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The role of *Caenorhabditis elegans* in the discovery of natural products for healthy aging†

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Covering: 2012 to 2023

The human population is aging. Thus, the greatest risk factor for numerous diseases, such as diabetes, cancer and neurodegenerative disorders, is increasing worldwide. Age-related diseases do not typically occur in isolation, but as a result of multi-factorial causes, which in turn require holistic approaches to identify and decipher the mode of action of potential remedies. With the advent of *C. elegans* as the primary model organism for aging, researchers now have a powerful *in vivo* tool for identifying and studying agents that effect lifespan and health span. Natural products have been focal research subjects in this respect. This review article covers key developments of the last decade (2012–2023) that have led to the discovery of natural products with healthy aging properties in *C. elegans*. We (i) discuss the state of knowledge on the effects of natural products on worm aging including methods, assays and involved pathways; (ii) analyze the literature on natural compounds in terms of their molecular properties and the translatability of effects on mammals; (iii) examine the literature on multi-component mixtures with special attention to the studied organisms, extraction methods and efforts regarding the characterization of their chemical composition and their bioactive components. (iv) We further propose to combine small *in vivo* model organisms such as *C. elegans* and sophisticated analytical approaches (“wormomics”) to guide the way to dissect complex natural products with anti-aging properties.

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1 Introduction

1.1. Historical development

Aging is a fundamental and fascinating process that affects all natural organisms.^{1–3} For humans, aging has always been a subject of curiosity and cultural reflection and it is addressed in religious, philosophical, and pop-culture contexts. It is a complex multi-factorial phenomenon manifesting as a decline of tissue and cell functions and an increase in the risks of many so-called age-related diseases, which include *e.g.*, Alzheimer's disease, Parkinson's disease, osteoporosis, osteoarthritis, type 2 diabetes, cardiovascular disease, chronic obstructive pulmonary disease, and glaucoma. Over the last decades, our understanding of aging has changed dramatically, with a paradigm shift from viewing it as a passive, entropic process to an actively

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regulated process influenced by a combination of hereditary, environmental, and lifestyle factors,⁴ which occur despite complex pathways of maintenance and repair.⁵

Hereby, invertebrate model organisms such as the rhabditid nematode *Caenorhabditis elegans* (*C. elegans*) have paved the way to effectively study the phenomenon of aging and its pathways.^{6,7} In the 1980s and early 1990s, research on *C. elegans* provided some remarkable results. It was discovered that mutations in highly conserved ancient genes such as the *daf-2* or *age-1* cause a doubling of life span depending on *daf-16*.^{8,9} There is strong evidence that the mammal orthologues of these genes, the insulin/insulin-like growth factor receptor, phosphatidylinositol-3-OH kinase and FOXO3 genes also contribute (negatively and positively) to mammal longevity.^{10–13} These early studies furthered the understanding of genes involved in the regulation of aging and revealed that the function of these gene products, hence aging, is open to interventions.^{6,14} Today we have identified several drugs that can increase mammal life span significantly.^{15–18} Although it is still

unclear how these approaches translate to humans,¹⁹ there is growing consensus that aging and late life multi-morbidity might be open to pharmaceutical intervention. That would provide the opportunity for holistic rather than individual intervention to combat the inevitable effects of aging like cancer, neurodegeneration, cerebral and cardiovascular diseases, blindness, sarcopenia and wrinkles – a regulatory pathway that gets these jobs done, all at once.

1.2. From lifespan to health span

For most of human history, human lifespan stagnated at around 20 to 40 years. Only recently with improvements in medicine, agriculture, sanitation and general living conditions life expectancy has been boosted to over 70 years.^{20,21} However, the proportion of life spent in good health has not increased to the same extent, meaning that more and more years are spent in poor health.^{22,23} Hence, multimorbidity and polypharmacy is the norm in old age.²⁴ Thus, the question



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Ulrike Grienke studied Pharmacy and received her PhD in 2011 from the University of Innsbruck, Austria. She has completed several postdocs/research stays in Austria, Ireland, and Brazil. In 2021, she received her habilitation in Pharmacognosy from the University of Vienna, Austria, where she currently works as senior scientist. Her research is focused on the discovery of

bioactive constituents from natural sources connecting different methodologies including NMR-based biochemometric approaches and supercritical fluid-based systems.



Julia Zwirchmayr studied Pharmacy at the University of Vienna, Austria, where she received her diploma in 2017 and her PhD degree in 2022. She conducted her doctoral thesis at the Division of Pharmacognosy, Department of Pharmaceutical Sciences under the mentorship of Ulrike Grienke and Judith M. Rollinger. During her stay as a post-doctoral researcher in the lab of professor Rollinger she

focused mainly on the discovery of bioactive secondary metabolites from Nature by implementing NMR-based biochemometric approaches and in vivo studies in Caenorhabditis elegans.



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based drug discovery. In her research she focuses on the interdisciplinary field of integrating big data analytics (cheminformatics, chemometrics) and phenotypic screening (Caenorhabditis elegans) as strategy for the discovery of novel lead structures against infections, metabolic syndrome, and inflammation.



arises as to whether potential life-prolonging therapy will only extend life, or will also compress morbidity? Interestingly, slowing aging does not necessarily mean reducing morbidity caused by age-related diseases such as cancer, cardiovascular disease, or Alzheimer's disease. Instead, research shows that mutations which extend lifespan can also come with an increased proportion of total life time spent healthy.^{25–28} However, there are also mutations where increased lifespan simply translates to an extended period of frailty.²⁹ In 2013, Rattan proposed using the term “healthy aging” instead of “anti-aging” to shift the understanding of the aging process as an evolving phenotype due to the failure of homeodynamics.⁵ According to the World Health Organization (WHO), the term “healthy aging” is defined “as the process of developing and maintaining the functional ability that enables well-being in older age” rather than just the absence of a disease.³⁰ The emerging field of geroscience is an interdisciplinary research area striving to understand the connection between aging and age-related diseases.^{23,31–33} The goal of geroscience is not to increase life span, but to understand the biology of aging in order to delay diseases and improve health span. In 2013, a conceptual framework of aging was established whereby nine common hallmarks of aging were defined.³⁴ Hereby primary hallmarks develop progressively with time. They comprise genomic instability, epigenetic alterations, telomere attrition and loss of proteostasis. Secondary hallmarks, which include mitochondrial dysfunction, deregulated nutrient-sensing and cellular senescence, evolve as a result. Together with tertiary hallmarks of altered cell communication and reduced stem cell turnover they produce the aged phenotype. A geroscientific hypothesis is that targeting the hallmarks of aging holistically would be an effective approach to delay the pathogenesis of age-related diseases.^{35,36} Thus, the early prevention of the onset of age-related diseases is considered as ideal approach for extending the health span and achieving healthy aging.³⁷ However, the framework of nine hallmarks of aging has been challenged in the last ten years.^{38,39} Despite much progress, many challenges remain in formulating an enhanced paradigm of aging.⁴⁰

Yet, it has been shown that the rate of aging, at least in part, can be delayed in mammalian model organisms by genetic, behavioural and also pharmacological means,^{15–18,41–45} and when aging is delayed, the rate of age-related diseases and conditions will also slow down.⁴⁶ Since aging is associated with a progressive deterioration of multiple organs, tissues and physiological functions, the usefulness of multicellular organisms for testing is undeniable in the search for aging-modulators. However, there are considerable and pervasive experimental challenges to studying aging in vertebrates and humans. This is especially true for natural products (NPs) including botanicals, because the challenges of sufficient and well-defined material are added. Short-lived animal models that are both, amenable to experimentation and miniaturization, are essential in aging research and provide meaningful guides to the biology of aging in humans.⁴⁷ Thus, the most-used experimental model organisms for screening the impact of NPs on

their age and health span extending properties are small invertebrates such as *Drosophila melanogaster* and *C. elegans*.

1.3. Current status

NPs are a recognized source of bioactive molecules. They contributed significantly to the arsenal of approved drugs.⁴⁸ Isolated NPs and their derivatives, such as rapamycin, resveratrol, and metformin, are probably among the most studied and pre-clinically advanced interventions against aging and age-related diseases. Botanicals and multi-component mixtures have also been studied early on⁴⁹ in the context of dietetics.^{50,51} The interface of healthy aging, *C. elegans* and NPs has grown immensely since then. A recent analysis of Saul and coworkers⁵² analysed *C. elegans* health span literature: according to their analysis, out of the 42 most studied samples in different aging phenotypes, 27 are natural compounds, 12 are extracts; only three samples are not from natural origin. Looking at the 34 most studied agents in test conditions (number of concentrations, temperatures, exposure timings, food and strains), 17 are natural compounds, 12 are extracts, three are NP derivatives (e.g., acetylsalicylic acid) and only two have no natural origin. This dominance of NPs in health span research is supposed to have manifold reasons, such as (i) their increased likelihood for bioactivity;^{53,54} (ii) the many advantages of *C. elegans*, especially for NP research;⁵⁵ (iii) the contradicted but popular free radical theory of aging has led many scientists to focus on antioxidants, such as polyphenols, which are widely found in nature;^{56–58} (iv) dietary interventions as a primary target for healthy aging and in case of botanicals only a blurry distinction between dietary and pharmacological intervention; (v) traditional knowledge on herbal remedies for symptoms related to healthy aging. (vi) Additionally, some NPs have been subject of anti-aging research through mammal models before the use of *C. elegans* became the premier model organism in this field;⁵⁹ (vii) according to the xenohormesis theory, naturally occurring metabolites, such as plant polyphenols might have an evolutionary role as signals to stimulate protective pathways in organisms consuming them.⁶⁰ Hence their dominance in the field stems from the high likelihood to have health span promoting effects due to a speculated evolutionary imprint.

The last decade has witnessed tremendous increase in research activities dealing with *C. elegans* used to search for anti-aging NP panaceas. The main aspects have been surveyed in the last ten years providing different perspectives on this research field, e.g., in 2013, Argyropoulou and colleagues⁶¹ gave a profound overview on natural compounds with anti-aging activities derived from phenotypic studies; Pallauf *et al.* 2017 (ref. 62) reviewed the lifespan-extending effects of flavonoids in different model organisms; Ding *et al.* 2017 (ref. 63) summarized treatments with reported activity in aging models; Matsunami, 2018 (ref. 64) reviewed literature on *C. elegans* as model for frailty. Chattopadhyay and Thirumurugan 2018,⁶⁵ who reviewed 18 dietary and medicinal plants, and Wang *et al.* 2021,⁶⁶ who focused on 23 plants from traditional Chinese medicine, both analyzed the longevity promoting effects of plant genera. Several reviews on the longevity promoting effect of natural compounds were also published.^{67,68} NPs active against transgenic *C. elegans* models of



Alzheimer's disease were surveyed by Navarro-Hortal *et al.*⁶⁹ Shen and coworkers^{70,71} reviewed the use of *C. elegans* as a model for researching bioactive compounds in food against aging, obesity and Alzheimer's disease. A review on the oxidative and anti-oxidative potential of NP in *C. elegans* was also published recently.⁷² In 2021, Saul *et al.*⁵² presented the "healthy worm database" and analyzed literature on health span promoting treatments, including extracts, in *C. elegans*. These reviews have summarized the outcome of hundreds of research initiatives dedicated to the identification of anti-aging NPs samples by means of *C. elegans*.

1.4. Scope of this review

A very important research question in pharmacognosy still remains on how to cope with the phenotypic effects from multi-component mixtures and to identify which molecules trigger those observed bioactivities. It is also imperative to understand how well a read-out from a whole organism, such as *C. elegans* wild type or mutant strains, can accelerate the search for efficient (herbal) drugs for the benefit for human health in particular with respect to an ever-increasing number of elderly people. The essentials of health span research in *C. elegans*, which includes worm biology, methods, and assays, were reviewed. In addition, a representative number of studies (169) on NPs research in *C. elegans* from 2012 onwards, were analyzed with special emphasis on the phytochemical aspects of extracts. This review presents past and present approaches for the discovery of anti-aging NPs implemented in pharmacognostic workflows as well as novel strategies to dissect complex outcomes.

2. *Caenorhabditis elegans* and healthy aging

C. elegans, a basal metazoan organism, offers several advantages as model organism for life sciences. Its use has contributed significantly to many important discoveries. Three of them were honored with noble prizes:⁷³ (i) the discovery of the genetic regulation of organ development and programmed cell death; (ii) the discovery of RNA interference and gene silencing with double-stranded RNA; and (iii) the development of green fluorescent protein as marker for gene expression. *C. elegans* is also an invaluable model organism especially in the field of health span research and for NP screens. Their small size allows for miniaturized high-throughput procedures at the scale of conventional cell culture methods,⁷⁴ which offers considerable advantages in terms of resources and the necessary quantities of test substances.⁷⁵ Large populations of genetically identical animals of synchronized age can be cultured and maintained easily within short time frames and with relatively simple procedures.⁷⁶ The worm has a rapid life cycle of two to three weeks under standard laboratory conditions and a completely sequenced and well-annotated genome with many well-established resources and protocols available.^{77,78} The nematode undergoes progressive, degenerative changes while aging, thus providing physiologically relevant data and insights into the underlying mechanism of human diseases.⁷⁹ Half of the human protein-coding genes have

recognizable orthologues in *C. elegans*.⁸⁰ Thus, the nematodes have the potential to bridge the gap between *in vivo* and *in vitro* approaches in the context of a whole-animal setting,^{81,82} and have become a popular animal model in aging research.⁸³ Moreover, an invertebrate model comes with certain advantages with respect to legal, regulatory and ethical issues.^{84,85}

This chapter briefly outlines main characteristics of *C. elegans* biology and anatomy, its use in aging research (Table 1), and pathways and genes known to affect its lifespan and health span (Table 2).

2.1. Biology

C. elegans is a soil dwelling non-parasitic nematode which feeds on bacteria and yeasts found in its natural habitat of compost heaps, rotting fruits and plants.^{86,87} In these microorganism-rich environments it interacts with a diversity of microorganisms and they not only serve as food but also as commensals. In the lab it is usually maintained on agar plates with axenic uracil auxotroph *Escherichia coli* as bacterial diet. Under these conditions, 99.9% of *C. elegans* are self-fertilizing hermaphrodites which can each produce more than 250 eggs.⁸⁸ The eggs are protected by a resilient shell, which allows them to resist even very adverse environments. After hatching, the life cycle of *C. elegans* consists of four larval stages L1 to L4. The end of each larval stage is characterized by a molt, until the reproductive adult stage is reached. In the absence of food, worms can arrest in L1 stage and survive for several days, or, if they have already reached the L2 stage, enter the dauer stage at which point they worms can survive for several months.⁸⁹ For longer periods of time, larvae can even be stored in a cryogenically frozen state at $-80\text{ }^{\circ}\text{C}$.^{76,90} Thus, large numbers of strains can be easily maintained. To generate an age-synchronized culture for lifespan experiments, worm cultures are bleached with an alkaline hypochlorite solution, whereby worms are sensitive to the bleach and disintegrate while the egg shell protects the embryos from death. The eggs are then agitated until they hatch, while in the absence of food they arrest at a synchronized L1 stage.⁹¹ Protocols that make use of filters to isolate eggs have also been described.⁹²

2.2. Anatomy

C. elegans (Fig. 1) is an unsegmented pseudocoelomate and lacks respiratory and circulatory systems. It is enveloped in a cuticle, a collagenous, extracellular, exoskeleton that shields the animal from its environment and maintains the morphology and integrity of the worm.⁹³ The cuticle is synthesized by a hypodermal cell layer and plays a critical role in body movement because it is attached to muscles.⁹⁴ The worm's mouth is at the anterior of its body. It is a small opening with a cavity that is separated from the intestinal lumen by the pharynx, a neuromuscular pump and one of the most complex organs of the worm. The pharynx is comprised of eight muscle cells, 20 neurons, and epithelial, support, and gland cells.^{95,96} The rhythmic contraction of pharyngeal muscles, referred to as pumping, sucks bacteria into the mouth of the worm, mechanically grinds them and transports the food into the intestinal lumen. The pumping rate is dependent on food availability, worm age and can be altered by compounds that modulate





Table 1 Most frequently employed assays used to analyze health span promoting NPs

Assay	Principle	Strains	Readout	Advantages	Limitations	Lit
Lifespan	Age-synchronized worms are incubated together with bacteria and test substances for several weeks. Living and dead worms are counted at certain times	N2, BA17, SS104	Kaplan–Meier plots	De facto standard in most laboratories most direct method to measure aging	Time consuming and tedious process	75 and 171–173
Automated lifespan	Scanner-based assays		Mean lifespan	Low technical requirements	No continuous observation	
Body movement	Flow-cell based assays Manual counting of body bends, bending angle or crawling speed Division of worms into movement classes	N2	Maximal lifespan. Death time 50% (DT ₅₀) % survival at specified time point Maximum velocity	Continuous observation, unbiased readout Most informative metric of health span	Researcher bias	122 121 123 and 174
Pharyngeal pumping	Counting of strokes per time which declines gradually with age	N2	Number of body bends % of population that can be assigned to defined classes <i>e.g.</i> , spontaneously moving, movement only after stimulation or no movements	Low technical requirements	High technical requirements, low throughput	148 and 175
Intestinal autofluorescence	Autofluorescent lipofuscin which accumulates with age is quantified	N2	Pumping rate	Good marker of health	Measures are noisy and observed effects can be independent of age	96 and 137–139
Reproductive period	Decline in reproductive ability is an early aging phenotype Counting of progenies manually/automated for several days (usually 3–5 days)	N2	Either by microscopy and image processing or with a plate reader Time span during reproduction is observed	Reproductive span is shorter than lifespan	Not aging-specific	105 and 176–179
Antioxidant capacity	Indirect measurement of ROS levels with dichlorodihydrofluorescein	N2	Relative fluorescence	Antioxidative effect <i>in vivo</i>	Regulated independently to other health parameters, not correlated with lifespan	180
Abiotic stress resistance	Worms are challenged with reactive oxygen species producing agents (H ₂ O ₂ , naphthoquinones, paraquat) or heat (35–37 °C)	N2	Kaplan–Meier plots, % survival at specified time point	Low technical requirements impaired stress resistance is a hallmark of aging	Many interferences, not specific for aging Mechanistic link of ROS and aging is debated not specific for aging	150, 151 and 181 182–185



Table 1 (Contd.)

Assay	Principle	Strains	Readout	Advantages	Limitations	Lit
Proteotoxicity	Alpha synuclein expression	NL5901, OW13	Quantification of YFP reporter marked alpha synuclein	Appropriate companion assays, link between longevity pathways and cellular processes (e.g., autophagy, vesicle trafficking and protein quality control systems) required for rescue of these phenotypes	Alpha synuclein is expressed in muscles and not in neural system Amyloid beta is expressed in muscles and not in neural system	186–188
	Amyloid beta expression	CL4176, CL2006, CL2659	Protection from paralysis induced by amyloid beta in body wall muscles			
		CL2331	Quantification of GFP reporter marked amyloid beta			
Transcription factor translocation	Measurement of nuclear translocation GFP-marked <i>daf-16</i> , <i>skr-1</i> or <i>hsf-1</i>	TJ356, LD1, MAH97, CF1824	Classification of phenotypes into nuclear translocated, intermediate and cytosolic forms	Mechanistic insight, low technical requirements	Subjective classification of intermediate forms	120 and 189

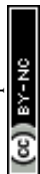
e.g., serotonin signaling.⁹⁷ The intestine consists of 20 cells and has various functions apart from digestion and nutrient absorption including nutrient storage and synthesis of macromolecules and yolk. Yolk particles are lipoproteins, which are packaged into oocytes in fertile worms. Four enteric muscles located at the posterior of the intestine and body work for defecation.⁹⁸ The gonad of the hermaphrodites is an ovotestis that produces sperm in the L4 stage and oocytes in the adult stage. As a result, the worm produces up to 300 progenies by self-fertilization within a few days, a process that involves self-destructive biomass repurposing.⁹⁹ Noteworthy features of the worm's anatomy are also six macrophage-like scavenger cells, the coelomocytes,¹⁰⁰ and 95 body wall muscle cells.¹⁰¹ For the worms' 302 neurons a complete map of synaptic connections, the connectome, is available.

