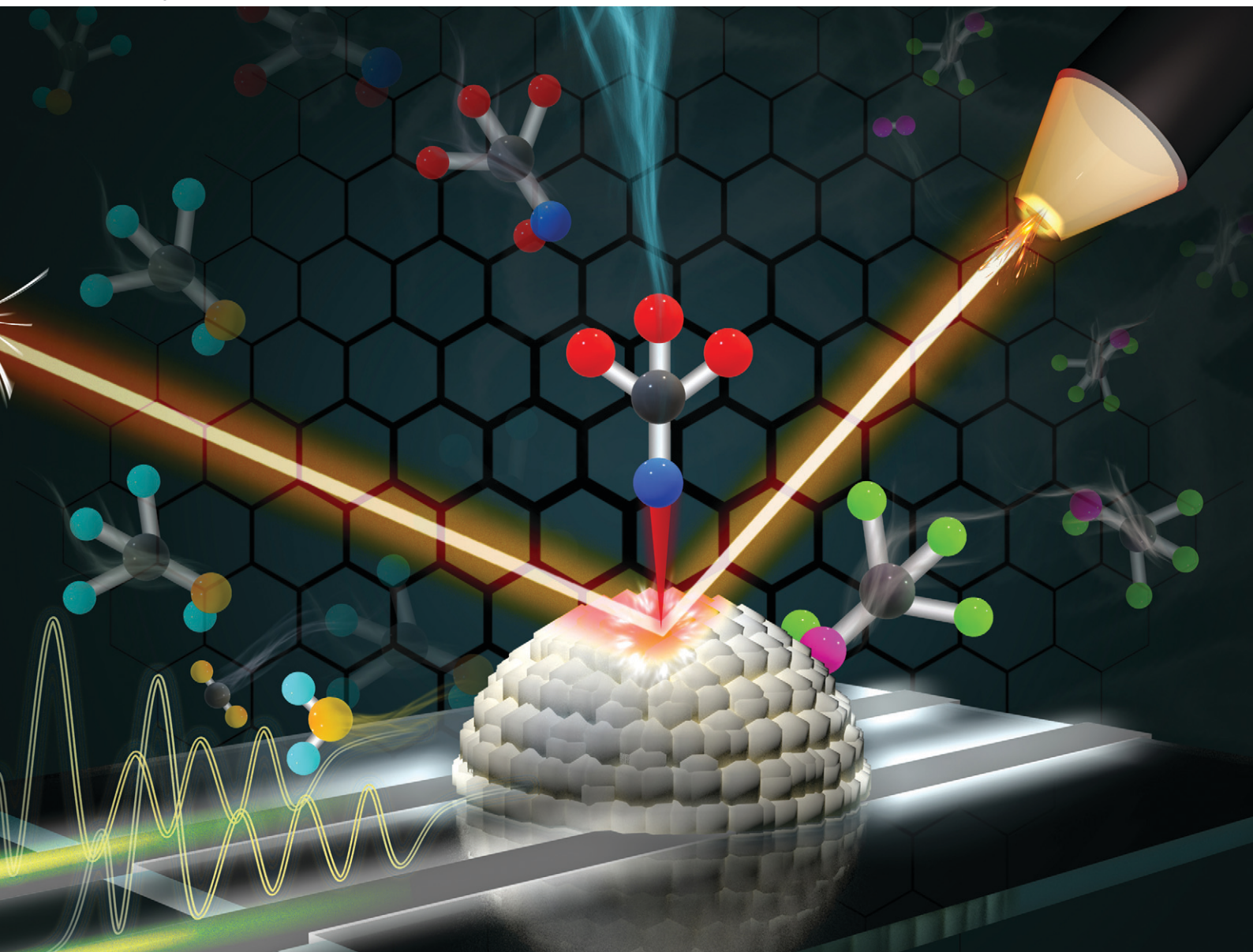


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Operando monitoring of a room temperature nanocomposite methanol sensor†

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The sensing of volatile organic compounds by composites containing metal oxide semiconductors is typically explained *via* adsorption–desorption and surface electrochemical reactions changing the sensor's resistance. The analysis of molecular processes on chemiresistive gas sensors is often based on indirect evidence, whereas *in situ* or *operando* studies monitoring the gas/surface interactions enable a direct insight. Here we report a cross-disciplinary approach employing spectroscopy of working sensors to investigate room temperature methanol detection, contrasting well-characterized nanocomposite (TiO₂@rGO-NC) and reduced-graphene oxide (rGO) sensors. Methanol interactions with the sensors were examined by (quasi) *operando*-DRIFTS and *in situ*-ATR-FTIR spectroscopy, the first paralleled by simultaneous measurements of resistance. The sensing mechanism was also studied by mass spectroscopy (MS), revealing the surface electrochemical reactions. The *operando* and *in situ* spectroscopy techniques demonstrated that the sensing mechanism on the nanocomposite relies on the combined effect of methanol reversible physisorption and irreversible chemisorption, sensor modification over time, and electron/O₂ depletion–restoration due to a surface electrochemical reaction forming CO₂ and H₂O.

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Introduction

Methanol, a volatile organic compound (VOC), is extensively used as an organic solvent in chemical, biomedical, pharmaceutical and other industries.^{1,2} It may also serve as a cost-efficient and clean-burning liquid fuel for automobiles, limiting emissions of sulphur oxides (SO_x), nitrogen oxides (NO_x) and particulate matter to the environment.³ However, methanol is toxic as it can be absorbed through the skin or lungs leading to methanol poisoning. Moreover, methanol toxicity increases *via* metabolism to formaldehyde and formic acid, which upon accumulation leads to adverse health effects.^{4,5} This illustrates the importance of an efficient methanol sensor capable of detection in an ambient environment.

Metal oxide semiconductors (MOSCs) such as In₂O₃, ZnO, WO₃, SnO₂, NiO, Co₃O₄, CuO, *etc.* have been widely used as

VOC gas sensing materials, based on their resistivity change upon exposure to analytes (chemiresistivity).^{6–12} To achieve higher sensitivity at relatively low working temperatures, MOSCs have been doped with noble metals such as Au, Ag, Pt, Pd, *etc.*^{13–16} Still, conventional MOSCs typically require working temperatures of 90–400 °C to desorb water chemisorbed on the sensor surface and to provide free sites for analyte chemisorption.¹⁷ High MOSC working temperatures maintain the sensitivity toward VOCs over repeated cycles,^{18,19} but induce high-power consumption, reduce the sensor lifetime and pose a security risk for sensor operation in a flammable environment.^{6–11}

The next generation of VOC sensors thus operates at room temperature (RT), benefitting from power-saving, eco-friendliness, sustainability, and higher safety.^{20,21} Recently, several materials have been tested as RT-VOC sensors, such as carbon-based materials including multiwall carbon nanotubes with polyaniline, carbon derivatives in polyetherimide with a liquid crystal polymer, and graphene doped polyaniline. These sensors were able to detect methanol vapor of 50–500 ppm, 300–1200 ppm, and 50–100 ppm, respectively.^{22–24} Moreover, MOSC based materials, such as CdO with polyaniline and a thin layer of Au, Ag, Pt or Cu on top of In₂O₃–SnO₂, have also shown detection limits of 100 ppm and 200–900 ppm, respectively. When In₂O₃ was combined with Ti₃C₂T_x (an MXene), methanol detection was further enhanced (5–100 ppm). Nevertheless, CdO and In₂O₃

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possess significant toxicity to humans: carcinogenic effect, renal toxicity, alveolar proteinosis, emphysema or interstitial fibrosis.^{25,26} Thus, in terms of biocompatibility, using TiO₂ nanomaterials as RT-VOC sensors has attracted attention. TiO₂ nanotubes and quantum dots exhibited promising methanol sensitivity of 100–300 ppm and 1000 ppm at RT, respectively.^{27,28} However, the sensors' response time and detection limit were rather high or required UV light for sufficient performance.

For MOSc-based VOC sensing devices, changes in electrical resistance and response–recovery times upon exposure to variable VOC concentrations have been reported repeatedly, but the sensing mechanism was often described rather hypothetically.¹⁷ To obtain a fundamental understanding of the sensor's functional properties rather requires real-time studies of the working sensor surface interacting with VOCs, *i.e.*, an *in situ* or *operando* approach hardly attempted before. Even more so as VOCs may irreversibly bind to MOScs, limiting the sensor functionality. For instance, methanol irreversibly adsorbs on nanocrystalline TiO₂ and requires ≈425 °C for removal of the adsorbed methoxy species.²⁹ Similarly, methanol adsorbs dissociatively on pure ZnO, as well as on ZnO decorated with Au nanoparticles, thus requiring 275–320 °C for operation. It was also reported that Au enhanced the ZnO reducibility and facilitated the generation of oxygen vacancies,³⁰ which is considered an important factor in VOC sensing by MOSc surfaces.

To improve the current understanding, we have combined and linked surface sensitive vibrational studies^{31–33} of metal/oxide nanomaterials with RT sensor technology, simultaneously monitoring methanol–surface interactions and the corresponding resistance changes in real time. *Operando* diffuse reflectance infrared Fourier transform spectroscopy (DRIFTS) and *in situ* attenuated total reflectance-Fourier transform infrared spectroscopy (ATR-FTIR) were applied to evaluate the methanol gas sensing performance of green-synthesized and well-characterized TiO₂ nanoparticles, reduced graphene oxide (rGO) and their nanocomposite (TiO₂@rGO-NC). Based on simultaneous mass spectroscopy, the mechanism of room temperature methanol gas sensing was directly confirmed. Furthermore, the multidisciplinary approach presented herein may stimulate analogous in-depth studies and the development of versatile RT-VOC sensors.

