

Cite this: *Dalton Trans.*, 2023, **52**,  
6152

## Multi-technique structural analysis of zinc carboxylates (soaps)†

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A series of medium- and long-chain zinc carboxylates (zinc octanoate, zinc nonanoate, zinc decanoate, zinc undecanoate, zinc dodecanoate, zinc pivalate, zinc stearate, zinc palmitate, zinc oleate, and zinc azelate) was analyzed by ultra-high-field <sup>67</sup>Zn NMR spectroscopy up to 35.2 T, as well as <sup>13</sup>C NMR and FTIR spectroscopy. We also report the single-crystal X-ray diffraction structures of zinc nonanoate, zinc decanoate, and zinc oleate—the first long-chain carboxylate single-crystals to be reported for zinc. The NMR and X-ray diffraction data suggest that the carboxylates exist in three distinct geometric groups, based on structural and spectroscopic parameters. The ssNMR results presented here present a future for dynamic nuclear polarization (DNP)-NMR-based minimally invasive methods for testing artwork for the presence of zinc carboxylates.

Received 19th January 2023,  
Accepted 14th April 2023

DOI: 10.1039/d3dt00184a

rsc.li/dalton

### 1. Introduction

Carboxylates of polyvalent metals, colloquially referred to as metallic soaps, are of significant commercial and scientific interest. Palmitates, stearates, oleates, and linoleates of lead, zinc, and aluminum are common additives to paints, varnishes, and lacquers.<sup>1–4</sup> Metal salts of fatty acids are widely used as precursors in the formation of colloidal semiconductor nanocrystals.<sup>5</sup> Soaps of certain heavy metals, such as those of zinc, form *via* saponification in oil paintings and are of concern to cultural heritage conservators.<sup>6–12</sup>

Zinc oxide is of particular interest to the technological/industrial sectors and art conservation community. ZnO has many applications such as gas detection for environmental protection,<sup>13</sup> modulation of light-emitting diodes,<sup>14</sup> and as

nanostructures in biomedical sensors.<sup>15</sup> Zinc oxide is a widely used pigment commonly known as zinc white.<sup>16</sup> Although zinc oxide has been known since antiquity as a byproduct of brass production, it was not used as a pigment until the late 18<sup>th</sup> century when concerns over the toxicity of its primary predecessor, lead white, began to grow.<sup>17</sup> Unlike technological applications that necessitate strict control of composition and environment, paint systems present often unknown environments regarding the form and composition of the zinc components that necessitates further study.

Paintings are non-equilibrium heterogeneous multilayer systems composed of various reactive organic and inorganic materials. Environmental conditions such as fluctuations in temperature (T), relative humidity (RH), and exposure to light, particularly UV, or treatment with aqueous or organic cleaning agents may trigger chemical changes, resulting in deterioration of the paints.<sup>18–23</sup> Metal soap formation is the cause of widespread visible deterioration of oil paintings dating from the 15<sup>th</sup> century to the modern era, including iconic works such as the *Anatomy Lesson of Dr. Nicolaes Tulp* by Rembrandt van Rijn from the 17<sup>th</sup> century<sup>24</sup> and *Couple with Their Heads Full of Clouds* by Salvador Dali from the early 20<sup>th</sup> century.<sup>25</sup>

Traditional oil paints are basically composed of pigments mixed with a binder. Traditional oil-based binders consist of drying oils, most commonly linseed oil.<sup>26</sup> Linseed oil, derived from the flax plant, consists of a mixture of triglycerides containing both unsaturated and saturated fatty acids. A typical composition of fresh pressed linseed oil is shown in Table 1; however composition can vary due to geographic origin of the

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† Electronic supplementary information (ESI) available. CCDC 2088849–2088851.

For ESI and crystallographic data in CIF or other electronic format see DOI:

<https://doi.org/10.1039/d3dt00184a>



**Table 1** Typical triglyceride fatty-acid composition of linseed oil<sup>26</sup>

Fatty acid	Percentage
Linolenic acid	57.6
Oleic acid	17.0
Linoleic acid	16.4
Palmitic acid	5.9
Stearic acid	3.1

flax plant.<sup>26</sup> The polymerization of unsaturated acyl chains *via* cross-linking reactions is a relatively rapid process and the paint layer cures (dries) within weeks.<sup>27</sup> The hydrolysis of triglyceride ester linkages is a much slower process. Saturated components like palmitic and stearic acid are not immobilized by cross-linking reactions, but rather are “free” to move about the paint matrix. Saponification occurs when these free fatty acids react with metal ions that have leached into the paint matrix or directly from pigment particles or additives. The soaps then aggregate and can form microscopic inclusions that deform the paint layers from within, or erupt through the paint surface as protrusions, or form on the surface as hazy films, or form between paint layers causing delamination and flaking.<sup>28–31</sup>

We<sup>32–35</sup> and others<sup>36–38</sup> have previously shown that one may identify and quantify lead soaps produced in paints containing lead white, 2PbCO<sub>3</sub>·Pb(OH)<sub>2</sub>, and pigments such as lead tin yellow type I (PbSnO<sub>4</sub>) *via* solid-state <sup>207</sup>Pb NMR spectroscopy, to specify the chemical structure of the lead-containing pigments and soaps, as well as to monitor their reactions with the linseed oil binder. Barannikov *et al.* have utilized a multi-technique analysis to assess the structure and reactivity of mercury soaps to model paintings containing cinnabar (HgS).<sup>39</sup> In this report, we provide the first <sup>67</sup>Zn NMR spectra of pure zinc soaps, demonstrating that high-field <sup>67</sup>Zn solid-state spectroscopy of soaps is an analytical technique to quantify and follow the formation of zinc soaps in paintings. <sup>67</sup>Zn (*I* = 5/2) is a low- $\gamma$  quadrupole nuclide with relatively low natural abundance (4.1%) and large quadrupolar moment ( $Q = 0.122 \times 10^{-28} \text{ m}^2$ ), which results in low sensitivity and broad resonances. Despite these difficulties, solid-state NMR studies of metallic zinc,<sup>40</sup> zinc salts,<sup>41–44</sup> organozinc complexes,<sup>45,46</sup> proteins,<sup>47,48</sup> and MOFs<sup>49,50</sup> have been reported. Although the interest in zinc carboxylates has been growing, there is a lack of single-crystal structural data for such compounds. Crystal growth of these compounds is difficult due to their low solubilities and long carboxylate chains. Furthermore, when crystals are produced, they are often small, thin, fragile plates that are both hard to handle and challenging to mount. We report the first single-crystal X-ray-determined structures of zinc nonanoate, zinc decanoate, and zinc oleate, representative of the distinct groups in which these solid structures may form.

## II. Experimental

### Synthesis of materials

Zinc carboxylates of octanoic (C8), nonanoic (C9), decanoic (C10), undecanoic (C11), dodecanoic (C12), hexadecenoic (pal-

mitic, C16), octadecanoic (stearic, C18), (9Z)-octadecenoic (oleic, Z-C18), nonanedioic (azelaic, Aze) and 2,2-dimethylpropanoic (pivalic, Piv) acids were synthesized *via* methods adapted from previously established protocols.<sup>18,32,51,52</sup> Typically, an appropriate acid was dissolved in 50 mL (C8, C9, Z-C18, Az) or 100 mL (C10, C11, C12, C18) of ethanol, or 50 mL water (Piv). This solution was mixed with an equimolar amount (5.0 mmol) of NaOH (1.0 M solution). The solution was heated to 70 °C. ZnCl<sub>2</sub> (in excess, dissolved in 50 mL water) was added to the reaction flask, at which point the appearance of the solution abruptly changed and a flocculant precipitate agglomerated at the surface of the solution. The solution was heated at 70 °C for 30–60 minutes. The precipitate was filtered and washed with water, ethanol, methanol, and acetone. All resulting materials, except Piv, were dried overnight at 40 °C and recrystallized in ethanol. The resulting material from Piv was washed with water, ethanol, and methanol, then dried overnight at 40 °C. The starting chemicals were purchased from Sigma Aldrich (zinc chloride: purity grade  $\geq 98\%$ ; octanoic acid: purity grade 98%; nonanoic acid: purity grade  $\geq 97\%$ ; decanoic acid: purity grade  $\geq 98\%$ ; undecanoic acid: purity grade 98%; dodecanoic acid: purity grade  $\geq 99\%$ ; palmitic acid: purity grade 98%; oleic acid: purity grade 90%; stearic acid: purity grade 95%; 2,2-dimethylpropanoic acid: purity grade 99%), Fisher Scientific (sodium hydroxide: purity grade 99.99%), Acros Organics (azelaic acid, purity grade  $>98\%$ ), and Decon Labs (ethanol: purity grade  $>99.2\%$ ).

### Analysis of materials

The purity of each compound was verified by attenuated total reflection infrared spectroscopy (ATR-FTIR) and powder X-ray diffraction (PXRD) analysis. The FTIR measurements were carried out with a Thermo Nicolet Nexus 470 FTIR spectrometer with a spectral resolution of 4 cm<sup>-1</sup>. The PXRD measurements were performed using a Bruker D8 XRD (LynxEye position-sensitive detector) operating with a Cu K $\alpha$ 1 X-ray generator ( $\lambda = 1.54 \text{ \AA}$ ) with a 40 kV beam voltage and 40 mA current. The samples were not rotated during measurement and were obtained with Bragg Brentano geometry.