2.3. *C. elegans* assays related to healthy aging

The aged phenotype of *C. elegans* develops dynamically over time until death. Many age-related changes on the molecular and tissue level become apparent at the level of the whole organism and can be easily assayed e.g., via behavioral phenotypes and morphological markers. These changes start from the first day of adulthood. Already at day 2 of adulthood learning and long-term memory worsens.¹⁰² Around day 4 of adulthood the hermaphrodite enters the post reproductive period and yolk lipoproteins start to accumulate ectopically in the body cavity.^{103,104} This is accompanied by gradual intestinal atrophy and an increase of auto-fluorescent pigments which represent probably advanced glycation end products.¹⁰⁵ The two posterior V5-derived (PVD) neurons for sensation of mechanical stimuli and cold temperatures degenerate.^{106,107} At day 7 of adulthood, sarcopenia is apparent histologically by a progressive disorganization of sarcomers,¹⁰⁸ and behaviorally by a reduced motor activity. The latter one is also caused by stiffening and thickening of the cuticle caused by an unregulated collagen biosynthesis.¹⁰⁸ Pharyngeal pumping also slows gradually due to an aging-related decline of pharyngeal muscles and other pathologic changes. While young adult worms have a pumping rate of more than 200 pumps per minute, old adults achieve less than 50 pumps per minute.^{109,110} Since the duration of contractions is extended, the grinder fails to break down the bacterial food. Hence, living bacteria increasingly colonize the intestinal lumen causing it to bloat.^{111,112} Whether and how this dysbiosis subsides with a more natural bacterial culture and a functional microbiome, however, still needs to be investigated. Around day 10, the body motions become sporadic. Their response to stimuli like plate shaking, light or touching decreases. Overall, old adults shorten and get thicker around the midbody area and their bodies begin to wrinkle, an indicator of physical deterioration.^{108,113} Death in *C. elegans* is usually assessed as non-response to harsh stimuli. A wave of necrotic cell death in the intestine and a burst of intense blue “death” fluorescence are reliable indicators of organismal death.^{112,114} The wormatlas has reviewed the aging of the worm and provides very good figures and video sequences that are available to the scientific community.¹¹⁵ To assess NPs-based biological effects in *C. elegans*, the following pharmacological endpoints can be determined by means of (i) survival (lifespan and lethality), (ii) behavioral

Table 2 Most relevant *C. elegans* genes involved in health span promoting effects in *C. elegans*

Gene	Influence	Human orthologue	Role	Significance	Exemplary NPs
<i>skn-1</i>	↑ ↑	NRF2	Transcription factor Cellular regulator of oxidative stress response	<i>skn-1</i> mutants show decreased resistance to oxidative stress and shortened lifespan ²²⁰ <i>skn-1</i> transgenic strains with constitutive nuclear localization exhibit increased oxidative stress resistance, improved health span parameters and mean lifespan is extended by 5–21% (ref. 221) <i>hsf-1</i> overexpression promotes longevity and delays age-related protein misfolding and proteotoxicity ^{232,233}	Urolithin A, ¹⁴⁰ curcumin, ²¹⁴ baicalein, ²²² withanolide A, ²²³ ginsenosides; ¹³² extracts of <i>Lycium barbarum</i> , ^{224,225} <i>Hibiscus sabdariffa</i> , ²²⁶ <i>Crataegus pinnatifida</i> , ²²⁷ <i>Apios americana</i> , ²²⁸ <i>Vaccinium corymbosum</i> , ²²⁹ <i>Anacardium occidentale</i> ²³⁰ and the essential oil of <i>Juniperus communis</i> , ²³¹ Trigonelline, ²¹² caffeine, ²³⁴ extracts of <i>Cratogeomys formosum</i> , ¹⁸⁷ <i>Coffea arabica</i> , ²³⁴ <i>Lonicera japonica</i> , ¹⁵⁸ and <i>Vicatia thibetica</i> ²³⁵
<i>hsf-1</i>	↑ ↑	HSF1	Transcription factor Protects against age-related proteotoxicity	Loss of function mutations in the insulin/insulin-like growth factor signaling (IIS) pathway extend lifespan two-fold ^{8,236}	Orientin, ²⁰² and naringin, ²⁰⁰ extracts of <i>Rubus idaeus</i> , ²³⁷ <i>Ribes fasciculatum</i> , ²³⁸ <i>Hedyotis diffusa</i> ¹⁹² and <i>Morus alba</i> ²³⁹
<i>daf-2</i>	↓ ↓ ↓	Insulin/insulin-like growth factor receptor (IR/IGF-1R)	Transmembrane tyrosine kinase receptor Various functions in metabolism, growth, and reproduction	Loss of function mutations in the insulin/insulin-like growth factor signaling (IIS) pathway extend lifespan two-fold ^{8,236}	Orientin, ²⁰² and naringin, ²⁰⁰ extracts of <i>Rubus idaeus</i> , ²³⁷ <i>Ribes fasciculatum</i> , ²³⁸ <i>Hedyotis diffusa</i> ¹⁹² and <i>Morus alba</i> ²³⁹
<i>age-1</i>	↓ ↓ ↓	Phosphoinositide-3-kinase (PI3K)	Kinase downstream to <i>daf-2</i>		
<i>daf-16</i>	↑ ↑ ↑	FOXO	Transcription factor Various functions in metabolism, cell proliferation and stress resistance	<i>daf-16</i> is required for increased life span and enhanced resistance to stress when the IIS pathway is downregulated ^{240,241}	Indicaxanthin, ¹²⁰ geniposide, ¹²⁷ 4-hydroxy-E-globularinin, ²⁴² hydroxytyrosol, ^{218,243} epigallocatechin gallate, ^{230,244–249} flavonoids ¹⁴⁹ and silymarin; ²¹¹ extracts of <i>Rhodiola</i> sp., ¹³¹ <i>Eugenia uniflora</i> , ²⁵⁰ <i>Warburgia salutaris</i> , ²⁴⁸ and <i>Punica granatum</i> , ²⁵¹ Stilbenes, ^{148,195} pentagalloylglucose, ¹⁹⁰ extracts of <i>Vaccinium uliginosum</i> , ²⁵⁴ and <i>Camellia sinensis</i> ^{255,256}
<i>sir-2.1</i>	↑ ↑	SIRT1	NAD ⁺ -dependent protein; senses cellular NAD ⁺ /NADH levels to regulate transcription, genome stability and many metabolic processes ²⁵²	Required for life span extension by caloric restriction; sirtuin overexpression increases longevity by up to 50% ²⁵³	Rapamycin ²⁵⁷ and extracts of <i>Vigna radiata</i> ¹³³ and <i>Ganoderma lucidum</i> ²⁵⁸
<i>let-363</i>	↑	TOR	Part of TORC1 and TORC2 kinase complexes; nutrient and energy sensor, tunes protein synthesis and autophagy	Adulthood knockdown of TORC1 pathway genes extends lifespan ~25% and enhances health span parameters such as motility, stress resistance and pharyngeal pumping ²⁵⁷	
<i>aak-1/aak-2</i>	↑	AMPK	Kinase; cellular sensor of energy levels, tunes cellular catabolic and anabolic processes	Couples lifespan to information about environmental stressors, energy levels and <i>daf-2</i> signaling ²⁵⁹ mediates lifespan extension through dietary restriction ^{259–261}	Trigonelline ²¹² and orientin; ²⁰² extracts of <i>Vigna radiata</i> ¹³³



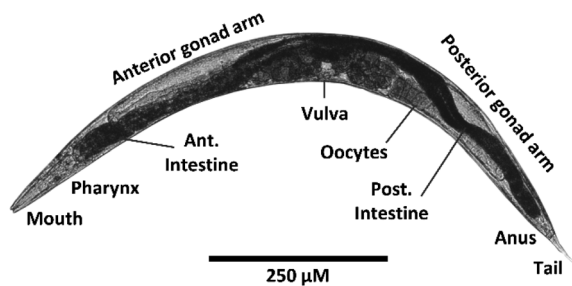


Fig. 1 Brightfield image of adult hermaphrodite *C. elegans* with scale and visible organs.

changes (pharyngeal pumping and locomotor activity), (iii) histological parameters (autofluorescence, muscle fiber organization), (iv) reproduction (morphologic deformity, brood size and larval development), (v) resistance to stressors (heat or reactive oxygen species (ROS)), (vi) biochemical markers, *e.g.*, measuring ROS levels, or (vii) gene/protein expression and cellular localization. Noteworthy assessments are also (viii) proteotoxicity assays that model amyloid beta, α -synuclein or polyglutamine toxicity (Table 1).^{72,116}

(i) Survival assays. Lifespan assays are probably the most direct method to determine the effect of test samples on aging. Hereby, cohorts of worms from the same synchronized populations are created by separating them into different wells or different petri dishes. Then they are challenged by interventions such as different diets, compound treatments, or double-strand RNA (dsRNA). Living and dead worms are counted at specified intervals (*e.g.*, every day, every second day). Usually, Kaplan–Meier survival plots are generated to illustrate the percentages of live worms in the cohorts over time, and the data are statistically analyzed.¹¹⁷ These assays can be performed either on solid agar or in liquid media. Both come with advantages and limitations reviewed elsewhere.^{117,118} Manual counting in these conventional lifespan assays is time-consuming and tedious and it comes with several limitations such as researcher-oriented bias and the exposure of worms repeatedly to light, heat or mechanical stresses. That is why more and more approaches to automate lifespan assays have been developed. The lifespan machine automatically captures sequential images of worm populations on agar plates. It determines death events when worms stop crawling for long periods of time with image processing software.^{119,120} Other methods employ flow cells where fresh media, food and interventions enter, while eggs, metabolites and debris exit the flow cell. Digital video recording allows for continuous observation.¹²¹ Beside death events, health span parameters like locomotion, morphology and behaviour can be assayed.¹²² Of course, this comes at the expense of high technical requirements and a decreased throughput compared to microwell-based approaches.

(ii) Behavioural assays. Maintaining mobility and physical fitness is an important indicator of healthy aging. In *C. elegans* it is also an important predictor of lifespan.^{110,123,124} The worm's locomotion gradually declines with age and is therefore a straightforward health span marker. The rate of body movement can be accessed with markers such as maximum bending amplitude, crawling speed on agar,^{125–127} number of body bends during fixed

intervals of time or bending angle.¹²⁸ The locomotion of worms can also be measured at the cohort level.^{129–131} In this case, cohorts of worms are generated and treated with samples, similar to life span studies. Then the nature of the worms' movements on successive days is examined, dividing the worms into motility classes as in the most unbiased way as possible *e.g.*, healthy class worms that exhibit symmetrical and spontaneous movements, aged class worms that exhibit uncoordinated, stiff movements, and very aged class worms that move only the tail or head in response to stimuli. In this way it can be shown how a treatment slows down the age-dependent motility decline.^{132,133} However, the classification of worms is prone to researchers' bias and body movements are noisy, because behavioural states are episodic. Food and neuro-modulators like exogenous 5-HT can substantially inhibit movements without necessarily affecting aging.^{124,134–136} A further measure of aging in *C. elegans* is the amplitude of pharyngeal pumping.¹³⁷ Hereby, pharyngeal contractions are counted during fixed intervals of time. As the pumping rate can have irregular rhythms, *e.g.*, showing pumping bursts,⁹⁷ it is necessary to count pumping of worms several times and calculate the mean pumping rate.¹³⁶ Technical requirements are low, a routine stereomicroscope and a hand counter is all that is needed.⁹⁶ However, the repeated measurements of pumping are cumbersome and prone to researcher bias. There are also methods to automate this measurement with image analysis of consecutive image frames¹³⁸ or by recording electropharyngeograms with electrodes.¹³⁹

(iii) Histological parameters. A histological marker of health span is the intensity of intestinal autofluorescence. Red channel fluorescence correlates well with the worm's remaining days of life, and is therefore a good marker of health.¹⁰⁵ Although not as frequently employed, muscle fibre organization is also a good surrogate parameter of health in aged worms. It can be assayed with transgene strains showing GFP-marked myosin (*e.g.*, strain RW1596).^{108,140,141}

(iv) Reproduction. Another frequently employed health span assay is the measurement of the reproductive period. In the L4 stage, the worms produce around 300 sperm cells. With the advent of adulthood, they switch to the production of oocytes which are continuously fertilized with sperm. When everything runs smoothly the worm can produce up to 300 progenies. The ability to produce eggs for a longer time period is sometimes regarded as health parameter but it is largely regulated independently of other health parameters and has a poor correspondence with lifespan.¹¹⁰ Nevertheless, it is useful to probe if health span is achieved *via* suppression of reproduction.^{142,143}

(v) Resistance to stressors. As most health span prolonging pathways also promote resistance to various abiotic factors like ROS generating chemicals, ultraviolet radiation or heat stress, assays that test for such properties are frequently used as surrogates to identify new health-promoting genes and agents.^{144–147} This stress resistance is often measured by exposing worms to elevated temperatures (*e.g.*, 35 °C) or a ROS generating chemical (*e.g.*, juglone, paraquat or peroxide) and comparing the survival of treated and untreated worms. This is also performed semi-automatically in plate-reader format using the Sytox®Green reagent which can only penetrate necrotic cells where it leads to a strong increase of fluorescence. Thus, SYTOX



fluorescence is positively correlated to worm mortality rate and negatively related to their stress resistance.^{148,149}

(vi) Biochemical markers. 2',7'-dichlorodihydrofluorescein (DCF) is a small-molecule fluorescent probe which is frequently used to assess intracellular ROS in *C. elegans*. Upon cellular uptake DCF is oxidized by ROS to the fluorescent product 2',7'-dichlorofluorescein. Thus, DCF fluorescence serves as an indicator of the accumulation of oxidative damage in worms, which is associated with senescence.¹⁵⁰ However, this oxidative stress theory of aging is increasingly challenged.^{151,152} In addition, most hits of these DCF assays are not followed up in detail and the documentation of the experimental protocol is sometimes inadequate. This is critical because the measurement of ROS and oxidative damage in general, and in particular with DCF, can be prone to many interferences and problems as summarized recently.¹⁵³

(vii) Protein expression and cellular localization. For a more mechanistic insight into health span promotion by treatments, western blot analysis, quantitative PCR or MS-based proteomics are used.^{83,154,155} The expression of several gene products which are associated with an improved health span *e.g.*, detoxification factors such as heat shock proteins, metallothioneins and super-oxidodismutase, can be monitored by transgene worms carrying GFP-reporters.^{156–159} Noteworthy are also strains carrying GFP fusion proteins with pro-longevity transcription factors such as *daf-16* and *skn-1* where a translocation to the nucleus can be monitored in response to treatments with fluorescence microscopy.¹⁶⁰

2.4. NPs in *C. elegans*-based drug screening

Treatment with test agents in *C. elegans* can be achieved by several methods which vary between studies. When worms are tested in a liquid buffer, test samples can be added to the medium in a given concentration similar to cell culture. However, *C. elegans* cultures on agar plates require mixing into or the spreading of the samples onto the agar. An alternative is to supplement the agents to the worm's food, the bacteria. The concentrations of test substance used and the concentrations achieved in the worm can often differ by several orders of magnitude. The agents can be absorbed by diffusion through the cuticle, the uptake *via* sensory cilia of neurons or through ingestion of bacteria which have taken up the samples. The latter approach is the most relevant for most drug-like molecules.¹⁶¹ This comes with some caveats as the living uracil-auxotroph *E. coli* has a considerable potential to metabolize test agents.^{162,163} The alternative of feeding inactivated bacteria, proposed by some researchers,¹⁶³ significantly impairs worm development.^{164,165} However, *C. elegans* also has a remarkable xenobiotic defense system with many xenobiotic detoxification enzymes and efflux pumps.^{166,167} It should therefore come as no surprise that drug concentrations several magnitudes higher than can be achieved in human plasma are used. Burns and colleagues reported that only 2% of bioactive compounds can produce a robust phenotype at a concentration of 25 μM .¹⁶⁸ However, certain NP classes such as flavonoids show good bioavailability.¹⁴⁹ Therefore, it is necessary to consider appropriate carrier concentrations, solubility and properties of test extracts and compounds, to prevent precipitation. We and other

groups have observed that DMSO concentrations up to 1% do not lead to significant changes of lifespan,^{75,169} although it is clear that high DMSO concentrations have impact regulatory pathways, physiologic rates and drug bioavailability.¹⁷⁰

Drug delivery, dosage, and time of administration also have major effects and may also be a major cause for contradictory results in the literature. Just to name two examples: for instance, when resveratrol is added to worms at L1 stage, it extends lifespan.^{190–192} However, if resveratrol is added later in life at L4 or adult stage, it is reported to have highly variable outcomes with either a significant lifespan extension or not.^{148,193–196} Another example is naphthazarin treatment, which in concentrations of up to 500 μM causes a lifespan extending effect on solid media,¹⁹⁷ while in liquid culture it exerts a potent nematotoxic effect already at 79 μM .¹⁹⁸ This controversial effect may be due to a greater uptake of compounds in liquid culture as it is reported for resveratrol and fluoxuridine.¹⁶¹ “Sola dosis facit venenum” – the dose makes the poison. This maxim attributed to Paracelsus applies very well to *C. elegans* lifespan modulation. Low doses of toxins can sometimes have beneficial effects on lifespan through not fully understood mechanisms which may include the up-regulation of stress response pathways. This was shown for naphthoquinones¹⁹⁷ and arsenite,¹⁹⁹ nor can the increase in lifespan be expected to be linear or exponential with the dose of supposedly non-toxic compounds. For most compounds inverted U-shaped dose response curves with highest effects at medium doses were shown for *e.g.*, naringin,²⁰⁰ withanolide A,²⁰¹ orientin²⁰² and quercetin.²⁰³ This non-linear dose–response poses a problem both for classical bioassay-guided fractionation but also for statistical correlations where active ingredients are identified based on the correlation of bioactivity and compound concentration.

Attentiveness is also warranted in the interpretation of results: *C. elegans* health span is highly sensitive to environmental variables like temperature, light, solid or liquid culture and composition of media. While these variables can be minimized with specialized equipment and handling, other parameters like standardization of the bacterial food source are more complex. Therefore, two-tiered cell banking systems, OD measurements for optimal harvesting points as well as precise harvesting and storage protocols need to be implemented to reduce variations. However, variation in adult survival of synchronized worms even in the same well can still be substantially different. Age-related pathologies appear to occur stochastically, and so do the resulting deaths.¹⁰⁸ According to the disposable soma theory the post-reproductive survival is not under evolutionary pressure. It is therefore more likely to vary. A high number of technical replicates and several parallel and independent experiments are necessary to confirm and quantitate lifespan increase.

