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# Reactivity of metal–oxo clusters towards biomolecules: from discrete polyoxometalates to metal–organic frameworks

David E. Salazar Marcano,  † Nada D. Savić,  † Kilian Declerck,  † Shorok A. M. Abdelhameed  and Tatjana N. Parac-Vogt  \*

Metal–oxo clusters hold great potential in several fields such as catalysis, materials science, energy storage, medicine, and biotechnology. These nanoclusters of transition metals with oxygen-based ligands have also shown promising reactivity towards several classes of biomolecules, including proteins, nucleic acids, nucleotides, sugars, and lipids. This reactivity can be leveraged to address some of the most pressing challenges we face today, from fighting various diseases, such as cancer and viral infections, to the development of sustainable and environmentally friendly energy sources. For instance, metal–oxo clusters and related materials have been shown to be effective catalysts for biomass conversion into renewable fuels and platform chemicals. Furthermore, their reactivity towards biomolecules has also attracted interest in the development of inorganic drugs and bioanalytical tools. Additionally, the structural versatility of metal–oxo clusters allows for the efficiency and selectivity of the biomolecular reactions they promote to be readily tuned, thereby providing a pathway towards reaction optimization. The properties of the catalyst can also be improved through incorporation into solid supports or by linking metal–oxo clusters together to form Metal–Organic Frameworks (MOFs), which have been demonstrated to be powerful heterogeneous catalysts. Therefore, this review aims to provide a comprehensive and critical analysis of the state of the art on biomolecular transformations promoted by metal–oxo clusters and their applications, with a particular focus on structure–activity relationships.

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## 1. Introduction

Living organisms are essentially highly complex bioreactors in which many biomolecular reactions take place simultaneously at all times to sustain the processes necessary for life. Therefore, the covalent bonds in biomolecules are regularly being broken and formed, typically using enzymes as catalysts.<sup>1–4</sup> For instance, the biological processes involved in the encoding, decoding, replication, and repair of genetic information rely on the formation and cleavage of phosphoester bonds that form the backbone of nucleic acids.<sup>5,6</sup> Furthermore, during translation of this genetic material, the information contained in messenger RNA is used to combine amino acids into proteins *via* peptide bond formation.<sup>7,8</sup> The amino acid building blocks needed for this process can be obtained by the reverse reaction, through the hydrolysis of peptide bonds in ingested or old proteins.<sup>9,10</sup> Moreover, the energy needed for many biological processes to occur, such as transcription and translation, is

obtained through hydrolysis of the phosphoanhydride bond in nucleotides.<sup>11,12</sup> Organisms can also store this energy within polysaccharides, composed of monosaccharides connected by glycosidic bonds, or lipids, such as triglycerides that consist of fatty acids linked to glycerol *via* ester bonds. The glycosidic and ester bonds in carbohydrates and lipids, respectively, can then be broken down to release monosaccharides and fatty acids, which can eventually be converted into energy.<sup>13–15</sup> These are just a few examples of the many biomolecular transformations involving common specific bond types that determine the structural composition of some of the most important biomolecules and their function. Hence, one of the main challenges in (bio)chemistry is being able to reproduce these biomolecular reactions *in vitro*, in order to understand the function, properties, and biochemical transformations of biomolecules. Such knowledge can be further leveraged for the formation of pharmaceuticals, biofuel production, and the synthesis of other industrially relevant products, among many other possible applications.<sup>16–20</sup>

Many biomolecular transformations in living organisms are performed by metalloenzymes, which incorporate metal cofactors within proteins. For instance, most hydrolytic reactions are

Department of Chemistry, KU Leuven, Celestijnenlaan 200F, 3001 Leuven, Belgium.

E-mail: tatjana.vogt@kuleuven.be

† These authors contributed equally.



catalyzed by zinc-containing proteins such as the enzyme carboxypeptidase A, which specifically removes C-terminal amino acid residues from a given peptide chain.<sup>21</sup>



**David E. Salazar Marcano**

*David E. Salazar Marcano is a post-doctoral researcher in the Laboratory of Bioinorganic Chemistry at KU Leuven. After obtaining a master's degree in Chemistry in 2018 from the University of Southampton, he completed his PhD in Chemistry at KU Leuven as an FWO fellow under the supervision of Prof. Tatjana N. Parac-Vogt. He is currently developing new hybrid (bio)materials based on polyoxometalates as well as investigating the reactivity of these nanoclusters towards proteins.*

Furthermore, metalloenzymes can employ single metallic centers or multimetallic clusters depending on the desired reactivity. For example, alkaline phosphatases use a combination of zinc and magnesium centers for dephosphorylation, while the multi-enzyme complex photosystem II contains a unique  $\text{Mn}_4\text{CaO}_5$  metal-oxo cluster that performs a water-splitting reaction during photosynthesis.<sup>22,23</sup> The presence of these metallic clusters and their composition is often key to achieving the desired reactivity. Hence, this has inspired the study of metallic clusters as biomimetic catalysts for a wide range of reactions.

