


 Cite this: *RSC Adv.*, 2023, **13**, 570

 Received 16th November 2022
 Accepted 9th December 2022

DOI: 10.1039/d2ra07294g

rsc.li/rsc-advances

Introduction

Streblus ilicifolius (S. Vidal) Corn. (Moraceae) is widely distributed in India, China, and South Asia.¹ Several phytochemical studies of *S. ilicifolius* have been carried out, leading to the identification of amide glycosides, coumarins, stilbenes, lignans, and polyphenols. In addition, the anti-tyrosinase, antimicrobial, and anti-inflammatory activities of these chemical constituents has been reported.^{2–4}

Our continued studies on the bioactivity-guided phytochemical investigation of medicinal plants for tyrosinase inhibitory activity^{5–9} has led to the identification of two coumarins (*Strebluses* A and B) and two stilbene-like derivatives (*Strebluses* C and D) from the stems of *Streblus ilicifolius*, among which *Streblus* C exhibited a remarkable inhibitory effect with an IC_{50} value of 0.01 μ M.^{10,11} Thus, this phytochemical study was continuously performed, leading to the isolation of four new stilbene-like derivatives, *Strebluses* E–H (1–4). Their structures were determined by NMR spectroscopic interpretation. In addition, their absolute configurations were identified based on the optical rotation calculation. Herein, the tyrosinase inhibitory activity assays and the molecular docking studies with the oxy-tyrosinase were performed.

Strebluses E–H, four new stilbene-like derivatives from the stems of *Streblus ilicifolius*[†]

Tho Huu Le,^{abc} Phu Hoang Dang,^{ab} Hai Xuan Nguyen,^{abc} Truong Nhat Van Do,^{abc} Nhan Trung Nguyen^{*abc} and Mai Thanh Thi Nguyen^{*abc}

Following bioactivity-guided isolation, four new stilbene-like derivatives, named *Strebluses* E–H, were isolated from the EtOAc-soluble fraction of the stems of *Streblus ilicifolius* (Moraceae). Their chemical structures were elucidated based on NMR spectroscopic data interpretation and optical rotation calculation. *Streblus* E possesses potent tyrosinase inhibitory activity with an IC_{50} value of 0.1 μ M. Oxy-tyrosinase has two bound Cu^{2+} ions and a peroxide group in the binding site, which has a role in the catalytic oxidation. Thus, a docking study of *Streblus* E with oxy-tyrosinase was performed to analyze the ligand–protein interactions. With *in silico* modelling, the *S* value and the ligand–protein interactions suggested that *Streblus* E showed lower binding affinity for oxy-tyrosinase than that of *Streblus* C.

Results and discussion

Structural elucidation of four new isolated compounds from *S. ilicifolius*

The EtOAc-soluble fraction of *S. ilicifolius* stems was chromatographed to obtain four undescribed stilbene-like derivatives, *Strebluses* E–H (1–4) (Fig. 1).

Compound 1, *Streblus* E, showed a molecular formula to be $C_{19}H_{22}O_5$ based on the quasi-molecular ion at m/z 331.1545 [$M + H$]⁺ (calcd for $C_{19}H_{23}O_5^+$, 331.1540) in the HRESIMS spectrum. The ¹H and ¹³C NMR spectra showed signals of the prenylated stilbene-like feature having the 5,6-dihydroxycyclohex-2-en-1-one moiety (Tables 1 and 2), which resembled those of *Streblus* C except for the absence of the acetonide group.¹⁰ The observed HMBC correlations (Fig. 2) indicated the presence of the 2,4-dihydroxyphenyl group in 1. The 5,6-dihydroxycyclohex-2-en-1-one substructure was established based on the HMBC

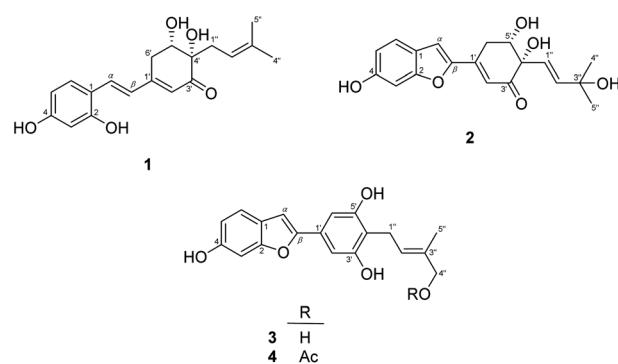


Fig. 1 The structure of compounds 1–4.

