RSC Advances



REVIEW

View Article Online
View Journal | View Issue



Cite this: RSC Adv., 2023, 13, 4436

Marine organisms as potential sources of natural products for the prevention and treatment of malaria

Walaa A. Negm, Da Shahira M. Ezzat Dbc and Ahmed Zayed xa

Vector-borne diseases (VBDs) are a worldwide critical concern accounting for 17% of the estimated global burden of all infectious diseases in 2020. Despite the various medicines available for the management, the deadliest VBD malaria, caused by *Plasmodium* sp., has resulted in hundreds of thousands of deaths in sub-Saharan Africa only. This finding may be explained by the progressive loss of antimalarial medication efficacy, inherent toxicity, the rise of drug resistance, or a lack of treatment adherence. As a result, new drug discoveries from uncommon sources are desperately needed, especially against multi-drug resistant strains. Marine organisms have been investigated, including sponges, soft corals, algae, and cyanobacteria. They have been shown to produce many bioactive compounds that potentially affect the causative organism at different stages of its life cycle, including the chloroquine (CQ)-resistant strains of *P. falciparum*. These compounds also showed diverse chemical structures belonging to various phytochemical classes, including alkaloids, terpenoids, polyketides, macrolides, and others. The current article presents a comprehensive review of marine-derived natural products with antimalarial activity as potential candidates for targeting different stages and species of *Plasmodium* in both *in vitro* and *in vivo* and in comparison with the commercially available and terrestrial plant-derived products, *i.e.*, quinine and artemisinin.

Received 14th December 2022 Accepted 26th January 2023

DOI: 10.1039/d2ra07977a

rsc.li/rsc-advances

Introduction

Vector-borne diseases (VBDs) are infectious diseases caused by parasites, bacteria, and viruses transmitted *via* vectors. About 700 000 deaths are reported officially by the World Health Organization (WHO) from these diseases per year worldwide, including malaria, dengue, schistosomiasis, human African trypanosomiasis, leishmaniasis, Chagas disease, chikungunya fever, Zika virus fever, yellow fever, West Nile fever, Japanese encephalitis, and onchocerciasis, Fig. 1. Based on the WHO reports released in 2020, VBDs are a worldwide concern that accounts for 17% of the estimated global burden of all infectious diseases.¹

VBDs are commonly associated with weather and climate, where the incidence of these diseases is mainly in the tropics and subtropical regions. The low hygiene, sanitation, waste management, and housing in these urban areas help also spread such diseases between the world's poorest people,

communities, and countries.² Various native to these regions as arthropods, including mosquitoes, ticks, sand flies, triatomine bugs, cockroaches, lice, fleas, and aquatic snails, are involved as mediators transmitting VBDs.^{3,4}

Particularly, malaria is the most challenging VBD that leads to health problems worldwide, especially in developing countries. It is a mosquito-borne infectious disease that affects humans and other animals. An estimated 405 000 malaria deaths worldwide were registered, along with 228 million cases in 2018, compared to 229 million cases and 409 000 deaths with more than 400 000 deaths in 2019, based on the WHO report. 5,6 In other words, malaria accounts for more than 50% of VBDs deaths.

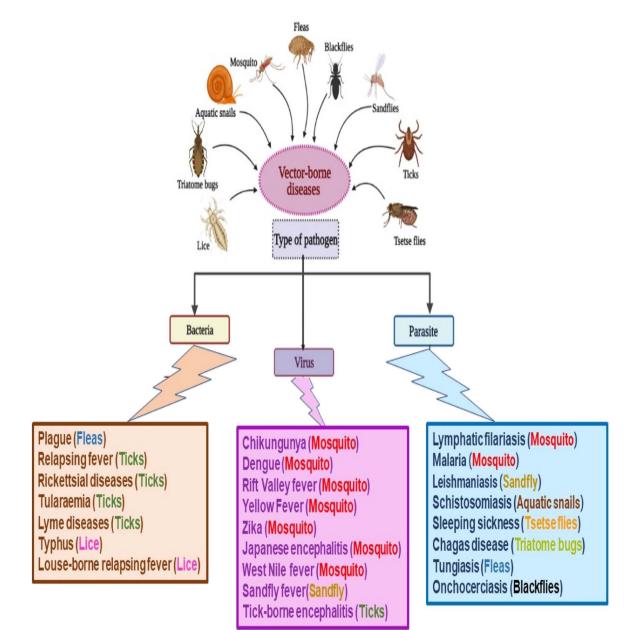
Malaria is transmitted through the bite of an infected Anopheles female mosquito. The infected mosquitoes carry one of several protozoans belonging to the genus *Plasmodium* (*P. falciparum*, *P. ovale*, *P. vivax*, *P. knowlesi*, and *P. malariae*).^{7,8} The parasite is then released into the bloodstream causing severe anemia and other signs and symptoms, including chills, fever, profuse sweating, headache, nausea, vomiting, abdominal pain, diarrhea, muscle pain, convulsions, coma, bloody stools.^{6,9,10} Serious complications or even death can occur in case of improper diagnosis or treatment.^{11–13} The life cycle of the malaria parasite of *Plasmodium* sp. is illustrated in Fig. 2. This figure is of great importance for helping drug discovery

^aDepartment of Pharmacognosy, Tanta University, College of Pharmacy, El-Guish Street, Tanta 31527, Egypt. E-mail: walaa.negm@pharm.tanta.edu.eg; ahmed. zayed1@pharm.tanta.edu.eg

^bDepartment of Pharmacognosy, Faculty of Pharmacy, Cairo University, Kasr El-Aini Street, Cairo 11562, Egypt. E-mail: shahira.ezzat@pharma.cu.edu.eg

Department of Pharmacognosy, Faculty of Pharmacy, October University for Modern Sciences and Arts (MSA), Giza 12451, Egypt

Review



Classification of vector-borne diseases (VBDs) according to pathogen types.

processes of novel drugs targeting critical stages in the parasite life cycle.

Chloroquine (CQ) and hydroxychloroquine are two existing chemical medicines that have limited usefulness and efficacy as antimalarials due to their high cost, unpleasant side effects, and evolution of multi-drug resistance associated with them.14 Hence, there has been an urgent need and continuous search for novel sources of more efficacious drugs to combat the disease. Nevertheless, natural products, including terrestrial medicinal plants, have a long history in the treatment of malaria owing to their relative efficacy, safety, reasonable cost, and availability. 15,16 The two most successful antimalarial drugs; namely artemisinin and quinine (Fig. 3), were sourced from medicinal plants of cinchona qinghao (Artemisia annua, Family Asteraceae) and (Cinchona officinalis, Family Rubiaceae), respectively, and have been used for hundreds of years and before the mosquito cycle was explored. Even today, in the fight against malaria, both quinine and artemisinin are still of prime importance.17

In contrast to terrestrial plants, marine organisms do not have a remarkable history of use in traditional medicine. However, recent advances in marine biology and engineering have helped investigation and scientific exploration of the marine environment to identify and isolate novel compounds, which have proven their potential bioactivities against lifethreatening diseases, including tumor and viral infections. 18-20 More than 30 000 compounds have been identified from about 240 000 known species of marine organisms. 21 Few of them have

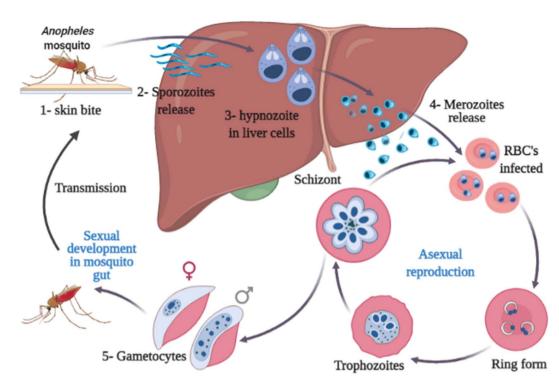


Fig. 2 Life cycle of the malaria parasite.

Fig. 3 Chemical structure of quinine (a) and artemisinin (b).

been approved by the Food and Drug Administration (FDA), including ziconotide (Prialt®) as a potent analgesic, trabectedin (Yondelis®), and cytarabine or ara-C (Cytosar-U®) as anti-tumor agents, vidarabine or ara-A (Vira-A®) and iota-carrageenan (Carragelose®) as an antiviral, and omega-3-acid ethyl ester (Lovaza®) for treating hypertriglyceridemia (Table 1).²²⁻²⁴

Recently, Nweze, et al. published a review article highlighting the potential of marine-derived natural products for the

treatment of some examples of diseases for neglected communities, including malaria, leishmaniasis, and trypanosomiasis.25 Although some of the previous studies could not identify the chemical structure of bioactive components that acted significantly against malaria,26 the current article focuses on malaria. It reviews the different chemical classes, i.e., alkaloids, terpenoids, endoperoxides, phosphotriesters, peptides and depsipeptides, and macrolides, derived from marine organisms, including sponges, cyanobacteria, actinomycete bacteria, soft corals, and algae. These bioactive have been confirmed to be potential candidates for managing malaria compared to commercially available products by targeting various stages in Plasmodium sp. life cycle. Moreover, the half maximum cytotoxic (CC50) and inhibitory concentration (IC50) against the different stages of the malaria parasite shall be highlighted, in addition to the possible mechanism of action and structureactivity relationships (SAR) in previous reports investigated the antiplasmodium activity. Hence, the current review may

Table 1 A list of some examples of approved marine-derived drugs currently on the market

Trade name	Scientific name	Source	Family	Indication	Ref.
Prialt [®]	Ziconotide	Cone snail species Conus magus	Conidae	potent analgesic	22-24
Yondelis [®]	Trabectedin	Candidatus Endoecteinascidia	Unclassified	Anti-tumor	22-24
		frumentensis	family candidatus		
		-	endolissoclinum		
Cytosar-U®	Cytarabine or ara-C	Cryptotethia crypta sponge	Tethyidae	Anti-tumor	22-24
Carragelose [®]	Iota-carrageenan	Eucheuma denticulatum sponge	Solieriaceae	Antiviral	22-24
Vira-A®	Vidarabine or ara-A	Tectitethya crypta sponges	Tethyidae	Antiviral	22-24

Table 2 A list of marine-derived antimalarial alkaloids showing their IC_{50} against various strains of *Plasmodium* sp., chemical structure and biogenic source^a

biogenic source ^a		•			
Compound	Antiplasmodial activity (IC ₅₀ value)	Structure	Source	Marine class	Ref.
Manzamine A (1)	$W2 = 0.015 \; \mu M \; D6$ = 0.0082 μM	H Z H OH OH		Sponge	27
8- Hydroxymanzamine (2)	$W2 = 0.014 \; \mu M \; D6 \\ = 0.010 \; \mu M$	N H OH		Sponge	27
Manzamine F (3)	$W2 = 2.93~\mu M~6 = 1.34~\mu M$	N H OH N H	Okinawan Haliclona	Sponge	27
6-Hydroxy- manzamine (4)	$W2=1.5~\mu M$ D6 $=$ 1.36 μM	OH N N N N N N N N N N N N N		Sponge	27

Compound	Antiplasmodial activity (IC_{50} value) Second	ructure	Source	Marine class	Ref.
Neo-kauluamine (5)	$D6 = 1.46 \ \mu M \ W2 = 2.41 \ \mu M$	HO O D H Z H O H D H Z H O H D H Z H O H D H D H D H D H D H D H D H D H D	Indo-pacific sponge	Sponge	28
12,34- Oxamanzamine A (6)	$D6=8.97~\mu M$	Z T T T T T T T T T T T T T T T T T T T		Sponge	28
Zamamidine A (7)	0.0008 to 0.016 μM	N H H N OH	Amphi- medon sp.	Sponge	35
Zamamidine B (8)				Sponge	

Compound	Antiplasmodial activity (IC ₅₀ value) Structure		Source	Marine class	Ref.
		NH H			
Zamamidine C (9)		N H N OH		Sponge	
Zamamidine D (10)		NH ₂ HN NH ₂ NH		Sponge	
Homofascaplysin (11)	K1 = 0.04 μM NF54 $= 0.07$ μM	N+ HO O	Hyrtios sponge	Sponge	38
Marinacarboline A (12) Marinacarboline B (13) Marinacarboline C (14)	$3D7$ and Dd2 IC $_{50}$ from 1.92 to 36.03 μM	$ \begin{array}{ccc} & & & & \\ & & & & \\ & & & & \\ & & & &$	Marin- actinospora thermoto- lerans	Actinomycete bacteria	39
Marinacarboline D (15)		(14) R= H			

