Organic acid evaporation kinetics from aqueous aerosols: implications for aerosol buffering capacity in the atmosphere†

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The acidity of atmospheric aerosols controls their impacts on heterogeneous and multiphase reactions, cloud formation, and human health. Recently, it has been shown that multiphase buffering can shift aerosol pH substantially compared to bulk solutions. Here, we highlight a unique type of multiphase buffering for aerosols that occurs when organic acids partition from aqueous salt aerosols upon acidification with a strong acid. In this case, rather than lowering the pH of the aerosol, titration with strong acids lowers the organic fraction within the aerosol while maintaining constant pH. We investigate evaporation rates for the model system lactic acid as well as other atmospherically-relevant species such as acetic, butyric, and methacrylic acids. We demonstrate that the timescales for evaporation of organic acids from aerosols are on the order of minutes, comparable to acidification rates in the atmosphere. The organic acid evaporation we observe for lactic acid in salt aerosols is enhanced compared to bulk measurements within what is expected based on differences in surface to volume ratios, indicating surface effects are important. In addition, we show that a salting-out effect drives small organic molecules to the surface, where they quickly evaporate, reducing acidity and causing a "superbuffering" effect. Our results can explain why aerosols in the pH range from 2 to 4 are able to resist further acidification by strong acids in the atmosphere. Overall, this work highlights unique properties of concentrated aerosols and demonstrates how inorganic ions and organic compounds together control multiphase buffering in the atmosphere.

Environmental significance

The pH of aerosols impacts heterogeneous atmospheric transformations, cloud formation and human health. Despite its importance, multiphase buffering, which controls aerosol pH, remains poorly studied. Here, we measured the evaporation rates of several small organic acids from single aqueous salt aerosols and found the timescale to be minutes, which is comparable to the rate of atmospheric acidification. These results reveal a unique type of titration process for aerosols, where pH remains constant but organic fraction decreases upon evaporation of the protonated organic acid following acidification with a strong acid. More generally, our findings show that evaporation rates from highly concentrated particles are dependent on particle viscosity, salting-out and surface effects. Our results highlight some additional considerations for atmospheric chemistry and climate models.

Introduction

Aerosol pH is a critical parameter for atmospheric chemistry. The pH of a particle impacts the surface propensity of fatty and amino acids, multiphase chemical reactions, and even the likelihood of the aerosol to seed a cloud. In addition, highly acidic aerosols have detrimental effects on human health. Since it is challenging to directly measure the pH of aerosols, which can have volumes smaller than one femtoliter, aerosol pH is commonly calculated by thermodynamic models which take aerosol and/or gas phase concentrations as inputs. These models can be useful, but they assume the aerosol is at equilibrium. Limitations of this assumption can potentially lead to erroneous results. Direct measurements of aerosol pH indicate that particles can be rapidly acidified to pH values of 2 to 4 depending on particle size, and several studies show that atmospherically aged aerosols can become very acidic with pH values < 0 while others remain in the pH 1 to 4 range. Another key aerosol parameter is organic fraction. For example, water-soluble organic compounds impact hygroscopicity of aerosols and in turn cloud condensation nuclei activity. Aerosols with a significant organic fraction can adopt a morphology of a highly viscous outer layer enclosing an aqueous core, which impacts aerosol reactivity. The identity of the organics as speciated by mass spectrometry has frequently

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been used for source apportionment to give insight into the origin of aerosols from pollution events and natural emissions. It is also important to track what organics partition from aerosols back into the atmosphere, as these species can then serve as gas-phase precursors for reactions producing additional secondary organic aerosols.

The partitioning of organic acids is of interest in particular as this partitioning influences both aerosol pH and organic fraction. While many studies have been conducted on the uptake of organics by aerosols, the loss of organics through partitioning from the aerosol into the gas phase has received much less attention. A study by Meng et al. used thermodynamic data to show that formic and acetic acids should be found overwhelmingly in the gas phase and are negligible in the aerosol phase. Some data indicate that these acids may be formed by aging aerosols, and thus, we would expect these acids to partition out of the aerosol. This process reduces the organic fraction and increases the pH of the system toward neutral pH by removing the organic acid.