Metal-oxo clusters (MOCs), which are nanostructures composed of multiple metal centers linked *via* bridging oxygen atoms, have attracted attention for their use as catalysts with promising enzyme-like activity in a wide array of reactions.<sup>24</sup> MOCs can be regarded as discrete units of metal oxides with similar structural and electronic properties, making them interesting model systems for metal oxide nanoparticles and surfaces, especially since they are typically relatively stable in solution as single molecules, and are, therefore, easier to study



**Nada D. Savić**

*Nada D. Savić is an FWO senior researcher in the Laboratory of Bioinorganic Chemistry at KU Leuven. After obtaining her PhD in Serbia in 2019, she moved to the group of Prof. Tatjana N. Parac-Vogt in Belgium. Her research interest is focused on creating regioselective and tunable synthetic proteases based on metal-substituted polyoxometalates and metal-oxo clusters for the hydrolysis of amphiphilic, insoluble and biologically related proteins.*



**Kilian Declerck**

*Kilian Declerck received his master's degree in Biochemistry & Biotechnology from KU Leuven in 2022. He is currently a PhD student in the Laboratory of Bioinorganic Chemistry, working on the development of discrete metal-oxo clusters and metal-organic layers as artificial nanozymes for catalysis of biochemical transformations, under the supervision of Prof. Tatjana N. Parac-Vogt.*



**Shorok A. M. Abdelhameed**

*Shorok A. M. Abdelhameed obtained her master's degree in Chemistry from KU Leuven. She then completed her PhD as an FWO fellow at KU Leuven in the Laboratory of Bioinorganic Chemistry under the supervision of Prof. Tatjana N. Parac-Vogt in 2022. Her PhD research centered around exploring the redox reactions of metal-substituted polyoxometalates towards proteins.*



**Tatjana N. Parac-Vogt**

*Tatjana N. Parac-Vogt is a full Professor of Chemistry and head of the Laboratory of Bioinorganic Chemistry at KU Leuven. She leads an interdisciplinary team performing research at the interface of inorganic chemistry, biochemistry, materials science, and catalysis. She is the recipient of the IUPAC 2023 Distinguished Women in Chemistry/Chemical Engineering award. Tatjana is also a Fellow of the Royal Society of Chemistry and has been elected as a Chemistry Europe Fellow (Class 2020/2021), the highest award given by an association of European chemical societies.*



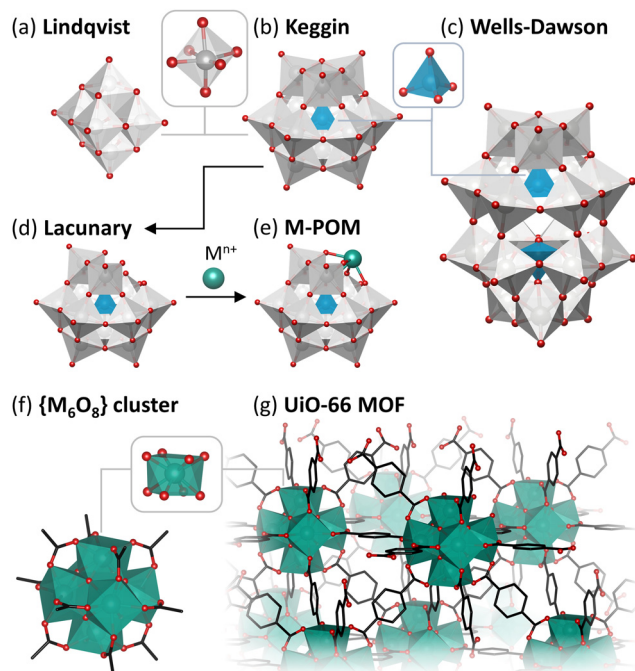
than insoluble and relatively complex  $\text{MO}_x$  solid materials.<sup>25–27</sup> Moreover, MOCs display significant structural versatility, which can be used to tune their catalytic properties. MOCs are typically categorized based on their charge as these clusters can be anionic, neutral, or cationic depending on their structure and composition.

Anionic MOCs are commonly known as polyoxometalates (POMs), which are typically formed by group V and VI transition metals in their highest oxidation states. POMs can take a wide variety of structures with different shapes and sizes that have distinct chemical and physical properties. Moreover, almost any other element in the periodic table can be incorporated into POM structures allowing for additional versatility in their properties. These anionic nanoclusters are primarily composed of metal centers (typically  $\text{V}^{5+}$ ,  $\text{Mo}^{6+}$ , or  $\text{W}^{6+}$ ) in octahedral coordination that are bound to terminal and bridging oxo ligands (formally  $\text{O}^{2-}$ ). Some of the most commonly explored POMs, especially for catalytic applications, are the Lindqvist ( $[\text{A}_6\text{O}_{19}]^{n-}$ ), Keggin ( $[\text{EA}_{12}\text{O}_{40}]^{n-}$ ), and Wells–Dawson ( $[\text{E}_2\text{A}_{18}\text{O}_{62}]^{n-}$ ) structures ( $\text{E} = \text{P}, \text{Si}, \text{etc.}$ ;  $\text{A} =$  typically  $\text{V}, \text{Mo}$ , or  $\text{W}$ ) shown in Fig. 1. Other transition metals can also be incorporated into POM structures *via* replacement of some of the metal centers and their corresponding oxo ligands to form metal-substituted POMs (M–POMs) *via* lacunary POMs (Fig. 1(d) and (e)). Due to this structural versatility, POMs have been developed as catalysts for a vast range of reactions such

as hydrogen evolution, water oxidation, and photoreduction of  $\text{CO}_2$ , which are key in developing sustainable solutions to climate change.<sup>28</sup> They have also been shown to be particularly suitable as acid catalysts towards a diverse set of molecules, by taking advantage of their Brønsted acidity or by exploiting the Lewis acidity of the metal centers in the POM structure.<sup>29</sup> Moreover, in recent years, the reactivity of POMs towards biomolecules has been increasingly investigated, demonstrating their ability to cleave many biologically relevant bonds such as phosphoester, peptide, phosphoanhydride, and glycosidic bonds.

