

PAPER

[View Article Online](#)
[View Journal](#) | [View Issue](#)Cite this: *Nanoscale Adv.*, 2023, 5, 3729

Seed free synthesis of polyethylene glycol stabilized gold nanoprisms exploiting manganese metal at low pH†

Kanika Bharti,  Md Azimuddin Sk and Kalyan K. Sadhu *

Manganese powder with a suitable potential ($E^0_{\text{Mn}/\text{Mn}^{2+}}$, -1.19 V) has never been investigated for the reduction of Au^{3+} ($E^0_{\text{Au}/\text{AuCl}_4^-}$, 1.00 V). In this study, we have utilized $E^0_{\text{Mn}/\text{Mn}^{2+}}$ and low pH dependent $E^0_{\text{Mn}^{2+}/\text{MnO}_2}$ for the polyethylene glycol stabilized gold nanoprism synthesis by reducing AuCl_4^- in the presence of thiol terminated polyethylene glycol as the stabilizing agent. The synthetic methodology for gold nanoprisms has been optimized by pH and Cl^- ion combination. Time dependent absorbance studies have been conducted to demonstrate the role of various reaction parameters such as the stabilizing agent, HCl concentration, temperature, and Mn metal. The synthesized gold nanoprism has been further utilized as a seed for nucleic acid and selected amino acid mediated edge and surface growth, respectively.

Received 2nd May 2023
Accepted 16th June 2023

DOI: 10.1039/d3na00292f

rsc.li/nanoscale-advances

1. Introduction

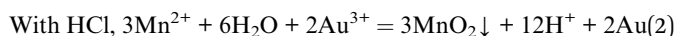
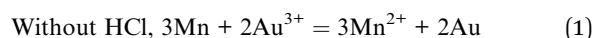
The journey of Mn metal as a reagent started 40 years ago through carbon–carbon bond formation reaction.¹ Within these 40 years, few Mn metal-based attempts were made directly for chemical syntheses such as Barbier and Reformatsky like reactions,² and adducts on fullerene.³ However, Mn metal as an additive to titanium has been known for 50 years^{4,5} and is currently also pursued in nickel^{6–8} catalyzed organic synthesis. Reactions exclusively with Mn metal are less in number as an oxide layer on the metal surface lowers the reducing ability of the Mn metal. The reducing ability was increased in freshly prepared Mn powder from the reduction on Mn^{2+} salts either with lithium naphthalenide^{9,10} or potassium graphite¹¹ or by the addition of catalytic amounts of PbCl_2 and Me_3SiCl ¹² or LiCl , PbCl_2 and InCl_3 additives.¹³ In this current decade, the mechanochemical approach using ball milling for Mn metal has drawn significant attention for organic transformations.^{14–16} The limited performance of Mn metal in synthetic chemistry restricts its utilization in the reduction of Au^{3+} . To the best of our knowledge, there is no report of gold nanoparticle synthesis using Mn metal powder.

Seed mediated growth of gold nanoprisms has drawn significant attention due to their implication in biomedical applications.¹⁷ In this study, kinetically controlled growth has been found to be important. A very recent study has also focused on the Au_{56} nanocluster of a seed sized gold nanoprism.¹⁸ To date synthesis of nanoprisms without seeds remains

challenging due to the parallel formation of rough gold nanoparticles.

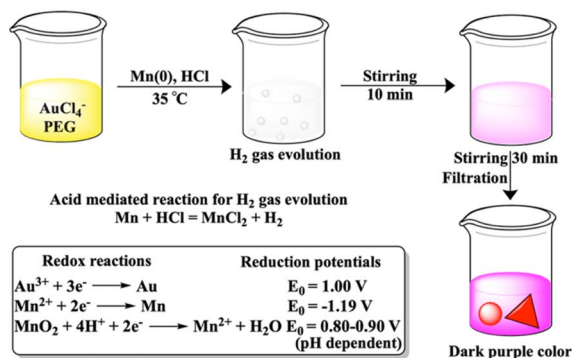
Selective growth along the face, edge and corners was observed by controlling the reduction potential of the reducing agent and tuning the concentration of the reducing agent and stabilising agent.¹⁹ Efforts have been made to understand the synthesis mechanism of established traditional methods, overcome their limitations and modify the synthesis methodologies in order to improve the yield.²⁰ Seed mediated²¹ and seedless synthetic methodologies²² are known for the synthesis of gold nanoprisms.

The growth mechanism of anisotropic nanoparticles has been influenced by reduction potential, which is indirectly governed by variations in seed concentration, pH, halide concentration, reducing agent, and surface passivating stabilizing agents.²³ In our previous reports with Fe and Zn, seed-mediated syntheses produced gold-iron oxide²⁴ or gold-zinc oxide nanocomposites.²⁵ In this study, we aimed to synthesize nanoparticles without the incorporation of the reducing metal in the final nano-structure. Here, we have utilized a seed free methodology for the gold nanostructure synthesis. In order to achieve this, we have explored three half reactions for gold nanoparticle synthesis using Mn metal (eqn (1) and (2) and Scheme 1). Thiol modified PEG can reduce Au^{3+} via thiol reduction and act as a stabilizing agent for the nanoparticle. Kinetically optimized condition produces gold nanoprisms as the major product by varying three reagents Mn metal, thiol containing polyethylene glycol (PEG) and pH of the solution for controlling Mn metal and Mn^{2+} in solution.



Department of Chemistry, Indian Institute of Technology Roorkee, Roorkee 247667, Uttarakhand, India. E-mail: sadhu@cy.iitr.ac.in

† Electronic supplementary information (ESI) available: Supporting figures and tables. See DOI: <https://doi.org/10.1039/d3na00292f>



Scheme 1 PEG-SH stabilized synthesis of gold nanoprisms through Mn metal-based reduction of Au³⁺.

2. Results and discussion

The reduction potential *vs.* pH graph for the Mn/Mn²⁺ couple is independent of proton concentration within acidic to neutral pH.²⁶ Considering the first two half reactions for Au³⁺ and Mn⁰, 1.5 equivalents of Mn metal is ideal for the gold nanoparticle formation. However, simple addition of 2 equivalents of Mn metal to the AuCl₄⁻ aqueous solution shows no generation of surface plasmon resonance (SPR) peak in absorbance, which confirms no gold nanoparticle formation. Prior to checking the role of the stabilizing agent in the gold nanoparticle formation *via* Mn metal based reduction, 0.5 equivalents of PEG (Au : PEG = 1 : 0.5) has been added to the Au³⁺ solution. The SPR peak has been generated at 630 nm within 10 min (Fig. S1a and S2, ESI†). This is possible due to the thiol-based reduction²⁷ of Au³⁺ to Au⁰ and nucleation takes place in the presence of PEG. However, the SPR peak intensity becomes broad with time (Fig. S1b, ESI†), which suggests the aggregation of the nucleated particles. The absorption nature remains unchanged in the presence of HCl. However, no reaction takes place by Mn and HCl combination in the absence of PEG (Fig. S3, ESI†).

