



Cite this: *Nanoscale Adv.*, 2023, **5**, 6017

Research progress related to thermosensitive hydrogel dressings in wound healing: a review

Ruting Gu,^a Haiqing Zhou,^a Zirui Zhang,^b Yun Lv,^c Yueshuai Pan,^d Qianqian Li,^e Changfang Shi,^f Yanhui Wang^{*g} and Lili Wei^{*h}

Wound healing is a dynamic and complex process in which the microenvironment at the wound site plays an important role. As a common material for wound healing, dressings accelerate wound healing and prevent external wound infections. Hydrogels have become a hot topic in wound-dressing research because of their high water content, good biocompatibility, and adjustable physical and chemical properties. Intelligent hydrogel dressings have attracted considerable attention because of their excellent environmental responsiveness. As smart polymer hydrogels, thermosensitive hydrogels can respond to small temperature changes in the environment, and their special properties make them superior to other hydrogels. This review mainly focuses on the research progress in thermosensitive intelligent hydrogel dressings for wound healing. Polymers suitable for hydrogel formation and the appropriate molecular design of the hydrogel network to achieve thermosensitive hydrogel properties are discussed, followed by the application of thermosensitive hydrogels as wound dressings. We also discuss the future perspectives of thermosensitive hydrogels as wound dressings and provide systematic theoretical support for wound healing.

Received 11th June 2023
Accepted 27th July 2023

DOI: 10.1039/d3na00407d
rsc.li/nanoscale-advances

Introduction

The skin is the first line of defense that protects the human body from external bacteria and viruses, and its integrity is extremely important for human safety.¹ When the integrity of the skin is compromised, patients experience discomfort, including pain, bleeding, inflammation, delayed healing, scarring, and even death.^{2,3} Wound healing is the basic stage of skin injury reconstruction. It is a dynamic and overlapping process, usually divided into four continuous and possibly overlapping processes: hemostasis, inflammation, proliferation, and remodeling (Fig. 1).^{4,5} These processes are regulated by complex networks comprising various cells and biological mediators.^{6,7}

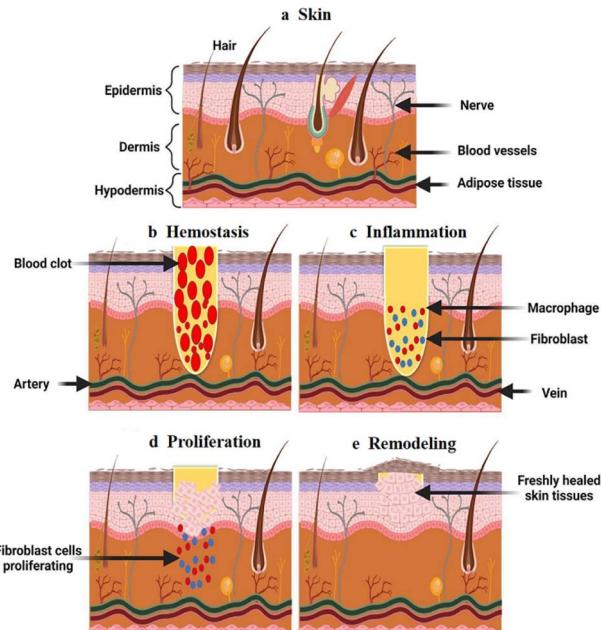


Fig. 1 Schematic diagram of the different wound healing phases: (a) normal skin; (b) hemostasis involves the promotion of platelets and formation of clots; (c) inflammation recruits leukocytes to the wound site to release cytokines; (d) the proliferation phase involves angiogenesis and extracellular matrix production; (e) the remodeling phase involves the organization and contraction of the extracellular matrix. This figure has been reproduced from ref. 4 with permission from Wiley Interdiscip. Rev.: Nanomed. Nanobiotechnol., copyright 2022.

^aDepartment of Thoracic Surgery, The Affiliated Hospital of Qingdao University, Qingdao 266000, China. E-mail: 13805424386@163.com

^bEmergency Departments, The Affiliated Hospital of Qingdao University, Qingdao 266000, China

^cSchool of Nursing, Qingdao University, Qingdao 266000, China

^dDepartment of Critical Care Medicine, The Affiliated Hospital of Qingdao University, Qingdao 266000, China

^eOphthalmology Department, The Affiliated Hospital of Qingdao University, Qingdao 266000, China

^fDepartment of Spinal Surgery, The Affiliated Hospital of Qingdao University, Qingdao 266000, China

^gDepartment of Oral Implantology, The Affiliated Hospital of Qingdao University, Qingdao 266000, China. E-mail: qdwyh06@163.com

^hOffice of the Dean, The Affiliated Hospital of Qingdao University, Qingdao, 266000, China. E-mail: weilili@qduhospital.cn



Studies have confirmed that temperature, pH, and other factors affect wound healing.^{8–12}

Wound dressings protect wounds from infection by absorbing osmotic fluid and isolating the external environment.^{13,14} Traditional dressings, such as gauze, cotton wool, and bandages can stop bleeding, absorb wound exudate, and protect the wound from bacterial infection; however, they do not have antibacterial or anti-inflammatory functions^{15,16} and may cause adhesion scabs and even hinder the wound healing process.^{17,18} Compared to traditional medical dressings, hydrogels have the advantages of a three-dimensional network structure, strong water absorption, and the maintenance of a moist environment in the wound to reduce the infection rate and accelerate wound healing;¹⁹ as such, they have become a research hotspot in the field of wound dressing. More importantly, hydrogel dressings can be endowed with excellent properties through structural design and functional integration,²⁰ thus playing an important role in the wound healing process.

Thermosensitive hydrogels are smart polymer hydrogels that undergo phase transitions under external temperature stimulation and have good biocompatibility and mechanical properties.²¹ The gelation of thermosensitive hydrogels is triggered by temperature, and they exhibit a phase transition at the critical solution temperature. Once the desired temperature is reached, known as the sol–gel transition, hydrogels form immediately.²² The temperature-induced sol–gel transition is safer *in vivo* and is more suitable for injection systems because it does not require denaturing crosslinkers,²³ leading to an easier and more economical process.^{24,25} They have excellent temperature sensitivity, drug delivery,^{26–28} cell loading,^{29,30} protein delivery,^{31,32} and gene therapy.^{33,34} As a free-flowing liquid before low-temperature gelation, the hydrogel precursor solution can be injected into the wound area to quickly fill irregular defects under physiological conditions, form a gel, and fill any irregular wound site or deep wound.³⁵ In addition, encapsulation in the flow state allows the therapeutic agent to be evenly dispersed in the hydrogel, whereas the rapid sol–gel transition at body temperature prevents the initial burst release of the therapeutic agent, thus controlling the release behavior.³⁶ These features are key to their application in human medicine,³⁷ particularly in the field of wound dressing. However, a systematic review of thermosensitive hydrogel wound dressings has not been conducted.

Herein, we summarize the progress in thermosensitive hydrogel wound dressings and discuss the polymers used in these dressings. The mechanisms of temperature sensitivity are reviewed and we discuss the different types of thermosensitive hydrogels and their applications in different wounds. Finally, we look ahead to the future development of thermosensitive hydrogels for wound-healing applications and make recommendations.

Common thermosensitive materials

An ideal wound dressing needs to be biocompatible and non-sensitive, with appropriate adhesion and mechanical properties, able to effectively cover the entire irregular wound, absorb

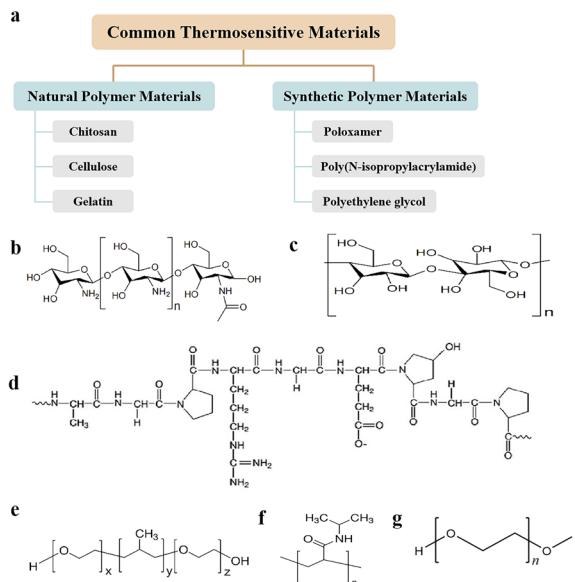


Fig. 2 (a) Common classification of thermosensitive materials; the structural formula of (b) chitosan; (c) cellulose; (d) gelatin; (e) poloxamer; (f) poly(*N*-isopropylacrylamide); (g) polyethylene glycol.

the exudate tissue fluid in time, ensure the environmental humidity of the wound, allow gas exchange, and prevent bacterial infection of the wound, so that it can be easily replaced to alleviate the pain of patients in the process of wound management.^{38,39}

Thermosensitive biomaterials are novel smart materials for preparing thermosensitive hydrogels. These sources can be divided into natural and synthetic polymer materials (Fig. 2a).⁴⁰

Natural bioactive polymers

Natural bioactive polymers are often used as wound dressings due to their biocompatibility and biodegradability.^{41,42} The most commonly used natural bioactive polymers for thermosensitive hydrogels are chitosan,^{43,44} cellulose^{45,46} and gelatin.^{47,48}

Chitosan (CS) is a polycation biopolymer that has a wide range of sources, and its structure consists of an *N*-acetylglucosamine group linked by a β -1,4-glycosidic bond and glucosamine residues (Fig. 2b).⁴⁹ CS exhibits good histocompatibility, biodegradability, adhesiveness, antimicrobial activity, and low cytotoxicity.^{50–53} Because of these valuable features, CS has been widely used in the fields of medicine and biology.^{52,54} However, because of the slow temperature response and low mechanical strength of chitosan, it usually needs to be improved by combining it with other substances.³⁶ Common chitosan thermosensitive hydrogels include chitosan–sodium glycerophosphate thermosensitive gels, chitosan graft gels, sulphydryl chitosan hydrogels, and chitosan/collagen gels.⁵⁵ Previous studies have shown that CS activates platelets and macrophages, resulting in excellent blood characteristics.⁵⁶ It can also promote the release of vascular endothelial growth factors, thereby promoting the formation of blood vessels and accelerating repair.⁵⁷ In addition, chitosan exhibits excellent antibacterial properties that can reduce the probability of wound



infection and accelerate wound healing.⁵⁸ The CS structure is easy to modify, easy to process, and can be made into a hydrogel. The CS hydrogel dressing has the characteristics of high repair efficiency, simple design and preparation, and has potential value in the field of wound dressing.⁵⁴

Cellulose, which has been used for a long time and has a long history, is the main material used in traditional gauzes. Cellulose derivatives are biological macromolecules containing glucose molecules connected by β -1,4-glycosidic bonds (Fig. 2c). Hydroxypropyl cellulose and methyl cellulose are representative polymers used in cellulose-based thermosensitive hydrogels. Cellulose-based thermosensitive hydrogels are widely used as biomedical materials because of their hydrophilicity, biocompatibility, biodegradability, and nontoxicity.²¹ At the structural level, cellulose is an ideal material for synthesizing biocompatible hydrogels because it easily modifies functional groups.^{59–61} The high biocompatibility and functional structure of cellulose polymer allows cellulose nanocomposites to be combined with various antibacterial agents.^{62,63} Oscar *et al.* developed a multifaceted formulation based on CNT-containing cellulose–chalcone, which can sustain the release of therapeutic substances with antimicrobial and wound-healing properties.⁶⁴ Hydrogels can potentially be utilized to treat complex wounds owing to their improved wound-healing properties and prevention of potential infections.

Gelatin is a natural protein derived from the hydrolysis of collagen; its molecular chain contains a large number of active functional groups (Fig. 2d). Gelatin is highly biocompatible and biodegradable in a physiological environment.^{47,65} Although mainly derived from animals, the digestive process confers gelatin very low antigenicity, with the formation of harmless metabolic products upon degradation while in body.^{66,67} When the temperature is below 25 °C, an aqueous gelatin solution solidifies due to the formation of triple helices and a rigid three-dimensional network.⁶⁸ At the physiological temperature, the physically formed hydrogels are dissolved.⁶⁹ Therefore, gelatin is often chemically crosslinked or combined with other polymers resulting in hydrogels that are stable at 37 °C. Studies have reported thermoresponsive hydrogel systems for drug delivery by combining gelatin with a chitosan/Pluronic mixture^{70,71} to promote wound healing.

Synthetic polymers

Synthetic materials include poloxamers,^{72,73} poly(*N*-isopropylacrylamides)^{74,75} and polyethylene glycol.^{76,77} Poloxamer is a synthetic amphiphilic copolymer^{78,79} consisting of a poly(ethylene oxide)–poly(propylene oxide)–poly(ethylene oxide) (PEO–PPO–PEO) triblock copolymer (Fig. 2e),⁸⁰ and the number of molecules can be adjusted to affect their hydrophilicity or hydrophobicity. Poloxamer, copolymers with good amphiphilicity, double responsiveness, and biocompatibility, have important applications in the field of synthetic dressings.^{81,82} Poloxamer hydrogel dressings have good elasticity and plasticity and can closely fit wounds to isolate bacteria. Their transparent texture can also be used to observe the condition of the wound while simultaneously keeping the wound moist, which is

especially suitable for dry wounds, such as burns.^{83,84} Poloxamer hydrogels have good dual responsiveness (pH and temperature) and biocompatibility and can bind to the phospholipid bilayer of cells to protect the cell membrane and promote the expression of growth factors and the proliferation of fibroblasts, thereby accelerating wound healing.^{85–87}

Poly(*N*-isopropylacrylamide) (PNIPAM) is a temperature-sensitive polymer polymerized from the monomer *N*-isopropyl acrylamide (NIPAM) *via* free-radical solution polymerization.⁸⁸ The molecular chain of PNIPAM contains both hydrophobic isopropyl ($-\text{CH}(\text{CH}_3)_2$) groups and hydrophilic amide ($-\text{CONH}-$) groups in its structure (Fig. 2f).⁸⁹ Polymer molecules form hydrogen bonds with each other because of hydrogen bonding with water molecules. PNIPAM dissolves in water to form a solution at room temperature and precipitates after increasing the temperature.^{90,91} This property makes PNIPAM suitable for biomedical applications, such as controlled wound dressings, drug delivery systems, and engineering scaffolds.^{92–94} Jiang *et al.* developed a new hydrogel, acryloyl-lysine (A-Lys), which was incorporated into poly(ethylene glycol)-crosslinked PNIPAM.⁹⁵ It exhibits good biological activity while maintaining its swelling properties and thermal reaction behavior. Polyhexamethylene biguanide, an antimicrobial agent, was loaded onto hydrogels. These materials can promote wound healing and provide active antimicrobial agents that inhibit infections.

