



Cite this: *RSC Adv.*, 2023, 13, 26102

$\text{Ni}_3\text{V}_2\text{O}_8$ nanoflower mimetic enzyme constructs a dual-signal system for ascorbic acid detection

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Ascorbic acid is a nutritional small molecule essential to human life activities and health, playing a vital role in many physiological processes. Fresh fruits and beverages can provide abundant AA to maintain human metabolic balance. Therefore, it is of great significance to develop a nanomaterial with superior nanozyme activity for rapid and convenient detection of ascorbic acid (AA) in fruits and beverages. Herein, a dual-signal sensing platform based on UV-vis absorption and test strip chromaticity for the quantitative determination of AA is presented. The sensing platform is based on the horseradish peroxidase-like activity of $\text{Ni}_3\text{V}_2\text{O}_8$ nanoflowers, which catalyzes the oxidation of 3,3',5,5'-tetramethylbenzidine (TMB) by hydrogen peroxide to the blue oxide TMB (ox TMB). The ox TMB produced by the oxidation has a characteristic absorption peak at 650 nm. In the presence of AA, the blue ox TMB is reduced to colorless TMB, and the quantitative detection of AA can be achieved by detecting the decrease in intensity of the absorption peak by UV-Vis spectrophotometry. Under the optimal experimental conditions, the sensing platform exhibited excellent sensitivity and selectivity. A wide linear range of 0.1 μM to 40 μM with a detection limit of 0.032 μM was obtained. The linear equation is $\Delta A = 0.02513c + 0.1164$ with a correlation coefficient of 0.9979. It showed excellent properties in the detection of real samples of fruit juices and beverages, meanwhile, a method for the rapid detection of AA based on chromaticity change of test strips was constructed with high sensitivity and convenience. The linearity range for the ascorbic acid was 1–50 μM with LOD of 0.42 μM . The developed sensing platform has the capability to quickly and accurately detect ascorbic acid (AA) in fresh fruits and beverages. This proposed method offers a new and promising approach for the rapid and cost-effective detection of ascorbic acid, which has a wide range of potential applications.

Received 22nd June 2023
Accepted 24th August 2023

DOI: 10.1039/d3ra04202b

rsc.li/rsc-advances

Introduction

Ascorbic acid (AA), also known as vitamin C, is found in a wide variety of fruits and vegetables¹ and is one of the most abundant biological small molecules in the human body. In addition to its antioxidant properties, it is an important neuromodulator. It is an essential small molecule nutrient for human life and body health, and plays a crucial role in many physiological processes.² The World Health Organization recommends 45 mg of ascorbic acid per day or 300 mg per week. Normal levels of ascorbic acid in human blood and urine are approximately 2–14 mg L^{−1}. The human body cannot synthesize ascorbic acid so it must be supplemented by regular intake to help.^{3,4} Therefore, the analysis of ascorbic acid levels in foods is of great significance. Currently, the detection and analysis methods for AA mainly include: titrimetric analysis,⁵ spectrophotometric methods,^{6,7} high performance liquid chromatography (HPLC)^{8,9}

and ultra performance liquid chromatography (UPLC)¹⁰ and electrochemical methods.^{11,12} Although these methods are highly sensitive and reliable for the accurate determination of AA, the expensive instruments, specialized operators, and complicated pre-treatment processes have severely limited the widespread development of instrumental methods in practical applications. To overcome these limitations, it is essential to develop a simple, rapid and *in situ* technique for the sensitive and selective detection of AA.¹³ The colorimetric method has clear advantages in practical applications as it allows for an intuitive and rapid indication of the target analyte concentration through color reactions.¹⁴ The paper strip analysis method based on the colorimetric method has the advantages of lower test dose, convenient portability and simple operation, and can realize the rapid and efficient detection of target objects.¹⁵

Enzymes are substances that lower the activation energy of a reaction, thereby influencing the reaction rate. They play a crucial role in the functioning of living organisms. Natural enzymes are highly efficient and specific. However, they are susceptible to external factors that can lead to denature. In recent years, some nanomaterials with similar activity to natural enzymes have attracted attention. These nanomaterials exhibit

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greater stability than natural enzymes across a wide range of temperature and pH conditions. Nanomaterials that possess similar activity to natural enzymes are referred to as mimetic enzymes. These advantages such as lower cost, good stability, good catalytic effect, reproducibility, availability for further modification, larger surface area and better reaction rate make mimetic enzymes an emerging hot material in environmental chemistry and biomedicine, *etc.* Horseradish peroxidase (HRP) is capable of oxidizing a variety of reaction substrates in the presence of H_2O_2 . HRP is generally immobilized on the dielectric material to provide effective electron transfer between HRP and the substrate, which greatly increases the sensitivity of the biosensor while also reducing the detection limit. Researchers are now focusing their attention on mimicking horseradish peroxidase-mimicking enzymes, and a variety of HRP-mimicking materials have been synthesized for the detection of glucose,¹⁶ cysteine,¹⁷ alkaline phosphatase,¹⁸ and others. However, the activity of existing catalysts needs to be further improved, so the development of novel nanomimetic enzymes is crucial. Various metal oxides have been developed for the detection of organics.¹⁹ Current research focus on the development of new materials. Recently, various mixed metal oxides have been explored to synergistically enhance the peroxidase-like activity.^{20,21} Three-dimensional (3D) nanoflowers catalytic materials have been shown to be rich in active sites on a large surface area for radical adsorption, which greatly enhances the catalytic capacity.^{22,23} Yang developed a simple one-step hydrothermal method to synthesize $\text{Ni}_3\text{V}_2\text{O}_8$ nanoflowers, which exhibited excellent electrochemical performance as an anode material for Li-ion batteries (LIBs).²⁴ These results provide an impetus to develop a rapid and low-cost sensing platform by using the $\text{Ni}_3\text{V}_2\text{O}_8$ nanoflowers with excellent catalytic activity as the sensing probe. However, the preparation of 3D nanomimetic enzymes to serve as nanozyme for AA detection has not been reported much.

