

## PAPER

View Article Online  
View Journal | View Issue



Cite this: *Org. Biomol. Chem.*, 2023, **21**, 768

# Sulfurized diterpenoids in amber as diagenetic indicators of sulfate-reducing processes in past depositional environments†

Lauriane Lenen,<sup>a</sup> Alice Fradet,<sup>a</sup> Philippe Schaeffer,<sup>a</sup> <sup>a</sup> Bernard Gomez<sup>b</sup> and Pierre Adam <sup>a</sup>\*

Two novel compounds isolated from an amber sample from the Santonian of Piolenc (Vaucluse, SE France) were identified using nuclear magnetic resonance and high-resolution mass spectrometry as sulfurized analogues of diterpenic acids from the isopimaric series originating from ancient conifers possibly related to the Cupressaceae family. These two compounds are members of a diterpenoid series corresponding to early diagenetic transformation products of resin diterpenoids. They were likely formed once plant resin comes into contact with reduced sulfur species originating from bacterial sulfate reduction occurring in anaerobic settings such as mangroves or marshes. They represent the first evidence of sulfurization processes affecting plant resin prior to diagenetic transformation into amber. Given their mode of formation, these compounds may be used as indicators of sulfate-reducing processes in past depositional environments.

Received 4th November 2022,  
Accepted 19th December 2022

DOI: 10.1039/d2ob02017c

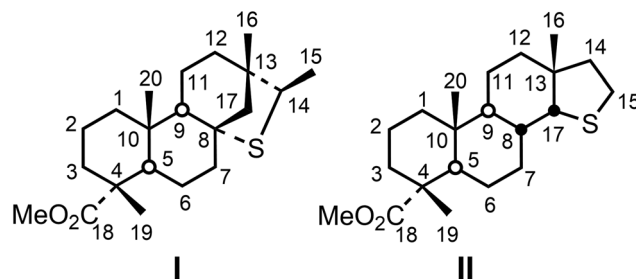
rsc.li/obc

## A. Introduction

Amber results from the fossilization of tree resin. Although the most ancient amber identified is dated back to the Carboniferous period (359–299 Ma),<sup>1</sup> most of the amber was formed much later in the geological record and the vast majority of amber is found in Cretaceous and younger sedimentary rocks.<sup>2,3</sup> The molecular assemblages that constitute amber are generally dominated by terpenic compounds occurring as monomeric entities and polymeric moieties occurring in macromolecular structures.<sup>4</sup> These assemblages are inherited from the plants that have produced the resin at the time of origin of the amber. Therefore, molecular investigation of amber using, for instance, gas chromatography coupled with mass spectrometry (GC-MS) or pyrolysis-GC-MS can furnish valuable information concerning the botanical origin of the amber.<sup>2,4–11</sup> The molecular signature of amber is, however, generally significantly modified compared to that of the genuine resin by various transformation processes during burial (*i.e.*, diagenesis). As the parent compounds are sometimes absent in amber, a correlation between the molecular

signal and a specific biological source is difficult to establish. In this respect, precise identification of molecular biomarkers from amber can provide new molecular tools allowing for a more reliable identification of the botanical sources of amber.<sup>11–16</sup> It can also potentially provide useful information on the palaeoenvironmental conditions prevailing in the ecosystems where the resin-producing trees have lived and on the alteration processes associated with these environments.

In this context, the molecular analysis of an amber sample from the Santonian (86.3–83.6 Ma) of Piolenc (Vaucluse, SE France) using GC-MS revealed the occurrence of an unreported series of compounds characterized by a molecular mass of 350 Da. Isolation of two members of this series (**I** and **II**, Fig. 1) led to their identification as sulfurized analogues of diterpenic acids from the isopimaric series using GC-MS, high resolution



**Fig. 1** Structures of sulfurized diterpenes identified in an amber sample from Piolenc.

<sup>a</sup>Université de Strasbourg, CNRS, Institut de Chimie de Strasbourg UMR 7177, F-67000 Strasbourg, France. E-mail: padam@unistra.fr

<sup>b</sup>Université de Lyon, CNRS, ENS, Laboratoire de Géologie de Lyon, Terre, Planètes, Environnement UMR 5276, F-69622 Villeurbanne, France

† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d2ob02017c>

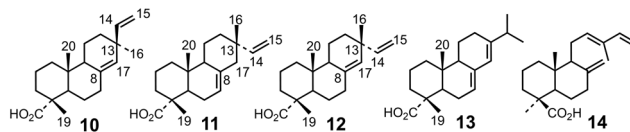


mass spectrometry (HR-MS) and comprehensive nuclear magnetic resonance (NMR) investigations.

