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2(1H)-Pyrazinones from acyclic building blocks: methods of synthesis and further derivatizations

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Pyrazinones (2(1H)-pyrazinones) are found as components of a range of natural substances and are involved in the preparation of a great number of bioactive molecules. Synthesis of such compounds, and analogues, requires knowledge of the heterocyclic properties of pyrazinones and, in particular, methods for their ring construction. This review deals with the strategies that have been developed for the synthesis of pyrazinones from acyclic precursors, especially α -amino acid-derived units, from the first examples in 1905 up to the most recent in 2021.

1 Introduction

Pyrazinones (strictly 2(1H)-pyrazinones) **1** have received considerable attention because they are present in a large number of natural products showing interesting biological activities (Fig. 1). These natural products include deoxyaspergillinic acid (**2a**) and flavacol (**2b**), isolated from *Aspergillus flavus*,^{1,2} phevalin (**2c**), tyrvalin (**2d**), leuvalin (**2e**) and phileucin (**2f**), from *Streptomyces* or *Staphylococcus* species,^{3–5} as well as arglecin (**3a**), argvalin (**3b**), and maremycin F (**4**), isolated from *Streptomyces*.^{6–8} Other examples are sorazinone B (**5**) and enhyppyrizinone A (**6**), which have been more recently isolated from myxobacteria species.^{9,10} The pyrazinone skeleton is also found in the bromotyrosine alkaloids ma'edamines A and B (**7a,b**), isolated from an Okinawan sponge *Suberea* sp.,¹¹ dragmacidin D (**8**), and the more elaborated dragmacidins E and F,^{12–14} which are present in several marine sponges species like *Dragmacidon*, *Halicortex*, *Spongosorites* and *Hexadella*, as well as the peptidyl metabolite JBIR-56 (**9**), isolated from a marine *Streptomyces*.¹⁵

Apart from constituting the central core in different natural products, the 2(1H)-pyrazinone scaffold has also been described as a key building block of great usefulness for many synthetic applications in drug design and medicinal chemistry programs, including the preparation of pharmacologically active derivatives (Fig. 2). In this regard, favipiravir (**10**) is an approved drug to target viral infections of influenza A and B, and avian influenza, and has recently been investigated for the treatment of Ebola virus and COVID-19.^{16,17} Another bioactive compound bearing a pyrazinone core is BMS-764459 (**11**), which has proven to be a potent antagonist of the receptor of corticotropin-

releasing factor-1 (CRF₁), elevated levels of which are associated with depression and anxiety-related disorders.¹⁸ The fungicide **12**, patented by DuPont, disrupts the polymerization of tubulin altering the cellular proliferation in fungal infections caused by phytopathogens and exhibits strong activity against human rhabdomyosarcoma cells.^{19,20}

Furthermore, synthetic pyrazinone derivatives have been described as inhibitors of different enzymes, which intervene in a wide range of diseases (Fig. 2). In this context, Heeres *et al.* developed a series of pyrazinone-containing derivatives among which **13** exhibits the best inhibitory activity against the reverse transcriptase, essential in the replication of human immunodeficiency virus (HIV).²¹ Besides, the inhibitory activity against thrombin of pyrazinone derivatives has been extensively studied with the identification of different candidates like **14a** or **14b**,²² which target trypsin-like serine protease thrombin (Factor IIa) involved in the blood coagulation system. More recently, Raubo *et al.* synthesised **15**, an inhibitor of p38 α mitogen-activated protein kinase, suitable for an inhaled method of administration.²³ This enzyme plays an important role in the regulation of pro-inflammatory signalling, relevant in different pathologies such as rheumatoid arthritis, Crohn's disease, or chronic obstructive pulmonary disease (COPD).

Owing to the importance of pyrazinones, many efforts have been devoted to the development of methods for their preparation and for the synthesis of derivatives. In particular, our interest in 2(1H)-pyrazinones lies in their use in dipolar cycloadditions of 1H-pyrazinium-3-olates (also known as 3-oxido-pyraziniums; referred to in chemical abstracts as "pyrazinium, 3,4-dihydro-1-R-3-oxo-, inner salts") **16** derived from 2(1H)-pyrazinones *via* selective imine-N-alkylation (\rightarrow **17**) followed by simple deprotonation (Scheme 1). These species undergo cycloadditions under mild conditions on reaction with dipolarophiles, forming 3,8-diazabicyclo[3.2.1]octanones **18**.^{24–29} For a general study of the process typified by **16** \rightarrow **18**, and for

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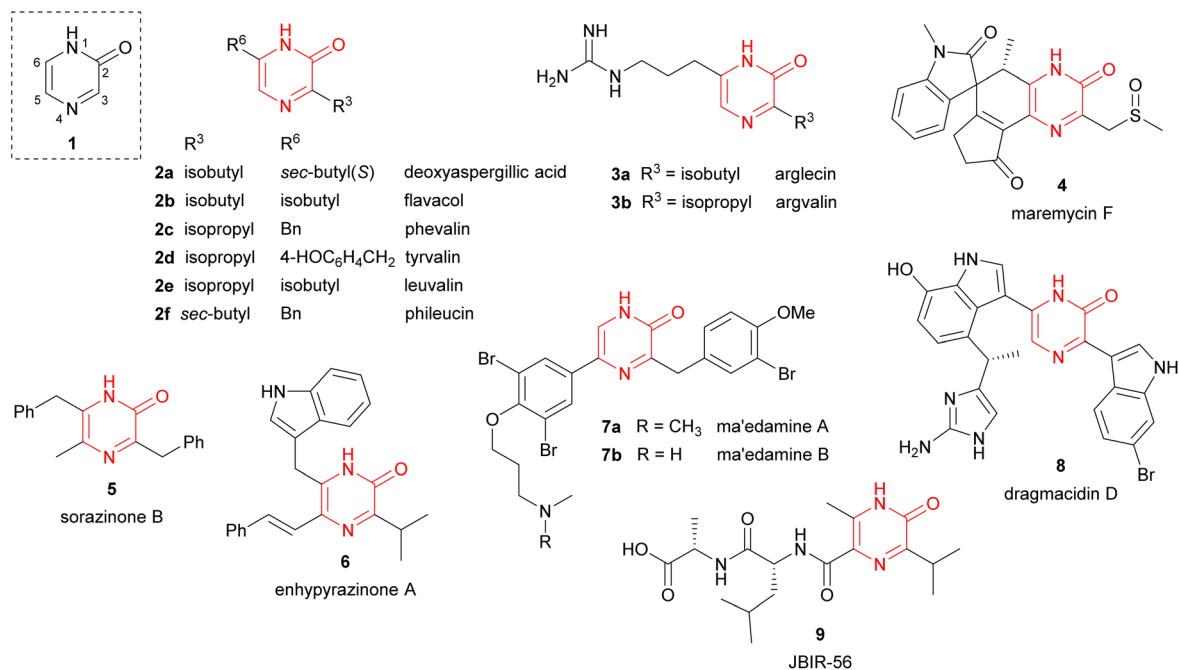


Fig. 1 2(1H)-Pyrazinone and representative natural derivatives.

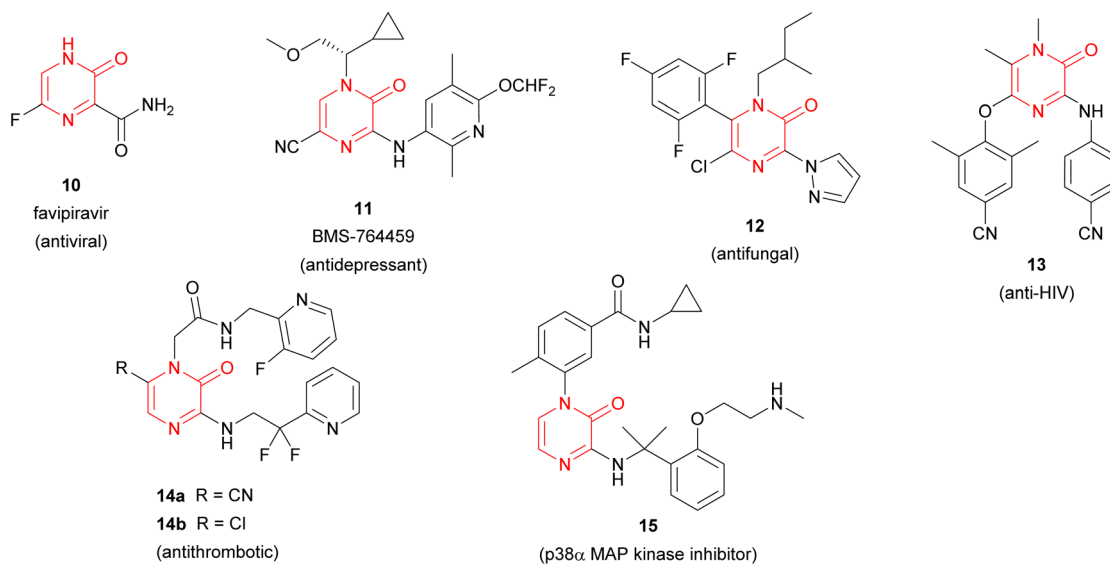


Fig. 2 Synthetic 2(1H)-pyrazinones with pharmacological activity.

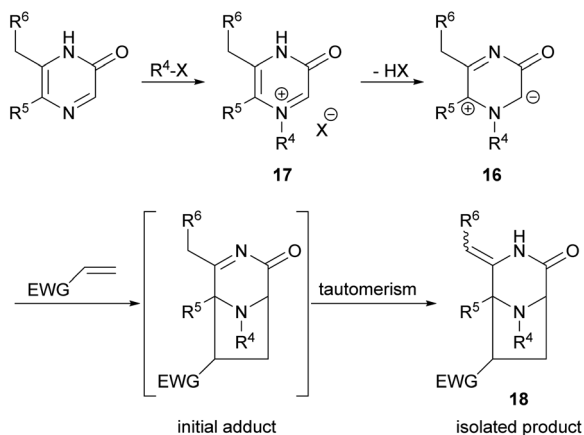
synthesis of pyrazinone-containing natural products, and analogues, access to variously substituted 2(1H)-pyrazinones is required.

In the early 20th century, a few papers described the first methods for the synthesis of pyrazinones.^{30–34} From the 1940s onwards, the preparation of pyrazinones was more extensively exploited due to their presence in natural products of biological interest. Since then, many authors have sought to overcome the limitations of the early methods and have designed new and

more versatile strategies to obtain a wide variety of pyrazinones following easier and more practicable approaches.

Despite the large number of articles published in this context, to the best of our knowledge, there is no review that covers the available methods for the synthesis of pyrazinones. Some of these methods involve the modification of an existing pyrazine scaffold, however in this review we deal with methods which start from the structural components of a pyrazinone, *i.e.*





Scheme 1 1,3-Dipolar cycloadditions of 3-oxidopyraziniums 16.

routes involving ring formation, though in some cases initial products require further modification.

2 Synthetic approaches

2.1 Synthesis from an α -amino ketone or α -amino aldehyde and an α -haloacetyl halide or α -aminoacetyl halide

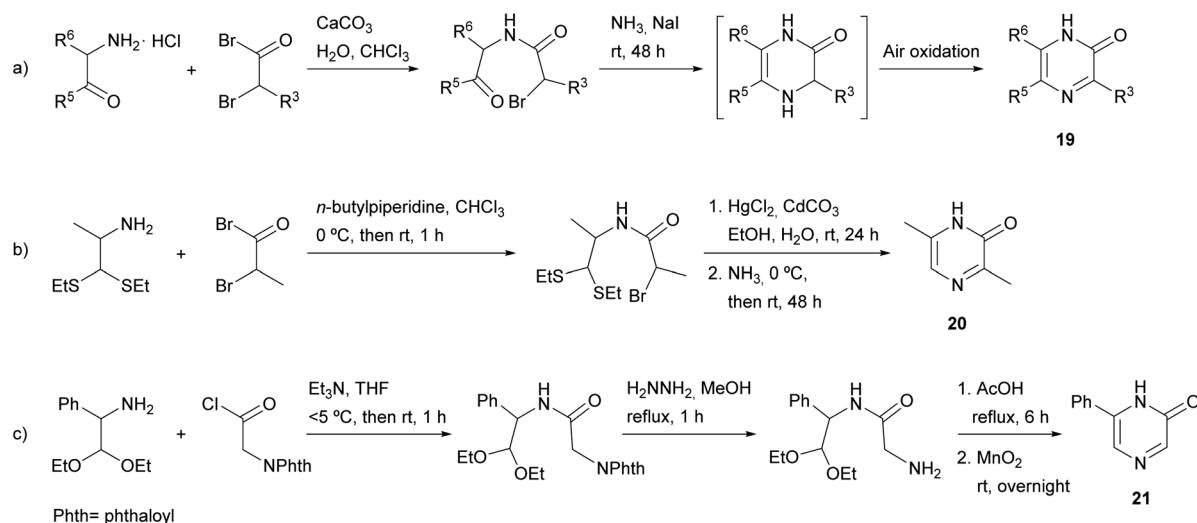
Prior to 1942 only a few methods for the preparation of substituted 2(1H)-pyrazinones had been described and they led to pyrazinones with identical groups at C-3 and C-6.^{30–34} Then, in 1942 Tota and Elderfield reported the preparation of 5,6-disubstituted and 3,5,6-trisubstituted 2(1H)-pyrazinones, in which the substituents can be different (Scheme 2a).³⁵ This was the first example in what has become a group of methods for the synthesis of 2(1H)-pyrazinones involving: (i) the condensation of two compounds, one providing N-1, C-5 and C-6, and the other, C-2 and C-3; (ii) reaction with a nitrogen reagent to supply N-4, and (iii) ring closure through the formation of the bond between N-4 and C-5.

In particular, the two starting compounds used by Tota and Elderfield were the hydrochloride of a 1,2-disubstituted α -amino ketone (affording N-1, C-5 and C-6) and a 2-bromo- or 2-chloro-substituted acid halide (supplying C-2 and C-3) (Scheme 2a). After their condensation, treatment of the resulting ketoamide with ammonia in the presence of sodium iodide afforded a dihydropyrazine, which yielded the corresponding pyrazinone 19 by air oxidation. The best results were obtained when using the most reactive bromoacetyl bromide. A drawback of this procedure is the self-condensation of the starting free α -amino ketone, which occurs very easily. To avoid it, the authors mixed the α -amino ketone and the α -bromoacetyl bromide in cold chloroform in the presence of a small amount of water and excess solid calcium carbonate.

Later, Baxter *et al.* published a modification of this method with the aim of preparing pyrazinones unsubstituted at position 5 (Scheme 2b).³⁶ This route involved the hydrochloride of an α -amino aldehyde or, alternatively, an α -amino aldehyde protected as an aminoacetal or aminothioacetal. 3,6-Dimethyl-2(1H)-pyrazinone (20) was only obtained when starting from 1,1-bis(ethylthio)propan-2-amine, which required an additional step to release the aldehyde functionality before reaction with ammonia. 6-Phenyl-2(1H)-pyrazinone (21) was also prepared following a similar approach, but in this case, the two starting components were an amino acetaldehyde and phthalimidoacetyl chloride, the latter providing N-4 after deprotection with hydrazine (Scheme 2c).³⁷ Ring closure and aromatisation were achieved by refluxing in acetic acid followed by oxidation with manganese dioxide.

2.2 Synthesis from an α -amino ketone or tryptamine and a glyoxylic acid derivative

Much more versatile and useful variations of the method were developed, in which the starting components are more readily available and avoid the self-condensation of the starting free α -amino ketone. With the aim of ensuring formation of 3-alkyl-5-

Scheme 2 Synthesis of 2(1H)-pyrazinones from an α -amino ketone or α -amino aldehyde and an α -haloacetyl halide or α -aminoacetyl halide.