Fig. 1 illustrates this effect and differences in the titration of a bulk solution and an aerosol. For both phases, the initial system is 1 m sodium acetate. Strong acid is then added to neutralize this conjugate base, according to eqn (1).

$$A^- (aq) + H^+(aq) \rightleftharpoons HA(aq)$$ (1)

For the bulk solution, the pH of the system follows a simple titration curve via the Henderson–Hasselbalch equation, shown by the dark line. The aerosol could follow this same pH profile. However, given that the acetic acid that is formed can partition out of the system, the particle could actually remain neutral if all of the formed acetic acid evaporates, as in eqn (2).

$$HA(aq) \rightleftharpoons HA(g)$$ (2)

In fact, the pH of the aerosol could lie anywhere in the shaded area during the titration depending on the rate at which the acetic acid evaporates and the concentration of acetic acid in the gas phase. For example, at 50% acid equivalent added, the pH could be as low as 4.75 (the $pK_a$ of acetic acid) or as high as 7 if all the acidic protons have been taken up by acetate that has then evaporated as acetic acid. At equilibrium, the extent to which eqn (2) occurs depends on Henry’s Law. Complete partitioning to the gas phase only occurs if there is little solubility or if the partial pressure in the surrounding atmosphere is near zero. Therefore, while the partitioning rates discussed here are helpful for estimating timescales of acid evaporation, it should be remembered that complete partitioning to the gas phase only occurs under the conditions described above.

Aerosols therefore have potential for a greater buffering capacity than a corresponding bulk solution of identical composition due to the greater importance of eqn (2). The difference between bulk and aerosol buffering has previously been highlighted by Zheng et al. who showed that the ammonia/ammonium conjugate pair, which buffers bulk solutions at an alkaline pH, has a more acidic peak buffering capacity in aerosols. This assessment was made on the assumption of aerosol equilibrium, but given Fig. 1, it is also important to consider the kinetics of partitioning as well. Recently, Li et al. quantified the kinetics of ammonia depletion for substrate-deposited aerosols mixed with nanoparticles to monitor droplet pH. They found evaporation to take ca. 10 minutes for 20-micron aerosols, showing that the process can be monitored on reasonable timescales. Kohli et al. measured the evaporation of aerosols containing various ethylene glycols and found evaporation on timescales of minutes to hours depending on the conditions. It is also known that small organics such as ethanol evaporate in seconds from single aerosols, and while this can be useful in introducing new species (dissolved in ethanol) to a levitated aerosol and then allowing the ethanol to evaporate, this process is too fast for a detailed kinetics study of organic evaporation.

In this study, we report measurements of the evaporation of organic acids from single, optically levitated micron-sized aqueous aerosols. Our aim is to provide insight into phenomena governing the timescales of the forward reaction in eqn (2), which in turn informs understanding of aerosol pH, organic mass fraction, and the partitioning of atmospherically relevant acids. Specifically, we have investigated the kinetics of evaporation of acetic, butyric and methacrylic acids from individual, optically levitated aerosols using cavity enhanced Raman spectroscopy. We also extensively studied lactic acid as a model system for a more viscous organic acid. Furthermore, we compared the aerosol evaporation rates to bulk solution rates to highlight differences between them. Overall, our goal in this study is to provide an upper limit for how quickly these acids evaporate from single aerosols and to show how different physicochemical processes impact the rate of evaporation due to the concentration of the organic acid and salts in the aerosol phase.
Environmental Science: Atmospheres

Experimental Methods

Solutions were prepared using milliQ water with resistivity > 18.1 MΩ. NaCl (Fisher, Certified ACS) and Na₂SO₄ (Fisher, Certified ACS) were baked at 200 °C for at least 48 h to remove some organic impurities, with remaining organics expected to have minimal impact on the Raman peaks of the highly concentrated samples we studied. Sodium nitrate (Sigma-Aldrich, ReagentPlus), hydrochloric acid (Fisher, Certified ACS, 5.95–6.05 N), methacrylic acid (Thermo Scientific, 99+%), acetic acid (Fisher, ReagentPlus), lactic acid (Fisher, 85% w/w in H₂O, Certified ACS), butyric acid (Fisher, >99%), and dimethylamino (Aldrich, 40 wt% in H₂O) were used without further purification. Samples were created by weighing the solids into a 20 mL scintillation vial with a Teflon cap, then adding the appropriate volume of solvents and dissolving the solids via agitation. Measurements of pH were carried out with a pH meter (Oakton Instruments) calibrated with pH 4 and 7 buffers.

