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## High-yield synthesis of HMF from glucose and fructose by selective catalysis with water-tolerant rare earth metal triflates assisted by choline chloride†

Fabrizio Olivito, \* Vincenzo Algieri, Matteo Antonio Tallarida, Antonio Jiritano, Paola Costanzo, Loredana Maiuolo and Antonio De Nino \*

The conversion of naturally occurring organic substances into value-added platform chemicals by simple, green, and efficient procedures represents one of the most accessible and sought-after routes towards sustainable chemistry. In the present work, we report the remarkable catalytic activity of rare-earth metal triflates in conjunction with choline chloride, a natural, low-cost, and available organic compound to selectively convert glucose and fructose into hydroxymethylfurfural (HMF). The hypothesized mechanism is based on the initial glycosylation of glucose assisted by scandium(III) triflate and choline chloride to produce a glycoside, which can evolve through an intramolecular rearrangement and subsequent dehydration to produce the final product HMF. A comparison with other types of catalysts is carried out with particular focus on the side reactions. The apparatus consists of a closed biphasic system and the excellent capacity of methyl propyl ketone (MPK) to extract HMF in only one cycle is proved. The process was conducted at 150 °C using 1.5 molar equivalents of choline chloride in which glucose was converted into HMF after three hours using the catalyst in 8% molar quantity, while fructose was converted in one hour employing the catalyst in 4% molar quantity. The best performance was obtained by employing scandium(III) triflate as a catalyst with an yield of 94% and 99% of HMF from glucose or fructose, respectively. We assumed a first-order reaction model for both glucose and fructose conversion into HMF. The R-squared values are greater than 0.9, demonstrating that our kinetic model fitted well with the experimental results. In addition, activation energies are 16.9 kJ mol<sup>-1</sup> for glucose and 9.31 kJ mol<sup>-1</sup> for fructose due to the longer reaction path of glucose. The catalytic system can be recycled up to five times with a HMF yield of over 80% for glucose and over 90% for fructose, maintaining the same selectivity.

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## 1. Introduction

The growing limitations of fossil resources have turned the spotlight onto innovative and sustainable technologies.<sup>1,2</sup> Renewable sources have gained enormous attention over the past years and this is largely due to the constant demand related to the expansion of the world population.<sup>3-5</sup> Biomass-derived substances represent the main source to draw renewable raw materials to be converted into useful products with a similar value to those obtained from fossil feedstocks.<sup>6-8</sup> Carbohydrate research grew fast in this direction and it is now a consolidated starting point to develop alternative chemistry

processes.<sup>9-11</sup> Glucose is not only abundant in nature and food-related products, but it also represents important energy storage for different living organisms.<sup>12,13</sup> It can be recovered from natural sources or can be produced from naturally occurring polymers, for instance, and non-food waste, like cellulose.<sup>14-16</sup> Fructose is less abundant than glucose but is also conspicuously present in nature as a renewable feedstock.<sup>17-19</sup> Furan biomass-derived molecules represent important targets for carbohydrate research.<sup>20</sup> Over the years, an extraordinary number of breakthroughs have been achieved regarding synthetic strategies and industrial applications, which have allowed these products to gain a precious market value.<sup>21-23</sup> Given the increasing use and multiple connections in various sectors, such as biofuels, textile industry, medical, pharmaceutical, and construction, these compounds have acquired the name of platform molecules.<sup>24-26</sup> HMF is one of the most investigated molecules belonging to this category with a broad network of interactions in different sectors.<sup>27,28</sup>

Department of Chemistry and Chemical Technologies, University of Calabria, Via P. Bucci, Cubo 12C, Rende, CS, Italy. E-mail: [denino@unical.it](mailto:denino@unical.it), [fabrizio.olivito@unical.it](mailto:fabrizio.olivito@unical.it)

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Furthermore, as a platform molecule, HMF represents a versatile intermediate to obtain various high-value derivatives, for example, deriving from its oxidation. In fact, 2,5-diformylfuran and 2,5-furandicarboxylic acid are promising intermediates for pharmaceuticals and very useful polymer-building blocks for the synthesis of functional polymers.<sup>29–31</sup>

HMF is a well-known product of sugar dehydration, studied and researched for decades.<sup>32,33</sup> Catalysis has played a central role in this process from the past years to the present day.<sup>34,35</sup> Conventionally, when the starting material is glucose, a Lewis acid is usually employed for its isomerization to fructose and then a Brønsted acid is necessary to push the dehydration towards the product.<sup>36,37</sup> Other reports have also hypothesized the direct dehydration of glucose to HMF by Brønsted acid without passing through fructose.<sup>38,39</sup> The interest in the selective production of HMF is always growing and this increases the publication of scientific papers.<sup>40,41</sup> A valid experimental procedure developed in this field consists of a biphasic system, in which the extraction solvent plays a central role together with other factors such as the catalyst loading, temperature, and reaction time.<sup>42,43</sup> Hydroxymethyl furfural is known to be unstable under aqueous conditions in the presence of Brønsted acids, because it can undergo rehydration to levulinic acid and formic acid or produce humin by autocondensation.<sup>44,45</sup> Metal halides usually employed as Lewis acids easily decompose in the presence of water or under harsh reaction conditions producing inorganic acids, such as hydrochloric acid, that can decompose HMF.<sup>46,47</sup> Choline chloride-based DES together with metal halide have been proven to be efficient methods to produce HMF from carbohydrates, but the main drawbacks are the need for large amounts of solvents to extract the product from the ionic phase and the toxicity of Lewis acid metals, for example, the use of chromium.<sup>48,49</sup> Rare earth metal halides and triflate have proved to be excellent Lewis acids as the catalytic system for HMF synthesis from glucose and fructose but the use of harmful solvents like *N,N*-dimethylacetamide (DMA), and DMSO was necessary to reach efficient yields.<sup>50,51</sup> In continuity with respect to the well-documented catalytic activity and water-tolerability of triflate lanthanides,<sup>52–54</sup> in this work, we proved the stability and the exceptional catalytic activity of rare-earth metal triflates to produce HMF through isomerization-dehydration of carbohydrates, with the participation of choline chloride, an abundant, cheap and biodegradable substrate that allows Fisher-type glycosylation on fructose before the dehydration steps. The used reaction apparatus consists of a biphasic system in which methyl propyl ketone (MPK) was proved to be an excellent extraction solvent for the recovery of HMF with respect to conventional ones. A comparison employing the relative metal halides was carried out and the yields and reaction by-products are discussed. The methodology furnished optimum selectivity and yields. Screening of different rare-earth metal cations was carried out, with the analysis of the used molar ratio, catalyst loading, temperature, and reaction time. The catalytic mechanism has been hypothesized and discussed. The mechanism is explained through

the high affinity of scandium for oxygen, the tendency of glucose to form the glucoside with choline, conditions suitable for dehydration, and numerous NMR and kinetic tests. HMF was extracted after only one reaction cycle while the recyclability of the catalytic system was proved to furnish excellent yields up to five times.

## 2. Experimental

### 2.1 Materials and methods

All chemicals and solvents were purchased from common commercial sources (see ESI†). Solvents were distilled before use. The analytical standard of HMF was purchased in analytical grade from Sigma Aldrich. All reactions were monitored by GC/MS analysis (Shimadzu, Kyoto, Japan). The GC-MS Shimadzu workstation was constituted by a GC 2010 (equipped with a 30 m-QUADREX 007-5MS capillary column, operating in the “split” mode, 1 mL min<sup>-1</sup> flow of He as carrier gas) and a 2010 quadrupole mass-detector. <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on Brüker spectrometers (Bruker Instrument, Inc., Zurich, Switzerland) at 300 MHz and 126 MHz, respectively, in CDCl<sub>3</sub> or D<sub>2</sub>O. Chemical shifts were reported in  $\delta$  units (ppm) with tetramethylsilane (TMS) as a reference.

