


 Cite this: *RSC Adv.*, 2023, 13, 33707

Integration of machine learning in 3D-QSAR CoMSIA models for the identification of lipid antioxidant peptides†

Thi Thanh Nha Tran, * Thi Dieu Thuan Tran and Thi Thu Thuy Bui

The comparative molecular similarity indices analysis (CoMSIA) method is a widely used 3D-quantitative structure–activity relationship (QSAR) approach in the field of medicinal chemistry and drug design. However, relying solely on the Partial Least Square algorithm to build models using numerous CoMSIA indices has, in some cases, led to statistically underperforming models. This issue has also affected 3D-CoMSIA models constructed for the ferric thiocyanate (FTC) dataset from linoleic antioxidant measurements. In this study, a novel modeling routine has been developed incorporating various machine learning (ML) techniques to explore different options for feature selection, model fitting, and tuning algorithms with the ultimate goal of arriving at optimal 3D-CoMSIA models with high predictivity for the FTC activity. Recursive Feature Selection and SelectFromModel techniques were applied for feature selection, resulting in a significant improvement in model fitting and predictivity (R^2 , R_{CV}^2 , and R^2_{test}) of 24 estimators. However, these selection methods did not fully address the problem of overfitting and, in some instances, even exacerbated it. On the other hand, hyperparameter tuning for tree-based models resulted in dissimilar levels of model generalization for four tree-based models. GB-RFE coupled with GBR (hyperparameters: learning_rate = 0.01, max_depth = 2, n_estimators = 500, subsample = 0.5) was the only combination that effectively mitigated overfitting and demonstrated superior performance (R_{CV}^2 of 0.690, R^2_{test} of 0.759, and R^2 of 0.872) compared to the best linear model, PLS (with R_{CV}^2 of 0.653, R^2_{test} of 0.575, and R^2 of 0.755). Therefore, it was subsequently utilized to screen potential antioxidants among a range of Tryptophyllin L tripeptide fragments, leading to the synthesis and testing of three peptides: F-P-5Htp, F-P-W, and P-5Htp-L. These peptides exhibited promising activity levels, with FTC values of 4.2 ± 0.12 , 4.4 ± 0.11 , and 1.72 ± 0.15 , respectively.

 Received 2nd October 2023
 Accepted 31st October 2023

DOI: 10.1039/d3ra06690h

rsc.li/rsc-advances

Introduction

The delicate balance between the production and neutralization of reactive species including reactive oxygen species (ROS), reactive nitrogen species (RNS), and reactive sulfur species (RSS) is essential to life.^{1,2} On the one hand, these reactive species participate in a variety of physiological processes within the mitochondria.³ On the other hand, the overproduction of these species, also known as oxidative stress, has been found to cause damage to many biomolecules including proteins, DNA (deoxyribonucleic acid), RNA (ribonucleic acid), and lipids.⁴ The supplement of external antioxidants has been suggested to maintain this delicate balance,^{5,6} with antioxidants being defined as “any substance that when present at low concentrations compared to that of an oxidizable substrate would significantly delay or prevent oxidation of that substrate”.^{7,8}

Antioxidant peptides have been extensively investigated in recent decades using both experimental and statistical approaches. A variety of antioxidant assays have been employed for peptide testing, with some commonly used including ABTS⁺ radical scavenging assay,^{9,10} ferric ion reducing antioxidant power (FRAP),^{11,12} 2,2-diphenyl-1-picrylhydrazyl radical-scavenging capacity (DPPH),^{13,14} oxygen radical absorbance capacity (ORAC),¹⁵ and the FTC method.¹⁶ Consequently, several datasets of antioxidant peptides have been made available to the public through databases.¹⁷ Three of these datasets have been frequently utilized for constructing QSAR models due to their favorable attributes in terms of peptide structure and bioassay homogeneity, which are typically required for statistical modeling. These datasets include the TEAC dataset consisting of 108 synthesized tripeptides, the FTC dataset containing 214 tripeptides,¹⁸ and the FRAP dataset comprising 172 tripeptides.¹⁹ Only a few studies have explored the application of other datasets with varying peptide lengths and bioassays for studying QSAR of antioxidant peptides, as these datasets necessitate special treatment to extract molecular features.^{20,21}

Faculty of Chemical Engineering, Industrial University of Ho Chi Minh City, 12 Nguyen Van Bao, Ho Chi Minh City, 700000, Vietnam. E-mail: tranthithanhnha@iuh.edu.vn

† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d3ra06690h>



QSAR modeling has emerged as an alternative approach to overcome the time and resource-intensive nature of biochemical methods, thereby contributing to the exploration of the chemical space of antioxidants.^{2,5,6} Among the various QSAR techniques, CoMFA and CoMSIA, have been widely utilized in studies exploring the QSAR of antioxidant peptides. In a notable study conducted in 2019, Y. Wang and colleagues developed 3D-QSAR CoMFA and 4-field CoMSIA models utilizing the FTC dataset comprising 198 peptides.²² Similarly, R. Zhang's group constructed CoMSIA models using a TEAC dataset of only 54 tryptophan-containing peptides.²³ Both studies utilized the Sybyl software with built-in Tripos force field and Gasteiger-Hückel charge for model derivation.

Our group has also employed CoMFA and CoMSIA techniques with the OPLS_2005 (Optimized Potentials for Liquid Simulations) force field to build 3D-QSAR models for predicting TEAC values. These models were subsequently employed to screen various tripeptide fragments derived from Tryptophyllin L peptides.^{24,25} This is a peptide family that possesses a characteristic Pro-Trp sequence extracted from the dorsal skin of the frog *Litoria rubella*, an Australian frog occupying a large area of central and northern Australia. Details of the extraction, structures and antioxidant activities of some peptides from this peptide family can be found in the following ref. 26 and 27 and also in our recent publications.^{24,25}

In our previous study, a Gaussian-based 3D-QSAR model constructed using the dataset of ABTS⁺ radical scavenging tripeptides was employed in a combined statistical and experimental approach to identify a range of antioxidant Tryptophyllin L peptides. These peptides exhibited excellent ABTS⁺, DPPH radical scavenging and reducing power. However, neither our research group nor any other has conducted an investigation into the lipid antioxidant properties of these peptides.

During exploration of the TEAC and FTC datasets, we observed a phenomenon that certain peptides displayed significant ABTS⁺ scavenging activity but showed negligible activity in the FTC assay, and *vice versa* (e.g. PWY, PWE, LHG).^{18,28} This observation prompted us to consider the reliability of relying solely on a single model built from just one dataset to predict antioxidant peptides within the Tryptophyllin L family. Consequently, we initiated our investigation into FTC 3D-QSAR modeling and experimental study of antioxidant Tryptophyllin L peptides. This study therefore aims to uncover potential lipid antioxidant Tryptophyllin L peptides that might have been overlooked by previous TEAC models and experiments.

Despite the notable accomplishments thus far, both CoMFA and CoMSIA methods suffer from inherent weaknesses. One such weakness is the excessive number of descriptors typically involved (often several thousands). Among these numerous descriptors, a significant portion is uninformative and irrelevant to biological activities, essentially introducing noise into the models. Without appropriate feature-selection techniques, the models' efficacy is compromised. Furthermore, the linear PLS estimator used as default in these methods, may not adequately capture the nonlinear nature of certain datasets,

leading to subpar statistical performance and predictive power.²⁹

Looking at the field of ML based 3D-QSAR, it can be seen that support vector machine (SVM) has been the most frequently utilized non-linear algorithm for developing field-based models. Several notable studies can be mentioned, such as the analysis of naphthyridone derivatives as ATAD2 bromodomain inhibitors, which employed least squares-support vector machine (LS-SVM) models based on CoMFA-field descriptors.³⁰ Additionally, investigations on reversible acetyl cholinesterase inhibitors were conducted using CoMFA and ligand protein interaction fingerprints.³¹ More recently, G. Floresta and V. Abbate constructed 3D-QSAR models to establish correlations between field descriptors calculated from the extended electron distribution (XED) force field and 5-HT_{2A}R activity. They employed four algorithms, namely *k*-nearest neighbor (kNN), SVM, random forest (RF), and relevance vector machine (RVC), provided by Forge software.³² To the best of our knowledge, there has been no previous research utilizing non-linear machine learning (ML) algorithms to construct 3D-QSAR CoMSIA models in the context of antioxidant activity.

Due to the aforementioned reasons, in this study, we have developed a more flexible approach to constructing 3D-CoMSIA regression models using the FTC dataset. The process of building the models, from data cleaning to feature selection and model construction, will be managed mainly through the utilization of Python scripts. A comprehensive comparison between traditional CoMSIA models and ML-based CoMSIA models will also be conducted. The former will be derived from two different force fields, namely OPLS_2005 and Tripos force fields. The latter will employ ML techniques to select field-similarity-index features and build models using the FTC dataset. The top-performing model will be employed to predict and guide the synthesis and subsequent evaluation of potential lipid antioxidant peptides from Tryptophyllin L family.²⁷ This investigation will also help determine the potential benefits and extent of applying different ML algorithms to 3D-CoMSIA QSAR models.

Material and methods

Data collection, optimization and alignment

The FTC dataset consisting of 214 peptides was collected from the published articles and presented in Table 1.^{18,28} The FTC values were shown as relative activities by adjusting the control to 1.0 (please see the Experimental section for more detail). The duplicates and values of less than 0.1 were then removed resulting in a dataset of 197 peptides.