Cationic MOCs are typically stabilized by organic or inorganic ligands, resulting in neutral structures that can contain a wide range of metal centers.<sup>25,30</sup> Such clusters are mainly reported for group IV metals ( $\text{Ti}, \text{Zr}, \text{Hf}$ ), which form a wide variety of structures that are highly Lewis acidic as well as less toxic and/or more stable than clusters of other transition metals (*e.g.*,  $\text{Pt}, \text{Pd}, \text{Ln}, \text{Fe}, \text{Cr}$ , *etc.*).<sup>31–36</sup> Hence, group IV clusters have been explored to a greater extent toward biologically relevant reactions. Unlike POMs,  $\text{Zr}$  and  $\text{Hf}$  centers in these clusters are typically 7- or 8-coordinated, largely resulting in tetragonal or cubic coordination geometries. These MOCs can drastically vary in size, ranging from the simplest  $\text{Zr}_4\text{O}_2(\text{OOMc})_{12}$  cluster to the much larger  $\text{Zr}_{70}(\text{SO}_4)_{58}(\text{O}/\text{OH})_{146}n(\text{H}_2\text{O})$  cluster.<sup>37,38</sup> Moreover, MOCs incorporating group IV metals have been extensively investigated as building units of Metal–Organic Frameworks (MOFs).<sup>39–41</sup> These MOFs are typically composed of polydentate carboxylate ligands connecting hexanuclear clusters,  $\{\text{M}_6\text{O}_8\}$ , in which each metal center is interconnected *via* four bridging oxo ligands leaving four coordination sites available for binding to other ligands (Fig. 1(f) and (g)).<sup>42</sup> As a result, MOCs have been mainly explored as building units for MOFs in many applications, such as catalysis, sensing, and imaging, while discrete neutral or cationic MOCs have been studied to a lesser extent.<sup>43–46</sup> Similarly to POMs, MOC-based materials have also been investigated as catalysts for promoting biomolecular transformations, with the notable difference of catalyzing these reactions heterogeneously and often with improved efficiency compared to other nanozymes. While a majority of reports on MOC-based materials involve MOFs, other MOC-based hybrid materials have also been investigated, such as POMs immobilized within MOFs or other scaffolds,<sup>47</sup> offering unique opportunities for the development of new heterogeneous catalysts for biomolecular transformations.<sup>48,49</sup>

Despite the significant progress that has been made in the development of metal–oxo clusters as catalysts for a wide range of reactions,<sup>29,45,50–52</sup> their reactivity towards biomolecules and the underlying molecular mechanisms have not been comprehensively reviewed in recent years. Yet, achieving such a mechanistic understanding is crucial due to the important role of the catalytic properties of MOCs in their biological activity as potential inorganic drugs, and in other biomedical applications, as described in multiple reviews.<sup>46,53–57</sup> The reactivity of POMs and MOFs have been recently reviewed by our group, mainly focusing on proteins as substrates.<sup>24,58,59</sup> However, a



**Fig. 1** Mixed ball-and-stick and polyhedral representation of (a) Lindqvist, (b) Keggin, (c) Wells–Dawson, (d) lacunary Keggin, and (e) M–POM Keggin structures as well as of (f) a discrete  $\{\text{M}_6\text{O}_8\}$  cluster capped with carboxylate ligands and (g) the UiO-66 MOF composed of  $\{\text{M}_6\text{O}_8\}$  clusters linked by bidentate carboxylate linkers. POM metal centers in grey, M–POM embedded metal centers and  $\text{M}_6$  cluster metal centers in teal, heteroatom in blue, oxygen in red, and carbon in black.





broader discussion of the role of MOCs and MOC-based materials as catalysts in biochemical transformations involving a wide range of biomolecules – such as nucleic acids, nucleotides, sugars, and lipids – is sorely lacking. This is particularly important as there is significant untapped potential regarding their reactivity with other biomolecules that has not been explored, which entails promising avenues for future research. Furthermore, the study of the catalytic activity of MOCs towards specific biomolecules could benefit from the insights gained from other biological systems. Therefore, this review discusses the reactivity of discrete MOCs, largely focusing on clusters formed by group IV to VI transition metals since they have been most extensively investigated. This is followed by a discussion on MOC-based materials, which mainly act as heterogeneous catalysts. For both classes of catalysts, subdivisions are made based on the type of reactivity they exhibit (*e.g.*, hydrolytic, oxidative, *etc.*) and the type of biomolecule involved in the catalytic reaction (*e.g.*, peptides, proteins, nucleic acids, carbohydrates, lipids, *etc.*). In all sections, the biomolecular reactivity of MOCs is discussed in the context of their potential medical, bio-analytical, and environmental applications. Furthermore, particular focus is given to the underlying mechanisms, the structure–activity relationships, and the structural properties of MOCs that can be tuned as a means of optimization towards specific future applications.