2.5. Pharmacological dissection

A major advantage of *C. elegans* is that after identification of health span promoting compounds, the deconvolution of required pathways and targets is feasible. Several pathways have been discovered that mediate the life-prolonging effects of NPs. The best studied is probably the insulin and insulin-like growth factor-1 (IIS) pathway with the tyrosine kinase receptor *daf-2* and the downstream



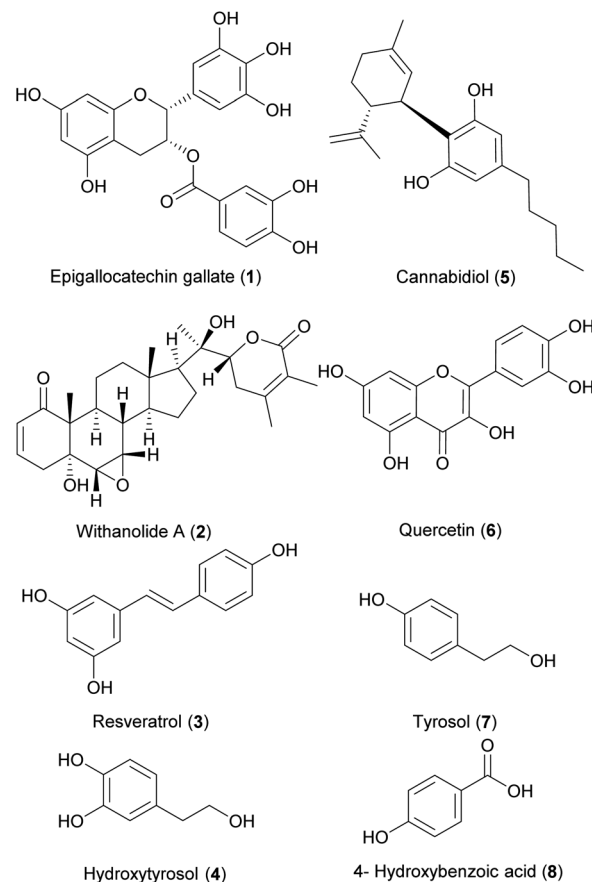
phosphoinositide-3-kinase *age-1*. When activated, this pathway inhibits the transcription factor *daf-16* and thus the expression of downstream genes involved in longevity. Knockdown of *daf-2* and *age-1* genes can prolong lifespan up to two times. There are other pathways frequently named in mediating lifespan prolonging activities of NPs. The putatively best studied ones are summarized in Table 2. NPs with a well demonstrated influence on such pathways have also shown to impact healthy aging in other model organisms and mammals *e.g.*, the urolithins.¹⁴⁰

The worms' simplicity and amenability to sophisticated, yet convenient, genetic techniques is a big advantage.²⁰⁴ There are long established genetic tools and resources available.^{78,79,205–208} The screening of nonsense and missense mutant strains, which are easily available (*e.g.*, from the *Caenorhabditis Genetics Center*)²⁰⁷ can foster the deconvolution of gene products that are required for life span extension.²⁰⁹ In this way, a plethora of NPs, including emodin, trigonelline, naringin, silymarin flavanols and hibiscus extract, have been shown to extend health span in a *daf-16*-dependent manner.^{200,202,210–212} Other NPs like acetylphenethylamine and ginsenosides require (also) the function of *str-2.1* or *skn-1*,^{132,213,214} whereas orientin and urolithin A require *aak-2*.^{140,202} Another possibility is to employ RNA interference which can be achieved by feeding bacteria expressing gene-specific dsRNA. It allows for knockdown in adults to avoid development defects (*e.g.*, in *daf-2* mutant). With specific promoters, tissue-specific RNA silencing is possible.^{215,216} The worms' transparent body allows for researchers to visualize and quantify molecular and cellular processes like the expression, localization and activity of proteins.²¹⁷ A frequently employed approach is to use transgene worms carrying fluorescent protein tagged transcription factors to test for nuclear translocation in response to treatment.²¹⁸ In this case *daf-16::gfp* (*e.g.*, strain TJ356) and *skn-1::gfp* (*e.g.* strain LD001) are useful to prove an involvement of these pathways. Also transgene worms expressing *hsp-16.2::gfp*, *sod-3::gfp* and *gst-7::gfp* are employed to test the induction of these antioxidant enzymes and stress reporters.²¹⁹ Gene expression levels can also be determined *via* western blot but larger populations (up to 1000 worms) or on RNA level *via* quantitative real-time polymerase chain reaction.¹⁷⁵ A shortcoming of these techniques is that there is no smoking gun in the sense of a discovered specific drug – target binding event. Since that event is causative for the modulation of protein function and the disorganization of a pathway, it results in lifespan extension. This knowledge could foster linking chemo-structural details to health span promoting phenomena at the organism level, which is valuable to structurally optimize compounds towards improved efficacy.

3. Natural compounds with anti-aging effects in *C. elegans*

A variety of natural compounds from almost all structural classes have been investigated in the past ten years for their effects on aging in *C. elegans*. They are presented in ESI Tables ESI1–14† and are grouped into structural classes without making a claim to completeness. Additionally, Tables ESI1–14† provide information on the experiments, their parameters (*e.g.*, dose, strain) in *C. elegans*, the findings of the respective studies and the report of

signalling pathways or genes involved in the observed mechanism. We have gathered data from 160 compounds reported in 85 representative studies that show healthy aging effects in *C. elegans*. The most researched compounds were epigallocatechin gallate (1, thirty experiments), resveratrol (3, seventeen experiments), withanolide A (2, fourteen experiments), hydroxytyrosol (4, fourteen experiments), cannabidiol (5, eleven experiments), quercetin (6, ten experiments), tyrosol (7, ten experiments) and 4-hydroxybenzoic acid (8, ten experiments).



A continued focus on structural classes with phenolic structures were observed (Fig. 2) with 92 out of 166 compounds being phenols. Most of these compounds can be ascribed to the compound classes of flavonoids (fourty compounds),

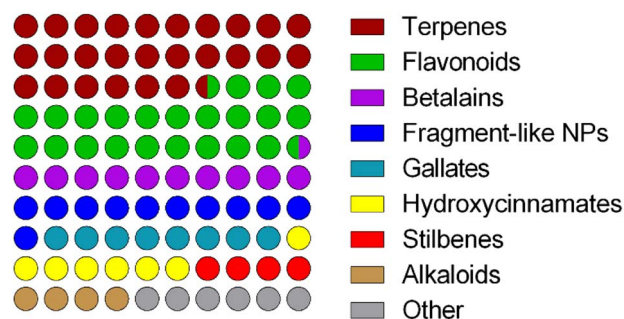


Fig. 2 Structural classes of NPs investigated in *C. elegans* for their health span promoting effects.



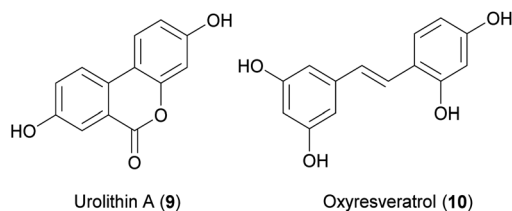
hydroxycinnamates (eleven compounds), tannins (thirteen compounds), and stilbenes (seven compounds). This focus can be attributed to the fact that dietary polyphenols, *e.g.*, from olives, tea, fruits, herbal teas and vegetables have long been suspected of preventing aging associated diseases.

It is also striking that almost one in four compounds were glycosides (thirty-nine compounds) with saponins, iridoids and flavonoids as frequent scaffolds. This is interesting because glycosylated NPs are usually not in the focus of interest for drug discovery projects,²¹⁹ due to their poor bioavailability in humans caused mainly by hydrolysis by the intestinal flora.²⁶² Whether there is a similar metabolism in the *C. elegans* – OP50 model remains elusive. Only two studies had looked into the fate of glycosylated NPs during incubation with bacteria.^{263,264} It would be interesting to determine which metabolites actually arrive in the worm and cause the observed effects.

However, there are also reports on healthy aging promoting terpenes, alkaloids, betalains, diarylheptanoids, anthraquinones and non-proteinogenic amino acids (Fig. 2). A considerable part of the collection was subsumed under the class “fragment-like NPs”.

3.1. Examples for investigated natural compounds

Urolithin A (**9**) is one of the major gut bacterial transformation products of ellagitannins *e.g.*, of pomegranate fruits and berries. In humans the endogenous formation of **9** varies dramatically depending on the individual microbiome composition.²⁶⁵ Therefore, it was suggested that **9** could also be supplemented directly to the diet. An application for **9** as a novel food ingredient in the European Union was filed in 2018.²⁶⁶ Supplemented **9** has shown to promote healthy aging across several species including *C. elegans*, *D. melanogaster*, and *Mus musculus*. The proposed mechanism of supplemented **9** is the induction of mitophagy, which comes with a contradictory improved mitochondrial function and proteostasis in later life, probably due to the elimination of dysfunctional mitochondria. Long-term treatment of *C. elegans* with urolithin A influences the regulation of mitophagy and mitochondrial biogenesis through the transcription factor *skn-1*. It increases mean and median lifespan but also improves other health span parameters like pharyngeal pumping, respiratory capacity, mitochondrial content, muscle fibre organization and locomotion in aged worms. The urolithin A mediated induction of mitophagy is conserved over species and also in aged rodents enhances muscle strength and running endurance.¹⁴⁰ A phase 1 clinical study confirms that urolithin A is bioavailable in humans. A long-term oral consumption (4 weeks period) has been demonstrated to be safe and shows signs of mitochondrial and cellular health improvements also in aged humans.²⁶⁷



The structure class of stilbenes, with its well-known representatives' resveratrol (*trans*-3,4',5-trihydroxystilbene, **3**), oxyresveratrol (**10**) and pterostilbene, has been repeatedly investigated in the context of aging across different species. A study on the life-extending effect of resveratrol in small metazoan organisms including *C. elegans* was published as early as 2004.²⁶⁸ Although resveratrol has been used as a positive control for lifespan experiments in many studies over the last 10 years,^{190,191,269} showing mainly positive effects, there are two studies that could not reproduce this lifespan-extending effect.^{148,193} Also, natural and synthetic derivatives of **3** were investigated: it has been shown that **10** has a stronger life-prolonging effect than **3**,^{195,196,270} whereas pterostilbene has no effect.¹⁴⁸ Several synthesized stilbene derivatives have been reported to increase the lifespan of *C. elegans* more robustly than **3**.¹⁴⁸

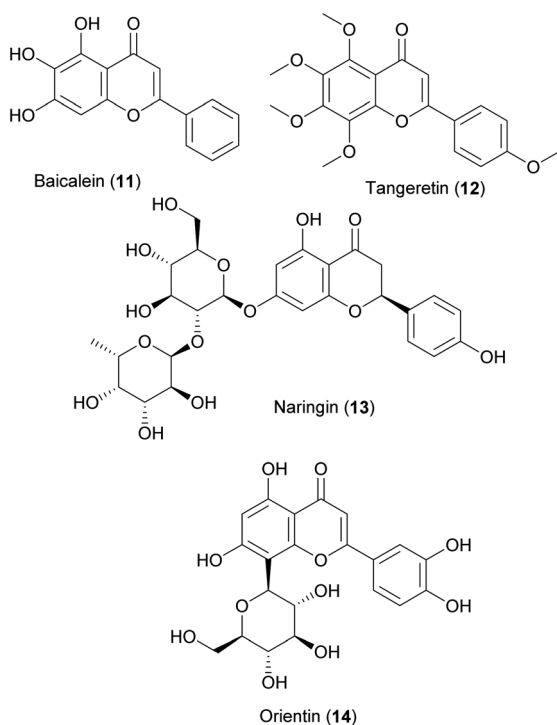
After nearly 20 years of anti-aging research with **3**, it is still highly controversial whether it can extend the lifespan of *C. elegans* or other model organisms.^{148,195,271} Two studies performed with rodents have found no statistically significant life extension upon treatment with **3**.^{45,272} One study reported that **3** improved life- and health span of mice on a high-calorie diet,²⁷³ another noted a positive influence on healthy aging on mice fed a standard diet.²⁷⁴ Sirtuins (*sir-2.1* in *C. elegans*) have been proposed to account for the health span promoting effects of **3** similar to a caloric restriction mimetic.²⁷⁵ Mammal SIRT1 activation of resveratrol was shown *in vitro* and *in vivo*.^{276,277} As caloric restriction can vary widely by genotype or diet, it is a conceivable cause of conflicting *in vivo* results.²⁷⁸ However, the interaction of **3** with sirtuins is also not without controversy as biochemical assays were called into question and results in model organisms are often not reproducible.^{271,279–281} **3** is contained in²⁸² larger amounts in berries of the genus *Vaccinium* (*e.g.*, cranberry, bilberry, cowberry)²⁸³ and redcurrant, as well as in smaller amounts in many other plants such as peanuts, pistachios, apples, tomatoes, grapes and cocoa.²⁸⁴ However, the concentrations of stilbenes in food and that of in this context often cited red wine^{284,285} are low compared to doses of preclinical and clinical studies. Considering also the poor bioavailability of **3**,²⁸⁶ claimed health benefits for different foods are questionable.

Flavonoids are consumed in substantial quantities by humans. Dietary intake is associated with positive effects in epidemiological studies against age-related diseases including diabetes mellitus type II,²⁸⁷ cardiovascular disease^{288,289} and cognition in elderly.²⁹⁰ Therefore, they were among the first research subjects tested on *C. elegans*,^{49,146,291} and they remain a prominent NP-class in the literature over recent years (Fig. 2). Baicalein (**11**), a flavonoid isolated from *Scutellaria baicalensis*, is known to be an activator of the transcription factor Nrf2 in mammalian cell lines.²²² In *C. elegans* it modulated stress-resistance against lethal thermal and sodium arsenite stress and dose-dependently extended the nematodes' lifespan *via* the Nrf2 orthologue *skn-1*. The FOXO orthologue *daf-16* was not involved in these processes.²⁹² The polymethoxylated flavonoid tangeretin (**12**), a constituent of citrus fruits, extended the mean lifespan *C. elegans* at 30 and 100 μ M. It slowed aging related functional declines and increased the resistance against heat-shock stress. On the mRNA expression level, *daf-16*, *hsp-16.2*, and *hsp-16.49* were upregulated. Tangeretin

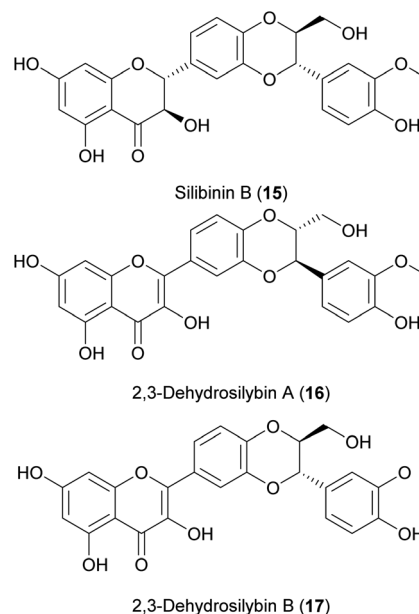


promoted the nuclear localization of *daf-16* and enhanced the expression of *hsp-16.2*. No effects were observed on the lifespan of *daf-2*, *age-1*, and *daf-16* mutants. Hence, it was suggested that besides an extension of lifespan, tangeretin enhances heat stress tolerance in an insulin/insulin-like growth factor signaling dependent manner.¹⁷⁵ Naringin (**13**), a flavanone-7-O-glycoside consisting of the flavanone moiety naringenin and the disaccharide neohesperidose, is another well-known constituent of citrus fruits, in particular of grapefruit. This compound was found to extend the lifespan of *C. elegans*. Moreover, it increased the thermal and oxidative stress tolerance and reduced the accumulation of lipofuscin. In Alzheimer's and Parkinson's disease models, it delayed their progression *via daf-16*.²⁰⁰ Orientin (**14**), the 8-C glucoside of the flavonoid luteolin, occurs in a wide range of medicinal plants, for instance in plants of the genus *Nasturtium*. In *C. elegans*, it increased the lifespan, improved heat, oxidative, and pathogenic stress resistance through the activation of stress responses, including *hsf-1*-mediated heat shock response, *skn-1*-mediated xenobiotic and oxidation response, mitochondria unfolded responses, endoplasmic unfolded protein response, and increased autophagy activity. Moreover, it activated AMPK and *daf-16*. In neurodegenerative disease models of *C. elegans*, it reduced the accumulation of toxic proteins (α -synuclein, β -amyloid, and poly-Q) and delayed the onset of the respective disease.²⁰²

Several studies have shown good bioavailability of flavonoid aglycones along with their extensive metabolism by conjugation in *C. elegans*.²⁹³⁻²⁹⁵ Treatment with quercetin-3-O-glucoside led to higher accumulation of **6** compared to treatment with the aglycone, which shows that the flavonoid glycoside was taken up and deglycosylated by the worm.²⁶⁴

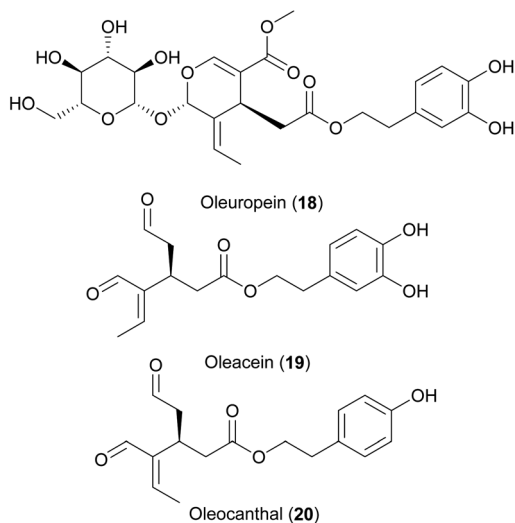


In 2017, Sciacca and co-workers, investigated four pure compounds of the NP flavonoid complex silymarin from milk thistle (*Silybum marianum* (L.) Gaertn.), *i.e.* silybin A, silybin B (**15**), 2,3-dehydrosilybin A (**16**) and 2,3-dehydrosilybin B (**17**), regarding their effects on the inhibition of A β amyloid growth and toxicity in *C. elegans*. Underlining the crucial role of stereochemistry, from all four constituents tested, silybin B was found to be the most effective in counteracting A β proteotoxicity.²⁹⁶ In the same year, Filippopoulou and co-workers published a study investigating the mixture of two flavonolignans: **16** and **17**. The mixture of the compounds was able to extend the lifespan of *C. elegans* depending on *fgt-1* and *daf-16*. In a nematode model of Alzheimer's disease, the compound mixture led to a decrease in the disease progression.²¹¹ However, in this study silybin A and B were not tested.

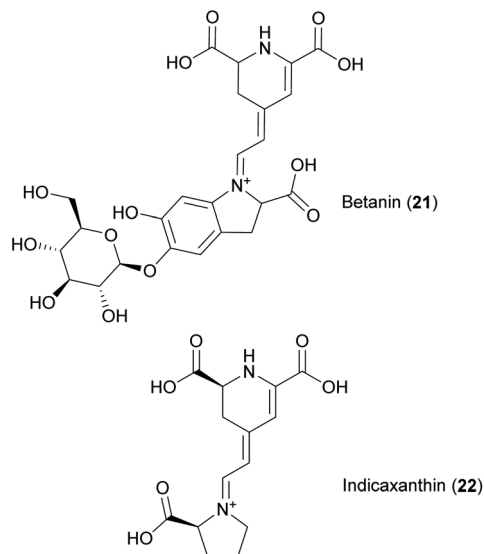


High consumption of extra virgin olive oil (*Olea europaea* L.) as the most representative food of the Mediterranean diet has been associated with longevity and reduced incidences of age-related diseases in many epidemiological and intervention studies.²⁹⁷⁻²⁹⁹ The triglyceride profile of olive oil with a particularly high proportion of esterified monounsaturated ω -9 fatty acids seems to be involved in these health-promoting effect.³⁰⁰ Supplementation of ω -9 fatty acids such as oleic acid can indeed promote longevity in *C. elegans*.³⁰¹ However, this effect seems to be dependent on the test protocol, as other groups did not show this effect.³⁰²⁻³⁰⁴ Olive fruits and leaves also contain an interesting profile of phenolic secondary metabolites which are responsible for bitter and pungent notes in olive oil and include hydroxytyrosol (**4**), tyrosol (**7**), as well as the secoiridoids oleuropein (**18**), oleacein (**19**) and oleocanthal (**20**). **4**, **7** and **18** have shown to improve *C. elegans* life- and health span in numerous studies dependent on *daf-16*, *hsf-1* and *skn-2*.^{218,243,269,305-308} The secoiridoids **19** and **20** have shown beneficial effects against aging and senescence in human fibroblasts and *D. melanogaster* but have never been evaluated in *C. elegans*.^{309,310}





Betalains are water-soluble nitrogen pigments responsible for the colour of different plant parts of *Caryophyllales*. Due to their stability and safety they are popular food additives. Several medicinal plants of the order *Caryophyllales* are used in traditional medicine and their biological actions are sometimes attributed to the betalain constituents. Betanin (betacyanin, 21), the pigment of beetroots, indicaxanthin (betaxanthin, 22) present in *Opuntia* fruits, and 15 other betalains were investigated in *C. elegans* survival assays using the automated lifespan machine and *in vivo* antioxidative assays.^{120,311} Treatment with 25 μM of betalain for 48 h prior to the reproductive period could prolong lifespan up to 30%.³¹¹ These effects were accompanied by increased *sir-2.1* and heat shock proteins expression. In response to betalain treatment, *daf-16* was nuclearized in the transgene worms TJ365, and lifespan extension was abolished in mutant *daf-16* worms. The two studies expand the alimentarium of lifespan prolonging NP with an established structure–activity relationship and mode of action.¹²⁰ However, their experimental setup with treatment restricted to the larval stages complicates comparison with other studies, where it is rather standard to start the treatments in the L4 or adult stadium.