Concentrations reported in this manuscript are for bulk solutions. Aerosols generated from these solutions can have notably higher concentrations, which we have quantified as enrichment factors (EFs) in previous work where we observed EFs of ca. 1 to 1.8 at an RH of 90%. Here, working at 80% RH, the EFs are likely slightly larger, since the equilibrium concentration of NaCl at 80% RH is 5.1 m.

For aerosol-phase experiments, single micron-sized aerosols were trapped using a commercial Aerosol Optical Tweezers 100 (Biral Inc.). Solutions were nebulized using an ultrasonic nebulizer (MicroAIR U22, OMRON) and self-coalesced in the optical trap to obtain a particle of the appropriate size, typically with a radius of 4 ± 1 µm. Precise control of aerosol size was not necessary since curvature effects have little importance in this micrometer size regime and past studies have indicated evaporation processes to be nearly size-independent even for submicron aerosols. Relative humidity (RH) was maintained at 80 ± 8% by a 30 sccm flow of a mixture of wet and dry N₂ gas. A faster flow rate was avoided to prevent unnecessary convection, which has previously been recognized as a potential source of degassing acceleration. Nebulized aqueous salt aerosols are emitted at ca. 100% RH, so water equilibration times to an 80% RH environment are expected to be less than one second. Temperature was typically 24.6 ± 0.1 °C and minimal heating from the 532 nm laser is expected. Once aerosols were trapped, Raman spectra were collected every second on the 1200 g mm⁻¹ setting, with the spectrum centered on either 570 nm to obtain kinetic data from the Raman band of interest (usually the C=C stretch, specifically the C=COOH peak, between 800 and 930 cm⁻¹) were normalized to this peak. The WGM interference from the kinetics data was also removed, and then the resulting dataset was fit to a ln(A₁/A₀) vs. time function, where A is the peak area at a given time and A₀ is the initial peak area. We note that spectral frequencies labeled on figures are provided for convenience and may shift by a few wavenumbers between experiments due to differences in ionic strength, solution composition, or bulk vs. aerosol phase. All post-Igor processing was carried out using custom MATLAB scripts.

The refractive index of solutions was measured with an ABBE-3L refractometer (Bausch & Lomb). About 0.5 mL of each solution was pipetted onto the refractometer prism and a sodium lamp was used to illuminate each sample for alignment. Pictures of the illuminated images were taken with a smartphone camera and processed in Inkscape to count the pixels per refractive index unit and calculate refractive index. We found this approach to be more precise and reproducible compared to estimating the refractive index by visual inspection. In the AOT, the refractive index was measured from the WGM bands on the water O–H stretch at 650 nm. Since bulk measurements were performed at the standard 589 nm, the AOT refractive index was converted to this wavelength using eqn (3).

\[ n_a = n_b + D_1 \left( \frac{1}{n_b} - \frac{1}{n_b} \right) + D_2 \left( \frac{1}{n_b} - \frac{1}{n_b} \right)^2 \]  \hspace{1cm} (3)

Here, \( n \) is the refractive index, \( \lambda \) is wavelength, and \( D_1 \) and \( D_2 \) are the first and second dispersion terms, which are also measured by...
the AOT. Refractive index data for lactic acid and lactate are shown in Fig. S1.

Surface tension data was obtained with an AquaPi tensiometer (Kibron) with Teflon sample cups. The tensiometer was calibrated with milliQ water to 72.8 ± 0.1 mN m⁻¹. Each pair of salt solution and salt solution + organic acid was measured repeatedly. The first reading from the tensiometer is designed to be discarded, so repeated measurements were performed after the first measurement until at least two measurements agreed within the instrument error of ± 0.1 mN m⁻¹. The tensiometer was cleaned with milliQ water and a butane torch between measurements.

