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Capillary electrophoresis mass spectrometry identifies new isomers of inositol pyrophosphates in mammalian tissues†

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Technical challenges have to date prevented a complete profiling of the levels of *myo*-inositol phosphates (InsPs) and pyrophosphates (PP-InsPs) in mammalian tissues. Here, we have deployed capillary electrophoresis mass spectrometry to identify and record the levels of InsPs and PP-InsPs in several tissues obtained from wild type mice and a newly created PPIP5K2 knockout strain. We observe that the mouse colon harbours unusually high levels of InsPs and PP-InsPs. Additionally, the PP-InsP profile is considerably more complex than previously reported for animal cells: using chemically synthesized internal stable isotope references and high-resolution mass spectra, we characterize two new PP-InsP isomers as 4/6-PP-InsP₅ and 2-PP-InsP₅. The latter has not previously been described in nature. The analysis of feces and the commercial mouse diet suggests that the latter is one potential source of noncanonical isomers in the colon. However, we also identify both molecules in the heart, indicating unknown synthesis pathways in mammals. We also demonstrate that the CE-MS method is sensitive enough to measure PP-InsPs from patient samples such as colon biopsies and peripheral blood mononuclear cells (PBMCs). Strikingly, PBMCs also contain 4/6-PP-InsP₅ and 2-PP-InsP₅. In summary, our study substantially expands PP-InsP biology in mammals.

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Inositol phosphates (InsPs) and pyrophosphates (PP-InsPs) are a complex signalling hub with diverse functions in eukaryotes.^{1–3} PP-InsPs have specialized physicochemical properties and biological functions that attract widespread interest.^{4–7} They occur as distinct isomers of differentially phosphorylated metabolites of InsP₆ (phytic acid and phytate). The current literature suggests that in yeast and mammals these phosphorylation reactions occur selectively and successively in the 5- and 1-positions (Fig. 1) leading to 5-PP-InsP₅ and 1,5-(PP)₂-InsP₄, respectively.^{8,9} In plants and slime-mold, 4/6-PP-

InsP₅ has been identified as the main isomer, with the absolute configuration of the biologically relevant enantiomer remaining unknown.^{10,11}

Kinases and phosphatases that synthesize and metabolize PP-IPs are distributed throughout all eukaryotic kingdoms.^{7,12} In mammals, there are three isoforms of IP6Ks that add a β -phosphate at position 5 and two isoforms of PPIP5Ks that add a β -phosphate at position 1.⁸

Most of the research into PP-InsP turnover in mammalian cells has relied on separation by HPLC of extracts of ³H-inositol radio-labeled cells, although in more recent years a more generally accessible PAGE technique has proved useful.^{13,14} This body of work has consistently concluded that 5-PP-InsP₅ is the most abundant PP-InsP isomer (generally <10% of InsP₆ levels). The levels of 1,5-(PP)₂-InsP₄ and 1-PP-InsP₅ are approximately 10-fold and 50-fold lower, which are below the PAGE detection limit.^{14–16} The relative ease with which 5-PP-InsP₅ abundance can be measured has in large part driven the field's primary focus on this isomer. For example, this PP-InsP has been reported to regulate insulin signalling, exocytosis, processing body formation, intracellular protein localization, and bioenergetic homeostasis.^{17–22}

More recently, 1,5-(PP)₂-InsP₄ has emerged as an independently regulated cellular signal. This facet of PP-InsP signalling first arose from kinetic assessments²³ of the PPIP5K kinase

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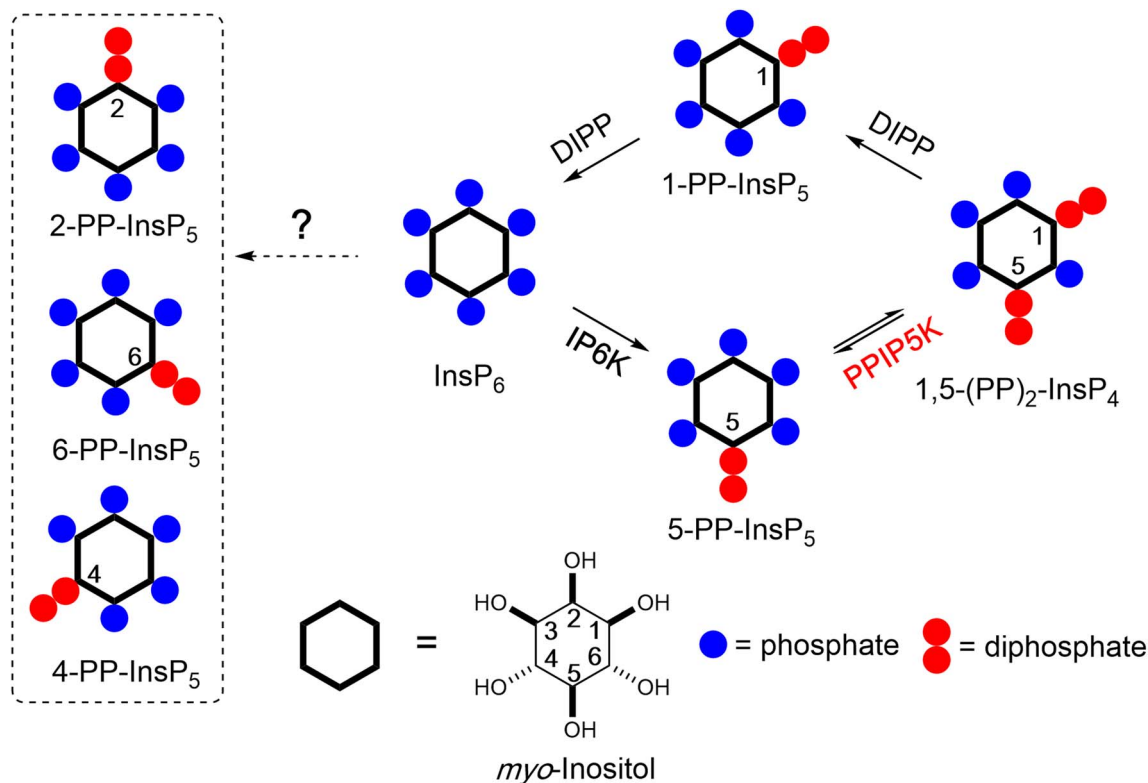


