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# PAPER

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# Investigating machine learning models to predict microbial activity during ozonation-biofiltration<sup>†</sup>

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Continuous online monitoring of water treatment process performance is an essential step in ensuring reliable water guality outcomes. In particular, it is important to ensure effective removal of microbial substances during advanced wastewater treatment processes. However, most microbial indicators cannot be continuously monitored by online processes. Therefore, it is necessary to monitor treatment process performance based on surrogate measures which can be reliably and continuously monitored. For example, water guality data such as colour, turbidity and chemical oxygen demand (COD) can be measured quickly and easily. In this study, a combined ozonation-biological media filtration process (O<sub>3</sub>/ BMF), was used to reduce microbial indicator concentration. After gathering water quality data and corresponding microbial indicator concentrations, we applied machine learning to develop models for predicting the amount of change in microbial indicator concentration following O<sub>3</sub>/BMF treatment. Three microbial indicators were studied, namely Clostridium perfringens, E. coli, and somatic coliphage. The most effective physico-chemical predictors for the removal of these microbial indicators were determined by means of mutual information. Associations between changes in the predictors' concentration during O<sub>3</sub>/ BMF and the reduction of the microbial indicators were identified using a range of supervised learning algorithms including Naïve Baves, random forest, support vector machines and generalised linear model. The impact of the type of prediction algorithm on prediction accuracy was investigated and the superior classifier was determined. Performance measures for microbial removal prediction were found to be superior for the support vector machines (SVM) classifier. Using SVM with a Gaussian kernel classifier, prediction accuracy for all microbial removal was above 75%. Moreover, other performance measures such as area under curve (AUC) and kappa statistics (KS) were higher in SVM compared to the other applied classifiers (AUC  $\ge$  0.80; KS  $\ge$  0.34). From this study, we have identified an objective and efficient method that can predict the effectiveness of the O<sub>3</sub>/BMF process in removing the three microbial indicators in water from a short list of commonly measured physico-chemical parameters

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To ensure water quality, online monitoring of pathogen removal is very important which is not possible with current technologies. Therefore, it is essential to find surrogate measures that can be easily monitored online. In this study, we have investigated machine learning techniques to predict microbial removal through surrogates such as water quality data in ozonation and biofiltration processes.

## Introduction

Previous studies have shown that ozonation can effectively reduce microbial concentrations, organic content, colour and trace chemicals in wastewater.<sup>1-4</sup> It has been observed that ozone increases the oxygen concentration and the biodegradability of organic material, facilitating more rapid biodegradation during subsequent biological filtration.

Current international guidelines for water recycling promote a risk management approach for the control of hazards with a primary emphasis on pathogens because of their potential acute, severe and widespread impacts.<sup>5–7</sup> These guidelines draw principles from Hazard Analysis Critical Control Point (HACCP) standards for the monitoring and control of hazards across a multi-barrier system. The

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performance of treatment barriers is evaluated through validation, which is the process of ensuring that the system can effectively control the hazards. For pathogens, this performance is measured by log 10 reduction values (LRV). Log removal value (LRV) is the percentage that a pathogen will be inactivated by a disinfection method. In mathematical for the transformation is the percentage of the transformation of transformation of the transformation of transformation of

A LRV equal to 1 means that the pathogen is reduced by 90% from its initial value. The monitoring of critical control points requires online parameters indicative of the hazard removal performance of the process. For ozonation and biological media filtration (BMF), however the water matrix characteristics play a crucial role in the disinfection effectiveness which has limited the use of indicators such as ozone dose and CT, applied elsewhere in water treatment.<sup>8-10</sup> This limitation has impeded the formal attribution of LRV credits to ozonation and BMF for wastewater in many jurisdictions. LRV credits have been awarded to postozonation at Melbourne Water's Eastern Treatment Plant (ETP) (the subject of this study) on the basis of CT disinfection. Post-ozonation CT disinfection is only possible due to the pre-treatment provided by pre-ozone and BMF primarily through satisfying ozone demand. There are, however reported studies on the use of alternative indicators for monitoring of microbial performance removal including bromate formation, O<sub>3</sub>:TOC ratio, and UVA reduction.<sup>11-13</sup> These studies have focussed on ozonation under ideal conditions, for example by removing suspended solids and using benchtop reactors.<sup>11</sup> The ability to obtain LRV credits for ozonation/BMF could translate into major cost savings for wastewater recycling projects by reducing the need for additional treatment processes such as high-energy photochemical processes, or by reducing chemical consumption for downstream disinfection processes. Alternatively, obtaining additional LRV credits for a treatment system can increase the resilience of recycled water production in the event of sub-optimal performance of one or more pathogen reduction barriers.