13-N-Demethylmet	s Ref.	Marine class	Source		Structure	Antiplasmodial activity (${ m IC}_{50}$ value)	Compound
Crambescidin 800 $3D7 = 0.16 \mu\text{M}$ FCR3 $= 0.24 \mu\text{M}$ $= $	е 38	Actinomycete bacteria	actinospora	$ \begin{array}{c} N \\ NH \\ OR_2 \end{array} $ $ \begin{array}{c} NH \\ OR_2 \end{array} $ $ \begin{array}{c} NH \\ OR_2 \end{array} $		$10.43~\mu M~Dd2 =$	methyl -pendolmycin (16) methyl- pendolmycin-14- <i>O</i> -
Crambescidin 800 $3D7 = 0.16 \mu\text{M}$ FCR3 $= 0.24 \mu\text{M}$ $= 0.24 \mu$				Me $R_2 = \alpha$ -glucose			
(19) crambescidin acid (20) 10.43 μ M Dd2 = 18.67 and 5.03 μ M (19) R=H (20) R=COOH Fromiamycalin (21) 3D7 = 0.24 μ M (19) R=H (20) R=COOH Unguiculin A (22) 3D7 = 12.86 μ M Ptilomycalins E (23) Prilomycalins F (24) Ptilomycalins G (25) and 0.46 μ M NH ON PTILOMYCALING F (24) PTILOMYCALING G (25) And 0.46 μ M NH ON PTILOMYCALING G (25) And 0.46 μ M				N N N	H_2N		
acid (20) 18.67 and 5.03 μM (19) R=H (20) R=COOH Fromiamycalin (21) 3D7 = 0.24 μM Unguiculin A (22) 3D7 = 12.86 μM Ptilomycalins E (23) Ptilomycalins G (25) and 0.46 μM NH NH NH NH NH NH NH NH NH N				ON H			
Fromiamycalin (21) $3D7=0.24~\mu M$ $\begin{array}{cccccccccccccccccccccccccccccccccccc$	39 ar			H R			
Fromiamycalin (21) $3D7=0.24~\mu M$ Unguiculin A (22) $3D7=12.86~\mu M$ Prilomycalins E (23) $3D7=0.35,0.23,$ and $0.46~\mu M$ NH NH NH NH NH NH NH NH NH N	40				(19) R=H		
Unguiculin A (22) $3D7=12.86~\mu M$ Ptilomycalins E (23) Ptilomycalins F (24) Ptilomycalins G (25) NH N				N	N	$3D7=0.24~\mu\text{M}$	Fromiamycalin (21)
Ptilomycalins E (23) Ptilomycalins F (24) Ptilomycalins G (25) Ptilomyc		Sponge		H NH ₂	* 13	$3D7=12.86~\mu\text{M}$	Unguiculin A (22)
Ptilomycalins F (24) $^{3D7}=0.35, 0.23,$ and $0.46~\mu M$ NH O							
	41			2 (24) R= H	.	3D/ = 0.35, 0.23,	Ptilomycalins F (24)
(25)R=H Ptilomycalins H (26) 3D7 = 0.46 μM				(25)R=H			

Table 2 (Contd.)

Compound	Antiplasmodial activity (IC ₅₀ value)	Structure	Source	Marine class	Ref.
		H ₂ N N N N N N N N N N N N N N N N N N N			
Opacaline B (27) Opacaline C (28)	<i>K</i> 1 range of 2.5–14 μM	Br NH NH ₂	Pseudodistoma opacum	New Zealand ascidian	42
Didemnidine A (29) Didemnidine B (30)	$K1=0.047~\mu M$	(27) n=2 (28) n=1 CF3CHO2 © NH NH NH ₃ *	Ascidian <i>Didemnum</i> sp.	Marine tunicate	43
		(29) R= H			
		(30) R=Br			
Salinosporamide A (31)	3D7 = 11.4 nM FCB = 19.6 nM	HO HN CI	Salinispora tropica	Marine actinomycete bacteria	44
Psammaplysin H (32) Psammaplysin F (33) Psammaplysin G (34)	$3D7 = 0.41, 1.92, \\ and 5.22 \; \mu M$	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Aplysinella strongylata	Sponge	52
		$(33) R= NHMe_3$			
		(34) $R = N(Me)CONH_2$			
Ceratinadin E (35)	$K1=0.9\;\mu M$	Br O OH O	Okinawan Pseudoceratina	Sponge	54
Ceratinadin F (36)	K1 > 8.16 μM	וט			

Compound	Antiplasmodial activity (IC ₅₀ value)	Structure	Source	Marine class	Ref
Compound	activity (1050 value)		Source	Watthe Class	KCI.
		Br HN O Br HN O Br O Br			
Tsitsikammamine C (37)	3D7 = 13 nM Dd2 = 18 nM	HN OH			55
Makaluvamine J (38)	3D7 = 25 nM Dd2 = 22 nM	N+ NH O OH			
Makaluvamine G (39) Makaluvamine L (40)	3D7 = 36, 40 nM Dd2 = 39, 21 nM	N+ NH OH	<i>Zyzzya</i> sp.	Sponge	
		$(39)R = CH_3$ (40) $R = H$			55
Makaluvamine K (41)	3D7 = 396 nM Dd2 = 300 nM	N N OH			
Damirone A (42) Damirone B (43)	3D7 = 1880 Dd2 = 360 nM	O N			
		$(42)R = CH_3$ (43) $R = H$			
Dihydro- discorhabdin B (44)	$D6 = 0.17 \mu M W2 = 0.13 \mu M$	O H N N Br HO Br	<i>Latrunculia</i> sp.	Sponge	56
Discorhabdin Y (45)	$EC_{50}=0.5~\mu M$				

Table 2 (Contd.)

Compound	Antiplasmodial activity (IC ₅₀ value) Structure	,	Source	Marine class	Ref.
		Br HN N			
Discorhabdin A (46)	$\begin{array}{l} D6 = 0.05 \; \mu M \; W2 = \\ 0.05 \; \mu M \end{array}$	HZ H S H			
Discorhabdin C (47)	D6 = 2.8 μ M W2 = 2.0 μ M	HZ HZ Br			
Discorhabdin E (48)	$W2=0.2\;\mu M$	HZ Br O			56
Discorhabdin L (49)	$W2=0.13~\mu M$	D TZ O O O O O O O O O O O O O O O O O O			
Dihydro- discorhabdin C (50)	D6 = 0.17 μM W2 = 0.13 μM	OH HZ N Br HO			
Girolline (51)	$FCM29 = 0.13~\mu\text{M}$	H ₂ N H OH	Cymbastela cantharella	Sponge	58
Thiaplakortone A (52)	3D7 = 51 Dd2 = 6.6 nM	CF3CO2 [©] NH ₃ NH ₃	Plakortis lita	Sponge	59

Table 2 (Contd.)

Compound	Antiplasmodial activity (${\rm IC}_{50}$ value) Structure		Source	Marine class	Ref.
Thiaplakortone B (53)	3D7 = 650 Dd2 = 92 nM	CF3CO2 \ominus			
Thiaplakortone C (54)	3D7 = 309 Dd2 = 171 nM	O CF3CO2 O NH (H)			
Thiaplakortone D (55)	3D7 = 279 Dd2 = 159 nM	NH O CF3CO2© NH O NH H			
Monam- philectine A (56)	$W2=600\ nM$		Hymeniacidon sp.	Sponge	60
Agelasine J (57)	$FcB1 = 6.6 \; \mu M$	N-N+N+N+2			61
Agelasine K (58)	$FcB1 = 8.3~\mu\text{M}$	CIT N N NH ₂	Agelas mauritiana	Sponge	
Agelasine L (59)	$FcB1 = 18 \; \mu M$	N N N N N N N N N N N N N N N N N N N			
Netamine G (60)		H ₂ N N H			
Netamine H (61)	NA	H ₂ N N H	Biemna laboutei	Sponge	62
Netamine I (62)		Å n —			

Compound	Antiplasmodial activity (IC ₅₀ value)	Structure		Source	Marine class Ref.
			H ₂ N N H		
Netamine J (63)			H ₂ N H		
Netamine K (64)			H ₂ N ⊕ HN = NH H		
Netamine L (65)	(64) $IC_{50} = 2.4 \mu M$		H ₂ N ⊕ HN NH H		
Netamine M (66)	NA for the other compounds (65–67)		H ₂ N ⊕ HN = NH H		
Netamine N (67)			H ₂ N ⊕ HN = NH H	Biemna labout	ei Sponge
^a NA:not available.					

open new frontiers for discovery and approval of novel potent drugs for this life-threatening disease.

2. **Alkaloids**

Various classes of marine-derived alkaloids have shown potent antimalarial activity. β-Carboline, indole, imidazole, and pyrrole alkaloids are mostly found. They showed bioactivities against different stages of the Plasmodium parasite with a unique mechanism of action. Fifteen classes represented by 67 compounds were reviewed. Among them are manzamine alkaloids which showed inhibitory activity against glycogen synthase 3 (GSK-3) topoisomerase. In addition, salinosporamide showed a potent protease inhibitory effect. Numerous marine-derived alkaloids shall be discussed in detail in the following sub-sections and Table 2, including their sources, IC₅₀, chemical structures, SAR, and mechanism of action.

2.1. Manzamines

Manzamines are polycyclic (7–8 rings) alkaloids containing a βcarboline moiety. Manzamine A (1) was first reported from an Okinawan sponge belonging to the Haliclona species (family Chalinidae).27 They are one of the essential antimalarial alkaloids. In addition to the lack of in vivo toxicity, the manzamines demonstrated greater effectiveness as antimalarial agents than the commonly used drugs artemisinin and CQ.28 The mechanism of manzamine alkaloids is not completely understood. Still, authors described β-carboline alkaloids as micromolar inhibitors of glycogen GSK-3 by malaria parasites and inhibitors of topoisomerase DNA through intercalation in DNA-base pairs.

Hence, a complete SAR investigation of manzamine alkaloids is necessary to understand the importance of each moiety (β-carboline and pentacyclic ring) and the influence of different substituents on antimalarial activity. Manzamines SAR is summarized into two objectives. The first is the effect of various substitutions on the β-carboline nucleus, and the other is the effect of substitutions on the pentacyclic ring.29 The β-carboline moiety of manzamine alkaloids is responsible for antimalarial activity. 9-N alkylation of the β-carboline ring decreases antimalarial activity, indicating that 9-NH is necessary for their antimalarial activity. Hydroxyl-group substitution at position 8 of the β-carboline skeleton does not significantly affect its antimalarial activity.

RSC Advances Review

Hence, 8-hydroxymanzamine (2) has the same effect as manzamine A, while manzamine F (3), a related derivative of manzamine A, is inactive. The eight-membered rings differ between the inactive manzamine F and the active manzamine A. The double bond reduction and/or the incorporation of a ketone group into the adjacent carbon is harmful to antimalarial activity. Likewise, hydroxyl group attachment at position 6 instead of position 8 has a negative effect on antimalarial activity, as shown by the lower potency of 6-hydroxy-manzamine A (4).²⁷ In vitro and in vivo studies, manzamines A and its 8-hydroxy derivative inhibited *P. falciparum* growth.³⁰ Several total syntheses of manzamines have been accomplished.^{29,31}

2.2. Neo-kauluamine

Neo-kauluamine (5) is a manzamine dimer, with a highly complex molecule composed of two units of manzamine fragmented by ether bonds between the eight-membered rings isolated from an unspecified genus of Indo-Pacific sponge (Petrosiidae, order Haplosclerida). Despite its structural complexity, neo-kauluamine displayed a strong efficacy *in vivo* and is considered an up-and-coming agent in malaria.²⁷ Although this structure, like manzamine F (3), lacks the double bond in the eight-membered ring, it showed the same activity as manzamine A. The lack of antimalarial activity for 12,34-oxamanzamine A (6) suggested that the C-12 hydroxyl, the C-34 methine, or the 8-ring conformation are of great importance for the antimalarial activity.²⁸

2.3. Zamamidines

Zamamidines A–D (7–**10**) are also manzamine alkaloids obtained from *Amphimedon* sp. sponge (*Niphatidae*). They had inhibitory activities against *P. falciparum* (IC₅₀ values from 0.0008 to 0.016 μM).^{32–34} Zamamidine C (**9**) is the most active one of the series. Zamamidine D (**10**) is the first manzamine alkaloid characterized by having a moiety of 2,2′-methylene bistryptamine instead of a unit of β-carboline.³⁵

2.4. Homofascaplysin A

Homofascaplysin A (11) is also β-carboline alkaloid. It was extracted from the *Hyrtios erecta* sponge (Thorectidae).³⁶ This alkaloid presented potent activity against CQ-resistant *P. falciparum* strains ($IC_{50} = 0.07 \, \mu M$) with approximately 10-fold less cytotoxicity.³⁷ This potent antiplasmodial activity of this compound demonstrated its potential as a lead structure among antimalarial agents and became a synthesis target for the production of other similar analogues.³⁸

2.5. Marinacarbolines

Marinacarbolines A–D (12–15), series of β -carboline alkaloids were obtained from the fermentation broth of the marine actinomycete bacteria *Marinactinospora thermotolerans* (Nocardiopsaceae). Marinacarbolines displayed antiplasmodial activities against 3D7 and Dd2 lines of *P. falciparum*, with IC₅₀ from 1.92 to 36.03 μ M.³⁹

2.6. Indolactam alkaloids

13-*N*-Demethyl-methylpendolmycin (**16**) and methylpendolmycin-14-*O-α*-glucoside (**17**) were derived from *Marinactinospora thermotolerans* (Nocardiopsaceae) fermentation broth. They were also found to exhibit moderate or weak activity against 3D7 ($IC_{50} = 20.75 \mu M$ and 10.43 μM) and Dd2 ($IC_{50} = 18.67 \mu M$ and 5.03 μM) strains of *P. falciparum*, respectively.³⁹

2.7. Crambescidins

Crambescidin 800 (18) was obtained from the Indonesian sponge (*Mycophora* sp. Crambeidae). Crambescidin 800 showed IC₅₀ of 160 nM and 240 nM, respectively, against the 3D7 and FCR3 lines of *P. falciparum*. Also, other alkaloids, including crambescidin 359 (19), crambescidin acid (20), and fromiamycalin (21), were extracted from the sponge *Monanchora unguiculate* collected in Madagascar. Crambescidin 359 was active against the 3D7 line of *P. falciparum* (IC₅₀ = 0.52 μ M). Additionally, Unguiculin A (22), an acyclic guanidine alkaloid, was detected in this sponge. In addition to four pentacyclic alkaloids ptilomycalins E-H (23–26) were also isolated. Among them, fromiamycalin (IC₅₀ = 0.24 μ M), unguiculin A (IC₅₀ = 12.86 μ M) ptilomycalins E (IC₅₀ = 0.35 μ M), F (IC₅₀ = 0.23 μ M), ptilomycalins G and H mixture (IC₅₀ = 0.46 μ M), respectively exhibited promising activity against *P. falciparum*.