## B. Results and discussion

The gas chromatogram of the apolar fraction isolated from the organic extract of an amber sample from Piolenc after methylation of carboxylic acids<sup>16,17</sup> is presented in Fig. 2. The distribution of the lipids is complex and several series of compounds comprising acids from sesquiterpenes (1 and 2) and diterpenes (3–7), as well as sesquiterpenic hydrocarbons (8 and 9), could be detected.

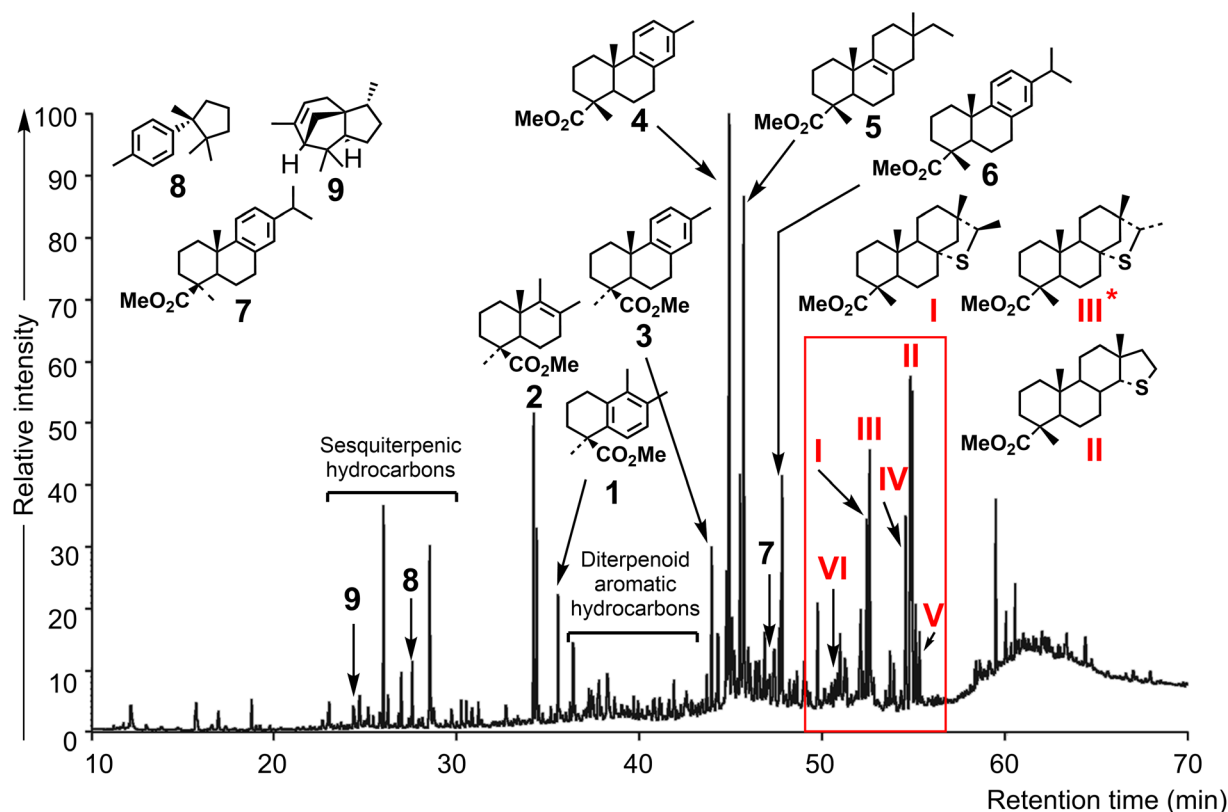
The tricyclic diterpenoid acids comprise, notably, dehydroabietic and callitrisic acids (6 and 7, respectively), and other diterpenic acids (*e.g.* 5) (Fig. 2) related to pimaric, isopimaric and sandaracopimaric acids (10, 11 and 12, Fig. 3). Diterpenoid aromatic hydrocarbons frequently encountered in amber have also been identified in the amber of Piolenc (Fig. 2) based on the comparison of their mass spectra with published spectra.<sup>2,11,14,15</sup> The occurrence of specific molecular markers in the organic extract of the Piolenc amber sample suggests that it must originate from resin exuded by conifers of the Cupressaceae family since such molecules occur in the living representatives of this family. In this



**Fig. 3** Structures of some common resin acids: pimaric acid 10, isopimaric acid 11, sandaracopimaric acid 12, abietic acid 13, and communic acid 14.

respect, callitrisic acid 7 (Fig. 2) has only been reported in the resin of the Cupressaceae genus *Callitris*<sup>18</sup> and is therefore considered a specific marker of this conifer family.<sup>5,19</sup> Similarly, cuparene 8 specifically occurs in the resin of Cupressaceae.<sup>20,21</sup>

