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Degradable glycopolymers for saRNA transfection†

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Gene delivery is a powerful technique that is often exploited in the treatment of several diseases. Currently, most gene vectors are expensive virus-based systems, which can trigger immune responses. As a cheaper and safer alternative to these systems, biodegradable polymers have been widely used to develop gene delivery systems due to their different desirable characteristics. In particular, poly(amido amine)s possess critical desirable characteristics, such as high cell transfection activity, that make them suitable and efficient for gene delivery. In this study, reducible poly(amido amine)s with different side chain lengths and glycopolymers have been developed to create polyplexes with self-amplifying RNA (saRNA). The cell transfection assay showed that sugar decorated poly(amido amine)s revealed better saRNA transfection activity than other non-decorated poly(amido amine)s. Nevertheless, no differences were obtained between the poly(amido amine)s with different side chain lengths. Overall, biodegradable poly(amido amine)s with different alcohols and sugars have been synthesized with high molecular weights and low molecular weight distributions to develop an optimal saRNA delivery system.

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Introduction

In recent years, considerable attention has been drawn towards gene therapies which have shown promising results in the treatment of a variety of illnesses including diabetes, blindness, Parkinson's disease and many types of cancer.^{1,2} Gene therapies, however, often rely on the use of viral systems to deliver biomolecules such as DNA and mRNA and can trigger an immune response.^{1,3,4} Consequently, the drive to find safer alternatives to modified viral vectors has increased considerably over the last years. Polymers are generally inexpensive, easy to prepare, non-toxic and possess good cellular uptake.^{5,6}

Extensive research on poly(amido amine)s prepared *via* the Michael polyaddition reaction has shown that these polymers possess powerful gene delivery properties.^{3,7} For examples, poly(amido amine)s have been demonstrated to readily form polyplexes with the negatively charged nucleic acid sequences since the nitrogen atoms present on the backbone undergo

protonation under physiological conditions.⁶ Moreover, they possess excellent water solubility as well as stability against hydrolysis.³ This is an essential characteristic since the nucleic acid sequences can stay protected within the vehicles and arrive safely at the desired target(s).^{3,7-9}

Overall, even though poly(amido amine)s are a promising class of gene delivery vehicles that have demonstrated high transfection efficiency, low cytotoxicity, and biodegradability, their suitability for specific gene delivery applications is limited due to their high charge density and molecular weight, which can result in toxicity and non-specific interactions with biological components such as cell membranes.

Lastly, poly(amido amine)s possess greater transfection efficiency compared to other polymers as the OH-containing side chains allow the polymers to form hydrogen bonds with the cell membrane resulting in enhanced cellular uptake.^{4,10} A very recent example of this is the preparation of poly(amido amine)s of up to 167 kDa from a reducible bisacrylamide (*N*, *N'*-bis(acryloyl)cystamine, BAC) and 4-amino-1-butanol.² The large poly(amido amine)s were able to form complexations with self-amplifying RNA (saRNA) which has the potential to be an excellent vaccine platform for various illnesses, viruses and cancer.⁶ Additionally, it was shown that the saRNA/poly(amido amine) complexes led to very high protein expression *in vivo* as well as high percentage of saRNA expressing cells *ex vivo* (human skin),² thus providing evidence that these polymers have real potential. Therefore, poly(amido amine)s-based glycopolymers with different carbohydrate moieties could

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advance this real potential in terms of their ability to efficiently transfect cells with low cytotoxicity, high stability and specific targeting properties.^{11,12}

In this work, the conditions for the aza-Michael polyaddition of different poly(amido amines) are explored to find the best conditions for the efficient preparation of high molecular weight polymers. The poly(amido amine)s were prepared from a reducible bisacrylamide, *N,N'*-bis(acryloyl)cystamine (BAC) and a range of amines with different spacer lengths and also with mannose and galactose amines. The polymers were protonated to form polyplexes with saRNA at different N/P ratios. The cell transfection studies have been carried out *via* different cell lines. Additionally, poly(amido amine)s with carbohydrate were tested in terms of their binding activity with two human lectins *via* Surface plasmon resonance (SPR) spectrometer.^{13–16}

Materials and methods

Materials

N,N'-Bis(acryloyl)cystamine (BAC), mannose amine and galactose amine were synthesised following literature procedures.^{40,41} Ethanolamine ($\geq 98\%$), 3-amino-1-propanol ($\geq 99\%$), 4-amino-1-butanol (98%), 5-amino-1-butanol ($\geq 92\%$), propargylamine (98%) were purchased from Sigma Aldrich and used as received.

Instruments and analysis

Proton nuclear magnetic resonance (^1H NMR) was measured on a Bruker DPX-300. The NMR samples were in $(\text{CD}_3)_2\text{SO}$ or CDCl_3 . For $\text{DMSO}-d_6$, the resonance signal of residual DMSO at 2.50 ppm (^1H) served as reference peaks for chemical shifts. The resonance signal of residual CHCl_3 in CDCl_3 is at 7.26 ppm. Gel permeation chromatography (GPC) measurements of polymers were carried out on an Agilent 1260 Infinity II-MDS instrument with two PLgel Mixed-D columns operating in DMF with 5 mM NH_4BF_4 or with PlAquagel Mixed-M columns in H_2O with 0.5% NaN_3 at 25 °C equipped with the following detectors: a refractive index (RI), viscometer, light scattering (LS), and variable wavelength detector (VWD). The instrument was calibrated with linear polyethylene glycol or poly(methyl methacrylate) standards (500–1 500 000 g mol $^{-1}$). The flow rate was 1 mL min $^{-1}$. All samples were passed through 0.2 micron nylon filters prior to GPC measurements. Dynamic light scattering (DLS) measurements were carried out on a Malvern nano-series DLS instrument. The measurements were carried out in distilled water at 25 °C using disposable cuvettes and were repeated three times. Surface plasmon resonance (SPR) measurements were performed on a T200 BIAcore system (GE Healthcare). The lectins MBL and CLEC10A (5 $\mu\text{g mL}^{-1}$) were immobilized on a gold CM5 chip *via* amino coupling reactions with *N*-hydroxysuccinimide and *N*-ethyl-*N'*-(dimethylaminopropyl)-carbodiimide at a flow rate of 5 $\mu\text{L min}^{-1}$ for 5 min at ambient temperature. The measurements were carried out in buffer solution (10 mM HEPES, 150 mM NaCl, and 5 mM CaCl_2) at pH = 7.4. The polymers were dis-

solved in the buffer, and their concentration varied between 10 and 0.63 μM . The regeneration was performed with the injection of a solution of 10 mM HEPES pH 7.4, 150 mM NaCl, 10 mM EDTA, and 0.01% Tween 20. The kinetic data of the bindings were determined from BIAevaluation 3.1 software. Z-average diameter and zeta potential of the polyplexes were analysed using a Zetasizer Nano ZS (Malvern Instruments, UK). Samples were equilibrated at room temperature and then diluted 1 : 100 in ultrapure H_2O , in a total of 850 μL . DTS1070 cuvettes were used, and the parameters as follows: material refractive index of 1.529, absorbance of 0.010, dispersant viscosity of 0.8872 cP, refractive index of 1.330 and dielectric constant of 79.

Step-growth polymerization *via* the aza-Michael reaction.