Samples were analysed using HPLC Agilent series 1100 equipped with an isocratic pump (Agilent technologies 1200 series) and UV-vis detector for HMF analysis and a refractive index detector for the detection of unreacted glucose and fructose. The reversed phase analytical column was a C18 Jupiter, with dimensions of 300 A, 250 × 4.60 mm, and 10 microns for particle size (Phenomenex, USA). The wavelength of the reference was 360 nm with a peak width of 100, while the wavelength for HMF analysis was set to 385 nm with a peak width of 8 mm. The chosen mobile phase was methanol (50%) and acetonitrile (50%), in the isocratic elution mode at a flow rate of 1 mL min<sup>-1</sup>. The sample injection volume was 20  $\mu$ L. The column was washed with methanol after 5 injections for 30 minutes. The quantitative analysis of HMF was performed by HPLC with a UV-vis detector, using the external analytical standard method. The unknown sample was diluted in water, filtered, and injected after removing the reaction solvent. The unreacted quantities of glucose and fructose were analysed by HPLC, dissolving the solid at the bottom of the biphasic system in water, using HPLC with a refractive index detector and a solution of 5 mM H<sub>2</sub>SO<sub>4</sub> as the mobile phase. Each test was done in triplicate and the values were expressed as mean values. Qualitative analysis of the products and the relative by-products in the solvent phase was evaluated by GC-MS, using a gradient temperature from 70 °C to 250 °C with an increase of 16 °C min<sup>-1</sup>.

Glucose conversion or fructose conversion (*x*), HMF yield (*y*), and HMF selectivity (*s*) were calculated based on the following eqn (1)–(3), in accordance with the previously reported studies:<sup>55</sup>

$$x = \frac{C_{\text{sugar}}^0 - C_{\text{sugar}}^t}{C_{\text{sugar}}^0} \times 100 \quad (1)$$



$$y = \frac{C_{\text{HMF}}^e}{C_{\text{HMF}}^t} \times 100 \quad (2)$$

$$s = \frac{y}{x} \times 100 \quad (3)$$

where  $C_{\text{sugar}}^0$  and  $C_{\text{sugar}}^t$  correspond to the initial concentration of glucose or fructose and the relative concentration at the reaction time  $t$  (in mol L<sup>-1</sup>).  $C_{\text{HMF}}^e$  is the effective concentration of HMF after the reaction time and  $C_{\text{HMF}}^t$  is the theoretical concentration assuming that glucose or fructose is totally converted.

For the recycling test, the catalytic system consisting of choline chloride/rare earth metal triflate was used without any pre-treatment for the next cycle, after removing the extraction solvent.

## 2.2 Preparation of rare earth metal triflates

Cerium(III) triflate, lanthanum(III) triflate, and ytterbium(III) triflate were prepared freshly in accordance with a procedure reported in the literature.<sup>56</sup> Cerium chloride heptahydrate, lanthanum chloride heptahydrate, and ytterbium chloride hexahydrate, were dissolved in distilled water and trifluoromethanesulfonic acid (3 molar equivalents) was added dropwise at room temperature. The reaction was allowed to stir at room temperature for two hours and after that, the solvent was removed under a vacuum. The solid was washed several times with diethyl ether and dried in an oven at 80 °C for one day and after that, the product was kept in a chemical drier for 15 days. Traces of water remained in the FT-IR spectrum in accordance with the literature.<sup>51</sup> The FT-IR spectra of the freshly synthesized cerium(III) triflate, lanthanum(III) triflate, and ytterbium(III) triflate were acquired using the Shimadzu IRAffinity-1S spectrometer (Shimadzu Corporation) in the spectral region of 375 and 4000 cm<sup>-1</sup> with a resolution of 1 cm<sup>-1</sup>, setting 50 scans for a single analysis and using the KBr pellet technique. The KBr pellets were obtained by mixing the sample with KBr powder (ratio 1:100) and pressing with a hydraulic press, at the pressure of 10 tons for 5 minutes. The resulting pellets were placed in the appropriate compartment of the instrument and exposed to the FT-IR light beam for analysis.

## 2.3 Reaction procedure

Glucose (40 mg) was mixed with choline chloride (1.5 molar equivalents) and the relative metal triflate (8% molar quantity with respect to glucose) in a closed Pyrex vial. Methyl propyl ketone (4 ml) was added, the vial was closed with a cap and the biphasic system was allowed to stir for three hours by using a magnetic bar at 150 °C. The solution was decanted and filtered through a sintered glass filter and the solvent was removed under a vacuum. The product in the form of a yellowish oil was analysed by GC-MS and HPLC to detect any trace of by-products and isolated with a yield of 94%. The unreacted solid was dissolved in water and analysed by HPLC to detect the trace of unreacted glucose. The purity of the product was

also confirmed by <sup>1</sup>H NMR spectroscopy in accordance with the spectra previously reported.<sup>57</sup>

Fructose (40 mg) was mixed with choline chloride (1.5 molar equivalents) and the relative metal triflate (4% molar quantity with respect to fructose) in a vial. Methyl propyl ketone (4 ml) was added, the vial was closed with a cap and the biphasic system was allowed to stir for one hour by using a magnetic bar at 150 °C. The solution was decanted and filtered through a sintered glass filter and the solvent was removed under a vacuum. The product in the form of yellowish oil was analysed by GC-MS and HPLC to detect any trace of by-products and isolated with a yield of 99%. The unreacted solid was dissolved in water and analysed by HPLC to detect the traces of unreacted fructose. The purity of the product was also confirmed by <sup>1</sup>H NMR spectroscopy in accordance with the spectra previously reported.<sup>48</sup>

## 3. Results and discussion

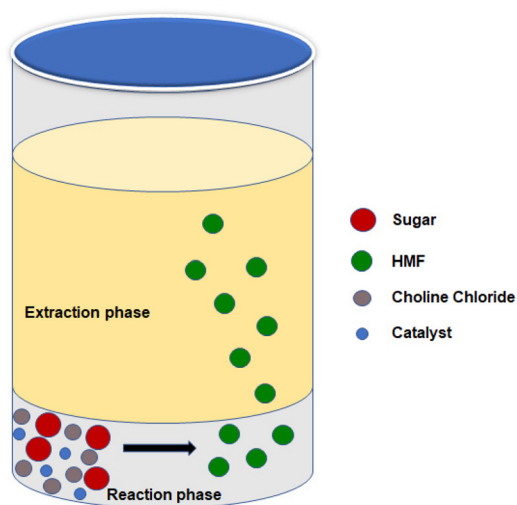
We focused our attention on the development of a synthetic strategy to produce HMF in high yields and selectivity, starting from fully renewable sources. Choline chloride is biocompatible and cheap molecule that was proved recently to possess a stabilizing effect for HMF production.<sup>58</sup> In particular, there are different reports on the physicochemical investigation of choline chloride-based DES with glucose and fructose or other sugars.<sup>59</sup> One of the most important limitations of the use of DES as a solvent, involves the difficulty to extract HMF quantitatively, due to the strong affinity of this molecule for these hydrogen-bond-rich phases.<sup>60</sup> K. De Oliveira Vigier *et al.*, recently focused attention on the role of choline chloride not only as a stabilizing agent but mostly as a reactive participant in the conversion of xylose into furfural.<sup>61</sup> With these important considerations in mind, we started the optimization of the process.

### 3.1 Optimization of the catalytic system

Initially, starting from glucose and fructose, we used an equimolar quantity of choline chloride in which these sugars were completely solubilized at 150 °C after some time. To evaluate any conversion, we used a closed vial, MIBK (methyl isobutyl ketone) as a conventional extraction solvent, to create a biphasic system,<sup>62</sup> as illustrated in Fig. 1. The latter is only a representative scheme, in which the real used volumes are not shown.