### Single-crystal X-ray diffraction

X-ray diffraction structures of zinc nonanoate, zinc decanoate, and zinc oleate were determined on single crystals grown in the laboratory through slow evaporation of solvent. Zinc decanoate was recrystallized from methanol. Zinc nonanoate and zinc oleate were recrystallized from ethanol. Crystals were mounted in inert oil. The crystal structures were analyzed in a manner similar to that of lead nonanoate<sup>33</sup> and lead octanoate.<sup>53</sup> Crystals were mounted using viscous oil onto a plastic mesh and cooled to the data collection temperatures, 150 K. Data were collected on a Bruker-AXS diffractometer with Cu-K $\alpha$  radiation ( $\lambda = 1.54178 \text{ \AA}$ ) focused with Goebel mirrors for zinc oleate, and on a Bruker-AXS D8 Venture Photon diffractometer with Cu-K $\alpha$  radiation ( $\lambda = 1.54178 \text{ \AA}$ ) focused with Goebel mirrors for zinc nonanoate and zinc decanoate. Unit cell parameters were obtained from 48 data frames, 2°  $\omega$ , from



different sections of the Ewald sphere or from  $180^\circ$   $\omega$  fast scans ( $1^\circ \text{ s}^{-1}$ ).<sup>54</sup> The systematic absences in the diffraction patterns were uniquely consistent with  $Pc$  or  $P2/c$  for zinc oleate and zinc nonanoate; whereas the systematic absences in the pattern gave possible structures for zinc decanoate in  $C2$ ,  $Cm$  or  $C2/m$ . In each case, an exhaustive exploration of the data uniquely yielded chemically reasonable and computationally stable results only with the space groups  $Pc$  for zinc oleate and zinc nonanoate, and  $C2$  for zinc decanoate. The data were treated with multi-scan absorption corrections.<sup>54</sup> The structures were solved using intrinsic phasing methods<sup>55</sup> and refined with full-matrix, least-squares procedures on  $F^2$ .<sup>3</sup> Zinc decanoate and zinc nonanoate were refined as 82/18, and 91/9 inversion twinned data sets, respectively. All non-H atoms were refined with anisotropic displacement parameters. H-atoms were constrained to idealized positions with isotropic parameters based on their attached atoms. Atomic scattering factors were contained in the SHELXTL program library.<sup>56</sup> The CIFs have been deposited at the Cambridge Structural Database under CCDC 2088849 for zinc oleate, 2088850 for zinc decanoate, and 2088851 for zinc nonanoate.†

### NMR spectroscopy

Zinc-67 NMR spectra were recorded with Bruker Avance NEO consoles and 3.2 mm probes designed and built at the National High Magnetic Field Laboratory (NHMFL) in Tallahassee, Florida, USA. Spectra at 35.2 T were acquired using the NHMFL series-connected hybrid (SCH) magnet at a frequency of  $\nu_0(^{67}\text{Zn}) = 93.87$  MHz. Spectra at 19.6 T were acquired on a 31 mm bore superconducting magnet at a frequency of  $\nu_0(^{67}\text{Zn}) = 52.02$  MHz. The spectra were recorded at ambient temperature, with either static powder samples or magic-angle spinning (MAS) at 15.625 kHz, and were externally referenced to a saturated solution of  $\text{Zn}(\text{NO}_3)_2$  in  $\text{D}_2\text{O}$  with  $\delta_{\text{iso}} = 0$  by using the frequency ratios of  $^{17}\text{O}$  in tap water and  $^{67}\text{Zn}$  reported in the IUPAC recommendations.<sup>57</sup> Spectra for ZnC8, ZnC9, and ZnC16 were obtained using a QCPMG sequence with a 4 and 8  $\mu\text{s}$   $\pi/2$ - and  $\pi$ -pulses and a six-rotor period inter-pulse delay  $\tau = 640$   $\mu\text{s}$  and a recycle delay of 0.5 s. The remaining spectra were obtained with a WURST-QCPMG sequence. For remaining spectra, the  $\pi/2$ - and  $\pi$ -pulses were increased to 5 and 10  $\mu\text{s}$ , respectively, with a WURST pulse applied before the QCPMG for signal enhancement. The WURST pulse was placed at +350 kHz frequency offset with sweep width equal to the MAS frequency of 15.635 kHz (or 16 kHz for ZnPIV) in  $\sim 2$  ms. The recycle delay was decreased to 0.2 s. The number of scans varied by sample and ranged from 256 for PIV to 327 680 for AZE.

Fitting of the spikelet patterns was carried out with Ssnake, an open-source NMR data processing software specifically aimed at solid-state NMR.<sup>58</sup> These compounds are dominated by quadrupole interactions. Visual fits were determined using SOLA in TopSpin 4.1.4 to establish a starting point for the quadrupole coupling constant ( $C_q$ ), asymmetry parameter ( $\eta_Q$ ), and the isotropic chemical shift ( $\delta_{\text{iso}}$ ) values. Refinement of the initial parameters were carried out using the quadrupole

fitting method in Ssnake. Fitting simulations were carried out until the values remained constant for 5 iterations. For figures, the line shapes of the Ssnake fitting parameters were simulated with solid line shape analysis (SOLA) in TopSpin v4.1. Comparison of the simulated spectra with the envelope of spikelets indicates only one zinc center per unit cell.

Cross polarization magic-angle-spinning (CPMAS) sequences similar to those used for lead soap analysis were used for carbon-13 analysis.<sup>32,34</sup> Carbon-13 NMR analyses were performed on a spectrometer with a Bruker AVIII console and a triple resonance 3.2 mm Z-GRD MAS probe.  $^{13}\text{C}$  spectra acquired at 14.1 T at a frequency of  $\nu_0(^1\text{H}) = 599.78$  MHz,  $\nu_0(^{13}\text{C}) = 150.81$  MHz at 295K with MAS at 14.0 kHz and were externally referenced through adamantane.<sup>59</sup> Additional spectra was required for zinc undecanoate and zinc oleate to resolve the doublet in the carboxylate region. These spectra were obtained on a Bruker AVIII console and a 4.0 mm HX MAS probe at 11.7 T at a frequency of  $\nu_0(^1\text{H}) = 500.13$  MHz,  $\nu_0(^{13}\text{C}) = 125.75$  MHz at 255K with MAS at 10.0 kHz, and externally referenced to adamantane. Acquisition parameters for these experiments included 1024 scans, 2 ms contact times, 5 s recycle delays,  $^{13}\text{C}$  pulse width 2.3  $\mu\text{s}$ , and proton decoupling at 70 kHz.

## III. Results and discussion

### X-ray diffraction structures

The structures determined by single-crystal X-ray diffraction data for a number of short-chain zinc soaps have been reported.<sup>60–67</sup> (Table 2) The local structure around the zinc ion in these structures is nearly tetrahedral, and the first coordination sphere consists of oxygens of four different carbonyl groups. Each carbonyl ligand forms a bidentate bridge with two zinc atoms in a *syn-anti* arrangement. Variations in the Zn–O bond length indicate an unsymmetrical bonding environment around the zinc centers. Through FTIR spectroscopy as well as single-crystal and powder X-ray diffraction, Taylor *et al.*<sup>80</sup> predicted at least two packing arrangements of hydrocarbon chains in the lattice, with the groups being characteristic of chains with eight or fewer carbons and chains with nine or more carbons. Further discussion of FTIR analysis is presented in the next section. Our single-crystal data presented in Table 3 indicate that nonanoate crystallizes in the monoclinic group  $Pc$ , the same space group reported for the octanoate,<sup>61</sup> indicating that these two compounds have similar packing arrangements.

Interestingly, we found that the oleate also crystallizes in the monoclinic  $Pc$  group. The structures are non-centrosymmetric, characterized by two crystallographically independent chains parallel to the ac plane. No evidence of interdigitation of sheets, as suggested by Nelson *et al.*,<sup>68</sup> was found for the nonanoate or the oleate. We therefore grouped zinc octanoate (ZnC8), zinc nonanoate (ZnC9), and zinc oleate (ZnZ-C18) together as group I.



Table 2 X-ray diffraction information on zinc carboxylates

Material	Unit formula	Space group	$\beta$	Zn–O (Å)	Cell volume (Å <sup>3</sup> )	Cell parameters (Å)	Ref.
Zinc acetate	Zn(C <sub>2</sub> H <sub>3</sub> O <sub>2</sub> ) <sub>2</sub>	<i>C2/c</i>	99.49	1.952, 1.960, 1.949, 1.965	1325.3(3)	<i>a</i> = 30.237(2) <i>b</i> = 4.799(1) <i>c</i> = 9.260(1)	60
Zinc propionate	Zn(C <sub>3</sub> H <sub>5</sub> O <sub>2</sub> ) <sub>2</sub>	<i>Pna2<sub>1</sub></i>	90	1.961, 1.977, 1.939, 1.948	849.7(1)	<i>a</i> = 9.2862(9) <i>b</i> = 4.7937(4) <i>c</i> = 19.0871(19)	62
Zinc butanoate	Zn(C <sub>4</sub> H <sub>7</sub> O <sub>2</sub> ) <sub>2</sub>	<i>P2<sub>1</sub>/c</i>	90.08	—	1056(2)	<i>a</i> = 23.48(3) <i>b</i> = 4.795(4) <i>c</i> = 9.380(9)	63
Zinc pentanoate	Zn(C <sub>5</sub> H <sub>9</sub> O <sub>2</sub> ) <sub>2</sub>	<i>P2<sub>1</sub>/a</i>	104.256	1.950, 1.947, 1.966, 1.963	1267.5(5)	<i>a</i> = 9.389(2) <i>b</i> = 4.7820(10) <i>c</i> = 29.126(7)	64
Zinc hexanoate	C <sub>24</sub> H <sub>44</sub> O <sub>8</sub> Zn <sub>2</sub>	<i>P1c1</i>	93.73	1.972, 1.947, 1.917, 1.945	1439.5(3)	<i>a</i> = 32.309(4) <i>b</i> = 4.7865(6) <i>c</i> = 9.3292(13)	65
Zinc heptanoate	Zn(C <sub>7</sub> H <sub>13</sub> O <sub>2</sub> ) <sub>2</sub>	<i>Pbc2<sub>1</sub></i>	90	1.975, 1.973, 1.980, 1.948 1.920, 1.946, 1.975, 1.976	1649.7(4)	<i>a</i> = 4.7651(6) <i>b</i> = 9.3404(15) <i>c</i> = 37.066(6)	67
Zinc octanoate	Zn(C <sub>8</sub> H <sub>15</sub> O <sub>2</sub> ) <sub>2</sub>	<i>Pc</i>	101.343	1.965, 1.966, 1.971, 1.988	897.78(1)	<i>a</i> = 21.093(2) <i>b</i> = 4.6905 <i>c</i> = 9.2544(9)	61

Table 3 Single crystal X-ray diffraction data and refinement parameters

	Zinc(II) nonanoate	Zinc(II) decanoate	Zinc(II) oleate
Chemical formula	Zn(C <sub>9</sub> H <sub>17</sub> O <sub>2</sub> ) <sub>2</sub>	Zn(C <sub>10</sub> H <sub>19</sub> O <sub>2</sub> ) <sub>2</sub>	Zn(C <sub>18</sub> H <sub>33</sub> O <sub>2</sub> ) <sub>2</sub>
Formula weight, g mol <sup>-1</sup>	379.82	407.87	1010.61
Temperature, K	100(2)	100(2)	150(2)
Crystal system	Monoclinic	Monoclinic	Monoclinic
Space group	<i>Pc</i>	<i>C2</i>	<i>Pc</i>
Cell dimensions			
<i>a</i> , Å	23.001(3)	7.7771(6)	41.827(3)
<i>b</i> , Å	4.6615(7)	5.5309(4)	4.6881(3)
<i>c</i> , Å	9.2225(14)	24.5099(17)	9.2475(5)
$\alpha$ , °	90	90	90
$\beta$ , °	95.957(3)	92.418(3)	90.714(4)
$\gamma$ , °	90	90	90
Volume, Å <sup>3</sup>	983.5(3)	1053.34(13)	1813.2(2)
<i>Z</i>	2	2	2
$\rho_{\text{calc}}$ , g cm <sup>-3</sup>	1.283	1.28	1.151
$\mu$ /mm <sup>-1</sup>	1.860	1.1771	1.190
<i>F</i> (000)	408.0	440.0	688.0
Reflections collected	7412	6074	21 258
Independent collections	2695	1839	6535
Data/restraints/parameters	2695/2/211	1839/1/116	6535/2/372
Goodness-of-fit	1.132	1.196	1.044
<i>R</i> [ <i>I</i> ≥ 2 $\sigma$ ( <i>I</i> )] <i>R</i> <sub>1</sub> / <i>wR</i> <sub>2</sub>	0.0480/0.1357	0.0337/0.0918	0.0761/0.1977
<i>R</i> indexes [all data] <i>R</i> <sub>1</sub> / <i>wR</i> <sub>2</sub>	0.0483/0.1359	0.0344/0.0934	0.0811/0.2016
Absolute structure parameter	0.09(8)	0.18(6)	0.01(5)
CCDC	2088851	2088850	2088849