Experimental

Materials

Titanium(IV) oxysulfate–sulfuric acid hydrate (TiOSO₄·xH₂SO₄·yH₂O) (Sigma-Aldrich®), graphite powder (Sigma-Aldrich®), methanol (analytical grade, by Sigma-Aldrich®), iso-propanol (Sigma-Aldrich®), potassium permanganate (>99%, Sigma-Aldrich®), L-ascorbic acid (99%, Sigma-Aldrich®), NaOH (Sigma-Aldrich®), deionized water (diH₂O) (Milli-Q®), hydrochloric acid solution (37%, Merck®), olive

leaf waste (kindly provided by Dr. Chiara Giosue, research fellow at Università Politecnica delle Marche, Italy).

A transparent glass substrate (dimensions: 3.2 cm × 2.4 cm), silver electrodes (purity = 99%, dimensions: 6 cm × 0.2 cm), precision digital multimeter (UNI-T® UT61E), sensor/electrode multimeter-connector (PalmSens®), hotplate magnetic stirrer-RH basic (IKA®, Germany), water bath ultrasonicator (FALC®, Italy), heating oven (LAPORTA®, Italy), electric grinder (KENWOOD® 500 W), centrifuge (ECCL31R Multispeed, Thermo Scientific®, USA), Brunauer–Emmett–Teller (BET) analyzer by Micromeritics®-ASAP-2020, X-ray powder diffractometer (Bruker®), TGA (Mettler®-851), UV/vis spectrometer (Lambda 750, PerkinElmer®), diffuse reflectance infrared Fourier transform spectroscopy (DRIFTS) using a Bruker Vertex 70 spectrometer with a DRIFTS cell (Pike), attenuated total reflectance-Fourier transform infrared (ATR-FTIR) spectrometer (Vertex-70, Bruker Optics®), ZnSe crystal (dimensions: 52 mm × 20 mm × 2 mm), quadrupole mass spectrometer (QMS) (Prisma Plus QMG 220, Pfeiffer Vacuum), high resolution transmission electron microscopy (HR-TEM) using an FEI TECNAI F20 field emission microscope.

Synthesis and characterization procedures

Synthesis of the sensor materials. Green synthesis of titanium oxide nanoparticles (TiO₂-NPs) and reduced graphene oxide (rGO) was achieved following a previously reported, but modified, method.^{34,35} Briefly, the green synthesis of TiO₂-NPs and a nanocomposite of TiO₂-NPs and rGO (TiO₂@rGO-NC) and the sensor production were accomplished by the following eight steps shown in Fig. 1.

i. Mature olive leaves, as a potential waste harvested in November, were thoroughly washed with dH₂O to remove solid particles (impurities) and dried at room temperature in the dark to avoid photodissociation of secondary metabolites. To ensure repeatability and to match the metabolic profile necessary for TiO₂-NP synthesis, it is important that the olive leaves are collected at a specific time of the year (preferentially November). The harvested leaves should be stored at >57% humidity at room temperature for further use.³⁶

ii. The dried olive leaves were ground to a fine powder. For water extract preparation, 25 g of the olive leaf powder was soaked in 500 mL of diH₂O for 12 h, and then heated to 85 °C for 3 h in a hot water bath. The suspension was allowed to cool to room temperature and was filtered using Whatman filter paper no. 1 to obtain an olive leaf extract.

iii. 500 mL of the prepared extract in a reaction flask was put on a hot plate magnetic stirrer at 85 °C and 150 rpm. Thereafter, 0.1 M TiOSO₄·xH₂SO₄·yH₂O was added to the preheated extract. After 20 min, the pH of the reaction mixture was adjusted to ~4 by dropwise addition of 3 M NaOH solution. Next, heating was turned off and the reaction mixture was cooled to room temperature while stirring at 150 rpm.



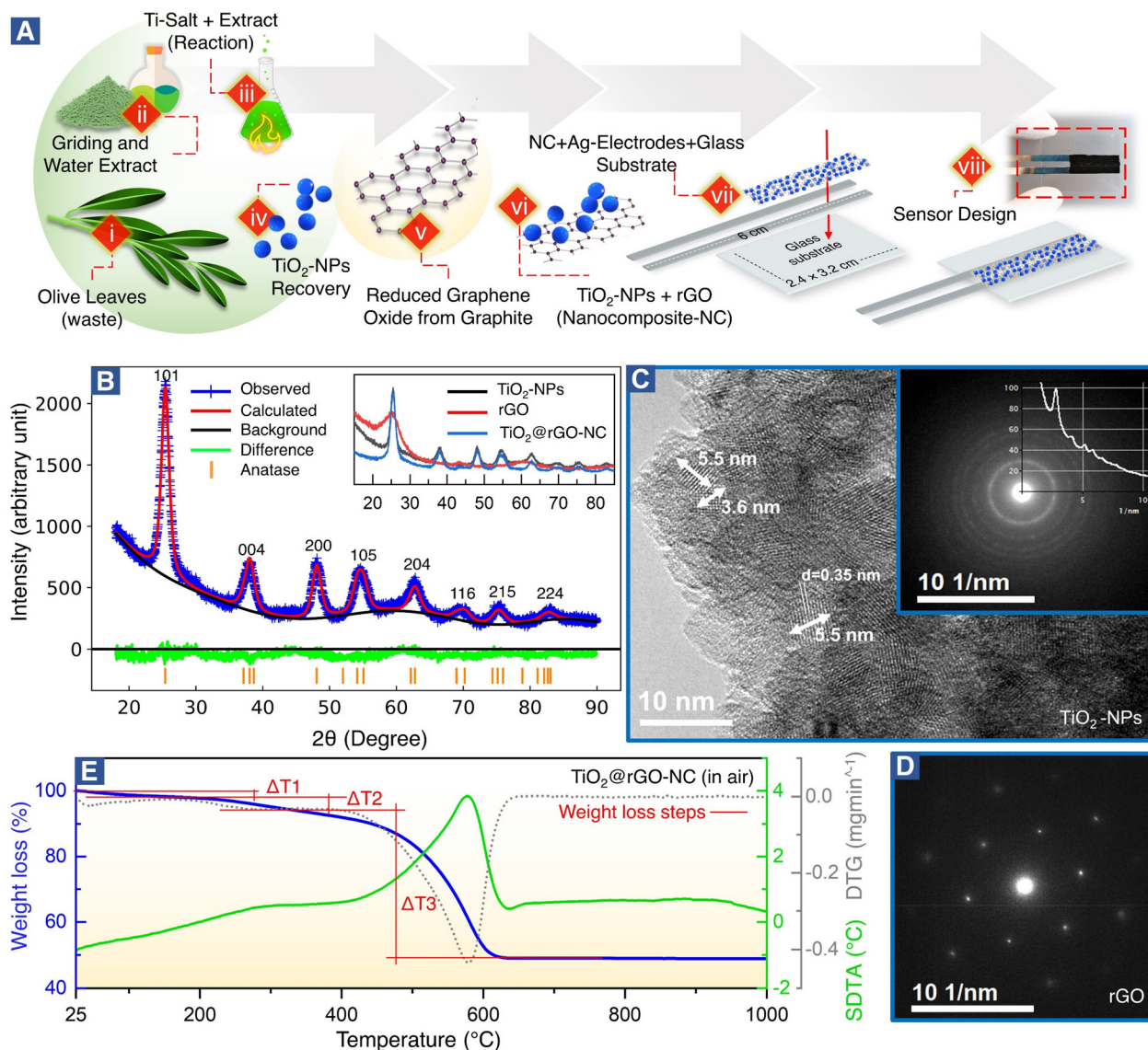


Fig. 1 (A) Schematics (steps i to viii) elaborating the green synthesis of titanium oxide nanoparticles (TiO₂-NPs), a nanocomposite (NC) of TiO₂-NPs and reduced graphene oxide (TiO₂@rGO-NC), and the sensor design (see Experimental for more details). (B) Crystal structure of TiO₂-nanoparticles (TiO₂-NPs) via Rietveld refinement. Inset: Measured diffractograms of the as-synthesized TiO₂-NPs, reduced-graphene oxide (rGO) and nanocomposite (TiO₂@rGO-NC). (C) HR-TEM image of unsupported TiO₂-NPs, inset: SAED pattern of TiO₂-NPs. (D) SAED pattern of rGO. (E) TG analysis (blue), DTG analysis (grey) and SDTA (green) of TiO₂@rGO-NC in air.

iv. TiO₂-NPs were collected by centrifugation at 8000 rpm and washed three times with diH₂O to remove uncoordinated secondary metabolites from the extract. TiO₂-NPs were dried in a hot air oven at 90 °C for 24 h. Next, the dried TiO₂-NPs were calcined at 350 °C for 3 h in a furnace before being kept in an airtight jar for further use.

v. The production of reduced graphene oxide (rGO) was achieved by adding 2 g of graphite powder in a reaction flask immersed in an ice-water bath and maintained below 10 °C. After this, 100 mL of concentrated H₂SO₄ was added under continuous stirring. Then, 6 g of KMnO₄ was gradually added. Next, the suspension was stirred at room temperature for 20 min followed by 10 min ultrasonication and stirring (process repeated 10 times). Subsequently, the reaction was

quenched by addition of 400 mL diH₂O and further ultrasonicated for 2 h. The suspension pH was adjusted at ~6 by addition of 1 M NaOH solution, which was further ultrasonicated for 1 h. Then, 20 g of L-ascorbic acid was dissolved in 200 mL diH₂O and slowly added to the suspension at room temperature while increasing the temperature to 95 °C for 1 h. The rGO was recovered by filtration using cellulose filter paper and washed with 1 M HCl solution and diH₂O to achieve neutral pH. Finally, the filtrate was vacuum-dried to obtain rGO pellets.

vi. Different weight ratios (25%, 50%, and 75%) of TiO₂-NPs were homogenized with rGO to prepare TiO₂@rGO-NC. Typically, TiO₂-NPs and rGO were mixed in the presence of diH₂O (ratio of 100 mg of solid in 2 mL of liquid) and



ultrasonicated for 15 min to ensure proper homogenization of TiO₂-NPs and rGO. One part of the as-prepared paste (TiO₂@rGO-NC) was deposited between two Ag-electrodes to produce a sensor. The remaining part was dried at 90 °C in an oven to remove water and kept for further use in characterization studies.

vii. The sensor design comprises 3 components: two Ag-electrodes (6 cm × 0.2 cm), a glass substrate (3.2 cm × 2.4 cm) and a thin layer (≈25 μm) of sensing material (*e.g.*, TiO₂@rGO-NC) deposited between the Ag-electrodes. The Ag-electrodes were immobilized with adhesive on the glass substrate and with the distance between the two Ag-electrodes matching the multimeter connector for resistance measurements.

viii. After deposition of the sensing material between the two Ag-electrodes, each sensor was stabilized and dried at 60 °C in an oven for 24 h. The prepared sensors were stored in a desiccator for further use.