After three hours, no trace of HMF was detected by GC-MS in the solvent phase. HMF is commonly produced by the dehydration reaction of sugars and for this reason, with the aim of creating a stable catalytic system, we used rare earth metal triflates because the water-tolerability of these species has been known for a long time.<sup>63,64</sup> We started with the optimization of the reaction conditions using glucose, choline chloride, and Er(OTf)<sub>3</sub> with MIBK as the extraction solvent. The results are reported in the following Table 1.





**Fig. 1** Schematic representation of the biphasic system used for HMF production.

**Table 1** The optimization of the reaction conditions for glucose conversion into HMF using erbium(III) triflate

Entry <sup>a</sup>	Catalyst (mol%)	Choline chloride (eq. mol)	HMF yield (%)
1	1%	1	5
2	2%	1	25
3	4%	1	55
4	8%	1	68
5	10%	1	68
6	8%	0.5	55
7	8%	1.5	78
8	8%	2	70

<sup>a</sup> Glucose (40 mg, 0.22 mmol), 4 ml of MIBK at 150 °C under magnetic stirring for 3 hours.

Using one equivalent of choline chloride, the glucose conversion into HMF increases with the increase of the percentage of the catalyst (entries 1–4). The percentage of the product is the same using 8% of the catalyst instead of 10% (entries 4 and 5). When half of the choline chloride equivalents are used (entry 6), the glucose conversion is lower. This result may demonstrate that choline chloride plays an essential role in the catalysis of this reaction and that it is not merely a reaction medium. In any case, the optimal equivalents of choline chloride are 1.5 because when 2 equivalents were used, the extraction process becomes worse, probably because the product has a stronger affinity with choline chloride with respect to the solvent (entries 7 and 8). Starting from fructose, the optimized conditions consist of 1.5 equivalent of choline chloride, 4% molar of the catalyst, and the quantitative yield is obtained after one hour. Both for glucose and fructose, we did not detect any by-products in the reaction phase by HPLC analysis.

At this point, a screening of different metal triflates starting from glucose and choline chloride was carried out and the resulting HMF yield is reported in Table 2.

**Table 2** Screening of different rare earth metal triflates on glucose

Entry <sup>a</sup>	Catalyst	HMF yield (%)
1	La(OTf) <sub>3</sub>	70
2	Ce(OTf) <sub>3</sub>	72
3	Ho(OTf) <sub>3</sub>	77
4	Er(OTf) <sub>3</sub>	78
5	Yb(OTf) <sub>3</sub>	81
6	Sc(OTf) <sub>3</sub>	84

<sup>a</sup> Glucose (40 mg, 0.22 mmol), choline chloride (46 mg, 0.33 mmol), 4 ml of MIBK, 8% molar equivalent of the catalyst, at 150 °C for 3 hours under magnetic stirring.

The HMF yields increase passing from lanthanum to ytterbium and scandium (entries 1–6), demonstrating that Lewis acid catalysis is fundamental in this process and is linked to electrons and the ionic radius of the element.<sup>65</sup>

The same evaluation of the different metal triflates was conducted starting from fructose and the yield of HMF is reported in Table 3.

The same trend of the metals is obtained from the reaction starting from fructose, with a yield increasing from lanthanum to scandium (entries 1–6). As can be seen, the yield in HMF is higher with respect to all the metals tested under the same reaction conditions on glucose.

In this reaction process, given the production of water molecules, the Lewis acidity of rare earth metals is also influenced by hydrolysis constants and high order of exchange rate constants for the substitution of inner-sphere water ligands (WERC), as reported in many studies, which are in agreement with the observed trend. In accordance with the literature,<sup>66</sup> the parameters are shown in Table 4.

Metal cations with  $pK_{\text{H}}$  values from 4.3 to 10.08 and WERC higher than  $3.2 \times 10^6 \text{ M}^{-1}\text{s}^{-1}$  show excellent performances in the reactions in which they act as Lewis acids. When the values are below this limit, metals are easily hydrolysable, while beyond this limit they are too stable to catalyse a reaction.<sup>67,68</sup>

From entries 1–6, it is evident that all the rare earth metals used are in this range of values and can be considered optimal Lewis acids.

At this point, considering the very good results obtained from both glucose and fructose, we decided to employ scandium(III) triflate as a catalyst for HMF production.

**Table 3** Screening of different rare earth metal triflates on fructose

Entry <sup>a</sup>	Catalyst	HMF yield (%)
1	La(OTf) <sub>3</sub>	78
2	Ce(OTf) <sub>3</sub>	80
3	Ho(OTf) <sub>3</sub>	99
4	Er(OTf) <sub>3</sub>	99
5	Yb(OTf) <sub>3</sub>	99
6	Sc(OTf) <sub>3</sub>	99

<sup>a</sup> Fructose (40 mg, 0.22 mmol), choline chloride (46 mg, 0.33 mmol), 4 ml of MIBK, 4% molar equivalent of the catalyst, at 150 °C for 1 hour under magnetic stirring.



**Table 4** Water hydrolysis constant and exchange rate constants for the substitution of inner-sphere water ligands of rare earth-metal cations used in this study

Entry	Cations	$pK_h$	WERC
1	La <sup>3+</sup>	7.6–8.5	$10^6$ – $10^8$
2	Ce <sup>3+</sup>	8.3	$2.7 \times 10^8$
3	Ho <sup>3+</sup>	8	$6.1 \times 10^7$
4	Er <sup>3+</sup>	7.9	$1.4 \times 10^8$
5	Yb <sup>3+</sup>	7.7	$8 \times 10^7$
6	Sc <sup>3+</sup>	4.3	$4.8 \times 10^7$

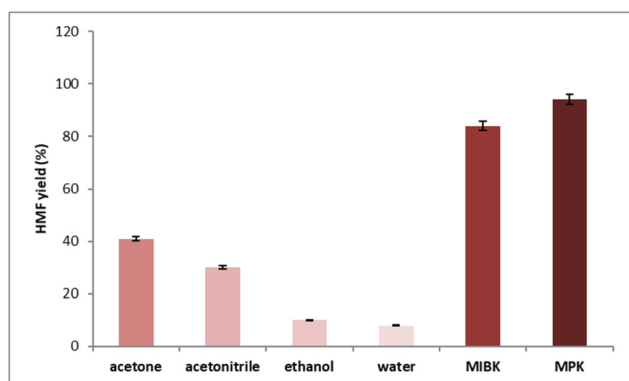
It is noteworthy that, in the presence of triflates as a catalyst, formic acid as a by-product was not detected. All of this is evident by the GC-MS chromatograms of crude that have showed only formation of HMF (see ESI<sup>†</sup>), also by using the operative conditions reported in literature to perform the quantitative detection of formic acid.<sup>69,70</sup>

### 3.2 The effect of the solvent

Starting from the promising results achieved with MIBK, we tested different solvents on the model reaction starting from glucose, choline chloride, and scandium(III) triflate. The results are presented in Fig. 2.

When polar aprotic solvents like acetone and acetonitrile were used, the yield of HMF was 41% and 30%, respectively. Probably the greater polarity of the solvent prevents the product formation because the reagents are solubilized and their contact was restricted. This deduction was confirmed when ethanol and water were used as polar protic solvents. The yields of 10% and 8%, respectively, are due to the complete dissolution and solvation of the reagents without the formation of a biphasic system in the vial.

When methyl propyl ketone (MPK) was used, the reaction yield increased to 94% (Table 5, entry 1) with respect to 84% (Table 2, entry 6) deriving from the use of conventional methyl isobutyl ketone (MIBK). This effect is justifiable because MPK



**Fig. 2** HMF yield using different solvents. Reaction conditions: glucose (40 mg, 0.22 mmol), choline chloride (46 mg, 0.33 mmol), 8% scandium(III) trifluoromethanesulfonate, 4 ml of solvent, 150 °C for 3 hours. The experiment was repeated three times and the value was expressed as a mean value.

