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Polymeric architecture as a tool for controlling the reactivity of palladium(II) loaded nanoreactors†

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Self-assembled systems, like polymeric micelles, have become great facilitators for conducting organic reactions in aqueous media due to their broad potential applications in green chemistry and biomedical applications. Massive strides have been taken to improve the reaction scope of such systems, enabling them to perform bioorthogonal reactions for prodrug therapy. Considering these significant advancements, we sought to study the relationships between the architecture of the amphiphiles and the reactivity of their Pd^{II} loaded micellar nanoreactors in conducting depropargylation reactions. Towards this goal, we designed and synthesized a series of isomeric polyethylene glycol (PEG)-dendron amphiphiles with different dendritic architectures but with an identical degree of hydrophobicity and hydrophilic to lipophilic balance (HLB). We observed that the dendritic architecture, which serves as the main binding site for the Pd^{II} ions, has greater influence on the reactivity than the hydrophobicity of the dendron. These trends remained constant for two different propargyl caged substrates, validating the obtained results. Density functional theory (DFT) calculations of simplified models of the dendritic blocks revealed the different binding modes of the various dendritic architectures to Pd^{II} ions, which could explain the observed differences in the reactivity of the nanoreactors with different dendritic architectures. Our results demonstrate how tuning the internal architecture of the amphiphiles by changing the orientation of the chelating moieties can be used as a tool for controlling the reactivity of Pd^{II} loaded nanoreactors.

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1. Introduction

Despite the vital role of water as a solvent in nature in general and especially in living systems, it did not make its way into mainstream organic chemistry. Even today, most organic synthesis is carried out in dry organic solvents since many of the chemicals and reagents used are sparingly soluble in water. Amphiphilic polymers that self-assemble in water, where the hydrophobic portions collapse together because of the hydrophobic effect, can help bridge this gap between organic and aqueous phase chemistry.¹ Polymeric assemblies, including micelles and vesicles, have gained increasing attention due to

their potential to serve as nanoreactors for conducting organic reactions in aqueous media.² These self-assembled structures have sizes in the nanometre range. They can host most hydrophobic substrates, effectively reducing the reaction volume and leading to a high local concentration of the substrate molecules. This phenomenon can eventually help in carrying out reactions in relatively milder conditions than conducting the corresponding reaction in an organic environment.^{3–5} Additional advantages of utilizing these systems include the reduced consumption of volatile organic solvents and reduced chemical load on the environment.

Although the development of nanoreactors is in its emergent state, significant contributions have already been made, especially in facilitating organometallic and organocatalytic reactions using self-assembled polymeric amphiphiles as reported by Meijer,^{6–8} Lipshutz,^{5,9–11} Weck,^{12–14} O'Reilly,^{15–18} Zimmerman,^{19–21} and many others.^{22–26} Several studies have focused on understanding the effect of different ligands,^{6–8,19,24,27} amphiphile's and substrate's hydrophobicity,^{7,26} and the type of co-surfactants,^{9,10} on the range and the type of the reactions that can be conducted in aqueous environment. In addition to these parameters, it is also important to further understand how the architecture of the amphiphiles and the resulting positioning and number of chelating moieties can affect the reactivity of the

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self-assembled nanoreactors. Hence, we decided to systematically tune the position of the chelating moieties and the number of end-groups in the dendritic part of the amphiphile while maintaining their overall molecular weight and hydrophilic to lipophilic balance (HLB).¹⁰ By using this approach, we aimed to isolate the effect of the dendritic architecture on reaction kinetics while minimizing the influence of other parameters. With this in mind, we synthesized a library of PEG-dendron amphiphiles with three, four, and six thioether containing end-groups in the dendritic part of the amphiphile (Fig. 1). The utilization of a dendritic structure as the lipophilic block allowed us to precisely tune the architecture and the lipophilicity of the dendritic block. This high molecular precision enabled us to synthesize amphiphiles with nearly identical molecular weights and HLB. In this study we used palladium-mediated *O*-propargyl cleavage^{27–30} as a model reaction due to its potential application as bioorthogonal uncaging approach for the activation of dyes and prodrugs.^{31–35} The suggested mechanisms of this palladium catalysed uncaging of propargylic substrates such as dyes and cytotoxic drugs has been recently reviewed by Domingos.³⁶

To evaluate the impact of the type of protecting group on the reaction rate, we synthesized two low molecular weight substrates containing different propargyl protecting groups (propargyl ether and propargyl carbamate). These substrates were used as model

prodrugs for conducting depropargylation reactions using Pd^{II}-loaded micellar nanoreactors with different dendritic architectures. This approach enabled us to investigate how the nano environment inside the micellar core, and the type of propargyl protecting group affect the reaction rates.

2. Results and discussion

Our amphiphiles were constructed from a commercial 5 kDa methoxy polyethylene glycol (mPEG_{5k}) as the hydrophilic block, and dendron as the lipophilic block. The dendritic architecture, and the number and length of the end-groups were varied to maintain the overall molecular weight and HLB of the amphiphiles to help us isolate the effect of their architecture on the kinetics.