The molecular distribution of the Piolenc amber sample also comprises a quantitatively abundant compound series characterized by a molecular weight of 350 Da (Fig. 2). To the best of our knowledge, the occurrence of the latter has only been reported previously twice in amber samples, either as “unknown” or as “unassigned”.<sup>22,23</sup> The electron impact (EI) mass spectra of compounds I–III and three other members of this series are shown in ESI Fig. 1S.† Since it was not possible to obtain more structural information solely based on the interpretation of the EI mass spectra, the isolation of compounds I and II was undertaken to perform detailed NMR



**Fig. 2** Gas chromatogram (GC-MS, EI, 70 eV) of the apolar fraction (FA) isolated from the methylated organic extract of a Piolenc amber sample. The area marked with the red rectangle corresponds to the zone of elution of compounds with a molecular weight of 350 Da (compounds I–VI). \*Tentative identification based on the similarity of the EI mass spectra of compounds I and III (*cf.* ESI Fig. 1S†).



structural investigations. These compounds were isolated using a chromatographic sequence comprising fractionation of the methylated organic extract on a silica gel column followed by further HPLC fractionation of the sub-fractions containing compounds **I** and **II** using normal phase (silica). Compounds **I** and **II** were isolated in sufficient amounts to allow for their identification using detailed NMR studies comprising 1D ( $^1\text{H}$ ,  $^{13}\text{C}$ , DEPT) NMR spectra as well as homo- ( $^1\text{H}$ - $^1\text{H}$ ; COSY, NOESY) and heteronuclear ( $^1\text{H}$ - $^{13}\text{C}$ ; HMBC, HSQC) correlation studies. In addition, the elemental composition of compounds **I** and **II** was determined using HR-MS.

### Identification of compound I

The  $[\text{M} + \text{H}]^+$  ion obtained by HR-MS using positive electrospray ionization (ESI) at  $m/z$  351.3251 corresponds to the molecular formula  $\text{C}_{21}\text{H}_{35}\text{O}_2\text{S}$  (calcd:  $m/z$  351.2352; ESI Fig. 2S<sup>†</sup>), indicating the contribution of a sulfur atom. This result was rather unexpected since organic sulfur compounds have, to the best of our knowledge, never been reported in amber. It should, however, be mentioned that amber samples from some specific geological sites have been shown to contain sulfur, although in low amounts.<sup>24</sup> The molecular formula obtained using HR-MS indicates the occurrence of five degrees of unsaturation as rings or double bonds.

According to the NMR data, it appears from the signal at 179.3 ppm in the  $^{13}\text{C}$ -NMR spectrum (Table 1) that one degree of unsaturation corresponds to the carbonyl group from a methyl ester functionality. This is corroborated by the occurrence of signals at 3.63 ppm and 52.0 ppm in the 1D  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR spectra (ESI Fig. 4S and 5S<sup>†</sup>), respectively, likely corresponding to the methyl from a carbomethoxy group. Compound **I** does not bear any double bond since no signals could be detected in the 110–160 ppm range in the  $^{13}\text{C}$ -NMR spectrum (Table 1). The remaining unsaturations, therefore, correspond to rings.

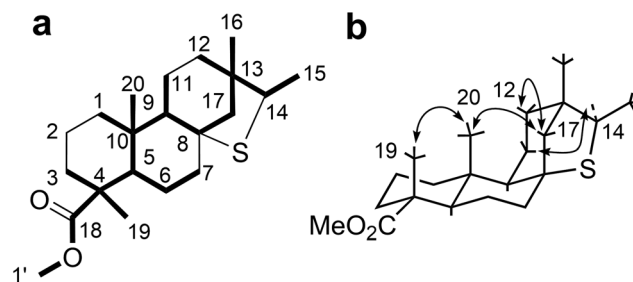
One deshielded proton was observed at 2.96 ppm which could correspond to a proton located on a sulfur-bearing carbon atom. This is supported by the chemical shift of the carbon atom bearing this proton deduced from the HSQC experiment (ESI Fig. 6S<sup>†</sup>): a tertiary C atom at 47.4 ppm. In addition, based on the comparison of the  $^{13}\text{C}$ -NMR and the DEPT spectra, it appears that there is one deshielded quaternary carbon atom with a chemical shift at 58.6 ppm, which indicates that it possibly bears a sulfur atom suggesting that the sulfur atom is incorporated into a thioether bridge between a tertiary and a quaternary carbon atom. This hypothesis was confirmed by the interpretation of the long-range ( $^{2,3}J$ )  $^1\text{H}$ - $^{13}\text{C}$  correlation pattern (HMBC experiment, ESI Fig. 7S<sup>†</sup>), and the carbon atom sequence of compound **I** could finally be established (Fig. 4a).