Recently cannabidiol (5) was discovered to influence the aging process of *C. elegans*. In the course of a toxicity screen, *C. elegans* was exposed to different concentrations of cannabidiol and it was revealed that instead of shortening, it increased lifespan up to 18% (at 40 μM). 5 doubled worm motility at old age, and increased their resilience to heat stress.³¹² Similar effects on *C. elegans* and *D. rerio* health span parameters were reported later.^{313–316} Especially, the age-associated decline of neuronal health was slowed by promotion of autophagy *via sir-2.1*, *bec-1* and *sqst-1*. Whether the endocannabinoid system of the worm is involved in these effects remains to be clarified. The *C. elegans* cannabinoid receptor NPR-19 is sensitive to the conserved endocannabinoids 2-arachidonoylglycerol and anandamide. *C. elegans* endocannabinoids are involved in monoaminergic signalling and feeding behaviour,^{317,318} axon regeneration,³¹⁹ development,³²⁰ and the coordination of nutrient status, metabolism and aging.³²¹

4. Multicomponent mixtures from natural sources with anti-aging effects in *C. elegans*

We have gathered data on over 255 different NP extracts/fractions from 163 different plant, fungal and marine species probed in *C. elegans* for healthy aging between 2012 and 2023. The extracts are presented in the ESI† grouped into marine invertebrate- (Table ESI15†), fungal- (Table ESI16†), and plant- (Table ESI17†) sourced samples. The tables provide information on chemical aspects (species, organ, extraction solvent, type of characterization, and description of multicomponent mixture), and information on the experiments in *C. elegans*, important parameters (*e.g.*, dose, strain), the findings of the respective studies and the report of signalling pathways or genes involved in the observed effect. Based on these data we classified the extracts regarding extraction solvent (Fig. 3A) and the level of chemical dissection of the multicomponent mixture (Fig. 3B).

Regarding extraction solvent, most studies used (hydro-) ethanol (40.0%) or (hydro-) methanol (13.9%) for extraction. 21.3% of all multi-component mixtures were aqueous extracts. Combined extracts with a broad polarity range of metabolites were also reported mainly using combined dichloromethane and methanol extracts (11.5%), as well as combined butyl-methyl ether and methanol extracts (3.4%). Only few studies test multi-component mixtures generated by midpolar and nonpolar solvents such as acetone (4.3%) and hexane (2.9%); ethyl acetate, dichloromethane or isopropanol are hardly represented in the literature.

Regarding the chemical profile, the multicomponent mixtures can be classified into five levels: (i) extracts chemically non-characterized, (ii) extracts with certain NP classes (*e.g.*, total phenol, flavonoid, or anthocyanins) quantified with a spectrophotometer, (iii) extracts partly characterized with several annotated constituents and/or main constituents quantified, (iv) extract qualitatively characterized (*e.g.*, with LC-MS(/MS), GC-MS, HPLC-DAD) to obtain a phytochemical profile, and (v) extracts subjected to bioactivity-guided fractionation. Nearly one third of the studies published between 2012 and 2023, do not provide a characterisation



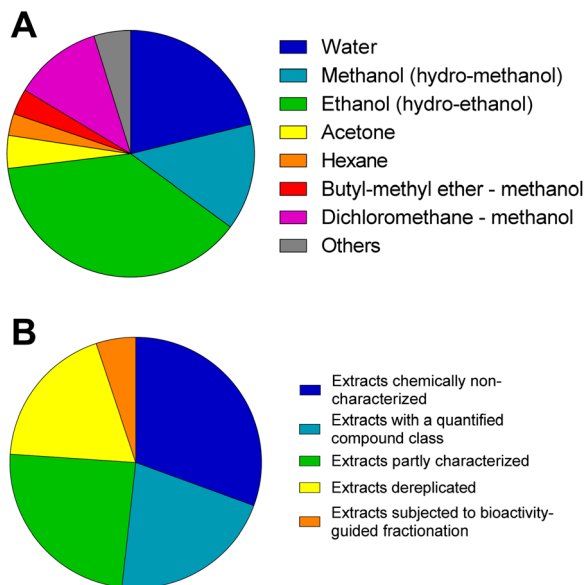


Fig. 3 Analysis of literature dealing with multicomponent mixtures regarding (A) the used solvents for extract preparation ($n = 208$) and (B) their degree of analytical characterization ($n = 255$).

of the tested multicomponent mixtures (30.6%). This is critical, as it severely compromises the reproducibility and interpretation of results – an essential prerequisite for the progress of the research field. A considerable part of the studies performed provide some information about the total phenolic and total flavonoid contents using *e.g.*, Folin–Ciocalteu assay (21.2%). 43.1% of the published literature contains at least one type of chromatographic analysis (*e.g.*, LC-DAD, LC-MS/MS, or GC-MS). In most of these analyses, the phytochemical profile of the respective extract is further characterised by compound annotation using databases or single reference compounds for comparison. The level of characterisation differs significantly among the studies. In most cases the tested extracts have not been further chemically investigated to isolate the constituents contributing to the observed effect. Admittedly, it requires knowledge and instrumentation in analytics, chromatography and characterization of isolated compounds or collaboration partners covering these requirements. In general, the chemical analysis of bioactive extracts is often biased towards well-known antioxidants (*e.g.*, investigation of the phenolic profile) or limited to well-known (and often well-investigated) chemical entities provided by suppliers. This generally inhibits research on novel, unexpected or minor compound classes. From the surveyed studies in this review it is obvious that extracts with already known potential radical scavenging *in vitro* activities are prioritized for *C. elegans* assays.^{57,322} However, approaches relying on *in vitro* antioxidant activities to screen for lifespan extending extracts has limitations. An extract screening study which included 30 different flower extracts concluded that the potency of *in vitro* radical scavenging activity shows no correlation to a prolongation of mean and median lifespan of nematodes.¹²⁷

Regarding the experimental parameters for the *C. elegans* tests, we observed an extremely wide range of tested extract concentrations ($0.1 \mu\text{g ml}^{-1}$ to 240 mg ml^{-1}).^{177,187} The median

tested concentration in *C. elegans* assays of $125 \mu\text{g ml}^{-1}$ is considerably higher than that reported for *in vitro* assays. About 30% of all tested concentrations were even above 10 mg ml^{-1} which raises questions on how these concentrations were achieved in the test media without precipitation, influencing pH and without causing osmotic stress on bacteria and worms. The validity of these experimental protocols and the reported effects have to be considered with caution.

4.1. Examples for investigated NPs

Among the investigated plant materials, a clear tendency towards edible plants can be observed. Especially from nutritional sciences, there is a huge interest in substantiating anti-aging claims of so called “super food”, nutraceuticals, and functional foods. In this respect many studies regarding anti-aging properties in *C. elegans* focus on the investigation of berries, *e.g.*, açai,^{169,257} goji,²⁵⁸ cranberries,^{259,260} blueberries,²⁶¹ raspberries,²⁶² black mulberries,²⁶³ and juniper berries.²⁶⁴ Interestingly, there is also a strong focus on botanicals consumed as beverages, *e.g.*, tea (black,²⁶⁵ green,²⁶⁶ oolong,²⁶⁷ rooibos,²⁶⁸ and mate^{269–271} tea), guaraná,²⁷² hops,²⁷³ and coffee.^{140,274} Another large portion of investigated extracts focuses on industrial waste products *e.g.*, coffee silver skin,²⁷⁵ corncob – a by-product of the corn industry,²⁷⁶ different fruit peels (*e.g.*, from apple^{277–279} or pomegranate^{280,281}). On species level, the most studied plants (number of experiments per species) in recent years was *Camellia sinensis*,^{127,255,256,323–326} followed by *Camellia tenuifolia*,³²⁷ *Ginkgo biloba*,^{187,328–330} *Lycium barbarum* (goji)^{224,225,331} and *Anacardium occidentale* (cashew).²³⁰ Also species of the genus *Vaccinium*^{229,254,332–334} and *Syzygium*^{55,335,336} were repeatedly investigated.

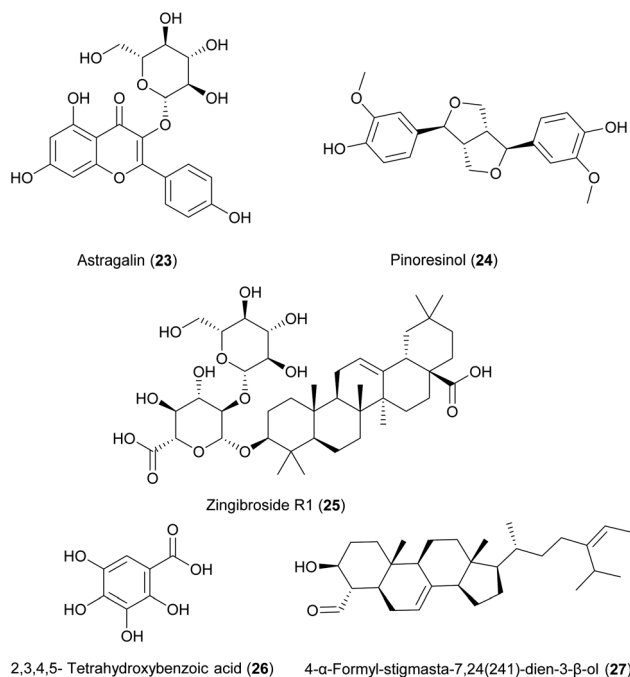
4.2. Following up studies of bioactive extracts

In a few studies, a bioactivity-guided isolation was performed using a *C. elegans* model for the bioactivity evaluation.^{257–259} These articles can be further divided into (i) studies that use *C. elegans* as an *in vivo* model for the confirmation of their *in vitro* results^{337,338} and (ii) studies where *C. elegans* is used as the primary model of bio-guided fractionation.^{339,340}

An example of the latter one is a study of Sayed and co-workers who compared the lifespan extending effects of *Cuscuta chinensis* and *Eucommia ulmoides* extracts generated with a mixture of butyl-methyl ether and methanol (1 : 1) followed by an extraction with 100% methanol. Both extracts improved the resistance towards oxidative stress, and decreased the intracellular level of ROS.¹²⁶ UPLC-Q-Exactive Orbitrap MS analysis allowed for metabolite identification and annotation. The authors also put efforts into unravelling the responsible single components. Hence, the extracts were fractionated *via* RP-HPLC and MPLC. 20 fractions of the *E. ulmoides* extract were again tested in the *C. elegans* heat stress assay. As a result, 7 fractions showed similar activity as the crude extract. 17 fractions of *C. chinensis* were further tested. From the extract of *C. chinensis*, enriched fractions containing astragalgin (23, purity by ¹H NMR ~75%), pinoresinol (24, purity by ¹H NMR ~75%), and zingibroside R1 (25, purity by ¹H NMR ~50%) were obtained. The fraction from the *C. chinensis* extract enriched in



zingibroside R1 improved the life span, the survival after heat stress, and the locomotion in a manner similar to the full *C. chinensis* extract and therefore zingibroside R1 could be (partly) responsible for the observed health benefits of *C. chinensis*. Furthermore, 2,3,4,5-tetrahydroxybenzoic acid (26) and the sterol lipid 4- α -formyl-stigmasta-7,24(241)-dien-3- β -ol (27) are abundantly present in the *C. chinensis* extract and its most bioactive fraction.



Another example of a bioactivity-guided fractionation is the study of Jia and coworkers who followed up on a bioactive extract of clams (*Meretrix meretrix*). They discovered that a peptide-rich extract of *M. meretrix* could increase the survival of worms and oxidative stress conditions induced by paraquat.³³⁹ They separated the extract based on molecular weight into two fractions by ultrafiltration whereby the low-molecular weight peptides showed better activity in the paraquat stress survival assay. Thus, the fraction was further separated into five subfractions by gel filtration chromatography. The first fraction (F1) showed the best activity which decreased over the consecutive fractions. By RP-nano-LC-MS/MS they identified 25 peptides in F1 and subsequently synthesized and tested them. Three peptides were shown to be responsible for the observed effect which was further explained by an increased *daf-16* dependent expression of *sod-3*, *ctl-1* and *ctl-2* after treatment.

4.3. *C. elegans* microbiome and natural products

Maintaining *C. elegans* in an unnatural but simplified and well-characterized culture with inactivated or dead OP50 as food sources is advantageous to directly probe the effects of NPs on worms. However, as in humans, in *C. elegans* its commensal microbial communities play a crucial physiological role.³⁴¹ Some naturally associated bacteria influence behaviour, stress- and infection resistance, fecundity, metabolism, and life span

of *C. elegans*;^{342–347} e.g., different bacterial food sources can cause different uptake/accumulation of flavonoids.²⁹³ Maintenance of *C. elegans* on *Bacillus subtilis* strains present in its natural habitat increased lifespan by more than 50% and increased heat shock survival by more than 200% compared to standard OP50 diet.³⁴⁸

It is well known that many NPs including dietary polyphenols interact with the human gut microbiome. Genuine NPs can be metabolized to different metabolites²⁶⁵ and *vice versa* NPs affect the composition of gut microbiota.^{349,350} Although *C. elegans* commensals are not identical to those of humans,³⁵¹ the worm is proposed as a model for microbiome research.³⁴¹ Recently an experimental microbiome modelling the native worm consisting of 14 bacterial strains was presented.³⁵² Future studies will show how this microbiome interacts with natural products and how this affects worm health.

4.4. Intrinsic complexity of multicomponent NP mixtures and the challenge of combinatorial effects

For the elucidation of biological networks, the discipline of systems biology has integrated -omics approaches (e.g., genomics, transcriptomics, proteomics, and metabolomics) focusing on large-scale data deriving from *C. elegans* assays.³⁵³ However, to answer questions regarding the anti-aging effects of multi-component NPs on *C. elegans*, further disciplines need to be involved. From the NP side, the bottleneck is still the identification and elucidation of the bioactive principle on the molecular level – which compounds are responsible for certain phenotypic effects (e.g., the prolongation of life span). Multi-component mixtures such as extracts generated from natural sources are known to contain hundreds of metabolites. Working with multicomponent mixtures thus requires holistic approaches on both the (phyto-)chemical as well as on the biological level. Regarding the screening of *C. elegans*, NP extracts are not often dissected in detail. Instead, specific (major) constituents known to be present in the respective mixture are exemplarily tested as representatives of the extract. Only in rare cases, the increase in lifespan observed for the single constituent is similar to that observed for the complex mixture, and thus only gives an incomplete picture.

Moreover, NPs are particularly prone to exert combinatorial effects including an amplification of activity (when $1 + 1 > 2$), potentiation (when $0 + 1 > 1$), or antagonism (when $0 + 0 < 0$). Another effect which can be observed is synergy (when $0 + 0 > 0$). In this instance, the combination of multiple constituents is active, while the constituents separately are inactive. Apart from chemical challenges (e.g., poor chromatographic separation, analytical problems), these phenomena have to be considered.^{354,355} A possible case where additive effects may occur, might happen if some extract constituents inhibit efflux pumps, while others are genuinely active but not stable in the organism. In a recent case study in our group, it was found that the action of ostruthin, one of the major active constituents of the extract of masterwort (*Peucedanum ostruthium*), is enhanced by other constituents such as imperatorin and isoimperatorin, which are themselves inactive and presumably act by inhibiting xenobiotic



defenses.³⁰² Such additive effects are the proposed mechanism of many botanical drugs. Traditional phytotherapy not only makes use of molecular mixtures from one plant organ, but also employs complex mixtures of several plants potentiating the chemical complexity. This approach is very similar to Nature's which also employs complex mixtures of metabolites with distinct strategies to modulate biological processes. *C. elegans* might be a key model to unravel the therapeutic effect of such complexity in a living organism with possible implications for mammals too.

When following up on a bioactive extract, the goal should be to minimize the isolation of unwanted, inactive, or already well-investigated compounds. Traditionally, bioactivity-guided fractionation is one of the most common techniques applied to isolate and identify the bioactive principle of a multicomponent mixture. Successful examples, where this technique was used in combination with a *C. elegans* set-up were described before.^{322,327,329,338–340,356} Interestingly, further techniques to dissect anti-aging multicomponent NPs using *C. elegans* as model organism are scarce in the scientific literature.

Useful approaches might be high-performance liquid chromatography (HPLC)-based (micro)-fractionation,³⁵⁷ MS-based techniques such as molecular networking,³⁵⁸ or biochemometric approaches, where bioactivity data is correlated with chemical data from spectroscopy (e.g., NMR) or spectrometry (e.g., MS). One such example is the biochemometric approach ELINA (Eliciting Nature's Activities) which correlates activity with ¹H NMR data to detect spectral features responsible for an observed effect.^{359–361} With this technique, it is possible to distinguish bioactives from inactives prior to isolation, as shown by the example of masterwort.³⁶²

5. Conclusions

An analysis of the literature shows that reports differ greatly in terms of the experimental focus. Many studies have elaborately established multiple assays to determine the function of the extracts not only on lifespan, but also on abiotic stress resistance, motility, proteotoxicity and more. Many mutant and transgenic *C. elegans* strains are widely established to unravel involved genes and pathways. However, a large part of studies does not deal with the chemical characterization or the isolation of constituents of the tested multicomponent mixtures. On the other hand, awareness has increased during the past years that extracts are chemically complex. Their composition can vary tremendously depending on the underlying material and its preparation. Accordingly, a chemical characterization of extracts used for any pharmacological investigation is mandatory.³⁶³

There is also a strong bias towards well-studied structural NP classes such as flavonoids, stilbenes and tannins. This bias is not unusual in NP drug discovery. The same structures are also overrepresented in hit lists of *in vitro* bioactivity screenings. These compounds are often termed pan-assay interference compounds (PAINS) or invalid metabolic panaceas (IMPs).^{364,365} The PAINS term was originally coined for high-throughput screenings employing recombinant enzymes in which flavonoids, catechol and other phenols frequently inhibited enzyme activity through aggregation or oxidation rather than by specific

interaction. Therefore, the PAINS concept cannot be translated to an *in vivo* model. However, there are indications that the effect of many phenols is also not a specific pharmacological effect. It is known that some polyphenols have moderate bactericidal effects,^{203,366,367} which might cause the lower availability of the bacterial nutritional source in *C. elegans* assays, at least in studies applying mg ml⁻¹ doses of NPs. Thus, observed lifespan effects are likely to be caused by dietary restriction or the mitigation of bacterial colonization of the intestine in aged worms. Interestingly, many polyphenols are investigated for anthelmintic effects caused by tanning of cuticle, buccal cavity and intestine²⁹⁷ which at certain concentrations might also lead to a dietary restriction mediated lifespan increase. However, the strong focus of scientific research on polyphenols is impacted by their essential role in food, as well as their relatively easy acquisition; quite the opposite of more "exotic" or novel natural compounds. Nevertheless, efforts should be at least partially directed towards new structural classes for health span increase in *C. elegans*.