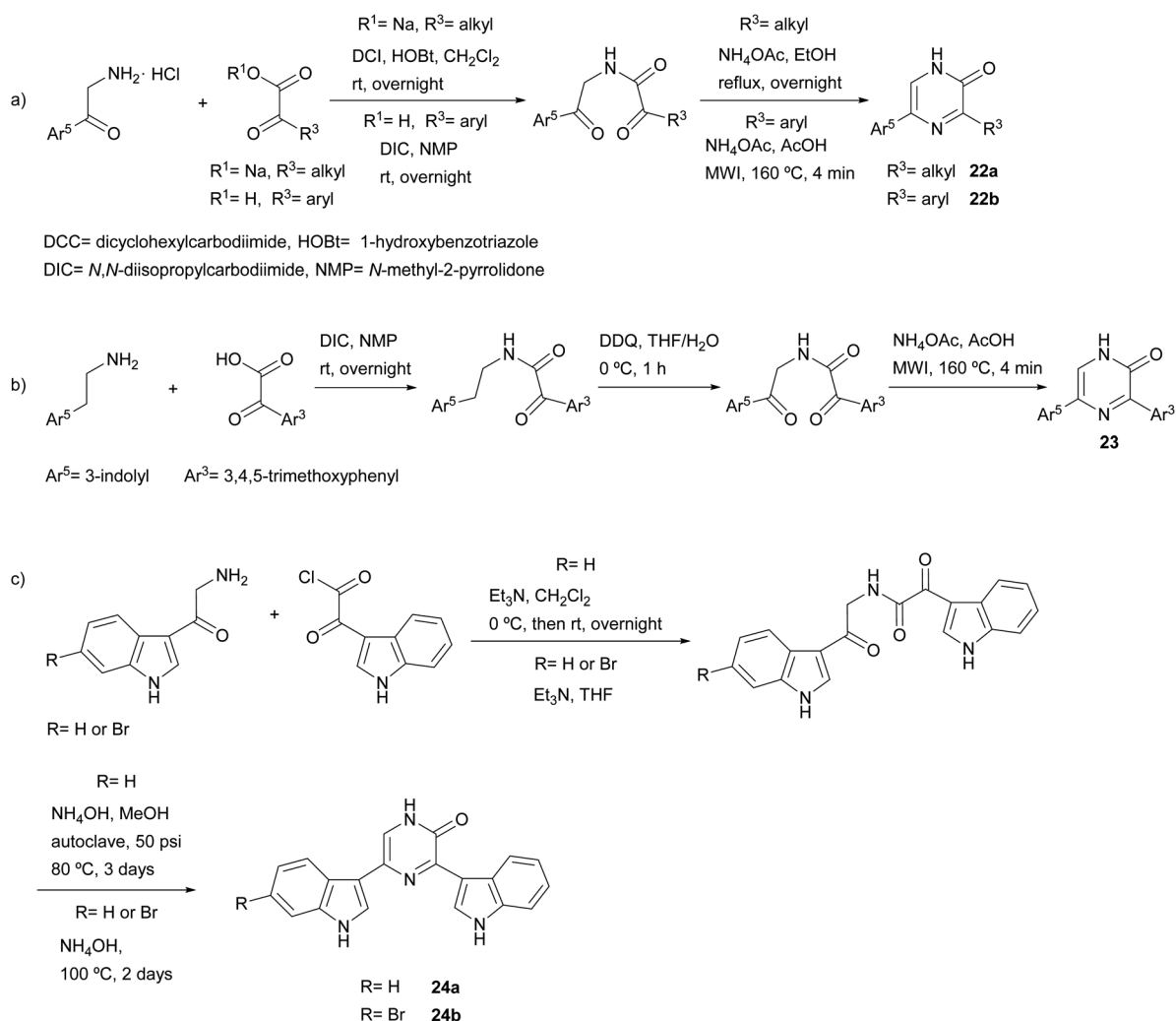
aryl-2(1*H*)-pyrazinones **22a** and avoiding the 3,6-disubstituted isomer, Bradbury *et al.* started from an α -amino ketone incorporating an aryl group, which was reacted with a sodium glyoxylate derivative. Ammonium acetate was used as nitrogen source leading to the cyclization of the resulting diketoamide in refluxing ethanol (Scheme 3a).³⁸ More recently, Johannes *et al.* extended this method to the synthesis of 3,5-diaryl-2(1*H*)-pyrazinones **22b** by using an arylglyoxylic acid and carrying out the cyclization under microwave irradiation (MWI) (Scheme 3a).³⁹ The versatility of this method allowed the preparation of a variety of derivatives incorporating different heterocyclic substituents with electron-donating and electron-withdrawing groups. In a study focussed on the identification of platelet-derived growth factor receptor (PDGFR) β -inhibitors, Johannes *et al.* also coupled a glyoxylic acid bearing a 3,4,5-trimethoxyphenyl with tryptamine, though an additional oxidation with 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ) before the cyclization step was necessary in order to obtain the corresponding 3,5-diaryl-2(1*H*)-pyrazinone **23** (Scheme 3b).^{40,41} 3,5-Bis(3-indolyl)-2(1*H*)-pyrazinones **24a–b** were also prepared by reacting 3-indolylglyoxylic chloride with an α -amino ketone

conveniently derivatized with an indolyl group, the cyclization being accomplished in excess of liquid ammonia at 80 °C under 50 psi for three days⁴² or in aqueous ammonia at 100 °C for two days (Scheme 3c).⁴³ This approach allowed the synthesis of precursors of 6'-debro- and 6',6''-didebro-*cis*-3,4-dihydrohamacanthin B.

2.3 Synthesis from diketopiperazines

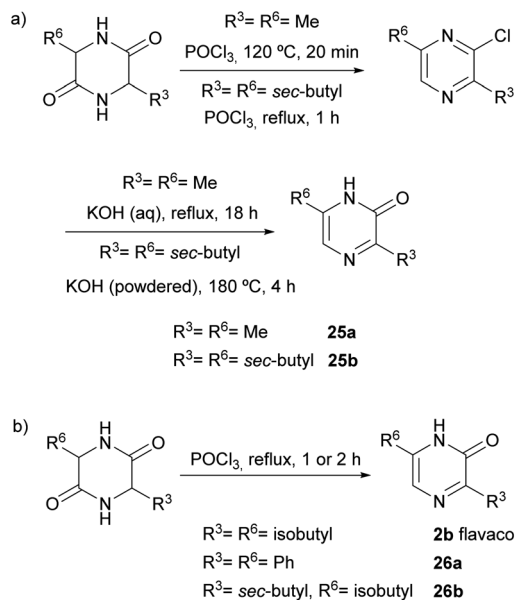
2(1*H*)-Pyrazinones can also be synthesized from diketopiperazines (2,5-piperazinediones). Unlike the other approaches included in this review, this method does not start from one or more acyclic products to form the pyrazinone ring. However, diketopiperazines can be prepared very easily through, for example, dehydration of amino acids, and so are readily available pyrazinone precursors.

This method, first developed by Baxter and Spring (1947),⁴⁴ involved the conversion of diketopiperazines into chloro substituted pyrazines, which were then transformed into pyrazinones (Scheme 4). Heating the diketopiperazines with phosphoryl chloride afforded a mixture of mono- and



Scheme 3 Synthesis of 2(1*H*)-pyrazinones from an α -amino ketone or tryptamine and a glyoxylic acid derivative.



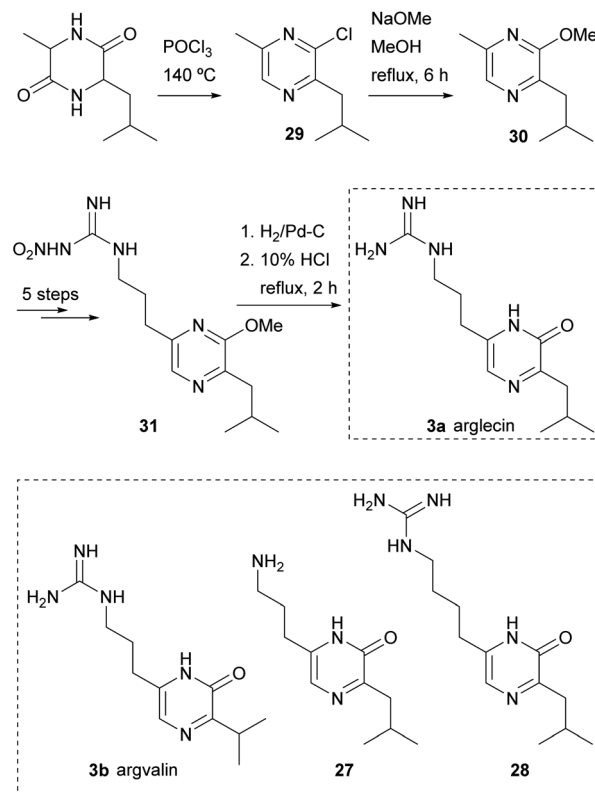


Scheme 4 Synthesis of 2(1H)-pyrazinones from diketopiperazines.

dichloropyrazines. The 2-chloropyrazine yielded a 2(1H)-pyrazinone by treatment with alkali or, alternatively, with sodium ethoxide followed by hydrolysis with hydrochloric acid. The first diketopiperazines used were symmetric, such as DL-alanine anhydride and DL-isoleucine anhydride, which provided 3,6-dimethyl-2(1H)-pyrazinone (**25a**) and 3,6-(di-sec-butyl)-2(1H)-pyrazinone (**25b**), respectively (Scheme 4a).⁴⁴ Two years later, Gallagher *et al.* reported the first example of direct transformation of a diketopiperazine into a 2(1H)-pyrazinone by exposure to phosphoryl chloride. In particular, DL-phenylglycine anhydride was converted to 3,6-diphenyl-2(1H)-pyrazinone (**26a**), presumably *via* loss of hydrogen chloride from a monochloro dihydropyrazine intermediate (Scheme 4b).⁴⁵ Treatment of DL-leucine anhydride with phosphoryl chloride also resulted in the direct dehydration of the diketopiperazine, affording 3,6-diisobutyl-2(1H)-pyrazinone (**2b**) together with a mixture of chloropyrazines (Scheme 4b).² This reaction allowed the synthesis and elucidation of the structure of flavacol (3,6-diisobutyl-2(1H)-pyrazinone) **2b**.

Unsymmetrical diketopiperazines were then used, one of the first being DL-leucyl-isoleucyl anhydride (Scheme 4b).⁴⁶ Subjecting this anhydride to phosphoryl chloride led to a mixture from which 3-(sec-butyl)-6-isobutyl-2(1H)-pyrazinone (**26b**) could be isolated. This pyrazinone is an isomer of deoxyaspergillitic acid (6-(sec-butyl)-3-isobutyl-2(1H)-pyrazinone) **2a**.

This procedure was also the first step in syntheses of arglecin **3a** and argvalin **3b** from DL-alanyl-leucyl anhydride and DL-alanyl-valyl anhydride, respectively, and of their analogues **27** and **28** (Scheme 5).^{47,48} As an example, the synthesis of arglecin involved heating DL-alanyl-leucyl anhydride in phosphoryl chloride at 140 °C. A mixture of chloropyrazines was obtained from which 2-chloro-3-isobutyl-6-methylpyrazine (**29**) was isolated. Refluxing **29** with sodium methoxide in methanol for 6 h yielded the 2-methoxyderivative **30**. The methyl group at

Scheme 5 Structure and synthesis of arglecin; structures of argvalin, and analogues **27** and **28**.

position 6 was subsequently derivatized as a nitroguanidino substituent in five steps, giving rise to **31**. Finally, hydrogenation of **31** followed by hydrolysis with 10% HCl afforded arglecin.⁴⁷

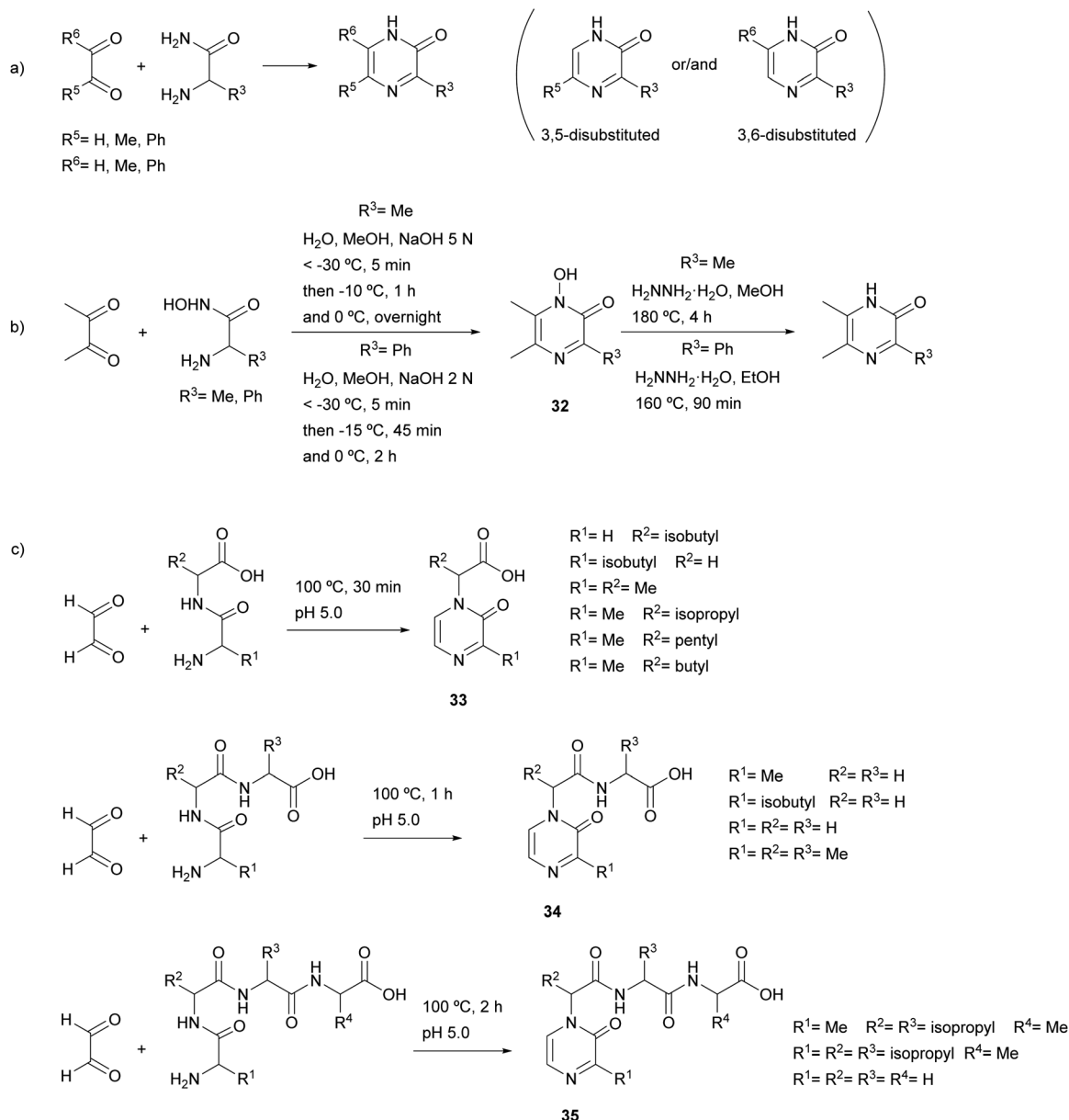
2.4 Synthesis from α -amino acid amides and 1,2-dicarbonyl compounds or analogues

2.4.1 Method of Jones and Karmas and Spoerri.

One of the most important approaches to 2(1H)-pyrazinones involves a one-pot condensation of an α -amino acid amide and a 1,2-dicarbonyl compound, leading to the formation of bonds between N-1 and C-6 and between N-4 and C-5 (Scheme 6). Since it was first described by R. G. Jones in 1949,⁴⁹ several studies have been carried out to optimise the reaction conditions, as well as to determine its regioselectivity and to prepare compounds with interesting applications.

In the original report by Jones, the reaction was performed using free α -amino acid amides and in the presence of a base (sodium or potassium hydroxide or piperidine) (general Scheme 6a).⁴⁹ The main drawback of this method was the preparation of the free α -amino acid amides, especially those with low molecular weight. Later, Karmas and Spoerri showed that the condensation also proceeds starting from the hydrohalides of the amino acid amides, which are easier to synthesise and, therefore, more readily accessible.⁵⁰ The condensation was also carried out in alkaline medium or, in case of decomposition of the amino acid amide, with a prior neutralization with silver



Scheme 6 Synthesis of 2(1H)-pyrazinones from α -amino acid amides using the method of Jones and Karmas and Spoerri.

oxide. Moreover, it was shown that the efficiency of the reaction depended on the temperature and the rate of the addition of the base, the best conditions for obtaining 2(1H)-pyrazinone being around $-50\text{ }^\circ\text{C}$ and a rate of addition of a NaOH solution between 8.0 and 8.6 mmol min^{-1} .⁵¹

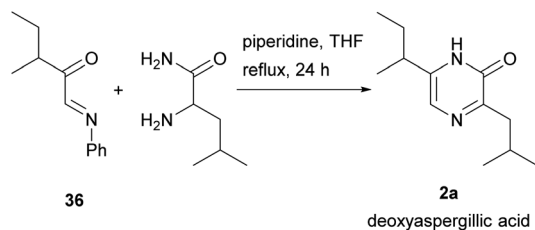
The scope of this reaction has been shown to be wide, having been conducted with many 1,2-diketones and 1,2-ketoaldehydes (glyoxal, methylglyoxal, diacetyl, phenylglyoxal, benzil) as well as α -amino acid amides (aminomalonyamide, glycine amide, alanine amide, methionine amide, tyrosine amide, α -aminophenylacetamide, leucine amide, α -aminobutyramide, norvaline amide and valine amide) (Scheme 6a). Moreover, the use of α -aminohydroxamic acids, instead of α -amino acid amides, allows the preparation of pyrazinones through the formation of

the corresponding 1-hydroxy-2(1H)-pyrazinones **32** and a subsequent reduction step with hydrazine (Scheme 6b).⁵²

Condensation of an α -amino acid amide with an unsymmetrical 1,2-dicarbonyl compound can occur in one or both of two directions, providing one of the two possible isomers or a mixture of both. In particular, when methylglyoxal reacted with a variety of α -amino acid amides the 5-methyl-2(1H)-pyrazinone was formed as a single or major isomer (Scheme 6a),^{49,50,53} whereas if the reaction mixture contained sodium bisulfite, the formation of the 6-methyl isomer was favoured.⁵⁴ In another study, the structure of the product resulting from methylglyoxal and alanine amide was demonstrated by X-ray crystallography to be 3,5-dimethyl-2(1H)-pyrazinone.⁵⁵

Glyoxal also reacted with dipeptides, tripeptides and tetrapeptides leading to 2(1H)-pyrazinones **33**, **34** and **35** bearing