In this study, three-dimensional $\text{Ni}_3\text{V}_2\text{O}_8$ nanoflowers were synthesized. The materials possess horseradish peroxidase activity and can oxidize TMB in the presence of H_2O_2 . While ascorbic acid can reduce the blue oxidized TMB (ox TMB) to colorless TMB. Thus, quantitative detection of ascorbic acid can be achieved. Based on the peroxidase-like nature of the material, a UV-Vis spectrophotometer and a dual-channel colorimetric test trip-smartphone method were constructed for the detection of AA. This study provides a new strategy for the rapid and efficient detection of AA. The preparation process of $\text{Ni}_3\text{V}_2\text{O}_8$ and the principle of ascorbic acid detection are shown in Scheme 1.

Experimental section

Reagents and apparatus

Chemical reagents and drugs were used directly without purification. Nickel(II) nitrate hexahydrate ($\text{Ni}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$), orthoboric acid (H_3BO_3), calcium chloride (CaCl_2), cupric chloride (CuCl_2), magnesium chloride (MgCl_2), sodium chloride (NaCl), sodium citrate ($\text{C}_6\text{H}_5\text{Na}_3\text{O}_7$) and ethanol ($\text{C}_2\text{H}_5\text{OH}$) were purchased from Tianjin Fengchuan Chemical Reagent Co., Ltd (Tianjin, China). Chitosan was purchased from Sigma-Aldrich (USA). Diammonium 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonate) (ABTS), *o*-phenylenediamine (OPD), 3,3',5,5'-tetramethylbenzidine (TMB), ascorbic acid ($\text{C}_6\text{H}_8\text{O}_6$), lithium hydroxide (LiOH), ammonium metavanadate (NH_4VO_3) and poly(vinyl alcohol) (PVA) were obtained from Ron Reagent Ltd (Shanghai, China). Sodium phosphate, dibasic (Na_2HPO_4), zinc chloride (ZnCl_2), ferric chloride (FeCl_3) and sucrose ($\text{C}_{12}\text{H}_{22}\text{O}_{11}$) were bought from Xilong Scientific Co., Ltd (Guangdong, China). $\text{L}(+)\text{-Tartaric acid}$ ($\text{C}_4\text{H}_6\text{O}_6$) was purchased from Shanghai Yuanye Bio-Technology Co., Ltd (Shanghai, China). 2-Amino-2-phenylacetic acid ($\text{C}_8\text{H}_9\text{NO}_2$) was bought from



Scheme 1 (A) Schematic diagram of the preparation process of $\text{Ni}_3\text{V}_2\text{O}_8$ and (B) principle of ascorbic acid detection.



Shanghai Adamas Reagent Co., Ltd (Shanghai, China). L-Phenylalanine ($C_9H_{11}NO_2$), L-isoleucine ($C_9H_{13}NO_2$), glycine ($C_2H_5NO_2$), L-threonine ($C_4H_9NO_2$) and *N*-[(*tert*-butoxy) carbonyl]-D-tryptophan ($C_{12}H_{12}N_2O_2$) were obtained from Shanghai Aladdin Biochemical Technology Co., Ltd (Shanghai, China). The ascorbic acid ELISA kit was purchased from Jiangsu Feiya Biotechnology Co., Ltd (Yancheng, China).

Scanning electron microscopy (SEM) images were obtained using an FEI emission scanning electron microscope (SU8020, Hitachi, Japan). UV-vis absorption spectra were recorded using an UV 2700 spectrometer (Shimadzu, Japan). X-ray photoelectron spectroscopy (XPS) measurements were performed using a Escalab 250Xi diffractometer (Thermo Fisher Scientific, America). The diffraction of X-ray diffraction (XRD) spectra were collected on a D/MAX-TTRIII (Rigaku, Japan).

Synthesis of $Ni_3V_2O_8$ nanoflowers

$Ni_3V_2O_8$ was synthesized following the previous literature.²² As shown in Scheme 1 A, 5.25 mmol of NH_4VO_3 was dissolved in 80 mL of deionized water and heat the water was heated to 80 °C. Then, 1.05 mmol of $Ni(NO_3)_2 \cdot 6H_2O$ was added and stirred to the above solution. Next, the pH of the solution was adjusted to about 9 with LiOH and stirred for 10 min. This mixture was transferred to a stainless-steel autoclave with 100 mL-capacity Teflon liner. The autoclave was then sealed and kept in an oven at 150 °C for 5 h. Then, centrifugation was performed to collect the precipitates, following by washing three times with distilled water and absolute ethanol and then vacuum drying in an oven overnight to obtain $Ni_3V_2O_8$ nanoflowers.

