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Unusually cyclized triterpenoids: occurrence, biosynthesis and chemical synthesis

Covering: 2009 to 2021

Biosynthetically, most of the syntheses of triterpenes follow the cascade cyclization and rearrangement of the acyclic precursors *viz.*, squalene (S) and 2,3-oxidosqualene (OS), which lead to the very well known tetra- and pentacyclic triterpene skeletons. Aside from these, numerous other triterpenoid molecules are also reported from various natural sources and their structures are derived from "S" and "OS" *via* some unusual cyclization operations which are different from the usual tetra- and pentacyclic frameworks. Numerous compelling advances have been made and reported in the identification of these unusual cyclized mono-, di-, tri- and tetracyclic triterpenes between 2009 and 2021. Besides a dramatic increase in the newly isolated uncommon cyclized triterpenoids, substantial progress in the (bio)-synthesis of these triterpenes has been published along with significant progress in their biological effects. In this review, 180 new unusual cyclized triterpenoids together with their demonstrated biogenetic pathways, syntheses and biological effects will be categorized and discussed.

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22 Notes and references

1 Introduction

Triterpenes represent a versatile group of terpenes comprised of six isoprene subunits and to date over 30 000 triterpenes have been identified from various natural sources.¹ Moreover, around 200 different triterpene skeletons have been reported to be derived from cyclization products of squalene (S), oxidosqualene (OS), or bis-oxidosqualene (BOS) catalysed by tripterpene synthases.² Apart from these, constantly new enzymes are identified which arrange precursors to these skeletons. Very recently, Tao *et al.*³ described the non-squalene-dependent triterpene biosynthesis and discovered two chimeric fungal class I triterpene synthases (TrTSs) for the conversion and production of hexaprenyl diphosphate into triterpenes.

Triterpenes are naturally present in fruit, vegetables and medicinal plants and are hence part of the daily human diet. A driving force behind triterpene research is the wide range of biological and pharmacological effects including cytotoxicity, inhibition of pathogenic microbes, anti-inflammatory, antiprotozoal, antitubercular, and antifeedant effects to name but a few. In addition, some triterpenes have illustrated enzymatic effects and have been shown to modulate specific receptor functions.⁴⁻⁷

The large majority of triterpenoids are 6-6-6-5 tetra-cycles, 6-6-6-6-5 pentacycles, or 6-6-6-6 pentacycles and these compounds are biosynthetically formed *via* cascade cyclizations and rearrangements of either squalene (S) or 2,3-oxidosqualene (OS). Although the majority of the triterpenes contain tetra- or pentacyclic skeletons, together with these triterpenes derived from cyclization processes there are a number that deviate from these mainstream polycyclization of S and OS to tetra- or pentacyclic compounds which have been reported from Nature. These triterpene structures, referred to as "unusually cyclized triterpenoids" are generated from unusual cyclizations of



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squalene (S) and 2,3-oxidosqualene (OS).^{2,8} These "unusually cyclized triterpenes" are biosynthesized via: (i) incomplete cyclization of the S-derived molecules whose cyclization processes start from the epoxide or terminal isopropylidene core and generate mono-, bi- and tricyclic triterpenoid skeletons. (ii) Cyclization of S or OS to generate polycyclic triterpenoid skeletons followed by cleavage of the already established ring systems. (iii) Two independent cyclizations of the S- or OSderived molecule.8

This review provides a comprehensive and critical coverage of 180 new unusual cyclized triterpenoids reported from January

2009 through to December 2021, and serves to complement the previous review of unusual cyclized triterpenoids published in Natural Product Reports.8 In addition, this review includes a few previously omitted articles published between 1998 and 2008. Notably, included are nor-iridal and nor-isomalabaricane terpenes which biosynthetically derived from iridal are and isomalabaricane-type triterpenoids. In much the same way as the previous review, the format focuses on the discovery of new unusual cyclized triterpenoids from natural sources along with their synthesis and biological effects. Additionally, recent studies of the fascinating biosynthesis of these unusual cyclized



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triterpenoids are covered in this review. There have been numerous synthetic strategies towards forming these unusual cyclized triterpenoids as end products and these studies are also covered in this review. Notably, characteristic features of unusual cyclized triterpenoids with their high degree of skeletal and structural diversification are also discussed.

2 Iridal triterpenes

2.1 Monocyclic iridals

The basic iridal nucleus is depicted in Fig. 1 and this skeleton forms from squalene epoxide. In addition, the majority of the iridal natural products are typified by a C-1 carbonyl function (mostly aldehyde), and an ester, or alcohol at C-3. Four monocyclic iridal triterpenes *viz.*, 8-hydroxylisoiridogermanal (1), belamcandane A (2), 3-*O*-acetyliridobelamal A (3), and belamcandal A (4) (Fig. 1) have been isolated from the rhizomes of *Belamcanda chinensis* collected from China. Iridals 1–3 feature a hydroxyl group at C-16 of the polyene side chain while triterpene 4 has a hydroxyl group at C-17.⁹ In addition, compounds 1 and 4 have an aldehyde function at C-1 and C-25, respectively. On the other hand, compound 3 is a rare iridal terpenoid which is lacking a carbonyl function at C-1 or the C-25 position. Bioassays for compounds 1–4 revealed inactivities towards various human cancer cells.⁹

Iriwattins A (5) and B (6) (Fig. 1), two unusual sixteen carbon comprising *nor*-iridal triterpenes were isolated from the flower *Iris wattii*. Furthermore, compounds 5 and 6 illustrated notable protein-tyrosine phosphatase 1B (PTP1B) enzyme inhibition of



Fig. 3 Structures of bicyclic iridals 20-29.

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52% and 14%, respectively.¹⁰ Iritectol D (7), 22-epiiritectol D (8), iritectol E (9), 22-epiiritectol E (10), iritectol F (11), and 22-epiiritectol F (12) were produced by *Iris tectorum*. All of them were isolated as pairs of C-22 epimers and all possess a hydroxyl group at C-16 of the polyene side chain while metabolites 11/12 have a C-14-ester group at the C-3 hydroxyl group of the side chain of the ring along with an additional hydroxyl moiety at C-23.¹¹

Three monocyclic iridal-type triterpenoids **13–15** (Fig. 2) were isolated from *Iris delavayi*. Iridal **13** is the geometrical isomer of **15**, with respect to the aldehyde group at C-1. In addition, metabolites **13–15** are rare iridals missing the C-10 tertiary hydroxyl group.¹² A literature survey revealed that only a few iridals described earlier are missing the C-10 tertiary hydroxyl group, such as marneral,¹³ marnerol,¹³ and 22,23-epoxy-10-deoxy-21-hydroxyiridal.¹⁴ Furthermore, compounds **13** and **15** contain an epoxide function at the C-18/C-19 position of the side chain.¹²

In another report, two iridal-type triterpenoids, named forrestins A (16) and B (17), were reported from *I. forrestii*. Both compounds are geometrical isomers of each other with respect to the aldehyde group. Moreover, compounds 16 and 17 have a tertiary hydroxyl group at C-19 of which the stereochemistry has not be established.¹⁵ Shi *et al.*¹⁶ reported iridojaponal C (18) from *I. japonica* and this compound illustrated moderate hepatoprotective effects. Moreover, iridal 18 has three hydroxyl groups in the polyene side chain. In 2018, Chen *et al.*¹⁷ reported the isolation of 17-hydroxyl-27-ene-iridal (19) from *I. confusa*. It features an exocyclic double bond between C-10/C-27 which is interestingly, also present in belamcandal A (4). Compounds having an exocyclic double at C-10/C-27 are very rare and were not reported in the previous review.⁸

2.2 Bicyclic iridals

Spirioiridoconfal C (20) along with isobelamcandal (21) (Fig. 3) were reported from related *I. confusa*. Metabolites 20 and 21 are spirioiridoconfals bearing C-19 and C-29 based exocyclic double bonds while compound 20 has an additional exocyclic bond at C-15 and C-28. Metabolite 20 (IC₅₀: 85 μ M; SI = 2.1) illustrated anti-HBV pronounced effects towards HBV DNA replication.¹⁷ A literature survey revealed that the earlier described iridotectoral B¹⁸ and hoogianal¹⁹ have a similar cyclopentane based spirocyclic core and were reported from the *Iris* species. *Belamcanda chinensis* rhizomes, collected from the Hebei province in China, produced two unusual iridal triterpenes named belamcanolide A (22), and belamcanoxide A (23). The latter compound demonstrated good cytotoxic effects towards colon (HCT-116), liver (HepG2), gastric (BGC-823), and lung (NCI-H1650) cancer cells with IC₅₀ ranging from 3.2 to 7.5 μ M.²⁰

Two *nor*-iridals named belamcanoxide B (24) and 16-O-acetylbelamcanoxide B (25) were isolated from *Belamcanda chinensis*. Furthermore, belamcanoxide B (24) illustrated good cytotoxic effects towards MCF-7 and HCT-116 cell lines with IC₅₀: 3.4 and 5.6 μ M respectively.⁹ In addition, this plant produces two further iridal lactones *viz.*, belamcanolides B (26), and C (27).⁹ Compounds 22, 26, and 27 are rare examples of a 5/ 6-fused bicyclic iridal lactone, in which these metabolites bear a furanone ring (lactone ring) attached between C-7 and C-8. Interestingly, compounds 23–25 feature an ether linkage between C-7 and C-10 of the 6-membered ring. In addition, molecule 23 is a structurally unique iridal having an isolated aldehyde in place of the α , β -unsaturated aldehyde moiety which is common for iridal-type triterpenoids. It is interesting to note that compounds 22 and 26 are C-8 epimers while iridals 22, 23, and 26 feature a hydroxyl group at C-16.⁹

Both geometrical isomers *viz.*, iritectols C (28)¹¹ and G (29)²¹ (Fig. 3), include the tetrahydrofuran moiety between C-16/C-19 in the polyene side chain, and are produced by *Iris tectorum*. Iritectol 28 illustrated good cytotoxic effects towards liver cancer (HePG2) with IC₅₀: 4.1 μ M. On the other hand, it also possessed moderate effects towards ovarian cancer (A2780: IC₅₀: 6.5 μ M), colon cancer (HCT-16: IC₅₀: 9.7 μ M), and lung cancer (NCI-H1650: IC₅₀: 9.8 μ M).²¹ Additionally, iritectol G (29) illustrated anti-epileptic effects *via* inhibiting spontaneous (IC₅₀: 8.2 μ M) and 4-aminopyridine-evoked calcium oscillations (IC₅₀: 12.5 μ M).²¹

2.3 Tricyclic iridals

Li *et al.*²² reported four iridal-type triterpenoids named polycycloiridals P-T (**30–34**) (Fig. 4) from *Belamcanda chinensis*, which is an ornamental plant of the Iridaceae family. These bicyclic metabolites have an additional cyclopentane core unit resulting from a C-18 and C-22 cyclization of the homofarnesyl chain. Furthermore, iridals **30/31** and the corresponding **32/33** are pairs of $\Delta^{2(7)}$ olefinic geometric isomers. Moreover, compound **34** has an ethoxy group at C-12 and based on extraction protocols, the authors proposed that this compound might be an artifact, although as yet, not proven.

Investigations of *B. chinensis* extracts yielded two unusual tricyclic-iridal triterpenoids named belamcandanes A (35) and B (36) (Fig. 4). It was found that these metabolites carry a rare α -terpineol moiety in their polyene side chain. Both compounds



Fig. 4 Structures of tricyclic iridals 30-36.