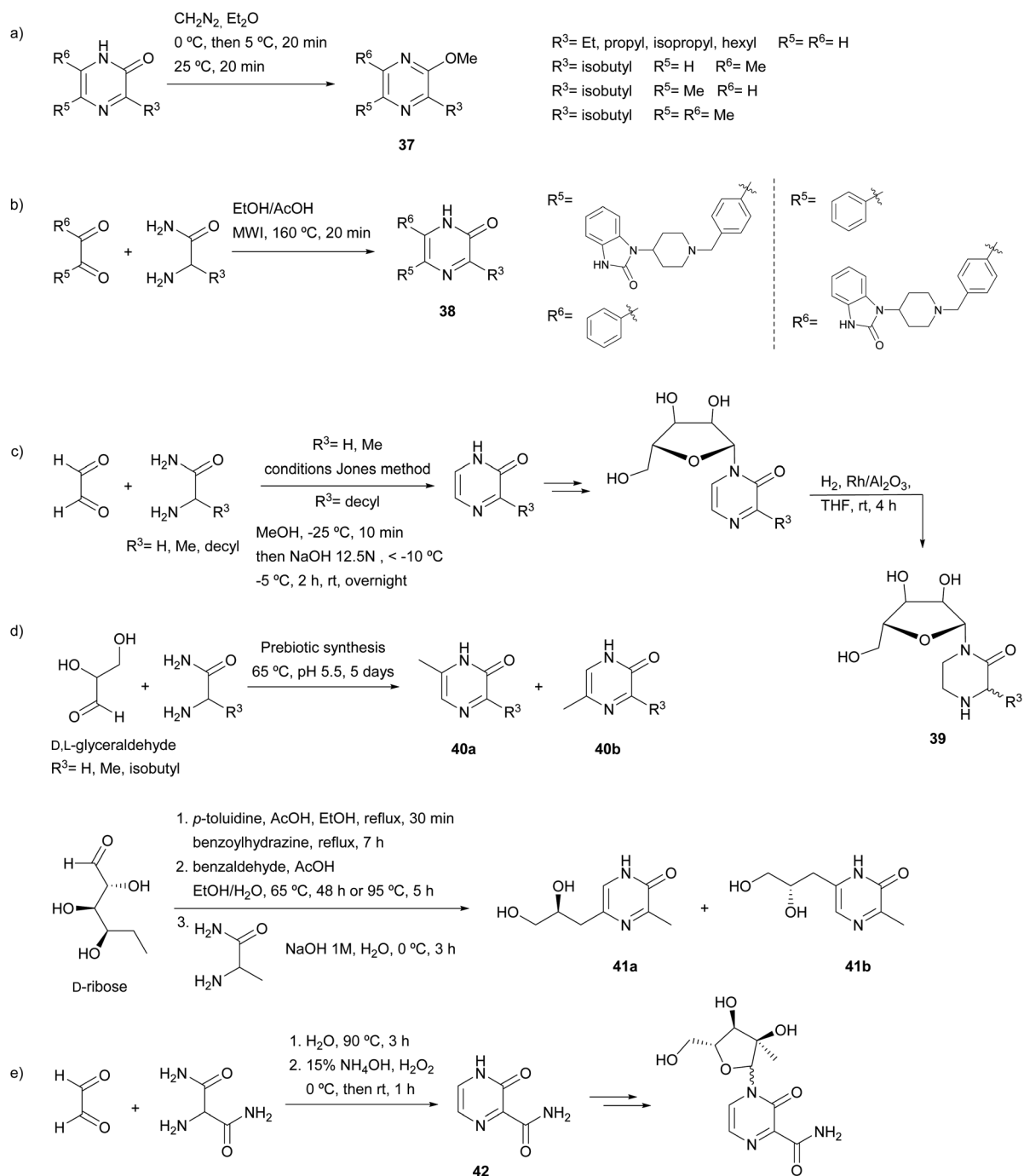




Scheme 7 Synthesis of 2(1*H*)-pyrazinones from α -aminoamides and 1,2-diketone mono Schiff bases.

a carboxylic acid, an acylamino acid or an acylpeptide at position 1, respectively (Scheme 6c).^{56,57} In these examples, the reaction took place between the two carbonyl groups of glyoxal and the N-terminal amino group and the amide nitrogen of the amino acid at position 1 in the peptide sequences.

2.4.2 Synthesis from α -amino acid amides and 1,2-diketone mono Schiff bases. Other authors have reported similar methods to access 2(1*H*)-pyrazinones based on the formation of bonds between N-1 and C-6 and between N-4 and C-5. In particular, Bergeron developed an effective method for



Scheme 8 2(1*H*)-Pyrazinones with potential applications prepared following Jones and Karmas and Spoerri procedure.



obtaining the 6-alkyl-2(1*H*)-pyrazinone isomer as a single isomer (Scheme 7).⁵⁸ This method took advantage of the different reactivity of the carbonyl and imino groups of ketoinmine **36**, which upon condensation with leucinamide yielded exclusively deoxyaspergillilic acid **2a** (3,6-diisobutyl-2(1*H*)-pyrazinone).

2.4.3 2(1*H*)-Pyrazinones with potential applications, prepared following Jones and Karmas and Spoerri procedure. Interestingly, the method described by Jones and Karmas and Spoerri^{49,50} has been applied to obtain numerous compounds with potential use as flavour ingredients or application in medicine (Scheme 8). In a search for analogues of the bell pepper compound 2-isobutyl-3-methoxypyrazine with strong odour and flavoring properties, pyrazinones were prepared from glyoxal and amino acid amides, and then reacted with diazomethane to be converted into 2-methoxy-3-alkylpyrazines **37** (Scheme 8a).^{59,60}

In the medicinal field, a collection of potential allosteric Akt kinase inhibitors bearing a 2(1*H*)-pyrazinone ring was synthesized (Scheme 8b).⁶¹ The two regioisomers **38** resulting from the reaction of a benzil derivative and different amino acid amides were screened for their Akt inhibitory activity. Derivatives selective for Akt1, Akt2 or both Akt1/Akt2 were identified. Moreover, 2(1*H*)-pyrazinones prepared following the Jones' method were employed for the synthesis of nucleosides. In a study concerning this topic, Benjahad *et al.* reported piperazin-2-one nucleosides **39** resulting from the reduction of pyrazinones linked to a ribofuranose (Scheme 8c).⁶² The synthesis of the corresponding pyrazinones was performed by treating glyoxal with glycine amide, alanine amide or 2-aminododecanamide. Two of these piperazin-2-one nucleosides bearing a decyl chain showed moderate activity against the Visna virus related to the human immunodeficiency virus. In another work, Weber *et al.* focussed their attention on the prebiotic synthesis of nucleosides (Scheme 8d). First, the synthesis of 2(1*H*)-pyrazinones from sugars (glyceraldehyde or erythrose) through reaction with an amino acid amide (glycine amide, alanine amide or leucine amide) was explored.⁶³ The corresponding 3,5- and 3,6-disubstituted pyrazinone isomers **40a** and **40b** were obtained. A few years later, aldopentoses, such as ribose, arabinose, xylose and lyxose, were subjected to similar conditions.⁶⁴ Although the results were promising, pyrazinones were not obtained in sufficient quantity to study their usefulness in pyrazine nucleic acid synthesis. Therefore, pyrazinones were prepared through traditional organic synthesis. In particular, *D*-ribose was reacted with alanine amide, which afforded the two possible regioisomers **41a** and **41b**. In another study, in an attempt to find new antiviral nucleoside analogues, 3-oxo-3,4-dihydropyrazine-2-carboxamide (**42**) was prepared from glyoxal and 2-aminopropanediamide, and then glycosylated with a ribofuranose derivative (Scheme 8e).⁶⁵

2.5 Synthesis from 2-chloro ketone oximes and α -amino acid esters

3,6-Disubstituted 2(1*H*)-pyrazinones were also synthesised from 2-chloroketone oximes and α -amino acid esters through

formation of bonds between N-1 and C-2 and between N-4 and C-5.⁶⁶ This method afforded pyrazinones **42** and it was also successfully applied to the preparation of deoxyaspergillilic acid (**2a**) (Scheme 9).⁶⁷ In the case of **2a**, reaction of 1-chloro-3-methyl-2-pentanone oxime and L-leucine ethyl ester led to the corresponding *N*-(2-oximinoalkyl)-amino acid ester **43**, which was then subjected to saponification of the ester, conversion of the oxime into a ketone, esterification of the carboxylic acid and treatment with ammonia with subsequent oxidation of the ring.

2.6 Synthesis from an amino acetal or amino alcohol, an oxalate and an amine

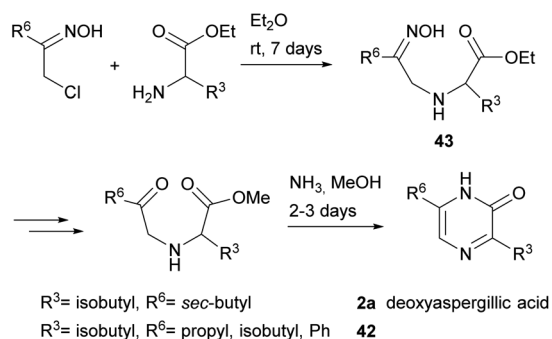
In 1975 Cheeseman *et al.* described a sequence for the preparation of the pyrazinone core as an alternative of the Strecker reaction, avoiding the high toxicity associated with trimethylsilyl cyanide (TMSCN). The key steps of this strategy involve the formation of the C-3–N-4 bond and a cyclization between N-1 and C-6.⁶⁸

This synthetic route starts with diethyl oxalate reacting with an equimolar quantity of 2,2-dimethoxyethylamine to afford oxamate **44** that is converted with benzylamine into the corresponding *N*-benzyl-*N'*-(2,2-dimethoxyethyl)oxamide **45** (Scheme 10a). This N-1-protected oxamide undergoes cyclization under reflux in acetic acid with a catalytic amount of HCl. The resulting pyrazinedione **46** is then treated with hexanemethyldisilazane (HDMS) to afford 1-benzyl-3-trimethylsilyloxy-2(1*H*)-pyrazinone (**47**), that can be used as a precursor of further functionalized pyrazinone scaffolds. Specifically, Cheeseman and co-workers used this strategy for the preparation of pyrazine nucleosides as analogues of the natural pyrimidine nucleosides with potential anti-tumoral activity. These compounds were obtained by the condensation of **47** with the corresponding acylglycosyl chloride, using silver perchlorate.⁶⁸

From this method, Fleitz *et al.* considered a similar approach for the construction of the pyrazinone core in a multi-kilogram scale synthesis (Scheme 10b).⁶⁹ In their approach, the cyclization precursor **48** contained an acetyl group instead of the dimethylacetal-protected aldehyde initially reported by Cheeseman, which after an acidic treatment afforded the 6-methyl-pyrazine-2,3-dione **49**. The oxamide **48** was synthesised from the oxidation of the corresponding alcohol **50**, which was previously obtained from a two-step condensation starting from ethyl oxalyl chloride. Moreover, the presence of an acetate group at N-1 in **49** allowed further functionalization after the selective hydrolysis of this ester. Likewise, the position 3 of the pyrazinedione **49** can be easily brominated using POBr₃. This halogenation allows nucleophilic substitution of amines to take place at this position. All in all, based on this strategy, the synthesis of the thrombin inhibitor **51a** was described.

Later, the incorporation of monocyclic⁷⁰ and bicyclic⁷¹ arginine derivatives at position 1 was also assessed to improve the selectivity and thrombin inhibitory activity of the previously reported **51a**, leading to the description of **51b** as the analogue with the best biological profile (Scheme 10b). This derivatization was achieved following a similar strategy: the cyclic





Scheme 9 Synthesis of 2(1H)-pyrazinones from 2-chloroketone oximes and α -amino acid esters.

arginine surrogates were bound to N-1 through the hydrolysis of the acetate group affording the carboxylic acid and subsequent coupling of the corresponding amine precursors mediated by 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDC), 1-hydroxybenzotriazole (HOBT) and Et_3N at room temperature.

The identification of the metabolic activation pathways of previously reported thrombin inhibitors led to the optimization of **51a** in order to increase the oral bioavailability and pharmacokinetics.^{72,73} Burgey *et al.* found that compounds with methyl groups at position 6 had low half-life in plasma because of oxidation of this position during metabolism.²² In contrast, when it was replaced by a chlorine or a nitrile group (**14a,b**), the inhibitory activity was enhanced and half-lives considerably increased. The synthetic pathway to introduce a nitrile group at position 6 is represented in Scheme 10c: starting from a 6-methylpyrazinedione **49** an oxidation of the 6-methyl with SeO_2 afforded the aldehyde, which was converted into the nitrile after forming the oxime and its subsequent reduction. On the other hand, the introduction of a chlorine at C-6 is achieved by reaction with *N*-chlorosuccinimide (NCS) in 1,2-dichloroethane (DCE) (Scheme 10d). From these intermediates, target molecules **14a** and **14b** are easily achieved following the strategy discussed previously.

2.7 Synthesis of 3,5-dihalo-2(1H)-pyrazinones from oxalic halides and α -aminonitriles: Hoonaert's method

Vekemans *et al.* reported in 1983 the most general method for the preparation of 3,5-dichloro- and 3,5-dibromo-2(1H)-pyrazinones.⁷⁴ This method involves the treatment of an α -aminonitrile **52** with an excess of the corresponding oxalyl halide in toluene or *o*-dichlorobenzene at 70–100 °C for 4–6 h or at room temperature for several days (Scheme 11). The α -aminonitrile provides the substituents at N-1 and C-6 of the pyrazinone core and it is usually obtained from a primary amine, an aldehyde and a cyanide source *via* a Strecker type reaction. If the primary amine is poorly nucleophilic, the α -aminonitrile **52** can also be prepared *via* a two-step procedure involving the synthesis of an imine followed by reaction with TMSCN (Scheme 11).²⁰ To form the pyrazinone, the salt of the α -aminonitrile must be used to avoid the formation of symmetrical oxamides. Free-base aminonitriles are employed only when they have poor nucleophilicity (*e.g.* $\text{R}^1 = \text{Ph}$).

The mechanism for the formation of the pyrazinone core involves first the acylation of the α -aminonitrile to form an oxamoyl halide **53** (bond between N-1 and C-2) (Scheme 12). Next, addition of HX to the nitrile, tautomerization to the enamine **54** and subsequent cyclization affords a cyclic pyrazine-2,3-dione **55** (bond between N-4 and C-3). This intermediate then reacts with an excess of the oxalyl halide allowing the introduction of the halogen atom at position 3 with release of CO_2 , CO and X^- . This step can be favoured by adding a tetraalkylammonium halide, acting as an additional halide source.

This method has been employed to obtain 3,5-dihalo-2(1H)-pyrazinones with a range of substituents at positions 1 and 6. However, it fails when R^1 is an acyl group and is difficult when R^6 is an electron-withdrawing group. In addition, when R^1 and/or R^6 are bulky groups, the yields are low because the conversion of the pyrazine-2,3-dione into the dihalopyrazinone is not efficient.⁷⁵ It has been reported that this halogenation step can be accelerated by adding a catalytic amount of DMF after the reaction of the α -aminonitrile with the oxalyl halide.^{20,75}

Applying this procedure, Tulinsky *et al.* described the asymmetric synthesis of atropisomeric 6-aryl-3,5-dichloro-2(1H)-pyrazinones (Scheme 13).^{76,77} They observed that the cyclization of the diastereomerically pure (*S,S*)- and (*R,R*)-aminonitriles **56a** and **56b**, derived from *o*-iodobenzaldehyde, (*R*)- or (*S*)- α -methylbenzylamine and NaCN , provided the diastereomerically pure pyrazinones (*aS,S*) **57a** and (*aR,R*) **57b**, respectively. They concluded that the configuration at the nitrile-bearing carbon determined the configuration of the asymmetric axis and that the steric bulk of the *ortho*-halo substituent influenced the selectivity.

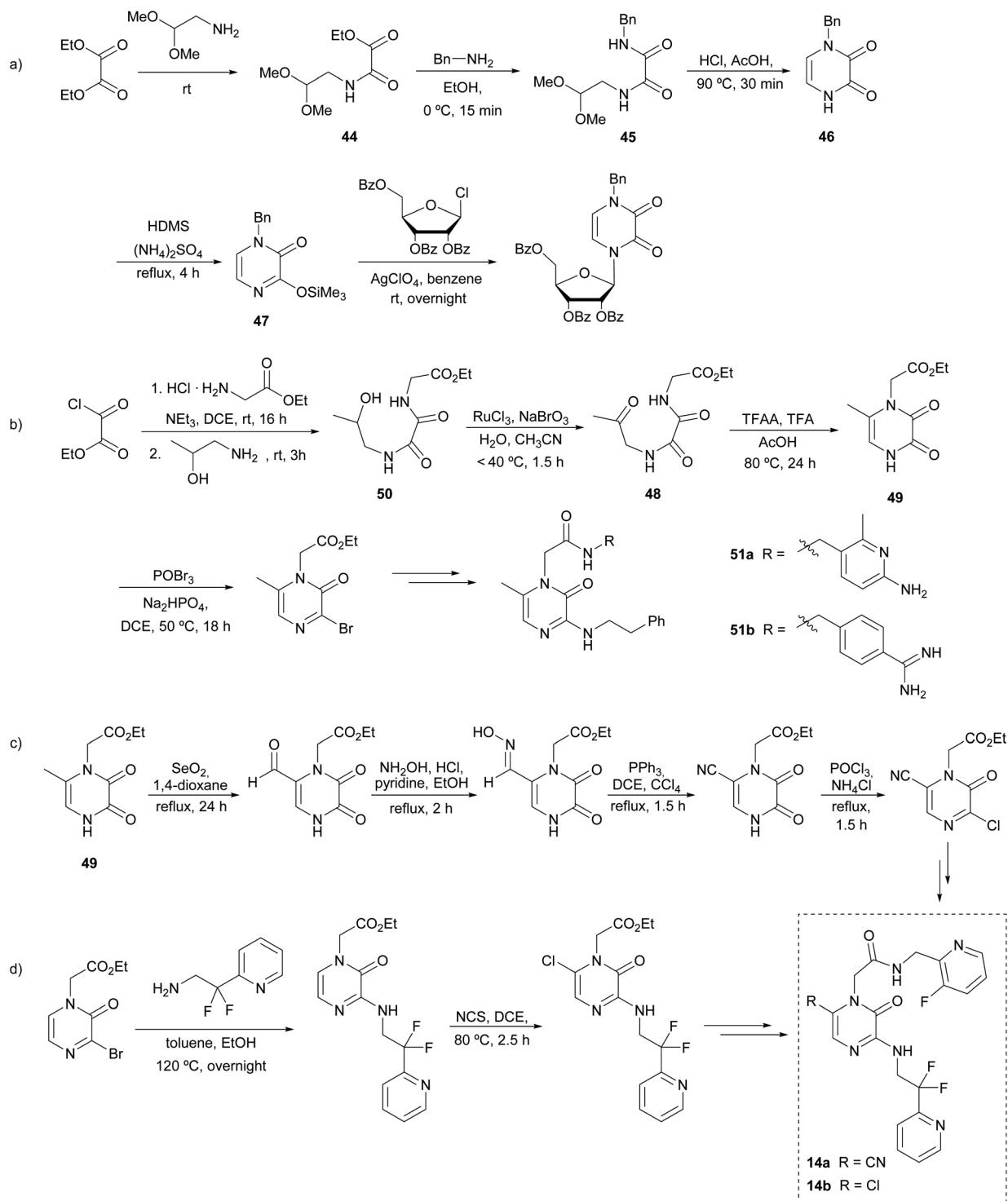
With the aim of taking advantage of combinatorial principles and to pave the way for the generation of libraries of biologically interesting structures, Kaval *et al.* transferred the Hoonaert's method to solid phase (Scheme 14).⁷⁸ A Wang amide resin was used as solid support which was treated with the corresponding aldehydes and TMSCN to yield the α -aminonitriles. Following acidification, the resulting α -aminonitrile salts were cyclized with an excess of oxalyl chloride in toluene providing the resin-bound 3,5-dichloro-2(1H)-pyrazinones bearing a range of substituents at position 6.