To achieve this goal, the authors highly recommend investigating promising extracts in more detail, e.g., through -omics studies. By introducing the term "wormomics", a new workflow for the discovery of anti-aging NPs could be established. The nematodes have multiple advantages that allow for setting up experiments to discover (new) bioactive constituents of extracts prior to their isolation, e.g., by MS/MS molecular networking, NMR- or HPLC/MS-based bioactivity profiling and biochemometric approaches. These advantages can best be exploited by high content miniaturized screenings in multiwell-plates to test samples in parallel in a higher throughput, but also by the wide range of well-established assays that are increasingly automated. It is expected that extracts with health-promoting effects in *C. elegans* warrant further employment in particular with respect to the use of high resolution analytical techniques and big data analysis to provide insight into new chemistries and thus to unveil hidden treasures in complex mixtures.

6. Conflicts of interest

There are no conflicts to declare.

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8. References

- 1 E. J. Stewart, R. Madden, G. Paul and F. Taddei, *PLoS Biol.*, 2005, **3**, e45.
- 2 A. Currais, *Ageing Res. Rev.*, 2017, **35**, 297–300.
- 3 R. Z. Moger-Reischer and J. T. Lennon, *Nat. Rev. Microbiol.*, 2019, **17**, 679–690.
- 4 G. Björklund, M. Shanida, R. Lysiuk, M. Butnariu, M. Peana, I. Sarac, O. Strus, K. Smetanina and S. Chirumbolo, *Molecules*, 2022, **27**, 7084.
- 5 S. I. S. Rattan, *Biogerontology*, 2013, **14**, 673–677.
- 6 C. J. Kenyon, *Nature*, 2010, **464**, 504–512.



- 7 M. R. Klass, *Mech. Ageing Dev.*, 1983, **22**, 279–286.
- 8 C. Kenyon, J. Chang, E. Gensch, A. Rudner and R. Tabtiang, *Nature*, 1993, **366**, 461–464.
- 9 D. B. Friedman and T. E. Johnson, *Genetics*, 1988, **118**, 75–86.
- 10 Y. Suh, G. Atzmon, M. O. Cho, D. Hwang, B. Liu, D. J. Leahy, N. Barzilai and P. Cohen, *Proc. Natl. Acad. Sci. U. S. A.*, 2008, **105**, 3438–3442.
- 11 B. J. Willcox, T. A. Donlon, Q. He, R. Chen, J. S. Grove, K. Yano, K. H. Masaki, D. C. Willcox, B. Rodriguez and J. D. Curb, *Proc. Natl. Acad. Sci. U. S. A.*, 2008, **105**, 13987–13992.
- 12 L. Pawlikowska, D. Hu, S. Huntsman, A. Sung, C. Chu, J. Chen, A. H. Joyner, N. J. Schork, W. C. Hsueh, A. P. Reiner, B. M. Psaty, G. Atzmon, N. Barzilai, S. R. Cummings, W. S. Browner, P. Y. Kwok and E. Ziv, *Ageing Cell*, 2009, **8**, 460–472.
- 13 M. Holzenberger, J. Dupont, B. Ducos, P. Leneuve, A. Gélöën, P. C. Even, P. Cervera and Y. Le Bouc, *Nature*, 2003, **421**, 182–187.
- 14 L. Guarente and C. Kenyon, *Nature*, 2000, **408**, 255–262.
- 15 A. Martin-Montalvo, E. M. Mercken, S. J. Mitchell, H. H. Palacios, P. L. Mote, M. Scheibye-Knudsen, A. P. Gomes, T. M. Ward, R. K. Minor, M. J. Blouin, M. Schwab, M. Pollak, Y. Zhang, Y. Yu, K. G. Becker, V. A. Bohr, D. K. Ingram, D. A. Sinclair, N. S. Wolf, S. R. Spindler, M. Bernier and R. de Cabo, *Nat. Commun.*, 2013, **4**, 2192.
- 16 R. Strong, R. A. Miller, A. Antebi, C. M. Astle, M. Bogue, M. S. Denzel, E. Fernandez, K. Flurkey, K. L. Hamilton, D. W. Lamming, M. A. Javors, J. P. de Magalhães, P. A. Martinez, J. M. McCord, B. F. Miller, M. Müller, J. F. Nelson, J. Ndikum, G. E. Rainger, A. Richardson, D. M. Sabatini, A. B. Salmon, J. W. Simpkins, W. T. Steegenga, N. L. Nadon and D. E. Harrison, *Ageing Cell*, 2016, **15**, 872–884.
- 17 R. Strong, R. A. Miller, C. J. Cheng, J. F. Nelson, J. Gelfond, S. K. Allani, V. Diaz, A. O. Dorigatti, J. Dorigatti, E. Fernandez, A. Galecki, B. Ginsburg, K. L. Hamilton, M. A. Javors, K. Kornfeld, M. Kaerberlein, S. Kumar, D. B. Lombard, M. Lopez-Cruzan, B. F. Miller, P. Rabinovitch, P. Reifsnyder, N. A. Rosenthal, M. A. Bogue, A. B. Salmon, Y. Suh, E. Verdin, H. Weissbach, J. Newman, F. Maccchiarini and D. E. Harrison, *Ageing Cell*, 2022, **21**, e13724.
- 18 D. E. Harrison, R. Strong, Z. D. Sharp, J. F. Nelson, C. M. Astle, K. Flurkey, N. L. Nadon, J. E. Wilkinson, K. Frenkel, C. S. Carter, M. Pahor, M. A. Javors, E. Fernandez and R. A. Miller, *Nature*, 2009, **460**, 392–395.
- 19 J. P. de Magalhães, M. Stevens and D. Thornton, *Trends Biotechnol.*, 2017, **35**, 1062–1073.
- 20 WHO, *World Health Statistics Overview 2019: Monitoring Health for the SDGs, Sustainable Development Goals*, 2019, <https://apps.who.int/iris/bitstream/handle/10665/311696/WHO-DAD-2019.1-eng.pdf>, accessed 13.01.2023.
- 21 J. Oeppen and J. W. Vaupel, *Science*, 2002, **296**, 1029–1031.
- 22 A. J. Scott, M. Ellison and D. A. Sinclair, *Nat. Aging*, 2021, **1**, 616–623.
- 23 Y.-C. Tsai, L.-H. Cheng, Y.-W. Liu, O.-J. Jeng and Y.-K. Lee, *Biosci. Microbiota, Food Health*, 2021, **40**, 1–11.
- 24 C. Ermogenous, C. Green, T. Jackson, M. Ferguson and J. M. Lord, *Drug Discovery Today*, 2020, **25**, 1403–1415.
- 25 K. A. Steinkraus, E. D. Smith, C. Davis, D. Carr, W. R. Pendergrass, G. L. Sutphin, B. K. Kennedy and M. Kaerberlein, *Ageing Cell*, 2008, **7**, 394–404.
- 26 E. Cohen, J. Bieschke, R. M. Perciavalle, J. W. Kelly and A. Dillin, *Science*, 2006, **313**, 1604–1610.
- 27 N. Arantes-Oliveira, J. R. Berman and C. Kenyon, *Science*, 2003, **302**, 611.
- 28 C. Selman, S. Lingard, A. I. Choudhury, R. L. Batterham, M. Claret, M. Clements, F. Ramadani, K. Okkenhaug, E. Schuster and E. Blanc, *FASEB J.*, 2008, **22**, 807–818.
- 29 J. A. Rollins, A. C. Howard, S. K. Dobbins, E. H. Washburn and A. N. Rogers, *J. Gerontol.*, 2017, **72**, 473–480.
- 30 J. R. Beard, A. Officer, I. A. de Carvalho, R. Sadana, A. M. Pot, J.-P. Michel, P. Lloyd-Sherlock, J. E. Epping-Jordan, G. M. E. E. Peeters, W. R. Mahanani, J. A. Thiagarajan and S. Chatterji, *Lancet*, 2016, **387**, 2145–2154.
- 31 B. K. Kennedy, S. L. Berger, A. Brunet, J. Campisi, A. M. Cuervo, E. S. Epel, C. Franceschi, G. J. Lithgow, R. I. Morimoto, J. E. Pessin, T. A. Rando, A. Richardson, E. E. Schadt, T. Wyss-Coray and F. Sierra, *Cell*, 2014, **159**, 709–713.
- 32 A. Yabluchanskiy, Z. Ungvari, A. Csiszar and S. Tarantini, *Physiol. Int.*, 2018, **105**, 298–308.
- 33 D. R. Seals, J. N. Justice and T. J. LaRocca, *J. Physiol.*, 2016, **594**, 2001–2024.
- 34 C. López-Otín, M. A. Blasco, L. Partridge, M. Serrano and G. Kroemer, *Cell*, 2013, **153**, 1194–1217.
- 35 L. M. DeVito, N. Barzilai, A. M. Cuervo, L. J. Niedernhofer, S. Milman, M. Levine, D. Promislow, L. Ferrucci, G. A. Kuchel, J. Mannick, J. Justice, M. M. Gonzales, J. L. Kirkland, P. Cohen and J. Campisi, *Ann. N. Y. Acad. Sci.*, 2022, **1507**, 70–83.
- 36 J. N. Justice, L. Niedernhofer, P. D. Robbins, V. R. Aroda, M. A. Espeland, S. B. Kritchevsky, G. A. Kuchel and N. Barzilai, *Cardiovasc. Endocrinol. Metab.*, 2018, **7**, 80–83.
- 37 S. I. Rattan, *Biogerontology*, 2013, **14**, 673–677.
- 38 V. Codd, C. P. Nelson, E. Albrecht, M. Mangino, J. Deelen, J. L. Buxton, J. J. Hottenga, K. Fischer, T. Esko, I. Surakka, L. Broer, D. R. Nyholt, I. Mateo Leach, P. Salo, S. Hägg, M. K. Matthews, J. Palmen, G. D. Norata, P. F. O'Reilly, D. Saleheen, N. Amin, A. J. Balmforth, M. Beekman, R. A. de Boer, S. Böhringer, P. S. Braund, P. R. Burton, A. J. de Craen, M. Denniff, Y. Dong, K. Douroudis, E. Dubinina, J. G. Eriksson, K. Garlaschelli, D. Guo, A. L. Hartikainen, A. K. Henders, J. J. Houwing-Duistermaat, L. Kananen, L. C. Karssen, J. Kettunen, N. Klopp, V. Lagou, E. M. van Leeuwen, P. A. Madden, R. Mägi, P. K. Magnusson, S. Männistö, M. I. McCarthy, S. E. Medland, E. Mihailov, G. W. Montgomery, B. A. Oostra, A. Palotie, A. Peters, H. Pollard, A. Pouta, I. Prokopenko, S. Ripatti, V. Salomaa, H. E. Suchiman,



- A. M. Valdes, N. Verweij, A. Viñuela, X. Wang, H. E. Wichmann, E. Widen, G. Willemsen, M. J. Wright, K. Xia, X. Xiao, D. J. van Veldhuisen, A. L. Catapano, M. D. Tobin, A. S. Hall, A. I. Blakemore, W. H. van Gilst, H. Zhu, J. Erdmann, M. P. Reilly, S. Kathiresan, H. Schunkert, P. J. Talmud, N. L. Pedersen, M. Perola, W. Ouwehand, J. Kaprio, N. G. Martin, C. M. van Duijn, I. Hovatta, C. Gieger, A. Metspalu, D. I. Boomsma, M. R. Jarvelin, P. E. Slagboom, J. R. Thompson, T. D. Spector, P. van der Harst and N. J. Samani, *Nat. Genet.*, 2013, **45**, 422–427.
- 39 B. A. Benayoun, E. A. Pollina and A. Brunet, *Nat. Rev. Mol. Cell Biol.*, 2015, **16**, 593–610.
- 40 D. Gems and J. P. de Magalhães, *Ageing Res. Rev.*, 2021, **70**, 101407.
- 41 D. F. Lawler, B. T. Larson, J. M. Ballam, G. K. Smith, D. N. Biery, R. H. Evans, E. H. Greeley, M. Segre, H. D. Stowe and R. D. Kealy, *Br. J. Nutr.*, 2008, **99**, 793–805.
- 42 J. A. Mattison, R. J. Colman, T. M. Beasley, D. B. Allison, J. W. Kemnitz, G. S. Roth, D. K. Ingram, R. Weindruch, R. de Cabo and R. M. Anderson, *Nat. Commun.*, 2017, **8**, 14063.
- 43 M. Blüher, B. B. Kahn and C. R. Kahn, *Science*, 2003, **299**, 572–574.
- 44 H. Kurosu, M. Yamamoto, J. D. Clark, J. V. Pastor, A. Nandi, P. Gurnani, O. P. McGuinness, H. Chikuda, M. Yamaguchi, H. Kawaguchi, I. Shimomura, Y. Takayama, J. Herz, C. R. Kahn, K. P. Rosenblatt and M. Kuro-o, *Science*, 2005, **309**, 1829–1833.
- 45 R. A. Miller, D. E. Harrison, C. M. Astle, J. A. Baur, A. R. Boyd, R. de Cabo, E. Fernandez, K. Flurkey, M. A. Javors, J. F. Nelson, C. J. Orihuela, S. Pletcher, Z. D. Sharp, D. Sinclair, J. W. Starnes, J. E. Wilkinson, N. L. Nadon and R. Strong, *J. Gerontol., Ser. A*, 2011, **66**, 191–201.
- 46 F. Sierra, *Cold Spring Harbor Perspect. Med.*, 2016, **6**, a025163.
- 47 B. M. Egan, A. Scharf, F. Pohl and K. Kornfeld, *Front. Pharmacol.*, 2022, **13**, 938650.
- 48 D. J. Newman and G. M. Cragg, *J. Nat. Prod.*, 2020, **83**, 770–803.
- 49 Z. Wu, J. V. Smith, V. Paramasivam, P. Butko, I. Khan, J. R. Cypser and Y. Luo, *Cell. Mol. Biol.*, 2002, **48**, 725–731.
- 50 J. A. Joseph, B. Shukitt-Hale, N. A. Denisova, R. L. Prior, G. Cao, A. Martin, G. Tagliamonte and P. C. Bickford, *J. Neurosci.*, 1998, **18**, 8047–8055.
- 51 M. A. Wilson, B. Shukitt-Hale, W. Kalt, D. K. Ingram, J. A. Joseph and C. A. Wolkow, *Ageing Cell*, 2006, **5**, 59–68.
- 52 N. Saul, S. Möller, F. Cirulli, A. Berry, W. Luyten and G. Fuellen, *Biogerontology*, 2021, **22**, 215–236.
- 53 H. van Hattum and H. Waldmann, *J. Am. Chem. Soc.*, 2014, **136**, 11853–11859.
- 54 S. C. Sukuru, J. L. Jenkins, R. E. Beckwith, J. Scheiber, A. Bender, D. Mikhailov, J. W. Davies and M. Glick, *J. Biomol. Screening*, 2009, **14**, 690–699.
- 55 B. Squiban and C. L. Kurz, *Curr. Drug Targets*, 2011, **12**, 967–977.
- 56 M. P. de Torre, R. Y. Cavero, M. I. Calvo and J. L. Vizmanos, *Antioxidants*, 2019, **8**, 142.
- 57 J. Chen, J. Zhang, Y. Xiang, L. Xiang, Y. Liu, X. He, X. Zhou, X. Liu and Z. Huang, *Food Funct.*, 2016, **7**, 943–952.
- 58 E. S. David, in *Bioactive Compounds in Phytomedicine*, ed. R. Iraj, IntechOpen, Rijeka, 2012, ch. 8, DOI: [10.5772/26102](https://doi.org/10.5772/26102).
- 59 J. A. Joseph, B. Shukitt-Hale, N. A. Denisova, D. Bielinski, A. Martin, J. J. McEwen and P. C. Bickford, *J. Neurosci.*, 1999, **19**, 8114–8121.
- 60 S. Suter and M. Lucock, *Explor. Res. Hypothesis Med.*, 2017, **2**, 1–7.
- 61 A. Argyropoulou, N. Aligiannis, I. P. Trougakos and A. L. Skaltsounis, *Nat. Prod. Rep.*, 2013, **30**, 1412–1437.
- 62 K. Pallauf, N. Duckstein and G. Rimbach, *Proc. Natl. Acad. Sci. U.S.A.*, 2017, **76**, 145–162.
- 63 A. J. Ding, S. Q. Zheng, X. B. Huang, T. K. Xing, G. S. Wu, H. Y. Sun, S. H. Qi and H. R. Luo, *Nat. Prod. Bioprospect.*, 2017, **7**, 335–404.
- 64 K. Matsunami, *Front. Nutr.*, 2018, **5**, 111.
- 65 D. Chattopadhyay and K. Thirumurugan, *Mech. Ageing Dev.*, 2018, **171**, 47–57.
- 66 L. Wang, X. Zuo, Z. Ouyang, P. Qiao and F. Wang, *J. Evidence-Based Complementary Altern. Med.*, 2021, 5591573.
- 67 Y. Ye, Q. Gu and X. Sun, *Compr. Rev. Food Sci. Food Saf.*, 2020, **19**, 3084–3105.
- 68 J. Martel, C.-Y. Wu, H.-H. Peng, Y.-F. Ko, H.-C. Yang, J. D. Young and D. M. Ojcius, *Microb. Cell*, 2020, **7**, 255–269.
- 69 M. D. Navarro-Hortal, J. M. Romero-Márquez, S. Osta, V. Jiménez-Trigo, P. Muñoz-Ollero and A. Varela-López, *Diseases*, 2022, **10**, 28.
- 70 P. Shen, Y. Yue and Y. Park, *Crit. Rev. Food Sci. Nutr.*, 2018, **58**, 741–754.
- 71 P. Shen, Y. Yue, J. Zheng and Y. Park, *Annu. Rev. Food Sci. Technol.*, 2018, **9**, 1–22.
- 72 A. Zhu, F. Zheng, W. Zhang, L. Li, Y. Li, H. Hu, Y. Wu, W. Bao, G. Li, Q. Wang and H. Li, *Antioxidants*, 2022, **11**, 705.
- 73 M. Greener, *Prescriber*, 2021, **32**, 29–32.
- 74 L. P. O'Reilly, C. J. Luke, D. H. Perlmutter, G. A. Silverman and S. C. Pak, *Adv. Drug Delivery Rev.*, 2014, **69**, 247–253.
- 75 J. Zwirchmayr, B. Kirchweiger, T. Lehner, A. Tahir, D. Pretsch and J. M. Rollinger, *Sci. Rep.*, 2020, **10**, 12323.
- 76 S. Brenner, *Genetics*, 1974, **77**, 71–94.
- 77 P. M. Meneely, C. L. Dahlberg and J. K. Rose, *Curr. Protoc. Essent. Lab. Tech.*, 2019, **19**, e35.
- 78 I. Antoshechkin and P. W. Sternberg, *Nat. Rev. Genet.*, 2007, **8**, 518–532.
- 79 T. Kaletta and M. O. Hengartner, *Nat. Rev. Drug Discovery*, 2006, **5**, 387–398.
- 80 W. Kim, R. S. Underwood, I. Greenwald and D. D. Shaye, *Genetics*, 2018, **210**, 445–461.
- 81 B. Chakravarty, *Nutr. Res.*, 2022, **106**, 47–59.
- 82 S. E. Hulme and G. M. Whitesides, *Angew. Chem., Int. Ed.*, 2011, **50**, 4774–4807.
- 83 S. J. Husson, S. Moyson, D. Valkenborg, G. Baggerman and I. Mertens, *Biochem. Biophys. Res. Commun.*, 2015, **468**, 519–524.