Poly(ethylene glycol) (PEG)-based polymers are water-swellable hydrophilic cross-linked polymers with high elasticity (Fig. 2g). Their hydrophilic and nonionic characteristics make them highly biocompatible and relatively resistant to protein adsorption;⁹⁶ thus, they are suitable for use in medical products, such as drug delivery and tissue engineering. The copolymerization of biocompatible polyesters and PEG forms hydrogel systems. By appropriately adjusting the lengths of the hydrophobic polyester and PEG blocks, a gel system with thermally responsive properties can be obtained, and its thermally responsive properties can be improved. Poly((lactic acid)-co-glycolic acid) and poly(ethylene glycol) (PLGA–PEG–PLGA) have the features of biocompatibility, thermal sensitivity and non-toxicity. The temperature sensitivity of the copolymer was controlled by hydrophobic (PLGA) and hydrophilic (PEG) groups in the polymer. Chen *et al.* prepared flexible synthetic hydrogel sealants using PEG. This study showed that PEG-based hydrogels have good biocompatibility and safety.⁹⁷ The effectiveness and convenience of using PEG-based hydrogels for wound closure and bleeding control in animal studies are discussed.

Mechanism of thermosensitive hydrogels

Thermosensitive hydrogels are water-soluble polymer systems mainly composed of a macromolecular backbone with hydrophilic and hydrophobic groups. They can change hydrophilic and hydrophobic properties and the size and volume of the gel network with a change in ambient temperature and undergo sol–gel or gel–sol phase transition.⁹⁸ At the molecular level, the

temperature sensitivity of thermosensitive hydrogels is due to hydrogen bonding between hydrophilic groups and water molecules and hydrophobic interactions between hydrophobic groups in the structure.⁹⁹ With a change in temperature, the hydrophilic and hydrophobic interactions of the hydrogel are affected, leading to a change in the internal network structure and overall volume of the gel.

Thermosensitive hydrogels are classified as thermal shrinkage- and thermal expansion-type thermosensitive hydrogels according to their swelling mechanism.¹⁰⁰ A heat-shrinkable thermosensitive hydrogel is defined as being in a state of contraction when the temperature is higher than the phase transition temperature, and expansion when the temperature is lower than the phase transition temperature. The opposite was observed for the thermally expanding thermosensitive hydrogel: it shrank at temperatures below the phase transition temperature and expanded at temperatures above the phase transition temperature. The corresponding critical temperature is called Volume Phase Transition Temperature (VPTT). These features are key to applications in drug delivery, cell culture, tissue engineering, protein delivery, gene therapy, wound healing *etc.*³⁶

The LCST represents the low critical solution temperature during phase separation.¹⁰¹ When the temperature is lower than the LCST, the molecular chain is mainly formed by hydrogen bonding between the hydrophilic group and the water molecule. The molecular chain stretches and opens, allowing the gel to fully absorb water and swell into a liquid shape. When heated above the LCST, intermolecular hydrogen bond interactions were destroyed, and hydrophobic interactions became dominant (Fig. 3a). At this time, the molecular chain contracted, and the gel was dehydrated and contracted. Around the upper critical solution temperature (UCST), thermally responsive polymers undergo phase transitions. When the environment is below the UCST, the material does not dissolve. When heated above the UCST, the material dissolves (Fig. 3b). The swelling and contraction of the thermosensitive hydrogel are shown in Fig. 3c.

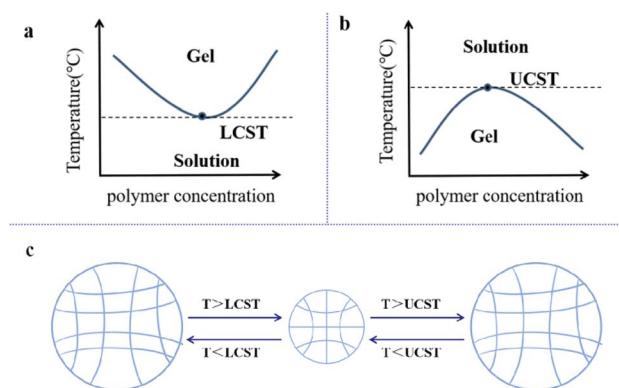


Fig. 3 (a) The LCST-type formulation undergoes sol–gel transition with an increase in temperature. (b) UCST-type formulation undergoes sol–gel transition as the temperature decreases. These figures have been reproduced from ref. 21 with permission from MDPI, copyright 2022. (c) Schematic of the swelling/contraction of the temperature-sensitive hydrogel.

Various types of thermosensitive hydrogel dressings

Thermosensitive hydrogels are of great interest in the biomedical field owing to their biodegradability, non-toxicity, inherent biocompatibility, wide availability, and embracing functionality. The excellent temperature sensitivity and injection characteristics of thermosensitive hydrogels make them suitable for drug, cell, protein, or factor delivery,^{102–104} and they can be used to fill any irregular wound site,¹⁰⁵ showing their characteristics and advantages for wound healing (Fig. 4). A summary of the applications, name of dressing composition, main features and other relevant information about thermosensitive hydrogel dressings is presented in Table 1.

Antibacterial dressing

Infection is a common complication of wounds, and its prevention is very important in wound repair. If the wound is infected, it will cause wound suppuration, prolong healing time, and even cause sepsis in severe cases, endangering life and health. Therefore, important requirements have been proposed for the antibacterial properties of wound dressings. Hydrogels with antibacterial activity, such as chitosan, contain antibacterial structures in their natural polymers and related derivatives. Ma *et al.* developed a novel *in situ* injectable hydroxypropyl chitin/TA/ferric ion (HPCH/TA/Fe) composite hydrogel.¹⁰⁶ The incorporation of TA provided hydrogen bonding and hydrophobic forces between the TA and HPCH, which improved the mechanical strength of the HPCH hydrogel. When the temperature is below the temperature of gelation (T_{gel}), the pre-cooled hydrogel precursor solution can be injected into an irregular wound area and the solution rapidly gels at a physiological temperature. Also, the TA in hydrogels can be used as a crosslinker to improve the mechanical properties of hydrogels. In addition, it can be used as an antibacterial agent and is released continuously for up to seven days in acidic environments (Fig. 5a). Similarly, a hybrid hydrogel was

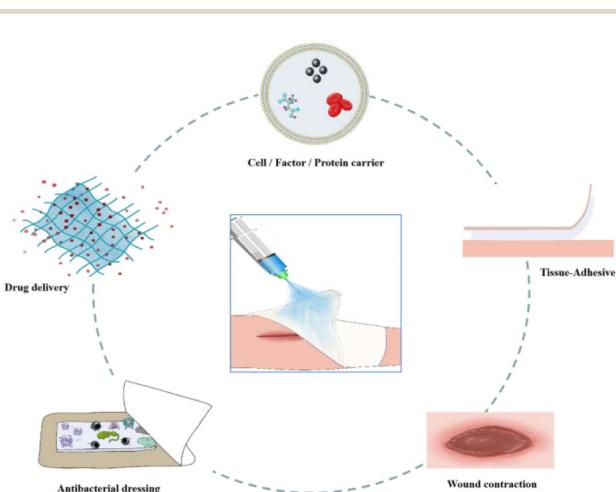


Fig. 4 The main advanced functions of thermosensitive hydrogel wound dressings.



Table 1 Summary of thermosensitive hydrogel dressings

Application	Name of dressing	Composition	Main features	$T_{\text{sol-gel}}$	Ref.	
Antibacterial dressing	HPCH/TA/Fe hydrogel	Chitin/tannic acid/ferric ion (HPCH/TA/Fe)	Thermosensitive, injectable, pH-sensitivity and good antibacterial properties	37 °C	106	
	PLELnBG–QCS-C hydrogel	The catechol-modified quaternized chitosan (QCS-C), poly(D,L-lactide)-poly(ethylene glycol)-poly(D,L-lactide) (PLEL), nano-scaled bioactive glass (nBG)	Thermosensitive, injectable and antibacterial properties	32.6 °C	107	
	CS hybrid hydrogels	Chitosan (CS) and poly(D,L-lactide)-poly(ethylene glycol)-poly(D,L-lactide) (PPP)	Thermosensitive, applied conveniently, good fluidity, hemostatic and antibacterial barrier	37 °C	108	
Drug delivery	HA–POL hydrogel	Hyaluronic acid (HA) – poloxamer 407(HA–POL)	Thermosensitive, bio-degradable, bacterial invasion, greater air permeability and effective moisturizing	30.0 °C	109	
	IB–Hep–PNIPAM hydrogel	Poly(N-isopropylacrylamide), <i>N</i> -acetylated heparin	Load and release of ibuprofen	37 °C	110	
	Curcumin–Alg–pNIPAM hydrogel	Poly(N-isopropylacrylamide), sodium alginate (Alg-g-pNIPAM)	Thermosensitive and injectable	27–42 °C	111	
	Cur-M-H	Curcumin (Cur), poly(ethylene glycol), PEG–PCL copolymer	Delivery of curcumin	Thermosensitive, injectable, anti-oxidant and anti-inflammatory properties	112	
	Dihydromyricetin-loaded hydrogel	Poloxamer, chitosan, hyaluronic acid	Load and release of Cur	37 °C	113	
	AuNPs hydrogel	Poloxamer 407, gold nanoparticles (AuNPs), polyethylene glycol (PEG), poly allyl amine hydrochloride (PAH)	Thermosensitive, good biocompatibility, non-toxic, antioxidant and anti-inflammatory	~35.5 °C	114	
	MSC&FMZC hydrogel	F127 hydrogel, MgO : ZnO/ chitosan hybrid particles (FMZC)	Load and release of AuNPs	3D mesenchymal stem cells (MSCs) delivery vehicle	37 °C	115
	Ni ₄ Cu ₂ /F127 hydrogel	F127 hydrogel, Ni ₄ Cu ₂ hollow nanospheres	Thermosensitive, suitable, nontoxic, higher antibacterial and antibiofilm activity	Load and release of therapeutic nanozymes	24 °C	116
	CP-nCur/gelatin hydrogel	Chitosan-P123 (CP), gelatin	Thermosensitive, sprayable, nontoxic, phase-change, porous, anti-inflammatory properties, angiogenesis	Thermosensitive, injectable, biocompatible, biodegradable	36.7 °C	70
	CH/GP/Cur hydrogel	Chitosan/β-glycerophosphate (CH/GP)	Load and release of curcumin (Cur)	Load and release of curcumin (Cur)	29 °C	117
	Icariin + PEG hydrogels	DL-Lactide, glycolide and polyethylene glycol	Thermosensitive, distinct anti-oxidative, antimicrobial and anti-nuclear factor-κB-signalling capacities	Thermosensitive, good mechanical properties and biocompatibility	30 °C	118
	PDA/P2-4@Ag gel	Poly(ε-caprolactone- <i>co</i> -glycolide)- <i>b</i> -poly(ethylene glycol)- <i>b</i> -poly(ε-caprolactone- <i>co</i> -glycolide) (PDA/P2) triblock copolymer	Load and release of Ag	Load and release of Ag	34–32 °C	119
	PNI/RA-Amps/E hydrogel	RA-Amps (RADA16-Acp-RRWRVIVKW), antibacterial peptide (Amps), PNIPAM	Thermosensitive, reactive oxygen species (ROS)-scavenging and antibacterial properties	Load and release of MGF E peptide	~32 °C	120
	PNIPAAm–CNC hybrid hydrogels	Poly(N-isopropylacrylamide) (PNIPAAm), cellulose nanocrystals (CNC)	Thermosensitive, injectable, good biocompatibility, good mechanical properties, and antibacterial and carrier functions	Load and release of metronidazole	36–39 °C	121



Table 1 (Contd.)

Application	Name of dressing	Composition	Main features	$T_{\text{sol-gel}}$	Ref.
Cell/factor/protein carrier	Hydrogel-BMSCs	<i>N,N'</i> -Bis(acryloyl)cystamine, <i>N</i> -isopropylacrylamide (NIPAM)	Load with bone marrow-derived mesenchymal stem cells (BMSCs) Thermosensitive, biocompatible, can provide a relatively aseptic and stable environment for the wounds and the transplanted BMSCs	37 °C	122
	SA/bFGF@pNIPAM/DS@p(NIPAM- <i>co</i> -AA) hydrogel	Sodium alginate (SA), poly(<i>N</i> -isopropylacrylamide) nanogels (pNIPAM NGs), p(<i>N</i> -isopropylacrylamide- <i>co</i> -acrylic acid) nanogels [p(NIPAM- <i>co</i> -AA) NGs]	Delivery of diclofenac sodium (DS) and basic fibroblast growth factor (bFGF) Thermosensitive, good mechanical properties, high fluid uptake ability, appropriate water vapor transmission rate, and no cytotoxicity	33 °C/40 °C	92
	Chitosan/collagen/β-GP hydrogels	Chitosan/collagen/β-glycerophosphate (β-GP)	Delivery of 3D Mesenchymal Stem Cell (3D MSCs) Thermosensitive, injectable and provides an environment conducive to the attachment and proliferation of encapsulated 3D MSCs	37 °C	123
	Hydrogel-hUCMSCs	Chitosan/glycerol phosphate sodium/cellulose nanocrystals CS/GP/CNC	Load with human umbilical cord-mesenchymal stem cells (hUCMSCs) Thermosensitive, injectable and low toxicity	32.5 °C	124
	PU/hydrogel	<i>N</i> -Isopropyl acrylamide (NIPAAm), acrylic acid (AAc)	Load and release of fibroblast growth factor-2 (FGF-2) Thermosensitive, injectable, easy to strip off and absorb exudates	32 °C	125
	PF-MC hybrid hydrogel	Gly-TETA (GT), Pluronic F-127 (PF) and methylcellulose (MC)	Load and release of siMMP-9 Thermosensitive, excellent biocompatibility and no toxicity	37 °C	126
	PF-127/hADSCs-Exos complexes	Pluronic F-127 (PF-127)	Release of hADSCs-Exos Thermosensitive, injectable and biocompatible	37 °C	127
	β-GP hydrogel	Amniotic membrane (AM), chitosan/β-glycerophosphate (β-GP), polylactic acid (PLA) microparticles	Load with H ₂ O ₂ Thermosensitive, injectable, nontoxic and oxygen-generating capacity	32.4 ± 2 °C	128
	CSMNA patch	Chitosan (CS), microneedless (MN), <i>N</i> -isopropylacrylamide (NIPAM)	Load and release of VEGF Thermosensitive, microneedle structure and antibacterial property	37 °C	129
	PLEL hydrogel	Poly(<i>D,L</i> -lactide)-poly(ethylene glycol)-poly(<i>D,L</i> -lactide) (PDLLA-PEG-PDLLA: PLEL), platelet-rich plasma (PRP) and growth factors (GFs)	Load and release of PRP GFs Thermosensitive, injectable and nontoxic	37 °C	130
	EGF-Cur-NP/H hydrogel	Nanoparticle/hydrogel (NP/H), growth factor (EGF) and curcumin (Cur)	Load and release of EGF and Cur Thermosensitive, injectable, low cytotoxicity, safety, biodegradable	37 °C	131
	CS-C/OPM/β-GP hydrogel	Catechol-functionalized chitosan (CS-C), β-sodium glycerophosphate (β-GP) and oyster peptide microspheres (OPM)	Load and release of oyster peptide Thermosensitive, no cytotoxicity, good cytocompatibility and blood compatibility	36 °C	132
	eLHBC hydrogel	L-DOPA and ε-poly-L-lysine, hydroxybutyl chitosan (HBC)	Delivery of mesenchymal stem cells (MSCs) Thermosensitive, antimicrobial properties and prevention of wound infection and inflammation response	17.68 °C	133
	sECM-MC hydrogels	Soluble extracellular matrix and methylcellulose (sECM), methylcellulose (MC)	Delivery of stem cells Thermosensitive, injectable and soluble	34 °C	134
	HBC/hPL hydrogel	Hydroxybutyl chitosan (HBC), human platelet lysate (hPL)	Load and release of hPL Thermosensitive and soluble	14 °C	135
	PF127-PT-b1 gel		Release of PT-b1	22 °C	136



Table 1 (Contd.)