Single-crystal X-ray data for the zinc decanoate (ZnC10) indicate a non-centrosymmetric structure with one crystallographically independent chain characterized by a lamellar stacking of sheets perpendicular to the *c* axis. As with the nonanoate and the oleate, there is no indication of interdigitation of sheets. ZnC10 was thus placed in group II. Rietveld-refined synchrotron powder X-ray diffraction data of the dodecanoate and tetradecanoate predict a monoclinic *C2* space group for both solids.<sup>69</sup> These structures match the crystallographically determined structure of the decanoate from this study. The

undecanoate was also refined using the Rietveld-refined method, from which it was suggested that it crystallizes in a structure, with two crystallographically independent chains in a manner similar to the short-chain carboxylates (group I).<sup>69</sup>

Spacing between CH<sub>3</sub>–CH<sub>3</sub> tail groups in opposite layers in the lamellae averaged 4.115 Å for the nonanoate and 4.119 Å for the oleate. In comparison, the CH<sub>3</sub>–CH<sub>3</sub> spacing was found to be substantially smaller at 3.741 Å for the decanoate. The smaller void spaces for the even-numbered chains allow more



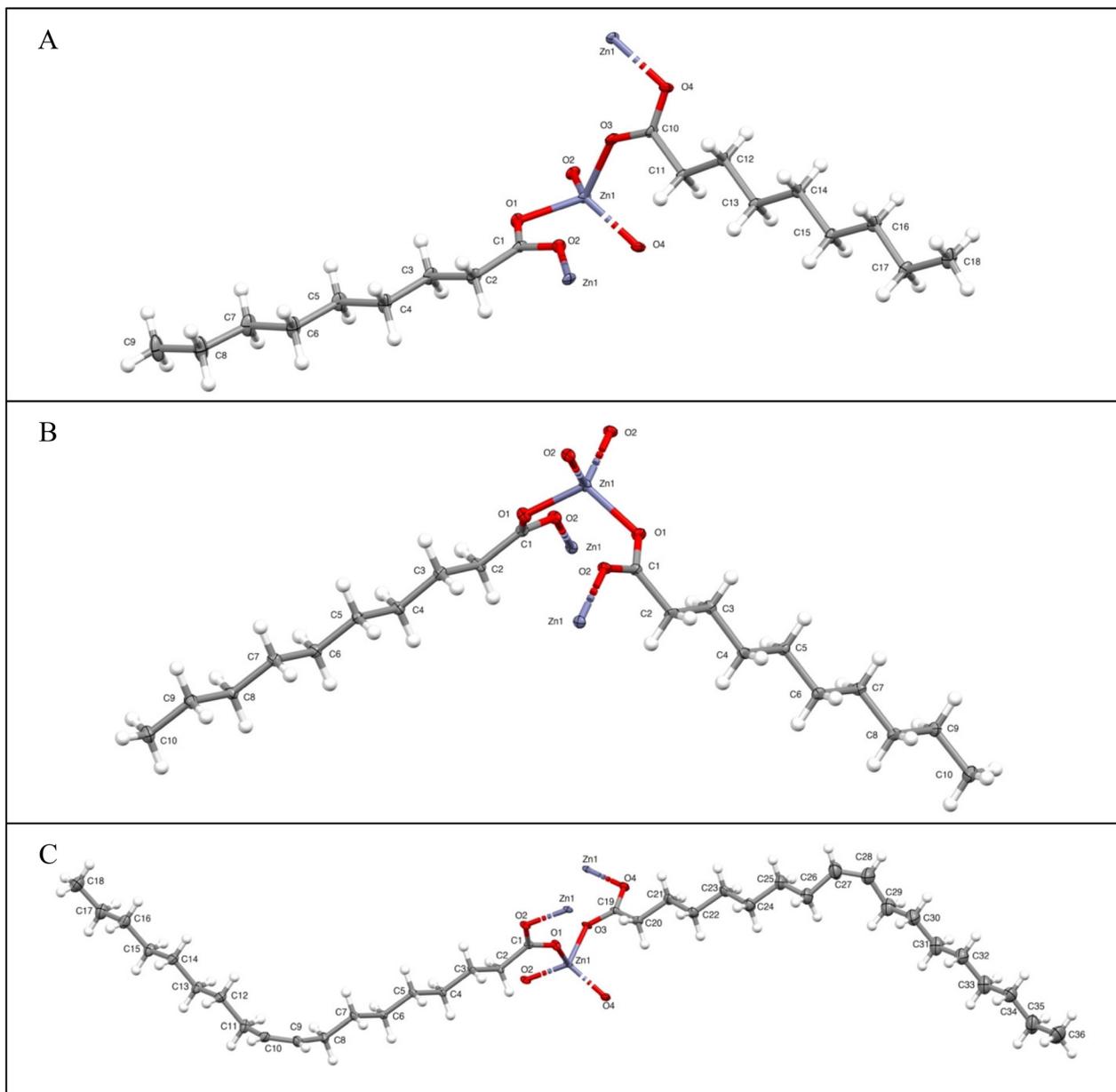


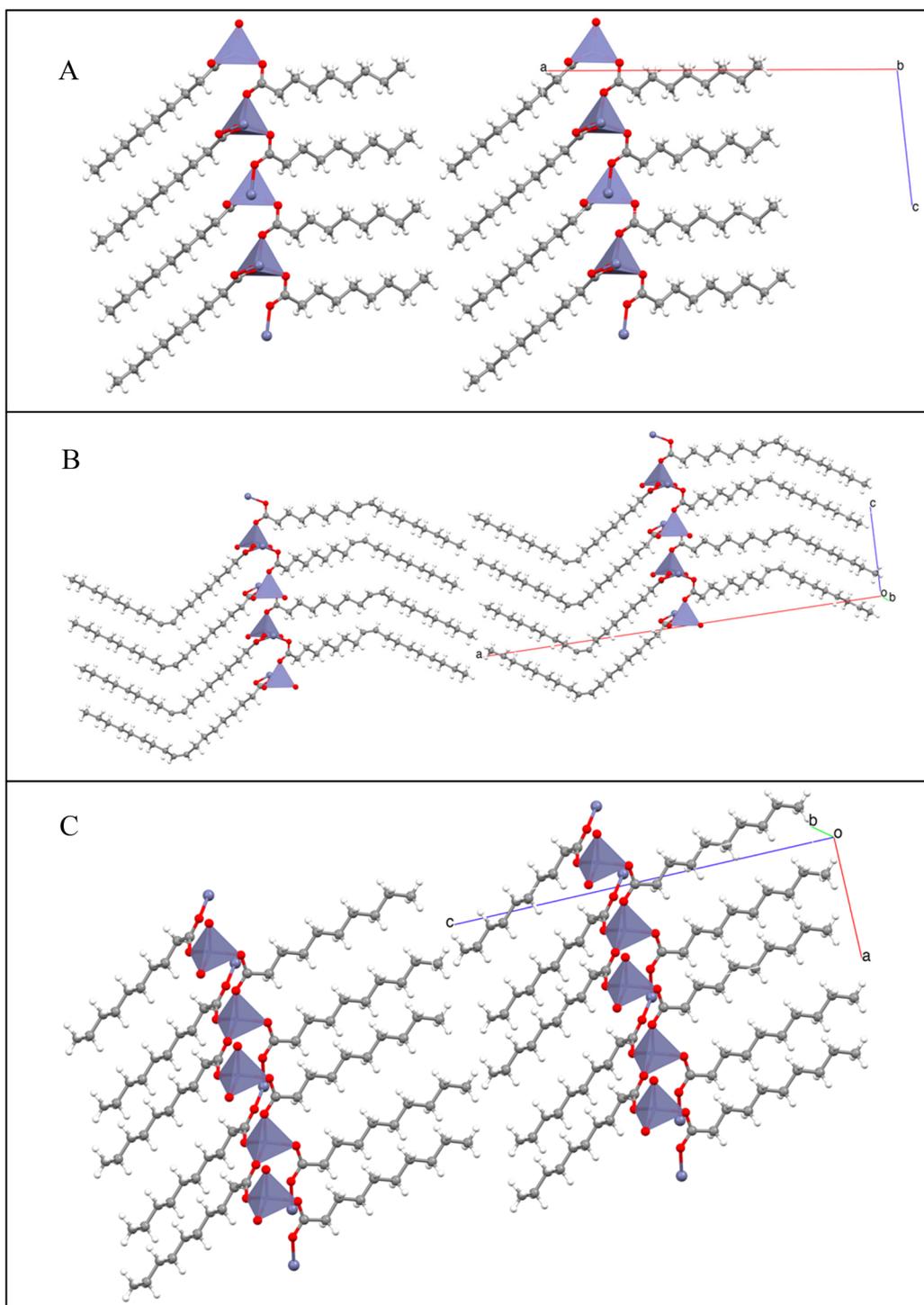
Fig. 1 X-ray-derived molecular structures of (A) zinc nonanoate, (B) zinc decanoate, and (C) zinc oleate, with 30% probability ellipsoids. H-atoms are depicted with standard bond lengths.

efficient packing and reduced distances between lamellar sheets seen in the crystal packing for the decanoate. Molecular structures are presented in Fig. 1. Group packing diagrams are presented in Fig. 2.