Results and analysis

Lactic acid experimental results

In order to quantify organic acid evaporation from aerosols, we first determined which Raman peaks should be monitored to detect to the presence of the acid, the conjugate base, and both species. Peak assignments used here are primarily based on previous studies. In the case of lactic acid, which we consider first, the stretching mode of the carbon–carbon single bond adjacent to the carboxylic acid group (i.e. the C-COOH stretching mode) is strong in the Raman spectra. Its frequency is 830 cm⁻¹ is well-resolved and there is a clear shift for the corresponding peak for the base (i.e. the C-COO⁻ stretching mode) at 856 cm⁻¹ (see Fig. 2a). Therefore, this was the peak used for monitoring the presence of lactic acid in the aerosol without interference from lactate. The 1730 cm⁻¹ peak due to the protonated C=O in the carboxylic acid group (C=OOH) is broad and weak in the Raman spectrum and therefore difficult to use for kinetic analysis. Nevertheless it is qualitatively useful to corroborate lactic acid evaporation from the aerosol (see Fig. 2b). We used sodium sulfate as our internal standard (980 cm⁻¹ band), which in our pH range (≈3.2) is non-volatile as a dianion and works well as a standard.

Lactic acid evaporates very slowly from bulk solutions which is unsurprising given its boiling point is greater than that of water. Spectra were collected over a 13 hours timescale. After ca. 11 hours of evaporation, lactic acid begins to crystallize, complicating the kinetics. In addition, water evaporation is significant on this timescale, which would increase the lactic acid concentration and hence the lactic acid signal. To account for this, we tracked the peak area of the C-COOH 830 cm⁻¹ band normalized to the sulfate internal standard (Fig. S2†). By evaluating only data before lactic acid crystallization, the bulk evaporation rate for 1.5 m lactic acid in a 1.5 m NaCl solution was determined to be 4.8 ± 0.3 × 10⁻⁶ s⁻¹.

Next, we performed experiments on the same solution but for micron-sized aerosols. Representative spectra are given in Fig. 2b. The decrease in the C-C peak area versus the internal standard peak area (see Experimental methods for details) fit first-order kinetics (Fig. 2e). The slope yields a rate constant k for the initial loss of lactic acid. From replicate experiments, k was found to be 1.2 ± 0.3 × 10⁻⁵ s⁻¹, which is 2.6 × 10⁻² faster than the corresponding bulk evaporation. The surface area to volume (hereafter S : V) ratio for the bulk system is ca. 3.1 cm⁻¹. The S : V ratio for a 4 micron aerosol is 7500 cm⁻¹. Thus, the aerosol S : V ratio is greater by a factor of 2.4 × 10³. This number is within the evaporation rate ratio of 2.6 ± 0.7 × 10³ with propagated error. Therefore, the enhancement seen for aerosol-phase evaporation is well-described by the high S : V ratio and is seen to scale with this ratio. We emphasize that this comparison is based on bulk solution concentrations. The aerosol is more concentrated in NaCl than the bulk, and future work comparing aerosols and bulk solutions at the same final salt concentrations may be insightful. However, since water evaporates faster than lactic acid from bulk solutions, the direct comparison of the two phases may not be exact and thus should not be overanalyzed (and would be challenging, as a 5.1 m NaCl solution would likely begin forming solid NaCl crystals by the time enough lactic acid evaporated for a reliable k value to be calculated). The key point here is that the increase in aerosol rate is comparable to the S : V ratios of the phases.

Lactic acid analysis: viscosity and salting-out

To gain insight into the factors controlling the rate of organic evaporation from aerosols, we performed experiments varying the concentration of lactic acid and sodium chloride in the bulk solutions used to prepare the aerosols. The results are shown in Fig. 3a.

At a given [NaCl], when the [lactic acid] decreases the rate constant increases. This may seem surprising, given that first-order rate constants are insensitive to concentration. However, here, a decrease in lactic acid substantially decreases the viscosity of the aerosol, which decreases the resistance experienced by a molecule moving through the aerosol and to the aerosol surface to evaporate. Viscosity and diffusion are discussed further in the section comparing our results with theory (vide infra).