Preparation of test strip

This kind of test strip was prepared by infiltrating the filter paper strips into the mixed solution containing $Ni_3V_2O_8$. Specifically, 1% polyvinyl alcohol (PVA) and 2% chitosan (CHI) were dissolved in 5 mL sodium acetate buffer solution and heated at 100 °C for 1 h. The solution was then slowly cooled to room temperature. Then, 1 mg mL⁻¹ $Ni_3V_2O_8$, 500 μL sodium acetate buffer and 500 μL TMB solution were added to the above solution. The model 102 filter paper was immersed in the solution for 2 hours. Next, filter paper was taken out and dried for 12 hours. The dried test paper was cut into 1 × 1 cm squares and stored in a cool, dark place.

The detection method of AA

As shown in Scheme 1B, based on the excellent horseradish peroxidase-like performance of $Ni_3V_2O_8$, AA was detected by UV-Vis absorbance spectra and test strip.

For the UV-Vis spectra detection method, ascorbic acid was detected under the optimized conditions. Briefly, 5 μL of 1 mg mL⁻¹ $Ni_3V_2O_8$, 10 μL of 10 mM TMB and 5 μL of 5 mM H_2O_2 were added to 160 μL of Na_2HPO_4 - $C_6H_8O_7$ buffer solution (pH 4.0) to react in dark for 10 min. The color of TMB changed to blue. Then, after adding various amounts of AA to the system, the absorbance was measured at 652 nm.

For the test strip detection method, prior to the ascorbic acid measurements, the obtained $Ni_3V_2O_8$ paper strips were cut into

1 × 1 cm squares and placed in the white PVC background plate. Different concentrations of AA solution were then dropped onto the small squares. The rapid color change of the small squares can be observed with the naked eye under ambient conditions, and the color photos were scanned by a smartphone equipped with a Fawn eyedropper APP. In this way, the sum of R + G + B (RGB color model) for each specific color can be quickly calculated, and the concentration of AA can be quickly obtained by using the calculator in the smartphone.

Results and discussion

X-ray diffraction (XRD) characterization of $Ni_3V_2O_8$

In this experiment, $Ni_3V_2O_8$ nanomaterial was characterized by XRD. As shown in Fig. 1, the peaks in the standing state for the 2 θ values of 15.5, 35.4, 43.4, 57.5, 62.1 and 64.9 were recorded as the (020), (132), (133), (260), (262) and (420) planes, respectively, which were in good agreement with the card PDF #70-1394 (red vertical line) and ref. 24 confirming the formation of $Ni_3V_2O_8$. Further, according to ref. 25 and 26, the average crystallite size could be calculated using the Debye-Scherrer formula (eqn (1)) from the obtained crystallographic planes, and the calculated average crystalline size from eqn (1) was found to be 3.7 nm. In eqn (1), D corresponds to the crystallite size, the X-ray wavelength ($\lambda = 1.5406 \text{ \AA}$), β is the full width at half maximum half (FWHM) of the more intense peak, and the Bragg diffraction angle is represented by θ .

$$D = 0.9\lambda/(\beta \cos \theta) \quad (1)$$

Microstructure characterization of $Ni_3V_2O_8$

The morphology of $Ni_3V_2O_8$ was characterized by scanning electron microscopy (Fig. 2). The SEM images showed that $Ni_3V_2O_8$ forms a large number of nanoflowers structures. The nanoflower structure of $Ni_3V_2O_8$ with about 100 nm diameter were shown on Fig. 2. The large surface area of the 3D

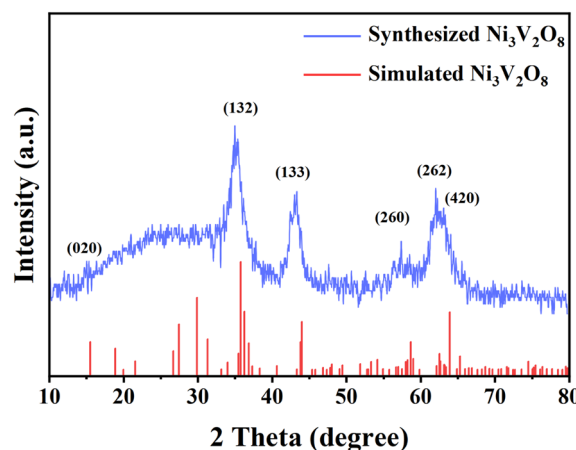


Fig. 1 XRD pattern of $Ni_3V_2O_8$ (red: simulated,²⁴ blue: synthesized in this experiment).



