

Cite this: *Polym. Chem.*, 2023, **14**, 1241PDMAEMA from α to ω chain ends: tools for elucidating the structure of poly(2-(dimethylamino)ethyl methacrylate)[†]Maria Rosella Telaretti Leggieri,^a Tahani Kaldéus,^{a,b} Mats Johansson^{a,b} and Eva Malmström^{a,b}

Poly(2-(dimethylamino)ethyl methacrylate) (PDMAEMA) is currently used for a wide range of applications, often involving the synthesis of block copolymers. Here, an in-depth characterization of PDMAEMA prepared by atom transfer radical polymerization (ATRP) is reported, with a focus on end group analysis. The structure of the polymer was elucidated by one- and two-dimensional NMR spectroscopy, which assessed the presence of deactivated chains and allowed for a quantification of their fraction. Detailed characterization by MALDI-TOF MS further provided insightful information about the chain end fidelity. On this basis, termination by disproportionation was found to be the main mechanism for the loss of active chain ends. The detailed characterization allowed for an estimation of the preserved chain end functionality (CEF) of PDMAEMA. Additionally, a chain extension experiment was conducted, using PDMAEMA as a macroinitiator for the polymerization of methyl methacrylate (MMA) by ATRP. The results of chain extension supported the estimation of CEF based on the data provided by NMR and MS. Although assessing the degree of polymerization of a block copolymer proves challenging when the amount of the initial block able to act as a macroinitiator is not known *a priori*, an accurate estimation of the DP and M_n of the obtained block copolymer was possible by total nitrogen analysis. The tools here provided for the characterization of PDMAEMA and its block copolymer architectures allow the obtainment of essential information about the extent of control over the homo- and copolymerization. Therefore, they are of high importance when well-defined structures are aimed for.

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Introduction

PDMAEMA is a polymethacrylate with attractive properties, the first being its solubility in aqueous media.¹ Furthermore, due to the presence of dimethylamino pendant groups in its polymer chain (Chart 1), PDMAEMA is both pH- and temperature-responsive in water. At pH \approx 8, the dimethylamino groups are partially protonated and the polymer displays a lower critical solution temperature (LCST) ranging between 32 and 57 °C. At pH \approx 10, the contracted state of completely non-ionized chains results in a decrease of the LCST to as low as 25 °C. At pH \approx 4, on the other hand, the polymer is fully proto-

nated and not thermoresponsive.^{1–3} The exact values of pH and LCST are dependent on the molecular weight. Additionally, the introduction of permanent charges by methylation of the amines can systematically increase the LCST.^{4,5} Fully quaternized PDMAEMA is a strong polyelectrolyte whose solubility in water is no longer affected by temperature or pH.

These tunable properties arising from the amine functionality, combined with PDMAEMA's biocompatibility, have opened up routes for applications in the biomedical field such as drug delivery with stimuli-driven controlled release^{6–8} and

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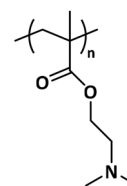


Chart 1 Repeating unit of poly(2-(dimethylamino)ethyl methacrylate) (PDMAEMA).



gene therapy, owing to its efficient DNA complexation.^{9,10} Moreover, the polymer is investigated for a wide spectrum of additional advanced applications such as water-dispersed nanolatexes,^{11,12} electrostatic adsorption and surface modification of negatively charged substrates,^{13,14} antimicrobial materials,^{15,16} biosensor and bioseparation systems based on stimuli-responsive grafted surfaces,^{17,18} water nanofiltration membranes,^{19,20} and stabilization of Pickering emulsions.^{21,22} The majority of these applications require a tailored design and a well-defined structure.

PDMAEMA as a homopolymer with narrow molecular weight distribution has been synthesized primarily by atom transfer radical polymerization (ATRP)^{23–25} and reversible addition–fragmentation chain-transfer (RAFT).^{26–28} The development of these reversible deactivation radical polymerization (RDRP) techniques has also led to the preparation of well-defined copolymers of PDMAEMA with various architectures, ranging from random, gradient, block and star copolymers^{29–31} to macromolecular brushes.^{27,32,33} Among these structures, amphiphilic block copolymers made of a hydrophilic PDMAEMA block and a hydrophobic block—often poly(methyl methacrylate) (PMMA), poly(butyl methacrylate) or polystyrene—have gained significant attention. Their most distinctive characteristic is the ability to spontaneously self-assemble in water, which has been extensively studied.^{34–36} PDMAEMA is an attractive choice of a hydrophilic corona in these micellar systems, not only promoting “smart” properties related to the response to varying pH values and temperatures,^{37–39} but also effective adsorption onto hydrophilic substrates.^{40–42}

Well-defined PDMAEMA has an advantageous structure in the context of a wide variety of research fields over the past three decades. However, its synthesis entails specific challenges which are not always addressed in application-driven studies: (1) the amines in DMAEMA units may be involved in unwanted side reactions. In the context of RDRP techniques involving the use of metal catalysts, the pendant amine groups can compete with the selected ligands (usually amines) in metal complexation. Consequently, achieving good control over the polymerization is challenging and reducing the amount of the metal catalyst employed is hardly feasible, although it has been explored with tailored methods.^{24,43} Moreover, the amines may participate in nucleophilic substitutions and redox reactions. (2) Thermally induced self-initiated free radical polymerization of DMAEMA is known to take place, although with a low rate of initiation, as first elucidated by Shalati *et al.*⁴⁴ (3) DMAEMA readily undergoes hydrolysis when polymerized in the presence of water, generating methacrylic acid which is then incorporated into the polymer chain, to an extent that can be limited by controlling the pH and quantified.⁴⁵ PDMAEMA, on the other hand, is not sensitive to hydrolysis,⁴⁶ unlike its acrylate counterpart poly(2-(dimethylamino)ethyl acrylate).^{47,48} (4) When synthesized in polar protic media, PDMAEMA's response to pH and temperature must be taken into account.

In studies where well-defined architectures are required, a thorough characterization of PDMAEMA and its copolymers is

needed. However, there is a lack of systematic knowledge on how to elucidate PDMAEMA's structure with commonly available techniques. To the authors' knowledge, during the past few decades only a limited number of studies have dwelled on PDMAEMA's detailed characterization by nuclear magnetic resonance (NMR) spectroscopy and mass spectrometry techniques,^{26,49–51} which can provide useful information about the degree of polymerization, molecular weight distribution and chain end fidelity. In the context of mass spectrometry, both matrix-assisted laser desorption ionization time-of-flight mass spectrometry (MALDI-TOF MS) and electrospray ionization mass spectrometry (ESI-MS) are widely used for the structural investigation of synthetic polymers, with different advantages and limitations,^{52,53} and the choice of one over the other is usually a matter of availability.⁵⁴ However, neither of them has been intensely used and thus optimized for PDMAEMA. Size-exclusion chromatography (SEC), on the other hand, is often used for estimating the molecular weight of PDMAEMA, although it does not give highly representative results in terms of molecular weight,^{29,31} as discussed later.

In this work, the aim is to provide adequate tools for the characterization of PDMAEMA, with an in-depth investigation of its structure by one- and two-dimensional NMR spectroscopy and MALDI-TOF MS. The objective is not to optimize experimental parameters for the synthesis, but to highlight important information that can be deduced from widely available analysis techniques. With this, we aim to further strengthen the potential of PDMAEMA and its copolymer architectures.

For this purpose, a commonly used method for the RDRP of DMAEMA was applied to prepare a model PDMAEMA that was extensively characterized: a typical protocol for ATRP mediated by CuBr, with a tetradentate ligand, in acetone. Similar protocols are widely used for the synthesis of PDMAEMA in view of various applications,^{3,32,49,55} in some cases targeting its chain extension to yield block copolymers.^{30,42,56} Especially for the latter goal, as well as for the purpose of chain end functionalization post-polymerization,⁵⁷ having information about the chain end fidelity is of primary importance. Traditional ATRP is most commonly mediated by CuBr. Copper bromide salts are preferred over copper chloride salts since alkyl bromides are more active than the corresponding alkyl chlorides.⁵⁸ However, bromine is considered a rather labile end group due to the low dissociation energy of the C–Br bond and its loss has been observed during polymerization at medium to high conversions.^{59,60}

Furthermore, samples of PDMAEMA were prepared by traditional ATRP mediated by CuCl (PDMAEMA_{Cl}), as well as with a very different ATRP system mediated by a photoredox catalyst (PDMAEMA_{UV,EBiB} and PDMAEMA_{UV,EBPA}), to provide insights into the different impacts of side reactions when varying the parameters of the ATRP system.

In this study, characterization aspects providing important information about the control and livingness of the polymerization are highlighted. The investigated characterization tools allow the obtainment of essential information about the pres-



ervation of chain end functionality (CEF) *i.e.* the fraction of chains bearing the active ω -end group which enables controlled propagation. CEF is often considered the most important parameter for assessing the livingness of ATRP systems.^{61,62} Additionally, a chain extension experiment was conducted to add evidence with respect to the CEF of the synthesized PDMAEMA.