Some previous studies have reported efforts to develop models for the prediction of microbial concentration based on some water quality data and operational parameters. Using an on-line UV absorbance analyser, Gerrity (2012)<sup>11</sup> developed a model to predict microbial inactivation during ozonation. Gamage  $(2013)^{14}$  used  $O_3$ : TOC ratio,  $\Delta UV254$  and  $\Delta$ TF to predict the inactivation of three surrogate microbes. Using a linear correlation for the prediction of microbial removal, Gamage reported that  $\Delta UV254$  and  $\Delta TF$  were able to most effectively predict microbial inactivation in ozone/H<sub>2</sub>O<sub>2</sub> systems. Gamage reported high variability in the prediction of E. coli concentration under different dosing conditions. However, traditional regression models might not be the optimum prediction tools for the complex relationships between microbial removal and operational or water quality data. Recently, multivariant predictive models based on Naïve Bayes have been developed to predict disinfection byproducts (DBPs) concentration in drinking water streams.<sup>15</sup> Although a few studies have investigated the prediction capacity of Naïve Bayes in the LRV performance of pathogens from wastewater streams,<sup>16</sup> there are still limited studies on the use of prediction tools for on-line monitoring of water treatment process performance.

The aim of this paper was to evaluate the use of several water quality and operational parameters as surrogates for the removal efficiency of microbial indicators during full-scale ozonation and biological media filtration of secondary treated wastewater. This research assessed previous reported indicators including colour and UVA. The key outcome is the presentation of a method for evaluating useful predictive variables through mutual information and calculating microbial LRVs using these predictors. This outcome is significant because it allows us to perform on-line monitoring to indicate the likely presence or removal of microbial substances in water samples without requiring cumbersome or time-consuming microbial measurements.

## Methodology

#### Full scale wastewater treatment plant

Melbourne Water's Eastern Treatment Plant (ETP) is located 30 km south east of Melbourne, Australia, and treats around 40% of Melbourne's sewage. The ETP is a tertiary wastewater treatment plant which produces around 400 ML per day of high-quality treated water which meets the requirements of both safe discharge to the receiving marine environment and "Class A" recycled water. The ETP process uses ozonation and biological media filtration as an advanced treatment train for secondary treated wastewater. A simplified process diagram of the  $O_3$ /BMF configuration at the ETP is depicted in Fig. 1. A dataset with 105 records of grab sample water quality from secondary effluent and BMF effluent was provided by Melbourne Water and used in developing prediction models.

LRV observations from the data shows that the reduction of microbial indicators *i.e.*, *Clostridium perfringens*, *E. coli*, and somatic coliphage was evidently successful through pre-ozonation and biological media filtration. Fig. 2 shows the average reduction of each microbial indicator after pre-ozonation and biological media filtration (sample point 2).

Water quality data from sample points 1 and 2 were also measured for a range of parameters, including microbial indicators, suspended solids (SS), alkalinity, nitrite, nitrate, ammonia, ultraviolet transmittance (UVT), and colour.

#### Analytical methods

A portable Hach HQ30d meter was used to measure pH and temperature, and a portable Hach 2100Q IS meter was used to measure the turbidity of samples. Hach colourimetric methods with a Hach DR1900 meter were used for the measurement of nitrite, nitrate, COD, and colour. UVT, UVA





and total suspended solids (TSS) were analysed by Australian Laboratory Services (ALS), Environmental Division, Melbourne, Australia. TOC was measured by Australian Water Quality Centre, South Australia. Bromate and Bromide were measured by National Measurement Institute (NMI), North Ryde, New South Wales, Australia. Microbial concentration of *C. perfringens* spores, *E. coli* and somatic coliphage was measured by Australian Water Quality Centre, South Australian Water Quality Centre, South Australia.

#### Prediction model development

The development of the prediction model was undertaken through a supervised learning scheme, as presented in Fig. 3. First, the predictive features and outcome of interest were pre-processed and went through discretisation steps. Then, in the feature selection step, the most useful (predictive) features were identified. The selected features along with the corresponding outcome labels were then used for the classification stage. This process was cross-validated 10-fold, and in each iteration of this validation, 80% of data were used for feature selection and training, while the remaining 20% of data were used for testing the classifier.

The modelling processes applied here incorporates a range of statistical methods. These are summarised, indicating common terminology, acronyms and abbreviations in Table 1.