Mesbah *et al.* state that the separation between structural archetypes occurs between ZnC9 (group I) and ZnC10 (group II), with the suggestion that odd-numbered chains with  $C > 10$  fall into the collection of group I structures.<sup>69</sup> Group I structures possess two orientations of the zinc tetrahedra within the unit cell. Group II structures possess only one orientation of the zinc tetrahedron. Both groups crystallize in an AB arrangement between layers. Expanded packing diagrams for

each group are provided in the ESI Fig. S1–S3† for visualization of the AB packing arrangement. Visualizations of group I and group II arrangements are given in Fig. 2. The oleate, although distinctly in the longer-chain category by number of carbons, falls within the group I crystal structure due to the fact that the chains on either side of the trans-double bond are nine carbons in length. This observation is particularly relevant for the chain bonded to the zinc center. The combination of the short chain around the immediate zinc center and the steric bulk of the bent oleyl chain results in a group I twisted tetrahedral arrangement, rather than the more ordered group II typical of the longer chains. The packing arrangement of the





**Fig. 2** Structures of group I represented by ZnC9 (A) and ZnZ-C18 (B), and group II represented via ZnC10 (C) highlighting the arrangement of tetrahedron about the zinc in each crystal type. Hydrogens have been removed for visual clarity.

zinc oleate is also provided in Fig. 2B. The single-crystal X-ray-diffraction-derived structural parameters of zinc(II) nonanoate, zinc(II) decanoate, and zinc(II) oleate are given in Table 3. The presence of two structural motifs has also been confirmed by FTIR, discussed further in the next section.<sup>70,71</sup>

X-ray powder patterns for the compounds are characterized by regularly spaced, higher-order  $00l$  reflections for low angles ( $2\theta < 15^\circ$ ). Reflections at mid and higher angles are irregularly spaced and of low intensity and match literature reports for metal soaps.<sup>3,19</sup> These variations are most notable in the com-



parisons of the single-crystal-generated and powder patterns for the ZnC9, ZnC10, and ZnZ-C18 compounds. It should be noted that the bulk powders were measured at room temperature while the single-crystal data was acquired at 150 K. The low-temperature acquisition reduces thermal motion, dynamic disorder, and librational effects.<sup>72</sup> The resulting patterns are of better resolution than those obtained at higher temperatures. This is most notable in the diffraction patterns of ZnZ-C18. The simulated pattern from the single-crystal data has clear resolution for the  $20^\circ \leq 2\theta \leq 25^\circ$ . The peaks in that region are greatly suppressed in the bulk, room temperature data likely due to a combination of thermal motion and multiple polytypes present in the powder sample. The single-crystal lattice parameters were used to generate powder patterns in excellent agreement with the experimental data in the main peaks below  $15^\circ$ . Experimental PXRD patterns for all compounds and their comparisons to the simulated patterns of the single crystals, where applicable, are found in the ESI (Fig. S4–S12†).

### FTIR spectroscopy

Hermans *et al.* observed both crystalline and amorphous domains of Zn carboxylates, zinc ions loosely coordinated to the ionomeric network of the cross-linked network, in a painting by van Gogh with infrared spectroscopy.<sup>73,74</sup> Similar results have been reported for paintings by Jackson Pollock<sup>75</sup> and Salvador Dalí.<sup>25</sup> The structural chemistry of zinc carboxylates has clear implications for art conservation.<sup>71,76</sup>

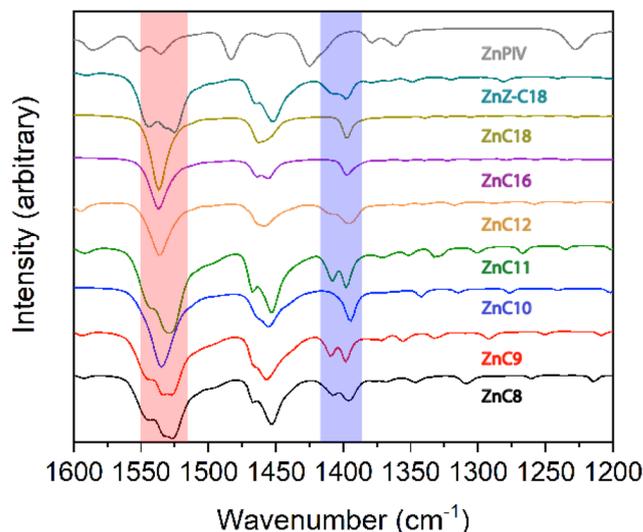
Several previous studies have used FTIR spectroscopy in conjunction with powder X-ray diffraction analysis to assess the coordination environment around the metal center in zinc carboxylates of varying lengths.<sup>63,65,68,70,71,77,78</sup> These studies have identified the characteristics of the bridging bidentate coordination of the carbonyl groups in these compounds. Splitting in the symmetric,  $\nu_s$ , and antisymmetric,  $\nu_{as}$ , vibrational bands of the carbonyl group is indicative of asymmetry in the zinc–oxygen bonds. The differences between the vibrational modes, tabulated in Table 4, agree with literature values for bidentate bridging ligands.<sup>79</sup>

Several studies have looked at the  $\nu_{as}\text{COOH}$  ( $\sim 1540\text{--}1520\text{ cm}^{-1}$ ) as an indicator of zinc coordination.<sup>70,71,80</sup>

Hermans *et al.* reported that short chain structures display split  $\nu_{as}\text{COOH}$  band indicating the presence of two distinct carboxylate groups, while the longer chain ( $C > 9$ ) structures display a single  $\nu_{as}\text{COOH}$  band. They theorize that the difference in packing is due to energetically zinc-coordination favored (short chain) *versus* van der Waals-favored (long chain) packing.<sup>70</sup> Nelson, Taylor, and Ellis propose a related theory also suggesting that the long chains pack in the group II arrangement due to increased van der Waals forces between the alkyl chains; however, the packing for the short chains is attributed to the electrostatic repulsion between the methyl tail groups of the chains.<sup>81</sup>

FTIR spectra of the zinc carboxylates in our study (Fig. 3 and S10;† Tables 4 and S1†) show a similar trend, with two marked differences from the literature. Both the undecanoate (ZnC11) and the oleate (ZnZ-C18) show splitting characteristic of an asymmetric zinc center. Consistent values for the rocking vibrations,  $\rho$ , of the carbonyl group (Table S1†) imply similar molecular structures for the entire series, which agrees with previously established literature values.<sup>80</sup> From analysis of the  $\nu_{as}\text{COOH}$  band, we confirm the placement of ZnC8, ZnC9, and ZnZ-C18 in group I and add ZnC11, ZnPIV, and ZnAZE to the group considering the splitting of the band. ZnC10 remains in group II while ZnC12, ZnC16, and ZnC18 are added due to the single band presented in their FTIR spectra.

Nelson *et al.*<sup>82</sup> found a variation in intensities of the  $\nu_{as}\text{CH}_3$  ( $\sim 2960\text{ cm}^{-1}$ ) and  $\nu_s\text{CH}_3$  ( $\sim 2970\text{ cm}^{-1}$ ) band between the short- and long-chain methyl tail groups. These variations were attributed to increased methyl–methylene interactions, and increasing bilayer overlap, with increasing chain length. We find no indication of this phenomenon in these FTIR spectra. All  $\nu_{as}(\text{CH}_3)$  intensities are more intense than the corresponding  $\nu_s(\text{CH}_3)$  intensities. Additionally, single-crystal



**Fig. 3** FTIR spectra of the zinc carboxylates, highlighting the  $\nu_s\text{ COO}$  ( $\sim 1540\text{--}1520\text{ cm}^{-1}$ ) highlighted in red and  $\nu_{as}\text{ COO}$  ( $1410\text{--}1395\text{ cm}^{-1}$ ) highlighted in blue.  $\Delta\nu$  for all samples is characteristic of bidentate ligands. The splitting of the resonances is indicative of asymmetric zinc centers.

**Table 4** Frequencies and intensities of carboxylate group FTIR bands

Compound	Frequency of bands $\text{cm}^{-1}$	
	$\nu_{as}\text{ COO}$	$\nu_s\text{ COO}$
ZnC8	1544, 1527 (s)	1408, 1396 (m, b)
ZnC9	1544, 1525 (vs)	1410, 1398 (m)
ZnC10	1535 (vs)	1396 (m)
ZnC11	1542, 1529 (vs)	1408, 1398 (m)
ZnC12	1535 (vs)	1408, 1394 (m, b)
ZnC16	1537 (s)	1396 (m, b)
ZnC18	1537 (vs)	1396 (m)
ZnZ-C18	1542, 1525 (s)	1408, 1398 (m, b)
ZnPiv	1550, 1521 (m)	1415 (m, b)
ZnAZE	1544, 1592 (vs)	1405, 1392 (s)

(vs) very strong; (s) strong; (m) medium; (w) weak; (b) broad.



X-ray diffraction data, discussed above, indicate a lack of overlap in the chain ends during packing. Medium to strong broad bands centered around  $1460\text{ cm}^{-1}$  are present for all compounds, except the pivalate, indicative of the mixing of  $\text{CH}_3$  bending and  $\text{CH}_2$  scissoring modes of different polymorphic forms, as discussed for the  $^{13}\text{C}$  NMR data. The absence of bands in the  $3500\text{--}3300\text{ cm}^{-1}$  region indicates that these materials are anhydrous. The absence of a carbonyl peak around  $1730\text{ cm}^{-1}$  and a shift of carbonyl peaks to lower frequencies implies the absence of free acid in these samples.

### NMR spectroscopy

$^{13}\text{C}$  NMR also provides information regarding the crystal packing of the zinc carboxylates (Fig. 4). ZnPIV exhibits a  $^{13}\text{C}$  spectrum distinct from the rest of the carboxylates, in agreement with the distinctive  $^{67}\text{Zn}$  NMR parameters. All  $^{13}\text{C}$  resonances for ZnPIV are less shielded than those of the pure acid and of the other zinc carboxylates. ZnAZE also exhibits a slightly different spectrum than the other carboxylates. The structure of azelaic acid consists of a carbon chain capped on each end with a carboxylic acid group producing two clear carboxylate resonances, one for each the bridging and terminal carboxylate groups. Fig. 5 shows a zoomed in view of the carboxylate region, spanning  $195\text{--}180\text{ ppm}$ . The bridging carboxylate resonance for ZnAZE at  $187.5\text{ ppm}$  shows a

shoulder not present in the resonance for the terminal carboxylate at  $186\text{ ppm}$ . This can be attributed to an unresolved doublet due to two carboxylate conformations around the zinc center. The carboxyl resonances for the short-chain ( $C \leq 9$ ) homologues around  $187\text{ ppm}$  are doubled, with a relative ratio of 1:1, indicating two unique carboxyl configurations in the unit cell. Doubling is also seen for the ZnZ-C18; however full resolution of the doublet was not possible. The carboxyl resonance is a singlet for the long-chain ( $C > 9$ ) molecules, indicating a single carboxylate configuration for these compounds.

These results are in agreement with the crystal structures of zinc nonanoate, zinc decanoate, and zinc oleate discussed above. Variations in the intensities and broadness of the resonances for the methylene carbons, between  $\sim 45\text{--}25\text{ ppm}$ , as well as shifts and doubling of the methyl carbon resonance at  $\sim 17\text{ ppm}$  indicate the possibility of multiple polymorphic forms in the powdered samples. This fact is corroborated by FTIR and PXRD data discussed above. While the  $^{13}\text{C}$  NMR data confirm the group placement for the majority of the carboxylates, the substantial variation in the ZnPIV spectrum necessitates separating it from the established grouping. A new group, group III, was thus established for ZnPIV.