Experimental

DMAEMA was polymerized by CuBr/HMTETA-mediated ATRP in acetone at 30 °C, using ethyl α -bromoisobutyrate (EBiB) as the initiator. The polymerization was conducted with the following molar ratios of monomer, initiator, catalyst, and ligand: $[M]/[I]/[Cu(I)]/[L] = 65/1/1/2$, in the absence of external stimuli. PDMAEMA was obtained at 52% conversion ($t = 135$ min); unless stated otherwise, the results of the characterization of this PDMAEMA sample were discussed. During the synthesis, purification and storage of PDMAEMA, heating was always avoided aiming to lower the risk of side reactions.

In a second step, PDMAEMA was chain-extended with PMMA, producing a PDMAEMA-*b*-PMMA block copolymer. The polymerization of MMA was carried out by CuCl/HMTETA-mediated ATRP in acetone at 50 °C, with the following molar ratios of monomer, initiator, catalyst, and ligand: $[M]/[I]/[Cu(I)]/[L] = 400/1/1/2$. PDMAEMA-*b*-PMMA was obtained at 55% conversion ($t = 280$ minutes); unless stated otherwise, the results of the characterization of this PDMAEMA-*b*-PMMA sample were discussed.

The reaction steps are visualized in Scheme 1. Further experimental details, as well as the synthetic procedures for preparing PDMAEMA_{Cl}, PDMAEMA_{UV,EBiB} and PDMAEMA_{UV,EBPA}, are reported in the ESI.†

Results and discussion

DMAEMA polymerization kinetics

PDMAEMA was synthesized by applying a commonly used ATRP protocol employing a CuBr/HMTETA catalyst complex. The reaction kinetics was monitored by ¹H-NMR and the polymerization was stopped at approximately 50% conversion, aiming for a limited extent of termination and for the preservation of the active chain end functionalities.⁶⁰ The latter is

specifically important to enable chain extension, from the perspective of designing well-defined polymer architectures for materials science applications. The ¹H-NMR of the polymerization mixture at final conversion is reported in Fig. S1.†

The polymerization kinetics is presented in Fig. 1A. The values of $\ln([M]_0/[M])$ were found to increase rather linearly with time after what could appear to be an induction period affecting the initial polymerization rate. A linear behavior of $\ln([M]_0/[M])$ is commonly considered a sign of good control over the reaction, indicating a stable concentration of active species. However, the slightly curved trend observed in the kinetics plot is likely to be the result of slow initiation, which is observed in the polymerization of methacrylates in traditional ATRP systems.^{63,64} The kinetics plot gives no direct indication of the chain end fidelity. Further investigations are essential to gain information about the fraction of chain end functionality (CEF) preserved when the polymerization is stopped.

The deviation from the linear behavior of the plot of number average molecular weights determined by SEC ($M_{n,SEC}$) as a function of conversion (Fig. 1B) can be correlated with slow initiation or side reactions that are hampering the control. Nonetheless, the evolution of $M_{n,SEC}$ is not regarded as highly sensitive to termination reactions.⁶⁵

An overlay of SEC traces revealing the evolution of molecular weight distributions (MWDs) with increasing monomer conversion is reported in Fig. S2.† To a certain extent, low molecular weight tailing was observed which may indicate loss of active chain ends in the course of the polymerization. The dispersity (D) values obtained, around 1.2 (Fig. 1B), are consistent with other reports of ATRP of DMAEMA at low

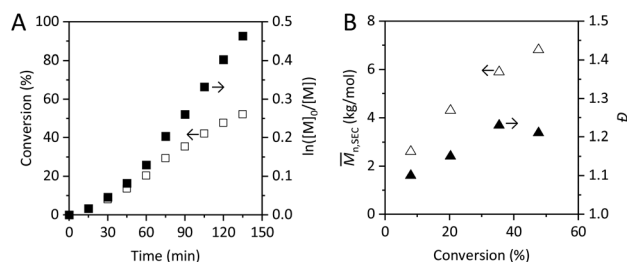
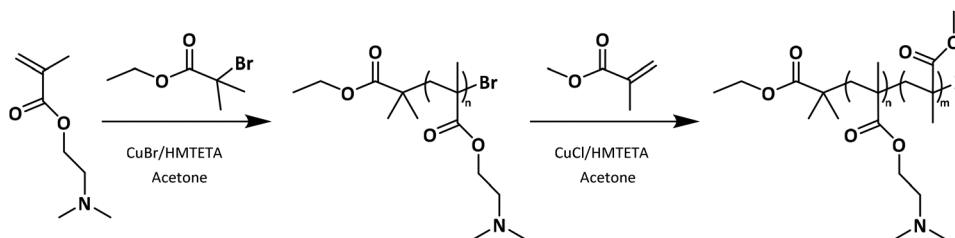


Fig. 1 (A) Kinetic plot of $\ln([M]_0/[M])$ (■) and conversion (□) over time for the ATRP of DMAEMA. (B) $M_{n,SEC}$ (△) and dispersity (▲) of PDMAEMA determined by SEC in DMF as a function of degree of conversion.



Scheme 1 Synthesis of PDMAEMA and subsequent chain extension with MMA. X = Br or Cl.



temperatures.^{24,25} This D , rather high in the context of RDRP systems, indicates the presence of side reactions; it is usually higher for DMAEMA polymerizations performed at higher temperatures.²³ However, SEC results are to be interpreted carefully, as discussed later.

From the final monomer conversion, the theoretical degree of polymerization of PDMAEMA was calculated as $DP_{\text{theo}} = 34$, resulting in an average molecular weight $M_{n,\text{theo}} = 5500 \text{ g mol}^{-1}$. This value was later confirmed by the end group analysis of PDMAEMA conducted by NMR after purification.

1D and 2D NMR analysis of PDMAEMA

A comprehensive and detailed interpretation of the NMR signals of PDMAEMA prepared by RDRP was not found in the literature. In fact, wrong interpretations have been reported because of this lack of accessible information. Here, an extensive investigation of the structure of the purified polymer was carried out by means of NMR spectroscopy, which allowed the determination of the experimental DP_{NMR} from end group analysis and estimation of CEF.

For the purpose of conducting an in-depth analysis of NMR spectra, acetone- d_6 was preferred over CDCl_3 and D_2O , as it provided the highest peak resolution while allowing for high concentrations of PDMAEMA. The signals in the ^1H -NMR spectrum (Fig. 2) and in the ^{13}C -NMR spectrum (Fig. S3†) were identified with the aid of two-dimensional NMR spectroscopy—correlation spectroscopy (COSY), heteronuclear single quantum coherence (HSQC) and heteronuclear multiple bond correlation (HMBC)—and distortionless enhancement by polarization transfer (DEPT). The resulting spectra, together with the respective assignments, are reported in Fig. S4–S9.†

In the ^1H -NMR spectrum of PDMAEMA initiated by EBiB, the signals arising from the α chain end (H_i , H_j , $\text{H}_{\text{I}(1)}$ and $\text{H}_{\text{I}(2)}$) were identified, providing useful information. The normalized integral ratio between the broad triplet arising from protons H_i and polymer signals, as shown in Fig. S10,† allowed the calculation of $DP_{\text{NMR}} = 34$ and consequently $M_{n,\text{NMR}} = 5500 \text{ g mol}^{-1}$. This value, coinciding with the calculated $M_{n,\text{theo}}$, is an indication of overall good control over the polymerization; possible side reactions have not led to a deviation from the targeted DP. The occurrence of self-initiation (which has been shown to