The step-growth polymers were synthesized following literature procedures. *N,N'*-Bis(acryloyl)cystamine (BAC) (200 mg, 0.77 mmol) and 3-amino-1-propanol (57.7 mg, 0.77 mmol) were weighed out in a microwave vial and dissolved in a solvent mixture of MeOH and water. Thereafter, TEA (15 μM , 0.11 mmol) was added dropwise to the mixture which was degassed for 15 minutes before the vial was placed in an oil bath at 45 or 80 °C. The reaction was allowed to go to completion and was typically stopped after 5–16 h by removal of the vial from the oil bath. Finally, the crude product was dialysed against water for 2 days and freeze-dried to obtain a polymer in the form of a white powder. In case of protonation, the polymers were firstly diluted in MeOH (20 mL) and protonated by adding a 1 M HCl solution until pH = 4 was achieved. Then, the polymers were dialyzed against acidic water (pH = 5) for 2 days.

Degradation of P(ABOL). P(ABOL) (50 mg, 0.01 mmol) was added to a microwave vial and dissolved in MeOH (0.2 mL). The reaction vial was purged with N_2 for ~ 15 min before the addition of a reducing agent. Thereafter, dithiothreitol (DTT) (7.7 mg, 0.05 mmol) was added to the vial containing P(ABOL). The reaction was stirred for approximately 2 h until the reaction mixture changed from yellow to colorless.

In vitro transcription of self-amplifying mRNA

Self-amplifying mRNA (saRNA) derived from VEEV alphavirus genome and encoding firefly luciferase (fLuc) was prepared by *in vitro* transcription. pDNA was linearised using MluI (New England BioLabs, UK) for 2 h at 37 °C, MluI was added again and incubated for another 1 h at 37 °C. Linearization was confirmed by agarose gel electrophoresis. For transcription into saRNA, 6 μl of linearised DNA template was synthesized into RNA transcripts *via* the mMessage Machine (Invitrogen, Thermo Fisher Scientific, UK) according to manufacturer's instructions. Transcripts were then purified by lithium chloride (LiCl) precipitation. Briefly, transcripts were frozen overnight at –20 °C and precipitated the next morning by centrifugation at 14 000 rpm for 20 min at 4 °C. Pellets were resuspended in 70% ethanol and centrifuged at 14 000 rpm for 5 min at 4 °C. The ethanol was removed, pellets were allowed to dry for 5 min, and transcripts were resuspended in ultrapure H_2O . RNA quantification was done using a NanoDrop One



(Thermo Fisher Scientific, UK) and RNA integrity was evaluated by RNA gel electrophoresis using a FlashGel™ System (Lonza, UK).

Formulation of polyplexes. Stock solutions of the step-growth polymers (**P1–P6**) at 2 mg mL⁻¹ were prepared in ultrapure H₂O. Polyplexes were prepared at different N/P ratios (1, 10, 20, 50, 100 and 200). The required amount of polymer at different N/P ratios was added to a fixed amount of RNA (20 µg). Polymers were added in a drop-wise manner to the RNA solution in either ultrapure H₂O or HEPES buffer 5% glucose (pH 7). Samples were mixed for 30 min at 500 rpm and at 20 °C, using a Thermomixer comfort (Eppendorf, Germany).

Cell line and culture conditions. HEK 293T/17 and HeLa cells (ATCC, US) were routinely grown in Dulbecco's modified Eagle's medium (DMEM) (Gibco, Thermo Fisher, UK) supplemented with 10% (v/v) fetal bovine serum (FBS), 1% (v/v) L-glutamine, and 1% (v/v) penicillin/streptomycin (Thermo Fisher, UK), at 37 °C under 5% CO₂. When confluent, cells were washed with DPBS 1X (Gibco, UK) and treated with trypsin (TrypLE Express 1X) (Gibco, UK) for seeding in new culture flasks (Corning, US).

THP-1 cells (ATCC, US) were routinely grown in RPMI-1640 Medium (Sigma, UK) supplemented with 10% (v/v) fetal bovine serum (FBS), 1% (v/v) L-glutamine, and 1% (v/v) penicillin/streptomycin (Thermo Fisher, UK), at 37 °C under 5% CO₂. When confluent, the whole cell suspension in culture media was centrifuged at 1750 rpm for 5 min, and the pellet was re-suspended in fresh RPMI-1640 medium for seeding in new culture flasks (Corning, US).

An immortalised cell line of human skeletal muscle cells (hSkMC) (PromoCell, UK) was routinely grown in Skeletal Muscle Cell Growth Medium (PromoCell, Germany), supplemented with SupplementMix (PromoCell, Germany). When confluent, cells were washed with DPBS 1X (Gibco, UK) and treated with trypsin. Neutralisation was done with DPBS 1X containing 10% FBS, and cells were centrifuged and re-suspended in the skeletal muscle cell growth medium for seeding in new culture flasks (Corning, US).

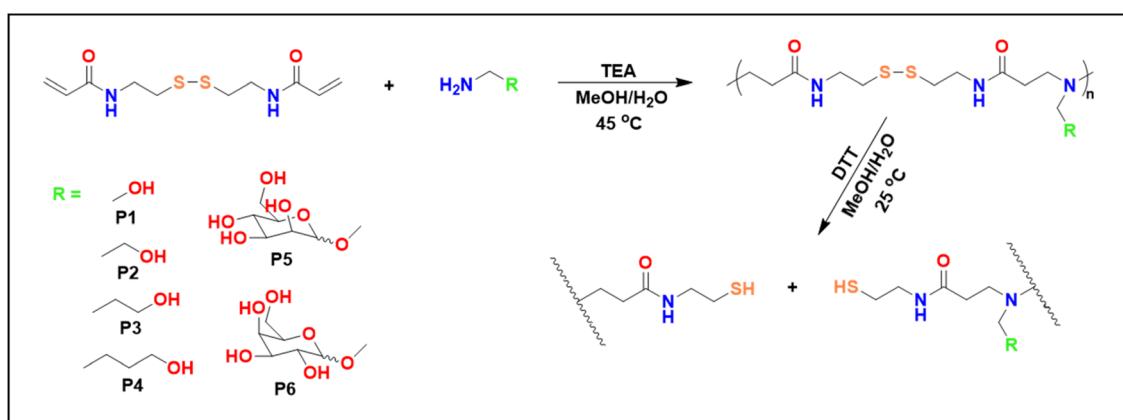
Cell transfection and luciferase assay. The cell experiments were carried out *via* the following procedure: Transfection assay was performed similar to as previously described by Blakney *et al.*,³ in which HEK 293T/17 and HeLa cells were seeded at 5 × 10⁴ cells per well in a 96-well plate 24 h prior to the experiment, THP-1 cells were seeded at 8 × 10⁴ cells per well, and the immortalised hSkMC, were seeded 10 × 10⁴ cells per well. On the day of the experiment, media was removed from the wells and replaced with 50 µL of transfection medium (DMEM with 1% (v/v) L-glutamine). Next, 100 ng of polyplexes in 100 µL of ultrapure H₂O or HEPES buffer 5% glucose was added to each well. Samples were allowed to transfect for 4 h. Media was then completely removed and 100 µL of supplemented DMEM per well was added. After 24 h of the initial transfection, the transfection efficiency was analysed by removing 50 µL of medium and adding 50 µL of ONE-Glo D-luciferin substrate (Promega, UK) to each well. The total volume was transferred to a white plate (Falcon®, US) and fluorescence intensity was analysed on a FLUOstar Omega plate reader (BMG LABTECH, UK).