2.8. Opacalines

Opacalines are alkyl guanidine-substituted β -carboline-containing metabolites obtained from the New Zealand ascidian *Pseudodistoma opacum* (Pseudodistomidae). Opacalines B (27) and C (28) displayed moderate activity against the CQ-resistant *P. falciparum* strain (IC₅₀ range of 2.5–14 μ M).⁴²

2.9. Spermidine

Two indole alkaloids, Didemnidines A (29) and B (30), were obtained from the New Zealand ascidian *Didemnum* sp. (Didemnidae). Among them, Didemnidine B showed mild activity (IC $_{50}=0.047~\mu M$) against *P. falciparum*.

2.10. Salinosporamide

Salinosporamide A (31) is a simple γ -lactam spiro-alkaloid isolated from an actinomycete bacteria belonging to the genus *Salinispora tropica* (Micromonosporaceae).⁴⁴ Salinosporamide A is also a cyclic depsipeptide (bicyclic β -lactone γ -lactam peptide).

Salinosporamide A showed potential as an antimalarial candidate. It exhibited a potent parasite proteasome inhibitor and antimalarial activity against P. falciparum in vitro (IC₅₀ = 11.4 nM) and in vivo against P. yoelii. ^{45,46} By controlling T cell proliferation and leading to cell cycle arrest, Salinosporamide A suppressed T cell activation and regulated the expression of cyclin-dependent kinases. ⁴⁷ Recently total synthesis for salinosporamide A molecule has been reported. ^{46,48,49}

Review RSC Advances

2.11. Bromotyrosine alkaloid

Several psammaplysin derivatives were obtained from the Indonesian marine sponge *Aplysinella strongylata* (*Aplysinellidae*). Psammaplysin H (32) is a bromotyrosine alkaloid from a marine sponge *Pseudoceratina* sp. and displayed more than 97% antimalarial activity (0.41 μM concentration). Psammaplysins H, F (33), and G (34) exhibited antimalarial activity, while Psammaplysins F and G displayed antimalarial activities against the drug-resistant strains of Plasmodium falciparum. Ceratinadins were bromotyrosine alkaloids that had the 1,6-dioxa-2-azaspiro[4.6] undecane skeleton, Ceratinadins E (35) and F (36), obtained from an Okinawan marine sponge *Pseudoceratina* sp. (Pseudoceratinidae). Ceratinadins E showed antimalarial activities against drug-resistant and drug-sensitive K1 P. falciparum strains.

2.12. Pyrrolo-iminoquinones

Pyrroloiminoquinone compounds, Tsitsikammamine C (37), makaluvamines J (38), G (39), L (40), K (41), damirones A (42), and B (43), were extracted from the Australian marine sponge Zyzzya sp. (Acarnidae, order Poecilosclerida). All compounds were investigated against 3D7 and Dd2 P. falciparum strains. Among them, tsitsikammamine C with $IC_{50} = 13$ and 18 nM, respectively, inhibited both ring and trophozoite stages of the malaria parasite life cycle. Makaluvamines J, G, and L showed a potent antimalarial activity (IC₅₀ < 100 nM) in vitro against both strains.⁵⁵ A class of two new brominated pyrroloiminoquinones; dihydrodiscorhabdin B (44) and discorhabdin Y (45), along with six pyrroloiminoquinone alkaloids, discorhabdins A (46), C (47), E (48), and L (49), dihydrodiscorhabdin C (50) were obtained from Alaskan sponge genus Latrunculia (Latrunculiidae) among them, discorhabdins A, C, and dihydrodiscorhabdin C displayed antimalarial in vitro activity against both D6 and W2 P. falciparum strains.56

2.13. Imidazole alkaloids

Girolline (51), a 2-aminoimidazol derivative initially isolated from a Caledonian sponge *Cymbastela cantharella* (Axinellidae)⁵⁷ showed antimalarial activity against *P. falciparum* (FCM29) strain of (IC₅₀ = 0.13 μ M) a high *in vivo* activity in a *P. vinckei petteri* rodent model.⁵⁸

2.14. Thiazine-derived alkaloids

Four thiazine-pyrroloquinone containing tricyclic quaternary alkaloids, Thiaplakortones A–D (52–55), were obtained from the Australian sponge *Plakortis lita* (Plakinidae). Thiaplakortones A–D exhibited potent inhibition against 3D7 CQ-sensitive (IC $_{50}$ = 51 nM, 0.65, 0.309, and 0.279 μ M) and Dd2 CQ-resistant *P. falciparum* (IC $_{50}$ = 6.6 nM 0.092, 0.171, and 0.159 μ M), respectively.⁵⁹

2.15. Terpenoid alkaloids

Several terpenoid alkaloids had been showing reasonable antimalarial activity. For instance, Monamphilectine A (56), a diterpenoid-lactam alkaloid from sponge *Hymeniacidon* sp., displayed action against the *P. falciparum* W2 strain ($IC_{50} = 0.60 \, \mu M$). Agelasines J (57), K (58), and L (59), three adenine terpenoids, were obtained from the sponge *Agelas mauritiana* (Agelasidae). They have weak antimalarial activity against the *P. falciparum* Columbian FcB1 strain ($IC_{50} = 6.6, 8.3$, and 18 μM , respectively). Netamines G–N ($^{60-67}$) and tricyclic alkaloids were obtained from the Madagascar sponge *Biemna laboutei* (Desmacellidae). Netamine K showed antimalarial activity ($IC_{50} = 2.4 \, \mu M$) against *P. falciparum*.

3. Terpenoids

All terpenes or terpenoids have fundamental repeating fivecarbon isoprene units. Terpenes are classified as hemiterpenes (C_5), monoterpenes (C_{10}), sesquiterpenes (C_{15}), diterpenes (C_{20}), sesterterpenes (C_{25}), triterpenes (C_{30}), and tetraterpenes/carotenoids (C_{40}). Marine-derived terpenoids have attracted potential interest similar to the terrestrial analogues represented by the sesquiterpene lactone artemisinin and isonitriles-containing terpenes. More than 30 compounds were isolated and showed antimalarial activity from marine organisms. Unique mechanisms were demonstrated, including the inhibitory activity against heme detoxification by isonitrile derivatives. They are discussed in the following subsections, and their chemical structures are shown in Table 3.

3.1. Sesquiterpene isonitriles

Sesquiterpenoid metabolites containing isonitrile, thiocyanate, and formamide were isolated for the first time from Axinella cannabina sponge (Axinellidae).66,67 Isonitrilecontaining compounds do their activity by blocking heme (FP) detoxification. The interaction of isonitrile with free heme was demonstrated by forming a coordination complex with the iron center. The drug carrier must possess a solid lipophilic molecular nucleus with at least a tricyclic structure that carries an isonitrile group and establishes further hydrophobic reactions above the ring plane. The interaction of marine isonitrile derivatives with heme can suppress the sequestration of FP into beta-hematin and prevent the peroxidative and glutathionemediated destruction of FP under conditions designed to imitate the environment inside the malaria parasite.68 Axisonitrile-1 (68) was the parent compound of the class of isonitrile containing sesquiterpenoids marine metabolites obtained from A. cannabina sponge. 67,69

Isolation of axisonitrile-1 was followed by other related sesquiterpenoids from the same sponge, as axamide-1 (69), axamide-2 (70), axamide-3 (71), axisonitrile-2 (72), axisonitrile-3 (73), axisothiocyanate-2 (74), axisothiocyanate-3 (75).⁷⁰⁻⁷² In 1978, (–)-8,15-diisocyano-11(20)-amphilectene (76) was first reported from *Hymeniacidon amphilecta* (Halichondriidae) and was subsequently shown to demonstrate potent anti-infective activity *in vitro*.^{73,74} The closely related axisothiocyanate-3 was inactive, indicating that the activity depends not only on the carbon skeleton's structural characteristics but also directly on the existence of the isonitrile group. The action was confirmed

 $\textbf{Table 3} \quad \text{A list of marine-derived terpenes-containing antimal arial drugs showing their IC}_{50} \text{ against various strains of } \textit{Plasmodium} \text{ sp., chemical } \text{ and } \text{ against various strains of } \text{ plasmodium} \text{ sp., chemical } \text{ against various strains of } \text{ plasmodium} \text{ sp., chemical } \text$ structure and biogenic source

Compound	Antiplasmodial activity (IC_{50} value)	Structure	Source	Marine class	Ref.
Axisonitrile-1 (68) Axamide-1 (69)		-N-C=C (68) R= H H			
Axamide-2 (70) Axisonitrile-2 (72) Axisothiocyanate-2 (74)	$W2 = 0.073 \ \mu M \ D6 \\ = 0.61 \ \mu M$	(69) R= \\ \tag{R} \\	Axinella cannabina	Sponge	67 and 69–72
Axamide-3 (71) Axisonitrile-3 (73) Axisothiocyanate-3 (75)		—N—C=C 			
(–)-8,15-Diisocyano- 11(20)-amphilectene (76)	W2 = 15 nM D6 = 16 nM K1 = 90 nM	(75) $R = $	Hymeniacidon amphilecta, Venzea flava	Sponge	73 and 74
Kalihinol A (77)	$FCR\text{-}3 = 12~\mu M$	⊝C v	Acanthella sp.	Sponge	68

Compound	Antiplasmodial activity (IC_{50} value)	Structure	Source	Marine class	Ref.
(8 <i>R</i>)-8-Bromo-10- epi-β-snyderol (7 8)	D6 = 0.012 and W2 = 0.017 μM	Br. OH	Laurencia obtusa	Red alga	77
au 1/a)		H_1 Q			
Chloroaureol (79) Aureol (80) Aureol acetate (81)	(79) D6 = 9.74 μ M (80) and (81) NA	(79) R ₁ =H, R ₂ = Cl	Smenospongia aurea	Sponge	78
		(80) $R_1 = R_2 = H$ (81) $R_1 = AC$, $R_2 = H$			
Pelorol (82)	NF54 = 0.005 and K1 = 0.0019 μ M	OH OH OH	Dactylospongia	Sponge	79
Ilimaquinone (83)	NF54 = 0.0026 K1 = $0.0048~\mu M$	OH O	elegans	. 0	79
Isocyanoclovane (84)		NC H			
2-Isocyanoclovene (85)	3D7 = 300, 290, and 260 nM Dd2 = 360, 830, and 870 nM	NC H	Phyllidia ocellate	Nudibranch	80
4,5-Epi-10- isocyanoi- sodauc-6-ene (86)		NC H			

Table 3 (Contd.)

