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Development and optimization of natural deep eutectic solvent-based dispersive liquid–liquid microextraction coupled with UPLC-UV for simultaneous determination of parabens in personal care products: evaluation of the eco-friendliness level of the developed method

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Dispersive liquid–liquid microextraction (DLLME) combined with ultra-high performance liquid chromatography–diode array detector (UHPLC-DAD) method has been developed and validated for the determination of parabens in personal care products. In this study, a natural deep eutectic solvent (NADES) composed of menthol and formic acid at a molar ratio of 1:2 was prepared and used as an extraction solvent. The influencing variables on the extraction efficiency such as extraction solvent type and volume, composition of NADES, salt addition, vortex and centrifugation time were investigated. The proposed method exhibited good linearity with determination coefficients of ≥ 0.9992 . The relative recoveries for the studied analytes ranged from 82.19 to 102.45%. Limits of detection and limits of quantification were in the range of 0.17–0.33 ng mL⁻¹ and 0.51–0.99 ng mL⁻¹, respectively. To evaluate the applicability of the developed method, it was successfully applied to determine four parabens in personal care products. Additionally, the eco-friendliness level of the presented method was evaluated using Eco-Scale Assessment, Green Analytical Procedure Index and Analytical GREenness metric. The developed method is simple, environmentally friendly and cost effective and it could be employed for determination of parabens in personal care products without harming the environment.

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1. Introduction

Parabens, esters of *p*-hydroxybenzoic acid, are a family of organic compounds that are widely used as preservatives for pharmaceutical products, food and cosmetics. The most commonly used parabens are methyl paraben (MP), ethyl paraben (EP), propyl paraben (EP) and butyl paraben (EP). Because of their inexpensive cost, broad spectrum activity and chemical stability over a wide pH range, parabens are used singly or in a combination.¹ Human are exposed to parabens on a daily

basis from various sources and it is well documented in the literature that parabens may pose a potential risk to human health by being linked to carcinogenic and estrogenic threats.^{2,3} It has been found that there is an association between exposure to parabens and the incidence of health adverse effects such as hearing loss,⁴ inflammatory bowel disease,⁵ chronic diarrheal symptoms and allergen sensitization⁶ and endocrine disrupting effects.⁷ In light of this, the European Union set a maximum concentration of (0.4% w/w) for individual parabens and a limit of (0.8% w/w) for total parabens in commercial products.⁸

A key step in the analytical processes is the selection of appropriate sample preparation technique because it reflects method accuracy and precision. Different sample preparation techniques have been introduced for the determination of parabens in different matrices including liquid–liquid extraction, dispersive liquid–liquid microextraction (DLLME), solid phase extraction and solid phase microextraction.^{9–12}

DLLME is a sample preparation technique that is characterized by its speed, ease of use and high extraction efficiency. However, the main drawback of conventional DLLME is the use

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of toxic chlorinated solvents such as chloroform and carbon tetrachloride as an extractant solvents.¹³ Greening pharmaceutical analysis has attracted growing interest aiming at minimizing the negative environmental impact and enhancing analysts' health and safety. In this regards, more benign solvents such as ionic liquids, supercritical fluids and deep eutectic solvents (DESs) have been introduced as green alternatives in DLLME.^{13,14} Recently, DESs have attracted great interest in different fields as extraction solvents because their production is simpler, less toxic, more affordable and biodegradable.^{15–17} DESs are prepared by mixing hydrogen bond donor (HBD) and hydrogen bond acceptor (HBA) at proper molar ratios forming hydrogen bond interactions.¹⁴ DESs exhibit much lower melting point than those of either individual components.¹⁴

Several studies have been reported for the utilization of various DESs in combination with DLLME for determination of parabens in different matrices such as food,^{18–20} pharmaceutical samples,²¹ cosmetics^{22–25} and environmental waters.²⁶ Most of DES-DLLME methods reported for determination of parabens in cosmetics utilized choline chloride *e.g.*^{22,24} or tetrabutylammonium chloride *e.g.*²⁵ However natural deep eutectic solvents (NADES) containing natural monoterpenes such as menthol are considered proper solvents for extraction due to its low water solubility and low price,^{27,28} their use in determination of parabens in cosmetics is still limited. Hence, this study aimed to develop a fast, simple, cost-effective and environmentally friendly menthol-based DES-DLLME method, followed by UHPLC-DAD for simultaneous determination of parabens in cosmetic samples. Additionally, three different metrics including Analytical Eco-Scale (AES),²⁹ Green Analytical Procedure Index (GAPI)³⁰ and the Analytical GREENness (AGREE) metric³¹ were used to assess the eco-friendliness profile of the developed method.

2. Material and methods

2.1. Chemicals, reagents and materials

Standards of four parabens including methylparaben (MP), ethylparaben (EP), propylparaben (PP) and butylparaben (BP) with purity of $\geq 98.0\%$ were supplied by Sigma-Aldrich (Steinheim, Germany). Menthol, formic acid, acetic acid, octanoic acid, oleic acid, decanoic acid and dodecanoic acid were purchased from Sigma-Aldrich (Steinheim, Germany). Stock solutions of parabens were prepared in methanol at a concentration of 1000 mg L^{-1} . Working standard solutions of different concentrations were prepared by dilution of the stock solutions in methanol and stored at -20°C until use. Methanol, ethanol and acetonitrile were of HPLC grade and supplied from Merck (Darmstadt, Germany). Samples were filtered using $0.2 \mu\text{m}$ PolyTetraFluoroEthylene (PTFE) – Thomson SINGLE StEP eXtreme filter vials (Restek, USA). Ultrapure water (Milli-Q, ultrapure grade) was obtained by a water purification system (ELGA, High Wycombe, UK).