In the following sections, "features" denote the system parameters which can be easily and continuously monitored (*e.g.*, operational parameters and physico-chemical parameters with online detection). The features included: amount of change (pre  $O_3/BMF$  – post  $O_3/BMF$ ) in total



Fig. 2 log removal reduction of three microbial indicator after preozonation and biological media filtration.

organic carbon (TOC), UV absorbance, UV transmittance, suspended solids, pH, alkalinity, colour, ammonia, nitrate and nitrite concentrations. "Outcome" denotes the amount of change (pre  $O_3/BMF$  – post  $O_3/BMF$ ) in microbial indicators, which are more difficult and laborious to continuously measure. The features are used to make a prediction model, which can predict the outcomes through a supervised learning process.

#### Pre-processing

In the pre-processing step, features with too many missing values were excluded. For a feature to be assessed as useful, at least 25% of the feature values had to be non-missing. For the remaining features, missing values were replaced with mean values from the non-missing values of that attribute.

#### Feature selection

The broad objective of feature selection is to identify the features with greatest predictive value. Finding the most characterising features is a crucial step in many pattern recognition applications.<sup>21,22</sup> Feature selection shows the importance of features for prediction, and how these features are related, and thus minimizes the number of predictors in the final machine learning model and improves the performance of model.<sup>23</sup> The central assumption when using a feature selection technique is that the data contain redundant or irrelevant features. Redundant features are those, which provide no more information than the currently selected features, and irrelevant features provide no useful information in any context.<sup>24,25</sup> The specific purpose of feature selection is to minimize the number of redundant and irrelevant parameters in the final machine learning model to improve performance and generalizability.<sup>26</sup> In this study, most predictive features were selected from the following water quality parameters: suspended solids (SS), alkalinity, nitrite, nitrate, ammonia, ultraviolet transmittance (UVT), and colour. By doing so, a reasonable ratio (1:15) between the number of features<sup>7</sup> and the number of data samples (105) was attained.

In this study we used the minimal-redundancy-maximalrelevance algorithm (mRMR) developed by Peng  $(2005)^{23}$ because of its advantages in terms of both feature selection complexity and feature classification accuracy. The focus of this method is on mutual-information-based feature selection. The mutual information between two random variables *X* and *Y* is defined based on their entropies and



Fig. 3 Flow chart of the data analysis scheme used in the present study.

their probabilistic density functions, as shown in (eqn 1(a)) to (1(c)):

$$I(X, Y) = H(X) - H(X|Y)$$
(1(a))

where H(X) represents the entropy of a discrete random variable *X*, and is defined as the average "surprise" in learning the value of *x* which is equal to the expectation of h(x) with respect to the probability distribution p(x) and is given by

 Table 1
 Key abbreviations and terminology relevant to various classifiers and model validation<sup>16-20</sup>

| Abbreviation | Meaning  | Explanation   |
|--------------|--|---|
| PA           | Prediction accuracy  | Quantifies the number of correctly predicted values divided by the total number of cases  |
| PE           | Prediction error   | Quantifies the number of incorrectly predicted values divided by the total number of cases  |
| KS           | Kappa statistic  | Measures the agreement between model predictions<br>and actual values as a metric in the range [-1, 1].<br>KS = 1 means perfect agreement, KS = 0 means that                            |
|              |  | agreement is equal to chance, and KS = -1 means<br>"perfect" disagreement   |
| AUC          | Area under the curve for the receiver operating characteristic curve (ROC) | AUC ranges between 0 and 1, where 1 represents perfect<br>matching, 0.5 reflects totally random models, and<br><0.5 indicates models generating predominantly inaccurate<br>predictions |
| TPR          | True positive rate   | Rate of correct positive predictions (high reductions)  |
| FPR          | False positive rate  | Failure to detect low reductions when they occurred   |
| TNR          | True negative rate   | Rate of correct negative predictions (low reductions)   |
| FNR          | False negative rate  | Failure to detect high reductions when they occurred  |
| NB           | Naïve Bayes  | Probabilistic classifier based on Bayes theorem   |
| GLM          | Generalised linear model   | Conventional linear regression models for a continuous response variable given continuous and/or categorical predictors   |
| RF           | Random forest  |   |
| SVM/bn       | Support vector machine/binary  | SVM algorithm can find a hyperplane in an <i>N</i> -dimensional space that distinctly classifies the data points using binary kernel function   |
| SVM/GK       | Support vector machine/Gaussian kernel                                     | SVM algorithm that uses Gaussian kernel   |
| SVM/PK       | Support vector machine/polynomial kernel                                   | SVM algorithm that uses polynomial kernel   |
| SVM/rbf      | Support vector machine/radial basis function                               | SVM algorithm that uses radial basis function as the kernel function  |

$$H(x) = -\sum_{x} p(x) \log_2 p(x)$$
(1(b))

In other words, *H* or the entropy of a random variable is the average level of "uncertainty" inherent in the variable's possible outcomes.