Connectivities of parts of the structure which could not be correlated based on the long-range ( $^{2,3}J$ )  $^1\text{H}$ - $^{13}\text{C}$  correlations could be established using, notably, the  $^1\text{H}$ - $^1\text{H}$  COSY and NOESY experiments. The structure of compound **I** could thus be related to that of diterpenoids from the pimaric or isopimaric acid series (which differ by the configuration at C-13; cf.

**Table 1**  $^1\text{H}$  (500 MHz) and  $^{13}\text{C}$  (125 MHz) chemical shifts for compounds **I** and **II** in  $\text{CDCl}_3$

Position <sup>a</sup>	<b>I</b>			<b>II</b>		
	$\delta_{\text{C}}$	$\delta_{\text{H}}$	$\delta_{\text{H}}$	$\delta_{\text{C}}$	$\delta_{\text{H}}$	$\delta_{\text{H}}$
1	39.6 <sup>a</sup>	1.79	0.92	38.2	1.69	1.04
2	17.8 <sup>b</sup>	1.58	1.50	18.0	1.51	1.51
3	36.7	1.69	1.51	36.7	1.72	1.51
4	47.6	—	—	47.6	—	—
5	49.9	1.72	—	49.2	1.74	—
6	24.1	1.44	1.08	24.4	1.39	1.05
7	40.1	1.84	1.84	34.9	1.59	1.20
8	58.6	—	—	34.9	1.69	—
9	57.1	1.51	—	47.1	1.38	—
10	39.2	—	—	36.0	—	—
11	19.5	2.14	1.65	20.1	1.49	1.11
12	39.4 <sup>a</sup>	1.39	1.39	30.0	1.60	1.07
13	45.6	—	—	43.2	—	—
14	47.4	2.97	—	45.4	1.81	1.72
15	21.6	1.12	—	27.0	2.73	—
16	24.1	0.96	—	23.2	1.05	—
17	46.9	1.92	1.49	60.8	3.15	—
18	179.3	—	—	179.4	—	—
19	16.4	1.12	—	16.9	1.15	—
20	17.9 <sup>b</sup>	0.99	—	14.6	0.86	—
1'	51.9	3.63	—	51.9	3.62	—

<sup>a</sup> cf. Fig. 1 for carbon numbering; <sup>a,b</sup>: may be interchanged.



**Fig. 4** (a) Carbon sequence (bold lines) established for compound **I** based on the  $^{2,3}J$   $^1\text{H}$ - $^{13}\text{C}$  correlation experiment (HMBC). (b) Main nuclear Overhauser effects (NOESY) observed for compound **I** allowing the correlation between **I** and diterpenoids from the isopimaric acid series to be made.

compounds **10** and **11** in Fig. 3) sulfurized at C-8 and C-14. The relative configuration of the stereocenters and the clear affiliation of compound **I** to tricyclic resin acids of the isopimaric series could be established based on the interpretation of the chemical shift values of specific protons and on the interpretation of the  $^1\text{H}$ - $^1\text{H}$  NOESY correlation pattern (Fig. 4b and ESI Fig. 8S<sup>†</sup>).

Thus, the  $^{13}\text{C}$  chemical shift of the methyl group located at C-4 at 16.4 ppm (Table 1) clearly indicates that this methyl group is axial due to the occurrence of a  $\gamma$ -gauche effect with  $\text{CH}_3$ -20.<sup>25</sup> Thus, the chemical shift of the axial methyl group at C-4 is around 16.8 ppm in the case of tricyclic resin acids such as abietic acid **13** (Fig. 3).<sup>26</sup> In contrast, the  $^{13}\text{C}$  chemical shift of the equatorial methyl groups at C-4 in the case of bicyclic diterpenoids related to communic acid **14** (Fig. 3) is significantly higher (around 28.5 ppm).<sup>16,27,28</sup> The relative configur-



ation at C-4 could be confirmed by the detection of spatial interactions between CH<sub>3</sub>-19 and CH<sub>3</sub>-20 (NOESY experiment, Fig. 4b and ESI Fig. 8S<sup>†</sup>), indicating that both methyl groups are located on the same side of the structure. In addition, the occurrence of nuclear Overhauser effects between one proton at C-17 and CH<sub>3</sub>-20 as well as with one proton of methylene CH<sub>2</sub>-12 (Fig. 4b) unambiguously shows that compound **I** is related to diterpenes of the isopimaric **11** acid series. Finally, all the <sup>1</sup>H and <sup>13</sup>C chemical shifts could be assigned (Table 1).