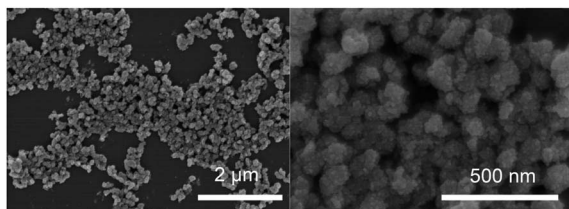


Fig. 2 Scanning electron microscope (SEM) images of $\text{Ni}_3\text{V}_2\text{O}_8$ with different magnifications.

nanoflowers has abundant free radical adsorption active sites, which provides the structural basis for the high catalytic capacity of the materials.

X-ray photoelectron spectroscopy (XPS) of $\text{Ni}_3\text{V}_2\text{O}_8$

In this experiment, the XPS of $\text{Ni}_3\text{V}_2\text{O}_8$ was further determined. As shown in Fig. 3A, the presence of Ni, V, and O elements in $\text{Ni}_3\text{V}_2\text{O}_8$ is evident. The peaks of nickel ions $\text{Ni } 2p_{1/2}$ and $\text{Ni } 2p_{3/2}$ were observed in the XPS spectrum of Ni 2p (Fig. 3B). The partial peak fitting of the Ni $2p_{1/2}$ and Ni $2p_{3/2}$ regions revealed that they contain Ni^{2+} (855.7 eV and 873.4 eV) and Ni^{3+} (857.1 eV and 875.1 eV). For the V 2p spectrum (Fig. 3C), it is also possible to fit to two chemical states of element V, V^{4+} (515.7 eV and 523.2 eV) and V^{5+} (516.8 eV and 524.3 eV), respectively. The mixed valence state of Ni and V may promote more redox reactions in the process. In addition, the C 1s spectrum (Fig. 3D) was split into four peaks, C=C (284.8 eV), C-O (285.8 eV), C=O (288.1 eV) and O=C-O (289.1 eV), respectively.

Steady-state kinetics evaluation and peroxidase-like activity analysis of $\text{Ni}_3\text{V}_2\text{O}_8$

The peroxidase-like activity of $\text{Ni}_3\text{V}_2\text{O}_8$ has been investigated. In the presence of H_2O_2 , common peroxidase chromogenic substrates, such as ABTS, OPD, and TMB, can be catalytically oxidized by $\text{Ni}_3\text{V}_2\text{O}_8$ to form blue-green, orange, and blue

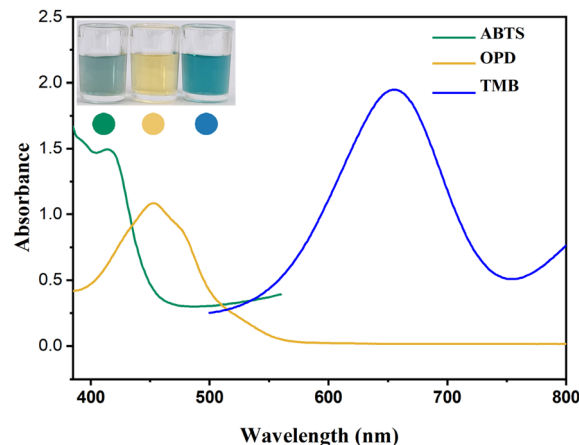


Fig. 4 Absorption curves of $\text{Ni}_3\text{V}_2\text{O}_8$ mimetic enzyme for different chromogenic substrates.

products, respectively. The products had characteristic absorption peaks at 420, 450, and 652 nm in the UV-Vis spectrum (Fig. 4), indicating that $\text{Ni}_3\text{V}_2\text{O}_8$ has peroxidase-like catalytic activity. In addition, TMB and OPD can produce more perfect absorption peaks. However, OPD is highly toxic, which is not conducive to laboratory safety, while TMB has higher sensitivity, better color product purity, reasonable stability and lower carcinogenicity. Therefore, TMB was chosen as the chromogenic substrate for mimetic enzyme $\text{Ni}_3\text{V}_2\text{O}_8$ in this work.

For the steady-state dynamics of TMB, the concentration of H_2O_2 is fixed (0.25 mM) and the concentration of TMB is varied from 0.015 mM to 0.3 mM. Similarly, for the steady-state dynamics of H_2O_2 , the concentration of TMB is fixed (0.5 mM) and the concentration of H_2O_2 is varied from 0.01 mM to 0.25 mM. The UV-vis absorption curves of various concentrations of TMB and H_2O_2 are shown in Fig. 5A and D. Fig. 5B and E shows the standard Michaelis–Menten curves for TMB and H_2O_2 , which were obtained by plotting a graph between the initial reaction rate *versus* concentration. For greater accuracy, the Lineweaver–Burk plot (Fig. 5C and F) can be used to calculate the Michaelis–Menten constant (K_m) and the maximal reaction velocity (V_{max}), and the results are presented in Table 1. K_m is often used as a measure of affinity of an enzyme to its substrate. The higher the affinity between the catalysts and the substrate, the lower the K_m . Table 1 illustrates that $\text{Ni}_3\text{V}_2\text{O}_8$ has a substantially lower K_m value with TMB and H_2O_2 than other mimetic enzymes and the natural enzyme HRP, indicating that $\text{Ni}_3\text{V}_2\text{O}_8$ has a much better affinity for two substrates than other mimetic enzymes and HRP.