Results and discussion

Synthesis of polymers *via* the aza-Michael reaction

Poly(amido amine)s have been synthesized *via* the aza-Michael addition reaction (Scheme 1), involving a series of primary amines which acted as Michael donors, and a disulfide containing bisacrylamide (BAC) which acted as a Michael acceptor (Table 1).

Firstly, all reactions were performed at 45 °C in a mixture of MeOH/H₂O (2 : 1) for about 16 h. It is worth noting, that even though a base is not required in the aza-Michael additions, triethylamine (TEA) was used to increase the rate of the reactions.¹⁷ Furthermore, a 1 : 1 molar ratio was aimed in all polymerizations. The reactions were monitored by ¹H NMR which generally showed full consumption of the vinyl groups (at 5.5–6.5 ppm) as well as a shift of the amide group from 8.4 to 8.2 ppm. The amines used in these studies were all aminoalco-



Scheme 1 Reaction scheme of synthesis of the polymers *via* the aza-Michael addition.



Table 1 Step-growth polymers prepared *via* the aza-Michael reaction using BAC and a series of different amines with different chain length. All polymers reached >99% conversion

Polymer	Amine	$M_{n, \text{GPC}}$ (Da)	$M_{w, \text{GPC}}$ (Da)	D
P1	Ethanolamine	3100	4500	1.4
P2	3-Amino-1-propanol	3900	5200	1.4
P3	4-Amino-1-butanol	4000	5400	1.4
P4	5-Amino-1-pentanol	5200	7100	1.4
P5	Mannosamine	16 000	46 000	3.2
P6	Galactosamine	12 000	20 000	2.4

hols with the spacer length between the primary amine and the hydroxyl group being the only difference between the amines (Scheme 1). Ethanolamine was the shortest aminoalcohol possessing two carbon atoms between the two functional groups. Conversely, 5-amino-1-propanol was the longest aminoalcohol with five carbon atoms between the amine and the hydroxyl groups. In all instances, the polymerizations were successful, and full consumption of the vinyl bonds (>99%) was observed in all reactions within 16 h. Furthermore, the formation of polymers was proven by the shift of the amide proton as well as the protons in the amines (Fig. 1B and Fig. S8†).

The polymers prepared using aminoalcohols were also analysed by DMF GPC analysis (Fig. 1A). The MWs of the polymers were found to be proportional to their amine chain length.

Thus, **P1** which had the shortest chain had an $M_{n, \text{GPC}} = 3100 \text{ g mol}^{-1}$, followed by **P2** with an $M_{n, \text{GPC}} = 3900 \text{ g mol}^{-1}$, **P3** with an $M_{n, \text{GPC}} = 4000 \text{ g mol}^{-1}$, and **P4** with an $M_{n, \text{GPC}} = 5200 \text{ g mol}^{-1}$ (Table 1). Although, **P1** resulted in the lowest $M_{n, \text{GPC}}$ value and **P4** provided the highest, the difference between all values was not significant. The small difference in the $M_{n, \text{GPC}}$ values between the polymers is likely to be due only to the different chain lengths of the amines rather than the polymer size. Additionally, the dispersities of all polymers with aminoalcohols (1.4) were found to be similar. Furthermore, as depicted in Fig. S10,† the sizes derived by the DLS measurements of the polymers with aminoalcohols are consistent with the M_n and M_w values derived from the GPC. Therefore, **P1** was found to be the smallest particle being 19.4 nm, followed by **P2** (34.1 nm), **P3** (36.3 nm) and **P4** (38.8).

As carbohydrates play key roles in biological recognition processes such as cell-cell adhesion, but also the attachment of viruses and bacteria, glycopolymers can be considered excellent candidates for targeted gene delivery for the treatment of various genetic diseases that are currently untreatable.^{18–21} Furthermore, carbohydrates are overexpressed on the surface of all cancer cells which means glycopolymers are an ideal platform for anticancer therapeutics.^{3,22} Therefore, sugar amines were synthesised from D-mannose and D-galactose and used to prepare glycopolymers (**P5** and **P6**) with BAC *via* the aza-Michael polyaddition (Scheme 1).

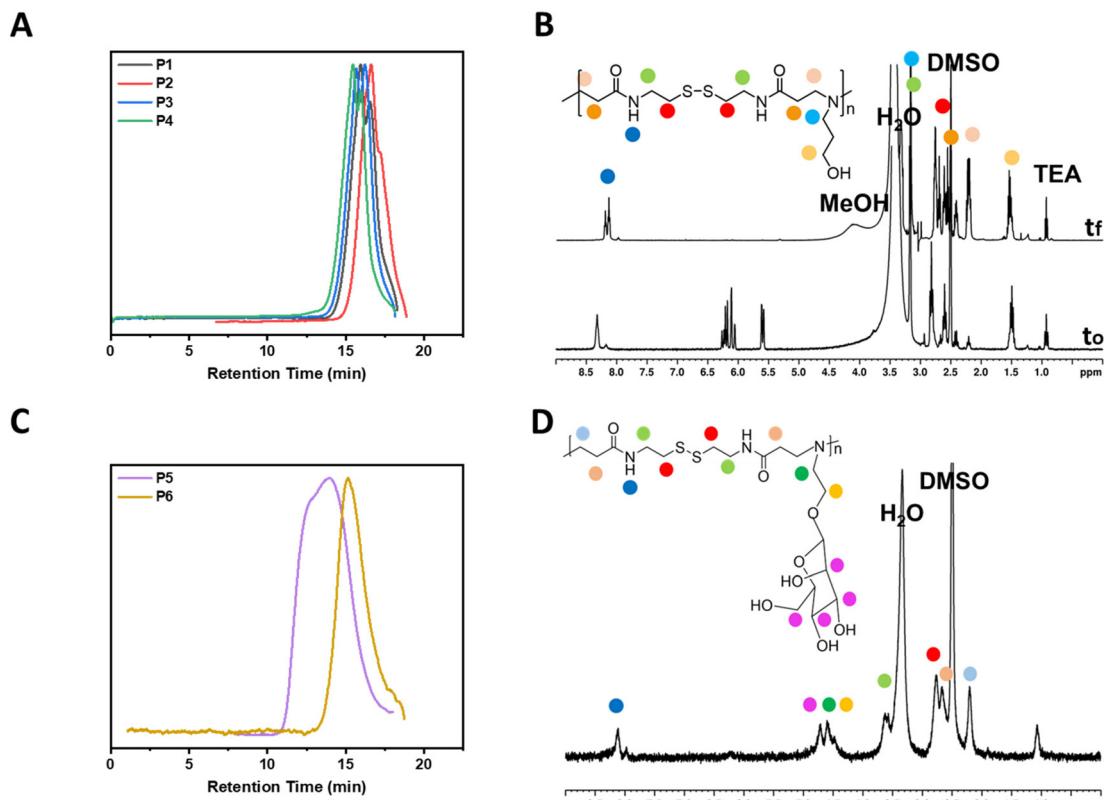


Fig. 1 DMF GPC traces of **P1**–**P4** (A), ¹H NMR spectra of **P2** (B), DMF GPC traces of **P5**–**P6** (C), and ¹H NMR spectra of **P5** (D).