[†]Faculty of Chemistry, University of Science, Ho Chi Minh City, 72711, Vietnam

[‡]Vietnam National University, Ho Chi Minh City, 71300, Vietnam. E-mail: nttmai@hcmus.edu.vn; ntphan@hcmus.edu.vn

[§]Research Lab for Drug Discovery and Development, University of Science, Ho Chi Minh City, 72711, Vietnam

[†] Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d2ra07294g>



Table 1 ^1H (500 MHz) NMR data (acetone- d_6) for compounds 1–4

Position	δ_{H} (J, Hz)			
	1	2	3	4
3	6.46, d (2.4)	7.00, d (2.1)	6.96, d (2.2)	6.95, d (2.0)
5	6.42, dd (8.5, 2.4)	6.87, dd (8.5, 2.1)	6.80, dd (8.4, 2.2)	6.80, dd (8.4, 2.0)
6	7.47, d (8.5)	7.52, d (8.5)	7.38, d (8.4)	7.38, d (8.4)
2'	5.97, brs	6.55, d (2.3)	6.92, s	6.92, s
5'	4.23, dd (5.6, 3.2)	4.15, dd (2.9, 2.5)		
6'	2.94–2.96, m	3.15, ddd (18.3, 2.9, 2.3) 3.08, dd (18.3, 2.5)	6.92, s	6.92, s
α	7.36, d (16.3)	7.33, s	6.91, brs	6.91, brs
β	7.00, d (16.3)			
1''	2.42, dd (14.7, 7.6) 2.34, dd (14.7, 6.9)	5.91, d (15.5)	3.43, d (7.3)	3.45, d (7.3)
2''	5.19, dd (7.6, 6.9)	6.10, d (15.5)	5.57, tq (7.3, 1.3)	5.66, tq (7.3, 1.3)
4''	1.67, s	1.22, s	3.90, d (6.2)	4.41, s
5''	1.57, s	1.19, s	1.80, d (1.3)	1.82, d (1.3)
2-OH	8.91, s			
4-OH	8.68, s	8.93, s	8.46, s	8.46, s
4'-OH	4.10, s	4.37, s		
5'-OH	3.65, s	4.00, s	8.33, s	8.42, s
3''-OH		3.62, s		
3'-OH			8.33, s	8.42, s
4''-OH			3.58, t (6.2)	
4''-OAc				1.98, s

Table 2 ^{13}C (125 MHz) NMR data (acetone- d_6) for compounds 1–4

Position	δ_{C} , type C			
	1	2	3	4
1	116.3	122.0	122.7	122.7
2	158.2	158.1	156.7	156.7
3	103.7	98.3	98.4	98.4
4	160.8	158.9	156.6	156.6
5	109.0	114.3	113.2	113.2
6	129.5	123.4	121.8	121.9
1'	154.9	144.2	130.0	130.2
2'	123.0	119.0	103.8	103.9
3'	201.1	199.1	157.3	157.3
4'	79.7	80.3	116.1	115.3
5'	72.9	74.5	157.3	157.3
6'	31.9	32.1	103.9	103.9
α	132.1	111.0	101.5	101.6
β	126.4	153.2	155.8	155.7
1''	35.5	124.6	22.7	22.8
2''	119.0	142.4	123.9	128.5
3''	134.8	70.3	135.7	130.6
4''	26.1	30.4	68.6	70.5
5''	18.1	30.2	13.9	14.1
4''-OCOCH ₃			20.8	
4''-OCOCH ₃			170.8	