Early model studies of Zn-soap formation in commercial linseed oil paints identified two distinct Zn oleate polymorphs,

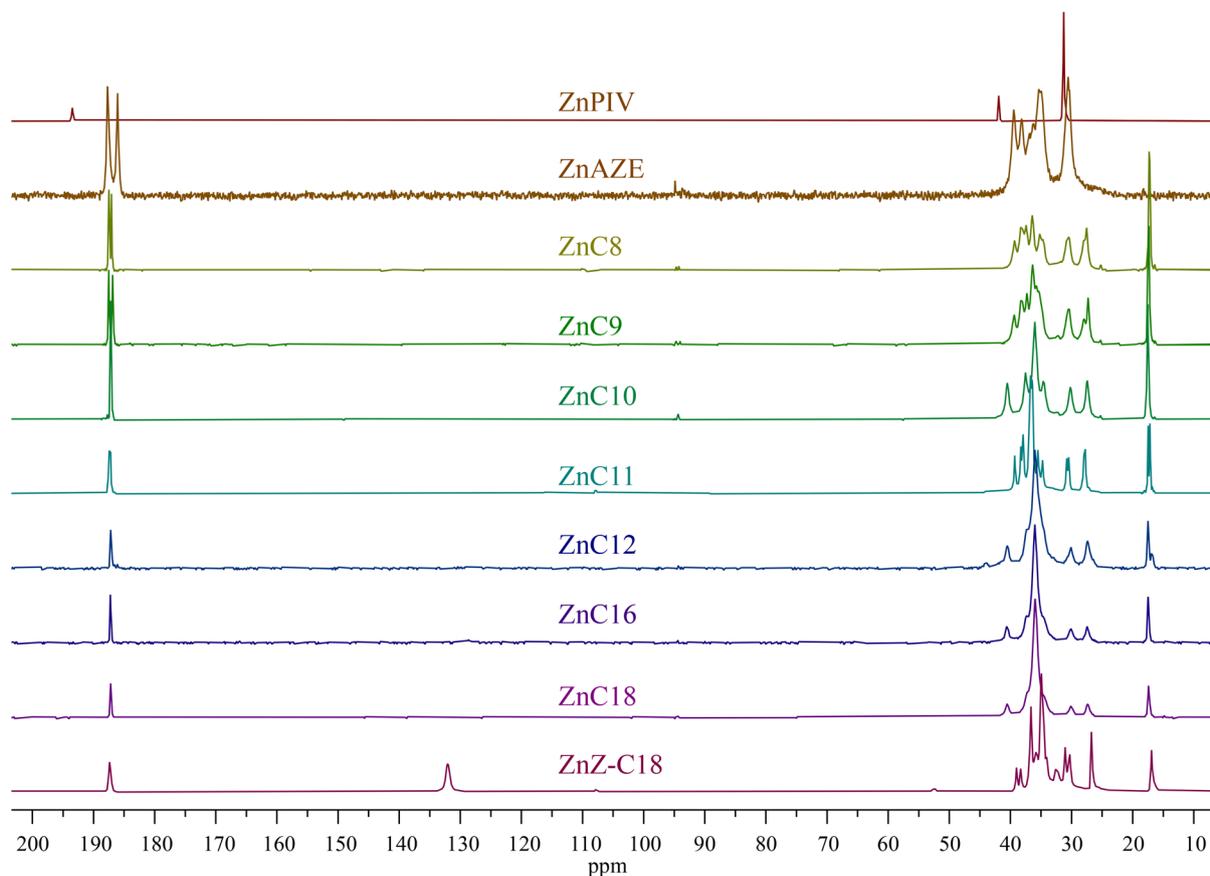


Fig. 4  $^{13}\text{C}$  NMR spectra for the zinc carboxylate series.



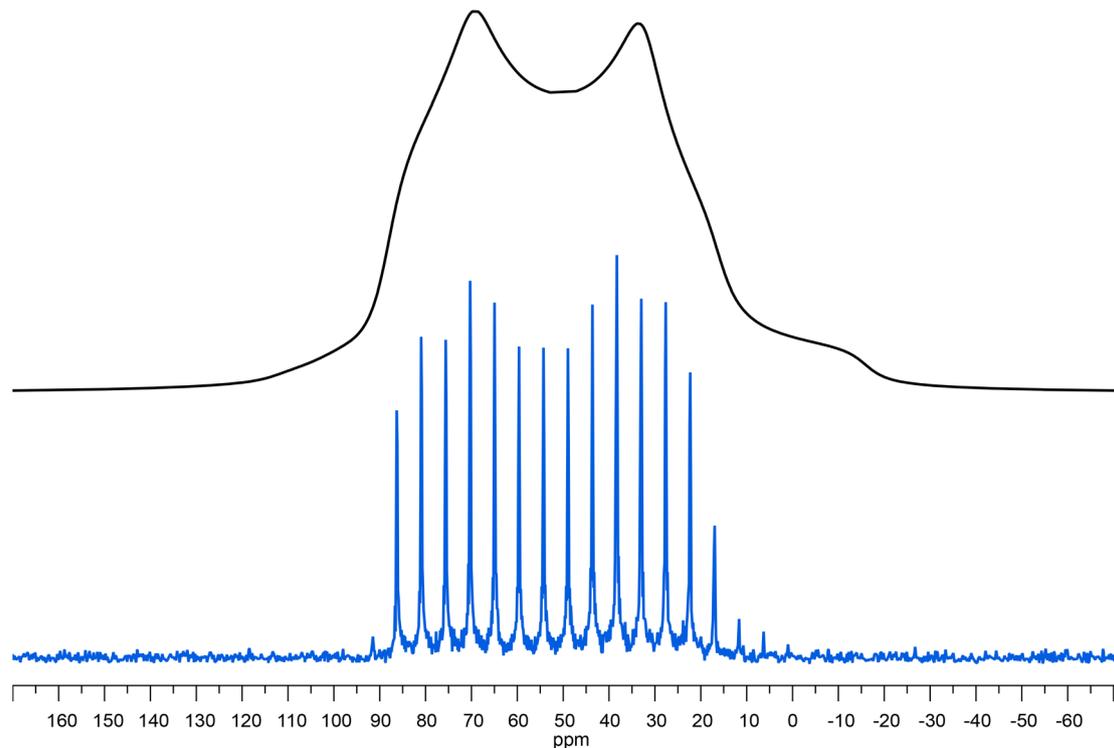


Fig. 5  $^{13}\text{C}$  NMR data for the carboxylate series, focusing on the carboxylate region.

depending on reaction conditions.<sup>8,70</sup> Unlike the corresponding free acids, zinc stearate and zinc oleate are miscible across the entire compositional range.<sup>83</sup> To accommodate the kink in the oleyl chain, neighboring palmitoyl chains were determined to be conformationally disordered, while more-distant palmitoyl chains remained in the all-trans conformation. Maines *et al.* have reported the intralayer cleavage (cohesive failure, rather than adhesive failure) of paint layers in modern paintings due to the formation of a Zn soap deposit, determined to be Zn oleate.<sup>84</sup> Lead oleate soaps, on the other hand, have not been observed, because oleic acid is rapidly converted to azelaic acid through oxidative cleavage. The structure of Zn oleate must somehow protect the double bond from oxidation.

Zinc-67 NMR parameters (quadrupolar coupling constant, anisotropic chemical shift) are diagnostic of non-spherical local electronic structure at the nuclear site, as is true for the

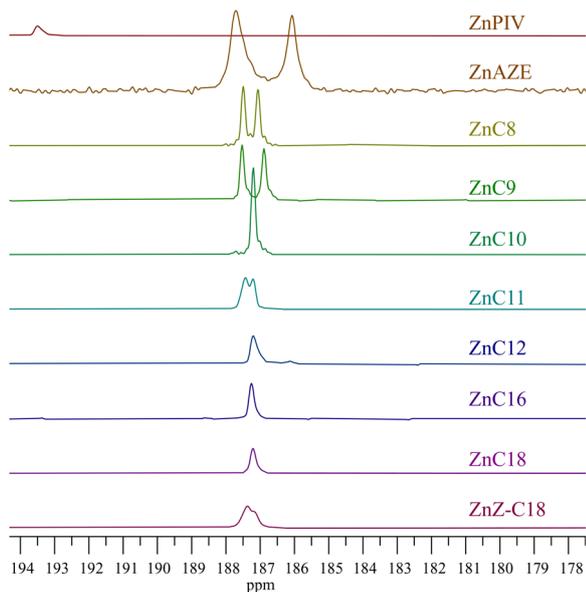
lead nuclei in lead-based soaps.<sup>32,34</sup>  $^{67}\text{Zn}$  NMR spectroscopy has been used to characterize the coordination geometry around  $\text{Zn}^{+2}$  ions in pure compounds.<sup>49,85–89</sup> In particular, the natural-abundance  $^{67}\text{Zn}$  NMR spectra of ZnO at 11.7 and 14.1 T have been reported.<sup>86,87</sup> The line shape of the central-transition is determined by second-order quadrupole effects, and for ZnO  $C_Q = 2.4$  MHz,  $\eta_Q = 0$ , and  $\delta_{\text{iso}} = 240$  ppm relative to a 0.1 M  $\text{ZnCl}_2$  aqueous solution. Comparison of various zinc-containing materials demonstrates that the NMR parameters are particularly sensitive to the local electronic environment.<sup>87</sup>

The electric-field gradient (EFG), as defined by the parameters  $C_Q$  and  $\eta_Q$ , is sensitive to the position and orientation changes in the local environment of the zinc center, particularly the placement of surrounding oxygen atoms.  $^{67}\text{Zn}$  NMR results for the series of carboxylates reported here indicate that the zinc environments fall into three groups. (Table 5)

Table 5  $^{67}\text{Zn}$  solid-state MAS NMR fitting parameters of solid zinc carboxylates, organized by group