### Identification of compound **II**

The mass spectrum of compound **II** obtained by GC-MS (EI) shows the same molecular mass, but a very distinct mode of fragmentation (*cf.* ESI Fig. 1bS<sup>†</sup>). The exact mass determined for compound **II** ([M + H]<sup>+</sup>: *m/z* 351.2354) (ESI Fig. 3S<sup>†</sup>) is compatible with the same molecular formula as for compound **I** (C<sub>21</sub>H<sub>35</sub>O<sub>2</sub>S). The signal at 3.62 ppm in the <sup>1</sup>H-NMR spectrum (ESI Fig. 9S<sup>†</sup>) and the signals at 179.4 and 51.9 ppm in the <sup>13</sup>C-NMR spectrum (ESI Fig. 10S<sup>†</sup>) show the presence of a carbomethoxy group (Table 1) as observed for compound **I**. Nevertheless, contrary to compound **I**, the occurrence of three deshielded proton signals at chemical shifts compatible with those of protons located  $\alpha$  to a sulfur atom (one at 3.15 ppm and two others at 2.73 ppm) suggests that compound **II** bears a thioether bridge located between a secondary and a tertiary carbon atom. The detailed investigation of the one-bond (<sup>1</sup>*J*) (ESI Fig. 11S<sup>†</sup>) and the long range (<sup>2,3</sup>*J*) (ESI Fig. 12S<sup>†</sup>) <sup>1</sup>H-<sup>13</sup>C correlations showed that compound **II** corresponds to a sulfurized derivative of pimaric, iso- or sandaracopimaric acid (**10**, **11** and **12**, Fig. 3) with a five-membered ring comprising a sulfur atom located between positions C-15 and C-17 (Fig. 5a). The relative configurations of the chiral centers were determined using a NOESY experiment (Fig. 5b and ESI Fig. 13S<sup>†</sup>).

Nuclear Overhauser effects were observed, notably, between H-20/H-19, H-20/H-8, H-8/H-16, H-17/H-16 and H-8/H-17 (Fig. 5b), indicating that all these protons are on the same side of the molecule. This unambiguously shows that the C/D ring junction is *cis* and that compound **II** is also related to the series of iso- or sandaracopimaric acid (**11** and **12**, respectively; Fig. 3), with the left moiety of the carbon skeleton being iden-

tical to that of compound **I**. Finally, all the <sup>1</sup>H and <sup>13</sup>C chemical shifts could be assigned (Table 1).

### Structures of compounds **III**–**VI**

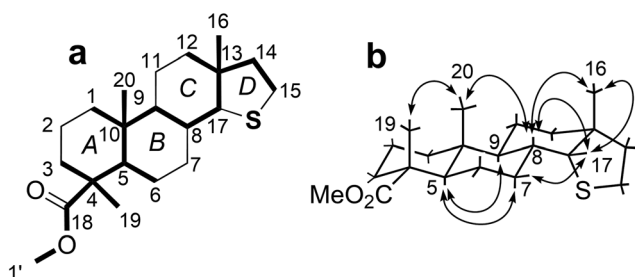
Based on the identification of the structures of compounds **I** and **II**, we propose that the other compounds with a molecular weight of 350 Da (comprising compounds **III**–**VI**) eluting in the range of retention times indicated by the red rectangle in Fig. 2 also correspond to diterpenoid sulfides. These compounds possibly derive from tricyclic diterpenoids from the isopimaric (with the sulfur atom located at other positions than in the case of compounds **I** and **II**), pimaric **10**, abietic **13** or communic **14** acid series. Among these sulfides, compound **III** possibly corresponds to an epimer at C-14 of compound **I** (*cf.* Fig. 2) given the strong similarities between their EI mass spectra (ESI Fig. 1S<sup>†</sup>). Only the retention time in gas chromatography and the relative intensities of the main fragments in the EI mass spectra are different.

### Origin and the mode of formation of compounds **I**–**II**

NMR investigation of the structures of **I** and **II** clearly showed that these compounds correspond to sulfurized diterpenoids related to isopimaric or sandaracopimaric acids **11** and **12**. The occurrence of such compounds in the amber sample from Piolenc is unexpected since the presence of organic sulfur compounds has, to the best of our knowledge, never been reported previously, neither in amber nor in any resin of the extant conifer species. In addition, since we have observed these compounds in amber samples from different geological periods and likely from different botanical origins, the possibility of a direct biosynthetic origin of these compounds is rather improbable. It can therefore be proposed that these compounds may rather correspond to alteration products of isopimaric or sandaracopimaric acids **11** and **12** formed *via* diagenetic sulfurization processes.