correlations from H-2' to C-4' and C-6', from H-5' to C-1' and C-3', from 5'-OH to C-5' and C-6', and from 4'-OH to C-3' and C-4'. The HMBC correlations from H- α to C-2, C-6, and C-1', from H- β to C-1, C-1', C-2', and C-6' were supportive of the C α -C1 and C β -C1' bonds. In addition, the C-4' prenyl group was assigned based on the HMBC correlations. The NOESY correlation between H-5'/H₂-1''/H-2'' deduced the presence of the *cis*-diol (Fig. 3). In

addition, the 3J coupling constants between H-5' and H₂-6' were 5.6 and 3.2 Hz, to suggest the equatorial configuration of H-5'.¹² The preferred conformers of *cis*-(*R,R*)-diol 1 were established by molecular mechanics calculation using MMFF94 force field.¹³ The obtained conformers were reoptimized using B3LYP density functional theory (DFT) method with the 6-31G* basis set, to obtain the most preferred conformer with 95.5% Boltzmann distribution. The optical rotation calculation at the sodium D line ($\lambda = 589.3$ nm) was carried out using DFT-B3LYP/6-311++G(2d,2p) function with the polarizable continuum model (PCM) for methanol.^{14,15} The calculated $[\alpha]_D$ value of *cis*-(*R,R*)-diol 1 was obtained as -694.12 , while the experimental value of $[\alpha]_D +630.0$ (*c* 0.01, MeOH) was the opposite in sign. Thus, an (*S,S*) absolute configuration was concluded for *Streblus E* (1), which was deacetonide-*ent*-*Streblus C*.

Compound 2, *Streblus F*, showed the sodium adduct molecular ion at m/z 367.1169 [M + Na]⁺ (calcd for C₁₉H₂₀O₆Na⁺, 367.1152) in the HRESIMS spectrum. The ^1H and ^{13}C NMR spectra of 2 closely resembled those of *Streblus D*, except for the presence of the 3-hydroxyisopent-1(*E*)-enyl group [δ_{H} 5.91 (d, $J = 15.5$ Hz, H-1''), 6.10 (d, $J = 15.5$ Hz, H-2''), 1.22 and 1.19 (s, $2 \times 3'$ -CH₃)] instead of the prenyl group in *Streblus D* (Tables 1 and 2). The observed NOESY correlation between H-5' and H-1'' confirmed the *cis*-orientation of the C-4' and C-5' hydroxy groups (Fig. 3). In addition, the equatorial configuration of H-5' was suggested based on its 3J coupling constants of 2.9 and 2.5 Hz.¹² The conformational analysis for (*R,R*)-2 was obtained seven conformers with a total Boltzmann weight >99%. The calculated $[\alpha]_D$ value of (*R,R*)-2 was +127.95, opposite in sign to its experimental value $[\alpha]_D -134.0$ (*c* 0.01, MeOH). Thus, the structure of *Streblus F* (2) was concluded as 4'*S*,5'*S*.



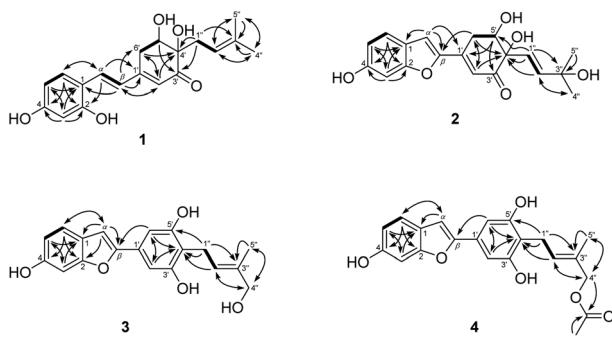


Fig. 2 ^1H - ^1H COSY (bold lines) and HMBC (solid arrows) correlations observed for 1-4.