And thus from (eqn 1(a)) and (1(b)):

$$I(X,Y) = \iint p(x,y) \log \frac{p(x,y)}{p(x)p(y)} dxdy \qquad (1(c))$$

I(X, Y) is the mutual information between two variable *X* and *Y*. Mutual information measures how much more is known about one random value (*Y*) when given another (*X*). In the context of this paper, *X* is a physico-chemical predictor like colour, and *Y* is a microbial indicator such as *E. coli*.

The mRMR method seeks to maximise the relevance of a feature set for a specific class and minimise the redundancy of all features in the feature set. Relevance is defined by the average value of all mutual information (MI) values between the individual feature  $(x_j)$  and the specific class (c). This is shown in formula (2), where  $(S_{m-1})$  is a feature set with m - 1 features. The task is to select the *m*th feature from the set { $X - S_{m-1}$ }. This is done through an incremental search method by selecting the feature that maximizes the condition inside the square brackets. Redundancy is the average value of all MI values (denoted with *I* in formula (1)) between the individual feature and every other feature in the set  $(x_i)$ .

$$\max_{x_{j} \in X - S_{m-1}} \left[ I\left(x_{j}; C - \frac{1}{m-1} \sum_{x_{i} \in X - S_{m-1}} I(x_{j}; x_{i})\right) \right]$$
(2)

The mRMR algorithm is suitable for unprocessed data, where the features selected in this way will have more or less correlation with each other. This is because mRMR does not intend to select features that are independent of each other. Instead, at each step, it tries to select a feature that minimises the redundancy and maximises the relevance.<sup>23</sup>

Using the mRMR method, for each outcome variable (*i.e.*, microbial indicator concentration), the top features were determined from a random 80% of the data and used for training the prediction model in the next step. Assessment of the number of features' effect on prediction accuracy showed that selection of the top four features resulted in the best performance (ESI† A).

#### Discretisation

The outcome variables (*i.e.*, microbial indicator concentrations) had continuous values. In order to perform multiclass classification, they needed to be discretised. Values of each outcome variable were divided into four quantiles, such that each quantile contained the same fraction of the total population. This approach was taken to ensure a balanced number of elements in each of the four classes.<sup>27</sup>

For an *n*-element vector *X*, quantiles were computed by using a sorting-based algorithm as follows:

1. The sorted elements in *X* are taken as the (0.5/n), (1.5/n), ..., ([n - 0.5]/n) quantiles. For example:

• For a data vector of five elements such as  $\{6, 3, 2, 10, 1\}$ , the sorted elements  $\{1, 2, 3, 6, 10\}$  respectively correspond to the 0.1, 0.3, 0.5, 0.7, 0.9 quantiles.

• For a data vector of six elements such as {6, 3, 2, 10, 8, 1}, the sorted elements {1, 2, 3, 6, 8, 10} respectively correspond to the (0.5/6), (1.5/6), (2.5/6), (3.5/6), (4.5/6), (5.5/6) quantiles.

2. Linear interpolation was used to compute quantiles for probabilities between (0.5/n) and ([n - 0.5]/n).

3. For the quantiles corresponding to the probabilities outside that range, the minimum or maximum values of the elements in *X* was assigned.

As a visual example, Fig. 4 shows the histogram of data values for log removal of variable *E. coli*. The bin edges were set such that it resulted in four quantiles, as explained above. The data that fell in the first quantile (first histogram bin) were labelled class 1, data in the second quantile were labelled class 2 and so on for classes 3 and 4.

For consistency, all the classification algorithms compared in this study used discretised data values.

#### Classifiers

After completing pre-processing, feature selection and discretisation steps, prediction was carried out through several widely used supervised learning classifiers, namely Naïve Bayes (NB), support vector machines (SVM) with binary, Gaussian, radial basis function, and polynomial kernels, generalized linear model (GLM), and random forest (bootstrap-aggregated) decision trees. The definition for each of these classifiers are described below.

**Naïve Bayes.** NB is a simple probabilistic classifier based on Bayes' theorem with the assumption of strong (naïve) independence between the features. NB has the important advantage of simplicity.

**Support vector machines.** SVMs are among the most robust supervised learning prediction methods. The objective of the support vector machine algorithm is to find a hyperplane in an *N*-dimensional space (*N*—the number of features) that distinctly classifies the data points.<sup>28</sup> SVMs have the advantage of being effective in high dimensional spaces, but also effective in cases where the number of dimensions is greater than the number of samples. SVMs are versatile, in that using different kernel functions, they can efficiently perform a non-linear classification, implicitly mapping their inputs into high-dimensional feature spaces.