2.2. Instrumentation

Chromatographic analysis of the investigated parabens was performed on UltiMate 3000 XRS UHPLC (Thermo Scientific

Dionex, Germany) equipped with an autosampler, a dual gradient pump, a thermostated column compartment and a diode array detector (DAD). The analysis was performed on Acquity BEH C18 column ($2.1 \text{ mm} \times 30 \text{ mm} \times 1.7 \mu\text{m}$) with a VanGuard precolumn (Waters, USA). The injection volume was $5 \mu\text{L}$ and the column temperature was set at 40°C . The flow rate was 0.6 mL min^{-1} . The gradient elution was utilized with a mobile phase composed of ethanol (B) and water (A) as follows: 0–8 min (5 to 100%, B), 8–10 min (5%, B), followed by 0.5 min post time for equilibration of the column. The quantitative analysis of parabens were performed at the wavelength of 254 nm .

For characterization of the prepared DES, the attenuated total reflection (ATR)-Fourier transform infrared (ATR-FTIR) spectra were recorded on Thermo Nicolet 50 spectrometer (USA) equipped with a DTGS detector and attenuated total reflectance (ATR) accessory. The spectra were measured from 4000 to 600 cm^{-1} . For recording the Proton Nuclear Magnetic Resonance (^1H NMR) spectra of the prepared DES, a 300 MHz Bruker Nuclear Magnetic Resonance (NMR) spectrometer (Bruker, Switzerland) was used. Dilution of samples were performed with deuterated dimethyl sulfoxide ($\text{d}_6\text{-DMSO}$).

2.3. Sample collection and preparation

Fifteen cosmetic samples including shampoos ($n = 4$), hair conditions ($n = 2$) and creams ($n = 7$) antiperspirant ($n = 2$) were collected from local markets in the Eastern province, Saudi Arabia. Each sample was accurately weighed (0.5 g) and transferred into a 50 mL polypropylene centrifuge tube, then 2 mL of methanol was added. The mixture was vortexed for 2 min , sonicated for 5 min and subjected to centrifugation for 10 min at 3500 rpm . The supernatant aqueous phase was then collected and diluted up to 25 mL with water. The prepared samples were vortexed and filtered with $0.45 \mu\text{m}$ membrane filter before subjecting to the developed DLLME procedure.

2.4. Preparation of deep eutectic solvents (DESs)

DESs were prepared by dissolving menthol as HBA and acids (formic acid, acetic acid, octanoic acid, oleic acid, decanoic acid and dodecanoic acid) as HBDs at proper molar ratio. The prepared DESs were stirred continuously for 15 min at 60°C until clear transparent solutions formed. After cooling to room temperature, the DESs were stored in a desiccator until required for DLLME.

2.5. Dispersive liquid–liquid microextraction (DLLME) procedures

Under optimum conditions, 5 mL of the sample solution was transferred into a 15 mL polypropylene centrifuge tube, then 300 mg of NaCl was added and vortexed till completely dissolved. Then, $80 \mu\text{L}$ of DES was added and vortexed for 3 min . The cloudy solution performed and centrifuged at 3500 rpm for 10 min . After removal of the lower aqueous phase, the remaining extraction phase was diluted with $120 \mu\text{L}$ methanol and then subjected to vortex and filtered using $0.2 \mu\text{m}$ of nano-filter vials before injected into the UHPLC-DAD system. Further



dilution in ethanol was performed in case the paraben concentration detected in real samples exceeded the upper linearity range.

Extraction recovery (ER%) and enrichment factor (EF) was used to evaluate the extraction efficiency using the following equations.

$$ER\% = \frac{C_{DES} \times V_{DES}}{C_{Sample} \times V_{Sample}} \times 100$$

Here, C_{Sample} and C_{DES} are the initial analyte concentrations in the sample solution and the extracted DES phase, respectively.

$$EF = \frac{C_{DES}}{C_{Sample}}$$

Here, V_{Sample} and V_{DES} are the volumes of the initial sample solution and extracted DES, respectively.

3. Results and discussion

Different extraction parameters should be considered in order to optimize the DLLME method. These parameters include type and volume of the extraction solvent (DES), molar ratio of HBA/HBD, vortex time, sonication time, centrifugation time, salt addition, solution pH and addition of disperser solvents. All the experiments were performed in triplicate using 5 mL of spiked samples (2 mg L^{-1}). Extraction recovery (%R) or enrichment factor (EF) were used to represent the extraction efficiency.

3.1. Selection of the extraction solvent (DES)

An extraction solvent should have important properties such as being immiscible in water, having a high affinity for analytes, and being a liquid at room temperature.³² In this study, six hydrophobic DESs were investigated (Table 1) to choose the most appropriate natural DES. At molar ratio of (1 : 1), five DES mixtures with homogenous liquid retained upon cooling were selected for further extraction recovery comparison (DES 1–5). The highest extraction recovery of MP (%), EP (%) and PP (%) was achieved by DES-1 (menthol and formic acid), whereas DES-3 exhibited the highest recovery for BP (%) (Fig. 1). Other DESs (DES-2, DES-4, and DES-5) showed no significant improvement in extraction recovery for the analytes studied. As a result, the DES-1 was chosen for further extraction procedures.