The structures of 197 peptides were generated using Chem-Draw Professional 15.1 software. Subsequently, each structure underwent optimization using the PM7 method from the Molecular Orbital Package (MOPAC) quantum chemistry program, as described in detail in the ref. 24.

In FTC dataset, three peptides YHY, YKY and YRY display the same highest lipid antioxidant activities (9.886) and share a common structure of two tyrosine at the first and the third



Table 1 The FTC dataset used for model building

No.	Peptide	Activity	No.	Peptide	Activity	No.	Peptide	Activity	No.	Peptide	Activity
1	LHA	3.918	51	PHT	6.247	101	RWN	2.404	151	RYY	2.257
2	LHD	3.593	52	PHV	3.335	102	RWQ	0.606	152	AYY	3.071
3	LHE	6.136	53	PHW	6.535	103	RWR	2.384	153	IYY	3.071
4	LHF	3.628	54	PHY	4.227	104	RWS	0.808	154	LYY	3.071
5	LHG	6.697	55	PWA	1.396	105	RWT	3.818	155	FYY	1.911
6	LHH	4.836	56	PWD	1.096	106	RWV	0.606	156	WYY	1.911
7	LHI	6.531	57	PWE	1.096	107	RWW	2.707	157	GYI	5.071
8	LHK	4.225	58	PWF	0.919	108	RWY	0.808	158	NYI	5.071
9	LHL	5.920	59	PWG	2.687	109	DHH	0.905	159	QYI	5.071
10	LHM	4.504	60	PWH	1.184	110	EHH	0.905	160	MYI	1.991
11	LHN	5.148	61	PWI	1.396	111	AHH	2.020	161	SYI	3.070
12	LHQ	4.136	62	PWK	0.407	112	IHH	2.020	162	TYI	3.070
13	LHR	5.184	63	PWL	1.096	113	FHH	1.803	163	CYI	0.470
14	LHS	4.293	64	PWM	0.796	114	WHH	1.803	164	YDI	3.047
15	LHT	5.584	65	PWN	2.104	115	YHH	1.803	165	YEI	3.047
16	LHV	3.481	66	PWQ	1.202	116	GHH	1.089	166	YHI	9.886
17	LHW	6.791	67	PWR	2.705	117	NHH	1.089	167	YKI	9.886
18	LHY	4.203	68	PWS	1.096	118	QHH	1.089	168	YRI	9.886
19	LWA	1.192	69	PWT	2.598	119	MHH	2.015	169	YAI	3.607
20	LWD	1.717	70	PWV	1.008	120	SHH	1.320	170	YII	3.607
21	LWE	1.717	71	PWW	2.899	121	THH	1.320	171	YLI	3.607
22	LWF	1.414	72	PWY	1.114	122	CHH	0.937	172	YFI	2.233
23	LWG	1.313	73	RHA	5.205	123	HDH	1.477	173	YWI	2.233
24	LWH	3.212	74	RHD	3.304	124	HEH	1.477	174	YGI	3.366
25	LWI	1.111	75	RHE	5.096	125	HAH	0.952	175	YNI	3.366
26	LWK	1.899	76	RHF	3.300	126	HIH	0.952	176	YQI	3.366
27	LWL	0.606	77	RHG	5.725	127	HLH	0.952	177	YMI	1.780
28	LWM	1.394	78	RHH	3.296	128	HFH	2.026	178	YSI	3.447
29	LWN	1.313	79	RHI	4.806	129	HWH	2.026	179	YTI	3.447
30	LWQ	2.505	80	RHK	2.694	130	HYH	2.026	180	YCI	3.087
31	LWR	2.909	81	RHL	3.501	131	HGH	0.832	181	YDI	4.116
32	LWS	2.020	82	RHM	3.218	132	HNH	0.832	182	YEI	4.116
33	LWT	2.020	83	RHN	5.713	133	HQH	0.832	183	YHI	5.303
34	LWV	1.616	84	RHQ	3.108	134	HMH	0.873	184	YKI	5.303
35	LWW	3.515	85	RHR	4.302	135	HSI	0.730	185	YRI	5.303
36	LWY	2.222	86	RHS	3.386	136	HTH	0.730	186	YAI	3.344
37	PHA	5.793	87	RHT	5.987	137	HCH	0.975	187	YDI	3.344
38	PHD	4.622	88	RHV	3.206	138	HDI	0.188	188	YLI	3.344
39	PHE	6.152	89	RHW	5.878	139	HHE	0.188	189	YFI	4.050
40	PHF	3.916	90	RHY	3.378	140	HHI	3.612	190	YWI	4.050
41	PHG	5.197	91	RWA	1.212	141	HHW	3.612	191	YGI	2.996
42	PHH	6.051	92	RWD	0.909	142	HHY	3.612	192	YNI	2.996
43	PHI	4.916	93	RWE	1.091	143	HHG	0.317	193	YQI	2.996
44	PHK	3.426	94	RWF	0.909	144	HHN	0.317	194	YMI	2.103
45	PHL	5.311	95	RWG	1.717	145	HHQ	0.317	195	YSI	3.983
46	PHM	3.714	96	RWH	1.091	146	HHC	0.128	196	YTI	3.983
47	PHN	6.061	97	RWI	1.232	147	DYI	3.417	197	YCI	0.637
48	PHQ	3.718	98	RWK	0.606	148	EYI	3.417			
49	PHR	4.751	99	RWL	3.212	149	HYI	2.257			
50	PHS	4.042	100	RWM	0.727	150	KYI	2.257			

amino acid and one basic amino acid at the second amino acid. As the structure alignment is of pivotal importance for 3D-QSAR modeling, in this study, two sets of aligned structures were prepared, employing either YRY or YHY as the reference molecule.

To align the optimized structures, the Flexible Ligand Alignment Panel in Maestro 11.5 software was utilized. Common scaffolds among the ligands were superimposed using the maximum common substructure and SMARTS, taking into account conformational variations in the side chains. A

round of manual alignment was then performed to ensure proper alignment of side chains not covered by the template molecule. The alignments of all 197 peptides with respect to YRY and YHY are illustrated in Fig. 1A and B.

For model development, the dataset was divided into a training set comprising 158 peptides (80%) and a test set comprising 39 peptides (20%). Different random seeds were employed to assess the reproducibility of the results (refer to ESI 1† for the train-set splits corresponding to different random seeds).



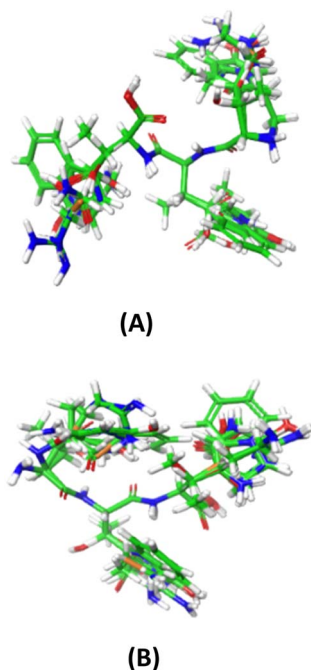


Fig. 1 The FTC aligned structures with (A) YRY and (B) YHY as molecular template.

Construction of 3D-CoMSIA models with OPLS_2005 and Tripos force fields

To assess the impact of two different force fields on the statistical performance of the models, we employed the OPLS_2005 and Tripos force fields on each set of aligned structures to generate CoMSIA models, referred to as OPLS-based and Tripos-based CoMSIA models, respectively. The construction of the OPLS-based models followed the procedure outlined in,^{24,25} while the construction of the Tripos-based models was guided by the Release Notes integrated into Sybyl X 2.1.

In brief, all aligned peptides were positioned within a 3D cubic lattice. Molecular similarity indices were calculated by comparing the similarity of each molecule to a common probe atom, placed at lattice points of the cubic lattice, which had a radius of 1 Å, a charge of +1, and a hydrophobicity of +1. The calculation involved five fields: steric, electrostatic, hydrophobic, hydrogen-bond donor, and hydrogen-bond acceptor. Energy values were truncated at a cutoff of 30 kcal mol⁻¹.

For the OPLS-based models, variables with a standard deviation less than 0.01, an absolute *t*-value smaller than 2, or within a distance of 2.0 Å from any training set atom were eliminated. For the Tripos-based models, the common dataset, exported from Maestro software as sdf files, was imported into Sybyl software. The CoMSIA descriptors were calculated using the calculate properties dialog, with the following parameters: charge calculation using Gasteiger-Hückel, attenuation set to 0.3, and automatic region creation. The models were constructed using the Partial Least Square Analysis module in the QSAR section in the MDE toolbar.

The optimal number of factors for constructing OPLS or Tripos models was determined following specific instructions

in the respective software. Using Maestro, a series of 20 models was created, gradually varying the number of PLS factors from 1 to 20 for each train-test split (random seed). The statistical metrics obtained from each model were compared to identify the most statistically reliable and robust model. These metrics encompassed the coefficient of determination (R^2), the cross-validation coefficient achieved through a leave-one-out approach (R_{LOO}^2), the external validation correlation coefficient (Q^2), the *F* value, the root-mean-square error in test set predictions (RMSE), and $R_{scramble}^2$, which is the coefficient of determination derived from a randomization test. While higher R^2 , R_{CV}^2 (R_{LOO}^2), Q^2 , and *F* values correspond to the higher reliability and predictability of the models, larger values of the remaining metrics suggest the opposite. Consequently, the optimal number of factors was selected to achieve a harmonious balance between these two sets of metrics.

