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Synthesis and biological evaluation of novel β -lactam-metallo β -lactamase inhibitors†

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β -lactamases are enzymes that deactivate β -lactam antibiotics through a hydrolysis mechanism. There are two known types of β -lactamases: serine β -lactamases (SBLs) and metallo β -lactamases (MBLs). The two existing strategies to overcome β -lactamase-mediated resistance are (a) to develop novel β -lactam antibiotics that are not susceptible to hydrolysis by these enzymes; or (b) to develop β -lactamase inhibitors that deactivate the enzyme and thereby restore the efficacy of the co-administered antibiotics. Many commercially available SBL inhibitors are used in combination therapy with antibiotics to treat antimicrobial resistant infections; however, there are only a handful of MBL inhibitors undergoing clinical trials. In this study, we present 11 novel potential MBL inhibitors (*via* multi-step chemical synthesis), that have shown to completely restore the efficacy of meropenem ($\leq 2 \text{ mg L}^{-1}$) against New Delhi metallo- β -lactamase (NDM) producing *Klebsiella pneumoniae in vitro*. These compounds contain a cyclic amino acid zinc chelator conjugated to various commercially available β -lactam antibiotic scaffolds with the aim to improve the overall drug transport, lipophilicity, and pharmacokinetic/pharmacodynamic properties as compared to the chelator alone. Biological evaluation of compounds **24b** and **24c** has further highlighted the downstream application of these MBLs, since they are non-toxic at the selected doses. Time-kill assays indicate that compounds **24b** and **24c** exhibit sterilizing activity towards NDM producing *Klebsiella pneumoniae in vitro* using minimal concentrations of meropenem. Furthermore, **24b** and **24c** proved to be promising inhibitors of VIM-2 ($K_i = 0.85$ and 1.87 , respectively). This study has revealed a novel series of β -lactam MBLIs that are potent, efficacious, and safe leads with the potential to develop into therapeutic MBLIs.

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Introduction

The World Health Organisation (WHO) has predicted that drug resistance will cause the deaths of 10 million people worldwide every year by 2050.^{1,2} Incorrect prescription, dispensing and usage of commercialised antibiotics have contributed significantly to antibiotic resistance.^{1,3,4} The Antimicrobial Resistance (AMR) National Strategy Framework (2018–2024), as well as the Global Research and Development priority setting for AMR, and

the Global Antibiotic Research and Development Partnership, prioritise research regarding antimicrobial treatment to help prevent obsolete antibiotics emanating as a result of mutations and bacterial evolution.^{5–7} Antibiotic resistance has worsened, due to the empirical treatment of hospitalised COVID-19 patients.^{3,8,9} There are several initiatives in place to either raise awareness, help reduce, or keep track of resistance, such as: the Global Action Plan on Antimicrobial Resistance (GAP); World Antimicrobial Awareness Week (WAAW); The Global Antimicrobial Resistance and Use Surveillance System (GLASS); Global Research and Development priority setting for AMR; Access Watch Reserve (AWaRe); and Global Antibiotic Research and Development Partnership (GARDP).⁷ Murray *et al.* estimated that 4.95 million deaths occurred in the year 2019 as a result of antibiotic resistance.⁶ Identified *Klebsiella pneumoniae*, as a common pathogen accounting for 29% of all reported bacterial infections.⁷ *Klebsiella pneumoniae* has recently been identified as a bacteria of concern, as mentioned in several reports, studies, and reviews, and is further substantiated by the resistance map (Fig. 1) generated from the CDDEP (Centre for Disease Dynamics, Economics and Policy).¹⁰ Unfortunately,

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Resistance of *Klebsiella pneumoniae* to Carbapenems

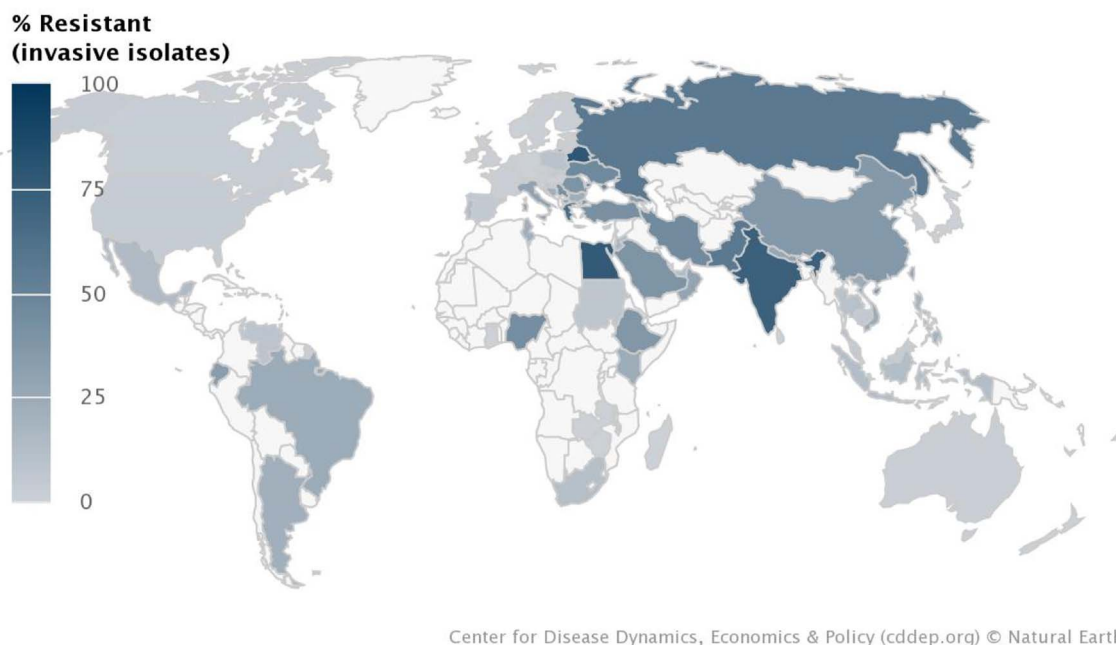


Fig. 1 World map indicating the incidence of carbapenem-resistant *Klebsiella pneumoniae* in each country.¹⁰

most countries, especially the lower-middle-income countries, either do not supply their data to surveillance systems such as GLASS, or do not track and identify the bacterial infections; therefore, the resistance percentages indicated in this map are an underestimation of the actual prevalence. These alarming trends call for urgent intervention to treat AMR infections.