Addition of 2.0 equivalents of Mn metal in the presence of PEG without any HCl generates a stable peak at 575 nm due to the formation of gold nanoparticles (Fig. S4, ESI†). The intensity of the absorption peak has not been affected much by varying the PEG or Mn concentration in the solution. The pH independent Mn/Mn²⁺ couple along with PEG is responsible for the reduction of Au³⁺ to Au⁰ in the absence of HCl (eqn (1)). In order to inspect pH dependent behavior of redox reaction between Mn and Au(III),²⁴ this has been added to the solution. Addition of 10 equivalents of HCl (Au : HCl = 1 : 10) to the reaction mixture shows the generation of effervescence as expected due to liberation of hydrogen gas after reaction with Mn metal (Scheme 1). The control reaction with separate treatment of hydrogen gas to the mixture of Au³⁺ and PEG does not show any SPR peak even after 1 h.

The SPR peak intensity at 536 increases significantly within 10 min (Fig. S5, ESI†). The time dependent studies for PEG reduction only and combination of all the three reagents PEG, Mn and HCl show the faster rate of SPR peak formation in the

second case (Fig. S6, ESI†). Interestingly, an additional broad peak has been observed in the NIR range. Monitoring the reaction for 6 h shows clear declining nature of both the peaks in the visible and NIR regions (Fig. S7, ESI†). The additional broad peak in the NIR range is a characteristic peak of gold nanoprisms due to in-plane dipole mode and the peak position depends on the prism thickness, edge length and dielectric environment.^{28,29} Kitaev *et al.* have reported gold nanoprisms with a tuneable shift of the same absorption peak within the visible to NIR range based on thiol- and halide-mediated reduction with hydrogen peroxide.³⁰

The reduction of Au³⁺ in the presence of Mn, HCl and PEG-SH develops dual absorption peaks, which are stable even after stirring the reaction mixture for 3 h (Fig. S5, ESI†). No SPR peak has been developed under the same reaction conditions in the absence of HCl and PEG as the stabilizing agent. In order to check the efficiency of the reduction in basic medium, we have also performed the reaction in the presence of NaOH. Absorbance spectra show a single SPR peak at 530 nm and the transmission electron microscopic (TEM) image after filtration confirms the formation of spherical gold nanoparticles at higher pH (Fig. S8 and S9, ESI†). In order to obtain a stable gold nanostructure with the maximum possible yield from the Mn metal-based reduction, concentrations of three reagents Mn, PEG and HCl have been further optimized.

In order to monitor the nanoparticle architecture in the presence of 10 equivalents of HCl, TEM images have been taken from the filtrate after 6 h of the reaction (Fig. S10, ESI†). TEM analysis confirms the polycrystalline behaviour of both larger spherical gold nanoparticles of 24.0 ± 1.4 nm diameter and smaller spherical gold nanoparticles of 10.9 ± 0.3 nm with fringe patterns (Fig. S11 and S12, ESI†).³¹ Increasing the HCl concentration from 14 to 32 equivalents, the SPR and NIR peaks have been found to be stable for long days. The pH working range for these stable peaks varies within 1.50 to 2.85, where the Mn²⁺/MnO₂ redox couple shows a reduction potential of 0.8 to 0.9 V.^{26,32}

TEM images in the presence of different concentrations of HCl confirm both the formation of spherical gold nanoparticles and their aggregation and nanoprisms (Fig. 1 and S13–S15, ESI†). These TEM images suggest that two parallel reactions have proceeded continuously within the solution. In one path spherical gold nanoparticles form with a SPR peak and their aggregation generates the NIR peak. On the other hand, a parallel process leads to the formation of 2D gold nanoprisms having dual absorption peaks. The height of gold nanoprisms has been confirmed from atomic force microscopic (AFM) images and found to be ~10 nm (Fig. 1e and f), which is significantly lower than the edge dimension of the gold nanoprism. These two reaction pathways have been monitored through changes in absorption intensities at 536 nm and 850 nm for 3 h. The 536 nm absorption has originated from the spherical gold nanoparticles and gold nanoprisms, whereas the 850 nm absorption is attributed to gold nanoprisms and aggregation of spherical gold nanoparticles. Similar absorbance spectra with dual peaks have been reported by Kitaev *et al.* for the thiol mediated reduction synthetic approach.³⁰ The 850 nm



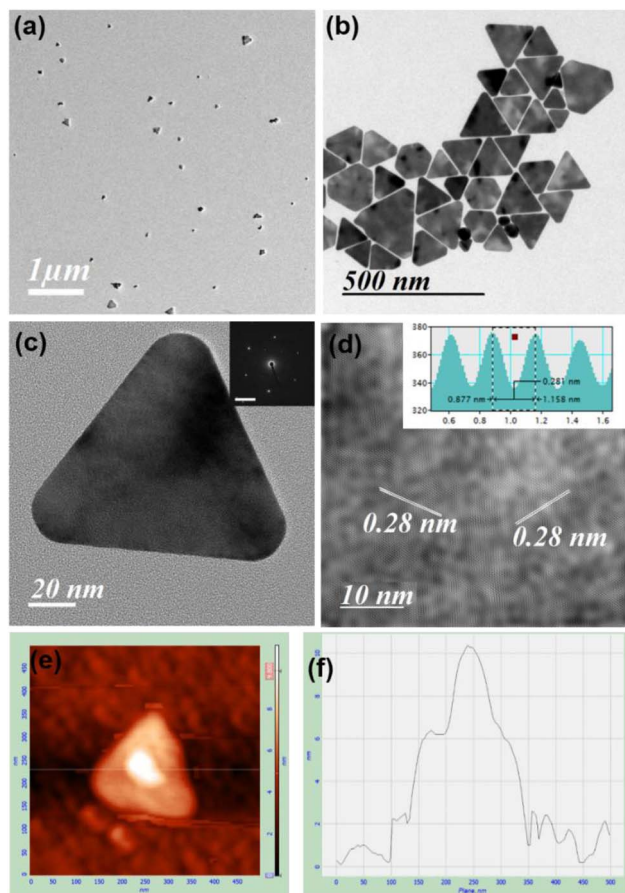


Fig. 1 TEM image (a) before and (b) after purification of gold nanoprisms obtained from the reaction between Au : Mn : PEG : HCl in 1 : 2 : 0.5 : 26 after 30 min; (c) HRTEM and SAED (inset) images and (d) fringe lines of nanoprisms and (e and f) AFM image and its analysis for the thickness of nanoprisms. Scale bar: (a) 1000 nm, (b) 500 nm, (c) 20 nm and (d) 10 nm.