**Generalized linear model.** GLM usually refers to conventional linear regression models for a continuous response variable given continuous and/or categorical predictors, and include multiple linear regression.<sup>29</sup> The advantage of GLM is being a flexible generalization of ordinary linear regression that allows for response variables that have error distribution models other than a normal distribution.

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Fig. 4 Histogram of log removal values for the outcome variable *E. coli*. Bin edges denote the boundaries of the four quantiles for discretisation.

**Random forest.** Random forest is one of the most popular and most powerful machine learning algorithms. Random forest is a type of bagging algorithm, which aims to reduce the complexity of models that over-fit the training data. The algorithm selects a random subset of predictors to use at each decision split as in the random forest algorithm.<sup>30</sup> Bootstrap-aggregated (bagged) decision trees combine the results of many decision trees, which reduces the effects of overfitting and improves generalization.

The premise is to train a computer model by giving it features along with their corresponding outcomes, such that the model can later predict an outcome based on input features only. The algorithms to train and test the computer model were developed in MATLAB Version (R2019b). The computer algorithms find patterns between features that are exclusively associated with the outcomes.

Cross-validation. The purpose of cross-validation is to test the ability of a machine learning model to predict new data, and to prevent problems like overfitting or selection bias. A 10-fold cross-validation method was used to train and test the classifiers. In other words, the prediction accuracy was computed as the average of ten iterations, where at each iteration, 20% of the data were randomly selected and left out as the test partition and the remaining 80% of the data were used for feature selection and training the classifier. The 80:20 split draws its justification from the well-known Pareto principle.<sup>31</sup> This procedure was repeated 10 times with the training and testing partitions selected randomly each time.17 The model performance was assessed with several performance parameters. These parameters included prediction accuracy, prediction error, kappa statistic, area under the receiver operating characteristic curve (AUC), true positive rate (TPR), false positive rate (FPR), true negative rate

(TNR) and false negative rate (FNR). Average  $\pm$  standard deviation of performance measures over the 10 iterations of cross-validation was calculated.

## Results and discussions

This study yielded a model that learned the multivariate associations between physico-chemical predictors and microbial indicators from a training dataset sampled from Melbourne Eastern Treatment Plant (Melbourne Water). Three specific findings resulting from this study were: 1) identifying the most predictive features; 2) discerning the predictor-outcome associations; and 3) assessing the impact of alternative prediction algorithms.

#### Identifying the most predictive features

Following the feature selection process, the top four features for each microbial indicator were determined as shown in Table 2. These features were the most frequently selected in the 10-fold cross validation. The frequency of each top feature after the cross-validation is also shown in Table 2. UVT, nitrite, nitrate, and colour were the most predictive

Table 2 Top four features for microbial removal during ozonation and biofiltration in Melbourne ETP. The number inside the brackets shows how many times in the 10 fold cross-validation that feature was selected. Features indicate  $\Delta$  values (pre O<sub>3</sub>/BMF – post O<sub>3</sub>/BMF)

| Clostridium perfringens removal | <i>E. coli</i> removal | Coliphage removal     |
|---------------------------------|------------------------|-----------------------|
| UVT <sup>10</sup>               | UVT <sup>10</sup>      | UVT <sup>10</sup>     |
| Nitrite <sup>10</sup>           | Nitrite <sup>10</sup>  | Nitrite <sup>10</sup> |
| Nitrate <sup>8</sup>            | Nitrate <sup>6</sup>   | Nitrate <sup>10</sup> |
| Colour <sup>6</sup>             | Colour <sup>6</sup>    | Colour <sup>9</sup>   |

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features for all of the studied microbial indicators. The reader is reminded that all features were  $\Delta$  values (pre O<sub>3</sub>/BMF – post O<sub>3</sub>/BMF).

#### Predictor-outcome association

The results from our model showed that an increase in  $\Delta$  nitrite (from <0.1 mg L<sup>-1</sup> to >0.4 mg L<sup>-1</sup>) with  $\Delta$ UVT greater than 24% was found to be associated with an increase of *Clostridium perfringens* LRV from 0.5 to 1.3. This is consistent with the fact that high levels of nitrite can stimulate the growth of bacteria, and therefore the effectiveness of the water treatment process in decreasing nitrite concentration is correlated with reduction in microbial concentration of water. An increase in  $\Delta$ UVT (from <19% to >33%), and colour reduction of 53 Pt–Co or more, with  $\Delta$  nitrate >0.34 mg L<sup>-1</sup> was found to be associated with increased *E. coli* LRV from 1.5 to 3.1. An increase in  $\Delta$  colour (from <75% to >87%), and  $\Delta$  nitrite of greater than 0.2 mg L<sup>-1</sup> was found to be associated with increase of coliphage LRV from 1.2 to 3.6.