**Table 5** Isolated yields of HMF in the optimum reaction system: MPK/Sc(OTf)<sub>3</sub>/choline chloride

Entry	Reagent	Conversion (%)	HMF yield (%)
1	Glucose	95	94
2	Fructose	100	99

is slightly more polar than MIBK, but has no solvation effect compared to the reactants. In addition, it has a lower boiling point, which allows easier removal from the product once formed. The reaction was also conducted in MPK for fructose, observing the same quantitative yield of 99%. Table 5 summarizes the obtained results for glucose and fructose as the starting materials.

To demonstrate the ability of MPK to extract the product, we calculated the partition coefficients of HMF ( $P_{\text{HMF}}$ ) in MPK/water and MIBK/water systems. Therefore, 5 mL of MPK or MIBK, and 5 mL of water containing 100 mM HMF, were mixed together and heated to the desired temperature. The partition coefficient ( $P_{\text{HMF}}$ ) was calculated by dividing the concentration of HMF ( $C_{\text{HMF}}$ ) in the extraction phase by  $C_{\text{HMF}}$  in the aqueous phase. The concentration was analysed by HPLC coupled with a UV detector. In Table 6 we reported the obtained data from 25 °C to 60 °C.

As reported in Table 6, the partition coefficient of HMF in MPK/water is higher than that of MIBK/water at all the tested temperatures (entries 1–3). This is another confirmation of the potential that this new solvent has if compared to the conventional ones, such as MIBK.<sup>71</sup>

### 3.3 Effect of reaction temperature

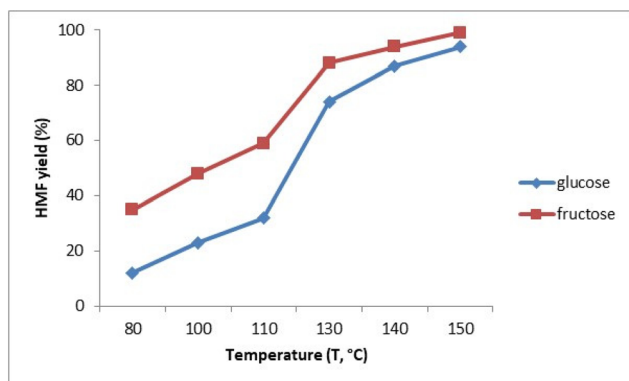
We evaluated the effect of temperature on the conversion of glucose and fructose into HMF, starting from the optimized reaction conditions. The values are reported in Fig. 3.

The production of HMF in this catalytic process is strongly dependent on the temperature. For glucose, the yield is about 12% at 80 °C until the maximum yield of 94% is reached 150 °C. From 150 °C onwards, the graph reached a plateau. It is remarkable how the product remains stable in the extraction solvent under the optimized reaction conditions even at high temperatures. The same trend was proved to start from fructose with reaction yields of HMF higher at all the tested reaction times. The yield is about 35% at 80 °C until a maximum yield of 99% at 150 °C and the value remained constant at higher temperatures. Both for glucose and fructose, a significant increase in the HMF yield between 110 °C and 130 °C was

**Table 6** The influence of temperature on  $P_{\text{HMF}}$  (in MPK or MIBK and water 1 : 1  $V_{\text{organic}}/V_{\text{aq}}$  and 0.1 mM HMF)

Entry	$T$ (°C)	$P_{\text{HMF}}$ (MPK)	$P_{\text{HMF}}$ (MIBK)
1	25	1.55	1.34
2	40	1.54	1.33
3	60	1.54	1.33





**Fig. 3** HMF yields from glucose at different reaction temperatures (blue). Reaction conditions: glucose (40 mg, 0.22 mmol), choline chloride (46 mg, 0.33 mmol), 8% scandium(III) trifluoromethanesulfonate, 4 ml of MPK, after 3 hours. The HMF yields from fructose at different reaction temperatures (red). Reaction conditions: fructose (40 mg, 0.22 mmol), choline chloride (46 mg, 0.33 mmol), 4% scandium(III) trifluoromethanesulfonate, 4 ml of MPK.

evident. Excellent selectivity was confirmed by no detection of by-products through the analysis of the reaction phase by HPLC for both carbohydrates.

### 3.4 Effect of reaction time

The dependence of the HMF yield on the reaction times is another critical parameter; the plots relative to the reaction starting from glucose (A) and fructose (B) at three different temperatures are shown in Fig. 4.

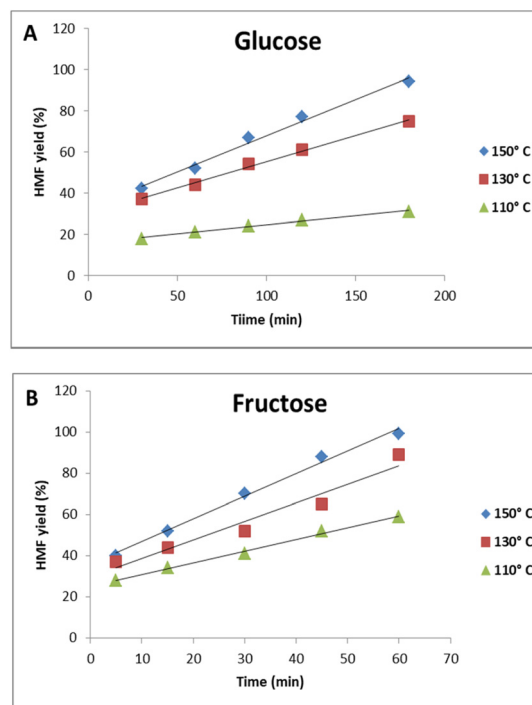
We began the investigation of the reaction kinetics relative to HMF production from glucose (Fig. 4A). The slope of the graph is higher at 130 °C and 150 °C respect to 110 °C, this is proof that temperature is crucial in this process. From 120 to 180 minutes, the process shows the best yield of 94% at 150 °C. Over the time of 180 minutes, the graph reached a plateau at all the temperatures used.

For the reaction conducted starting from fructose, the kinetics are remarkably different (Fig. 4B). The slope of the graph is similar at 130 °C and 150 °C and just a little lower at 110 °C. The reaction starting from fructose is already efficient in terms of yield. After only five minutes, the yield of HMF produced was 40% at 150 °C, the optimized yield of 99% was reached after 60 minutes at the same temperature. An identical yield was obtained after 90 minutes, demonstrating the stability of the product in the reaction medium. The difference in the reaction kinetics starting from glucose and fructose will be discussed in sections 3.6 and 3.7.

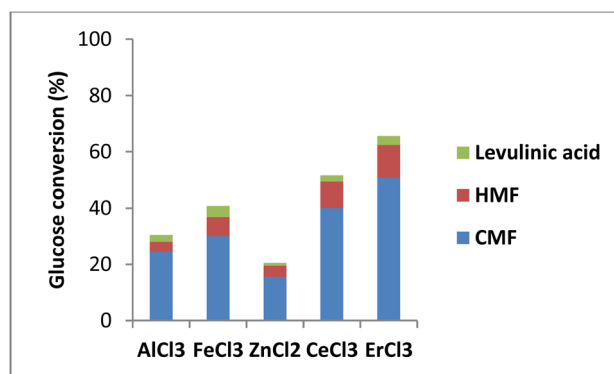
### 3.5 Product analysis using metal halides

At this point, we carried out an investigation about the use of the metal halides instead of triflates on the optimized reaction starting from glucose. The percentage of the products is reported in Fig. 5.

From the reported results, it is evident that the production of chloromethyl furfural (CMF) is the main compound in all



**Fig. 4** (A) HMF yields at different reaction times starting from glucose at three different temperatures. Reaction conditions: glucose (40 mg, 0.22 mmol), choline chloride (46 mg, 0.33 mmol), 8% scandium(III) triflate, 4 ml of MPK. (B) HMF yield at different reaction times starting from fructose at three different temperatures. Reaction conditions: fructose (40 mg, 0.22 mmol), choline chloride (46 mg, 0.33 mmol), 4% scandium(III) triflate, 4 ml of MPK.