absorbance in the NIR range is a characteristic peak for gold nanoprisms due to in-plane dipole mode²⁹ or aggregation of gold nanoparticles.³³ These kinetic data for 3 h have been followed as a combination of two factors of the well explored KJMA model^{34–37} and fitted as per eqn (1).

$$A = C_1 (1 - e^{k_1 t^{n_1}}) + C_2 (1 - e^{k_2 t^{n_2}}) \quad (3)$$

where C_1 and C_2 are the proportionality constants, k_1 and k_2 are the apparent rate constants, and n_1 and n_2 are the Avrami constants for the two parallel sets of reactions in the solutions. Fitting of the reactions with eqn (1) shows n_1 and n_2 values close to 2 and 1, respectively (Fig. S16 and Table S1, ESI†). For comparison of all the kinetic data with the same expression, eqn (1) has been modified as eqn (2) with $n_1 = 2$ and $n_2 = 1$.

$$A = C_1 (1 - e^{k_1 t^2}) + C_2 (1 - e^{k_2 t}) \quad (4)$$

HCl concentration dependent kinetics fitting (Fig. 2) with eqn (2) shows almost constant k_1 values for 536 nm data (Table 1) within 14–26 equivalents of HCl. However,

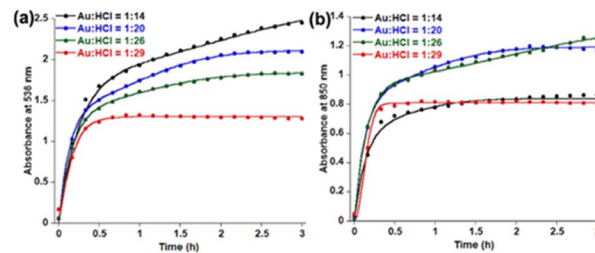


Fig. 2 Fitting of time dependent absorbance data at (a) 536 nm and (b) 850 nm in the presence of varying HCl concentration using eqn (2).

a significant enhancement of k_1 values has been observed by increasing the HCl concentration to 29 equivalents. In the case of 850 nm kinetics, the k_1 value decreases within 14–26 equivalents of HCl and has significantly enhanced at 29 equivalents HCl. On the contrary, k_2 values are of a similar order for both 536 nm and 850 nm in the presence of 14–29 equivalents of HCl. Comparing these kinetic data with TEM images, it is clear that $n_1 = 2$ deals with the spherical gold nanoparticles and $n_1 = 1$ is associated with gold nanoprism formation. The k_1 parameter for 850 nm decreases with the decreasing trend of aggregation of spherical gold nanoparticles in TEM. On the other hand, the k_2 parameter decreases when a smaller number of nanoprisms have been observed in TEM. A recent review on the KJMA model³⁴ suggests that the nanoprism formation in our case has been guided through diffusion-controlled growth without having any nucleation. Increasing the concentration of protons from 10 equivalents to 32 equivalents results in protonation of oxygen atoms in PEG, which is responsible for the continuous increase of ζ potential (Fig. S17, ESI†).

In order to obtain nanoprisms as the predominant product the reaction with 26 equivalents of HCl has been filtered after 30 min, where kinetic data changes its pattern. However, spherical gold nanoparticle formation cannot be avoided in this condition (Fig. 1a). The gold nanoprism and few planar twinned prisms have been collected as the predominant product (Fig. 1b and S18, S19, ESI†) after few rounds of washing. The high-resolution TEM (HRTEM) for gold nanoprisms confirms the curvature around the tips and presence of fringe lines with 0.28 nm interplanar distance (Fig. 1c and d). Lowering down the temperature from 35 °C to 10 °C, the k_2 value significantly decreases and the k_1 value remains almost constant (Fig. S20 and Table S2, ESI†). This trend is confirmed *via* TEM imaging study, where mostly spherical gold nanoparticles have been formed at 10 °C (Fig. S21, ESI†).

Initial slope calculations from 10 min reactions show a decreasing trend of rate with increasing HCl concentration (Fig. S22, ESI†) due to pH dependent non-elementary reaction³⁸ for nanoparticle formation. In order to check the end product of Mn in the reaction, the solution and insoluble precipitate have been analysed carefully. The point and area wise elemental mapping of gold nanoprisms in TEM confirms no significant incorporation of Mn in the solution (Fig. 3a–e) after filtration. XPS analysis of the insoluble product confirms the peaks at 641.9 eV and 653.7 eV for Mn 2p_{3/2} and 2p_{1/2}, respectively, due to Mn⁴⁺ oxidation state (Fig. 3f).³⁹ Survey scan of XPS confirms the

Table 1 Calculated rate constants from absorbance data in Fig. 2

Equiv. of HCl	536 nm		850 nm	
	Spherical nanoparticle formation	Nanoprism formation	Aggregation of spherical nanoparticle	Nanoprism formation
	k_1 (h^{-2})	k_2 (h^{-1})	k_1 (h^{-2})	k_2 (h^{-1})
14	0.14	3.89	1.11	6.67
20	0.58	7.19	0.72	7.34
26	0.51	5.95	0.16	6.51
29	23.31	6.29	35.97	6.82

presence of MnO_2 and trace amount of PEG in the precipitate (Fig. S23, ESI†).