Fig. 3a also shows the changes in the rate of lactic acid evaporation from the aqueous aerosol as a function of sodium chloride concentration. Faster rates are found for higher concentrations of NaCl. This likely has to do with a “salting-out” effect where inorganic ions and organic molecules compete for hydration and organics are thus more easily lost from the aqueous phase. The literature on salting out effects is complex. For NaCl, salting out has been observed for a wide range of molecules. For pure acetic acid aerosols, however, it has been found that adding NaCl does not impact the rate of organic evaporation, probably due to the already very high volatility of acetic acid. Theoretically, the equilibrium behavior caused by salting out can be quantified by Sechenov coefficients, but unfortunately these coefficients are scarce in the literature and not available for lactic acid. To determine if salting out was occurring here, we measured the surface pressure, π, for a series of solutions of 1 m lactic acid with increasing [NaCl] (Fig. 3b). Eqn (4) provides a definition of π:

\[ \pi = \gamma_0 - \gamma \]  

where, \( \gamma_0 \) is the surface tension of an NaCl solution and \( \gamma \) is the surface tension of the same solution with 1 m lactic acid. The data show that NaCl increases the surface activity of lactic acid.
In particular, in a solution with 4 m NaCl, the surface activity of lactic acid doubles compared to a solution with no NaCl. With this increased surface activity, the lactic acid is more likely to reside at the interface where it can readily evaporate from an aerosol. Furthermore, the increase in viscosity due to added NaCl is completely negated by the salting-out effect. Indeed, a 4 m NaCl solution has a viscosity of 1.35 cP, while a 2 m lactic acid solution has a viscosity of ca. 1.31 cP. While this level of viscosity decreased the rate of evaporation for lactic acid particles as discussed above, the rate of evaporation nevertheless increases with increasing [NaCl], indicating that salting-out is a key factor for the prediction of organic partitioning.

To determine whether salting-out by NaCl is enhanced in the aerosol phase, ideally, the kinetics of bulk solutions and aerosols free of NaCl should be compared. In practice, we found NaCl-free lactic acid aerosols to be too unstable to provide reliable data. Instead, we extrapolated the 2 m lactic acid solution data shown in Fig. 3a to a 0 m NaCl concentration and obtained a $k$ of $1.8 \pm 0.5 \times 10^{-3}$ s$^{-1}$. The corresponding bulk kinetics experiment with 2 m lactic acid and no NaCl gave a $k$ of $2.0 \pm 0.7 \times 10^{-6}$ s$^{-1}$. This corresponds to an aerosol-phase enhancement of 900. This enhancement is significantly below the 1.5 m lactic acid 1.5 m NaCl enhancement of 2600 $\pm$ 700. Therefore, at least for the case of NaCl, salting-out may be enhanced in the aerosol phase and could accelerate evaporation kinetics, although the S : V ratio remains the most important benchmark for determining enhancement.

**Results for other atmospherically-relevant molecules**

The behavior of methacrylic acid was found to be similar to lactic acid. Unfortunately, due to solubility limitations and the requirement of high concentrations in the aerosol phase to obtain quantitative data with Raman spectroscopy, it was not practical to carry out concentration- or salt-dependent studies for methacrylic acid. However, we were able to obtain reproducible data for aerosols generated from bulk solutions of 0.67 m methacrylic acid and 0.67 m sodium sulfate at pH 2.9. Representative spectra are given in Fig. 4 (see Fig. S3† for bulk spectra), where the loss of the 808 cm$^{-1}$ peak corresponds to the loss of methacrylic acid. From replicate experiments we obtained a first order rate constant of $0.029 \pm 0.014$ s$^{-1}$. The corresponding rate constant for a bulk solution (see data