β -lactamases are enzymes produced by the bacteria as a defence mechanism, which irreversibly deactivates β -lactam antibiotics by hydrolysing/cleaving the amide bond in the β -lactam ring before it has had the opportunity to bind to the penicillin-binding proteins (PBPs).^{3,11–17} These enzymes have been characterised by Ambler classes A, B, C and D, based on their amino acid homology and molecular properties.^{11–14} Classes A, C and D (SBLs) utilise a serine residue at the active site to effect the hydrolyses of the β -lactams while class B enzymes utilize either one or two zinc(II) ions at their active sites. The latter are referred to as MBLs and use a hydroxide ion from an activated water molecule as a nucleophile in the ring-opening process.^{11,14,18–21}

Several SBL inhibitors are used in combination therapy with β -lactam antibiotics *e.g.*, ampicillin with sulbactam, piperacillin with tazobactam, and a host of other combinations.¹² Unlike the SBLs, there are no MBLs with clinically approved inhibitors^{8,12,13} or enhancers.²² MBLs have been shown to deactivate all β -lactam-containing antibiotics, except for monobactams (specifically aztreonam).^{23,24} Aztreonam is used in combination with ceftazidime/avibactam as a treatment since most MBL-producing *Enterobacteriales* co-produce SBLs, which can hydrolyse aztreonam.^{11,23–29} A 2003 study found that metal chelators

inhibit the Verona Integron-encoded metallo- β -lactamase 2 (VIM-2) enzyme, presumably through complexation of its zinc ion.¹² This led to the hypothesis that metal chelators could be developed to quell resistance caused by MBLs.¹²

Metal chelating agents, such as 2,6-dipicolinic acid (DPA) and ethylenediaminetetraacetic acid (EDTA), have proven to inhibit the MBL's catalytic activity by isolating or removing the metal ions from its active site.^{17,30–32} One of the drawbacks of using these “metal stripping” agents for clinical use is the lack of selectivity and specificity towards MBLs as their use could result in the removal of essential metals and the deactivation of physiological metalloproteins.^{17,33}

Our group recently investigated the antibacterial effect of cyclic amino acidic zinc chelators when co-administered with the clinically-used carbapenem antibiotics, meropenem and imipenem (the most widely prescribed carbapenem antibiotics worldwide).^{12,34,35} We found that conjugation to existing β -lactam antibiotic scaffolds improved the transport of the chelators to the MBL active site, as well as their pharmacokinetics/pharmacodynamics (PK/PD) profile.^{36,37} Our rationale was based on the premise that the chelating moiety would be responsible for MBL inhibition whilst the β -lactam scaffold would provide for increased lipophilicity and selective transport to the bacteria.

Results and discussion

Following from our studies conducted for NOTA (1)^{12,34} and the reported BP series,^{36,37} we decided to investigate the effect of (i)



NOTA and (ii) β -lactam scaffolds as potential MBLIs. We envisaged that these changes could offer alternate PK/PD profiles of such MBLIs. We first investigated the effect of varying the chelator to having an additional donor atom for metal bonding and an additional carbon spacer for steric effects (Fig. 2).

For this to be achieved, one of the carboxylic acids from these zinc chelators was to be employed in an amide bond linkage to the lactam scaffold. The protected form of NOTA (7) or NODAGA (4) chelators were purchased and used for coupling to the commercially available 7-aminocephalosporanic acid (7-ACA) (Scheme 1). 7-ACA is a cephalosporin core which has been used to generate a number of cephalosporin antibiotics. Its solubility has proved to be very poor in solvents such as dimethylformamide (DMF) and acetonitrile (ACN), which are commonly employed for amide couplings. 7-ACA had to be dissolved in a mixture of acetone and water with base, then coupled to 4 or 7 using the peptide coupling agent 1-[bis(dimethylamino)methylene]-1*H*-1,2,3-triazolo[4,5-*b*]pyridinium 3-oxid

hexafluorophosphate (HATU) at -10°C , to furnish compounds 5 and 8 in yields of 40% and 25%, respectively, after purification with preparative supercritical fluid chromatography (prep-SFC). Since compound 4 formed an asymmetric centre, compound 5 and its subsequent products were obtained as an epimeric mixture. Attempts to improve the yield with higher temperatures resulted in the formation of undesired by-products. Compounds 5 and 8 were deprotected with trifluoroacetic acid (TFA), using thioanisole as a scavenger to afford the target compounds 6 and 9 in 60% and 70% yields respectively.

Unlike NOTA and NODAGA, protected NODASA could not be obtained commercially or synthetically. Challenges with its synthesis are discussed in the ESI.† An alternative route was envisaged whereby we first attach the β -lactam to the chelator, followed by the alkylation in order to provide enough steric hindrance to prevent the over-alkylation step. We synthesised 11 using a procedure reported by Dutta *et al.*,³⁸ 1,4,7-triazanonane (10) underwent a Michael-addition reaction with mono-methyl fumarate to afford 11 in excellent yields (>95%) (Scheme 2). Compound 11 was Boc-protected to afford 12 in yields of 85–96% after purification on a silica gel column. Compound 12 was coupled to 7-ACA to afford 13 with yields of 30–50%, following similar protocols adopted for the coupling of NOTA and NODAGA. Similar to compound 4 all compounds obtained after the coupling reaction of 11 with enantiomerically pure β -lactams were also obtained as epimeric mixtures. The Boc-groups were removed at room temperature using TFA and anisole as the scavenger in dichloroethane (DCE) and washed in ether to afford the TFA salt of 14 in excellent yields (>90%), without the need for further purification. Compound 14 was alkylated with *tert*-butyl bromoacetate to afford 15, without any observable over-alkylation and was purified *via* prep-SFC to provide the pure product in 46–50% yields. Compound 15 was