Table 1 The composition of DESs investigated

Abbreviation	HBA	HBDs	Ratio (HBA : HBD)	Observation
DES-1	Menthol	Formic acid	1 1	Homogenous
DES-2		Acetic acid		
DES-3		Octanoic acid		
DES-4		Oleic acid		
DES-5		Decanoic acid		
DES-6		Dodecanoic acid		Heterogenous

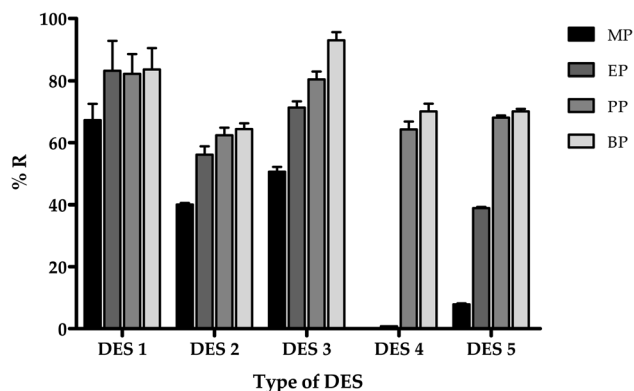


Fig. 1 Effect of the type of DESs on the extraction recovery for the studied parabens.

3.2. Characterization

The formation of DESs is driven by the intermolecular hydrogen bonding between menthol and formic acid. Hydrogen bond interactions in DES are widely explored using FTIR and ^1H -NMR.³³ The FTIR spectra of DES-1, formic acid and menthol are shown in Fig. 2. The absorption peak at 1685 cm^{-1} in formic acid belongs to $\text{C}=\text{O}$ and was shifted to 1716 cm^{-1} in DES-1 spectra. The presence of very broad band at 2945 cm^{-1} belongs to carboxylic $-\text{OH}$ in formic acid. The characteristic peak of hydroxyl group in menthol at 3237 cm^{-1} was attributed for free hydroxyl group. The association of this $-\text{OH}$ in hydrogen bonding with formic acid and the change in the vibration indicated the formation of DES-1.^{18,26}

Menthol has secondary alcohol in an electron rich cyclohexane structure and is expected to be a good hydrogen bond acceptor and formic acid would act a strong hydrogen bond donor in the binary complex. ^1H NMR was performed on the reference compounds alone and on their expected complex and the spectra were compared and interpreted. Fig. 3 displays the ^1H NMR spectra of formic acid, menthol, and DES-1. As shown, broadness of the COOH peak of formic acid was noted in the complex spectrum which would suggest hydrogen bonding and change in the environment of the carboxylic acid proton. The alpha proton in menthol ($\text{CH}-\text{OH}$) appears downfield at 4.6 MHz in the complex spectrum instead of 3.1 MHz in the reference structure which is due to the hydrogen bonding of oxygen that makes it more electron withdrawing. Similarly the alcoholic proton is becoming more deshielded and is shifted to 8.2 MHz again due to the involvement of the alcohol oxygen in hydrogen bonding as hydrogen acceptor with formic acid.

3.3. Effect of molar ratio

Different molar ratios of the selected DES (menthol-formic acid) were investigated (Fig. 4). As shown in the figure, the extraction efficiency for all analytes showed a similar trend for all prepared DESs. For all target analytes, it was found that the ratio of 1 : 2 provides the highest extraction efficiency. Therefore, the ratio of 1 : 2 (menthol-formic acid) was chosen for further steps.