## 2. Reactivity of discrete metal–oxo clusters

### 2.1. Amino acids, peptides, and proteins

The peptide bond, which connects amino acid residues into complex three-dimensional functional structures to form proteins, is highly stable, the hydrolysis half-life under physiological conditions being estimated at *ca.* 600 years.<sup>60</sup> Therefore, cleaving peptide bonds is an extremely challenging task. Yet, the selective cleavage of peptide bonds is essential in many biological and biomedical studies, particularly in the context of proteomics, which is the large-scale study of the structure, function, and localization of proteins.<sup>61–63</sup> Furthermore, peptide bond hydrolysis is also important in determining enzymatic active sites,<sup>64</sup> mapping metal and ligand binding sites,<sup>65</sup> footprinting,<sup>66</sup> designing therapeutic drugs,<sup>67</sup> and in studying protein folding.<sup>68</sup> Natural enzymes, known as peptidases or proteases, are commonly used to catalyze peptide bond cleavage, with trypsin being the most frequently used. However, these enzymes suffer from several shortcomings as their catalytic activity is mainly preserved under very narrow experimental conditions (temperature and pH)<sup>69</sup> and trypsin especially tends to produce many short peptide fragments, limiting the accurate analysis of the proteins' structural composition.<sup>70</sup> Natural proteases are also prone to self-digestion, thereby contaminating the analyte, resulting in loss of structural information about the protein under study. Considering the importance of peptide bond cleavage, the development of new metal-based catalysts with peptidase-like

properties has attracted significant interest.<sup>71–74</sup> In this respect, MOCs, and particularly POMs, have been shown to be a promising class of compounds for the hydrolysis of peptide bonds in a wide range of different substrates.

**2.1.1. Hydrolysis of peptides.** Studies on protein hydrolysis are often limited to assessing the activity and selectivity of the catalyst since proteins are generally too large and complex to obtain accurate information at a molecular level. Therefore, to investigate the reactivity of MOCs towards peptide bonds, oligopeptides have been used as simpler targets that serve as good model systems.

Initially, molybdate and vanadate oxo-clusters were investigated as catalysts for the hydrolysis of several dipeptides.<sup>75,76</sup> Peptide bond cleavage was determined to be purely hydrolytic in nature since neither the formation of paramagnetic species nor oxidative modifications on the resulting amino acid products were observed. The hydrolysis of dipeptides was investigated under various experimental conditions since the speciation of molybdates and vanadates depends on the concentration, pH, temperature, and ionic strength of the solution.<sup>77</sup> These studies indicated that the monomeric orthovanadate ( $[\text{VO}_4]^{3-}$ ) and orthomolybdate ( $[\text{MoO}_4]^{2-}$ ) species were the most active in promoting peptide bond hydrolysis (Fig. 2). Furthermore, increasing the ionic strength of the solution increased the rate of hydrolysis in the presence of  $[\text{MoO}_4]^{2-}$ , but the opposite effect was observed when using  $[\text{VO}_4]^{3-}$ . This observation was linked to the different effects of the ionic strength on the speciation of molybdates and vanadates as well as on their interactions with dipeptides. In addition, a detailed mechanistic study revealed that dipeptides bind to  $[\text{VO}_4]^{3-}$  *via* bidentate coordination of the terminal amino nitrogen and the amide carbonyl oxygen, while tridentate coordination was proposed for  $[\text{MoO}_4]^{2-}$  *via* the terminal amino nitrogen, amide oxygen, and carboxylate oxygen (Fig. 2). In both cases, coordination-induced polarization of the peptide carbonyl bond activated it towards nucleophilic attack, but the tridentate coordination of the dipeptide to  $[\text{MoO}_4]^{2-}$  gave rise to better reactivity.<sup>76</sup> The hydrolytic activity is also strongly dependent on the nature of the amino acid side chains of the dipeptides (Table 1) and the greatest reactivity was observed for dipeptides

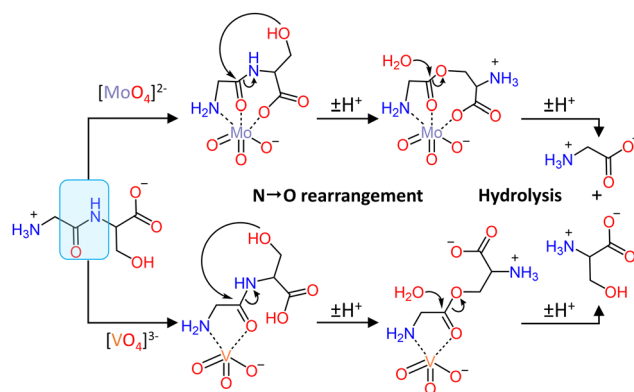


Fig. 2 Proposed mechanism of the hydrolysis of Gly-Ser catalyzed by  $[\text{MoO}_4]^{2-}$  (top) or  $[\text{VO}_4]^{3-}$  (bottom) *via* an *N,O*-acyl rearrangement.