Characterization of the sensor materials. X-ray diffraction (XRD) was carried out at RT using a Cu-Kα radiation source ($\lambda = 1.5406 \text{ \AA}$) at an operating voltage of 40 kV (current of 30 mA). The XRD measurements were recorded at the angle of diffraction (2θ) between 10° and 90°. The crystallographic parameters of the sensor materials were identified through Rietveld refinement using HighScore Plus® (v2021) connected with the ICDD® database. The XRD results were plotted using GSAS®-II (v4776) and OriginPro® (v2021).

The morphology and crystal structure of the various materials (TiO₂-NPs, TiO₂@rGO-NC) were evaluated by high resolution-transmission electron microscopy (HR-TEM), electron energy loss spectrometry (EELS) and selected area electron diffraction (SAED), using an FEI TECNAI F20 field emission microscope equipped with a GATAN GIF Tridiem energy filter and a GATAN Rio16 CMOS camera. For HR-TEM, the samples were prepared as follows: 5 mg TiO₂-NPs (or TiO₂@rGO-NC) and 5 mL diH₂O were ultrasonicated for 20 min. Using a micropipette, 1 drop of the ultrasonicated suspension was deposited on a commercial TEM copper grid covered with a lacy carbon film. Before each measurement, the TEM sample was vacuum dried for 10 min.

Brunauer-Emmett-Teller (BET) analysis of the as-prepared materials (TiO₂-NPs, rGO and TiO₂@rGO NC) was carried out on a Micromeritics surface area and porosity analyzer. To determine the specific surface area (SSA), N₂ adsorption at -196 °C was performed on an ASAP 2020 Micromeritics apparatus on 0.5 g sample, preheated under vacuum (<0.013 mbar) at 50 °C for 3 h. Evaluation of the SSA was based on the linear part of the BET analysis, and pore size distributions were obtained by applying the Barrett-Joyner-Halenda (BJH) equation to the isotherm desorption branch, and the total pore volume was estimated from the N₂ uptake at a P/P_0 of 0.99.

Thermogravimetry (TG), derivative thermogravimetry (DTG) and simultaneous differential thermal analysis (SDTA) of the as-prepared TiO₂-NPs, rGO and TiO₂@rGO NC were used to monitor the thermal properties in terms of mass

change and exothermicity/endothemicity, both in air and nitrogen. The gas flow was set to 60 mL min⁻¹ in the temperature range of 25–1000 °C, with a rate of 10 °C min⁻¹. For each sample, 5–20 mg of the dried powder sample were measured in an alumina-150 μL sample holder.

Room temperature methanol gas sensing and *operando* spectroscopic analysis

The setup for combined room temperature methanol gas sensing and (quasi) *operando* DRIFTS, shown in Fig. 2, is described below.

Design of the methanol gas sensing chamber

Chemiresistive methanol gas sensing measurements were performed at room temperature in an enclosed glass chamber under a continuous flow (60 mL min⁻¹) of dry synthetic air (SA). Before starting each measurement, the sensor was placed inside the chamber, connected to the multimeter-connector, and allowed to stabilize under a continuous SA flow for 24 h. The desired concentration (3×10^3 – 2.1×10^4 ppm) of methanol was introduced with the steady SA flow using microinjection, with 1 μL methanol yielding 3000 ppm. For lower concentrations of 75–150 ppm, methanol was diluted with diH₂O (ratio of 1:36).

The methanol molecules (gas) *vs.* chamber air molecules were calculated accordingly:

$$\text{Density of methanol (room temperature)} = 0.7913 \text{ g mL}^{-1}$$

$$1 \text{ } \mu\text{L} = 0.001 \text{ mL}$$

$$0.001 \text{ mL} \times 0.7913 \text{ g mL}^{-1} = 0.0007913 \text{ g} = 0.7913 \text{ mg}$$

$$X = 0.7913 \text{ mg} \times 6.022 \times 10^{23} \text{ molecules per mol} / 32.040 \text{ mg}$$

$$X = 1.4875 \times 10^{19} \text{ molecules}$$

calculating air molecules:

$$Y = 6.022 \times 10^{23} \text{ molecules per mol} \times 200 \text{ mL} / 24.460 \text{ mL}$$

$$Y = 4.9240 \times 10^{21} \text{ molecules}$$

So, the methanol molecules per air molecules are

$$1.4875 \times 10^{19} / 4.9240 \times 10^{21} = 0.003 \text{ (3000 ppm)}$$

The electrical resistance R was measured using a high-resistance multimeter, read out by UT61E software (v.4.01).



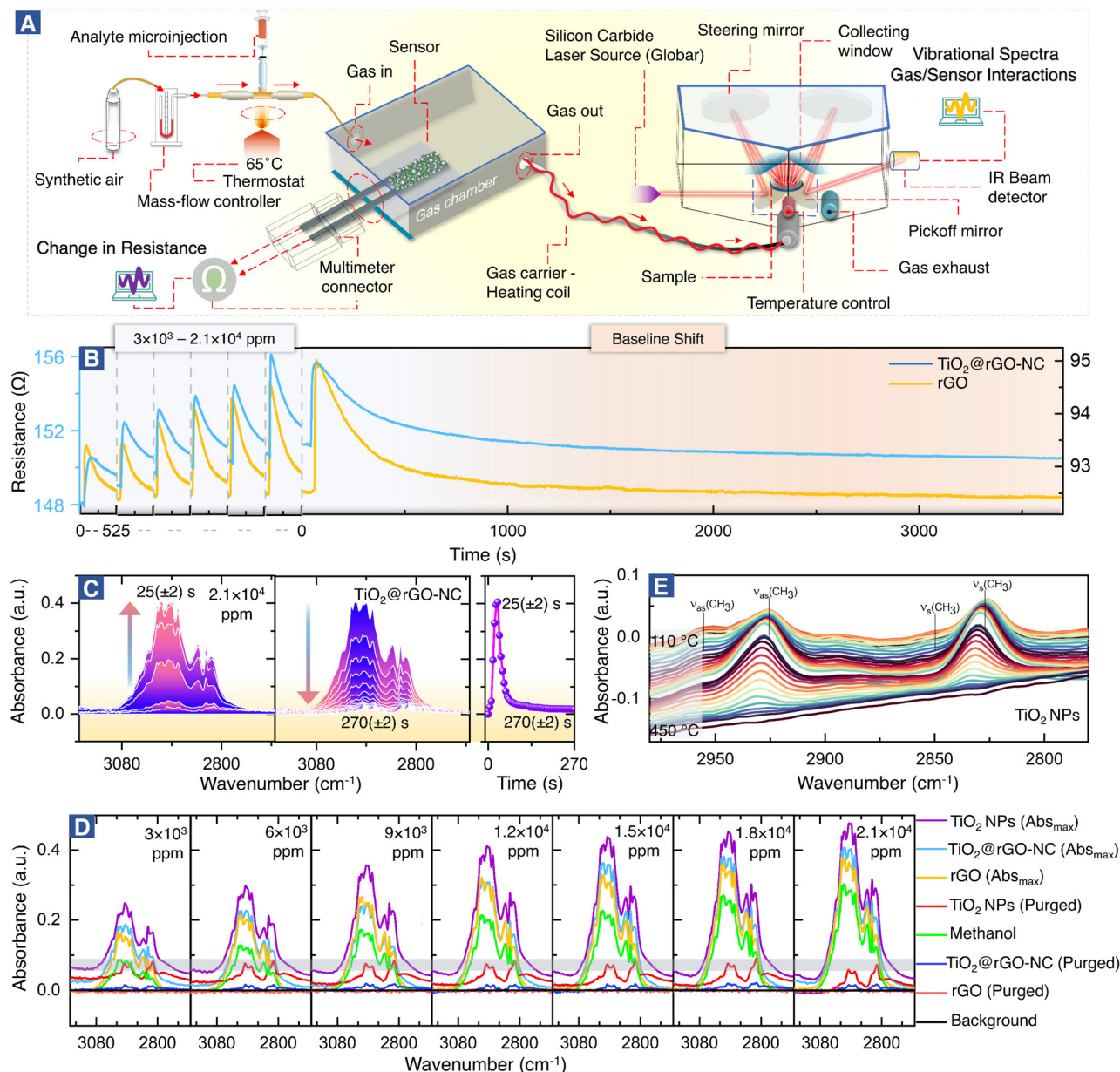


Fig. 2 Quasi *operando* resistance/DRIFTS studies of room temperature methanol gas sensing by different sensor materials. (A) Experimental design, (B) methanol sensing and initial baseline shift, (C) time-resolved DRIFTS, adsorption-desorption studies of methanol over the 50% NC sensor, (D) *operando* DRIFTS spectra of methanol adsorption at different concentrations (3×10^3 to 2.1×10^4 ppm), and (E) temperature programmed desorption (TPD) of surface adsorbed methanol on TiO_2 -NPs monitored by DRIFTS.