While these findings are noteworthy and useful as a guideline to the influence of changes in physico-chemical predictors on changes in microbial indicator concentration, it should be noted that the predictors are surrogate measures of the treatment performance and that there is not necessarily a direct relationship between the measurement and the presence of the microbial indicator.

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Many more hidden and intricate associations may exist between these variables that cannot be detected from simple assessment of data. Pattern recognition and sophisticated machine learning algorithms were used to detect those associations and be able to predict the amount of microbial LRV due to  $O_3$ /BMF solely based on measurement of the four predictors before and after the water treatment process.

#### Effect of prediction algorithm

The prediction model, developed in the form of a MATLAB script, took the difference between four physico-chemical measurements (namely UVT, colour, nitrite and nitrate) before and after the  $O_3$ /BMF process as inputs and predicted the microbial removal value range associated with those inputs. Classification was performed using a range of prediction algorithms, which were described in the classifiers section. The performance measures for each of these classifiers are presented in Table 3. Eight performance matrices were compared in Table 3 (abbreviations are described in Table 1). These values were calculated based on 10-fold cross validation for each of the seven prediction algorithms. The calculation method for the reported performance measures is explained in ESI<sup>†</sup> B.

The AUC score varies from 0–1, with 0.5 indicating a totally random model and 1 no error in prediction. AUC < 0.5 denotes models predicting erroneously most of the time.