The step growth polymerizations of *D*-mannosamine and *D*-galactosamine with BAC were carried out at 45 °C in a mixture of MeOH/H₂O and the reaction mixtures were stirred for 15 days. Considering the results obtained for **P2** using different reaction conditions, a slight excess of BAC (1.2 equiv.) was used for the polymerizations of **P5** and **P6** to achieve larger polymers. A lower reaction temperature (45 °C) was chosen, because sugar derived monomers are suspected to be more sensitive to higher temperatures for prolonged periods. Additionally, the polymerizations were monitored by GPC analysis (Fig. 1C) and stopped after 15 days.

Although the polymerizations of the sugar amines proceeded slowly, large glycopolymers, **P5** and **P6**, were successfully obtained. **P5**, which was prepared from *D*-mannosamine, was the largest glycopolymer having an $M_{n, GPC} = 16\,000$ g mol⁻¹. On the contrary, **P6**, prepared from *D*-galactosamine, was smaller than **P5** and had an $M_{n, GPC} = 12\,000$ g mol⁻¹. Additionally, the glycopolymers had larger polydispersity values (3.2 for **P5** and 2.4 for **P6**) than those shown previously for the other step growth polymers (**P1–P4**). However, this was expected since an increase in the polydispersity values is directly proportional to an increase in the MW of polymers prepared following this method.

Effect of solvent, temperature and stoichiometry on step-growth polymerization. Although the set-up of the Michael addition is extremely facile, it is not ideal for industrial scale-up since long reaction times are needed to obtain high MW

polymers.¹⁷ Thus, different reaction conditions for the polymerization of **P2** were investigated. In the ESI, Table S1† shows a summary of the results obtained from changing different variables including temperature, solvent, and stoichiometry.

The polymerizations of poly(amido amine)s are generally carried out below 50 °C.^{3,7,23,24} However, even though good results are obtained using these conditions, longer reaction times (>24 h) are normally required. To decrease the long polymerization times, the Michael addition of 3-amino-1-propanol and BAC was carried out at 80 and 100 °C and the monomer conversion was monitored by ¹H NMR and GPC analysis. During this investigation, it was observed that varying the temperature of the reaction had a massive influence in the polymerization rate but did not have an influence on any other aspect of the polymerization. Thus, the polymer size by GPC ($M_{n, GPC} = \sim 4000$ g mol⁻¹, $M_{w, GPC} = \sim 5000$ g mol⁻¹) remained constant and no real difference was observed between the reactions carried out at 45, 80 or 100 °C. The polymerization, however, proceeded significantly faster at 80 °C and was completed within 4 h, with 74% of the vinyl groups being consumed after only 1 h from the start of the reaction (Fig. 2A and B). The rate of the polymerization was further increased when the reaction was carried out at 100 °C, allowing a 98% reduction of the vinyl groups after 1 h from the beginning of the reaction and complete disappearance of the vinyl groups within 2 h (Fig. 2C).

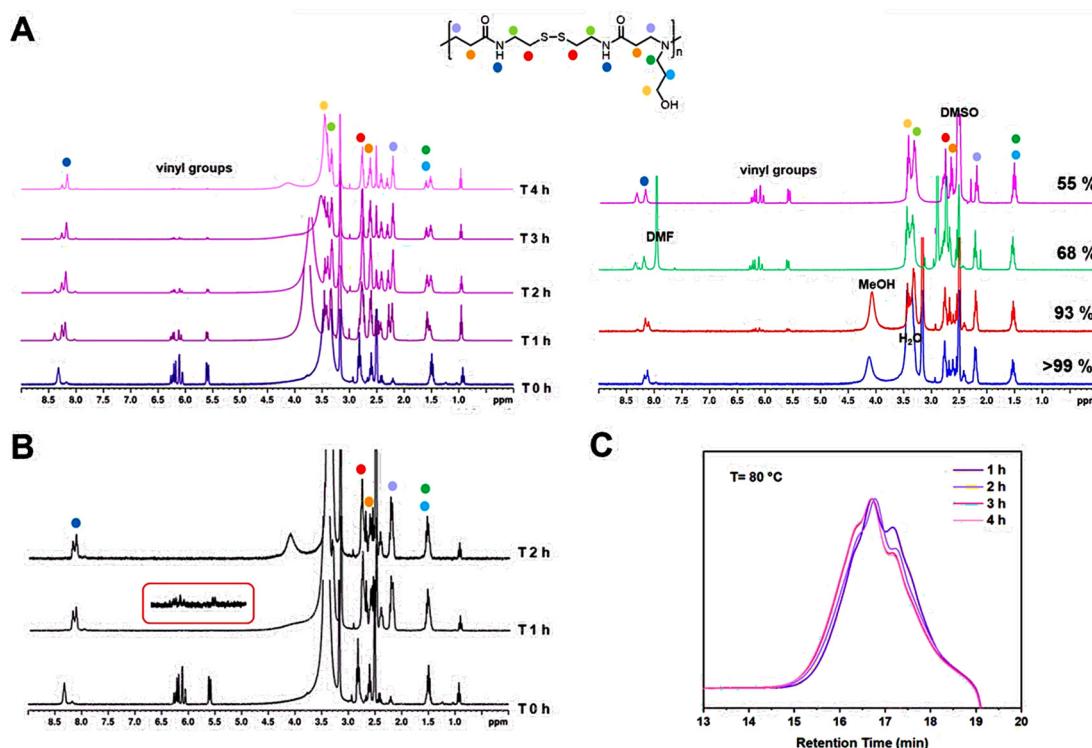


Fig. 2 ¹H NMR spectra showing different phases of the polymerization of **P2** at 80 °C (A, left), ¹H NMR traces of the polymerization of **P2** in different solvents after 16 h at 45 °C (A, right), DMF GPC traces of the polymerization of **P2** at 80 °C (B), and ¹H NMR spectra of the polymerization at 100 °C, which show nearly full conversion of the vinyl bonds after 1 h from the start of the reaction (C).



The next variable explored during the investigation was the effect of the solvent in the polymerization. The solvents selected for this part of the study were DMSO, DMF and MeOH and were compared to the original solvent mixture (MeOH/H₂O (2 : 1)). For the first part of this investigation, the reactions were carried out at 45 °C and were stopped after approximately 16 h from the start of the reaction. ¹H NMR samples were taken at the end of the 16 h to quantify the reduction of the vinyl groups (Fig. 2A, right). The original solvent mixture MeOH/H₂O afforded the best results and >99% reduction of the vinyl groups was observed after 16 h. Good results were also obtained for the polymerization in MeOH which led to a

reduction of 93% after 16 h. Conversely, the polymerizations in the aprotic solvents, DMF and DMSO, were less efficient and led to lower values of 68% and 55%, respectively.