2.1. Tuning the number of end-groups in dendron

Synthesis and characterization of amphiphiles. Three amphiphiles were synthesized: mPEG_{5k}-D-(C14)₃ (C14×3) (calculated $M_n = 6081$ Da), mPEG_{5k}-D-(C11)₄, (3,5) (C11×4 (3,5)) (calculated $M_n = 6084$ Da) and mPEG_{5k}-D-(C6)₆ (C6×6) (calculated $M_n = 6094$ Da). The C14×3 amphiphile was synthesized by coupling mPEG_{5k}-NH₂ to an activated 4-nitrophenol ester of 3,4,5-tris(allyloxy)benzoic acid³⁷ to get mPEG_{5k}-triene followed by a thiol-ene reaction using 1-tetradecanethiol to obtain the amphiphile. For making the four-armed (3,5) amphiphile, C11×4 (3,5), mPEG_{5k}-NH₂ was coupled with an activated 4-nitrophenol ester of 3,5-bis(prop-2-yn-1-yloxy) benzoic acid³⁷ to get mPEG_{5k}-di-yne (3,5), followed by thiol-yne reaction with 1-undecanethiol. To make the C6×6 amphiphile, first the complete C6×6 dendron was synthesized by propargylating gallic acid,³⁷ which was then coupled to 1-hexanethiol using thiol-yne chemistry. Finally, the C6×6 dendron was conjugated to mPEG_{5k}-NH₂ to get the final amphiphile (Scheme 1). ¹H-NMR, High-Performance Liquid Chromatography (HPLC), and Gel Permeation Chromatography (GPC) were used to verify the conversion, product's purity, molecular weight and polydispersity, respectively. All experimental results showed excellent correlations with the expected values, as can be seen in the ESI.†

After synthesizing the amphiphiles, their self-assembly into micelles in aqueous media (phosphate buffered saline-PBS, pH 7.4, 37 °C) was examined by measuring their critical micelle concentration (CMC) values using the Nile red method.³⁸ The CMC values of all three amphiphiles were nearly identical (Table 1 and Fig. S20, S21 and S23†). Next, we used dynamic light scattering (DLS) to estimate the size of the micelles in the presence and absence of Pd^{II} salt. To prepare the metal loaded micelles, the Pd^{II} salt and the different amphiphiles were dissolved separately in acetone, mixed and stirred briefly, followed by the evaporation of acetone to form a thin film, which was rehydrated in PBS to yield aqueous micellar solutions ([amphiphile] = 320 μM). The diameters of the micelles measured by DLS were found to be around 20 nm (for the empty micelles), while slightly smaller diameters were

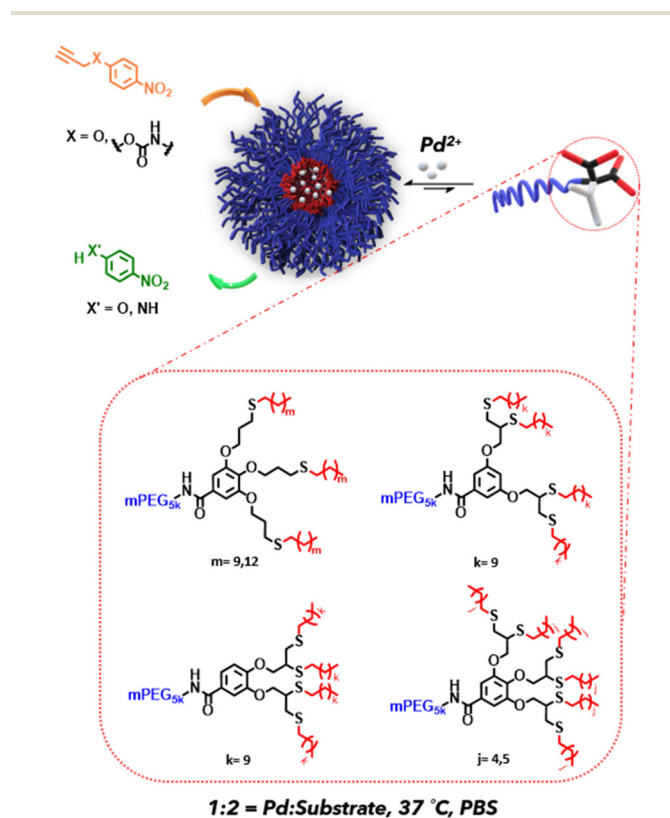
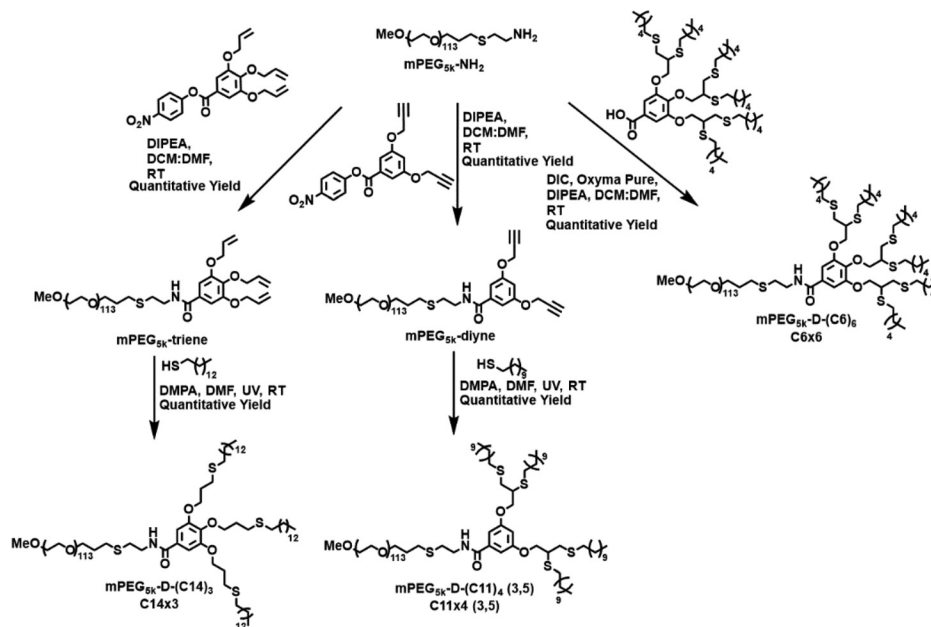