**Fig. 5** Glucose conversion using metal halides. Reaction conditions: glucose (40 mg, 0.22 mmol), choline chloride (46 mg, 0.33 mmol), 8% of the catalyst, 4 ml of MPK, after 3 hours at 150 °C.

obtained mixtures. This molecule is another important platform molecule with high potential.<sup>72</sup> The production of CMF can be rationalized with the fact that such catalysts are unstable under the conditions used in this study. The production of water together with high temperature and pressure promotes the decomposition of the metal halides.<sup>73</sup> This decomposition is due to the hydrolysis of the species with the



consequent production of hydrochloric acid. The presence of an inorganic acid together with the availability of chloride ions coming from both the acid and the choline chloride converts the produced HMF into the respective halogenated molecule CMF (see ESI†). Another indicator of this result is the presence of levulinic acid in the mixture. This acid is a common degradation product of HMF, mostly in the presence of water and a Brønsted acid like hydrochloric acid.<sup>74</sup>

### 3.6 Hypothesis of the catalytic mechanism

After we have analysed and discussed the critical parameters of the process, we supposed the mechanism underlying this new synthetic method for the production of HMF starting from glucose or fructose in the presence of Sc(III) triflate/choline chloride as a catalytic system, as depicted in Fig. 6.

Due to the substantial difference between the kinetics of the reaction starting from glucose and the same reaction start-

ing from fructose, we supposed an initial isomerization of glucose to fructose in the first step, which justifies the longer reaction times.<sup>75</sup>

Furthermore, considering the known affinity of lanthanides and rare earth elements, in general, to coordinate with oxygen atoms,<sup>76</sup> we supposed the initial chelation of scandium(III) triflate with hydroxyls on C1 and C2 of glucose. Then, choline chloride can react with glucose anomeric hydroxyl by Fischer-type glycosidation to produce glycoside **1**, which can evolve to fructofuranosyl oxonium ion **2** through an intramolecular rearrangement.<sup>77</sup> In the subsequent dehydration step, scandium(III) triflate is the fundamental key to promoting double water elimination and producing the final product HMF **3**.

To support this hypothesis, we recorded <sup>13</sup>C NMR spectra on the glucose/Sc(OTf)<sub>3</sub>/choline chloride mixture in which we did not observe any C=O group of the open-chain form of glucose.<sup>78</sup> Instead, in this mixture, we noted a slight shift of

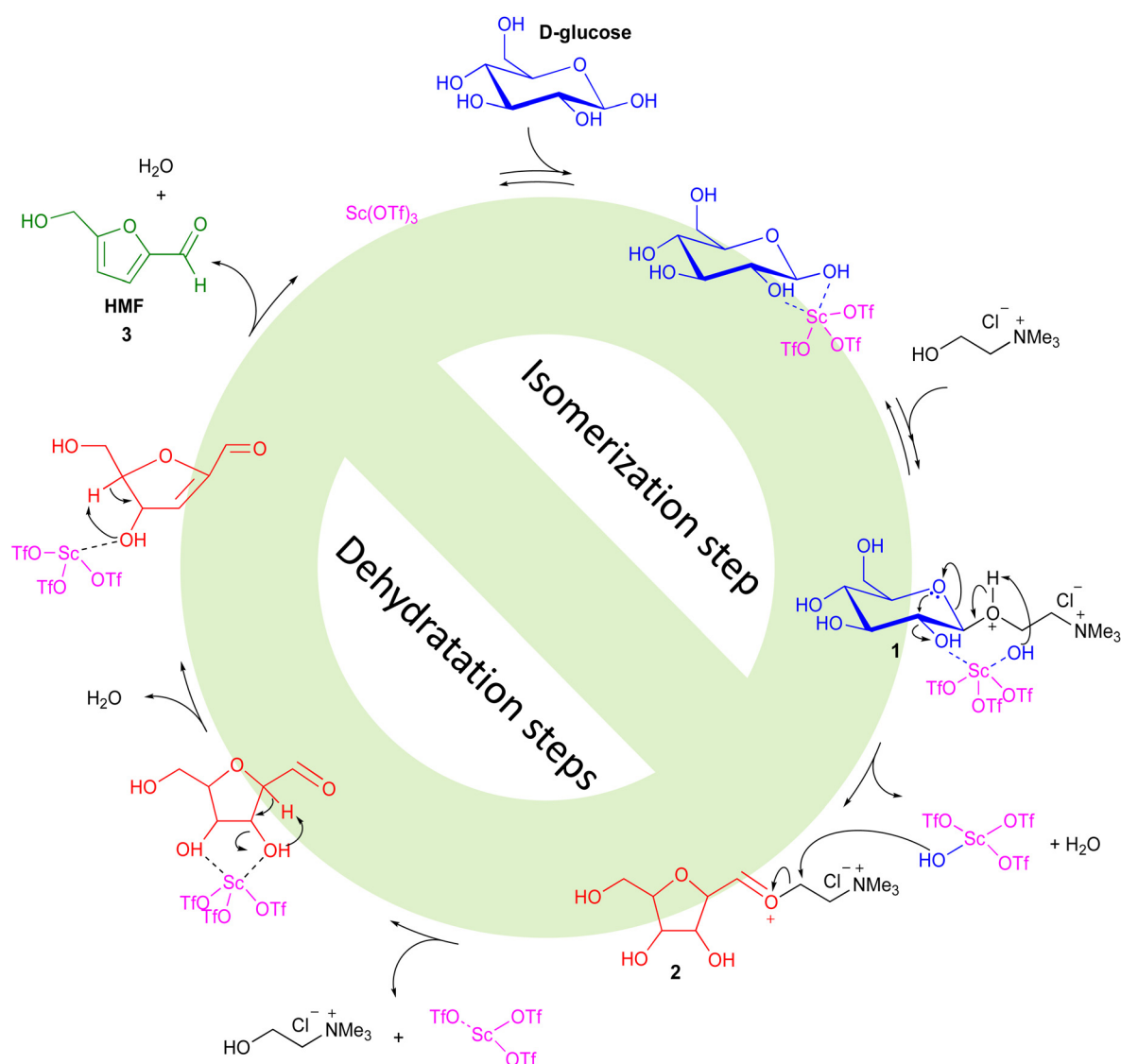


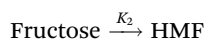
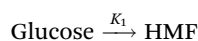
Fig. 6 The supposed catalytic mechanism starting from glucose.



signal of the glucose anomeric carbon with respect to the classical value of only glucose, which may confirm the formation of glucoside **1** (see ESI for spectra†).

### 3.7 Reaction kinetic models of HMF synthesis from glucose and fructose

There are few studies on the production of HMF from glucose, fructose, or other carbohydrates in a biphasic system.<sup>79,80</sup> In other procedures starting from cellulose, the production of HMF passing from glucose is described through consecutive first-order reaction models and the rate constant for glucose isomerization to fructose is commonly neglected.<sup>81,82</sup> We used a biphasic system and no trace of fructose was detected during the reaction investigation. In addition, we did not detect other common by-products such as levulinic acid or formic acid.<sup>83</sup> For these reasons, we assumed that the production of HMF from glucose and fructose follows a first-order model with the relative rate constants  $k_1$  and  $k_2$ , reported as follows:



The experiments were conducted at three different reaction temperatures of 110, 130, and 150 °C for the conversion of both glucose and fructose. Due to the different reaction kinetics, we decided to use a different sampling interval, from 0 to 60 minutes for fructose and from 0 to 200 minutes for glucose. We reported the derived reaction rate equations for glucose conversion into HMF:

$$\frac{dC_g}{dt} = -k_1 C_g \quad (4)$$

$$\frac{dC_{\text{HMF}}}{dt} = -k_1 C_g \quad (5)$$

where  $C_g$  is the glucose concentration and  $C_{\text{HMF}}$  is the HMF concentration.