Fig. 1 Main metabolic reactions that determine the turnover of inositol pyrophosphates in mammalian cells. Three isoforms of IP6Ks phosphorylate the 5-position of InsP₆ and two isoforms of PIP5Ks phosphorylate the 1-position with a preference for 5-PP-InsP₅ over InsP₆. The question mark indicates that an unknown pathway is responsible for the synthesis of the 4/6-PP-InsP₅ and 2-PP-InsP₅ identified in the current study (inside dotted box, note that 4/6-PP-InsP₅ are enantiomers). PIP5Ks also harbour a phosphatase domain catalyzing dephosphorylation 1,5-(PP)₂-InsP₄. DIPPs are specialized phosphatases that degrade the phosphoric anhydrides in PP-InsPs.

domain that phosphorylates 5-PP-InsP₅ to 1,5-(PP)₂-InsP₄ and the separate phosphatase domain that degrades 1,5-(PP)₂-InsP₄ back to 5-PP-InsP₅ (see Fig. 1). Moreover, the phosphatase activity is inhibited by elevations in the cellular levels of inorganic phosphate (P_i), thereby enhancing net 1,5-(PP)₂-InsP₄ production independently of any changes in 5-InsP₇ levels.^{23,24} As a consequence, the net kinase and phosphatase activities are tied to cellular energy and phosphate homeostasis.^{3,25} It has since been demonstrated that 1,5-(PP)₂-InsP₄ stimulates P_i efflux from mammalian cells through an interaction with an SPX domain on the transmembrane XPR1 protein.^{26,27} Moreover, pharmacologic inhibition of IP6Ks in mammals (rodents and monkeys), which restrains PP-InsP₅ and 1,5-(PP)₂-InsP₄ synthesis (see Fig. 1), leads to attenuation of systemic hyperphosphatemia through inactivation of XPR1; these findings are an important milestone for potential pharmacological treatment of chronic kidney disease.²⁸ Naturally occurring human variants of PPIP5K2 have been associated with deafness²⁹ and keratoconus.³⁰ Recently, [³H]inositol-radiolabeling of a hematopoietic stem cell line from a PPIP5K2^{-/-} mouse indicated that 1,5-(PP)₂-[³H]InsP₄ levels are no different from those in a typical culture medium (data for PPIP5K2^{-/-} cells were not reported).³¹ In such circumstances, it has become more important to be able to accurately assay dynamic fluctuations in 1,5-(PP)₂-InsP₄ concentrations.

A portfolio of additional methods has been introduced that can assay the mass levels of (PP)-InsPs in extracts of mammalian and plant cells, including using transition metals (*e.g.* Fe and Y) and absorbance detection (metal dye detection, MDD)^{32–34} and the coupling of in-line mass spectrometry to hydrophilic interaction liquid chromatography (HILIC) and metal-free C₁₈ reversed phase columns.^{28,35,36} NMR detection with ¹³C enriched inositol is another recent and promising addition to the analytical portfolio.³⁷ In 2020, capillary electrophoresis (CE) with mass spectrometry compatible buffers was reported for PP-InsP analytics, with only nanoliter sample consumption and accurate isomer assignment and quantitation by using stable isotope internal reference compounds.³⁸

We now significantly expand the value of our new PP-InsP profiling techniques through our identification of substantial cellular quantities of mammalian 4/6-PP-InsP₅ and 2-PP-InsP₅ (see Fig. 1) based on comigration with reference compounds and high-resolution mass spectra. This conclusion is facilitated by adapting a recently developed ¹⁸O phosphate labelling approach³⁹ in order to stereoselectively synthesize 4-PP-InsP₅ to use as a heavy internal standard. Finally, it was our goal to optimize CE-MS to monitor the complete array of PP-InsPs from human patient tissues. For this work, we selected colon biopsies and peripheral blood mononuclear cells including enriched T cell subpopulations (PBMCs, CD8⁺). Strikingly, we also identify 4/6-PP-InsP₅ and 2-PP-InsP₅ in PBMCs that are particularly



enriched in a CD8⁺ T cell preparation. Overall, this dramatic increase in the complexity of PP-InsP metabolism indicates that their biological significance has been greatly underestimated.

Results

With an established protocol that uses TiO₂ beads, we extracted and enriched InsPs and PP-InsPs from different mouse tissues.^{14,38}

The enriched samples were analyzed by CE-QQQ using the same background electrolyte (35 mM ammonium acetate adjusted to pH 9.7 with NH₄OH, *i.e.*, BGE-A) that we deployed in our previous study.³⁸ Samples were spiked with internal heavy isotope reference compounds (¹³C labels) of several different InsPs and PP-InsPs for assignment and quantitation. This is the first time that this method has been applied to any animal tissue for the quantification of the levels of the least abundant PP-IPs, namely 1,5-(PP)₂-