Gising *et al.* demonstrated that the speed of Hoonaert's method can be improved using MWI.^{79–81} They developed a rapid one-pot, two-step protocol that involved: (i) a microwave-assisted Strecker reaction to provide the α -aminonitrile and (ii) addition of HCl gas, then oxalyl chloride and cyclization under MWI (Scheme 15).

Researchers from Bristol-Myers Squibb described an optimized procedure that allowed the synthesis of 3,5-dihalo-2(1H)-pyrazinones in multigram quantities, involving lower reaction temperatures and the use of lower boiling point solvents.^{82,83} This procedure consists of the preparation of the α -aminonitrile *via* the nucleophilic substitution of chloroacetonitrile with a primary amine in the presence of KI and K_2CO_3 in MeCN followed by acidification and final treatment with oxalyl chloride or oxalyl bromide in toluene (Scheme 16).

2.7.1 Application of 3,5-dihalo-2(1H)-pyrazinones. 3,5-Dihalo-2(1H)-pyrazinones, synthesized using the procedures

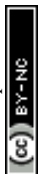


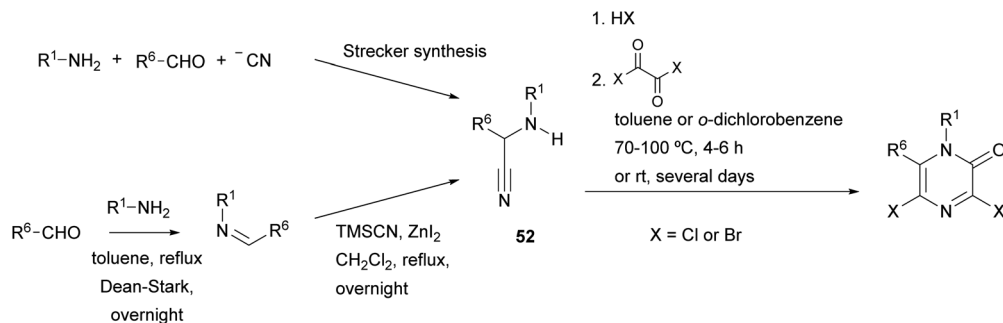


Scheme 10 Synthesis of 2(1H)-pyrazinones from an aminoacetal or amino alcohol, an oxalate, and an amine, and further derivatizations.

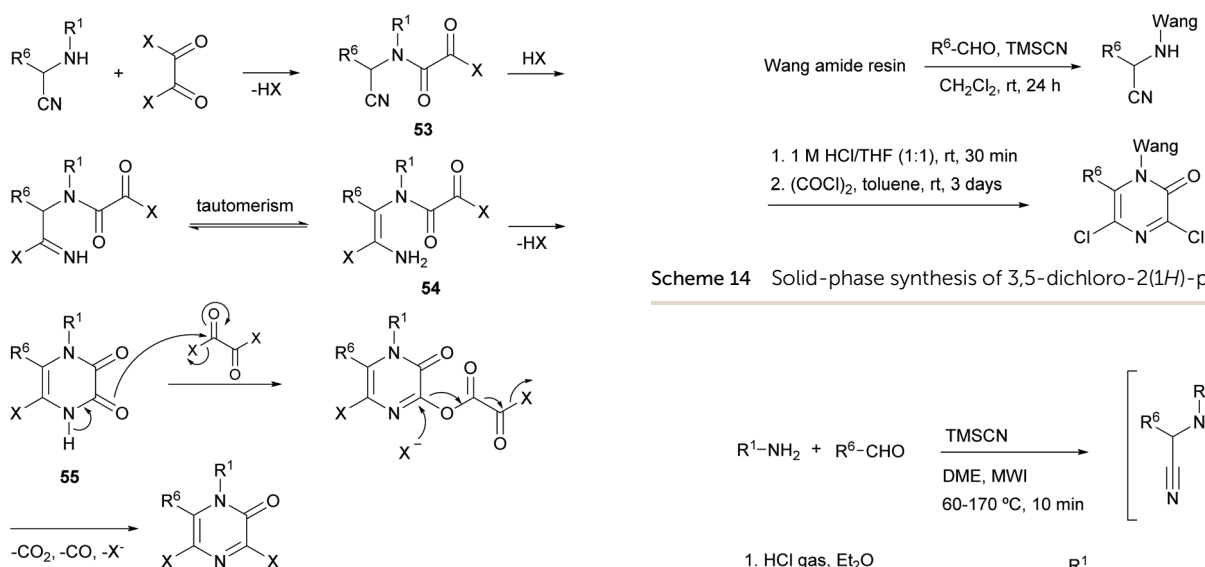
described above, have been shown to be versatile building blocks for the preparation of a variety of biologically relevant compounds. Position C-3, being an imidoyl chloride moiety, can be selectively derivatized with a variety of nucleophiles *via* addition/elimination reactions. In addition, metal-catalyzed reactions in solution as well as on solid phase have been used to incorporate aryl, heteroaryl, alkyl, alkenyl or alkynyl groups at

C-3. The functionalization of the C-5-vinylhalo position is more difficult, but once position 3 is derivatized, substituents can be introduced at C-5 through a metal-catalyzed cross-coupling reaction. Alternatively, both positions can be dehalogenated *via* a palladium-catalyzed reaction with sodium formate. It has also been demonstrated that the azadiene system of the pyrazinone system can undergo intra- and intermolecular

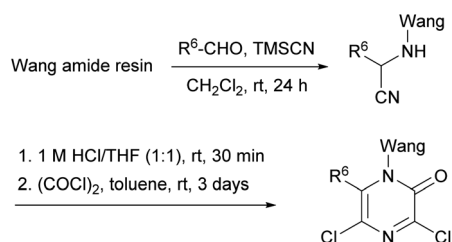




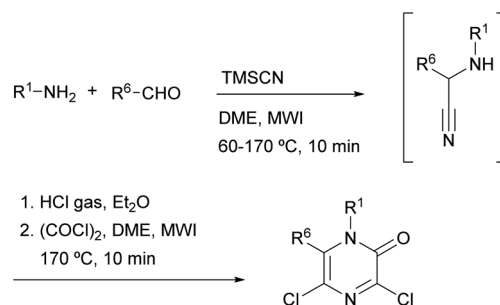
Scheme 11 Synthesis of 3,5-dihalo-2(1H)-pyrazinones using Hoornaert's method.



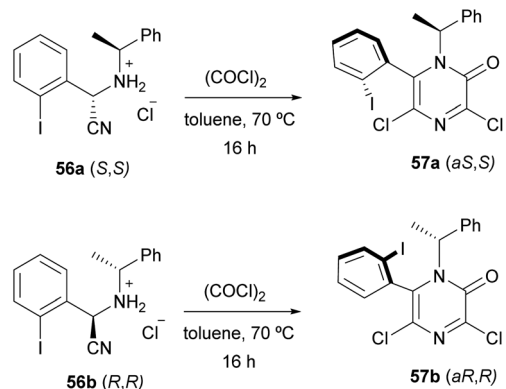
Scheme 12 Mechanism for the formation of 3,5-dihalo-2(1H)-pyrazinones.



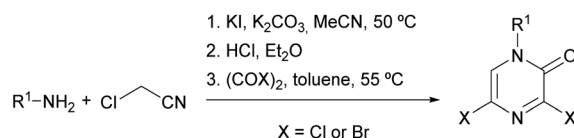
Scheme 14 Solid-phase synthesis of 3,5-dichloro-2(1H)-pyrazinones.



Scheme 15 Synthesis of 3,5-dichloro-2(1H)-pyrazinones under MWI.



Scheme 13 Synthesis of atropisomeric 6-aryl-3,5-dichloro-2(1H)-pyrazinones.



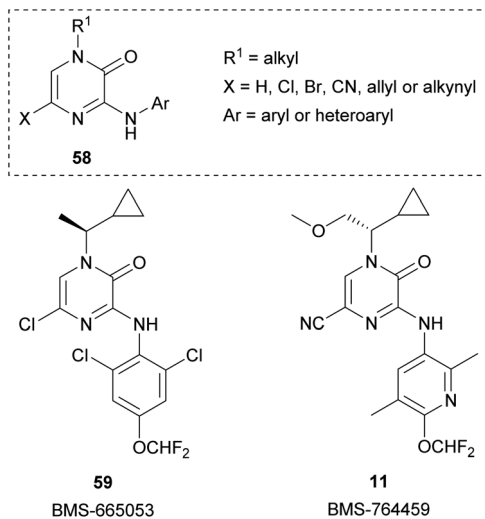
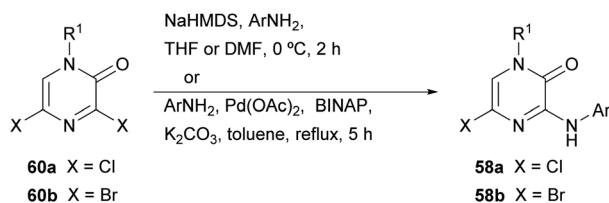
Scheme 16 Synthesis of 3,5-dihalo-2(1H)-pyrazinones starting from a primary amine and chloroacetonitrile.

cycloaddition reactions with a range of olefins and acetylenes to provide (fused)heterocyclic systems. All this reactivity, both in solution and on solid phase, has been previously reviewed.⁸⁴⁻⁸⁷

Herein, the developments made in this field during the ten years after 2010 are described.

Starting from 3,5-dihalo-2(1H)-pyrazinones, Bristol-Myers Squibb researchers developed a family of *N*-3-(hetero)aryl-2(1H)-pyrazinones as corticotropin-releasing factor-1 receptor (CRF₁R) antagonists that could be useful for the treatment of psychiatric disorders.^{18,82,83,88-90} The general structure **58** of this family of 2(1H)-pyrazinones and two of the most representative members, BMS-665053 (**59**) and BMS-764459 (**11**), are depicted in Fig. 3. Both compounds are highly potent (IC₅₀ ≤ 1.0 nM) and



Fig. 3 2(1H)-Pyrazinones developed as CRF₁R antagonists.Scheme 17 Synthesis of 5-halo-2(1H)-pyrazinones **58**.

selective CRF₁R antagonists with good pharmacokinetic properties and efficacy in rats, but BMS-764459 shows an improved metabolic profile.^{88,89} Based on this activity as CRF₁R antagonists, later they prepared fluorinated analogues and explored their potential as Positron Emission Tomography (PET) radioligands.^{91,92}

The synthesis of this family of compounds, with general structure **58**, was achieved *via* a base-mediated reaction or a palladium-catalyzed site-selective coupling of the corresponding (hetero)aryl amine with the 3,5-dihalo-2(1H)-pyrazinone **60a** or **60b** (Scheme 17),^{18,82,83,88–90} which were prepared

as shown in Scheme 16. 5-Bromo-2(1H)-pyrazinone **58b** served as a starting material for the incorporation of substituents at C-5 of the pyrazinone *via* palladium-catalyzed cross-coupling reactions (Scheme 18).¹⁸ The *des*-halo analogue was obtained by hydrogenation in presence of Pd/C.¹⁸ Analogues bearing a methyl group at position 5 were also prepared following an alternative method that involved the synthesis of a 3-chloro-5-methyl-2(1H)-pyrazinone intermediate by the condensation of an α -amino ketone and ethyl chlorooacetate using a similar approach to that discussed in Section 2.2.¹⁸

Sandström's group developed a family of achiral inhibitors of the hepatitis C virus NS3 protease based on 2(1H)-pyrazinones with general structure **61** (Fig. 4). They chose this scaffold for its ability to retain the H-bonding pattern of the peptide backbone and to act as a β -strand inducer. The starting point of their design was tripeptide **62**, and their first study led to the identification of compounds **61a** and **61b** (Fig. 4).⁸⁰ The

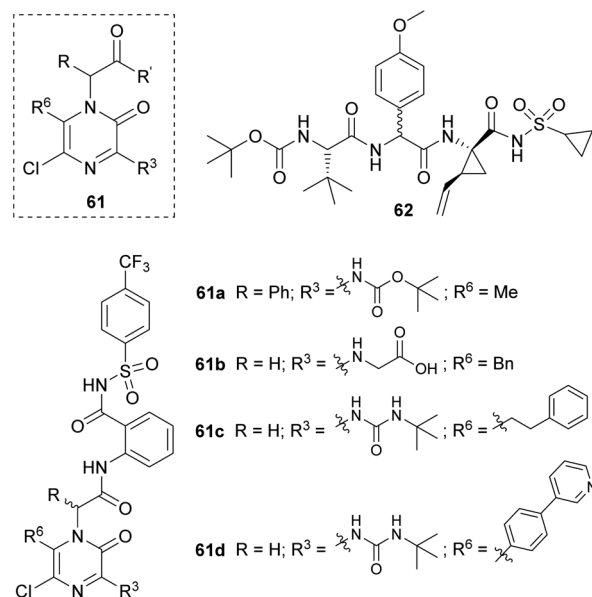
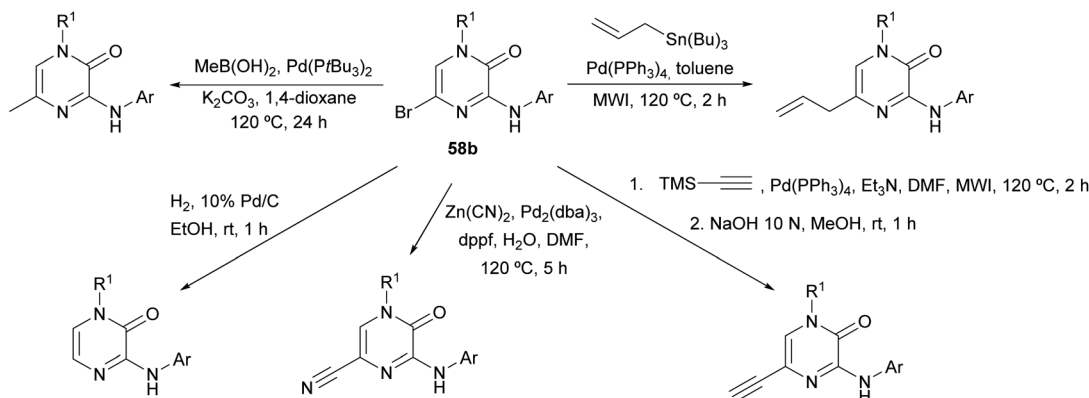


Fig. 4 2(1H)-Pyrazinones developed as inhibitors of the hepatitis C virus NS3 protease.

Scheme 18 Derivatization of 5-bromo-2(1H)-pyrazinone **58b**.

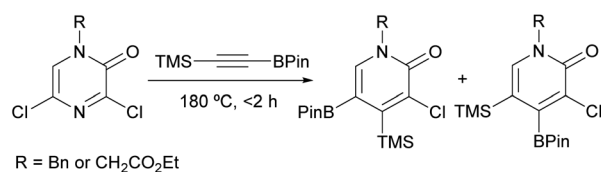
former was taken as a lead for further optimization.⁸¹ This second study provided **61c** and **61d** as two of the best 2(1*H*)-pyrazinone-based inhibitors and showed the important role of elongated substituents at C-6 for the activity.

The synthesis of the pyrazinone scaffold was performed *via* Hoornaert's method using MWI (Scheme 15).⁷⁹ The resulting 3,5-dichloro-2(1*H*)-pyrazinone, incorporating the required substituents at positions 1 and 6, was selectively derivatized at position 3 with *tert*-butyl carbamate or *tert*-butylurea *via* a palladium-catalyzed Buchwald *N*-arylation coupling using Pd(OAc)₂, Xantphos and Cs₂CO₃ in 1,2-dimethoxyethane (DME) under MWI. Alternatively, this position could be derivatized with an amino group *via* a nucleophilic aromatic substitution with the corresponding amine in presence of DIEA in CH₃CN.^{80,81}

In another study, Harker *et al.* described the synthesis of 2-pyridone boronic acid derivatives in multigram quantities *via* a [4 + 2] cycloaddition between a 3,5-dichloro-2(1*H*)-pyrazinone and a trimethylsilyl alkynylboronate (Scheme 19).⁹³ The corresponding pyrazinones were prepared on multigram scale following the procedure described by Bristol Myers Squibb researchers (Scheme 16).^{82,83} The cycloaddition was carried out at 180 °C in the absence of solvent for <2 h and provided the expected 2-pyridone boronic acid derivatives as a 2 : 1 mixture of regioisomers. These could be separated using high performance counter-current chromatography (HPLC) and were further derivatized to provide a range of functionalized 2-pyridones.

