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Enhancing solubility and stability of sorafenib through cyclodextrin-based inclusion complexation: *in silico* and *in vitro* studies†

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Sorafenib (SOR) is an oral multikinase inhibitor that effectively hampers the growth and spread of cancer cells by targeting angiogenesis and proliferation. However, SOR tablets (Nexavar) have limited oral bioavailability, ranging from 38% to 49%, due to their low water solubility. To address this issue, cyclodextrins (CDs), widely used to enhance the solubility and stability of lipophilic drugs by encapsulating them within their molecular structure, were considered in this study. We focused on β -cyclodextrin (β CD) and its derivatives, including hydroxypropyl- β -cyclodextrin (HP β CD), dimethyl- β -cyclodextrin (DM β CD), sulfobutylether- β -cyclodextrin (SBE β CD), and compared them with γ -cyclodextrin (γ CD) for generating inclusion complexes with SOR. The 200 ns molecular dynamics simulations revealed that SOR could form inclusion complexes with all CDs in two possible orientations: pyridine group insertion (P-form) and chlorobenzotrifluoride group insertion (C-form), primarily driven by van der Waals interactions. Among the four β CD derivatives studied, SOR exhibited the highest number of atom contacts with SBE β CD and demonstrated the lowest solvent accessibility within the hydrophobic cavity of SBE β CD. These findings correlated with the highest binding affinity of SOR/SBE β CD complex determined by SIE, MM/GBSA, and MM/PBSA methods. Experimental results further supported our computational predictions, in which SBE β CD exhibited a stability constant of 940 M⁻¹ at 25 °C, surpassing β CD's stability constant of 210 M⁻¹. Taken together, our results suggest that the modified CDs, particularly SBE β CD, hold promising potential as an efficient molecular encapsulating agent for SOR, offering improved solubility and stability for this lipophilic drug.

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1. Introduction

Sorafenib (SOR), also known as 4-(4-(3-(4-chloro-3-(trifluoromethyl)phenyl)ureido)phenoxy)-*N*-methylpicolinamide, is currently the only FDA-approved treatment for hepatocellular carcinoma (HCC). It functions as an oral multikinase inhibitor by targeting several kinases and growth factors, including Raf kinases, vascular endothelial growth factor receptor (VEGFR), Fms-like tyrosine kinase 3 (FLT-3), and c-Kit.^{1,2} Its mechanism of action involves inhibiting angiogenesis, suppressing cell proliferation, and impeding the spread of cancer cells, making it effective against various tumor types.^{3,4} Despite its efficacy,

sorafenib has poor water solubility,⁵ which decreases its bioavailability (38–49%) when administered in tablet form, compared to an oral solution.⁶ This limited solubility, combined with its high cytotoxicity to normal tissues, can lead to suboptimal treatment outcomes for many cancer types.⁷ Additionally, the use of sorafenib may give rise to various adverse effects, including dry skin, itching, acne, diarrhoea, hair loss, and vomiting.⁸

Various strategies have been explored to enhance the solubility of poorly water-soluble drugs and substances, including the use of co-solvents, cyclodextrins (CDs) complexation, and nano-engineered delivery systems.^{9–10} Among these approaches, the encapsulation of hydrophobic molecules using CDs has gained significant attention due to its cost-effectiveness, high drug loading capacity, improved bioavailability, commercial availability, favorable pharmacochemical properties, enhanced dissolution rate, and potential to improve the stability of lipophilic compounds, along with excellent biocompatibility.^{11,12} CDs are cyclic oligosaccharides naturally derived from starch through enzymatic breakdown.¹³ They consist of α -D-glucopyranose units connected by α -1,4 glycosidic bonds and possess a cone-shaped structure with a hydrophilic outer surface and a hydrophobic

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inner cavity. The most commonly encountered natural CDs are α CD, β CD, and γ CD, which contain 6, 7, and 8 α -D-glucopyranose units, respectively.¹⁴ CDs allow guest molecules to enter its nanocavity through van der Waals forces.¹⁵ β CD derivatives such as sulfobutylether- β CD (SBE β CD) and 2-hydroxypropyl- β CD (HP β CD) offer several advantages over natural β CD. They exhibit higher water solubility, form more substantial complexes with other molecules, and possess lower toxicity, making them more suitable for pharmaceutical applications.^{16–19} Modified forms of β CD, such as methylated β CD (M β CD) and HP β CD, are commonly employed for drug encapsulation due to their ability to form inclusion complexes and their significantly higher water solubility (>500 mg mL^{−1}) compared to β CD.²⁰ In addition, γ CD has been widely utilized in various industries due to its higher water solubility, larger internal cavity, and more bioavailability.^{21,22} Previous studies have demonstrated the significant enhancement of pharmacokinetic, biodistribution, and pharmaceutical properties of regorafenib through the use of mannose- γ CD (M γ CD).²³

Despite some existing research on the inclusion complexation of SOR with β CD and γ CD,²⁴ there is still a lack of knowledge regarding the specific structural details of these complexes. Furthermore, the inclusion complexation of SOR with CD derivatives remains poorly understood. Therefore, the main objective of this study is to enhance our understanding of the structural characteristics of inclusion complexes formed between SOR and six host molecules (β CD, DM β CD, HP β CD, SBE β CD, γ CD, and M γ CD). All-atom molecular dynamics (MD) simulations and free energy calculations have been employed to investigate the structural dynamics of inclusion complexes in an aqueous solution. Additionally, this work aims to identify the most effective host molecule among the six studied CDs for enhancing the water solubility of SOR. To complement the computational analysis, the solubility and stability of the inclusion complexes were also investigated experimentally.

2. Computational details

2.1. Preparation of 3D structures of SOR and CDs

The HF/6-31G* level of theory was used to optimize the 3D structure of the SOR using the Gaussian09 (ref. 25) program as per the standard protocol.^{26–28} The protonation state of SOR was characterized using MarvinSketch²⁹ at a pH of 7.0. It was discovered that HP β CD with high (7.76) or medium (6.16) degrees of substitution resulted in reduced solubility and increased nephrotoxicity, compared to HP β CD with a low degree of substitution (4.55).³⁰ HP β CD with four substitutions on the primary rim (O6) of β CD has been reported to exhibit less probability of cavity self-closure.³¹ Accordingly, this study utilized HP β CD with four HP substitutions on its primary rim. SBE β CD with a degree of substitution of 7 was found to be the most effective host for rasagiline among the different degrees of substitution of SBE β CD evaluated.³² The 3D structures of β CD, γ CD, and HP β CD were obtained from previous studies.^{31,33,34} The M γ CD with a single mannose substitution on the primary rim of γ CD was constructed, as reported previously.²³

2.2. Molecular docking study

The CDOCKER module in Accelrys Discovery Studio 2.5 (Accelrys Software Inc., San Diego, CA, USA) was used to generate the inclusion complex between SOR and CDs. The binding affinity of SOR towards CDs was evaluated by conducting a docking process using a 10 Å sphere. From the top 100 hits, their percentage of docked conformations (%DCs) was recorded, and the docked complexes with the lowest binding interaction energy were selected as the starting structures for further MD simulations.