Due to our assumption, the equations are similar for fructose conversion:

$$\frac{dC_f}{dt} = -k_2 C_f \quad (6)$$

$$\frac{dC_{\text{HMF}}}{dt} = k_2 C_f \quad (7)$$

where  $C_f$  and  $C_{\text{HMF}}$  correspond to fructose and HMF concentrations, respectively.

The activation energy was calculated using the Arrhenius equation:

$$k = A \exp\left(-\frac{E_a}{RT}\right) \quad (8)$$

In Fig. 7, we reported the Arrhenius plots for glucose and fructose conversion into HMF.

The  $R$ -squared values for both glucose and fructose conversions are greater than 0.9, which demonstrates that our assumed model fits well with the experimental results. The

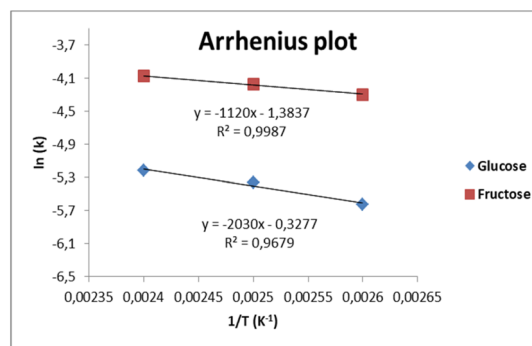


Fig. 7 Arrhenius plot for glucose conversion into HMF (blue) and fructose conversion into HMF (red).

Table 7 Kinetic parameters for glucose and fructose conversion into HMF in a biphasic system

Temperature (K)	383.15	403.15	423.15
Rate constant (min <sup>-1</sup> )			
$K_1$	$3.6 \times 10^{-3}$	$4.7 \times 10^{-3}$	$5.4 \times 10^{-3}$
$K_2$	$1.4 \times 10^{-2}$	$1.5 \times 10^{-2}$	$1.7 \times 10^{-2}$
Activation energy (kJ mol <sup>-1</sup> )			
$E_{a1}$	16.9		
$E_{a2}$	9.31		

value is just a little lower for glucose, despite the assumption of neglecting the isomerization step. The values of the activation energies are reported in Table 7.

The value of the activation energy of 16.9 kJ mol<sup>-1</sup> is higher for glucose conversion with respect to fructose conversion (9.31 kJ mol<sup>-1</sup>). These findings are in accordance with our hypothesized mechanism in which glucose passes for fructose through isomerization and then dehydrates, while for fructose, the reaction path is the direct dehydration to HMF. These values are of the same order but lower than the ones reported in similar studies.<sup>75,84–87</sup>

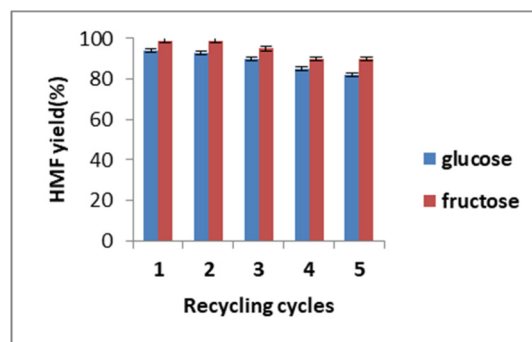


Fig. 8 Recycling of the catalytic system for glucose and fructose. Reaction conditions: glucose (20 mg, 0.11 mmol), 4 ml of solvent, 150 °C for 3 hours. Fructose (20 mg, 0.22 mmol), 4 ml of solvent, 150 °C for 1 hour. The experiments were repeated three times and the values expressed are the mean values.



**Table 8** A comparison with the HMF yield reported in the literature

Entry	Reagent	Catalyst	Time (min)	<i>T</i> (°C)	HMF yield (%)	Ref.
1	Glucose	NbO/NbP	120	152	26.5	88
2	Glucose	HNbWO <sub>6</sub>	180–1440	120–140	36–44	89
3	Fructose	WO <sub>3</sub> /SnO <sub>2</sub>	120	120	93	90
4	Fructose	PWAl-200	240	170	61.7	91
5	Glucose	DES/CrCl <sub>3</sub>	40	70	80	92
6	Fructose	HCl/KBr	1	(MW) 160	91	93 <sup>a</sup>
7	Glucose	AlCl <sub>3</sub> /HCl	20	175	54.5	94 <sup>a</sup>
8	Glucose	Phosphated titania	60	150	66	95
9	Fructose	[C <sub>4</sub> SO <sub>3</sub> Hmim] [HSO <sub>4</sub> ]	180	120	94.6	96 <sup>a</sup>
10	Fructose	Sulfur-doped peanut shell	120	130	94.6	97
11	Glucose	Sc(OTf) <sub>3</sub>	180	150	94	This study
12	Fructose	Sc(OTf) <sub>3</sub>	60	150	99	This study

<sup>a</sup> Reactions in which biphasic systems are employed.

### 3.8 Recycling test

To demonstrate the further use of the method, we evaluated the recyclability of the catalytic system for up to five cycles.

After the fresh reaction, a polar phase containing the catalytic system (scandium(III) triflate/choline chloride) was added with sugar (glucose or fructose) and MPK, and the process was repeated four times. The results are shown in Fig. 8.

The reaction yield for glucose is 90% after three cycles and 82% after five cycles, while for fructose, the yield is 95% after three cycles and 90% after five cycles. The different behaviour of the catalytic system for glucose may be due to a greater probability of deactivation due to prolonged reaction temperature and pressure. Anyway, the results are satisfactory with respect to the literature values reported in the following paragraph.

### 3.9 A comparison with literature data

In Table 8, we show a comparison with significant works from the past years on the catalytic conversion of glucose or fructose into HMF.

The HMF yield from glucose is lower for all the reported studies using different solid acid catalysts (entries 1 and 2), metal halides (entries 5 and 7), and a microreactor with phosphated titania catalyst (entry 8) with respect to the present work (entries 11 and 12). The same result was obtained considering the study on fructose conversion, using solid acid catalysts (entries 3 and 4), water-soluble inorganic acid (entry 6), acid-functionalized ionic liquid (entry 9), and sulphur-doped peanut shell catalysts in ionic liquid (entry 10) with respect to the results of the present study (entries 11 and 12).

## 4. Conclusions

In this work, we presented an innovative strategy to produce HMF with excellent yields and selectivity starting from glucose or fructose. The methodology employs a reusable catalytic system consisting of rare earth metal(III) trifluoromethanesulfonates (*i.e.* scandium(III) triflate) and choline chloride. We

tested different solvents and methyl propyl ketone furnished the best performance. This solvent has several advantages, including a low boiling point, which allows easy removal from the product. In addition, it is stable under the temperature and pressure used in this study and can be recycled after evaporation. During the discussion, we analysed all the critical parameters related to this synthetic procedure, including the choice and catalyst loading, reaction time, and temperature. Due to the difference in the reaction kinetics for glucose and fructose conversion, we hypothesized a catalytic mechanism. The role of the rare-earth metal Lewis acid is crucial, both for glucose isomerization into fructose and also for fructose dehydration. We also hypothesized the important role of choline chloride in glucose glycosylation. The best results in HMF yield were obtained using scandium(III) triflate. In this work, we have provided a method for obtaining HMF from glucose and fructose in short reaction times and in very high yields. Rare earth metal triflates can be purchased or produced from their relative metal halides, while choline chloride is an abundant and cheap reagent. The high selectivity of the developed methodology avoids further purification practices, since after the reaction, it is sufficient to remove only the extraction solvent to obtain pure HMF that can be used in subsequent applications. Finally, this methodology can also be applied to biomass-derived sugars, demonstrating the high eco-compatibility of the process.