In 2006, DuPont patented the tubulin-modulating fungicide **12**, containing a pyrazinone core (Fig. 5).¹⁹ Later, they performed a structure–activity relationship analysis incorporating a range of modifications at positions N-1, C-3, C-5 and C-6 of the 2(1*H*)-pyrazinone ring.²⁰ Best derivatives, highlighting **63a–d** and **64a–b** included in Fig. 5, generally contained a branched alkyl group at N-1, a N-linked pyrazole at position 3, a chloro group at position 5 and a 2,4,6-trisubstituted phenyl ring at position 6. These compounds showed fungicidal activity against economically important plant pathogenic fungi and also exhibited activity similar to that of placlitaxel against rhabdomyosarcoma cells.

The substituent present at position 3 was introduced by treating the corresponding 3,5-dichloro-2(1*H*)-pyrazinone, obtained as described in Scheme 11, with a variety of nucleophiles or by subjecting it to a metal-catalyzed cross-coupling reaction, yielding compounds containing an N- or a C-linked heterocycle.²⁰ Amide groups were also introduced by treatment of the 3,5-dichloro-2(1*H*)-pyrazinone with *N*-cyanomethylbenzotriazoles in the presence of a base and subsequent oxidation with peracetic acid.



Scheme 19 Synthesis of 2-pyridones from 2(1*H*)-pyrazinones.

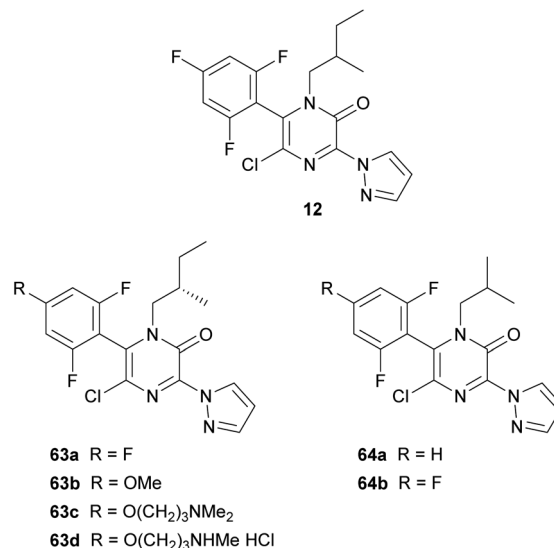


Fig. 5 2(1*H*)-Pyrazinones developed as tubulin-modulating compounds.

Yang *et al.* synthesized a collection of 1,3,5-trisubstituted-2(1*H*)-pyrazinones **65** and studied their activity as PB2 inhibitors (Fig. 6).⁹⁴ PB2 is a subunit of influenza RNA-dependent RNA polymerase and it is an important target for the design of anti-influenza drugs. The structure of these new 2(1*H*)-pyrazinones was based on VX787 (**66**), a PB2 inhibitor that has entered phase III clinical trials, by replacing the pyrimidine ring with a pyrazinone moiety. The best analogue **65a** is depicted in Fig. 6.

This family of compounds was prepared either from the commercially available 3,5-dibromo-1-methyl-2(1*H*)-pyrazinone or from a 3,5-dibromo-2(1*H*)-pyrazinone prepared following the procedure described in Scheme 16. This step provided the group at C-1. The substituent at C-3 was incorporated by reaction with the corresponding amine in presence of DIEA whereas the substituent at C-5 was introduced *via* a Suzuki–Miyaura cross-coupling using the required 7-azaindole-3-boronic acid pinacol ester.

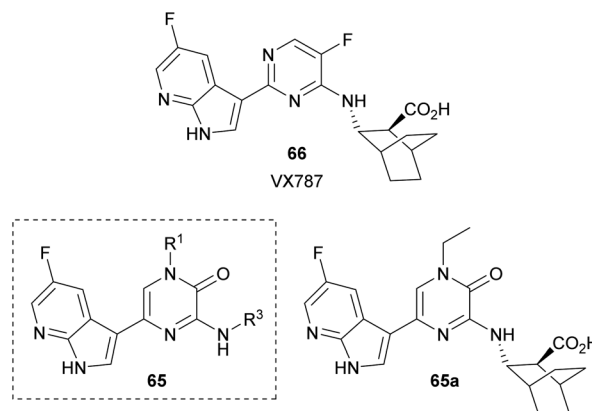


Fig. 6 2(1*H*)-Pyrazinones developed as PB2 inhibitors.



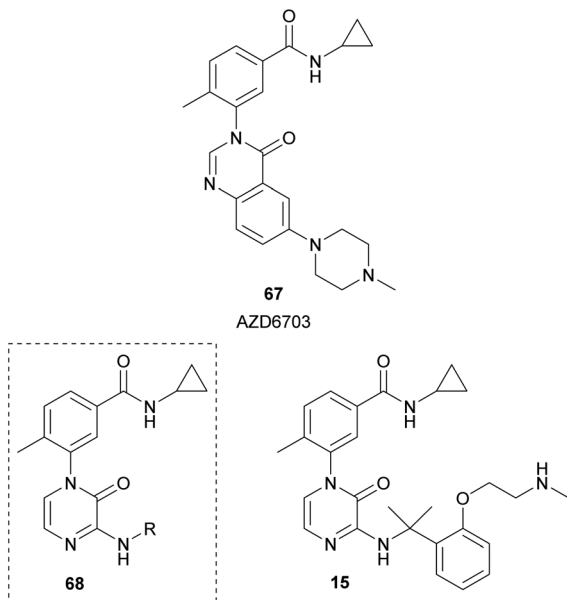


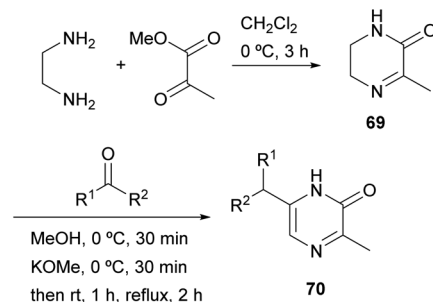
Fig. 7 2(1H)-Pyrazinones developed as p38 α MAP kinase inhibitors.

Researchers from AstraZeneca, starting from their lead compound AZD6703 (**67**), designed analogues **68** containing a 2(1H)-pyrazinone core instead of the quinazolinone scaffold (Fig. 7).²³ Their aim was the development of selective p38 α mitogen-activated protein kinase (MAPK) inhibitors. p38 α plays a relevant role in the modulation of the pro-inflammatory signalling. They performed a structure–activity relationship analysis by incorporating a variety of aminoalkyl groups at position 3 of the pyrazinone ring and identified compound **15** with *in vitro* and *in vivo* biological properties for an inhaled drug candidate.

The synthesis of this family of compounds was carried out from the required 1-aryl-3,5-dibromo-2(1H)-pyrazinone, which was obtained as depicted in Scheme 16. The aminoalkyl group at position 3 was incorporated *via* a nucleophilic aromatic substitution with the corresponding amine in presence of Et₃N. The bromine at position 5 was removed through hydrogen transfer hydrogenation using ammonium formate and Pd/C in EtOH under MWI.

2.8 Synthesis from ethylenediamine and methyl pyruvate

Masuda *et al.* described in 1981 another strategy in which the 1,2- and 4,3-bonds of the final pyrazinone are made (Scheme 20).⁹⁵ Methyl pyruvate was condensed with ethylenediamine to afford 3-methyl-5,6-dihydro-2(1H)-pyrazinone **69** *via* imine and amide formation. In the next step, a strange alkylation was achieved by reaction of **69** with an aldehyde or ketone under alkaline conditions. Although the authors first reported that reaction had taken place at position 5, it was later demonstrated by NOE experiments and X-ray crystallography that 6-alkyl-3-methyl-2(1H)-pyrazinones **70** were obtained.⁹⁶ The process probably involves deprotonation, prototropy and 6-alkylation at 5,6-enamine. Compounds **70** were further converted into



Scheme 20 Synthesis of 2(1H)-pyrazinones from ethylenediamine and methyl pyruvate.

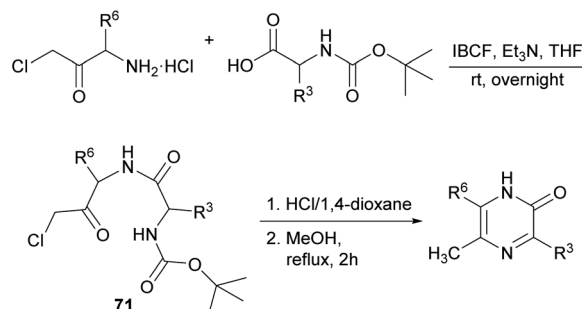
pyrazines bearing an alkoxy, a phenoxy, an alkylthio or a phenylthio group with interesting odour properties.

2.9 Synthesis from dipeptides

2.9.1 Synthesis from dipeptidyl chloromethyl ketones. A simple pathway for the preparation of 5-methyl-2(1H)-pyrazinones from dipeptidyl chloromethyl ketones **71** was first reported by Okada *et al.* (Scheme 21).^{97,98} Starting from the coupling of a *tert*-butyloxycarbonyl (Boc)-protected amino acid with a chloromethyl α -amino ketone using isobutyl chloroformate (IBCF), the key part of this approach involves a deprotection-cyclization sequence of **71** in which the Boc group is removed with HCl/1,4-dioxane and, subsequently, the pyrazinone ring is constructed under MeOH reflux by the formation of the N-4–C-5 bond.

For the last step, the presence of the chlorine atom in the dipeptidyl derivative plays a key role as it notably increases the electrophilicity of the carbonyl group, and it is easily released as a good leaving group favouring the formation of the pyrazinone. Moreover, the reaction can occur independently of the stereoisomerism of the starting reagent.⁹⁹ From this route, 2-acetoxypyrazines can be easily accessed, which have been proposed as useful acetylating reagents, reacting selectively with amino groups of amino acids in the presence of hydroxy groups.¹⁰⁰

This method constitutes an efficient strategy to provide highly substituted rings in high yields under mild conditions, depending on the amino acids employed that determine R³ and R⁶ in the final pyrazinone core: when aspartate or glutamate are



Scheme 21 Synthesis of 2(1H)-pyrazinones from Boc-protected amino acids and a chloromethyl amino ketone.



used as components of the starting dipeptidyl chloromethyl ketone, pyrazinone carboxylates can be obtained. Nevertheless, in contrast to that observed for position 6, carboxymethyl groups at R³ rapidly decarboxylate due to the low electron density of this position.¹⁰¹

In addition, amino groups can be easily introduced as substituents at positions 3 and/or 6, protected with a benzyloxycarbonyl (Z) group prior to the cyclization. Although the catalytic hydrogenation of the Z group of aminomethyl attached to position 6 causes instantaneous and selective deamination of this position,¹⁰² when larger aminoalkyl chains are introduced, the Z group can be removed and the amine further functionalized by coupling to various amino acids.

Based on this strategy, the development of opioid mimetics constitutes an interesting application for pyrazinone-containing compounds. Thus, Okada *et al.* synthesised a series of analogues of the pentapeptide enkephalin, a natural ligand of opioid receptors.^{103–105} Specifically, they incorporated a pyrazinone ring with aminoalkyls of different lengths as R³ or R⁶, at which a tyrosine was attached. Based on the results which pointed to a strong binding to the μ -opioid receptor and good lipophilicity given by the pyrazinone core, several modifications were prepared. Among them, the inclusion of 2',6'-dimethyl-L-tyrosine (Dmt) as in **72** (Fig. 8) has been reported to enhance receptor affinity, and to exhibit higher analgesic and antinociceptive activities.^{106–112} Moreover, Li *et al.* reported that *N*-allylation of Dmt in these scaffolds allows the conversion of μ -opioid agonists into potent μ -opioid antagonists.¹¹³

An additional example of an application related to pyrazinone derivatives synthesised following this strategy was reported by Miyazaki *et al.* whose work was aimed at the development of analogues of the somatostatin, a cyclic tetradecapeptide involved in the inhibition of growth hormone secretion.^{114,115} Particularly, they designed novel cyclic

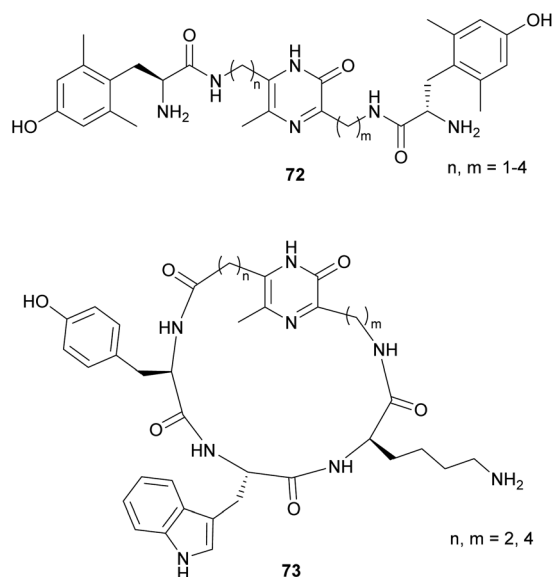
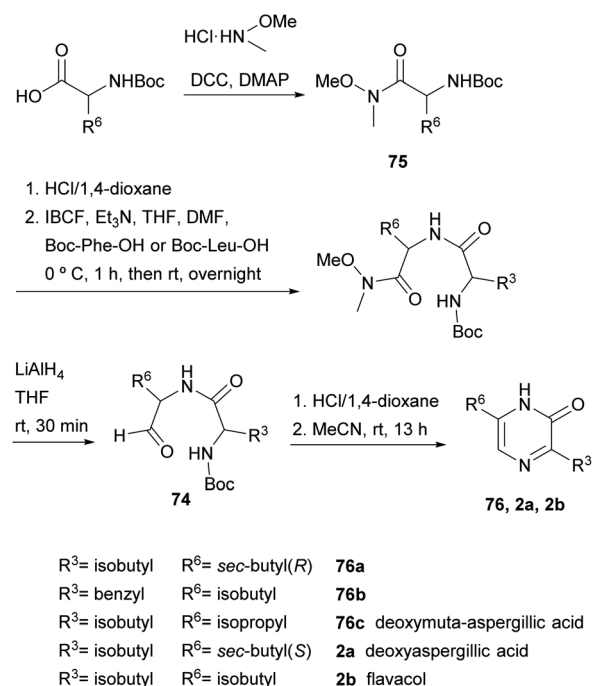


Fig. 8 General structures of pyrazinone-containing opioid derivatives with Dmt and somatostatin analogues.

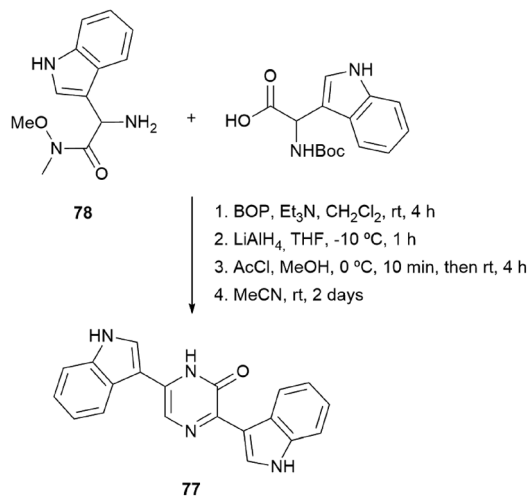
analogues which included a pyrazinone ring instead of the disulfide bond. The resulting compounds, with a general structure **73**, exhibited antiproliferative activity on tumour cells, but also presented interesting properties: pyrazinone-containing peptides are able to cross the blood–brain barrier, as well as the epithelial membrane of gastrointestinal tracts. Also, the presence of the heterocycle increases the lipophilicity, enhancing the interaction with cell membranes, and offers higher resistance to enzymatic degradation.

2.9.2 Synthesis from dipeptidyl aldehydes, ketones or carboxamides. The cyclization of dipeptidyl chloromethyl ketones following the preceding approach constitutes an easy and convenient strategy for the synthesis of 2(1*H*)-pyrazinones, however its use is restricted to pyrazinones bearing a methyl group at position 5. A similar but much more versatile approach for the preparation of pyrazinones, which also involves the formation of the bond between N-4 and C-5, is the cyclisation of dipeptides with a C-terminal aldehyde, methyl or phenyl ketone, or carboxamide.

In fact, in 1996, the research group that first developed the cyclization of dipeptidyl chloromethyl ketones, devised a route to pyrazinones without a substituent at position 5 starting from a dipeptidyl aldehyde **74** (Scheme 22).¹¹⁶ This aldehyde was obtained from a Boc protected α -amino Weinreb amide **75**, which was subjected to Boc removal followed by coupling with the second Boc amino acid in the sequence and final selective reduction of the Weinreb amide to an aldehyde. After Boc removal of **74**, cyclization was promoted at room temperature in MeCN. Following this approach, besides the 2(1*H*)-pyrazinones **76a–b**, the natural products deoxymuta-aspergillilic acid **76c**, deoxyaspergillilic acid **2a** and flavacol **2b** were prepared.



Scheme 22 Synthesis of 2(1*H*)-pyrazinones from dipeptidyl aldehydes.