Experimental feasibility analysis

To verify the feasibility of the experimental design, the catalytic oxidation capacity to TMB of $\text{Ni}_3\text{V}_2\text{O}_8$ in the presence of H_2O_2 was investigated (Fig. 6). When only TMB and H_2O_2 were added, the absorbance of the system at 652 nm was 0.048, which can be ignored. When only TMB and $\text{Ni}_3\text{V}_2\text{O}_8$ were added, the absorbance of the system at 652 nm was 0.366. When TMB, $\text{Ni}_3\text{V}_2\text{O}_8$

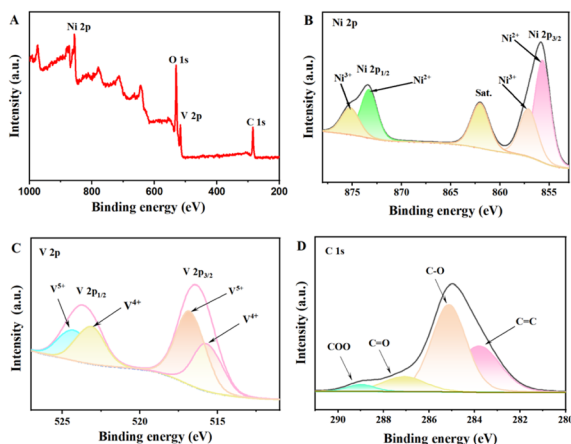


Fig. 3 XPS spectrum of $\text{Ni}_3\text{V}_2\text{O}_8$: (A) full spectrum; high resolution spectrum of Ni 2p (B), V 2p (C) and C 1s (D).



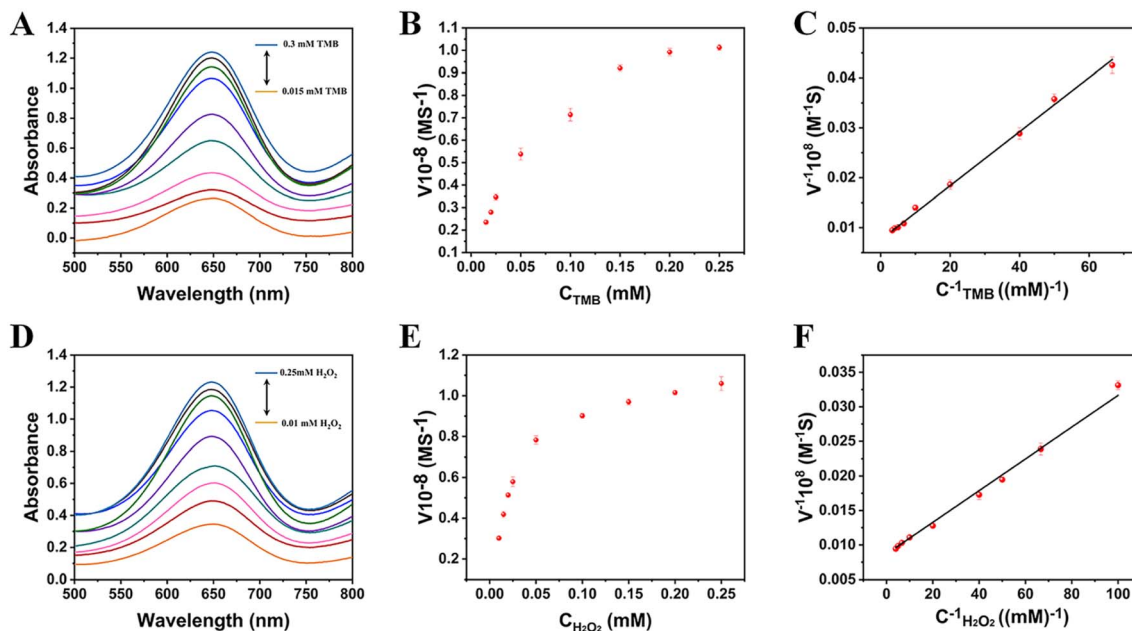


Fig. 5 Steady-state kinetic analysis of $\text{Ni}_3\text{V}_2\text{O}_8$ (the concentrations of TMB varied from 0.015 to 0.3 mM with a fixed amount of H_2O_2 (0.25 mM) (A–C) and the concentrations of H_2O_2 varied from 0.01 to 0.25 mM with a fixed amount of TMB (0.5 mM) (D–F). (C) and (F) is corresponding double reciprocal curves for TMB and H_2O_2 . Error bars represent the standard deviation for three measurements).

and H_2O_2 were added, the absorbance of the system at 652 nm was 1.280, indicating that $\text{Ni}_3\text{V}_2\text{O}_8$ material has good properties of horseradish peroxidase simulating enzyme. After adding TMB and $\text{Ni}_3\text{V}_2\text{O}_8$ materials for color reaction with H_2O_2 for 10 min, the absorbance of the sensing platform at 652 nm with AA is only 0.80, indicating that AA can reduce ox TMB. The higher the concentration of AA, the more TMB has been reduced, the lower the absorbance of the system. Therefore, the concentration of AA can be measured indirectly. This analysis shows that the proposed scheme is reasonable and feasible.