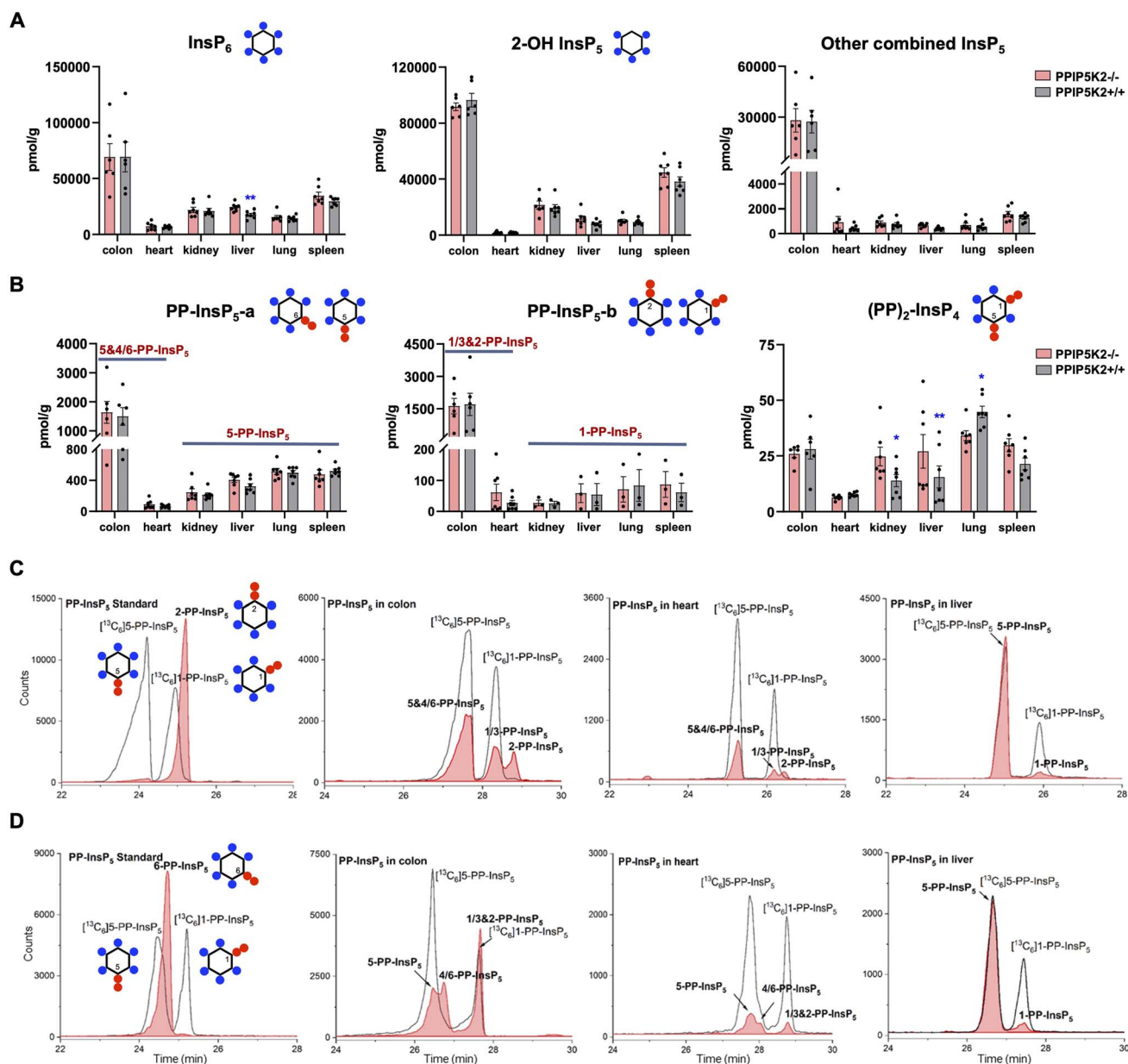


Fig. 2 Profiling of PP-InsPs in mouse tissues (wild type vs. PPIP5K2 knockout) and observation of new isomers. (A) Profiling of InsP₆, 2-OH InsP₅, and the total for all other InsP₅ isomers (4/6-OH InsP₅, 1/3-OH InsP₅, and 5-OH InsP₅). (B) PP-InsP₅-a and PP-InsP₅-b refer to two base-line resolved peaks. The tentative identification of the components of each peak (in bold font) is described in the text. Data for (A) and (B) are indicated as means \pm SEM, $n = 7$ and for the colon, $n = 6$. * $P < 0.05$ and ** $P < 0.01$, Student's *t*-test. (C) Extracted ion electropherograms (EIEs) of [¹³C] labeled internal reference compounds (black lines) plus a 2-PP-InsP₅ standard (red trace) or PP-InsP₅ extracts from the mouse colon, heart and liver, resolved with BGE A containing 35 mM ammonium acetate titrated with ammonium hydroxide to pH 9.7. D EIEs of [¹³C]-labeled internal reference compounds (black lines) plus (left panel) a 6-PP-InsP₅ standard (red trace) or PP-InsP₅ extracts from the mouse colon, heart and liver (red trace), resolved with BGE B containing 40 mM ammonium acetate titrated with ammonium hydroxide to pH 9.0.



InsP₄ and 1-PP-InsP₅ (for representative examples see Fig. 2C and ESI Fig. 1†).

We also used this method to compare InsP and PP-InsP levels in multiple mouse tissues, including the colon, heart, kidney, liver, lung and spleen (Fig. 2A–C). These molecules were generally least abundant in the heart. It is worth mentioning that other minor InsP₅ isomers including 4/6-OH InsP₅, 1/3-OH InsP₅, and 5-OH InsP₅ have also been identified and quantified (see representative examples obtained from the mouse colon and mouse heart; ESI Fig. 2†), while 2-OH InsP₅ was always by far the predominant isomer in all investigated mouse tissues (Fig. 2A).