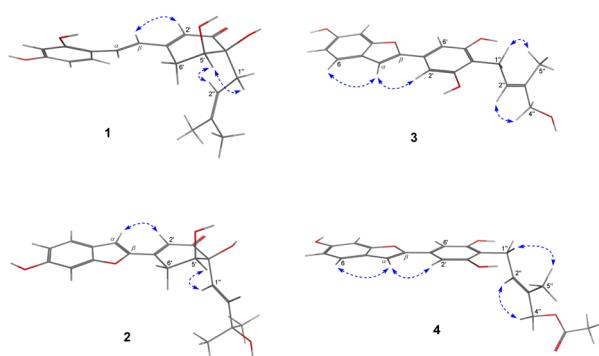


Fig. 3 NOESY correlations observed for 1-4.

Streblus G (3) showed the molecular formula $\text{C}_{19}\text{H}_{18}\text{O}_5$, as deduced from the negative HRESIMS spectrum at m/z 325.1087 [$\text{M} - \text{H}$] $^-$ (calcd for $\text{C}_{19}\text{H}_{17}\text{O}_5^-$, 325.1081). The ^1H spectrum showed signals for a 1,3,4-trisubstituted [δ_{H} 7.38 (d, $J = 8.4$ Hz, H-6), 6.96 (d, $J = 2.2$ Hz, H-3), 6.80 (dd, $J = 8.4, 2.2$ Hz, H-5)] and a 1,3,4,5-tetrasubstituted [δ_{H} 6.92 (s, H-2' and H-6')] aromatic rings, an olefinic proton [δ_{H} 6.91 (brs, H- α)], a hydroxylated prenyl group [δ_{H} 3.43 (d, $J = 7.3$ Hz, H-2''), 5.57 (tq, $J = 7.3, 1.3$ Hz, H-2''), 3.90 (d, $J = 6.2$ Hz, H-4''), 1.80 (d, $J = 1.3$ Hz, H-3''), 3.58 (t, $J = 6.2$ Hz, 4''-OH)], and two hydroxy groups [δ_{H} 8.46 (s, 6-OH), 8.33 (s, 3'-OH and 5'-OH)]. The ^{13}C NMR data exhibited resonances for 14 aromatic carbons [δ_{C} 98.3-157.2] and a hydroxylated prenyl group [δ_{C} 135.5 (C-3''), 123.7 (C-2''), 68.5 (C-4''), 22.5 (C-1''), 13.7 (C-5'')] (Table 2). These data resembled closely those of moracin M, except for the presence of the hydroxylated prenyl group at C-4'.¹⁶ The HMBC correlations from H-6 to C- α , C-4, and C-2, from H-5 to C-1 and C-3, from H-3 to C-1 and C-5, from H- α to C- β , C-1, and C-2, from H-2'/H-6' to C- β permitted the structural assignment of 3 as shown (Fig. 2). The C-3' and C-5' hydroxy groups were identified by the HMBC correlations with two corresponding oxygenated aromatic carbons. In addition, the HMBC correlations from H-2'' to C-4', from H-2'' and H-3'' to C-4'' indicated the location of the 4''-hydroxyprenyl group at C-4'. The NOESY correlations between H-6/H- α /H-2'(6'), H-2''/H-3'', and H-2''/H-2'' confirmed the presence of the 2-phenylbenzofuran moiety and

the (2''E)-4''-hydroxyprenyl group (Fig. 3). Thus, the structure of *Streblus G* (3) was concluded as 4''-(4''-hydroxyprenyl)moracin M.

Streblus H (4) showed a sodiated molecular ion peak at m/z 391.1151 [$\text{M} + \text{Na}$] $^+$ (calcd for $\text{C}_{21}\text{H}_{20}\text{O}_6\text{Na}$, 391.1152) in the HRESIMS spectrum. The ^1H and ^{13}C NMR spectra of 4 resembled closely those of 3, except for the presence of signals for an acetyl group [δ_{H} 2.08; δ_{C} 170.8, 20.8] (Table 2). The HMBC correlation between the H-2'' oxymethylene protons and the acetoxy carbonyl at δ_{C} 170.8 suggested that the acetylation happened at C-4'' (Fig. 2). In addition, the NOESY correlations between H-2''/H-3'' and H-2''/H-2'' supported the *E* configuration of the double bond in 4''-acetoxypropenyl group (Fig. 3). Thus, the structure of *Streblus H* (4) was assigned as 4''-acetyl-*streblus G*.