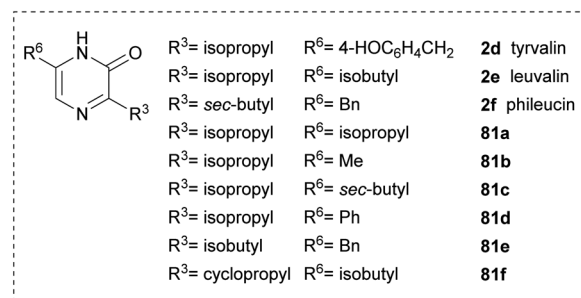
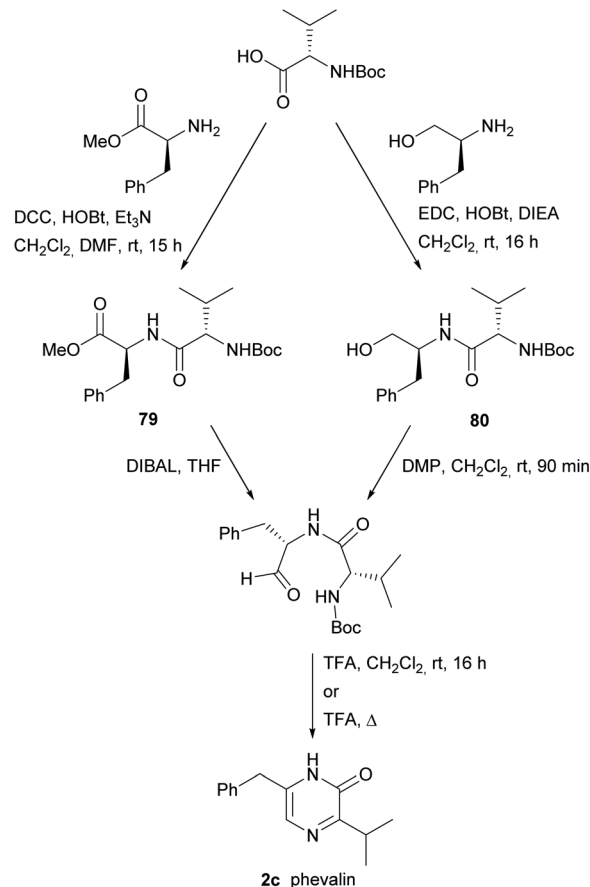
Scheme 23 Synthesis of 3,6-bis(3-indolyl)-2(1H)-pyrazinone.

Moreover, the stereochemistry of the chiral carbon atom of deoxyaspergillinic acid **2a** was determined to be *S*. Taking advantage of this method, Jiang and Gu prepared 3,6-bis(3-indolyl)-2(1H)-pyrazinones (**77**), corresponding to the core of dragsmacidin D, the key starting amino acids in this case being *N*-Boc-indolylglycine and the Weinreb amide **78** also derived from indolylglycine (Scheme 23).⁴²

Alternative approaches to obtain the dipeptidyl aldehydes were reported, thus Aubé *et al.* used a dipeptidyl ester **79** that was reduced to aldehyde with DIBAL or a dipeptidyl alcohol **80** that was oxidized to aldehyde with Dess-Martin periodinane (DMP) (Scheme 24). Phevalin (**2c**) was obtained following both approaches.^{117,118} Tyrvalin (**2c**), leuvalin (**2d**), phileucine (**2e**) and the six synthetic analogs **81a–f** were also prepared using an amino alcohol as starting material for the corresponding dipeptidyl aldehydes.¹¹⁸ Formation of the pyrazinones was achieved in trifluoroacetic acid (TFA) *via* a one-pot Boc removal, ring closure and spontaneous oxidation.

In 2004, Adam *et al.* and Matsushita *et al.* took this method a step further. On the one hand, the former authors prepared 2(1H)-pyrazinones again substituted at position 5, this time by cyclising dipeptidyl ketones **82** (Scheme 25).¹¹⁹ These intermediates were readily obtained through coupling Boc-protected amino acids with α -amino ketones or with α -amino secondary alcohols, the latter followed by an additional oxidation with DMP. After Boc removal of **82** with HCl, stirring the resulting dipeptidyl ketones in pyridine at 80 °C afforded the expected 5-phenyl or 5-methyl-2(1H)-pyrazinones **83** in high yields.

Matsushita *et al.* carried out N–H insertion reactions of α -diazo- β -ketoesters with Boc amino acid amides in the presence of a rhodium octanoate catalyst to afford α -acylamino- β -ketoesters **84** as precursors of pyrazinones (Scheme 26).¹²⁰ Their subsequent treatment with TFA at 100 °C led to Boc removal and cyclisation, yielding 2(1H)-pyrazinones **85** with a methyl or phenyl at position 5 and with a carboxylate substituent at position 6. To gain more insight into this method, the authors extended it to the solid phase. Accordingly, α -diazo- β -ketoesters

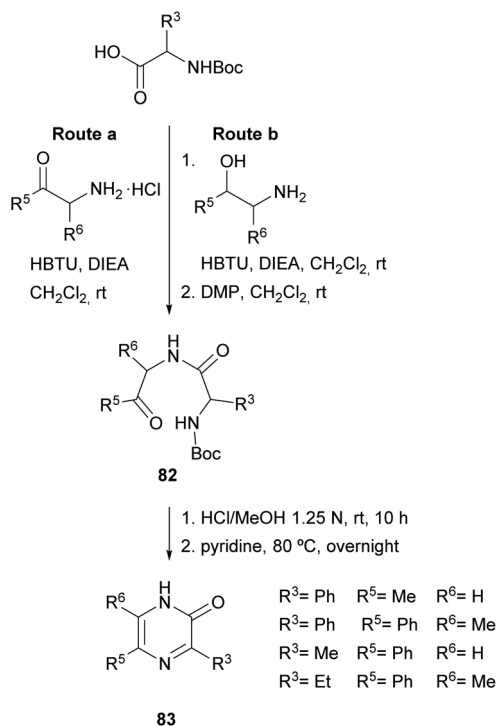


Scheme 24 Two routes for the preparation of dipeptidyl aldehydes as precursors of 3,6-disubstituted pyrazinones.

linked to a JandaJel polymer were used and converted to the resin-bound pyrazinones **86**. The procedure as described in solution was followed, except for the cyclization which was performed in acetic acid after Boc removal with TFA. Before cleavage from the resin, these pyrazinones **86** were then alkylated at N-1 or converted into pyrazines through reaction with POBr₃ and subsequent solid-phase Suzuki coupling with aryl boronic acids.

Dipeptides bearing a carboxamide group at the C-terminus have also been converted into 3,5,6-trisubstituted-2(1H)-pyrazinones, the substituent at position 5 being a carboxamide. With the aim of obtaining dipeptidyl carboxamides **87**, Basso *et al.* used the PADAM strategy, based on a Passerini multi-component reaction, amine deprotection and acyl migration (Scheme 27).¹²¹ In particular, they reacted an *N*-Fmoc α -amino



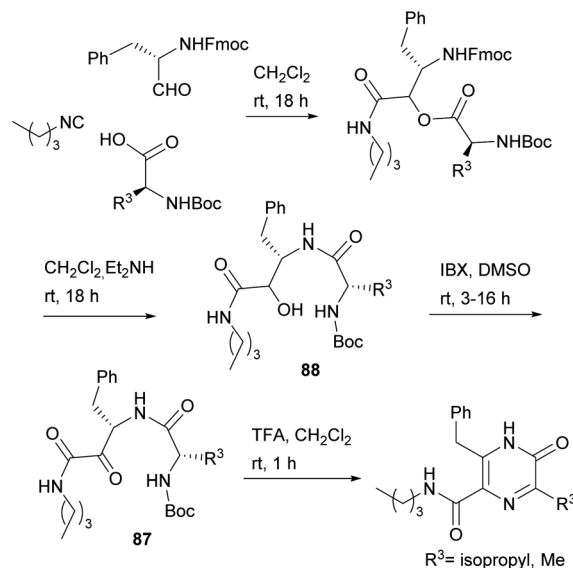


Scheme 25 Synthesis of 2(1H)-pyrazinones from dipeptidyl ketones.

aldehyde with *N*-Boc α -amino acids and an isocyanide, and then removed the Fmoc group with Et₂NH, which also led to spontaneous intramolecular acyl migration affording α -hydroxy amides **88**. The hydroxyl group of **88** was then oxidized with 2-iodoxybenzoic acid (IBX) to reach the desired dipeptidyl carboxamides **87**. Final treatment of **87** with TFA not only allowed Boc removal and *in situ* cyclization, but also the unexpected aromatisation of the ring, giving rise to the corresponding 3,5,6-trisubstituted-2(1H)-pyrazinones.

2.10 Synthesis through an Ugi condensation

Faggi *et al.* proposed a novel strategy for the synthesis of pyrazinones, with a carboxamide at position 6, through an Ugi four-component condensation between an arylglyoxal (R⁵ = Ar), a glyoxylic acid, an isocyanide, and a primary amine (Scheme

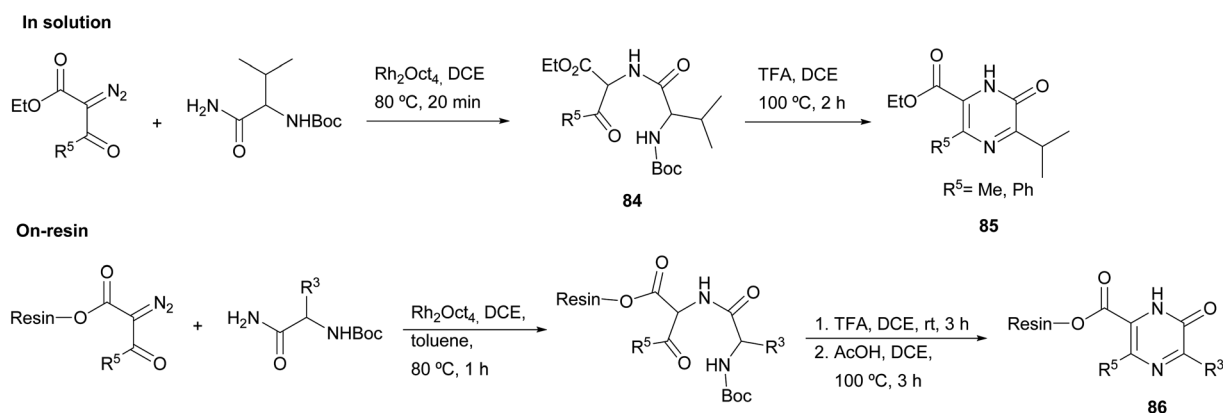


Scheme 27 Synthesis of 2(1H)-pyrazinones from dipeptidyl carboxamides.

28a).¹²² The condensation affords the intermediate **89**, which in the presence of ammonia undergoes a Davidson cyclization providing polysubstituted pyrazinone-6-carboxamides **90**.

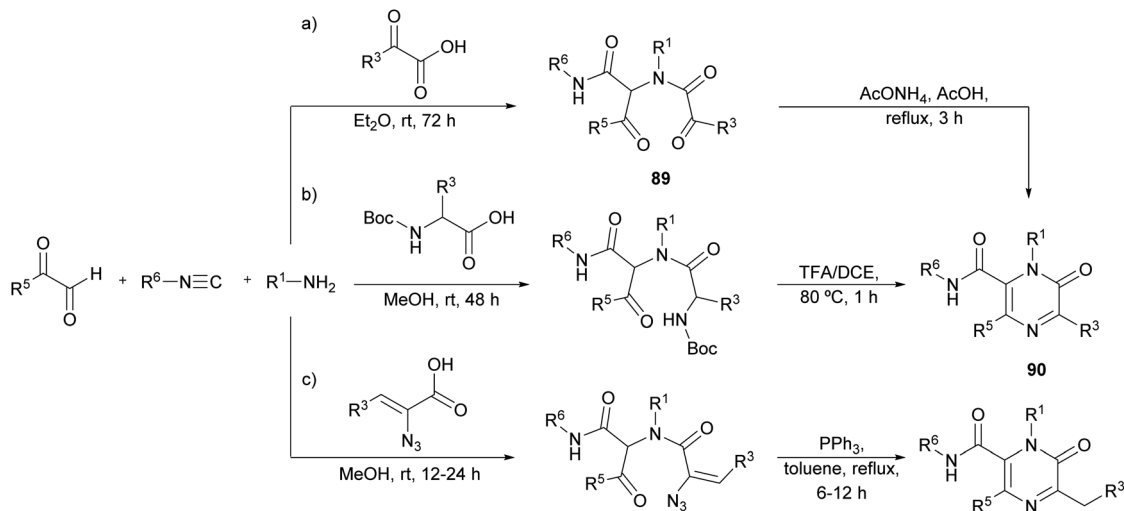
Later, Azuaje *et al.* further developed this Ugi-based strategy employing a *N*-Boc protected amino acid instead of the glyoxylic acid (Scheme 28b).¹²³ In their one-pot approach, after the four-component condensation, a subsequent step involving a deprotection/cyclization strategy with TFA/DCE provides pyrazinone **90**. Interestingly, they reported that when 2 N HCl/ether at 0 °C was employed to cleave the Boc group, a 3,4-dihdropyrazin-2(1H)-one was isolated instead of the target pyrazinone. The resulting pyrazinones have been proven to be efficient adenosine receptor antagonists.¹²⁴

More recently, Wu *et al.* reported a similar approach employing α -azidovinyl acids (Scheme 28c). After the Ugi condensation, the resulting intermediate was treated with PPh₃ leading to a one-pot Staudinger/Aza-Wittig/isomerization cascade giving 2(1H)-pyrazinones in an efficient way.¹²⁵



Scheme 26 The use of N-H insertion reactions for the preparation of 2(1H)-pyrazinones either in solution or on solid phase.





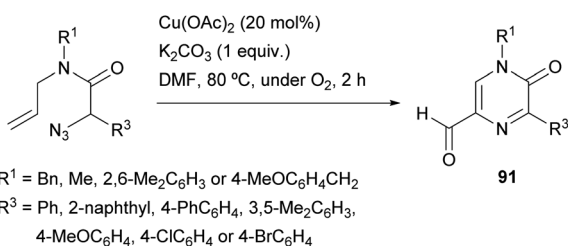
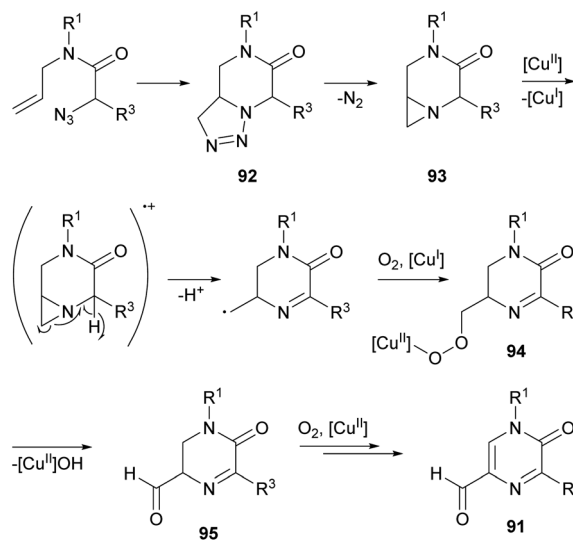
Scheme 28 Synthesis of 2(1H)-pyrazinones by means of Ugi reactions.

These strategies based on four-component reactions allow the assembly of pyrazinones with a wide variety of substituents depending on the readily available substrates involved.

2.11 Synthesis from α -azido-*N*-allylamides

Zhang *et al.* reported a synthesis of 5-formyl-2(1H)-pyrazinones starting from α -azido-*N*-allylamides *via* a copper(II)-catalyzed reaction under an O₂ atmosphere, that involves the formation of the bond between N-4 and C-5 (Scheme 29).¹²⁶ This procedure provided, in moderate to good yields, 5-formyl-2(1H)-pyrazinones **91** bearing an electron-rich aryl, a phenyl, a 4-chlorophenyl or a 4-bromophenyl at position 5 and a methyl, a benzyl or a 2,6-dimethylphenyl at N-1.

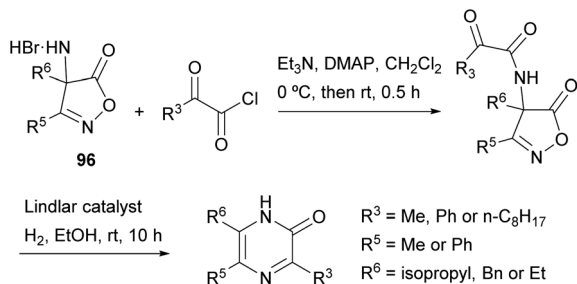
The proposed pathway for this transformation involves first a 1,3-dipolar cycloaddition between the azido group and the alkene moiety to yield a bicyclic triazolone **92** which could undergo denitrogenative fragmentation giving a bicyclic aziridine **93** (Scheme 30). Next, single-electron oxidation of this intermediate with Cu(OAc)₂ and deprotonation would result in ring opening of the aziridine yielding a primary alkyl radical, which would be transformed into a copper peroxy species **94** by reaction with O₂. Finally, elimination of copper hydroxide and oxidation of the resulting 5-formyl dihydropyrazinone **95** would give the corresponding 5-formyl-2(1H)-pyrazinone **91**.

Scheme 29 Synthesis of 2(1H)-pyrazinones from α -azido-*N*-allylamides.Scheme 30 Proposed mechanism for the synthesis of 2(1H)-pyrazinones from α -azido-*N*-allylamides.

2.12 Synthesis from 3,4-disubstituted 4-amino-5(4H)-isoxazolones

Becalli and Marchesini described a synthesis of 3,5,6-trisubstituted 2(1H)-pyrazinones from 3,4-disubstituted 4-amino-5(4H)-isoxazolones (Scheme 31).¹²⁷ This method is an alternative to the protocol first described by Jones for the preparation of 2(1H)-pyrazinones starting from α -amino acid amides and 1,2-dicarbonyl compounds, discussed in Section 2.4. First,⁴⁹ the corresponding 3,4-disubstituted 4-amino-5(4H)-isoxazolone **96** is treated with a 2-oxoacyl chloride in the presence of Et₃N and 4-(dimethylamino)pyridine (DMAP). Next, hydrogenation of the resulting intermediate, using the Lindlar catalyst, results in the reduction of the N–O bond, decarboxylation and intramolecular cyclization, yielding the 3,5,6-trisubstituted 2(1H)-pyrazinone.