Fig. 5 Structures of tricyclic iridals 37-49.

possessed moderate hepatoprotective effects while they were not active towards NCI-H1650, HepG2, BGC 823, HCT-116, and A2780 cancer cells.²³ Iridals **30–34** feature a spiro furan ring formed *via* the C-13 hydroxyl group attack to a C-26 aldehyde group. A literature survey revealed that iridotectoral A,¹⁸ (13*R*)iridotectoral C,²⁴ and D,¹⁸ have a similar furan based spiro cyclic core reported from other *Iris* species.

A further three iridal-type triterpenoids, iritectol G (37), iritectol H (38) and 19-*epi*-iritectol H (39) (Fig. 5) bearing a rearranged homofarnesyl side chain and a 6/7/5 core system, were also reported from *I. tectorum*. Moreover, compound 37 is the second compound having been assigned the same name *viz.*, iritectol G because compound 29 has already been reported with the same name. Iridals 38 and 39 are C-19 epimers and this diastereomeric mixture was shown to possess moderate neuroprotective effects.²⁵

In another report, iridojaponal A (40) and B (41) were produced by *Iris japonica*. Notably, iridojaponal A (40) and B (41) are sixteen carbon comprising degraded iridals. Iridals 40 and 41 demonstrated moderate hepatoprotective effects.¹⁶ Chen *et al.*¹⁷ reported that the spirioiridoconfals A (42) B (43) were isolated from *I. confusa*. Metabolites 42 and 43 are spiro-iridals bearing a C-16 and C-26 based tetrahydrofuran ring.

Six iridal-type triterpenoids, named spirioiridotectals A-F



Fig. 6 Structures of tetracyclic iridals 50-64.



Fig. 7 Structures of miscellaneous iridals 65-68.

(44–49) representing three pairs of geometrical isomers (with respect to aldehyde group), were produced by *I. tectorum*. Interestingly, compounds 44–49 feature a 3,6-dihydro-2*H*-pyran core between C-17 and C-26. Moreover, iridals 44 and 45 are geometrical isomers of compounds 46 and 48 with respect to the *exo*-C-18/19-double bond only. Furthermore, metabolites 44, 45 and 49 possess moderate neuroprotective effects towards PC12 cell.²⁶ In addition, spirioiridotectal D (45) suppressed NO production with an IC₅₀: 0.54 μ M.²⁷

2.4 Tetracyclic iridals

Four new iridal polycycloiridals A–D (**50–53**) (Fig. 6), having an unusual α -terpineol group were isolated from *I. tectorum*. Iridals **50** and **52** have a distinct absolute configuration at the α -terpineol group. Moreover, compounds **50–53** illustrate hepatoprotective effects towards D-galactosamine-induced toxicity with activity ranging between 65 to 96%.²⁸ In another report, polycycloiridals

E–J (54–59) were reported from *I. tectorum*. These compounds possess a 6/5/7 tricyclic core, as observed earlier. In addition, metabolites 54–59 possess a cyclopentane ring which is suggested to be formed *via* cyclization between C-18 and C-22 of the homofarnesyl chain. Furthermore, these compounds contain two *E* configured double bonds at $\Delta^{14(15)}$ and $\Delta^{16(17)}$.²⁷ Li *et al.*²² reported five tricyclic iridal-type triterpenoid analogs bearing the 6/5/7-core named polycycloiridals K–O (60–64) from *Belamcanda chinensis* and these metabolites possess a cyclopentane core resulting *via* cyclization between C-18 and C-22 of the homofarnesyl chain. Moreover, these compounds illustrated weak PTP1B inhibitory effects.

2.5 Miscellaneous iridal triterpenoids

Three spiroiridal-isoflavone hybrid compounds named belamcandanins A–C (**65–67**) (Fig. 7), have been isolated from *B. chinensis*. Metabolites **65–67** represent rare iridal-flavone hybrid compounds. Biological investigation of these compounds demonstrated that they were not active towards lung cancer (NCI-H1650), hapetocelluar cancer (HepG2), stomach cancer (BGC 823), colon cancer (HCT-116), and breast cancer (MCF-7).²⁰ An unusual dimeric iridal-type triterpenoid, named dibelamcandal A (**68**) having two iridal type triterpenoid units and linked *via* a cyclohexene ring (gives rise to dimeriziaton *via* a [4 + 2]-cycloaddition), was reported from *B. chinensis*. Furthermore, this compound illustrates promising molluscicide effects towards *Pomacea canaliculate* with an LC₅₀: 1.3 µg mL⁻¹.²⁹

3 Biosynthesis

3.1 Biosynthesis of monocyclic iridals

Oxidosqualene cyclases are key enzymes which are responsible for the intriguing structural diversity of triterpenes derived from oxidosqualene (**69**)^{30,31} (Scheme 1). Over 100 triterpene skeletons have been reported from oxidosqualene (**69**) *via* catalytic variability in cation– π annulation, ring expansion, hydride or 1,2-methyl shifts, and deprotonation. In 2006, Xiong *et al.*³⁰ experimentally proved that oxidosqualene (**69**) is converted into bicyclic **72** *via* cyclization, hydride and 1,2-methyl shifts, which subsequently undergoes a Grob-type fragmentation to achieve monocyclic iridal skeleton **73**. The latter compound could be reduced or oxidized in order to achieve iridals **74a,b** (see also



Scheme 1 Biosynthesis of monocyclic iridals 1–19. Reproduced from ref. 30 and ³¹ with permission from the John Wiley and Sons.



Fig. 1). The study further demonstrated that the *Arabidopsis* 77 *thaliana* gene At5g42600 is responsible to encode an oxial dosqualene cyclase acting as a catalyst for the Grob-type fragmentation. In addition, further hydroxylation and dehydration, the and dehydrogenation in the polyene side chains provided iridals **1–19**. (7

3.2 Biosynthesis of bicyclic iridals

Plausible biosynthetic pathways for some bicyclic iridals were proposed as shown in Schemes 2 and 3.²³ Upon oxidation of the Me-26 groups in iridal **75a** and its geometric isomer **75b**,^{23,32,33} nucleophilic attack of the C-14 double bond to the C-26 aldehyde carbonyl (**76a,b**) forms the 6/5 spiro bicyclic skeletons 77**a,b** in an ene-type reaction. Subsequently, further functionalization would be possible at the unsaturated polyene side chain *via* hydroxylation, dehydrogenation, and acetylation and thus could provides iridals **20** and **21**.

After oxidation of the C-25 aldehyde moiety in iridobelamal A (77) along with C-8 hydroxylation, the subsequent lactonization between the CO₂H group at C-25 and the C-8 hydroxyl group to belamcanolide A (22) is initiated (Scheme 3). Moreover, belamcanoxide B (24) and 16-*O*-acetylbelamcanoxide B (25) are suggested to be derived from belamcanoxide A (79), which would involve an oxa-Michael addition, and acetylation to lead to the target compounds 24 and 25.²⁰ On the other hand, iritectols C (28) and G (29) could biosynthetically be derived from iridobelamal A (77) and its isomeric isoiridogermanal (79),



Scheme 3 Plausible biosynthesis of bicyclic iridals 22, 24, 25, 28, and 29. Reproduced from ref. 11 and ²⁰ with permission from the Elsevier and Royal Society of Chemistry respectively.

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Scheme 4 Plausible biosynthesis of tricyclic iridals 30-33, 35, and 36. Reproduced from ref. 23 with permission from the Elsevier.

which after epoxidation at C-18/C-19 following by nucleophilic attack of the C-16 hydroxyl group to C-19 is formed.11

Biosynthesis of tri- and tetracyclic iridals 3.3

The biosynthesis of tri- and tetracyclic iridals 30-33 is initiated by C-13 hydroxylation in compound 76 to yield intermediate 82a,b, followed by nucleophilic attack of the C-13 OH group to the C-26 aldehyde carbonyl to achieve the 6/5 spiro bicyclic hemiacetals 83a,b (Scheme 4). Subsequently, epoxidation at C-18/C-19 which is consequently opened by hydroxyl attack at C-23 is followed by the cascade reaction resulting in attack of the resulting C-22 nucleophile at C-18 of the epoxide ring leading to the C-22/C-18 cyclopentane ring systems (85a,b). The latter compound could then undergo a dehydration of the tertiary alcohol to produce iridals 30-33. On the other hand the C-22/C-23 double in the side chain could

be epoxidized to form 86a,b and the iridals 35/36 could then be formed through cyclization of the unsaturated side chain via nucleophilic attack of the C-16/C-17 double to C-22 followed by C-13 hydroxylation.23

The biosynthesis of iridals 37-39 and 54-63 is initiated by acetal formation between the C-3 hydroxyl group and C-26 hemiacetal in 87a,b affording ketal 88a,b which is followed by epoxidation at C-18/C-19 to epoxide 89a,b (Scheme 5).25 Further conversion steps viz., nucleophilic addition of an OH at C-23, nucleophilic attack of the double bond to C-18, and dehydration lead to iridals 54-63. The final formation to iridals 37-39 could be considered by an additional epoxidation at C-15/C-16 to form 90a,b followed by attack at the epoxide ring and cleavage of the cyclopentane ring leading to the ring-opened hydroxy ketones 37-39.25



Hypothetical biosynthesis of tricyclic iridals 37–39 and 54–63. Reproduced from ref. 25 with permission from the Elsevier. Scheme 5



Scheme 6 Hypothetical biosynthesis of tricyclic iridals 40-43 and 48-53. Reproduced from ref. 16 with permission from the Elsevier.

For the formation of iridals 40-43, 48-53, spirocyclic 77a,b (Scheme 6) undergoes a 5-exo trig cyclization followed by C-15/C-28 epoxidation and then resulting epoxide hydrolysis could provide iridals 42 and 43. Similarly, the same intermediate might also undergo 6-endo trig cyclization followed by C-15/C-28 epoxidation and dehydration to produce iridals 48 and 49. According to Baldwin's rule, 5-exo trig and 6-endo trig cyclizations are favored but enzymes can possibly catalyze the chemically disfavoured pathways as well.34 Moreover, the biosynthetic precursor of iridals 50-53 might possibly be traced back to either epi-anhydrobelachinal or iso-anhydrobelachinal (91a,b),28,35 which could undergo a series of conversions such as epoxidation at C-22/C-23, nucleophilic hydroxyl addition at C-14, and intramolecular cyclization to form 50–53 (Scheme 6). The biosynthetic route of iridals 40 and 41 could commence from iridals 50-53 via acid catalysis to produce the key intermediates 94a,b. Subsequently, intermediates 94a,b might undergo C-13/C-14 bond oxidative cleavage to afford 95a,b. Finally, dehydration of the later compound could form the degraded iridals, iridojaponals A (40) and B (41).16

3.4 Biosynthesis of miscellaneous iridals

Belamcandal (96) or its geometrical isomer could initially undergo deacetylation, double bond isomerization, and C-17

hydroxylation. Moreover chlorination of the resulting hydroxyl group could afford the chlorinated product **98**, which might undergo dechlorination to provide cation **99** (Scheme 7). Subsequent, Friedel–Crafts alkylation between later cation **99** with isoflavone **100** *via* typical Friedel–Crafts alkylation mechanism would produce belamcandanins A-C (**65–67**).²⁰ On the other hand, dibelamcandal A (**68**) could be biosynthesized through the $[4 + 2]_{\pi}$ Diels–Alder reaction at C-29/C-29' and C-17/C-19' between the two monomers **101** and **102** (Scheme 7).²⁹

4 Tricyclic 6-6-5 triterpenes

4.1 Isomalabaricanes-type terpenes

Literature revealed that the genus *Rhabdastrella*, which can be found in tropical oceans³⁶ produced various isomalabaricane triterpenes with intriguing chemical diversity and demonstrating a wide range of biological effects.^{37–39} Isomalabaricanes are a group of tricyclic triterpenes bearing a 6/6/5 *trans-syn-trans* tricyclic core attached to a polyene side chain (Table 1, structure "B"). The difference between isomalabaricanes and malabaricanes is in the stereochemistry of the B–C ring junction: *trans-syn-trans* (iso) and *trans-anti-trans*, respectively. The overwhelming number of structural features distinguishing these metabolites can be found in the terpenoid side chain, which



Scheme 7 Plausible biosynthesis of miscellaneous iridals 65–68. Reproduced (with slight modification from 97 to 65–67) from ref. 20 and ²⁹ with permission from the Royal Society of Chemistry and American Chemical Society respectively.

consists of acyclic isoprene-units up to sesquiterpenes. The side chains remain acyclic, or depending on the oxidation state, have been cyclized into gamma and delta lactones. Carbocyclic moieties within the side-chains most likely result from ene- or aldol reactions.