The response time and recovery time of the sensor were calculated using eqn (1) and (2) which are shown in Fig. 4B,

$$\text{Response time} = T_{\text{max}} - T_{\text{start}} \quad (1)$$

$$\text{Recovery time} = T_{\text{baseline}} - T_{\text{max}} \quad (2)$$

Response time is defined as the time of maximum (100%) alteration in resistance in the presence of methanol (T_{max}) minus the time at which the methanol pulse was started (T_{start}). Recovery time is defined as the time needed by the sensor to achieve the baseline (resistance in air) (T_{baseline})

after pulsing methanol minus the time of maximum (100%) alteration in resistance upon methanol pulsing (T_{max}).

Operando-diffuse reflectance infrared Fourier transform spectroscopy (DRIFTS) and temperature programmed desorption (TPD)

Molecular interactions of methanol gas with the sensing materials (e.g. rGO or TiO_2 @rGO-NC) were analyzed in real time using a downstream DRIFTS cell under the same experimental conditions as those of the resistance measurements in the environmental chamber (Fig. 1). The IR



spectrometer was equipped with a silicon carbide IR source (Globar®), a liquid nitrogen-cooled mercury cadmium telluride (MCT) detector and a commercial DRIFTS mirror unit. Briefly, 50 mg of sensing material was placed in a porous ceramic sample cup and mounted in the DRIFTS cell. The spectra of the sensing material in SA served as the background and the spectra of the DRIFTS cell without the sensing material but only methanol gas (under the same SA flow) were also acquired. After a methanol gas pulse (3×10^3 – 2.1×10^4 ppm) was introduced, IR-spectra (64 scans) were recorded in 5 s intervals *via* the OPUS® (v6.5) software. For each measurement, the Abs_{max} value is defined as the maximum IR absorption for a particular methanol concentration, while the purged value is defined as the IR absorption when there is no further change over time.

Finally, for temperature-programmed methanol desorption, the sample was heated to 450 °C (10 °C min^{-1}) in the DRIFTS cell while recording the IR spectra.

***In situ* attenuated total reflectance-Fourier transform infrared (ATR-FTIR) spectroscopy**

Interactions of liquid methanol with powdered sensing materials (*e.g.*, rGO, TiO₂-NPs or TiO₂@rGO-NC) were examined at room temperature in real time by *in situ* ATR-FTIR. The spectrometer was equipped with a liquid nitrogen-cooled mercury cadmium telluride (MCT) detector and a commercial ATR unit. The samples prepared in diH₂O were deposited over the surface of the ZnSe crystal and air dried. The spectra of the sensing material in a steady flow of He (8 mL min^{-1}) were taken as the background, while the spectra without the sensing material and with only methanol were taken as the positive control. IR spectra (64 scans) were recorded at 5 s intervals *via* the OPUS® software (v6.5) monitoring liquid methanol (1 mL) injection in a helium (He) environment inside the ATR-FTIR cell. IR spectra were recorded until there was no further signal change.

***Operando* mass spectroscopy (MS) of room temperature methanol gas sensing**

The mechanism of room temperature methanol gas sensing, previously proposed to occur *via* a surface electrochemical reaction of methanol removing charge carriers, was directly examined by *operando*-MS paralleled by resistance measurements. Thus, the methanol gas sensing chamber was connected to a quadrupole mass spectrometer (Prisma Plus QMG 220, Pfeiffer Vacuum; Fig. 5). The experimental conditions were the same as those for sensor performance measurements, but for improved detection of methanol gas and molecular species resulting from the surface electrochemical reaction, the methanol concentration was increased to $3\text{--}9 \times 10^5$ ppm (injection volume = 0.1–0.3 mL). Molecular species were detected using the QMS equipped with a secondary electron multiplier (SEM) detector.

Results and discussion

Green synthesis and characterization of sensor materials

Fig. 1A(steps i–viii) shows the green synthesis of titanium oxide nanoparticles (TiO₂-NPs), a nanocomposite (NC) of TiO₂-NPs and reduced graphene oxide (TiO₂@rGO-NC), and the sensor fabrication (for further details see the Experimental section). Olive leaves, an organic waste, are a rich source of naturally occurring metal ions chelating secondary metabolites, including secoiridoids and flavonoids. Notably, oleuropein and quercetin are highly reactive compounds and possess proven potential for reduction of metal ions to metal nanoparticles.^{35,37} In the present study, the water extract of olive leaves was used as a natural reagent to react with titanium oxydisulfate in the synthesis of TiO₂-NPs. Moreover, when the reaction conditions (*i.e.*, extract concentration, reaction temperature and pH) were compared to the classical green synthesis, it was realized that the extract of mature plant leaves was more reactive than that of young plant leaves.³⁴ A possible reason may be the accumulation of nitrogenous organic compounds (NOCs) upon ageing. Also, various types of NOCs are well known to assist in producing small nanoclusters (*e.g.*, Au₁₀, Au₁₂ and Au₁₄).³⁸ In the current study, in addition to oleuropein and quercetin, NOC participation was advantageous for achieving TiO₂-NPs of small size. Thus, using olive leaf waste in the synthesis of NPs as an alternative to synthetic reagents is not only efficient, but also represents a sustainable and environmentally friendly route. The advantages of the green synthesis method reported herein over the previously reported ones are detailed in Table S1.†

The crystallography of the as-prepared TiO₂-NPs, rGO and a nanocomposite of TiO₂-NPs and rGO (TiO₂@rGO-NC) was examined by X-ray diffraction (XRD). The broad diffraction peaks (Fig. 1B) revealed the small crystallite size of the green synthesized TiO₂-NPs. The Bragg peaks were in good agreement with ICDD 04-016-2837,³⁹ indicating that TiO₂-NPs were anatase.²⁸ The crystal structure of TiO₂-NPs was further evaluated *via* Rietveld refinement (Fig. 1B and S1†), showing unit cell parameters of $a = 3.79\text{ Å}$, $b = 3.79\text{ Å}$ and $c = 9.48\text{ Å}$. The average crystallite size of TiO₂-NPs was calculated using Debye–Scherrer's equation:

$$D = 0.9\lambda/\beta \cos \theta \quad (3)$$

where D is the average crystallite size (nm), λ is the X-ray wavelength (1.5406 Å), β is the full width at half maximum (FWHM) in radians, and θ is the Bragg angle. The resulting average crystallite size of TiO₂-NPs was 4.1 nm, further detailed in Table S2.†

The XRD patterns of rGO and TiO₂@rGO-NC are shown as an inset of Fig. 1B. The broad peak of rGO at $2\theta = 25.27^\circ$ indicates that thick carbon stacks of graphite segregated into a few layers of rGO, as commonly observed during exfoliation of graphite into graphene, graphene oxide or rGO.⁴⁰ The results are also consistent with the powder UV-vis spectra of



rGO (Note S1 and Fig. S2†). Furthermore, XRD confirmed that the structural properties of TiO_2 -NPs remained unchanged upon preparation of the nanocomposite (TiO_2 @rGO-NC).

The grain size of anatase (Fig. 1C), as determined by high-resolution transmission electron microscopy (HR-TEM) under extended Scherzer conditions, was found to be <6 nm, in good agreement with XRD. The fine-grained polycrystalline areas (Fig. 1C) were identified to be anatase by means of selected area electron diffraction (SAED; inset of Fig. 1C) and *via* the energy loss near edge structure (ELNES). The recorded ELNES, as shown in Fig. S3A,† is clearly characteristic of anatase- TiO_2 (more details in Note S2†). The observed lattice planes are attributed to the (101) planes of anatase.

TiO_2 -NPs possess tremendous surface catalytic potential;^{41,42} however, green synthesized, homogeneous and small sized TiO_2 -NPs have been scarcely reported in the literature.^{43,44} Recently, anatase- TiO_2 quantum dots with a crystallite size of 4.2 nm were obtained by chemical synthesis in a time-consuming and complex procedure utilizing acetylacetone, *n*-butanol, and 4-dodecylbenzene sulfonic acid.²⁸ In the current study, TiO_2 -NPs with a small crystallite size (~ 4 nm) and a homogeneous morphology seem to result from the synergistic effect of secondary metabolites from olive leaves, reaction temperature, and optimized pH. Maurya *et al.* described green synthesized TiO_2 NPs of 13 nm in size of the pure anatase phase, whereas NPs prepared by a chemical synthesis method were 16 nm large.⁴⁵ Also, Tsega *et al.* reported pure anatase- TiO_2 -NPs in a pH range between 4.4 and 6.8. A drastic change in crystallite size from 24 to 8 nm was observed when the pH was lowered to ~ 3.2 ; but it also resulted in a loss of phase purity.⁴⁶ A similar effect of low pH on the TiO_2 -NP phase purity was observed by Isley *et al.*⁴⁷ Hence, a pH of ~ 4 seems to be the borderline to achieve pure anatase TiO_2 -NPs.⁴⁴

The large flakes of graphene (rGO) (Fig. S3B†) yielded a single crystalline SAED pattern (Fig. 1D), showing the typical hexagonal symmetry of (0001) orientation. The TiO_2 @rGO nanocomposite (Fig. S3B†) exhibited a homogeneous distribution of TiO_2 -NPs within the rGO matrix.