Fig. 1 Schematic illustration of Pd^{II}-loaded micellar nanoreactors based on PEG-dendron amphiphiles with varied architectures for the depropargylation of substrates with different propargyl protecting groups.





Scheme 1 Synthetic route for C14x3, C11x4 (3,5) and C6x6 amphiphiles.

Table 1 Molecular properties of the C14x3, C11x4 (3,5) and C6x6 amphiphiles and their micellar assemblies

| Hybrid | Name | End-group | M_n^a (kDa) | M_n^b (kDa) | D^b | CMC ^c (μ M) | D_h^d (nm) [Pd] = 0 μ M | D_h^e (nm) [Pd] = 75 μ M | D_h^f (nm) [Pd] = 150 μ M |
|--|-------------|---|---------------|---------------|-------|-----------------------------|----------------------------------|-----------------------------------|------------------------------------|
| mPEG _{5k} -D-(C14) ₃ | C14x3 | <i>n</i> -C ₁₄ H ₂₉ | 6.1 | 6.3 | 1.03 | 4 ± 1 | 24 ± 4 | 22 ± 2 | 20 ± 7 |
| mPEG _{5k} -D-(C11) ₄ (3,5) | C11x4 (3,5) | <i>n</i> -C ₁₁ H ₂₃ | 6.1 | 6.3 | 1.04 | 4 ± 1 | 17 ± 1 | 15 ± 1 | 15 ± 2 |
| mPEG _{5k} -D-(C6) ₆ | C6x6 | <i>n</i> -C ₆ H ₁₃ | 6.1 | 6.0 | 1.04 | 3 ± 1 | 20 ± 1 | 16 ± 1 | 16 ± 2 |

^a Calculated based on a 5 kDa mPEG and the expected exact mass of the dendrons. ^b Measured by GPC using PEG commercial standards. ^c Determined using the Nile red method. ^d Hydrodynamic diameter measured by DLS (% volume) of micelles formed from micelles only ([amphiphile] = 320 μ M). ^e Hydrodynamic diameter measured by DLS (% volume) of micelles ([amphiphile] = 320 μ M) with the encapsulated Pd(OAc)₂ salt [75 μ M]. ^f Hydrodynamic diameter measured by DLS (% volume) of micelles ([amphiphile] = 320 μ M) with the encapsulated Pd(OAc)₂ salt [150 μ M].

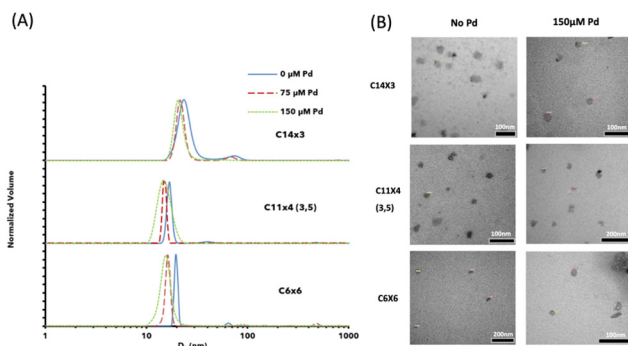


Fig. 2 (A) DLS measurements of micelles with different dendritic architectures ([amphiphile] = 320 μ M) without encapsulated Pd(OAc)₂ (blue solid lines), micelles with encapsulated Pd^{II} ([Pd(OAc)₂] = 75 μ M) (red large-dashed lines) and micelles with encapsulated Pd^{II} ([Pd(OAc)₂] = 150 μ M) (green dashed lines) in PBS (pH = 7.4) at 37 °C. (B) TEM images of the micelles based on the C14x3, C11x4 (3,5) and C6x6 amphiphiles, with (right column) and without (left column) Pd(OAc)₂ salt.

