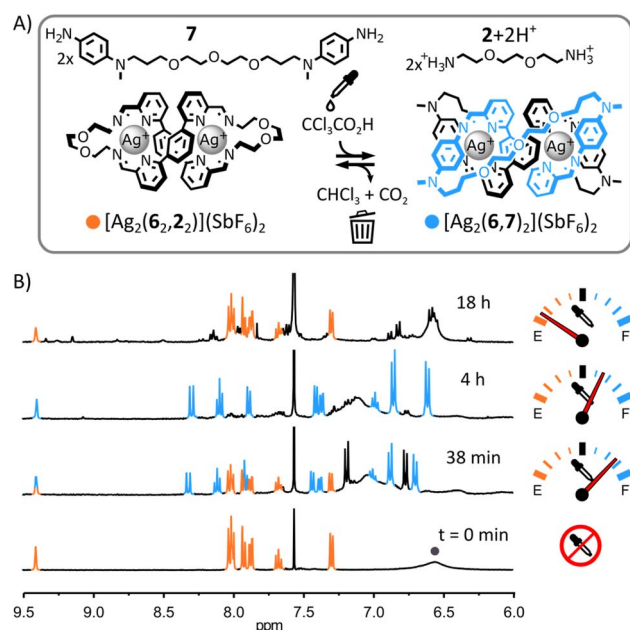


Fig. 6 (A) Transient rearrangement of macrocycle $[\text{Ag}_2(\mathbf{6}_2, \mathbf{2}_2)]^{2+}$ into $[\text{Ag}_2(\mathbf{6}, \mathbf{7})_2]^{2+}$. Conditions: CCl_3COOH (10 eq.), $\text{CD}_3\text{CN}:\text{CDCl}_3$ 3 : 1, r. t. (B) Partial ^1H NMR spectra (400 MHz, $\text{CD}_3\text{CN}:\text{CDCl}_3$ 3 : 1, 297 K) showing the evolution of the crude reaction mixture upon addition of CCl_3COOH ; the diagnostic signals of $[\text{Ag}_2(\mathbf{6}_2, \mathbf{2}_2)]^{2+}$ and $[\text{Ag}_2(\mathbf{6}, \mathbf{7})_2]^{2+}$ are coloured in orange and blue, respectively, two of the diagnostic signals of **7** are highlighted by a grey circle.

preferential incorporation of an arylamine or an alkylamine component in a metal complex, enabling the transient expression of complexes otherwise unfavourable in the absence of trichloroacetic acid.

The electronic properties of the arylamine component appeared to be crucial to the robustness of the transient rearrangement process, whereas the nature of the alkylamine component and tetrahedral metal ions had little influence. Transient processes can temporarily simplify the output of a self-assembly process by populating a different, more structured, equilibrium state. The properties of metal complexes can be controlled in time *via* the transient modification of their compositions. We showed that the mechanical properties of a macrocyclic complex could be temporarily altered by rearranging it into a topologically more complex [2]catenane.

By providing a generalizable method to transiently access different equilibrium states in systems of metal–organic complexes (or architectures), the information gleaned from our investigation will facilitate the development of new adaptive behaviours in these systems; behaviours exploiting both the capabilities of the default equilibrium (or kinetically trapped) state of the system and the capabilities of transient equilibrium states accessible *via* energy-dissipation.

Data availability

The synthetic procedures, characterization, and spectral data supporting this article have been uploaded as part of the ESI.† CCDC 2162663 and CCDC 2162664 contain the supplementary crystallographic data for this paper.

Author contributions

J. F. A. conceived and conducted the experiments. L. K. and N. K. solved and refined the X-ray structures. J. F. A. and B. B. supervised the project. The manuscript was written by J. F. A. All authors reviewed the manuscript.

Conflicts of interest

The authors declare no conflicts of interest.