Tyrosinase inhibitory activity and docking studies

All isolated compounds were tested for their tyrosinase inhibitory activities.¹⁷ Kojic acid was used as a positive control. *Streblus E* (1) showed potent inhibitory effect with an IC_{50} value of 0.1 μM as compared to that of kojic acid (IC_{50} , 44.6 μM). All remaining compounds were inactive ($\text{IC}_{50} > 100$ μM). The docking study of 1 was performed with MOE following our previous procedure.⁹ In the binding pocket of the top-rank pose, 1 showed the H-donor interaction from the C-4' hydroxy group to peroxide bridge PER404, and from the C-2 hydroxy group to ASP45 residue (Fig. 4). The aromatic ring had the π -cation interaction with ARG55 residue. In addition, the prenyl group showed the σ - σ and π - σ interactions with VAL195 and TRP184, respectively.

The genus *Streblus* is a small deciduous shrub, which includes 20 species and mainly distributed in South China and South Asia. The phytochemical studies of *Streblus ilicifolius* were carried out to obtain various class of compounds. In our previous studies, we reported the structure and the anti-tyrosinase evaluation of four new compounds, in which *Streblus C* showed noteworthy inhibitory activity.^{10,11} With these

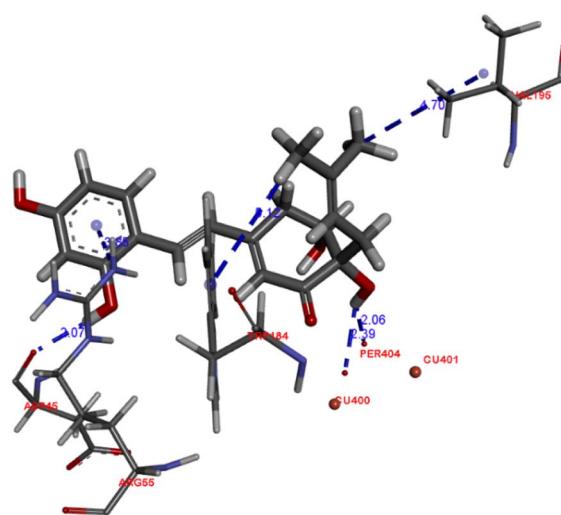


Fig. 4 Docked pose of best ranked docking score of compound 1.



interesting results, we continued to carry out the bioactivity-guided isolation, leading to the identification of four new stilbene-like derivatives were isolated as well as their *in vitro* and *in silico* tyrosinase inhibitory activity were reported. In this study, *Streblus* E (**1**) showed potent inhibitory effect with an IC_{50} value of 0.1 μ M. The 2,4-resorcinol subunit highly contributed to inhibitory activity.¹⁸ In addition, the formation of the five-membered ring gave rise to the loss of inhibitory effect.¹⁹ With *in silico* modelling, the *S* value and the ligand–protein interactions suggested that **1** showed lower binding affinity for oxy-tyrosinase than that of *Streblus* C.¹⁰ This result was used to clarify the remarkable difference in IC_{50} values of *Streblus* C (0.01 μ M) and E (0.1 μ M).

Conclusions

Four new stilbene-like derivatives, named *Streblus* E–H (**1**–**4**), were isolated from the EtOAc-soluble fraction of the stems of *Streblus ilicifolius* (Moraceae). Their structures were elucidated based on NMR spectroscopic data interpretation with the aid of optical rotation calculation. *Streblus* E (**1**) showed potent tyrosinase inhibitory effect with an IC_{50} value of 0.1 μ M.