2.3. Molecular dynamics simulations

The Glycam-06 (ref. 35) and the general AMBER force fields³⁶ were employed to simulate the behavior of CDs and SOR, respectively. Water molecules (TIP3P) were added to the model, with a 15 Å spacing distance, to solvate the SOR/CD complexes. The added water was minimized using a combination of 1000 steps of steepest descent and 3000 steps of conjugated gradient. The entire model underwent an overall minimization using the same methods. The simulations assumed periodic boundary conditions and a 2 fs time step. Initially, the complexes were heated from 10 K to 298 K for 100 ps, followed by three individual all-atom molecular dynamics simulations in the NPT ensemble, with a temperature of 298 K and pressure of 1 atm, utilizing the AMBER20 (ref. 37) software package. The SHAKE algorithm³⁸ was applied to constrain hydrogen-involved bonds, and the Particle Mesh Ewald³⁹ method was used with a cutoff of 12 Å to calculate charge–charge interactions. Three replicates of all-atom MD simulations (MD#1–3) were conducted for each system, with a duration of 200 ns. To assess the stability of the system, the root-mean-squared displacement (RMSD) was calculated, and the last 100 ns of simulation data was selected for further analyses. The preferred binding orientation of each complex was determined by analyzing the distances between the SOR components and CDs. The most representative structures of the inclusion complex were identified through RMSD clustering and the use of DBSCAN density-based clustering algorithms.⁴⁰ The number of contacts between SOR and CDs was determined using a cutoff value of 3.0 Å. The accessibility of water to SOR was analyzed by calculating the solvent-accessible surface area (SASA) of the complex. The potential energy surface (PES) was calculated to gain insight into the structure of CDs during the simulation. Binding affinity was assessed through solvated interaction energy (SIE),⁴¹ molecular mechanics/generalized Born surface area (MM/GBSA) and mechanics/Poisson–Boltzmann surface area (MM/PBSA) based binding free energy calculations.⁴²

3. Experimental studies

3.1. Materials

SOR was purchased from Sigma-Aldrich (St. Louis, MO, USA). β CD was obtained from FUJIFILM Wako Pure Chemicals Corporation, Osaka, Japan, while SBE7 β CD with a degree of substitution (DS) ranging from 6.0 to 7.1 was acquired from Medchem Express (Monmouth Junction, NJ, USA).



3.2. Phase solubility study

The previously reported methods by Higuchi and Connors⁴³ were used to conduct a phase solubility study to analyse the host–guest behaviors at 25 °C. An excess amount of SOR prepared in pure water was added to different concentrations of β CD and SBE β CD, ranging from 0 to 10 mM, followed by vortexing and sonication to ensure consistent mixing of components throughout. Note that the samples were unbuffered. It is worth noting that light protection was necessary during the preparation of SBE7 β CD and SOR solutions in water. The mixtures were incubated for 72 hours at 25 °C with constant shaking at 250 rpm. Following the incubation, the suspension was subjected to centrifugation at 12 000 rpm for 15 minutes. The resulting saturated supernatants were filtered through 0.45 μ m membrane filters to remove any undissolved SOR. The filtrate was then appropriately diluted with a 1 : 1 (v/v) mixture of ethanol and water. The solution's SOR concentration was then measured at 260 nm. The stability constant (K_c) for encapsulation was determined using the formula:

$$K_c = \frac{\text{slope}}{S_0(1 - \text{slope})}$$

where S_0 is the y-intercept indicating how much SOR can dissolve in water without any β CDs.⁴⁴ The experiment was performed three times independently, and the findings were reported as mean \pm standard error of the mean (SEM).

4. Results and discussion

4.1. Binding patterns of SOR inside the cavity of CDs

Based on 100 docking runs, it was discovered that SOR had two favorable positions within the hydrophobic interior of CDs. The insertion of the pyridine group of SOR into the CDs' cavity was referred to as the P-form, while chlorobenzotrifluoride group

insertion was named as C-form (Fig. 1c). The data in Fig. 2 shows %DCs of P-form and C-form which highlights that the C-form is more prevalent in all complexes compared to the P-form. This finding agrees well with the previously reported 2D NOESY spectra of SOR/ β CD and SOR/ γ CD, where SOR/ β CD displayed correlations between the –OH and H2 protons of β CD and the protons on the ureido group (C-form) of SOR.²⁴ In other reported study, in the most stable complex, the fluorine atoms are oriented towards the hydrophobic primary rim of amphiphilic cyclodextrin (aCD) while the oxygen-rich portion of the SOR faces the hydrophilic secondary rim.⁴⁵ C. Phan *et al.* also confirmed the encapsulation of SOR with CDs, where they recorded the ¹H-NMR spectrum which indicates signals in the region of 9.5–7.0 ppm for the SOR protons and the signals in the range of 5.5–3.0 ppm to the intrinsic proton peaks of SOR/ β CD and SOR/ γ CD.²⁴ It is noteworthy, that the signal intensity of the protons in SOR was smaller than that in CDs, which may be due to the modification or shielding in the cavities of CDs after complexation.²⁴ The interaction energies of both P-form (ranging from –28 to –37 kcal mol^{–1}) and C-form (ranging from –28 to –39 kcal mol^{–1}) are similar; thus, both orientations were selected as the starting structures for MD simulations.

4.2. System stability

The stability of the inclusion complexes was evaluated by calculating the all-atom RMSD (Fig. 3). Results showed that most complexes remained stable during the simulation, with fluctuations in RMSD ranging from 2–6 Å, while a few systems got stable after 100 ns. Previous study shows that RMSD maps of MD simulation ranging from 1 Å to 4 Å, indicating the majority of the complex formations.⁴⁵ Based on the fact that all inclusion complexes reach equilibrium after 100 ns, further analyses were conducted using the last 100 ns (100–200 ns) of MD simulations.

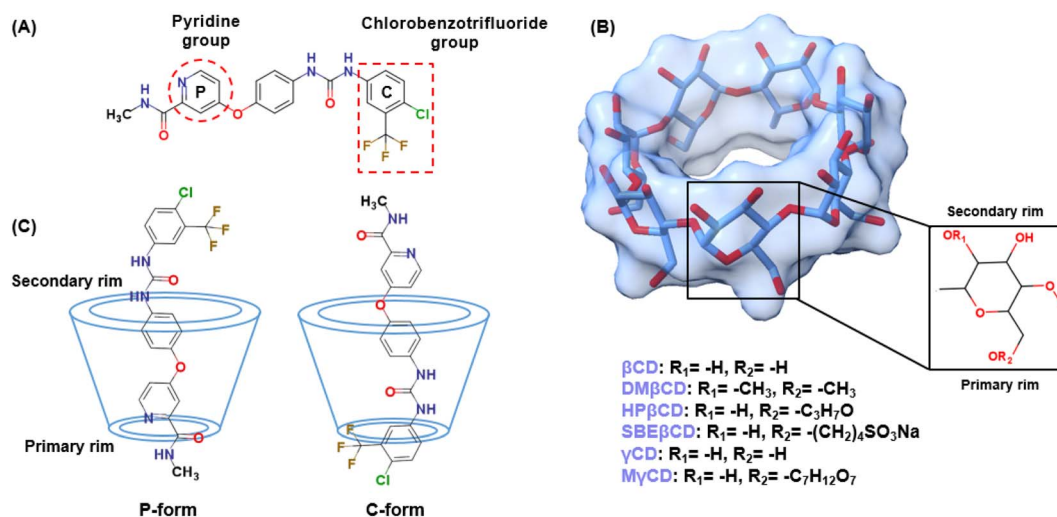


Fig. 1 (A) Chemical structure of SOR containing pyridine group and chlorobenzotrifluoride group. (B) 3D structure of all studied CDs. (C) The possible orientations of SOR in complex with CDs: pyridine group insertion (P-form) and chlorobenzotrifluoride group insertion (C-form).