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Notes and references

- For selected resources on self-assembly:(a) G. M. Whitesides and B. A. Grzybowski, *Science*, 2002, **295**, 2418–2421; (b)

K. Das, L. Gabrielli and L. J. Prins, *Angew. Chem., Int. Ed.*, 2021, **60**, 20120–20143.

- For selected general resources on stimuli-responsive self-assembly:(a) J.-M. Lehn, *Supramolecular Chemistry: Concepts and Perspectives*, John Wiley & Sons, Ltd, 1995; (b) *Intelligent Stimuli-Responsive Materials*, ed. Q. Li, John Wiley & Sons, Ltd, 2013; (c) J. W. Steed and J. L. Atwood, *Supramolecular Chemistry*, Wiley, 3rd edn, 2022; (d) H. Ivan and L. Jean-Marie, *Proc. Natl. Acad. Sci. U. S. A.*, 1997, **94**, 2106–2110; (e) O. Sijbren, R. L. E. Furlan and J. K. M. Sanders, *Science*, 2002, **297**, 590–593.
- For recent review articles covering applications of stimuli-responsive self-assembly:(a) J.-M. Lehn, *Angew. Chem., Int. Ed.*, 2015, **54**, 3276–3289; (b) S. Erbas-Cakmak, D. A. Leigh, C. T. McTernan and A. L. Nussbaumer, *Chem. Rev.*, 2015, **115**, 10081–10206; (c) V. Blanco, D. A. Leigh and V. Marcos, *Chem. Soc. Rev.*, 2015, **44**, 5341–5370; (d) A. J. McConnell, C. S. Wood, P. P. Neelakandan and J. R. Nitschke, *Chem. Rev.*, 2015, **115**, 7729–7793; (e) A. J. R. Amaral and G. Pasparakis, *Polym. Chem.*, 2017, **8**, 6464–6484; (f) J. F. Stoddart, *Angew. Chem., Int. Ed.*, 2017, **56**, 11094–11125; (g) J.-F. Ayme and J.-M. Lehn, in *Advances in Inorganic Chemistry*, ed. R. van Eldik and R. Puchta, Academic Press, 2018, vol. 71, pp. 3–78; (h) M. Grzelczak, L. M. Liz-Marzán and R. Klajn, *Chem. Soc. Rev.*, 2019, **48**, 1342–1361; (i) A. Goswami, S. Saha, P. K. Biswas and M. Schmittel, *Chem. Rev.*, 2020, **120**, 125–199; (j) S. M. Jansze and K. Severin, *J. Am. Chem. Soc.*, 2019, **141**, 815–819; (k) S. M. Jansze, G. Cecot and K. Severin, *Chem. Sci.*, 2018, **9**, 4253–4257.
- For selected general resources on energy-consumption in biological processes:(a) B. Alberts, A. Johnson, J. Lewis, D. Morgan, M. Raff, K. Roberts and P. Walter, *Molecular Biology of the Cell*, W.W. Norton & Company, 2015; (b) S. Mann, *Angew. Chem., Int. Ed.*, 2008, **47**, 5306–5320.
- For selected general resources on man-made dissipative self-assembly:(a) M. Fialkowski, K. J. M. Bishop, R. Klajn, S. K. Smoukov, C. J. Campbell and B. A. Grzybowski, *J. Phys. Chem. B*, 2006, **110**, 2482–2496; (b) A. Sorrenti, J. Leira-Iglesias, A. J. Markvoort, T. F. A. de Greef and T. M. Hermans, *Chem. Soc. Rev.*, 2017, **46**, 5476–5490; (c) S. A. P. van Rossum, M. Tena-Solsona, J. H. van Esch, R. Eelkema and J. Boekhoven, *Chem. Soc. Rev.*, 2017, **46**, 5519–5535; (d) M. Kathan and S. Hecht, *Chem. Soc. Rev.*, 2017, **46**, 5536–5550; (e) G. Ragazzon and L. J. Prins, *Nat. Nanotechnol.*, 2018, **13**, 882–889; (f) M. Baroncini, S. Silvi and A. Credi, *Chem. Rev.*, 2020, **120**, 200–268; (g) N. Singh, G. J. M. Formon, S. de Piccoli and T. M. Hermans, *Adv. Mater.*, 2020, **32**, 1906834; (h) C. Biagini and S. Di Stefano, *Angew. Chem., Int. Ed.*, 2020, **59**, 8344–8354; (i) B. Rieß, R. K. Grötsch and J. Boekhoven, *Chem*, 2020, **6**, 552–578; (j) S. Amano, S. Borsley, D. A. Leigh and Z. Sun, *Nat. Nanotechnol.*, 2021, **16**, 1057–1067; (k) L. S. Kariyawasam, M. M. Hossain and C. S. Hartley, *Angew. Chem., Int. Ed.*, 2021, **60**, 12648–12658; (l) H. Lee, J. Tessarolo, D. Langbehn, A. Baksi, R. Herges and G. H. Clever, *J. Am. Chem. Soc.*, 2022, **144**, 3099–3105; (m) R. G. DiNardi,