Optimization of the experimental conditions

The effect of the concentration of $\text{Ni}_3\text{V}_2\text{O}_8$, buffer type, pH and the reaction time on the catalytic activity of $\text{Ni}_3\text{V}_2\text{O}_8$ were investigated. The influence of different concentrations of $\text{Ni}_3\text{V}_2\text{O}_8$ on the difference of absorbance values is shown in Fig. 7A, with the increase of $\text{Ni}_3\text{V}_2\text{O}_8$ concentration, the difference of absorbance ΔA ($\Delta A = A_x - A_0$, where A_x and A_0 are the absorbance of the system at 652 nm in the presence and

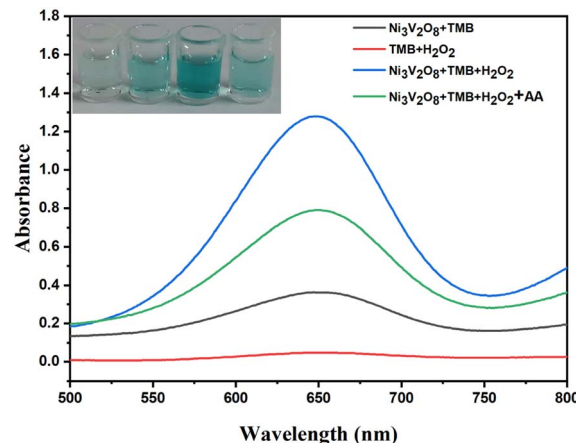


Fig. 6 Experimental feasibility analysis.

Table 1 Comparison of kinetic parameters of different nanozymes

Catalyst	K_m (TMB), mM	V_{\max} (TMB) (10^{-8} M s^{-1})	K_m (H_2O_2), mM	V_{\max} (H_2O_2) (10^{-8} M s^{-1})	Reference
$\text{Fe}_3\text{O}_4@\text{C}/\text{Ni NW}$	0.10	3.90	0.059	3.62	27
Pd nanostructures	0.16	201	1064	443	28
$\alpha\text{-Fe}_2\text{O}_3/\text{Fe}_3\text{O}_4$	0.23	5.52	2.85	3.87	29
Co_9S_8	1.64	99.0	7.39	35.0	30
Por- NiCo_2S_4	0.3	34.86	4.5	4.32	31
HRP	0.434	10.0	3.7	8.71	32
$\text{Ni}_3\text{V}_2\text{O}_8$	0.0714	131.91	0.0263	115.00	This work



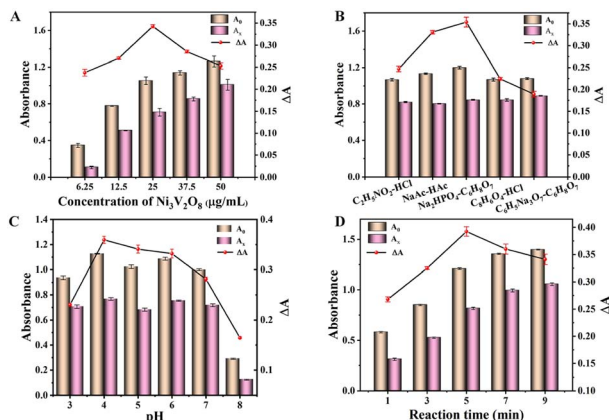


Fig. 7 Influences of (A) concentration of $\text{Ni}_3\text{V}_2\text{O}_8$ (6.25–50 $\mu\text{g mL}^{-1}$), (B) buffer of the reaction system, (C) pH of the reaction system (3–8), and (D) time (1–9 min) for absorbance values.

absence of AA respectively) increases. When the mass concentration of $\text{Ni}_3\text{V}_2\text{O}_8$ reached 25 $\mu\text{g mL}^{-1}$, the activity of the peroxidase-like enzyme reached the highest level. This finding could be mainly due to the limitation of dissolved oxygen and hydrogen peroxide concentration in the system. The catalytic activity of $\text{Ni}_3\text{V}_2\text{O}_8$ in $\text{C}_2\text{H}_5\text{NO}_2\text{-HCl}$, $\text{C}_6\text{H}_5\text{Na}_3\text{O}_7\text{-C}_6\text{H}_8\text{O}_5$, NaAc-HAc , $\text{Na}_2\text{HPO}_4\text{-C}_6\text{H}_8\text{O}_7$, $\text{C}_8\text{H}_6\text{O}_4\text{-HCl}$ buffer solution were also investigated (Fig. 7B). Among the five buffers, the value of ΔA was the highest in the $\text{Na}_2\text{HPO}_4\text{-C}_6\text{H}_8\text{O}_7$ buffer. Therefore, in this experiment, disodium phosphate citrate buffer was selected as the best buffer for the sensing platform. Catalytic studies in buffer solutions were studied in the pH range of 3.0–8.0 to find the optimum pH. As shown in Fig. 7C, the catalytic activity increased initially as the pH rises from 3.0 to 4.0, but then decreased as the pH increased further. This can be attributed to the decomposition of H_2O_2 at a high pH,³³ resulting in a reduction in its oxidation activity towards TMB. Therefore, pH 4.0 was chosen as the optimal pH of the buffer solution. The effect of reaction times on the difference of absorbance values was shown in Fig. 7D. It can be seen that with the increase of the color development reaction time, the difference of the absorbance values showed a trend of first increasing and then slowly decreasing, and the difference of the absorbance values reached the maximum at 5 min. Therefore, 5 min was selected as the optimal color development reaction time. It can be concluded that $\text{Ni}_3\text{V}_2\text{O}_8$ had high catalytic activity under the optimum conditions of $\text{Ni}_3\text{V}_2\text{O}_8$ 25 $\mu\text{g mL}^{-1}$, $\text{Na}_2\text{HPO}_4\text{-C}_6\text{H}_8\text{O}_7$ buffer pH 4.0, and 5 min of reaction time.