As sensor materials may need higher temperatures for operation or reactivation, the thermal properties of the prepared nanomaterials were analyzed by thermogravimetry (TG), derivative thermogravimetry (DTG) and simultaneous differential thermal analysis (SDTA) (Fig. 1E and S4†). The temperature of maximum decomposition (T_n), the relative weight loss (Δm) and the temperature of 5% mass loss (defined as the decomposition temperature, T_d) are summarized in Table S3† for each material. The thermal analysis of TiO_2 @rGO-NC (Fig. 1E) was distinctively different from that of TiO_2 -NPs and rGO (Fig. S4†), showing intermediate values. In air, the first weight loss until 150 °C is due to the removal of water. This endothermic process overlaps at $T > 200$ °C with the exothermic thermal decomposition of organic compounds and a maximum weight loss at $T = 290$ °C. At $T > 400$ °C, due to the presence

of oxygen, a sharp oxidative decomposition of graphene occurred with an exothermic peak and a T_n of 576 °C. The presence of TiO_2 -NPs increased the thermal stability of graphene in air, increasing the temperature of maximum weight loss from $T = 531$ °C to 576 °C, while the T_d increased from 290 °C to 314 °C. For $T > 650$ °C no further weight loss was recorded. This demonstrates that the TiO_2 @rGO-NC is thermally stable up to at least 300 °C and would be compatible with high temperature sensing. The thermal stability of the other sensor materials is presented in Note S3, Fig. S4 and Table S3.†

The specific surface area (SSA) of the sensing materials is an important factor for the interaction with analyte molecules and was measured using N_2 adsorption-desorption isotherms, calculated by the Brunauer-Emmett-Teller (BET) and Barrett-Joyner-Halenda (BJH) (dV/dw) methods. The BET surface areas of TiO_2 -NPs, rGO and 50 wt% TiO_2 @rGO-NC were $101.5 \text{ m}^2 \text{ g}^{-1}$, $36.4 \text{ m}^2 \text{ g}^{-1}$ and $70.8 \text{ m}^2 \text{ g}^{-1}$, respectively. Moreover, the BJH cumulative pore volume and pore width of TiO_2 -NPs, rGO and 50% TiO_2 @rGO-NC were $0.11 \text{ cm}^3 \text{ g}^{-1} - 5.9 \text{ nm}$, $0.13 \text{ cm}^3 \text{ g}^{-1} - 37.4 \text{ nm}$ and $0.08 \text{ cm}^3 \text{ g}^{-1} - 6.8 \text{ nm}$, respectively, as depicted in Fig. S5 (more details in Note S4†). Accordingly, upon preparation of the TiO_2 @rGO-NC the SSA was preserved. Not unexpectedly, the high SSA of TiO_2 -NPs is beneficial for MeOH sensing.^{28,48,49}

Operando-DRIFTS and *in situ*-ATR-FTIR spectroscopy during room temperature methanol sensing

Fig. 2A shows the setup for simultaneous room temperature measurements of the resistance R and the corresponding DRIFTS spectra of the sensor materials (for a pulse, the time delay between measurements is only ≤ 5 s). In analogy with heterogeneous catalysis, when spectroscopy is paralleled with measurements of catalytic performance, the current combined resistance/DRIFTS analysis is termed (quasi) *operando*. Fig. 2B shows how the different sensor materials respond to methanol pulses for a range of concentrations. Selected time-resolved DRIFTS spectra of methanol adsorption/desorption on TiO_2 @rGO-NC are shown in Fig. 2C (for other concentrations and rGO see Fig. S6 and S7†). The panel on the right shows the time-evolution of the peak maximum. Note that for TiO_2 @rGO-NC the response times of resistance and IR absorption coincide very well (Fig. S8†). In contrast, for rGO alone the IR response was much shorter (10–20 s) while the resistance response was delayed (35–45 s). Moreover, the recovery time differed, suggesting different bonding of the adsorbates.

Fig. 2D shows the DRIFTS spectra for various methanol concentrations and sensor materials, displaying the maximum peak intensities of each (the relative absorbance scales nearly linear with the methanol concentration; Fig. S9†), as well as those after purging. The spectra in the C–H stretching vibration range, with intensities in the order TiO_2 -NPs $>$ TiO_2 @rGO-NC $>$ rGO, characterize the adsorbed



species. The absorption bands between 2800 and 3000 cm^{-1} result from $\nu(\text{CH}_3)$ vibrations of physisorbed methanol and chemisorbed methoxy (CH_3O). The spectra of methanol in the absence of a sensor (on the aluminum mirror; positive control) and of the background (sensor in pure synthetic air (SA); negative control) are included in Fig. 2D for reference. Interestingly, after sample purging by flowing synthetic air, all peaks disappeared for rGO, whereas some remained for TiO_2 and $\text{TiO}_2@\text{rGO-NC}$. This clearly demonstrates the important role of TiO_2 -NPs, and that most of the methanol adsorption was still reversible. To fully remove the irreversible CH_3O species from TiO_2 *via* recombinative desorption, temperatures $>400^\circ\text{C}$ are needed (Fig. 2E and S10†). This also explains the initial baseline shifts in resistance upon methanol sensing, which are attributed to irreversible methoxy adsorption modifying the sensor resistance. Fig. S10† further validates the correlation between the concentration of TiO_2 -NPs (0, 25, 50 and 75% in rGO) *versus* the initial baseline shift (sensor stability) resulting from irreversible CH_3O adsorption on TiO_2 -NPs at room temperature. The higher the TiO_2 -NPs concentration, the higher the baseline distortion, thus affecting the sensor performance over time.

To study the interaction of the sensor materials with the analyte in more detail, liquid methanol was examined by *in situ* ATR-FTIR in the range of 1000–3600 cm^{-1} , as shown in Fig. 3 (with magnified insets of selected ranges) and Fig. S11–S13.† Once more, TiO_2 -NPs enhanced the interaction, confirming the results for gaseous methanol.

After drying in He, the ATR of methanol adsorbed on TiO_2 showed four resonances at 2821, 2850, 2923, and 2950 cm^{-1} . The first two peaks are assigned to the $\nu_s(\text{CH}_3)$ mode of chemisorbed methoxy (CH_3O) and physisorbed CH_3OH on TiO_2 , respectively. The latter two peaks result from the Fermi resonance of chemisorbed methoxy (ν_{F1} , CH_3O) and physisorbed methanol (ν_{F2} , CH_3OH), respectively.^{50–53} Manzoli *et al.* reported that the bands between 2000 and 1000 cm^{-1} may be related to differently coordinated methoxy species on TiO_2 , with the band at 1156 cm^{-1} characterizing on-top methoxy species on Ti^{3+} near an oxygen vacancy. The band at 1135 cm^{-1} may be due to on-top species on Ti^{4+} sites.⁵⁴ The shoulder at 1025 cm^{-1} and the peak at 1120 cm^{-1} were previously assigned to $\nu(\text{C}-\text{O})$ of methanol physisorbed and chemisorbed on TiO_2 , respectively.⁵⁵

Sensor performance

To evaluate the sensor performance in more detail (Fig. 4), each sensor was stabilized in dry synthetic air for 24 h, and then exposed to the methanol gas stimulus to establish the adsorption-induced baseline shift described above (*cf.* Fig. 2B). Once the sensor was stabilized, the room temperature methanol gas sensing performance of $\text{TiO}_2@\text{rGO-NC}$ and rGO (as reference) was tested for 3×10^3 – 2.1×10^4 ppm of methanol in a steady flow of SA using the chemiresistive methanol gas sensing setup (Fig. 2A).

Fig. 4A shows the relationship between the methanol gas stimulus at room temperature and the induced resistance change. It is apparent that the response increased linearly with methanol concentration. As shown in Fig. 4B, the response time of the sensor is defined in terms of time needed by the sensor to achieve 100% alteration in resistance for a particular methanol gas stimulus, while the recovery time refers to the time taken to recover from 100% to the baseline. Overall, 50% $\text{TiO}_2@\text{rGO-NC}$ showed the shortest response time of ~ 15 s for both 1.5×10^4 and 1.8×10^4 ppm. Moreover, it is also clear from Fig. 4A and C that addition of TiO_2 -NPs to rGO not only shortened the sensor response time, but also the recovery time, with the latter being equally important for RT sensor operation.²⁷ Apart from the linearity of resistance *vs.* methanol concentration and the response–recovery behavior, the signal reproducibility at both 3×10^3 and 2.1×10^4 ppm methanol was tested (Fig. 4D), with both sensing materials ($\text{TiO}_2@\text{rGO-NC}$ and rGO) showing excellent reproducibility. Additionally, the estimated limit of detection (LOD defined as 3 times the standard deviation SDV of the sensor in SA (baseline values without methanol gas), divided by the slope of the linear part of the calibration curve)⁵⁶ was ~ 213 and ~ 847 ppm for the 50% $\text{TiO}_2@\text{rGO-NC}$ sensor and rGO, respectively. Moreover, the $\text{TiO}_2@\text{rGO-NC}$ also exhibited methanol sensitivity at much lower methanol concentrations of 75 to 150 ppm, as shown in Fig. S14.†

In light of previous studies (Fig. 4E), the current TiO_2 -NPs and the nanocomposite exhibit promising properties for methanol sensing, outperforming many previously reported materials in the response time. When synthesized by a chemical method, TiO_2 -quantum dots were able to detect 1000 ppm of methanol vapor at room temperature with a response time of ~ 200 s.²⁸ Similarly, Ti as a two-dimensional metal carbide (MXene) could detect 100 ppm of methanol gas at room temperature with a response time of ~ 300 s.⁵⁷ The detailed comparison in Table S4 shows the relevance of the $\text{TiO}_2@\text{rGO-NC}$ sensor obtained *via* a facile green synthesis, exhibiting good methanol sensitivity, short response/recovery time and reproducibility/stability, with the methanol sensing process even examined by *operando* spectroscopy herein.