**Table 1** Selected conditions, percentage conversion, and rate constants ( $k_{\text{obs}}$ ) reported for the hydrolysis of peptides promoted by different (poly)oxometalates catalysts (cat.)

| Catalyst                          | Substrate  | Reaction conditions                 | Conversion/%            | $k_{\text{obs}}/\times 10^{-6} \text{ s}^{-1}$ | Ref.  |
|-----------------------------------|--|-------------------------------------|-------------------------|--|-------|
| [MoO <sub>4</sub> ] <sup>2-</sup> | Gly-X<br>(X = Ser, Thr, Ala, Tyr, Gly, Asn, Met)   | pD 7, 60 °C, 60 h, 60 eq. cat.      | 2 (Met)–68 (Ser)        | 5.9 (X = Ser)                                  | 75    |
|                                   | X-Gly<br>(X = Ser, Cys, Asp, Lys)  | pD 5–7, 60 °C, 60 h, 60 eq. cat.    | 2 (Lys)–17 (Cys)        | 0.84 (Asp; pD 7)<br>2.34 (Asp; pD 5)           | 75,78 |
|                                   | His-Ser  | pD 7, 60 °C, 60 h, 60 eq. cat.      | 91                      |  | 75    |
|                                   | Leu-Ser  | pD 7, 60 °C, 60 h, 60 eq. cat.      | 43                      |  | 75    |
|                                   | Asp-Ala  | pD 7, 60 °C, 60 h, 60 eq. cat.      | 3                       |  | 75    |
|                                   | Ala-His  | pD 7, 60 °C, 60 h, 60 eq. cat.      | 5                       |  | 75    |
|                                   | Gly-Ser-Phe  | pD 7, 60 °C, 60 h, 60 eq. cat.      | 25                      |  | 75    |
|                                   | Glu-Cys-Gly  | pD 7, 60 °C, 60 h, 60 eq. cat.      | 4                       |  | 75    |
|                                   | Gly-X<br>(X = Ser, Thr, Ala)   | pD 4.4, 60 °C, 140 h, 12.5 eq. cat. | 5 (Ser)–15 (Thr)        |  | 76    |
|                                   |  | pD 7.4, 60 °C, 140 h, 12.5 eq. cat. | 7 (Ala)–52 (Ser)        | 0.07 (Ala)–1.3 (Ser)                           |       |
| [VO <sub>4</sub> ] <sup>3-</sup>  |  | pD 7.4, 37 °C, 140 h, 12.5 eq. cat. |                         | 0.09 (Ser)                                     |       |
|                                   | Ser-Gly  | pD 4.4–7.4, 60 °C, 140 h, 12.5 eq.  | 3 (pD 7.4)–6 (pD 4.4)   | 0.05 (pD 7.4)                                  | 76    |
|                                   | His-Ser  | pD 4.4–7.4, 60 °C, 140 h, 12.5 eq.  | 25 (pD 4.4)–40 (pD 7.4) | 1.2  | 76    |
|                                   | Leu-Ser  | pD 4.4–7.4, 60 °C, 140 h, 12.5 eq.  | 12 (pD 4.4)–39 (pD 7.4) | 1.0  | 76    |
|                                   | Gly-Ser-Phe  | pD 4.4, 60 °C, 140 h, 12.5 eq. cat. | 4 (pD 4.4)–16 (pD 7.4)  | 0.3 (pD 7.4)                                   | 76    |
|                                   |  | pD 5, 60 °C, 1 eq. cat.             |                         | 0.08 (Glu)–3.5 (Ser)                           | 79,80 |
|                                   | Gly-X<br>(X = Gly, Ala, Val, Leu, Ile, Phe, Asp, Asn, Glu, Gln, Ser, Thr, Tyr, Lys, Arg, His)      |                                     |                         |  |       |
|                                   | X-Gly<br>(X = Ala, His, Ser)   | pD 5, 60 °C, 1 eq. cat.             |                         | 0.34 (Ser)–0.5 (Ala)                           | 79,80 |
|                                   | X-Ser<br>(X = Leu, Ile)  | pD 5, 60 °C, 1 eq. cat.             |                         | 0.35 (Ile)<br>3.53 (Leu)                       | 79,80 |
|                                   | Gly-X<br>(X = Gly, Ala, Val, Leu, Ile, Phe, Ser, Thr, His, Lys, Arg, Asp, Asn, Gln)                | pD 7.4, 60 °C, 1 eq. cat.           |                         | 0.3 (Gly)                                      | 81    |
| Zr-WD 1 : 2                       |  | pD 7.4, 60 °C, 6 eq. cat.           |                         | 0.01 (Ile)–2.3 (Ser)                           |       |
|                                   |  | pD 5.4, 60 °C, 6 eq. cat.           |                         | 0.4 (Gly)                                      |       |
|                                   | Gly-Gly-Gly  | pD 7.4, 60 °C, 4 weeks, 6 eq. cat.  | 45% Gly                 |  | 82    |
|                                   | Gly-Gly-Gly-Gly  | pD 7.4, 60 °C, 4 weeks, 6 eq. cat.  | 45% Gly                 |  | 82    |
|                                   | Gly-Ser-Phe  | pD 7.4, 60 °C, 10 days, 6 eq. cat.  | 31% Gly                 |  | 82    |
|                                   | Gly-Ser  | pD 5.4, 60 °C, 1 eq. cat.           |                         | 0.3  | 83    |
|                                   | Gly-X<br>(X = Gly, Ser, Thr, Tyr, Asp, Met, Glu, Asn, Gln, His, Lys, Arg, Ala, Val, Leu, Ile, Phe) | pD 5.4, 60 °C, 1 eq. cat.           |                         | 0.08 (Ile)–6.3 (Ser)                           | 83,84 |
|                                   | X-Gly<br>(X = Ser, Cys, Asp, His)  | pD 5.4, 60 °C, 1 eq. cat.           |                         | 0.07 (Asp)–0.8 (Ser)                           | 84    |
|                                   | X-Ser<br>(X = Leu, Ile, Tyr)   | pD 5.4, 60 °C, 1 eq. cat.           |                         | 0.5 (Tyr)–4.5 (Leu)                            | 84    |
|                                   | Gly-Gly-Gly  | pD 5.4, 60 °C, 4 weeks, 2 eq. cat.  | 65% Gly                 | 0.7  | 82    |