observed for Pd^{II}-loaded micelles (Table 1 and Fig. 2). Transmission electron microscopy (TEM) images of empty, and Pd^{II} loaded micelles further confirmed the presence of nano-sized spherical particles (Fig. 2 and Fig. S31[†]), which correlates with the data that was obtained from DLS.

The two substrates *para*-nitrophenyl propargyl ether (PNPPE)²⁶ and *N*-*para*-nitrophenyl *O*-propargyl carbamate (PNACAPE)³⁹ with propargyl ether and propargyl carbamate protecting groups, respectively, were synthesized as previously reported. The depropargylation reactions were conducted by adding the substrate, PNPPE (300 μ M) and PNACAPE (150 μ M) to micellar solutions containing 150 μ M and 75 μ M Pd^{II} respectively. The ratio of Pd^{II} to substrate was 1 : 2 in both cases. The reaction progress at 37 °C was monitored by HPLC (Fig. 3, and Fig. S34–S37, S40 and S41[†]).

To evaluate the rate of depropargylation, the area under the curve (AUC) of the substrate's peak in each chromatogram was measured (Fig. 3A) and the decrease in AUC (in %) was plotted as a function of time (Fig. 3B and C). To assess the reaction



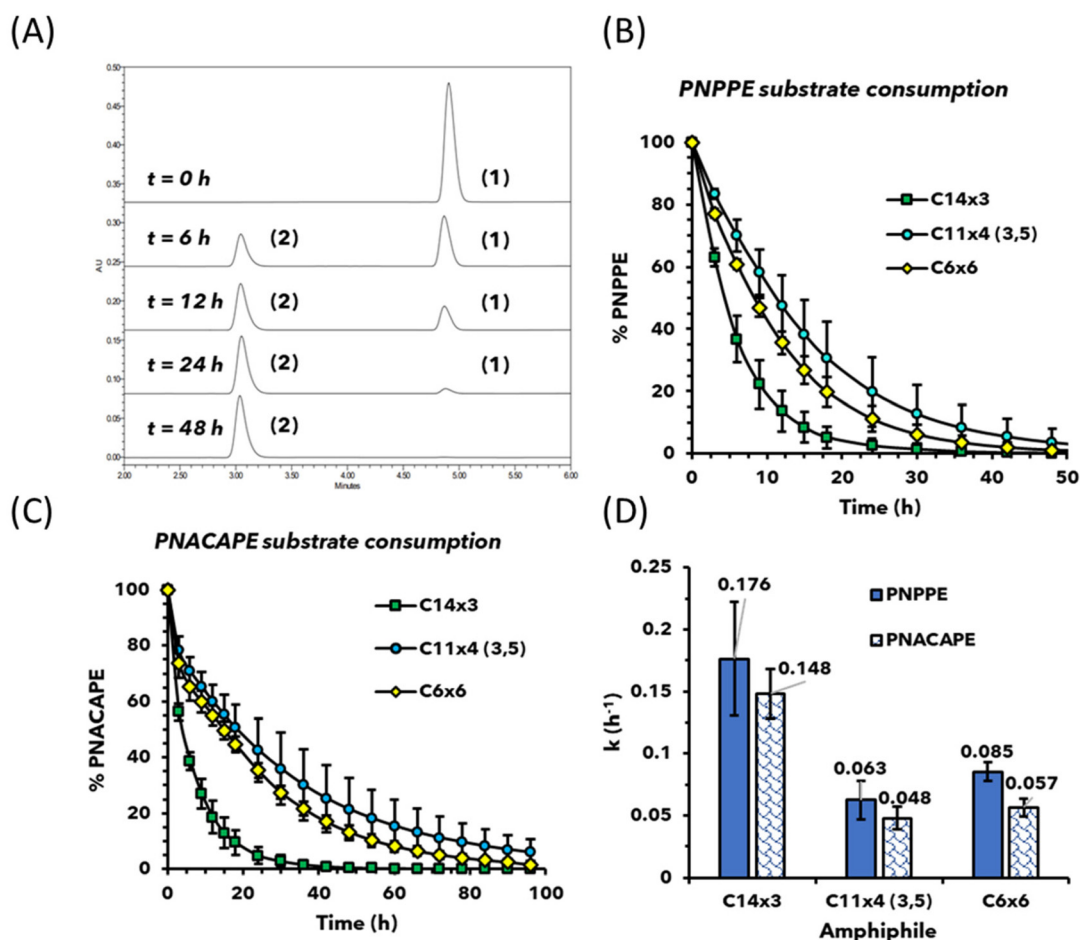


Fig. 3 (A) Representative HPLC chromatogram overlay (taken at 307 nm), showing the transformation of PNPPE (1) to *para*-nitrophenol (PNP) (2) using Pd^{II} loaded C14x3 amphiphilic micelles ([Amphiphile] = 320 μ M, [PNPPE] = 300 μ M, [Pd(OAc)₂] = 150 μ M, [Pd(OAc)₂:Substrate] = 1:2). Normalized substrate consumption of (B) PNPPE and (C) PNACAPE over time in the presence of C14x3 (square, green), C11x4 (3,5) (circle, deep sky blue) and C6x6 (diamond, yellow) metallic micelles. (D) Rate constants of PNPPE (blue, solid fill) and PNACAPE (blue, shingle pattern) depropargylation in presence of C14x3, C11x4 (3,5) and C6x6 metal loaded micelles.

kinetics, a natural log of the experimental data was plotted against time which provided a linear equation correlating with a pseudo first-order reaction $\ln[A] = -kt + \ln[A]_0$. The rate constant (k) values were calculated from the slope of the kinetic data, and the theoretical half-life ($t_{1/2}$) values were calculated using the equation $t_{1/2} = \ln(2)/k$. The rate constant k and calculated values of $t_{1/2}$ are presented alongside the experimental value of $t_{1/2}$ in Table S1.†

The kinetic results showed that micelles made with C14x3 amphiphile were two and a half times faster in performing depropargylation than micelles made from C6x6 and C11x4 (3,5) amphiphiles, while the C11x4 (3,5) being the slowest of the three (Fig. 3B and C). The difference in depropargylation rates between C6x6 and C11x4 (3,5) amphiphiles was small compared to C14x3 (Fig. 3D).