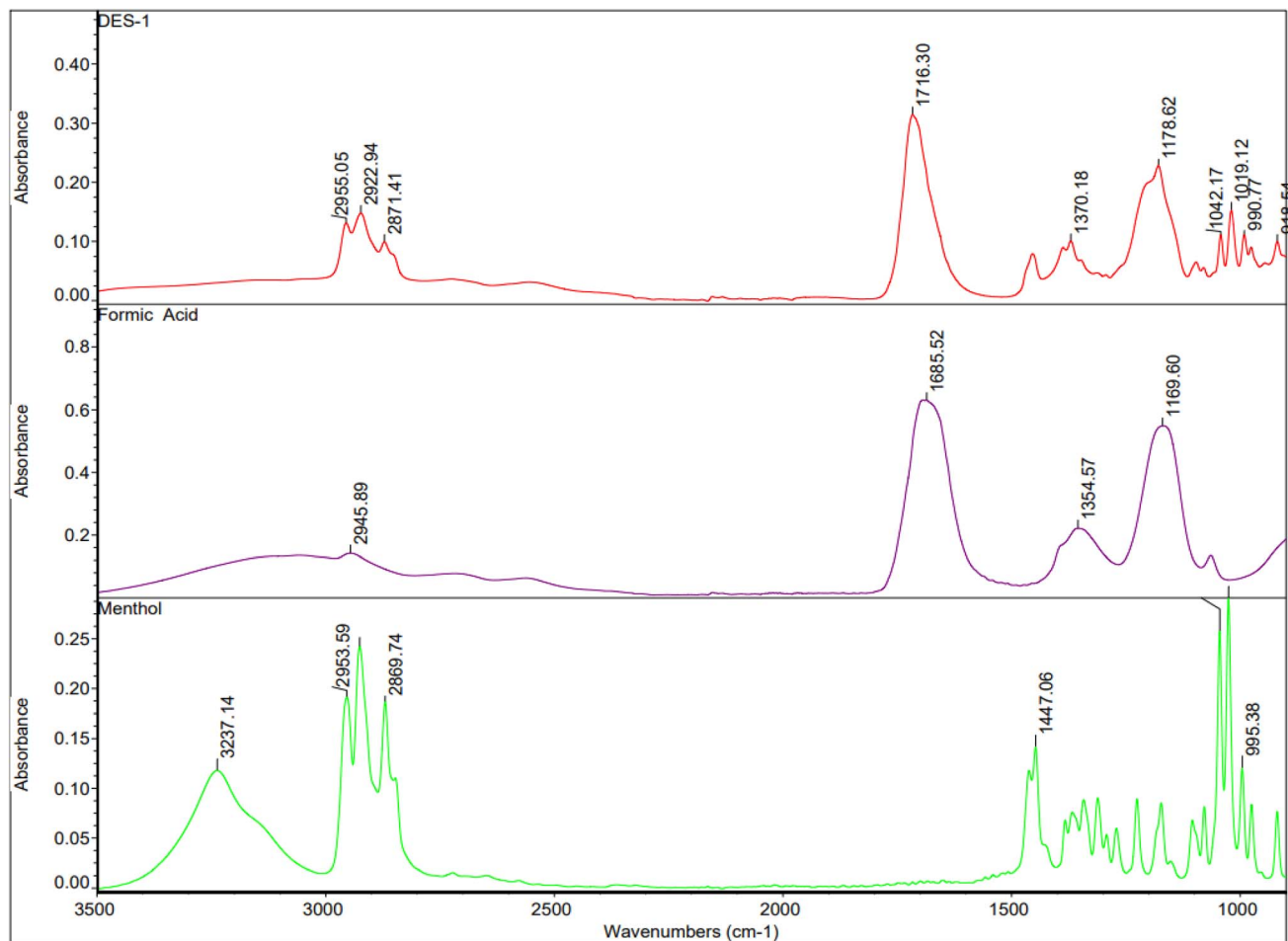


Fig. 2 FTIR spectra of the prepared DES-1 and its individual components (menthol and formic acid).

3.4. Effect of DES volume and sample volume

After selecting the best DES and the optimal molar ratio, the effect of DES volume between 80 μL and 300 μL on the extraction recovery of analytes were assessed as shown in Fig. 5a. The optimum extraction recovery for all studied parabens was achieved by using 80 μL of DES-1. Thus, it was selected for subsequent experiments. Also, the effect of sample volume on extraction efficiency was evaluated. Different sample volumes between 2 mL to 15 mL were investigated as shown in Fig. 5b. The enrichment factors (EFs) for all analytes were found to be increased to their maximum at a volume of 5 mL and then decreased by increasing sample volume. This could be attributed to the reduction in extraction recovery with higher sample volumes. Therefore, 5 mL was used as an optimum sample volume.

3.5. Effect of pH

For the effect of pH on the extraction performance, previous studies reported that pH has a negligible effect on the extraction recovery of parabens,^{18,26} therefore the pH was not adjusted in this study.

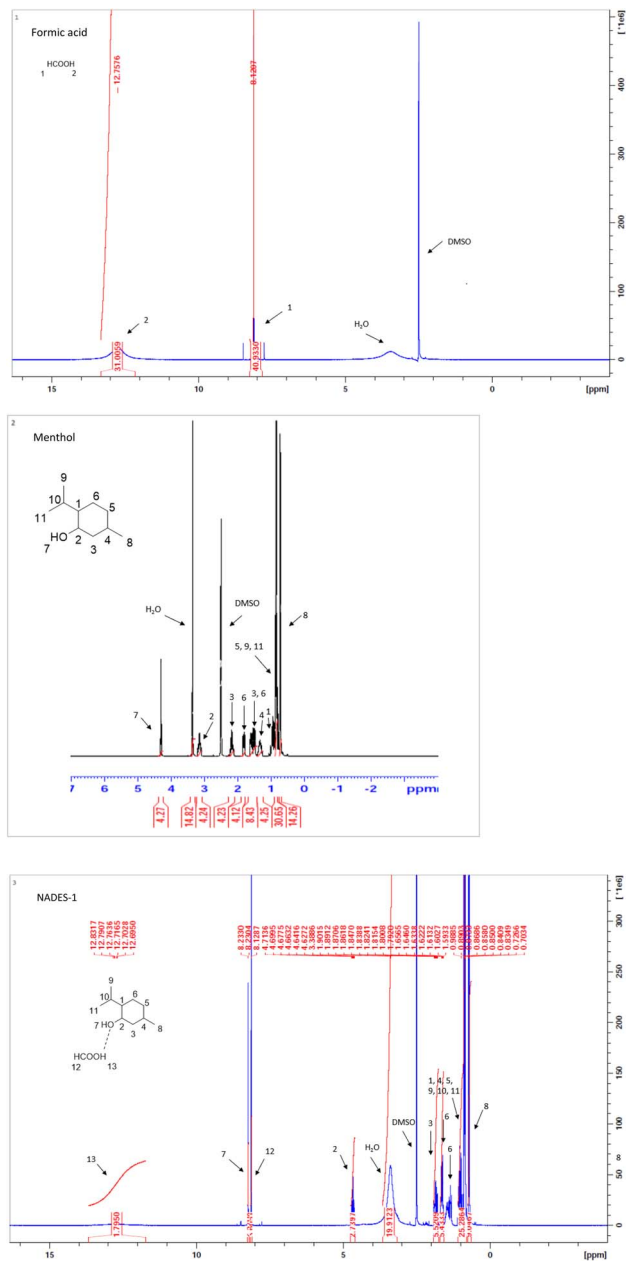
3.6. Effect of vortex, sonication and centrifugation time

The importance of the vortex agitation to increase partitioning of the targeted compounds from the extraction solvent into

aqueous phase and enhancing the extraction performance has been reported in several DLLME studies.^{34–36} In this study, different vortex time from 0 to 300 s were investigated at fixed rotational speed (2800 rpm) (Fig. 6a). The results show that a significant increase in the extraction efficiency achieved at vortex time of 30 s was gradually decreased by increasing the vortex time from 60 to 120 s. The optimal vortex time was found to be at 180 s. Sonication has also been reported to increase the solvent extractability and improve the mass transfer between two phases, thus affecting the extraction efficiency.³⁷ In this study, no significant improvement in extraction efficiency was observed by sonication (0 min–10 min) as shown in Fig. 6b. As a result, it was not used in any further experiments.