In the case of the Tripos-based models, the optimal number of PLS factors was determined automatically by the software based on the best cross-validation result from 20 models. This optimal PLS number was then employed for all subsequent PLS analyses.

The formulas for the determination of all the aforementioned statistical parameters were presented in ESI 2.† To ensure the consistency of any comparisons, these formulas were employed uniformly for all models developed in this work, unless otherwise stated.

ML based CoMSIA models

The process of constructing ML-based CoMSIA models is depicted in Fig. 2. The model building was performed using Python 3.10.11 on a computer (HP 340S G7 Notebook PC) equipped with an Intel Core i7 1.30 GHz CPU. All subsequent steps, from data pre-processing to model building, were carried out in Jupyter Notebook using various Python modules from the Sklearn library.

Feature extraction

The peptide YRY was chosen in further ML based modeling as it has the longest side-chain at the second amino acid making the alignment more consistent at this position compared to YKY and YHY (two other peptides having the same FTC values).

The CoMSIA descriptors were extracted using the Manage CoMFA module of the QSAR MenuBar in Sybyl X 2.1. A total of 6480 CoMSIA variables were collected (see ESI 3†) and served as the independent variables for building the ML models. These variables were organized into feature columns, with columns 1–1296 corresponding to the steric field, columns 1297–2592 to the electrostatic field, columns 2593–3888 to the hydrophobic field, columns 3889–5184 to the hydrogen-bond acceptor field, and columns 5185–6480 to the hydrogen-bond donor field. Each column was marked to track the contribution of its respective field to the final models. All the required features and estimators were imported from the scikit-learn (<https://scikit-learn.org/>) and XGBoost package (<https://xgboost.readthedocs.io/en/stable/>) for implementation.^{33,34}



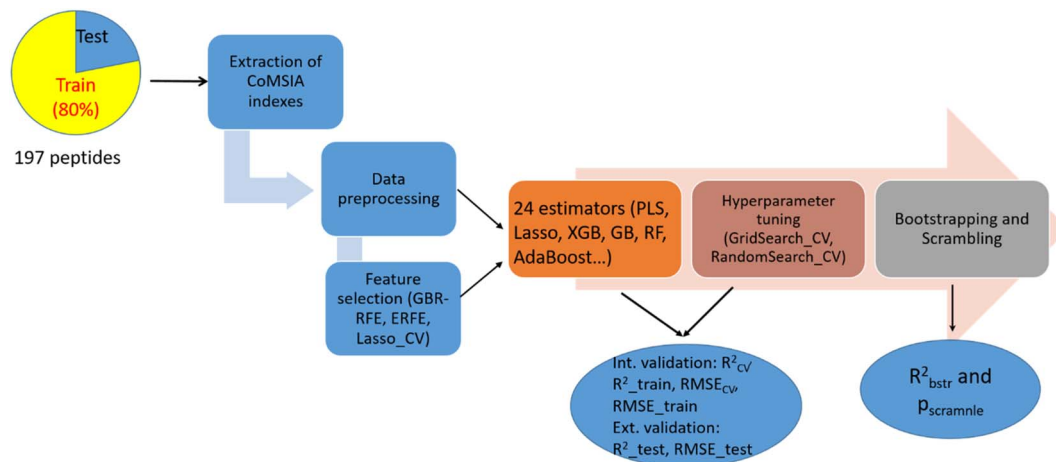


Fig. 2 The process of constructing ML-based CoMSIA models.

Data pre-processing

Several pre-processing steps were applied to the CoMSIA-variable dataset. Firstly, any column with missing values was dropped from the dataset. Additionally, columns containing fewer than 5 values were also removed. Next, a correlation analysis was conducted to identify features (variables) with a correlation greater than 0.95. Among these highly correlated features, the ones that had the least correlation with the target variable were eliminated from the dataset.

Feature selection

Following the data pre-processing step, two feature selection approaches were employed: SelectFromModel-LassoCV and GradientBoosting-Recursive Feature Elimination (GB-RFE). These two methods were deliberately chosen to assess the impact of LassoCV, known for its linear supportiveness, and RFE, which relies on feature importance or coefficient attributes, on the statistical performance of the models.

The SelectFromModel-LassoCV method is a feature selection technique in scikit-learn that selects important features from a dataset based on the coefficients derived from the LassoCV (Lasso Cross-Validation) algorithm. LassoCV is a variant of Lasso regression that incorporates cross-validation to automatically select the regularization parameter. On the other hand, GradientBoosting Recursive Feature Elimination (GB-RFE) is a feature selection technique that combines the Gradient Boosting algorithm with a recursive feature elimination process. It aims to identify and select the most important features by iteratively training a Gradient Boosting model and eliminating the least significant features (using feature importance attributes).

Construction of ML based CoMSIA models

After the data splitting into X_{train} and X_{test} , the training set (X_{train}) was utilized to construct models. During each iteration of the cross-validation loop, a pipeline consisting of a StandardScaler and an estimator was employed to build the models. The `cross_val_score` function was used to split the

X_{train} dataset into internal training and testing subsets for the current fold. The StandardScaler within the pipeline performed two operations on the internal training and testing subsets. Firstly, it fit the scaler on the internal training data ($X_{train_internal}$) and then transformed it to obtain the scaled version. Next, it applied the same scaling transformation on the internal testing data ($X_{test_internal}$) using the parameters learned from the internal training data. The scaled $X_{train_internal}$ and $X_{test_internal}$ were used for training and evaluating the model, respectively, within the current fold of cross-validation. This ensured that the scaling was performed independently for each fold and prevented any data leakage from the testing set to the training set, which is crucial for proper evaluation during cross-validation.

In this study, three groups of models were constructed sequentially, each serving different analytical purposes that will be discussed in the Result and discussion section. The first group of models was built after the data pre-processing step using 24 estimators with default hyperparameters and without any feature selection (Script S1†). The second group of models was constructed using one of the aforementioned feature selection methods (Script S2.1–2.2†). In the first two groups, several parameters, including R_{CV}^2 , Root Mean Squared Error (RMSE), Std_RMSE, and R^2 (coefficient of determination for the training set), were computed to compare the performance and generalization of the models. R^2_{test} (coefficient of determination for the test set) was used to assess their predictability.

Finally, several models with the best cross-validation (CV) statistics from the second group were selected for hyperparameter tuning, leading to the creation of the third group of models (Script S3.1–3.4†). Grid search and random search techniques were employed, along with five-fold cross-validation, to identify the optimal hyperparameters for the models. These models were trained and evaluated on the inner folds of the training set using different hyperparameter combinations, and the best hyperparameters were chosen based on their CV performance. GridSearch_CV has also been used to derive the optimum number of PLS components using each phase of feature selection (Script S4.1–4.4†).



RFE selection methods utilizing different estimators including RandomForest, XGBoost, and AdaBoost (Embedded-RFE of RF, XGB, Ada), were executed to assess the impact of these selection methods on the overall model performance (Script S5.1–5.3†). Bootstrapping (R_{bstr}^2) and scrambling (p value) evaluation were also performed to assess the robustness and reliability of the ML-based models (Script S6.1–6.2†).

Experimental

Materials

All peptides were prepared using L-isomers of each amino acid by solid-phase synthesis with the fluorenylmethoxycarbonyl (Fmoc) strategy. The synthesis was conducted by GL Biochem Co., Ltd. The purities were approximately 95% as evidenced by high-performance liquid chromatography (HPLC) and mass spectrometry (MS) data. Linoleic acid (~95%) was purchased from Sigma Chemical (St. Louis, MO). Ammonium thiocyanate, ferrous chloride and other reagents were obtained with the analytical grade. Thermo Scientific Genesys 20 served as the equipment in all UV-Vis measurements.

Ferric thiocyanate assay

The ferric thiocyanate assay was conducted following the procedure described in the ref. 18. Test samples dissolved in 0.5 mL of deionized water were combined with linoleic acid emulsion (1.0 mL, 50 mM) and phosphate buffer (1.0 mL, 0.1 M) in 5 mL glass test tubes. The final concentration of test sample is 40 μM . The test tubes were tightly sealed with silicon rubber caps and placed in a dark environment at 60 °C. Throughout the incubation period, small aliquots (50 μL) of the reaction mixtures were extracted at various intervals.

To assess the extent of oxidation, sequential additions of ethanol (2.35 mL, 75%), ammonium thiocyanate (50 μL , 30%), and ferrous chloride (50 μL , 20 mM in 3.5% HCl) were made to the extracted reaction mixtures. After allowing the mixture to stand for 3 minutes, the absorbance of the solution was measured at 500 nm. A control sample, excluding the peptides but containing the same components as the test sample, was prepared. The induction period, denoting the time required to reach an absorbance of 0.3, was calculated. The relative activities of the test samples were determined by dividing their respective induction periods by that of the control sample. All experiments were conducted in triplicate, and the average values were recorded.