Compared to other tissues, the colon is notable for containing substantially higher levels of InsP₆ (2- to 5-fold), 2-OH-InsP₅ (2- to 10-fold) and the sum of the remaining, quantitatively more minor InsP₅ isomers (19- to 52-fold). The colon also contains much higher levels of PP-InsP₅ isomers (Fig. 2A–C). In most of the studied tissues (kidney, liver, lung, and spleen), two baseline-resolved PP-InsP₅ signals were observed (labeled 'a' and 'b'), which co-eluted precisely with internal standards of [¹³C₆]5-PP-InsP₅ and [¹³C₆]1-PP-InsP₅, respectively, in each of two different BGE conditions (Fig. 2C and ESI Fig. 3A†). In these tissues, the relative proportion of 1-PP-InsP₅ to 5-PP-InsP₅ (approximately 1 to 7) is higher than that determined by our previous CE analysis of a line of immortalized HCT116 cells (1 to 13);⁴⁰ a ratio of only 1 to 50 was previously obtained by HPLC analysis of [³H]inositol-labeled extracts of immortalized cells.⁴¹

An unexpected outcome of the EIE obtained using BGE-A was that the PP-InsP-b signals derived from the colon and heart split into two approximately equally sized peaks that are incompletely resolved; the earlier-eluting peak comigrated with an internal standard of [¹³C₆]1-PP-InsP₅ (Fig. 2C). The elution time of the second peak corresponds precisely to the elution time of a replicate sample spiked with an internal standard of 2-PP-InsP₅ (ESI Fig. 4†). In addition, there is an indication that the PP-InsP-a signal derived from the colon also separates into two incompletely resolved peaks (Fig. 2C). To pursue the latter observation, we reran the samples with the background electrolyte adjusted to 40 mM ammonium acetate titrated with ammonium hydroxide to pH 9.0 (*i.e.*, BGE-B). This procedure extended the peak-to-peak resolution within the PP-InsP-a signal to the extent that its two components are also visible in the extracts prepared from the colon and heart (Fig. 2D). Note that, in contrast, the use of BGE-B did not perturb the coelution of internal standards of [¹³C₆]5-PP-InsP₅ and [¹³C₆]1-PP-InsP₅ with PP-InsP-a and PP-InsP-b signals, respectively, that were prepared from the kidney, liver, lung and spleen (Fig. 2C and D; ESI Fig. 3A and B†). However, we do not exclude that matrix effects in other tissues would blur the presence of low levels of additional PP-InsP isomers.

In this set of experiments with BGE-B, the first component of PP-InsP-a extracted from the colon comigrates with the internal standard of [¹³C₆]5-PP-InsP₅ and the second component of PP-InsP-a has an elution time that matches that of a standard of 6-PP-InsP₅ from separate runs (Fig. 2D). Thus, we tentatively identify the second component of PP-InsP-a as 4/6-PP-InsP₅ and by a process of elimination we suggest that the second

component of PP-InsP-b is 2-PP-InsP₅. Moreover, the proposed nature of 1/3-PP-InsP₅, 2-PP-InsP₅, 5-PP-InsP₅ and 4/6-PP-InsP₅ from the colon is also consistent with their high-resolution mass spectra collected by using a CE-qTOF system (ESI Fig. 5†). Other potential candidates with an identical mass, such as triphosphates of inositol-tetrakisphosphates (*e.g.* 5-PPP-InsP₄), have been described so far only *in vitro*.⁴² The *myo*-configuration for these new PP-InsPs seems likely, since there is no prior identification of any other multiply phosphorylated inositol stereoisomers in mammals.

It is notable that in the colon we estimate that the levels of 1-PP-InsP₅ (*i.e.*, half of PP-InsP-b) and 5-PP-InsP₅ (*i.e.*, half of PP-InsP-a) are approximately equivalent (Fig. 2C and D); this observation implies that we must profoundly modify prior perceptions of 1-PP-InsP₅ as a quantitatively minor constituent of mammalian cells and/or consider the possibility that the enantiomer 3-PP-InsP₅ is also present. Currently applied methods do not resolve the enantiomers.

