



Cite this: *Org. Biomol. Chem.*, 2023, **21**, 3761

Received 28th March 2023,
Accepted 17th April 2023

DOI: 10.1039/d3ob00476g

rsc.li/obc

First total synthesis of type II abyssomicins: (\pm)-abyssomicin 2 and (\pm)-neoabyssomicin B \dagger

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The intramolecular Diels–Alder reaction (IMDA) of a butenolide derivative, as an entry to the type II abyssomicin scaffold, and the total synthesis of (\pm)-abyssomicin 2 and (\pm)-neoabyssomicin B are reported for the first time. A facile route to the IMDA precursor, the formation of a type I intermediate and two paths to (\pm)-neoabyssomicin B are also discussed.

The family of abyssomicins is a subclass of class I spirotetronate polyketides with fascinating architectures and engaging biological activities, comprising more than 40 members to date.^{1,2} According to their structure, abyssomicins are classified as type I, e.g. abyssomicin C (1), or their “enantiomeric” counterparts at C-15 type II, e.g. abyssomicin 2 (2) and neoabyssomicin B (3) (Fig. 1).^{1,3} Abyssomicin C (1) was the first member to captivate scientific interest due to its inhibitory activity against methicillin-resistant *Staphylococcus aureus* and other Gram-positive bacteria.⁴ Recently, type II abyssomicins have also garnered increasing attention. Thus, abyssomicin 2 (2) was discovered during screening to identify HIV-1 reactivators exhibiting latent HIV activation selectively and antimicrobial activity against Gram-positive bacteria, including methicillin-resistant *Staphylococcus aureus* (MRSA).⁵ Also, many studies have been performed on the biosynthetic pathways leading to both types of compounds,^{6–8} and many impressive syntheses have been reported for abyssomicin C (1) and type I analogues.^{9–11} However, no reported synthetic methods for type II abyssomicins exist.

A biomimetic strategy towards both types of compounds implies the construction of key-intermediates 4 (type I) or 6 (type II) from the corresponding substituted butenolides 5 or 7 via a stereoselective intramolecular Diels–Alder reaction

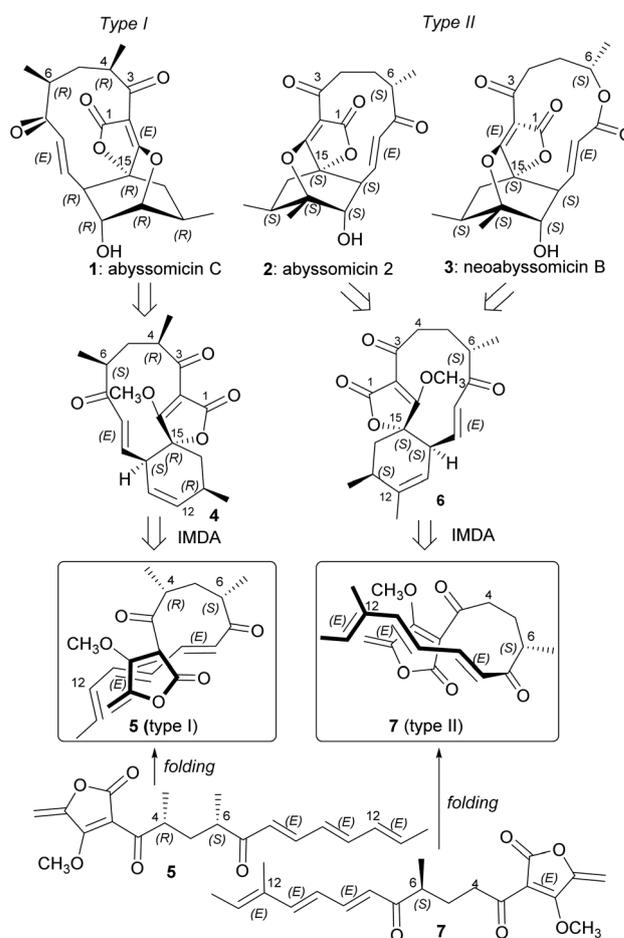


Fig. 1 Representative type I and type II abyssomicins and biomimetic approaches.