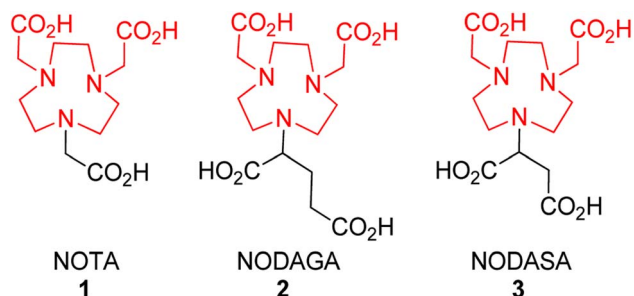
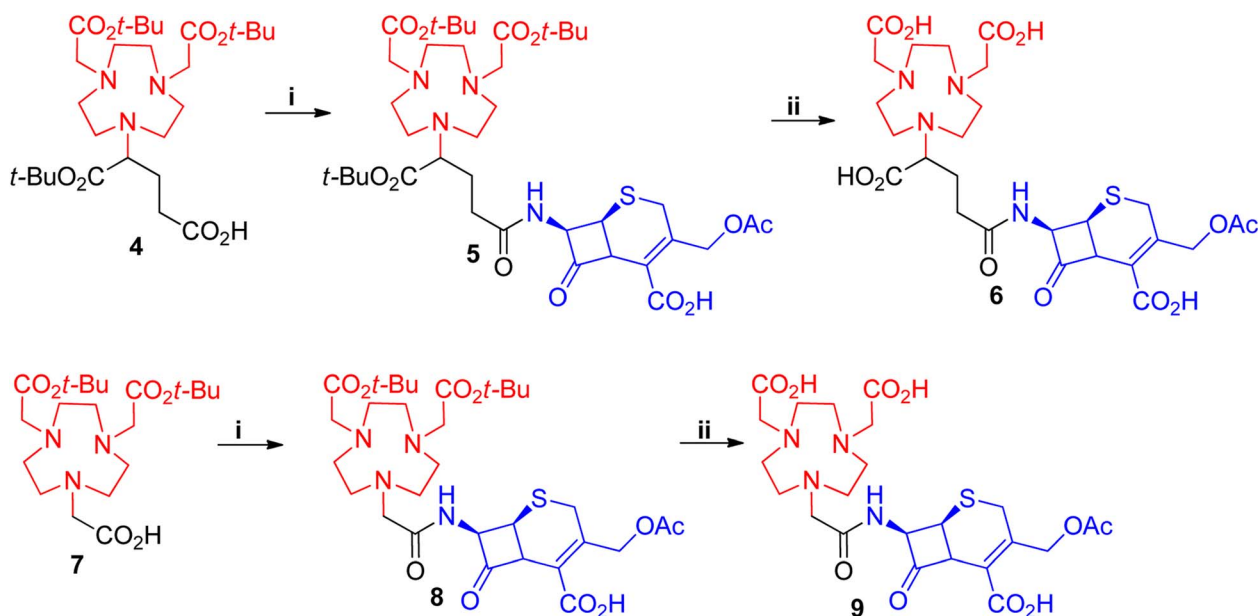
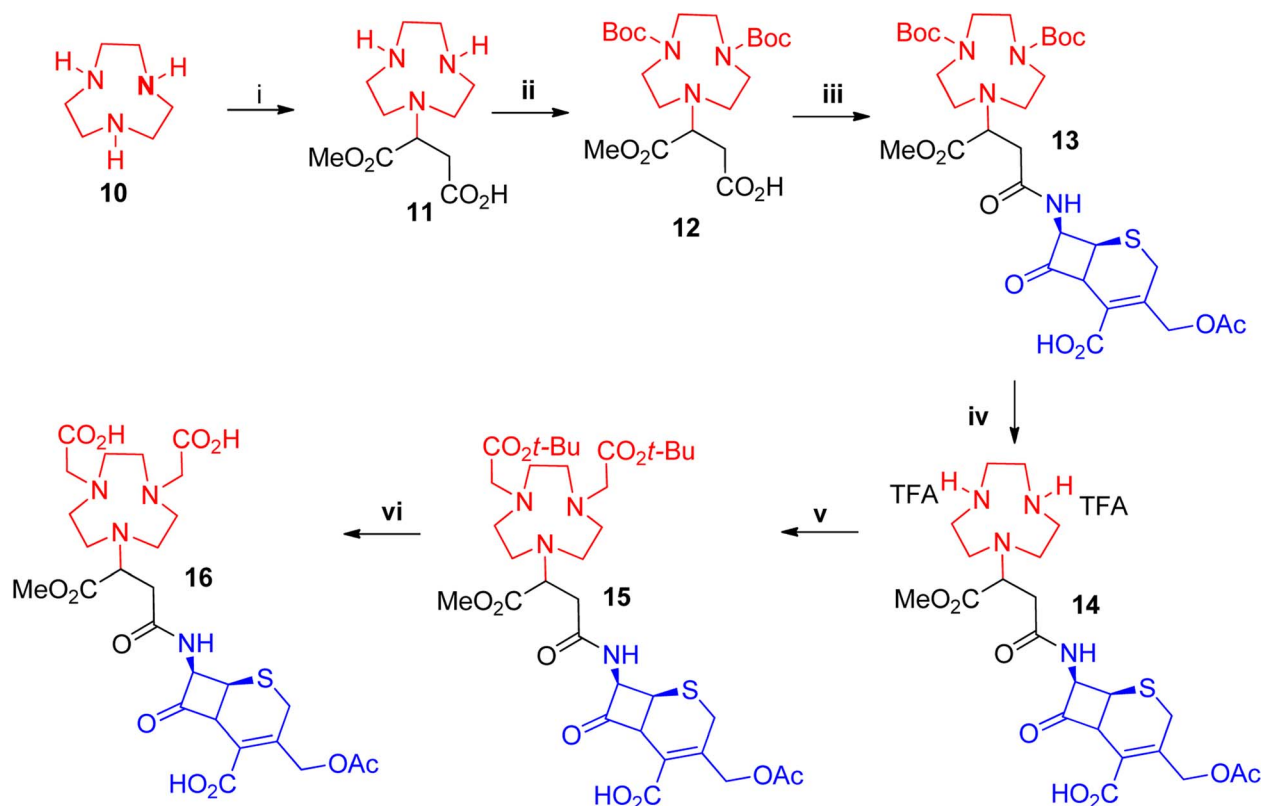


Fig. 2 1,4,7-Triazacyclononane chelators: NODASA derivative, NOTA and NODAGA.



Scheme 1 Synthesis of NOTA-7-ACA and NODAGA-7-ACA. i = HATU, DIEA, ACN, 7-ACA (dissolved in a mixture of NaHCO_3 , H_2O , acetone, ACN), -20°C , 1 h ii = thioanisole, TFA, rt, 3 h.