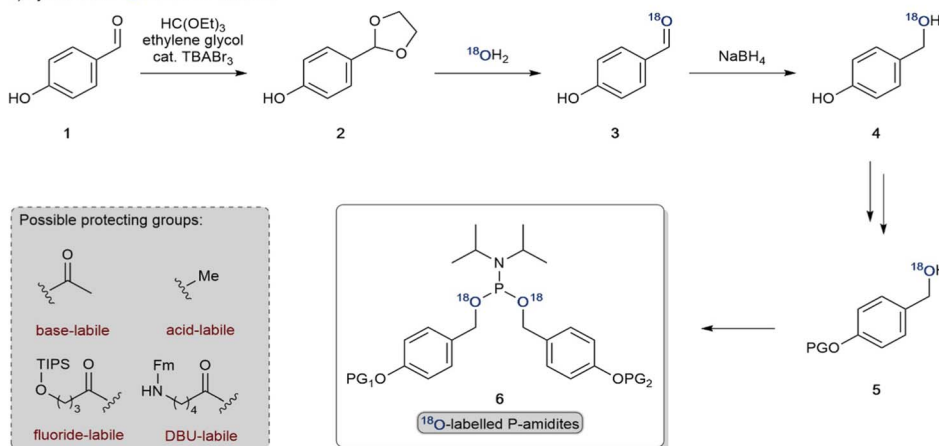
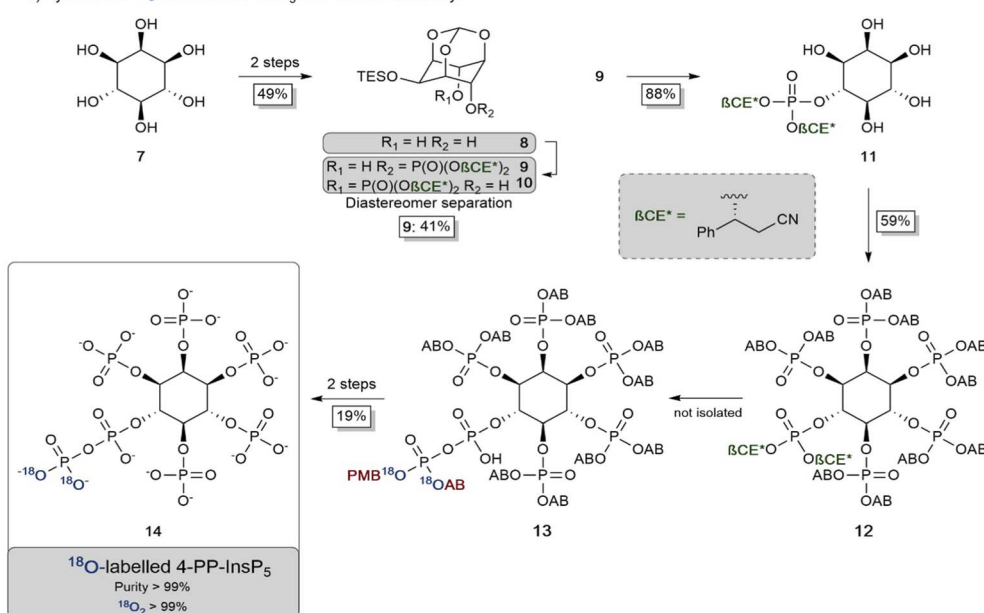
The 2-PP-InsP₅ isomer has not previously been identified in any biological material, possibly because it is both unexpected and only present at relatively low levels. In contrast, 4/6-PP-InsP₅ was recently discovered to be a major PP-InsP isomer in plants.¹⁰ Clearly, the latter is also a quantitatively important isomer in the mouse colon and heart (Fig. 2), and so it was particularly important to further validate its nature. Thus, we have developed a synthetic route to the preparation of enantiomerically pure [¹⁸O₂]4-PP-InsP₅ to deploy as an internal standard for additional chromatographic resolutions (see Fig. 3).

We have also recorded 1,5-(PP)₂-InsP₄ levels in mouse tissues (Fig. 2B). These varied over a 5-fold range, with the lowest levels in the heart and the highest in the lung; as far as we are aware, no previous study has provided such data. This accomplishment enabled us to determine the impact upon 1,5-(PP)₂-InsP₄ levels in a newly created PPIP5K2 knockout mouse (ESI Fig. 6 and 7†). No abnormal phenotype in the KO was observed. The litter size and gender distribution were not modified. No gross anatomical phenotype was observed during tissue collection. Food intake or energy expenditure (metabolic rate and physical activity) were unchanged (ESI Fig. 8†). We did not conduct any behavioral phenotyping.

The knockout only resulted in a statistically significant reduction in 1,5-(PP)₂-InsP₄ levels in the lung tissue (Fig. 2B). In fact, 1,5-(PP)₂-InsP₄ levels trended higher in several PPIP5K2 knockout tissues compared to the wild-type, and in the kidney and liver this effect was statistically significant. Although this might initially seem a counter-intuitive outcome, it is possible that in these two tissues the loss of the PPIP5K2 1,5-(PP)₂-InsP₄-phosphatase domain may have a larger metabolic effect than the loss of the 5-PP-InsP₅ kinase domain. The knockout did not elicit a statistically significant impact on 1,5-(PP)₂-InsP₄ levels in either the colon or the heart. The observation of tissue dependent variability in PP-InsP signaling brought about by PPIP5K2 knockout may depend in part on the extent to which PPIP5K1 compensates for the deletion of PPIP5K2 catalytic activity, although no such effect was evident in the liver (ESI Fig. 6 and 7†). Note also that the PPIP5K2 KO did not have off-target effects on any of the other InsPs and PP-



A

a) Synthesis of ^{18}O -labelled P-amidites:b) Synthesis of ^{18}O -labelled 4-PP-InsP₅ via P-amidite chemistry:

B

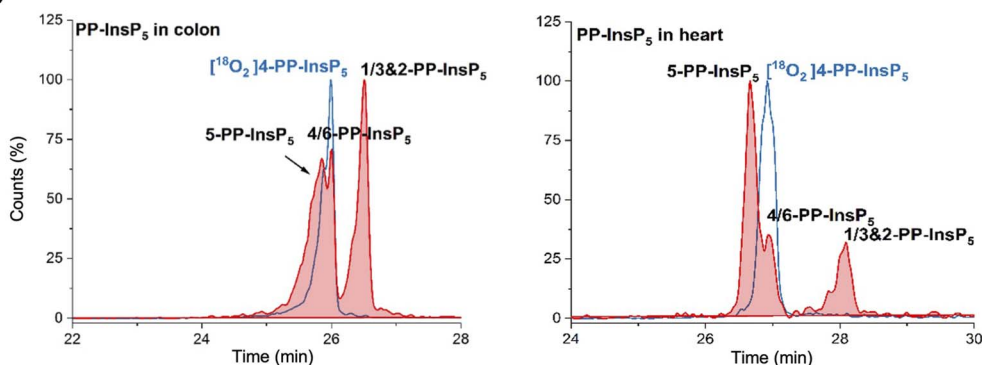


Fig. 3 (A) Synthesis and application of ^{18}O labeled P-amidites with diverse protecting group patterns and their application to a late-stage labeling 4-PP-InsP₅ synthesis. AB: acetoxybenzyl and PMB: *para*-methoxybenzyl. (B) Separation of 5-PP-InsP₅ and 4/6-PP-InsP₅ (filled red plots) from mouse colon and heart samples using BGE-B and assignment of the isomer with the new internal reference compound [$^{18}\text{O}_2$] 4-PP-InsP₅ (blue plot) as either 4-PP-InsP₅ or 6-PP-InsP₅. EIEs (PP-InsP₅ and [$^{18}\text{O}_2$] PP-InsP₅) are scaled to the largest peak indicated as 100%.