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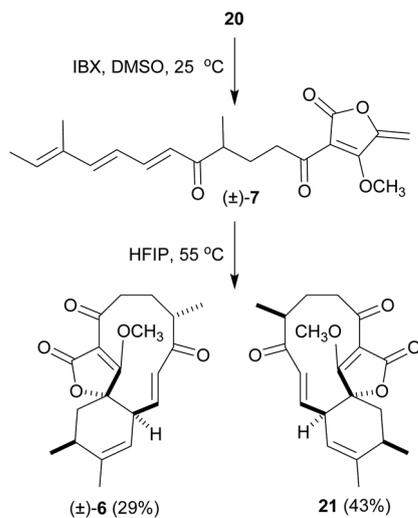
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\dagger Electronic supplementary information (ESI) available: Experimental details, compound characterization, copies of ^1H and ^{13}C and 2D NMR spectra, and X-ray analysis. CCDC 2250166. For ESI and crystallographic data in CIF or other electronic format see DOI: <https://doi.org/10.1039/d3ob00476g>





Scheme 2 Oxidation-IMDA reaction of 20.

mixture. The formation of two isomers was observed, the type I (\pm)-6 (29% yield) and the type II 21 (43% yield), respectively, indicating that precursor (\pm)-7 folds in two ways during cycloaddition. It should be noted that HFIP proved superior to other solvents, such as chloroform and toluene, resulting in lower yields and (\pm)-6/21 ratios. Although a systematic study and a thorough computational and experimental investigation are required to explore the reasons that affect the stereochemical results of this crucial reaction, it is noteworthy that divergent facial selectivities are observed in the IMDA reaction of the two precursors 5 and (\pm)-7, *i.e.*, by simply moving a methyl unit from C4 to C12. The mixture of the two isomers was inseparable by simple flash column chromatography, and preparative thin layer chromatography (prep TLC) was required for their complete separation.

The structure of 21 was further confirmed by X-ray analysis (Fig. 2).

Unfortunately, compound (\pm)-6 was sensitive to purification. It was isolated as a \sim 5:1 inseparable mixture with a cycloaddition product whose formation was observed after prep TLC.‡ However, (\pm)-6 was successfully epoxidized with *m*-CPBA to afford 22, isolated as a single isomer, in 73% yield

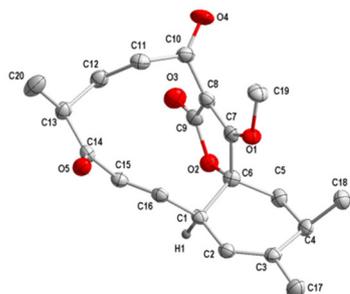
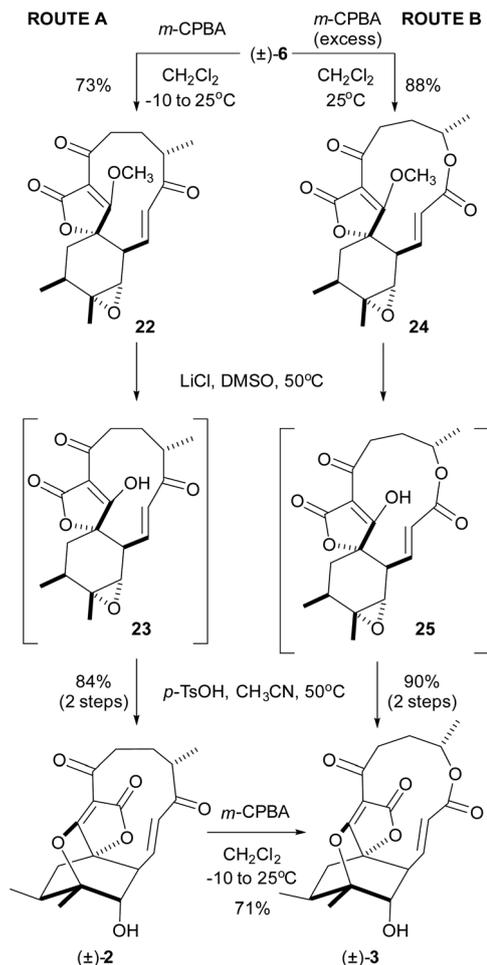


Fig. 2 Structure of 21 by X-ray analysis.