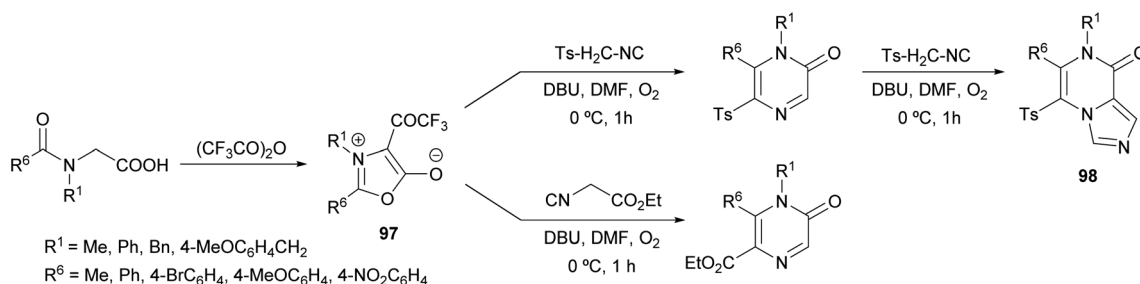




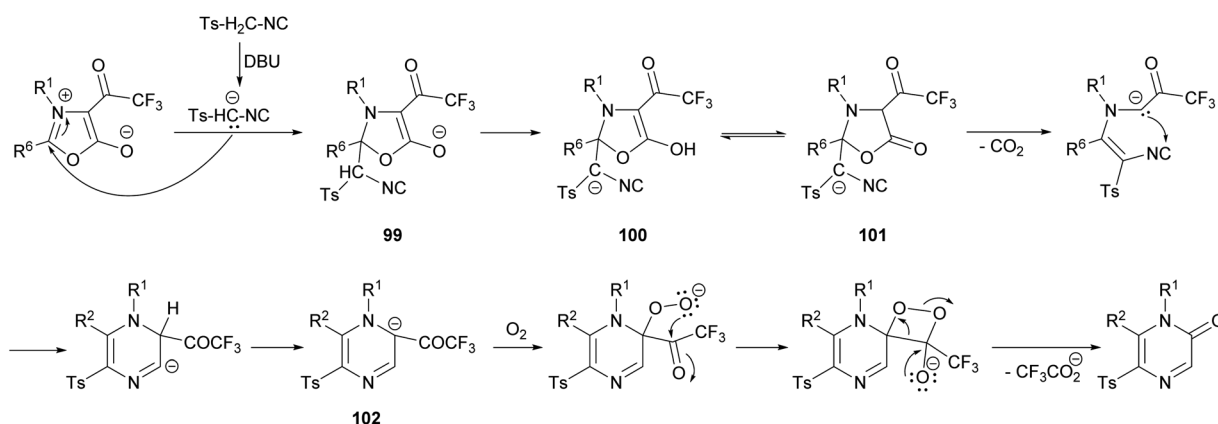
Scheme 31 Synthesis of 2(1H)-pyrazinones from 4-amino-5(4H)-isoxazolones.

2.13 Synthesis from mesoionic oxazoles and an isocyanide

Saijo *et al.* described a novel strategy to afford 2(1H)-pyrazinones by the ring transformation of mesoionic 1,3-oxazolium-5-olates by a base-mediated reaction with *p*-toluenesulfonylmethyl isocyanide (TosMIC) (Scheme 32).¹²⁸ Later, they found that this methodology could also be applied to ethyl isocyanoacetate as a reagent bearing an active methylene group.¹²⁹ In both cases, the 4-trifluoroacetyl-1,3-oxazolium-5-olate **97** is obtained by the cyclodehydration of a *N*-acyl-*N*-alkylglycine with trifluoroacetic anhydride. Moreover, if an excess of TosMIC is used, an imidazo[1,4-*a*]pyrazin-8(7H)-one derivative **98** is produced, due to the high electrophilic character of C-3 of the 2(1H)-pyrazinone core.



Scheme 32 Preparation of mesoionic oxazoles and their use in the synthesis of 2(1H)-pyrazinones.



Scheme 33 Proposed mechanism for the reaction of mesoionic oxazoles and an isocyanide.

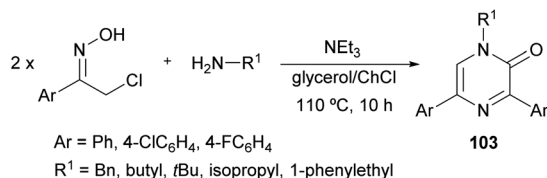
The mechanism of this reaction would involve the initial deprotonation of the active methylene group of TosMIC by 1,8-diazabicyclo(5.4.0)undec-7-ene (DBU) and a nucleophilic attack of the resulting carbanion to the C-2 position of the oxazolium affording the intermediate **99**, which is converted into **100** by proton transfer (Scheme 33).¹²⁸ This intermediate is in equilibrium with **101**, which after a decarboxylation promotes a nucleophilic attack causing the ring closure. After a proton transfer, intermediate **102** would react with O₂ forming a superoxide complex, which suffers a recombination affording the pyrazinone after the elimination of the trifluoroacetate anion.

2.14 Synthesis from 2-chloro ketone oximes and an amine in a deep eutectic solvent

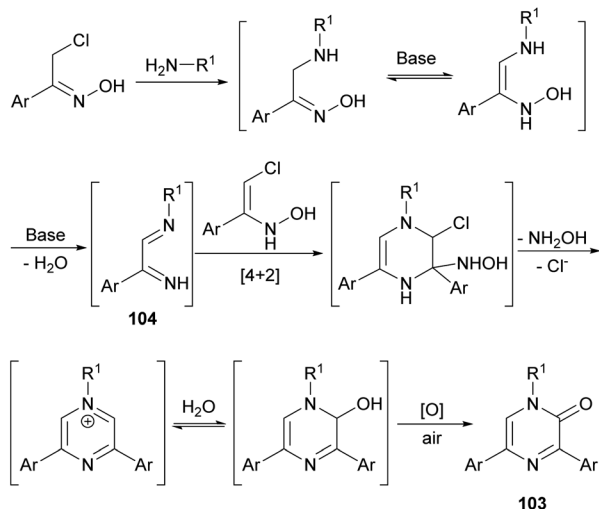
In 2017, Perrone *et al.* reported the first synthesis of 1-alkyl-3,5-diaryl trisubstituted 2(1H)-pyrazinones (**103**) in deep eutectic solvents (DESS) by means of the coupling between aromatic α -chloro oximes with aliphatic primary amines (Scheme 34).¹³⁰ α -Chloro ketone oximes, which are reported to be highly reactive, were prepared by a straightforward condensation of α -chloro ketones with hydroxylamine hydrochloride.

Regarding the optimization of the reaction conditions, they found that the use of choline chloride (ChCl) based DESSs, such as the combination glycerol/ChCl or urea/ChCl, resulted in better yields than using organic solvents like THF or DMF. In all cases, reactions were carried out at 110 °C for 10 h with an excess of the primary amine, and after a simple work-up,





Scheme 34 Synthesis of 2(1*H*)-pyrazinones from α -chloro oximes and an amine.



Scheme 35 Proposed mechanism for the formation of pyrazinones from α -chloro oximes and an amine.

pyrazinones were obtained and the DES mixtures could be recovered.

This route constitutes a green and simple way to obtain 2(1*H*)-pyrazinones. However, this methodology cannot be applied using aliphatic α -chlorinated oximes or aromatic amines as starting reagents. The mechanism of formation could involve an initial nucleophilic attack of the primary amine to the α -chloro oxime, a subsequent base-catalysed tautomerization and a water elimination *via* N–O bond breaking, affording the 1,4-diaza-1,3-diene **104** (Scheme 35).¹³⁰ This intermediate reacts with the enolic form of another α -chlorinated oxime through a [4 + 2] cycloaddition. The release of hydroxylamine and chloride favours the aromatization of the ring and a nucleophilic attack of water. A final oxidation step provides the pyrazinone **103**.

3 Concluding remarks

The synthesis of the 2(1*H*)-pyrazinone scaffold can be achieved through the combination of a wide variety of acyclic components as key building blocks, especially α -amino acids (and esters), α -amino ketones (and aldehydes), α -amino acid amides, glyoxylic acid derivatives, α -haloacyl halides, 1,2-dicarbonyl compounds, 2-chloro ketone oximes, α -aminonitriles, dipeptidyl chloromethyl ketones, dipeptidyl aldehydes, ketones and carboxamides, isocyanides, primary amines, and mesoionic

oxazoles. The review deals with the various synthetic strategies, starting from the earliest, published at the turn of the 20th century, up to those described in recent years. The ways in which the components are assembled to construct the heterocyclic ring, as well as the details of reaction conditions, are reviewed for each of these. All these approaches pave the way for the preparation of highly functionalized 2(1*H*)-pyrazinones with interesting biological applications.

Conflicts of interest

There are no conflicts to declare.

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References

- J. D. Dutcher, *J. Biol. Chem.*, 1947, **171**, 321–339.
- G. Dunn, G. T. Newbold and F. S. Spring, *J. Chem. Soc.*, 1949, 2586–2587.
- M. E. Álvarez, C. B. White, J. Gregory, G. C. Kydd, A. Harris, H. H. Sun, A. M. Gillum and R. Cooper, *J. Antibiot.*, 1995, **48**, 1165–1167.
- M. Zimmermann and M. A. Fischbach, *Chem. Biol.*, 2010, **17**, 925–930.
- N. Böhringer, M. Götschow, G. M. König and T. F. Schäberle, *Nat. Prod. Commun.*, 2017, **12**, 107–109.
- S. Umezawa, K. Tatsuta, T. Tsuchiya, H. Umezawa and H. Naganawa, *Tetrahedron Lett.*, 1971, **12**, 259–262.
- K. Tatsuta, K. Fujimoto, M. Yamashita, T. Tsuchiya, S. Umezawa and H. Umezawa, *J. Antibiot.*, 1973, **26**, 606–608.
- Y.-Q. Tang, I. Sattler, R. Thiericke, S. Grabley and X.-Z. Feng, *Eur. J. Org. Chem.*, 2001, **2**, 261–267.
- R. Jansen, S. Sood, K. I. Mohr, B. Kunze, H. Irschik, M. Stadler and R. Müller, *J. Nat. Prod.*, 2014, **77**, 2545–2552.
- F. Zhang, D. R. Braun, S. R. Rajski, D. DeMaria and T. S. Bugni, *Mar. Drugs*, 2019, **17**, 698.
- K. Hirano, T. Kubota, M. Tsuda, K. Watanabe, J. Fromont and J. Kobayashi, *Tetrahedron*, 2000, **56**, 8107–8110.
- A. E. Wright, S. A. Pomponi, S. S. Cross and P. McCarthy, *J. Org. Chem.*, 1992, **57**, 4772–4775.
- R. J. Capon, F. Rooney, L. M. Murray, E. Collins, A. T. R. Sim, J. A. P. Rostas, M. S. Butler and A. R. Carroll, *J. Nat. Prod.*, 1998, **61**, 660–662.
- A. Cutignano, G. Bifulco, I. Bruno, A. Casapullo, L. Gomez-Paloma and R. Riccio, *Tetrahedron*, 2000, **56**, 3743–3748.
- K. Motohashi, K. Inaba, S. Fuse, T. Doi, M. Izumikawa, S. T. Khan, M. Takagi, T. Takahashi and K. Shin-Ya, *J. Nat. Prod.*, 2011, **74**, 1630–1635.
- Y. Furuta, K. Takahashi, Y. Fukuda, M. Kuno, T. Kamiyama, K. Kozaki, N. Nomura, H. Egawa, S. Minami, Y. Watanabe, H. Narita and K. Shiraki, *Antimicrob. Agents Chemother.*, 2002, **46**, 977–981.



- 17 S. Joshi, J. Parkar, A. Ansari, A. Vora, D. Talwar, M. Tiwaskar, S. Patil and H. Barkate, *Int. J. Infect. Dis.*, 2021, **102**, 501–508.
- 18 R. A. Hartz, V. T. Ahuja, A. G. Arvanitis, M. Rafalski, E. W. Yue, D. J. Denhart, W. D. Schmitz, J. L. Ditta, J. A. Deskus, A. B. Brenner, F. W. Hobbs, J. Payne, S. Lelas, Y.-W. Li, T. F. Molski, G. K. Mattson, Y. Peng, H. Wong, J. E. Grace, K. A. Lentz, J. Qian-Cutrone, X. Zhuo, Y.-Z. Shu, N. J. Lodge, R. Zaczek, A. P. Combs, R. E. Olson, J. J. Bronson, R. J. Mattson and J. E. Macor, *J. Med. Chem.*, 2009, **52**, 4173–4191.
- 19 J. F. Berezna, P. L. Sharpe, R. B. Sheth, T. M. Stevenson, A. E. Taggi, C.-P. Tseng and W. Zhang, *PCT Application WO 2006089060*, E. I. DuPont De Nemours, 2006.
- 20 A. E. Taggi, T. M. Stevenson, J. F. Berezna, P. L. Sharpe, S. Gutteridge, R. Forman, J. J. Bisaha, D. Cordova, M. Crompton, L. Geist, P. Kovacs, E. Marshall, R. Sheth, C. Stavis and C.-P. Tseng, *Bioorg. Med. Chem.*, 2016, **24**, 435–443.
- 21 J. Heeres, M. R. de Jonge, L. M. H. Koymans, F. F. D. Daeyaert, M. Vinkers, K. J. A. Van Aken, E. Arnold, K. Das, A. Kilonda, G. J. Hoornaert, F. Compennolle, M. Cegla, R. A. Azzam, K. Andries, M.-P. de Béthune, H. Azijn, R. Pauwels, P. J. Lewi and P. A. J. Janssen, *J. Med. Chem.*, 2005, **48**, 1910–1918.
- 22 C. S. Burgey, K. A. Robinson, T. A. Lyle, P. E. J. Sanderson, S. D. Lewis, B. J. Lucas, J. A. Krueger, R. Singh, C. Miller-Stein, R. B. White, B. Wong, E. A. Lyle, P. D. Williams, C. A. Coburn, B. D. Dorsey, J. C. Barrow, M. T. Stranieri, M. A. Holahan, G. R. Sitko, J. J. Cook, D. R. McMasters, C. M. McDonough, W. M. Sanders, A. A. Wallace, F. C. Clayton, D. Bohn, Y. M. Leonard, T. J. Detwiler, J. J. Lynch, Y. Yan, Z. Chen, L. Kuo, S. J. Gardell, J. A. Shafer and J. P. Vacca, *J. Med. Chem.*, 2003, **46**, 461–473.
- 23 P. Raubo, R. Evans and P. Willis, *Bioorg. Med. Chem. Lett.*, 2020, **30**, 127412.
- 24 P. A. Allway, J. K. Sutherland and J. A. Joule, *Tetrahedron Lett.*, 1990, **31**, 4781–4782.
- 25 N. D. Yates, D. A. Peters, P. A. Allway, R. L. Beddoes, D. I. C. Scopes and J. A. Joule, *Heterocycles*, 1995, **40**, 331–347.
- 26 D. A. Peters, N. D. Yates, D. I. C. Scopes and J. A. Joule, *Heterocycles*, 1995, **40**, 983–991.
- 27 M. Helliwell, Y. You and J. A. Joule, *Heterocycles*, 2006, **70**, 87–91.
- 28 E. R. Ashley, E. G. Cruz and B. M. Stoltz, *J. Am. Chem. Soc.*, 2003, **125**, 15000–15001.
- 29 K. M. Allan and B. M. Stoltz, *J. Am. Chem. Soc.*, 2008, **130**, 17270–17271.
- 30 F. R. Japp and J. Knox, *J. Chem. Soc., Trans.*, 1905, **87**, 701–707.
- 31 H. McCombie and E. Parry, *J. Chem. Soc., Trans.*, 1909, **95**, 584–590.
- 32 C. Gastaldi, *Gazz. Chim. Ital.*, 1921, **51**, 233–255.
- 33 B. H. Ingham, *J. Chem. Soc.*, 1927, 692–700.
- 34 M. Busch and W. Foerst, *J. Prakt. Chem.*, 1928, **119**, 287.
- 35 Y. A. Tota and R. C. Elderfield, *J. Org. Chem.*, 1942, 313–319.
- 36 R. A. Baxter, G. T. Newbold and F. S. Spring, *J. Chem. Soc.*, 1947, 370–372.
- 37 N. Sato, *J. Heterocycl. Chem.*, 1978, **15**, 665–670.
- 38 R. H. Bradbury, D. Griffiths and J. E. Rivett, *Heterocycles*, 1990, **31**, 1647–1653.
- 39 E. Johannes, R. Horbert, J. Schlosser, D. Schmidt and C. Peifer, *Tetrahedron Lett.*, 2013, **54**, 4067–4072.
- 40 B. Pinchuk, E. Johannes, S. Gul, J. Schlosser, C. Schaechtele, F. Totzke and C. Peifer, *Mar. Drugs*, 2013, **11**, 3209–3223.
- 41 R. Horbert, B. Pinchuk, E. Johannes, J. Schlosser, D. Schmidt, D. Cappel, F. Totzke, C. Schächtele and C. Peifer, *J. Med. Chem.*, 2015, **58**, 170–182.
- 42 B. Jiang and X. H. Gu, *Heterocycles*, 2000, **7**, 1559–1568.
- 43 F. Y. Miyake, K. Yakushijin and D. A. Horne, *Org. Lett.*, 2002, **4**, 941–943.
- 44 R. A. Baxter and F. S. Spring, *J. Chem. Soc.*, 1947, **25**, 1179–1183.
- 45 J. J. Gallagher, G. T. Newbold, F. S. Spring and J. C. Woods, *J. Chem. Soc.*, 1949, 910–912.
- 46 J. J. Gallagher, G. T. Newbold, W. Sharp and F. S. Spring, *J. Chem. Soc.*, 1952, 4870–4874.
- 47 A. Ohta, Y. Aoyagi, Y. Kurihara, K. Yuasa, M. Shimazaki, T. Kurihara and H. Miyamae, *Heterocycles*, 1987, **26**, 3181–3191.
- 48 A. Ohta, Y. Aoyagi, Y. Kurihara, A. Kojima, K. Yuasa and M. Shimazaki, *Heterocycles*, 1988, **27**, 437–444.
- 49 R. G. Jones, *J. Am. Chem. Soc.*, 1949, **71**, 78–81.
- 50 G. Karmas and P. E. Spoerri, *J. Am. Chem. Soc.*, 1952, **74**, 1580–1584.
- 51 T. Konakahara and Y. Takagi, *Heterocycles*, 1978, **9**, 1733–1739.
- 52 G. Dunn, J. A. Elvidge, G. T. Newbold, D. W. C. Ramsay, F. S. Spring and W. Sweeny, *J. Chem. Soc.*, 1949, 2707–2712.
- 53 J. Maguire, D. Paton and F. L. Rose, *J. Chem. Soc. C*, 1969, 1593–1597.
- 54 F. L. Muehlmann and A. R. Day, *J. Am. Chem. Soc.*, 1956, **78**, 242–244.
- 55 M. Helliwell, Y. Yun and J. A. Joule, *Acta Crystallogr.*, 2006, **62**, o955–o956.
- 56 N. van Chuyen, T. Kurata and M. Fujimaki, *Agric. Biol. Chem.*, 1972, **36**, 1257–1258.
- 57 N. van Chuyen, T. Kurata and M. Fujimaki, *Agric. Biol. Chem.*, 1973, **37**, 1613–1618.
- 58 R. J. Bergeron and P. G. Hoffman, *J. Org. Chem.*, 1980, **45**, 163–165.
- 59 R. M. Seifert, R. G. Buttery, D. G. Guadagni, D. R. Black and J. G. Harris, *J. Agric. Food Chem.*, 1970, **18**, 246–249.
- 60 R. M. Seifert, R. G. Buttery, D. G. Guadagni, D. R. Black and J. G. Harris, *J. Agric. Food Chem.*, 1972, **20**, 135–137.
- 61 C. W. Lindsley, Z. Zhao, W. H. Leister, R. G. Robinson, S. F. Barnett, D. Defeo-Jones, R. E. Jones, G. D. Hartman, J. R. Huff, H. E. Huber and M. E. Duggan, *Bioorg. Med. Chem. Lett.*, 2005, **15**, 761–764.
- 62 A. Benjahad, R. Granet, P. Krausz, C. Bosgiraud and S. Delebassée, *Nucleosides Nucleotides*, 1996, **15**, 1849–1861.
- 63 A. L. Weber, *Orig. Life Evol. Biosph.*, 2008, **38**, 279–292.