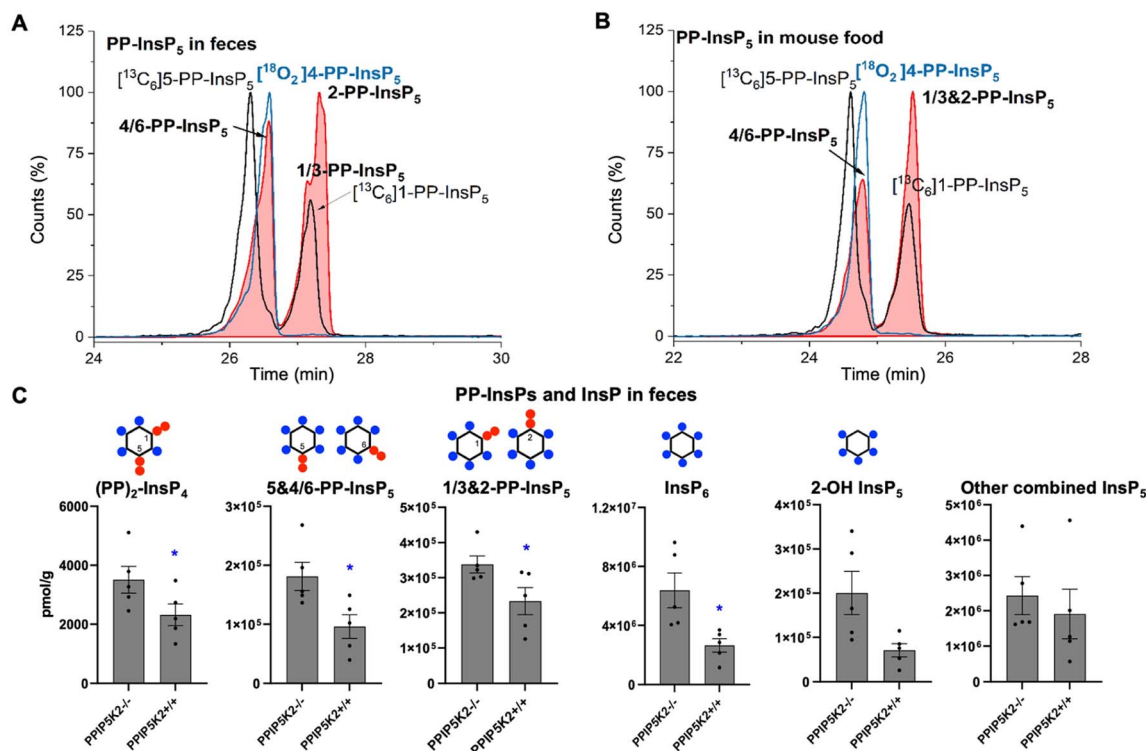


Fig. 4 (A) CE-MS analysis of PP-InsP₅ in mouse feces (filled red plots) with internal references of [¹³C₆]5-PP-InsP₅ and [¹³C₆]1-PP-InsP₅ (black plot) and [¹⁸O₂]4-PP-InsP₅ (blue plot) using BGE B. 4/6-PP-InsP₅ isomer is identified in mouse feces as well. (B) Analysis of PP-InsP₅ in mouse food the same as in (A), which contains high levels of this 4/6-PP-InsP₅ isomer. (C) Profiling of PP-InsPs and InsPs in mouse feces (wild type vs. PPIP5K2 knockout). Data are indicated as means ± SEM, *n* = 5. **P* < 0.05, and Student's *t*-test.

InsPs analyzed in this study (Fig. 2A and B), except that InsP₆ was increased in the PPIP5K KO liver.

We could not derive sufficient purified amounts of the putative 4/6-PP-InsP₅ for NMR analysis to further corroborate the identity of this isomer. So instead, we generated a reference compound with a heavy isotope label to serve as an internal standard for CE-MS. We reasoned that the comigration of this compound under different separation conditions would serve as a strong indication that it is indeed 4/6-PP-InsP₅ in its *myo*-configuration. The enzymes for plant 4/6-PP-InsP₅ synthesis are not yet known and so an enzymatic synthesis starting from InsP₆ of the reference compound with ¹³C labels was not possible.⁴³ A fully chemical synthesis from expensive ¹³C glucose in a multi-step linear approach was deemed not feasible.³⁷ We thus relied on our recently developed ¹⁸O phosphate labeling approach in which the expensive isotopic label can be introduced in the penultimate step of the synthesis.³⁹

In brief, ¹⁸O labeled phosphoramidites (P-amidites) with high ¹⁸O/¹⁶O ratios are key to the synthesis. These high ratios can be obtained by the strategy shown in Fig. 3A(a). *Para*-hydroxybenzaldehyde is transformed into its acetal **2**, which is then hydrolyzed in the presence of 99% ¹⁸O enriched water. The aldehyde **3** is directly reduced to stable alcohol **4**, which can then be protected on the phenol with diverse protecting groups (in the case described here simply acetate giving the acetoxybenzyl (AB) protecting group). The alcohols **5** are then transformed into P-amidites of the general structure **6**, enabling diverse protecting

group patterns and high ¹⁸O enrichment. The inositol structure is assembled as reported previously,^{44–47} as shown in Fig. 3A(b). While strictly a desymmetrization was not required and the generation of racemic 4/6-PP-InsP₅ would have been sufficient, we still generated the enantiomerically pure compound for potential future applications. Desymmetrization was achieved from intermediate protected diol **8**, which was reacted with an unsymmetric P-amidite containing chiral protecting groups (β-CE*, an arylated enantiomerically pure variant of the β-cyanoethyl protecting group). The obtained diastereomeric mixture was separated and then the inositol protecting groups were removed giving pentaol **11**. **11** was phosphorylated to protected InsP₆ **12** with orthogonal protecting groups (β-CE*) in the 4-position.⁴⁴ Selective deprotection in that position then enables the introduction of the ¹⁸O labeled phosphate bearing two ¹⁸O oxygen atoms (*M* + 4). Global deprotection gave [¹⁸O]₂ 4-PP-InsP₅ **14** in 99% purity with >99% isotopic enrichment as determined by CE-MS. This reference compound was then dissolved in water and its concentration was determined by quantitative ¹H- and ³¹P-NMR.