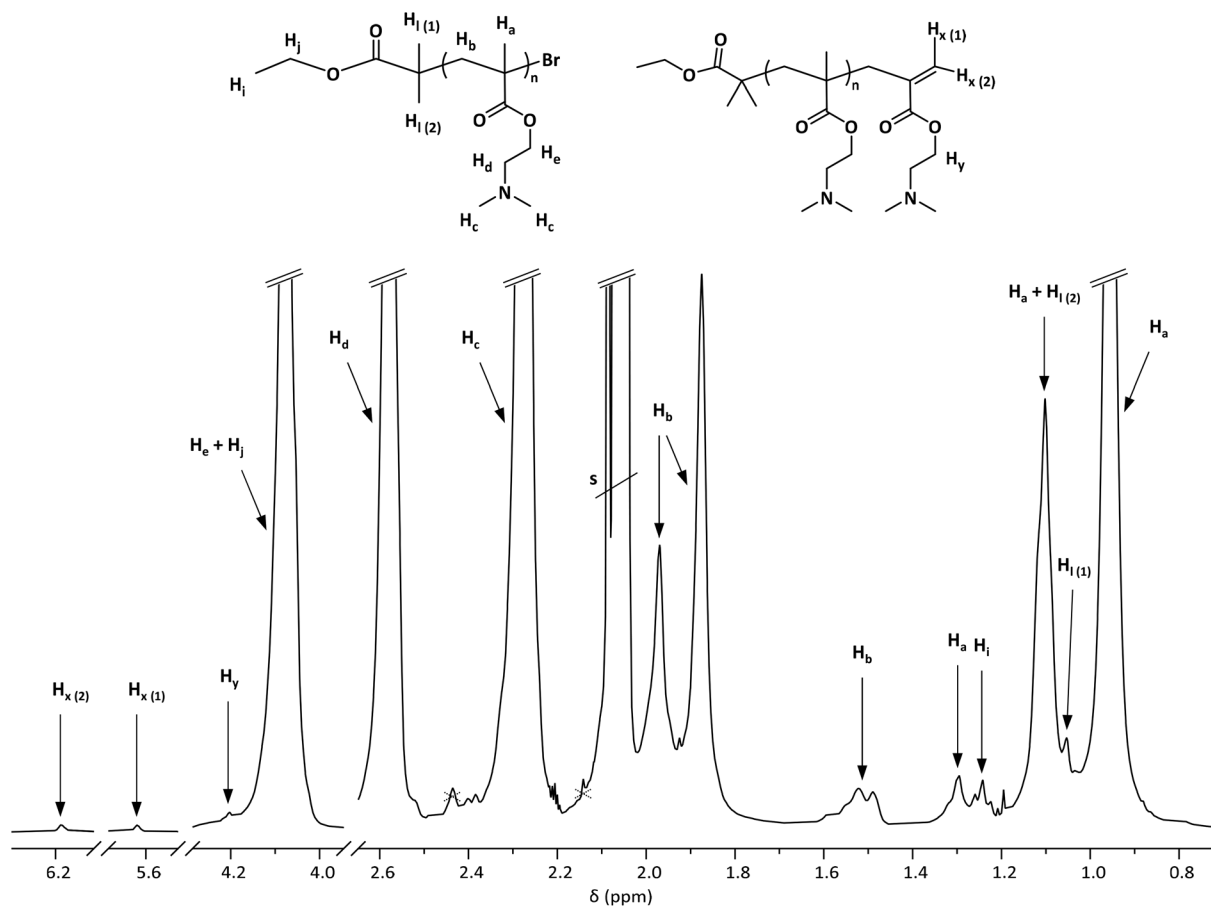


Fig. 2 Enlargement of the ^1H -NMR of PDMAEMA in acetone- d_6 . The full spectrum is reported in Fig. S11,† and the assignments are supported by 2D NMR spectra (Fig. S4–S9†). An “s” notation is used to mark the multiplet arising from acetone- d_6 (2.05 ppm) along with the singlet arising from residual acetone (2.09 ppm). The signal at 2.44 ppm, marked with a dotted cross, is a satellite of the intense H_c signal, as it is evident in the HMBC spectrum (Fig. S8†); the second satellite overlaps with the solvent signal.



take place at higher polymerization temperature than that selected in this study⁴⁴) would affect this calculation; however, the absence of self-initiation was confirmed by MALDI TOF-MS analysis.

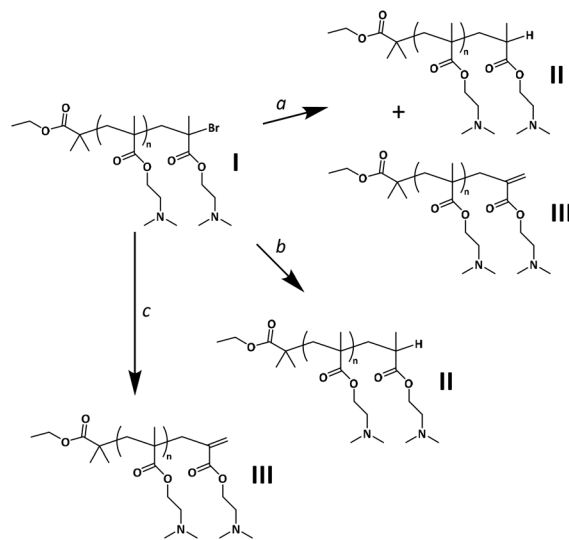
The interpretation of the region of the ¹H-NMR spectrum at a low chemical shift is often neglected when spectra of PDMAEMA are reported. However, since the signals of the α chain end are commonly found in this region, it is important to report a comprehensive analysis. The methyl and methylene protons in the backbone of PDMAEMA give rise to signals both in ¹H-NMR and ¹³C-NMR spectra which are split in distinct peaks associated with different configuration sequences. In the ¹H-NMR spectrum, in agreement with the peaks that have been described for PMMA,⁶⁶ the methyl group on the polymer backbone (H_a) gives three signals: 0.95 ppm (syndiotactic triad, rr), 1.09 ppm (heterotactic triad, mr/rm), and 1.30 ppm (isotactic triad, mm). The content of triads with different tacticity configurations in the polymer chain is proportional to the areas of the respective peaks, when well resolved.⁶⁷ Based on the signals arising from H_a , it is evident that isotactic triads are present only in low amounts, whereas syndiotactic triads are prevalent. The signals arising from the methylene protons in the backbone (H_b) appear at 1.50, 1.90 and 1.96 ppm, with a more complex configuration-based splitting compared to H_a .

Similarly, the ¹³C-NMR spectrum of PDMAEMA presented in Fig. S3† shows tacticity-split signals of methyl (C_a), methylene (C_b), quaternary (C_c) and carbonyl carbons in the backbone, with patterns that are typical of polymethacrylates.^{51,66}

While the signals of the initiator segment at the α chain end were identified, ω -end group analysis by means of NMR was limited. Evidence of Br-terminated PDMAEMA chains is hard to find in NMR spectra because of overlapping signals. The protons of the methyl and methylene groups geminal to the Br-end group cannot be distinguished for this reason, as already pointed out in a previous study.²⁵ Similarly, in the ¹³C-NMR spectrum the signal of the quaternary carbon at the chain end, expected at approximately 58 ppm in analogy to Br-terminated PMMA,⁶⁸ overlaps with the C_d resonance (Fig. S3†).

On the other hand, small characteristic signals of ω -unsaturated polymer chains were detected in the NMR spectra of PDMAEMA, as a consequence of side reactions affecting the ω chain end. After proving that they did not originate from monomer residues, a quantitative analysis of CEF was conducted based on their integration, made possible by the selection of suitable parameters for the NMR analysis (*i.e.* the sample concentration and the number of scans).

Two signals of alkene protons were identified in the ¹H-NMR spectrum at 5.61 and 6.18 ppm, denoted $H_{x(1)}$ and $H_{x(2)}$ in Fig. 2. These signals were found to correlate with each other in the COSY spectrum (Fig. S5†) and to correlate with carbon signals of low intensity in the HMBC spectrum, attributed to the ω chain end (Fig. S8†). Based on their chemical shift and on the HMBC long-range C–H correlations, the signals were attributed to the alkene protons of structure III shown in Scheme 2. Their chemical shift is extremely similar to that of



Scheme 2 Possible side reactions leading to active end group loss during the Cu(I)-mediated ATRP of DMAEMA. ^a Disproportionation. ^b Chain transfer. ^c Elimination of HBr.

the alkene protons of the DMAEMA monomer, but diffusion-ordered spectroscopy (DOSY) allowed their assignment to polymer units and not the residual monomer. In the DOSY spectrum of the polymer (Fig. S12†), protons $H_{x(1)}$ and $H_{x(2)}$ indeed have a diffusion coefficient of approximately $1 \times 10^{-6} \text{ cm}^2 \text{ s}^{-1}$, in contrast with the diffusion coefficient of $2 \times 10^{-5} \text{ cm}^2 \text{ s}^{-1}$ displayed by the monomer signals (Fig. S13†).

Side reactions affecting the control over the polymerization are known to occur during polymerizations *via* traditional ATRP, especially when the reaction is stopped at medium to high conversions.⁶⁰ In Scheme 2, the major possible pathways towards the formation of PDMAEMA “dead” chains, under the current experimental conditions, are summarized as follows: (a) termination *via* disproportionation, producing an ω -unsaturated chain (structure III)—*via* abstraction of a methyl proton from the propagating radical chain—and an ω -hydrogenated chain (structure II) in a 1:1 ratio. On the other hand, the elimination product originating from the abstraction of a methylene proton (structure III-A in Fig. S14A†), is commonly neglected,^{54,69,70} since structure III is largely favored due to lower steric hindrance.⁷¹ Nonetheless, the formation of structure III-A is not expected to alter the results of this study in terms of CEF assessment: in MS, the mass would be equal to that of structure III; in ¹H-NMR, the signal of the resulting alkene proton is expected to overlap with the signal of proton $H_{x(2)}$ of structure III, and therefore it would be accounted for. (b) Chain transfer *via* hydrogen radical abstraction, which has been shown to occur under certain conditions with the ligand acting as the hydrogen radical donor (usually at very high conversions or in the presence of excess ligand; however, in this study it cannot be excluded that DMAEMA amines may lead to a similar reaction).^{72–74} (c) Additionally, it has been reported that the



elimination of HBr occurs during ATRP at medium to high conversion,^{60,75} also leading to structure III.