## Author contributions

Conceptualization, A. D. N., F. O.; methodology, F. O., V. A., A. J. and M. A. T.; validation, F. O., and P. C.; formal analysis, F. O., V. A. and P. C.; investigation, F. O., V. A., A. J. and M. A. T.; resources, A. D. N., L. M., and P. C.; data curation, F. O., A. J., L. M.; writing—original draft preparation, A. D. N., F. O., and L. M.; writing—review and editing, A. D. N., F. O. and L. M.; supervision, A. D. N., and L. M.; project administration, A. D. N., and L. M.; funding acquisition, A. D. N., L. M., and P. C. All authors have read and agreed to the published version of the manuscript.



## Conflicts of interest

There are no conflicts to declare.

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## References

- P. Marion, B. Bernela, A. Piccirilli, B. Estrine, N. Patouillard, J. Guilbot and F. Jérôme, *Green Chem.*, 2017, **19**, 4973.
- V. G. Zuin, I. Eilks, M. Elschami and K. Kümmerer, *Green Chem.*, 2021, **23**, 1594.
- J. Kühnborn, J. Groß and T. Opatz, *Nat. Prod. Rep.*, 2020, **37**, 380.
- C. Claver, M. B. Yeamin, M. Reguero and A. M. Masdeu-Bultó, *Green Chem.*, 2020, **22**, 7665.
- S. E. Motika and P. J. Hergenrother, *Nat. Prod. Rep.*, 2020, **37**, 1395.
- M. Hiloidhari, D. C. Baruah, M. Kumari, S. Kumari and I. S. Thakur, *Cleaner Prod. Lett.*, 2019, **220**, 931.
- F. Shen, X. Xiong, J. Fu, J. Yang, M. Qiu, X. Qi and D. C. W. Tsang, *Renewable Sustainable Energy Rev.*, 2020, **130**, 109944.
- K. Alper, K. Tekin, S. Karagöz and A. J. Ragauskas, *Sustainable Energy Fuels*, 2020, **4**, 4390.
- P. R. Jensen and S. Meier, *Chem. Commun.*, 2020, **56**, 6245.
- A. Kumar and R. Srivastava, *Sustainable Energy Fuels*, 2019, **3**, 2475.
- J. Remón, F. Santomauro, C. J. Chuck, A. S. Matharu and J. H. Clark, *Green Chem.*, 2018, **20**, 4507.
- A. Cazor, C. Deborde, A. Moing, D. Rolin and H. This, *J. Agric. Food Chem.*, 2006, **54**, 4681.
- T. Bauchop and S. R. Elsdén, *J. Gen. Microbiol.*, 1960, **23**, 457.
- A. Nayak, I. N. Pulidindi and C. S. Rao, *Renewable Energy*, 2020, **159**, 215.
- I. Romeo, F. Olivito, A. Tursi, V. Algieri, A. Beneduci, G. Chidichimo, L. Maiuolo, E. Sicilia and A. De Nino, *RSC Adv.*, 2020, **10**, 34738.
- J. Wang, J. Xi and Y. Wang, *Green Chem.*, 2015, **17**, 737.
- S. R. Taylor, S. Ramsamoj, R. J. Liang, *et al.*, *Nature*, 2021, **597**, 263.
- A. Kriebs, *Nat. Rev. Endocrinol.*, 2020, **16**, 402.
- S. R. Taylor, S. Ramsamoj, R. J. Liang, *et al.*, *Nature*, 2021, **597**, 263.
- Z. Zhang and G. W. Huber, *Chem. Soc. Rev.*, 2018, **47**, 1351.
- R. Gogar, S. Viamajala, P. A. Relue and S. Varanasi, *ACS Sustainable Chem. Eng.*, 2021, **9**, 3428.
- B. Saha and M. M. Abu-Omar, *ChemSusChem*, 2015, **8**, 1133.
- H. Chang, M. S. Kim, G. W. Huber and J. A. Dumesic, *Green Chem.*, 2021, **23**, 9479.
- L. Maiuolo, F. Olivito, V. Algieri, P. Costanzo, A. Jiritano, M. A. Tallarida, A. Tursi, C. Sposato, A. Feo and A. De Nino, *Polymers*, 2021, **13**, 2802.
- R. S. Assary, T. Kim, J. J. Low, J. Greeley and L. A. Curtiss, *Phys. Chem. Chem. Phys.*, 2012, **14**, 16603.
- M. Mascal, *ACS Sustainable Chem. Eng.*, 2019, **7**, 5588.
- A. A. Rosatella, S. P. Simeonov, R. F. M. Frade and C. A. M. Afonso, *Green Chem.*, 2011, **13**, 754.
- M. G. Davidson, S. Elgie, S. Parsons and T. J. Young, *Green Chem.*, 2021, **23**, 3154.
- P. Kisszekelyi, R. Hardian, H. Vovusha, B. Chen, X. Zeng, U. Schwingenschlgl, J. Kupai and G. Szekely, *ChemSusChem*, 2020, **13**, 3127.
- G. Totaro, L. Sisti, P. Marchese, M. Colonna, A. Romano, C. Gioia, M. Vannini and A. Celli, *ChemSusChem*, 2022, **15**, e202200501.
- A. Al Ghatta and J. P. Hallett, *Green Chem.*, 2022, **24**, 3309.
- D. Mastrocola and M. Munari, *J. Agric. Food Chem.*, 2000, **48**, 3555.
- J. Y. G. Chan and Y. Zhang, *ChemSusChem*, 2009, **2**, 731.
- C. Carlini, M. Giuttari, A. M. Raspolli Galletti, G. Sbrana, T. Armaroli and G. Busca, *Appl. Catal. A Gen.*, 1999, **183**, 295.
- C. Lin, C. Chai, Y. Li, J. Chen, Y. Lu, H. Wu, L. Zhao, F. Cao, K. Chen, P. Wei and P. Ouyang, *Green Chem.*, 2021, **23**, 2058.
- B. Saha and M. M. Abu-Omar, *Green Chem.*, 2014, **16**, 24.
- H. Li, Z. Xia, P. Yan and Z. C. Zhang, *Catal. Today*, 2022, **402**, 10.
- W. Guo, T. Kortenbach, W. Qi, E. Hensen, H. J. Heeres and J. Yue, *Appl. Catal.*, 2022, **301**, 120800.
- T. Istasse and A. Richel, *RSC Adv.*, 2020, **10**, 23720.
- G. Parameswaram and S. Roy, *RSC Adv.*, 2018, **8**, 28461.
- J. Slak, B. Pomeroy, A. Kostyniuk, M. Grilc and B. Likozara, *Chem. Eng. J.*, 2022, **429**, 132325.
- Q. Hou, W. Li, M. Zhen, L. Liu, Y. Chen, Q. Yang, F. Huang, S. Zhang and M. Ju, *RSC Adv.*, 2017, **7**, 47288.
- B. Saha and M. M. Abu-Omar, *Green Chem.*, 2014, **16**, 24.
- B. Girisuta, L. P. B. M. Janssen and H. J. Heeres, *Green Chem.*, 2006, **8**, 701.
- R. Weingarten, W. C. Conner Jr. and G. W. Huber, *Energy Environ. Sci.*, 2012, **5**, 7559.
- M. Hartman, O. Trnka and O. Šolcová, *Ind. Eng. Chem. Res.*, 2005, **44**, 6591.
- C. W. Koch, A. Broido and B. B. Cunningham, *J. Am. Chem. Soc.*, 1952, **74**, 2349.
- X. Guo, H. Zhu, Y. Si, X. Lyu, Y. Cheng, L. Zheng, L. Wang and X. Li, *Ind. Eng. Chem. Res.*, 2022, **61**, 7216–7224.
- C. Rosenfeld, J. Konnerth, W. Sailer-Kronlachner, P. Solt, T. Rosenau and H. W. G. van Herwijnen, *ChemSusChem*, 2020, **13**, 3544.