Scheme 2 Synthesis of NODASA-7-ACA. i = mono-methyl fumarate, DIEA, DCM, rt, 4 h ii = Boc_2O , DIEA, DCM, rt, 4 h iii = HATU, DIEA, ACN, 7-ACA (dissolved in a mixture of NaHCO_3 , H_2O , acetone, ACN), -20°C , 1 h iv = anisole, TFA, DCE, rt, 20 min. v = *tert*-butyl bromoacetate, DIEA, DCE, $0-25^\circ\text{C}$, 1 h vi = thioansole, TFA, rt, 3 h.

subjected to a hydrolysis reaction of the *tert*-butyl groups with TFA to afford the target compound **16** in 65% yield. All reactions were monitored with LC-MS and characterized using standard techniques.

Analogues **6**, **9** and **16** were evaluated, in combination with meropenem, for their ability to inhibit NDM harbouring strains of *Escherichia coli*³⁹ and *Klebsiella pneumoniae*.¹⁰ Meropenem is a carbapenem antibiotic in clinical use and currently displays little-to-no activity against carbapenemase-producing bacteria. Hence, it was chosen to assess the synergistic effect of these new MBLIs (Table 1, entry 1).⁴⁰ The compounds were evaluated using the checkerboard method and the results are shown in Table 1. Unless otherwise stated, the minimum inhibitory concentrations (MIC) were obtained by co-administration of meropenem with an inhibitor and all experiments were performed in triplicate.

Table 1 shows that meropenem alone did not inhibit the growth of neither *E. coli* NDM-1 nor *Klebsiella pneumoniae* NDM at a concentration of 128 mg L^{-1} . Co-administration with high concentrations of 7-ACA resulted in only a modest regain of meropenem activity against both strains. However, co-administration with chelator compounds **1** and **2** drastically enhanced the activity of meropenem to achieve MICs between $0.06-0.125\text{ mg L}^{-1}$ with low chelator concentrations of $4-16\text{ mg L}^{-1}$. Although the results for chelators **1** and **2** were excellent, they are not viable options for therapy due to the low

bioavailability, non-specificity towards metal chelators, and off-target activities that often accompanies these types of compounds. The synthesized compounds **6**, **9**, and **16** were found to lack any antibacterial activities on their own, even with concentrations greater than 128 mg L^{-1} . Combination of these chelators with meropenem restored its MIC efficacies which is

Table 1 The MIC activity of meropenem with zinc chelators and when coupled to 7-ACA different against MBL-producing bacterial species

Entry	Inhibitor	Minimum inhibitory concentration ^a - mg L^{-1}			
		<i>Escherichia coli</i> NDM-1		<i>Klebsiella pneumoniae</i> NDM	
		Meropenem	MBLI	Meropenem	MBLI
1	None	128	0	128	0
2	7-ACA	2	64	2	32
3	NOTA (1)	0.06	4	0.125	4
4	NODAGA (2)	0.06	4	0.06	16
5	6	0	>128	0	>128
6	9	0	>128	0	>128
7	16	0	>128	0	>128
8	6	0.25	32	0.25	16
9	9	0.125	16	0.125	16
10	16	1	16	2	8
11	NOTA-Zn	>128	>128	>128	>128

^a The checkerboard assays were conducted in triplicate $p < 0.01$.



Table 2 The MICs of meropenem with NODASA- β -lactam inhibitors coupled against two MBL producing bacterial strains

Entry	Inhibitor	Minimum inhibitory concentration (mg L^{-1}) ^a			
		<i>Escherichia coli</i> NDM-1		<i>Klebsiella pneumoniae</i> NDM	
		Meropenem	MBLI	Meropenem	MBLI
1	9	0.125	16	0.125	16
2	20a	0.125	16	0.25	16
3	20b	0.5	16	0.5	16
4	24a	0.06	16	0.125	8
5	24b	0.125	8	0.25	8
6	24c	0.06	8	0.25	8
7	24d	0.125	16	0.125	16

^a All assays were conducted in triplicate $p < 0.001$.**Table 3** MIC of compounds **24b** and **24c** against NDM, VIM and IMP expressing bacteria

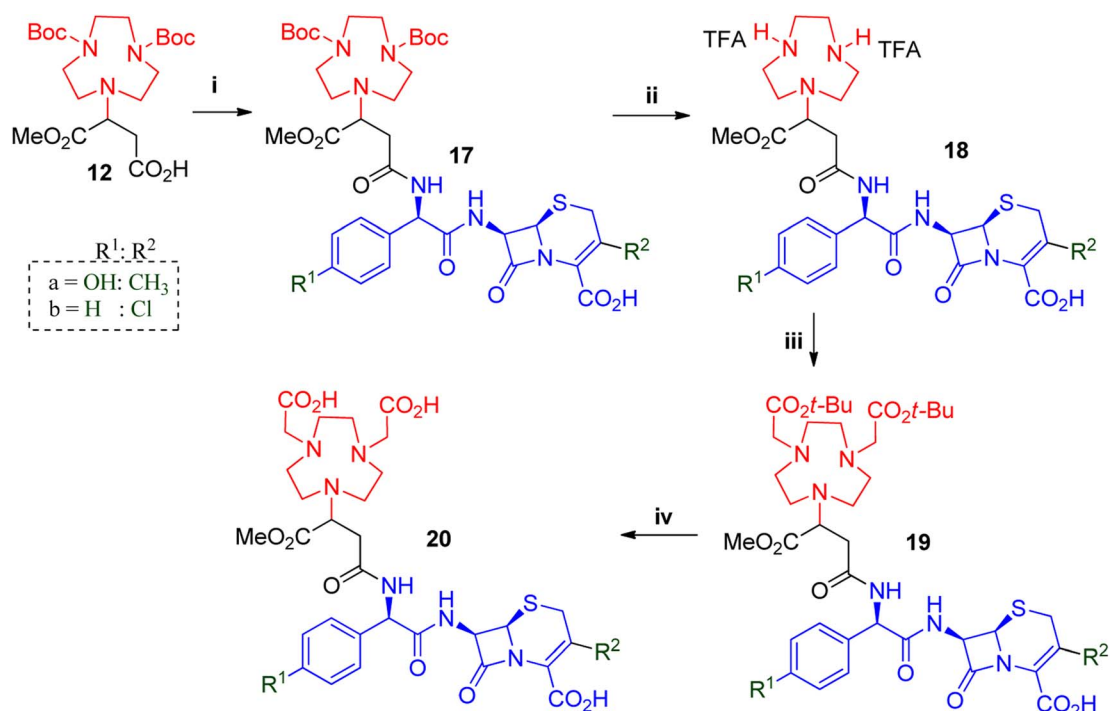
MBL-producing bacteria	Minimum inhibitory concentration (mg L^{-1}) ^a			
	Meropenem	24b	Meropenem	24c
<i>E. coli</i> NDM-4	0.06	8	0.25	8
<i>E. coli</i> IMP-1	0.03	8	0.5	8
<i>E. coli</i> IMP-8	0.125	8	0.25	8
<i>Enterobacter cloacae</i> NDM-1	0.25	8	0.25	8
<i>E. cloacae</i> VIM-1	0.5	8	0.25	8