| Zr-WD 4 : 2                       | Gly-Gly-Gly-Gly  | pD 7.4, 60 °C, 4 weeks, 2 eq. cat.  | 55% Gly                 | 0.7  | 82    |
|                                   | Gly-Ser-Phe  | pD 7.4, 60 °C, 10 days, 2 eq. cat.  | 55% Gly                 | 1.0  | 82    |
|                                   | Gly-X<br>(X = Ser, Gly, Thr)   | pD 7.4, 60 °C, 1 eq. cat.           |                         | 0.8 (Ser)–0.03 (Gly)                           | 85    |
|                                   | X-Gly<br>(X = Cys, His, Ser)   | pD 7.4, 60 °C, 1 eq. cat.           |                         | 0.03   | 85    |
|                                   | His-Ser  | pD 7.4, 60 °C, 1 eq. cat.           |                         | 1  | 85    |
|                                   | Leu-Ser  | pD 7.4, 60 °C, 1 eq. cat.           |                         | 0.5  | 85    |
|                                   | Gly-X<br>(X = Ser, Gly, Thr, Ala, Leu, Ile, Tyr)   | pD 7.4, 60 °C, 1 eq. cat.           |                         | 0.02 (Ile)–1.5 (Ser)                           | 86    |
|                                   | Ser-Gly  | pD 7.4, 60 °C, 1 eq. cat.           |                         | 0.1  | 86    |
|                                   | His-Gly  | pD 7.4, 60 °C, 1 eq. cat.           |                         | 0.5  | 86    |
|                                   | His-Ser  | pD 7.4, 60 °C, 1 eq. cat.           |                         | 9.5  | 86    |
| Zr-K 1 : 2                        | Leu-Ser  | pD 7.4, 60 °C, 1 eq. cat.           |                         | 1.3  | 86    |
|                                   | Gly-Gly-Gly  | pD 5.4, 60 °C, 4 weeks, 2 eq. cat.  | 30% Gly                 |  | 82    |
|                                   | Gly-Gly-Gly-Gly  | pD 5.4, 60 °C, 4 weeks, 2 eq. cat.  | 30% Gly                 |  | 82    |
|                                   | Gly-Ser-Phe  | pD 5.4, 60 °C, 2 eq. cat.           | 20% Gly                 |  | 82    |
|                                   | Gly-X<br>(X = Ser, Gly, Thr)   |                                     |                         |  |       |
|                                   | X-Gly<br>(X = Cys, His, Ser)   |                                     |                         |  |       |
|                                   | His-Ser  |                                     |                         |  |       |
|                                   | Leu-Ser  |                                     |                         |  |       |
|                                   | Gly-X<br>(X = Ser, Gly, Thr, Ala, Leu, Ile, Tyr)   |                                     |                         |  |       |
|                                   | Ser-Gly  |                                     |                         |  |       |
| Ce-K 1 : 2                        | His-Gly  |                                     |                         |  |       |
|                                   | His-Ser  |                                     |                         |  |       |
|                                   | Leu-Ser  |                                     |                         |  |       |
|                                   | Gly-Gly-Gly  |                                     |                         |  |       |
|                                   | Gly-Gly-Gly-Gly  |                                     |                         |  |       |
|                                   | Gly-Ser-Phe  |                                     |                         |  |       |
|                                   | Gly-X<br>(X = Ser, Gly, Thr)   |                                     |                         |  |       |
|                                   | X-Gly<br>(X = Cys, His, Ser)   |                                     |                         |  |       |
|                                   | His-Ser  |                                     |                         |  |       |
|                                   | Leu-Ser  |                                     |                         |  |       |
| Zr-L 1 : 1                        | Gly-X<br>(X = Ser, Gly, Thr, Ala, Leu, Ile, Tyr)   |                                     |                         |  |       |
|                                   | Ser-Gly  |                                     |                         |  |       |
|                                   | His-Gly  |                                     |                         |  |       |
|                                   | His-Ser  |                                     |                         |  |       |
|                                   | Leu-Ser  |                                     |                         |  |       |
|                                   | Gly-Gly-Gly  |                                     |                         |  |       |
|                                   | Gly-Gly-Gly-Gly  |                                     |                         |  |       |
|                                   | Gly-Ser-Phe  |                                     |                         |  |       |
|                                   | Gly-X<br>(X = Ser, Gly, Thr)   |                                     |                         |  |       |
|                                   | X-Gly<br>(X = Cys, His, Ser)   |                                     |                         |  |       |