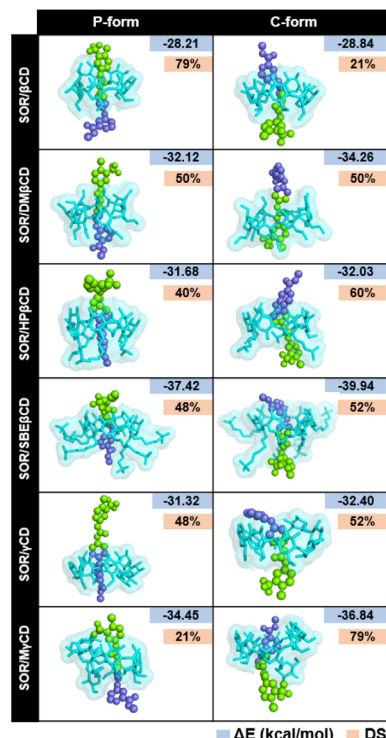


Fig. 2 %Docked conformations (%DCs) and CDOCKER interaction energy (ΔE) of all docked complexes.

4.3. SOR mobility inside the CD cavity

The dynamic behavior of the encapsulated SOR inside the hydrophobic interior of CDs was analyzed over time by calculating the distance between the center of mass of pyridine group, ring of chlorobenzotrifluoride group of SOR and the center of mass of the primary rim of CDs, ignoring the functional substituents (Fig. 4). In the case of SOR/βCD, SOR/γCD, and SOR/MγCD in both orientations (P-form and C-form), SOR moves towards the primary rim and secondary rim throughout the simulation because of the flexible pyridine group and the larger size of SOR. Considering SOR/SBEβCD (C-form) inclusion complex, SOR moves towards the narrow rim of SBEβCD and remains inside the side chains of SBEβCD for all three MD

simulations. All complexes were further confirmed by taking snapshots of each complex throughout the simulation, as illustrated in Fig. S1–S3 in ESI.†

To verify the results, the RMSD clustering was performed by combining the final 100 ns trajectories of all three MD simulations, based on 10 000 snapshots, depicted in Fig. 5. The results revealed that there were three groups of inclusion complexes, including cluster 1 (red) being the most prevalent for all studied systems, cluster 2 (green) was the second largest population, and cluster 3 (purple) was the lowest population. It was observed that the distribution of populations among the clusters was significantly distinct. In the case of SOR/βCD, SOR/γCD, and SOR/MγCD, all three clusters were found and the remaining complexes only showed two clusters. These outcomes justify the distance analysis results (Fig. 4), specifically in the case of SOR/SBEβCD (C-form) which apparently indicated to be oriented as P-form. Fig. 5 also clarifies that SOR was consistently located within the core of CDs in all complexes, indicating the formation of inclusion complexes.

4.4. Atomic contacts and solvent accessibility toward inclusion complex

To investigate the encapsulation of SOR by CDs through MD simulations, the number of atomic contacts between SOR and CDs using a cutoff of 3 Å distance was determined (Fig. 6). A high level of molecular contact indicates a favorable and close interaction between the host and guest molecule, while a low level of contact indicates a weaker interaction.⁴⁶ Fig. 6 highlights that the average atom contacts of the SOR/SBEβCD inclusion complex (C-form) were the highest (54.2 ± 9.1) among all studied complexes followed by C-form of SOR/HPβCD (50.0 ± 9.6). The second most atom contacts were observed in the P-form of SOR/SBEβCD (49.7 ± 9.8) followed by the P-form of SOR/DMβCD (49.3 ± 8.0). Table S1 in ESI† shows the interactions of SOR with CDs, which is in favor of the statement that SBEβCD could be the best encapsulating agent for SOR.

To validate these results, we further investigate the water accessibility towards SOR (Fig. 6). The idea is that when SOR creates an inclusion complex with CDs, the amount of water molecules surrounding the SOR decreases and the interaction

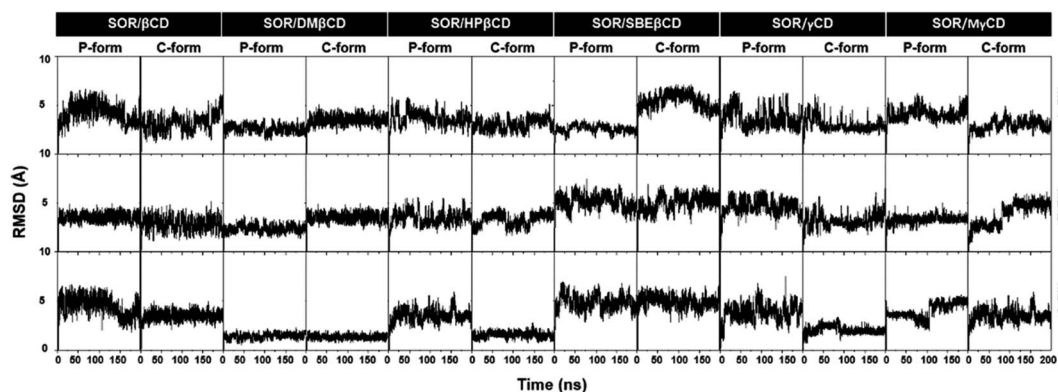


Fig. 3 All-atom RMSD profiles of all complexes in both P-form and C-form, plotted over 200 ns in three replicates (MD1–3).

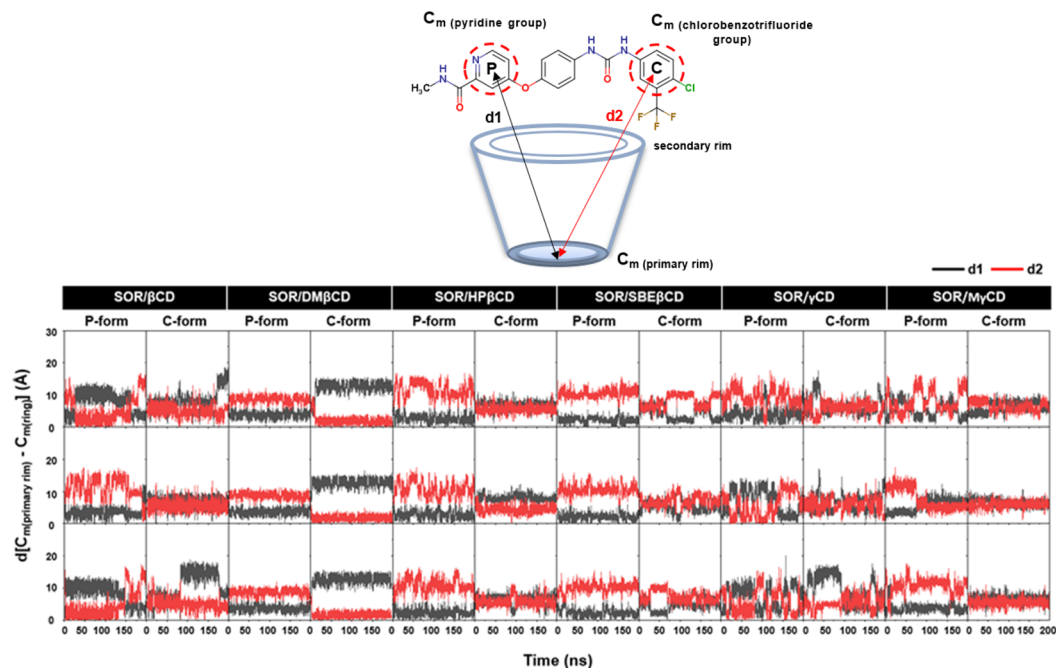


Fig. 4 Distance analysis of all inclusion complexes plotted over 200 ns, where $d[C_m(\text{primary rim}) - C_m(P)]$ (d_1) and $d[C_m(\text{primary rim}) - C_m(C)]$ (d_2) are represented in black and red, respectively.

between the host and guest molecule increases. The lowest SASA value of $250.4 \pm 25.3 \text{ \AA}^2$ was observed in C-form of SOR/SBE β CD inclusion complex followed by P-form of SOR/SBE β CD ($273.7 \pm 27.9 \text{ \AA}^2$). The second lowest SASA value was seen in the C-form of SOR/HP β CD ($282.2 \pm 19.4 \text{ \AA}^2$), followed by the P-form of SOR/HP β CD ($296.4 \pm 25.6 \text{ \AA}^2$). The highest SASA values were found in the P-form of SOR/ β CD ($338.9 \pm 17.8 \text{ \AA}^2$) and the C-form of SOR/ β CD ($342.3 \pm 21.2 \text{ \AA}^2$), which indicates that SOR/ β CD is more likely to interact with water molecules as compared to other complexes. Table S1 in ESI† clearly represent as the SASA values decreases, atom contacts between SOR and CDs increases.⁴⁷ In conclusion, low SASA values support that SBE β CD could be the best host molecule for SOR.