The products obtained from the polymerizations in different solvents were also analysed by GPC (Fig. 3A). From the GPC traces, it appears that the reactions in DMF and in DMSO were only at partial conversion after the 16 h reaction time and the low molecular weight shoulder shows leftover BAC monomer, consistent with the NMR data. The $M_{w, GPC}$ values from the polymerizations in DMF and DMSO are 2300 and 1900 g mol⁻¹, respectively, and are significantly lower than that obtained in a mixture of MeOH/H₂O (2 : 1) ($M_{w, GPC}$ =

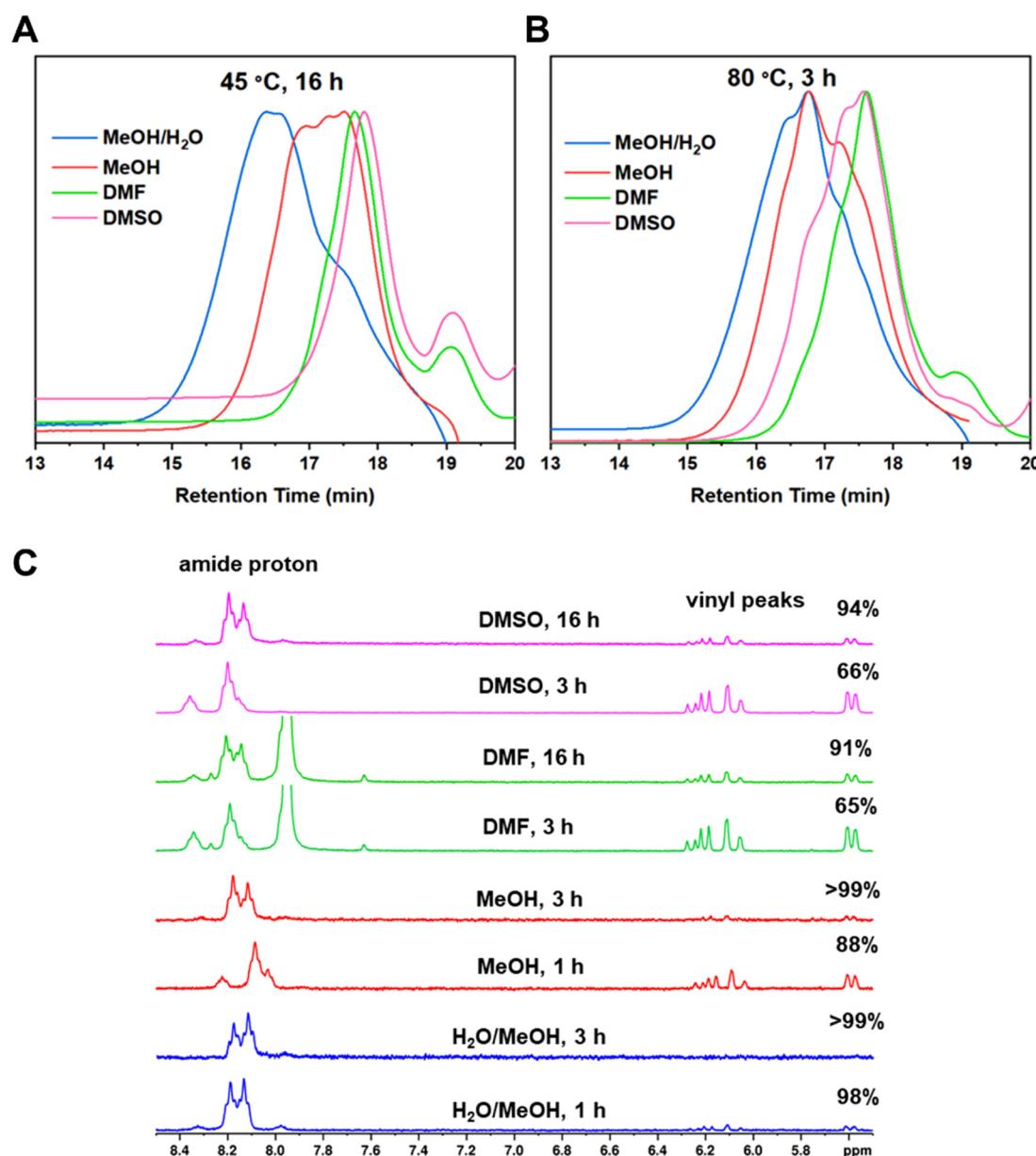


Fig. 3 DMF GPC traces of the polymerization of P2 in different solvents at 45 °C (A) and at 80 °C (B), ¹H NMR spectra of the area between 5–8.5 ppm of the Michael additions in different solvent at 80 °C show the consumption of the vinyl groups as well as the shift of the amide proton (C).



5200 g mol⁻¹). Even though monodisperse polymer peaks can be observed for low molecular weight species within step growth polymers, it is likely that the species formed were oligomers and not polymers, which supports the obtained $M_{w, GPC}$ values. On the contrary, the product from the polymerization in MeOH had an $M_{w, GPC} = 3400$ g mol⁻¹ and the GPC traces indicate that larger species have started to form. This is because more species of different sizes are present in later stages of a step growth polymerization, resulting in asymmetrical peaks.

To improve the polymerization in MeOH, DMF and DMSO, the temperature was increased to 80 °C. Again, the reactions were monitored by ¹H NMR (Fig. 3C) to determine the monomer consumption. It was found that even though the reaction in MeOH/H₂O was completed within 1 h, the reaction in pure MeOH went to full conversion after 3 h. As depicted in Fig. 3C, the full disappearance of the BAC vinyl groups and a complete shift of the amide proton from 8.2 to 8.1 ppm were observed after 3 h. It is worth noting that a small amide peak at 8.2 ppm is still visible in the ¹H NMR spectrum after 1 h from the start of the polymerization, when the monomer conversion was 96%. Additionally, the $M_{w, GPC}$ value (5000 g mol⁻¹) and GPC trace (Fig. 3B) obtained in this reaction were similar to those obtained when water was used in the polymerization ($M_{w, GPC} = 5100$ g mol⁻¹). Thus, the use of water in the reaction might not be necessary to achieve high conversions. However, water seems to accelerate the polymerization rate and its role as a potential catalyst in other Michael addition reactions has been previously reported.²⁵ This is because water molecules allow for hydrogen bond formation which makes the amine more nucleophilic and the carbonyl group more electrophilic.²⁶ However, because methanol is needed to solubilise the monomers, a combination of both solvents is ideal. The polymerizations in DMF and DMSO did not show a significant reduction of the vinyl groups at 80 °C after 3 h (Fig. 3C). Thus, the reactions were kept stirring for a total of 16 h. Increasing the temperature and the reaction time led to higher monomer conversions and $M_{w, GPC}$ values. The two step growth polymers obtained in DMF and DMSO had an $M_{w, GPC} = 3200$ and 3900 g mol⁻¹ with a monomer conversion of 91% and 94%, respectively.

Step-growth polymerization is a well-established process that is dependent on the concentration and stoichiometry of the reactants. This is because steric effects and incomplete reactions can limit the chain-chain addition steps, thereby affecting the polymerization process. It is possible to tune the molecular weight of polymers *via* adjusting the stoichiometry of the reactants. An equimolar ratio was used for the optimization reactions and only 2 reactions with a 20% excess of BAC were monitored at 45 °C for 16 and 48 hours. While the 16 hours reaction resulted in a 90% conversion, the obtained polymer was larger than those obtained at the equimolar ratio. The highest molecular weight in the synthesized polymers was obtained after 48 hours, with a dispersity of 1.2. The reason could be that using an excess of BAC may promote the formation of the polymer by shifting the equilibrium of the reaction towards the polymerization mechanism.