- 64 M. Gately, S. Wong, J. Peoples, D. Galamay, G. Delgado, A. L. Weber and T. Campbell, *Nucleosides, Nucleotides Nucleic Acids*, 2020, **39**, 866–891.
- 65 C. Pierra, C. Counor, R. Storer and G. Gosselin, *Collect. Czech. Chem. Commun.*, 2011, **76**, 1327–1333.
- 66 M. Masaki and M. Ohta, *Bull. Chem. Soc. Jpn.*, 1963, **36**, 1177–1179.
- 67 Y. Chigira, M. Masaki and M. Ohta, *Bull. Chem. Soc. Jpn.*, 1966, **39**, 1014–1016.
- 68 G. W. H. Cheeseman, A. J. Freestone, R. A. Godwin and T. L. Hough, *J. Chem. Soc. Perkin Trans.*, 1975, **1**, 1888–1891.
- 69 F. J. Fleitz, T. A. Lyle, N. Zheng, J. D. Armstrong III and R. P. Volante, *Synth. Commun.*, 2000, **30**, 3171–3180.
- 70 J. E. Reiner, D. V. Siev, G. L. Araldi, J. J. Cui, J. Z. Ho, K. M. Reddy, L. Mamedova, P. H. Vu, K.-S. S. Lee, N. K. Minami, T. S. Gibson, S. M. Anderson, A. E. Bradbury, T. G. Nolan and J. E. Semple, *Bioorg. Med. Chem. Lett.*, 2002, **12**, 1203–1208.
- 71 J. J. Cui, G.-L. Araldi, J. E. Reiner, K. M. Reddy, S. J. Kemp, J. Z. Ho, D. V. Siev, L. Mamedova, T. S. Gibson, J. A. Gaudette, N. K. Minami, S. M. Anderson, A. E. Bradbury, T. G. Nolan and J. E. Semple, *Bioorg. Med. Chem. Lett.*, 2002, **12**, 2925–2930.
- 72 R. Singh, M. V. Silva Elipse, P. G. Pearson, B. H. Arison, B. K. Wong, R. White, X. Yu, C. S. Burgey, J. H. Lin and T. A. Baillie, *Chem. Res. Toxicol.*, 2003, **16**, 198–207.
- 73 P. E. J. Sanderson, K. J. Cutrona, D. L. Dyer, J. A. Krueger, L. C. Kuo, S. D. Lewis, B. J. Lucas and Y. Yan, *Bioorg. Med. Chem. Lett.*, 2003, **13**, 161–164.
- 74 J. Vekemans, C. Pollers-Wieers and G. Hoornaert, *J. Heterocycl. Chem.*, 1983, **20**, 919–923.
- 75 W. M. De Borggraeve, B. M. P. Verbist, F. J. R. Rombouts, V. G. Pawar, W. J. Smets, L. Kamoune, J. Alen, E. V. Van der Eycken, F. Compennolle and G. J. Hoornaert, *Tetrahedron*, 2004, **60**, 11597–11612.
- 76 J. Tulinsky, S. A. Mizesak, W. Watt, L. A. Dolak, T. Judge and R. B. Gammill, *Tetrahedron Lett.*, 1995, **12**, 2017–2020.
- 77 J. Tulinsky, B. V. Cheney, S. A. Mizesak, W. Watt, F. Han, L. A. Dolak, T. Judge and R. B. Gammill, *J. Org. Chem.*, 1999, **64**, 93–100.
- 78 N. Kaval, W. Dehaen and E. Van der Eycken, *J. Comb. Chem.*, 2005, **7**, 90–95.
- 79 J. Gising, P. Örtqvist, A. Sandström and M. Larhed, *Org. Biomol. Chem.*, 2009, **7**, 2809–2815.
- 80 P. Örtqvist, J. Gising, A. E. Ehrenberg, A. Vema, A. Borg, A. Karlén, M. Larhed, U. H. Danielson and A. Sandström, *Bioorg. Med. Chem.*, 2010, 6512–6525.
- 81 J. Gising, A. K. Belfrage, H. Alogheli, A. Ehrenberg, E. Åkerblom, R. Svensson, P. Artursson, A. Karlén, U. H. Danielson, M. Larhed and A. Sandström, *J. Med. Chem.*, 2014, **57**, 1790–1801.
- 82 R. A. Hartz, V. T. Ahuja, M. Rafalski, W. D. Schmitz, A. B. Brenner, D. J. Denhart, J. L. Ditta, J. A. Deskus, E. W. Yue, A. G. Arvanitis, S. Lelas, Y.-W. Li, T. F. Molski, H. Wong, J. E. Grace, K. A. Lentz, J. Li, N. J. Lodge, R. Zaczek, A. P. Combs, R. E. Olson, R. J. Mattson, J. J. Bronson and J. E. Macor, *J. Med. Chem.*, 2009, **52**, 4161–4172.
- 83 J. Li, D. Smith, S. Krishnananthan, R. A. Hartz, B. Dasgupta, V. Ahuja, W. D. Schmitz, J. J. Bronson, A. Mathur, J. C. Barrish and B.-C. Chen, *Org. Process Res. Dev.*, 2012, **16**, 156–159.
- 84 N. Kaval, K. Bisztray, W. Dehaen, C. O. Kappe and E. Van der Eycken, *Mol. Divers.*, 2003, **7**, 125–133.
- 85 V. G. Pawar and W. M. de Borggraeve, *Synthesis*, 2006, 2799–2814.
- 86 N. Kaval, P. Appukkuttan and E. Van der Eycken, *Top Heterocycl. Chem.*, 2006, **1**, 267–304.
- 87 V. P. Mehta, P. Appukkuttan and E. Van der Eycken, *Curr. Org. Chem.*, 2011, **15**, 265–283.
- 88 R. A. Hartz, V. T. Ahuja, X. Zhuo, R. J. Mattson, D. J. Denhart, J. A. Deskus, V. M. Vrudhula, S. Pan, J. L. Ditta, Y.-Z. Shu, J. E. Grace, K. A. Lentz, S. Lelas, Y.-W. Li, T. F. Molski, S. Krishnananthan, H. Wong, J. Qian-Cutrone, R. Schartman, R. Denton, N. J. Lodge, R. Zaczek, J. E. Macor and J. J. Bronson, *J. Med. Chem.*, 2009, **52**, 7653–7668.
- 89 X. Zhuo, R. A. Hartz, J. J. Bronson, H. Wong, V. T. Ahuja, V. M. Vrudhula, J. E. Leet, S. Huang, J. E. Macor and Y.-Z. Shu, *Drug Metab. Dispos.*, 2010, **38**, 5–15.
- 90 R. A. Hartz, V. T. Ahuja, W. D. Schmitz, T. F. Molski, G. K. Mattson, N. J. Lodge, J. J. Bronson and J. E. Macor, *Bioorg. Med. Chem. Lett.*, 2010, 1890–1894.
- 91 J. A. Deskus, D. D. Dischino, R. J. Mattson, J. L. Ditta, M. F. Parker, D. J. Denhart, D. Zuev, H. Huang, R. A. Hartz, V. T. Ahuja, H. Wong, G. K. Mattson, T. F. Molski, J. E. Grace Jr., L. Zueva, J. M. Nielsen, H. Dulac, Y.-W. Li, M. Guaraldi, M. Azure, D. Onthank, M. Hayes, E. Wexler, J. McDonald, N. J. Lodge, J. J. Bronson and J. E. Macor, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 6651–6655.
- 92 D. J. Denhart, D. Zuev, J. L. Ditta, R. A. Hartz, V. T. Ahuja, R. J. Mattson, H. Huang, G. K. Mattson, L. Zueva, J. M. Nielsen, E. S. Kozlowski, N. J. Lodge, J. J. Bronson and J. E. Macor, *Bioorg. Med. Chem. Lett.*, 2013, **23**, 2052–2055.
- 93 W. R. R. Harker, P. M. Delaney, M. Simms, M. J. Tozer and J. P. A. Harrity, *Tetrahedron*, 2013, **69**, 1546–1552.
- 94 J. Yang, J. Du, C. Huang, T. Wang, L. Huang, S. Yang and L. Li, *Bioorg. Med. Chem. Lett.*, 2019, **29**, 1609–1613.
- 95 H. Masuda, M. Yoshida and T. Shibamoto, *J. Agric. Food Chem.*, 1981, **29**, 944–947.
- 96 S. Mihara and H. Masuda, *J. Agric. Food Chem.*, 1990, **38**, 1032–1034.
- 97 H. Taguchi, T. Yokoi, F. Kasuya, Y. Nishiyama, M. Fukui and Y. Okada, *J. Chem. Soc. Chem. Commun.*, 1994, 247.
- 98 Y. Okada, H. Taguchi, Y. Nishiyama and T. Yokoi, *Tetrahedron Lett.*, 1994, **35**, 1231–1234.
- 99 H. Taguchi, T. Yokoi, M. Tsukatani and Y. Okada, *Tetrahedron*, 1995, **51**, 7361–7372.
- 100 H. Taguchi, K. Hirano, T. Yokoi, K. Asada and Y. Okada, *Chem. Pharm. Bull.*, 1995, **43**, 1336–1339.



- 101 H. Taguchi, T. Yokoi and Y. Okada, *Chem. Pharm. Bull.*, 1996, **44**, 2037–2041.
- 102 Y. Okada, Y. Fujisawa, A. Morishita, K. Shiotani, A. Miyazaki, Y. Fujita, T. Li, Y. Tsuda, T. Yokoi, S. D. Bryant and L. H. Lazarus, *Tetrahedron Lett.*, 2002, **43**, 8137–8139.
- 103 Y. Okada, M. Tsukatani, H. Taguchi, T. Yokoi, S. D. Bryant and L. H. Lazarus, *Chem. Pharm. Bull.*, 1998, **46**, 1374–1382.
- 104 Y. Okada, A. Fukumizu, M. Takahashi, T. Yokoi, Y. Tsuda, S. D. Bryant and L. H. Lazarus, *Chem. Pharm. Bull.*, 1999, **47**, 1193–1195.
- 105 Y. Okada, A. Fukumizu, M. Takahashi, J. Yamazaki, T. Yokoi, Y. Tsuda, S. D. Bryant and L. H. Lazarus, *Tetrahedron*, 1999, **55**, 14391–14406.
- 106 Y. Jinsmaa, A. Miyazaki, Y. Fujita, T. Li, Y. Fujisawa, K. Shiotani, Y. Tsuda, T. Yokoi, A. Ambo, Y. Sasaki, S. D. Bryant, L. H. Lazarus and Y. Okada, *J. Med. Chem.*, 2004, **47**, 2599–2610.
- 107 Y. Jinsmaa, Y. Okada, Y. Tsuda, K. Shiotani, Y. Sasaki, A. Ambo, S. D. Bryant and L. H. Lazarus, *J. Pharmacol. Exp. Ther.*, 2004, **309**, 432–438.
- 108 T. Li, Y. Fujita, K. Shiotani, A. Miyazaki, Y. Tsuda, A. Ambo, Y. Sasaki, Y. Jinsmaa, E. Marczak, S. D. Bryant, S. Salvadori, L. H. Lazarus and Y. Okada, *J. Med. Chem.*, 2005, **48**, 8035–8044.
- 109 K. Shiotani, T. Li, A. Miyazaki, Y. Tsuda, S. D. Bryant, A. Ambo, Y. Sasaki, L. H. Lazarus and Y. Okada, *Bioorg. Med. Chem. Lett.*, 2006, **16**, 5793–5796.
- 110 Y. Koda, K. Shiotani, I. Toth, Y. Tsuda, Y. Okada and J. T. Blanchfield, *Bioorg. Med. Chem. Lett.*, 2007, **17**, 2043–2046.
- 111 K. Shiotani, A. Miyazaki, T. Li, Y. Tsuda, T. Yokoi, A. Ambo, Y. Sasaki, S. Bryant, Y. Jinsmaa, L. Lazarus and Y. Okada, *Med. Chem.*, 2007, **3**, 583–598.
- 112 K. Shiotani, T. Li, A. Miyazaki, Y. Tsuda, T. Yokoi, A. Ambo, Y. Sasaki, S. D. Bryant, L. H. Lazarus and Y. Okada, *Bioorg. Med. Chem. Lett.*, 2007, **17**, 5768–5771.
- 113 T. Li, Y. Jinsmaa, M. Nedachi, A. Miyazaki, Y. Tsuda, A. Ambo, Y. Sasaki, S. D. Bryant, E. Marczak, Q. Li, H. S. Swartzwelder, L. H. Lazarus and Y. Okada, *Bioorg. Med. Chem.*, 2007, **15**, 1237–1251.
- 114 A. Miyazaki, T. Yokoi, Y. Tachibana, R. Enomoto, E. Lee, G. Bokonyi, G. Keri, Y. Tsuda and Y. Okada, *Tetrahedron Lett.*, 2004, **45**, 6323–6327.
- 115 A. Miyazaki, Y. Tsuda, S. Fukushima, T. Yokoi, T. Vántus, G. Bökönyi, E. Szabó, A. Horváth, G. Kéri and Y. Okada, *Bioorg. Med. Chem. Lett.*, 2008, **18**, 6199–6201.
- 116 Y. Okada, H. Taguchi and T. Yokoi, *Tetrahedron Lett.*, 1996, **37**, 2249–2252.
- 117 Y. Zeng, Q. Li, R. P. Hanzlik and J. Aubé, *Bioorg. Med. Chem. Lett.*, 2005, **15**, 3034–3038.
- 118 R. Ramesh, M. T. Bovino, Y. Zeng and J. Aubé, *J. Org. Chem.*, 2019, **84**, 3647–3651.
- 119 I. Adam, D. Orain and P. Meier, *Synlett*, 2004, 2031–2033.
- 120 H. Matsushita, S. H. Lee, K. Yoshida, B. Clapham, G. Koch, J. Zimmermann and K. D. Janda, *Org. Lett.*, 2004, **6**, 4627–4629.
- 121 A. Basso, L. Banfi, G. Guanti, R. Riva and P. Tosatti, *Synlett*, 2011, 2009–2012.
- 122 C. Faggi, M. García-Valverde, S. Marcaccini, R. Pepino and M. C. Pozo, *Synthesis*, 2003, 1553–1558.
- 123 J. Azuaje, A. El Maatougui, J. M. Pérez-Rubio, A. Coelho, F. Fernández and E. Sotelo, *J. Org. Chem.*, 2013, **78**, 4402–4409.
- 124 J. Azuaje, C. Carbajales, M. González-Gómez, A. Coelho, O. Caamaño, H. Gutiérrez-De-Terán and E. Sotelo, *Future Med. Chem.*, 2015, **7**, 1373–1380.
- 125 J. Wu, L. Zhao, M.-L. Yang and M.-W. Ding, *J. Org. Chem.*, 2021, **86**, 10755–10761.
- 126 L. Zhang, J.-Y. Lee, N. Yamazaki and S. Chiba, *Synlett*, 2011, 2167–2170.
- 127 E. M. Becalli and A. Marchesini, *Synthesis*, 1991, 861–862.
- 128 R. Saijo, K.-I. Kurihara, K. Akira, H. Uno and M. Kawase, *Tetrahedron Lett.*, 2013, **54**, 4418–4421.
- 129 R. Saijo, H. Sekiya, E. Tamai, K.-I. Kurihara, J. Maki, H. Sakagami and M. Kawase, *Chem. Pharm. Bull.*, 2017, **65**, 365–372.
- 130 S. Perrone, M. Capua, F. Messa, A. Salomone and L. Troisi, *Tetrahedron*, 2017, **73**, 6193–6198.