4.5. CDs conformations in relation to PES calculations

The evaluation of the structural distortions in CDs upon the binding of SOR was performed through PES analysis. This analysis was based on the measurement of two distances: (1) the distance between adjacent oxygen atoms at the wider rim $d_n[O_{3n} - O_{2n+1}]$, and (2) the distance between adjacent glucopyranose units $d_n[O_{4n} - O_{4n+1}]$. These distances were used to determine the strength of the intramolecular hydrogen bonds and the ellipticity of the CDs, respectively. Eqn (1) was applied to calculate probability distributions of these distances and get the value of free energy, $F(x,y)$:

$$F(x,y) = -K_B T \log[P(x,y)] \quad (1)$$

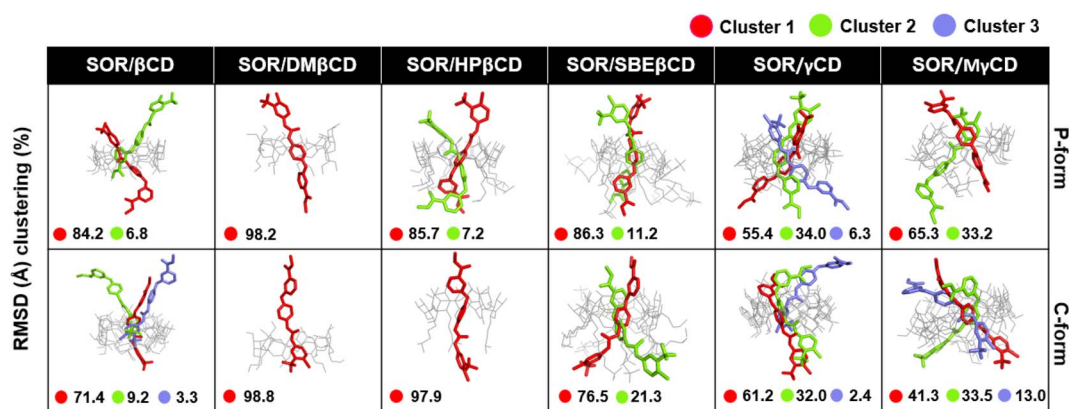


Fig. 5 Representative structures of host-guest inclusion of each RMSD cluster for both P-form and C-form of SOR/CDs inclusion complexes. Clusters population was labelled in a percentage.

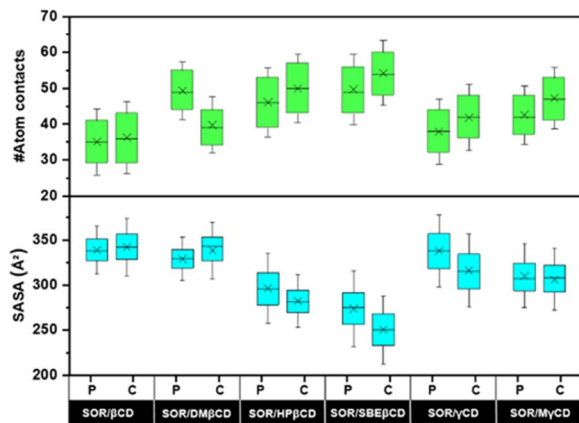


Fig. 6 The number of atomic interactions and SASA calculated over last 100 ns for all inclusion complexes. Green and cyan boxes cover the area between the 25th and 75th percentiles. Mean values are represented by a cross, while whiskers determine the standard deviation.

where k_B is the Boltzmann constant, T is the absolute temperature (298 K), and $P(x,y)$ is the probability of x for $d_n[O3_n - O2_{n+1}]$ and y for $d_n[O4_n - O4_{n+1}]$ distances.

In the PES study of complexes, two local minima (M1 and M2) were observed, as depicted in Fig. 7. The contour graphs display the probability values for distances, colors ranging from red to blue, and darker shades of blue indicating lower free energy level. The encapsulation process resulted in a stable conformation of CDs, which was demonstrated by the presence of a distinct M1 region in all analyzed complexes. This stability

was achieved due to the formation of hydrogen bonds between $O3_n$ and $O2_{n+1}$. The M2 region found in SOR/βCD, SOR/γCD, and SOR/MγCD in both P and C-form indicates the distortion of the glucopyranose structures but totally disappeared in the remaining complexes, indicating the most stable complexes. The Intensity of M2 region of SOR/γCD and SOR/MγCD is clearly increased when compared to SOR/βCD, which is also reported previously that the network of hydrogen bonds of $O3-O2$ in γCD is relatively less than βCD due to the distortion of glucopyranose structures in γCD.⁴⁸ The molecular encapsulation of SOR towards these CDs clearly indicates the formation of hydrogen bonds (higher proportion of $O3_{(n)} - O2_{(n+1)}$ with a distance of 2–3 Å) on the wider rim, which are in good agreement with the previous studies.^{49–51}

4.6. System compactness

To gain further insight into the compactness of the system, we calculated radius of gyration (R_g). The collapse in specific structures, such as polymers, proteins, and micelles formation, can be measured by means of R_g .⁵² The probability of the R_g of all complexes is shown in Fig. 8. In previous study, the average R_g value of DMβCD, HPβCD, and SBEβCD was 6.60–7.00, 6.85, and 8.03 Å, respectively.^{31,50,53} The average R_g value of SOR/βCD, SOR/DMβCD, SOR/HPβCD, SOR/SBEβCD, SOR/γCD, and SOR/MγCD was 6.40 ± 0.40 , 6.70 ± 0.70 , 6.61 ± 0.18 , 7.59 ± 0.17 , 7.01 ± 0.20 , and 7.00 ± 0.30 Å, respectively. The decline in R_g values observed in the studied complexes are due to the interaction between SOR and CDs, which results in a decreased flexibility of the CDs' conformation upon inclusion complex formation. R_g values of P-form and C-form were not significantly