Calibration curve of the UV-vis sensing platform for AA

The absorbance of the system was determined under optimum conditions with AA in the concentrations range of 0.0–40 μM . As shown in Fig. 8A, the absorbance intensity reduced at 652 nm decreased with increasing AA. A linear relationship was established between the absorbance difference and the concentration of AA in the range of 0.1–40 μM , with the linear equation of $Y = 0.02513X + 0.1164$, with a linear correlation coefficient of

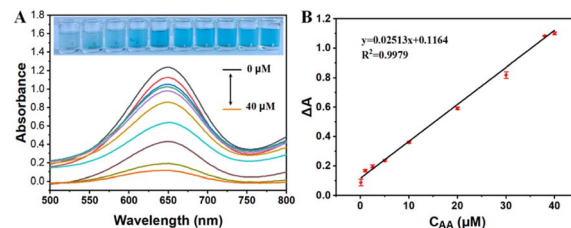


Fig. 8 The UV-vis absorption curves of different concentrations of AA (A) and calibration curve (B).

0.9979, as shown in Fig. 8B. The LOD of AA was calculated to be 0.032 μM at 3σ .

Calibration curve of test strip for AA

As shown in Fig. 9A, the $\text{Ni}_3\text{V}_2\text{O}_8$ test strips were sensitive response to the solutions containing different concentrations of AA. The R + G + B value of each color was then calculated and correlated with the concentrations of AA (Fig. 9B). As a result, a calibration curve of R + G + B was obtained in the range of AA concentrations from 1 to 50 μM , with the linear equation of $Y = 3.872X + 397.3$ with a linear correlation coefficient of 0.9908. The LOD for AA was calculated to be 0.42 μM at 3σ . Thus, in combination with a smartphone, portable, economical, and visual test strips have been developed for the detection of AA *in situ*.

Furthermore, as shown in Table 2, compared to other reported AA detection systems, the constructed sensing platform has a wide linear range and a lower detection limit. This may be attributed to the good catalytic effect of the $\text{Ni}_3\text{V}_2\text{O}_8$ material. Therefore, the established colorimetric sensor has the potential for practical sample analysis.

The selectivity of the sensing platform

A range of biologically disruptive chemicals including a variety of amino acid molecules (L-glycine, L-threonine, tryptophan, glutamate, L-phenylalanine, L-phenylglycine, L-isoleucine,

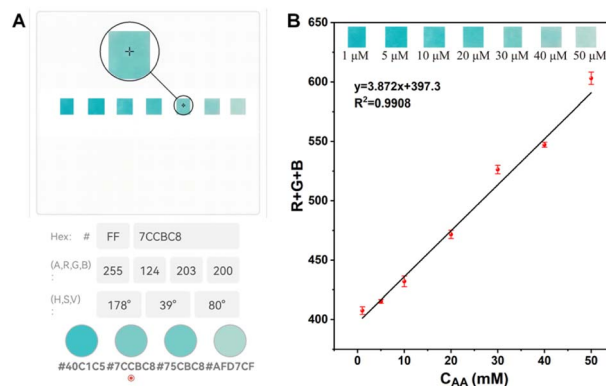


Fig. 9 (A) Schematic diagram of recognition RGB by Fawn eyedropper; (B) RGB calibration curve of the test strip.

Analytical method	Material	Linear range	LOD	Reference
Electrochemistry	CaCO ₃	12–500 μM	10.8 μM	34
Electrochemistry	PPYNTs/Co ₃ O ₄	5–80 μM	0.23 μM	35
Fluorometry	LDH-GQD	5–300 μM	1.73 μM	36
Fluorometry	SGQDs-MnO ₂	0.5–20 μM	0.16 μM	37
Colorimetry	Fe–Mn	8–56 μM	0.88 μM	38
Colorimetry	MIL-88	2.57–10.10 μM	1.03 μM	39
Colorimetry	Mn@Co ₃ O ₄ NSs	1–8 μM	0.4 μM	40
Colorimetry	Ni ₃ V ₂ O ₈	0.1–40 μM	0.03182 μM	This work
RGB	Ni ₃ V ₂ O ₈	1–50 μM	0.42 μM	This work