Importantly, the kinetic trends among the three amphiphiles remained the same for both PNPPE and PNACAPE substrates, although the depropargylation of the latter was slower for all three micellar nanoreactors.

ICP-MS measurements were conducted to ensure that all the micellar nanoreactors had similar Pd^{II} concentrations for a particular substrate and that the obtained trends resulted solely from the architectural changes in the amphiphiles (Fig. S45 and S46†).

2.2. The effect of the length of the end-group on the kinetics of depropargylation

Next, we aimed to investigate if the number of carbon atoms in the end-groups affected the kinetic results. Towards this, mPEG_{5k}-D-(C7)₆, *i.e.*, C7x6, was synthesized and compared with C14x3 to assess their ability to conduct the depropargylation reactions, since both C7x6 and C14x3 amphiphiles have a total of forty-two carbon atoms in their end-groups. The C7x6 amphiphile was synthesized using the same method employed for the C6x6 amphiphile by preparing the dendron followed by coupling it to mPEG_{5k}-NH₂ (Fig. S10†). As seen in Tables 1 and 2, C7x6 and C14x3 based micelles had nearly identical CMCs and diameters (4 μ M and ~20 nm respectively).



Table 2 Molecular properties of the C7×6, C11×3, and C11×4 (3,4) amphiphiles and their micellar assemblies

| Hybrid | Name | End-group | M_n^a (kDa) | M_n^b (kDa) | D^b | CMC ^c (μM) | D_h^d (nm) [Pd] = 0 μM | D_h^e (nm) [Pd] = 75 μM | D_h^f (nm) [Pd] = 150 μM |
|--|--------------------|---|---------------|---------------|-------|-----------------------|-----------------------------|------------------------------|-------------------------------|
| mPEG _{5k} -D-(C7) ₆ | C7×6 | <i>n</i> -C ₇ H ₁₅ | 6.2 | 5.7 | 1.07 | 4 ± 1 | 21 ± 2 | 20 ± 1 | 19 ± 3 |
| mPEG _{5k} -D-(C11) ₃ | C11×3 | <i>n</i> -C ₁₁ H ₂₃ | 6.0 | 5.7 | 1.04 | 5 ± 1 | 15 ± 1 | 15 ± 1 | 15 ± 1 |
| mPEG _{5k} -D-(C11) ₄ (3,4) | C11×4 (3,4) | <i>n</i> -C ₁₁ H ₂₃ | 6.1 | 6.6 | 1.12 | 6 ± 1 | 20 ± 1 | 20 ± 1 | 18 ± 2 |

^a Molecular weight calculated based on a 5 kDa mPEG and the expected exact mass of the dendrons. ^b Molecular weight measured by GPC using PEG commercial standards. ^c Determined using the Nile red method. ^d Hydrodynamic diameter measured by DLS (% volume) of micelles formed from micelles only ([amphiphile] = 320 μM). ^e Hydrodynamic diameter measured by DLS (% volume) of micelles ([amphiphile] = 320 μM) with the encapsulated Pd(OAc)₂ salt [75 μM]. ^f Hydrodynamic diameter measured by DLS (% volume) of micelles ([amphiphile] = 320 μM) with the encapsulated Pd(OAc)₂ salt [150 μM].