with a serine residue downstream of the peptide bond (X-Ser where X = an amino acid residue). This is due to the intramolecular nucleophilic attack of the hydroxyl group of serine on the peptide bond, which leads to an *N,O*-acyl rearrangement and the formation of a more hydrolytically labile ester bond (Fig. 2).<sup>75,76</sup> Therefore, hydrolysis of dipeptides by vanadates and molybdates depends on several factors: (i) the speciation, (ii) the nature of the metal and its binding to the dipeptide, and

(iii) the amino acid sequence. However, the kinetic lability of molybdate and vanadate oxo-clusters in solution limits their broader applicability and complicates their investigation in catalytic reactions.

Further research into the hydrolytic activity of POMs shifted towards polyoxotungstate clusters since they are less labile compared to vanadates and molybdates in solution. However, the lower lability of tungsten centers also hinders their ability



to effectively coordinate to dipeptides. This problem can be overcome by using M-POMs that contain other transition metal centers incorporated into the POM structure (Fig. 1(e)). Therefore, Wells–Dawson polyoxotungstate M-POMs with different embedded metals ( $M = \text{Mn}^{3+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Co}^{2+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Y}^{3+}$ ,  $\text{Eu}^{3+}$ ,  $\text{Yb}^{3+}$ ,  $\text{Ce}^{4+}$ ,  $\text{Zr}^{4+}$ , and  $\text{Hf}^{4+}$ ) were investigated toward peptide bond hydrolysis of several dipeptides (Ala-Gly and Gly-X where  $X = \text{Gly}$ , Ala, Val, Leu, Ile, Phe, Ser, Thr, Tyr, Glu, Gln, Asn, Asp, His, Lys, Arg, Met, and Cys).<sup>79,80</sup> The  $\text{Zr}^{4+}$ - and  $\text{Hf}^{4+}$ -substituted Wells–Dawson M-POMs were shown to be considerably more catalytically active towards peptide bond hydrolysis in comparison to all other M-POMs (Table 1) due to the high Lewis acidity of  $\text{Zr}^{4+}$  and  $\text{Hf}^{4+}$ , as well as their high coordination number, redox inactivity, and oxophilicity.<sup>80</sup> The importance of Lewis acidic metal ions in the hydrolytic process was confirmed by the addition of different carboxylic acid inhibitors, which competitively bind to the embedded metal center and, therefore, decreased the rate of peptide bond hydrolysis. The POM framework also plays an essential role since it prevents the formation of insoluble gels, which typically occurs when using  $\text{Zr}^{4+}$  and  $\text{Hf}^{4+}$  salts.<sup>70,80</sup> Hence, polyoxotungstate M-POMs are promising catalysts for peptide bond hydrolysis that exploit the high Lewis acidity of the embedded metal center and the aqueous stability of the POM framework as an inorganic ligand.

$\text{Zr}^{4+}$  and  $\text{Hf}^{4+}$  M-POMs are often present in solution as dimers (Fig. 3(a)) where two POM units bind to one metal center (e.g.,  $[\text{M}(\alpha_2\text{-P}_2\text{W}_{17}\text{O}_{61})_2]^{16-}$ ; M-WD 1:2) or two M-POMs are linked *via*  $\mu\text{-OH}$  ligands (e.g.,  $[\{\text{M}(\mu\text{-OH})(\text{H}_2\text{O})(\alpha_2\text{-P}_2\text{W}_{17}\text{O}_{61})\}_2]^{16-}$ ; M-WD 2:2). In these dimers the embedded metal center in the dimer with a 1:2 metal:POM ratio has no free coordination sites, while each metal center in the 2:2 dimer only has one free coordination, which is typically occupied by a labile water ligand that can be exchanged with the incoming substrate. However, the dimeric POMs are typically in equilibrium with a monomeric POM species ( $[\text{M}(\alpha_2\text{-P}_2\text{W}_{17}\text{O}_{61})]^{6-}$ ; M-WD 1:1), which forms upon dissociation of the dimers (Fig. 3(a)). This monomeric species is believed to be the catalytically active species since it has four free coordination sites on  $\text{Zr}^{4+}/\text{Hf}^{4+}$ .<sup>80</sup> Interestingly, the dimeric tetrazirconium-substituted Wells–Dawson M-POM ( $[\text{Zr}_4(\mu_3\text{-O})_2(\text{OH})_2(\text{H}_2\text{O})_4(\text{P}_2\text{W}_{16}\text{O}_{59})_2]^{14-}$ ; Zr-WD 4:2), composed of two Wells–Dawson POM units linked by a  $\{\text{Zr}_4(\mu_3\text{-O})_2(\text{OH})_2(\text{H}_2\text{O})_4\}^{10+}$  cluster (Fig. 3(b)), was also shown to have considerable activity towards Gly-X dipeptides (Table 1) although its dissociation into a monomer has not been reported.<sup>81</sup> This is likely due to the ability of dipeptides to coordinate in a

bidentate fashion to  $\text{Zr}^{4+}$  centers bearing two water ligands,  $\{\text{Zr}(\text{H}_2\text{O})_2\}$ , by displacing those labile waters without affecting the overall structure of the POM.

The proposed mechanism of peptide bond hydrolysis promoted by M-POMs involves coordination of the terminal amine nitrogen and the peptide carbonyl oxygen to a metal center, resulting in activation of the carbonyl carbon towards nucleophilic attack by solvent water.<sup>80,87</sup> Furthermore, it was found that the uncoordinated terminal carboxylic acid has an important role in the hydrolytic process since it acts as a general base that can remove a proton from the attacking water nucleophile.<sup>81,87</sup> Hence, although M-POMs have a tendency to form dimeric structures, for them to be catalytically active there needs to be several free coordination sites that allow for bidentate binding of the dipeptide, which may be achieved through dissociation of the dimer in solution.