The results of commercial enzyme immunoassay kits and $\text{Ni}_3\text{V}_2\text{O}_8$ sensing platform for the determination of AA in tablets and human serum were compared. For the kit, the UV-vis absorbance increased gradually increased of the target AA concentration, and showed a good linear relationship in the range of 2.5–40 μM with the linear equation of $Y = 0.04332X + 0.4552$ with a linear correlation coefficient of 0.9902, as shown in Fig. 12B. Subsequently, the sensing platform constructed by this work and ELISA kit were used to detect the absorbance of different concentrations of tablets respectively. Each group was tested three times in parallel, and the AA concentration was calculated according to the calibration linearity curve of the system. The results are shown in Table 3. These results indicated that the proposed method for the determination of AA is accurate and can be applied to the assay of real samples.

Practical application is one of the important standards for investigating the application value of sensors. To confirm the feasibility of the sensing platform for practical applications, the sensor was used to detect the AA content in fresh fruit juices and drinks. Five kinds of juice and four kinds of drinks were selected for the experiment: cherry tomatoes, strawberry, orange, longan, kiwi, vita lemonade, green tea, kumquat and yiquan lemonade. In the experiment, all fruit juices and drinks were diluted 10 times for testing. As shown in Fig. 13, the highest AA content in the fruit was kiwi, while in the drinks was vita lemonade. The results show that the sensing platform can be used to detect real samples and has good practical application value.

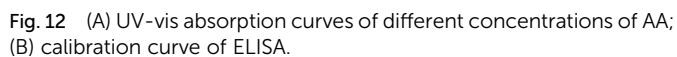
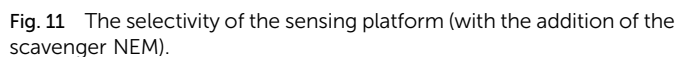


Table 3 Comparison of different detection methods for actual samples ($n = 3$)^a

Real sample		VC tablet			Human serum		
AA ELISA KITS	Found (μM)	18.35	18.39	18.42	7.38	7.29	7.25
	Average (μM)		18.39			7.31	
	RSD (%)		0.19			0.97	
U-V	Found (μM)	18.86	18.95	18.91	7.27	7.35	7.23
	Average (μM)		18.91			7.28	
	RSD (%)		0.22			0.86	
	Relative error (%)		2.83			−0.41	
RGB	Found (μM)	19.03	18.78	18.26	7.67	8.19	7.41
	Average (μM)		18.69			7.76	
	RSD (%)		2.11			5.08	
	Relative error (%)		1.63			6.12	

^a Relative error (%) = $(X - Y)/Y$, X is the experimental measurement value, Y is the ELISA measurement value.

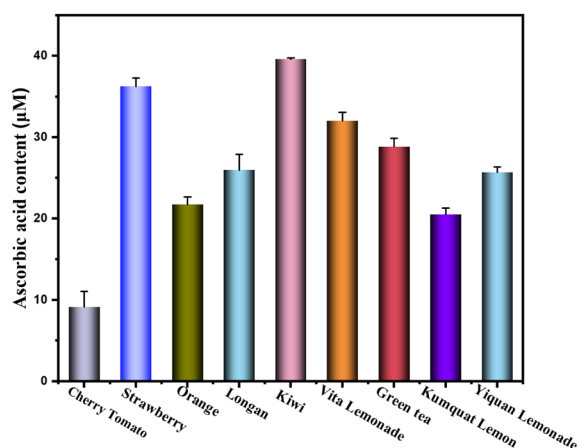


Fig. 13 Determination of the actual samples.

Conclusions

In this work, $\text{Ni}_3\text{V}_2\text{O}_8$ nanoflowers mimetic enzyme was synthesized, and a sensing platform was constructed for the detection of ascorbic acid (AA) based on the dual signal of UV-vis absorption and chromaticity change of test strips. For the UV-vis spectra detection method, in the range of 0.1–40 μM , the absorbance difference (ΔA) shows a good linear relationship with the AA concentration, and the detection limit is 0.03182 μM . And the constructed test strip assay has the advantage of easy detection of real samples with a lower detection limit of detection of 0.42 μM . In addition, the $\text{Ni}_3\text{V}_2\text{O}_8$ nanoflowers show an excellent selectivity for 22 different interferences including ions, small molecules, and amino acids commonly found in real samples. Furthermore, the sensing platform was successfully applied to determine of AA content in five fruit juices and four commercial beverages. The proposed detection method has good selectivity and high sensitivity, which can be used for the rapid and low-cost detection of AA in the field. Further, from the results of this study, the sensing platforms especial test strip can be a prominent tool for determining the AA levels in fruit juices and beverages. The work reported in this paper represents only preliminary information, and we are

continuing to work on these models in order to bring our sensor based on $\text{Ni}_3\text{V}_2\text{O}_8$ for real-time detection of AA into commercial use. We are currently working on further optimizing this sensing platforms to make it cost-effective, robust and more accurate.

Conflicts of interest

The authors declare there are no conflicts of interest regarding the publication of this paper.

Acknowledgements

This work was supported by the National Natural Science Foundation of China (Grant No. 21765026).

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