Result and discussion

Data distribution

The distribution of activities for 197 peptides in the FTC dataset was shown in Fig. 3, revealing a right-skewed pattern with a skewness of 0.943, where the number of peptides with lower activities is significantly higher than those with larger activities.

However, it is important to note that linear regression remains robust to deviations from normality in the target variable itself, as long as the residuals or prediction errors meet the

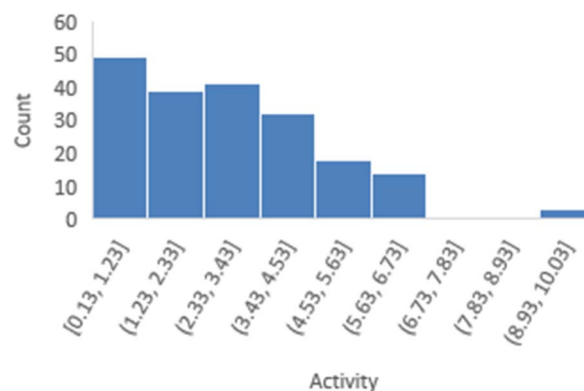


Fig. 3 The distribution of activities for 197 peptides in the FTC dataset.

assumption of a normal distribution. This robustness is supported by the Central Limit Theorem, making it less necessary for the target variable to follow a normal distribution.³⁵ Therefore, in the case of skewness less than 1, we have chosen not to transform the target variable as it will complicate the explanation of the predicted results, and we will discuss the distribution of prediction errors in the next section.

OPLS-based CoMSIA models

Fig. 4 illustrates the correlation between key statistical parameters obtained from OPLS-3D-CoMSIA models as the number of factors varied from 1 to 20. A compromise is achieved between the highest R^2 , R_{CV}^2 (R_{LOO}^2), Q^2 values, and the lowest R_{Scramble}^2 and RMSE_test values at 3 factors for both YHY (Fig. 4A) and YRY (Fig. 4B) aligned datasets, using the same random seed.

The R^2 , R_{CV}^2 , Q^2 values for the first superimposed set of structures are 0.63, 0.55, and 0.63, while R_{Scramble}^2 and RMSE_test are 0.12 and 1.22, respectively. Similarly, for the YRY aligned dataset, the R^2 , R_{CV}^2 , and Q^2 values are 0.64, 0.54, and 0.46, with R_{Scramble}^2 and RMSE_test at 0.13 and 1.47, respectively. These results indicate the performances of the OPLS-based models are moderate for both superimposed datasets in terms of model fitting and predictivity.

The correlation pattern remains generally consistent across five different data splits for each aligned set, resulting in all OPLS-based CoMSIA models having a common number of 3 PLS factors for optimal statistics. Observation of cross-validation (R_{CV}^2) and prediction coefficients (Q^2) for 5 random seeds in both YHY and YRY alignments reveals that the change in the molecular template does not affect the statistical results of OPLS-based models substantially, and these values do not mutually exceed 0.6. This reaffirms the moderate performance of all OPLS-based models. For detailed statistical information related to factor selection and PLS analysis in OPLS-based models, please refer to ESI 4.†

To assess the reliability of the 3-factor-CoMSIA models, distribution plots and skewness calculations for prediction errors were conducted using various random seeds. Fig. 5 depicts a distribution plot of prediction errors for random seed 1 (YHY-reference). This distribution illustrates a general normal distribution centered around 0 but still exhibits a long left tail,



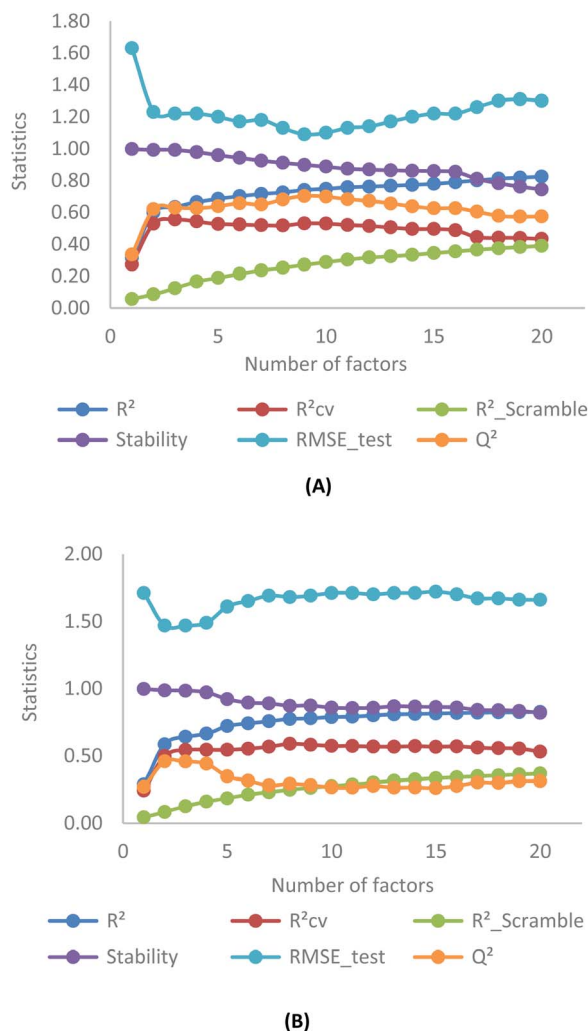


Fig. 4 Statistics obtained from OPLS-3D-CoMISA models with YHY (A) and YRY (B) as templates.

highlighting two primary characteristics of this model type: the reliability of statistical inference, and the model's limitation in adequately predicting structures with extreme activities. Similar distribution patterns were also observed for five other train-test splits, as presented in ESI 1.†

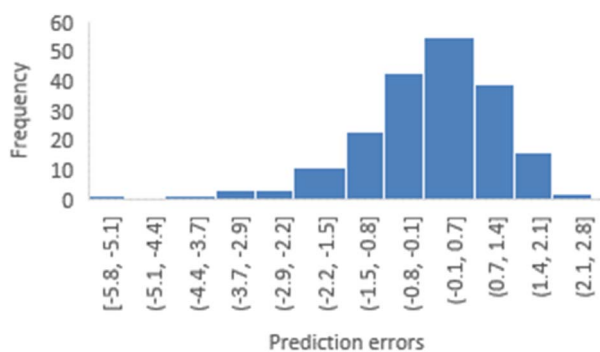


Fig. 5 Distribution of prediction errors derived from the 3-factor-CoMISA model with YRY as the reference.

Table 2 Statistics of the selected Tripos CoMSIA-based models^a

Tripos-CoMSIA model	Optimal factor	R^2	R_{LOO}^2	R_{bstr}^2	SEP	Q^2
YHY_ref	11	0.756	0.446	0.781	1.432	0.547
YRY_ref	18	0.828	0.531	0.684	1.350	0.339

^a SEP: standard error of prediction.

Tripos-based CoMSIA models

It is important to note that the cross-validation coefficient is denoted in this study as R_{LOO}^2 instead of Q_{LOO}^2 , as provided by the Sybyl X software's output. This choice was made to prevent any potential confusion with the variance explained in external prediction, which is also referred to as Q^2 . Additionally, the evaluation of a model based only on R^2 and Q_{LOO}^2 as reported by a number of studies^{22,23} is insufficient as it could not represent the performance of the model on the unseen data.^{36–38} For that reason, in this study, parameters for both internal and external assessment were evaluated to arrive at conclusions regarding each model's performance.

To facilitate direct comparison between the OPLS and Tripos-based CoMSIA models, all the Tripos models were constructed using the same random seed used for OPLS models ranging from 1 to 5, thereby maintaining consistent train-test compositions (ESI 1†). The statistical results for two of the Tripos-based CoMSIA models built with random seed 1 are presented in Table 2.

As the optimal number of factors (PLS number) was determined solely based on R_{CV}^2 , the PLS number in Tripos-based CoMSIA models is significantly higher than that in OPLS models when using the same random seed and molecular template. This method of PLS factor optimization also leads to variation in the PLS number across different random seeds, as it responds promptly to changes in the composition of the training set.

Furthermore, the Tripos-based CoMSIA models tend to exhibit overfitting, as indicated by substantial discrepancy between R^2 (0.756 and 0.828) and R_{CV}^2 (0.446 and 0.531) for YHY and YRY aligned sets, respectively. For comprehensive statistical details regarding Tripos-based models generated with five different random seeds during factor selection and PLS analysis, please refer to ESI 5.†

Predictability plots for the OPLS and Tripos based models

The predictability plots for the OPLS models are illustrated in Fig. 6, while those for the Tripos force field are displayed in Fig. 7. These plots indicate that both models have inadequate predictive performance, severely underestimating the FTC values of peptides in the high activity range. The root-mean-square error in the test-set predictions from the OPLS model reaches as high as 1.460, which is similar to the standard error of prediction from the Tripos model (1.432). These errors are considered significant considering the range of FTC activity in the dataset only up to nearly 10. All prediction performances of OPLS and Tripos-based models can be found in ESI 4 and 5.† To the best of our knowledge, the only study constructing 3D-



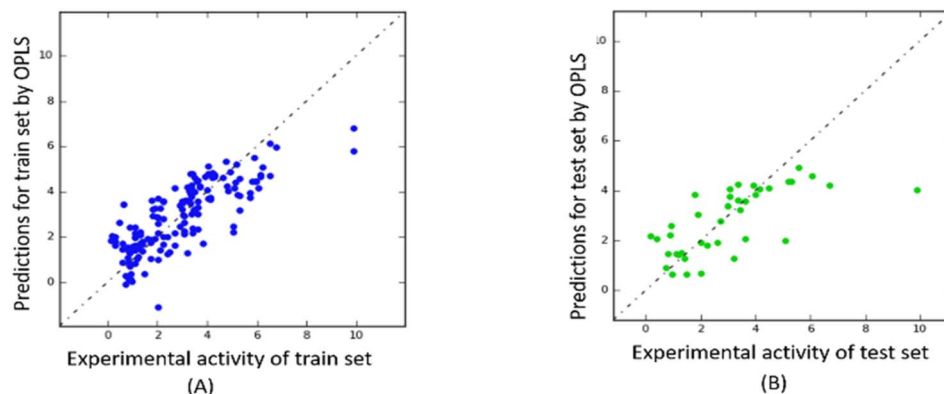


Fig. 6 Predictability plots of the OPLS models for the training set (A) and for test set (B), random seed 1, YRY reference.