Mechanistic study of room temperature methanol sensing by *operando*-resistance/mass spectroscopy

Based on previous studies and the current *operando* and *in situ* spectroscopy of the working sensor, the mechanism of methanol gas sensing in terms of the ionosorption model¹⁵⁸ can be evaluated.

The sensor in air

The conduction channel in the sensor is through rGO, but the resistance is modified by TiO_2 , which improves the response and recovery time. Generally, TiO_2 shows an n-type response to gas stimuli, but in the current case it exhibits a p-type response which increases the resistance upon gas exposure,^{28,59} likely due to structure defects in the



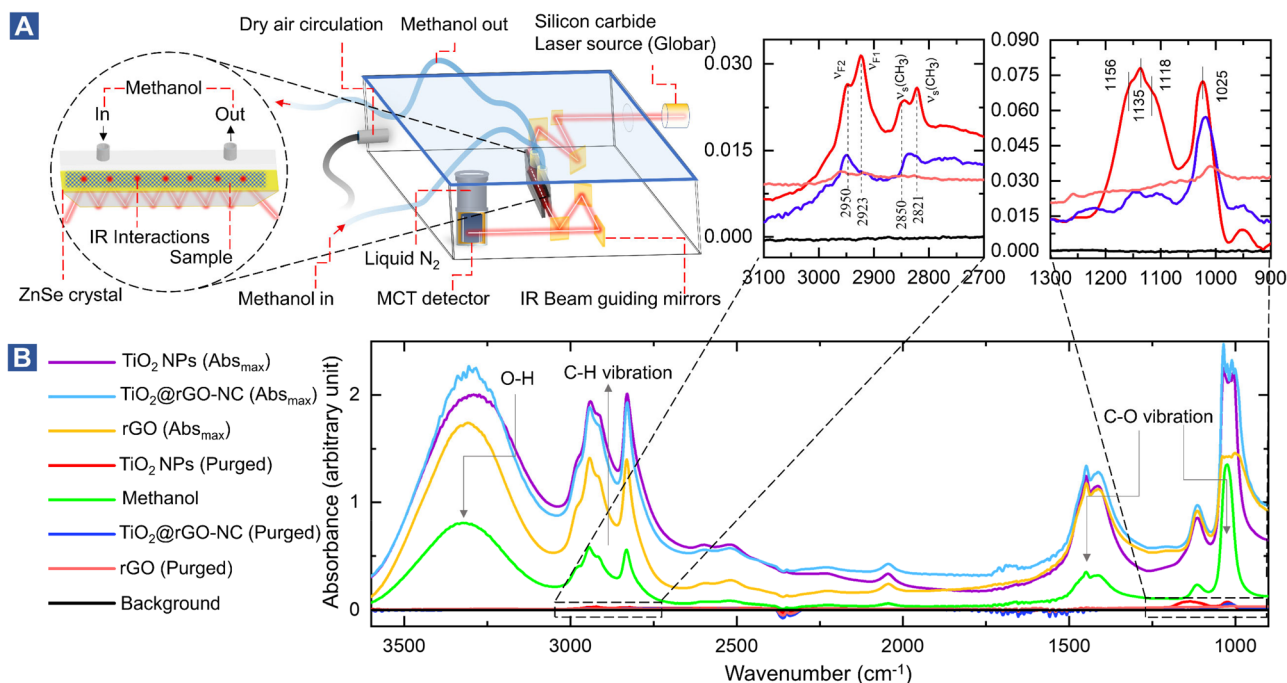
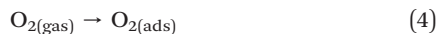


Fig. 3 (A) Experimental design of *in situ* ATR-FTIR: vibrational spectroscopy at the liquid/solid interface (methanol on the sensor materials), (B) *in situ* ATR-FTIR spectra of 1 mL MeOH (liquid-phase) injection.

nanocrystals.^{60,61} Green-synthesized TiO₂-NPs behave like a p-type MOSc with hole carriers,²⁸ i.e., when exposed to the dry air carrier gas, oxygen adsorbs on the surface of TiO₂-NPs and captures electrons, forming O⁻ and O²⁻ (eqn (4)–(7)).²⁸ As a result, the charge carrier density on the TiO₂-NP surface increases by formation of a conduction layer (hole accumulation layer, HAL), providing the basis for improved sensing. In this state, the sensor surface is considered stable at room temperature in the SA carrier gas.



Reversible and irreversible methanol adsorption

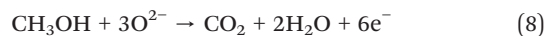
The initial exposure of the sensor to methanol is the critical process, representing the core finding of this study. Using *operando* and *in situ* spectroscopy, it was observed that the sensing materials (TiO₂@rGO-NC and rGO) behaved differently upon methanol gas stimulation. In the case of rGO, the first methanol exposure showed reversible adsorption and physisorbed CH₃OH desorbed when the methanol pulse vanished upon steady air flow. As apparent from (quasi) *operando*-DRIFTS (Fig. 2D), rGO exhibited weaker methanol adsorption than the other sensing material,

due to the low hydrophilicity of rGO, resulting in weak surface adsorption.⁶² This also explains the longer response time of rGO (Fig. 4C), with reversible methanol adsorption on rGO yielding a stable baseline.

The resistance of the rGO was around 100 ohms with a p-type response, and addition of TiO₂ increased the resistance to around 200 Ω. Even more importantly, addition of TiO₂-NPs to the rGO matrix significantly improved the sensor performance. The first methanol exposure resulted in physisorbed CH₃OH and chemisorbed methoxy (CH₃O), the latter being responsible for the baseline shift. Hence, reversible and irreversible RT methanol adsorption on the surface of the sensing materials controls the baseline shift and stability over time.

Surface electrochemical reaction and interface effects

As suggested by previous reports,⁵⁸ the interaction of methanol with TiO₂@rGO-NC initiates a surface electrochemical reaction between CH₃OH and the ionosorbed oxygen species (eqn (8)). This releases free electrons which neutralize the holes in TiO₂ by electron-hole recombination (eqn (9)). Hence, the decrease in the availability of holes results in an increase in sensor resistance, as observed experimentally. Moreover, deposition of TiO₂-NPs on rGO may further accelerate methanol oxidation at the interface, leading to an increased charge carrier recombination rate, also documented by previous observations.⁶³



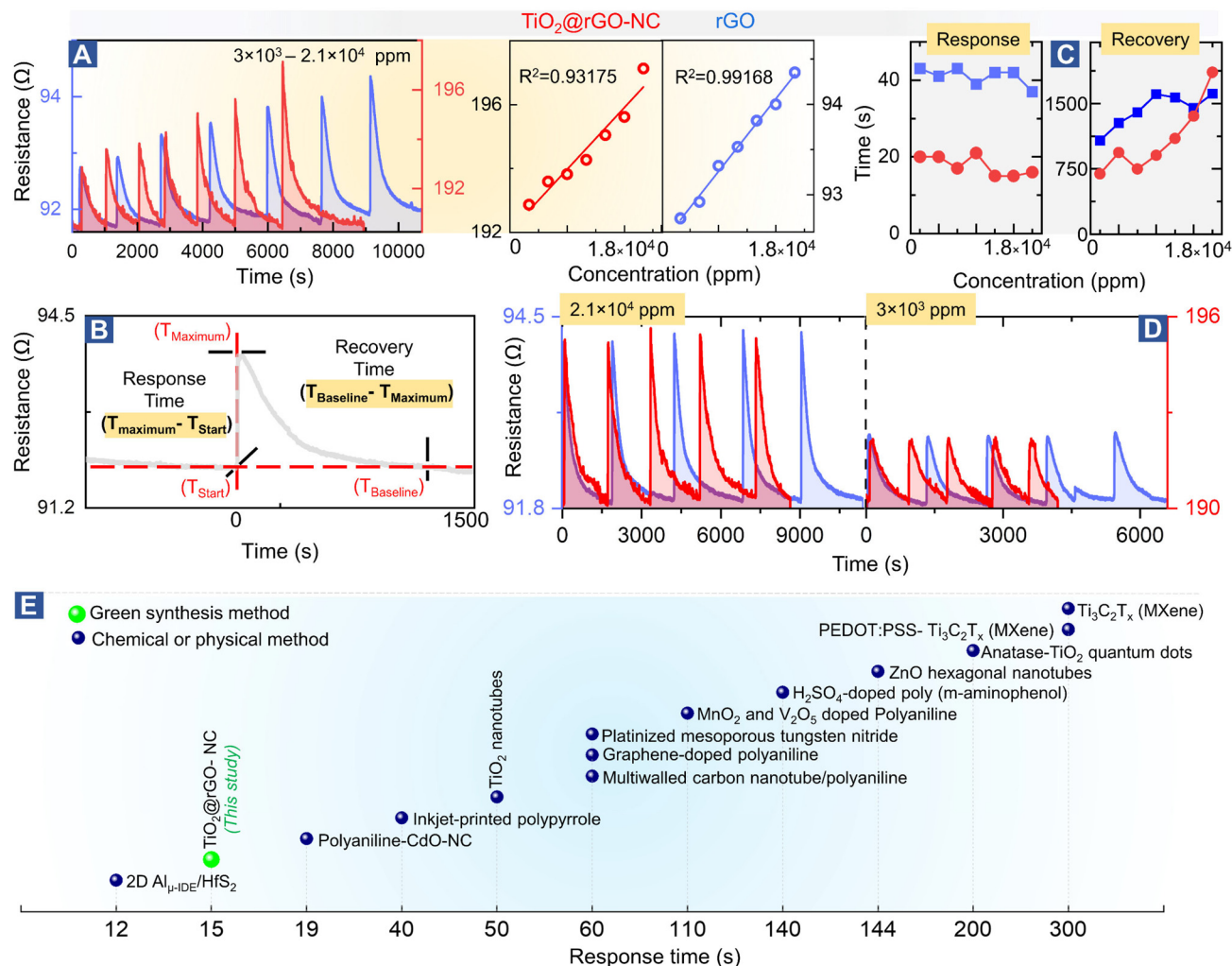


Fig. 4 Methanol gas sensing performance of different materials: (A) room temperature methanol sensing by 50%TiO₂@rGO-NC and rGO at different concentrations (left) and linear fit (right), (B) illustration of the sensor response and recovery time measurement, (C) sensor performance with respect to the response and recovery time, (D) reproducibility of room temperature methanol gas sensing for 50%TiO₂@rGO and rGO at higher and ~10-times lower methanol concentration. For even lower concentrations, see Fig. S14,† and (E) comparison of the room temperature methanol sensing response time of the current and previously reported materials (for more details, see Table S4†).