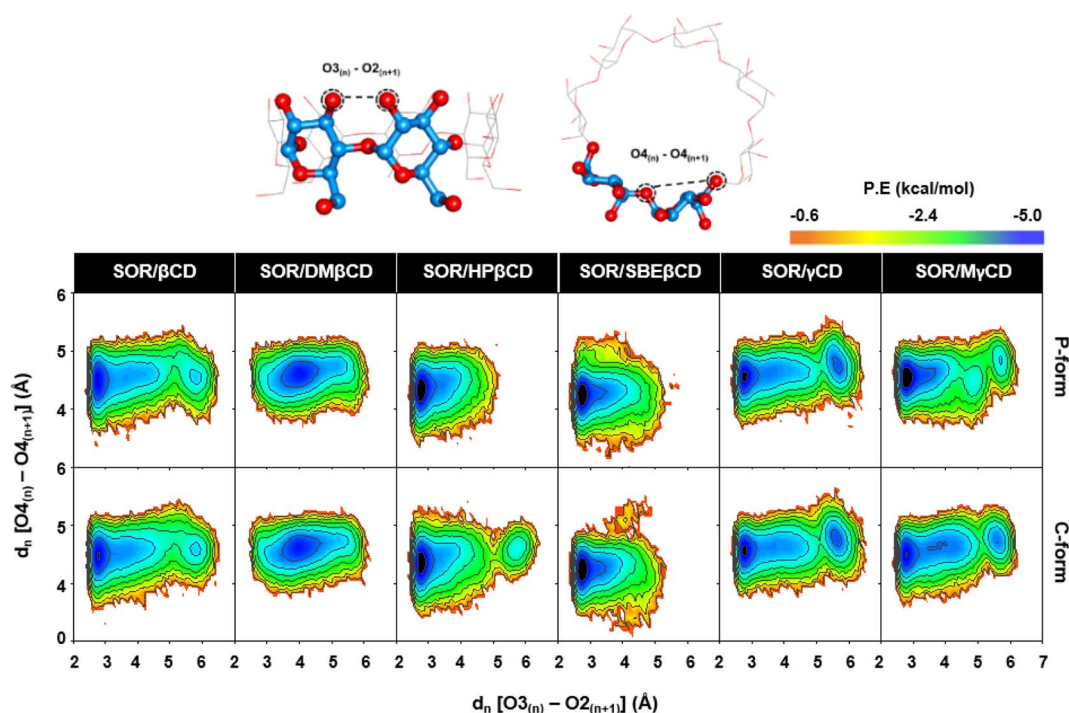


Fig. 7 PES plots generated for all studied CDs in complex with SOR, calculated over last 100 ns.



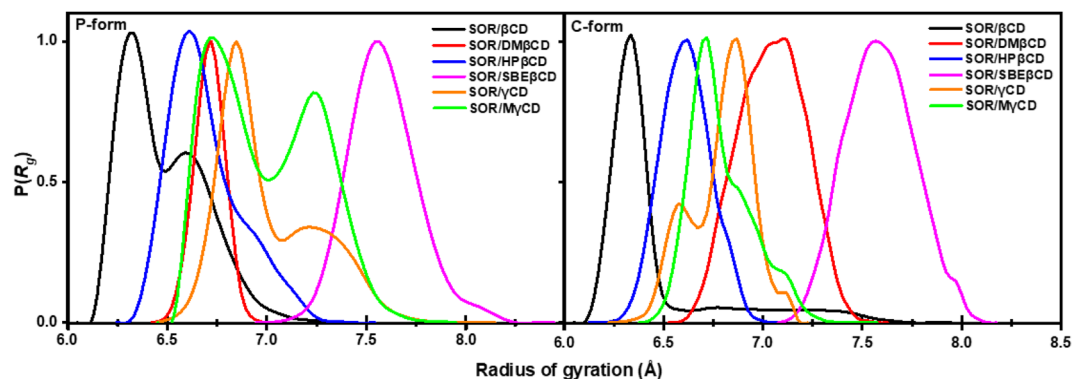


Fig. 8 Probability of the radius of gyration of all inclusion complexes both in P-form and C-form.

different from each other. We also found the fluctuation in SOR/ β CD (P-form), SOR/ γ CD, and SOR/M γ CD in both P-form and C-form which is due to the distortion of glucopyranose structures of CDs. These findings also support the PES calculation results (Fig. 7). In general, the investigation of molecular compactness provides additional evidence for the formation of an inclusion complex between SOR and CDs.

4.7. Binding free energy calculation

To assess the binding strength between host and guest molecules in various inclusion complexes, 10 000 snapshots taken from the last 100 ns MD simulations were utilized for SIE calculations using sietraj,⁴¹ depicted in Table 1. Compared to other complexes, the value obtained for SOR/SBE β CD (-7.17 to -7.84 kcal mol⁻¹) was the lowest. The observed energy value for the SOR/ γ CD complex (-6.16 to -6.50 kcal mol⁻¹) was lower than that of the SOR/ β CD complex (-5.80 to -6.24 kcal mol⁻¹), which aligns with the previous study where SOR/ γ CD shows higher stability constant (K_c) value (172.50 M⁻¹) than SOR/ β CD (68.30 M⁻¹).²⁴

In order to conduct a more in-depth study of β CD and its derivatives, binding free energy (ΔG_{bind}) was determined using MM/GBSA and MM/PBSA-based calculations. All energy components are shown in Fig. 9. The formation of inclusion complexes was predominantly influenced by the van der Waals force, which resulted in energy values of approximately -25 to -29 kcal mol⁻¹ for SOR/ β CD, -27 to -31 kcal mol⁻¹ for SOR/DM β CD, -34 to -38 kcal mol⁻¹ for SOR/HP β CD and -38 to -41 kcal mol⁻¹ for SOR/SBE β CD respectively. These findings align with previous studies that have highlighted the critical

role of the van der Waals force in driving the formation of inclusion complexes.^{54,55} The calculated $\Delta G^{\text{MM/GBSA}}$ for all complexes can be ranked as SOR/SBE β CD > SOR/HP β CD > SOR/DM β CD > SOR/ β CD, with the value of -10.22 ± 0.97 to -16.32 ± 1.10 kcal mol⁻¹, -7.99 ± 0.80 to -14.21 ± 1.10 kcal mol⁻¹, -11.26 ± 0.90 to -12.12 ± 0.59 kcal mol⁻¹, and -3.22 ± 1.10 to -7.02 ± 1.10 kcal mol⁻¹ respectively. The $\Delta G^{\text{MM/PBSA}}$ exhibited the same trend as $\Delta G^{\text{MM/GBSA}}$ with the value of -4.79 ± 1.00 to -11.14 ± 1.10 kcal mol⁻¹, -1.75 ± 0.90 to -6.25 ± 1.20 kcal mol⁻¹, -2.00 ± 1.10 to -3.00 ± 0.70 kcal mol⁻¹ and 1.00 ± 1.10 to -1.00 ± 0.70 kcal mol⁻¹ respectively. Studies have provided evidence that SBE β CD exhibits greater inclusion capacity than parent β CD because of the long hydrocarbon chain and hydrophobic butyl moiety present in its structure, which increases its hydrophobicity.^{56,57}

In addition, to understand the interactions of SOR with β CD and its derivatives, their percentage of interactions was calculated for the last 100 ns using LigandScout.⁵⁸ Fig. 10 highlighted the contribution of individual glucopyranose units or substituted groups toward SOR. It was found that the interactions of SOR/SBE β CD complex in both P-form and C-form were higher (up to 73% in the case of hydrophobic interactions and up to 33% in the case of hydrogen bonding) than SOR/HP β CD in both P-form and C-form (hydrophobic interaction; 36% and hydrogen bonding; 15%), SOR/DM β CD in both P-form and C-form (hydrophobic interaction; 41% and hydrogen bonding; 27%) and SOR/ β CD in both P-form and C-form (hydrogen bonding; 32%), which is consistent with ΔG_{bind} calculations (Fig. 9). When interacting with substituted groups on β CD, SOR