Addition of HCl not only makes the solution acidic, but also introduces more chloride ions in the solution. The pH has been reported to control the crystallographic orientation and growth.⁴⁰ At lower pH, growth is predominating over nucleation. However, at very low pH reduction of Au^{3+} to Au^0 has been inhibited.⁴¹ In our case, anisotropic gold nanoparticles have been formed in the acidic medium after the growth of the nucleated particle with time. In addition to controlling the pH of the solution by HCl, Cl^- ion plays an important role through Au^+ -halide complexes.⁴² The Mn based reductions have been checked with other acids, such as HNO_3 , H_2SO_4 and HBr . In the presence of 26 mM HNO_3 and H_2SO_4 the formation of isotropic gold nanoparticles as major products is confirmed in TEM images (Fig. S24 and S25, ESI†). Cl^- ion from HAuCl_4 in all these cases is responsible for few gold nanoprism formation. In the presence of 11 mM or higher HBr , the SPR peak has been destabilized (Fig. S26, ESI†). At 8 mM HBr concentration, both the spherical and prism shaped nanoparticles are observed after the synthesis (Fig. S27, ESI†). This study confirms the role of halide ions in the synthesis of anisotropic nanoparticles, similar to another report.⁴³

In order to have better understanding of the growth mechanism, HRTEM images have been monitored at different time intervals. The TEM image after performing the reaction for 10 minutes shows the aggregated gold nanoparticles (Fig. 4a). Only a few nanoprisms have been observed at the same TEM grid. Aggregation and coalescence driven growth pathway⁴⁴ have

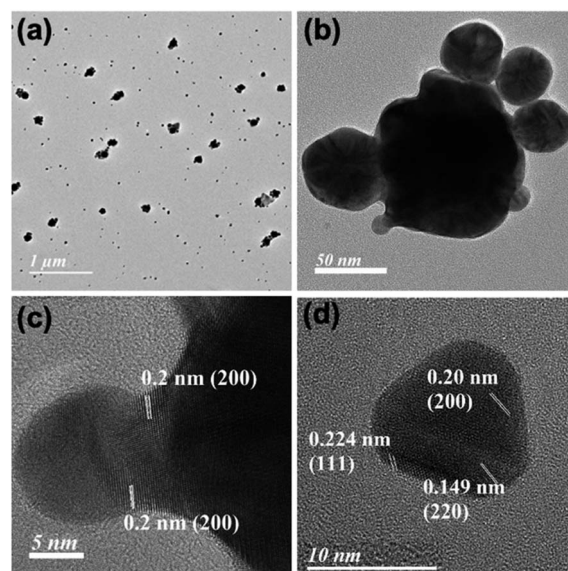


Fig. 4 TEM analysis just after 10 min of the reaction showing (a) aggregates, (b) the formation of the spherical nucleated particle, (c) their merging assembly through (200) planes of the gold nanoparticle and (d) nanoprism with (200) and (111) planes along the face and edge, respectively. Scale bar: (a) 1 μm , (b) 50 nm, (c) 5 nm, and (d) 10 nm.

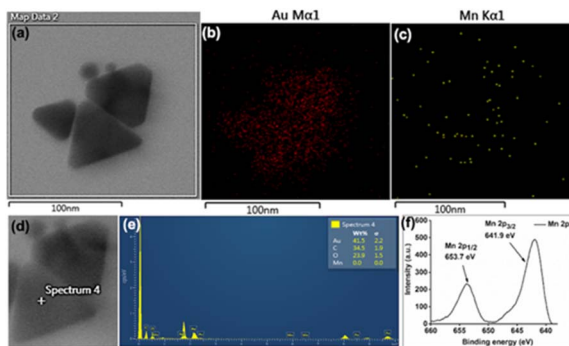
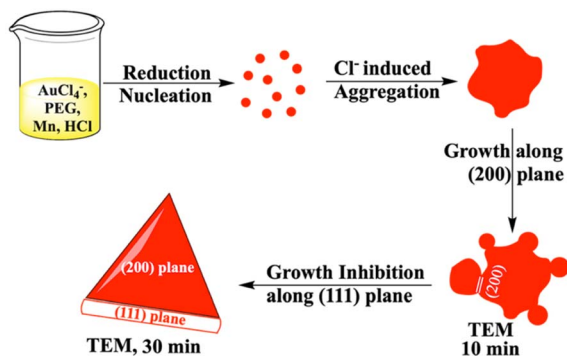


Fig. 3 (a–c) Area wise mapping and (d and e) point wise EDX for Au and Mn in gold nanoprisms; (f) XPS data for Mn obtained from the residue after 3 h of reaction. Scale bar: (a–c) 100 nm.

been clearly reflected in the TEM image (Fig. 4b). HRTEM analysis suggests oriented attachment along (200) crystallographic planes (Fig. 4c) as observed in another study by us.⁴⁵ The growth takes place by assembling and merging⁴⁶ and finally transforming into gold nanoprisms after continuing the reaction for 30 minutes (Scheme 2). The HRTEM of gold nanoprisms shows the presence of the (111) plane along the edges and the (200) plane along the surface (Fig. 4d and 5a).

The equivalent of PEG has been optimized by monitoring 536 nm and 850 nm absorbance in the presence of 0.5–1.2 equivalents of PEG with respect to Au^{3+} amount in the reaction mixture (Fig. S28, ESI†). Similar to the HCl variation, k_1 and k_2 values at both the wavelengths have been compared (Table S3, ESI†) and 0.6 equivalents of PEG has been optimized for nanoprism formation. HRTEM analysis of nanoprisms shows hexagonal close packed *hcp* (2H) unit cells with AB–AB type packing (Fig. 5).^{47–49} The SAED pattern (Fig. 5b) has been found to be similar to experimental data observed for a cubic Au cell and a hexagonal cell.⁴⁹ The twinned structure provides





Scheme 2 Mechanism of gold nanoprism synthesis from Mn based reduction in the presence of PEG and HCl.

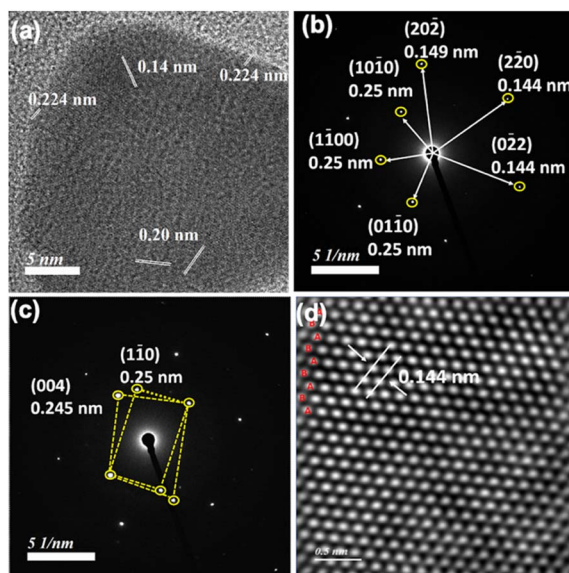


Fig. 5 (a) HRTEM, (b) SAED, (c) twinned structure and (d) high-angle annular dark field (HAADF) images showing *hcp* (2H) unit cells with AB–AB stacking for nanoprisms obtained from Au : Mn : PEG : HCl in 1 : 2 : 0.6 : 26 after 30 min. Scale bar: (a) 5 nm and (d) 0.5 nm.

a different SAED pattern due to its different crystal orientation. The interplanar distance for this packing has been found to be 0.14 nm, which arises due to (022) planes.⁵⁰ SAED patterns confirm the presence of the twinned structure.^{51,52} In the presence of 1.2 equivalents of PEG, the number of nanoprisms has significantly dropped down and small nanoclusters have been observed around the corner of the nanoprisms (Fig. S29, ESI†).