Among the isomalabaricanes isolated from R. aff. distincta (103-108), only rhabdastrellin A (103) showed moderate cytotoxic effects toward leukemia (HL-60: IC_{50} : 4.2 µg mL⁻¹), which may be related to the 2-pyranone modified side chain and which closely resembles previously reported Jaspolides A and B.40 Rhabdastrellins B-F (104-108) contain only acyclic side chains altering in their oxidized positions, although the stereochemistry of the C-24 hydroxyl group of the side chain in Rhabdastrellin B (104) could not be unambiguously established. For the relative stereoconfiguration of isomalabaricanes, it was demonstrated during isolation, that these triterpenoids are isomerized at C-13 via exposure to light and thus, it was suggested that the 13-Z isomers might be artefacts rather than new entities.36 In another report, Kiem et al.41 reported isomalabaricane derived nor-terpenoids named rhabdastrellins G-K (109-113) from the sponge R. providentiae. Compounds 109-111 possessed cytotoxic effects towards HepG2, MCF-7, LU-1, SK-Mel2, and HL-60 with IC_{50} ranging from 11 to 85 $\mu M.$ Notably, the 20(22)-Z isomer 110 illustrated much better cytotoxic effects (IC₅₀: from 11 to 16 μ M) in comparison with the 20(22)-*E* isomer **109** (IC₅₀: from 56 to 87 μM).

The marine sponge *R. globostellata* produced the side-chain shortened isomalabaricane triterpenes globostelletins B–I (**114–121**) (Table 1).⁴² Compounds **114–121** were tested against various cancer cell lines with only moderate activities. Again,

among the 13*E*/13-*Z* isomers, globostelletins H (120) and I (121), isomer 121 (13-*Z*) illustrated better inhibitory effects towards A2780 (IC₅₀ = 7.6 μ M) than its *E* isomer *viz.*, compound 120 (13-*E*) (A2780: IC₅₀ = 8.1 μ M).

The same organism also produced globostelletins J–R (122–130) bearing cyclopentane modified side chains. Natural products 122–126 have the five-membered cyclopentane core formed between C-15 and C-23 in the polyene side chain while compounds 127 and 128 (Table 1) feature a 25-dimethyltetrahydrofuran moiety, which was joined to another cyclopentane core such as 20-hydroxy-20-methylcyclopentane at the C-17 and C-24 positions.⁴³ Moreover, compounds 129 and 130 bearing a substituted cyclopentane core feature a hydroxyl group at C-22. Globostelletins K (123) and L (124) illustrated moderate inhibition towards focal adhesion kinase (FAK), protein kinases (ALK), Aurora-B, and VEGF-R2. On the other hand, the remaining compounds were not active towards sixteen tumor-related protein kinase enzymes.⁴³

In another report the sponge *Stelletta* sp. collected from Hainan, China, produced the isomalabaricane triterpene stellettin N (131).⁴⁴ This single member was associated with a natural product family resulting from isolation studies done on *St. tenuis*. In this regard Li *et al.*⁴⁵ isolated isomalabaricane triterpenes *viz.*, stellettins N–P (132–134) from this marine sponge. Unfortunately, one of the compounds *viz.*, 132 was again published as "stellettin N" because metabolite 131 was already named as "stellettin N".⁴⁴ Some features of compound 132 are unique such as the C-13 β-hydroxyl group as well as the saturated C-13/C-14 bond. The vast majority of this family have fully conjugated polyene side chains. Very few isomalabaricane

12	22 17 17	27			°
19 3 5 10 8 10 8 10 10 8 10 10 10 10 10 10 10 10 10 10	B baricane skeleton	²⁶ R ¹ / ₁ R ² 103-13 and 15	H H 3 1, 133-153 5-166		
Compound	R ¹	R^2	R ³	R ⁴	Source
Rhabdastrellin A (103)	Н	ОН	CH ₂ OH	× ~ ~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	Rhabdastrella aff. distincta ³⁶
Rhabdastrellin B (104)	Н	ОН	CH ₂ OH	132 OH	Rhabdastrella aff. distincta ³⁶
Rhabdastrellin C (105)	Н	ОН	CH ₂ OH	13.34 HOH	Rhabdastrella aff. distincta ³⁶
Rhabdastrellin D (106)	Н	ОН	CH ₂ OH	Starton Con	Rhabdastrella aff. distincta ³⁶
Rhabdastrellin E (107)	Н	ОН	Ме	13.24 CO ₂ Me	Rhabdastrella aff. distincta ³⁶
Rhabdastrellin F (108)	Н	ОН	Ме	13 2 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5	Rhabdastrella aff. distincta ³⁶
Rhabdastrellin G (109)	0	—	Me		Rhabdastrella providentiae ⁴¹
Rhabdastrellin H (110)	О	_	Ме	×~>	Rhabdastrella providentiae ⁴¹
Rhabdastrellin I (111)	Ο	_	Ме	HO OF	Rhabdastrella providentiae ⁴¹
Rhabdastrellin J (112)	Н	OAc	Ме	СНО	Rhabdastrella providentiae ⁴¹
Rhabdastrellin K (113)	Н	OAc	Ме	СНО	Rhabdastrella providentiae ⁴¹
Globostelletin B (114)	0	—	Me	=§CO2H	Rhabdastrella globostellata ⁴²
Globostelletin C (115)	0	_	Ме	135 CHO	Rhabdastrella globostellata ⁴²
Globostelletin D (116)	О	—	Me	135 CHO 132	Rhabdastrella globostellata ⁴²
Globostelletin E (117)	0	_	Ме	13 5 5 500 CO ₂ H 13E	Rhabdastrella globostellata ⁴²
Globostelletin F (118)	0	_	Me	135 CO ₂ H	Rhabdastrella globostellata ⁴²
Globostelletin G (119)	0	_	Ме	13-34 CO ₂ H	Rhabdastrella globostellata ⁴²
Globostelletin H (120)	О	_	Me	13 CO ₂ H	Rhabdastrella globostellata ⁴²

13*E*

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Table 1 (Contd.)

19 1 1 10 10 10 10 10 10 10 10	3 30 18 B Iabaricane skeleton	27 25 26 R ¹ / ₂ R ² 103-13 and 15	0 H H 3 1, 133-153 5-166		
Compound	\mathbb{R}^{1}	\mathbb{R}^2	R ³	R^4	Source
Globostelletin I (121)	Ο	_	Ме	13 CO ₂ H	Rhabdastrella globostellata ⁴²
Globostelletin J (122)	0	_	Ме		Rhabdastrella globostellata ⁴³
Globostelletin K (123)	0	_	Ме		Rhabdastrella globostellata ⁴³
Globostelletin L (124)	0	_	Me	s 	Rhabdastrella globostellata ⁴³
Globostelletin M (125)	0	_	Ме	=3 H Ac	Rhabdastrella globostellata ⁴³
Globostelletin N (126)	0	_	Ме	$= \begin{cases} S_{1} \\ S_{2} \\ S_{1} \\ S_{2} \\ $	Rhabdastrella globostellata ⁴³
Globostelletin O (127)	Н	ОН	CH ₂ OH		Rhabdastrella globostellata ⁴³
Globostelletin P (128)	Н	ОН	CH₂OH		Rhabdastrella globostellata ⁴³
Globostelletin Q (129)	Н	ОН	CH ₂ OH	HO, HIII HIII HIII HIII OH	Rhabdastrella globostellata ⁴³
Globostelletin R (130)	Н	ОН	CH ₂ OH	HO, HIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	Rhabdastrella globostellata ⁴³
Stellettin N (131)	Н	OAc	Ме	-25	Stelletta sp. ⁴⁴
Stellettin O (133)	0	_	Ме	When the second	Stelletta tenuis ⁴⁵
Stellettin P (134)	0	_	Ме	Me OH	Stelletta tenuis ⁴⁵
Stellettin Q (135)	Н	OAc	Ме		Stelletta sp. ⁴⁷

Table 1 (Contd.)

$ \begin{array}{c} & & & & & & & & & & & & & & & & & & &$					
Compound	R^1	R^2	R ³	R^4	Source
Stellettin R (136)	Н	OAc	Me		Stelletta sp ⁴⁷
Auroral 137	ОН	Н	CH ₂ OH	132 CHO	Rhabdastrella globostellata ⁴⁸
Auroral 138	ОН	Н	CH ₂ OH	13 5 John CHO 13E	Rhabdastrella globostellata ⁴⁸
Auroral 139	ОН	Н	CH ₂ OH	135 55 CHO 13Z	Rhabdastrella globostellata ⁴⁸
Auroral 140	ОН	Н	CH ₂ OH	13 5 13E CHO	Rhabdastrella globostellata ⁴⁸
Jaspiferin C (141)	0	_	Me		Jaspis stellifera ⁴⁹
Jaspiferin D (142)	Н	ОН	CH ₂ OAc	13 the second second	Jaspis stellifera ⁴⁹
Jaspiferin E (143)	Н	ОН	CH ₂ OAc	132	Jaspis stellifera ⁴⁹
Jaspiferin F (144)	Н	ОН	Me		Jaspis stellifera ⁴⁹
Jaspiferin G (145)	Н	ОН	CH ₂ OH	= € - E H OH	Jaspis stellifera ⁵⁰
Jaspiferin H (146)	0	_	Ме	HO HO	Jaspis stellifera ⁵¹
Jaspiferin I (147)	Н	ОН	Ме	73 5 55 mm CHO 13Z	Jaspis stellifera ⁵¹
Jaspiferin J (148)	Н	ОН	Ме	13 5 - 5 - 5 - 5 - 5 - 5 - 5 - 5 - 5 - 5	Jaspis stellifera ⁵¹
Rhabdaprovidine A (149)	Н	OAc	Me	13 5 OMe 13 7 OMe 132	Rhabdastrella providentiae ⁵²
Rhabdaprovidine B (150)	Н	OAc	Ме	OMe 13 5 13 5 13 13 13 13 13 13 13 13 13 13 13 13 13	Rhabdastrella providentiae ⁵²

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Table 1 (Contd.)