Termination *via* recombination, on the other hand, is known to have a minor impact on the polymerization of methacrylates, compared to disproportionation. To mention a largely studied example, approximately 70% of termination occurs *via* disproportionation for PMMA at 25 °C.^{69,76,77} However, disproportionation is known to be even more dominant for polymer chains with a bulkier ester alkyl side group, such as PDMAEMA;⁷⁸ model studies have shown that the ratio of k_{td}/k_{tr} (where k_{td} is the rate constant of termination *via* disproportionation and k_{tr} is the rate constant of termination *via* recombination) is approximately twice as high for butyl methacrylate (BMA) compared to MMA.⁷¹ In this study, recombination was thus not expected to be a significant side reaction in the polymerization of DMAEMA, which was confirmed by MALDI-TOF MS.

From the ratio between the integrated signals of the $H_{x(1)}$ or $H_{x(2)}$ alkene protons at unsaturated ω chain ends and the signal of α chain end protons H_i (Fig. S10[†]), it was found that approximately 27% of all PDMAEMA chains are terminated with a C=C double bond (although small signals only allow for a rough estimation). If disproportionation was the main mechanism responsible for the formation of chains of structure III, a population of structure II should be present in an equal amount, resulting in CEF \approx 46% (approximately 54% of all PDMAEMA chains are devoid of the active ω -end group). This assumption was further supported by the results of MALDI-TOF MS.

The singlet of the terminal proton in structure II could not be identified in the ¹H-NMR spectrum, therefore a direct quantification of structure II was not possible by means of NMR spectroscopy. Structure II was, however, detected by MALDI MS measurements.

Overall, highly valuable data were obtained from NMR analysis. The assignment of the signal of proton H_i at the α chain end allowed the determination of the $M_{n,NMR}$. Moreover, in combination with the unequivocal assignment of protons $H_{x(1)}$ and $H_{x(2)}$ at the ω chain end, it enabled a preliminary estimation of CEF.

When comparing the ¹H-NMR spectrum of PDMAEMA shown in Fig. 2, obtained in acetone-*d*₆, with a spectrum of the same polymer in D₂O (Fig. S15[†]), it is evident that less information is provided by the latter.

The results highlight a rather poor chain end fidelity and a large impact of side reactions over the polymerization mechanism, despite the fairly good data obtained in terms of kinetics, the agreement between the targeted and experimental degree of polymerization, as well as dispersity of the polymer. In the current case, the end group loss was apparently slow enough for it not to alter the agreement between the targeted and experimental DP. Stopping the polymerization at higher conversion would most likely lead to more evident effects. However, a low CEF obviously has a detrimental effect on the possibility of chain-extending PDMAEMA, as confirmed later, and therefore it is important to highlight which tools are available to assess it.

The polymerization conditions may be tuned to improve the control over the side reactions, but this was beyond the scope of this work.

MALDI-TOF MS analysis of PDMAEMA

Structural investigation of PDMAEMA was further conducted by mass spectrometry, which can provide in-depth insight into the chain end fidelity. In this regard, soft ionization MS techniques are a powerful asset, since they can display specific populations of polymer chains. Quantitative analyses are, however, challenging due to limitations such as mass bias.⁵²

MS spectra of PDMAEMA have rarely been reported in the literature, and the use of this analysis should be further optimized. The interpretation of the spectra was not straightforward, and some important limitations were encountered when using MALDI-TOF MS for the characterization of PDMAEMA. The following discussion is expected to be a valuable basis for further studies.

An enlargement of the MALDI-TOF MS spectrum of PDMAEMA (isolated at 52% conversion) is presented in Fig. 3A. A population corresponding to structure I (Scheme 2) was not detected in the spectrum. Two main populations were observed, as Na⁺ adducts. The consecutive peaks in each of these populations are separated by a mass interval corresponding to the monoisotopic mass of DMAEMA repeating units ($\Delta = 157.1 \pm 0.1$), hence the peaks are to be attributed to polymer chains. This provides proof that no side reactions have affected the units in the main populations of polymer chains. Additionally, smaller peaks due to adducts with H⁺ and K⁺ are present.

The masses of the peaks in the major series correspond to the masses of the overlapping isotopic distributions of structures II and III shown in Scheme 2. The presence of ω -hydrogenated and ω -unsaturated chains devoid of the Br-end group functionality is in agreement with the findings deduced from NMR analysis, although MALDI-TOF MS provides significantly more detailed information. The population of deactivated chains appears to be predominant in the mass spectrum.

The absence of the expected end group in the masses of at least one of the main populations detected by MALDI-TOF MS is often observed for polymethacrylates and polyacrylates prepared by RDRP techniques, confirming the presence of side reactions during polymerization such as those displayed in Scheme 2.^{72,74,79–81} It must be noted that dehalogenation *via* gas phase elimination of HBr or HCl during the MALDI ionization process, and thus not correlated with reactions occurring during polymerization, has also been proposed in this context.^{25,82–85} Nonaka *et al.* proved that under their experimental conditions, the appearance of peaks due to deactivated chains of poly(methyl methacrylate), poly(methyl acrylate) and polystyrene was linked to the high laser power and decomposition during the flight in the reflector mode.⁸⁶ In this study, the minimum laser power required to generate a spectrum was used, and the ratio between the peak intensities of the two main populations observed when mass spectra were acquired



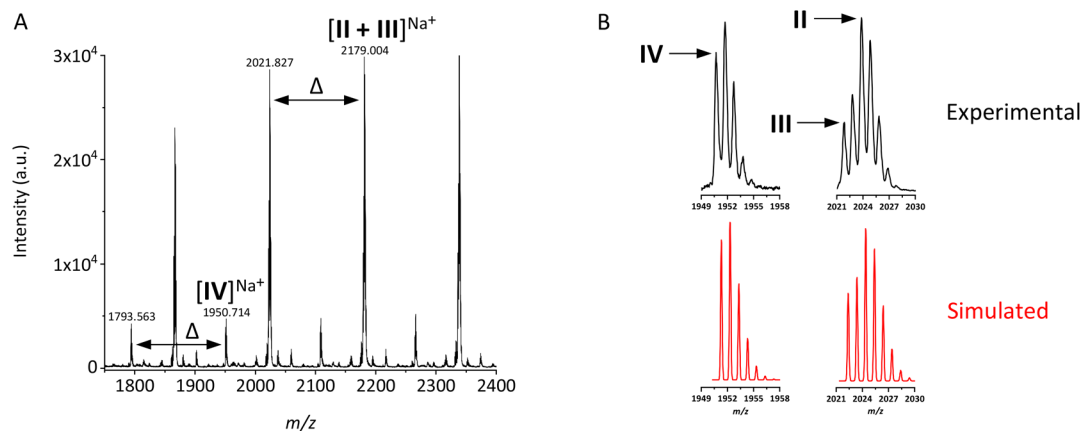


Fig. 3 (A) Enlargement of the MALDI-TOF MS spectrum of PDMAEMA acquired in the reflector mode, where monoisotopic masses of different populations are reported. In the case of populations II and III, whose signals overlap, the monoisotopic mass of population III is reported. The full spectrum is shown in Fig. S17C.† (B) Comparison between the observed isotope distributions and those simulated *via* EnviPat,⁸⁸ with arrows pointing at the monoisotopic masses of different populations for DP = 12.

in the reflector mode (Fig. 3A) is similar to that obtained in the linear mode (Fig. S16B†) with the latter displaying lower isotopic resolution. Moreover, complementary evidence of ω -unsaturated structures was found in the NMR spectra. Therefore, the mass populations assigned to structures II and III were believed to exist in the sample prior to analysis, at least partially; a contribution of gas phase HBr elimination during the MALDI process could not be excluded.

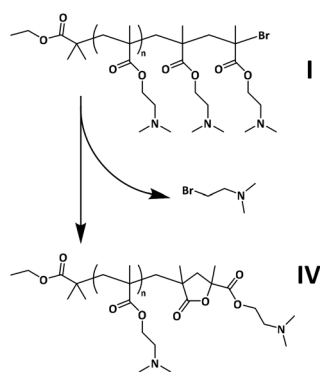
A MALDI-induced dehalogenation process involving cyclization at the ω -chain end is proposed here to account for the presence of the population of lower intensity identified in the mass spectrum, whose masses are equal to those of chains of structure IV, depicted in Scheme 3. It has been shown previously that the C–X (X = Br or Cl) bond in polymethacrylates may be unstable under the conditions of the MALDI ionization process, in contrast with polyacrylates.⁸⁷ Irvine and co-workers demonstrated that decomposition *via* the loss of methyl bromide and formation of a five-membered lactone end group can occur for halogen-terminated PMMA.⁶⁸ Evidence of PMMA with this end group in MALDI-TOF MS analysis has since been

highlighted in several studies.^{89–91} Here, a similar mechanism is proposed for the end group decomposition of PDMAEMA during MALDI, with loss of 2-bromo-*N,N*-dimethylethanamine and cyclization as depicted in Scheme 3.

The interpretation of the masses allowed the identification of the main populations of PDMAEMA chains, with the support of well-resolved isotope distributions. The isotope distributions of the peaks assigned to populations IV and II + III were compared to simulated isotope distributions, as shown in Fig. 3B (where the simulated isotope distribution arising from population II + III was based on a 1 : 1 intensity ratio between the two populations). The similarity of the observed patterns with simulated patterns is a strong confirmation of the assignment of the peaks to the proposed structures.