^a All assays were conducted in triplicate $p < 0.001$.

evidence for their MBL inhibitory activities. Importantly, zinc pre-complexed NOTA was also evaluated as a potential MBL inhibitor and as expected, produced no bacterial inhibition, confirming that zinc chelation is essential to restore the activity of meropenem. Additionally, the compounds in Table 1 displayed an absence of activity towards SBL expressing bacteria *Serratia marcescens* (KPC-2) and *E. coli* (OXA-28), indicating specific activity towards MBLs; NDM (Tables 1–3), IMP and VIM (Table 3).

The biological activity confirmed that 7-ACA, coupled with various zinc chelators, could restore the activity of meropenem towards MBL-expressing pathogens. The high cost of commercial starting materials **7** and **4**, prompted us to pursue the structural optimisation with the more affordable NODASA (**3**) derivatives.

The synthesis of further derivatives was based on related and commercially available cephalosporins: cefadroxil; cefaclor; ceftizoxime; ceftibuten; ceftiofur and ceftazidime. Structurally similar β -lactams cefadroxil and cefaclor were used to synthesise compounds **20a** and **20b** (Scheme 3). Unlike 7-ACA, cefadroxil had excellent solubility in DMF was therefore the first choice solvent for its HATU facilitated coupling to **12**. However, poor yields were observed due to incompleteness of the reaction. We anticipated that the reaction could benefit from longer reaction times and employed the more stable coupling reagent combination of EDC.HCl with HOBT to get yields of 40–55% for compound **17a** after prep-SFC purification. Cefaclor displayed better solubility and gave optimum couplings to **12** in acetonitrile with COMU in just two hours to afford **17b** in 50–70% yields.

**Scheme 3** General synthesis of NODASA- β -lactam antibiotic used in studies A. i = β -lactam, HOBt, EDC.HCl, DMF/DMSO, rt, 6–12 h or COMU, DIEA, ACN, rt, 2 h ii = anisole, TFA, DCE, rt, 2–6 h iii = *tert*-butyl bromoacetate, DIEA, DCE, 0–25 °C, 2–6 h iv = thioansole, TFA, rt, 3–6 h.

Once the β -lactams were coupled to the chelator to afford **17a** and **17b**, their solubility properties became similar. Therefore, the amine deprotection to afford **18a** and **18b** was similar to that of **13**, with the only difference being the reaction times. The reactions with **17a** took between two and three hours while **17b** required just one hour to complete, before affording compounds **18a** and **18b** in quantitative yields. Thereafter, compounds **18a** and **18b** were washed with diethyl ether, and alkylated similarly to **14**, with varying reaction times. Compounds **19a** and **19b** were purified using prep-SFC to give products with yields of 30–45%. Thereafter, **19a** and **19b** underwent hydrolysis with TFA to afford the desired compounds **20a** and **20b** respectively.

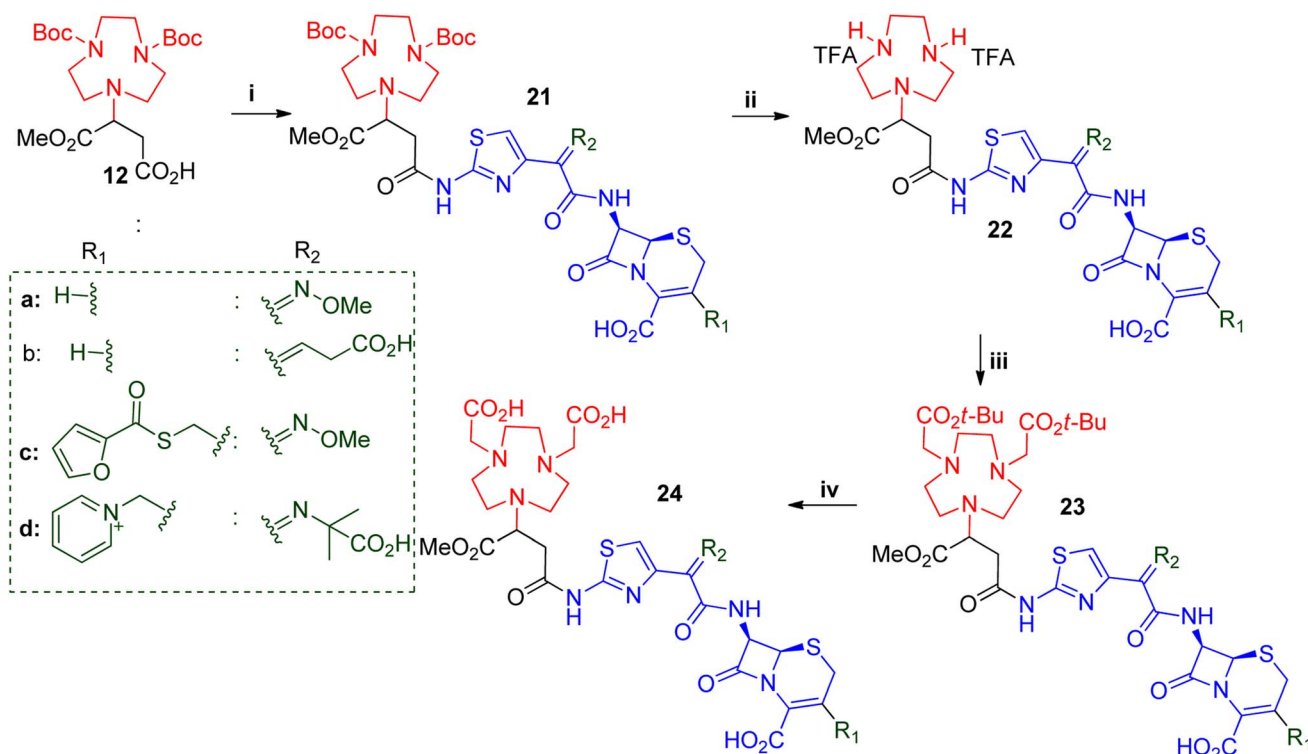
Another set of structurally related β -lactams was also explored, namely ceftizoxime, ceftibuten, ceftiofur and ceftazidime. These were used to synthesise compounds **24a–d**, respectively (Scheme 4). Solubility tests showed that all of the starting β -lactams were only applicable in DMF and DMSO. Compound **12** was used to couple to each of these β -lactams with EDC.HCl and HOBT in dry DMF/DMSO. Pure compounds **21a–d** were obtained after prep-SFC purification with yields ranging from 50–65%. Compounds **21a–d** were Boc-protected, using TFA with anisole as a scavenger in DCE. All of the reactions were completed in 1–2 hours with quantitative yields obtained after the TFA was removed over a stream of nitrogen gas and washed with diethyl ether. The crude salts of compounds **22a–d** were alkylated with *tert*-butyl bromoacetate to afford **23a–d**. The reactions were performed in DCM in