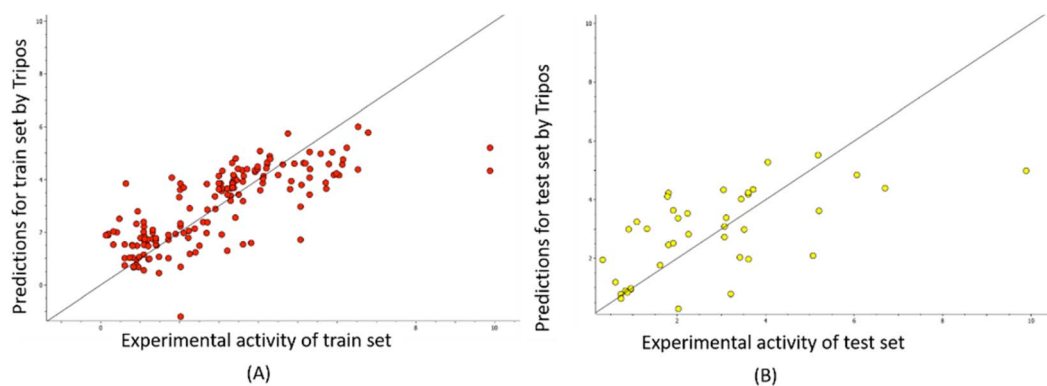


Fig. 7 Predictability plots of the Tripos models for the training set (A) and for test set (B), random seed 1, YRY reference.

CoMSIA models using the FTC dataset was implemented with YRY being the alignment template using the Sybyl X software. The models were evaluated based on only two statistical parameters (R^2 of 0.914 and Q^2 of 0.733) and no evaluation of external prediction was concluded.²² Thus, in our opinion, there are no reliable 3D-CoMSIA models for this data set have been reported before this study.

ML based CoMSIA models without variable selection

The primary reason for using Python scripts, instead of fixed functions within commercial software, to construct 3D-CoMSIA models for the FTC dataset was to allow for more flexibility in experimenting with various feature selection methods and regression algorithms. This approach aimed to mitigate the impact of the abundant number of CoMSIA variables and the imbalanced distribution of the FTC activities that could not be handled successfully by the traditional PLS modeling routine.

The training dataset containing 6840 CoMSIA variables underwent preprocessing, resulting in 1282 variables that were used as inputs for the first group of models. The statistics of these models are presented in Table 3. Without employing any feature selection method, all the models performed below statistical expectations. The primary factor contributing to this poor performance is the high dimensionality of the features and the small sample size.

Among 24 regression estimators, the GradientBoosting-Regressor (GBR) exhibited the best performance, with an R_{CV}^2 value of 0.500 and a Root Mean Square Error for cross-validation ($RMSE_{CV}$) of 1.326. Generally, the tree-based, Lasso and Bayesian Ridge outperformed other regressors, which can be attributed to the inherent feature-selection nature of these algorithms. This feature has helped to mitigate the impact of the high number of CoMSIA indices derived from the FTC dataset. The PLS model with 3 components demonstrated less effective cross-validation estimation compared to the OPLS_2005 and Tripos models, achieving an R_{CV}^2 of 0.301 and an $RMSE_{CV}$ of 1.515.

Interestingly, the coefficients of determination for the test set (R^2_{test}) and training set (R^2) were remarkably higher than the R_{CV}^2 for most models, particularly for the tree models. This suggests the presence of overfitting in these models. For example, the GBR model exhibited an R_{CV}^2 of 0.500 and an R^2 of 0.995, indicating the need for feature selection to achieve more reliable models.

ML based CoMSIA models with feature selection

The GB-RFE method was applied to the 1282 variables obtained from the data preprocessing step for feature selection from the YRY-aligned structure set. The elimination of variables was performed iteratively, with 20 features being removed at each



Table 3 Performance parameters of 24 ML-based 3D-CoMSIA models without feature selection

Regression algorithm	R_{CV}^2	RMSE _{CV}	R^2_{test}	RMSE _{test}	R^2	RMSE
GradientBoostingRegressor()	0.500	1.290	0.812	0.872	0.995	0.125
RandomForestRegressor()	0.465	1.332	0.812	0.870	0.930	0.489
Lasso(alpha = 0.1)	0.415	1.391	0.534	1.371	0.719	0.979
BayesianRidge()	0.413	1.377	0.688	1.123	0.760	0.904
HistGradientBoostingRegressor()	0.400	1.400	0.754	0.996	0.970	0.319
TweedieRegressor()	0.371	1.420	0.666	1.161	0.807	0.811
AdaBoostRegressor()	0.344	1.442	0.824	0.844	0.866	0.675
SVR(epsilon = 0.2)	0.306	1.520	0.309	1.670	0.589	1.183
PLSRegression(n_components = 3)	0.301	1.500	0.646	1.195	0.702	1.008
BaggingRegressor(base_estimator = SVR())	0.297	1.531	0.292	1.691	0.568	1.213
XGBRegressor(booster = None)	0.230	1.569	0.832	0.824	1.000	0.000
ElasticNet()	0.221	1.603	0.190	1.808	0.272	1.575
DecisionTreeRegressor()	0.122	1.669	0.504	1.415	1.000	0.000
KNeighborsRegressor(n_neighbors = 2)	0.032	1.749	0.497	1.426	0.657	1.081
NuSVR(nu = 0.1)	0.018	1.789	0.103	1.903	0.186	1.666
QuantileRegressor()	-0.042	1.850	0.000	2.010	-0.002	1.848
MLPRegressor()	-0.372	1.964	0.628	1.226	0.999	0.052
HuberRegressor()	-0.835	2.246	0.219	1.776	0.845	0.727
GaussianProcessRegressor()	-2.700	3.430	-2.175	3.580	1.000	0.000
XGBRegressor(booster = 'gblinear')	-3.393	3.328	-0.270	2.265	0.885	0.625
Ridge()	-6.160	3.901	-0.918	2.782	0.897	0.593
KernelRidge()	-8.350	4.958	-2.352	3.679	-1.580	2.965
LinearSVR(random_state = 1, tol = 1 × 10 ⁻⁵)	-8.630	4.607	-0.903	2.772	0.852	0.710
LinearRegression()	-702.889	40.070	-116.058	21.739	1.000	0.000

iteration. No significant difference was observed in the resulting models when varying the number of variables eliminated at a time (20, 10, or 5). To optimize computational efficiency, the removal of 20 features per iteration was chosen. This recursive process continued until the desired number of features was reached. Among the pruned dataset, twelve variables (feature columns) were selected for the training dataset, specifically columns [2240, 2946, 3077, 3251, 3257, 4857, 5566, 5657, 5688,

5831, 5961, and 5987]. These variables corresponded to the lowest Root Mean Square Error (RMSE) for the associated GBR model, indicating the importance of hydrophobicity and hydrogen-bond donor in relation to FTC activity.