Experimental

General experimental procedures

Optical values were measured on a Shimadzu UV-1800 spectrophotometer (Shimadzu Pte., Ltd., Singapore). NMR spectra were acquired on a Bruker Avance III 500 spectrometer (Bruker BioSpin AG, Bangkok, Thailand). Chemical shifts are expressed as δ values. HRESIMS data were acquired on Bruker microTOF-QII mass spectrometer (Bruker Singapore Pte., Ltd., Singapore). Column chromatography (CC) was carried out using silica gel 60, 0.06–0.2 mm (Scharlau, Barcelona, Spain) and LiChroprep RP-18, 40–63 μ m (Merck KGaA, Darmstadt, Germany). Kieselgel 60 F₂₅₄ or RP-18 F₂₅₄ plates for thin-layer chromatography (TLC) were purchased from Merck (Merck KGaA, Darmstadt, Germany). Mushroom tyrosinase (EC 1.14.18.1; 3933 U mL⁻¹) and L-dihydroxyphenylalanine (L-DOPA) were obtained from Sigma-Aldrich (Sigma-Aldrich Pte Ltd, Singapore). Other chemicals were of the highest grade available.

Plant material

The stems of *Streblus ilicifolius* were collected at Hoai Nhon District, Binh Dinh Province, Vietnam, in October 2017. Its scientific name was identified by Dr Rer. Nat. Anh Tuan Dang Le, Faculty of Biology and Biotechnology, University of Science, Ho Chi Minh City, Vietnam. A sample (MCE0052) has been deposited at the Department of Medicinal Chemistry, Faculty of Chemistry, University of Science, Ho Chi Minh City, Vietnam.

Extraction and isolation

The dried powdered stems of *S. ilicifolius* (7.0 kg) were exhaustively extracted in a Soxhlet extractor with *n*-hexane, EtOAc, and MeOH to yield *n*-hexane – (64.8 g), EtOAc – (117.2 g),

and MeOH – (378.0 g) soluble fractions, respectively. The EtOAc-soluble fraction was chromatographed by silica gel CC (15 \times 150 cm) and eluted with MeOH–CHCl₃ mixtures (v/v, 0 : 100 \rightarrow 100 : 0) to afford 18 fractions (Fr.1–Fr.18). Fraction Fr.8 (0.8 g) was subjected to further silica gel CC and was eluted with MeOH–CHCl₃ (v/v, 0 : 100 \rightarrow 100 : 0) mixtures to yield 2 fractions (Fr.8.1 and Fr.8.2). Fraction Fr.8.1 (445 mg) was chromatographed over a silica gel column with EtOAc–CHCl₃ mixtures (v/v, 0 : 100 \rightarrow 100 : 0) mixtures to obtain five fractions (Fr.8.1.1 and Fr.8.1.5). Fraction Fr.8.1.4 (50.9 mg) was purified by preparative TLC with an EtOAc–CHCl₃ mixture (v/v, 30 : 70) to afford compound **2** (3.0 mg). Fraction Fr.11 (4.7 g) was subjected to silica gel CC with MeOH–CHCl₃ mixtures (v/v, 0 : 100 \rightarrow 100 : 0) to obtain six fractions (Fr.11.1–Fr.11.6). Fraction Fr.11.5 (849 mg) was separated by silica gel CC with EtOAc–CHCl₃ mixtures (v/v, 0 : 100 \rightarrow 100 : 0) to obtain four fractions (Fr.11.5.1–Fr.11.5.4). Fraction Fr.11.5.2 (118 mg) was purified by CC with Me₂CO–*n*-hexane mixtures (v/v, 0 : 100 \rightarrow 100 : 0), then the resulting fraction was purified by preparative TLC with an EtOAc–*n*-hexane mixture (v/v, 50 : 50) to afford compound **4** (4.0 mg). Fraction Fr.14 (19.6 g) was separated by silica gel CC with MeOH–CHCl₃ mixtures (v/v, 0 : 100 \rightarrow 100 : 0) to obtain 11 fractions (Fr.14.1–Fr.14.11). Fraction Fr.14.9 (8.8 g) was loaded onto a silica gel column and eluted with Me₂CO–CHCl₃ mixtures (v/v, 0 : 100 \rightarrow 70 : 30) to give 20 fractions (Fr.14.9.1–Fr.14.9.20). Fraction Fr.14.9.5 (41.4 mg) was chromatographed over a silica gel column with a MeOH–CHCl₃ mixture (v/v, 10 : 90) to afford compound **3** (4.0 mg). Fraction Fr.14.9.8 (46.8 mg) was separated by CC with Me₂CO–CHCl₃ mixtures (v/v, 0 : 100 \rightarrow 70 : 30), then the resulting fractions were purified by preparative reversed-phase TLC with a H₂O–MeOH mixture (v/v, 30 : 70) to afford compound **1** (3.0 mg).