Compound	Antiplasmodial activity (IC_{50} value)	Structure	Source	Marine class	Ref.
8a,11-Dihydroxy- pachydictoyl A (87)		OH OH			
4,18-Dihydroxy- dictyolactone (88)	$\mathit{K}1 = 10.0~\mu\text{M}$	HO, H	Dictoyta sp.	Brown alga	83
Laevigatol A (89)	Dd2 IC ₅₀ < 5.0 μM	о <u>н</u> Н О О О О О О О О О О О О О О О О О О О	Pseudopterogorgia elisabethea	Soft coral	84 and 85
Pseudopterosin V (90)	$Dd2 = 2.2~\mu M$	OH OH OH OH	Pseudopterogorgia elisabethea	Soft coral	86
Dorisenone D (91)	$K1=1.3~\mu M$		Dysidea arenaria	Sponge	87

by comparison among the activities of related compounds. It showed that the biological activity generally depends on the isocyanide functionality and the carbon skeleton structural features. The isocyanide group's location also has a pivotal role.⁷⁵

Bis-isonitrile-containing product and isocyanoterpene members, Kalihinols, were isolated from a Guam sponge, *Acanthella* sp. 68,76 Many natural Kalihinol products are potent inhibitors of *P. falciparum*. Hence its importance, several Kalihinol analogs were synthesized and investigated using drugsensitive and resistant *P. falciparum* for blood-stage antimalarial activity. 68 Kalihinol A (77) showed increased potency and activity against the FCR-3 strain (Kalihinol A EC₅₀ = 1.2 nM). 67 . (8*R*)-8-Bromo-10-epi-β-snyderol (78), a sesquiterpene isolated from *Laurencia obtuse* (Rhodomelaceae) red alga displayed

activity against *P. falciparum* D6 and W2 strains (D6 = 0.012 and W2 = 0.017 μ M), respectively.⁷⁷

6'-Chloroaureol (79), aureol (80) and aureol acetate (81), sesquiterpene-phenol were obtained from *Smenospongia aurea* sponge.⁷⁸ These derivatives displayed reasonable activity against *P. falciparum* D6 strain. Another marine sesquiterpene, pelorol (82), and sesquiterpene quinone, ilimaquinone (83), were extracted from the sponge *Dactylospongia elegans* (Thorectidae). Both showed moderate antiplasmodial activity against K1 and NF54 strains of *P. falciparum*.⁷⁹

Sesquiterpenes, 2-isocyanoclovene (84) and 2-isocyanoclovane (85) were obtained from the Australian nudibranch *Phyllidia ocellate* (Phyllidiidae). They showed activity against 3D7 ($IC_{50} = 300, 290, and 260 nM, respectively)$

Review **RSC Advances**

and Dd2 P. falciparum clones (IC₅₀ = 360, 830, and 870 nM, respectively).80

3.2. Diterpene isonitriles

A phytochemical investigation of the Cymbastela hooperi sponge provided 15 diterpenes containing isonitriles isocyanate, isothiocyanate, and isonitrile functionalities, which showed higher antimalarial effect and moderate toxicity.81 The activity of analogues with isocyanate and isothiocyanate functionality was up to ten times lower, demonstrating that the isonitrile group improved activity. An analogue containing only the formamide functional group but no isonitrile group was also ineffective against P. falciparum, implying that the formamide group isn't necessary for antiplasmodial efficacy.82 Monamphilectine A (56) was a diterpenoid β-lactam alkaloid separated from a Hymeniacidon sp. sponge (Halichondriidae). Monamphilectine A displayed a potent antimalarial activity,.60 While (-)-8,15diisocyano-11(20)-amphilectene (86), re-isolated from the Svenzea flava sponge (Scopalinidae), was used as a precursor to synthesize five new products, all of which were tested against laboratory colonies of P. falciparum and Mycobacterium tuberculosis H₃₇Rv.⁷⁵

In addition, 8a,11-dihydroxypachydictoyl A (87), and 4,18dihydroxydictyolactone (88) were diterpenoids isolated from Dictoyta sp. of the brown alga, which displayed antimalarial activity (IC₅₀ = 10.0 μ M) against K1 strain of *P. falciparum*.⁸³

Moreover, soft corals and echinoderms living in Vietnamese seas provided several diterpenes. Among them is laevigatol A (89), which had a moderately antiplasmodial activity with an IC_{50} < 5.0 μ M.^{84,85} Also, a series of diterpene glycosides was obtained from the Caribbean soft coral Pseudopterogorgia elisabethea (Gorgoniidae). Among them pseudopterosin V (90) exhibited an antimalarial activity (IC₅₀ = $2.2 \mu M$) against CQresistant P. falciparum colonies.86

3.3. Other terpenoids

Screening of marine sponges extracts from Spongia, and Ircinia genera revealed thr presence of broad-spectrum antiprotozoal meroterpenes, linear triterpenoid, and squalene, with inhibitory effects on P. falciparum and Trypanosoma. The dorisenone D (91), a dimeric C21 meroterpenoid obtained from Dysidea arenaria sponge (Dysideidae), may become a promising antiplasmodial compound.87

Endoperoxide-containing compounds

One of the most fundamental advances in malaria chemotherapy was the discovery and manufacturing of endoperoxidecontaining drugs. Undoubtedly, the artemisinin discovery was the beginning of research in this area. Artemisinin is a cadinane sesquiterpene lactone establishing a 1,2,4-trioxane moiety isolated from sweet wormwood Artemisia annua L. leaves (Asteraceae). Artemisinin showed nanomolar potency against COresistant Plasmodium strains. The endoperoxide linkage is essential for antimalarial activity. One of the artemisinin

derivatives lacking the endoperoxide bridge showed no antimalarial activity.88

These drugs containing endoperoxide were purported to interact via endoperoxide bond with the iron(II) center of the heme unit released in the food vacuole during the digestion of hemoglobin and lead to peroxide bridge cleavage the consequent formation of oxygen-centered radicals. Because of an intramolecular rearrangement, these reactive species were converted into free C-centered radicals, toxic to the parasite through the alkylation of sensitive macromolecular targets. A sarco-endoplasmic reticulum Ca2+ dependent ATPase of P. falciparum has been proposed as a possible target for these active species.89,90 Although, most likely, artemisinin activity is not mediated by interaction with a single enzyme. It had been proposed that the Fe²⁺-containing species interacting with the endoperoxide bond is not heme.⁹¹ The marine antimalarial drug-containing endoperoxide was divided according to their structural feature into peroxyketal and non-peroxyketal, as demonstrated below, and the chemical structures are listed in Table 4.

4.1. Peroxyketal

Peroxyplakoric acids are the parent compound in the class of peroxyketals/3-alkoxy-1,2-dioxane derivatives, and its methyl esters; peroxyplakoric acids A₃ (92) and B₃ (93). It was extracted from the Okinawan sponge of *Plakortis* sp. (Plakinidae). 92,93 Peroxyketals derivatives showed potent activity in vitro (IC₅₀ = 150 and 120 nM against P. falciparum FCR3) and a good selective toxicity index. The long alkyl side chain in these derivatives is important for antimalarial activity as the synthetic analog containing methyl group instead of the nonadienyl group was completely nonactive. It has been observed that transforming the ester group into an amide group increases in vivo antimalarial potency.94

4.2. Non-peroxyketal

Plakortin (94) is a simple 1,2-dioxane metabolite, and was isolated from Plakortis halichondroides.95 Plakortin and its analogues, named dihydroplakortin (95), 3-epiplakortin (96), plakortide Q (97), plakortide E (98), were re-isolated from the Caribbean sponge Plakortis simplex.96 All these compounds, except plakortide E, displayed good antimalarial activity against (D10) CQ-sensitive and (W2) CQ-resistant strains of P. falciparum, with no cytotoxicity and a more potent activity on the (W2) strain (IC₅₀ in D6 = 1.37 nM, and W2 = 1.11 nM). Plakortide E was found to be inactive. It could be ascribed to a fivemembered ring presence instead of a six-membered ring and/ or the crowded substituents at carbons flanking the endoperoxide linkage. 97,98 Currently, plakortin is among pre-clinically investigated antiplasmodial candidates.99

On the other side, plakortide L (99), was extracted from a Jamaican sponge Plakortis sp.97,98 Plakortide O (100) and plakortide P (101), were isolated from plakortis halichondrioides, presented mild antimalarial activity in vitro (IC₅₀ > 0.023 μ M). Despite of their similarities with the plakortin, the configurational changes around the dioxane ring and/or the differences RSC Advances Review

Table 4 A list of marine-derived endoperoxide-containing antimalarial drugs a

Compound	Antiplasmodial activity (IC ₅₀ value)	Structure	Source	Marine class	Ref.
Peroxyplakoric acids A ₃ (92)	FCR3 = 150 and 120 nM, respectively	H ₃ COOC H ₃ O-O OCH ₃	<i>Plakortis</i> sp.	Sponge	92 and 93
Peroxyplakoric acid B ₃ (93)		H ₃ COOC			
Plakortin (94)	W2 = 0.16 nM	0-0-0			
Dihydroplakortin (95)	D10 = 1.12 μ M W2 = 0.76 μ M	0-0-0			96
3-Epiplakortin (96)	NA	0-0	Plakortis halichondroides, Plakortis simplex	Sponge	96
Plakortide Q (97)	D10 = 1000 nM W2 = 520 nM	0-0 -0			
Plakortide E (98)	D6 = 1.37 nM W2 = 1.11 nM	CO ₂ Me			
Plakortide L (99)	NA	0-0			
Plakortide O (100)	NA	0-0	<i>Plakortis</i> sp.	Sponge	97 and 98
Plakortide P (101)	NA	0-0			
Plakortin diol (102)	FcM29 = 800 nM	Ç			101

Table 4 (Contd.)

Compound	Antiplasmodial activity (IC_{50} value)	Structure	Source	Marine class Ref.
		OHO CO ₂ Me		
		O-O OR		
Plakortin hydroxyl (103) Plakortin methoxy (104)	D10 = 1.26 nM	(103) R = H		
Plakortin acetoxy (105)		$(104) R = CH_3$		
		$(105) R = COCH_3$		
^a NA: not available.				

in the alkyl side chains are responsible for the observed decrease of activity. ¹⁰⁰ It was established that the role of the endoperoxide system in the antimalarial activity of plakortin derivatives is pivotal, as the diol derivative (102) with an open peroxide ring exhibited no antimalarial activity, when the ester group substitution to the corresponding hydroxyl (103), methoxy (104), or acetoxy (105) derivatives affected the antimalarial activity or the selectivity of these derivatives. ¹⁰¹

5. Quinones, polyketides, phenols, and acids

Several phenolic compounds with antimalarial activity have been isolated from different marine sources. For instance, from *Hyrtios* sponge (Thorectidae), 5-oxopuupehenol (**106**) was isolated. It exhibited antimalarial activity against D6 and W2 clones of *P. falciparum*. In addition, three xanthones were isolated from the marine fungi *Chaetomium* sp. (Chaetomiaceae), *i.e.*, chaetoxanthones A (**107**), B (**108**), and C (**109**). Chaetoxanthones A and B contain a dioxane-tetrahydropyran moiety, while chaetoxanthone C is a chlorinated xanthone containing a tetrahydropyran ring. Among them, chaetoxanthone B exhibited a significant antimalarial activity (IC $_{50} = 1.4 \text{ nM}$).

A family of monoterpene-quinones, xestoquinone, and alisiaquinones, showed promising antimalarial activities. Xestoquinone (110) was obtained from a Vanuatu Pacific marine sponge *Xestospongia* sp., while alisiaquinone A (111), B (112), C (113), and alisiaquinol (114) were extracted from an unidentified deep-water Caledonian sponge. Xestoquinone was a selective active inhibitor of a protein kinase (Pfnek-1) of *P. falciparum* with $IC_{50} = 1 \mu M$. Alisiaquinone C presented activity against F32, FcB1, and FcM29 of *P. falciparum* (IC₅₀ = 0.15, 0.21, and 0.08 μM , respectively). ^{104,105} From the marine-derived fungus

Fusarium sp., a series of secondary metabolites such as 9α -hydro-xyhalorosellinia A (115), bostrycin (116), nigrosporin B (117), javanicin (118), and anhydrofusarubin (119) were isolated.¹⁰⁶

Among marine fungi, phytochemical investigation of Halorosellinia oceanica BCC 5149 resulted in isolation of different compounds such as 2-hexylidene-3-methylsuccinic acid, cytochalasin Q, 5-carboxymellein, 2-hexylidene-3-methylsuccinic acid 4-methyl ester, and halorosellinic acid. Cytochalasin Q, 5carboxymellein, halorosellinic acid, and its acetonide derivative showed antimalarial activity with IC₅₀ values of 17, 4, 13, and 19 μg mL⁻¹, respectively. 107 Also, the tetramic acid called vermelhotin was isolated from an unidentified fungus CRI247-01 (a member of the order Pleosporales). Vermelhotin exhibited moderate antiplasmodial activity 1-10 μg mL⁻¹.108 Besides, marine sponges (e.g., a Vanuatu marine sponge Pseudoceratina sp.) produced homogentisic acid derivative acting as protein kinase inhibitors Pfnek-1 with an IC_{50} about 1.8 μM and moderately active in vitro against a FcB1 P. falciparum strain $(IC_{50} = 12 \mu M)$. Homogentisic acid is among pre-clinically investigated antiplasmodial candidates.99

Furthermore, (*S*)-curcuphenol (**120**) is a sesquiterpene phenol isolated from the Jamaican sponge *Didiscus oxeata*, displayed *in vitro* activity against D6 and W2 strains of *P. falci-parum* (MIC = 0.017 and 0.008 μ M, respectively). (2*Z*,6*R*,8*R*,9*E*)[3-ethyl-5-(2-ethyl-hex-3-enyl)-6-methyl-5*H*-furan-2-ylidene]-acetic acid methyl ester (**121**), a polyketide was isolated from *Plakortis angulospiculatus* sponge. It showed mild antimalarial activity against D6 and W2 *P. falciparum* colonies (IC₅₀ = 6.6 nM on both strains). 111,112

Also, gracilioether A–C (122–124), a polyketal peroxide obtained from the crude extract of the deep sea *Agelas gracilis* sponge (Agelasida), presented an antimalarial activity ($IC_{50} = 28.22 \mu M$).¹¹³

RSC Advances Review

Table 5 A list of marine-derived quinones, polyketides, and phenols antimalarial drugs showing their IC_{50} against various strains of *Plasmodium* sp., chemical structure and biogenic source

Compound	Antiplasmodial activity (${ m IC}_{50}$ value)	Structure	Source	Marine class	Ref.
5-Oxopuupehenol (106)	D6 = 5.8 μ M W2 = 3.8 μ M	OH OH	<i>Hyrtios</i> sp.	Sponge	102
Chaetoxanthone A (107) Chaetoxanthone B (108)	K1 = 1.4 nM	(107) R= OH (108) R= H	<i>Chaetomium</i> sp.	Marine fungi	103
Chaetoxanthone C (109)	NA	O OH HO			
Xestoquinone (110)	$FcB1 = 3.0 \; \mu M$		Xestospongia sp.	Sponge	104 and 105
Alisiaquinone A (111) alisiaquinone B (112)	FcMC29 = 8.50, 2.60 µM FcB1 = 7.40, 8.40 µM F32 = 9.10, 7.10 Mm, respectively	R			
		(111) $R = H$ (112) $R = OMe$	Unidentified deep-water	Con a series	104 and
Alisiaquinone C (113)	FcMC29 = 0.08 μ M FcB1 = 0.21 μ M F32 = 0.15 μ M	S O O O O O O	caledonian sponge	Sponge	105
Alisiaquinol (114)	FcMC29 = 7.90 μM FcB1 = 6.40 μM F32 = 9.90 μM	OH OH			
9α-Hydro- xyhalorosellinia A (115)	$K1 = 25 \; \mu M$	OH OH HOHOH	Fusarium sp.	Marine fungi	106
Bostrycin (116) Nigrosporin B (117)	$K1 = 9.8 \mu M (116)$ NA for (117)				

Table 5 (Contd.)