- 50 F. Wang, A.-W. Shi, X.-X. Qin, C.-L. Liu and W.-S. Dong, *Carbohydr. Res.*, 2011, **346**, 982–985.
- 51 K. Beckerle and J. Okuda, *J. Mol. Catal. A: Chem.*, 2012, **356**, 158–164.
- 52 O. Bortolini, A. De Nino, A. Garofalo, L. Maiuolo, A. Procopio and B. Russo, *Appl. Catal., A*, 2010, **372**, 124.
- 53 A. Procopio, R. Dalpozzo, A. De Nino, M. Nardi, B. Russo and A. Tagarelli, *Synthesis*, 2006, **2**, 332.
- 54 S. Kobayashi, S. Nagayama and T. Busujima, *J. Am. Chem. Soc.*, 1998, **120**, 8287.
- 55 P. Zhao, Y. Zhang, Y. Wang, H. Cui, F. Song, X. Suna and L. Zhanga, *Green Chem.*, 2018, **20**, 155.
- 56 P. H. Smith, Z. E. Reyes, C.-W. Lee and K. N. Raymond, *Inorg. Chem.*, 1988, **27**, 4154.
- 57 A. Dibenedetto, M. Aresta, L. di Bitonto and C. Pastore, *ChemSusChem*, 2016, **9**, 118.
- 58 A. R. Mankar, A. Pandey, A. Modak and K. K. Pant, *Renewable Energy*, 2021, **177**, 643.
- 59 R. Häkkinen and A. Abbott, *Green Chem.*, 2019, **21**, 4673.
- 60 P. Makoś, E. Słupek and J. Gębicki, *J. Mol. Liq.*, 2020, **308**, 113101.
- 61 S. Jiang, C. Verrier, M. Ahmar, J. Lai, C. Ma, E. Muller, Y. Queneau, M. Pera-Titus, F. Jérôme and K. De Oliveira Vigier, *Green Chem.*, 2018, **20**, 5104.
- 62 A. Pande, P. Niphadkar, K. Pandare and V. Bokade, *Energy Fuels*, 2018, **32**, 3783.
- 63 S. Meier, *Catal. Sci. Technol.*, 2020, **10**, 1724.
- 64 S. Kobayashi and I. Hachiya, *J. Org. Chem.*, 1994, **59**, 3590.
- 65 T. Kishida, T. Yamauchi, Y. Kubota and Y. Sugi, *Green Chem.*, 2004, **6**, 57.
- 66 S. Kobayashi, S. Nagayama and T. Busujima, *J. Am. Chem. Soc.*, 1998, **120**, 8287.
- 67 C. F. Baes Jr. and R. Mesmer, *The Hydrolysis of Cations*, Wiley, New York, 1976.
- 68 K. B. Yatsimirskii and V. P. Vasil'ev, *Instability Constants of Complex Compounds*, Pergamon, New York, 1960.
- 69 M.-A. del Barrio, J. Hu, P. Zhou and N. Cauchon, *J. Pharm. Biomed. Anal.*, 2006, **41**, 738.
- 70 M. Stupak, V. Kocourek, I. Kolouchova and J. Hajslova, *Food Control*, 2017, **80**, 307.
- 71 E. Weingart, L. Teevs, R. Krieg and U. Prüße, *Energy Technol.*, 2018, **6**, 432.
- 72 M. Mascal, *ACS Sustainable Chem. Eng.*, 2019, **7**, 5588.
- 73 M. Hartman, O. Trnka and O. Šolcová, *Ind. Eng. Chem. Res.*, 2005, **44**, 6591.
- 74 R. Weingarten, J. Cho, R. Xing, W. C. Conner Jr. and G. W. Huber, *ChemSusChem*, 2012, **5**, 1280.
- 75 I. Delidovich and R. Palkovits, *ChemSusChem*, 2016, **9**, 547.
- 76 T. Moeller, D. F. Martin, L. C. Thompson, R. Ferrús, G. R. Feistel and W. J. Randall, *Chem. Rev.*, 1965, **65**, 1.
- 77 S. Jiang, C. Verrier, M. Ahmar, J. Lai, C. Ma, E. Muller, Y. Queneau, M. Pera-Titus, F. Jérôme and K. De Oliveira Vigier, *Green Chem.*, 2018, **20**, 5104.
- 78 X. Zhang, P. Murria, Y. Jiang, W. Xiao, H. I. Kenttämaa, M. M. Abu-Omarb and N. S. Mosier, *Green Chem.*, 2016, **18**, 5219.
- 79 L. Atanda, M. Konarova, Q. Ma, S. Mukundan, A. Shrotri and J. Beltramini, *Catal. Sci. Technol.*, 2016, **6**, 6257–6266.
- 80 W. Guo, Z. Zhang, J. Hacking, H. J. Heeres and J. Yue, *Chem. Eng. J.*, 2021, **409**, 128182.
- 81 N. S. Mosier, A. Sarikaya, C. M. Ladisch and M. R. Ladisch, *Biotechnol. Prog.*, 2001, **17**, 474–480.
- 82 L. Vanoye, M. Fanselow, J. D. Holbrey, M. P. Atkins and K. R. Seddon, *Green Chem.*, 2009, **11**, 390–396.
- 83 H. S. Kim, S.-K. Kim and G.-T. Jeong, *J. Ind. Eng. Chem.*, 2018, **63**, 48–56.
- 84 N. A. S. Ramli and N. A. S. Amin, *Chem. Eng. J.*, 2018, **335**, 221–230.
- 85 K. Kumar, S. Pathak and S. Upadhyayula, *J. Cleaner Prod.*, 2020, **256**, 120292.
- 86 K.-L. Chang, Q. T. Huynh, C.-T. Zhong, W.-R. Chen, H.-Y. Wang, P. Phitsuwan, Y.-C. Lin and G. C. C. Yang, *Environ. Technol.*, 2022, **28**, 102844.
- 87 J. Wang, H. Cui, J. Wang, Z. Li, M. Wang and W. Yi, *Chem. Eng. J.*, 2021, **415**, 128922.
- 88 M. N. Catrinck, E. S. Ribeiro, R. S. Monteiro, R. M. Ribas, M. H. P. Barbosa and R. F. Teófilo, *Fuel*, 2017, **210**, 67.
- 89 A. Takagaki, *Catalysts*, 2019, **9**, 818.
- 90 G. Raveendra, M. Surendarb and P. S. Sai Prasad, *New J. Chem.*, 2017, **41**, 8520.
- 91 X. Wang, T. Lv, M. Wu, J. Sui, Q. Liu, H. Liu, J. Huang and L. Jia, *Appl. Catal., A*, 2019, **574**, 87.
- 92 Q. Li, Q. Li, Q. Li, K. Sun, Y. Shao, S. Zhang, Z. Yan, L. Zhang, Q. Liu, Y. Wang and X. Hu, *Ind. Eng. Chem. Res.*, 2020, **59**, 17554.
- 93 P. Wrigstedt, J. Keskiaväli and T. Repo, *RSC Adv.*, 2016, **6**, 18973.
- 94 C. Wang, L. Zhang, T. Zhou, J. Chen and F. Xu, *Sci. Rep.*, 2017, **7**, 40908.
- 95 W. Guo, T. Kortenbach, W. Qi, E. Hensen, H. Jan Heeres and J. Yue, *Appl. Catal., B*, 2022, **301**, 120800.
- 96 F. Tao, H. Song and L. Chou, *RSC Adv.*, 2011, **1**, 672.
- 97 K.-L. Chang, S. C. Muega, B. Ivan, G. Ofrasio, W.-H. Chen, E. G. Barte, R. R. M. Abarca, M. Daniel and G. de Luna, *Chemosphere*, 2022, **291**, 132829.