19 10 10 10 10 10 10 10 10 10 10	15 17 10 10 10 10 10 10 10 10 10 10	27 25 26 R ¹ , R ² , 103-131 and 155	0 H H 133-153 5-166		
Compound	R^1	R^2	R ³	R^4	Source
Rhabdaprovidine C (151)	Н	OAc	Ме	OMe =§=√OMe	Rhabdastrella providentiae ⁵²
Rhabdaprovidine D (152)	0	_	Ме	-ξ=<⊂ ^{CO} 2Me	Rhabdastrella providentiae ⁵³
Rhabdaprovidine E (153)	0	_	Ме	-ξ=<⊂ ^{OH}	Rhabdastrella providentiae ⁵³
Stelliferin J (155)	Н	OAc	Ме	H CH CH	Rhabdastrella globostellata ⁴⁶
Stelliferin K (156)	Н	OAc	Ме	HO OH	Rhabdastrella globostellata ⁴⁶
Stelliferin L (157)	Н	OAc	Me	жон Гон	Rhabdastrella globostellata ⁴⁶
Stelliferin M (158)	Н	OAc	Me	A Contraction of the second se	Rhabdastrella globostellata ⁴⁶
Stelliferin N (159)	Н	OAc	Ме	же сторинание и сторина Сторинание и сторинание и сторинан	Rhabdastrella globostellata ⁴⁶
Rhabdastin A (160)	Н	OAc	Ме	-§=⟨CO₂Me	Rhabdastrella globostellata ⁵⁴
Rhabdastin B (161)	Н	ОН	Ме	=⋛=<⊂ ^{CO} 2Me	Rhabdastrella globostellata ⁵⁴
Rhabdastin C (162)	Н	OAc	CO ₂ Me	=§=<	Rhabdastrella globostellata ⁵⁴
Rhabdastin D (163)	Н	OAc	Me	H H H	Rhabdastrella globostellata ⁵⁴
Rhabdastin E (164)	Н	OAc	Me	H H H H H H H H H H H H H H H H H H H	Rhabdastrella globostellata ⁵⁴
Rhabdastin F (165)	Н	OAc	Me	H 136 H OH	Rhabdastrella globostellata ⁵⁴
Rhabdastin G (166)	Н	OAc	Ме	H H H H H H H H H H H H H H H H H H H	Rhabdastrella globostellata ⁵⁴

triterpenes have been reported which have only one or two of the normally unsaturated bonds saturated *viz.*, stelliferins J–N and globostelletins J–P.^{43,46} Compounds **132–134** were evaluated against lung cancer (A549), gastric cancer (AGS), and glioblastoma (U-251MG) where it was found that these compounds were only active towards AGS with IC₅₀: in the range from 4.5 to 9.6 μ M.⁴⁵ Stellettins Q (**135**) and R (**136**) were isolated from *Stelletta* sp. and both compounds have a cyclopentane ring in the polyene side chain formed between C-15 and C-22. Moreover, isomalabaricane triterpenes **135** and **136** (Table 1) are a 13-*Z*/*E* geometric pair of the same 15*R*,23*S* isomers.⁴⁷

Isomalabaricanes such aurorals **137–140** are produced by the sponge *R. globostellata*. Biological studies revealed that aurorals **137** and **138** possessed good cytotoxic effects towards epidermoid carcinoma (KB cells) with ID_{50} : 0.2 µg mL⁻¹. On the other hand, compounds **139** and **140** displayed moderate effects with an ID_{50} : 8 µg mL⁻¹.⁴⁸

Jaspiferins C-F (141-144) were isolated from the marine sponge Jaspis stellifera and jaspiferin C (141) has a sixmembered carbon core formed between C-15 and C-23 which is rather rare among isomalabaricane triterpenes.⁴⁹ An isomalabaricane triterpenoid, jaspiferin G (145), comprising a cyclopentane ring in the pendant polyene side chain was produced by the sponge Jaspis stellifera.50 Jaspiferins H-J (146-148) were obtained from J. stellifera, among which Jaspiferin H (146) has a five membered lactone ring (γ -lacone).⁵¹ From the sponge R. providentiae Dung et al.52,53 reported rhabdaprovidines A-E (149-153) and G (154). The latter one, rhabdaprovidine G (154) has an additional pyran ring formed between C-12 and C-15. Notably, this metabolite possesses a novel chemical structure of the 6-6-5-6-tetracyclic core along with nine chiral carbon centres which is quite unique among isomalabaricane skeletons.53 Bioactivity studies revealed that rhabdaprovidines A-C (149-151) inhibited NO production in LPS in which the IC₅₀: ranged from 17.5 to 46.8 μ M with 150 (Table 1) being the most potent IC50: 17.5 µM.52

Stelliferins J–N (155–159) were isolated from the sponge *R. globostellata* collected from Hainan Island, China. The configuration of the stereogenic centres in triterpenoids 155–159 were determined *via* CD and NMR measurements using Mosher's method. All these natural products possess a diol (secondary/tertiary hydroxyl group) in the polyene side chains with terpenes 155 and 156 featuring a acyclic side chain and 157–159 a cyclopentane diol unit. Metabolites 157 and 159 illustrated antimicrobial effects towards *Bacillus subtilis* with IC₅₀ 8 µg mL⁻¹.⁴⁶

Rhabdastins A–G (**160–166**, Table 1) were produced by the sponge *R. globostellata*. Furthermore, compounds **160–166** have a cyclopentane group generated in the conjugated polyene side chains.⁵⁴ Compounds **163**, **164**, and **166** demonstrated modest cytotoxicity on leukemia cells (HL-60) with IC₅₀: 21, 29, and 44 μ M, respectively. Isomalabaricanes **163** and **164** have been studied in detail revealing to cause caspase 3 cleavage in HL-60 cells at 10 μ M without effecting expression of Bcl-2 in HL-60 cells.⁵⁴ Among the latest natural products of this class, Lai *et al.*⁵⁵ isolated rhabdastins H (**167**) and (**168**) (Fig. 8) from the sponge *Rhabdastrella* sp. These isolates demonstrate antiproliferative effects towards leukemic cells (Molt4 and K562)



Fig. 8 Structures of isomalabaricane-type triterpenoids 167,168 and dimeric isomalabaricane 169,170.



Fig. 9 Structures of isomalabaricane-derived *nor*-terpenoids 171–177.

with IC₅₀ values ranging from 9.8 to 16.5 μ M. Jaspolides G (169) and H (170), two unusual bisisomalabaricanes isomers were reported from the marine sponge *Jaspis* sp.⁵⁶ The intriguing structures not only involve two isomalabaricane moieties, they also possess 2-pyrone heterocycles, which makes the C₆₀ natural product even more impressive.

4.2 Isomalabaricane-derived nor-terpenoids

Two C-19-nor isomalabaricane triterpenes named cyclobutastellettolides A (**171**) and B (**172**) were produced by *Stelletta* sp. (Fig. 9). The bioassay-guided isolation revealed them to significantly enhance reactive oxygen species (ROS) generation in murine macrophages. Notably, these two compounds comprise four fused rings, of which one is a cyclobutanone and this is indeed a rare feature in isomalabaricane triterpenes.⁵⁷

In another report, isomalabaricane-derived *nor*-terpenoids, stellettins S–V (**173–176**) were isolated from the marine sponge *Stelletta* sp. Stellettin S (**173**) represents an intriguing example

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because this compound has an alkyne group and this is the first example of an isomalabaricane triterpene which features this rare functionality as a substituent.⁴⁷ The marine sponge *R. globostellata* produces *nor*-isomalabaricanes triterpene globostelletin A (177) which has a rather unique structure.⁴²

4.3 Malabaricane type triterpenes

Malabaricanes are a group of tricyclic triterpenes bearing a 6-6-5 trans-anti-trans tricyclic core attached to a polyene side chain (Fig. 10, structure "C"). Six malabaricane type triterpenes 178, 179 with cyclobutane modified side chains, and 182-185 (Fig. 10) were reported from the oleoresin of Ailanthus malabarica.58 During the time covered by this report, Duan et al.59 demonstrated that triterpenes 178 and 179 with the suggested unusual cyclobutane modifications have the same NMR data (in particular ¹³C NMR data) as the dammarane-type triterpenenes ocotillone (180) and gardaubryone C (181), respectively, leading to a revision of the formerly proposed structures. Of note, their structural reassignment was established by further using DFT chemical shift analysis and the CASE algorithm. Bio-guided isolation studies on Commiphora africana provided the malabaricane triterpene commafric A (186) bearing a methyl group at C-13 instead of C-8. It illustrated significant cytotoxic effects

towards lung cancer lines (A549: IC_{50} : 4.52 $\mu g~mL^{-1})$ along with moderate effects towards ovarian cancer (A2780), stomach cancer (SNU638), and pancreatic cancer (MIA-PaCa-2), with $IC_{50} \sim 10~\mu g~mL^{-1}.^{60}$

Modified side chains have been identified in ailanthusins F (187) and G (188) (Fig. 10), which are produced by the stem of *Ailanthus triphysa*, a tree commonly found in Thailand and Australia. Moreover, these metabolites were not active in cytotoxicity screening towards MOLT-3, HepG2, MDA-MB-321, HuCCA-1, T47-D, A549, HeLa, and MRC-5.⁶¹ In another report, Messina *et al.*⁶² isolated malabaricatrienone (189) and malabaricatrienol (190) from the methanol extract of the resin of *Bursera microphylla* which is called "elephant tree" and is widely present in Mexico. In addition, these metabolites were not active against various murine and human cancer cells.

5 Biosynthesis of isomalabaricane and malabaricane trietrpenoids

Isomalabaricanes and malabaricanes feature 6-6-5 *trans-syntrans* and *trans-anti-trans* tricyclic cores while both skeletons have different stereochemistry at the C-8 and C-9 positions (Table 1 and Fig. 10). The basic core skeleton of isomalabaricane **195** is thought to be biosynthetically generated



Fig. 10 Structures of malabaricane-type triterpenoids 178, 179, 182-190.

through cyclizations of 2,3-oxidosqualene (69) *via* chair– \Box boat–chair (191) transition states (Scheme 8).^{57,63,64} Similarly, it has been suggested that the biosynthesis of the basic core skeleton of malabaricane triterpenoids also proceeds through cyclizations of 2,3-oxidosqualene (69) *via* a chair–chair–chair (193) transition state^{57,63,64} and triterpene synthases are responsible to generate the malabaricane basic skeleton 196 (Scheme 8).^{64,65}

It has been reported that two *Arabidopsis thaliana* genes including At5g4810 and At4g15340 encode OSCs which furnish arabidiol and thalianol.^{8,66,67} In addition, numerous

malabaricanes have been obtained *via* site-directed mutagenesis of *A. acidocaldarius* squalene hopene cyclase which in turn comprise point mutants of Tyr420, Phe601 and Ile261.^{8,68,69} This suggests that the malabaricanes are produced in nature with custom tailored enzymes. Lodeiro *et al.*⁶⁵ demonstrated that the isomalabaricane core arises from a lanosterol cyclase mutant (Tyr510) while the native enzyme has yet to be discovered. Stelletin E (**197**) (Scheme 9) could be the plausible precursor to generate globostelletins B (**114**), D (**116**), F (**118**), and I (**121**) *via* oxidative degradation reactions.^{42,57,70}



Scheme 8 Hypothetical biosynthesis of isomalabaricane (195) malabaricane (196) basic skeletons. Reproduced from ref. 64 and ⁶⁵ with permission from the American Chemical Society.



Scheme 9 Hypothetical biosynthesis of isomalabaricane triterpenoids 114, 116, 118, 121, 171 and 177. Reproduced from ref. 57 with permission from the American Chemical Society.

8



Scheme 10 Hypothetical biosynthesis of isomalabaricane triterpenoids 122–124 and 127–129. Reproduced from ref. 43 with permission from Elsevier.