Compound	Antiplasmodial activity (IC ₅₀ value)	Structure	Source	Marine class	Ref.
		O R ₁ OH			
		(116) $R_1=R_2=OH$ (117) $R_1=R_2=OH$			
Javanicin (118)	$K1=12\;\mu M$	O OH			
Anhydrofusarubin (119)	$K1=14~\mu M$	O OH			
(S)-Curcuphenol (120)	$\begin{array}{l} D6 = 0.017 \; \mu M \; W2 \\ = 0.008 \; \mu M \end{array}$	OH	Didiscus oxeata	Sponge	110
(2 <i>Z</i> ,6 <i>R</i> ,8 <i>R</i> ,9 <i>E</i>)[3-ethyl-5-(2-ethyl-hex-3-enyl)-6-methyl-5 <i>H</i> -furan-2-ylidene]-acetic acid methyl ester (121)	D6 = 6.6 nM W2 = 6.6 nM		Plakortis angulospiculatus	Sponge	111 and 112
Gracilioether A (122)	$ItG=28.22~\mu M$	0-0, OH			
Gracilioether B (123)	$ItG=1.56\;\mu M$		Agelas gracilis	Sponge	113
Gracilioether C (124)	$ItG=31.02~\mu M$	HO			
Actinosporin E (125)	$IC_{50} = 0.019$ to 0.028 μM	но он	Actinokineospora spheciospongiae	Actinomycete bacteria	114

Table 5 (Contd.)

Compound	Antiplasmodial activity (IC_{50} value)	Structure		Source	Marine class Ref.
Actinosporin H (126)			HOOH		
Actinosporin G (127)			HO OH		
Tetrangulol (128)			HO		
Capillasterquinone B (129)			HO O OH		

Recently, the angucyclines Actinosporins E, H, G (125–127), Tetrangulol (128), and Capillasterquinone B (129) have been isolated based on antimalarial guided fractionation of the cocultured fermentation of the marine bacterium Actinokineospora spheciospongiae (Pseudonocardiaceae). Nevertheless, these compounds have not been reported under axenic conditions. Upon antimalarial screening, they displayed activity ranging from $\rm IC_{50}=0.019$ to $0.028~\mu M.^{114}$ The chemical structure and source are summarized in Table 5.

6. Macrolides

Bastimolide A, a 40-membered ring polyhydroxy macrolide with 10 stereocentres and a rare *tert*-butyl terminus (130), was obtained from *Okeania hirsuta* marine *Cyanobacterium*. It displayed highly potent activity against four multidrug-resistant TM90-C2A, TM90-C2B, W2, and TM91-C235 (IC $_{50}=0.089, 0.11, 0.18, 0.34$ nM, respectively) *P. falciparum* strains. ¹¹⁵ Bastimolide B (131), polyhydroxy macrolide with a long aliphatic chain containing terminal *tert*-butyl group. It exhibited strong antimalarial activity against CQ-sensitive *P. falciparum* strain HB3 (IC $_{50}=5.7~\mu$ M). ¹¹⁶ X-ray crystallography was used to determine the macrocyclic lactone's planar structure and absolute

configuration, which consists of a 1,3-diol, one 1,3,5-triol, and six 1,5-diols. The rare *tert*-butyl group near the lactone ester in the bastimolide structure is thought to protect the lactone ring against hydrolysis.¹¹⁷

In addition, Palstimolide A, a polyhydroxy macrolide, was obtained from a tropical cyanobacterium Leptolyngbya sp. (Leptolyngbyaceae). Palstimolide A (132) had structural similarities to bastimolides. It displayed potent antimalarial activity against the blood stage of P. falciparum Dd2 strain ($IC_{50} = 172.5$ nM).¹¹⁸

Moreover, bromophycolides A (133), D (134), R (135), S (136), T (137), and U (138), diterpene benzoates macrolides were obtained from *Callophycus serratus* red algae (Onagraceae). Bromophycolides having 15- and 16-membered rings have been demonstrated to be effective antimalarials. No significant effects of the lactone ring size on activity were observed between 15- and 16-membered lactone rings. Some bromophycolides target haem crystallization, implying that haemozoin production is inhibited as a mechanism of action. ¹¹⁹ Bromophycolide A presented good activity against drug-resistant Dd2 strain (IC $_{50}$ = 377 nM), drug-sensitive 3D7, and HB3 strains (IC $_{50}$ = 499 and 493 nM), respectively. ¹²⁰ The chemical structures of these compounds belonging to macrolides are traced in Table 6.

Table 6 A list of marine-derived macrolides antimalarial drugs showing their IC_{50} against various strains of *Plasmodium* sp., chemical structure and biogenic source

Compound	Antiplasmodial activity (${\rm IC}_{50}$ value)	Structure	Source	Marine class	Ref.
Bastimolide A (130)	TM90-C2A = 0.089TM90-C2B = 0.11 W2 = 0.18 TM91-C235 = 0.34 nM	HO OH OH OH	Okeania hirsuta	Cyano- bacteria	115
Bastimolide B (131)	$HB3 = 5.7 \; \mu M$	OH OH OH OH OH OH			116
Palstimolide A (132)	Dd2 = 172.5 nM	OH OH OH OH	<i>Leptolyngbya</i> sp.	Cyan- obacteria	118
Bromophycolide A (133)		Br OH Br			
Bromophycolide D (134)	Dd2 = 377 nM, 3D7 = 499 nM HB3 = 493 nM	HO Br	Callophycus serratus	Red alga	119
Bromophycolide R (135)		HO HO			
Bromophycolide S (136)					

Antiplasmodial activity (IC $_{50}$ Marine Compound value) Structure Source class Ref.

Bromophycolide T (137)

Bromophycolide U (138)

Bromophycolides have been mentioned among drugs that are investigated in pre-clinical trials.⁹⁹

Peptides and depsipeptides

Peptides are short cyclic or acyclic chains between two and fifty amino acids, linked by amide covalent bonds (peptidic bonds). At the same time, depsipeptides are cyclic or acyclic compounds of α -amino and α -hydroxycarboxylic acids linked to each other by esters and amides units. ¹²¹ Several peptides and depsipeptides from marine sources were reported for antiplasmodial activity are, summarized in Table 7.

Even though a good number of peptides and depsipeptides have presented good antimalarial activity, their mechanism of action is not well understood. Some of them displayed a strong inhibitory effect on some key enzymes present in the malaria parasite; moreover, the relation between the inhibition and their antimalarial activity remains unestablished.⁵⁸

7.1. Cyclic peptides

Three new heterocyclic macrocyclic peptides, Balgacyclamides A–C (139–141), were isolated from *Microcystis aeruginosa* (Microcystaceae). Balgacyclamides A and B displayed potent activity against the CQ-resistant strain K1 of *P. falciparum* (IC $_{50}$ = of 9.0 and 8.2 μ M), respectively. ¹²² A class of modified hexacyclopeptides, aerucyclamides A–D (142–145) were obtained from the cyanobacterium *Microcystis aeruginosa*. They presented a potent activity against the K1 strain of *P. falciparum* (IC $_{50}$ = 5.0, 0.7, 2.3, and 6.3 μ M, respectively). ^{123,124} Mollamide B (146),

thiazoline hexapeptides found in the Indonesian tunicate *Didemnum mole* (Didemnidae). It showed a moderate antimalarial activity against D6 and W2 strains (IC₅₀ = 0.0029 and 0.003 μ M, respectively) of *P. falciparum*.¹²⁵

7.2. Acyclic peptides

Dolastatin 10 (147), an acyclic peptide extracted from cyanobacterium Symploca species^{126,127} exhibited strong activity against P. falciparum 3D7 colonies, with $IC_{50} = 74$ nM. In addition, four acyclic lipopeptides, dragonamides A (148), B (149), dragomabin (150), and carmabin A (151) have been isolated from the cyanobacterium Moorea producens (Cyanobacteriaceae) (formerly Lyngbya majuscula). dragomabin, carmabin A and dragonamide A displayed good antimalarial activity ($IC_{50} = 6.0$, 4.3 and 7.7 μ M, respectively). $I^{128,129}$

Morover, malyngamide X (152) is the first (7R)-lyngbic acid connected to a new tripeptide backbone. It was obtained from *Bursatella leachii* (Aplysiidae), a Thai sea hare, presented a moderate antimalarial activity with a half effective dose (ED₅₀) = 5.44 μ M against *P. falciparum* (K1) multidrug-resistant strain. ¹³⁰

7.3. Cyclic depsipeptides

Romidepsin (153), a cyclic depsipeptide histone deacetylase (HDAC) inhibitor, is responsible for the observed anti-Plasmodium activity of *Chromobacterium* species. ^{131,132} Venturamides A (154) and B (155), two compounds from thiazole and oxazole cyclodepsipeptides class obtained from the

A list of marine-derived peptides and depsipeptides antimalarial drugs showing their IC₅₀ against various strains of Plasmodium sp., chemical structure and biogenic source^a This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

Table 7

Ref.		122		123 and 124
Marine class		Cyanobacteria		Cyanobacteria
M				
Source		Microcystis aeruginosa		Microcystis aeruginosa
		NH HN H	O I I I O O O O	S Z H N T
Structure				
Antiplas- modial activity (IC ₅₀ value)	K1 I $G_{50}=$ of 9.0 and 8.2 μM , respectively		NA	$K1 = 5.0, 0.7, 2.3,$ and 6.3 μ M, respectively
Compound	Balgacyclamide A (139)	B (140)	Balgacyclamide C (141)	Aerucyclamide A (142)

Compound	Antiplas- modial activity (IC ₅₀ value)	Structure		Source	Marine class	Ref.
Aerucyclamide B (143)		,,,, O I				
Aerucyclamide C (144)		III O I				
Aerucyclamide D (145)		"" O E	O N H N H			
Mollamide B (146)	$D6 = 0.0029 \ \mu M \ W2 = 0.003 \ \mu M$	N H		Didemnum mole	Tunicate	125

Table 7 (Contd.)					
Compound	Antiplas- modial activity (IC ₅₀ value)	Structure	Source	Marine class	Ref.
Dolastatin 10 (147)	$\mathrm{FCH5 \cdot C2} = 0.1 \; \mathrm{nM}$	S NH O O O O O O O O O O O O O O O O O O	Symploca sp.	Cyanobacteria	126 and 127
Dragonamide A (148)		N N N N N N N N N N N N N N N N N N N			
Dragonamide B (149)	3D7 = 7.7, 7.0, and 6.0 µM	O N N N N N N N N N N N N N N N N N N N	Moorea producens	Cyanobacteria	128 and 129
Dragomabin (150)					

Carmabin A (151)

 $3D7 = 4.3 \, \mu M$

Open Access Article. Published on 02 February 2023. Downloaded on 12/5/2025 1:09:30 PM.	This article is licensed under a Creative Commons Attribution 3.0 Unported Licence	
Open A	(cc)) BY	

Compound	Antiplas- modial activity (IC ₅₀ value)	Structure	Source	Marine class	Ref.
		O-ZOO			
Malyngamide X (152)	$\mathrm{K1ED_{50}} = 5.44~\mu\mathrm{M}$	O HN HO O N	Bursatella leachii	Thai sea hare	130
Romidepsin (153)	~150 nM	O HN O HN O S	Violaceous sp	Cyanobacteria	131 and 132
Venturamide A (154) Venturamide B (155)	$W2=8.2$ and $5.2~\mu M$	O NH NHO O O O O O O O O O O O O O O O O	Oscillatoria sp.	Cyanobacteria	133

•	1	7	٠
٠	,		
	١		
•	2	•	
	ζ	1	
(r	-	١
,	١	•	
١			l
	2	_	
ı		L	
١		7	١
	•	1	
	_	•	
	(•	١
٠	7	ï	
ı	0	(
ı	۰		

Compound	Antiplas- modial activity (IC ₅₀ value)	Structure		Source	Marine class	Ref.
Companeramide A (156)	270 Mrs 052 - 20		HO NI	Donomonia monine		
Companer- amide B (157)	= 1000 and 230 nM		TZ O N N N N N N N N N N N N N N N N N N	Cyanobacterium sp.	Cyanobacteria	134

RSC Advances

Open Access Article. Published on 02 February 2023. Downloaded on 12/5/2025 1:09:30 PM.