In DLLME, centrifugation could improve the two phase separation and shorten the sample preparation time. Therefore, different centrifugation times (5 to 30 min) were investigated as illustrated in Fig. 5c. The obtained results showed a significant improvement in the extraction efficiency at 10 min. By increasing the centrifugation time (>10 min), the extraction performance was kept constant. Therefore, 10 min was selected as the optimum centrifugation time.





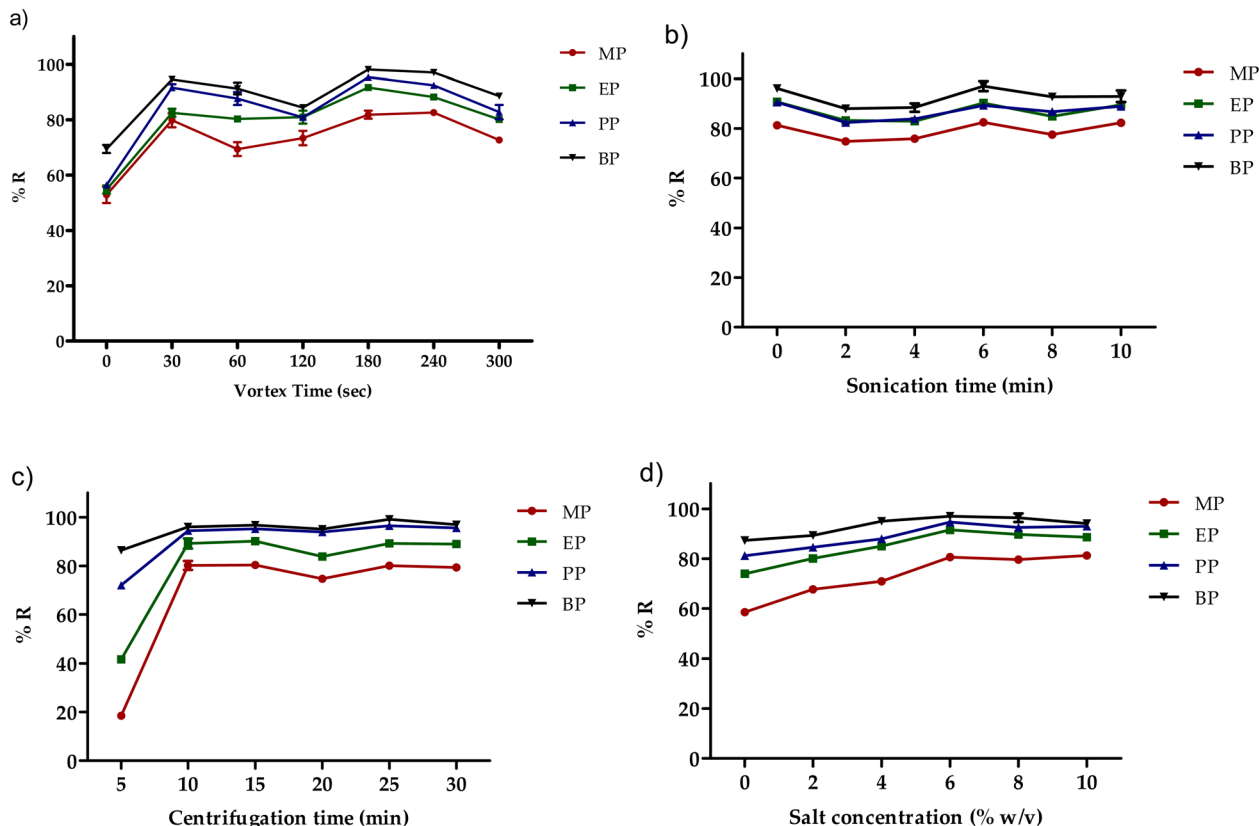


Fig. 6 Effect of (a) vortex time, (b) sonication time, (c) centrifugation time and (d) salt addition on the extraction recovery of the studied parabens.

illustrated in Fig. 7, no significant improvement in the extraction recovery of the targeted parabens was observed when dispersive solvents were used. Therefore, the experiments were performed without using any disperser solvents.

3.9. Optimization of the chromatographic separation

Taking into account the principles of green analytical chemistry, chromatographic separation was optimized to obtain the maximum separation efficiency, sensitivity and selectivity. In this regard, a short, narrow bore column packed with sub-2 μm

particles was utilized to achieve rapid analysis while consuming less solvent and producing less waste. To adhere with the principles of green analytical chemistry, ethanol was used as a green biodegradable organic solvent in the mobile phase. Gradient mode was also employed to enhance resolution, peak shape, and to ensure complete elution of the hydrophobic DES. To achieve the best resolution in the shortest possible analysis time, various gradient systems were tried. Under the optimised condition, DES-1 and all studied analytes were completely eluted in 4.7 min (Fig. 8).

3.10. Analytical performance of the developed method

Different validation parameters were investigated to demonstrate the suitability of the developed method including linearity, specificity, limit of detection (LOD), limit of quantification (LOQ), intra-day and inter-day precision and accuracy.