- A. O. Douglas, R. Tian, J. R. Price, M. Tajik, W. A. Donald and J. E. Beves, *Angew. Chem. Int. Ed.*, 2022, **61**, e202205701; (n) V. W. L. Gunawardana, T. J. Finnegan, C. E. Ward, C. E. Moore and J. D. Badjić, *Angew. Chem., Int. Ed.*, 2022, **61**, e202207418.
- 6 (a) J. Boekhoven, A. M. Brizard, K. N. K. Kowli, G. J. M. Koper, R. Eelkema and J. H. van Esch, *Angew. Chem., Int. Ed.*, 2010, **49**, 4825–4828; (b) J. P. Wojciechowski, A. D. Martin and P. Thordarson, *J. Am. Chem. Soc.*, 2018, **140**, 2869–2874; (c) N. Singh, B. Lainer, G. J. M. Formon, S. de Piccoli and T. M. Hermans, *J. Am. Chem. Soc.*, 2020, **142**, 4083–4087.
- 7 (a) M. R. Wilson, J. Solà, A. Carlone, S. M. Goldup, N. Lebrasseur and D. A. Leigh, *Nature*, 2016, **534**, 235–240; (b) J. A. Berrocal, C. Biagini, L. Mandolini and S. Di Stefano, *Angew. Chem., Int. Ed.*, 2016, **55**, 6997–7001; (c) E.-C. Sundus, S. D. P. Fielden, U. Karaca, D. A. Leigh, C. T. McTernan, D. J. Tetlow and M. R. Wilson, *Science*, 2017, **358**, 340–343; (d) C. Biagini, S. D. P. Fielden, D. A. Leigh, F. Schaufelberger, S. Di Stefano and D. Thomas, *Angew. Chem., Int. Ed.*, 2019, **58**, 9876–9880; (e) D. Mariottini, D. Del Giudice, G. Ercolani, S. Di Stefano and F. Ricci, *Chem. Sci.*, 2021, **12**, 11735–11739; (f) A. Goswami, S. Saha, E. Elramadi, A. Ghosh and M. Schmittel, *J. Am. Chem. Soc.*, 2021, **143**, 14926–14935; (g) A. Ghosh, I. Paul and M. Schmittel, *J. Am. Chem. Soc.*, 2021, **143**, 5319–5323; (h) S. Amano, S. D. P. Fielden and D. A. Leigh, *Nature*, 2021, **594**, 529–534; (i) D. Thomas, D. J. Tetlow, Y. Ren, S. Kassem, U. Karaca and D. A. Leigh, *Nat. Nanotechnol.*, 2022, **17**, 701–707; (j) S. Borsley, E. Kreidt, D. A. Leigh and B. M. W. Roberts, *Nature*, 2022, **604**, 80–85.
- 8 (a) C. S. Wood, C. Browne, D. M. Wood and J. R. Nitschke, *ACS Cent. Sci.*, 2015, **1**, 504–509; (b) L. S. Kariyawasam and C. S. Hartley, *J. Am. Chem. Soc.*, 2017, **139**, 11949–11955; (c) F. Rispoli, E. Spatola, D. Del Giudice, R. Cacciapaglia, A. Casnati, L. Baldini and S. Di Stefano, *J. Org. Chem.*, 2022, **87**, 3623–3629.
- 9 (a) S. Maiti, I. Fortunati, C. Ferrante, P. Scrimin and L. J. Prins, *Nat. Chem.*, 2016, **8**, 725–731; (b) B. G. P. van Ravensteijn, W. E. Hendriksen, R. Eelkema, J. H. van Esch and W. K. Kegel, *J. Am. Chem. Soc.*, 2017, **139**, 9763–9766; (c) R. K. Grötsch, C. Wanzke, M. Speckbacher, A. Angi, B. Rieger and J. Boekhoven, *J. Am. Chem. Soc.*, 2019, **141**, 9872–9878.
- 10 (a) S. M. Morrow, I. Colomer and S. P. Fletcher, *Nat. Commun.*, 2019, **10**, 1011; (b) A. H. J. Engwerda, J. Southworth, M. A. Lebedeva, R. J. H. Scanes, P. Kukura and S. P. Fletcher, *Angew. Chem., Int. Ed.*, 2020, **59**, 20361–20366; (c) S. Yang, G. Schaeffer, E. Mattia, O. Markovitch, K. Liu, A. S. Hussain, J. Ottelé, A. Sood and S. Otto, *Angew. Chem., Int. Ed.*, 2021, **60**, 11344–11349; (d) A. H. J. Engwerda, J. Southworth, M. A. Lebedeva, R. J. H. Scanes, P. Kukura and S. P. Fletcher, *Angew. Chem., Int. Ed.*, 2020, **59**, 20361–20366.