Furthermore, the SelectFromModel-LassoCV method with a threshold of 0.01 identified 37 variables that contributed significantly to the Lasso models. Among these variables, the variable corresponding to the hydro-bond donor field still

Table 4 Performance of 20 ML based CoMSIA models with two different feature selection strategies

GB-RFE	R_{CV}^2	RMSE _{CV}	R^2_{test}	RMSE _{test}	R^2	RMSE
GradientBoostingRegressor(random_state = 1)	0.644	1.072	0.756	0.993	0.977	0.280
RandomForestRegressor()	0.638	1.094	0.760	0.985	0.950	0.413
XGBRegressor(booster = None)	0.624	1.095	0.721	1.061	1.000	0.001
AdaBoostRegressor()	0.607	1.131	0.629	1.224	0.853	0.708
HistGradientBoostingRegressor()	0.529	1.245	0.668	1.158	0.916	0.535
BaggingRegressor(base_estimator = SVR())	0.449	1.358	0.331	1.643	0.565	1.218
SVR(epsilon = 0.2)	0.447	1.360	0.337	1.636	0.578	1.200
Lasso(alpha = 0.1)	0.377	1.416	0.472	1.461	0.509	1.293
BayesianRidge()	0.355	1.420	0.476	1.454	0.542	1.249
KNeighborsRegressor(n_neighbors = 2)	0.328	1.484	0.510	1.406	0.783	0.860
SelectFromModel_LassoCV						
PLSRegression(n_components = 3)	0.653	1.058	0.575	1.310	0.744	0.934
TweedieRegressor()	0.649	1.068	0.596	1.277	0.598	1.170
BayesianRidge()	0.621	1.112	0.646	1.196	0.662	1.074
GradientBoostingRegressor()	0.514	1.252	0.700	1.101	0.974	0.300
RandomForestRegressor()	0.514	1.266	0.727	1.049	0.934	0.473
Lasso(alpha = 0.1)	0.506	1.278	0.617	1.244	0.606	1.158
KNeighborsRegressor(n_neighbors = 2)	0.503	1.263	0.434	1.511	0.800	0.825
MLPRegressor()	0.495	1.245	0.666	1.162	0.655	1.084
XGBRegressor(booster = None)	0.485	1.300	0.667	1.159	1.000	0.001
AdaBoostRegressor()	0.476	1.316	0.597	1.276	0.803	0.819



showed essential importance. Additionally, less impacts on FTC activity were observed for 3 other fields including electrostatic, hydrophobic and hydro-bond acceptor, as indicated by the ratios of selected variables representing these fields. The selected variables were as follows: [209, 664, 693, 913, 1414, 1458, 1582, 1833, 1906, 1928, 2216, 2345, 3060, 3191, 3251, 3299, 3442, 3535, 3620, 4262, 4463, 4664, 4723, 4747, 4866, 5277, 5421, 5545, 5566, 5670, 5831, 5843, 5855, 5891, 6004, 6125, and 6403].

The top ten models selected based on their cross-validation R_{CV}^2 and $RMSE_{CV}$ for each feature selection method are presented in Table 4. Several noteworthy points can be highlighted. Firstly, the implementation of feature selection has improved the cross-validation estimation of all models. Secondly, the GB-RFE feature selection method has yielded advantages for tree-based models (GradientBoosting, RandomForest, XGBoost, and AdaBoost), while the SelectFromModel-LassoCV has shown benefits for linear models (PLS, Lasso, and BayesianRidge). The GBR model stood out as the best performer in the first group, achieving an R_{CV}^2 of 0.644, $RMSE_{CV}$ of 1.072, R^2_{test} of 0.756, and R^2 of 0.977. In the second group, the PLS model showcased the highest R_{CV}^2 of 0.653, $RMSE_{CV}$ of 1.058, R^2_{test} of 0.575, and R^2 of 0.755. However, there was a significant overlap of estimators between the two top-ten model groups, indicating that these estimators are suitable for developing 3D CoMSIA models of FTC dataset.

Among the 24 distinct regression techniques examined, non-linear regression methods consistently demonstrated superior fitting performance when assessed across the two feature selection methods. This underscores the presence of non-linearity in predicting antioxidant activity and, in turn, provides an explanation for the subpar model performance observed in the previous section and prior studies that relied on linear regression methods.²²

For a small dataset like FTC, the superior performance of GB-RFE tree-based models compared to Lasso-PLS can be attributed to their intrinsic algorithms. PLS is particularly sensitive to outliers because the linear regression line is directly influenced by the mean of the target variable during fitting. In contrast, extreme target values only affect local trees and the local splitting decisions in tree-based models, resulting in an improved overall perform for GBR, RandomForest, XGBoost, and AdaBoost.

ML based CoMSIA models with hyperparameter tuning

The disparity of R_{CV}^2 , R^2_{test} , and R^2 in Table 4 suggests that the feature selection only improved the cross-validation performance but still could not fix the problem of overfitting completely for tree-based models. Hyperparameter tuning using GridsearchCV was carried out on hundreds of hyperparameter combinations to explore the improvement of generalization ability for the four tree-based models.

Fig. 8 visually illustrates the superior performance of tree-based models when compared to the LassoCV-PLS model (with $n = 3$ components). Specifically, while the cross-validation R_{CV}^2 of the PLS model is comparable to that of the four tree-based models, the tree-based models with tuned hyperparameters clearly outperform the LassoCV-PLS model in predictivity.

After hyperparameter tuning, the GBR model with specific settings (learning_rate = 0.01, max_depth = 2, n_estimators = 500, subsample = 0.5) showed the most significant improvement in model generalization, with notable reductions in the differences between R_{CV}^2 , R^2 and R^2_{test} (0.690, 0.872, and 0.759, respectively). Likewise, the $RMSE_{CV}$, $RMSE$ and $RMSE_{test}$ values for the GBR model decreased to 1.042, 0.66, and 0.987, respectively.

On the other hand, the other tree-based models, including RandomForest, XGB, and AdaBoost, unexpectedly did not achieve the same level of improvement, as indicated by larger disparities between their coefficients. There are various reasons contributing to the dissimilar response between GBR and the other three models to hyperparameter tuning, with common factors being feature selection, data size, and incomplete hyperparameter tuning. Since testing all possible measures is not feasible, we conducted an additional experiment involving Embedded RFE (ERFE) feature selection coupled with hyperparameter tuning with GridSearchCV.

The ERFE feature selection was applied with three different estimators (RF, XGB, and Ada). However, the results indicated that the ERFE method did not improve the overfitting issue; in fact, it exacerbated it, as shown in Fig. 9. The application of the Embedded XGB model revealed a notable escalation in overfitting, evidenced by a rise in R^2 to 1.000. This increase further widened the gap between this parameter and R_{CV}^2 (0.645) as well as R^2_{test} (0.764). Similar trends were observed in the RF and Ada models, where the discrepancy of these coefficients also intensified.

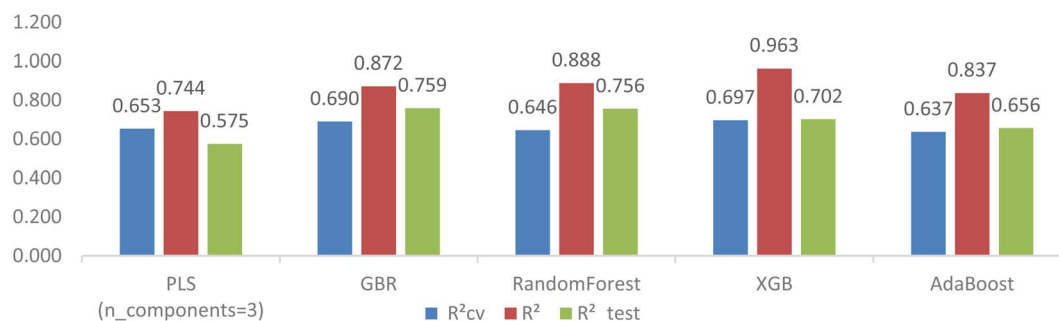


Fig. 8 Comparison of PLS with GBR-RFE tree-based models after hyperparameter tuning.



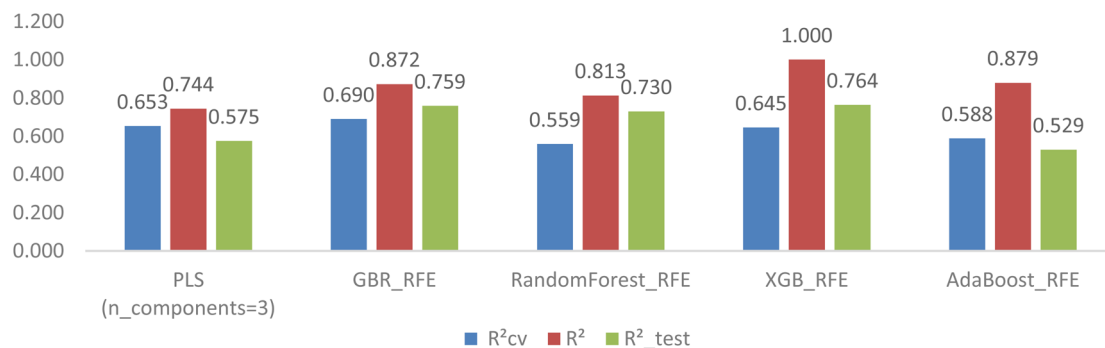


Fig. 9 Comparison of PLS with ERFE tree-based models after hyperparameter tuning.

Bootstrapping and Y-scrambling evaluation for the GB-RFE GBR model

The bootstrapping evaluation has generated R_{bstr}^2 of 0.703 (standard deviation (SD): 0.056), average MSE: 1.200 (SD: 0.227). These results are similar to R^2_{test} of the GBR model, indicating that the model's performance is consistent, robust, and not dependent on the specific sample of data used for training.

The result of Y-scrambling activity for the GBR model has shown a p value <0.001 after 100 iterations, suggesting that the model's performance is not due to random chance and strengthening its reliability and applicability.

Predictability of the GB-RFE GBR model for the FTC dataset

Fig. 10 shows the correlation between predicted and experimental FTC values for the training and test sets obtained from the GBR model. The plot demonstrates the excellent fitting of the GBR model compared to the Triplos and OPLS-based CoMSIA models for the training set, even at the highest activity levels. However, its performance was comparatively less efficient for the test set, particularly in the activity range higher than 6 (please refer to ESI 6† for all the predictions on the FTC dataset using the RFE-GBR model with the optimized hyperparameters).