The intensity ratio between peaks of populations IV and II + III was not used for an estimation of CEF, for two reasons. First, the mechanism shown in Scheme 3 might not be the only process of MALDI-induced end group decomposition occurring for PDMAEMA. Second, a strong effect of mass discrimination towards shorter chains was observed, especially when mass spectra were acquired in the reflector mode (*e.g.* the spectrum reported in Fig. 3A), but probably also affecting the spectra obtained in the linear mode (Fig. S16A†). For this reason, MS was not used for assessing the average molecular weight and dispersity of PDMAEMA. Mass discrimination towards shorter chains has been highlighted and thoroughly studied for more common polymethacrylates, such as PMMA.^{92–94} On the other hand, the data available on matrix selection and optimization of sample preparation for PDMAEMA are still very scarce. In this work, a limited screening of commonly used matrices was carried out before selecting 2,3-dihydrobenzoic acid (DHB) as the preferred matrix. Further studies may provide improvements in this regard.

The issue of mass discrimination obviously implies that the amount of terminated polymer chains is overestimated in the mass spectra. Accordingly, the intensity of the peaks assigned



Scheme 3 Proposed mechanism of PDMAEMA ω -end group decomposition during MALDI-TOF MS analysis.



to population IV, even lower than expected on the basis of the NMR results, was not directly correlated to the amount of Br-terminated PDMAEMA chains in the sample prior to analysis.

The relative intensity of the signals of populations II *versus* III, on the other hand, provides information about the ratio between the different side reactions discussed above and displayed in Scheme 2. Since both populations II and III are present in the mass spectrum, and with a roughly 1:1 intensity ratio, it is reasonable to assume that disproportionation is the dominating side reaction leading to end group loss. A contribution from both elimination of HBr (mechanism c) and chain transfer (mechanism b) to the formation of populations III and II, respectively, could be additionally present. There are several studies investigating end group analysis of various polymers by mass spectrometry where only ω -hydrogenated chain ends have been detected,^{72,79,81,95} hence suggesting the occurrence of chain transfer reactions. In contrast, when populations of ω -unsaturated and ω -hydrogenated chains are detected with equal intensity, disproportionation is the most likely cause.⁸⁰

Since MALDI-TOF MS spectra indicate that ω -hydrogenated polymer chains are present in a 1:1 ratio with respect to ω -unsaturated chains, the results corroborate the calculation of CEF \approx 46% based on NMR analysis.

To gain some qualitative insight into the evolution of CEF with conversion of PDMAEMA, samples isolated at increasing monomer conversions were analyzed by MALDI-TOF MS. The mass spectrum of the polymer at 14% conversion (Fig. S17A[†]), when $M_{n,theo} = 1400 \text{ g mol}^{-1}$, shows a single distribution of peaks with masses corresponding to those of population IV, and therefore attributed to active chains with the preserved Br end group. It can be concluded that, at this early stage of the polymerization, termination was not significant. At 35% conversion (Fig. S17B[†]), when $M_{n,theo} = 3600 \text{ g mol}^{-1}$, active and deactivated chains generated two populations of roughly equal intensity. Finally, at 52% conversion (Fig. S17C[†]), when $M_{n,theo} = 5500 \text{ g mol}^{-1}$, the population of deactivated chains became predominant. Although the last two measurements were affected by mass discrimination to some extent, the results correlate with a gradual loss of bromine functionality during the synthesis, starting at rather low values of conversion.

The results of MALDI-TOF MS analysis on PDMAEMA confirmed the significant presence of deactivated chains, as first suggested by the NMR spectra. Furthermore, mass spectra allowed the obtainment of insightful information about the mode of end group loss and thus confirmed the estimation of CEF \approx 46%. Finally, it is worth pointing out two additional considerations. First, a population with masses corresponding to self-initiated chains was not detected in the spectra, suggesting control over the initiation mechanism. Second, a population with masses corresponding to bimolecular termination *via* recombination was not present in the mass spectra acquired in the linear mode (less affected by mass discrimination), suggesting that this termination route is not plausible for PDMAEMA under the current experimental conditions.

Comparison with different ATRP systems

PDMAEMA synthesized with a traditional EBiB/CuBr/HMTETA ATRP system, and recovered at approximately 50% conversion, was herein used as a model PDMAEMA on which an extensive end group analysis was conducted. Since a variety of ATRP methods exist, with significant differences, it is relevant to apply the characterization tools investigated here to PDMAEMA synthesized using different ATRP systems, especially from the perspective of highlighting the value of CEF, and the impact of different side reactions by means of MALDI-TOF MS. For this reason, the analysis of PDMAEMA prepared using two additional ATRP systems is discussed here.

First, PDMAEMA_{Cl} was synthesized using CuCl while keeping all other experimental conditions previously used for the CuBr-mediated ATRP constant. Chlorine is regarded as a less labile RDRP end group compared to bromine, due to the higher dissociation energy of the C–Cl bond, therefore the use of CuCl is expected to result in less significant side reactions, as opposed to CuBr.^{96,97} The dispersity of PDMAEMA_{Cl}, determined by SEC in DMF ($D = 1.2$) was similar to that observed for PDMAEMA synthesized by CuBr-mediated ATRP.

In the mass spectrum recorded for PDMAEMA_{Cl} (Fig. S18A[†]), the same major populations observed for PDMAEMA synthesized with CuBr were detected (IV and II + III). However, the intensity ratio of the peaks of populations IV over II + III was found to be significantly higher. This supported the assumption that population IV corresponds to the population of chains with a preserved halogen end group: a population which is present in higher number in the case of PDMAEMA_{Cl} due to the higher stability of the Cl end group. It is thus reasonable to propose that halogen-terminated PDMAEMA, under the current experimental conditions, undergoes the end group decomposition pathway presented in Scheme 3 during the MALDI process, both in the case of Br- and Cl-terminated chains. It is hypothesized that the pattern of isotope distribution attributed to populations II + III for PDMAEMA_{Cl} (Fig. S18B[†]) differs from that shown in Fig. 3B for PDMAEMA prepared by CuBr-mediated ATRP due to a smaller contribution of the elimination of HCl, compared to HBr.

The ¹H-NMR spectrum of PDMAEMA_{Cl}, with integrated signals, is reported in Fig. S19.[†] Signals originating from protons in unsaturated chain ends were detected. By applying the same calculation carried out before, a CEF \approx 34% was estimated for PDMAEMA_{Cl}, lower than that of PDMAEMA synthesized by CuBr-mediated ATRP.

To provide an additional comparison, PDMAEMA_{UV,EBiB} and PDMAEMA_{UV,EBPA} were prepared by photomediated ATRP using 10-phenylphenothiazine (PTH) as a photoredox catalyst, according to a method adapted from that described by Treat *et al.*⁹⁸ (however, a different light source was used). The two polymers were prepared by using two different initiators, EBiB and ethyl α -bromophenylacetate (EBPA), respectively. A high dispersity was determined by SEC in DMF when analyzing PDMAEMA_{UV,EBiB} and PDMAEMA_{UV,EBPA} ($D = 2.4$, in both cases), indicating very low control over the polymerization.



In the MALDI-TOF MS spectra of the polymers (Fig. S20 and S21†), two major populations are present. The population with the highest intensity corresponds to terminated chains of structure II + III. Interestingly, both for PDMAEMA_{UV,EBiB} and PDMAEMA_{UV,EBPA} the masses of the signals of the population of lower intensity correspond to those of self-initiated PDMAEMA chains devoid of the Br end group. It is proposed that, under the current experimental conditions, the amine group of DMAEMA is involved in a photoredox reaction resulting in the abstraction of a hydrogen atom from the carbon adjacent to the amine group, with a mechanism analogous to that reported by Allushi *et al.* for tertiary amines used as “co-initiators” for photomediated ATRP in the presence of thioxanthone photocatalysts.⁹⁹ Under these conditions, the resulting DMAEMA radical (Fig. S14B†) is thus able to initiate polymerization, competing with the initiator. It is evident that the choice of experimental set-up for photomediated ATRP (*e.g.* choice of photocatalyst, wavelength of the light source and light intensity) is particularly important, to achieve polymers with narrow MWD and to control the initiation mechanism. Since self-initiated chains are present in the samples of PDMAEMA_{UV,EBiB} and PDMAEMA_{UV,EBPA}, it is not possible to calculate CEF on the basis of the ratio of integrated signals of ω and α chain ends. However, since neither a population of chains having the Br end group nor a population of chains corresponding to structure IV were identified in the mass spectra, CEF can be expected to be extremely low.

It is evident that the results of mass spectrometry conducted on PDMAEMA samples prepared by these additional ATRP systems raise insightful considerations, although more extensive investigations are needed to draw conclusions with respect to the polymerization mechanisms. CuCl can ensure higher control over the polymerization, compared to CuBr. Photomediated ATRP of DMAEMA, on the other hand, can lead to a notable extent of self-initiation of DMAEMA and a high dispersity.