The UHPLC-DAD chromatograms of the spiked and unspiked samples (Fig. 8) showed that the investigated parabens are entirely separated from other interfering components and revealed the absence of matrix interferences, reflecting a good specificity of the developed DLLME method. Calibration curves were evaluated by using the peak areas of spiked samples at five different concentration levels of MP, EP, PP and BP ($n = 3$). The calibration curves exhibit a good linearity over the range of 20–4000 ng mL^{-1} with satisfactory correlation coefficients ($r^2 \geq 0.9992$). The LODs, based on signal-to-noise

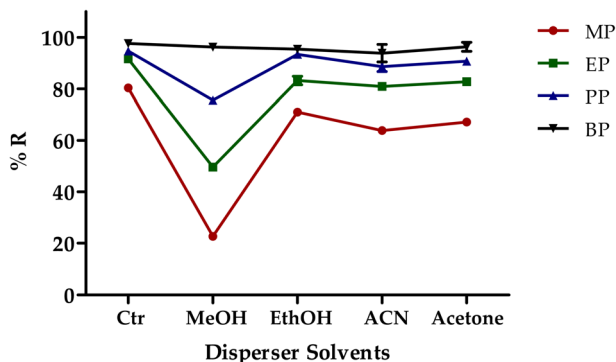


Fig. 7 Effect of the dispersive solvents on the extraction recovery of the studied parabens.

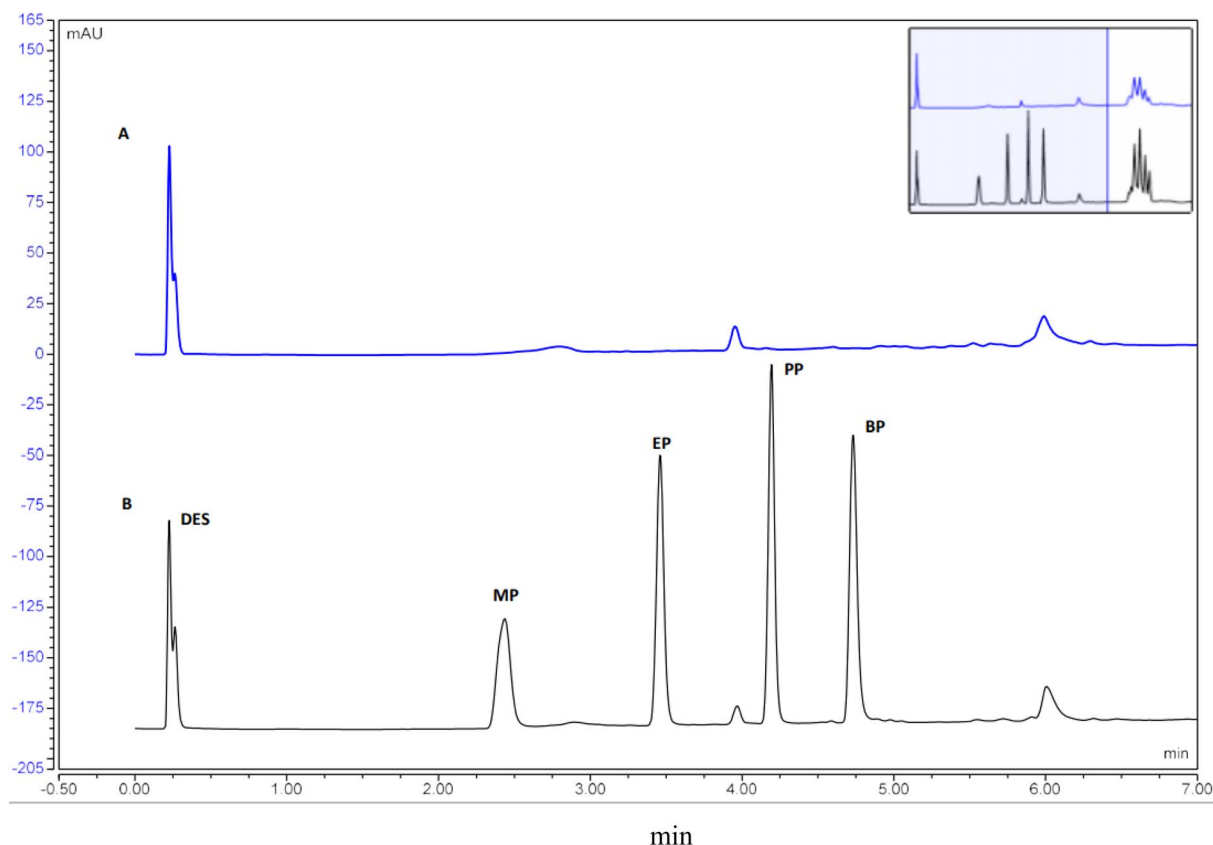


Fig. 8 UHPLC-DAD chromatograms of (A) non-spiked sample extracted by DLLME and (B) a spiked sample extracted by DLLME.

ratio of 3 were in the range of 0.17 to 0.33 ng mL⁻¹. The LOQs, based on signal-to-noise ratio of 10 were in the range of 0.51 to 0.99 ng mL⁻¹.

Precision was evaluated at three concentration levels for each analyte (40 ng mL⁻¹, 2000 ng mL⁻¹ and 4000 ng mL⁻¹). The enrichment factors (EFs) for all studied analytes were in the range of 52 to 62 (Table 2). The intra-day and inter-day precisions of the developed method were tested by performing three

replicates of the spiked samples within a day and on three consecutive days, respectively. The relative recoveries were in the range of 82.19–102.45%. The RSDs were ≤0.43% and 3.33% for intra-day and inter-day precision, respectively which indicated good precision of the developed method. The analytical performance of the developed DES-based DLLME method is presented in Table 2.