Predictions of FTC activity of Tryptophyllin L tripeptides

The GBR model, employing the GB-RFE selection method and specific hyperparameters (learning_rate = 0.01, max_depth = 2,

$n_{estimators} = 500$, subsample = 0.5), was identified as the most optimal 3D-CoMSIA model constructed using FTC data, and therefore was used to predict the lipid antioxidant activity of 13 Tryptophyllin L peptide fragments.

The two most important field effects on lipid antioxidant capability, including hydrophobicity and hydrogen bond donor, were illustrated in Fig. 11. A notable observation is the positive impact of hydrophobicity in the first amino acid position on overall activity, but interestingly, this effect reverses when the

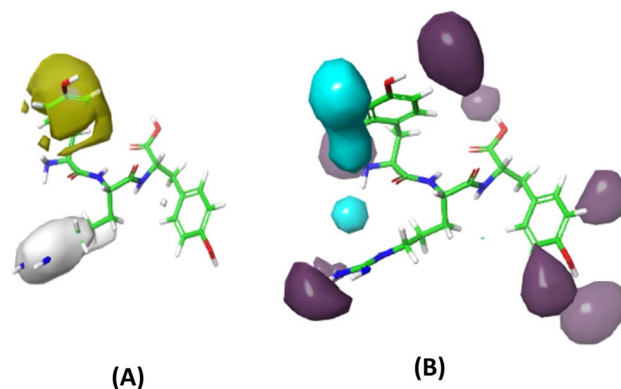


Fig. 11 Contour maps of field contribution to lipid antioxidant activity (A) hydrophobicity (yellow: positive, white: negative) and (B) hydrogen-bond donor (purple: positive, cyan: negative).

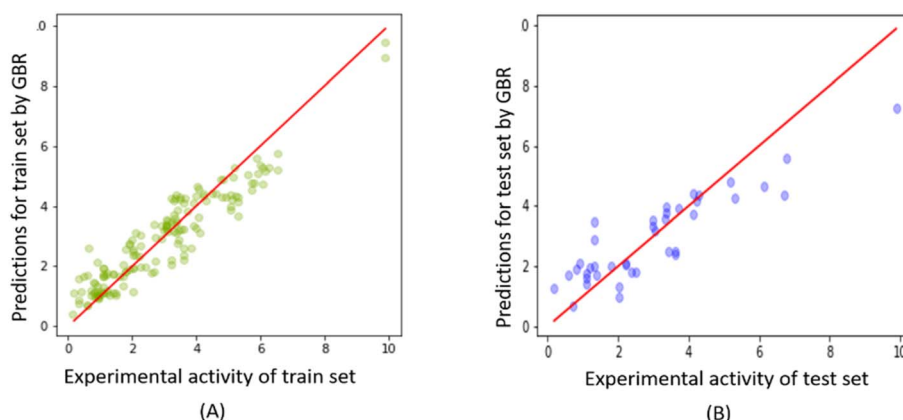


Fig. 10 Experimental versus predicted FTC values derived from the GBR model for (A) FTC training and (B) test set.



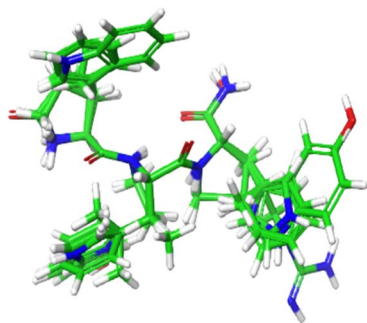


Fig. 12 Structures of 13 Tryptophyllin L tripeptides superimposed on the YRY reference.

hydrophobic amino acid is in the second position. Additionally, hydrogen-bond donor groups are found to be beneficial to FTC activity when located on the second and third amino acid positions, while they have a mixed effect on the first amino acid.

Structures of 13 Tryptophyllin L tripeptides superimposed on the YRY reference are displayed in Fig. 12. The CoMSIA indices from these peptides were subjected to the RFE-GBR model to predict the FTC activities of these peptide fragments (ESI 7[†]). Table 5 presents 13 Tryptophyllin L tripeptides along with their corresponding predicted FTC values. Based on the predictions from the GBR models, the peptides F-P-W and F-P-5Htp were identified as the highest FTC activities.

The most interesting observation from the predictions on Tryptophyllin L tripeptides was the combined effect of phenylalanine at the first amino acid and tryptophan at the third amino acid on the FTC activity. This combination provides hydrophobicity for the first position and hydrogen-bond donor for the C-terminal leading to the highest FTC activity in the list. The FTC value does not change significantly when tryptophan is replaced by 5-hydroxytryptophan (5Htp) explained by the subtle difference in hydrogen-bond donor ability of 5Htp compared with that of tryptophan.

Three tripeptides were synthesized and tested using the FTC assay, including FPW, F-P-5Htp, and P-5Htp-L. The latter two

tripeptides are derived from the Tryptophyllin L F-P-5Htp-L, is of special interest as containing 5-Htp one of the tryptophan metabolites. Experimental results yielded FTC values of 4.2 ± 0.12 , 4.4 ± 0.11 , and 1.72 ± 0.15 for F-P-5Htp, F-P-W, and P-5Htp-L, respectively. These experimental results generally align with the predictions of the RFE-GBR model, suggesting the potential lipid antioxidant properties of these three peptides.

Conclusion

In conclusion, this study conducted a comprehensive comparison of two modeling approaches to determine the most statistically reliable and robust model for predicting the lipid antioxidant activities of tripeptides. The first approach utilized the traditional PLS algorithm within Maestro 11.5 and Sybyl X 2.1, while the second approach employed various machine learning algorithms for model selection.

The analysis of the FTC CoMSIA dataset using traditional methods revealed several key findings. Firstly, changing the molecular template for alignment had no significant impact on the CoMSIA models' statistics. However, switching between force fields resulted in notable differences. OPLS-based models exhibited more stable PLS numbers and statistical parameters compared to Tripos force field models, which displayed variations in PLS numbers across different data splits and tended to overfit. Secondly, the CoMSIA models from both force fields showed low predictive capability (Q^2) with a strong inclination to underestimate FTC activity, particularly in the high activity range.

The ML-based modelling routine was intentionally designed to process the data into three stages. The ML-based 3D-CoMSIA models without feature selection have revealed better performances for four tree-based regressors (RandomForest, XGB, AdaBoost and GBR), Lasso and Bayesian Ridge Regressor as a result of the feature self-selected nature of these estimators. This feature is beneficial greatly in alleviating the detrimental impact of the abundance of CoMSIA indices on the models.

Out of the 24 distinct regression techniques investigated, non-linear regression methods consistently exhibited better model fitting than the linear methods when evaluated using the two feature selection methods GBR-RFE and Select-FromModel_LassoCV. This highlights the presence of non-linearity in predicting antioxidant activity, thus offering an explanation for the less favorable performance of linear models previously.

The competition of four tree-based models at the final stage of hyperparameter tuning has revealed different levels of generalization for these tree-based models, although improvement in terms of cross-validation and predictivity were observed for all of these models. Among those, the GBR model with the GBR-RFE selection method and specific hyperparameters (learning_rate = 0.01, max_depth = 2, n_estimators = 500, subsample = 0.5) was selected as the best model for predicting the FTC activity of tripeptides when displaying smallest disparity in internal and external statistics. It is therefore statistically reliable to be used for screening of lipid-antioxidant tripeptides.

Table 5 FTC predictions by GBR mode^a

No.	Title	FTC predictions by GBR model
1	FPW	3.882
2	pEFP	3.476
3	FPF	2.556
4	IPW	3.225
5	FLP	1.996
6	PWF	1.287
7	PWF(NH ₂)	1.304
8	FHR(NH ₂)	2.119
9	PWP	1.772
10	PFP	1.755
11	WFH	2.443
12	P-5Htp-L	1.818
13	F-P-5Htp	3.694

^a 5Htp: 5 hydroxytryptophan, pE: pyroglutamic acid.



Overall, the ML-based modeling approach has demonstrated greater efficiency compared to the traditional CoMSIA method when modeling 3D-field similarity models for the FTC dataset. This conclusion is based not only on the comparison of models constructed in this study but also on a review of results from previous research.

The predicted FTC values of 13 Tryptophyllin L peptide fragments by the RFE-GBR model have guided the experimental testing of three tripeptides, yielding FTC values of 4.2 ± 0.12 , 4.4 ± 0.11 , and 1.72 ± 0.15 for F-P-5Htp, F-P-W, and P-5Htp-L, respectively. The experimental results generally aligned with the model predictions, suggesting that these peptides have great potential as lipid antioxidants and should be further tested on food-based samples.

Finally, this study offers a collection of adaptable Python scripts (please refer to the attached Script S1–S6† for relevant code) that can be used to model various bioactivities or properties, requiring only an input CoMSIA dataset as a minimum requirement. These scripts enable model construction using 24 estimators, both with and without feature selection. Notably, the scripts incorporate features such as variable selection (GBR-RFE, ERFE, and SelectFromModel) and hyperparameter tuning through GridSearch_CV and RandomSearch_CV, scrambling and bootstrapping evaluation, allowing the estimation of model consistency and reliability.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

We express our gratitude to the Faculty of Chemical Engineering at the Industrial University of Ho Chi Minh City for their generous provision of instruments and invaluable support during the antioxidant testing conducted in this study.