![Fig. 2 Lactic acid evaporation. (a) Comparison of aerosol (top traces) and bulk (bottom traces) Raman spectra. Bulk solutions were 1 m lactic acid, neutralized with KOH if lactate was needed, and aerosol solutions were created by nebulizing these solutions. Several Raman frequencies are labeled (see Discussion S1† for peak assignments). (b) Raman spectra are collected as a function of time from an aerosol generated from a 1.5 m lactic acid 1.5 m NaCl solution. It is seen that lactic acid evaporates in 150 s as indicated by the 830 cm$^{-1}$ C–COOH peak. Sulfate (980 cm$^{-1}$) was used as the internal standard. Sharp frequencies in the bottom spectrum correspond to WGMs. (c) First-order kinetic fit to the loss of the C–COOH peak area in lactic acid at 830 cm$^{-1}$. The fit has a slope of $-0.016$ s$^{-1}$ and an $R^2$ of 0.97.](image-url)
provided in ESI† is 3.69 ± 0.06 × 10⁻⁵ s⁻¹. This represents an aerosol-phase acceleration factor of ca. 800, which is fully accounted for by the difference in the S:V ratios.

We anticipate salting-out to be less important for methacrylic acid. Based on surface tension measurements (Fig. S4†), methacrylic acid has a higher π value than lactic acid for the same concentration of sodium chloride. However, the slope of the π vs. [NaCl] plot is smaller, showing that methacrylic acid is less sensitive to salting out as [NaCl] increases. This illustrates the possibility that molecules which are already highly surface active will be less sensitive to salting-out, which makes sense given the propensity of these molecules to be at the surface where there is a limited hydration sphere at the air–water interface.

Next, we consider aerosol evaporation data from two other carboxylic acids, namely acetic and butyric acid. The bulk Raman spectra for butyric acid is shown in Fig. 5a. The carbon–carbon symmetric stretch observed for the C–COOH and C–COO⁻ groups in butyric acid and butyrate at 868 and 878 cm⁻¹, respectively, changes slightly over time. There are also several peaks due to the symmetric and asymmetric bending modes of CH₂ and CH₃ groups in the 1400 to 1480 cm⁻¹ range. Rather than resolve them individually, we simply note the ratio of the maxima at 1414 and 1453 cm⁻¹ is sensitive to pH as shown in Fig. 5a. An additional note for these experiments is that we chose sodium nitrate as the internal standard due to its very strong Raman band at 1050 cm⁻¹ which is useful for experiments involving coalescence (see ESI†).

The C–COOH and C–COO⁻ stretching mode peaks at 868 and 878 cm⁻¹, respectively, can be resolved by peak fitting in confocal spectroscopy, but due to lower resolution of the Raman spectrometer in the aerosol optical tweezers, these peaks are found to be less reliable for quantitative analysis. The presence of butyric acid can be qualitatively seen by the 868 cm⁻¹ shoulder which decreases in subsequent spectra as the acid evaporates. This phenomenon can be more clearly seen in the ratio of the 1414 and 1453 cm⁻¹ maxima. When the aerosol is first trapped, the 1453 cm⁻¹ peak has greater intensity, indicating the system contains more butyric acid than butyrate. After 70 seconds, the peaks have nearly equal intensity. At this
At any moment in time, ca. 7.3% of the molecules are within 100 nm of the interface, while for the bulk phase in our conditions, only 0.003% are. This leads to more variations for bond environments in the aerosol phase. A clear example of the varying peak prominence between phases can be seen in Fig. 5 in the 1715 cm$^{-1}$ COOH peak, corresponding to butyric acid, which is more visible in the bulk phase. Nevertheless, in order to determine a lower bound rate constant, we assumed the ratio of the peaks corresponded to the same acid to base ratio as the bulk and calculated a rate constant of 0.02 s$^{-1}$. Comparing this to the bulk rate of 2.70 ± 0.05 × 10$^{-5}$ s$^{-1}$, the aerosol evaporation rate is enhanced by a factor of ca. 103.

Acetic acid was found to evaporate faster than butyric acid. We were unable to obtain individual spectra giving evidence that acetic acid was present in the aerosol phase, and the acidification of sodium acetate led to loss of all organic signal faster than the time required for the aerosol to stabilize (ca. 3 seconds). We therefore conclude that acetic acid evaporation likely occurs on the timescale of milliseconds or faster and that it is unlikely to be detected in atmospheric aerosols, consistent with predictions. Acetic acid may be detected in aerosols with higher organic fractions and at lower RH conditions.