Application	Name of dressing	Composition	Main features	$T_{\text{sol-gel}}$	Ref.
Tissue-adhesive hydrogels	Gel-PCLA hydrogel	Pluronic F-127 (PF127), antimicrobial peptidomimetics (AMPMs)	Thermosensitive, antibacterial properties and absorbable		
	NIPAAm-DAA-HA hydrogel	Gelatins (Gs), <i>b</i> -poly(caprolactone- <i>co</i> -lactide) (PCLA)	Thermosensitive, injectable, tissue-adhesive, porous structure, biocompatibility and biodegradability	37 °C	137
	DN gel hydrogel	<i>N</i> -Isopropyl acrylamide, dopamine acrylate (DAA) and stearyl methacrylate (C18)	Thermosensitive, adhesive, self-healing, antibacterial properties and dual injection system	25~32 °C	138
	DG-loaded HP hydrogel	Catechol-Fe ³⁺ and NIPAAm-methacryloyl	Thermosensitive, mechanical flexibility, good adhesive strength, injectable, self-healing, antibacterial, and hemostatic properties	37 °C	139
	GA-HGC hydrogels	Hydroxypropyl chitosan (HPCS) and poly(<i>N</i> -isopropylacrylamide) (PNIPAM)	Load and release of dipotassium glycyrrhizinate (DG)	37 °C	140
Wound contraction	QCS/rGO-PDA/PNIPAm hydrogel	Hexanoyl glycol chitosan (HGC), gallic acid (GA)	Thermosensitive, injectable, highly ductile, self-healable, biocompatible and antimicrobial activity	35 °C	141
	PNIPAm-AA/QCS-CD/PPY hydrogels	Quaternized chitosan- <i>graft</i> -cyclodextrin, adenine, and polypyrrole nanotubes	Thermosensitive, self-healing properties, high compressive strength, strong tissue adhesive, strength biodegradability, excellent biocompatibility	33 °C	142
	Rg3-gel	Ginsenoside Rg3, poloxamer 407	Thermosensitive, suitable mechanical properties, self-healing, on-demand removal, antioxidant, hemostasis, and photothermal/intrinsic antibacterial activity	32 °C	143
	Reticular supramolecular hydrogels	Poly(<i>N</i> -isopropylacrylamide) (PNIPAM)	Thermosensitive, injectable, good network structures, swelling water retention capacity, excellent biocompatibility, good antibacterial, antioxidant properties and wound contraction	22 °C	144
	GNA hydrogel	Methacrylic anhydride modified gelatin (GelMA), acrylic acid (AA) and <i>N</i> -isopropylacrylamide (NIPAM)	Thermosensitive, self-healing, contraction, low cytotoxicity and good biocompatibility	31 °C	145

prepared using CS and poly(*D,L*-lactide)-poly(ethylene glycol)-poly(*D,L*-lactide) (PPP) through a simple process for convenient application.¹⁰⁸ PPP molecules established interaction with CS through H-bonds and formed complexes as loose long chains, in contact with each other, responding to body temperature and rapidly forming hydrogels within approximately 5 s. The optimal formula included a 7% CS PPP micellar solution, exhibiting excellent liquidity and allowing optional application as a spray. The formed hydrogels showed tight adherence to the human skin, rapidly ceased bleeding, effectively inhibited bacterial growth (Fig. 5b), and accelerated wound healing. Moreover, this simple hybrid hydrogel can be easily prepared and applied and has potential applicability in clinical wound treatment. Although hydrogel dressings can protect wounds

from infection, they can be deformed by external forces, leading to infection. Thermosensitive hydrogel materials offer a solution to this problem; when the external environment changes, a phase transition occurs, which can quickly fill irregular wound areas and form gels under physiological conditions. Therefore, the use of thermosensitive hydrogels with antibacterial properties for wound healing is highly desirable.

Drug delivery

Drugs play an important role in wound healing.¹⁴⁷ Thermosensitive hydrogels have been used as drug carriers in the biomedical field^{148~150} because of their injectability, ability to carry therapeutic agents for specific site delivery, prolonged



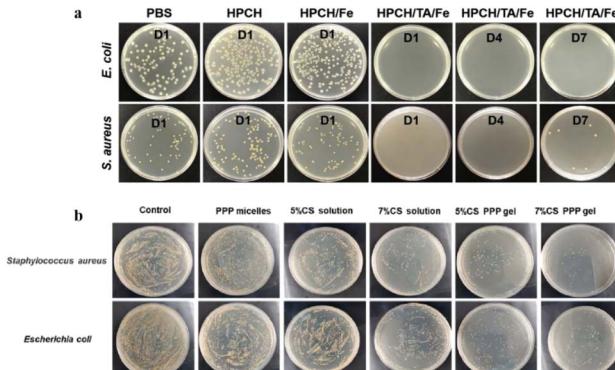


Fig. 5 (a) Bacterial colonies of *E. coli* and *S. aureus* after coculture with PBS, HPCH, HPCH/Fe (day 1), and HPCH/TA/Fe (days 1, 4, 7) hydrogels, respectively. This figure has been reproduced from ref. 106 with permission from Elsevier, copyright 2020. (b) Inhibition of bacterial growth in a rat liver hemorrhage model. This figure has been reproduced from ref. 108 with permission from Elsevier, copyright 2021.

drug effects, improved patient compliance, and reduced systemic toxicity. Thermosensitive hydrogels can be used as drug carriers for local delivery and activate immune cells to promote wound healing.

Heparin was coupled to poly(*N*-isopropyl acrylamide) by developing an injectable *in situ* gel-forming polymer, and ibuprofen was encapsulated in the hydrogel.¹¹⁰ Hep-PNIPAM hydrogel had the highest swelling ratio and formed a hydrogel at 37 °C. Ibuprofen is a non-steroidal anti-inflammatory drug (NSAID) that is critical during the inflammatory phase of healing; however, it has a short half-life. Ibuprofen was encapsulated in the Hep-PNIPAM hydrogel by simple mixing and was formulated as an injectable solution. When the temperature was up to 37 °C, ibuprofen was released mainly by diffusion and was followed by hydrogel erosion. The ibuprofen release profiles from the Hep-PNIPAM hydrogels are shown in Fig. 6a. The drug release curve revealed that the cumulative release was ~70% and sustained ibuprofen release lasted for 1 week. Hydrogels create a moist environment on wounds, absorb exudates, and prevent dehydration. In combination with anti-pain drugs, they create an environment conducive to wound healing.

Curcumin is one of the most potent natural antioxidants.¹⁵¹ Therefore, a thermosensitive graft copolymer of alginate pNIPAM was synthesized for curcumin in the wound area.¹¹¹

The polymer showed excellent thermal gelation properties at different pH and concentration ranges, with good rheological properties. These characteristics allowed the formulation of an injectable dressing of curcumin with easy application to all types of wounds and lesions with different surface geometries and depths. The optimum copolymer showing the gelling temperature at 35 °C, Alg-g-NIPAM(15) was selected for curcumin loading. The cumulative release percentage of curcumin from Cur-F is presented in Fig. 6b. The formulation showed a triphasic release pattern at PBS pH 7.4, the curcumin release time increased up to 72 h (92.4% ± 5.4). This prolonged release of curcumin can be due to both a lag time being needed for the system to absorb water and a slow process of polymer erosion

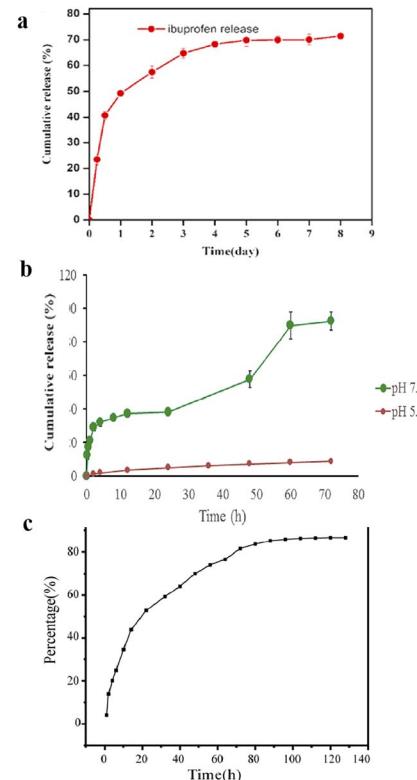


Fig. 6 (a) *In vitro* drug release profiles for the ibuprofen-loaded Hep-PNIPAM hydrogel at pH 7.4 and 37 °C. This figure has been reproduced from ref. 110 with permission from MDPI, copyright 2020. (b) The cumulative *in vitro* release of curcumin from the formulation as a function of time and pH. This figure has been reproduced from ref. 111 with permission from Elsevier, copyright 2021. (c) *In vitro* release of dihydromyricetin. This figure has been reproduced from ref. 113 with permission from Elsevier, copyright 2022.

leading to the release of curcumin molecules encapsulated within more internal parts of the gel. This extended release is beneficial for wound healing.

Zhao *et al.* designed a 37 °C-sensitive hydrogels system using poloxamer 407 as a thermosensitive matrix with chitosan and hyaluronic acid, loaded with dihydromyricetin as a wound dressing.¹¹³ They took the gelation time and temperature as response values to optimize the proportion of various materials and obtained the formula that can be used as a gel at the temperature closest to the human body. Then, 10% dihydromyricetin was added to the gel for structural characterization. Scanning electron microscopy showed that the dihydromyricetin hydrogels performed better in terms of homogeneous distribution and structural quantity, enabling dihydromyricetin to enter cells and be released. The hydrogel was able to release myricetin dihydrate at 37 °C for up to 100 h, and the cumulative release rate was 86% according to the *in vitro* drug release test (Fig. 6c); therefore, it can reduce the frequency of repeated dressing changes in clinical practice.

Thermosensitive hydrogels for drug delivery were developed by mixing thermosensitive polymers with other natural polymers. The critical gelation transition temperature is close to the physiological environmental temperature of the human body



and can be regulated, playing the role of continuous drug release at the right time and target site, thereby improving its suitability and sustainability as a drug carrier. As a physically cross-linked temperature-sensitive hydrogel, there are no organic solvents or nontoxic initiators in the preparation and use processes, which makes it safer. Therefore, thermosensitive hydrogel materials are ideal for sustained drug-release systems.

Cell/factor/protein carrier

Dressings loaded with cells, factors, or proteins can accelerate wound healing. Hydrogels have a high water content and a structure similar to that of the natural extracellular matrix, which can be used as a carrier for cell delivery.¹⁵² In addition, thermosensitive hydrogels can effectively control and continuously release proteins, avoiding adverse side effects that may occur due to the instability of proteins and the need for repeated dosing over time.¹⁵³

The treatment of local wounds with mesenchymal stem cells (MSC) is widely used as a promising approach to enhance tissue regeneration.^{154–156} A new strategy was proposed in which three-dimensional mesenchymal stem cell (3D MSC) spheroids were combined with an injectable thermosensitive chitosan/collagen/β-glycerol phosphate (β-GP) hydrogel to accelerate chronic wound healing by enhanced vascularization and paracrine effects.¹²³ (Fig. 7a). Chitosan/collagen/β-GP solution mixed with 3D MSC spheroids was rapidly turned into a gel at body temperature by physical cross-linking, completely covering the wound. The

hydrogel provided an environment conducive to the attachment and proliferation of encapsulated MSCs, particularly by accelerating the proliferation and paracrine factor secretion of 3D MSC spheroids, thereby promoting wound healing in mice.

Lin *et al.* developed a polyurethane (PU)/hydrogel, a copolymer of the thermosensitive *N*-isopropyl acrylamide (NIPAAm) and acrylic acid (Aac).¹²⁵ NIPAAm was simultaneously copolymerized with Aac, the hydrogel was grafted onto porous PU using-ray irradiation, and fibroblast growth factor-2 (FGF-2) was loaded into the composite. The release of FGF-2 can last for at least 7 days (Fig. 7b), because the interactions between the negative charge of Aac (with COO⁻) and the positive charge of FGF-2 maintain its slow release. This prevents the fast release of FGF-2 due to its high diffusibility. Therefore, the temperature-sensitive feature of the hydrogel can be used to control the release of the active ingredient, as well as to absorb the wound exudate and to keep the wound in a moist environment. The PU/hydrogel composite strips are easier to use than commercial wound dressings due to their thermal sensitivity properties.