An earlier report suggests that there is a strong influence of pH as well as halide on the nanoprism morphology.⁵³ We have also observed that the optimized amount of HCl concentration is required to balance the pH and halide concentration for the maximum yield of nanoprisms. XRD analysis (Fig S30, ESI†) of the gold nanoprism obtained by varying HCl concentration shows similar planes (111), (002), (022), (311), and (222) corresponding to the *fcc* structure. Similar to the observed HAADF data (Fig. 5d), we have also observed (002 h) and (101 h) planes corresponding to the *hcp* structure reported⁴⁷ for gold nanoparticles.

Similar to HCl variation, the initial rate of the reaction decreases with increasing concentration of PEG (Fig. S31, ESI†). Changing PEG (Mn = 2000) with other sulphur containing ligands such as terminated thiol in polystyrene, lipoic acid, oxidized and reduced glutathione, 3-mercaptopropanoic acid, 6-mercaptoheptanoic acid and other PEG (Mn = 800 and 6000) shows the generation of prominent SPR peaks in few cases. However, the characteristic dual peaks due to nanoprisms are missing for these ligands (Fig. S32, ESI†).

In order to check the role of Mn, Au : Mn has been varied from 1 : 1.5 to 1 : 20. A minimum of 1.5 equivalents of Mn has been introduced for reducing Au³⁺ to Au⁰. To overcome the poor stability of dual peaks by reduction with 1.5 equivalents of Mn, 2 equivalents of Mn metal has been added. A further increase up to 20 equivalents only increases the amount of spherical gold nanoparticles in the solution (Fig. S33–S35, ESI†). More interestingly, unlike with HCl or PEG variations, the initial rate of SPR peak formation does not vary by increasing Mn metal concentration (Fig. S36, ESI†).

Rough surface gold nanotriangles with abundant hotspots on their surface show better SER performance as compared to gold nanotriangles with a smooth surface.⁵⁴ We have planned to utilize the nanoprism as a seed for further growth reaction with a standard reducing agent, such as hydroxyl amine. The growth reactions of gold nanoprisms were carried out in the presence of nucleic acids and amino acids with positively charged side chains. In contrast to the preferred ligand binding at the tip of the

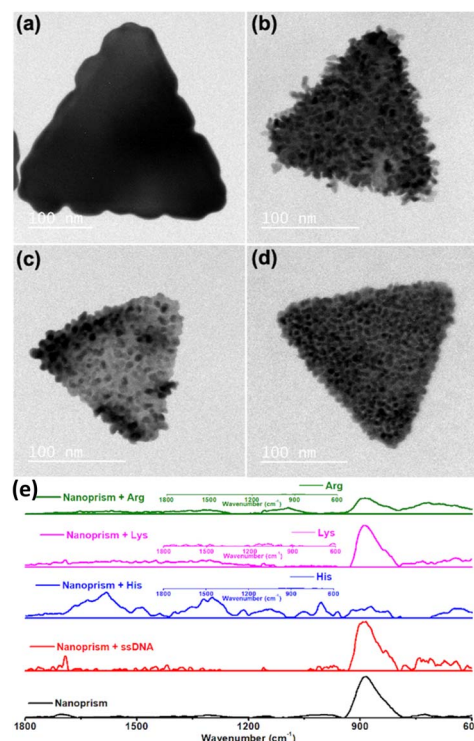
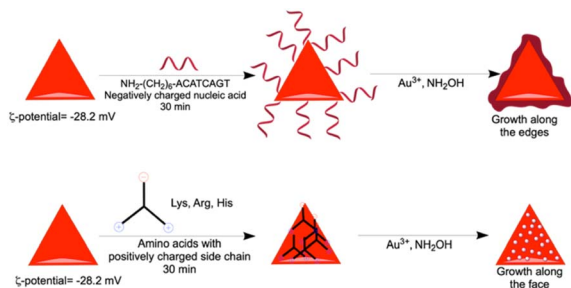


Fig. 6 Growth of gold nanoprisms in the presence of (a) amine modified single strand nucleic acid, (b) His, (c) Lys and (d) Arg; (e) Raman spectra of the nanoprism before and after growth reactions. Scale bar: (a–d) 100 nm.





Scheme 3 Nanoprism edge and face growth mechanism for nucleic acid and amino acids with positive charge, respectively.

nanoprism,⁵⁵ our studies show that growth of gold nanoprisms takes place around the edges or flat surface in the presence of an amine modified single strand nucleic acid sequence ($\text{NH}_2\text{-(CH}_2\text{)}_6\text{-ACATCAGT}$) or amino acids (His, Lys and Arg) with positively charged side chains, respectively (Fig. 6a–d). After the growth reactions, only His shows almost no shift of visible and NIR peaks except enhanced absorbance (Fig. S37–S40, ESI†). The TEM image for His after growth shows formation of longer particles on the prism surface. This overall unique growth pattern in the case of His has been selectively reflected in Raman spectroscopy (Fig. 6e) for the characteristic τ tautomer and π tautomer with a hydrated or dehydrated amide around 1650 cm^{-1} .⁵⁶

In order to find out the mechanism of different growth reactions at the edge and face, we have carried out the ζ -potential measurement of the purified gold nanoprism. In order to avoid the repulsion, the edge of the negatively charged gold nanoprism (ζ -potential -28.2 mV) having less surface area of the (111) plane has been targeted by the negatively charged nucleic acids during incubation (Scheme 3). On the other hand, the face of the nanoprism with more surface area of the (200) plane has been targeted with amino acids having positively charged side chains (Scheme 3).