To observe this process in real time, at least in part, the methanol gas sensing chamber was connected to a mass spectrometer (MS), as shown in Fig. 5A. This allowed us to link resistance changes upon methanol stimulation with simultaneous gas phase analysis, once more in an *operando* mode (Fig. 5B). The MS detection allowed identification of products of the surface electrochemical reaction. At varying concentrations of methanol (3×10^5 – 9×10^5 ppm), H₂O and CO₂ were traced as products of methanol reacting with pre-adsorbed oxygen, according to eqn (8) and Fig. 5C.

Methanol conversion to CO₂ and H₂O due to the surface electrochemical reaction was thus evident from MS, with the tailing MS signals indicating persistent room temperature methanol oxidation. This once more highlights the importance of methanol adsorption-desorption on the surface and interfaces of TiO₂-NPs for the resistance changes. Electron paramagnetic resonance (EPR) spectroscopy could

even directly detect the formation of reactive oxygen species.⁶⁴

However, the production of H₂O by the surface electrochemical reaction or a high humidity at room temperature may affect the sensor performance. To address this issue, methanol gas together with vaporized H₂O (1:36) was introduced to the sensing chamber. Under highly humid conditions (75 ppm methanol in 2925 ppm H₂O), one expects a loss of functionality, but the sensor still detected up to 150 ppm methanol in 5850 ppm H₂O before collapsing (Fig. S14†). This confirms that the sensing material is highly selective towards MeOH over H₂O, as well as resistant towards co-dosed H₂O.

Conclusions

In summary, a nanocomposite methanol sensor material, consisting of TiO₂ nanoparticles and reduced graphene oxide

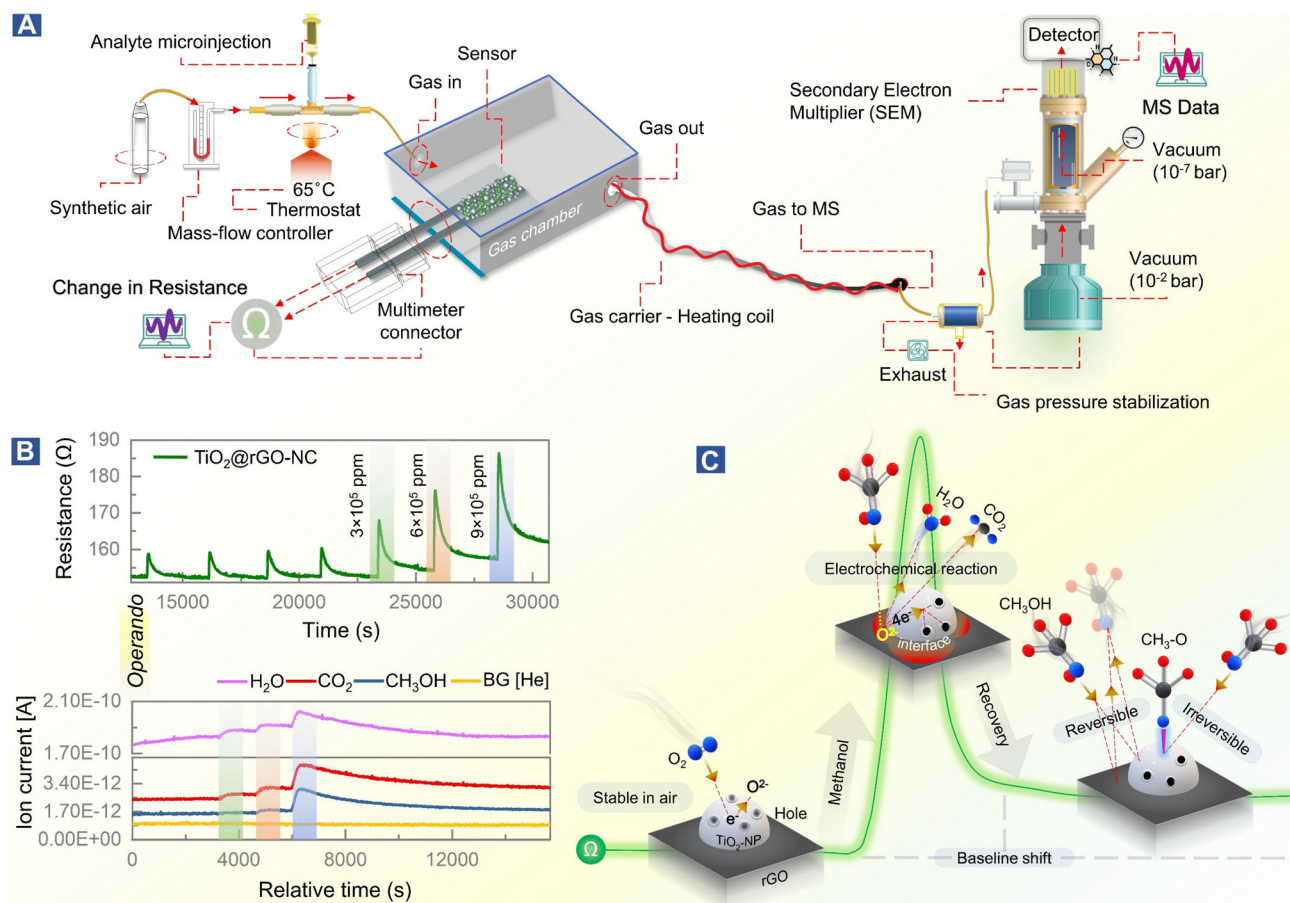


Fig. 5 Room temperature methanol gas sensing on 50% TiO₂@rGO-NC monitored by *operando* mass spectrometry (MS). (A) Experimental design of *operando*-resistance/mass spectroscopy of the surface electrochemical reaction upon methanol gas exposure, (B) change in resistance due to the methanol stimulus with simultaneous detection of product molecules, and (C) illustration of the methanol sensing mechanism; see text for details.

(rGO), was prepared by facile green chemistry routes, employing an organic olive leaf waste extract as an alternative to synthetic reagents. The TiO₂-NPs were 4–6 nm in size and had an anatase structure. Although intended for room temperature sensing, the nanocomposite is even stable to at least 300 °C. Using a setup for (quasi) *operando*-resistance/DRIFTS measurements, the interaction of methanol with the various sensor materials was examined in real time. Methanol exposure increased the resistance, with TiO₂@rGO-NC showing the shortest response and recovery times. The resistance scaled linearly with the methanol concentration and had a high degree of reproducibility. The room temperature molecular interactions between CH₃OH and the sensor surface were identified as reversible and irreversible methanol adsorption, the latter due to methoxy CH₃O formation especially on TiO₂-NPs, leading to a resistance baseline shift. The same interactions were observed for liquid methanol by *in situ* ATR-FTIR. The mechanism of methanol gas sensing in terms of the ionosorption model was confirmed by *operando* mass spectroscopy/resistance measurements, identifying CO₂ and H₂O as products of a surface electrochemical reaction. Methanol adsorption

releases free electrons neutralizing the holes in TiO₂ and thus increasing the overall sensor resistance, which includes a conduction channel through rGO. The presented work may open new opportunities in *operando* monitoring of working sensor materials derived from eco- and environmentally-friendly synthesis approaches.

Author contributions

Qaisar Maqbool: conceptualization, methodology, validation, software, formal analysis, investigation, data curation, writing – original draft, writing – review & editing. Nevzat Yigit: conceptualization, methodology, validation, formal analysis, investigation, data curation, writing – review & editing. Michael Stöger-Pollach: methodology, validation, formal analysis, investigation, writing – review & editing. Maria Letizia Ruello: validation, writing – review & editing. Francesca Tittarelli: validation, resources, writing – review & editing, supervision. Günther Rupprechter: conceptualization, validation, resources, writing – review & editing, supervision.