Even though organic sulfur compounds have yet not been reported from amber/resin, numerous organic sulfur compounds have been identified in the sedimentary record and their formation has been extensively investigated during the last decades.<sup>29–31</sup> They revealed to be formed by abiotic early diagenetic processes involving functionalized biological lipids and inorganic reduced sulfur species resulting from microbial sulfate reduction in anaerobic environments.<sup>29–31</sup> Carbonyls and double bonds are the most liable functionalities to be sulfurized in such a context.<sup>31–36</sup> In this respect, it appears that isopimaric or sandaracopimaric acids bear double bonds at positions potentially allowing for sulfurization at C-8, C-14, C-15 and C-17 by the addition of H<sub>2</sub>S or polysulfides of microbial origin. The mechanisms illustrated in Fig. 6 can therefore be proposed to account for the formation of compound **I** from isopimaric or sandaracopimaric acids **11** and **12** and that of **II** from sandaracopimaric acid **12**.

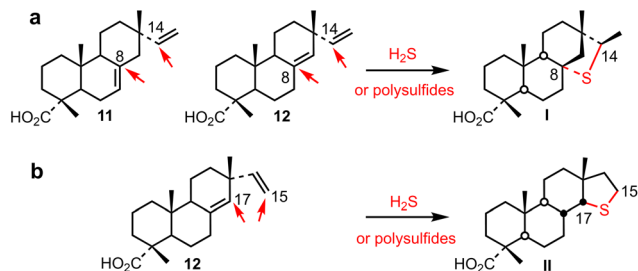
Compounds **I** and **II** present different sites of sulfurization (C-8/C-14 vs. C-15/C-17), indicating a different selectivity in the sulfurization processes. In the case of compound **I**, the sulfur atom is located at the most substituted positions, which con-



**Fig. 5** (a) Carbon sequence (bold lines) established for compound **II** based on the <sup>2,3</sup>*J* <sup>1</sup>H-<sup>13</sup>C correlation experiment (HMBC). (b) Main nuclear Overhauser effects (NOESY) observed for compound **II** allowing the correlation between **II** and diterpenoids from the isopimaric acid series to be made.



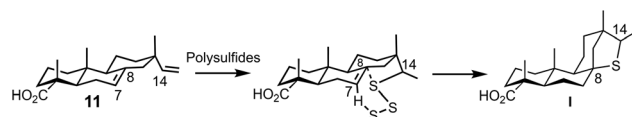




**Fig. 6** Diagenetic pathway of the formation of (a) compound **I** from isopimaric acid **11** or sandaracopimaric acid **12** and (b) compound **II** from sandaracopimaric acid **12** by reduced sulfur species ( $\text{H}_2\text{S}$ , polysulfides) formed by bacterial sulfate reduction in sediments.

trasts with the case of compound **II**, where the sulfur atom is located at the least substituted positions. Therefore, two modes of sulfurization might be considered to account for the formation of **I** and **II**. A radical process involving  $\text{H}_2\text{S}$  and sandaracopimaric acid **12** can be proposed to explain the formation of compound **II** since such a mechanism would result in the sulfurization of the least substituted positions of the double bonds (anti-Markovnikov-type selectivity).<sup>37</sup> Such a mechanism cannot be considered for the formation of compound **I** since the sulfur atom is located at the most substituted positions of potential biological precursors as frequently observed in the case of sedimentary organic sulfur compounds formed by abiotic processes.<sup>32–35</sup> An acid-catalyzed process involving diterpenic acids and  $\text{H}_2\text{S}$  can also be excluded considering the strongly acidic conditions required for such a reaction,<sup>38,39</sup> which are not compatible with the conditions prevailing in natural environmental settings. It can therefore be proposed that sulfurization occurred following a concerted reaction involving polysulfides. Thus, it has been shown that sulfurization of isolated alkenes by polysulfides results in the formation of thiols or polysulfides with the sulfur atom located at the most substituted positions (Markovnikov-type selectivity).<sup>39</sup> In addition, under these conditions, sulfurization of non-conjugated dienes leads to the formation of thianes or thiolanes with the preferential formation of the smallest sulfur-containing rings.<sup>33–35,39</sup> Such a mechanism can be proposed to play a key role in sedimentary sulfurization processes, based on laboratory experiments and on the fact that thiolanes and thianes in sedimentary organic matter are often sulfurized at the most substituted positions.<sup>32–35,39</sup> This mechanism might therefore perfectly account for the formation of compound **I** by sulfurization of isopimaric acid **11** by polysulfides as illustrated in Fig. 7.