3. Conclusion

In summary, Mn metal has been activated by HCl at lower pH and in combination with PEG, AuCl_4^- has been reduced to form PEG stabilized gold nanoprisms. Kinetic optimization study not only guides the better yield of nanoprisms from their competitive spherical nanoparticle synthesis, but also helps to find out the concentration window for HCl and PEG for this synthesis. Interesting morphologies are obtained after growth of the nanoprism around the edge or the surface in the presence of nucleic acid or positively charged amino acids, respectively, as ligands. Among the studied amino acids, the selective growth pattern in the presence of His clearly shows the typical amide vibrations through Raman spectroscopy.

4. Experimental details

4.1 Materials required

The chemicals gold(III) chloride trihydrate, different polymers such as poly(ethylene glycol) methyl ether thiol ($\text{Mn} = 2000, 800, \text{ and } 6000$), and polystyrene thiol terminated, 3-

mercaptopropanoic acid, and 6-mercaptohexanoic acid were purchased from Sigma-Aldrich, and trisodium citrate dihydrate was purchased from Merck chemicals. α -Lipoic acid was purchased from TCI India. Hydroxylamine hydrochloride ($\text{NH}_2\text{OH}\cdot\text{HCl}$), glutathione (oxidized and reduced forms), and Mn, Fe, Cr and Zn metal powders were purchased from SISCO Research Laboratory. Lysine, arginine and histidine were purchased from Himedia Laboratories Pvt. Ltd. All the oligonucleotides used in the study were purchased from GeneX India Bioscience Pvt. Ltd.

4.2 Characterization

All the kinetic experiments were performed using a Cary UV-Vis-Multicell Peltier (Agilent) within the $200\text{--}1100\text{ nm}$ range. The TEM images were taken using an FEI, Technai G2 20 STWIN. The ζ -potential was measured using a Zetasizer Nano ZS90 (Malvern Instruments). XPS experiments were performed with a PHI 5000 Versa Prob II, FEI Inc., and a C60 sputter gun was used for characterization. An FE-SEM (Zeiss Gemini SEM model) was used to analyze the morphological features of the nanoprism. A HRTEM with $a \geq 200\text{ kV}$ Schottky Field Emission Gun (FEG) including a High Voltage Supply Unit was used for taking high resolution TEM images of the nanoprism. HAADF, STEM detectors & imaging modes (maximum magnification up to $\geq 150\text{ Mx}$, HAADF resolution $= 0.15\text{ nm}$) used to analyse the crystal packing.

4.3 Reaction condition for time dependent absorbance studies

All the syntheses were performed in 3 mL UV cuvettes in a Cary UV-Vis spectrophotometer at 35°C and 800 rpm adjusting the final reaction volume to 2.5 mL . Poly(ethylene glycol) methyl ether thiol ($\text{Mn} = 2000$) was used for all the kinetics.

4.4 Synthesis of gold nanoprisms with varying HCl concentration

Time dependent absorbance studies were performed by varying HCl concentration from $\text{Au}:\text{HCl} = 1:14$ to $1:29$ without a seed. PEG-SH solution was prepared in water (12 mg mL^{-1}) and 260 mM HCl solution was prepared. 1 mg (0.0025 mmol) HAuCl_4 was added in water and kept stirring. After that $250\text{ }\mu\text{L}$ of PEG-SH (3 mg) solution was added, followed by different volumes of 260 mM HCl concentration stock. 0.3 mg (0.005 mmol) Mn metal powder was added finally and the final reaction volume was maintained at 2.5 mL in each case. The absorbance spectra were recorded at an interval of 10 min with continuous stirring for 360 minutes . Similarly the reaction was performed at higher pH using $\text{Au}:\text{NaOH} = 1:26$. All the TEM images have been taken from the filtrate after filtration.

4.5 Synthesis of gold nanoprisms with varying PEG-SH amount

Time dependent absorbance studies were performed by varying the PEG-SH amount from 0.50 to 1.20 equivalents amount with respect to Au^{3+} . PEG-SH solution was prepared in water (12 mg mL^{-1}) and 260 mM HCl solution was prepared. 1 mg (0.0025



mmol) HAuCl_4 was added in water and kept stirring. After that varying volume of PEG-SH solution was added, followed by 250 μL 260 mM HCl concentration from stock maintaining the final volume at 2.5 mL. The HCl concentration in the final reaction volume is 26 mM. Finally, 0.3 mg (0.005 mmol) Mn was added and absorbance spectra were recorded at an interval of 10 minutes for 360 minutes with continuous stirring. Control reactions were performed using other thiol containing stabilising agents such as α -lipoic acid, oxidised and reduced glutathione, PEG-SH (Mn = 800 and 6000), polystyrene thiol terminated, 3-mercaptopropanoic acid, and 6-mercaptophexanoic acid. The 0.0015 mmol concentration used was equivalent to 3 mg PEG-SH (Mn = 2000). All the TEM images have been taken from the filtrate after filtration.

4.6 Synthesis of gold nanoprisms with varying Au : Mn ratio

Time dependent absorbance studies were performed by varying the Au : Mn ratio (1 : 1.5, 1 : 2, 1 : 4, 1 : 8, 1 : 14, and 1 : 20). PEG-SH solution was prepared in water (12 mg mL^{-1}) and 260 mM HCl solution was prepared. 1 mg (0.0025 mmol) HAuCl_4 was added in water and kept stirring. After that 250 μL PEG-SH solution (3 mg) was added, followed by 250 μL 260 mM HCl concentration. The HCl concentration in the final reaction volume was 26 mM. Finally varying amount of Mn was added and the final reaction volume was maintained at 2.5 mL in each case. The absorbance spectra were recorded at an interval of 10 minutes for 360 minutes with continuous stirring. All the TEM images have been taken from the filtrate after filtration.

4.7 Gold nanoprism synthesis on a large scale

20 mg (0.05 mmol) HAuCl_4 was added to 40 mL water in a round bottom flask. The reaction mixture was stirred at 800 rpm and the temperature was maintained at 35 $^{\circ}\text{C}$. 5 mL (60 mg) PEG-SH solution was added to the reaction during stirring followed by 5 mL 260 mM HCl such that the HCl concentration in the reaction was 26 mM. Finally, 6 mg Mn metal powder was added to the reaction mixture. The final reaction volume was maintained at 50 mL. The reaction was continued for 30 minutes. After filtration, the filtrate was centrifuged and washed several times with water. The FE-SEM and TEM images were taken before further growth reactions.