Fig. 3B demonstrates the first application of this newly generated isotopologue. Briefly, both colon and heart samples were spiked with the new reference and we utilized the optimized BGE-B that is capable of 5-PP-InsP₅ and 4/6-PP-InsP₅ separation. Masses were recorded and an identical migration of the unknown analyte with our reference in the same matrix was found, strongly suggesting that it is indeed 4/6-PP-InsP₅ that has been measured for the first time in mammalian tissues.



To understand the complexity of the profiles of InsPs and PP-InsPs in the colon in an organismal context, we additionally analyzed mouse feces and found them to contain very high levels of most analytes (Fig. 4A and C, ESI Fig. 9†). Moreover, neither PP-InsP peak co-eluted precisely with internal standards of either 5-PP-InsP₅ or 1-PP-InsP₅ again pointing towards the existence of both 4/6-PP-InsP₅ and 2-PP-InsP₅. In fact, the two new isomers are the most abundant analytes we detect (Fig. 4A). Interestingly, the PPIP5K knockout contained increased levels of all analytes in feces. We excluded that this was due to differences in food intake (ESI Fig. 8†). It may be possible that such changes are caused by different expression of digestive enzymes of PP-InsPs.

We next investigated if the mouse laboratory diet might contribute to the unprecedented complexity of the colonic PP-InsP profile. We provided mice with the “Rodent NIH-31 Open Formula Autoclavable Diet”, much of which is of plant origin. This is significant because recent work has determined that the quantitatively most important PP-InsP isomer in plants is one that had previously been overlooked, namely 4/6-PP-InsP₅.¹⁰ Indeed, our internal standards allowed us to conclude that large amounts of 4/

6-PP-InsP₅ were present in the mouse diet, although a precise quantification was hindered by insufficient separation of the 4/6- and 5-PP-InsP₅ peaks from the PP-InsP₅-a peak (Fig. 4B). Nevertheless, the latter was smaller than the PP-InsP₅-b peak, which likely comprises a mixture of 1-PP-InsP₅ and 2-PP-InsP₅.

Our results present the possibility that the diet might be the source of the colon's unusually high levels of InsP₆ and PP-InsPs, as well as the more complex PP-InsP profile. Furthermore, 2-OH InsP₅ is the minor InsP₅ isomer in mouse feces and also in mouse food (ESI Fig. 10†), in contrast to it being the major InsP₅ in the colon. This result suggests that the exceptional PP-InsPs and InsP₆ profile in the colon are not due to contamination from feces during sample preparation. In this case, endocytosis of dietary InsP₆ and PP-InsPs by colonic epithelial cells should be considered as a viable possibility.

Finally, in order to demonstrate the sensitivity of the method and its potential in translational research, we obtained human samples for enrichment and profiling. We analyzed one 18 mg wet tissue colon biopsy, which was sufficient to profile the main PP-InsP and InsP contents (Fig. 5A). Only canonical isomers were identified, *i.e.* 5-PP-InsP₅, InsP₆, and 2-OH InsP₅. We