This process might comprise first the sulfurization of the vinyl side-chain at the C-14 position due to lower steric hindrance, followed by cyclization at the C-8 position according to a concerted mechanism. The latter would involve a cyclic intermediate, addition of sulfur and hydrogen atoms on the same side of the  $\Delta^7$  double bond and the formation of the smallest ring as described previously in the case of other substrates.<sup>33,35</sup>



**Fig. 7** Possible mechanism of sulfurization of isopimaric acid **11** with polysulfides leading to compound **I**.

### Sulfurized diterpenoids in amber: geochemical implications

The sulfurized diterpenoids identified in the Piolenc amber sample were possibly formed after the deposition of the resin from Cupressaceae conifer shrubs or trees within sediments. Since the formation of these organo-sulfur compounds required the presence of reduced sulfur species produced by bacterial sulfate reduction, it can be anticipated that the conditions which prevailed in the sediments at the time of deposition were oxygen-depleted, as is the case in environments such as marshes or mangroves. Thus, in such organic-rich environments dominated by plant inputs, the conditions are favorable for the development of anaerobic bacteria, including sulfate-reducing bacteria which will produce  $\text{H}_2\text{S}$  or polysulfides able to react with organic compounds. It is unlikely that the sulfurization of the diterpenoids occurred when the resin had already been transformed into amber since amber is a solid material which probably prevents sulfur species from diffusing into it. Therefore, sulfurization most likely occurred at an early stage of burial when the resin is sufficiently fluid to allow the diffusion of reduced sulfur species into the resin drops and their reaction with diterpenoids prior to further diagenetic transformation into amber upon deeper burial.

## C. Conclusions

Two fossil diterpenoids from a Cretaceous amber sample located in the city of Piolenc (Vaucluse, SE France) were identified as sulfides of diterpenic acids using GC-MS, HR-MS and detailed NMR investigations. These compounds are related to isopimaric and sandaracopimaric acids which can be predominant constituents of conifer resins. A possible diagenetic pathway for their formation is proposed and it involves the sulfurization of isopimaric and sandaracopimaric acids by reduced sulfur species ( $\text{H}_2\text{S}$  or polysulfides) originating from bacterial sulfate reduction following two different mechanisms. Sulfurization processes leading to the formation of **I** and **II** will be further investigated using laboratory experiments involving resin acids and reduced sulfur species under geochemically relevant conditions. It is likely that these compounds formed at the earliest stages of diagenesis, when the fresh conifer resin entered a sedimentary setting depleted of oxygen (e.g., marsh, mangrove), and came into contact with the reduced sulfur species generated by sulfate-reducing bacteria. Therefore, these novel organo-sulfur compounds represent the first evidence of sulfuriza-



tion processes affecting plant resin prior to its diagenetic transformation into amber. Thus, these compounds may be used as indicators of sulfate-reducing processes in past depositional environments. In this respect, it should be mentioned that compounds **I–II** and related structures with the same molecular weight have also been detected in variable amounts, but not systematically, in other amber samples from different origins or ages. Investigation of their presence and significance in terms of palaeoenvironments of deposition is currently underway.

## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

We would like to thank Estelle Motsch and Stéphanie Coutin (University of Strasbourg) for the mass spectrometry analyses, and Maurice Coppe (University of Strasbourg) for the NMR analyses. The “Centre National de la Recherche Scientifique” and the “Université de Strasbourg” financially supported this research.