Globostelletin A (177) is assumed to be generated by oxidative degradation (loss of CO₂) from globostelletin B (114) and therefore it is also considered to be a member of the isomalabaricane class⁴² (Scheme 9). Moreover, globostelletin A (177) is considered as the putative biosynthetic precursor of cyclobutastellettolide A (171) (Scheme 9). Globostelletin A (177), or its hydrate 202 may be converted into cyclobutastellettolides A (171) *via* an intramolecular dehydration followed by δ -elimination. The ΔG calculation results for possible biosynthetic intermediates indicated that the seven-membered ring formation in tricyclic 171 is preferred to happen prior to the formation of the fused γ -lactone, in contrast to the simultaneous generation of the cyclobutane-fused lactone.⁵⁷ Cyclobutastellettolide B (172) was presumed to be generated by the same protocol from stellettin H or I (3-OAc derivatives of stellettin E).^{57,71}

Stellettin E (196), which co-existed in the same specimen (R. globostellata), is thought to be considered as the putative precursor for globostelletins J-L (122-124), which bear an unusual cyclopentane derived side chain (Scheme 10).43 Formation of this unit might be initiated via a 15,23-electrocyclization followed by 20,22-epoxidation to afford the cyclohexane unit 204. Moreover, the later intermediate could then undergo a semi-pinacol rearrangement, 17-hydroxylation, dehydration and methylation to generate the globostelletine triterpenes 122-124. On the other hand, 3-epi-stelletin J (206) could be derived from stellettin E (196) and possibly be the precursor for the biosynthetic formation of globostelletins O-Q (127–129). Considering the very likely epoxidation at C-24/C-25 to form epoxide 207 followed by attack by the nucleophilic $\Delta^{17(20)}$ double bond and hydroxylation of the resulting tertiary carbocation at C-20 (208), this will eventually provide globostelletin Q (129). Globostelletins O (127) and P (128) are more than likely to be derived from 129 through nucleophilic attack of the C-25 hydroxyl group at the C-16 double bond to form the pyran ring.

Stelliferins can be considered to be starting materials for another modified isomalabaricane, Jaspiferin C (141). It contains a cyclohexenyl moiety in the side chain which is most likely derived from stellettin E (196). Its biosynthesis could initially be formed by lactonization between the C-25 carboxylic



Scheme 11 Plausible biosynthesis of isomalabaricane triterpenoid 141. Reproduced from ref. 49 with permission from the Taylor ϑ Francis.

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Scheme 12 Hypothetical biosynthesis of isomalabaricane triterpenoid 163–166. Reproduced from ref. 54 with permission from the American Chemical Society.

acid group attacking the C-24 double bond followed by dehydrogenation to afford 2-pyrone-modified stelletin B (209) (Scheme 11). The resulting lactone ring in the stelletin B (209) could reasonably be ring-opened *via* an hydroxyl attack to generate intermediate 210. The later intermediate features an enolate tautomer. Finally, the cyclohexenyl moiety may be generated between C-15 and C-23 to form the resulting cyclohexenone core unit in the polyene side chain of jaspiferin C (141).⁴⁹



Scheme 13 Hypothetical biosynthesis of isomalabaricane triterpenoids 169 and 170. Reproduced from ref. 56 with permission from the Elsevier.

Rhabdastins A–C (**160–162**) are thought to be biogenetically generated *via* oxidative cleavage of stelliferin derivatives as described for globostelletin B (**114**) (Scheme 9). A proposed biogenetic route for rhabdastins D–G (**163–166**) *via* oxidation and cyclization of stelliferins is depicted in Scheme 12.⁵⁴ Stelliferin D (**212**) formation could proceed by initial C-3 acetylation and C-21 hydroxylation followed by epoxidation of the double bond at C-24 in stelliferin A (**213**)⁷² to form epoxide **214** at C-24/ C-25. Thereafter, attack by the $\Delta^{17(20)}$ double bond in later compound may cleave the epoxide and provide a tertiary carbocation at C-20 (**215**).

This could then be followed by nucleophilic attack by water from the α -face to produce rhabdastin G (166) which in turn could be followed by the C-25 OH group being lost *via* a dehydration to give rhabdastin E (164). On the other hand, rhabdastin F (165) may be a photoisomer of 164, because this later compound was slowly transformed into its C-13 isomer 165 under room light conditions. In addition, rhabdastin D (163) may presumably be formed through a similar route from stelliferin B (218)⁷² including epoxidation, epoxide ring-opening, α face nucleophilic attack by water as described for compounds 164–166. Additionally, alternative biogenetic routes for compounds 163–166 *via* formation of stelliferin C (216) from stelliferin A (213)⁷² are also depicted in Scheme 12.

Jaspolides G (169) and H (170) are the first examples of isomalabaricane based dimers and these are geometrical isomers with respect to the C-13 double bond.⁵⁶ The authors postulated that the structure of jaspolide G (169) could be biosynthetically formed between stellettin A (221)⁷³ and isogeoditin A (223)⁷⁴ which are representative isomalabaricane type triterpenes and a *nor*-triterpenes, respectively⁵⁶ (Scheme 13). Initially, hydroxylation might occur at C-13 of stellettin A (221) which would afford the intermediate 222 with a concomitant migration of the double bond at C-14/C-18. A double bond between C-23/C-24 of geoditin A (223) and the conjugated double bonds at C-15/C-16 and C-14/C-18 of the intermediate 222 might then undergo a [4



Fig. 11 Structures of triterpenes 224-228.

+ 2]_{π} cycloaddition to afford the corresponding dimer containing the newly formed cyclohexene ring. The C-13 isomerization *viz.*, *Z*-form (**169**) to *E*-form (**170**) is frequently present in isomalabaricanes due to their thermal sensitivity.

6 Tetranordammarane and polypodane triterpenoids

Three unusual triterpene saponins named nototronesides A–C (224–226) (Fig. 11), that feature a rare 6/6/9 tricyclic tetranordammarane core, are produced by *Panax notoginseng*, commonly referred Chinese ginseng. They were screened for their neuroprotective effects where it was found that metabolite 225 enhanced the cell viability remarkably by 79%.⁷⁵ *Commiphora mukul* gum resin, also known as guggul, has been employed to treat obesity, inflammation, and lipid disorders in Ayurvedic medicine. Myrrhanol C (227) and its corresponding keto analog myrrhanone C (228), were reported from this guggul gum resin.^{76,77} Furthermore, myrrhanol C (227) was reported from *Pistacia lentiscus* resin and found to possess potent effects towards prostate cancer cells.⁷⁸

7 Biosynthesis of tetranordammaranes

The aglycon of nototroneside A (224) (nototrone A, 238) represents the first example of an unusual tricyclic dammarane-type triterpene.⁷⁵ A proposed biosynthetic pathway of nototrone A (238) is depicted in Scheme 14.75 In this scheme the authors proposed that tricyclic (238) is biosynthetically derived from protopanaxadiol (229), which is a reported aglycon of numerous saponins from Panax plants.75,79,80 Initially, dehydration of the C-12 hydroxyl group of protopanaxadiol (229) would generate a double bond at the $\Delta^{12,13}$ position, which could subsequently undergo epoxidation via an enzyme-catalyzed epoxidation to afford the corresponding oxidized analog 231. This latter derivative could then undergo a fragmentation of the highly strained epoxide intermediate by cleavage of the C-13/C-14 bond to produce a nine-membered enol 232.81 The intermediate enol 232 is thought to then undergo keto-enol a plausible tautomerisation which would lead to the generation the keto anion 233 and upon a deprotonation lead to the formation of tricyclic triterpene 234. Enzymatic oxidation of 234 to acid 235, its decarboxylation and C-13 oxidation would ultimately lead to nototrone A (238).

8 Schiglautone and volvalerane type triterpenes

Chemical investigation of the evergreen shrubs *Schisandra glaucescens* (Schisandraceae) provided the schiglautane-type triterpene named schiglautone A (239) (Fig. 12), which feature an unusual 6/7/9-fused tricyclic core. The structure of this



Scheme 14 Hypothetical biosynthesis of nototroneside A (224) sapogenin nototrone A (238). Reproduced from ref. 75 with permission from the American Chemical Society.

unusual scaffold was confirmed *via* spectroscopic as well as Xray analysis. Compound **239** with an unusual 7,9-membered bicyclic moiety seems to have no precedent among reported secondary metabolites or synthetic compounds and is clearly different from numerous skeletons reported from the Schisandraceae family. Moreover, it illustrated weak cytotoxic effects towards HeLa, Hep G2, and SGC-7901.⁸² Volvalerenol A (**240**), which features a 7/12/7 tricyclic ring system, was obtained from the EtOH extracts of the roots of *Valeriana hardwickii*.⁸³



ig. 12 Structures of triterpenes 239 and 240.

9 Biosynthesis of schiglautone and volvalerane type triterpenes

As could be expected, the biosynthesis of these very unusual triterpenes was of tremendous interest. The starting point for schiglautone A (239) is thought to be derived from anyuweizic acid (241) (Scheme 15),82 which was also reported from the same plant species in large quantities. The authors postulated that triterpene 239 might conceivably be generated from 241 via enzymatic conversion reactions which include oxidation, acidification hydroxylation, and most importantly ring expansion. The key biosynthetic conversions include two ring expansions *i.e.* degradation of one isoprene unit in the purple color in intermediate 243 to form a nine-membered ring and ring expansion via the shifting of one carbon unit of the isoprene unit in green color (245) to form a seven member ring (246).82 On the other hand, the 7/12/7 tricyclic comprising triterpene volvalerenol A (240) could be biogenetically derived from two units of FPP (farnesyl pyrophosphate, 250). Initially FPP 250 could undergo head-to-tail condensation followed by a series of



Scheme 15 Hypothetical biosynthesis for schiglautone A (239) and volvalerenol A (240). Reproduced from ref. 82 and ⁸³ with permission from the American Chemical Society.

8



Fig. 13 Structures of triterpenes 255–257.

enzymatic reactions including epoxidation, dehydrogenation, electrophilic addition, and oxidation reactions (Scheme 15).⁸³

10 Unusual tricyclic seco/*nor*-lanostane

Chemical investigation of traditionally used *Kadsura coccinea*, which was collected from China, yielded one unusual tricyclic triterpenoid, kadcotrione A (255) (Fig. 13) bearing an 18(13 \rightarrow 12)-abeo-13,17-seco-14 β -lanostane skeleton.⁸⁴ Notably, metabolite 255 bears a 6/6/6-fused skeleton having a β -oriented C-14

methyl group which is not consistent with the methyl group at the C-28 position present in lanostane-type triterpenoids. In addition, this compound also features a C₁₁ side chain attached to C-14. This side chain comprises a keto, methyl, and carboxylic group. Furthermore, its absolute configuration was established *via* both the calculated and experimental ECD spectra.⁸⁴

The norlanostane triterpenoid named kadcotrione B (256), having a 14(13 \rightarrow 12)-abeo-13,18-dinorlanostane core was also produced by *K. coccinea*. Additionally, this plant produced kadcotrione C (257) which features a 14(13 \rightarrow 12)-abeo-12,13-secolanostane core system. Both triterpenes 256 and 257 have a 6/6/5 ring scaffold system which is configurationally similar to the isomalabaricane skeleton but is different from that of isomalabaricane, because of a C₁₃ or C₁₁ side chain along with a Me group at the 28 position. Furthermore, only compound 256 illustrated weak anti-HIV-1 effects with EC₅₀: 30.2 μ M. In addition, compounds 255 and 256 were not active towards SMMC-7721, A-549, MCF-7, and SW480.⁸⁴

11 Biosynthesis of seco/norlanostane

The structures of kadcotriones A-C (255–257) possess the same ring A-B *trans*-relationship to that of 12 β -hydroxycoccinic acid (258), a lanostane triterpene which was also isolated from *K*. *coccinea* together with triterpenes 255–257 (Scheme 16).⁸⁴ Based



Scheme 16 Hypothetical biosynthesis of kadcotriones A-C (255–257). Reproduced from ref. 84 with permission from the American Chemical Society.