the presence of DIEA at 0 °C to room temperature and the times varied from 4–7 hours. The crude reactions were purified using SFC to afford **23a–d** with 30–40% yields. Subsequently, the *tert*-butyl groups of **23a–d** were cleaved using TFA to afford the desired compounds **24a–d** in 89–95% yields.

All six synthesised derivatives **20a–b** and **24a–d** (Fig. 3), were evaluated as MBLIs against *Escherichia coli* NDM-1 (ref. 39) and *Klebsiella pneumoniae* NDM,¹⁰ in combination with meropenem.

The data in Table 2 demonstrates that all of the evaluated compounds, **20a–b** and **24a–d**, successfully restored the activity of meropenem to MICs ranging from 0.06–0.5 mg L⁻¹ with inhibitor concentrations between 8 and 16 mg L⁻¹. When comparing the activity between the synthesised MBLIs, **24b** and **24c** showed the best activities.

All six derivatives, **20a–b** and **24a–d**, fully restored meropenem's efficacy (≤ 2 mg L⁻¹) according to the EUCAST breakpoints.⁴² These derivatives demonstrated similar activities when compared to the lead inhibitor in Table 1, *i.e.*, compound **9**, while the thiazole-containing compounds' (**24a–d**) MICs produced better results than the non-thiazole-containing compounds (**20a–b**). It is also noted that compounds **24a–d** have a longer distance between the chelator moiety and the lactam moiety than **20a** and **20b**. The binding free energies of the two families of compounds (**20** versus **24**) were previously calculated by our group using advanced molecular dynamics studies. In general, the system with the longer chain distance between the chelator and the lactam moieties was superior.^{36,37} It is also possible that a thiazole ring, which is present in



Scheme 4 General synthesis of NODASA- β -lactam antibiotic used in studies B. i = β -lactam, HOBT, EDC.HCl, DMF or DMSO, rt, 6–12 h ii = anisole, TFA, DCE, rt, 2–6 h iii = *tert*-butyl bromoacetate, DIEA, DCE, 0–25 °C, 2–6 h iv = thioansole, TFA, rt, 3–8 h.



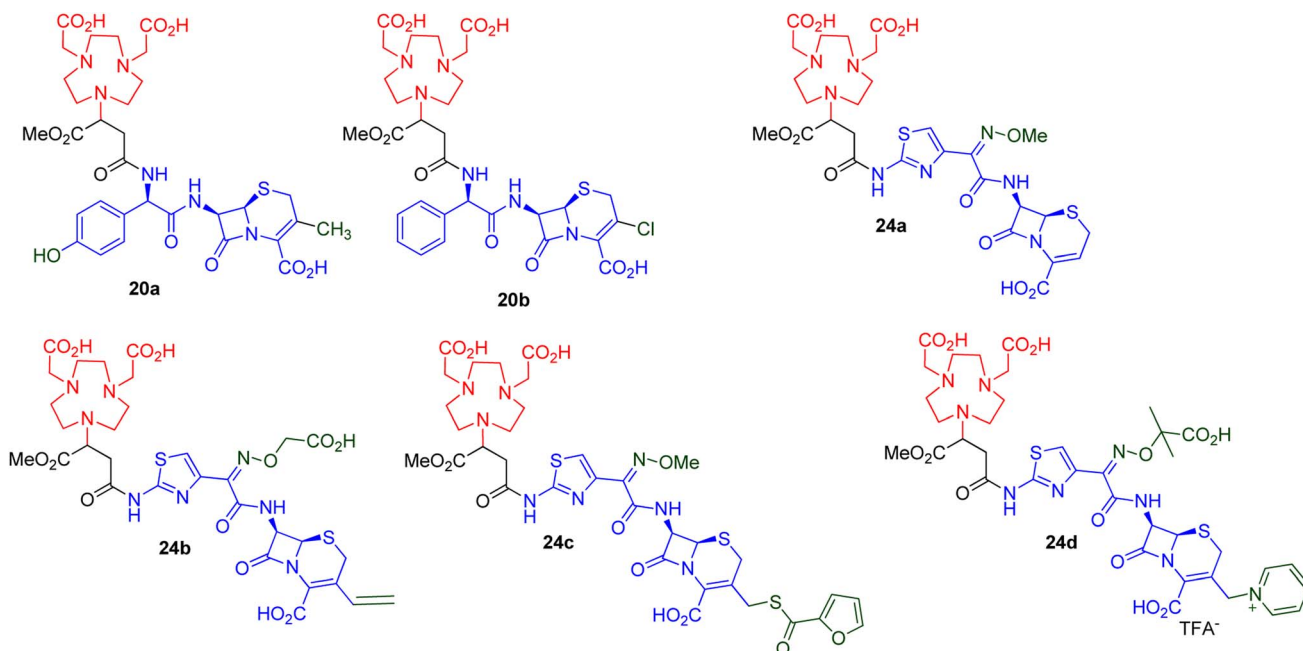


Fig. 3 Synthesized NODASA derivatives as MBLIs.⁴¹

compounds **24**, is necessary to enhance the MBL inhibition. This could be investigated further by trying different approaches to insert unsubstituted carbon chain linkers with varying chain lengths between compounds **20** or a structurally similar compound; a linker containing a phenyl group; or a linker containing a thiazole group, and comparing the activities of these compounds (beyond the scope of this manuscript).