Biodegradation properties of poly(amido amine)s. Polymer degradation under physiological conditions is a crucial necessity for different types of biomedical applications in terms of biocompatibility, controlled drug/gene release and short-term accumulation since the body is capable of metabolising and excreting small molecules more efficiently than large molecules. This is known as the glomerular filtration barrier and is the process by which the kidneys filter blood to retain large proteins (6–7 nm) and excrete smaller particles.²⁷ Additionally, it is essential that whilst the polymers retain good extracellular stability during delivery, they should degrade at the desired targets.²⁸ Poly(amido amine)s prepared using BAC are a good example of degradable polymers due to the presence of the disulfide bond. The disulfide bond in the polymers undergoes rapid degradation in the presence of thiol-based reducing agents. Therefore, this class of polymers is likely to be reduced by the natural occurring glutathione (GSH).^{28–31} This approach is often employed to achieve intracellular drug delivery and can be considered a form of triggered release rather than targeted degradation. By incorporating specific linkages or chemical moieties that are responsive to the reducing environment, such as disulfide bonds, degradation can be triggered in the presence of high levels of GSH. Thus, the degradation of **P2** was carried out using dithiothreitol (DTT) (Scheme 1), an effective disulfide-reducing agent could work similarly to GSH.

The degradation was carried out at room temperature using an excess of DTT which allowed for a rapid reduction of the polymer disulfide groups. This resulted in polymer fragmentation into smaller molecules with thiol end groups and the oxidation of DTT to form a stable cyclic molecule.³² The reaction was monitored by ¹H NMR and GPC which showed degradation within 2 h from the start of the reaction. Fig. 4A shows a shift to the left of the amide proton around the 8.0–8.3 ppm area, but also a shift to the left of the protons on the polymer side chains in the area between 1.2–1.6 ppm, indicating successful degradation. Degradation was also observed in the GPC traces in Fig. 4B, which shows a massive shift towards the right (lower apparent molecular weight) representing the degraded polymer. The degraded product had an $M_{n, GPC} = 1900$ g mol⁻¹ and an $M_{w, GPC} = 2300$ g mol⁻¹ and thus, **P2** was disintegrated into a much smaller molecule. It is worth noting that the degradation was monitored for 3 days, but no further shifts in the GPC were observed after the initial 2 h sample.

Lectin binding properties of glycopolymers. The interaction ability of the synthesized glycopolymers were investigated using Surface plasmon resonance (SPR) through towards two human lectins, mannose-binding lectin (MBL) and C-type lectin domain family 10 member A (CLEC10A). According to the literature, these two lectins show specificity towards different sugar types. While MBL binds with a range of sugars including mannose, glucose, fucose and *N*-acetyl-mannosamine (ManNAc),³³ CLEC10A exhibits high specificity towards *N*-acetyl-galactosamine (GalNAc).³⁴ Both MBL and CLEC10A are responsible for a variety of defence mechanisms in the human body, and particularly, they have been demonstrated to be key components in the development of certain cancer types.



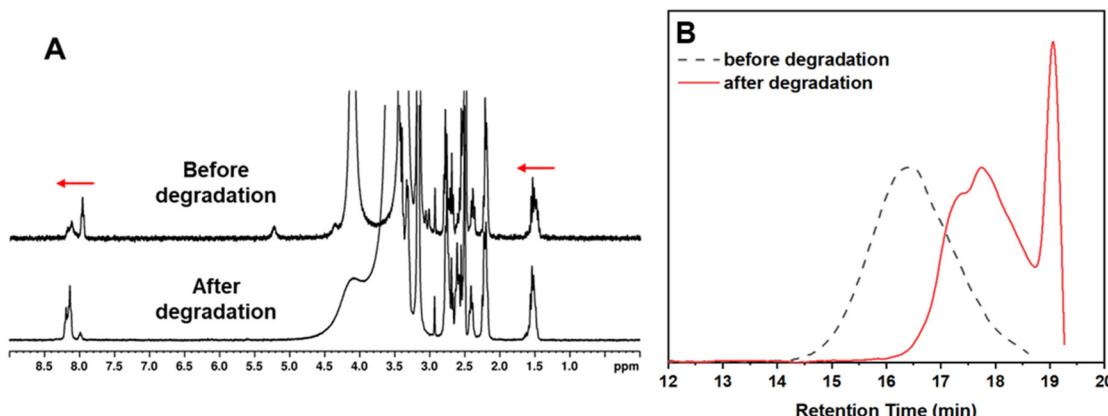


Fig. 4 ^1H NMR of the reaction mixture before and after degradation had occurred (A), DMF GPC traces of **P2** before and after degradation (B).

For example, various studies have demonstrated the over-expression of MBL in patients suffering from colorectal and ovarian cancers.^{35–37} Additionally, recent research on breast cancer led to the discovery that CLEC10A binds to GalNAc residues found in breast cancer tissue.^{38,39} Therefore, it seems obvious that incorporating sugars in nucleic acid vectors would be advantageous for delivery for the treatment of specific cancer types.

The different concentrations of **P5** and **P6** between 20 μM and 1.25 μM were measured to determine the binding effectiveness between the glycopolymers and the lectins (Fig. 5). As expected, MBL only showed specificity towards **P5** at all concentrations used, whereas it did not show any binding with **P6** at its highest concentration (20 μM). Conversely, CLEC10A showed interactions with **P6** at all concentrations, but no interactions with **P5** at 20 μM were observed. Generally, **P5** and **P6** showed high affinity towards MBL and CLEC10A, respectively, and bound rapidly with the lectins. Moreover, the binding of **P5** with MBL occurred in a more gradual manner compared to that of **P6** with CLEC10A. For this reason, the SPR curves

obtained from the binding of **P6** with CLEC10A are sharper than those obtained from the measurements of **P5** with MBL. As shown in Table S2,[†] this is also confirmed by the association constant, k_a , which is significantly higher for **P6** (3.05×10^3) than for **P5** (122). Although it is important to clarify that in both cases, strong interactions with the lectins were obtained and that the binding showed saturation within 5 minutes from the injection of the polymers. However, the dissociation constant (k_d) was higher for **P6** (1.51×10^{-3}) than for **P5** (6.79×10^{-5}), indicating that not all binding between **P6** and CLEC10A was retained with the injection of buffer. This is also visible in Fig. 5B, where the SPR curves appear to decline rapidly at 250 s which is the point in which buffer was injected. Nevertheless, interactions between **P6** and CLEC10A are still existent after that point, despite the decline.