SEC analysis of PDMAEMA

Size-exclusion chromatography, performed using *N,N*-dimethylformamide (DMF) as the eluent and poly(methyl methacrylate) standard calibration, was used to provide information about the polymerization kinetics, but its use is not relevant for giving representative values of molecular weight of PDMAEMA. SEC analysis on purified PDMAEMA determined $M_{n,SEC}$ values significantly deviating from $M_{n,theo}$. When the analysis was repeated, traces with varying $M_{n,SEC}$ values were obtained, ranging from 7000 to 9600 g mol⁻¹. Polymers containing amine functionalities such as PDMAEMA are difficult to characterize using conventional SEC systems. Many authors have highlighted that the plausible interaction of PDMAEMA with the columns may lead to a broadening of the MWD, especially at low molecular weights, and additionally the $M_{n,SEC}$ obtained is highly affected by different hydrodynamic volumes in relation with the standards.^{29,31,100} Addition of triethylamine in the mobile phase may help in avoiding interactions with the columns¹⁰¹ and other existing solutions (*e.g.*

converting the polymer into poly(methyl methacrylate) for the purpose of the analysis³ and using more complex SEC systems²⁶).

Chain extension of PDMAEMA

A chain extension experiment produced additional evidence of the extent of chain end fidelity of PDMAEMA. PDMAEMA was used as a macroinitiator for the synthesis of PDMAEMA-*b*-PMMA by ATRP mediated by CuCl. CuCl was used for chain extension since the high activity of the Br end group of the macroinitiator, more labile than Cl which is expected to become predominant in the propagation step, should ensure initiation efficiency.¹⁰² The kinetics plot of the reaction, reported in Fig. 4A, reveals a trend similar to that observed in the polymerization of DMAEMA. The plots of $M_{n,SEC}$ and D over conversion are shown in Fig. S22.† The rather linear trend observed in the $\ln([M]_0/[M])$ might be the result of a balance between slow initiation and termination, since it is not corroborated by the evidence of high control over the polymerization.

In agreement with the presence of deactivated chains revealed by NMR and MS, the MWDs of aliquots withdrawn from the polymerization mixture at 34% and 43% conversion, plotted in dotted and dashed blue lines in Fig. 4B, show bimodality. The peak at lower molecular weight, overlapping with the trace of the PDMAEMA macroinitiator, corresponds to the fraction of PDMAEMA dead chains, unable to initiate copolymerization.

During copolymerization, the presence of dead PDMAEMA, unable to initiate chain extension, could also be immediately detected by NMR, using a DOSY experiment. In Fig. S23,† the DOSY spectrum of the sample withdrawn from the polymerization mixture at 34% conversion is reported. The spectrum reveals the presence of the PDMAEMA homopolymer, whose protons show distinctive signals with a lower diffusion coefficient compared to the protons of the copolymer.

The selected method of purification of the copolymer (precipitation in MeOH) ensured the complete removal of unreacted PDMAEMA, as it is evident from the molecular

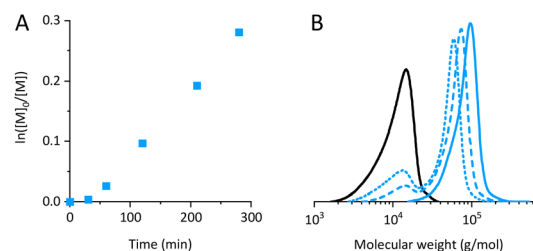


Fig. 4 (A) Kinetic plot of $\ln([M]_0/[M])$ over time during ATRP of MMA initiated by PDMAEMA. (B) Overlay of MWDs obtained by SEC in DMF: PDMAEMA macroinitiator (black, solid line); samples from the PDMAEMA-*b*-PMMA reaction mixture at $t = 150$ min, 34% conversion (blue, dotted line) and at $t = 210$ min, 43% conversion (blue, dashed line); purified PDMAEMA-*b*-PMMA isolated at $t = 280$ min, 55% conversion (blue, solid line).



weight distribution of the purified copolymer plotted in a solid blue line in Fig. 4B. The block copolymer showed $D = 1.2$, suggesting that a functional material can be obtained despite the non-ideal character of the polymerization.

When discussing the results of chain extension of PDMAEMA, an additional consideration can be elaborated. The consequence of a significant amount of disproportionation is that the ω -unsaturated chains are not truly dead; in fact, they can be involved in chain transfer reactions¹⁰³ and may even react further as macromonomers (although with low reactivity) and thus be incorporated into the copolymer structure. This mechanism might contribute to the chain extension; however, further investigations should be conducted in this regard.

Molecular weight determination of PDMAEMA-*b*-PMMA

Assessing the molecular weight of the PDMAEMA-*b*-PMMA copolymer proved challenging. The presence of deactivated chains affects the monomer/macromonomer ratio, hindering the correlation between targeted and experimental DP. Since the amount of the initial block able to act as the macroinitiator was not known *a priori*, the DP_{theo} of PMMA initiated by PDMAEMA (and, consequently, $M_{n,theo}$) was an underestimated value. Moreover, after purification of the block copolymer, determining $M_{n,NMR}$ based on the ratio between integrated signals of the extension block and of the initiating block was not possible, due to the different solubility of the two blocks in the NMR solvent leading to the overestimation of one over the other. The PMMA/PDMAEMA integrated signal ratio in the ¹H-NMR spectrum of the copolymer obtained in acetone-*d*₆, reported in Fig. S24,† would lead to a DP_{NMR} of 1470 for PMMA, resulting in a $M_{n,NMR}$ of approximately 150 000 g mol⁻¹ which is not a reliable estimation.

To determine the molecular weight of PDMAEMA-*b*-PMMA, the amine in the repeating units of PDMAEMA was an advantageous asset. Elemental analysis (EA) was conducted on the copolymer using a total nitrogen analyzer. With this method, the ratio of mol_{DMAEMA} per $g_{copolymer}$ to mol_{MMA} per $g_{copolymer}$ was determined, from which it was possible to estimate the $DP_{EA} = 553$ of PMMA and, consequently, the average molecular weight $M_{n,EA}$ as presented in Table 1. The obtained value of $M_{n,EA}$ was considered highly representative of the average molecular weight of the purified copolymer, in contrast with $M_{n,SEC}$ which is highly affected by the difference in the hydrodynamic volume compared to the standards and thus not accurate.

Table 1 Molecular weight of PDMAEMA-*b*-PMMA assessed via different methods

$M_{n,theo}^a$	$M_{n,EA}^b$	$M_{n,SEC}^c$
27 400	60 900	75 900

^a Calculated on the basis of conversion assessed by ¹H-NMR.

^b Determined by calculations based on total nitrogen analysis.

^c Determined by SEC with DMF as the eluent.

$M_{n,EA}$ strongly deviates from $M_{n,theo}$, confirming that deactivated PDMAEMA chains were largely present in the polymerization mixture. Since PMMA's DP_{EA} is 2.5 times the DP calculated from the conversion of MMA assessed by ¹H-NMR ($DP_{theo} = 219$), an estimation of CEF $\approx 40\%$ is deduced. This value is slightly lower than that calculated on the basis of NMR combined with MS analysis (CEF $\approx 46\%$), but very close to it. This shows that the estimation of CEF based on the data provided by NMR and MS is rather reliable. When designing the block copolymerization of PDMAEMA, the calculation of CEF may allow the adjustment of the ratio of the monomer to PDMAEMA for a better prediction of the final degree of polymerization.

Conclusions

In response to a lack of information available in the literature, an in-depth characterization of PDMAEMA prepared by ATRP was carried out by using NMR and MALDI-TOF MS techniques. The detailed investigation of PDMAEMA's structure, especially the chain ends, provides insightful information about future studies where PDMAEMA-based architectures are prepared.

A comprehensive one- and two-dimensional NMR characterization allowed the assignment of the signals originating from PDMAEMA and consequent verification of the experimental degree of polymerization of the polymer, as well as the presence of side reactions lowering the fraction of chain end functionality (CEF).

An interpretation of the peaks in MALDI-TOF MS spectra was given, with the conclusion that a MALDI-induced fragmentation—with the formation of a lactone at the ω chain end—affects the observed masses of halogen-terminated PDMAEMA chains (both Br- and Cl-terminated). Based on the mass spectra, the loss of the active end group was found to be mainly due to disproportionation during polymerization. MALDI-TOF MS also enabled a qualitative assessment of the evolution of the loss of the active end group throughout the course of the polymerization. It was observed that Br-terminated chains were gradually converted into chains devoid of the expected end group, already with a significant end group loss at low values of conversion.

The selected analyses allowed the examination of aspects of the non-ideal character of the conducted polymerization of DMAEMA and subsequent chain extension. Additionally, the combination of the data collected from NMR and MS led to estimate CEF $\approx 46\%$. Providing tools for conducting these analyses will be useful especially in view of the synthesis of block copolymers of PDMAEMA by ATRP.

By varying the ATRP system used, different samples of PDMAEMA were analyzed and compared. MALDI-TOF MS and NMR analyses provided evidence that replacing CuBr with CuCl leads to higher control over the ω chain end. Furthermore, when samples of PDMAEMA synthesized by photomediated ATRP were analyzed, MALDI-TOF MS allowed the assessment of the presence of a self-initiated polymer.