- 84 K. Horvath, D. Angeletti, G. Nascetti and C. Carere, *Ann. Ist. Super. Sanita*, 2013, **49**, 9–17.
- 85 A. Desalermos, M. Muhammed, J. Glavis-Bloom and E. Mylonakis, *Expert Opin. Drug Discovery*, 2011, **6**, 645–652.
- 86 B. S. Samuel, H. Rowedder, C. Braendle, M.-A. Félix and G. Ruvkun, *Proc. Natl. Acad. Sci. U.S.A.*, 2016, **113**, E3941–E3949.
- 87 M.-A. Félix and F. Duveau, *BMC Biol.*, 2012, **10**, 59.
- 88 D. H. Hall and Z. F. Altun, *C. elegans Atlas*, Cold Spring Harbor Laboratory Press, 2007.
- 89 L. R. Baugh, *Genetics*, 2013, **194**, 539–555.
- 90 J. J. Stastna, A. D. Yiapanas, A. A. Mandawala, K. E. Fowler and S. C. Harvey, *Cryobiology*, 2020, **92**, 86–91.
- 91 M. Porta-de-la-Riva, L. Fontrodona, A. Villanueva and J. Cerón, *J. Visualized Exp.*, 2012, **64**, e4019.
- 92 H. B. Atakan, F. Ayhan and M. A. M. Gijs, *Lab Chip*, 2020, **20**, 155–167.
- 93 H. Xiong, C. Pears and A. Woollard, *Sci. Rep.*, 2017, **7**, 9839.
- 94 A. P. Page and I. Johnstone, *WormBook*, 2007, DOI: [10.1895/wormbook.1.138.1](https://doi.org/10.1895/wormbook.1.138.1).
- 95 B.-M. Song and L. Avery, *Worm*, 2013, **2**, e21833.
- 96 D. Raizen, B. M. Song, N. Trojanowski and Y. J. You, *WormBook*, 2012, DOI: [10.1895/wormbook.1.154.1](https://doi.org/10.1895/wormbook.1.154.1).
- 97 K. S. Lee, S. Iwanir, R. B. Kopito, M. Scholz, J. A. Calarco, D. Biron and E. Levine, *Nat. Commun.*, 2017, **8**, 14221.
- 98 J. D. McGhee, *Wiley Interdiscip. Rev.: Dev. Biol.*, 2013, **2**, 347–367.
- 99 M. Ezcurra, A. Benedetto, T. Sornda, A. F. Gilliat, C. Au, Q. Zhang, S. van Schelt, A. L. Petrache, H. Wang, Y. de la Guardia, S. Bar-Nun, E. Tyler, M. J. Wakelam and D. Gems, *Curr. Biol.*, 2018, **28**, 2544–2556.
- 100 A. Buis, S. Bellemin, J. Goudeau, L. Monnier, N. Loiseau, H. Guillou and H. Aguilaniu, *Cell Rep.*, 2019, **28**, 1041–1049.
- 101 D. G. Moerman and A. Fire, in *C. elegans II*, ed. D. L. Riddle, T. Blumenthal, B. J. Meyer and J. R. Priess, Cold Spring Harbor Laboratory Press Copyright © 1997, Cold Spring Harbor Laboratory Press, Cold Spring Harbor (NY), 1997.
- 102 A. L. Kauffman, J. M. Ashraf, M. R. Corces-Zimmerman, J. N. Landis and C. T. Murphy, *PLoS Biol.*, 2010, **8**, e1000372.
- 103 T. Sornda, M. Ezcurra, C. Kern, E. R. Galimov, C. Au, Y. de la Guardia and D. Gems, *J. Gerontol.*, 2019, **74**, 1180–1188.
- 104 C. C. Kern, S. Townsend, A. Salzmänn, N. B. Rendell, G. W. Taylor, R. M. Comisel, L. C. Foukas, J. Bähler and D. Gems, *Nat. Commun.*, 2021, **12**, 5801.
- 105 T. Komura, M. Yamanaka, K. Nishimura, K. Hara and Y. Nishikawa, *npj Aging Mech. Dis.*, 2021, **7**, 12.
- 106 L. E. T. Zhou, S. Koh, M. Chuang, R. Sharma, N. Pujol, A. D. Chisholm, C. Eroglu, H. Matsunami and D. Yan, *Neuron*, 2018, **97**, 125–138.
- 107 C. J. Smith, J. D. Watson, W. C. Spencer, T. O'Brien, B. Cha, A. Albeg, M. Treinin and D. M. Miller, *Dev. Biol.*, 2010, **345**, 18–33.
- 108 L. A. Herndon, P. J. Schmeissner, J. M. Dudaronek, P. A. Brown, K. M. Listner, Y. Sakano, M. C. Paupard, D. H. Hall and M. Driscoll, *Nature*, 2002, **419**, 808–814.
- 109 D. K. Chow, C. F. Glenn, J. L. Johnston, I. G. Goldberg and C. A. Wolkow, *Exp. Gerontol.*, 2006, **41**, 252–260.
- 110 C. Huang, C. Xiong and K. Kornfeld, *Proc. Natl. Acad. Sci. U.S.A.*, 2004, **101**, 8084–8089.
- 111 J. J. Collins, C. Huang, S. Hughes and K. Kornfeld, in *Wormbook*, 2007, pp. 1–21.
- 112 M. D. McGee, D. Weber, N. Day, C. Vitelli, D. Crippen, L. A. Herndon, D. H. Hall and S. Melov, *Aging Cell*, 2011, **10**, 699–710.
- 113 L. Herndon, C. Wolkow, M. Driscoll and D. Hall, in *WormAtlas*, 2017, pp. 9–39, DOI: [10.1007/978-3-319-44703-2_2](https://doi.org/10.1007/978-3-319-44703-2_2).
- 114 C. Coburn, E. Allman, P. Mahanti, A. Benedetto, F. Cabreiro, Z. Pincus, F. Matthijssens, C. Araiz, A. Mandel, M. Vlachos, S. A. Edwards, G. Fischer, A. Davidson, R. E. Pryor, A. Stevens, F. J. Slack, N. Tavernarakis, B. P. Braeckman, F. C. Schroeder, K. Nehrke and D. Gems, *PLoS Biol.*, 2013, **11**, e1001613.
- 115 L. Herndon, C. Wolkow, M. Driscoll and D. Hall, in *WormAtlas*, 2018, DOI: [10.3908/wormatlas.8.4](https://doi.org/10.3908/wormatlas.8.4).
- 116 L. Gonzalez-Moragas, A. Roig and A. Laromaine, *Adv. Colloid Interface Sci.*, 2015, **219**, 10–26.
- 117 H.-E. H. Park, Y. Jung and S.-J. V. Lee, *Mol. Cells*, 2017, **40**, 90–99.
- 118 F. R. G. Amrit, R. Ratnappan, S. A. Keith and A. Ghazi, *Methods*, 2014, **68**, 465–475.
- 119 N. Stroustrup, B. E. Ulmschneider, Z. M. Nash, I. F. López-Moyado, J. Apfeld and W. Fontana, *Nat. Methods*, 2013, **10**, 665–670.
- 120 M. A. Guerrero-Rubio, S. Hernandez-Garcia, J. Escibano, M. Jimenez-Atienzar, J. Cabanes, F. Garcia-Carmona and F. Gandia-Herrero, *Food Chem.*, 2020, **330**, 127228.
- 121 B. Xian, J. Shen, W. Chen, N. Sun, N. Qiao, D. Jiang, T. Yu, Y. Men, Z. Han, Y. Pang, M. Kaeberlein, Y. Huang and J.-D. J. Han, *Aging Cell*, 2013, **12**, 398–409.
- 122 D. P. Felker, C. E. Robbins and M. A. McCormick, *Transl. Med. Aging*, 2020, **4**, 1–10.
- 123 J.-H. Hahm, S. Kim, R. DiLoreto, C. Shi, S.-J. V. Lee, C. T. Murphy and H. G. Nam, *Nat. Commun.*, 2015, **6**, 8919.
- 124 A. Jushaj, M. Churgin, B. Yao, M. De La Torre, C. Fang-Yen and L. Temmerman, *PLoS One*, 2020, **15**, e0229583.
- 125 H. N. Kim, H. W. Seo, B. S. Kim, H. J. Lim, H. N. Lee, J. S. Park, Y. J. Yoon, J. W. Oh, M. J. Oh, J. Kwon, C. H. Oh, D. S. Cha and H. Jeon, *Nat. Prod. Sci.*, 2015, **21**, 128–133.
- 126 S. M. A. Sayed, S. Alseekh, K. Siems, A. R. Fernie, W. Luyten, C. Schmitz-Linneweber and N. Saul, *Nutrients*, 2022, **14**, 4199.
- 127 Q. Chen, X. Yang, E. Capanoglu, A. T. Amrouche, L. Wu, J. Luo, Y. Zhu, Y. Wang, X. Jiang, D. Zhang and B. Lu, *Food Funct.*, 2023, **14**, 457–470.
- 128 Y.-X. Chen, P. T. N. Le, T.-T. Tzeng, T.-H. Tran, A. T. Nguyen, I. H.-J. Cheng, C.-Y. F. Huang, Y.-J. Shiao and T.-T. Ching, *Nutrients*, 2021, **13**, 4317.
- 129 K. Koch, N. Weldle, S. Baier, C. Buechter and W. Waetjen, *Eur. J. Nutr.*, 2020, **59**, 137–150.
- 130 W. Zhang, B. Zheng, N. Deng, H. Wang, T. Li and R. H. Liu, *J. Food Sci.*, 2020, **85**, 4367–4376.



- 131 S. Jiang, N. Deng, B. Zheng, T. Li and R. H. Liu, *Food Funct.*, 2021, **12**, 4471–4483.
- 132 H. Wang, S. Zhang, L. Zhai, L. Sun, D. Zhao, Z. Wang and X. Li, *Food Funct.*, 2021, **12**, 6793–6808.
- 133 M. Tao, R. Li, T. Xu, Z. Zhang, T. Wu, S. Pan and X. Xu, *Food Funct.*, 2021, **12**, 8196–8207.
- 134 R. J. McCloskey, A. D. Fouad, M. A. Churgin and C. Fang-Yen, *J. Neurophysiol.*, 2017, **117**, 1911–1934.
- 135 S. Dernovici, T. Starc, J. A. Dent and P. Ribeiro, *Dev. Neurobiol.*, 2007, **67**, 189–204.
- 136 B. Kirchweger, L. C. Klein-Junior, D. Pretsch, Y. Chen, S. Cretton, A. L. Gasper, Y. V. Heyden, P. Christen, J. Kirchmair, A. T. Henriques and J. M. Rollinger, *Front. Neurosci.*, 2022, **16**, 826289.
- 137 M. A. Bolanowski, R. L. Russell and L. A. Jacobson, *Mech. Ageing Dev.*, 1981, **15**, 279–295.
- 138 M. Scholz, D. J. Lynch, K. S. Lee, E. Levine and D. Biron, *J. Neurosci. Methods*, 2016, **274**, 172–178.
- 139 S. R. Lockery, S. E. Hulme, W. M. Roberts, K. J. Robinson, A. Laromaine, T. H. Lindsay, G. M. Whitesides and J. C. Weeks, *Lab Chip*, 2012, **12**, 2211–2220.
- 140 D. Ryu, L. Mouchiroud, P. A. Andreux, E. Katsyuba, N. Moullan, A. A. Nicolet-Dit-Félix, E. G. Williams, P. Jha, G. Lo Sasso, D. Huzard, P. Aebischer, C. Sandi, C. Rinsch and J. Auwerx, *Nat. Med.*, 2016, **22**, 879–888.
- 141 P. J. Campagnola, A. C. Millard, M. Terasaki, P. E. Hoppe, C. J. Malone and W. A. Mohler, *Biophys. J.*, 2002, **82**, 493–508.
- 142 L. Amigoni, M. Stuknyte, C. Ciaramelli, C. Magoni, I. Bruni, I. De Noni, C. Airolidi, M. E. Regonesi and A. Palmioli, *J. Funct. Foods*, 2017, **33**, 297–306.
- 143 L. A. Harrington and C. B. Harley, *Mech. Ageing Dev.*, 1988, **43**, 71–78.
- 144 E. de Castro, S. Hegi de Castro and T. E. Johnson, *Free Radical Biol. Med.*, 2004, **37**, 139–145.
- 145 G. J. Lithgow, T. M. White, S. Melov and T. E. Johnson, *Proc. Natl. Acad. Sci. U.S.A.*, 1995, **92**, 7540–7544.
- 146 A. Kampkötter, T. Pielarski, R. Rohrig, C. Timpel, Y. Chovolou, W. Wätjen and R. Kahl, *Pharmacol. Res.*, 2007, **55**, 139–147.
- 147 S. Murakami and T. E. Johnson, *Genetics*, 1996, **143**, 1207–1218.
- 148 N. Fischer, C. Buechter, K. Koch, S. Albert, R. Csuk and W. Waetjen, *J. Pharm. Pharmacol.*, 2017, **69**, 73–81.
- 149 C. Büchter, D. Ackermann, S. Havermann, S. Honnen, Y. Chovolou, G. Fritz, A. Kampkötter and W. Wätjen, *Int. J. Mol. Sci.*, 2013, **14**, 11895–11914.
- 150 C. F. Labuschagne and A. B. Brenkman, *Ageing Res. Rev.*, 2013, **12**, 918–930.
- 151 T. J. Schulz, K. Zarse, A. Voigt, N. Urban, M. Birringer and M. Ristow, *Cell Metab.*, 2007, **6**, 280–293.
- 152 V. I. Pérez, A. Bokov, H. Van Remmen, J. Mele, Q. Ran, Y. Ikeno and A. Richardson, *Biochim. Biophys. Acta*, 2009, **1790**, 1005–1014.
- 153 M. P. Murphy, H. Bayir, V. Belousov, C. J. Chang, K. J. A. Davies, M. J. Davies, T. P. Dick, T. Finkel, H. J. Forman, Y. Janssen-Heininger, D. Gems, V. E. Kagan, B. Kalyanaraman, N.-G. Larsson, G. L. Milne, T. Nyström, H. E. Poulsen, R. Radi, H. Van Remmen, P. T. Schumacker, P. J. Thornalley, S. Toyokuni, C. C. Winterbourn, H. Yin and B. Halliwell, *Nat. Metab.*, 2022, **4**, 651–662.
- 154 D.-E. Jeong, Y. Lee and S.-J. V. Lee, in *Hypoxia: Methods and Protocols*, ed. L. E. Huang, Springer New York, New York, 2018, pp. 213–225.
- 155 W. N. Tawe, M.-L. Eschbach, R. D. Walter and K. Henkle-Dührsen, *Nucleic Acids Res.*, 1998, **26**, 1621–1627.
- 156 P. Link and M. Wink, *Phytomedicine*, 2019, **55**, 119–124.
- 157 P. Link, K. Roth, F. Sporer and M. Wink, *Molecules*, 2016, **21**, 871.
- 158 Z.-Z. Yang, Y.-T. Yu, H.-R. Lin, D.-C. Liao, X.-H. Cui and H.-B. Wang, *Free Radical Biol. Med.*, 2018, **129**, 310–322.
- 159 D. Pretsch, J. M. Rollinger, A. Schmid, M. Genov, T. Woehrer, L. Krenn, M. Moloney, A. Kasture, T. Hummel and A. Pretsch, *Sci. Rep.*, 2020, **10**, 11707.
- 160 S. T. Henderson and T. E. Johnson, *Curr. Biol.*, 2001, **11**, 1975–1980.
- 161 S.-Q. Zheng, A.-J. Ding, G.-P. Li, G.-S. Wu and H.-R. Luo, *PLoS One*, 2013, **8**, e56877.
- 162 A. P. García-González, A. D. Ritter, S. Shrestha, E. C. Andersen, L. S. Yilmaz and A. J. M. Walhout, *Cell*, 2017, **169**, 431–441.
- 163 S. Beydoun, H. S. Choi, G. Dela-Cruz, J. Kruempel, S. Huang, D. Bazopoulou, H. A. Miller, M. L. Schaller, C. R. Evans and S. F. Leiser, *Commun. Biol.*, 2021, **4**, 258.
- 164 D. H. Kim, *Annu. Rev. Genet.*, 2013, **47**, 233–246.
- 165 I. Lenaerts, G. A. Walker, L. Van Hoorebeke, D. Gems and J. R. Vanfleteren, *J. Gerontol., Ser. A*, 2008, **63**, 242–252.
- 166 L. Larigot, D. Mansuy, I. Borowski, X. Coumoul and J. Dairou, *Biomolecules*, 2022, **12**, 342.
- 167 T. H. Lindblom and A. K. Dodd, *J. Exp. Zool.*, 2006, **305**, 720–730.
- 168 A. R. Burns, I. M. Wallace, J. Wildenhain, M. Tyers, G. Giaever, G. D. Bader, C. Nislow, S. R. Cutler and P. J. Roy, *Nat. Chem. Biol.*, 2010, **6**, 549–557.
- 169 A. AlOkda and J. M. Van Raamsdonk, *MicroPubl., Biol.*, 2022, DOI: [10.17912/micropub.biology.000634](https://doi.org/10.17912/micropub.biology.000634).
- 170 X. Wang, X. Wang, L. Li and D. Wang, *Biochem. Biophys. Res. Commun.*, 2010, **400**, 613–618.
- 171 N. Wu, Y.-C. Ma, X.-Q. Gong, P.-J. Zhao, Y.-J. Jia, Q. Zhao, J.-H. Duan and C.-G. Zou, *Nat. Commun.*, 2023, **14**, 240.
- 172 H. Peixoto, M. Roxo, S. Krstin, T. Roehrig, E. Richling and M. Wink, *J. Agric. Food Chem.*, 2016, **64**, 1283–1290.
- 173 L. Gao, R. Zhang, J. Lan, R. Ning, D. Wu, D. Chen and W. Zhao, *J. Nat. Prod.*, 2016, **79**, 3039–3046.
- 174 T. E. Johnson, *Proc. Natl. Acad. Sci. U. S. A.*, 1987, **84**, 3777–3781.
- 175 Y. Liu, Z. Zhou, L. Yin, M. Zhu, F. Wang, L. Zhang, H. Wang, Z. Zhou, H. Zhu, C. Huang and S. Fan, *BioFactors*, 2022, **48**, 442–453.
- 176 B. Gerstbrein, G. Stamatas, N. Kollias and M. Driscoll, *Ageing Cell*, 2005, **4**, 127–137.
- 177 Q. Ruan, Y. Qiao, Y. Zhao, Y. Xu, M. Wang, J. Duan and D. Wang, *J. Ethnopharmacol.*, 2016, **177**, 101–110.