Streblus E (**1**): yellow, amorphous powder; ¹H and ¹³C NMR (500 MHz, acetone-*d*₆, see Tables 1 and 2); HRESIMS *m/z* 331.1545 [M + H]⁺ (calcd for C₁₉H₂₃O₅⁺, 331.1540).

Streblus F (**2**): yellow, amorphous powder; ¹H and ¹³C NMR (500 MHz, acetone-*d*₆, see Tables 1 and 2); HRESIMS *m/z* 367.1169 [M + Na]⁺ (calcd for C₁₉H₂₀O₆Na⁺, 367.1152).

Streblus G (**3**): yellow, amorphous powder; ¹H and ¹³C NMR (500 MHz, acetone-*d*₆, see Tables 1 and 2); HRESIMS *m/z* 325.1087 [M – H]⁻ (calcd for C₁₉H₁₇O₅⁻, 325.1081).

Streblus H (**4**): yellow, amorphous powder; ¹H and ¹³C NMR (500 MHz, acetone-*d*₆, see Tables 1 and 2); HRESIMS *m/z* 391.1151 [M + Na]⁺ (calcd for C₂₁H₂₀O₆Na⁺, 391.1152).

Tyrosinase inhibitory assay

All pure compounds were dissolved in DMSO and tested at concentrations of 0.01–100 μ M. Assay mixtures in 0.1 M phosphate buffer pH 6.8 were prepared immediately before use, consisting of 100 μ L of tyrosinase solution (15 U mL⁻¹) and 1900 μ L of test solution. These mixtures were preincubated at 32 °C for 30 min, followed by 1000 μ L of L-DOPA 1.5 mM in pH 6.8 phosphate buffer, and incubated at 32 °C for 7 min. The absorbance (*A*) at 475 nm was acquired on Shimadzu UV-1800 spectrophotometer. The inhibitory percentage (*I*%) was calculated according to the formula: $I\% = [(A_{\text{control}} - A_{\text{sample}})/A_{\text{control}}]$



× 100%. All experiments were performed in triplicate and data were represented as the mean of three samples with standard deviation.

Optical rotation calculation

The conformational searches were performed on Spartan'18 (Wave function, Inc., Irvine, USA) using Merck molecular force field (MMFF). All conformers with Boltzmann weight > 1% were optimized using DFT method at the B3LYP/6-31G* level in the gas phase. The optical rotation calculations at 589.3 nm were carried out using the DFT-B3LYP with 6-311++G(2d,2p) basis set in IEFPCM solvation model for methanol. These calculations were performed on Gaussian 16 Rev. C.01 (Gaussian, Inc., Wallingford, USA). The calculated optical rotation values were expressed as the Boltzmann-weighted average of all output data.

Molecular docking

Docking studies were performed with Molecular Operating Environment 2019 (MOE 2019.0102) suite (Chemical Computing Group ULC, Montreal, Canada). All structures were minimized up to 0.0001 gradients using the Amber12:EHT force field. The oxy-tyrosinase structure (1WX2) was prepared following our previous procedure. The molecular docking procedure was carried out with Triangle Matcher placement, Induced Fit refinement, and two scoring methods (London dG and GBVI/WSA dG). Five top results with the negative binding free energy value (S value) were selected to show up. The ligand interactions were carried out using BIOVIA Discovery Studio Visualizer 2016 (Dassault Systèmes Americas Corp., Waltham, USA).