| Pathogen                    | Performance measure | NB              | GLM                               | RF                                | SVM/bn                            | SVM/GK          | SVM/PK          | SVM/rbf         |
|-----------------------------|---------------------|-----------------|-----------------------------------|-----------------------------------|-----------------------------------|-----------------|-----------------|-----------------|
| Clostridium perfringens LRV | PA                  | $0.73 \pm 0.03$ | $0.92 \pm 0.00$                   | $0.77 \pm 0.03$                   | $0.76 \pm 0.03$                   | $0.78 \pm 0.02$ | $0.75\pm0.02$   | $0.77 \pm 0.03$ |
|                             | PE                  | $0.27 \pm 0.03$ | $\textbf{0.08} \pm \textbf{0.00}$ | $0.23 \pm 0.03$                   | $0.24 \pm 0.03$                   | $0.22 \pm 0.02$ | $0.25\pm0.02$   | $0.23 \pm 0.03$ |
|                             | KS                  | $0.26\pm0.10$   | $0.00\pm0.00$                     | $0.34 \pm 0.09$                   | $0.32\pm0.07$                     | $0.37\pm0.06$   | $0.31\pm0.05$   | $0.33 \pm 0.09$ |
|                             | AUC                 | $0.81 \pm 0.01$ | $0.79 \pm 0.01$                   |                                   | $\textbf{0.71} \pm \textbf{0.04}$ | $0.80 \pm 0.11$ | $0.90\pm0.01$   | $0.76\pm0.08$   |
|                             | TPR                 | $0.47 \pm 0.07$ | $0.00\pm0.00$                     | $0.56\pm0.07$                     | $0.54\pm0.06$                     | $0.61\pm0.05$   | $0.49 \pm 0.05$ | $0.60 \pm 0.08$ |
|                             | FPR                 | $0.18 \pm 0.02$ | $0.02\pm0.00$                     | $0.14 \pm 0.02$                   | $0.14 \pm 0.02$                   | $0.13 \pm 0.01$ | $0.16\pm0.02$   | $0.13 \pm 0.02$ |
|                             | TNR                 | $0.82\pm0.02$   | $0.98 \pm 0.00$                   | $0.86 \pm 0.02$                   | $0.86 \pm 0.02$                   | $0.87 \pm 0.01$ | $0.84 \pm 0.02$ | $0.87 \pm 0.02$ |
|                             | FNR                 | $0.53 \pm 0.07$ | $\textbf{1.00} \pm \textbf{0.00}$ | $\textbf{0.44} \pm \textbf{0.07}$ | $\textbf{0.46} \pm \textbf{0.06}$ | $0.39 \pm 0.05$ | $0.51\pm0.05$   | $0.40 \pm 0.08$ |
| E. coli LRV                 | PA                  | $0.74 \pm 0.02$ | $0.93 \pm 0.00$                   | $0.74 \pm 0.03$                   | $0.75\pm0.03$                     | $0.75\pm0.02$   | $0.75\pm0.03$   | $0.73 \pm 0.03$ |
|                             | PE                  | $0.26 \pm 0.02$ | $\textbf{0.07} \pm \textbf{0.00}$ | $0.26 \pm 0.03$                   | $0.25 \pm 0.03$                   | $0.25 \pm 0.02$ | $0.25 \pm 0.03$ | $0.27 \pm 0.03$ |
|                             | KS                  | $0.29 \pm 0.07$ | $0.00\pm0.00$                     | $0.31 \pm 0.07$                   | $0.34 \pm 0.07$                   | $0.34 \pm 0.06$ | $0.32\pm0.09$   | $0.28 \pm 0.08$ |
|                             | AUC                 | $0.79 \pm 0.01$ | $0.82 \pm 0.01$                   |                                   | $0.73 \pm 0.06$                   | $0.88 \pm 0.01$ | $0.70\pm0.11$   | $0.88 \pm 0.01$ |
|                             | TPR                 | $0.49 \pm 0.05$ | $0.00\pm0.00$                     | $0.49 \pm 0.05$                   | $0.50\pm0.05$                     | $0.52\pm0.05$   | $0.50\pm0.06$   | $0.47 \pm 0.06$ |
|                             | FPR                 | $0.17 \pm 0.02$ | $0.02\pm0.00$                     | $0.17 \pm 0.02$                   | $0.17 \pm 0.02$                   | $0.16\pm0.02$   | $0.17\pm0.02$   | $0.18 \pm 0.02$ |
|                             | TNR                 | $0.83 \pm 0.02$ | $0.98 \pm 0.00$                   | $0.83 \pm 0.02$                   | $0.83 \pm 0.02$                   | $0.84 \pm 0.02$ | $0.83 \pm 0.02$ | $0.82 \pm 0.02$ |
|                             | FNR                 | $0.51\pm0.05$   | $1.00\pm0.00$                     | $0.51\pm0.05$                     | $0.50\pm0.05$                     | $0.48 \pm 0.05$ | $0.50\pm0.06$   | $0.53 \pm 0.06$ |
| Coliphage                   | PA                  | $0.75\pm0.02$   | $0.92\pm0.00$                     | $0.75 \pm 0.04$                   | $0.76\pm0.02$                     | $0.78 \pm 0.02$ | $0.76\pm0.02$   | $0.76 \pm 0.03$ |
|                             | PE                  | $0.25\pm0.02$   | $0.08\pm0.00$                     | $0.25 \pm 0.04$                   | $0.24 \pm 0.02$                   | $0.22\pm0.02$   | $0.24\pm0.02$   | $0.24 \pm 0.03$ |
|                             | KS                  | $0.33 \pm 0.07$ | $0.00\pm0.00$                     | $0.34 \pm 0.09$                   | $0.35\pm0.06$                     | $0.41 \pm 0.05$ | $0.37\pm0.06$   | $0.37 \pm 0.08$ |
|                             | AUC                 | $0.83 \pm 0.01$ | $0.85 \pm 0.01$                   |                                   | $0.82\pm0.04$                     | $0.91 \pm 0.01$ | $0.75 \pm 0.14$ | $0.92\pm0.01$   |
|                             | TPR                 | $0.49 \pm 0.04$ | $0.00\pm0.00$                     | $0.51 \pm 0.07$                   | $0.51 \pm 0.04$                   | $0.56 \pm 0.04$ | $0.53 \pm 0.05$ | $0.53 \pm 0.06$ |
|                             | FPR                 | $0.17 \pm 0.02$ | $0.02\pm0.00$                     | $0.16\pm0.02$                     | $0.16\pm0.01$                     | $0.15\pm0.01$   | $0.16\pm0.01$   | $0.16 \pm 0.02$ |
|                             | TNR                 | $0.83 \pm 0.02$ | $0.98 \pm 0.00$                   | $0.84 \pm 0.02$                   | $0.84 \pm 0.01$                   | $0.85\pm0.01$   | $0.84 \pm 0.01$ | $0.84 \pm 0.02$ |
|                             | FNR                 | $0.51 \pm 0.04$ | $1.00\pm0.00$                     | $0.49 \pm 0.07$                   | $0.49 \pm 0.04$                   | $0.44\pm0.04$   | $0.47 \pm 0.05$ | $0.47 \pm 0.06$ |

 Table 3
 Arithmetic mean ± standard deviation of performance measures from the 10-fold cross validation for three pathogen removal from the dataset of Melbourne ETP (abbreviations in the table are explained in the footnote)

NB: Naïve Bayes, GLM: generalized linear model, RF: random forests, SVM/bn: support vector machines with binary kernel, SVM/GK: support vector machines with Gaussian kernel, SVM/PK: support vector machines with polynomial kernel, SVM/rbf: support vector machines with radial basis function kernel, LRV: log removal, PA: prediction accuracy, PE: prediction error, KS: kappa statistic, AUC: area under curve, TPR: true positive rate, FPR: false positive rate, TNR: true negative rate, FNR: false negative rate.