Fig. 14 Structures of sipholanes and siphonellines 268–273.

on their structural similarities, the triterpene **258** could be considered to be the biosynthetic precursor and thus might undergo a series of enzymatic conversions including dehydration, 1,2-migration of a methyl group, and oxidation/oxidative degradations.⁸⁴ Furthermore, the key step in the biosynthesis of kad-cotriones A–C (**255–257**) is a fragmentation of the C–D ring in intermediate **261**, and oxidative cleavage of the C-13/C-17 double bond in **265** (to generate compound **255**), and, similarly, tricyclic **267** in order to biosynthesize triterpenes **256** and **257**.

12 Sipholanes and siphonellines

The sipholane skeleton may be described as a *cis*-fused cyclopentane-cycloheptane ethylene linked hexane-oxepane.⁸⁵ Investigations of the sponge *Siphonochalina siphonella* yielded the two benzoxepine-azulene linked sipholane type triterpenes, sipholenols N (**268**) and O (**269**) (Fig. 14).⁸⁶ On the other hand sipholane type triterpenoids also include the octahydro-chromene-azulene system linked⁸⁷ *via* a two carbon bridge. Jain *et al.* reported sipholane **270** from *Siphonochalina siphonella*, which possessed no cytotoxic effects toward epidermoid cancer cells such as KB-C2 and KB-3-1.⁸⁷

Siphonellines comprising of decahydro tetramethylbenzooxepine ring systems attached to cyclohexenol analogs *via* an ethylene bridge are also known. *Callyspongia* sp. produced siphonelline triterpenes named siphonellinol D (271), E (272), and siphonellinol C-23-hydroperoxide (273) (Fig. 14) and these triterpenes were inactive toward the two epidermoid cancer cells (KB-C2 and KB-3-1).⁸⁷

13 Sodwanones and neviotines

Sodwanones have structural similarities to the sipholanes as well as possess a hexane-oxepane subunit attached to a variety



Fig. 15 Structures of sodwanones and neviotines 274-280

of decalins. Chemical examination of the red sea sponge *S.* siphonella yielded five sodwanones **274–278** (Fig. 15) and these triterpenes proved to be inactive toward the two epidermoid cancer cell lines KB-C2 and KB-3-1.⁸⁷ Neviotine type triterpenoids are comprised of a pentacyclic core system including a bicyclic (octahydro-1*H*-indene) ethylene linked with a tricyclic (dodecahydro-2*H*-indeno[5,4-*b*]oxepine). Siphonochalina siphonella produced the two neviotine triterpenes named neviotines C (**279**)⁸⁸ and D (**280**).⁸⁶ Moreover, compound **280** illustrated significant inhibition of RANKL induced osteoclastogenesis with IC₅₀: 12.8 μ M.⁸⁶ On the other hand, neviotines C (**279**) illustrated moderate cytotoxic effects towards MCF-7, PC-3 and A549.⁸⁸

14 Biosynthesis of sipholanes, siphonellines, sodwanones, and neviotines

Sipholanes, siphonellines, sodwanones, and neviotines could feasibly be biosynthesized *via* two separate cascade cyclizations starting from the multiple-oxidized di- (282) or triepoxysqualenes (283). In all triterpene skeletons, cyclization of the doubly epoxidized half could be initiated by acid catalyzed enzymatic opening of the epoxide moiety in 282 and 283 (Scheme 17).⁸⁹ On the other hand, for the siphonellinoles the western part appears to follow a cascade-cyclization protocol *via* an electrophilic protonation of a double bond (Scheme 17), while the western part in sodwanones could be biosynthesized after protonation of the western half carrying a monoepoxidized terminal moiety.

A more different and complex biosynthetic route could involve the biogenesis of neviotine type triterpenes (Scheme 17).^{89,90} The starting di-epoxide in **282** could be the same as for the biosynthesis of sipholanes, and sodwanones, with an interrupted cyclization cascade, delivering the tricyclic core



Scheme 17 Hypothetical biosynthesis of sipholane, siphonelline, sodwanone and neviotine skeletons. Reproduced from ref. 89 and ⁹⁰ with permission from the American Chemical Society and Springer Nature respectively.



Fig. 16 Structures of onocerane and belamchinane-type triterpenoids 286–288.

structure **284**. Subsequently, the eastern half cyclization could be initiated *via* late-stage epoxidation of the C-15 methylene group to form epoxide **285**, thus setting up the second cascade cyclization to generate the neviotane skeleton.

15 Onocerane and belamchinanetype triterpenoids

Two onocerane triterpenoids named reinereins A (**286**) and B (**287**) (Fig. 16), were isolated *via* a bioassay guided chromatographic protocol from the bark of *Reinwardtiodendron cinereum*, found in Malaysia and Indonesia. Triterpene **286** illustrated moderate cytotoxic effects towards HL-60, A549, MCF7, and HepG2 cancer cells with IC₅₀ ranging from 11.5 to 27.8 μ M. On the other hand, compound **287** was only active towards HL-60 and A549 with IC₅₀: 27.8 and 29.7 μ M, respectively.⁹¹

In another report, the belamchinane-type triterpenoid named belamchinenin A (**288**), exhibiting an unusual 6/5/6fused tricyclic skeleton has been reported from *Belamcanda chinensis*, whose root extract is widely used in China and found successful in curing various diseases including cancer. Notably, metabolite **288** bears a half-caged tricyclic core along with a flexible geranyl group. As could be expected by the activity guided fractionation studies, compound **288** demonstrated cytotoxic effects towards NCI-H1650, HepG2, BGC 823, HCT-116, and MCF-7 cancer cells in which the IC₅₀: ranged from 2.29 to 4.47 μ M.⁹²

16 Biosynthesis of onocerane and belamchinane-type triterpenoids

Members of the natural product family of onoceranes have been isolated previously, *e.g.* α -onocerin (**291**) was discovered in 1855



Scheme 18 Hypothetical biosynthesis of onoceranes 286 and 287. Reproduced from ref. 96 and ⁹⁷ with permission from the John Wiley and Sons and American Chemical Society respectively.

and its complete structure was established in 1955.⁹³ Later, its biosynthetic route was postulated to involve two cyclizations starting from 2,3,22,23-dioxidosqualene (**289**).^{94,95} Despite the fact that these findings on α -onocerin (**291**) were established over six decades ago, the nature of the enzymes or genes that catalyze the two right and left cyclizations remained elusive. In 2016, Kushiro and his team^{96,97} investigated the biosynthesis of α -onocerin (**291**) (Scheme 18) and identified the two elusive genes from the fern *Lycopodium clavatum*, LCC and LCD of the oxidosqualene cyclase (OSC) family. The findings demonstrated that dioxidosqualene cyclase LCC catalyzed the first cyclization to get a decalin-intermediate (ring A,B) pre- α -onocerin (**290**) and subsequently onocerin synthase LCD catalyzed the second cyclization to afford α -onocerin (**291**). Moreover, reinereins A (**286**) and B (**287**) could be biosynthetically derived from α - onocerin (291) through biosynthetic conversions including oxidative ring A and D cleavage, oxidation and double bond isomerization.

Most of the iridal-type triterpenoids have the same absolute configurations at C-6, C-10, and C-11 (*e.g.* iridobelamal A (77), Scheme 19).^{92,98} Furthermore, belamchinenin A (288), which feature a novel 6/5/6-tricyclic core system, has stereochemistry at C-6, C-10, and C-11 being the same as for iridal-type triterpenoids, indicating that triterpene 288 could likely be derived from iridal-type triterpenoids. The iridal-type triterpenoid iridobelamal A (77) could be considered to be the biogenetic precursor, which after dehydratation to 293 might be converted into tricyclic 294 through intramolecular [4 + 2] cyclisation followed by oxidative and dehydrogenative steps to afford the scaffold of belamchinenin A (288).⁹²



Scheme 19 Biosynthesis of belamchinenin A (288). Reproduced from ref. 92 with permission from the Royal Society of Chemistry.



Scheme 20 Diastereoselective synthesis of 21-deoxy-iridogermanal (295).

17 Chemical synthesis of unusual cyclized triterpenoids

17.1 Isomalabaricane-type triterpenoids

17.1.1 Synthesis of 21-desoxy-iridogermanal. A welldesigned synthetic approach for the accomplishment of monocyclic triterpene 21-desoxy-iridogermanal (295)99 was established by Arseniyadis and co-workers.¹⁰⁰ The synthesis of this natural product started with the preparation of the tetrasubstituted cyclohexane 298 with two adjacent quaternary carbon centers (Scheme 20). This cyclohexane was synthesized through the intermediate 297 from the key octahydro-1Hindene analog 296 in three steps.¹⁰¹ Next, benzyl protection of the C-13 hydroxyl group in 298, followed by a selective acetal cleavage produced diol 299, which was further converted into 300 via selective protection of the primary alcohol group as its TBS derivative. The remaining secondary hydroxyl group was subsequently oxidized with the Dess-Martin protocol to afford ketone 300 which then underwent fluoride-mediated desilylation to produce a mixture of enone 301, together with C6(S) 302 which was equilibrated to the C6(R) 303 epimer.

Precursor **303** on reaction with (2-propenyl)lithium furnished the corresponding carbinol **304** by Swern oxidation of the primary hydroxl group at C-5 provided aldehyde **305**. The latter aldehyde was subjected to a Wittig olefination to afford the α , β -unsaturated ester **306** which upon a Dauben–Michno oxidative rearrangement¹⁰² gave a mixture of the aldehydes **307** *EZ* and **307** *EE*. Reduction of aldehyde **307** *EZ* under Luche reduction conditions and a subsequent silylation of the resulting primary hydroxyl group afforded **308**. Next, selective Mg/ MeOH mediated reduction of the double bond of the acrylate moiety of **308**¹⁰³ followed by LiAlH₄ mediated reduction of the resulting alcohol at C-3 provided the advanced intermediate **309**.

Debenzylation of **309** with Li/NH₃ afforded the resulting C-13 alcohol which was subsequently transformed to iodide **310**. Moreover, *E*-vinyl iodide **311** which was produced from geranyl acetone according to a published two step procedure¹⁰⁴ and using the Marshall protocol¹⁰⁵ for a Suzuki–Miyaura reaction.¹⁰⁶ The latter compound was coupled to the key iodide **310** to produce the sp²–sp³ coupled **312** which contains the complete iridal triterpene skeleton. Synchronized acidic deprotection of all protecting groups and then PCC-mediated selective allylic oxidation provided the desired (+)-iridal **295** in 53% isolated yield. Throughout the synthesis, the chiral quaternary center at C-4a was evident in governing the diastereoselective synthesis.

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17.2 Isomarabarican-type triterpenes

17.2.1 Total syntheses of (\pm) -rhabdastrellic acid A and (\pm) -stelletins A and E. Sarlah and co-workers accomplished a 14-step total synthesis of the highly cytotoxic isomalabaricane triterpenoids *i.e.* (\pm) -stelletin E (196) and (\pm) -rhabdastrellic acid A (313) from commercially available geranylacetone (314) (Scheme 21).¹⁰⁷ Accordingly, the epoxynitrile 315 was prepared from 314 via a modified van Leusen reductive cyanation¹⁰⁸ followed by an NBS mediated selective epoxidation of the terminal double bond.¹⁰⁹ Next, bicyclic 316 was prepared via reductive radical polyene cyclization catalyzed by Cp₂TiCl₂ as the key step,¹¹⁰ which was followed by silvlation of the resulting C-3 alcohol to afford the target intermediate in a 5:1 epimeric mixture at C-8. The bicyclic ketone 316 was then converted to

the key alkyne precursor 317 by virtue of a two-step protocol including conversion of the ketone functionality to an α,β unsaturated aldehyde by employing dichloromethyllithium followed by lithium acetylide addition and an in situ pivaloate protection.