Despite having the same number of carbon atoms in the end-groups, C14×3 based micelles were almost twice as fast as compared to C7×6 based micelles at depropargylating both the substrates (Fig. 4). Additionally, the C7×6 based micelles were approximately 10–15% faster than micelles of C6×6. This result is in good correlation with our previous work, which showed that increase in the length of the end-group for a particular architecture leads to faster depropargylation.²⁶ This highlighted that the architecture had greater influence on the kinetics than the length of the end-groups and the hydrophobicity of the dendron.

Following this comparison, we synthesized an additional amphiphile, mPEG_{5k}-D-(C11)₃ (C11×3), and conducted depropargylation experiments using its micelles, which were then compared with the C11×4 (3,5) based micelles. The C11×3 amphiphile was synthesized using the same method employed for making the C14×3 amphiphile, using a thiol–ene coupling reaction of mPEG_{5k}-triene with 1-undecanethiol to obtain the final amphiphile (Fig. S1†). As seen Fig. 4, it was observed that C11×3 based micelles were twice as fast as C11×4 (3,5) in depropargylating both the substrates. The overall rates obtained for C11×3 were closer to C14×3 than to C11×4 (3,5), reinforcing the hypothesis that the architecture had a greater influence on the observed kinetics than the length of the end-groups. In this case as well, the depropargylation rates of C11×3 based micelles were approximately 25% slower than those obtained for C14×3 based micelles, which aligns well

with our current and previous findings on the contribution of end-group hydrophobicity for the same polymeric architecture.²⁶

2.3. Changing the position of the end-groups

To understand the influence of the position of the branches of the dendron on the kinetics, a different isomer of the four-armed amphiphile was synthesized with the 1,2 dimercaptoether containing end-groups in the third and the fourth position, yielding the mPEG_{5k}-D-(C11)₄ (3,4) amphiphile, *i.e.*, C11×4 (3,4). The synthetic procedure was similar to the C6×6 amphiphile where the C11×4 (3,4) dendron was first synthesized and then subsequently coupled to mPEG_{5k}-NH₂ to obtain the final C11×4 (3,4) amphiphile (Fig. S6†). As observed in Fig. 5, the C11×4 (3,4) based micelles exhibited the fastest kinetics among all the polymers studied. This result was unexpected as the C6×6 amphiphile, has two of its three 1,2 dimercaptoether moieties located at the third and the fourth position of the aromatic branching unit, similar in their substitution pattern to the two 1,2 dimercaptoether groups of the C11×4 (3,4).

2.4. Anisotropy measurements

To understand the reason for the observed trends in kinetic rates, we aimed to investigate whether the difference in mobility of guest molecules inside the micelles can be correlated

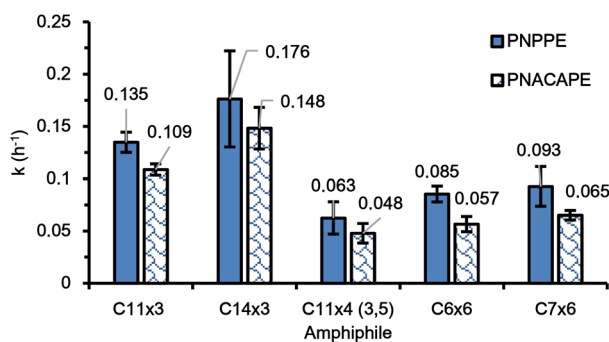


Fig. 4 Rate constants of PNPPE (blue, solid fill) and PNCAPE (blue, shingle pattern) depropargylation in presence of C11×3, C14×3, C11×4 (3,5), C6×6 and C7×6 metallic micelles.

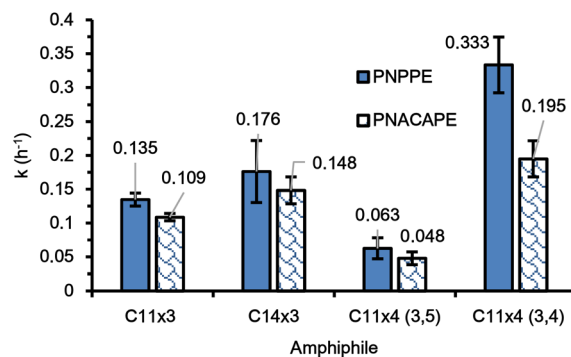


Fig. 5 Rate constants of PNPPE (blue, solid fill) and PNCAPE (blue, shingle pattern) depropargylation in presence of C11×3, C14×3, C11×4 (3,5) and C11×4 (3,4) metallic micelles.