- 178 Z. Pincus, T. C. Mazer and F. J. Slack, *Aging*, 2016, **8**, 889–898.
- 179 G. Di Rosa, G. Brunetti, M. Scuto, A. T. Salinaro, E. J. Calabrese, R. Crea, C. Schmitz-Linneweber, V. Calabrese and N. Saul, *Int. J. Mol. Sci.*, 2020, **21**, 3893.
- 180 S. Li, H. A. Stone and C. T. Murphy, *Lab Chip*, 2015, **15**, 524–531.
- 181 D. S. Yoon, M.-H. Lee and D. S. Cha, *Bio-Protoc.*, 2018, **8**, e2774.
- 182 P. Back, B. P. Braeckman and F. Matthijssens, *Oxid. Med. Cell. Longevity*, 2012, 608478.
- 183 G. J. Lithgow and G. A. Walker, *Mech. Ageing Dev.*, 2002, **123**, 765–771.
- 184 J. R. Cypser, P. Tedesco and T. E. Johnson, *Exp. Gerontol.*, 2006, **41**, 935–939.
- 185 T. E. Johnson, E. de Castro, S. Hegi de Castro, J. Cypser, S. Henderson and P. Tedesco, *Exp. Gerontol.*, 2001, **36**, 1609–1617.
- 186 K. M. Van Pelt and M. C. Truttmann, *Transl. Med. Aging*, 2020, **4**, 60–72.
- 187 R. Keowkase and N. Weerapreeyakul, *Planta Med.*, 2016, **82**, 516–523.
- 188 W. Cai, J. Wu, X. Wang, J. Huang, Z. Shen and X. Chen, *Tradit. Med. Mod. Med.*, 2019, **2**, 19–25.
- 189 S. Govindan, M. Amirthalingam, K. Duraisamy, T. Govindhan, N. Sundararaj and S. Palanisamy, *Biomed. Pharmacother.*, 2018, **102**, 812–822.
- 190 Y. Chen, B. Onken, H. Chen, S. Xiao, X. Liu, M. Driscoll, Y. Cao and Q. Huang, *J. Agric. Food Chem.*, 2014, **62**, 3422–3431.
- 191 A. Upadhyay, J. Chompoo, N. Taira, M. Fukuta and S. Tawata, *Biosci., Biotechnol., Biochem.*, 2013, **77**, 217–223.
- 192 J. Li, D. Liu, D. Li, Y. Guo, H. Du and Y. Cao, *Chem. Biodiversity*, 2022, **19**, e202100685.
- 193 W. Chen, L. Rezaizadehnajafi and M. Wink, *J. Pharm. Pharmacol.*, 2013, **65**, 682–688.
- 194 M. Lucanic, W. T. Plummer, E. Chen, J. Harke, A. C. Foulger, B. Onken, A. L. Coleman-Hulbert, K. J. Dumas, S. Guo, E. Johnson, D. Bhaumik, J. Xue, A. B. Crist, M. P. Presley, G. Harinath, C. A. Sedore, M. Chamoli, S. Kamat, M. K. Chen, S. Angeli, C. Chang, J. H. Willis, D. Edgar, M. A. Royal, E. A. Chao, S. Patel, T. Garrett, C. Ibanez-Ventoso, J. Hope, J. L. Kish, M. Guo, G. J. Lithgow, M. Driscoll and P. C. Phillips, *Nat. Commun.*, 2017, **8**, 14256.
- 195 J. Lee, G. Kwon, J. Park, J.-K. Kim and Y.-H. Lim, *Exp. Biol. Med.*, 2016, **241**, 1757–1763.
- 196 J. Li, Z. Lin, X. Tang, G. Liu, Y. Chen, X. Zhai, Q. Huang and Y. Cao, *Food Funct.*, 2020, **11**, 6595–6607.
- 197 P. R. Hunt, T. G. Son, M. A. Wilson, Q.-S. Yu, W. H. Wood, Y. Zhang, K. G. Becker, N. H. Greig, M. P. Mattson, S. Camandola and C. A. Wolkow, *PLoS One*, 2011, **6**, e21922.
- 198 A. S. Arampatzis, O. Tsave, B. Kirchweiger, J. Zwirchmayr, V. P. Papageorgiou, J. M. Rollinger and A. N. Assimopoulou, *Front. Pharmacol.*, 2022, **13**, 909285.
- 199 S. Schmeisser, K. Schmeisser, S. Weimer, M. Groth, S. Priebe, E. Fazius, D. Kuhlow, D. Pick, J. W. Einax, R. Guthke, M. Platzer, K. Zarse and M. Ristow, *Aging Cell*, 2013, **12**, 508–517.
- 200 Q. Zhu, Y. Qu, X.-G. Zhou, J.-N. Chen, H.-R. Luo and G.-S. Wu, *Oxid. Med. Cell. Longevity*, 2020, 6069354.
- 201 B. A. Akhooon, S. Pandey, S. Tiwari and R. Pandey, *Exp. Gerontol.*, 2016, **78**, 47–56.
- 202 Y. Qu, L. Shi, Y. Liu, L. Huang, H.-R. Luo and G.-S. Wu, *Oxid. Med. Cell. Longevity*, 2022, 8878923.
- 203 K. Pietsch, N. Saul, S. Chakrabarti, S. R. Stürzenbaum, R. Menzel and C. E. W. Steinberg, *Biogerontology*, 2011, **12**, 329–347.
- 204 J. Nance and C. Frøkjær-Jensen, *Genetics*, 2019, **212**, 959–990.
- 205 L. R. Girard, T. J. Fiedler, T. W. Harris, F. Carvalho, I. Antoshechkin, M. Han, P. W. Sternberg, L. D. Stein and M. Chalfie, *Nucleic Acids Res.*, 2007, **35**, D472–D475.
- 206 *WormAtlas*, ed. Z. F. Altun, L. A. Herndon, C. A. Wolkow, C. Crocker, R. Lints and D. H. Hall, pp. 2002–2023, <https://www.wormatlas.org>.
- 207 *Caenorhabditis Genetics Center (CGC)*, <https://cgc.umn.edu>, accessed 05.03.2023.
- 208 *Worm Base*, <https://wormbase.org/#012-34-5>, accessed 05.03.2023.
- 209 O. Thompson, M. Edgley, P. Strasbourger, S. Flibotte, B. Ewing, R. Adair, V. Au, I. Chaudhry, L. Fernando, H. Hutter, A. Kieffer, J. Lau, N. Lee, A. Miller, G. Raymant, B. Shen, J. Shendure, J. Taylor, E. H. Turner, L. W. Hillier, D. G. Moerman and R. H. Waterston, *Genome Res.*, 2013, **23**, 1749–1762.
- 210 X. Zhao, L. Lu, Y. Qi, M. Li and L. Zhou, *Biosci., Biotechnol., Biochem.*, 2017, **81**, 1908–1916.
- 211 K. Filippopoulou, N. Papaevgeniou, M. Lefaki, A. Paraskevopoulou, D. Biedermann, V. Kren and N. Chondrogianni, *Free Radical Biol. Med.*, 2017, **103**, 256–267.
- 212 W.-Y. Zeng, L. Tan, C. Han, Z.-Y. Zheng, G.-S. Wu, H.-R. Luo and S.-L. Li, *Oxid. Med. Cell. Longevity*, 2021, 7656834.
- 213 J. H. Kim, I. H. Bang, Y. J. Noh, D. K. Kim, E. J. Bae and I. H. Hwang, *Int. J. Mol. Sci.*, 2020, **21**, 2212.
- 214 C. W. Yu, C. C. Wei and V. H. C. Liao, *Free Radical Res.*, 2014, **48**, 371–379.
- 215 A. Le Bras, *Lab. Anim.*, 2020, **49**, 311.
- 216 J. S. Watts, H. F. Harrison, S. Omi, Q. Guenthers, J. Dalelio, N. Pujol and J. L. Watts, *G3: Genes, Genomes, Genet.*, 2020, **10**, 4167–4176.
- 217 P. Shah, Z. Bao and R. Zaidel-Bar, *Genetics*, 2022, **221**, iyac068.
- 218 Y. Wang, S. Luo, Z. Xu, L. Liu, S. Feng, T. Chen, L. Zhou, M. Yuan, Y. Huang and C. b. Ding, *Arabian J. Chem.*, 2021, **14**, 103149.
- 219 Y. Gong, Y. Luo, J.-A. Huang, J. Zhang, Y. Peng, Z. Liu and B. Zhao, *J. Funct. Foods*, 2012, **4**, 988–993.
- 220 J. H. An and T. K. Blackwell, *Genes Dev.*, 2003, **17**, 1882–1893.
- 221 J. M. A. Tullet, M. Hertweck, J. H. An, J. Baker, J. Y. Hwang, S. Liu, R. P. Oliveira, R. Baumeister and T. K. Blackwell, *Cell*, 2008, **132**, 1025–1038.



- 222 S. Havermann, R. Rohrig, Y. Chovolou, H. U. Humpf and W. Wätjen, *J. Agric. Food Chem.*, 2013, **61**, 2158–2164.
- 223 J. Nass, S. Abdelfatah and T. Efferth, *Phytomedicine*, 2021, **84**, 153482.
- 224 Y. Niu, J. Liao, H. Zhou, C.-C. Wang, L. Wang and Y. Fan, *Molecules*, 2022, **27**, 4952.
- 225 J. Liu, J. Meng, J. Du, X. Liu, Q. Pu, D. Di and C. Chen, *Molecules*, 2020, **25**, 3511.
- 226 K. Koch, N. Weldle, S. Baier, C. Büchter and W. Wätjen, *Eur. J. Nutr.*, 2020, **59**, 137–150.
- 227 X. Wang, X. Li, L. Li, X. Yang, J. Wang, X. Liu, J. Chen, S. Liu, N. Zhang, J. Li and H. Wang, *Food Funct.*, 2022, **13**, 10680–10694.
- 228 F. Yan, Y. Yang, L. Yu and X. Zheng, *J. Agric. Food Chem.*, 2017, **65**, 7457–7466.
- 229 H. Wang, J. Liu, T. Li and R. H. Liu, *Food Funct.*, 2018, **9**, 5273–5282.
- 230 C. Duangjan, P. Rangsinth, X. Gu, M. Wink and T. Tencomnao, *Oxid. Med. Cell. Longevity*, 2019, 9012396.
- 231 S. Pandey, S. Tiwari, A. Kumar, A. Niranjana, J. Chand, A. Lehri and P. S. Chauhan, *Ind. Crops Prod.*, 2018, **120**, 113–122.
- 232 A. L. Hsu, C. T. Murphy and C. Kenyon, *Science*, 2003, **300**, 1142–1145.
- 233 J. F. Morley and R. I. Morimoto, *Mol. Biol. Cell*, 2004, **15**, 657–664.
- 234 J. Brunquell, S. Morris, A. Snyder and S. D. Westerheide, *Cell Stress Chaperones*, 2018, **23**, 65–75.
- 235 W. Liu, Y. Guan, S. Qiao, J. Wang, K. Bao, Z. Mao, L. Liao, A. Moskalev, B. Jiang, J. Zhu, C. Xia, J. Li and Z. Hu, *Oxid. Med. Cell. Longevity*, 2021, 9942090.
- 236 Y.-P. Zhang, W.-H. Zhang, P. Zhang, Q. Li, Y. Sun, J.-W. Wang, S. O. Zhang, T. Cai, C. Zhan and M.-Q. Dong, *Nat. Commun.*, 2022, **13**, 6339.
- 237 B. Song, B. Zheng, T. Li and R. H. Liu, *Food Funct.*, 2020, **11**, 3598–3609.
- 238 H. Jeon and D. S. Cha, *Chin. J. Nat. Med.*, 2016, **14**, 335–342.
- 239 F. Yan, X.-A. Chen and X. Zheng, *Food Res. Int.*, 2017, **102**, 213–224.
- 240 S. Ogg, S. Paradis, S. Gottlieb, G. I. Patterson, L. Lee, H. A. Tissenbaum and G. Ruvkun, *Nature*, 1997, **389**, 994–999.
- 241 K. Lin, J. B. Dorman, A. Rodan and C. Kenyon, *Science*, 1997, **278**, 1319–1322.
- 242 V. Shukla, D. Yadav, S. C. Phulara, M. M. Gupta, S. K. Saikia and R. Pandey, *Free Radical Biol. Med.*, 2012, **53**, 1848–1856.
- 243 J. M. Romero-Marquez, M. D. Navarro-Hortal, V. Jimenez-Trigo, P. Munoz-Ollero, T. Y. Forbes-Hernandez, A. Esteban-Munoz, F. Giampieri, I. Delgado Noya, P. Bullon, L. Vera-Ramirez, M. Battino, C. Sanchez-Gonzalez and J. L. Quiles, *Antioxidants*, 2022, **11**, 629.
- 244 M. Roxo, H. Peixoto, P. Wetterauer, E. Lima and M. Wink, *Oxid. Med. Cell. Longevity*, 2020, 7590707.
- 245 C. Duangjan, P. Rangsinth, X. Gu, S. Zhang, M. Wink and T. Tencomnao, *Phytomedicine*, 2019, **64**, 153061.
- 246 W. B. Bakrim, A. D. R. Nurcahyanti, M. Dmirieh, I. Mahdi, A. M. Elgamal, M. A. El Raey, M. Wink and M. Sobeh, *Oxid. Med. Cell. Longevity*, 2022, 3486257.
- 247 N. M. Hegazi, M. Sobeh, S. Rezaq, M. A. El-Raey, M. Dmirieh, A. M. El-Shazly, M. F. Mahmoud and M. Wink, *Sci. Rep.*, 2019, **9**, 1–12.
- 248 M. A. O. Abdelfattah, M. Dmirieh, W. Ben Bakrim, O. Mouhtady, M. A. Ghareeb, M. Wink and M. Sobeh, *J. Ethnopharmacol.*, 2022, **292**, 115187.
- 249 S. Thabit, H. Handoussa, M. Roxo, B. Cestari de Azevedo, N. S. E. El Sayed and M. Wink, *Molecules*, 2019, **24**, 2633.
- 250 A. L. Tambara, L. de Los Santos Moraes, A. H. Dal Forno, J. R. Boldori, A. T. Goncalves Soares, C. de Freitas Rodrigues, L. R. B. Mariutti, A. Z. Mercadante, D. S. de Avila and C. C. Denardin, *Food Chem. Toxicol.*, 2018, **120**, 639–650.
- 251 J. Zheng, D. Heber, M. Wang, C. Gao, S. B. Heymsfield, R. J. Martin, F. L. Greenway, J. W. Finley, J. H. Burton, W. D. Johnson, F. M. Enright, M. J. Keenan and Z. Li, *Int. J. Vitam. Nutr. Res.*, 2017, **87**, 149–158.
- 252 M. S. Bonkowski and D. A. Sinclair, *Nat. Rev. Mol. Cell Biol.*, 2016, **17**, 679–690.
- 253 H. A. Tissenbaum and L. Guarente, *Nature*, 2001, **410**, 227–230.
- 254 M. Maulik, S. Mitra, S. Hunter, M. Hunstiger, S. R. Oliver, A. Bult-Itto and B. E. Taylor, *Sci. Rep.*, 2018, **8**, 1–13.
- 255 L.-G. Xiong, J.-A. Huang, J. Li, P.-H. Yu, Z. Xiong, J.-W. Zhang, Y.-S. Gong, Z.-H. Liu and J.-H. Chen, *J. Agric. Food Chem.*, 2014, **62**, 11163–11169.
- 256 D. J. Deusing, S. Winter, A. Kler, E. Kriesl, B. Bonnlaender, U. Wenzel and E. Fitzenberger, *Fitoterapia*, 2015, **102**, 163–170.
- 257 S. Robida-Stubbs, K. Glover-Cutter, D. W. Lamming, M. Mizunuma, S. D. Narasimhan, E. Neumann-Haefelin, D. M. Sabatini and T. K. Blackwell, *Cell Metab.*, 2012, **15**, 713–724.
- 258 V. T. Cuong, W. Chen, J. Shi, M. Zhang, H. Yang, N. Wang, S. Yang, J. Li, P. Yang and J. Fei, *Exp. Gerontol.*, 2019, **117**, 99–105.
- 259 J. Apfeld, G. O'Connor, T. McDonagh, P. S. DiStefano and R. Curtis, *Genes Dev.*, 2004, **18**, 3004–3009.
- 260 B. Onken and M. Driscoll, *PLoS One*, 2010, **5**, e8758.
- 261 H. J. Weir, P. Yao, F. K. Huynh, C. C. Escoubas, R. L. Goncalves, K. Burkewitz, R. Laboy, M. D. Hirschey and W. B. Mair, *Cell Metab.*, 2017, **26**, 884–896.
- 262 L. Chen, H. Cao, Q. Huang, J. Xiao and H. Teng, *Crit. Rev. Food Sci. Nutr.*, 2022, **62**, 7730–7742.
- 263 B. Spanier, R. Lang, D. Weber, A. Lechner, T. Thoma, M. Rothner, K. Petzold, T. Lang, A. Beusch, M. Bösl, V. Schlagbauer, H. Daniel and T. Hofmann, *J. Agric. Food Chem.*, 2019, **67**, 4774–4781.
- 264 M. Dueñas, F. Surco-Laos, S. González-Manzano, A. M. González-Paramás, E. Gómez-Orte, J. Cabello and C. Santos-Buelga, *Pharmacol. Res.*, 2013, **76**, 41–48.
- 265 F. A. Tomás-Barberán, R. García-Villalba, A. González-Sarrias, M. V. Selma and J. C. Espín, *J. Agric. Food Chem.*, 2014, **62**, 6535–6538.