The α -fluoro hydrazone 318 was prepared via an efficient annulation and functionalization protocol by reacting 317 with a cationic gold(1) catalyst in the presence of Selectfluor to give the C-ring α -fluoro enone as a single diastereomer. Notably, in *p*-toluenesulfonyl hydrazone formation facilitates situ construction of the α -fluoro hydrazone **318**. The key *trans-syn*trans intermediate 319 was obtained from 318 first by treating the latter with triethylamine in methanol to promote a conjugate addition of the methanol solvent and produce an



Scheme 22 Synthesis of (\pm) -stelletin A (327).

BY-NC

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Synthesis of 13E,17E-globostellatic acid X methyl ester mimic 332 Scheme 23

azoalkene followed by stereospecific addition of hydrogen via a retro-ene rearrangement of the allylic diazene. Next, 319 was transformed into the enone 320 by employing reductive zirconation and copper-catalyzed cross-coupling with acetyl chloride as C-2 unit. The triketone skeleton 321 was synthesized by the relay hydroboration of the double bond of the ketone 320 followed by an *in situ* silyl group deprotection and IBX oxidation. The key tricyclic intermediate 322 was synthesized as a single isomer from ketone 321 by bromination using a Vilsmeier reagent. The next core skeleton to be synthesized was the polyene side chain 326. In order to prepare polyene side chain 326, initially, stannane dienal 324 was synthesized from 3picoline (323) according to a published procedure.¹¹¹

Compound 324 was coupled with phosphonate 325 (Scheme 21) under typical Horner-Wadsworth-Emmons olefination conditions to furnish the polyene coupling partner 326. Coupling between the tricyclic vinyl bromide 322 and 326 was achieved to produce the methyl ester of rhabdastrellic acid A which upon saponification produces the (\pm) -rhabdastrellic acid A (313). The geometric isomer stelletin E (196) was prepared upon irradiation of (\pm) -rhabdastrellic acid A (313).

In order to prepare stelletin A (327),¹¹² initially, gibepyrone C (330) was prepared for the pyrone side-chain in both target molecules (Scheme 22). Dimerization of tiglic acid (328)¹¹³ provided lactone 329 which was subsequently converted into gibepyrone C (330) through a Johnson-Lemieux oxidation to

afford a ketone. This is followed by ethoxy vinyl zinc addition and hydrolysis to afford the target aldehyde 330. This in turn was subjected to a Boron-Wittig reaction¹¹⁴ to furnish the pyrone coupling partner 331. Finally, Suzuki coupling of diketone 321 and the pyrone derivative 331 gave stelletin A (327). Unfortunately, stelletin A (327) could not been converted into stelletin A (209) via the photoisomerization protocol due to significant decomposition under UV light.112

17.2.2 Synthesis of BC-ring model of globostellatic acid X methyl esters. Kobayashi et al.115 prepared a biological mimic of 13E,17E-globostellatic acid X methyl ester 332 (Scheme 23) in a 10-step synthetic approach starting from the commercially available Hajos-Wiechert ketone (333). The four parent globostellatic acid X methyl esters have been reported from the sponge Rhabdastrella globostellata.¹¹⁶ Accordingly, ketone 333 was stereoselectively reduced to trans-hydrindanedione 334, employing DIBAL-H followed by conversion into the ethylene ketal 335 to protect the less-hindered ketone. Notably, a single E-configured isomer 336, was produced by the enolate olefination after treatment with ethyl 2, 3-dibromopropionate and iodomethane. Moreover, SeO2-mediated allylic oxidation of 336 furnished the racemic allylic alcohol 337 which was followed by ester reduction with DIBALH and oxidation of the resulting two hydroxyl groups which afforded the keto-aldehyde 338. Unfortunately, the precursor 338 could not be transformed into 339 via an attempted Takai olefination.



Scheme 24 Total synthesis of (+)-myrrhanol C (227).

An alternative route was therefore developed. Thus, reduction of the ester group in 336 *via* DIBAL-H provided methacrolein 340 which was further successfully converted into the vinyl iodide **341** *via* a Takai olefination. Notably, the keto group in ring C of the latter compound was generated by using $CrO_3/$ 3,5-dimethylpyrazole and this was followed by ketal



Scheme 25 Total synthesis of (+)-myrrhanols A (365), B (366), (+)-myrrhanones A (367), and (+)-B (368).

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deprotection to afford the coupling fragment **342**. The aldehyde **349** which was required for installation of the polyene side chain, was prepared in six steps from methylmalonate (**343**) (Scheme 23). Wittig reaction between aldehyde **349** and tributylphosphonium salt **350** established the side chain precursor **351**. Lastly, the desired BC-ring **332** was achieved *via* Stille cross-coupling between fragments **342** and **351**.¹¹⁶

17.3 Polypodanes

17.3.1 Total synthesis of (+)-myrrhanol C. Domingo *et al.*¹¹⁷ prepared (+)-myrrhanol C (227) for which the detailed synthesis is depicted in Scheme 24. In the course of this synthesis, the bicyclic diol fragment 352 was prepared using ex-chiral pool precursors including (-)-sclareol, (-)-sclareolide, or farne-sol.¹¹⁸⁻¹²⁰ The bicyclic diol fragment 352 was converted to the trihydroxy intermediate 353 *via* a biotransformation based C-H oxidation. The primary alcohol group in 353 was chemoselectively protected with the pivaloyl protecting group to provide diol 354 followed by the secondary hydroxyl group being protected as its TBS ether (355) and thirdly the tertiary hydroxyl group was protected as its MEM ether to the fully alcohol protected 356.

The advanced intermediate **359** was synthesized by *via* primary alcohol **357**, formed by reduction of the pivaloyl ester of **356** and Swern oxidation to the corresponding aldehyde **358**. This latter compound was subjected to a Wittig olefination employing methyltriphenylphosphonium bromide to afford the bicyclic fragment **359**. Vinyl halide **363** was prepared by coupling geranyl bromide **360** and **361** followed by Zirconium-catalyzed carboalumination of the acetylene group and iodination. Suzuki–Miyaura coupling of precursor **359** and iodide **363**

provided bicyclic 364 and lastly, acid hydrolysis furnished myrrhanol C (227).

17.3.2 Total synthesis of (+)-myrrhanols A, B and (+)-myrrhanones A, B. The triterpenes (+)-myrrhanols A (365), B (366), (+)-myrrhanones A (367), and (+)-B (368) were prepared by Barrero and co-workers (Scheme 25).121 The synthesis of intermediate 359 started from farnesyl acetate 369 which was converted into the bicyclic precursor 371 via a two step sequence including the Sharpless epoxidation protocol and an Ti(III)-mediated cyclization.122 Furthermore, the secondary hydroxyl group at C-3 was protected as its TBS ether to furnish 372 and then deprotection of the acetoxy group provided alcohol 373. Next, the tertiary hydroxyl group in 374 was produced as the only diastereoisomer through epoxidation of the exocyclic double bond with *m*CPBA followed by LiAlH₄ mediated reductive opening of the epoxide ring. Diol 374 was then converted into precursor 357 via a three step protocol including protection of the primary alcohol group as its acetyl, the tertiary hydroxyl group as its MEM ether and deacetylation.

Precursor 357 was transformed to the bicyclic synthon 359 by Swern oxidation of the primary alcohol followed by a Wittig olefination of with resulting aldehyde. Preparation of the coupling partner, vinyl iodide 379, was accomplished using geranyl acetate and employing the same synthetic protocol described for iodide 363. The key intermediates, bicyclic synthon 359 and vinyl iodide 379 were coupled using the Suzuki-Miyaura protocol to produce 380 which was finally converted to the target compound, (+)-myrrhanol A (365) upon deprotection of the three protecting groups. Following on to this, (+)-myrrhanol A (365) was converted to (+)-myrrhanone A (367) employing a Dess-Martin oxidation of the C-3 hydroxyl group. The (+)-myrrhanone A (367) could be converted to



Scheme 26 Synthesis of tricyclic core 392.

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(+)-myrrhanone B (**368**) upon further Dess–Martin oxidation to produce firstly the corresponding aldehyde and after further oxidation, the corresponding carboxylic acid. Formation of the final target (+)-myrrhanol B (**366**) was achieved by chemo- and stereoselective reduction of (+)-myrrhanone B (**368**) with NaBH₄.¹²¹

17.4 Schiglautanes

17.4.1 Total synthesis of an atropisomer of schiglautone A. The structural features of this natural product are quite unique and it possesses a cyclohexyl-fused bicyclo-[6.4.1]tridecane skeleton, including six stereogenic centers, three of them being quaternary. This natural product appears as an atropisomer and the diastereoselective synthesis was achieved by Ding and coworkers. Preparation of the tricyclic enone 392 (Scheme 26),¹²³ which carries the entire cyclic carbon skeleton, commenced with the regioselective epoxidation of trans-nerolidol (381) followed by acetylation to furnish epoxypolyprene 382. Furthermore, titanium(m)-catalyzed radical cyclization¹²⁴ of 382 afforded the desired trans-fused bicyclic precursor 383. mCPBA mediated epoxidation of ring double bond of 383 followed by inversion of the C-3 hydroxyl group through oxidation (Ley-Griffith oxidation) and reduction (Meerwein-Ponndorf-Verley reduction) protocols, and benzylic protection of the resulting hydroxyl group generated epoxide 385.

The lactone **387** was constructed from **385** by employing the Gansäuer protocol,¹²⁵ which upon reduction, Swern oxidation, subsequent chemoselective Grignard addition and silylation, provided bicyclic ketone **388**. The O-allylated **389** was generated after treatment of **388** with LiHMDS/allyl bromide and this was followed by a Claisen rearrangement to construct diene **390**.

This rearrangement afforded the C-1 bridging unit. The tricyclic carbon skeleton **391** was prepared from diene **390** *via* a Grubbs II catalyst based ring closing metathesis, yielding a *cis*-configured double bond. The correctly folded and substituted **392** was generated from ketone **391** through a Jones oxidation along with an addition of methyl lithium and a formal allylic ([3,3]-rearrangement) oxidation *via* the Dauben–Michno protocol.

Michael addition of the CuBr-modified Grignard reagent 393 to 392 (Scheme 27) followed by a subsequent Rubottom oxidation with DMDO resulted in formation of the hydroxylated and epoxidized 394. Treatment of the epoxide of 394 with the strong oxidant PCC afforded the 'diol'-cleaved aldehyde 395. Horner-Wadsworth-Emmons reaction of the latter aldehyde with 396 provided the Z-configured 397 selectively upon addition of crown ethers. This triketone was first treated with methyl acrylate to give the correct side-chain moiety and then with DMDO to afford the ene-dione 398 in a one-pot procedure. After cleavage of the C-3 benzyl group, reduction of the C-12 ketone was achieved via Luche conditions and delivered the major diastereomer 399 along with hemiketal 400 and aldehyde 401. Lastly, desilylation of 399 with TBAF afforded diastereoselectively 402. Surprisingly, NMR data of 402 did not accurately resemble that of natural schiglautone A (239)82 and based on NMR and X-ray data the structure of the synthesized compound 402 was established as a diastereomeric atropisomer of 239.