Complexation of saRNA with poly(amido amine)s. Self-amplifying RNA (saRNA) used in this work for polyplex formation and transfection studies derives from the Venezuelan Equine Encephalitis virus (VEEV) alphavirus genome and encodes firefly luciferase (fLuc). saRNA is an unique vaccine

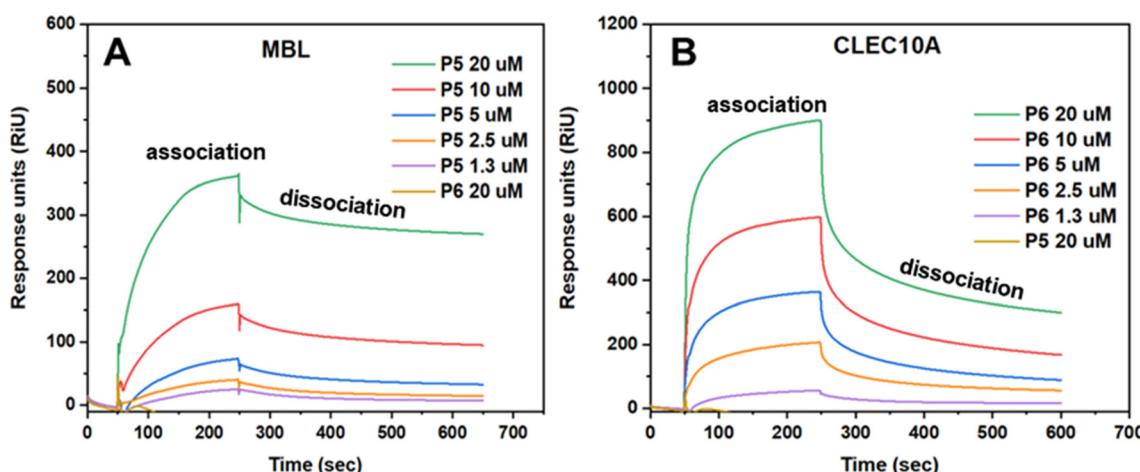


Fig. 5 SPR measurements of **P6** with MBL (A) and **P5** with CLEC10A (B).



model because it can self-replicate once in the cytoplasm. This means that higher protein expression can be achieved compared to traditional mRNA vaccines.³ The polyplexes were formed by direct mixing of different positively charged polymer amine groups to negatively-charged nucleic acid phosphate groups (N/P) ratios and a fixed amount of saRNA (20 μ g) in ultrapure water (Fig. 6A).

While the size of the polyplexes ranged between 200 and 300 nm, the polyplexes formed at N/P = 1 were around 600 nm for **P2**, **P3** and **P4**. The reason could be that higher quantities of negative charges result in greater electrostatic repulsion which affects the size of the polyplexes. This was also confirmed by the zeta potential measurements (Fig. 6B) which showed that the N/P ratios of 1–50 resulted in more negatively charged polyplexes, indicating that more positively charged polymer was needed. For this reason, the N/P = 100–200 were more suitable for the formulations of polyplexes using **P1–P4**. Cell transfection studies of the polyplexes at all N/P ratios were performed in HEK 293/T17 cells to investigate their potential in gene delivery (Fig. 7). The results obtained from the cell transfection studies are comparable to those reported in the literature.¹⁷ The polyplexes made from **P1–P4** showed

similar transfection efficacy implying that different side chain lengths do not have a significant impact on cell transfection activity.

Further studies with different cell lines were performed using **P1–P4** showing a variety of results. The polyplexes were tested for transfection activities in HeLa cells, THP-1 cells and human skeletal muscle cells (Fig. 7B). Whilst the results with the HeLa cells were similar to those obtained with HEK293 t/17 cells, the polyplexes showed higher levels of transfection in THP-1 cells, but a decrease in transfection activity in human skeletal muscle cells.

Thereafter, polyplexes of the glycopolymers **P5** and **P6** with saRNA were also prepared. The size and zeta potential of polyplexes prepared from **P5** and **P6** at N/P = 100 are similar to those obtained from **P1–P4** at this concentration and are shown in Fig. 8A and B. Nonetheless, the cell transfection results obtained with **P5** and **P6** showed that they have greater transfection efficacy than **P1–P4** (Fig. 8C). This might be due to a much greater number of hydroxyl groups present in **P5** and **P6** which may help with the formation of hydrogen bonding on the cell surface for better uptake of polyplexes of **P5** and **P6**. In addition, the $M_{w, GPC}$ of **P5** and **P6** is significantly higher

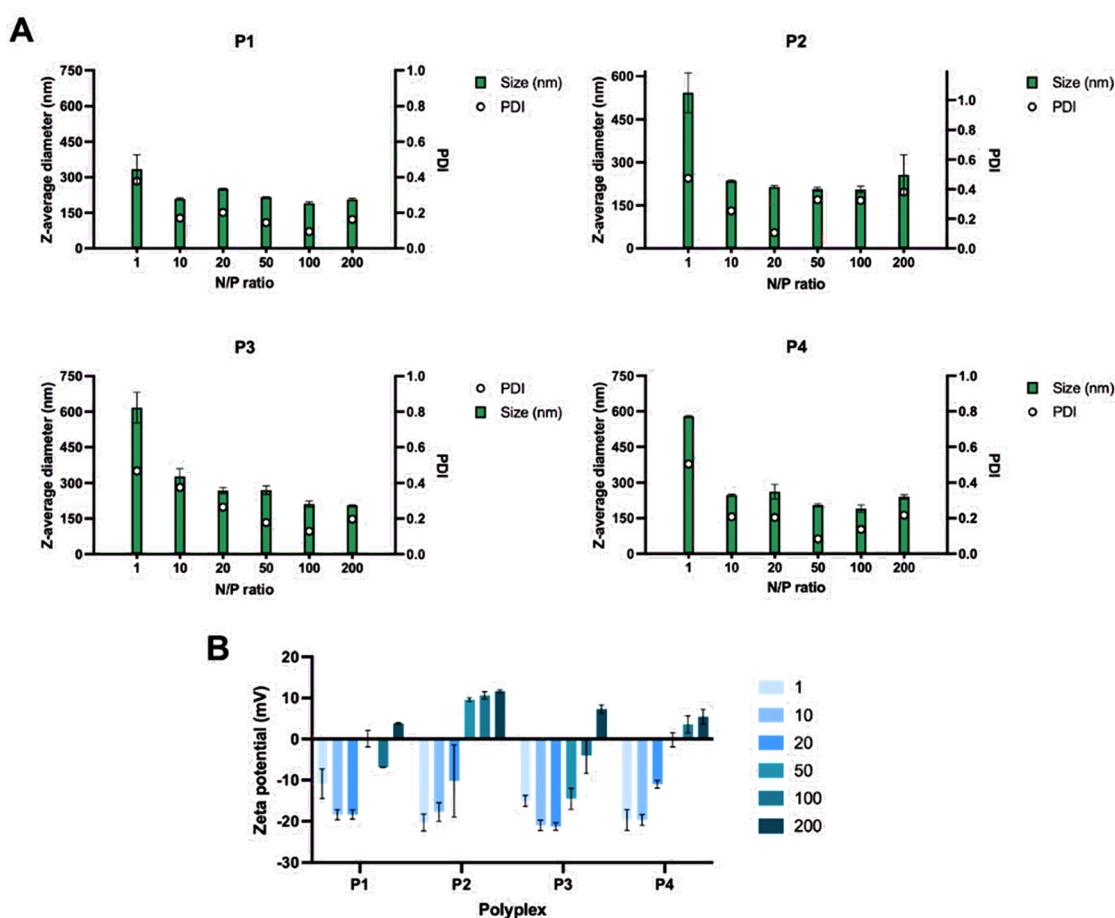


Fig. 6 Size of the polyplexes formed using different N/P ratios of **P1–P4** (A) and zeta potential measurements of the polyplexes at different N/P ratios (B).