Material	$B_0$ (T)	$\delta_{\text{iso}}$ (ppm)	$C_Q$ (MHz)	$\eta_Q$	LB (Hz)
ZnAZE	19.6	$101.3 \pm 0.3$	$6.92 \pm 0.02$	$0.375 \pm 0.003$	$1400 \pm 8$
ZnC8	35.2	$96.0 \pm 0.5$	$8.04 \pm 0.04$	$0.2979 \pm 0.004$	$740 \pm 27$
ZnC9	35.2	$92.1 \pm 0.6$	$8.06 \pm 0.05$	$0.342 \pm 0.006$	$530 \pm 16$
ZnC11	19.6	$86.0 \pm 0.9$	$8.2 \pm 0.1$	$0.26 \pm 0.01$	$1500 \pm 100$
ZnZ-C18	19.6	$51.3 \pm 0.2$	$7.109 \pm 0.005$	$0.217 \pm 0.006$	$484 \pm 4$
ZnC10	19.6	$103.9 \pm 0.3$	$2.44 \pm 0.06$	$0.51 \pm 0.02$	$229 \pm 30$
ZnC12	19.6	$104.1 \pm 0.1$	$2.454 \pm 0.003$	$0.4469 \pm 0.0001$	$232 \pm 2$
ZnC16	35.2	$102.1 \pm 0.5$	$2.57 \pm 0.06$	$0.67 \pm 0.02$	$200 \pm 35$
ZnC18	19.6	$80.3 \pm 0.4$	$2.63 \pm 0.04$	$0.63 \pm 0.01$	$160 \pm 28$
ZnPIV	35.2	$95 \pm 3$	$11.0 \pm 0.1$	$0.799 \pm 0.008$	$500 \pm 80$



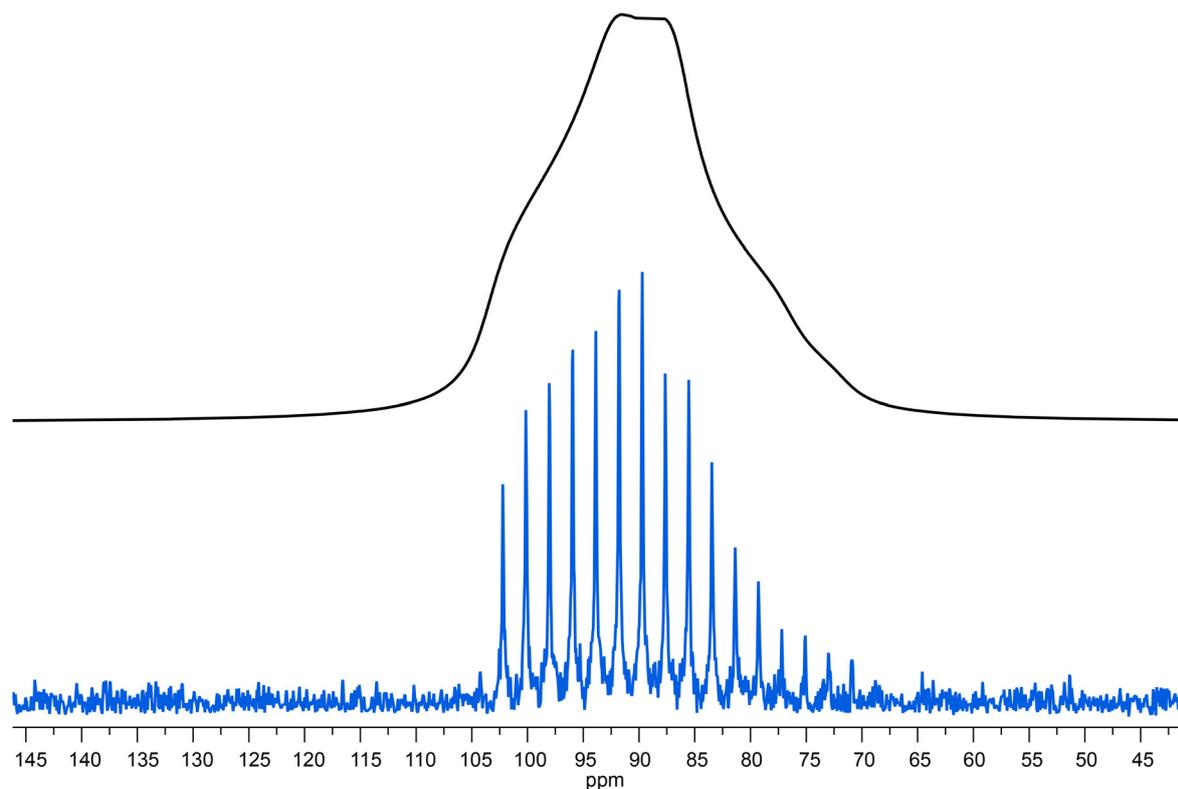


**Fig. 6**  $^{67}\text{Zn}$  spectra for ZnC8. (Top, Black) Snake fitting parameters visualized in TopSpin; (Bottom, Blue) WURST-QCPMG spectrum at 15 625 Hz MAS.  $B_0$  35.2 T.

Fig. 6, 7, and 8 shows spikelet spectra for molecules representative of the three distinct groups, together with their respective simulated powder patterns. Group I, which includes

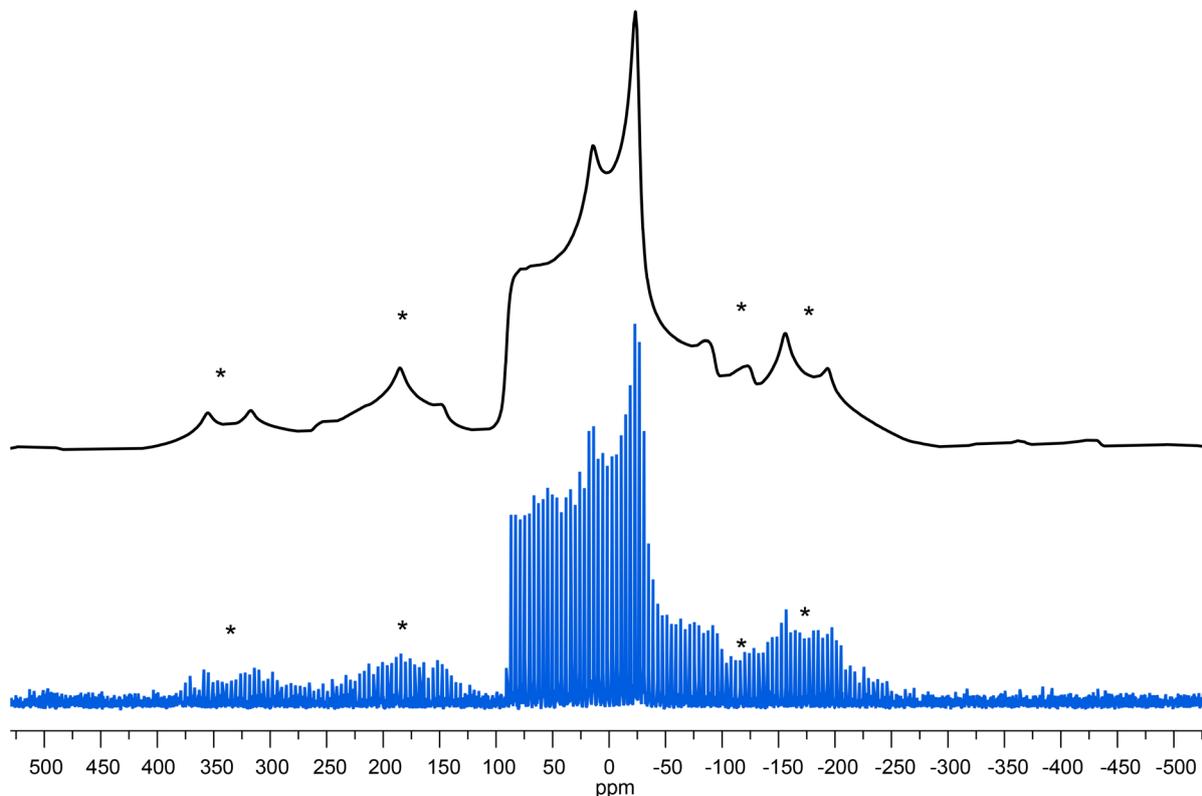
shorter-chain homologues as well as zinc undecanoate and zinc oleate, evinces a quadrupole coupling constant  $\sim 8$  MHz with a moderate asymmetry parameter ( $\sim 0.30$ ). Group II is characterized by smaller quadrupole coupling constants ( $\sim 2.5$  MHz) with a larger asymmetry parameter ( $\sim 0.60$ ) and includes the longer chain carboxylates ( $C > 9$ ). The large quadrupole coupling constant for groups I and II are indicative of strongly asymmetric local environments around the zinc center. Group III is characterized by the largest quadrupole coupling constant ( $\sim 11.0$  MHz) and includes only zinc pivalate.

Similarly, the NMR spectra of group I compounds (Fig. 6, S14–S16<sup>†</sup>) suggest an asymmetric placement of oxygens around the zinc center, but the smaller magnitude of  $C_Q$  suggests a lesser effect than for the pivalate. The large  $C_Q$  indicates a distorted electronic environment around the metal centers. This supports the theory that these structures are likely influenced by methylene–methylene interactions between layers of the lamellae in the structures as suggested by Nelson, Taylor, and Ellis<sup>81</sup> rather than a preferential coordination geometry for zinc as proposed by Hermans *et al.*<sup>70</sup> The relatively short chain lengths of molecules of group II result in structures dominated by electrostatic repulsion between the methyl tail groups of the chains, as can be seen in the single-crystal diffraction information discussed above. The methyl tail groups necessitate a distortion of the tetrahedral arrangement around zinc to accommodate these repulsive forces.<sup>81</sup> In



**Fig. 7**  $^{67}\text{Zn}$  spectra for ZnC10. (Top, Black) Snake fitting parameters visualized in TopSpin; (Bottom, Blue) WURST-QCPMG spectrum at 15 625 Hz MAS.  $B_0$  19.6 T.





**Fig. 8**  $^{67}\text{Zn}$  spectra for ZnPIV. (Top, Black) Ssnake fitting parameters visualized in TopSpin; (Bottom, Blue) WURST-QCPMG spectrum at 16 000 Hz MAS. Sideband regions denoted with asterisks.  $B_0$  35.2 T.

zinc oleate, the *cis*-double bond between C9 and C10 prevents the all *trans* arrangement of the aliphatic chains found in the other long-chain carboxylates, which affects the structural alignment.

Group II consists of even-numbered long-chain ( $C > 9$ ) carboxylates (Fig. 7, S17–S19†). The structures of these carboxylates are dominated by geometric packing. The straight, all *trans* arrangement of the chains allows relatively easy packing with only minimal distortion of the tetrahedral structure around the zinc. This arrangement increases the van der Waals interactions between the chains which outweighs the electrostatic repulsion from the methyl tail groups. The longer chains allow greater distribution of electrostatic charge, minimizing the effect on the structure. Literature calculations of the molecular spacing for the long chain carboxylates indicate that the void spacing between chains is greater for the odd-numbered chains.<sup>68</sup> This increase in spacing would result in disorder in the packing and thus distortions in the tetrahedral structure. This effect can be seen in the spectrum of zinc undecanoate, whose  $^{67}\text{Zn}$  spectrum falls within group I.