Author contributions

MTTN and NTN designed the work. THL, HXN, and TNVD directed the experiments. THL and TNVD performed tyrosinase inhibitory assay. PHD performed molecular docking and optical rotation calculations. The manuscript was written by THL, PHD, MTTN, and NTN.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This research is funded by Vietnam National University Ho Chi Minh City (VNU-HCM) under grant number NCM2020-18-01.

Notes and references

- B. Singh, A. Chettri, D. Adhikari and S. K. Barik, *J. Bot. Res. Inst. Tex.*, 2012, **23**, 611–614.
- S. Dej-adisai, K. Parndaeng and C. Wattanapiromsakul, *Trop. J. Pharm. Res.*, 2016, **15**, 497–506.
- G. Zhang, L. Hao, D. Zhou, W. Liu, C. Li, S. Su, X. Xu, X. Huang and J. Li, *Biochem. Syst. Ecol.*, 2019, **87**, 103962.
- Y. Huang, X. Huang, G. Tian, W. Zhang, S. Su, X. Xu, J. Li and B. Liu, *Nat. Prod. Res.*, 2022, **36**, 1485–1493.
- N. T. Nguyen, M. H. K. Nguyen, H. X. Nguyen, N. K. N. Bui and M. T. T. Nguyen, *J. Nat. Prod.*, 2012, **75**, 1951–1955.
- H. X. Nguyen, N. T. Nguyen, M. H. K. Nguyen, T. H. Le, T. N. Van Do, T. M. Hung and M. T. T. Nguyen, *Chem. Cent. J.*, 2016, **10**, 2.
- P. H. Dang, T. T. Nguyen, T. H. Le, H. X. Nguyen, M. T. T. Nguyen and N. T. Nguyen, *Nat. Prod. Res.*, 2018, **32**, 1745–1750.
- P. H. Dang, L. T. T. Nguyen, H. T. T. Nguyen, T. H. Le, T. N. V. Do, H. X. Nguyen, N. D. Le, M. T. T. Nguyen and N. T. Nguyen, *Nat. Prod. Res.*, 2019, **33**, 2883–2889.
- P. H. Dang, T. H. Le, T. N. V. Do, H. X. Nguyen, M. T. T. Nguyen and N. T. Nguyen, *Evidence-Based Complementary Altern. Med.*, 2021, **2021**, 8872920.
- N. T. Nguyen, P. H. Dang, H. X. Nguyen, T. N. V. Do, T. H. Le, T. Q. H. Le and M. T. T. Nguyen, *Evidence-Based Complementary Altern. Med.*, 2021, **2021**, 5561176.
- N. T. Nguyen, H. X. Nguyen, T. H. Le, D. H. Nguyen, T. N. V. Do, P. H. Dang and M. T. T. Nguyen, *Nat. Prod. Res.*, 2022, **36**, 4967–4972.
- Z. Lin, T. Zhu, Y. Fang, Q. Gu and W. Zhu, *Phytochemistry*, 2008, **69**, 1273–1278.
- T. Lewis-Atwell, P. A. Townsend and M. N. Grayson, *Tetrahedron*, 2021, **79**, 131865.
- P. J. Stephens, F. J. Devlin, J. R. Cheeseman and M. J. Frisch, *J. Phys. Chem. A*, 2001, **105**, 5356–5371.
- T. Aharon and M. Caricato, *J. Chem. Theory Comput.*, 2020, **16**, 4408–4415.
- H. X. Li, J. U. Park, X. D. Su, K. T. Kim, J. S. Kang, Y. R. Kim, Y. H. Kim and S. Y. Yang, *Molecules*, 2018, **23**, 2559.
- E. T. Arung, I. W. Kusuma, Y. M. Iskandar, S. Yasutake, K. Shimizu and R. Kondo, *J. Wood Sci.*, 2005, **51**, 520–525.
- S. Khatib, O. Nerya, R. Musa, M. Shmuel, S. Tamir and J. Vaya, *Bioorg. Med. Chem.*, 2005, **13**, 433–441.
- Z. P. Zheng, K. W. Cheng, Q. Zhu, X. C. Wang, Z. X. Lin and M. Wang, *J. Agric. Food Chem.*, 2010, **58**, 5368–5373.