The process of wound healing is closely related to the dynamic balance of extracellular matrix (ECM).¹⁵⁷ Matrix metalloproteinases (MMPs) play an important role in regulating the metabolic balance between ECM synthesis and degradation.¹⁵⁸ A hybrid thermosensitive hydrogel dressing was developed for the localized and prolonged delivery of siMMP-9.¹²⁶ The thermosensitive hydrogel consisted of Pluronic F-127 (PF) and methylcellulose (MC), and the GT/siMMP9 complex was loaded onto the hydrogel. The siMMP-9 was first complexed with GT *via* electrostatic interaction, and then GT/siMMP-9 was encapsulated in PM hydrogel. The thermosensitive hybrid hydrogel could be formed *in situ* and then form a strong interface with arbitrarily shaped wounds. In addition, the hybrid hydrogel released GT/siMMP9 continuously through temperature-sensitive control for up to seven days, resulting in significant MMP-9 silencing (Fig. 7c) and accelerated wound healing in diabetic rats.

Traditional thermal gels often exhibit poor physical properties and low binding affinities for cells or proteins, which limit their practical application. Modified thermosensitive hydrogels can be used as carriers of cells or proteins for wound healing owing to their good biocompatibility, biodegradability, and physical stability.

Tissue-adhesive hydrogels

Although various types of hydrogel dressings have been used in soft tissue regeneration, the high water content of the hydrogel polymer network makes them brittle.¹⁵⁹ Hydrogels with superior mechanical properties meet the requirements for tissue regeneration. Therefore, it is necessary to develop adhesive, elastic, and biodegradable hydrogels. Many researchers have developed synthetic hydrogels that mimic biological tissues. However, the damaged covalent bonds in the hydrogel network cannot be easily recombined when the hydrogels are subjected to cyclic loading or deformation,^{160–162} and exhibit poor adhesion or self-healing abilities. Biocompatible and biodegradable polymer materials can be used in composite hydrogel wound dressings for rapid tissue adhesion and closure.

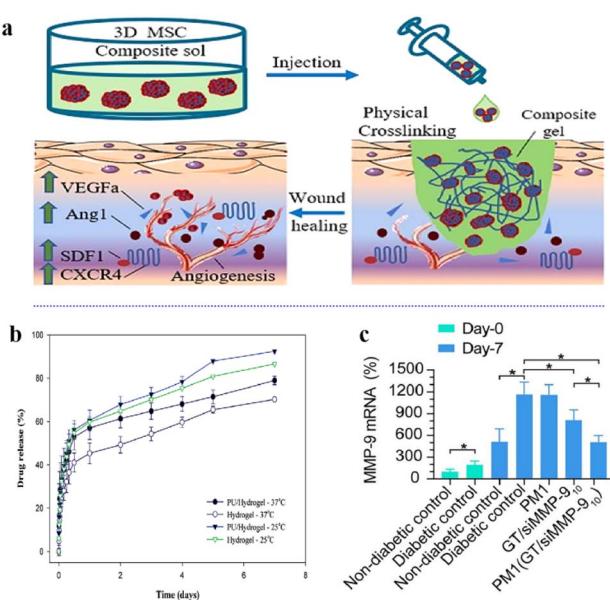


Fig. 7 (a) Schematic illustration of the formation of injectable material. The hydrogel-3D MSCs combination. This figure has been reproduced from ref. 123 with permission from Elsevier, copyright 2022. (b) The release profile of FGF-2 from the hydrogel and PU/hydrogel composite at 25 °C and 37 °C. This figure has been reproduced from ref. 125 with permission from the Royal Society of Chemistry, copyright 2015. (c) MMP-9 protein levels in skin tissues of rats by western blot at day-0 and day-7 after respective treatments. This figure has been reproduced from ref. 126 with permission from BMC, copyright 2021.



b-Poly(caprolactone-*co*-lactide) (PCLA-*b*-PEG-*b*-PCLA, referred to as PCLA), a bioabsorbable temperature-sensitive block copolymer, is conjugated to the backbone of injectable gelatins (IGs) to form a dynamic coordination network.¹³⁷ The investigator adhered an aqueous solution of PCLA copolymer hydrogel and IG conjugate hydrogel to the rat skin (Fig. 8a). The higher number of PCLA block polymer in the conjugates creates a hydrophobic balance to maintain the gel state at body temperature (37 °C). In addition, the existence of functional groups in the Gel-PCLA conjugate can make hydrogen bonding that contributes to the improvement of gel strength. The PCLA copolymers showed temperature-sensitivity and the freely flowing PCLA copolymer sols at low temperature were transformed into stable hydrogels. Therefore, the *in situ* formation of IGs can be considered as promising wound dressing materials for the treatment of multiple wounds.

An injectable multifunctional hydrogel of a mussel-inspired adhesive and self-healing hydrogel was shown to be suitable as a wound-healing dressing for treating deep wounds.¹³⁸ This was inspired by ocean mussels, which can use catechol groups and metal ions to attach to the surfaces of wet objects in fluid environments and self-heal after taking damage.^{163,164} Using a thermally sensitive material as the hydrogel base, the sol-gel transformation can be controlled according to temperature changes in the human body. Methacrylate (C18) was added to

improve the mechanical properties. Furthermore, chelation was conducted between dopamine acrylate with a catechol group and iron ions (Fe^{3+}) in a ferric chloride solution. A sol-gel transition can be quickly achieved through the thermosensitive properties and action of catechol-iron ions, which produces adhesion and self-healing effects. The bioadhesion experimental setup is shown in Fig. 8b. The results showed that as the DAA- Fe^{3+} ratio increased, the adhesiveness of the hydrogel increased (Fig. 8c). The actual adhesion properties of the gel are presented, including its successful adhesion to a glove, polystyrene plate, and glass slide (Fig. 8d).

The hydrogel covered the wound while absorbing exudate and blood from the wound, creating a protective barrier against bacterial infection. However, owing to the high water content of the hydrogel, the wet wound environment, and continuous exudation, it is difficult for hydrogel dressings to adhere to the wound tissue. Tissue adhesion in healing dressings is critical for deep-tissue wounds. Thus, injectable hydrogels that keep a wound moist while maintaining high adhesion can be used for further applications in wound healing.

Wound contraction

Wound closure is a key step in surgery and trauma handling and it allows for the biological event of healing through the joining of wound edges.¹⁶⁵ The hydrogels' thermoresponsive self-contraction ability assists wound closure in the early stages of wound healing. When the hydrogel adheres tightly to the surrounding skin, the contraction of the hydrogel triggers the wound closure. Therefore, a kind of novel multifunctional wound dressing that has biochemical self-contraction ability while promoting wound closure is desired for wound-healing application.

A recent study reported a two-pronged strategy of biomechanically active and biochemically functional hydrogel wound dressing.¹⁴² They used a series of biomechanically active injectable self-healing hydrogels based on quaternized chitosan (QCS), polydopamine-coated reduced graphene oxide (rGO-PDA), and poly(*N*-isopropylacrylamide) (PNIPAm) as multifunctional wound dressings. QCS, rGO-PDA, and NIPAM monomers were fully mixed, and the QCS/rGO-PDA/PNIPAm hydrogels were synthesized with the addition of initiator APS/TEMED and cross-linker BIS *via* free radical polymerization. The researchers applied a hydrogel with outstanding tissue adhesion properties on the wound. Subsequently, because of the thermoresponsive self-contraction ability, when the temperature increased above 33 °C (LCST of PNIPAm), the volume of the hydrogel decreased rapidly, which was beneficial for the initial stage of wound healing through the contraction force-assisted closure of cracked wounds (Fig. 9a). Hydrogels adhere strongly to the skin and assist wound closure by actively contracting wounds through self-contraction.

Yu *et al.*¹⁴³ developed a novel supramolecular poly(*N*-isopropylacrylamide) hybrid hydrogel dressing composed of quaternized chitosan-*graft*-cyclodextrin, adenine, and polypyrrole nanotubes *via* host-guest interactions and hydrogen bonds. Adenine was introduced into the PNIPAm polymer backbone *via*

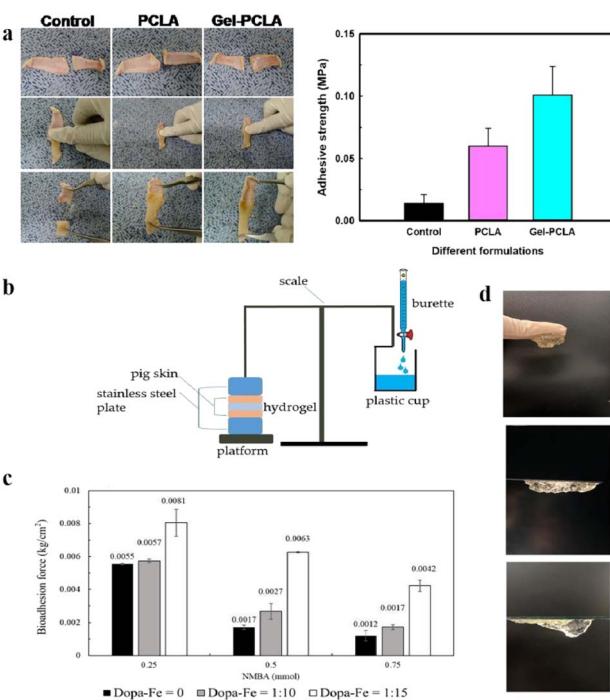


Fig. 8 (a) The adhesive strength of PCLA and IG hydrogels was estimated through a universal testing machine. This figure has been reproduced from ref. 137 with permission from the American Chemical Society, copyright 2019. (b) A bioadhesion testing device by gravimetric measurement. (c) Comparison of hydrogel bioadhesion with DAA- Fe^{3+} ratios and different concentrations of crosslinking agent (NMBA). (d) Image of adhesive hydrogels on the glove, polystyrene, and glass surfaces. These figures have been reproduced from ref. 138 with permission from MDPI, copyright 2022.



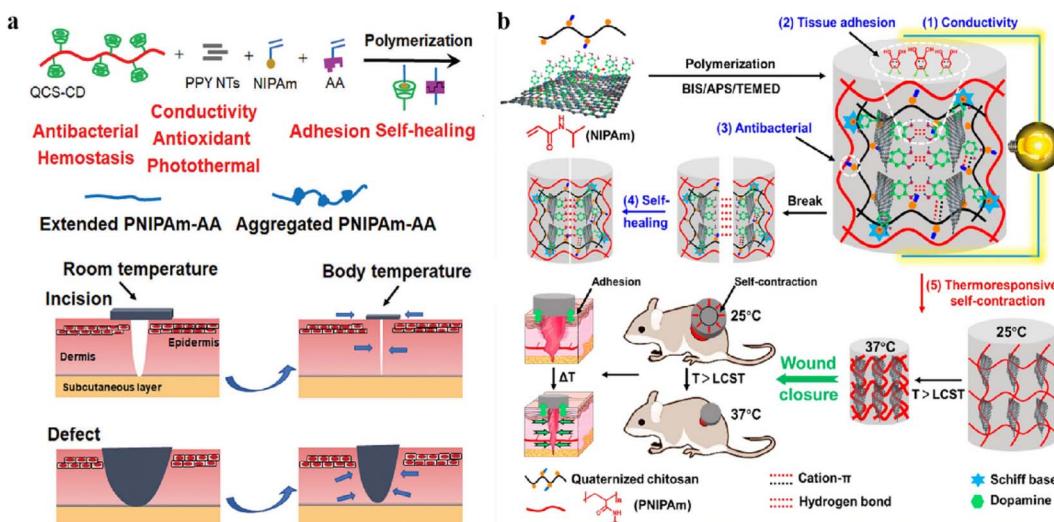


Fig. 9 (a) Schematic diagram of the hydrogels assisting wound closure by thermoresponsive self-contraction. This figure has been reproduced from ref. 142 with permission from the American Chemical Society, copyright 2022. (b) Schematic representation of the PNIPAm-AA/QCS-CD/PPY hydrogel via a dual physical crosslinking strategy and the illustration of simultaneous wound closure and dressing assisted by PNIPAm-AA/QCS-CD/PPY hydrogel. This figure has been reproduced from ref. 143 with permission from Wiley-VCH GmbH, copyright 2022.

free-radical polymerization forming PNIPAm-AA. The researchers added QCS for its biocompatibility, high solubility, inherent antibacterial activities, and synergistic tissue adhesion. At the same time, PPY NTs were introduced to further impart antioxidant properties, antibacterial capacity, and conductivity to the hydrogels. Physical crosslinking, including host-guest interactions and hydrogel bonding, was employed to fabricate the multifunctional hydrogels in this work (Fig. 9b). The hydrogel demonstrated a thermal contraction of 47% of the remaining area after 2 h at 7 °C and tissue adhesion of 5.74 kPa, which are essential for noninvasive wound closure. PNIPAm-AA/QCS-CD/PPY hydrogels adhere strongly to the skin and assist wound closure by actively contracting wounds through self-contraction to promote wound closure and wound healing.

Thermosensitive hydrogel for diverse wounds

Skin wound healing results from a series of complex biological and molecular events, including angiogenesis, cell adhesion, migration, proliferation, differentiation, and extracellular

matrix (ECM) deposition.¹⁶⁶ Several factors affect the process and duration of wound healing. Appropriate wound dressings play important roles in promoting wound healing. Thermosensitive hydrogels are characterized by their excellent temperature sensitivity and injection characteristics and can be used to fill irregular wound sites. Simultaneously, they can be loaded with bioactive molecules or drugs to make functional complementation dressings. Thermosensitive hydrogels can be used to treat chronic diabetic, burn, and surgical wounds (Fig. 10). A summary of thermosensitive hydrogel dressings for diverse wounds is presented in Table 2.

Chronic diabetic wounds

Diabetes is a common clinical disease. According to the findings of the International Diabetes Federation (IDF), approximately 537 million adults worldwide will be living with diabetes in 2021.¹⁶⁷ Diabetic wounds are stimulated by high blood glucose, often accompanied by a persistent inflammatory response, oxidative stress injury, angiogenesis disorders and a high risk of bacterial infection, resulting in skin ulcer, tissue necrosis, wound infection and other problems, and its treatment is complex and expensive, which is a worldwide problem.¹⁶⁸ Therefore, in addition to the original properties of hydrogel dressings, an ideal diabetic wound hydrogel should have anti-inflammatory, antibacterial, antioxidant, pro-angiogenic, and hypoglycemic effects.

In chronic wounds, repeated tissue damage causes an excessive release of cytokines, continuously stimulating and recruiting immune cells to the injured site, resulting in an overactive inflammatory response and the prevention of wound healing.^{169,170} Chen *et al.* developed a biodegradable, multifunctional crosslinker and an *N*-isopropylacrylamide (NIPAM)-based, thermosensitive hydrogel to carry BMSCs to treat diabetic ulcers.¹⁷¹ The crosslinker contained an RGD-like motif that



Fig. 10 Thermosensitive hydrogel for diverse wounds.