4.8 Synthesis of gold nanoparticles with HNO_3 , H_2SO_4 and HBr

PEG-SH solution was prepared in water (12 mg mL^{-1}) and 260 mM H_2SO_4 and HNO_3 solution was prepared. 1 mg (0.0025 mmol) HAuCl_4 was added in water and kept stirring. After that 250 μL of PEG-SH (3 mg) solution was added, followed by 250 μL of 260 mM acid concentration stock. The acid concentration was 26 mM in the final reaction volume. 0.3 mg (0.005 mmol) Mn metal powder was added finally and the final reaction volume was maintained at 2.5 mL in each case. The absorbance spectra were recorded at an interval of 10 min with continuous stirring. The TEM images were taken after filtering the reaction mixture. The reaction was performed using HBr following a similar procedure using 8 mM and 11 mM final

concentrations. The absorbance spectra were recorded at an interval of 10 min with continuous stirring. The TEM image was taken after performing reaction using 8 mM HBr for 80 min.

4.9 Growth reaction performed using DNA (amine-5'-ACATCAGT-3') concentration

The gold nanoprism seed stock prepared was diluted with water such that the intensity of absorbance was kept constant at 0.45. 20 mL 200 mM hydroxylamine hydrochloride ($\text{NH}_2\text{OH}\cdot\text{HCl}$) solution was prepared with the pH adjusted to 5 and stored in a refrigerator. 300 μL gold nanoprism seed was incubated with DNA (50 nM) for 30 minutes. 3 μL 200 mM NH_2OH was added to the seed solution and vigorously stirred for 10 minutes. 5 μL of 0.8% w/v HAuCl_4 was then added to the solution. More intense purple colour was observed after the reaction and absorbance spectra were recorded.

4.10 Growth reaction performed in the presence of amino acids

300 μL gold nanoprism seed was incubated with 9 mM amino acid for 30 minutes. 3 μL 200 mM NH_2OH was added to the seed solution and vigorously stirred for 10 minutes. 5 μL of 0.8% w/v HAuCl_4 was added to the solution. More intense purple colour was observed after the reaction and absorbance spectra were recorded.

Author contributions

K. K. S. has conceived the idea and written the manuscript. K. B. performed all the experiments related to nanoprism synthesis, Raman experiment and written the manuscript. M. A. S. performed the growth of gold nanoprism in presence of amino acids. All authors discussed the results and commented on the manuscript.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

K. K. S. acknowledges the SERB-DST grant (CRG/2018/000269) for funding.

References

- 1 T. Hiyama, M. Sawahata and M. Obayashi, *Chem. Lett.*, 1983, 1237–1238.
- 2 G. Cahiez and P.-Y. Chavant, *Tetrahedron Lett.*, 1989, **30**, 7373–7376.
- 3 W. Si, X. Zhang, S. Lu, T. Yasuda, N. Asao, L. Han, Y. Yamamoto and T. Jin, *Sci. Rep.*, 2015, **5**, 13920.
- 4 M. L. H. Green and C. R. Lucas, *J. Chem. Soc., Dalton Trans.*, 1972, 1000–1003.
- 5 R. S. P. Coutts, P. C. Wailes and R. L. Martin, *J. Organomet. Chem.*, 1973, **47**, 375–382.