Based on the synthetic viability from Table 2, compounds **24b** and **24c** were further screened for activity against pathogens harbouring VIM and IMP MBLs. Table 3 indicates that **24b** and **24c** restored the efficacy of meropenem against pathogens harbouring *bla*_{NDM-4}, *bla*_{VIM-1}, *bla*_{IMP-1} and *bla*_{IMP-8} genes, in addition to bacteria expressing *bla*_{NDM-1} genes (Tables 1 and 2).

Compounds **24b** and **24c** are therefore effective inhibitors of the most clinically relevant subclass, B1 MBLs.

The cytotoxicity using a human liver (HepG2) cell line was evaluated for **24b** and **24c**. From the cytotoxicity data, IC₅₀ values of 42.34 mg mL⁻¹ and 51.83 mg mL⁻¹ were obtained for **24b** and **24c**, respectively (Fig. 4). In general, compound **24b** reduced cell viability compared to the control, with 10 µg mL⁻¹ being significant; compound **24c** increased cell viability compared to the control, with 200 µg mL⁻¹ being significant (Fig. 4A). The methyl thiazol tetrazolium (MTT) assay measures cellular metabolic output and compound **24c** increases metabolic output. It is known that cancer cells utilise anaerobic glycolysis for ATP generation and minimises mitochondrial

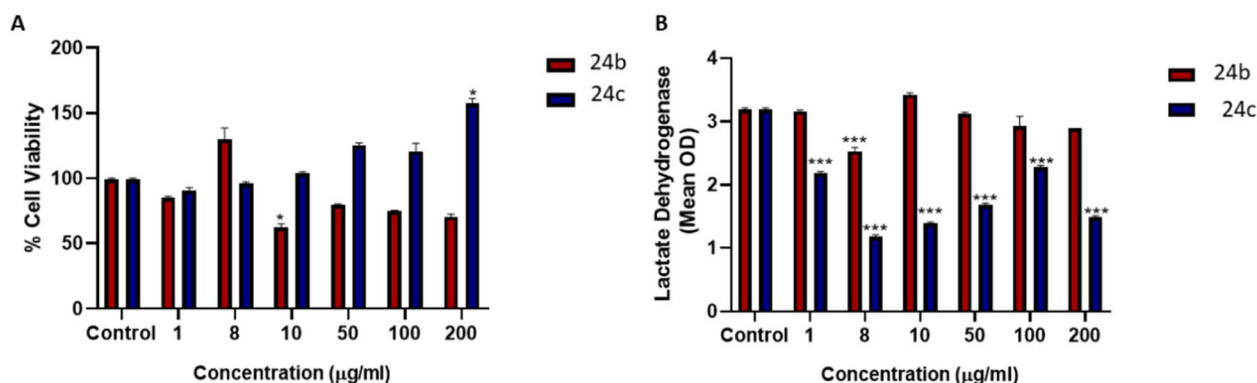


Fig. 4 Cell viability studies conducted on HepG2 cells using varying concentrations of **24b** and **24c**. (A) Cell viability was reduced at 1 µg L⁻¹ and 10–200 µg L⁻¹ with proliferation observed at 8 µg L⁻¹ for **24b**. **p* < 0.5 and ****p* < 0.01, relative to the control. **24c** exhibited a dose-dependent increase in the cell viability of HepG2 cells. However, cell viability was only significantly altered at 200 µg L⁻¹ **p* < 0.05 relative to control. (B) LDH levels were significantly reduced at 8 µg L⁻¹ and remained unaffected at 1 µg L⁻¹ and 10–200 µg L⁻¹, indicating **24b** does not induce necrosis in HepG2 cells after exposure. Similarly, **24c** also did not induce necrosis, as LDH was significantly reduced at all concentrations. ****p* < 0.001 relative to control. All assays were conducted in triplicate.

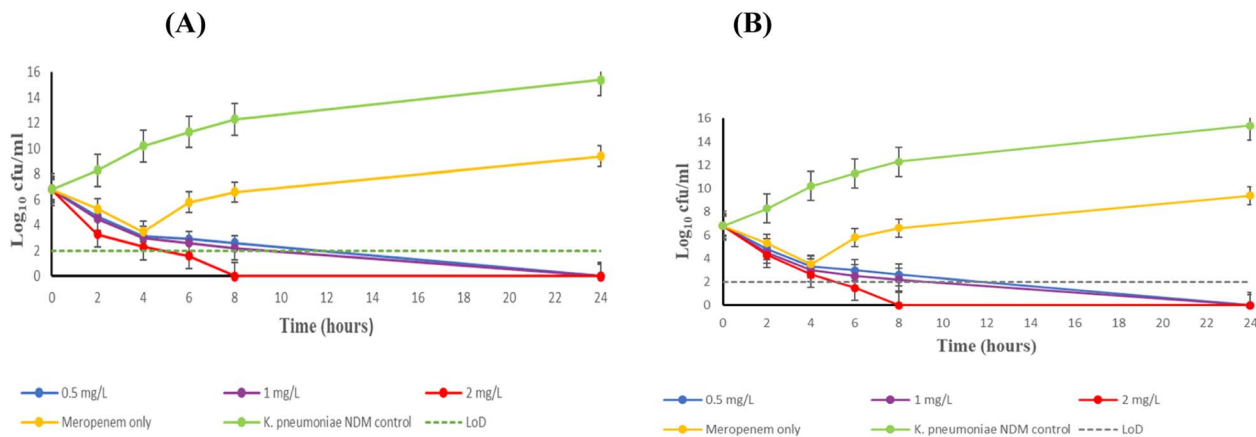


Fig. 5 Time-kill kinetic study of *K. pneumoniae* NDM. (A) The combination of **24b** + meropenem. (B) The combination of **24c** + meropenem. All assays were conducted in triplicate. The combination of 32 mg L^{-1} **24c** + 0.5 mg L^{-1} meropenem produced $p < 0.0441$ when compared to monotherapy and $p < 0.0013$ for no therapy.

output as this will activate apoptosis. The lactate dehydrogenase (LDH) assay further confirmed the non-toxic properties of both compounds **24b** and **24c** with the former significantly decreasing LDH membrane leakage at $8 \mu\text{g mL}^{-1}$ (Fig. 4B). Compound **24c** significantly decreased LDH membrane leakage at all evaluated concentrations (Fig. 4B). The chelator concentrations selected for drug susceptibility screenings ($\leq 32 \text{ mg L}^{-1}$) were below the IC_{50} 's, which allowed us to proceed safely with biological evaluation.