BY This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

Table 7 (Contd.)						
Compound	Antiplas- modial activity (IC ₅₀ value)	Structure		Source	Marine class	Ref.
Lagunamide A (158)			HO NH			
Lagunamide B (159)	NF54 $= 190,910$ and 290 nM		HOHO.	Lyngbya majuscule	Cyanobacteria	135 and 136
Lagunamide C (160)		·	HO			

1	7	
	ζ	
•	۲	
	ž	
(0220	
١	_	
1		١
	0	
1	0	
Ī	a	į
ŀ	Ė	

(00)					
Compound	Antiplas- modial activity (IC ₅₀ value)	Structure	Source	Marine class	Ref.
Mollemycin A (161)	3D7 = 7 nM Dd2: 9 nM	HO O O O O O O O O O O O O O O O O O O	Streptomyces sp.	Marine actinomycetes	137
Symplocamide A (162)	$W2=0.95~\mu M$	Br O HN O HN O O HN O O HN O O O O O O O O	Streptomyces ballenaensis and S. bangulaensis	Marine actinomycetes	138 and 139
Actinoramide A (163)	HB3 = 190 nM Cp250 = 210 nM Dd2 = 220 nM 7G8 = 160 nM GB4 = 340 nM				140

-	-
1	c
1	222
i	ĉ
(_
`	Ξ
1	
	٩
÷	c
ŀ	ά
•	

date (contra.)					
Compound	Antiplas- modial activity (IC ₅₀ value)	Structure	Source	Marine class	Ref.
		O HO O O O NZH			
Symplostatin 4 (164)	$3\mathrm{D7~ED_{50}} = 74~\mathrm{nM}$	OCH3	Symploca sp	Cyanobacteria	141 and 142
Gallinamide A (165)	$3D7 = 50.1 \pm 7.6 \text{ nM W2} = 8.4 \mu\text{M}$	NH O IN O	Schizothrix sp.	Cyanobacteria	144
Viridamide A (166)	$W2=5.8~\mu\mathrm{M}$	NH HN O	Oscillatoria nigro-viridis	Cyanobacteria	145

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence. Open Access Article. Published on 02 February 2023. Downloaded on 12/5/2025 1:09:30 PM.

Table 8 A list of marine-derived phosphotriesters, polyether, and steroidal glycosides antimalarial drugs showing their IC50 against various strains of Plasmodium sp., chemical structure and biogenic source^a

Compound	Antiplasmodial activity (IC ₅₀ value)	Structure	Source	Marine class	Ref.
Salinipostin A (167) Salinipostin F (172) Salinipostin I (175)	W2 = 50, 266, and 126 nM	(167) R= Bu (172) R= Pr			
Salinipostin B (168) Salinipostin D (170) Salinipostin G (173) Salinipostin J (176)	W2 = 139, 82, 1.52 µM, respectively (176) NA	(173) $R = Et$ (168) $R = Bu$ (170) $R = i - Bu$ (173) $R = Pr$ (176) $R = Et$	Salinospora sp.	Marine actinomycetes	146
Salinipostin C (169) Salinipostin E (171) Salinipostin H (174) Salinipostin K (177)	W2 = 415, 3.22, 8.70 μM, respectively (177) NA	(169) $R = Bu$ (171) $R = i-Bu$			
Monensin (178)	D6 and W2 0.15 to 0.3 nM	HOOH OF OF OH OF OH	Streptomyces cinnamonensis	Marine actinomycetes	147 and 148

Compound	Antiplasmodial activity (IC ₅₀ value)	Structure	Source	Marine class	Ref.
Pandaroside E (179)		HO HO HO HO HO			
Pandaroside G (180)	$W2 = 0.78.0.05$ and $0.038~\mu M$	HO OH OINX	Pandaros acanthifolium	Sponge	150
Pandaroside H (181)		HO OH HO OH HO OH			

 a NA: not available.

Review RSC Advances

cyanobacterium *Oscillatoria* sp., presented activity against (IC_{50} = 8.2 and 5.2 μ M, respectively) *P. falciparum* W2 strain. ¹³³

Two cyclodepsipeptides, companeramides A (156) and B (157), were obtained from a Panamanian marine Cyanobacterium sp. (Cyanobacteriaceae). Exhibited strong antiplasmodial activity against D6 strain in vitro (IC₅₀ = 0.57 and 0.22 μ M, respectively).134 Three cytotoxic cyclic depsipeptides, lagunamides A-C (158-160), were isolated from Lyngbya majuscule (Oscillatoriaceae). The planar lagunamide macrocyclic scafold consists of peptide and polyketide substructures, and the main diferences are in the polyketide part. Lagunamides A and B are 26-membered macrocycles, while lagumanide C has an additional methylene carbon in the polyketide structure. Lagunamides A-C, showed potent activity against P. falciparum NF54 strain (IC₅₀ = 0.19, 0.91, and 0.29 μ M, respectively).^{135,136} The double bond in the side chain of lagunamide B might be responsible for the lower activity.117 Mollemycin A (161), a glycol-hexadepsipeptide-polyketide isolated from a marinederived Streptomyces sp. CMBM0244 (Streptomycetaceae), exhibited a potent and selective growth inhibitory activity against drug-sensitive 3D7 and multidrug-resistant Dd2 clones of P. falciparum (IC₅₀ = 7.0 and 9.0 nM, respectively). ¹³⁷ Symplocamide A (162), cyclodepsipeptide was extracted from the marine Cyanobacterium symploca sp. (Cyanobacteriaceae), showed potent antimalarial activity against W2 strain (IC₅₀ = 0.95 μM) of *P. falciparum*. ^{138,139} Cyclic tetrapeptide, actinoramide A (163) was isolated from marine actinomycetes Streptomyces ballenaensis, and S. bangulaensis (Streptomycetaceae). This tetrapeptide had potent activity against clones of drug-resistant P. falciparum including Cp250, Dd2, 7G8, and GB4 ($IC_{50} = 210$, 220, 160, and 340 nM, respectively) and drug-sensitive HB3 clone (IC₅₀ = 190 nM). 140

7.4. Acyclic depsipeptides

Symplostatin 4 (164), an acyclic depsipeptide extracted from Symploca sp. (Microcoleaceae). Symplostatin 4 showed a significant activity against 3D7 strain (ED₅₀ = 74 nM) of P. falciparum. It displayed its activity on P. falciparum falcipains in infected red blood cells, indicating inhibition of the hemoglobin degradation pathway as a possible mode of action. 143

A further cyanobacterial acyclic depsipeptide derivative, named gallinamide A (165) obtained from tropical reef *Schizothrix* sp. (Cyprinidae). Gallinamide A showed a moderate *in vitro* antimalarial activity against the W2 strain ($IC_{50} = 8.4 \mu M$) of *P. falciparum*.¹⁴⁴ Viridamide (166), a lipodepsipeptide obtained from the cyanobacterium *Oscillatoria nigro-viridis* (Oscillatoriaceae), showed the activity against *P. falciparum* ($IC_{50} = 5.8 \mu M$).¹⁴⁵

8. Phosphotriesters

A new class of antimalarials with long-chain bicyclic phosphotriesters, salinipostins A–K (167–177), Table 8, were obtained from *Salinospora* sp. bacteria (Micromonosporaceae). SAR fndings indicated that an increase in alkyl chain length attached to the phosphoester oxygen and vinyl carbon led to

increased activity while branching of the alkyl causes a slight reduction in activity. The most active compound salinipostin A, did not afect parasite schizonts, indicating that it acts by disrupting the processes required for the establishment or growth of intracellular parasites. Salinispostin A did not inhibit haemozoin formation but cause cellular disorganization and disintegration of internal structure. These compounds showed different activity against *P. falciparum* W2 strain. Salinipostins A and D displayed the most potent activity (IC $_{50}=50$ and 82 nM) followed by salinipostins I, B, F, C, G, E and H (IC $_{50}=0.126,\,0.139,\,0.266,\,0.1415,\,1.52,\,3.22,\,8.70\,\mu\text{M})$, respectively. Only, salinipostins K and J displayed weak activity. The salinipostins K and J displayed weak activity.

9. Miscellaneous compounds

9.1. Polyethers

A polyether ionophore isolated from *Streptomyces cinnamonensis* (Streptomycetaceae) named Monensin (178), Table 8. Monensin has been displayed a strong antimalarial activity against *P. falciparum*. ^{147,148} In a recent study, a polyether metabolite was extracted from Streptomyces sp. strain H668. This polyether displayed *in vitro* antimalarial activity against both D6 and W2 strains of P. falciparum with IC_{50} values from 0.15 to 0.3 nM. ¹⁴⁹

9.2. Steroid glycosides

The steroidal glycosides pandaroside E (179), G (180), and H (181) (Table 8), had been isolated from the Caribbean sponge $Pandaros\ acanthifolium\ (Microcionidae)$. They strongly inhibited the growth of $P.\ falciparum$ at low sub-micromolar concentrations. ¹⁵⁰

10. Conclusion

Malaria is among the crucial VBDs affecting the global health, based on the WHO official reports. It is easily progressed to impairment of important human body organs, including the liver, and death in case improper diagnosis and treatment. This emergency has acquired a special interest among health care providers and researchers to find more effective and safer medicaments, especially against the multi-resistant strain of Plasmodium sp. for the people of the developing countries. Particularly, natural-derived treatment of infectious diseases, including malaria is still the most convenient, safe, effective, and diverse. Marine organisms have attracted great potential in the last few decades as a promising and non-traditional source of bioactive compounds. Moreover, recent technological advances have led to isolate and identify thousands of marinederived compounds belonging to various chemical classes. A total of 181 compounds derived from different marine sources, including sponges, cyanobacteria, marine algae, and actinomycetes, were reviewed in the current research possessing potential antimalarial activities with unique SAR and targeting different growth stages, including ring and trophozoite stage. More than half of the compounds belong to three major chemical classes comprising alkaloids, terpenoids, and polyketides. Such chemical diversity, potency, and less cytotoxicity **RSC Advances** Review

are recognized as great start point for further SAR and clinical investigations of antimalarial candidates. The current article assumed that marine-derived natural products can also open up novel resources of bioactive compounds for novel candidates for management of other infectious diseases, exploring the oceans and seas treasures. Three compounds, including bromophycolides, plakortin, and homogentisic acid, are investigated as antimalarial drugs in pre-clinical trials and may be approved and marketed soon.

Conflicts of interest

Authors declare that there are no known conflicts of interest associated with this publication and there has been no significant financial support for this work that could have influenced its outcome.

List of abbreviations

CQ Chloroquine

 EC_{50} Half maximal effective concentration

Half maximal effective dose ED_{50} FΒ Iron-protoporphyrin IX

Food and Drug Administration **FDA**

GSK-3P Glycogen synthase 3

 IC_{50} Half-maximal inhibitory concentration MIC minimum inhibitory concentration SAR Structure-activity relationships

VBDs Vector-borne diseases P. falciparum D6 A West African clone P. falciparum W2 The Indochina clone W2 WHO World Health Organization

References

- 1 WHO, Global Vector borne diseases, World Health Organization, 2020.
- 2 S. Degroote, K. Zinszer and V. Ridde, Infect. Dis. Poverty, 2018, 7, 96.
- 3 T. R. Zolnikov, in Autoethnographies on the Environment and Human Health, Springer International Publishing, Cham, 2018, pp. 113-126, DOI: 10.1007/978-3-319-69026-1_9.
- 4 K. D. Lafferty, Ecology, 2009, 90, 888-900.
- 5 World Health Organization WHO, World malaria report, World Health Organization, Geneva, Switzerland, Licence: CC BY-NC-SA 3.0 IGO, 2019.
- 6 WHO, Key facts; news room, Malaria, 2020.
- 7 E. Fattorusso and O. Taglialatela-Scafati, Mar. Drugs, 2009, 7, 130-152.
- 8 C. J. Sutherland, N. Tanomsing, D. Nolder, M. Oguike, C. Jennison, S. Pukrittayakamee, C. Dolecek, T. T. Hien, V. E. Do Rosário and A. P. Arez, J. Infect. Dis., 2010, 201, 1544-1550.
- 9 C. Fançony, A. Soares, J. Lavinha, H. Barros and M. Brito, Int. J. Environ. Res. Public Health, 2019, 16, 466.
- 10 F. Di Gennaro, C. Marotta, P. Locantore, D. Pizzol and G. Putoto, Trop. Med. Int. Health, 2020, 5, 141.