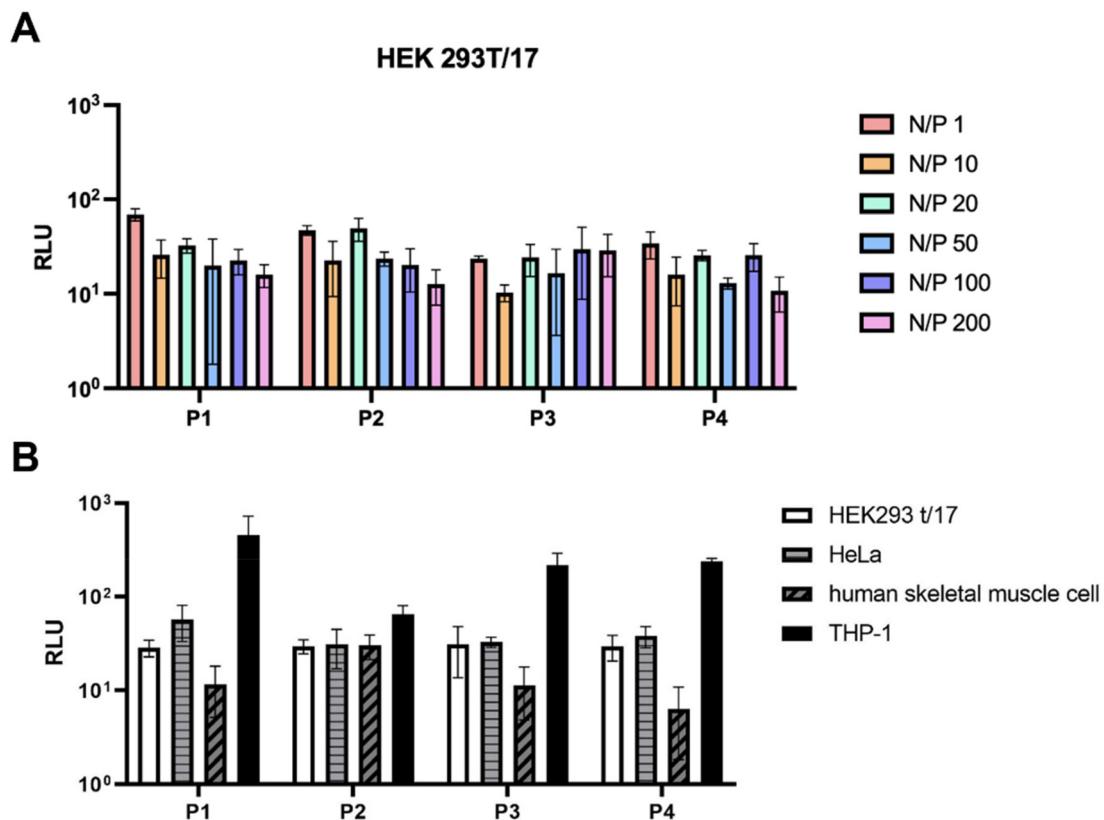


Fig. 7 Cell transfection efficiency in HEK 293T/17 cells of the polyplexes prepared with fLuc-saRNA and P1–P4 at different N/P ratios (A), cell transfection efficiency in different cell lines at N/P = 100 (B). RLU: relative luminescence unit.

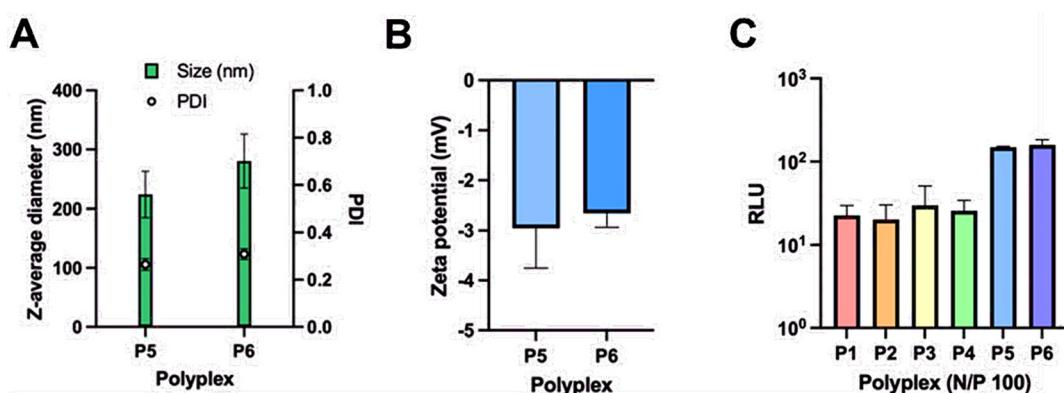


Fig. 8 DLS measurements showing the size of the polyplexes formed from P5 and P6 with fLuc-saRNA (A), zeta potential measurements of the P5 and P6 polyplexes (B) and cell transfection results in HEK 293/T17 cells obtained from all polymers (P1–P6) at N/P = 100 (C). RLU: relative luminescence unit.

than that obtained from the other polymers, which might enhance the encapsulation efficiency of heavy genes like saRNA (~9500 nt).³ Finally, the HEK 293/T17 cells did not show any specificity between P5 and P6 which performed equally well in these studies, despite containing different sugars. However, more studies using multiple cells would need to be carried out to determine any specificity achieved from using different sugars.

Conclusions

The step growth polymerization *via* the Michael polyaddition of different aminoalcohols and a reducible monomer, BAC, was succeeded in this work. Firstly, different aminoalcohols in terms of chain length were used to produce reducible step growth polymers. The polymerization reactions were monitored by ¹H NMR and the obtained polymers were character-

ized by DMF GPC. Secondly, different reaction conditions by changing temperature, solvent and stoichiometry have been studied to understand and improve the Michael addition process more. It was found that a temperature increase accelerates the reaction rate drastically, reducing the time of the polymerization from 16 h to 2–4 h. Furthermore, even though the reaction was demonstrated to work more efficiently in protic solvents, the presence of water accelerates the polymerization rate. Conversely, the reaction in aprotic solvents such as DMF and DMSO was significantly slower than protic solvents. Varying the stoichiometry by adding a slight excess of BAC allowed the formation of larger polymers. Additionally, the obtained polymers were degraded by reduction of the disulfide bonds using DTT. A very rapid degradation was observed, which is a highly desirable characteristic of polymeric gene carriers.

Furthermore, mannose and galactose amines were synthesized to prepare glycopolymers *via* the aza-Michael polyaddition. The incorporation of sugar is desirable for targeted gene delivery which could help the treatment of specific types of cancer. The high MW biodegradable glycopolymers were synthesized successfully and then their binding affinities towards two human lectins have been investigated.

Finally, the step growth polymers were used to prepare polyplexes with saRNA which is a promising vaccine platform. The cell transfection studies of the polyplexes showed that while there is not a considerable effect of different aminoalcohols as a side chain, glycopolymers exhibited slightly higher transfection levels than aminoalcohol based polymers in HEK 293/T17 cells. Although, no difference was obtained between glycopolymers due to sugar type in terms of selective uptake, the results demonstrated that the addition of these molecules to polyplexes may lead to more efficient gene delivery systems and warrants further research.

Overall, biodegradable poly(amido amine)s with different sugars and alcohols with high molecular weights and molecular weight distributions have been synthesized to develop saRNA delivery system. The polymerization conditions were improved, and it can be easily scaled up with a small batch-batch variability.

Conflicts of interest

There are no conflicts to declare.

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