Table 2 Analytical performance of the developed DES-based DLLME method

Analyte	Added (ng mL ⁻¹)	Linearity equation	r^2	LOD (ng mL ⁻¹)	LOQ (ng mL ⁻¹)	ER	Intra-day		Inter-day	
							R%	RSD%	R%	RSD%
MP	40	$y = 0.0099x + 0.3693$	0.9997	0.17	0.51	52	83.12	2.60	82.19	1.83
	2000						82.57	0.63	82.35	1.00
	4000						82.97	1.04	84.75	3.33
EP	40	$y = 0.0149x + 0.2104$	0.9993	0.20	0.60	58	91.07	0.95	92.95	0.43
	2000						92.55	1.13	91.42	2.23
	4000						93.06	1.35	91.61	1.73
PP	40	$y = 0.0173x - 0.092$	0.9992	0.22	0.65	60	95.73	1.83	96.84	1.87
	2000						96.63	1.85	94.67	2.07
	4000						95.14	0.57	95.13	1.30
BP	40	$y = 0.0172x + 0.1379$	0.9993	0.33	0.99	62	102.45	2.19	99.62	0.93
	2000						98.93	1.62	99.47	2.14
	4000						99.11	0.43	101.13	1.55



3.11. Application to real samples

Under the optimized extraction conditions, the developed DES-based DLLME method was employed to determine the target parabens in 15 different personal care products from the local market. Parabens were found in 40% of the analyzed samples (Table 3). MP was the most frequently detected paraben with concentrations ranged from 18.3 to 702.2 mg kg⁻¹. EP and PP were found in two samples (221.9 and 622.5 mg kg⁻¹) and four samples (0.84 to 341.6 mg kg⁻¹), respectively. BP was found in only one sample at a concentration of 0.17 mg kg⁻¹. These findings are in an agreement with other previous studies reported for determination of parabens in personal care products *e.g.*¹²

3.12. Comparison with other previous methods

A comparison with other reported DES-DLLME methods for determination of parabens was performed to assess the performance of the developed method. As illustrated in Table 4, the analytical performance of the proposed method is comparable with the reported methods in terms of linearity, accuracy and limits of detection. The main advantage of the developed method is the rapidity and simplicity. As presented in Table 4, the run times for all reported methods ranged from 10–35 min, compared to the 4.8 min achieved in this study. The proposed method is also notable for its minimal waste production and low solvent consumption. Additionally, the presented method utilized monoterpenes (*e.g.* menthol) for sample extraction which are low in water solubility, toxicity, and cost. Moreover, the proposed method was successfully applied for determination of parabens in cosmetic products from different types including hair care, skin care and body care products. In conclusion, the developed method outperforms other reported methods with comparable analytical performance in terms of speed and cost.

3.13. Assessment of the eco-friendliness level of the developed method

To assess the eco-friendliness level of the developed method and evaluate its effects on the environment, safety and human health, three assessment tools were utilized. These tools included Analytical Eco-Scale (AES),²⁹ Green Analytical Procedure Index (GAPI)³⁰ and Analytical GREENness metric.³¹

The AES allocates penalty points (PPs) to many parameters such as reagents (amounts and hazards), energy consumption, occupational hazard and waste generated that lower the method's greenness. These PPs are then subtracted from 100 and the remaining points are used to classify the method's level of greenness as excellent (>75), acceptable (>50) or insufficiently green (50).²⁹ In the presented work, the total number of AES calculated penalty points was 17 and when these points were deducted from 100, the resulting points were 83 which indicated that the developed method is excellent green (>75) (Table 5). Combining two or more assessment tools play an important role in providing a clear image about the degree of eco-friendliness level of the developed method.³⁹ Therefore, the Green Analytical Procedure Index (GAPI) was employed. The results of GAPI index are displayed as pentagrams in three different colors ranging from green to yellow to red, corresponding to method greenness levels of high, medium, and low, respectively.³⁰ As shown in Table 5, the developed method's GAPI pictograms have seven green parts and only three red parts, indicating that the developed method has a low impact on the environment and can be applied to both qualitative and quantitative determination without endangering human health.

The AGREE metric was also employed to provide a thorough review of the proposed method's level of eco-friendliness utilizing a numerical scale from 0 to 1. Values close to one indicate high level of greenness.³¹ The proposed method's computed AGREE score of 0.72 demonstrated its high level of eco-friendliness.

Table 3 The application of the developed DES-DLLME method on real samples^a

Analyte	MP		EP		PP		BP	
Sample	Found (mg kg ⁻¹)	RSD	Found (mg kg ⁻¹)	RSD	Found (mg kg ⁻¹)	RSD	Found (mg kg ⁻¹)	RSD
S1	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S2	702.2	5.32	622.5	3.62	n.d.	n.d.	0.17	2.69
S3	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S4	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S5	494.4	0.06	n.d.	n.d.	341.6	0.18	n.d.	n.d.
S6	319.7	0.39	221.9	0.80	n.d.	n.d.	n.d.	n.d.
S7	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S8	n.d.	n.d.	<LOQ	n.d.	n.d.	n.d.	n.d.	n.d.
S9	n.d.	n.d.	<LOQ	n.d.	n.d.	n.d.	n.d.	n.d.
S10	n.d.	n.d.	<LOQ	n.d.	n.d.	n.d.	n.d.	n.d.
S11	n.d.	n.d.	<LOQ	n.d.	<LOQ	n.d.	n.d.	n.d.
S12	n.d.	n.d.	<LOQ	n.d.	0.85	4.37	n.d.	n.d.
S13	18.3	3.90	n.d.	n.d.	0.84	4.62	n.d.	n.d.
S14	173.1	6.12	n.d.	n.d.	86.5	4.89	n.d.	n.d.
S15	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.