The chain extension of PDMAEMA with MMA confirmed the poor chain end fidelity highlighted by NMR and MALDI-TOF MS; only a fraction of the PDMAEMA chains initiated the block copolymerization, as shown by SEC and DOSY. The obtained copolymer was, however, successfully purified from any unreacted PDMAEMA. The challenge of determining the DP of the extension block, not knowing *a priori* the percentage of active PDMAEMA chains, was overcome by means of elemental analysis.

PDMAEMA is a polymer with highly tunable properties, which are attractive in the context of a variety of advanced applications. It is often employed in the synthesis of tailor-made copolymer architectures. The characterization tools provided in this work are important in view of verifying the extent of control over PDMAEMA polymerization and chain extension, and highlighting how well-defined the polymer structures are. Therefore, these tools are believed to strengthen the versatility of PDMAEMA and its copolymer architectures, especially in the contexts where well-defined structures are needed.

Conflicts of interest

There are no conflicts to declare.

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References

- V. Bütün, S. P. Armes and N. C. Billingham, *Polymer*, 2001, **42**, 5993–6008.
- G. M. Liu, D. Wu, C. C. Ma, G. Z. Zhang, H. F. Wang and S. H. Yang, *ChemPhysChem*, 2007, **8**, 2254–2259.
- F. A. Plamper, M. Ruppel, A. Schmalz, O. Borisov, M. Ballauff and A. H. E. Muller, *Macromolecules*, 2007, **40**, 8361–8366.
- M. Vamvakaki, G. F. Unali, V. Butun, S. Boucher, K. L. Robinson, N. C. Billingham and S. P. Armes, *Macromolecules*, 2001, **34**, 6839–6841.
- R. Yanez-Macias, I. Alvarez-Moises, I. Perevyazko, A. Lezov, R. Guerrero-Santos, U. S. Schubert and C. Guerrero-Sanchez, *Macromol. Chem. Phys.*, 2017, **218**, 7.
- C. L. Peng, H. M. Tsai, S. J. Yang, T. Y. Luo, C. F. Lin, W. J. Lin and M. J. Shieh, *Nanotechnology*, 2011, **22**, 11.
- A. Car, P. Baumann, J. T. Duskey, M. Cham, N. Bruns and W. Meier, *Biomacromolecules*, 2014, **15**, 3235–3245.
- M. Farshbaf, S. Davaran, A. Zarebkohan, N. Annabi, A. Akbarzadeh and R. Salehi, *Artif. Cells, Nanomed., Biotechnol.*, 2018, **46**, 1872–1891.
- P. van de Wetering, J. Y. Cherng, H. Talsma, D. J. A. Crommelin and W. E. Hennink, *J. Control. Release*, 1998, **53**, 145–153.
- S. Agarwal, Y. Zhang, S. Maji and A. Greiner, *Mater. Today*, 2012, **15**, 388–393.
- B. M. Reis, S. P. Armes, S. Fujii and S. Biggs, *Colloids Surf., A*, 2010, **353**, 210–215.
- J. Engström, F. L. Hatton, L. Wågberg, F. D'Agosto, M. Lansalot, E. Malmström and A. Carlmark, *Polym. Chem.*, 2017, **8**, 1061–1073.
- M. Wählander, F. Nilsson, A. Carlmark, U. W. Gedde, S. Edmondson and E. Malmström, *Nanoscale*, 2016, **8**, 14730–14745.
- T. Kaldéus, M. R. Telaretti Leggieri, C. Cobo Sanchez and E. Malmström, *Biomacromolecules*, 2019, **20**, 1937–1943.
- A. Munoz-Bonilla and M. Fernandez-Garcia, *Prog. Polym. Sci.*, 2012, **37**, 281–339.
- L. A. B. Rawlinson, S. M. Ryan, G. Mantovani, J. A. Syrett, D. M. Haddleton and D. J. Brayden, *Biomacromolecules*, 2010, **11**, 443–453.
- K. Nagase, J. Kobayashi, A. Kikuchi, Y. Akiyama, H. Kanazawa and T. Okano, *Biomacromolecules*, 2008, **9**, 1340–1347.
- O. Schepelina and I. Zharov, *Langmuir*, 2008, **24**, 14188–14194.
- A. R. Roudman and F. A. DiGiano, *J. Membr. Sci.*, 2000, **175**, 61–73.
- R. H. Du and J. S. Zhao, *J. Membr. Sci.*, 2004, **239**, 183–188.
- J. T. Tang, M. F. X. Lee, W. Zhang, B. X. Zhao, R. M. Berry and K. C. Tam, *Biomacromolecules*, 2014, **15**, 3052–3060.
- T. Saigal, H. C. Dong, K. Matyjaszewski and R. D. Tilton, *Langmuir*, 2010, **26**, 15200–15209.
- X. Zhang, J. H. Xia and K. Matyjaszewski, *Macromolecules*, 1998, **31**, 5167–5169.
- H. Dong and K. Matyjaszewski, *Macromolecules*, 2008, **41**, 6868–6870.
- R. A. Cordeiro, N. Rocha, J. P. Mendes, K. Matyjaszewski, T. Guliashvili, A. C. Serra and J. F. J. Coelho, *Polym. Chem.*, 2013, **4**, 3088–3097.
- M. Sahnoun, M. T. Charreyre, L. Veron, T. Delair and F. D'Agosto, *J. Polym. Sci., Part A: Polym. Chem.*, 2005, **43**, 3551–3565.
- A. Zengin, G. Karakose and T. Caykara, *Eur. Polym. J.*, 2013, **49**, 3350–3358.
- Q. F. Xiong, P. H. Ni, F. Zhang and Z. Q. Yu, *Polym. Bull.*, 2004, **53**, 1–8.
- A. P. Narrainen, S. Pascual and D. M. Haddleton, *J. Polym. Sci., Part A: Polym. Chem.*, 2002, **40**, 439–450.
- S. B. Lee, A. J. Russell and K. Matyjaszewski, *Biomacromolecules*, 2003, **4**, 1386–1393.
- C. Bruce, I. Javakhishvili, L. Fogelström, A. Carlmark, S. Hvilsted and E. Malmström, *RSC Adv.*, 2014, **4**, 25809.