Table 1 Predicted ΔG^{SIE} of SOR in complex with all studied CDs, $n = 3$

	P-form			C-form		
	MD1	MD2	MD3	MD1	MD2	MD3
SOR/ β CD	-6.14 ± 0.34	-6.24 ± 0.41	-6.11 ± 0.35	-6.19 ± 0.54	-6.13 ± 0.35	-5.80 ± 0.51
SOR/DM β CD	-6.69 ± 0.32	-6.66 ± 0.33	-6.68 ± 0.32	-6.35 ± 0.34	-6.30 ± 0.32	-6.28 ± 0.30
SOR/HP β CD	-6.85 ± 0.43	-6.79 ± 0.41	-6.89 ± 0.43	-7.26 ± 0.41	-7.40 ± 0.41	-7.14 ± 0.41
SOR/SBE β CD	-7.23 ± 0.41	-7.17 ± 0.38	-7.24 ± 0.37	-7.78 ± 0.39	-7.84 ± 0.44	-7.74 ± 0.48
SOR/ γ CD	-6.16 ± 0.42	-6.30 ± 0.59	-6.25 ± 0.37	-6.50 ± 0.43	-6.25 ± 0.45	-6.29 ± 0.50
SOR/M γ CD	-6.58 ± 0.44	-6.97 ± 0.39	-6.60 ± 0.35	-6.92 ± 0.38	-6.84 ± 0.35	-6.65 ± 0.58



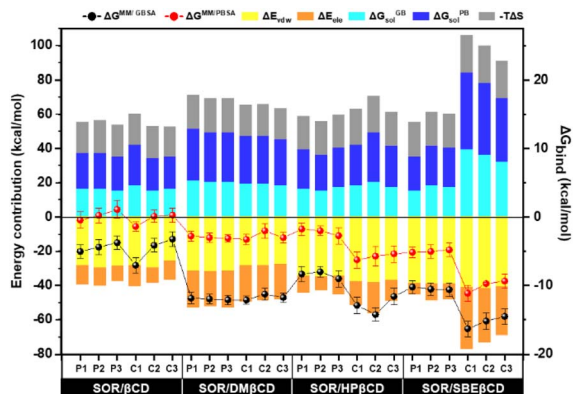


Fig. 9 Binding free energy calculation based on the MM/GBSA and MM/PBSA methods and their energy contribution of SOR in complex with CDs, calculated over last 100 ns simulation, $n = 3$.

exhibits hydrophobic interactions, while with the glucopyranose units of β CD, it forms hydrogen bonds.

4.8. Phase solubility study

The interaction between SOR and β CDs was analyzed experimentally using a phase solubility diagram where the

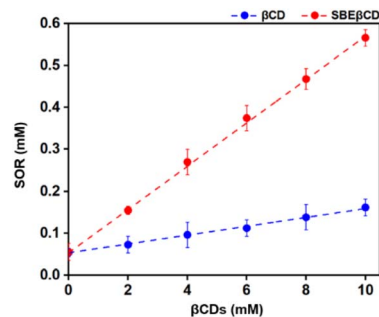


Fig. 11 The solubility behavior of the complex formed between SOR and β CDs at 25 °C as shown in the phase solubility plot.

concentration of β CDs was plotted along the x-axis and SOR concentration on y-axis. The results (Fig. 11) revealed that the concentration of SOR increased linearly with the increasing concentration of CDs. This finding suggests that SOR and the two CDs, β CD and SBE β CD can form inclusion complexes in a 1 : 1 stoichiometry ratio between the host and guest acquiring A_L type plot.²⁴ Previous studies have demonstrated that CDs can increase the stability of several drugs/compounds such as amoxicillin,⁵⁹ baicalein,⁶⁰ dicloxacillin,⁶¹ and eugenol.⁶² By formation of inclusion complexes, CDs provide a protective

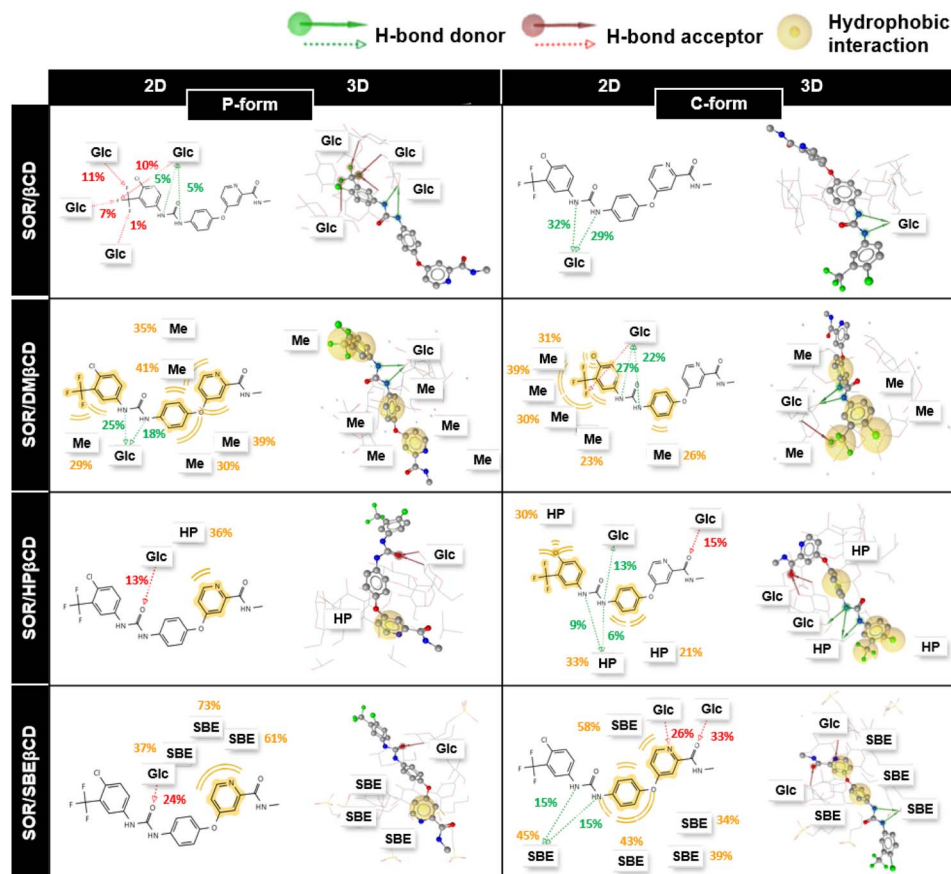


Fig. 10 The 2D and 3D guest–host interactions of β CDs in both orientations (P-form and C-form). Glc, Me, HP and SBE represent glucopyranose unit, methyl, hydroxypropyl, and sulfobutylether substitutions, respectively.



Table 2 The stability constants (K_c) of SOR/ β CD and SOR/SBE β CD determined at 25 °C

	Type	Slope	R^2	Intercept	K_c (M^{-1})	Reported K_c (M^{-1})
SOR/ β CD	A _L	0.011	0.996	0.052	210	68, ²⁴ 735 (ref. 66)
SOR/SBE β CD	A _L	0.051	0.999	0.058	940	—
SOR/ γ CD	B _S	—	—	—	—	172 (ref. 24)

shield around drug molecules, shielding them from environmental factors that could lead to degradation or loss of efficacy.⁶³ The effect of complexation on the chemical stability of drugs in terms of stability constant has been reported in previous studies for amoxicillin,⁵⁹ cephalotin,⁶⁴ and doxorubicin.⁶⁵ As shown in Table 2, the stability constant (K_c) of SOR/SBE β CD displays much better than SOR/ β CD with the values of 940 and 207 M^{-1} , indicating that SBE β CD is a better host than β CD for inclusion complexation with SOR.