Notes and references

- 1 N. Wiernsperger, Oxidative stress as a therapeutic target in diabetes: revisiting the controversy, *Diabetes Metab.*, 2003, **29**(6), 579–585.
- 2 M. Carocho and I. C. Ferreira, A review on antioxidants, prooxidants and related controversy: natural and synthetic compounds, screening and analysis methodologies and future perspectives, *Food Chem. Toxicol.*, 2013, **51**, 15–25.
- 3 M. Ebadi, Antioxidants and free radicals in health and disease: an introduction to reactive oxygen species, oxidative injury, neuronal cell death and therapy in neurodegenerative diseases, *Crit. Rev. Toxicol.*, 2001, **38**, 13–71.
- 4 H. Sies, C. Berndt and D. P. Jones, Oxidative stress, *Annu. Rev. Biochem.*, 2017, **86**, 715–748.
- 5 C. Vassalle, M. Maltinti and L. Sabatino, Targeting oxidative stress for disease prevention and therapy: where do we stand, and where do we go from here, *Molecules*, 2020, **25**(11), 2653.
- 6 H. J. Forman and H. Zhang, Targeting oxidative stress in disease: promise and limitations of antioxidant therapy, *Nat. Rev. Drug Discovery*, 2021, **20**(9), 689–709.
- 7 B. Halliwell, M. A. Murcia, S. Chirico and O. I. Aruoma, Free radicals and antioxidants in food and in vivo: what they do and how they work, *Crit. Rev. Food Sci. Nutr.*, 1995, **35**(1–2), 7–20.
- 8 B. Halliwell and M. Whiteman, Measuring reactive species and oxidative damage in vivo and in cell culture: how should you do it and what do the results mean?, *Br. J. Pharmacol.*, 2004, **142**(2), 231–255.
- 9 R. Re, N. Pellegrini, A. Proteggente, A. Pannala, M. Yang and C. Rice-Evans, Antioxidant activity applying an improved ABTS radical cation decolorization assay, *Free Radicals Biol. Med.*, 1999, **26**(9–10), 1231–1237.
- 10 L. Zheng, M. Zhao, C. Xiao, Q. Zhao and G. Su, Practical problems when using ABTS assay to assess the radical-scavenging activity of peptides: Importance of controlling reaction pH and time, *Food Chem.*, 2016, **192**, 288–294.
- 11 I. F. Benzie and J. J. Strain, The ferric reducing ability of plasma (FRAP) as a measure of “antioxidant power”: the FRAP assay, *Anal. Biochem.*, 1996, **239**(1), 70–76.
- 12 C. Sonklin, N. Laohakunjit and O. Kerdchoechuen, Assessment of antioxidant properties of membrane ultrafiltration peptides from mungbean meal protein hydrolysates, *PeerJ*, 2018, **6**, e5337.
- 13 O. P. Sharma and T. K. Bhat, DPPH antioxidant assay revisited, *Food Chem.*, 2009, **113**(4), 1202–1205.
- 14 M. S. Blois, Antioxidant determinations by the use of a stable free radical, *Nature*, 1958, **181**, 1199–1200.
- 15 P. Manual, *OxiSelect™ Oxygen Radical Antioxidant Capacity (ORAC) Activity Assay*, Cell Biolabs, Inc.
- 16 R. Chapman and K. Mackay, The estimation of peroxides in fats and oils by the ferric thiocyanate method, *J. Am. Oil Chem. Soc.*, 1949, **26**(7), 360–363.
- 17 P. Minkiewicz, J. Dziuba, A. Iwaniak, M. Dziuba and M. Darewicz, BIOPEP database and other programs for processing bioactive peptide sequences, *J. AOAC Int.*, 2008, **91**(4), 965–980.
- 18 K. Saito, D.-H. Jin, T. Ogawa, K. Muramoto, E. Hatakeyama, T. Yasuhara, *et al.*, Antioxidative properties of tripeptide libraries prepared by the combinatorial chemistry, *J. Agric. Food Chem.*, 2003, **51**(12), 3668–3674.
- 19 M. Tian, B. Fang, L. Jiang, H. Guo, J. Cui and F. Ren, Structure-activity relationship of a series of antioxidant tripeptides derived from β -Lactoglobulin using QSAR modeling, *Dairy Sci. Technol.*, 2015, **95**, 451–463.
- 20 T. H. Olsen, B. Yesiltas, F. I. Marin, M. Pertseva, P. J. García-Moreno, S. Gregersen, *et al.*, AnOxPePred: using deep learning for the prediction of antioxidative properties of peptides, *Sci. Rep.*, 2020, **10**(1), 21471.
- 21 Y.-W. Li and B. Li, Characterization of structure-antioxidant activity relationship of peptides in free radical systems using QSAR models: key sequence positions and their amino acid properties, *J. Theor. Biol.*, 2013, **318**, 29–43.
- 22 H. Guo, Y. Wang, Q. He, Y. Zhang, Y. Hu, Y. Wang, *et al.*, In silico rational design and virtual screening of antioxidant



- tripeptides based on 3D-QSAR modeling, *J. Mol. Struct.*, 2019, **1193**, 223–230.
- 23 W. Yan, G. Lin, R. Zhang, Z. Liang and W. Wu, Studies on the bioactivities and molecular mechanism of antioxidant peptides by 3D-QSAR, in vitro evaluation and molecular dynamic simulations, *Food Funct.*, 2020, **11**(4), 3043–3052.
- 24 T. T. N. Tran, D. P. Tran, V. C. Nguyen, T. D. T. Tran, T. T. T. Bui and J. H. Bowie, Antioxidant activities of major tryptophyllin L peptides: a joint investigation of Gaussian-based 3D-QSAR and radical scavenging experiments, *J. Pept. Sci.*, 2021, **27**(4), e3295.
- 25 T. T. N. Tran, D. P. Tran, T. M. A. Nguyen, T. H. Tran, N. N. A. Phan, V. C. Nguyen, *et al.*, Virtual screening and rational design of antioxidant peptides based on tryptophyllin L structures isolated from the *Litoria rubella* frog, *J. Pept. Sci.*, 2022, **28**(4), e3380.
- 26 S. T. Steinborner, P. A. Wabnitz, R. J. Waugh, J. H. Bowie, C. Gao, M. J. Tyler, *et al.*, The structures of new peptides from the Australian red tree frog '*Litoria rubella*'. The skin peptide profile as a probe for the study of evolutionary trends of amphibians, *Aust. J. Chem.*, 1996, **49**(9), 955–963.
- 27 T. T. N. Tran, *Structural and mechanistic studies of post-translationally modified peptides and proteins*, School of Chemistry and Physics: The University of Adelaide, 2014.
- 28 Y.-W. Li, B. Li, J. He and P. Qian, Quantitative structure–activity relationship study of antioxidative peptide by using different sets of amino acids descriptors, *J. Mol. Struct.*, 2011, **998**(1–3), 53–61.
- 29 J. B. Ghasemi and H. Tavakoli, Improvement of the prediction power of the CoMFA and CoMSIA models on histamine H3 antagonists by different variable selection methods, *Sci. Pharm.*, 2012, **80**(3), 547–566.
- 30 B. Sepehri, Z. Rasouli, Z. Hassanzadeh and R. Ghavami, Molecular docking and QSAR analysis of naphthyridone derivatives as ATAD2 bromodomain inhibitors: application of CoMFA, LS-SVM, and RBF neural network, *Med. Chem. Res.*, 2016, **25**, 2895–2905.
- 31 H. Ghafouri, M. Ranjbar and A. Sakhteman, 3D-QSAR studies of some reversible Acetyl cholinesterase inhibitors based on CoMFA and ligand protein interaction fingerprints using PC-LS-SVM and PLS-LS-SVM, *Comput. Biol. Chem.*, 2017, **69**, 19–27.
- 32 G. Floresta and V. Abbate, Machine learning vs. field 3D-QSAR models for serotonin 2A receptor psychoactive substances identification, *RSC Adv.*, 2021, **11**(24), 14587–14595.
- 33 F. Pedregosa, G. Varoquaux, A. Gramfort, V. Michel, B. Thirion, O. Grisel, *et al.*, Scikit-learn: Machine learning in Python, *J. Mach. Learn. Res.*, 2011, **12**, 2825–2830.
- 34 T. Chen and C. Guestrin, Xgboost: A scalable tree boosting system, *Proceedings of the 22nd ACM SIGKDD International Conference on Knowledge Discovery and Data Mining*, 2016.
- 35 S. G. Kwak and J. H. Kim, Central limit theorem: the cornerstone of modern statistics, *Korean J. Anesthesiol.*, 2017, **70**(2), 144–156.
- 36 V. Consonni, D. Ballabio and R. Todeschini, Comments on the definition of the Q₂ parameter for QSAR validation, *J. Chem. Inf. Model.*, 2009, **49**(7), 1669–1678.
- 37 R. Veerasamy, H. Rajak, A. Jain, S. Sivadasan, C. P. Varghese and R. K. Agrawal, Validation of QSAR models-strategies and importance, *Int. J. Drug Des. Discovery*, 2011, **3**, 511–519.
- 38 P. Gramatica, On the development and validation of QSAR models, *Comput. Toxicol.*, 2013, 499–526.