- 11 (a) M. Tena-Solsona, C. Wanzke, B. Riess, A. R. Bausch and J. Boekhoven, *Nat. Commun.*, 2018, **9**, 2044; (b) M. M. Hossain, J. L. Atkinson and C. S. Hartley, *Angew. Chem., Int. Ed.*, 2020, **59**, 13807–13813; (c) C. M. E. Kriebisch, A. M. Bergmann and J. Boekhoven, *J. Am. Chem. Soc.*, 2021, **143**, 7719–7725; (d) M. G. Howlett, R. J. H. Scanes and S. P. Fletcher, *JACS Au*, 2021, **1**, 1355–1361; (e) J. Deng and A. Walther, *Nat. Commun.*, 2020, **11**, 3658; (f) D. Del Giudice, M. Valentini, G. Melchiorre, E. Spatola and S. Di Stefano, *Chem. – Eur. J.*, 2022, e202200685; (g) P. S. Schwarz, M. Tena-Solsona, K. Dai and J. Boekhoven, *Chem. Commun.*, 2022, **58**, 1284–1297.
- 12 For selected review article covering complex functions in systems of dynamic molecules: (a) F. B. L. Cougnon and J. K. M. Sanders, *Acc. Chem. Res.*, 2012, **45**, 2211–2221; (b) M. E. Belowich and J. F. Stoddart, *Chem. Soc. Rev.*, 2012, **41**, 2003–2024; (c) Y. Jin, C. Yu, R. J. Denman and W. Zhang, *Chem. Soc. Rev.*, 2013, **42**, 6634–6654; (d) J. Li, P. Nowak and S. Otto, *J. Am. Chem. Soc.*, 2013, **135**, 9222–9239; (e) A. Herrmann, *Chem. Soc. Rev.*, 2014, **43**, 1899–1933; (f) N. Roy, B. Bruchmann and J.-M. Lehn, *Chem. Soc. Rev.*, 2015, **44**, 3786–3807; (g) D. A. Roberts, B. S. Pilgrim and J. R. Nitschke, *Chem. Soc. Rev.*, 2018, **47**, 626–644; (h) F. Beuerle and B. Gole, *Angew. Chem., Int. Ed.*, 2018, **57**, 4850–4878; (i) P. Chakma and D. Konkolewicz, *Angew. Chem., Int. Ed.*, 2019, **58**, 9682–9695; (j) D. Zhang, T. K. Ronson, Y.-Q. Zou and J. R. Nitschke, *Nat. Rev. Chem.*, 2021, **5**, 168–182; (k) I. Jahović, Y.-Q. Zou, S. Adorinni, J. R. Nitschke and S. Marchesan, *Matter*, 2021, **4**, 2123–2140; (l) N. Zheng, Y. Xu, Q. Zhao and T. Xie, *Chem. Rev.*, 2021, **121**, 1716–1745.
- 13 For selected research article on complex functions in systems of dynamic molecules: (a) H. Ivan and L. Jean-Marie, *Proc. Natl. Acad. Sci. U. S. A.*, 1997, **94**, 2106–2110; (b) O. Sijbren, R. L. E. Furlan and J. K. M. Sanders, *Science*, 2002, **297**, 590–593; (c) A. D. W. Kennedy, N. de Haas, H. Iranmanesh, E. T. Luis, C. Shen, P. Wang, J. R. Price, W. A. Donald, J. Andréasson, F. Huang and J. E. Beves, *Chem.–Eur. J.*, 2019, **25**, 5708–5718; (d) M. He and J.-M. Lehn, *J. Am. Chem. Soc.*, 2019, **141**, 18560–18569; (e) A. Osypenko, S. Dhers and J.-M. Lehn, *J. Am. Chem. Soc.*, 2019, **141**, 12724–12737; (f) J.-F. Ayme and J.-M. Lehn, *Chem. Sci.*, 2020, **11**, 1114–1121; (g) J.-F. Ayme, S. Dhers and J.-M. Lehn, *Angew. Chem., Int. Ed.*, 2020, **59**, 12484–12492; (h) J.-F. Ayme, J.-M. Lehn, C. Bailly and L. Karmazin, *J. Am. Chem. Soc.*, 2020, **142**, 5819–5824; (i) B.-N. T. Nguyen, J. D. Thoburn, A. B. Grommet, D. J. Howe, T. K. Ronson, H. P. Ryan, J. L. Bolliger and J. R. Nitschke, *J. Am. Chem. Soc.*, 2021, **143**, 12175–12180; (j) O. Borodin, Y. Shchukin, C. C. Robertson, S. Richter and M. von Delius, *J. Am. Chem. Soc.*, 2021, **143**, 16448–16457; (k) R. Gu and J.-M. Lehn, *J. Am. Chem. Soc.*, 2021, **143**, 14136–14146; (l) N. F. König, D. Mutruc and S. Hecht, *J. Am. Chem. Soc.*, 2021, **143**, 9162–9168.
- 14 (a) D. Schultz and J. R. Nitschke, *J. Am. Chem. Soc.*, 2006, **128**, 9887–9892; (b) M. Hutin, C. A. Schalley, G. Bernardinelli and J. R. Nitschke, *Chem. – Eur. J.*, 2006, **12**, 4069–4076.
- 15 The distribution of products observed by ^1H NMR spectroscopy minutes after the addition of the arylamine