- 32 J. Pietrasik, B. S. Sumerlin, R. Y. Lee and K. Matyjaszewski, *Macromol. Chem. Phys.*, 2007, **208**, 30–36.
- 33 Y. Y. Xu, S. Bolisetty, M. Drechsler, B. Fang, J. Y. Yuan, M. Ballauff and A. H. E. Müller, *Polymer*, 2008, **49**, 3957–3964.
- 34 V. V. de Souza, M. L. D. Noronha, F. L. A. Almeida, C. A. R. Prado, A. C. Doriguetto and F. H. Florenzano, *Polym. Bull.*, 2011, **67**, 875–884.
- 35 G. L. Xiao, Z. B. Hu, G. P. Zeng, Y. Q. Wang, Y. Q. Huang, X. L. Hong, B. L. Xia and G. Y. Zhang, *J. Appl. Polym. Sci.*, 2012, **124**, 202–208.
- 36 F. L. Baines, S. P. Armes, N. C. Billingham and Z. Tuzar, *Macromolecules*, 1996, **29**, 8151–8159.
- 37 B. P. Binks, R. Murakami, S. P. Armes and S. Fujii, *Angew. Chem., Int. Ed.*, 2005, **44**, 4795–4798.
- 38 J. C. P. de Souza, A. F. Naves and F. H. Florenzano, *Colloid Polym. Sci.*, 2012, **290**, 1285–1291.
- 39 V. Chrysostomou and S. Pispas, *J. Polym. Sci., Part A: Polym. Chem.*, 2018, **56**, 598–610.
- 40 H. Walter, C. Harrats, P. Muller-Buschbaum, R. Jerome and M. Stamm, *Langmuir*, 1999, **15**, 1260–1267.
- 41 J. Engström, T. Benselfelt, L. Wågberg, F. D'Agosto, M. Lansalot, A. Carlmark and E. Malmström, *Nanoscale*, 2019, **11**, 4287–4302.
- 42 E. Larsson, C. Cobo Sanchez, C. Porsch, E. Karabulut, L. Wågberg and A. Carlmark, *Eur. Polym. J.*, 2013, **49**, 2689–2696.
- 43 I. Zaborniak, M. Sroka and P. Chmielarz, *Polymer*, 2022, **254**, 9.
- 44 M. D. Shalati and R. M. Scott, *Macromolecules*, 1975, **8**, 127–130.
- 45 L. Carlsson, A. Fall, I. Chaduc, L. Wågberg, B. Charleux, E. Malmström, F. D'Agosto, M. Lansalot and A. Carlmark, *Polym. Chem.*, 2014, **5**, 6076–6086.
- 46 P. van de Wetering, N. J. Zuidam, M. J. van Steenberg, O. van der Houwen, W. J. M. Underberg and W. E. Hennink, *Macromolecules*, 1998, **31**, 8063–8068.
- 47 P. Cotanda, D. B. Wright, M. Tyler and R. K. O'Reilly, *J. Polym. Sci., Part A: Polym. Chem.*, 2013, **51**, 3333–3338.
- 48 N. P. Truong, Z. F. Jia, M. Burges, N. A. J. McMillan and M. J. Monteiro, *Biomacromolecules*, 2011, **12**, 1876–1882.
- 49 L. Mespouille, M. Vachaud, F. Suriano, P. Gerbaux, W. Van Camp, O. Coulembier, P. Degee, R. Flammang, F. Du Prez and P. Dubois, *React. Funct. Polym.*, 2008, **68**, 990–1003.
- 50 V. Barboiu, G. L. Ailiese, A. G. Grigoras, E. Rusu and E. S. Dragan, *Int. J. Polym. Anal. Charact.*, 2019, **24**, 610–618.
- 51 J. Niskanen, C. Wu, M. Ostrowski, G. G. Fuller, S. Hietala and H. Tenhu, *Macromolecules*, 2013, **46**, 2331–2340.
- 52 T. Gruendling, S. Weidner, J. Falkenhagen and C. Barner-Kowollik, *Polym. Chem.*, 2010, **1**, 599–617.
- 53 A. H. Soeriyadi, M. R. Whittaker, C. Boyer and T. P. Davis, *J. Polym. Sci., Part A: Polym. Chem.*, 2013, **51**, 1475–1505.
- 54 M. Buback, F. Günzler, G. T. Russell and P. Vana, *Macromolecules*, 2009, **42**, 652–662.
- 55 K. Matyjaszewski, H. C. Dong, W. Jakubowski, J. Pietrasik and A. Kusumo, *Langmuir*, 2007, **23**, 4528–4531.
- 56 A. Boujemaoui, C. Cobo Sanchez, J. Engström, C. Bruce, L. Fogelström, A. Carlmark and E. Malmström, *ACS Appl. Mater. Interfaces*, 2017, **9**, 35305–35318.
- 57 D. Zhou, L.-W. Zhu, B.-H. Wu, Z.-K. Xu and L.-S. Wan, *Polym. Chem.*, 2022, **13**, 300–358.
- 58 P. Krys and K. Matyjaszewski, *Eur. Polym. J.*, 2017, **89**, 482–523.
- 59 V. Coessens, T. Pintauer and K. Matyjaszewski, *Prog. Polym. Sci.*, 2001, **26**, 337–377.
- 60 J. F. Lutz and K. Matyjaszewski, *Macromol. Chem. Phys.*, 2002, **203**, 1385–1395.
- 61 M. Zhong and K. Matyjaszewski, *Macromolecules*, 2011, **44**, 2668–2677.
- 62 F. Lorandi and K. Matyjaszewski, *Isr. J. Chem.*, 2020, **60**, 108–123.
- 63 S. Karanam, H. Goossens, B. Klumperman and P. Lemstra, *Macromolecules*, 2003, **36**, 3051–3060.
- 64 A. K. Nanda and K. Matyjaszewski, *Macromolecules*, 2003, **36**, 8222–8224.
- 65 J. Qiu, B. Charleux and K. Matyjaszewski, *Polimery*, 2001, **46**, 453–460.
- 66 A. S. Brar, G. Singh and R. Shankar, *J. Mol. Struct.*, 2004, **703**, 69–81.
- 67 G. Greyling and H. Pasch, *Anal. Chem.*, 2015, **87**, 3011–3018.
- 68 C. D. Borman, A. T. Jackson, A. Bunn, A. L. Cutter and D. J. Irvine, *Polymer*, 2000, **41**, 6015–6020.
- 69 Y. Nakamura and S. Yamago, *Macromolecules*, 2015, **48**, 6450–6456.
- 70 Y. Nakamura, T. Ogihara and S. Yamago, *ACS Macro Lett.*, 2016, **5**, 248–252.
- 71 G. Moad and D. H. Solomon, in *The Chemistry of Radical Polymerization*, Elsevier Science Ltd, Amsterdam, 2nd edn, 2005, pp. 233–278.
- 72 M. Bednarek, T. Biedron and P. Kubisa, *Macromol. Chem. Phys.*, 2000, **201**, 58–66.
- 73 F. Nystrom, A. H. Soeriyadi, C. Boyer, P. B. Zetterlund and M. R. Whittaker, *J. Polym. Sci., Part A: Polym. Chem.*, 2011, **49**, 5313–5321.
- 74 A. Anastasaki, C. Waldron, P. Wilson, R. McHale and D. M. Haddleton, *Polym. Chem.*, 2013, **4**, 2672–2675.
- 75 J. F. Lutz and K. Matyjaszewski, *J. Polym. Sci., Part A: Polym. Chem.*, 2005, **43**, 897–910.
- 76 C. Bamford, G. Eastmond and D. Whittle, *Polymer*, 1969, **10**, 771–783.
- 77 T. Kashiwagi, A. Inaba, J. E. Brown, K. Hatada, T. Kitayama and E. Masuda, *Macromolecules*, 1986, **19**, 2160–2168.
- 78 G. Odian, in *Principles of polymerization*, John Wiley & Sons, 4th edn, 2004, pp. 236–237.
- 79 K. Matyjaszewski, S. M. Jo, H. J. Paik and D. A. Shipp, *Macromolecules*, 1999, **32**, 6431–6438.
- 80 E. Frick, A. Anastasaki, D. M. Haddleton and C. Barner-Kowollik, *J. Am. Chem. Soc.*, 2015, **137**, 6889–6896.



- 81 M. Bednarek, T. Biedron and P. Kubisa, *Macromol. Rapid Commun.*, 1999, **20**, 59–65.
- 82 S. Coca, C. B. Jasieczek, K. L. Beers and K. Matyjaszewski, *J. Polym. Sci., Part A: Polym. Chem.*, 1998, **36**, 1417–1424.
- 83 K. Matyjaszewski, Y. Nakagawa and C. B. Jasieczek, *Macromolecules*, 1998, **31**, 1535–1541.
- 84 A. Can, E. Altuntas, R. Hoogenboom and U. S. Schubert, *Eur. Polym. J.*, 2010, **46**, 1932–1939.
- 85 X. D. Liu, L. F. Zhang, Z. P. Cheng and X. L. Zhu, *Polym. Chem.*, 2016, **7**, 689–700.
- 86 H. Nonaka, M. Ouchi, M. Kamigaito and M. Sawamoto, *Macromolecules*, 2001, **34**, 2083–2088.
- 87 L. Charles, *Mass Spectrom. Rev.*, 2014, **33**, 523–543.
- 88 M. Loos, C. Gerber, F. Corona, J. Hollender and H. Singer, *Anal. Chem.*, 2015, **87**, 5738–5744.
- 89 N. K. Singha, S. Rimmer and B. Klumperman, *Eur. Polym. J.*, 2004, **40**, 159–163.
- 90 A. T. Jackson, A. Bunn, I. M. Priestnall, C. D. Borman and D. J. Irvine, *Polymer*, 2006, **47**, 1044–1054.
- 91 N. K. Singha and A. L. German, *J. Appl. Polym. Sci.*, 2007, **103**, 3857–3864.
- 92 B. C. Guo, H. Chen, H. Rashidzadeh and X. Liu, *Rapid Commun. Mass Spectrom.*, 1997, **11**, 781–785.
- 93 S. J. Wetzel, C. M. Guttman and J. E. Girard, *Int. J. Mass Spectrom.*, 2004, **238**, 215–225.
- 94 G. Montaudo, F. Samperi and M. S. Montaudo, *Prog. Polym. Sci.*, 2006, **31**, 277–357.
- 95 A. Herberg, X. Q. Yu and D. Kuckling, *Polymers*, 2019, **11**, 23.
- 96 D. A. Shipp, J. L. Wang and K. Matyjaszewski, *Macromolecules*, 1998, **31**, 8005–8008.
- 97 K. Matyjaszewski, D. A. Shipp, J. L. Wang, T. Grimaud and T. E. Patten, *Macromolecules*, 1998, **31**, 6836–6840.
- 98 N. J. Treat, H. Sprafke, J. W. Kramer, P. G. Clark, B. E. Barton, J. Read de Alaniz, B. P. Fors and C. J. Hawker, *J. Am. Chem. Soc.*, 2014, **136**, 16096–16101.
- 99 A. Allushi, C. Kutahya, C. Aydogan, J. Kreutzer, G. Yilmaz and Y. Yagci, *Polym. Chem.*, 2017, **8**, 1972–1977.
- 100 X. Zhang and K. Matyjaszewski, *Macromolecules*, 1999, **32**, 1763–1766.
- 101 S. Creutz, P. Teyssié and R. Jérôme, *Macromolecules*, 1997, **30**, 6–9.
- 102 C. H. Peng, J. Kong, F. Seeliger and K. Matyjaszewski, *Macromolecules*, 2011, **44**, 7546–7557.
- 103 C. L. Moad, G. Moad, E. Rizzardo and S. H. Thang, *Macromolecules*, 1996, **29**, 7717–7726.