Conflicts of interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Notes and references

- 1 F. Dalena, A. Senatore, M. Basile, S. Knani, A. Basile and A. Iulianelli, *Membranes*, 2018, **8**, 98.
- 2 S. S. Araya, V. Liso, X. Cui, N. Li, J. Zhu, S. L. Sahlin, S. H. Jensen, M. P. Nielsen and S. K. Kær, *Energies*, 2020, **13**, 596.
- 3 G. A. Olah, *Angew. Chem., Int. Ed.*, 2005, **44**, 2636–2639.
- 4 Z. Nekoukar, Z. Zakariaei, F. Taghizadeh, F. Musavi, E. S. Banimostafavi, A. Sharifpour, N. Ebrahim Ghuchi, M. Fakhar, R. Tabaripour and S. Safanavaei, *Ann. Med. Surg.*, 2021, **66**, 102445.
- 5 M. L. Frenia and J. L. Schauben, *Ann. Emerg. Med.*, 1993, **22**, 1919–1923.
- 6 R. Xing, L. Xu, J. Song, C. Zhou, Q. Li, D. Liu and H. W. Song, *Sci. Rep.*, 2015, **5**, 1–14.
- 7 L. Zhu, W. Zeng, H. Ye and Y. Li, *Mater. Res. Bull.*, 2018, **100**, 259–264.
- 8 S. C. Yeow, W. L. Ong, A. S. W. Wong and G. W. Ho, *Sens. Actuators, B*, 2009, **143**, 295–301.
- 9 S. K. Ayyala and J. A. Covington, *Chemosensors*, 2021, **9**, 247.
- 10 H. Nguyen and S. A. El-Safty, *J. Phys. Chem. C*, 2011, **115**, 8466–8474.
- 11 P. Patil, U. T. Nakate, Y. T. Nakate and R. C. Ambare, *Mater. Sci. Semicond. Process.*, 2019, **101**, 76–81.
- 12 U. T. Nakate, Y. T. Yu and S. Park, *Microelectron. Eng.*, 2022, **251**, 111662.
- 13 Z. Feng, Y. Ma, V. Natarajan, Q. Zhao, X. Ma and J. Zhan, *Sens. Actuators, B*, 2018, **255**, 884–890.
- 14 N. G. Patel, P. D. Patel and V. S. Vaishnav, *Sens. Actuators, B*, 2003, **96**, 180–189.
- 15 V. Postica, A. Vahl, D. Santos-Carballal, T. Dankwort, L. Kienle, M. Hoppe, A. Cadi-Essadek, N. H. De Leeuw, M. I. Terasa, R. Adelung, F. Faupel and O. Lupan, *ACS Appl. Mater. Interfaces*, 2019, **11**, 31452–31466.
- 16 Y. M. Choi, S. Y. Cho, D. Jang, H. J. Koh, J. Choi, C. H. Kim and H. T. Jung, *Adv. Funct. Mater.*, 2019, **29**, 1808319.
- 17 T. Lin, X. Lv, Z. Hu, A. Xu and C. Feng, *Sensors*, 2019, **19**, 233.
- 18 J. van den Broek, S. Abegg, S. E. Pratsinis and A. T. Güntner, *Nat. Commun.*, 2019, **10**, 1–8.
- 19 K. Singh, R. C. Singh and S. Sharma, *J. Phys.: Conf. Ser.*, 2021, **1849**, 012034.
- 20 G. J. Thangamani, K. Deshmukh, T. Kovářík, N. A. Nambiraj, D. Ponnammam, K. K. Sadasivuni, H. P. S. A. Khalil and S. K. K. Pasha, *Chemosphere*, 2021, **280**, 130641.
- 21 X. Li, Z. An, Y. Lu, J. Shan, H. Xing, G. Liu, Z. Shi, Y. He, Q. Chen, R. P. S. Han, D. Wang, J. Jiang, F. Zhang and Q. Liu, *Adv. Mater. Technol.*, 2021, 2100872.
- 22 A. Bora, K. Mohan, D. Pegu, C. B. Gohain and S. K. Dolui, *Sens. Actuators, B*, 2017, **253**, 977–986.
- 23 S. Bag, K. Rath and K. Pal, *Nanotechnology*, 2017, **28**, 205501.
- 24 R. Pal, S. L. Goyal, I. Rawal and S. Sharma, *Iran. Polym. J.*, 2020, **29**, 591–603.
- 25 E. M. Bomhard, *Environ. Toxicol. Pharmacol.*, 2018, **58**, 250–258.
- 26 J. Y. Lee, M. Tokumoto and M. Satoh, *FASEB J.*, 2021, **35**, e21236.
- 27 P. Bindra and A. Hazra, *Sens. Actuators, B*, 2019, **290**, 684–690.
- 28 A. Šutka, R. Eglitis, A. Kuzma, K. Smits, A. Zukuls, J. D. Prades and C. Fàbrega, *ACS Appl. Nano Mater.*, 2021, **4**, 2522–2527.
- 29 A. Dahal, N. G. Petrik, Y. Wu, G. A. Kimmel, F. Gao, Y. Wang and Z. Dohnálek, *J. Phys. Chem. C*, 2019, **123**, 24133–24145.
- 30 K. Kähler, M. C. Holz, M. Rohe, J. Strunk and M. Muhler, *ChemPhysChem*, 2010, **11**, 2521–2529.
- 31 M. A. Bañares, M. O. Guerrero-Pérez, J. L. G. Fierro and G. G. Cortez, *J. Mater. Chem.*, 2002, **12**, 3337–3342.
- 32 K. Föttinger and G. Rupprechter, *Acc. Chem. Res.*, 2014, **47**, 3071–3079.
- 33 G. Rupprechter, *Small*, 2021, **17**, 2004289.
- 34 Q. Maqbool, G. Barucca, S. Sabbatini, M. Parlapiano, M. L. Ruello and F. Tittarelli, *J. Hazard. Mater.*, 2022, **423**, 126958.
- 35 Q. Maqbool, M. Nazar, S. Naz, T. Hussain, N. Jabeen, R. Kausar, S. Anwaar, F. Abbas and T. Jan, *Int. J. Nanomed.*, 2016, **11**, 5015–5025.
- 36 M. N. M. Esther, C. T. Cristina, O. José, S. M. Rosario and G. L. Alonso, *Antioxidants*, 2021, **10**, 1963.
- 37 A. K. Mittal, S. Kumar and U. C. Banerjee, *J. Colloid Interface Sci.*, 2014, **431**, 194–199.
- 38 A. W. Cook and T. W. Hayton, *Acc. Chem. Res.*, 2018, **51**, 2456–2464.
- 39 V. O. Odhiambo, T. Le Ba, Z. Kónya, C. Cserhádi, Z. Erdélyi, M. C. Naomi and I. Miklós Szilágyi, *Mater. Sci. Semicond. Process.*, 2022, **147**, 106699.
- 40 S. Abdolhosseinzadeh, H. Asgharzadeh and H. S. Kim, *Sci. Rep.*, 2015, **5**(1), 1–7.
- 41 H. Huang, J. Lin, G. Zhu, Y. Weng, X. Wang, X. Fu and J. Long, *Angew. Chem., Int. Ed.*, 2016, **55**, 8314–8318.
- 42 Q. Gu, J. Long, L. Fan, L. Chen, L. Zhao, H. Lin and X. Wang, *J. Catal.*, 2013, **303**, 141–155.
- 43 A. Sidorowicz, V. Margarita, G. Fais, A. Pantaleo, A. Manca, A. Concas, P. Rappelli, P. L. Fiori and G. Cao, *PLoS One*, 2022, **17**, e0274753.
- 44 Q. Maqbool, N. Czerwinska, C. Giosue, S. Sabbatini, M. L. Ruello and F. Tittarelli, *J. Cleaner Prod.*, 2022, 133853.



- 45 I. C. Maurya, S. Singh, S. Senapati, P. Srivastava and L. Bahadur, *Sol. Energy*, 2019, **194**, 952–958.
- 46 M. Tsega and F. B. Dejene, *Heliyon*, 2017, **3**, e00246.
- 47 S. L. Isley and R. L. Penn, *J. Phys. Chem. C*, 2008, **112**, 4469–4474.
- 48 S. Singh and S. Sharma, *Sens. Actuators, B*, 2022, **350**, 130798.
- 49 K. Wu, W. Zhang, Z. Zheng, M. Debliquy and C. Zhang, *Appl. Surf. Sci.*, 2022, **585**, 152744.
- 50 A. A. Liu, S. Liu, R. Zhang and Z. Ren, *J. Phys. Chem. C*, 2015, **119**, 23486–23494.
- 51 R. R. Feng, A. A. Liu, S. Liu, J. J. Shi, Y. Liu and Z. F. Ren, *Chin. J. Chem. Phys.*, 2015, **28**, 11.
- 52 C. Y. Wang, H. Groenzin and M. J. Shultz, *J. Phys. Chem. B*, 2004, **108**, 265–272.
- 53 X. Li, G. H. Deng, R. J. Feng, K. Lin, Z. Zhang, Y. Bai, Z. Lu and Y. Guo, *Chin. Chem. Lett.*, 2016, **27**, 535–539.
- 54 M. Manzoli, A. Chiorino and F. Boccuzzi, *Appl. Catal., B*, 2005, **57**, 201–209.
- 55 P. F. Rossi and G. Busca, *Colloids Surf.*, 1985, **16**, 95–102.
- 56 S. Srinives, T. Sarkar and A. Mulchandani, *Electroanalysis*, 2013, **25**, 1439–1445.
- 57 E. Lee, A. Vahidmohammadi, B. C. Prorok, Y. S. Yoon, M. Beidaghi and D. J. Kim, *ACS Appl. Mater. Interfaces*, 2017, **9**, 37184–37190.
- 58 Q. Rong, Y. Zhang, J. Hu, K. Li, H. Wang, M. Chen, T. Lv, Z. Zhu, J. Zhang and Q. Liu, *Sci. Rep.*, 2018, **8**, 1–12.
- 59 L. Xie, Z. Li, L. Sun, B. Dong, Q. Fatima, Z. Wang, Z. Yao and A. A. Haidry, *Front. Mater.*, 2019, **6**, 96.
- 60 M. Wang, Y. Zhu, D. Meng, K. Wang and C. Wang, *Mater. Lett.*, 2020, **277**, 128372.
- 61 S. Wang, L. Pan, J. J. Song, W. Mi, J. J. Zou, L. Wang and X. Zhang, *J. Am. Chem. Soc.*, 2015, **137**, 2975–2983.
- 62 K. Phasuksom, W. Prissanaroon-Ouajai and A. Sirivat, *RSC Adv.*, 2020, **10**, 15206–15220.
- 63 E. Antolini, *Appl. Catal., B*, 2018, **237**, 491–503.
- 64 A. Brückner, *Chem. Soc. Rev.*, 2010, **39**, 4673–4684.