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Values of 0.5-0.7 indicate poor classification performance; values of 0.7-0.9 indicate fair classification performance, and values higher than 0.9 indicate excellent classification performance. Prediction accuracy is calculated as the total number of correct predictions divided by the total number of cases. This metric ranges between 0 and 100% with higher values indicating better prediction. Cohen's kappa statistic measures the agreement between model predictions and actual values as a metric in the range [-1, 1] considering adjustment due to chance effects.<sup>32</sup> Kappa = 1 means perfect agreement, kappa = 0 means that agreement is equal to chance, and kappa = -1 means "perfect" disagreement.<sup>32</sup> Distinct levels of agreement in the range between 0 and 1 have been defined for kappa coefficient:  $^{33} < 0.2 =$  slight; 0.2-0.4 = fair; 0.4-0.6 = moderate; 0.6-0.8 = substantial; and >0.8 = almost perfect.

Assessing the prediction performance for three microbial communities showed that with the exception of the GLM classifier, the prediction accuracy for other classifiers (including NB, RF and all types of SVM) is around 75%, which is very promising. However, in order to evaluate the performance of a classifier, all performance measures should be considered simultaneously. Performance of a classifier is reliable when the value of both the true positive rate (TPR) and true negative rate (TNR) are greater than 50%. The higher KS and AUC is also representative of a better prediction performance.

Although prediction accuracy of the GLM classifier was above 90%, TPR was around zero and TNR is around 1 which means that relying on prediction accuracy was not sufficient and GLM was not an effective classifier for prediction of microbial removal during these water treatment processes.

With the TPR of less than 50% (47%, 49% and 47% for *clostridium perfringens*, *E. coli* and coliphage LRV, respectively), the Naïve Bayes model was also not a good classifier for any of the microbial communities. On the other hand, support vector machine with Gaussian kernel had above 50% TPR for all three microbial indicators. FPR was also very promising. Moreover, AUC and KS were the highest compared to the values of other classifiers.

For coliphage LRV, although the value of PA, TPR and TNR were almost similar in RF and all types of SVM, however, the AUC and KS was higher in SVM/GK. Therefore, SVM/GK was considered the most suitable classifier among all other classifiers.

The significance of results lies in the capacity of the SVM/GK model to predict the microbial indicator log removal value of a sample with unknown microbial concentration based on its previously learned knowledge and four simple measurements (*i.e.*, UVT, colour, nitrite and nitrate) before and after the  $O_3$ /BMF process. As shown in ESI† A, optimal prediction accuracy resulted with these four predictors. This has great implications for faster and more cost-effective assessment of the efficacy of  $O_3$ /BMF water treatment process for microbial activity removal. The prediction model, developed in the form of a MATLAB

script, takes the four physico-chemical measurements as inputs and calculates the microbial removal value range associated with those inputs.

## Conclusions

Using mutual information and support vector machines, we showed that several surrogate measures can efficiently predict the level of microbial indicator removal during ozonation–biofiltration (O<sub>3</sub>/BMF).

Key findings from this study are:

• Three microbial indicators; *Clostridium perfringens*, *E. coli*, and somatic coliphage have been efficiently removed by the combination of ozonation and biological media filtration.

• Removal of three microbial indicators *Clostridium perfringens*, *E. coli*, and somatic coliphage can be predicted based on physico-chemical measurements.

• Feature selection based on mutual information showed that the top four physico-chemical predictors of microbial indicator removal were UVT, colour, nitrite and nitrate concentrations.

• The best prediction algorithm was found to be support vector machines with Gaussian kernel (SVM/GK), followed by SVM with radial basis function, and random forests.

• Using the SVM/GK classifier, prediction accuracy for all microbial removals was above 75%, AUC  $\geq$  0.80, and kappa statistic (KS)  $\geq$  0.34.

• This prediction model, developed in the form of a MATLAB script, takes the four physico-chemical measurements as inputs, and calculates the microbial removal value range associated with those inputs. Therefore, this model can be used to assess the performance of other systems based on changes in the surrogate measures from pre- to post-water treatment process.

While removal of most microbial indicators during O3/ BMF cannot be continuously monitored by online processes, the methodology discussed in this study provides a fast and cost-effective alternative based on surrogate measures. This is important because continuous online monitoring of water treatment process performance is an essential step in ensuring reliable water quality outcomes.

## Conflicts of interest

There are no conflicts to declare.

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