Scheme 3 Final transformations and completion of synthesis of (\pm)-abyssomicin 2 ((\pm)-2) and (\pm)-neoabyssomicin B ((\pm)-3).

(Scheme 3, route A). These conditions proved superior to the use of dimethyldioxirane,⁹ which led to a mixture of products and incomplete conversion of (\pm)-6 to 22. Moreover, a one-pot procedure, including demethylation of 22 with LiCl/DMSO and intramolecular epoxide opening, established the oxygen bridge and led to (\pm)-abyssomicin 2 ((\pm)-2) in 84% yield. Lastly, a Baeyer-Villiger oxidation of the latter with *m*-CPBA led to (\pm)-neoabyssomicin B ((\pm)-3) in 71% yield (43.5% from (\pm)-6). The above sequence of transformations agrees with those proposed biosynthetically.⁸ However, an alternative route to (\pm)-3 from (\pm)-6, where the Baeyer-Villiger oxidation of the 11-membered ring ketone precedes the formation of the oxygen bridge, was ensured by using an excess of *m*-CPBA, followed by demethylation and *in situ* intramolecular epoxide opening of the resulting epoxide lactone 24 (Scheme 3, route B). This sequence of transformations led to (\pm)-3 in substantially higher yield (79% from (\pm)-6) than route A. The pathway *via* route B introduces an alternative sequence of transformations to neoabyssomicin B, which is worth investigating biosynthetically.



Conclusions

In conclusion, the first total synthesis of type II abyssomicins was achieved *via* an efficient methodology based on a biomimetic approach involving the IMDA of a suitably substituted butenolide derivative. A facile racemic synthetic route was developed for a proof of concept. Thus, the synthesis of the IMDA precursor (\pm)-7 was based on an efficient coupling of tetronate derivative **19** with aldehyde **18**, prepared *via* a convenient high-yielding barium-assisted Wadsworth–Horner–Emmons reaction to establish the sensitive keto–triene side chain. In contrast to the IMDA of abyssomicin C precursor **5**, providing type I isomer **4** as a single isomer, the cycloaddition of (\pm)-7 gives rise to type II and type I isomers (\pm)-**6** and **21**, respectively, providing an entry to both types of abyssomicin systems. Thus, the key-intermediate (\pm)-**6** led to (\pm)-abyssomicin **2** ((\pm)-**2**), which was prepared in 16 steps from **8** and 6% overall yield. Also, two sequences of transformations from (\pm)-**6** led to (\pm)-neoabyssomicin B ((\pm)-**3**) manoeuvring a Baeyer–Villiger reaction after or before the formation of the oxygen bridge, with the second one being more efficient. Thus, (\pm)-**3** was prepared from **8** in 17 steps and 4.3% overall yield *via* (\pm)-abyssomicin **2** ((\pm)-**2**), as proposed biosynthetically, or in 16 steps and 7.8% overall yield *via* the epoxide–lactone **24**, introducing a possible different biosynthetic pathway. The stereoselectivity of the IMDA reaction and the subsequent transformations to the final products also allow application of this approach to the asymmetric synthesis of **2** and **3** starting from **7**. The information obtained from studying the above systems enriched our knowledge of the exciting world of abyssomicins. The factors that affect the stereochemical outcome of the IMDA reaction in order to control it in one direction and the development of an asymmetric route to these natural products and new abyssomicin systems are under investigation and will be reported in due course.

Author contributions

A. C. methodology, investigation, data curation, validation, writing part of original draft, review, editing, funding acquisition. G. D. A. investigation, validation; V. P., C. P. R. formal analysis and methodology; J. M. H. data curation; V. M. data curation, writing – original draft; A. S. F. investigation, data curation, review and editing; E. A. C. writing part of the original draft, review and editing; V. P. V. conceptualization, project administration, supervision, resources, data curation, visualization, writing – original draft, review and editing.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This research is co-financed by Greece and the European Union (European Social Fund- ESF) through the Operational

Programme “Human Resources Development, Education and Lifelong Learning” in the context of the project “Strengthening Human Resources Research Potential *via* Doctorate Research – 2nd Cycle” (MIS-5000432), implemented by the State Scholarships Foundation (IKY).

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† The structure of this by-product could not be fully elucidated, but it was presumably attributed to the formation of a ring similar to the abyssomicin D system. See ESI† for details.

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