Fig. 5 represents the effect exhibited by chelator **24b/24c** + meropenem over 24 hours against the virulent *K. pneumoniae* NDM-expressing bacteria, respectively. Both chelators displayed excellent bactericidal activity against this strain, with meropenem concentrations of 0.5, 1 and 2 mg L^{-1} . A $> 3 \log_{10}$ decrease in the bacterial load was observed 6 hours post-inoculation for combination therapy, relative to meropenem monotherapy. Although a sharp decrease of bacteria was observed at 4 hours post-inoculation with meropenem monotherapy, this effect could not be sustained and *K. pneumoniae* NDM continued to proliferate. This was expected, as the chelator is essential to potentiate the activity of meropenem to susceptible levels. 32 mg L^{-1} of compounds **24b** and **24c** were needed to achieve complete bactericidal activity with 0.5 mg L^{-1} meropenem. Increasing the meropenem dose to 2 mg L^{-1} resulted in faster sterilizing activity from 8 hours post-inoculation, without any regrowth in a 24 hour period.

To understand the kinetics of MBLs; VIM-2 and IMP-1, enzyme analyses were undertaken utilizing Michaelis–Menten conditions and Lineweaver–Burk plots (Fig. S1†). The kinetics parameters of both MBLs (Table 4), shared a similarity with regards to the V_{max} and catalytic turn over numbers. The resultant K_{m} values, however, were not similar, demonstrating that VIM-2 was a more catalytically efficient enzyme between the two MBLs. This infers that pathogens expressing VIM-2 can neutralize antibiotics even at low concentrations, much faster as compared to IMP-1, that requires a larger antibiotic concentration to reach V_{max} and produce optimal hydrolysis. Therefore, much higher doses of the antibiotic may need to be

administered in treatment regimens to eradicate infectious disease caused by VIM expressing bacteria. Conversely these required doses are not safe to administer and have therefore contributed to the soaring resistance rates.

To further assess the potential of **24b/24c**, inhibition of enzyme kinetics was performed, to generate the compounds' inhibitory constant (K_{i}). The activity plots of **24b/24c** followed a graphical pattern characteristic of non-competitive inhibitors (Fig. S2 and S3†). The \ln (% residual activity) values of each enzyme decreased over time, with the highest inhibitor concentration having the lowest residual activity values. The K_{obs} values were observed to increase as the inhibitor concentration increased. The K_{obs} values were deduced from the residual activity plot and the K_{i} values (Table 5) were determined by interpolation of the K_{obs} versus inhibitor concentration plot.

Table 5 conveys that stronger inhibition was exhibited by **24b** against both IMP-1 and VIM-2, as compared to **24c**. This finding

Table 4 The steady-state kinetic constants of IMP-1 and VIM-2 with nitrocefins as substrate

Kinetics parameter	IMP-1	VIM-2
V_{max} ($\mu\text{mol min}^{-1}$)	2.41 ± 0.19	2.62 ± 0.11
K_{m} (μM)	60.42 ± 0.54	4.08 ± 0.04
k_{cat} (s^{-1})	0.40	0.44
$k_{\text{cat}}/K_{\text{m}}$ ($\mu\text{M}^{-1} \text{s}^{-1}$)	6.64×10^{-3}	10.70×10^{-2}

Table 5 The inhibition kinetics parameters for **24b/24c** against VIM-1 and IMP-1^a

Enzymes (MBLs)	24b	24c
	K_{i} (μM)	K_{i} (μM)
IMP-1	23.87 ± 0.00	74.44 ± 0.00
VIM-2	0.85 ± 0.00	1.87 ± 0.00

^a All the data is shown as means \pm SD, $n = 3$.



concur with the results of the antimicrobial susceptibility tests in which **24b** achieved a 1–4 2-fold dilution decrease in the meropenem MIC, whilst utilizing the same inhibitor concentration for both **24b** and **24c** (Table 3). Furthermore, compound **24b** and **24c** displayed stronger inhibition towards VIM-2 as compared to IMP-1. Compounds **24b** and **24c** were superior in potency to predecessor compound BP1 ($K_{iapp} = 24.8 \mu\text{M}$).³⁶

Conclusion

The study has shown that β -lactam antibiotics covalently linked to cyclic amino acidic zinc chelators can potentially re-sensitize meropenem and target metallo- β -lactamase resistance in *E. coli* NDM-1 and *K. pneumoniae* NDM. Between 8–32 mg L⁻¹ of compounds, **6**, **9**, **16**, **20a–b** and **24a–d** were able to restore *E. coli* NDM-1 and *K. pneumoniae* NDM sensitivity towards meropenem at MICs ranging from 0.06–2 mg L⁻¹. Subsequent biological evaluation of the potent **24b** and **24c** revealed the compounds to be safe to administer, as neither cell death nor reduced cell viability was observed at the concentrations investigated herein. An added advantage of compound **24c** (increases metabolic output and decreases LDH leakage) will aid eukaryotic cells to increase ATP production and help fight bacterial insult. Time-kill studies demonstrated that compounds **24b** and **24c** produced sterilizing activity at 24 hours post-inoculation, utilizing a minimal meropenem dose of 0.5 mg L⁻¹. Bactericidal activity was achieved faster (from 8 hours post-inoculation) by increasing the meropenem dose to 2 mg L⁻¹. Enzyme inhibition studies indicated **24b** and **24c** are non-competitive, potent inhibitors of VIM-2. The overall findings of this study are that the novel series of β -lactam MBLIs reported herein, are potent, viable, and non-toxic candidates, with promising potential. Our research group is further exploring the scope of this class of MBLIs.

Conflicts of interest

There are no conflicts to declare.

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