Zinc pivalate (Fig. 8) consists of a small, but sterically bulky *t*-butyl ligand connected to the carboxylate group which, in turn, bridges two zinc centers. The steric hindrance caused by the four bridging groups bonded to each zinc results in a distorted tetrahedron around the zinc centers, which gives a rather large  $C_Q$ . This is consistent with the crystallographic data obtained by Clegg *et al.*<sup>52</sup>

## IV. Conclusions

Insight into the structure of medium and long chain zinc carboxylates can be obtained *via* spectroscopic and X-ray diffraction analysis. The series can be subdivided into three groups based on their spectroscopic and diffraction parameters.

From the spectroscopic and X-ray diffraction analysis, we conclude that group I consists of short chain ( $C < 9$ ) carboxylates, odd-numbered long chain ( $C > 10$ ) carboxylates, and zinc oleate; group II consists of even-numbered long chain ( $C > 10$ ) carboxylates; and group III consists of a relatively small molecule (ZnPIV) containing sterically bulky ligands. Based on the similarity of the NMR parameters, FTIR resonances, and powder X-ray diffraction data, we propose that the crystal structures of zinc nonanoate and zinc oleate can serve as models for group I compounds and the crystal structure of zinc decanoate can serve as a model for group II compounds.

For the group I compounds, the  $^{67}\text{Zn}$  NMR spectra have large  $C_Q$  ( $\sim 8$  MHz) indicating a distorted arrangement around the zinc center. The  $^{13}\text{C}$  NMR resonances for the carboxylate carbon present as a doublet centered at  $\sim 187$  ppm and show that there are two unique carboxylates around the zinc center. FTIR spectra show doubling in the symmetric and asymmetric carboxylate stretches at  $1550$ – $1525$   $\text{cm}^{-1}$  and  $1408$ – $1394$   $\text{cm}^{-1}$ , respectively, also indicative of multiple carboxylate environments. The X-ray crystal structures of both zinc nonanoate and zinc oleate confirm these findings. Both crystals possess struc-



tures with slightly distorted tetrahedra around the zinc centers as shown by the varying Zn–O bond lengths and two crystallographically independent chains.

The  $^{67}\text{Zn}$  NMR spectra for the group II compounds display much smaller values of  $C_Q$  ( $\sim 2.5$  MHz) than group I indicating a more symmetric arrangement around the zinc center. The  $^{13}\text{C}$  NMR resonance for the carboxylate carbon is a singlet indicating a single carboxylate environment. FTIR spectra show no doubling in the symmetric or asymmetric carboxylate stretches. The single crystal structure of zinc decanoate displays a symmetric tetrahedron around the zinc center with essentially equivalent Zn–O bond lengths and two crystallographically equivalent chains. Regular progression is seen in the region assigned to  $\text{CH}_2$  twisting modes ( $1350\text{--}1150\text{ cm}^{-1}$ ) for all chain lengths, with the exception of zinc pivalate, suggesting systematic vibrational motion as expected for chains in the all *trans* conformation.

Group III consists of zinc pivalate—a short, but sterically bulky ligand. The  $^{67}\text{Zn}$  NMR data are consistent with a large  $C_Q$  value ( $\sim 11.0$  MHz) indicating a distorted arrangement around the zinc center. The  $^{13}\text{C}$  NMR resonance for the carboxylate carbon is a singlet more deshielded than the corresponding resonances for the other compounds in the series. The FTIR spectrum shows characteristics of a bridging bidentate ligand with  $\nu_s - \nu_{as} = \sim 140\text{ cm}^{-1}$ .

The addition of solid-state NMR analyses to the traditional FTIR and X-ray diffraction techniques allows for the more exclusive grouping of the zinc carboxylates studied here. The  $C_Q$  is highly sensitive to the local environment around the zinc center for each carboxylate. The application of this technique and the expansion into dynamic nuclear polarization (DNP)-based NMR techniques to enhance the zinc signal in zinc white containing paint films could provide invaluable information to the conservation community by probing the local zinc structure relating to the formation of the zinc-ionomer network and the subsequent formation of zinc soaps.

## Author contributions

Molly Wagner: Writing-original draft; conceptualization; formal analysis; investigation; visualization; Roberta Pigliapochi: conceptualization; writing-review & editing; Valeria Di Tullio: conceptualization; writing-review & editing; Jaclyn Catalano: conceptualization; writing-review & editing; Nicholas Zumbulyadis: conceptualization; funding acquisition; writing-review & editing; Silvia A. Centeno: conceptualization; funding acquisition; writing-review & editing; Xiaoling Wang: investigation; writing-review & editing; Kuizhi Chen: investigation; writing-review & editing; Ivan Hung: investigation; writing-review & editing; Zhehong Gan: funding acquisition; investigation; supervision; writing-review & editing; Michael R. Dworzak: investigation; formal analysis; writing-review & editing; Glenn P. A. Yap: funding acquisition; formal analysis; supervision; writing-review & editing; Cecil Dybowski: concep-

tualization; funding acquisition; supervision; writing-review & editing.

## Conflicts of interest

There are no conflicts of interest to declare.

## Acknowledgements

The work described here was supported by the National Science Foundation under grants DMR-1608594 and DMR-1608366 to The Metropolitan Museum of Art and the University of Delaware. A portion of this work was performed at the National High Magnetic Field Laboratory, which is supported by the National Science Foundation Cooperative Agreement No. DMR-1644779 and the State of Florida. Development of the SCH magnet and NMR instrumentation was supported by NSF (DMR-1039938 and DMR-0603042) and NIH P41 GM122698. GPAY thanks the National Institutes of Health for grant S10 OD026896A. The authors acknowledge Gerald E. Decker and Casey A. Rowland for assistance in obtaining the X-ray structures.

## References

- 1 R. Bossert, *J. Chem. Educ.*, 1950, **27**(1), 10–15.
- 2 E. Ordonez and J. Twilley, *Anal. Chem.*, 1997, **69**(13), 416A–422A.
- 3 M.-C. Corbeil and L. Robinet, *Powder Diffr.*, 2002, **17**(1), 52–60.
- 4 M. L. Kastens and F. R. Hansen, *Ind. Eng. Chem.*, 1949, **41**(10), 2080–2090.
- 5 S. Basel, K. Bhardwaj, S. Pradhan, A. Pariyar and S. Tanang, *ACS Omega*, 2020, **5**, 6666–6675.
- 6 C. Higgitt, M. Spring and D. Saunders, *Natl. Gallery Tech. Bull.*, 2003, **24**, 75–95.
- 7 J. van der Weerd, M. Geldof, L. S. van der Loeff, R. M. A. Heern and J. J. Boon, *Zeitschrift für Kunsttechnologie und Konservierung.*, 2003, **17**, 407–416.
- 8 A. E. Jacobsen and W. H. Gardner, *Ind. Eng. Chem.*, 1941, **33**, 1254–1256.
- 9 K. Helwig, J. Poulin, M.-C. Corbeil, E. Moffatt and D. Duguay, in *Issues in Contemporary Oil Paint*, ed. K. Jan van den Berg, A. Burnstock, M. de Keijzer, J. Krueger, T. Learner, A. de Tagle and G. Heydenreich, Springer, Switzerland, 2014, pp. 167–184.
- 10 G. Osmond, Zinc Soaps: An Overview of Zinc Oxide Reactivity and Consequences of Soap Formation in Oil-Based Paintings, in *Metal Soaps in Art, Conservation and Research*, ed. F. Casadio, K. Keune, P. Noble, A. Van Loon, E. Hendriks, S. A. Centeno and G. Osmond, Springer, Switzerland, 2019, pp. 25–46.
- 11 P. Noble, in *Metal Soaps in Art, Conservation and Research*, ed. F. Casadio, K. Keune, P. Noble, A. Van Loon, E.