Similarly, the previous studies reported that the phase solubility curve for SOR and β CD was an A_L-type, with the K_c value of 68.3 M^{-1} and 735.8 M^{-1} .^{24,66} However, the phase solubility curve for SOR and γ CD was reported to be a B_S-type, with a K_c value of 172.5 M^{-1} .²⁴

5. Conclusion

In this study, we conducted 200 ns MD simulations to observe the behavior of SOR when bound to CDs. Our findings suggest that there are two potential binding modes (referred to as P-form and C-form) for the SOR-CD complex, as indicated by molecular docking. According to the analysis of water accessibility and atomic contacts, the SBE β CD has higher encapsulation efficiency with SOR than other CDs. PES calculation indicated that all systems have a significant population of stable M1 region, but a distortion of glucose units (M2) was only observed in SOR/ β CD, SOR/ γ CD, and SOR/M γ CD systems. In the MM/GBSA and MM/PBSA calculations, SOR/SBE β CD exhibited the lowest ΔG_{bind} values, followed by SOR/HP β CD, SOR/DMBCD, and SOR/ β CD, respectively. In the analysis of MM energy components, the inclusion complex is primarily formed by the van der Waals interaction. Phase solubility studies indicated that β CD and SBE β CD were able to enhance the solubility of SOR. Moreover, the observed A_L-type profile implied a 1 : 1 interaction between the host and guest molecules, and a greater K_c was observed in SOR/SBE β CD. Based on the structural evidence and experimental results, it can be concluded that CDs, particularly SBE β CD, have the potential as effective drug carrier for SOR to enhance its water solubility and stability.

Conflicts of interest

The authors report no conflict of interest, financial or otherwise.

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References

- 1 M. L. Bondi, A. Scala, G. Sortino, E. Amore, C. Botto, A. Azzolina, D. Balasus, M. Cervello and A. Mazzaglia, *Biomacromolecules*, 2015, **16**, 3784–3791.
- 2 S. M. Wilhelm, C. Carter, L. Tang, D. Wilkie, A. McNabola, H. Rong, C. Chen, X. Zhang, P. Vincent, M. McHugh, Y. Cao, J. Shujath, S. Gawlak, D. Eveleigh, B. Rowley, L. Liu, L. Adnane, M. Lynch, D. Auclair, I. Taylor, R. Gedrich, A. Voznesensky, B. Riedl, L. E. Post, G. Bollag and P. A. Trail, *Cancer Res.*, 2004, **64**, 7099–7109.
- 3 J. M. Llovet, A. M. Di Bisceglie, J. Bruix, B. S. Kramer, R. Lencioni, A. X. Zhu, M. Sherman, M. Schwartz, M. Lotze, J. Talwalkar, G. J. Gores and Panel of Experts in HCC D. C. T., *J. Natl. Cancer Inst.*, 2008, **100**, 698–711.
- 4 P. T. White and M. S. Cohen, *Expert Opin. Drug Discovery*, 2015, **10**, 427–439.
- 5 X. Q. Wang, J. M. Fan, Y. O. Liu, B. Zhao, Z. R. Jia and Q. Zhang, *Int. J. Pharm.*, 2011, **419**, 339–346.
- 6 European Medicines Agency, *Nexavar (Sorafenib): Summary of Product Characteristics*, https://www.ema.europa.eu/en/documents/product-information/nexavar-epar-product-information_en.pdf.
- 7 M. Pulkkinen, J. Pikkarainen, T. Wirth, T. Tarvainen, V. Haapa-aho, H. Korhonen, J. Seppala and K. Jarvinen, *Eur. J. Pharm. Biopharm.*, 2008, **70**, 66–74.
- 8 Z. Izadiyan, M. Basri, H. R. Fard Masoumi, R. Abedi Karjiban, N. Salim and K. Kalantari, *Mater. Sci. Eng., C*, 2019, **94**, 841–849.
- 9 A. S. Deshmukh, K. J. Tiwari and V. R. Mahajan, *Int. J. Pharm. Sci. Nanotechnol.*, 2017, **10**(8), 3701–3708.
- 10 A. Shanmuga Priya, J. Sivakamavalli, B. Vaseeharan and T. Stalin, *Int. J. Biol. Macromol.*, 2013, **62**, 472–480.
- 11 E. M. Del Valle, *Process Biochem.*, 2004, **39**, 1033–1046.
- 12 S. Tommasini, D. Raneri, R. Ficarra, M. L. Calabrò, R. Stancanelli and P. Ficarra, *J. Pharm. Biomed. Anal.*, 2004, **35**, 379–387.
- 13 C. Muankaew and T. Loftsson, *Basic Clin. Pharmacol. Toxicol.*, 2018, **122**, 46–55.