matched that obtained when the same system of components was left to equilibrate for 18 h at 60 °C from an identical or a different starting point (see ESI† Section 3), indicating that the system has reached equilibrium—or is close to equilibrium—in this short time span.

- 16 (a) G. Schwarzenbach, *Helv. Chim. Acta*, 1952, **35**, 2344–2359; (b) D. Schultz and J. R. Nitschke, *Angew. Chem., Int. Ed.*, 2006, **45**, 2453–2456.
- 17 (a) E. C. Constable, S. M. Elder, M. J. Hannon, A. Martin, P. R. Raithby and D. A. Tocher, *J. Chem. Soc., Dalton Trans.*, 1996, 2423–2433; (b) E. C. Constable, A. J. Edwards, G. R. Haire, M. J. Hannon and P. R. Raithby, *Polyhedron*, 1998, **17**, 243–253; (c) J. R. Price, Y. Lan and S. Brooker, *Dalton Trans.*, 2007, 1807–1820.
- 18 (a) E. J. Corey, J. O. Link and Y. Shao, *Tetrahedron Lett.*, 1992, **33**, 3435–3438; (b) Y. Li, T. Zheng, W. Wang, W. Xu, Y. Ma, S. Zhang, H. Wang and Z. Sun, *Adv. Synth. Catal.*, 2012, **354**, 308–312.
- 19 (a) M. A. Murcko and R. A. DiPaola, *J. Am. Chem. Soc.*, 1992, **114**, 10010–10018; (b) J.-F. Ayme, J. E. Beves, D. A. Leigh, R. T. McBurney, K. Rissanen and D. Schultz, *Nat. Chem.*, 2012, **4**, 15–20.
- 20 (a) J.-F. Ayme, J. E. Beves, C. J. Campbell and D. A. Leigh, *Chem. Soc. Rev.*, 2013, **42**, 1700–1712; (b) G. Gil-Ramírez, D. A. Leigh and A. J. Stephens, *Angew. Chem., Int. Ed.*, 2015, **54**, 6110–6150.
- 21 The distribution of products observed by ¹H NMR spectroscopy minutes after the addition of diarylamine 7 to [Ag₂(**6**₂,**2**₂)]²⁺ matched that obtained during the selective self-assembly of [Ag₂(**6**₂,**2**₂)]²⁺ from a mixture of Ag(SbF₆), **7**, **6** and **2** at 60 °C for 18 h (see ESI† Section 3.2), indicating that the system has reached equilibrium—or is close to equilibrium—in this short time span.
- 22 J. P. Carpenter, C. T. McTernan, J. L. Greenfield, R. Lavendomme, T. K. Ronson and J. R. Nitschke, *Chem*, 2021, **7**, 1534–1543.