with the difference in depropargylation rates for amphiphiles with different architectures. The mobility of a fluorescent molecule inside the micelles could provide a proxy for the freedom of movement of a substrate molecule inside the core thus ultimately affecting the rates of depropargylation. To calculate the anisotropy factors, we encapsulated Nile Red dye ([Dye] = 21 nM, [amphiphile] = 320 μ M), along with various concentrations of Pd(OAc)₂ ([Pd] = 0, 75, 150 μ M). The

measurements were conducted by placing light polarizers at the source and the detector in different combinations of vertical and horizontal orientations (section 4.3 in ESI†).⁴⁰

The results showed an increase in the anisotropy factor with increasing concentration of Pd(OAc)₂ but the value remained the same for all micelles at a given concentration of Pd(OAc)₂ with a slight variation of around 5% (Fig. 6). Thus, it was concluded that the Nile Red had approximately the same mobility in the core of the micelles irrespective of their amphiphile's architecture, suggesting that mobility inside the core is not the influence behind the observed trends in the depropargylation kinetics.

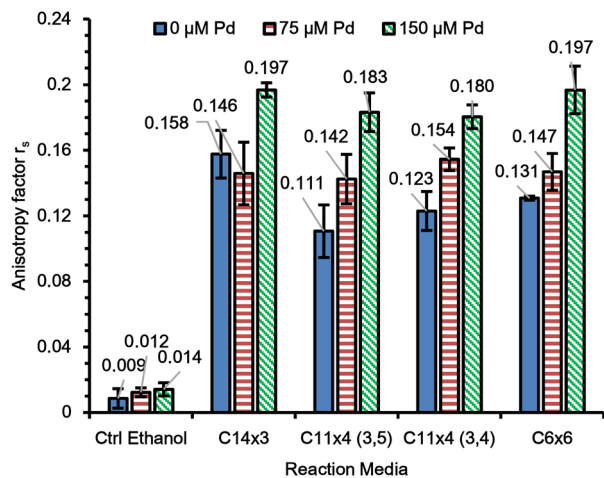


Fig. 6 Anisotropy factor of Nile red dye in amphiphilic micellar environment ([Amphiphile] = 320 μ M) at different concentrations of encapsulated palladium acetate. [Pd^{II}] = 0 μ M (blue, solid fill), 75 μ M (orange, horizontal stripes), 150 μ M (green, downward stripes).

2.5. DFT computed binding modes of thioethers/Pd^{II} complexes in different dendrons

To better understand the observed kinetic trends, DFT calculations were carried out using the BP86-D3 method,^{41,42} with Ahlrichs' def2-SVP basis set,⁴³ and with the relativistic effect of palladium that was accounted for by the Stuttgart–Dresden (SDD) effective core potential (ECP).^{44,45} Before DFT measurements, we simplified our ligand/Pd^{II} structures based on the results from our previous NMR titration experiments with PEG-dendron amphiphile (3,5-based architecture) and Pd(OAc)₂, which showed that only protons adjacent to the sulphur, oxygen, and the aromatic ring, got affected when coordinating with the Pd^{II} salt.²⁶ Therefore, for these model systems, the structure of the amphiphile was simplified by replacing the PEG chain and dendritic aliphatic end-groups with methyl groups, while preserving the exact architecture of

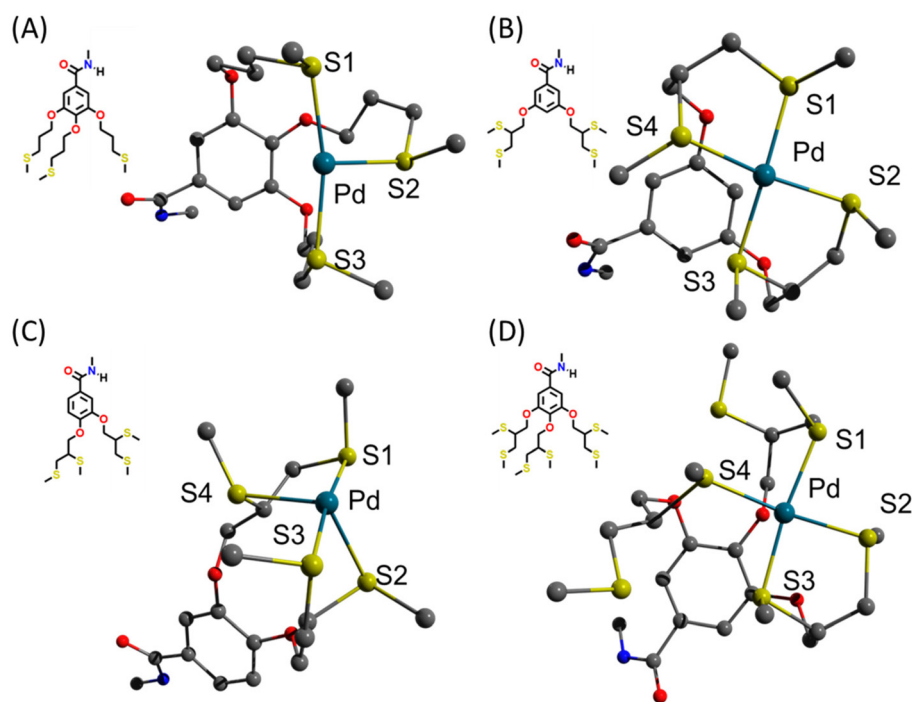


Fig. 7 Simulated results of the predicted orientation of thioethers around the palladium atom (green) in the dendritic structures of (A) three-armed architecture, (B) four-armed (3,5) architecture, (C) four-armed (3,4) architecture and (D) six-armed architecture.