- 11 S. Suleman, T. Beyene Tufa, D. Kebebe, S. Belew, Y. Mekonnen, F. Gashe, S. Mussa, E. Wynendaele, L. Duchateau and B. De Spiegeleer, J. Ethnopharmacol., 2018, 213, 262-279.
- 12 A. Mpimbaza, S. Nayiga, G. Ndeezi, P. J. Rosenthal, C. Karamagi and A. Katahoire, PLoS One, 2019, 14, e0217262.
- 13 G. M. de Jong, M. B. B. McCall, W. A. Dik, R. T. Urbanus, L. J. Wammes, R. Koelewijn, R. W. Sauerwein, A. Verbon, J. J. van Hellemond and P. J. J. van Genderen, Cytokine, 2020, 125, 154838.
- 14 F. R. Ochsendorf, J. Dtsch Dermatol. Ges., 2010, 8, 829-845.
- 15 P. F. Uzor, V. D. Prasasty and C. O. Agubata, J. Evidence-Based Integr. Med., 2020, 2020, 9385125.
- 16 D. G. I. Kingston and M. B. Cassera, Prog. Chem. Org. Nat. Prod., 2022, 117, 1-106.
- 17 F. Mojab, Avicenna J. Phytomed., 2012, 2, 52.
- 18 C. Alves, J. Silva, S. Pinteus, H. Gaspar, M. C. Alpoim, L. M. Botana and R. Pedrosa, Front. Pharmacol., 2018, 9,
- 19 A. Zayed, T. Hahn, D. Finkelmeier, A. Burger-Kentischer, S. Rupp, R. Krämer and R. Ulber, Process Biochem., 2019, 81, 182-187.
- 20 A. Zayed, K. Muffler, T. Hahn, S. Rupp, D. Finkelmeier, A. Burger-Kentischer and R. Ulber, Mar. Drugs, 2016, 14, 79.
- 21 U. Lindequist, Biomol. Ther., 2016, 24, 561-571.
- 22 D. G. Nair, R. Weiskirchen and S. K. Al-Musharafi, Acta Pharmacol. Sin., 2015, 36, 158-170.
- 23 S. A. Dyshlovoy and F. Honecker, Mar. Drugs, 2020, 18(1),
- 24 A. Zayed, PhD thesis, TU Kaiserslautern, Kaiserslautern, Germany, 2018.
- 25 J. A. Nweze, F. N. Mbaoji, Y.-M. Li, L.-Y. Yang, S.-S. Huang, V. N. Chigor, E. A. Eze, L.-X. Pan, T. Zhang and D.-F. Yang, Infect. Dis. Poverty, 2021, 10, 9.
- 26 P. J. McCarthy, B. F. Roberts, A. Carbonell, J. Roberts, A. E. Wright and D. Chakrabarti, Trop. Med. Int. Health, 2019, 4, 103.
- 27 T. Kubota, S.-i. Kurimoto and J. i. Kobayashi, Alkaloids Chem. Biol., 2020, 84, 1-124.
- 28 J. Dai, W. Dan, U. Schneider and J. Wang, Eur. J. Med. Chem., 2018, 157, 622-656.
- 29 P. Ashok, S. Ganguly and S. Murugesan, Drug Discovery Today, 2014, 19, 1781-1791.
- 30 D. Skropeta, N. Pastro and A. Zivanovic, Mar. Drugs, 2011, 9, 2131-2154.
- 31 P. Ashok, H. Lathiya and S. Murugesan, Eur. J. Med. Chem., 2015, 97, 928-936.
- 32 N. H. Shady, M. A. Fouad, M. Salah Kamel, T. Schirmeister and U. R. Abdelmohsen, Mar. Drugs, 2019, 17, 19.
- 33 Y. Takahashi, T. Kubota, J. Fromont and J. i. Kobayashi, Org. Lett., 2009, 11, 21-24.
- 34 M. Yamada, Y. Takahashi, T. Kubota, J. Fromont, A. Ishiyama, K. Otoguro, H. Yamada, S. Ōmura and J. i. Kobayashi, Tetrahedron, 2009, 65, 2313-2317.

Review

35 T. Kubota, K. Nakamura, S.-i. Kurimoto, K. Sakai, J. Fromont, T. Gonoi and J. i. Kobayashi, *J. Nat. Prod.*, 2017, **80**, 1196–1199.

- 36 N. H. Shady, M. A. Fouad, S. Ahmed, S. M. Pimentel-Elardo, J. R. Nodwell, M. S. Kamel and U. R. Abdelmohsen, J. Antibiot., 2018, 71, 1036–1039.
- 37 G. Kirsch, G. M. König, A. D. Wright and R. Kaminsky, *J. Nat. Prod.*, 2000, **63**, 825–829.
- 38 M. E. Zhidkov, M. A. Sidorova and I. A. Lyakhova, *Tetrahedron Lett.*, 2018, 59, 1417–1420.
- 39 H. Huang, Y. Yao, Z. He, T. Yang, J. Ma, X. Tian, Y. Li, C. Huang, X. Chen and W. Li, J. Nat. Prod., 2011, 74, 2122–2127.
- 40 Y. Shi, Y. Moazami and J. G. Pierce, *Bioorg. Med. Chem.*, 2017, 25, 2817–2824.
- 41 P.-E. Campos, J.-L. Wolfender, E. F. Queiroz, L. Marcourt, A. Al-Mourabit, M. Frédérich, A. Bordignon, N. De Voogd, B. Illien and A. Gauvin-Bialecki, *J. Nat. Prod.*, 2017, 80, 1404–1410.
- 42 S. T. Chan, A. N. Pearce, M. J. Page, M. Kaiser and B. R. Copp, J. Nat. Prod., 2011, 74, 1972–1979.
- 43 R. Finlayson, A. N. Pearce, M. J. Page, M. Kaiser, M.-L. Bourguet-Kondracki, J. L. Harper, V. L. Webb and B. R. Copp, J. Nat. Prod., 2011, 74, 888–892.
- 44 K. Dineshkumar, A. Vasudevan and W. Hopper, *Comb. Chem. High Throughput Screening*, 2017, **20**, 3–19.
- 45 J. Prudhomme, E. McDaniel, N. Ponts, S. Bertani, W. Fenical, P. Jensen and K. Le Roch, *PLoS One*, 2008, 3, e2335.
- 46 Y. Lee, C. Phat and S.-C. Hong, Peptides, 2017, 95, 94-105.
- 47 H.-S. Lee and G.-S. Jeong, *Molecules*, 2020, 25, 5031.
- 48 L. B. Marx and J. W. Burton, *Chem.-Eur. J.*, 2018, **24**, 6747–6754.
- 49 H. Gholami, A. Kulshrestha, O. K. Favor, R. J. Staples and B. Borhan, *Angew. Chem.*, 2019, **131**, 10216–10219.
- 50 E. T. Oluwabusola, J. N. Tabudravu, K. S. Al Maqbali, F. Annang, G. Pérez-Moreno, F. Reyes and M. Jaspars, *Chem. Biodiversity*, 2020, 17, e2000335.
- 51 B. D. Hikmawan, S. Wahyuono and E. P. Setyowati, *J. Appl. Pharm. Sci.*, 2020, **10**, 142–157.
- 52 M. Xu, K. T. Andrews, G. W. Birrell, T. L. Tran, D. Camp, R. A. Davis and R. J. Quinn, *Bioorg. Med. Chem. Lett.*, 2011, 21, 846–848.
- 53 X. Yang, R. A. Davis, M. S. Buchanan, S. Duffy, V. M. Avery, D. Camp and R. J. Quinn, J. Nat. Prod., 2010, 73, 985–987.
- 54 S.-i. Kurimoto, T. Ohno, R. Hokari, A. Ishiyama, M. Iwatsuki, S. Ōmura, J. i. Kobayashi and T. Kubota, *Mar. Drugs*, 2018, 16, 463.
- 55 R. A. Davis, M. S. Buchanan, S. Duffy, V. M. Avery, S. A. Charman, W. N. Charman, K. L. White, D. M. Shackleford, M. D. Edstein and K. T. Andrews, J. Med. Chem., 2012, 55, 5851–5858.
- 56 M. I. Bilan, A. A. Grachev, A. S. Shashkov, M. Kelly, C. J. Sanderson, N. E. Nifantiev and A. I. Usov, *Carbohydr. Res.*, 2010, 345, 2038–2047.
- 57 B. Singh, G. Mal, S. K. Gautam and M. Mukesh, in *Advances* in *Animal Biotechnology*, Springer, 2019, pp. 429–439.

- 58 J. Fotie, in *Discovery and Development of Therapeutics from Natural Products Against Neglected Tropical Diseases*,
 Elsevier, 2019, pp. 7–47.
- 59 R. A. Davis, S. Duffy, S. Fletcher, V. M. Avery and R. J. Quinn, J. Org. Chem., 2013, 78, 9608–9613.
- 60 E. Avilés and A. D. Rodríguez, Org. Lett., 2010, 12, 5290–5293.
- 61 W. Balansa, S. I. M. Wodi, F. J. Rieuwpassa and F. G. Ijong, *Biodiversitas*, 2020, 21(2), 699–706.
- 62 E. Gros, A. Al-Mourabit, M.-T. r. s. Martin, J. Sorres, J. Vacelet, M. Frederich, M. Aknin, Y. Kashman and A. Gauvin-Bialecki, J. Nat. Prod., 2014, 77, 818–823.
- 63 A. Ludwiczuk, K. Skalicka-Woźniak and M. Georgiev, in *Pharmacognosy*, Elsevier, 2017, pp. 233–266.
- 64 J. Emsermann, U. Kauhl and T. Opatz, *Mar. Drugs*, 2016, 14, 16.
- 65 J. Achan, A. O. Talisuna, A. Erhart, A. Yeka, J. K. Tibenderana, F. N. Baliraine, P. J. Rosenthal and U. D'Alessandro, *Malar. J.*, 2011, 10, 144.
- 66 S. Sumarna, M. Azis, S. S. Sangadji and I. N. Idrus, *Techno: Jurnal Penelitian*, 2019, **8**, 328–333.
- 67 K. Kyei-Baffour, D. C. Davis, Z. Boskovic, N. Kato and M. Dai, *Bioorg. Med. Chem.*, 2020, 28, 115678.
- 68 M. E. Daub, J. Prudhomme, C. Ben Mamoun, K. G. Le Roch and C. D. Vanderwal, ACS Med. Chem. Lett., 2017, 8, 355– 360.
- 69 F. Le Bideau, M. Kousara, L. Chen, L. Wei and F. o. Dumas, *Chem. Rev.*, 2017, **117**, 6110–6159.
- 70 B. Di Blasio, E. Fattorusso, S. Magno, L. Mayol, C. Pedone,C. Santacroce and D. Sica, *Tetrahedron*, 1976, 32, 473-478.
- 71 E. Fattorusso, S. Magno, L. Mayol, C. Santacroce and D. Sica, *Tetrahedron*, 1974, 30, 3911–3913.
- 72 E. Fattorusso, S. Magno, L. Mayol, C. Santacroce and D. Sica, *Tetrahedron*, 1975, **31**, 269–270.
- 73 N. Manivel, S. K. Shukla and S. Muthuraman, *J. Mar. Biotechnol.*, 2020, 4, 2365–2409.
- 74 E. Avilés, J. Prudhomme, K. G. Le Roch, S. G. Franzblau, K. Chandrasena, A. M. S. Mayer and A. D. Rodríguez, *Bioorg. Med. Chem. Lett.*, 2015, 25, 5339–5343.
- 75 K. Nieves, J. Prudhomme, K. G. Le Roch, S. G. Franzblau and A. D. Rodríguez, *Bioorg. Med. Chem. Lett.*, 2016, 26, 854–857.
- 76 M. Dilrukshi, S. Hettiarachi and E. Edirisinghe, 10th International Research Conference 2017, General Sir John Kotelawala Defence University, 2017, pp. 452–459.
- 77 M. T. Cabrita, C. Vale and A. P. Rauter, *Mar. Drugs*, 2010, 8, 2301–2317.
- 78 N. Chadha and O. Silakari, *Eur. J. Med. Chem.*, 2017, **134**, 159–184.
- 79 S. S. Ebada, N. de Voogd, R. Kalscheuer, W. E. Müller and P. Proksch, *Phytochem. Lett.*, 2017, 22, 154–158.
- 80 A. M. White, G. K. Pierens, T. Skinner-Adams, K. T. Andrews, P. V. Bernhardt, E. H. Krenske, E. Mollo and M. J. Garson, *J. Nat. Prod.*, 2015, 78, 1422–1427.
- 81 A. D. Wright, A. McCluskey, M. J. Robertson, K. A. MacGregor, C. P. Gordon and J. Guenther, *Org. Biomol. Chem.*, 2011, 9, 400–407.