- 266 EFSA, *Novel Food Authorisation*, https://open.efsa.europa.eu/question/EFSA-Q-2018-00724_2023, accessed 03.08.2023.
- 267 P. A. Andreux, W. Blanco-Bose, D. Ryu, F. Burdet, M. Ibberson, P. Aebischer, J. Auwerx, A. Singh and C. Rinsch, *Nat. Metab.*, 2019, **1**, 595–603.
- 268 J. G. Wood, B. Rogina, S. Lavu, K. Howitz, S. L. Helfand, M. Tatar and D. Sinclair, *Nature*, 2004, **430**, 686–689.
- 269 S. Feng, C. Zhang, T. Chen, L. Zhou, Y. Huang, M. Yuan, T. Li and C. Ding, *Antioxidants*, 2021, **10**, 1697.
- 270 A. Matencio, M. A. Guerrero-Rubio, F. Caldera, C. Cecone, F. Trotta, F. Garcia-Carmona and J. M. Lopez-Nicolas, *Int. J. Pharm.*, 2020, **589**, 119862.
- 271 T. M. Bass, D. Weinkove, K. Houthoofd, D. Gems and L. Partridge, *Mech. Ageing Dev.*, 2007, **128**, 546–552.
- 272 R. Strong, R. A. Miller, C. M. Astle, J. A. Baur, R. de Cabo, E. Fernandez, W. Guo, M. Javors, J. L. Kirkland, J. F. Nelson, D. A. Sinclair, B. Teter, D. Williams, N. Zaveri, N. L. Nadon and D. E. Harrison, *J. Gerontol., Ser. A*, 2013, **68**, 6–16.
- 273 J. A. Baur, K. J. Pearson, N. L. Price, H. A. Jamieson, C. Lerin, A. Kalra, V. V. Prabhu, J. S. Allard, G. Lopez-Lluch, K. Lewis, P. J. Pistell, S. Poosala, K. G. Becker, O. Boss, D. Gwinn, M. Wang, S. Ramaswamy, K. W. Fishbein, R. G. Spencer, E. G. Lakatta, D. Le Couteur, R. J. Shaw, P. Navas, P. Puigserver, D. K. Ingram, R. de Cabo and D. A. Sinclair, *Nature*, 2006, **444**, 337–342.
- 274 K. J. Pearson, J. A. Baur, K. N. Lewis, L. Peshkin, N. L. Price, N. Labinsky, W. R. Swindell, D. Kamara, R. K. Minor, E. Perez, H. A. Jamieson, Y. Zhang, S. R. Dunn, K. Sharma, N. Pleshko, L. A. Woollett, A. Csiszar, Y. Ikeno, D. Le Couteur, P. J. Elliott, K. G. Becker, P. Navas, D. K. Ingram, N. S. Wolf, Z. Ungvari, D. A. Sinclair and R. de Cabo, *Cell Metab.*, 2008, **8**, 157–168.
- 275 D. Chen and L. Guarente, *Trends Mol. Med.*, 2007, **13**, 64–71.
- 276 N. L. Price, A. P. Gomes, A. J. Ling, F. V. Duarte, A. Martin-Montalvo, B. J. North, B. Agarwal, L. Ye, G. Ramadori, J. S. Teodoro, B. P. Hubbard, A. T. Varela, J. G. Davis, B. Varamini, A. Hafner, R. Moaddel, A. P. Rolo, R. Coppari, C. M. Palmeira, R. de Cabo, J. A. Baur and D. A. Sinclair, *Cell Metab.*, 2012, **15**, 675–690.
- 277 B. P. Hubbard, A. P. Gomes, H. Dai, J. Li, A. W. Case, T. Considine, T. V. Riera, J. E. Lee, S. Y. E, D. W. Lamming, B. L. Pentelute, E. R. Schuman, L. A. Stevens, A. J. Y. Ling, S. M. Armour, S. Michan, H. Zhao, Y. Jiang, S. M. Sweitzer, C. A. Blum, J. S. Disch, P. Y. Ng, K. T. Howitz, A. P. Rolo, Y. Hamuro, J. Moss, R. B. Perni, J. L. Ellis, G. P. Vlasuk and D. A. Sinclair, *Science*, 2013, **339**, 1216–1219.
- 278 K. Pallauf, I. Günther, G. Kühn, D. Chin, S. de Pascual-Teresa and G. Rimbach, *Adv. Nutr.*, 2021, **12**, 995–1005.
- 279 C. Burnett, S. Valentini, F. Cabreiro, M. Goss, M. Somogyvári, M. D. Piper, M. Hoddinott, G. L. Sutphin, V. Leko, J. J. McElwee, R. P. Vazquez-Manrique, A. M. Orfila, D. Ackerman, C. Au, G. Vinti, M. Riesen, K. Howard, C. Neri, A. Bedalov, M. Kaerberlein, C. Soti, L. Partridge and D. Gems, *Nature*, 2011, **477**, 482–485.
- 280 M. Kaerberlein, T. McDonagh, B. Heltweg, J. Hixon, E. A. Westman, S. D. Caldwell, A. Napper, R. Curtis, P. S. DiStefano, S. Fields, A. Bedalov and B. K. Kennedy, *J. Biol. Chem.*, 2005, **280**, 17038–17045.
- 281 C. Brenner, *Life Metab.*, 2022, **1**, 122–133.
- 282 S. T. Soukup, B. Spanier, G. Grünz, D. Bunzel, H. Daniel and S. E. Kulling, *PLoS One*, 2012, **7**, e46914.
- 283 S. Ehalá, M. Vaheer and M. Kaljurand, *J. Agric. Food Chem.*, 2005, **53**, 6484–6490.
- 284 S. Weiskirchen and R. Weiskirchen, *Adv. Nutr.*, 2016, **7**, 706–718.
- 285 H. Yuan and R. Marmorstein, *Science*, 2013, **339**, 1156–1157.
- 286 T. Nunes, L. Almeida, J.-F. Rocha, A. Falcão, C. Fernandes-Lopes, A. I. Loureiro, L. Wright, M. Vaz-da-Silva and P. Soares-da-Silva, *J. Clin. Pharmacol.*, 2009, **49**, 1477–1482.
- 287 R. Zamora-Ros, N. G. Forouhi, S. J. Sharp, C. A. González, B. Buijsse, M. Guevara, Y. T. van der Schouw, P. Amiano, H. Boeing, L. Bredsdorff, G. Fagherazzi, E. J. Feskens, P. W. Franks, S. Grioni, V. Katzke, T. J. Key, K. T. Khaw, T. Kühn, G. Masala, A. Mattiello, E. Molina-Montes, P. M. Nilsson, K. Overvad, F. Perquier, M. L. Redondo, F. Ricceri, O. Rolandsson, I. Romieu, N. Roswall, A. Scalbert, M. Schulze, N. Slimani, A. M. Spijkerman, A. Tjonneland, M. J. Tormo, M. Touillaud, R. Tumino, A. D. van der, G. J. van Woudenberg, C. Langenberg, E. Riboli and N. J. Wareham, *J. Nutr.*, 2014, **144**, 335–343.
- 288 M. G. Hertog, E. J. Feskens, D. Kromhout, P. Hollman and M. Katan, *Lancet*, 1993, **342**, 1007–1011.
- 289 M. G. Hertog, D. Kromhout, C. Aravanis, H. Blackburn, R. Buzina, F. Fidanza, S. Giampaoli, A. Jansen, A. Menotti and S. Nedeljkovic, *Arch. Intern. Med.*, 1995, **155**, 381–386.
- 290 T. M. Holland, P. Agarwal, Y. Wang, K. Dhana, S. E. Leurgans, K. Shea, S. L. Booth, K. B. Rajan, J. A. Schneider and L. L. Barnes, *Neurology*, 2023, **100**, e694.
- 291 A. Kampkötter, C. Timpel, R. F. Zurawski, S. Ruhl, Y. Chovolou, P. Proksch and W. Wätjen, *Comp. Biochem. Physiol., Part B*, 2008, **149**, 314–323.
- 292 S. Havermann, H.-U. Humpf and W. Waetjen, *Fitoterapia*, 2016, **113**, 123–127.
- 293 B. Ayuda-Durán, E. Sánchez-Hernández, S. González-Manzano, C. Santos-Buelga and A. M. González-Paramás, *Front. Nutr.*, 2022, **9**, 989427.
- 294 G. Grünz, K. Haas, S. Soukup, M. Klingenspor, S. E. Kulling, H. Daniel and B. Spanier, *Mech. Ageing Dev.*, 2012, **133**, 1–10.
- 295 F. Surco-Laos, J. Cabello, E. Gómez-Orte, S. González-Manzano, A. M. González-Paramás, C. Santos-Buelga and M. Dueñas, *Food Funct.*, 2011, **2**, 445–456.
- 296 M. F. M. Sciacca, V. Romanucci, A. Zarrelli, I. Monaco, F. Lolicato, N. Spinella, C. Galati, G. Grasso, L. D'Urso, M. Romeo, L. Diomedede, M. Salmona, C. Bongiorno, G. Di Fabio, C. La Rosa and D. Milardi, *ACS Chem. Neurosci.*, 2017, **8**, 1767–1778.
- 297 G. Buckland and C. A. Gonzalez, *Br. J. Nutr.*, 2015, **113**, S94–S101.



- 298 R. Estruch, E. Ros, J. Salas-Salvadó, M.-I. Covas, D. Corella, F. Arós, E. Gómez-Gracia, V. Ruiz-Gutiérrez, M. Fiol and J. Lapetra, *N. Engl. J. Med.*, 2018, **378**, e34.
- 299 M. A. Martínez-González, M. Bes-Rastrollo, L. Serra-Majem, D. Lairon, R. Estruch and A. Trichopoulou, *Nutr. Rev.*, 2009, **67**, S111–S116.
- 300 L. G. Gillingham, S. Harris-Janz and P. J. Jones, *Lipids*, 2011, **46**, 209–228.
- 301 S. Han, E. A. Schroeder, C. G. Silva-García, K. Hebestreit, W. B. Mair and A. Brunet, *Nature*, 2017, **544**, 185–190.
- 302 J. Goudeau, S. Bellemin, E. Toselli-Mollereau, M. Shamalnasab, Y. Chen and H. Aguilaniu, *PLoS Biol.*, 2011, **9**, e1000599.
- 303 S.-J. Lee, A. B. Hwang and C. Kenyon, *Curr. Biol.*, 2010, **20**, 2131–2136.
- 304 R. Ratnappan, F. R. G. Amrit, S.-W. Chen, H. Gill, K. Holden, J. Ward, K. R. Yamamoto, C. P. Olsen and A. Ghazi, *PLoS Genet.*, 2014, **10**, e1004829.
- 305 J. C. Garcia-Moreno, M. Porta de la Riva, E. Martínez-Lara, E. Siles and A. Cañuelo, *Neurobiol. Aging*, 2019, **82**, 60–68.
- 306 A. Cañuelo, B. Gilbert-López, P. Pacheco-Liñán, E. Martínez-Lara, E. Siles and A. Miranda-Vizuete, *Mech. Ageing Dev.*, 2012, **133**, 563–574.
- 307 G. Di Rosa, N. Saul, C. Schmitz-Linneweber and V. Calabrese, *Pathophysiology*, 2018, **25**, 203.
- 308 L. Diomedede, S. Rigacci, M. Romeo, M. Stefani and M. Salmona, *PLoS One*, 2013, **8**, e58893.
- 309 M. Katsiki, N. Chondrogianni, I. Chinou, A. J. Rivett and E. S. Gonos, *Rejuvenation Res.*, 2007, **10**, 157–172.
- 310 T. Nikou, V. Liaki, P. Stathopoulos, A. D. Sklirou, E. N. Tsakiri, T. Jakschitz, G. Bonn, I. P. Trougakos, M. Halabalaki and L. A. Skaltsounis, *Food Chem. Toxicol.*, 2019, **125**, 403–412.
- 311 M. A. Guerrero-Rubio, S. Hernandez-Garcia, F. Garcia-Carmona and F. Gandia-Herrero, *Food Chem.*, 2019, **274**, 840–847.
- 312 M. H. Land, M. L. Toth, L. MacNair, S. A. Vanapalli, T. W. Lefever, E. N. Peters and M. O. Bonn-Miller, *Cannabis Cannabinoid Res.*, 2020, **6**, 522–527.
- 313 Z. Wang, P. Zheng, Y. Xie, X. Chen, N. Solowij, K. Green, Y. L. Chew and X. F. Huang, *FASEB J.*, 2021, **35**, e21537.
- 314 A. P. Vanin, W. A. Tamagno, C. Alves, L. Mesacasa, L. F. Santin, N. T. Sutorillo, D. Bilibio, C. Müller, L. Galon and R. R. Kaizer, *Sci. Rep.*, 2022, **12**, 15376.
- 315 Z. Wang, P. Zheng, X. Chen, Y. Xie, K. Weston-Green, N. Solowij, Y. L. Chew and X.-F. Huang, *Geroscience*, 2022, **44**, 1505–1524.
- 316 Z. Pandelides, C. Thornton, A. S. Faruque, A. P. Whitehead, K. L. Willett and N. M. Ashpole, *Geroscience*, 2020, **42**, 785–800.
- 317 D. O. Mitchell, L. Wen Jing, C. Tobias, A. B. Bruce and K. Richard, *J. Neurosci.*, 2017, **37**, 2859.
- 318 A. Levichev, S. Faumont, R. Z. Berner, Z. Purcell, A. M. White, K. Chicas-Cruz and S. R. Lockery, *Curr. Biol.*, 2023, **33**, 1625–1639.
- 319 S. I. Pastuhov, K. Matsumoto and N. Hisamoto, *Genes Cells*, 2016, **21**, 696–705.
- 320 N. Harrison, M. A. Lone, T. K. Kaul, P. Reis Rodrigues, I. V. Ogungbe and M. S. Gill, *PLoS One*, 2014, **9**, e113007.
- 321 M. Lucanic, J. M. Held, M. C. Vantipalli, I. M. Klang, J. B. Graham, B. W. Gibson, G. J. Lithgow and M. S. Gill, *Nature*, 2011, **473**, 226–229.
- 322 S.-C. Cheng, W.-H. Li, Y.-C. Shi, P.-L. Yen, H.-Y. Lin, V. H.-C. Liao and S.-T. Chang, *J. Agric. Food Chem.*, 2014, **62**, 4159–4165.
- 323 A. Takahashi, T. Watanabe, T. Fujita, T. Hasegawa, M. Saito and M. Suganuma, *Biosci., Biotechnol., Biochem.*, 2014, **78**, 1206–1211.
- 324 P. Yuan, L.-Y. Pan, L.-G. Xiong, J.-W. Tong, J. Li, J.-A. Huang, Y.-S. Gong and Z.-H. Liu, *Food Funct.*, 2018, **9**, 3798–3806.
- 325 S. Abbas and M. Wink, *Antioxidants*, 2014, **3**, 129.
- 326 S. Zhang, C. Duangjan, T. Tencomnao, J. Liu, J. Lin and M. Wink, *Food Funct.*, 2020, **11**, 8179–8192.
- 327 C.-C. Wei, C.-W. Yu, P.-L. Yen, H.-Y. Lin, S.-T. Chang, F.-L. Hsu and V. H.-C. Liao, *J. Agric. Food Chem.*, 2014, **62**, 10701–10707.
- 328 G. Dominguez-Rodriguez, D. Ramon Vidal, P. Martorell, M. Plaza and M. L. Marina, *J. Agric. Food Chem.*, 2022, **70**, 7993–8009.
- 329 J. S. Sangha, D. Fan, A. H. Banskota, R. Stefanova, W. Khan, J. Hafting, J. Craigie, A. T. Critchley and B. Prithviraj, *J. Funct. Foods*, 2013, **5**, 1180–1190.
- 330 N. Shen, W. Zeng, F. Leng, J. Lu, Z. Lu, J. Cui, L. Wang and B. Jin, *Food Funct.*, 2021, **12**, 12395–12406.
- 331 L. Xiong, N. Deng, B. Zheng, T. Li and R. H. Liu, *Food Funct.*, 2021, **12**, 7851–7866.
- 332 S. Guha, O. Nataraja, C. G. Murbach, J. Dinh, E. C. Wilson, M. Cao, S. Zou and Y. Dong, *Nutrients*, 2014, **6**, 911–921.
- 333 S. Guha, M. Cao, R. M. Kane, A. M. Savino, S. Zou and Y. Dong, *Age (Dordr)*, 2013, **35**, 1559–1574.
- 334 C. Scerbak, E. Vayndorf, A. Hernandez, C. McGill and B. Taylor, *GeroScience*, 2018, **40**, 151–162.
- 335 M. Sobeh, A. Esmat, G. Petruk, M. A. O. Abdelfattah, M. Dmirieh, D. M. Monti, A. B. Abdel-Naim and M. Wink, *J. Funct. Foods*, 2018, **41**, 223–231.
- 336 M. I. Prasanth, J. M. Brimson, S. Chuchawankul, M. Sukprasansap and T. Tencomnao, *Oxid. Med. Cell. Longevity*, 2019, 7024785.
- 337 A.-G. Wu, J.-F. Teng, V. K.-W. Wong, X.-G. Zhou, W.-Q. Qiu, Y. Tang, J.-M. Wu, R. Xiong, R. Pan, Y.-L. Wang, B. Tang, T.-Y. Ding, L. Yu, W. Zeng, D.-L. Qin and B. Y.-K. Law, *Phytomedicine*, 2019, **65**, 153088.
- 338 B. Tsolmon, Y. Fang, T. Yang, L. Guo, K. He, G.-Y. Li and H. Zhao, *Food Chem.*, 2021, **343**, 128392.
- 339 W. Jia, Q. Peng, L. Su, X. Yu, C. W. Ma, M. Liang, X. Yin, Y. Zou and Z. Huang, *Mar. Drugs*, 2018, **16**, 444.
- 340 E. B. Lee, M. M. Xing and D. K. Kim, *Arch. Pharmacol Res.*, 2017, **40**, 825–835.
- 341 F. Zhang, M. Berg, K. Dierking, M.-A. Félix, M. Shapira, B. S. Samuel and H. Schulenburg, *Front. Microbiol.*, 2017, **8**, 485.
- 342 C. Heintz and W. Mair, *Cell*, 2014, **156**, 408–411.



- 343 I. Gusarov, L. Gautier, O. Smolentseva, I. Shamovsky, S. Eremina, A. Mironov and E. Nudler, *Cell*, 2013, **152**, 818–830.
- 344 Q. Sun, N. M. Vega, B. Cervantes, C. P. Mancuso, N. Mao, M. N. Taylor, J. J. Collins, A. S. Khalil, J. Gore and T. K. Lu, *Mol. Syst. Biol.*, 2022, **18**, e9933.
- 345 S. Urquiza-Zurich, V. A. Garcia-Angulo, P. Burdisso, M. F. Palominos, L. Fernandez-Hubeid, P. A. Harcha, J. P. Castillo and A. Calixto, *mBio*, 2023, **14**, e0340222.
- 346 R. Saito, N. Sato, Y. Okino, D. S. Wang and G. Seo, *Biosci. Microbiota, Food Health*, 2023, **42**, 124–130.
- 347 A. L. Gaeta, K. Willicott, C. W. Willicott, L. E. McKay, C. M. Keogh, T. J. Altman, L. C. Kimble, A. L. Yarbrough, K. A. Caldwell and G. A. Caldwell, *iScience*, 2023, **26**, 106859.
- 348 V. Donato, F. R. Ayala, S. Cogliati, C. Bauman, J. G. Costa, C. Leñini and R. Grau, *Nat. Commun.*, 2017, **8**, 14332.
- 349 M. C. Rodríguez-Daza, E. C. Pulido-Mateos, J. Lupien-Meilleur, D. Guyonnet, Y. Desjardins and D. Roy, *Front. Nutr.*, 2021, **8**, 689456.
- 350 E.-M. Pferschy-Wenzig, M. R. Pausan, K. Ardjomand-Woelkart, S. Röck, R. M. Ammar, O. Kelber, C. Moissl-Eichinger and R. Bauer, *Nutrients*, 2022, **14**, 2111.
- 351 M. Shapira, *Curr. Opin. Microbiol.*, 2017, **38**, 142–147.
- 352 P. Dirksen, A. Assié, J. Zimmermann, F. Zhang, A. M. Tietje, S. A. Marsh, M. A. Félix, M. Shapira, C. Kaleta, H. Schulenburg and B. S. Samuel, *G3: Genes, Genomes, Genet.*, 2020, **10**, 3025–3039.
- 353 R. Van Assche, V. Broeckx, K. Boonen, E. Maes, W. De Haes, L. Schoofs and L. Temmerman, *J. Mol. Biol.*, 2015, **427**, 3441–3451.
- 354 L. K. Caesar and N. B. Cech, *Nat. Prod. Rep.*, 2019, **36**, 869–888.
- 355 A. Panossian, E.-J. Seo and T. Efferth, *Phytomedicine*, 2018, **50**, 257–284.
- 356 Y. Fu and W. Zhao, *J. Nat. Prod.*, 2020, **83**, 505–515.
- 357 O. Potterat and M. Hamburger, *Planta Med.*, 2014, **80**, 1171–1181.
- 358 A. E. Fox Ramos, L. Evanno, E. Poupon, P. Champy and M. A. Beniddir, *Nat. Prod. Rep.*, 2019, **36**, 960–980.
- 359 U. Grienke, P. A. Foster, J. Zwirchmayr, A. Tahir, J. M. Rollinger and E. Mikros, *Sci. Rep.*, 2019, **9**, 11113.
- 360 J. Langeder, K. Doring, H. Schmietendorf, U. Grienke, M. Schmidtke and J. M. Rollinger, *J. Nat. Prod.*, 2022, **86**, 8–17.
- 361 J. J. Kellogg, D. A. Todd, J. M. Egan, H. A. Raja, N. H. Oberlies, O. M. Kvalheim and N. B. Cech, *J. Nat. Prod.*, 2016, **79**, 376–386.
- 362 J. Zwirchmayr, C. D. Cruz, U. Grienke, P. Tammela and J. M. Rollinger, *iScience*, 2023, 107523.
- 363 M. Heinrich, B. Jalil, M. Abdel-Tawab, J. Echeverria, Ž. Kulić, L. J. McGaw, J. M. Pezzuto, O. Potterat and J.-B. Wang, *Front. Pharmacol.*, 2022, **13**, 953205.
- 364 J. Bisson, J. B. McAlpine, J. B. Friesen, S. N. Chen, J. Graham and G. F. Pauli, *J. Med. Chem.*, 2016, **59**, 1671–1690.
- 365 J. B. Baell, *J. Nat. Prod.*, 2016, **79**, 616–628.
- 366 T. T. Cushnie and A. J. Lamb, *Int. J. Antimicrob. Agents*, 2011, **38**, 99–107.
- 367 K.-T. Chung, Z. Lu and M. Chou, *Food Chem. Toxicol.*, 1998, **36**, 1053–1060.