- Hendriks, S. A. Centeno and G. Osmond, Springer, Switzerland, 2019, pp. 1–22.
- 12 A. Burnstock, in *Metal Soaps in Art, Conservation and Research*, ed. F. Casadio, K. Keune, P. Noble, A. Van Loon, E. Hendriks, S. A. Centeno and G. Osmond, Springer, Switzerland, 2019, pp. 243–262.
- 13 . A. Sadek, S. Choopun, W. Wlodarski, S. Ippolito and K. Kalantar-zadeh, *IEEE Sens. J.*, 2007, 7, 919–924.
- 14 A. Tsukazaki, A. Ohtomo, T. Onuma, M. Ohtani, T. Makino, M. Sumiya, K. Ohtani, S. Chichibu, S. Fuke, Y. Segawa, H. Ohno, H. Koinuma and M. Kawaski, *Nat. Mater.*, 2005, 4, 42–46.
- 15 V. P. Shetti, S. D. Bukkitgar, K. R. Reddy, C. V. Reddy and T. M. Aminabhavi, *Biosens. Bioelectron.*, 2019, 141, 111417.
- 16 H. Kühn, in *Artists' Pigments A Handbook of Their History and Characteristics*, ed. R. Feller, Archetype Publications, London, 1986, vol. 1, pp. 169–186.
- 17 N. Eastaugh, V. Walsh, T. Chaplin and R. Siddall, *Pigment Compendium. A Dictionary of Historical Pigments*, Elsevier Butterworth-Heinemann, Burlington, 2004.
- 18 J. Catalano, A. Murphy, Y. Yao, N. Zumbulyadis, S. Centeno and C. Dybowski, *Understanding the Dynamics and Structure of Lead Soaps in Oil Paintings Using Multinuclear NMR in Metal Soaps in Art: Conservation and Research*, ed. F. Casadio, K. Keune, P. Noble, A. van Loon, E. Hendriks, S. Centeno and G. Osmond, Springer, Heidelberg, 2019. DOI: [10.1007/978-3-319-90617-1](https://doi.org/10.1007/978-3-319-90617-1).
- 19 L. Robinet and M.-C. Corbeil, *Stud. Conserv.*, 2003, 48, 23–40.
- 20 J. van der Weerd, A. van Loon and J. J. Boon, *Stud. Conserv.*, 2005, 50, 3–22.
- 21 M. J. Plater, B. De Silva, T. Gelbrich, M. B. Hursthouse, C. L. Higgitt and D. R. Saunders, *Polyhedron*, 2003, 22, 3171–3179.
- 22 L. Baij, J. Hermans, B. Ormsby, P. Noble, P. Iedema and K. Keune, *Heritage Sci.*, 2020, 8, 1–23.
- 23 S. Garrappa, E. Kočí, S. Švarcová, P. Bedzička and D. Hradil, *Microchem. J.*, 2020, 156, 104842.
- 24 K. Keune and J. J. Boon, *Stud. Conserv.*, 2007, 52, 161–176.
- 25 K. Keune and G. Boevé-Jones, in *Issues in Contemporary Oil Paint*, ed. K. J. van den Berg, A. Burnstock, M. de Keijzer, J. Kruger, T. Learner, A. de Tagle and G. Heydenreich, Springer International Publishing, Switzerland, 2014, pp. 283–294.
- 26 J. D. J. van den Berg, N. D. Vermist, L. Carlyle, M. Holcapek and J. J. Boon, *J. Sep. Sci.*, 2004, 27, 181–199.
- 27 M. Lazzari and O. Chiantore, *Polym. Degrad. Stab.*, 1999, 65, 303–313.
- 28 J. J. Boon, F. Hoogland and K. Keune, in *34<sup>th</sup> Annual Meeting of the American Institute for Conservation of Historic and Artistic Works*. Ed. P. H. Mar, AIC Postprints, Providence, RI, 2006, pp. 16–23.
- 29 S. A. Centeno and D. Mahon, *The Metropolitan Museum of Art Bulletin, Summer*, The Metropolitan Museum of Art, New York, 2009, pp. 12–19.
- 30 K. Keune, A. van Loon and J. J. Boon, *Microsc. Microanal.*, 2011, 17, 690–701.
- 31 A. Artesani, *Mater. Chem. Phys.*, 2020, 255, 123640.
- 32 J. Catalano, Y. Yao, A. Murphy, N. Zumbulyadis, S. A. Centeno and C. Dybowski, *Appl. Spectrosc.*, 2014, 68, 280–286.
- 33 J. Catalano, A. Murphy, Y. Yao, F. Alkan, N. Zumbulyadis, S. A. Centeno and C. Dybowski, *J. Phys. Chem. A*, 2014, 118, 7952–7958.
- 34 J. Catalano, A. Murphy, Y. Yao, G. P. A. Yap, N. Zumbulyadis, S. A. Centeno and C. Dybowski, *Dalton Trans.*, 2015, 44, 2340–2347.
- 35 T. Kobayashi, F. A. Perras, A. Murphy, Y. Yao, J. Catalano, S. A. Centeno, C. Dybowski, N. Zumbulyadis and M. Pruski, *Dalton Trans.*, 2017, 46, 3535–3540.
- 36 E. Kočí, J. Rohlíček, L. Kobera, J. Plocek, S. Švarcová and P. Bezdička, *Dalton Trans.*, 2019, 48, 12531.
- 37 S. Švarcová, E. Kočí, P. Bezdička, S. Garrappa, L. Kobera, J. Plocek, J. Brus, M. Št'astny and D. Hradil, *Dalton Trans.*, 2020, 49, 5044.
- 38 S. Švarcová, E. Kočí, J. Plocek, A. Zhankina, J. Hradilová and P. Bezdička, *J. Cult. Herit.*, 2019, 38, 8–19.
- 39 R. Barannikov, E. Kočí, P. Bezdička, L. Kobera, A. Mahun, J. Rohlíček, J. Plocek and S. Švarcová, *Dalton Trans.*, 2022, 51, 4019.
- 40 T. J. Bastow, *J. Phys.: Condens. Matter*, 1996, 8, 11309–11315.
- 41 G. Wu, S. Kroeker and R. E. Wasylshen, *Inorg. Chem.*, 1995, 34, 1595–1598.
- 42 A. C. Kunwar, G. L. Turner and E. Oldfield, *J. Magn. Reson.*, 1986, 69, 124–127.
- 43 S. Sham and G. Wu, *Can. J. Chem.*, 1999, 77, 1782–1787.
- 44 R. Ida and G. Wu, *J. Phys. Chem. A*, 2002, 106, 11234–11239.
- 45 F. H. Larsen, A. S. Lipton, H. J. Jakobsen, N. C. Nielsen and P. D. Ellis, *J. Am. Chem. Soc.*, 1999, 121, 3783–3784.
- 46 A. S. Lipton, C. Bergquist, G. Parkin and P. D. Ellis, *J. Am. Chem. Soc.*, 2003, 125, 3768–3772.
- 47 A. S. Lipton, R. W. Heck and P. D. Ellis, *J. Am. Chem. Soc.*, 2004, 126, 4735–4739.
- 48 A. S. Lipton, R. W. Heck, G. R. Staeheli, M. Valiev, W. A. De Jong and P. D. Ellis, *J. Am. Chem. Soc.*, 2008, 130, 6224–6230.
- 49 A. Sutrisno, V. Terskikh, Q. Shi, Z. Song, J. Dong, S. Ding, W. Wang, B. Provost, T. Daff, T. Woo and Y. Huang, *Chem. – Eur. J.*, 2012, 18, 12251–12259.
- 50 R. Madsen, A. Qiao, J. Sen, I. Hung, K. Chen, Z. Gan, S. Sen and Y. Yue, *Science*, 2020, 367, 1473–1476.
- 51 M.-C. Corbeil and L. Robinet, *Powder Diffr.*, 2002, 17, 52–60.
- 52 W. Clegg, D. Harbron, C. Homan, P. Hunt, I. Little and B. Straughan, *Inorg. Chim. Acta*, 1991, 186, 51–60.
- 53 Y. Yao, G. P. A. Yap, J. Catalano, N. Zumbulyadis, S. A. Centeno and C. Dybowski, *CSD Commun.*, 2018, 1576707.
- 54 Apex3, Bruker AXS Inc., Madison, WI, 2015.
- 55 G. M. Sheldrick, *Acta Crystallogr., Sect. A: Found. Adv.*, 2015, 71, 3–8.



- 56 G. M. Sheldrick, *Acta Crystallogr., Sect. C: Struct. Chem.*, 2015, **71**, 3–8.
- 57 R. K. Harris, E. D. Becker, S. M. C. De Menezes, R. Goodfellow and P. Granger, *Pure Appl. Chem.*, 2001, **73**, 1795–1818.
- 58 S. G. J. van Meerten, W. M. J. Franssen and A. P. M. Kentgens, *J. Magn. Reson.*, 2019, **301**, 56–66.
- 59 C. Morcombe and K. Zilm, *J. Magn. Reson.*, 2003, **162**, 479–486.
- 60 W. Clegg, I. Little and B. P. Straughan, *Acta Crystallogr., Sect. C: Cryst. Struct. Commun.*, 1986, **42**, 1701–1703.
- 61 F. Lacouture, J. Peultier, M. Francois and J. Steinmetz, *Acta Crystallogr., Sect. C: Cryst. Struct. Commun.*, 2000, **56**, 556–557.
- 62 W. Clegg, I. R. Little and B. R. Straughan, *Acta Crystallogr., Sect. C: Cryst. Struct. Commun.*, 1987, **43**, 456–457.
- 63 J. Blair, R. A. Howie and J. L. Wardell, *Acta Crystallogr., Sect. C: Cryst. Struct. Commun.*, 1993, **49**, 219–221.
- 64 R. A. Taylor and H. A. Ellis, *Acta Crystallogr., Sect. E: Struct. Rep. Online*, 2008, **64**, m895.
- 65 R. A. Taylor, H. A. Ellis, P. Maragh and N. White, *J. Mol. Struct.*, 2006, **787**, 113–120.
- 66 P. Segedin, N. Lah, M. Zefram, I. Leba and L. Golic, *Acta Chim. Slov.*, 1999, **46**, 173–184.
- 67 J. Peultier, M. Francois and J. Steinmetz, *Acta Crystallogr., Sect. C: Cryst. Struct. Commun.*, 1999, **55**, 2064–2065.
- 68 P. Nelson and R. Taylor, *Appl. Petrochem. Res.*, 2014, **4**, 253–285.
- 69 A. Mesbah, C. Juers, E. François, R. Steinmetz and J. Steinmetz, *Z. Kristallogr.*, 2007, **26**, 593–598.
- 70 J. J. Hermans and K. Helwig, *Appl. Spectrosc.*, 2020, **74**(12), 1505–1514.
- 71 M. Beerse, K. Keune, P. Iedema, S. Wouterson and J. Hermans, *ACS Appl. Polym. Mater.*, 2020, **2**, 5674–5685.
- 72 A. E. Goeta and J. A. K. Howard, *Chem. Soc. Rev.*, 2004, **33**, 490–500.
- 73 J. Hermans, K. Keune, A. van Loon and P. D. Iedema, *J. Anal. At. Spectrom.*, 2015, **30**, 1600–1608.
- 74 J. Hermans, K. Keune, A. van Loon and P. D. Iedema, *Phys. Chem. Chem. Phys.*, 2016, **18**, 10896–10905.
- 75 F. Gabrieli, F. Rosi, A. Vichi, L. Cartechini, L. P. Buemi, S. G. Kazarian and C. Miliani, *Anal. Chem.*, 2017, **89**, 1283–1289.
- 76 J. J. Hermans, L. Baij, M. Koenis, K. Keune, P. Iedema and S. Woutersen, *Sci. Adv.*, 2019, **5**, EAAW3592.
- 77 T. Ishioka, K. Maeda and I. Kanesaka, in *Spectroscopy of Biological Molecules: New Directions*, ed. J. Greve, G. J. Puppels and C. Otto, Springer, Dordrecht, 1999, pp. 313–314.
- 78 T. Ishioka, K. Maeda, I. Watanabe, S. Kawauchi and M. Harada, *Spectrochim. Acta, Part A*, 2000, **56**, 1731–1737.
- 79 N. Alcock, V. Tracy and T. Waddington, *Dalton Trans.*, 1976, **21**, 2243–2246.
- 80 R. Taylor and H. Ellis, *Spectrochim. Acta, Part A*, 2007, **68**, 99–107.
- 81 P. Nelson, R. Taylor and H. Ellis, *J. Mol. Struct.*, 2013, **1034**, 75–83.
- 82 P. Nelson, H. Ellis and R. Taylor, *J. Mol. Struct.*, 2010, **982**, 10–15.
- 83 S. Barman and S. Vasudevan, *J. Phys. Chem. B*, 2007, **111**, 5212–5217.
- 84 C. A. Maines, D. Rogala, S. Lake and M. Mecklenberg, Deterioration in Abstract Expressionist Paintings: Analysis of Zinc Oxide Paint Layers in Works from the Collection of the Hirshhorn Museum and Sculpture Garden, Smithsonian Institution, *MRS Proc. Online*, 2011, **1319**, DOI: [10.1557/opl.2011.733](https://doi.org/10.1557/opl.2011.733).
- 85 Y. Huang and A. Sutrisno, *Annu. Rep. NMR Spectrosc.*, 2014, **81**, 1–46.
- 86 S. F. Dec, M. F. Davis, G. E. Maciel and C. E. Bronniman, *Inorg. Chem.*, 1993, **32**, 955–959.
- 87 G. Wu, *Chem. Phys. Lett.*, 1998, **298**, 375–380.
- 88 F. H. Larsen, J. Skibsted, H. J. Jakobsen and N. Nielsen, *J. Am. Chem. Soc.*, 2000, **122**, 7080–7086.
- 89 K. H. Mroué and W. P. Power, *J. Phys. Chem. A*, 2010, **114**, 324–335.