Table 2 Summary of thermosensitive hydrogel dressings for diverse wounds

Application	Name of dressing	Composition	Main features	$T_{\text{sol-gel}}$	Ref.
Chronic diabetic wounds	SOD-PP hydrogels	Poly(<i>N</i> -isopropyl-acrylamide)/poly(γ -glutamic acid) (PP)	Load and release of superoxide dismutase (SOD) Thermosensitive, injectable, antioxidant and good biocompatibility	28 °C	75
	PM (GT/siMMP-9) hydrogel	Pluronic F-127 (PF) and methylcellulose (MC)	Load and release of Gly-TETA (GT)/matrix metalloproteinase 9 siRNA (siMMP-9) Thermosensitive, excellent biocompatibility, no toxicity and provided a moist environment	37 °C	126
	BMSCs + hydrogel	<i>N</i> -Isopropylacrylamide (NIPAM), poly(amidoamine) (PAA) and bone marrow mesenchymal stem cells (BMSCs)	Delivery of BMSCs Thermosensitive, injectable, anti-inflammatory and biocompatible	33 °C	171
	<i>L. lactis</i> thermo-sensitive hydrogel	Poloxamer	Delivery lactic acid Thermosensitive, anti-inflammatory, good biocompatibility	37 °C	172
	CS + LG hydrogel	L-Glutamate acid (LG) and chitosan (CS)	Thermosensitive, good swelling, thermal stability, smooth surface topography, and controlled biodegradability	37 °C	199
	ADSCs-PF127 hydrogel	Pluronic F127 (PF127)	Delivery of adipose-derived stem cells (ADSCs) Thermosensitive, injectable and good biocompatibility	37 °C	173
	PBNPs@PLEL	Poly(D,L -lactide)-poly(ethylene glycol)-poly(D,L -lactide) (PDLLA-PEG-PDLLA) hydrogel (PLEL) and Prussian blue nanoparticles (PBNPs)	Release of PBNPs Thermosensitive, injectable, anti-inflammatory, ROS scavenging and mitochondrial function restoration	~28.2 °C	176
	PPCN + SDF-1 hydrogel	Poly(polyethylene glycol citrate- <i>n</i> -isopropyl acrylamide) (PPCN)	Load and release of stromal cell-derived factor-1 (SDF-1) Thermosensitive, injectable and antioxidant	37 °C	177
	Poxin hydrogel	Poloxamox 407 and insulin (Poxin)	Delivery of insulin Thermosensitive, injectable, nontoxic and good biocompatibility	25 °C	181
	ROS-sensitive hydrogel	<i>N</i> -Isopropylacrylamide (NIPAAm), 2-hydroxyethyl methacrylate (HEMA) and benzoyl peroxide (BPO)	Delivery of recombinant human MG53 (rhMG53) protein Thermosensitive, injectable and ROS-scavenging	19 °C	200
	CRMs-hydrogel	Pluronic F127, Pluronic F68	Load and release of curcumin and rifampicin Thermosensitive, excellent intracellular and extracellular bacterial	33 °C	201
	CNPs@GMS/ hydrogel	Gelatin microspheres (GMS), cur nanoparticles (CNPs) and Pluronic F127 and F68	Load and release of curcumin (Cur), MMP9-responsive Thermosensitive, efficient, safe, anti-oxidants	32.97 \pm 0.52 °C	71
	CBP/GMs@Cel&INS hydrogel	Matrix metalloproteinase-9 (MMP-9), polyvinyl alcohol (PVA) and chitosan, phenylboric acid	Load and release of insulin (INS) and celecoxib Thermosensitive, injectable, self-adaptive, remodelling and self-healing properties	25 °C	202
	hUCMSC-exos-PF-127 hydrogel	Pluronic F-127 (PF-127), human umbilical cord MSC (hUCMSC)-derived exosomes (hUCMSC-exos)	Delivery of hUCMSC-exos Thermosensitive and injectable	12.4–17.8 °C	203
	hASCs-hydrogel	Human decellularized adipose matrix (hDAM)	Delivery of human adipose-derived stem cells (hASCs) Thermosensitive and provide a favorable microenvironment for hASCs	31 °C	204
	Nr-CWS hydrogel	Nocardia rubra cell wall skeleton (Nr-CWS), poloxamers 407 and 188	Load and release of Nr-CWS Thermosensitive and good affinity	20 °C	205



Table 2 (Contd.)

Application	Name of dressing	Composition	Main features	$T_{\text{sol-gel}}$	Ref.
Burn wounds	CLD-hFGF2 hydrogel	Camelina lipid droplets (CLD), poloxamer P188 and P407	Load and release of human fibroblast growth factor 2 (hFGF2) Thermosensitive, injectable, high bioactivity	24–25 °C	190
	Chitosan-PDEAAM hydrogel	Chitosan, poly(<i>N,N</i> -diethylacrylamide) (PDEAAM)	Load and release of incorporated antibiotics Thermosensitive, easy to remove and antibacterial properties	37 °C	191
	ZnMet-PF127 hydrogel	Pluronic F127 (PF127)	Load and release of ZnMet Thermosensitive, sprayable, anti-inflammatory and antibacterial effects	37 °C	192
	AgNPs-MC hydrogel	Methylcellulose (MC) hydrogel, silver oxide nanoparticles (NPs) and silver acetate (CH_3COOAg)	Release of Ag ions Thermosensitive, injectable, excellent antimicrobial activity	33.6–37 °C	206
	AgSD/NS hydrogel	Silver sulfadiazine/nanosuspensions (AgSD/NS) and poloxamer 407	Load and release of AgSD/NS Thermosensitive, non-toxic and antibacterial	29.8 °C	207
	CAT-Gel-Alg hydrogel	Gelatin (Gel)-alginate (Alg) biocompatible hydrogel (Gel-Alg)	Delivery of catalase (CAT) and ROS scavenging Thermosensitive, antioxidant-biocompatible, safe and sustainable	20 °C	208
	PLX/CS/HA-VM hydrogel	Poloxamer 407 (PLX), chitosan (CS), hyaluronic acid (HA), melatonin (MLT), vitamin A, D3 and E	Delivery of MLT, vitamins A, D3 and E Thermosensitive, antibacterial and antioxidant	30 ± 0.2 °C	209
	FA-loaded hydrogels	Ferulic acid (FA), chitosan/gelatin/glycerol phosphate	Load and release of FA Thermosensitive, excellent antioxidant and biocompatible	37 °C	210
	MSC-CM/hydrogel	Conditioned medium (CM), chitosan, collagen, β -glycerophosphate (β -GP) and mesenchymal stem cells (MSCs)	Delivery of MSCs Thermosensitive, good biocompatibility and low cytotoxicity	37 °C	211
	CEAllo hydrogel	Poloxamer 407	Thermosensitive, safe and allows for the application of the cells thinly	20 °C	212
Surgical wounds	RHC-chitosan hydrogels	Chitosan, recombinant human collagen-peptide (RHC)	Release of RHC Thermosensitive, injectable, superior mechanical strength and excellent attachment substrate for cells	15/17/24 °C	213
	AgSPHA hydrogel	Hyaluronic acid (HA), corn silk extract (CSE) and silver nanoparticles (Ag NPs)	Thermosensitive, injectable, good mechanical properties, antibacterial properties and biocompatibility	32.5 °C	214
	GT-SA-TPFX hydrogel	Sodium alginate (SA), gelatin (GT), ferric ions and protocatechualdehyde	Thermosensitive, injectable, sufficient mechanical, adhesive strength, good biocompatibility, self-healing capacity and antibacterial and antioxidation properties	~43 °C	197
	TRS hydrogel	<i>N</i> -Isopropylacrylamide, butylacrylate	Thermosensitive, injectable, shape adaptability, safety, effectiveness and easy to remove	32 °C	198
	mXG/HBC hydrogel	Galactose-modified xyloglucan (mXG) and hydroxybutyl chitosan (HBC)	Thermosensitive, injectable, excellent cytocompatibility, hemocompatibility and antibacterial properties	mXG/HBC ratio	215
	ZnG/rhEGF@Chit/Polo hydrogels	Chitosan/poloxamer-based thermosensitive hydrogels, zinc gluconate/recombinant human epidermal growth factor (ZnG/rhEGF)	Load and release of ZnG/rhEGF Thermosensitive, good cytocompatibility, mild non-irritating and biocompatible	34.85 °C	216
	PTX-loaded PECE hydrogel	Poly(ethylene glycol)-poly(3-caprolactone)-poly(ethylene glycol) (PEG-PCL-PEG, PECE)	Release of paclitaxel Thermosensitive, injectable and biocompatible	37 °C	217
	HBC-PSB hydrogel	Hydroxybutyl chitosan (HBC) and poly(sulfobetaine methacrylate) (PSBMA)	Thermosensitive, injectable, non-toxic, good synergistic antibacterial, excellent self-healing behaviors and anti-adhesion	PSBMA content	218



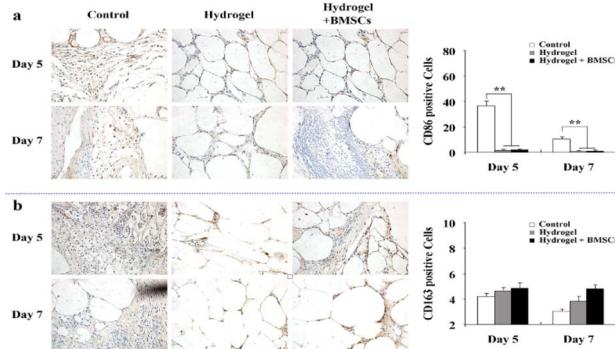


Fig. 11 (a) The expression of M1 macrophages. (b) The expression of M2 macrophages. These figures have been reproduced from ref. 171 with permission from Nature, copyright 2015.

promoted the attachment and differentiation of BMSCs. The polymer was in the sol state at room temperature; after immersion in a 33 °C water bath, it experienced sol–gel transition in minutes. Researchers seeded BMSCs on the surface of hydrogels, which can adhere to and proliferate on the hydrogel. The results showed that the expression of pro-inflammatory M1-type macrophages in the experimental group was lower than that in the control group, and the expression of M2-type macrophages was higher (Fig. 11a and b). A hydrogel loaded with BMSCs can promote the adhesion and differentiation of BMSC, which is conducive to wound healing and improves the inflammatory microenvironment of diabetic wounds. In another study, a thermosensitive hydrogel dressing loaded with *Lactobacillus lactis* was prepared.¹⁷² The hydrogel material produced and delivered lactic acid *in situ*, both *in vitro* and *in vivo*, promoted the transformation of macrophages from M1 to M2, and accelerated the healing of chronic diabetic wounds.

Neovascularization plays a crucial role in wound healing. However, microvascular formation in the wounds of patients with diabetic feet is impaired, resulting in wounds being unable to obtain sufficient oxygen, nutrients, and growth factors.¹⁷¹ In a recent study, rat adipose-derived mesenchymal stem cells (ADSCs) were loaded into Pluronic F127 (PF127) to prepare a novel thermosensitive hydrogel material, and the effect of PF127 hydrogel-loaded ADSCs on angiogenesis in large diabetic skin wounds was observed.¹⁷³ Immunohistochemistry, micro-vessel counting, wound area measurement, and VEGF measurement showed that the experimental group had more new capillaries and faster wound healing than the control group ($P < 0.05$). PF127 hydrogel-loaded ADSCs accelerated wound healing by promoting the formation of new capillaries in diabetic rat wounds.

High-reactive oxygen species (ROS) concentrations are one of the reasons for chronic wound healing caused by diabetes.¹⁷⁴ Dressings that scavenge ROS can effectively reduce the concentration of reactive oxygen species in a wound, improve its inflammatory microenvironment, and promote wound healing.¹⁷⁵ Xu *et al.* prepared a thermosensitive poly(D,L-lactide)-poly(ethylene glycol)-poly(D,L-lactide) (PDLLA-PEG-PDLLA) hydrogel (PLEL)-based wound dressing in which PBNPs were

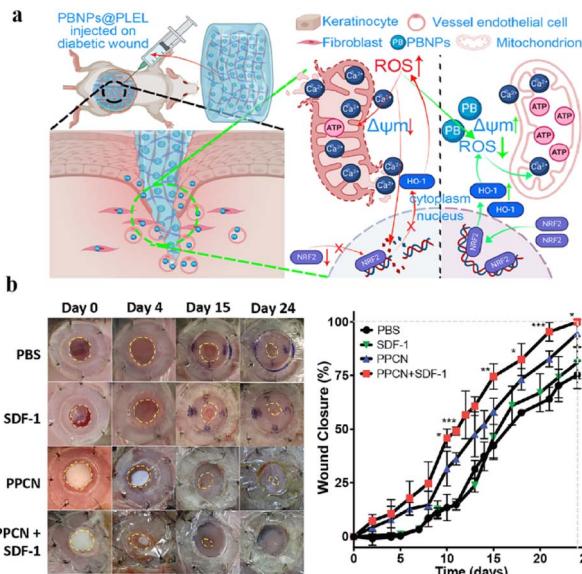


Fig. 12 (a) Schematic overview of the use of PBNPs@PLEL for the enhancement of diabetic wound healing due to its ability to scavenge ROS and to promote the functional recovery of mitochondria. This figure has been reproduced from ref. 176 with permission from ACS Publications, copyright 2022. (b) PPCN + SDF-1 promotes the healing of diabetic wounds. This figure has been reproduced from ref. 177 with permission from Elsevier, copyright 2016.

homogeneously incorporated. The system at 37 °C was in a gel state. This injectable PBNPs@PLEL system can adapt to various wound types and form a gel within 30 s *in vivo*.¹⁷⁶ Once placed, PBNPs@PLEL exhibited sustained and controlled release behavior. PBNPs were slowly released and exerted sustained antioxidant activity after the injection of PBNPs@PLEL into the wound site (Fig. 12a). Zhu *et al.* synthesized poly(polyethylene glycol citrate-*n*-isopropyl acrylamide) (PPCN) *via* sequential polycondensation and free radical polymerization reactions.¹⁷⁷ PPCN has intrinsic antioxidant properties due to the diol-citrate esters in its backbone that may reduce oxidative stress at the wound after gelatinization above the LCST (37 °C). A thermosensitive hydrogel material with sustained release of SDF-1 was prepared, which not only played an antioxidant role but also accelerated the regeneration of wound granulation tissue and promoted rapid wound healing (Fig. 12b). SDF-1 was introduced to promote angiogenesis and cell migration. When the temperature dropped to 37 °C, there was sustained release of SDF-1 from the hydrogel.