To demonstrate that the observed evaporation was not a function of other experimental variables and conditions such as surrounding water vapor slowly replacing organics in the aerosol, we performed experiments with the dimethylammonium cation, DMAH, a charged protonated species that does not evaporate from the aerosol or bulk solution, as a control experiment. Dimethylamine, DMA, has been observed as a significant component of particles and cloud water. Given that the pKa of DMA is ca. 10.7, it is expected to exist almost exclusively as DMAH in atmospheric aerosols. We aerosolized a 2 m DMA/DMAH solution with an initial pH of 10.6, corresponding to slightly more DMAH than DMA in the initial solution. By comparison to the bulk spectrum of this mixture (Fig. S7†), it can be seen that DMAH but not DMA is present in the aerosol. We infer that DMA partitioned away during the nebulization process, leaving only DMAH in the aerosol phase.

Fig. 5 Raman spectra of butyric acid. (a) Confocal spectra with insets showing spectral differences at various pH levels. The broad boxed COOH band at 1715 cm$^{-1}$ also shows pH-dependence. From top to bottom, the pH values are 5.60, 5.07, 4.82, 4.77, 4.64, 4.27, 4.07 and 3.41. The uncertainty for each is ± 0.04 pH units. (b) Aerosol phase traces (averages of 11 spectra each) showing butyric acid evaporation and pH increase. The sharp features correspond to WGMs.

Fig. 6 Aerosol spectra of dimethylammonium cation at time t = 0 and t = 110 min. Spectral bands do not significantly change over 110 minutes. Traces show an average of 70 spectra each to show peaks more clearly, which creates the oscillation-like artifact in the 110 min spectrum due to movement of WGMs.
Over 110 minutes, the DMAH signal does not significantly change, indicating that this ion does not partition into the gas phase, nor does transient DMA (from the dynamic equilibrium of DMAH with DMA) escape at a fast enough rate to slowly deplete the aerosol of DMAH (Fig. 6). Our finding is therefore consistent with the observation of DMAH in real atmospheres and cloud water.

**Comparison of results with theory**

We have shown here that organic acids can partition from aerosols on fast timescales, often on the order of minutes, potentially contributing to multiphase buffering. To put this timescale in context, we calculated a theoretical rate of acidification for a 4-micron radius aqueous aerosol. We considered only ambient concentrations of the strong acids HCl at 1 ppb, HNO₃ at 1 µg m⁻³, and H₂SO₄ at 10⁶ molecules/cm⁻³. We used a wind speed of 15 km h⁻¹, just above the minimum associated with the formation of waves and hence e.g. sea spray aerosols. This allowed the calculation of the number of acidic molecules the aerosol would collide with as it travels through a volume equal to the cylinder swept out by its area. Assuming no buffering, this creates an aerosol at pH 4.4 in 1 second and pH 2.3 in 2 minutes. This means that acidification can be slow enough that the evaporation of newly-protonated organic acids would be slow enough that the evaporation of newly-protonated organic acids

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correlation, this $\Delta H_v$ value corresponds to a saturation concentration of $2.8 \times 10^5$ kg m$^{-3}$, which follows from the fact that lactic acid is both volatile (b.p. = 122 °C) and miscible in water.\textsuperscript{57,58} We also note that our simplified model does not include temperature effects. For an atmospheric aerosol, evaporation could substantially cool the aerosol, decreasing the rate of evaporation. This effect would lead to an increase in agreement between these measurements and model results.