the dendritic cores (Fig. 7). Subsequently, we optimized the geometries of the four model systems and the resulting geometry around Pd^{II} was analysed for each system.

The optimized structure of the Pd^{II}-complex with the three-armed dendron (Fig. 7A) exhibits a T-shaped geometry of the Pd^{II}-center with vacant fourth position that could potentially serve to bind the substrate to the palladium center. The optimized structure of the Pd^{II}-complex with the four-armed (3,4) dendron has a seesaw geometry around Pd^{II}-center, which is atypical and unfavoured in the case of Pd^{II} ion (Fig. 7C). The optimized structures of the Pd^{II}-complex with the four-armed (3,5) and six-armed dendrons both have a square planar geometry, which typical for Pd^{II}-complexes (Fig. 7B and D).^{46,47}

The calculated results (Fig. 7) provided important insights into the different reactivities that were observed in the kinetic experiments (Fig. 4 and Fig. 5). While nanoreactors made from four-armed (3,5) and the six-armed dendritic amphiphiles were slowest among the four studied systems, both architectures were shown to have square planar geometry architectures around the Pd^{II}-center, which is a more stable geometry for the Pd^{II}-complex.

On the other hand, the dendritic cores of the other two amphiphiles, which were significantly more reactive, showed atypical seesaw geometry (for the four-armed (3,4) architecture) and a T-shaped geometry (for the three-arm architecture). These unique geometries can be expected to be less stable and hence allows for a better substrate binding, which enables increased reactivity. Overall, the calculated geometries could provide an explanation for the different reactivities observed for the four systems, indicating that the more stable the chelating of the Pd^{II} ions, the less reactive the nanoreactors become. Nevertheless, it is important to note that despite the different geometries, all amphiphiles were shown to maintain the same amount of Pd^{II} ions, demonstrating the critical role of the dendritic core's architecture on the reactivity of the micellar nanoreactors.

3. Conclusions

This work was aimed at understanding the influence of the hydrophobic dendritic core's architecture on Pd^{II}-mediated depropargylation reaction in micellar nanoreactors. Our micellar nanoreactors were based on self-assembled PEG-dendron amphiphiles with palladium acetate as the catalytic species. The ability to synthesize the dendritic block with a high precision enabled us to construct PEG dendron amphiphiles with different dendritic architectures while controlling their HLB and overall molecular weight by changing the number, position and length of the end-groups. These nanoreactors were used to perform depropargylation reactions in PBS with two different propargyl caged substrates, which could potentially be replaced by a propargyl caged prodrug in the future.

The obtained results highlighted how changing the dendritic architecture of the amphiphiles has a greater influence on the depropargylation kinetics than the number and length of

the end-groups. In our observations, the micelles based on C11×4 (3,4) exhibited the fastest kinetics, surpassing the speed of C11×4 (3,5)-based micelles by four to five times. Remarkably, this significantly greater efficiency was observed despite both amphiphiles being isomers with identical atomic composition. The reason for such a drastic difference in the reactivity of micelles with different architectures was elucidated with the help of DFT calculations of the dendritic core of the amphiphile around the Pd^{II} ion. These calculations suggested that CX×4 (3,4) architecture should have a highly unlikely seesaw geometry, while the three-armed architecture showed less stable T-shape geometry for their Pd^{II} complexes. These unusual geometries, in contrast to the square planar geometry obtained for the CX×4 (3,5) and six-arm amphiphiles, are predicted to be more labile. As a result, they allow Pd^{II} to participate more actively in the depropargylation process compared to the other architectures.

This study highlights how subtle changes in the polymeric architecture can potentially lead to enormous changes in the reaction kinetics for conducting abiotic reactions in aqueous media. Furthermore, when designing micellar nanoreactors for biorthogonally activating prodrugs, we believe these results can serve as a valuable tool for controlling the prodrug activation rate, allowing it to be customized according to the medical requirements of patients.

Author contributions

SW and RJA conceived and designed the project. SW and PR synthesized the amphiphiles and performed the depropargylation experiments. KV helped with performing depropargylation experiments. ST synthesized the PNPPE substrate. RD performed the DFT calculations. AG performed the ICP-MS characterizations. All authors contributed to the interpretation of results and experimental design. SW wrote the manuscript, and all authors revised the final version.

Conflicts of interest

There are no conflicts to declare.

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