^a n.d.: not detected.

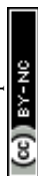
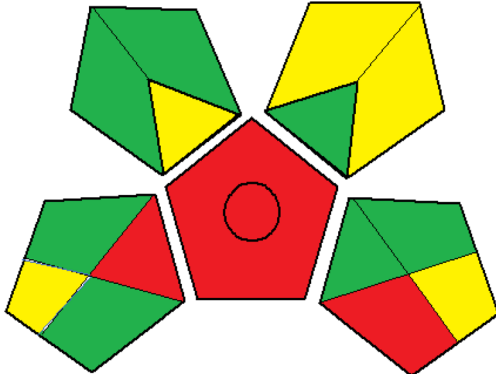





Table 4 Comparison of the developed method with other reported methods for determination of parabens using DES-DLLME

Analytes	Matrix	Separation technique	DES type and molar ration	Run time	r^2	Linearity range	LOD	R%	Ref.
MP, EP, PP, BP	Cosmetic oil	HPLC-UV	Choline chloride-ethylene glycol (1 : 2)	35 min	0.9992–0.9995	0.1–100 $\mu\text{g mL}^{-1}$	0.049–0.061 $\mu\text{g mL}^{-1}$	>84%	22
MP, EP, PP, BP	Edible oil	HPLC-UV	Tetrabutylammonium chloride–water (1 : 5)	10 min	>0.9986	1.5–500 $\mu\text{g L}^{-1}$	0.2–0.4 $\mu\text{g L}^{-1}$	85.1–106.8%	20
MP, EP, PP, BP	Liquid pharmaceutical samples	HPLC-UV	Choline chloride–glucose	—	0.9934–0.9962	0.1–5000 ng mL^{-1}	0.04–0.15 ng mL^{-1}	80.95–103.12%	21
MP only	Cosmetic products	UV-vis	Proline, malic acid and water (1 : 2 : 3)	—	0.9978	15–650 $\mu\text{g L}^{-1}$	4.5 $\mu\text{g L}^{-1}$	88.30%	23
MP, EP, PP, BP	Fruit juice samples, cosmetic and two pharmaceutical products	spectrophotometry HPLC-UV	Menthol and polyethylene glycol (1 : 1)	14 min	0.9995–0.9999	1–1000 ng mL^{-1}	0.3–2 ng mL^{-1}	66.3–97.5%	18
MP, EP, PP, BP, benzoic acid, sorbic acid	Beverages	HPLC-UV	Tetrabutylammonium bromide and acetic acid (1 : 2)	18 min	>0.9998	0.05–50.0 mg L^{-1}	0.02 to 0.05 mg L^{-1}	78.41–100.92%	19
MP, EP, PP, BP	Tap, river, lake and wastewater	HPLC-UV	Menthol and decanoic acid (2 : 1)	15 min	0.9961–0.9989	3–1000 ng mL^{-1}	0.6–0.8 ng mL^{-1}	69.1–78.5%	26
MP, EP, PP, BP	Cosmetic baby oil	HPLC-UV	Choline chloride-ethylene glycol (1 : 2)	—	0.9992–0.9995	0.1–100 $\mu\text{g mL}^{-1}$	0.049–0.061 $\mu\text{g mL}^{-1}$	82.02–101.95%	24
MP, EP, PP, BP	Toothpaste	HPLC-UV	Tetrabutylammonium chloride–ethanol (1 : 5)	11 min	≥ 0.9995	0.05–10 $\mu\text{g g}^{-1}$	0.01–0.02 $\mu\text{g g}^{-1}$	79.4–108.6%	25
MP, EP, PP, BP	Personal care products	HPLC-UV	Menthol–formic acid (1 : 2)	4.8 min	0.9992–0.9997	20–4000 ng mL^{-1}	0.17 to 0.33 ng mL^{-1}	82.19–102.45%	This work

Table 5 Investigation on the eco-friendliness level of the developed method using eco-scale, GAPI and AGREE tools

Eco-scale		GAPI pictogram	AGREE pictogram
Reagents	PPs		
Ethanol	4		
Formic acid	6		
Menthol	4		
NaCl	0		
Instruments			
UHPLC-DAD	0		
Occupational hazard	0		
Waste	3		
Total PPs	17		
Eco-scale	83		

4. Conclusion

In this study, a green analytical method for the simultaneous determination of four parabens in personal care products utilizing menthol-based DES-DLLME was developed and optimized. The presented method demonstrated good linearity ($r^2 \geq 0.9992$) and satisfactory recoveries (82.19–102.45%) with RSD $\leq 3.33\%$. The suggested method is environmentally friendly, quick, sensitive, and cost-effective. The method presented here could be successfully applied for simultaneous determination of parabens in personal care products without endangering human health or the environment.

Author contributions

Abdulmalik M. Alqarni: conceptualization, validation, formal analysis, writing – original draft & editing, investigation, project administration, visualization, supervision. Ahmed Mostafa: validation, formal analysis, resources, writing – review & editing. Heba Shaaban: validation, Formal analysis, writing – review & editing, funding acquisition. Mohamed S. Gomaa: review & editing. Danyah Albashrayi: formal analysis. Batool Hasheeshi: formal analysis. Nujud Bakhawain: formal analysis. Atheer Aseeri: formal analysis. Abdulaziz Alqarni: formal analysis. Abdulgani A. Alamri: visualization, formal analysis. Mohammad A. Alrofaidi: visualization, formal analysis.

Conflicts of interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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