The application of topical insulin has also been shown to be of great help in improving wound healing.^{178,179} However, due to molecular instability and sustained delivery effects, local insulin administration still faces great challenges.¹⁸⁰ A previous study reported that a thermosensitive hydrogel combined with insulin injection improved skin wound healing in diabetic mice.¹⁸¹ Poloxamer 407 is widely employed for drug delivery because it is reported to be nontoxic and can form gels at 25 °C. Poloxamer 407 and insulin were injected into the wounds of diabetic mice and compared with those of other controls to measure the rate of wound healing at 28 days intervals



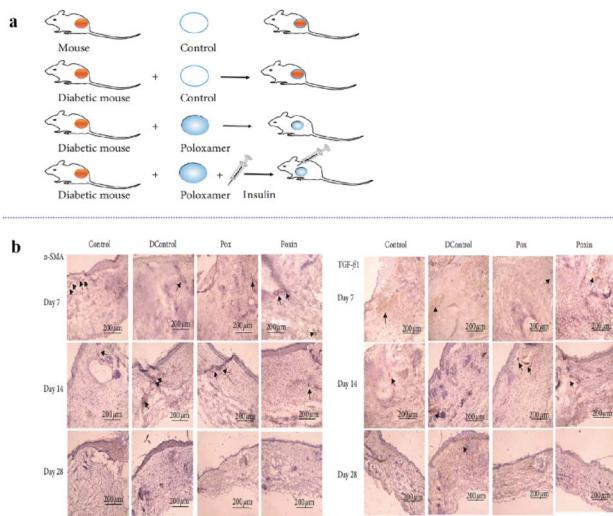


Fig. 13 (a) Experimental mice model: control, diabetic mouse (dControl), poloxamer (Pox), and poloxamer plus insulin (Poxin). (b) α -SMA and TGF- β 1 antigen expression in the wound bed on days 7, 14, and 28 after wounding: samples, dControl, Pox, and Poxin groups. Black arrows show examples of secretion sites of the antigens. These figures have been reproduced from ref. 181 with permission from Hindawi, copyright 2022.

(Fig. 13a). The results showed that the expression of both transforming growth factor beta 1 (TGF- β 1) and α -smooth muscle actin (α -SMA) increased significantly in the early stages of diabetic wounds after insulin injection (Fig. 13b). Poloxamer + insulin-treated mice displayed enhanced wound healing.

The emergence of thermosensitive hydrogels has provided a new approach to the management of diabetic wounds. Thermosensitive hydrogels can be attached to different materials to adapt to different types of wounds. The sustained release of drugs or bioactive molecules from thermosensitive hydrogels is important for promoting chronic wound healing. The generation of chronic diabetic wounds is a multifaceted and multi-linked process. Based on the different stages of diabetic wound healing, more targeted and individualized thermosensitive hydrogel dressings with additional functions can be designed.

Burn wounds

Burns are a common type of injury in humans and are generally caused by exposure to hot liquids, vapors, or gases.^{182,183} Different classifications were performed according to the depth of the burn wounds.¹⁸⁴ Burn wound dressings need to be replaced frequently, which aggravates pain and discomfort.¹⁸⁵ Traditional burn wound dressings easily adhere to the wound after absorbing the wound exudate. When dressings are removed, the wound is injured again, and new tissue tears easily.^{186,187} Therefore, an ideal dressing for the treatment of burn wounds should provide a local moist wound-healing environment for the body, adapt to different wound shapes, and improve the therapeutic effect of wound-healing drugs.

Human fibroblast growth factor 2 (hFGF2) can promote fibroblast proliferation, migration, and differentiation; the local

administration of hFGF2 in burn wounds can promote granulation tissue regeneration and angiogenesis,¹⁸⁸ but it can easily lose its biological activity.¹⁸⁹ In order to provide a supporting environment for hFGF2 and controls its release in a stable manner. Zhang *et al.* encapsulated camelina lipid droplet (CLD)-hFGF2 in thermosensitive hydrogels.¹⁹⁰ The poloxamer composite hydrogels were synthesized from P188 and P407. The shell of these spherical micelles consists of a polyoxyethylene nucleus and an expanded polyoxyethylene chain. With the increase in temperature, the hydrophobic polyoxyethylene segment in the hydrogel loses water and polymerizes into micelles. As the temperature continues to rise, the formation arrangement of micelles forms hydrogels. They can quickly gel on the surface of burn wounds, reduce drug flow loss and improve the drug utilization rate. The prepared CLD-hFGF2 hydrogel can gel with changes in temperature (Fig. 14a) and undergo *in vitro* degradation. CLD-hFGF2 exhibited an initial burst release from the hydrogels after 12 h (approximately 30%), followed by a slow release over 72 h (almost 60%) after drug delivery, CLD-hFGF2 promoted the proliferation and migration of NIH3T3 cells. *In vivo* experiments also showed that the CLD-hFGF2 hydrogel accelerated extracellular matrix (ECM) regeneration and angiogenesis, significantly improved burn wound

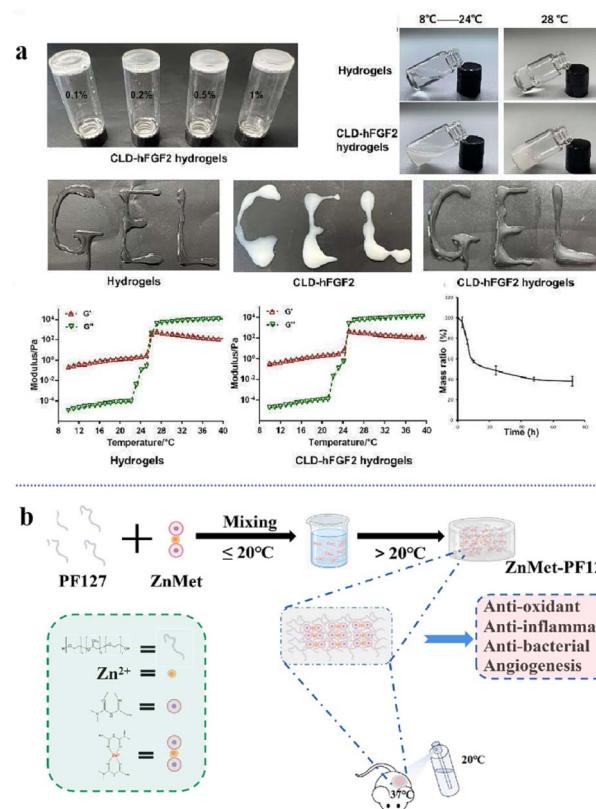


Fig. 14 (a) The thermosensitive property of CLD-hFGF2 hydrogels and *in vitro* degradability. This figure has been reproduced from ref. 190 with permission from MDPI, copyright 2022. (b) Schematic diagram of the ZnMet-PF127 preparation process and application. This figure has been reproduced from ref. 192 with permission from Elsevier, copyright 2022.



healing rates, and reduced the number of dressing changes. In another study, a heat-responsive wound dressing was synthesized using chitosan and poly(*N,N*-diethylacrylamide) (PDEAAm).¹⁹¹ It has temperature-dependent and swelling properties and is easy to separate at lower temperatures. This ability to modulate adhesion will facilitate a non-traumatic manner of removal from wound sites, especially burn wounds. Antimicrobial activity studies have demonstrated the ability of the dressing to control the release of incorporated antibiotics and enhance wound healing by inhibiting bacterial growth.

Burn wounds vary in depth and area and their shapes are usually irregular. Moreover, owing to the complexity and severity of burn wounds, traditional dressings cannot meet the requirements for burn wound healing. In a recent study, the thermosensitive hydrogel Pluronic F127 (PF127) was used for the first time as a sprayer for skin wound healing (Fig. 14b).¹⁹² A coordination complex of zinc and metformin (ZnMet) was loaded onto PF127. The polymer did not change the properties of PF127, and the porous structure allowed the uniform release of the drug in the tissue. Sprayable ZnMet-PF127 evenly filled irregular skin defects in the liquid state. After ZnMet-PF127 was sprayed on the skin wound, it immediately changed from liquid to semi-solid due to its thermosensitive characteristics, isolating the skin wound from the external environment and reducing wound infection. It can also provide a wet environment for burn wounds as well as anti-inflammatory and anti-bacterial effects to promote wound healing.

Thermosensitive hydrogels can quickly gel at body temperature. When dressings are changed, a gel-solution thermosensitive conversion can be used to facilitate fading and reduce unnecessary pain. At the same time, it is not limited by the shape and depth of the wound and can completely cover the wound due to its injectable features. In addition, thermosensitive hydrogels loaded with drugs or bioactive substances exhibit good biocompatibility, which can promote burn wound healing and reduce the frequency of dressing changes through the sustained release of drugs or factors. These properties make thermosensitive hydrogels promising materials for a wide range of applications in burn wounds.

Surgical wounds

Traditional closure methods for surgical wounds include the use of sutures or staples, which are usually invasive and may cause unsatisfactory tissue integration, trauma, and leakage of content.^{193–196} Thermosensitive hydrogels exhibit good biocompatibility, tissue adhesion, and temperature-sensitivity. They can avoid secondary injury to the surgical incision, reduce the frequency of dressing changes, and promote wound healing, providing a new choice for surgical wound healing. Recently, a novel bio-inspired injectable wound dressing with superior mechanical properties, adhesion strength, temperature adhesion, and self-healing ability was developed.¹⁹⁷ Ferric ions were added to a multifunctional adhesive composed of sodium alginate (SA), gelatin (GT) and protocatechualdehyde for sutureless post-wound closure. The thermal-response phase transition

of GT, the Schiff base bond, and the catechol-Fe-coordinate bond in the network give the hydrogel injectable, shape-adaptable, and strong adhesion and self-healing abilities at body temperature. Rat skin incisions were created, and then sutures, biomedical glue, and adhesive hydrogel were used to heal the incision; one without treatment was set as the control group. Compared with the other control groups, the mechanical strength of the skin tissue treated with the hydrogel group was stronger, the incision was smoother, and the incision closure effect was better than that of the other groups (Fig. 15a). This is primarily because the temperature-dependent adhesion of the hydrogel makes the sealant easy to operate. The hydrogel can separate fault-tolerant adhesion from bad adhesion on the instrument by sensing the local temperature. In addition, its good self-healing ability allows the reopened incision to be closed repeatedly.

Good adhesive strength and injection performance can effectively close the incision and promote healing. This multifunctional hydrogel may contribute to the healing of surgical incisions, and the fast and effective closure of surgical wounds is critical. If not treated promptly, injuries can rapidly escalate

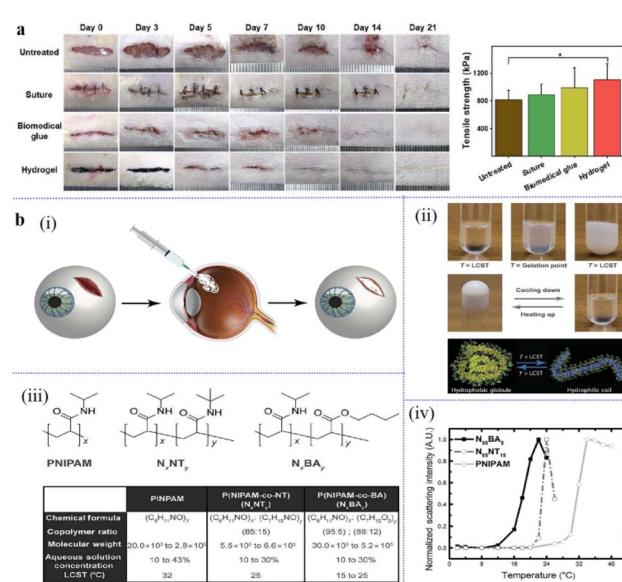


Fig. 15 (a) Images of the incisions closed by suturing and the tensile strength of the healed skin tissues on day 21. This figure has been reproduced from ref. 197 with permission from *Nano-Micro Lett.*, copyright 2022. (b) The design of a thermoresponsive hydrogel to seal scleral perforation. (i) A schematic depicting the implementation of a temperature-mediated adhesive hydrogel that adapts to wound shape and a tool for its deployment at a perforation site in the sclera. (ii) Images of the changes in physical properties as the sealing hydrogel transitions from hydrophilic coils to hydrophobic globules at its lower critical solution temperature (LCST). (iii) Molecular structures of formulations of poly(*N*-isopropylacrylamide) (PNIPAM) with butylacrylate (BA) or *N*-tert-butylacrylamide (NT) and a table of resulting alterations of molecular properties and LCST values. (iv) Normalized scattering intensity as a function of temperature for PNIPAM, poly(*N*-IPAM-*co*-NT) (N85NT15), and poly(*N*-IPAM-*co*-BA) (N95BA5). A.U., arbitrary units. These figures have been reproduced from ref. 198 with permission from the American Association for the Advancement of Science, copyright 2017.



and produce adverse outcomes. Bayat *et al.* designed a system for temporarily closing open eye lesions that restored the intraocular pressure (IOP) and was easily removed (Fig. 15b).¹⁹⁸ Through the reversible and thermal response properties of PNIPAM, its thermal response behavior and mechanical strength were adjusted to form a hydrogel that could fill irregular edges and seal the wound. Their thermal-sensitive behavior allows thermal sealants (TRS) to be easily removed by applying cold water without additional trauma to the surrounding tissue during placement or removal. Hydrogels have the advantages of rapid reversible wound occlusion, shape adaptability, safety, and effectiveness, as well as translational potential as intelligent wound sealants, which can be used as intelligent wound dressings for the temporary occlusion of surgical incisions or wounds. Thermosensitive hydrogels can be thermally responsive through biodynamic bonding with rapid injectable filling, good adhesive strength, responsive adhesion ability, and easy placement or removal properties, making them promising candidates for surgical incision dressings.

Conclusions and further perspectives

Wound healing is a complex dynamic biological process. With the development and integration of biomedicine and materials science, the research and development of new hydrogels with high efficiency, intelligence, and microenvironmental adaptability provide new ideas and opportunities for wound repair. Thermosensitive hydrogels make use of the specificity of polymer materials in response to external temperatures so that the polymer undergoes reversible conformational changes under external temperature changes and realizes the transformation process from solution to gel. It is an excellent carrier for loading various bioactive substances or drugs in liquid form and can promote the transdermal absorption of drugs after being turned into a gel. Simultaneously, good mechanical ability and tissue adhesion of thermosensitive hydrogels, easy removal, and other characteristics are crucial for wound management. A few points should be emphasized when designing new thermosensitive hydrogels for wound healing as follows. (a) The mechanical stiffness of the thermosensitive hydrogels formed *in situ* has a great influence on their practical application. (b) The selection of the thermosensitive polymer types will affect the biocompatibility and biodegradability of gel systems. (c) Appropriate temperature-sensitive materials should be selected according to different wound types. (d) The duration of gelation time should be controlled reasonably according to the therapeutic needs to avoid the sudden release of drugs, factors, proteins, etc.