To highlight the importance of enhanced partitioning, we next compared our experimental findings to predictions from E-AIM Model III.\textsuperscript{39-61} We used the ratio of moles from each bulk solution and an RH of 80%. Lactic acid was input as a singly dissociating acid using a $p\mathcal{K}_a$ of 3.86 and a Henry’s Law constant of $7.1 \times 10^7$ mol kg$^{-1}$ atm$^{-1}$.\textsuperscript{62,63} Using this value and Henry’s Law, a 1.5 m lactic acid solution would have a vapor pressure of 5.1 $\times$ 10$^{-8}$ atm. For our systems, E-AIM predicted (molality-based) pH values ranging from 1.43 to 1.71 (see Table S1). This is striking, given that this is an equilibrium model, and our data indicates that the majority of lactic acid partitions away by the time the aerosol reaches equilibrium. Even assuming no further lactic acid evaporates past our limit of detection (which is unlikely), the aerosol pH would be at least as high as 2.4 and could theoretically even be neutral. The situation is similar for methacrylic acid, with an E-AIM predicted pH of 2.5 and a minimum experimental pH of 2.8. This demonstrates that thermodynamic models can underestimate pH when Henry’s Law and volatility are both high. While E-AIM is likely predicting a pH close to what would be observed in a bulk solution with the majority of lactic acid remaining in the aqueous phase, we have found that lactic acid can be efficiently salted out of aerosols and therefore is primarily present in the gas phase. Thus, care should be taken when assessing the final composition of aerosols with large fractions of volatile components. Indeed, while equilibrium partitioning may describe the formation of organic-containing aerosols well, the properties of the formed aerosol can be quite different from equilibrium predictions.\textsuperscript{64}

Using our data, we can categorize the lifetime of an aerosol based on its pH. Using sea spray aerosols as an example, these particles are rapidly acidified to pH 2–4 within minutes.\textsuperscript{11,12} At this point, the aerosol pH drops below the $p\mathcal{K}_a$ value of many organic acids and their volatilization causes multiphase buffering, preventing the aerosol pH from dropping further. Buffering from nonvolatile species such as charged amino acids may help maintain the aerosol pH of ca. 2.\textsuperscript{66,67} This can explain how both fresh and aged sea spray aerosols have similar pH levels. Neutralization by ammonia may also occur, although depending on the aerosol liquid water content, due to partitioning, the peak buffering capacity of the ammonia/ammonium system may be at an acidic pH.\textsuperscript{68} Over many days, the multiphase buffering capacity may eventually be exhausted, allowing aerosols to equilibrate with strong acids in the atmosphere and reach pH < 0.9. Thus, for areas producing aerosols with a high fraction of small organic acid molecules, the pH can be expected to remain near the average $p\mathcal{K}_a$ of these acids for most of the aerosol’s lifetime since organic partitioning is so fast that the process will rarely be kinetically limited. For aerosols with a low fraction of these organic acids or their conjugate bases, the particle will more quickly drop to a very acidic pH and have more harmful effects on human health.

These findings give some additional insights into experimental methods that utilize small organic molecules. Some mass spectrometry and liquid jet methods rely on organics as solvents for analytes.\textsuperscript{69} The fast evaporation of these organics, and the resulting concentration of the analytes, should be accounted for when considering the data. Recently, the phenomenon of reaction acceleration in the aerosol phase has been a topic of considerable interest.\textsuperscript{70-72} We have shown that organic acids can evaporate from aqueous salt aerosols faster than would be predicted from a $S:V$ or viscosity analysis alone, with salt-out likely contributing to this enhancement. Therefore, salt-out effects should be quantified when comparing reaction rates in aerosols or liquid jets to bulk solutions.

Conclusions and implications

We have shown that small organic acids evaporate rapidly from aqueous salt aerosols in conditions of low gas phase concentrations over timescales ranging from milliseconds to minutes depending on the specific organic acid. Evaporation rate can be slowed by increases in viscosity but, despite the fact that increases in NaCl increase viscosity, evaporation rate is accelerated by NaCl due to salting-out effects. Given the lifetime of typical atmospheric particles, small organic acids leave very quickly.\textsuperscript{55} This illustrates the possibility of a distinct type of titration that can occur for aqueous aerosols. In a typical bulk-aqueous phase titration of a conjugate base, as acid is added, the pH decreases and the organic fraction remains constant. For an aerosol composed of conjugate base, however, the pH actually remains constant as the organic fraction decreases since the organic acid partitions rapidly into the gas phase. The released organic acids could then react with atmospheric amines to form secondary organic aerosols.\textsuperscript{39}

Author contributions

Kyle Angle and Vicki Grassian designed the experiments. Kyle Angle and Christopher Nowak performed the experiments. Kyle Angle analyzed the data and carried out calculations. Kyle Angle and Vicki Grassian prepared the manuscript.

Conflicts of interest

The authors declare no competing interests.

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Environmental Science: Atmospheres


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