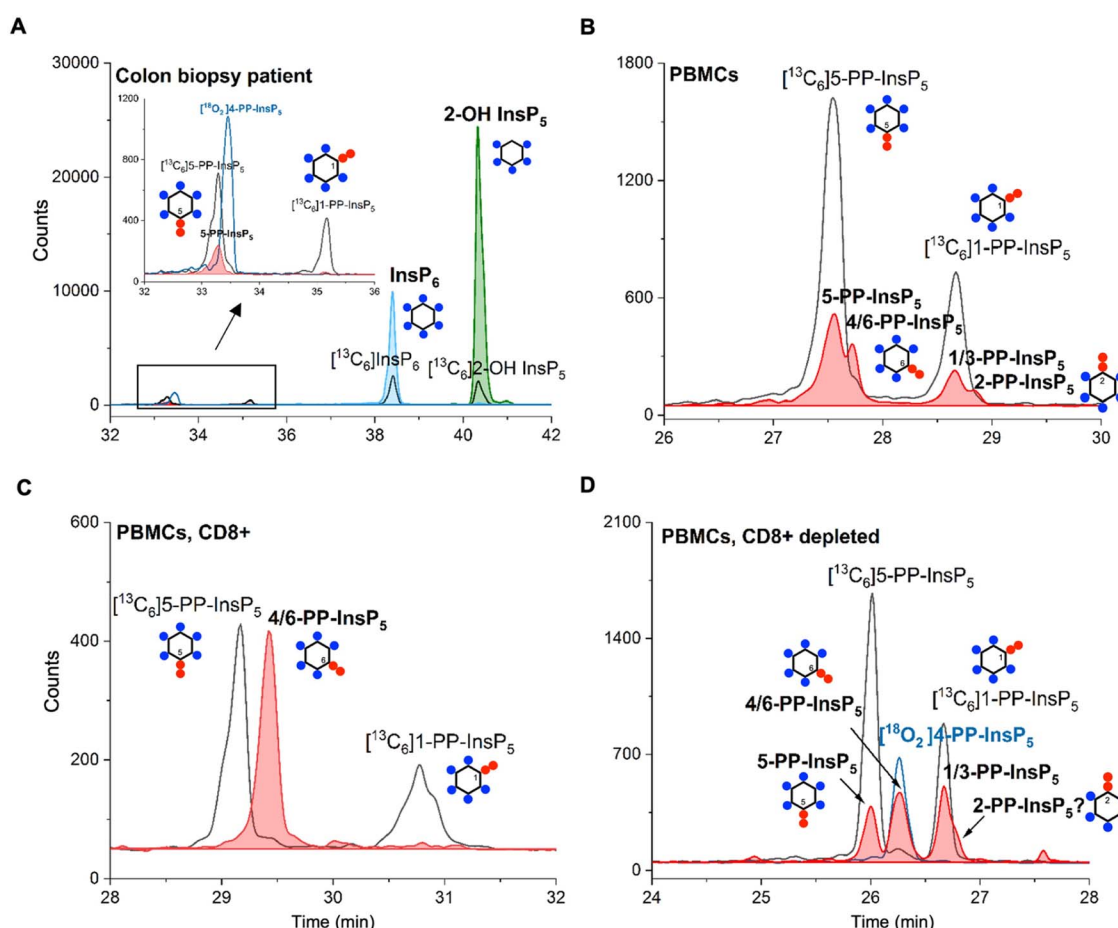


Fig. 5 (A) CE-MS analysis of a human colon tissue biopsy (18 mg) enables the identification of several important inositol phosphate (InsP₆ and InsP₅) and pyrophosphate (5-PP-InsP₅) isomers. (B) 4/6-PP-InsP₅ is identified in PBMCs by CE-MS analysis. The electropherograms are representative of independent biological triplicates giving comparable results. (C) 4/6-PP-InsP₅ is enriched in a CD8⁺ T-cell preparation and is also present in the CD8⁺ depleted PBMC pool (D). It is assigned by its exact same migration time as [¹⁸O₂] 4-PP-InsP₅. PP-InsP₅ (filled red plot) isomer identification is achieved with the aid of [¹³C₆]5-PP-InsP₅, [¹³C₆]1-PP-InsP₅ (black plot) and [¹⁸O₂] 4-PP-InsP₅ (blue plot).



additionally analyzed peripheral blood mononuclear cells (PBMCs; Fig. 5B) from donors, and also CD8⁺ T-cells enriched from the PBMC pool by FACS (see the ESI†). Strikingly, in one such enriched sample, we identified 4/6-PP-InsP₅ as the sole PP-InsP isomer (Fig. 5C). Of note, the CD8⁺ depleted PBMC pool (Fig. 5D) also contained 4/6-PP-InsP₅ as well as 5-PP-InsP₅ and the latter was identified as the minor isomer. Moreover, a peak comigrating with 2-PP-InsP₅ was identified in PBMCs (Fig. 5B) and can be tentatively assigned to a shoulder of the peak of the CD8⁺ depleted fraction (Fig. 5D). CD8⁺ enrichment did not provide enough material for analysis in all samples studied, so it remains unclear whether the surprising 4/6-PP-InsP₅ enrichment is generally found in CD8⁺ cells from different donors. However, our analysis now firmly establishes that this new isomer is of mammalian origin.

Conclusions

We have applied CE-MS profiling to delineate a more sophisticated picture of InsP and PP-InsP distributions in metazoan samples. Thus, inositol pyrophosphate signalling appears even more complex than previously thought. The CE-MS method also has sufficient sensitivity to profile for the first time biopsies from human patients and PBMCs including isolated CD8⁺ T-cells from human blood. We obtain several unexpected results based on the high separation efficiency of capillary electrophoresis that have gone undetected with recently developed LC-MS approaches.^{28,35,36} In particular, we identify very high levels of PP-InsPs in colon tissue, which are potentially endocytosed from the laboratory diet, including large quantities of the putative noncanonical 4/6 and 2-PP-InsP₅ isomers. Our data therefore represent a paradigm shift in our understanding of dietary influences upon PP-InsP metabolism and signaling in the colon. While 4/6-PP-InsP₅ and 2-PP-InsP₅ in the colon could possibly originate from the endocytosis of food constituents, this phenomenon cannot apply to heart samples as well as human PBMCs. Consequently, it appears that 4/6-PP-InsP₅ and 2-PP-InsP₅ can also be synthesized by mammals.

Our new isomer assignments are based on the exact mass determination and exact comigration with standards of both PP-InsPs, including a novel synthetic 4-PP-InsP₅ bearing two ¹⁸O oxygen isotope labels. Future studies must now address the enantiomeric identity of the new metazoan 4/6-PP-InsP₅ as well as a complete structural assignment of 2-PP-InsP₅ by NMR to firmly establish *myo*-configuration and exclude other potential constitutional isomers of the same mass and identical migration during CE. The colonic uptake, dynamic regulation, unknown enzymology and functions of these new isomers will be productive directions for future research, including their presence in the central nervous system. With the ability to profile PP-InsPs from human biopsies and blood samples, their establishment as potential disease biomarkers will also become an important future endeavor.

Data availability

Additional data can be found in the ESI.†

Author contributions

DQ and CG designed, performed, and evaluated analytical experiments. CG was responsible for the animal experiments. SBS supervised animal experiments. GL and VE performed biopsy extractions and analytics. KR conducted the chemical synthesis. TB and LS isolated and extracted PBMCs. BB supervised human tissue extractions and designed experiments. HJJ and SBS conceived the idea of the project and designed experiments. HJJ, DQ, CG, and SBS wrote the manuscript. All authors provided feedback on experimental design and contributed to manuscript revisions.

Conflicts of interest

There are no conflicts to declare.

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