Although thermosensitive hydrogels have achieved some success in wound treatment as wound-healing dressings, some problems remain. For example, most studies are based on animal and tissue models, which are different from those of the human body. Conducting research in a clinic requires further research and testing. In addition, the shortcomings of thermosensitive hydrogels include slow temperature response, poor biocompatibility and low mechanical properties, and will affect the expansion in the field of wound healing application. Therefore, the development of a successful thermosensitive

hydrogel for wound healing depends on the development of sophisticated material engineering. The future direction of thermosensitive hydrogels for wound healing is to optimize and improve the characteristics of hydrogels to satisfy the treatment needs, and to prepare smart hydrogels with lower cost and lower toxicity, which is expected to realize personalized and refined treatment of different wounds and have broad development potential and clinical application prospects. Therefore, this study can provide a reference for the design and development of smart hydrogel wound dressings.

Author contributions

Ruting Gu: conceptualization, data curation, methodology, writing – original draft. Haiqing Zhou: data curation, methodology, validation. Zirui Zhang: methodology, formal analysis. Yun Lv: supervision, validation. Yueshuai Pan: data curation. Qianqian Li: data curation. Changfang Shi: visualization. Yan-hui Wang: supervision, writing – review & editing. Lili Wei: project administration, writing – review & editing.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The authors thank for the financial support of Shandong Provincial Natural Science Foundation (ZR2022QE273).

References

- 1 E. S. Chambers and M. Vukmanovic-Stejic, *Immunol.*, 2020, **160**, 116–125.
- 2 J. Larouche, S. Sheoran and K. Maruyama, *Adv. Wound Care*, 2018, **7**, 209–231.
- 3 K. Singh, E. Camera and L. Krug, *Nat. Commun.*, 2018, **9**, 3425.
- 4 E. Kolanthai, Y. Fu and U. Kumar, *Wiley Interdiscip. Rev.: Nanomed. Nanobiotechnol.*, 2022, **14**, e1741.
- 5 J. Koehler, F. P. Brandl and A. M. Goepferich, *Eur. Polym. J.*, 2018, **100**, 1–11.
- 6 P. Martin, *Science*, 1997, **276**, 75–81.
- 7 G. Luo, Y. Lu and C. Huang, *Zhonghua Shaoshang Zazhi*, 2023, **39**, 9–14.
- 8 R. Derwin, D. Patton and P. Avsar, *Int. Wound J.*, 2022, **19**, 1397–1408.
- 9 G. Gethin, J. D. Ivory and D. Sezgin, *Wound Repair Regen.*, 2021, **29**, 843–847.
- 10 T. M. Honnegowda, U. E. Padmanabha and P. Rao, *World J. Plast. Surg.*, 2016, **5**, 265–273.
- 11 L. Y. Zhu, S. X. Guo and P. Wu, *Zhonghua Shaoshang Zazhi*, 2018, **34**, 829–832.
- 12 Y. Liang, M. Li and Y. Yang, *ACS Nano*, 2022, **16**, 3194–3207.
- 13 J. Rojas, M. Bedoya and Y. Ciro, *Cellulose*, 2015, **32**, 193–228.

14 Y. Li, X. Jiang and Z. Qin, *J. Xi'an Jiaotong Univ., Med. Sci.*, 2022, **43**, 943–951.

15 C. Canal, F. Gaboriau and S. Villeger, *Int. J. Pharm.*, 2009, **367**, 155–161.

16 K. Chen, F. Wang and S. Liu, *Int. J. Biol. Macromol.*, 2020, **148**, 501–509.

17 S. Hu, S. Bi and D. Yan, *Carbohydr. Polym.*, 2018, **184**, 154–163.

18 H. Peng, X. Ning and G. Wei, *Carbohydr. Polym.*, 2018, **195**, 349–355.

19 A. R. Abbasi, M. Sohail and M. U. Minhas, *Int. J. Biol. Macromol.*, 2020, **155**, 751–765.

20 Y. Liang, J. He and B. Guo, *ACS Nano*, 2021, **15**, 12687–12722.

21 R. Fan, Y. Cheng and R. Wang, *Polymers*, 2022, **14**, 2379.

22 P. Patel, A. Mandal and V. Gote, *J. Polym. Res.*, 2019, **26**, 1–11.

23 Y. K. Du, U. P. Shinde and B. Yeon, *Prog. Polym. Sci.*, 2013, **38**, 672–701.

24 M. Calderón, M. A. Quadir and M. Strumia, *Biochimie*, 2010, **92**, 1242–1251.

25 C. Kojima, *Expert Opin. Drug Delivery*, 2010, **7**, 307–319.

26 W. Zhang, X. Jin and H. Li, *Carbohydr. Polym.*, 2018, **186**, 82–90.

27 M. F. Jéssica, F. M. Simone, A. Gizelda Maria, S. Danilo Martins Dos, C. F. Sérgio Paulo and S. Amilton Martins Dos, *Colloids Surf., B*, 2018, **175**, 73–83.

28 L. Pham, L. H. Dang and M. D. Truong, *Biomed. Pharmacother.*, 2019, **117**, 109183.

29 L. Zhang, S. Gurankit, M. Zhang, S. X. Chen, K. G. Xu, P. C. Xu, X. Wang, Y. H. Chen, L. Zhang and L. Zhang, *Mater. Sci. Eng., C*, 2018, **90**, 159–167.

30 C. Qu, Z. Bao and X. Zhang, *Int. J. Biol. Macromol.*, 2019, **125**, 78–86.

31 B. Tang, J. shan and T. Yuan, *Carbohydr. Polym.*, 2019, **209**, 198–206.

32 M. Dhrubojoyti, A. Md and S. Santhosh, *J. Drug Delivery Sci. Technol.*, 2018, **46**, 498–510.

33 J. I. Lee, H. S. Kim and H. S. Yoo, *Int. J. Pharm.*, 2009, **373**, 93–99.

34 A. Paul, A. Hasan and H. A. Kindi, *Acs Nano*, 2014, **8**, 8050–8062.

35 J. Qu, X. Zhao and Y. Liang, *Biomaterials*, 2018, **183**, 185–199.

36 H. Huang, X. Qi and Y. Chen, *Saudi Pharm. J.*, 2019, **27**, 990–999.

37 M. Wang, M. Chen and W. Niu, *Biomaterials*, 2020, **261**, 120301.

38 D. Ma, *Preparation and Properties of Multifunctional Composite Hydrogel-based Wound Dressings*, Jilin University, 2022.