- 6 F. Wang, Y. Chen, W. Rao, L. Ackermann and S.-Y. Wang, *Nat. Commun.*, 2022, **13**, 2588.
- 7 R. Río-Rodríguez, L. Blanco, A. Collado, J. A. Fernández-Salas and J. Alemán, *Chem.–Eur. J.*, 2022, **28**, e202201644.
- 8 Y.-Z. Yang, Y. Li, G.-F. Lv, D.-L. He and J.-H. Li, *Org. Lett.*, 2022, **24**, 5115–5119.
- 9 S.-H. Kim, M. V. Hanson and R. D. Rieke, *Tetrahedron Lett.*, 1996, **37**, 2197–2200.
- 10 R. D. Rieke and S.-H. Kim, *J. Org. Chem.*, 1998, **63**, 5235–5239.
- 11 A. Fürstner and H. Brunner, *Tetrahedron Lett.*, 1996, **37**, 7009–7012.
- 12 K. Takai, T. Ueda, T. Hayashi and T. Moriwake, *Tetrahedron Lett.*, 1996, **37**, 7049–7052.
- 13 Z. Peng and P. Knochel, *Org. Lett.*, 2011, **13**, 3198–3201.
- 14 W. I. Nicholson, J. L. Howard, G. Margi, A. C. Seastram, A. Khan, R. R. A. Bolt, L. C. Morrill, E. Richards and D. L. Browne, *Angew. Chem., Int. Ed.*, 2021, **60**, 23128–23133.
- 15 A. C. Jones, J. A. Leitch, S. E. Raby-Buck and D. L. Browne, *Nat. Synth.*, 2022, **1**, 763–775.
- 16 R. Takahashi, P. Gao, K. Kubota and H. Ito, *Chem. Sci.*, 2023, **14**, 499–505.
- 17 S. Stangherlin, N. Cathcart, F. Sato and V. Kitaev, *ACS Appl. Nano Mater.*, 2020, **3**, 8304–8318.
- 18 Y. Song, Y. Li, M. Zhou, H. Li, T. Xu, C. Zhou, F. Ke, D. Huo, Y. Wan, J. Jie, W. W. Xu, M. Zhu and R. Jin, *Nat. Commun.*, 2022, **13**, 1235.
- 19 Y. Li, H. Lin, W. Zhou, L. Sun, D. Samanta and C. A. Mirkin, *Sci. Adv.*, 2021, **7**, eabf1410.
- 20 L. Scarabelli and L. M. Liz-Marzán, *ACS Nano*, 2021, **15**, 18600–18607.
- 21 Z. Sun, A. Umar, J. Zeng, X. Luo, L. Song, Z. Zhang, Z. Chen, J. Li, F. Su and Y. Huang, *ACS Appl. Nano Mater.*, 2022, **5**, 1220–1231.
- 22 L. Chen, F. Ji, Y. Xu, L. He, Y. Mi, F. Bao, B. Sun, X. Zhang and Q. Zhang, *Nano Lett.*, 2014, **14**, 7201–7206.
- 23 M. A. Wall, S. Harmsen, S. Pal, L. Zhang, G. Arianna, J. R. Lombardi, C. M. Drain and M. F. Kirche, *Adv. Mater.*, 2017, **29**, 1605622.
- 24 S. A. Lone, P. Sanyal, P. Das and K. K. Sadhu, *ChemistrySelect*, 2019, **4**, 8237–8245.
- 25 K. Bharti, S. A. Lone, A. Singh, S. Nathani, P. Roy and K. K. Sadhu, *Front. Chem.*, 2021, **9**, 639090.
- 26 B. Douglas, D. McDaniel and J. Alexander, *Concepts and Models of Inorganic Chemistry*, Wiley & Sons, New York, 3rd edn, 1994, Chapter 8.
- 27 S. Palmal, S. K. Basiruddin, A. R. Maity, S. C. Ray and N. R. Jana, *Chem.–Eur. J.*, 2013, **19**, 943–949.
- 28 J. E. Millstone, S. J. Hurst, G. S. Me'traux, J. I. Cutler and C. A. Mirkin, *Small*, 2009, **5**, 646–664.
- 29 L. Liu, C. A. Burnyeat, R. S. Lepsenyi, I. O. Nwabuko and T. L. Kelly, *Chem. Mater.*, 2013, **25**, 4206–4214.
- 30 S. Stangherlin, N. Cathcart, F. Sato and V. Kitaev, *ACS Appl. Nano Mater.*, 2020, **3**, 8304–8318.
- 31 S. I. Stoeva, B. L. V. Prasad, S. Uma, P. K. Stoimenov, V. Zaikovski, C. M. Sorensen and K. J. Klabunde, *J. Phys. Chem. B*, 2003, **107**, 7441–7448.
- 32 S. Nijjer, J. Thonstad and G. M. Haarberg, *Electrochim. Acta*, 2000, **46**, 395–399.
- 33 T. J. Norman, C. D. Grant, D. Magana, J. Z. Zhang, J. Liu, D. Cao, F. Bridges and A. V. Buuren, *J. Phys. Chem. B*, 2002, **106**, 7005–7012.
- 34 J. S. Blázquez, F. J. Romero, C. F. Conde and A. Conde, *Phys. Status Solidi B*, 2022, **259**, 2100524.
- 35 T. S. Sabir, D. Yan, J. R. Milligan, A. W. Aruni, K. E. Nick, R. H. Ramon, J. A. Hughes, Q. Chen, R. S. Kurti and C. C. Perry, *J. Phys. Chem. C*, 2012, **116**, 4431–4441.
- 36 S. P. Shields, V. N. Richards and W. E. Buhro, *Chem. Mater.*, 2010, **22**, 3212–3225.
- 37 P. N. Njoki, J. Luo, M. M. Kamundi, S. Lim and C.-J. Zhong, *Langmuir*, 2010, **26**, 13622–13629.
- 38 V. Onesto, F. Gentile, M. Russo, M. Villani, P. Candeloro, G. Perozziello, N. Malara, E. D. Fabrizio and M. L. Coluccio, *Langmuir*, 2019, **35**, 14258–14265.
- 39 J. Jiang and A. Kucernak, *Electrochim. Acta*, 2002, **47**, 2381–2386.
- 40 M. R. Rahman, F. S. Saleh, T. Okajima and T. Ohsaka, *Langmuir*, 2011, **27**, 5126–5135.
- 41 C. G. Yuan, C. Huo, S. Yu and B. Gui, *Physica E Low Dimens Syst Nanostruct.*, 2017, **85**, 19–26.
- 42 M. R. Langille, M. L. Personick, J. Zhang and C. A. Mirkin, *J. Am. Chem. Soc.*, 2012, **134**, 14542–14554.
- 43 J. S. DuChene, W. Niu, J. M. Abendroth, Q. Sun, W. Zhao, F. Huo and W. D. Wei, *Chem. Mater.*, 2013, **25**, 1392–1399.
- 44 B. Huang, L. Miao, J. Li, Z. Xie, Y. Wang, J. Chai and Y. Zhai, *Nat. Commun.*, 2022, **13**, 1402.
- 45 J. K. Sahu, S. A. Lone and K. K. Sadhu, *Langmuir*, 2022, **38**, 5865–5873.
- 46 J. Lee, J. Yang, S. G. Kwon and T. Hyeon, *Nat. Rev. Mater.*, 2016, **1**, 16034.
- 47 X. Huang, S. Li, Y. Huang, S. Wu, X. Zhou, S. Li, C. L. Gan, F. Boey, C. A. Mirkin and H. Zhang, *Nat. Commun.*, 2011, **2**, 292.
- 48 Z. Fan, M. Bosman, X. Huang, D. Huang, Y. Yu, K. P. Ong, Y. A. Akimov, L. Wu, B. Li, J. Wu, Y. Huang, Q. Liu, C. E. Png, C. L. Gan, P. Yang and H. Zhang, *Nat. Commun.*, 2015, **6**, 7684.
- 49 M. C. Mendoza-Ramirez, H. G. Silva-Pereyra and M. Avalos-Borja, *Mater. Charact.*, 2020, **164**, 110313.
- 50 W. Li, W. Tong, A. Yadav, E. Bladt, S. Bals, A. M. Funston and J. Etheridge, *Chem. Mater.*, 2021, **33**, 9152–9164.
- 51 X. Liu, T. H. Man, J. Yin, X. Lu, S. Q. Guo, T. Ohmura and D. H. Ping, *Sci. Rep.*, 2018, **8**, 14454.
- 52 Z. C. Canbek, R. Cortes-Huerto, F. Testard, O. Spalla, S. Moldovan, O. Ersen, A. Wisnet, G. Wang, J. Goniakowski and C. Noguera, *Cryst. Growth Des.*, 2015, **15**, 3637–3644.
- 53 T. H. Ha, H. J. Koo and B. H. Chung, *J. Phys. Chem. C*, 2007, **111**, 1123–1130.
- 54 S. Z. Qu, Y. X. Zhao, H. S. Kang, J. W. Zou, L. Ma, S. J. Ding and X. B. Chen, *ACS Omega*, 2022, **7**, 48438–48446.
- 55 A. Kim, S. Zhou, L. Yao, S. Ni, B. Luo, C. E. Sing and Q. Chen, *J. Am. Chem. Soc.*, 2019, **141**, 11796–11800.
- 56 A. Ghosh, M. J. Tucker and F. Gai, *J. Phys. Chem. B*, 2014, **118**, 7799–7805.