- 14 A. Popielec and T. Loftsson, *Int. J. Pharm.*, 2017, **531**, 532–542.
- 15 P. Wongpituk, *Mol. Simul.*, 2017, **43**(13–16), 1356–1363.
- 16 S. Gould and R. C. Scott, *Food Chem. Toxicol.*, 2005, **43**, 1451–1459.
- 17 Y. Nagase, M. Hirata, H. Arima, S. Tajiri, Y. Nishimoto, F. Hirayama, T. Irie and K. Uekama, *J. Pharm. Sci.*, 2002, **91**, 2382–2389.
- 18 D. R. Luke, K. Tomaszewski, B. Damle and H. T. Schlamm, *J. Pharm. Sci.*, 2010, **99**, 3291–3301.
- 19 T. Irie and K. Uekama, *J. Pharm. Sci.*, 1997, **86**, 147–162.
- 20 T. Loftsson and D. Duchêne, *Int. J. Pharm.*, 2007, **329**, 1–11.
- 21 Z. Li, M. Wang, F. Wang, Z. Gu, G. Du, J. Wu and J. Chen, *Appl. Microbiol. Biotechnol.*, 2007, **77**, 245–255.
- 22 X. Zuobing, W. Hou, Y. Kang, N. Yunwei and X. Kou, *Food Hydrocolloids*, 2019, **97**, 105202.
- 23 H. Bai, J. Wang, C. U. Phan, Q. Chen, X. Hu, G. Shao, J. Zhou, L. Lai and G. Tang, *Nat. Commun.*, 2021, **12**(1), 759.
- 24 C. Phan, Z. Zheng, J. Wang, Q. Wang, X. Hu, G. Tang and H. Bai, *Biomater. Sci.*, 2019, **7**, 4758–4768.
- 25 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, G. A. Petersson, H. Nakatsuji, X. Li, M. Caricato, A. Marenich, J. Bloino, B. G. Janesko, R. Gomperts, B. Mennucci, H. P. Hratchian, J. V. Ortiz, A. F. Izmaylov, J. L. Sonnenberg, D. Williams-Young, F. Ding, F. Lipparini, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson, D. Ranasinghe, V. G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, K. Throssell, J. A. Montgomery, Jr, J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, T. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, J. M. Millam, M. Klene, C. Adamo, R. Cammi, J. W. Ochterski, R. L. Martin, K. Morokuma, O. Farkas, J. B. Foresman, and D. J. Fox, *Gaussian 09, Revision A.02*, Gaussian, Inc., Wallingford CT, 2016.
- 26 M. Klaewkla, T. Charoenwongpaiboon and P. Mahalapbutr, *J. Mol. Liq.*, 2021, **335**, 116537.
- 27 T. Aiebchun, P. Mahalapbutr, A. Auepattanapong, O. Khaikate, S. Seetaha, L. Tabtimmai, C. Kuhakarn, K. Choowongkamon and T. Rungrotmongkol, *Molecules*, 2021, **26**, 2211.
- 28 K. Sanachai, T. Aiebchun, P. Mahalapbutr, S. Seetaha, L. Tabtimmai, P. Maitarad, I. Xenikakis, A. Geronikaki, K. Choowongkamon and T. Rungrotmongkol, *RSC Med. Chem.*, 2021, **12**, 430–438.
- 29 MarvinSketch, Chemaxon, <https://chemaxon.com/marvin>.
- 30 P. Li, J. Song, X. Ni, Q. Guo, H. Wen, Q. Zhou, Y. Shen, Y. Huang, P. Qiu, S. Lin and H. Hu, *Int. J. Pharm.*, 2016, **513**, 347–356.
- 31 K. Kerdpol, J. Kicuntod, P. Wolschann, S. Mori, C. Rungrim, M. Kunaseth, H. Okumura, N. Kungwan and T. Rungrotmongkol, *Polymers*, 2019, **11**, 145.
- 32 Z.-I. Szabó, R. Ludmerczki, B. Fiser, B. Noszá and G. Tóth, *Electrophoresis*, 2019, **40**, 1897–1903.
- 33 P. Mahalapbutr, P. Wongsanan, T. Charoenwongpaiboon, M. Prousoontorn, W. Chavasiri and T. Rungrotmongkol, *Biomolecules*, 2019, **9**, 545.
- 34 J. Q. Zhang, D. Wu, K. M. Jiang, D. Zhang, X. Zheng, C. P. Wan, H. Y. Zhu, X. G. Xie, Y. Jin and J. Lin, *Carbohydr. Res.*, 2015, **406**, 55–64.
- 35 K. N. Kirschner, A. B. Yongye, S. M. Tschampel, J. González-Outeirino, C. R. Daniels, B. L. Foley and R. J. Woods, *J. Comput. Chem.*, 2008, **29**, 622–655.
- 36 J. Wang, R. M. Wolf, J. W. Caldwell, P. A. Kollman and D. A. Case, *J. Comput. Chem.*, 2004, **25**, 1157–1174.
- 37 D. A. Case, K. Belfon, I. Y. Ben-Shalom, S. R. Brozell, D. S. Cerutti, T. E. Cheatham, III, V. W. D. Cruzeiro, T. A. Darden, R. E. Duke, G. Giambasu, M. K. Gilson, H. Gohlke, A. W. Goetz, R. Harris, S. Izadi, S. A. Izmailov, K. Kasavajhala, A. Kovalenko, R. Krasny, T. Kurtzman, T. S. Lee, S. LeGrand, P. Li, C. Lin, J. Liu, T. Luchko, R. Luo, V. Man, K. M. Merz, Y. Miao, O. Mikhailovskii, G. Monard, H. Nguyen, A. Onufriev, F. Pan, S. Pantano, R. Qi, D. R. Roe, A. Roitberg, C. Sagui, S. Schott-Verdugo, J. Shen, C. L. Simmerling, N. R. Skrynnikov, J. Smith, J. Swails, R. C. Walker, J. Wang, L. Wilson, R. M. Wolf, X. Wu, Y. Xiong, Y. Xue, D. M. York and P. A. Kollman, *AMBER 2020*, University of California, San Francisco, 2020.
- 38 J.-P. Ryckaert, G. Ciccotti and H. J. C. Berendsen, *J. Comput. Phys.*, 1977, **23**, 327–341.
- 39 B. A. Luty and W. F. Van Gunsteren, *J. Phys. Chem.*, 1996, **100**, 2581–2587.
- 40 M. Ester, H.-P. Kriegel, J. Sander and X. Xu, *A density-based algorithm for discovering clusters in large spatial databases with noise*, 1996, vol. 96, pp. 226–231.
- 41 T. Sulea, Q. Cui and E. O. Purisima, *J. Chem. Inf. Model.*, 2011, **51**, 2066–2081.
- 42 S. Genheden and U. Ryde, *Expert Opin. Drug Discovery*, 2015, **10**, 449–461.
- 43 T. Higuchi, *Adv. Anal. Chem. Instrum.*, 1965, **4**, 117–212.
- 44 P. Saokham, C. Muankaew, P. Jansook and T. Loftsson, *Molecules*, 2018, **23**, 1161.
- 45 G. Raffaini, A. Mazzaglia and M. Catauro, *Macromol. Symp.*, 2021, **395**, 2000201.
- 46 P. Mahalapbutr, T. Charoenwongpaiboon, C. Phongern, N. Kongtaworn, S. Hannongbua and T. Rungrotmongkol, *J. Mol. Liq.*, 2021, **337**, 116394.
- 47 B. Ren, H. Gao, Y. Cao and L. Jia, *J. Hazard. Mater.*, 2015, **285**, 148–156.
- 48 C. W. Yong, C. Washington and W. Smith, *Pharm. Res.*, 2008, **25**, 1092–1099.
- 49 J. Kicuntod, W. Khuntawee, P. Wolschann, P. Pongsawasdi, W. Chavasiri, N. Kungwan and T. Rungrotmongkol, *J. Mol. Graphics Modell.*, 2016, **63**, 91–98.
- 50 A. Oo, K. Kerdpol, P. Mahalapbutr and T. Rungrotmongkol, *J. Mol. Liq.*, 2022, **347**, 118002.
- 51 P. Mahalapbutr, B. Nutho, P. Wolschann, W. Chavasiri, N. Kungwan and T. Rungrotmongkol, *J. Mol. Graphics Modell.*, 2018, **79**, 72–80.



- 52 M. Lobanov, N. S. Bogatyreva and O. V. Galzitskaia, *Mol. Biol.*, 2008, **42**, 701–706.
- 53 T. Charoenwongpaiboon, A. Oo, S. Nasoontorn, T. Rungrotmongkol, S. Kanokmedhakul and P. Mahalapbutr, *Int. J. Mol. Sci.*, 2022, **23**, 9776.
- 54 H. M. H. Soe, S. Chamni, P. Mahalapbutr, N. Kongtaworn, T. Rungrotmongkol and P. Jansook, *Carbohydr. Res.*, 2020, **498**, 108190.
- 55 W. Chen, C.-E. Chang and M. K. Gilson, *Biophys. J.*, 2004, **87**, 3035–3049.
- 56 M. Fukuda, D. A. Miller, N. A. Peppas and J. W. McGinity, *Int. J. Pharm.*, 2008, **350**, 188–196.
- 57 O. Das, V. M. Ghate and S. A. Lewis, *Indian J. Pharm. Sci.*, 2019, **81**, 589–600.
- 58 G. Wolber and T. Langer, *J. Chem. Inf. Model.*, 2005, **45**, 160–169.
- 59 S. Hidaka, T. Tokumura, K. Tomono, T. Suzuki, H. Ueda, T. Nagai, M. Nagaoka, R. Nakane and Y. Machida, *Yakugaku Zasshi*, 2010, **130**, 889–893.
- 60 N. Bourkaib, J. Zhou, J. Yao, Z. Fang and O. Mezghrani, *Drug Dev. Ind. Pharm.*, 2013, **39**, 918–927.
- 61 M. M. Echezarreta-Lopez, I. Otero-Mazoy, H. L. Ramírez, R. Villalonga and J. J. Torres-Labandeira, *Curr. Drug Discovery Technol.*, 2008, **5**, 140–145.
- 62 F. Kayaci, Y. Ertas and T. Uyar, *J. Agric. Food Chem.*, 2013, **61**, 8156–8165.
- 63 L. Dahabra, G. Broadberry, A. Le Gresley, M. Najlah and M. Khoder, *Molecules*, 2021, **26**, 1698.
- 64 T. Loftsson, H. Friðriksdóttir, E. Stefansson, S. Thorisdottir, Ö. Guðmundsson and T. Sigthorsson, *J. Pharm. Pharmacol.*, 1994, **46**, 503–504.
- 65 M. E. Brewster, T. Loftsson, K. S. Estes, J.-L. Lin, H. Fridriksdóttir and N. Bodor, *Int. J. Pharm.*, 1992, **79**, 289–299.
- 66 M. R. Donthi, S. R. Munnangi, K. V. Krishna, S. A. Marathe, R. N. Saha, G. Singhvi and S. K. Dubey, *AAPS PharmSciTech*, 2022, **23**, 254.