17.4.2 Synthesis of an advanced intermediate of schiglautone A. Chapelain *et al.*¹²⁶ prepared the advanced intermediate 403 *via* a clever convergent enantioselective route and this synthesis is presented in Scheme 28. Thus α -methylation of ketone 404 followed by hydroxylation furnished keto-alcohol



405. The keto group in the latter molecule was reduced with NaBH₄, and the resulting diol underwent Pb(OAc)₄ mediated oxidative cleavage to furnish keto-aldehvde 406. The aldehvde 406 was transformed into alkyne 407 through the Seyferth-Gilbert protocol127 and this latter compound was next subjected to a Baeyer-Villiger oxidation which delivered acetate 408. The precursor 408 was elaborated to aldehyde 409 through a two step protocol including cleavage of the acetate group and oxidation of the resulting primary hydroxyl group into an aldehvde.

The preparation of ketone 419 commenced from (-)-citronellene (410), which was initially transformed to aldehyde 411 in two steps¹²⁸ (Scheme 29). The latter molecule was subjected to a Grignard addition (derived from 5-bromo-1-pentene) to



Scheme 29 Synthesis of fragment 417.

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afford alcohol 412 and this step was followed by oxidation to the ketone and then followed by a second Grignard addition to furnish triene 413. Grubbs I catalyst based ring-closing metathesis between the terminal primary alkene groups delivered allylic alcohol 414 and oxidative transposition provided enone 415. Luche reduction of enone 415 provided allylic racemic alcohol 416. This mixture was subjected to enzymatic resolution to afford allylic alcohol 417.

The precursor 417 was subjected to chemoselective cyclopropanation to furnish alcohol 418 (Scheme 30) and PCC mediated oxidation to the ketone along with Birch reduction mediated cyclopropane ring opening provided ketone 419. Aldol addition between aldehyde 409 and ketone 419 provided enone 420 which was elaborated into ketone 421 through conjugate reduction with LAH/CuI. The thermodynamically stable silvl enol ether of 421 was constructed and subjected to Furukawa's cyclopropanation reagent¹²⁹ to provide the desired product 422 together with side products 423-425. Lastly, the desired precursor 422 was transformed into ketone 403 through cyclopropane ring opening with NaOH/EtOH.

17.5 Unusual tricyclic seco/nor-lanostane

17.5.1 Studies towards the total synthesis of kadcotrione B. The synthesis of kadcotrione B (256) started from construction of the bicyclic precursor 434 depicted in Scheme 31.130 Michael addition of diketone 426 and ethyl vinyl ketone 427 afforded triketone 428 which was subsequently elaborated organocatalytically into trans-decalone 429 via the Hagiwara protocol.131 Regioselective ketal protection of ketone 429 as its ketal 430 was followed by reductive methylation incorporating Birch reduction to furnish the α, α -dimethyl trans-decalone 431. LiAlH₄ mediated C-3 ketone reduction furnished the sec-alcohol 432. The resulting hydroxyl group was protected as its benzyl ether to afford 433. The ketal group was cleaved in 433 via acid hydrolysis to ketone 434. In order to construct 6/6/5-tricyclic ketone 437, the bicyclic ketone 434 was then subjected to a four step procedure developed by Smith and coworkers132 which provided diketone 435 and was subsequently elaborated into tricyclic ketone 436 through NaH mediated internal aldol condensation. Notably, insertion of the methyl group in the 5membered ring in 436 was achieved at the α -position to the keto group via treatment with LiHMDS and MeI to afford the tricyclic core 437.

The synthesis of aliphatic fragment 448, as depicted in Scheme 32, commenced with 1,5-pentanediol (438) via the protected aldehyde 439 by following the Tsuji protocol.133 This latter compound was elaborated into the Z-methacrylate derivative 441 with the Still-Gennari phosphonate 440 under the Horner-Wadsworth-Emmons reaction protocol. The ester group in 441 was reduced to the primary alcohol via DIBAL-H to afford alcohol 442 which was followed by protection of the resulting hydroxyl group as its benzyl ether and then cleavage of the TBS group afforded 443. Furthermore, the primary alcohol group in the latter molecule was transformed into the corresponding carboxylic acid through two oxidation procedures viz., Swern and Pinnick oxidations to afford 444.134





Precursor **444** was next transformed into amide **446** *via* a coupling reaction with (*R*)-4-benzyloxazolidin-2-one (**445**) and diastereoselective methylation of **446** was achieved by following the Evans protocol¹³⁵ to deliver molecule **447**. The amide **447** was converted into the main fragment **448** through a three step procedure including chiral auxiliary cleavage, transformation into a Weinreb amide and treatment with vinylmagnesium bromide. With the key fragments **448** and **437** in hand, initial attempts to combine them through Michael addition and further elaboration into Kadcotrione B (**256**) *via* a two-step sequence including the deprotection of the benzyl group, and

oxidation of primary and secondary alcohols in a single step were investigated but proved inadequate to deliver the desired **449**. The various reaction conditions that were investigated (NaOEt, *t*-BuOK, LDA, KHMDS, NaHMDS, LiHMDS) for the crucial Michael addition between **437** and **448** resulted in the decomposition of starting materials.

17.6 Abudinols

17.6.1 Total synthesis *ent*-abudinol B. McDonald and coworkers¹³⁶ reported on the synthesis of an enantiomer of the marine natural product abudinol B described as *ent*-abudinol



Scheme 32 Synthesis of fragment 448.



Scheme 33 Total syntheses ent-abudinol B (ent-460).

(*ent*-460) (Scheme 33). In the course of their synthesis, the precursor, farnesyl *p*-nitrobenzoate 450, was transformed into diepoxide 451 *via* regio- and enantioselective epoxidation followed by elaboration into allylic bromide 452 *via* a standard series of reactions including ester saponification, mesylation, and bromide substitution. The other key component *i.e.* alphasilyl ketone 455 was prepared by alkylation of geranyl bromide

(454) with the metalloenamine derived from 453 followed by imine hydrolysis.

Furthermore, ketone **455** was elaborated into diepoxyenolsilane **456** through two steps including addition of vinylmagnesium chloride to the acylsilane **455** and Brook rearrangement. The TMSOTf mediated regio- and stereoselective tandem tricyclization of **456** provided the *trans-anti-trans*-fused



tricyclic core **457** which upon treatment with the classical Wittig reagent, transformed it into the disubstituted alkene **458**. Diepoxide *ent*-**459** α was subsequently prepared from alkene **458** through regio- and stereoselective double epoxidation. Lastly, TMSOTf catalyzed cyclization of diepoxide *ent*-**459** α produce *ent*-abudinol B (*ent*-**460**) in low yield (15%).¹³⁶

17.7 Muzitones

17.7.1 Total synthesis of muzitone and *ent*-muzitone. McDonald and co-workers¹³⁶ presented (Scheme 34) an elegant research investigation to reassess the structure of muzitone that had previously been incorrectly assigned.^{137,138} The synthesis commenced with the preparation of diepoxide **461**, which was constructed by reaction between allylic bromide **452** and imine **453**. The diepoxide **461** was further elaborated into the precursor **462** by treatment with vinylmagnesium bromide and geranyl bromide (**454**) through a Brook rearrangement. Moreover, TMSOTf mediated cyclization of **462** produced the tricyclic ketone **463** having a free tertiary hydroxyl group which upon treatment with *p*-nitrobenzoyl chloride, afforded the corresponding *p*-nitrobenzoate ester **464**. After removal of the *p*nitrobenzoate group, installation of the 30th carbon was achieved through a classic Wittig reaction to produce the triene **465**. Following on to this, the triene **465** was converted into diepoxide **466** by employing the Shi epoxidation protocol and this was followed by TMSOTf mediated cyclization to afford the pentacyclic core **467**.

Protection of the two free hydroxyl groups as their acetates in the latter compound and migration of the double bond from the C-13/C-14 into the C-14/C-25 position was accomplished after treatment with HI to afford isomer **469**. The compound *ent*-**470**, which is considered as "pre muzitone" was achieved by deacetylation of **469** employing K_2CO_3 in methanol. Finally, *ent*muzitone (*ent*-**472**) was constructed *via* the oxidative cleavage of the tetrasubstituted alkene in *ent*-**470** with O_3/Me_2S or indirectly by from **469** by producing diol **471** initially through osmylation followed by reduction and then deacetylation along with Pb(OAc)₄ mediated oxidative cleavage which produces the same desired *ent*-muzitone (*ent*-472). In addition, X-ray analysis of bis-*p*-nitrobenzoate analog *ent*-472 established the *ent*-472 is truly *ent*-muzitone. Unfortunately, Boone *et al.*¹³⁶ observed some discrepancies in their NMR data for *ent*-472 and the original natural muzitone.¹³⁷ Based on this synthetic investigation, the structure of muzitone assigned by Kashman group is consequently incorrect.

18 Conclusions

This review provides the second comprehensive overview on some intriguing unusual cyclized triterpenes. Marine organisms, in particular sponges, produce unusual cyclized triterpenoids with remarkable chemical diversity. Advanced scientific studies into these unusually cyclized triterpenes establishes a sound strategy in the importance of these molecules. These triterpenoids have shown a broad spectrum of biological effects including anti-epileptic, cytotoxic, hepatoprotective, neuroprotective, anti-inflammatory, and antidiabetic effects.

The majority of these unusual triterpenes include malabaricane, isomalabaricane, and iridal-type triterpenoids. Overall, isomalabaricanes were reported from four genera of sponges including *Rhabdastrella*, *Jaspis*, *Geodia*, and *Stelletta*. However, taxonomic identification of these sponges remains questionable.^{139,140} On the other hand, malabaricane triterpenoids were present in higher plants, fungi, ferns, and bottom sediments.¹⁴⁰ Moreover, iridals were found in the Iridaceae family, mainly in the genus *Iris*. However only a few were reported from the genera *Tigridia* and *Belamcanda*. Furthermore, the unusual and quite fascinating structures have challenged the esteemed synthetic chemists to construct these demanding molecules with the correct absolute configuration and which has required them establishing new synthetic methodologies.

There is no doubt that nature has established fascinating pathways to construct natural products which has unfortunately resulted in less effort by scientists being focused on investigating biosynthetic pathways of unusual cyclized triterpenoids and also understanding the structures along with mechanisms of SC and OSC enzymes which generate these molecules. On the positive side, Xiong et al.³⁰ investigated how oxidosqualene is converted into the monocyclic iridal skeleton. In addition, two Arabidopsis thaliana genes encode OSCs which furnish the malabaricane skeleton^{8,66-69} Lodeiro et al.65</sup> demonstrated that the isomalabaricane core arises from a lanosterol cyclase mutant (Tyr510) while the native enzyme has yet to be discovered. Moreover, Kushiro and his team96,97 investigated the biosynthesis of the α -onocerin skeleton and successfully identified the two genes from the fern Lycopodium clavatum, LCC and LCD of the oxidosqualene cyclase (OSC) family.

Thus, more intensive investigations on cyclizing enzymes which generate these triterpenes, will be an intriguing and challenging research field for future natural product research. In addition, there are still numerous fundamental questions about enzyme mechanisms which still need to be fully investigated. Notably, the relationship between active-site structure and cyclization mechanism of the numerous SC and OSC enzymes presents a formidable task. Further studies are required for the enzyme proteins structure-based engineering along with manipulating substrates. Finally, future research needs to focus on the genomics and synthetic biology for identifying and diversifying these unusual triterpenoid molecules because we believe that this will facilitate remarkable progress in finding new lead compounds for potential biomedical applications.

19 Author contributions

H. H. and B. W.: conceptualized, supervised, and edited the review process. J. X. and A. A. wrote the manuscript and drew the figures/schemes. I. R. G. wrote and edited the manuscript.

20 Conflicts of interest

There are no conflicts to declare.

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