RSC Advances Review

- 82 E. Avilés, J. Prudhomme, K. G. Le Roch and A. D. Rodríguez, Tetrahedron, 2015, 71, 487-494.
- 83 J. Jongaramruong and N. Kongkam, J. Asian Nat. Prod. Res., 2007, 9, 743-751.
- 84 N. P. Thao, J. H. No, B. T. T. Luyen, G. Yang, S. Y. Byun, J. Goo, K. T. Kim, N. X. Cuong, N. H. Nam and C. Van Minh, Molecules, 2014, 19, 7869-7880.
- 85 N. P. Thao, B. T. T. Luyen, R. Brun, M. Kaiser, P. Van Kiem, C. Van Minh, T. J. Schmidt, J. S. Kang and Y. H. Kim, Molecules, 2015, 20, 12459-12468.
- 86 I. I. Rodríguez, Y.-P. Shi, O. J. García, A. D. Rodríguez, A. M. Mayer, J. A. Sánchez, E. Ortega-Barria and J. González, J. Nat. Prod., 2004, 67, 1672–1680.
- 87 W.-H. Jiao, B.-H. Cheng, G.-D. Chen, G.-H. Shi, J. Li, T.-Y. Hu and H.-W. Lin, Org. Lett., 2018, 20, 3092-3095.
- 88 C.-x. Liu, Chin. Herb. Med., 2017, 9, 101-114.
- 89 P. M. O'neill, V. E. Barton and S. A. Ward, Molecules, 2010, **15**, 1705–1721.
- 90 F. L. Wang, Y. B. Ji and B. Yang, Exp. Ther. Med., 2020, 20, 630-636
- 91 E. Fattorusso and O. Taglialatela-Scafati, Phytochem. Rev., 2010, 9, 515-524.
- 92 B.-N. Han, L.-L. Hong, B.-B. Gu, Y.-T. Sun, J. Wang, J.-T. Liu and H.-W. Lin, in Symbiotic Microbiomes of Coral Reefs Sponges and Corals, Springer, 2019, pp. 329-463.
- 93 D. W. Triningsih, J. Tanaka and A. Trianto, Res. J. Biotechnol., 2019, 14, 126-130.
- 94 N. Murakami, M. Kawanishi, H. M. Mostagul, J. Li, S. Itagaki, T. Horii and M. Kobayashi, Bioorg. Med. Chem. Lett., 2003, 13, 4081-4084.
- 95 T. Schirmeister, S. Oli, H. Wu, G. Della Sala, V. Costantino, E.-J. Seo and T. Efferth, Mar. Drugs, 2017, 15, 63.
- 96 C. Campagnuolo, Ε. Fattorusso, O. Taglialatela-Scafati, N. Basilico, S. Parapini and D. Taramelli, Eur. J. Org. Chem., 2005, 5077-5083.
- 97 E. Fattorusso, S. Parapini, C. Campagnuolo, N. Basilico, O. Taglialatela-Scafati and D. Taramelli, J. Antimicrob. Chemother., 2002, 50, 883-888.
- 98 S. Oli, U. R. Abdelmohsen, U. Hentschel and T. Schirmeister, Mar. Drugs, 2014, 12, 2614-2622.
- 99 C. Prashar, N. Thakur, S. Chakraborti, S. S. A. Hussain, K. Vashisht and K. C. Pandey, Front. Drug Discovery, 2022, 2, 1065231.
- 100 M. Jiménez and S. Garzon, J. Nat. Prod., 2003, 66(5), 655-
- 101 C. Fattorusso, G. Campiani, B. Catalanotti, M. Persico, N. Basilico, S. Parapini, D. Taramelli, C. Campagnuolo, E. Fattorusso and A. Romano, J. Med. Chem., 2006, 49, 7088-7094.
- 102 H.-S. Wang, H.-J. Li, X. Nan, Y.-Y. Luo and Y.-C. Wu, J. Org. Chem., 2017, 82, 12914-12919.
- 103 A. Pontius, A. Krick, S. Kehraus, R. Brun and G. M. König, J. Nat. Prod., 2008, 71, 1579-1584.
- 104 D. Desoubzdanne, L. Marcourt, R. Raux, S. Chevalley, D. Dorin, C. Doerig, A. Valentin, F. Ausseil and C. Debitus, J. Nat. Prod., 2008, 71, 1189-1192.

- 105 D. Laurent, V. Jullian, A. Parenty, M. Knibiehler, D. Dorin, S. Schmitt, O. Lozach, N. Lebouvier, M. Frostin and F. Alby, Bioorg. Med. Chem., 2006, 14, 4477-4482.
- 106 K. Trisuwan, N. Khamthong, V. Rukachaisirikul, S. Phongpaichit, S. Preedanon and J. Sakayaroj, J. Nat. Prod., 2010, 73, 1507-1511.
- 107 M. Chinworrungsee, P. Kittakoop, M. Isaka, A. Rungrod, M. Tanticharoen and Y. Thebtaranonth, Bioorg. Med. Chem. Lett., 2001, 11, 1965-1969.
- 108 C. Kasettrathat, N. Ngamrojanavanich, S. Wiyakrutta, S. Ruchirawat and P. Kittakoop, C. Mahidol, Phytochemistry, 2008, 69, 2621-2626.
- 109 N. Lebouvier, V. Jullian, I. Desvignes, S. Maurel, A. Parenty, D. Dorin-Semblat, C. Doerig, M. Sauvain and D. Laurent, Mar. Drugs, 2009, 7, 640-653.
- 110 M. Gautam, D. Acharya, Z. Ali Bhat and D. Kumar, Nat. Prod. I., 2017, 7, 84-96.
- 111 X.-F. Liu, Y. Shen, F. Yang, M. T. Hamann, W.-H. Jiao, H.-J. Zhang, W.-S. Chen and H.-W. Lin, Tetrahedron, 2012, 68, 4635-4640.
- 112 R. d. A. Epifanio, L. S. Pinheiro and N. C. Alves, J. Braz. Chem. Soc., 2005, 16, 1367-1371.
- 113 R. Ueoka, Y. Nakao, S. Kawatsu, J. Yaegashi, Y. Matsumoto, S. Matsunaga, K. Furihata, R. W. van Soest and N. Fusetani, J. Org. Chem., 2009, 74, 4203-4207.
- 114 H. A. Alhadrami, B. Thissera, M. H. Hassan, F. A. Behery, C. J. Ngwa, H. M. Hassan, G. Pradel, U. R. Abdelmohsen and M. E. Rateb, Mar. Drugs, 2021, 19, 109.
- 115 C.-L. Shao, R. G. Linington, M. J. Balunas, A. Centeno, P. Boudreau, C. Zhang, N. Engene, C. Spadafora, T. S. Mutka and D. E. Kyle, J. Org. Chem., 2015, 80, 7849-7855.
- 116 C.-L. Shao, X.-F. Mou, F. Cao, C. Spadafora, E. Glukhov, L. Gerwick, C.-Y. Wang and W. H. Gerwick, J. Nat. Prod., 2018, 81, 211-215.
- 117 N. Tajuddeen and F. R. Van Heerden, Malar. J., 2019, 18, 1-62.
- 118 L. Keller, J. L. Siqueira-Neto, J. M. Souza, K. Eribez, G. M. LaMonte, J. E. Smith and W. H. Gerwick, Molecules, 2020, 25, 1604.
- 119 E. P. Stout, S. Cervantes, J. Prudhomme, S. France, J. J. La Clair, K. Le Roch and J. Kubanek, ChemMedChem, 2011, 6, 1572.
- 120 A.-S. Lin, E. P. Stout, J. Prudhomme, K. L. Roch, C. R. Fairchild, S. G. Franzblau, W. Aalbersberg, M. E. Hay and J. Kubanek, J. Nat. Prod., 2010, 73, 275-278.
- 121 A. Gupta, A. Mishra and N. Puri, J. Biotechnol., 2017, 259,
- 122 C. Portmann, S. Sieber, S. Wirthensohn, J. F. Blom, L. Da Silva, E. Baudat, M. Kaiser, R. Brun and K. Gademann, J. Nat. Prod., 2014, 77, 557-562.
- 123 C. Portmann, J. F. Blom, K. Gademann and F. Jüttner, J. Nat. Prod., 2008, 71, 1193-1196.
- 124 C. Portmann, J. F. Blom, M. Kaiser, R. Brun, F. Jüttner and K. Gademann, J. Nat. Prod., 2008, 71, 1891-1896.

- 125 M. S. Donia, B. Wang, D. C. Dunbar, P. V. Desai, A. Patny, M. Avery and M. T. Hamann, *J. Nat. Prod.*, 2008, 71, 941– 945.
- 126 L. T. Tan and M. Y. Phyo, Molecules, 2020, 25, 2197.
- 127 B. Nowruzi, G. Sarvari and S. Blanco, *InHandb. Algal Sci. Microbiol. Technol. Med*, 2020, 441–454.
- 128 Y. Mi, J. Zhang, S. He and X. Yan, Mar. Drugs, 2017, 15, 132.
- 129 N. Engene, E. C. Rottacker, J. Kaštovský, T. Byrum, H. Choi, M. H. Ellisman, J. Komárek and W. H. Gerwick, *Int. J. Syst. Evol. Microbiol.*, 2012, **62**, 1171.
- 130 S. Suntornchashwej, K. Suwanborirux, K. Koga and M. Isobe, *Chem. Asian J.*, 2007, 2, 114–122.
- 131 R. G. Saraiva, C. R. Huitt-Roehl, A. Tripathi, Y.-Q. Cheng, J. Bosch, C. A. Townsend and G. Dimopoulos, *Sci. Rep.*, 2018, 8, 1–14.
- 132 K. M. VanderMolen, W. McCulloch, C. J. Pearce and N. H. Oberlies, *J. Antibiot.*, 2011, **64**, 525–531.
- 133 R. G. Linington, J. González, L.-D. Urena, L. I. Romero, E. Ortega-Barría and W. H. Gerwick, J. Nat. Prod., 2007, 70, 397–401.
- 134 O. B. Vining, R. A. Medina, E. A. Mitchell, P. Videau, D. Li, J. D. Serrill, J. X. Kelly, W. H. Gerwick, P. J. Proteau and J. E. Ishmael, J. Nat. Prod., 2015, 78, 413–420.
- 135 A. Tripathi, J. Puddick, M. R. Prinsep, M. Rottmann, K. P. Chan, D. Y.-K. Chen and L. T. Tan, *Phytochemistry*, 2011, 72, 2369–2375.
- 136 A. Tripathi, J. Puddick, M. R. Prinsep, M. Rottmann and L. T. Tan, *J. Nat. Prod.*, 2010, 73, 1810–1814.
- 137 R. Raju, Z. G. Khalil, A. M. Piggott, A. Blumenthal, D. L. Gardiner, T. S. Skinner-Adams and R. J. Capon, *Org. Lett.*, 2014, **16**, 1716–1719.
- 138 R. G. Linington, D. J. Edwards, C. F. Shuman, K. L. McPhail, T. Matainaho and W. H. Gerwick, *J. Nat. Prod.*, 2008, **71**, 22–27.

- 139 J. Huang, Y. Zhang, L. Dong, Q. Gao, L. Yin, H. Quan, R. Chen, X. Fu and D. Lin, *J. Ethnopharmacol.*, 2018, 213, 280–301.
- 140 K. C.-C. Cheng, S. Cao, A. Raveh, R. MacArthur, P. Dranchak, G. Chlipala, M. T. Okoneski, R. Guha, R. T. Eastman and J. Yuan, J. Nat. Prod., 2015, 78, 2411– 2422.
- 141 T. Conroy, J. T. Guo, N. H. Hunt and R. J. Payne, *Org. Lett.*, 2010, **12**, 5576–5579.
- 142 H. Luesch, W. Y. Yoshida, R. E. Moore, V. J. Paul, S. L. Mooberry and T. H. Corbett, *J. Nat. Prod.*, 2002, 65, 16–20.
- 143 S. C. Stolze, E. Deu, F. Kaschani, N. Li, B. I. Florea, K. H. Richau, T. Colby, R. A. van der Hoorn, H. S. Overkleeft and M. Bogyo, *Chem. Biol.*, 2012, 19, 1546–1555.
- 144 R. G. Linington, B. R. Clark, E. E. Trimble, A. Almanza, L.-D. Ureña, D. E. Kyle and W. H. Gerwick, *J. Nat. Prod.*, 2009, 72, 14–17.
- 145 T. L. Simmons, N. Engene, L. D. Ureña, L. I. Romero, E. Ortega-Barría, L. Gerwick and W. H. Gerwick, *J. Nat. Prod.*, 2008, **71**, 1544–1550.
- 146 C. J. Schulze, G. Navarro, D. Ebert, J. DeRisi and R. G. Linington, *J. Org. Chem.*, 2015, **80**, 1312–1320.
- 147 C.-Y. Lin, Y. Zhang, J.-H. Wu, R.-H. Xie, J. Qiao and G.-R. Zhao, *Microorganisms*, 2020, **8**, 271.
- 148 V. Rajendran, H. S. Ilamathi, S. Dutt, T. Lakshminarayana and P. C. Ghosh, *Curr. Top. Med. Chem.*, 2018, **18**, 1976–1986.
- 149 M. Na, D. A. Meujo, D. Kevin, M. T. Hamann, M. Anderson and R. T. Hill, *Tetrahedron Lett.*, 2008, **49**, 6282–6285.
- 150 E. L. Regalado, D. Tasdemir, M. Kaiser, N. Cachet, P. Amade and O. P. Thomas, J. Nat. Prod., 2010, 73, 1404–1410.