## References

- 1 P. S. Bray and K. B. Anderson, *Science*, 2009, **326**, 132–134.
- 2 Y. A. Nohra, V. Perrichot, L. Jeanneau, L. Le Pollès and D. Azar, *J. Nat. Prod.*, 2015, **78**, 1284–1293.
- 3 G. Pastorelli, *Archaeological Baltic amber: Degradation mechanisms and conservation measures*, PhD thesis, Università di Bologna, 2009.
- 4 J. B. Lambert, J. A. Santiago-Blay and K. B. Anderson, *Angew. Chem., Int. Ed.*, 2008, **47**, 9608–9616.
- 5 K. B. Anderson, *Geochem. Trans.*, 2006, **7**, 2, DOI: [10.1186/1467-4866-7-2](#).
- 6 O. O. Sonibare, T. Hoffmann and S. F. Foley, *Org. Geochem.*, 2012, **51**, 55–62.
- 7 R. Pereira, I. S. Carvalho, B. R. T. Simoneit and D. A. Azevedo, *Org. Geochem.*, 2009, **40**, 863–875.
- 8 J. S. Mills, R. White and L. J. Gough, *Chem. Geol.*, 1984, **47**, 15–39.
- 9 B. R. T. Simoneit, P. G. Hatcher and A. Nissenbaum, *Org. Geochem.*, 1988, **13**, 677–690.
- 10 O. O. Sonibare, R. J. Huang, D. E. Jacob, Y. Nie, E. Kleine-Benne, T. Hoffmann and S. F. Foley, *J. Anal. Appl. Pyrol.*, 2014, **105**, 100–107.
- 11 C. Menor-Salvan, B. R. T. Simoneit, M. Ruiz-Bermejo and J. Alonso, *Org. Geochem.*, 2016, **93**, 7–21.
- 12 Y. Wang, W. Jiang, Q. Feng, H. Lu, Y. Zhou, J. Liao, Q. Wang and G. Sheng, *Org. Geochem.*, 2017, **113**, 90–96.
- 13 J. Jossang, H. Bel-Kassaoui, A. Jossang, M. Seuleiman and A. Nel, *J. Org. Chem.*, 2008, **73**, 412–417.
- 14 K. Kimura, Y. Minamikawa, Y. Ogasawara, Y. Yoshida, K. Saitoh, H. Shinden, Q. Y. Yue, S. Takahashi, T. Miyakawa and H. Koshino, *Fitoterapia*, 2012, **83**, 907–912.
- 15 T. Kawamura, H. Koshino, T. Nakamura, Y. Nagasawa, H. Nanao, M. Shirai, S. Uesugi, M. Ohno and K. Kimura, *Org. Geochem.*, 2018, **120**, 12–18.
- 16 N. de Lama Valderrama, P. Schaeffer, A. Leprince, S. Schmitt and P. Adam, *Org. Geochem.*, 2022, **167**, 104372.
- 17 H. Brechbühler, H. Büchi, E. Hatz, J. Schreiber and A. Eschenmoser, *Angew. Chem., Int. Ed. Engl.*, 1963, **2**, 212–213.
- 18 B. R. T. Simoneit, R. E. Cox, D. R. Oros and A. Otto, *Molecules*, 2018, **23**, 3384, DOI: [10.3390/molecules23123384](#).
- 19 K. Knight, P. Bingham, D. Gimaldi, K. Anderson, R. Lewis and C. Savrda, *Cretaceous Res.*, 2010, **31**, 85–93.
- 20 A. Otto and V. Wilde, *Bot. Rev.*, 2001, **67**, 141–238.
- 21 P. J. Grantham and A. G. Douglas, *Geochim. Cosmochim. Acta*, 1980, **44**, 1801–1810.
- 22 T. C. Fischer, O. O. Sonibare, B. Aschauer, E. Kleine-Benne, P. Braun and B. Meller, *Palaeontology*, 2017, **60**, 743–759.
- 23 E. C. Stout, C. W. Beck and K. B. Anderson, *Phys. Chem. Miner.*, 2000, **27**, 665–678.
- 24 F. Riquelme, P. Northrup, J. L. Ruvalbaca-Sil, V. Stajonoff, D. P. Siddons and J. Alvarado-Ortega, *Appl. Phys. A*, 2014, **116**, 97–109.
- 25 H. Beierbeck, J. K. Saunders and J. W. Apsimon, *Can. J. Chem.*, 1977, **55**, 2813–2828.
- 26 B. Gigante, L. Santos and M. J. Marcelo-Curto, *Magn. Reson. Chem.*, 1995, **33**, 318–321.
- 27 J. M. Fang, Y. C. Sou, Y. H. Chiu and Y. S. Cheng, *Phytochemistry*, 1993, **34**, 1581–1584.
- 28 D. Zinkel and W. B. Clarke, *Phytochemistry*, 1985, **24**, 1267–1271.
- 29 J. S. Sinninghe Damsté and J. W. de Leeuw, *Org. Geochem.*, 1990, **16**, 1077–1101.
- 30 P. Schaeffer, C. Reiss and P. Albrecht, *Org. Geochem.*, 1995, **23**, 567–581.
- 31 P. Adam, P. Schneckeburger, P. Schaeffer and P. Albrecht, *Geochim. Cosmochim. Acta*, 2000, **64**, 3485–3503.
- 32 A. Behrens, P. Schaeffer and P. Albrecht, *Tetrahedron Lett.*, 1997, **38**, 4921–4924.
- 33 J. Poinot, P. Schneckeburger, P. Adam, P. Schaeffer, J. Trendel and P. Albrecht, *J. Chem. Soc., Chem. Commun.*, 1997, 2191–2192.
- 34 S. Gug, P. Schaeffer, P. Adam, S. Klein, E. Motsch and P. Albrecht, *Org. Geochem.*, 2009, **40**, 876–884.
- 35 P. Adam, P. Schaeffer, S. Gug, E. Motsch and P. Albrecht, *Org. Geochem.*, 2009, **40**, 885–894.



- 36 Y. Hebting, P. Schaeffer, A. Behrens, P. Adam, G. Schmitt, P. Schneckenburger, S. Bernasconi and P. Albrecht, *Science*, 2006, **312**, 1627–1631.
- 37 F. Dénès, M. Pichowicz, G. Povie and P. Renaud, *Chem. Rev.*, 2014, **114**, 2587–2693.
- 38 W. C. van Zijll Langhout and H. I. Waterman, *J. Appl. Chem.*, 1954, **4**, 285–288.
- 39 W. De Graaf, J. S. Sinninghe Damsté and J. W. de Leeuw, *J. Chem. Soc., Perkin Trans. 1*, 1995, 635–640.