39 J. S. Boateng, K. H. Matthews and H. N. Stevens, *J. Pharm. Sci.*, 2008, **97**, 2892–2923.

40 S. Han, M. Li and X. Liu, *Colloids Surf., B*, 2013, **102**, 833–841.

41 S. Alven, S. Peter and Z. Mbese, *Polymers*, 2022, **14**, 724.

42 M. Prasathkumar and S. Sadhasivam, *Int. J. Biol. Macromol.*, 2021, **186**, 656–685.

43 A. M. Mohammed, S. K. Osman and K. I. Saleh, *AAPS PharmSciTech*, 2020, **21**, 131.

44 W. Zheng, J. Wang and L. Xie, *J. Mater. Sci.: Mater. Med.*, 2019, **30**, 106.

45 R. Nigmatullin, V. Gabrielli and C. Juan, *Cellulose*, 2019, **26**, 529–542.

46 E. Dashtimoghadam, H. Salimi-Kenari and R. Nasseri, *Colloids Surf., A*, 2020, **590**, 124489.

47 S. H. Nezhadi, P. F. Choong and F. Lotfipour, *J. Drug Targeting*, 2009, **17**, 731–738.

48 L. Klouda, *Eur. J. Pharm. Biopharm.*, 2015, **97**, 338–349.

49 C. N. Schnell, M. V. Galván and M. A. Zanuttini, *Cellulose*, 2019, 1465–1481.

50 Z. Lu, J. Gao and Q. He, *Carbohydr. Polym.*, 2017, **156**, 460–469.

51 N. Sayari, A. Sila and B. E. Abdelmalek, *Int. J. Biol. Macromol.*, 2016, **87**, 163–171.

52 L. Pighinelli and M. Kucharska, *Carbohydr. Polym.*, 2013, **93**, 256–262.

53 M. A. Matica, F. L. Aachmann and A. Tøndervik, *Int. J. Mol. Sci.*, 2019, **20**, 5889.

54 D. Zhang, S. Lu and Z. Hu, *Appl. Chem. Ind.*, 2019, **48**, 1734–1739.

55 B. Zhang, Z. Zou and X. Nie, *Shandong Chem. Ind.*, 2020, **49**, 49–51.

56 H. Ueno, F. Nakamura and M. Murakami, *Biomaterials*, 2001, **22**, 2125–2130.

57 Z. Mao, H. Shi and R. Guo, *Acta Biomater.*, 2009, **5**, 2983–2994.

58 O. M. Dragostin, S. K. Samal and M. Dash, *Carbohydr. Polym.*, 2016, **141**, 28–40.

59 L. H. Fu, C. Qi and M. G. Ma, *J. Mater. Chem. B*, 2019, **7**, 1541–1562.

60 Y. Pan, X. Zhao and X. Li, *Polymers*, 2019, **11**, 1846.

61 F. Feizabadi, M. Minaiyan and A. Taheri, *Curr. Drug Delivery*, 2018, **15**, 840–849.

62 M. Alavi, *e-Polym.*, 2019, **19**, 103–119.

63 F. Wang, Y. Pan and P. Cai, *Bioresour. Technol.*, 2017, **241**, 482–490.

64 O. Forero-Doria, E. Polo and A. Marican, *Carbohydr. Polym.*, 2020, **242**, 116383.

65 Y. Huang, L. Bai and Y. Yang, *J. Colloid Interface Sci.*, 2022, **608**, 2278–2289.

66 M. Santoro, A. M. Tatara and A. G. Mikos, *J. Controlled Release*, 2014, **190**, 210–218.

67 A. O. Elzoghby, *J. Controlled Release*, 2013, **172**, 1075–1091.

68 L. Klouda and A. G. Mikos, *Eur. J. Pharm. Biopharm.*, 2008, **68**, 34–45.

69 C. Joly-Duhamel, D. Hellio and M. Djabourov, *Langmuir*, 2002, **18**, 7208–7217.

70 L. Pham, L. H. Dang and M. D. Truong, *Biomed. Pharmacother.*, 2019, **117**, 109183.

71 J. Liu, Z. Chen and J. Wang, *ACS Appl. Mater. Interfaces*, 2018, **10**, 16315–16326.



72 W. Boonlai, V. Tantishaiyakul and N. Hirun, *AAPS PharmSciTech*, 2018, **19**, 2103–2117.

73 B. Shriky, A. Kelly and M. Isreb, *J. Colloid Interface Sci.*, 2020, **565**, 119–130.

74 X. Zhai, Z. Cui and Y. Li, *Heliyon*, 2022, **8**, e12063.

75 Y. Dong, H. Zhuang and Y. Hao, *Int. J. Nanomed.*, 2020, **15**, 1939–1950.

76 B. Darabian, H. Bagheri and S. Mohammadi, *Prog. Biomater.*, 2020, **9**, 45–64.

77 L. Huang, Z. Zhu and D. Wu, *Carbohydr. Polym.*, 2019, **225**, 115110.

78 L. Du, L. Tong and Y. Jin, *Wound Repair Regen.*, 2012, **20**, 904–910.

79 E. Gioffredi, M. Boffito and S. Calzone, *Procedia CIRP*, 2016, **49**, 125–132.

80 C. Fu, F. Ren and Q. Zhang, *Colloid Polym. Sci.*, 2015, **293**, 2191–2200.

81 T. Li, Q. Bao and J. Shen, *Int. J. Pharm.*, 2020, **580**, 119238.

82 P. S. Sundaran, A. Bhaskaran and S. T. Alex, *J. Mater. Sci.: Mater. Med.*, 2017, **28**, 88.

83 J. Wasiak, H. Cleland and F. Campbell, *Cochrane Database Syst. Rev.*, 2013, **2013**, CD002106.

84 J. L. Soriano-Ruiz, A. C. Calpena-Campmany and M. Silva-Abreu, *Int. J. Biol. Macromol.*, 2020, **142**, 412–422.

85 M. Kim, F. Heinrich and G. Haugstad, *Langmuir*, 2020, **36**, 3393–3403.

86 V. Kant, A. Gopal and D. Kumar, *Acta Histochem.*, 2014, **116**, 5–13.

87 T. A. Kwiatkowski, A. L. Rose and R. Jung, *Am. J. Physiol.: Cell Physiol.*, 2019, **318**, C253–C262.

88 J. Zhou, G. Chen and D. Chen, *Acta Mater. Compositae Sin.*, 2022, 1–9.

89 M. J. Ansari, R. R. Rajendran and S. Mohanto, *Gels*, 2022, **8**, 454.

90 L. Tang, L. Wang and X. Yang, *Prog. Mater. Sci.*, 2020, **115**, 100702.

91 W. J. Chuang and W. Y. Chiu, *Polymer*, 2012, **53**, 2829–2838.

92 X. Lin, X. Guan and Y. Wu, *Mater. Sci. Eng., C*, 2020, **115**, 111123.

93 Y. Hu, Y. Shin and S. Park, *Polymers*, 2022, **15**, 122.

94 S. Ziane, S. Schlaubitz and S. Miraux, *Eur. Cells Mater.*, 2012, **23**, 147–160, discussion 160.

95 B. Jiang, J. C. Larson and P. W. Drapala, *J. Biomed. Mater. Res., Part B*, 2012, **100**, 668–676.

96 T. Kardan, R. Mohammadi and S. Taghavifar, *Int. J. Lower Extremity Wounds*, 2021, **20**, 263–271.

97 S. L. Chen, R. H. Fu and S. F. Liao, *Cell Transplant.*, 2018, **27**, 275–284.

98 M. H. Park, M. K. Joo and B. G. Choi, *Acc. Chem. Res.*, 2012, **45**, 424–433.

99 T. Thambi, Y. Li and D. S. Lee, *J. Controlled Release*, 2017, **267**, 57–66.

100 Z. Li and J. Guan, *Expert Opin. Drug Delivery*, 2011, **8**, 991–1007.

101 S. Chatterjee and P. C. Hui, *Polymers*, 2021, **13**, 2086.

102 R. Yang, V. Sabharwal and N. Shlykova, *JCI Insight*, 2018, **3**, e123415.

103 R. Censi, T. Vermonden and M. J. van Steenbergen, *J. Controlled Release*, 2009, **140**, 230–236.

104 H. Li, Q. Ji and X. Chen, *J. Biomed. Mater. Res., Part A*, 2017, **105**, 265–273.

105 Y. J. Dong, J. F. Liao and Z. C. Kong, *Appl. Catal., B*, 2018, **237**, 9–17.

106 M. Ma, Y. Zhong and X. Jiang, *Carbohydr. Polym.*, 2020, **236**, 116096.

107 Z. Zheng, S. Bian and Z. Li, *Carbohydr. Polym.*, 2020, **249**, 116826.

108 X. Jin, Q. Fu and Z. Gu, *Int. J. Biol. Macromol.*, 2021, **184**, 787–796.

109 X. Li, A. Li and F. Feng, *Anim. Models Exp. Med.*, 2019, **2**, 107–113.

110 Y. H. Chang, *Polymers*, 2020, **12**, 2619.

111 M. Zakerikhoob, S. Abbasi and G. Yousefi, *Carbohydr. Polym.*, 2021, **271**, 118434.

112 C. Gong, Q. Wu and Y. Wang, *Biomaterials*, 2013, **34**, 6377–6387.

113 Y. Zhao, X. Liu and X. Peng, *Int. J. Biol. Macromol.*, 2022, **216**, 475–486.

114 N. N. Mahmoud, S. Hikmat and G. D. Abu, *Int. J. Pharm.*, 2019, **565**, 174–186.

115 M. Rafie and A. Meshkini, *Eur. J. Pharm. Sci.*, 2021, **167**, 106031.

116 X. Jin, W. Zhang and J. Shan, *ACS Appl. Mater. Interfaces*, 2022, **14**, 50677–50691.

117 Y. Zhao, J. G. Liu and W. M. Chen, *Exp. Ther. Med.*, 2018, **15**, 1304–1313.

118 Y. Y. Teng, M. L. Zou and S. Y. Liu, *Front. Bioeng. Biotechnol.*, 2022, **10**, 902894.

119 P. Ge, S. Chang and T. Wang, *Nanoscale*, 2023, **15**, 644–656.

120 T. Feng, H. Wu and W. Ma, *J. Mater. Chem. B*, 2022, **10**, 6143–6157.

121 K. Zubik, P. Singhsa and Y. Wang, *Polymers*, 2017, **9**, 119.

122 Z. Lei, G. Singh and Z. Min, *Mater. Sci. Eng., C*, 2018, **90**, 159–167.

123 M. Yang, S. He and Z. Su, *ACS Omega*, 2020, **5**, 21015–21023.

124 H. Xu, S. Huang and J. Wang, *Int. J. Biol. Macromol.*, 2019, **137**, 433–441.

125 Y. J. Lin, G. H. Lee and C. W. Chou, *J. Mater. Chem. B*, 2015, **3**, 1931–1941.

126 B. Lan, L. Zhang and L. Yang, *J. Nanobiotechnol.*, 2021, **19**, 130.

127 Y. Zhou, X. L. Zhang and S. T. Lu, *Stem Cell Res. Ther.*, 2022, **13**, 407.

128 T. F. Dadkhah, I. Shabani and A. Shabani, *Carbohydr. Polym.*, 2022, **281**, 119020.

129 J. Chi, X. Zhang and C. Chen, *Bioact. Mater.*, 2020, **5**, 253–259.

130 M. Qiu, D. Chen and C. Shen, *Int. J. Mol. Sci.*, 2016, **17**, 1001.

131 X. Li, X. Ye and J. Qi, *Int. J. Nanomed.*, 2016, **11**, 3993–4009.

132 D. Zhang, Q. Ouyang and Z. Hu, *Int. J. Biol. Macromol.*, 2021, **173**, 591–606.

133 M. P. Tian, A. D. Zhang and Y. X. Yao, *Carbohydr. Polym.*, 2021, **261**, 117878.



134 E. J. Kim, J. S. Choi and J. S. Kim, *Biomacromolecules*, 2016, **17**, 4–11.

135 T. Lim, Q. Tang and Z. Zhu, *J. Biomed. Mater. Res., Part A*, 2020, **108**, 2111–2122.

136 X. Chen, C. Zhou and J. Wang, *Adv. Healthcare Mater.*, 2022, **11**, e2200546.

137 M. H. Turabee, T. Thambi and D. S. Lee, *ACS Appl. BioMater.*, 2019, **2**, 2500–2510.

138 K. Y. Chang, Y. N. Chou and W. Y. Chen, *Polymers*, 2022, **14**, 3346.

139 M. Hou, X. Wang and O. Yue, *Biomater. Adv.*, 2022, **134**, 112556.

140 D. Y. Zhu, Z. P. Chen and Z. P. Hong, *Acta Biomater.*, 2022, **143**, 203–215.

141 S. G. Park, M. X. Li and W. K. Cho, *Carbohydr. Polym.*, 2021, **260**, 117808.

142 M. Li, Y. Liang and J. He, *Chem. Mater.*, 2020, **32**, 9937–9953.

143 R. Yu, M. Li and Z. Li, *Adv. Healthcare Mater.*, 2022, **11**, e2102749.

144 X. Peng, C. Ding and Y. Zhao, *Front. Bioeng. Biotechnol.*, 2022, **10**, 831007.

145 D. Y. Zhu, X. J. Chen and Z. P. Hong, *ACS Appl. Mater. Interfaces*, 2020, **12**, 22534–22542.

146 Z. Li, J. Huang and Y. Jiang, *Adv. Healthcare Mater.*, 2022, **11**, e2201878.

147 T. Maver, S. Hribernik, T. Mohan and K. Stana-Kleinschek, *RSC Adv.*, 2015, **5**, 77873–77884.

148 M. T. Cook, P. Haddow and S. B. Kirton, *Adv. Funct. Mater.*, 2020, **31**, 2008123.

149 T. Sarwan, P. Kumar and Y. E. Choonara, *Front. Mater.*, 2020, **7**, 1–17.

150 W. Treesuppharat, P. Rojanapanthu and C. Siangsanoh, *Biotechnol. Rep.*, 2017, **15**, 84–91.

151 T. Ak and I. Gülcin, *Chem.-Biol. Interact.*, 2008, **174**, 27–37.

152 H. Liu, J. Liu, C. Qi, Y. P. Fang, L. Zhang, R. X. Zhuo and X. L. Jiang, *Acta Biomater.*, 2016, **35**, 228–237.

153 E. Redwan, *Hum. Antibodies*, 2007, **16**, 137–158.

154 J. Guo, H. Hu and J. Gorecka, *Am. J. Physiol.: Cell Physiol.*, 2018, **315**, C885–C896.

155 J. Y. Li, K. K. Ren and W. J. Zhang, *Stem Cell Res. Ther.*, 2019, **10**, 247.

156 C. Yue, Z. Guo and Y. Luo, *Stem Cells Int.*, 2020, **2020**, 7430968.

157 S. A. Eming, P. Martin and M. Tomic-Canic, *Sci. Transl. Med.*, 2014, **6**, 265sr6.

158 P. Zhu, *Dynamic Changes of MMP-9 and TIMP-1 in Skin Wound Healing of Diabetic Rats*, Sun Yat-sen University, 2006.

159 P. Calvert, *Adv. Mater.*, 2009, **21**, 743–756.

160 Y. Yang and M. W. Urban, *Chem. Soc. Rev.*, 2013, **42**, 7446–7467.

161 M. H. Turabee, T. Thambi and J. S. Lym, *Biomater. Sci.*, 2017, **5**, 837–848.

162 L. Han, L. Yan and K. Wang, *NPG Asia Mater.*, 2017, e372.

163 H. W. Chien, H. Y. Lin and C. Y. Tsai, *Polymers*, 2020, **12**, 2008.

164 H. W. Chien and T. H. Chiu, *Eur. Polym. J.*, 2020, **130**, 109654.

165 C. K. Sen, G. M. Gordillo and S. Roy, *Wound Repair Regen.*, 2009, **17**, 763–771.

166 H. Yaseen and M. Khamaisi, *Cell. Immunol.*, 2020, **356**, 104154.

167 D. J. Magliano and E. J. Boyko, *IDF Diabetes Atlas*, International Diabetes Federation, Brussels, 2021.

168 R. M. Stoekenbroek, J. Lokin and M. M. Nielsen, *Diabetologia*, 2017, **60**, 1271–1275.

169 R. Frykberg and J. Banks, *Adv. Wound Care*, 2015, **4**, 560–582.

170 M. Phillipson and P. Kubes, *Trends Immunol.*, 2019, **40**, 635–647.

171 S. Chen, J. Shi and M. Zhang, *Sci. Rep.*, 2015, **5**, 18104.

172 Y. Lu, J. Deng and J. Wang, *Chin. J. Burns*, 2020, **36**, 13.

173 K. Lin, L. Fan and S. Wang, *Chin. J. Diabetes*, 2017, **25**, 449–453.

174 J. H. Kim, B. Yang and A. Tedesco, *Sci. Rep.*, 2019, **9**, 19318.

175 W. Cao and C. Gao, *Chin. J. Burns*, 2021, **37**, 9.

176 Z. Xu, Y. Liu and R. Ma, *ACS Appl. Mater. Interfaces*, 2022, **14**, 14059–14071.

177 Y. Zhu, R. Hoshi and S. Chen, *J. Controlled Release*, 2016, **238**, 114–122.

178 O. Rezvani, E. Shabbak and A. Aslani, *Ostomy/Wound Manag.*, 2009, **55**, 22–28.

179 M. C. Ribeiro, V. Correia and F. Silva, *Eur. J. Pharm. Sci.*, 2020, **150**, 105330.

180 L. I. Moura, A. M. Dias and E. C. Leal, *Acta Biomater.*, 2014, **10**, 843–857.

181 L. Fang, H. Wu and X. Li, *Int. J. Endocrinol.*, 2022, **2022**, 7847011.

182 N. Tian, J. Wei and Y. Li, *J. Colloid Interface Sci.*, 2020, **566**, 69–78.

183 Y. Dong, M. Cui and J. Qu, *Acta Biomater.*, 2020, **108**, 56–66.

184 A. Markiewicz-Gospodarek, M. Koziol and M. Tobiasz, *Int. J. Environ. Res. Public Health*, 2022, **19**, 1338.

185 K. S. Romanowski, J. Carson and K. Pape, *J. Burn Care Res.*, 2020, **41**, 1129–1151.

186 Y. Huangfu, S. Li and L. Deng, *ACS Appl. Mater. Interfaces*, 2021, **13**, 59695–59707.

187 S. Chakrabarti, B. Mazumder and J. Rajkonwar, *Sci. Rep.*, 2021, **11**, 3357.

188 X. Fu, Z. Shen and Y. Chen, *Lancet*, 1998, **352**, 1661–1664.

189 Z. Xiao, X. Zheng and Y. An, *Biomater. Sci.*, 2021, **9**, 882–891.

190 Y. Zhang, W. He and S. Zhang, *Int. J. Mol. Sci.*, 2022, **23**, 12716.

191 L. M. Geever, K. E. Mcevoy and J. I. Ngadaonye, *Int. J. Polym. Mater.*, 2014, **63**, 873–883.

192 Z. Liu, W. Tang and J. Liu, *Bioact. Mater.*, 2023, **20**, 610–626.

193 S. D. Li, N. Chen, X. P. Li, Z. P. Xie, Z. Y. Ma, J. Zhao, X. Hou and X. B. Yuan, *Adv. Funct. Mater.*, 2020, **30**, 2000130.

194 X. Zhao, Y. Liang and Y. Huang, *Adv. Funct. Mater.*, 2020, **30**, 1910748.

195 Y. Liang, Z. Li and Y. Huang, *ACS Nano*, 2021, **15**, 7078–7093.



196 Y. Ma, J. Yao and Q. Liu, *Adv. Funct. Mater.*, 2020, **30**, 1001820.

197 Y. Liang, H. Xu and Z. Li, *Nano-Micro Lett.*, 2022, **14**, 185.

198 N. Bayat, Y. Zhang and P. Falabella, *Sci. Transl. Med.*, 2017, **9**, eaan3879.

199 P. Thangavel, B. Ramachandran and S. Chakraborty, *Sci. Rep.*, 2017, **7**, 10701.

200 H. Niu, H. Li and Y. Guan, *Bioact. Mater.*, 2022, **18**, 104–115.

201 Q. Zhao, J. Liu and S. Liu, *ACS Appl. Mater. Interfaces*, 2022, **14**, 46224–46238.

202 W. Zhou, Z. Duan and J. Zhao, *Bioact. Mater.*, 2022, **17**, 1–17.

203 J. Yang, Z. Chen and D. Pan, *Int. J. Nanomed.*, 2020, **15**, 5911–5926.

204 Z. Chen, B. Zhang and J. Shu, *J. Biomed. Mater. Res., Part A*, 2021, **109**, 1418–1428.

205 J. Wang, B. Zhao and L. Sun, *PLoS One*, 2022, **17**, e0279727.

206 M. H. Kim, H. Park and H. C. Nam, *Carbohydr. Polym.*, 2018, **181**, 579–586.

207 X. Liu, H. Gan and C. Hu, *Int. J. Nanomed.*, 2019, **14**, 289–300.

208 H. M. Abdel-Mageed, E. A. A. Abd and R. B. Abdel, *3 Biotech*, 2022, **12**, 73.

209 J. L. Soriano, A. C. Calpena and M. J. Rodríguez-Lagunas, *Pharmaceutics*, 2020, **13**, 8.

210 C. Y. Tsai, L. C. Woung and J. C. Yen, *Carbohydr. Polym.*, 2016, **135**, 308–315.

211 P. Zhou, X. Li and B. Zhang, *BioMed Res. Int.*, 2019, **2019**, 5768285.

212 H. Yim, H. T. Yang and Y. S. Cho, *Burns*, 2014, **40**, 1642–1649.

213 A. Deng, Y. Yang and S. Du, *Mater. Sci. Eng., C*, 2021, **119**, 111555.

214 P. Makvandi, G. W. Ali and S. F. Della, *Carbohydr. Polym.*, 2019, **223**, 115023.

215 E. Zhang, Q. Guo and F. Ji, *Acta Biomater.*, 2018, **74**, 439–453.

216 S. Lin, L. Pei and W. Zhang, *Mater. Sci. Eng., C*, 2021, **130**, 112450.

217 N. Lei, C. Gong and Z. Qian, *Nanoscale*, 2012, **4**, 5686–5693.

218 X. Dong, F. Yao and L. Jiang, *J. Mater. Chem. B*, 2022, **10**, 2215–2229.