The hydrolytic activity of Lindqvist and Keggin Zr-POMs towards dipeptides has also been investigated. The Keggin Zr-POMs were shown to effectively catalyze the hydrolysis of glycylserine (Gly-Ser) and glycylglycine (Gly-Gly) dipeptides to produce single amino acids as the main products (Fig. 4(a)), along with small amounts of cyclic dipeptide side-products (cGly-Ser and cGly-Gly).<sup>83</sup> The cyclization reaction was shown to be reversible as the Zr-POMs hydrolyzed cGly-Ser and cGly-Gly back into the dipeptides, which were further hydrolyzed to give amino acid products (Fig. 4(a)). Comparison of two dimeric Keggin M-POMs (Table 1), showed that the rate constant for the hydrolysis of Gly-Ser was 20 times higher for  $[\{\text{Zr}(\mu\text{-OH})(\text{H}_2\text{O})(\alpha\text{-PW}_{11}\text{O}_{39})_2\}^{8-}]$  (Zr-K 2:2) compared to  $[\text{Zr}(\alpha\text{-PW}_{11}\text{O}_{39})_2]^{10-}$  (Zr-K 1:2). This demonstrates the importance of the  $\text{Zr}^{4+}$  centers for the hydrolysis reaction, since dissociation of Zr-K 2:2 gives two hydrolytically active monomeric POMs ( $[\text{Zr}(\alpha\text{-PW}_{11}\text{O}_{39})]^{3-}$ ; Zr-K 1:1) while only one Zr-K 1:1 is formed from dissociation of Zr-K 1:2 together with a catalytically inactive lacunary POM.<sup>83</sup> Furthermore, Zr-K 1:1 was confirmed to be the catalytically active species since binding to Zr-K 2:2, which could in principle occur *via* replacement of coordinated  $\text{H}_2\text{O}$ , was found to be thermodynamically unfavorable by Density Functional Theory (DFT) calculations.<sup>87</sup> However, even though the formation of Zr-K 1:1 is favored at lower pH, the rate constant of Gly-Gly hydrolysis catalyzed by Zr-K 2:2 was observed to vary with the pH following a bell-shaped profile reaching a maximum at around pH 5.5. This is because, at higher pH, Zr-K 2:2 was converted to Zr-K 1:2 or formed an inactive complex with the dipeptide, while at lower pH protonation of the amine nitrogen hindered effective peptide

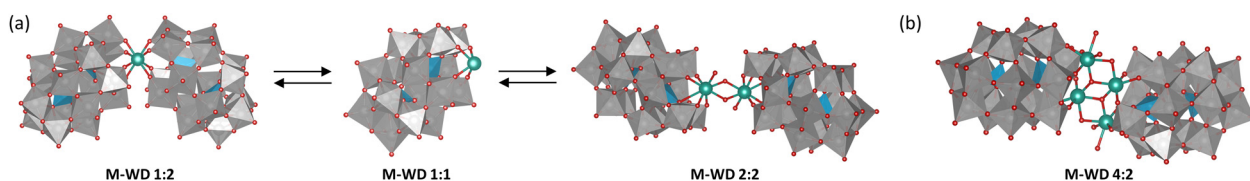


Fig. 3 (a) Equilibrium between the dimeric  $[\text{M}(\alpha_2\text{-P}_2\text{W}_{17}\text{O}_{61})_2]^{16-}$  (M-WD 1:2), the monomeric  $[\text{M}(\alpha_2\text{-P}_2\text{W}_{17}\text{O}_{61})]^{6-}$  (M-WD 1:1), and the dimeric  $[\{\text{M}(\mu\text{-OH})(\text{H}_2\text{O})(\alpha_2\text{-P}_2\text{W}_{17}\text{O}_{61})\}_2]^{16-}$  (M-WD 2:2) Wells–Dawson M-POMs ( $M = \text{Zr}^{4+}$  or  $\text{Hf}^{4+}$ ). (b) Structure of the dimeric tetrazirconium-substituted Wells–Dawson M-POM ( $[\text{Zr}_4(\mu_3\text{-O})_2(\text{OH})_2(\text{H}_2\text{O})_4(\text{P}_2\text{W}_{16}\text{O}_{59})_2]^{14-}$ ; Zr-WD 4:2). Tungsten in gray, oxygen in red, phosphorus in blue, zirconium/hafnium in teal.



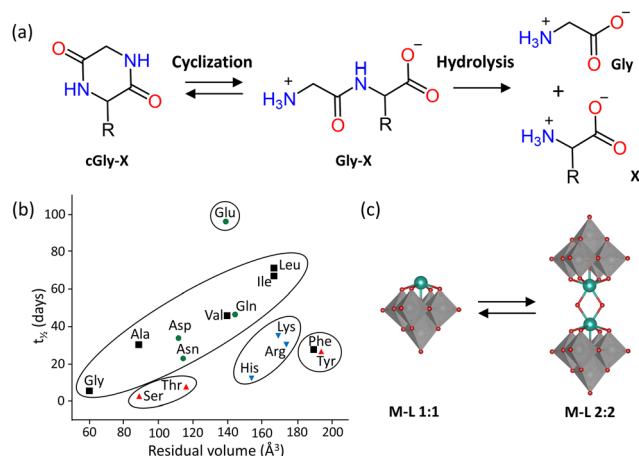


Fig. 4 (a) Hydrolysis and cyclization of a Gly-X dipeptide. (b) Plot of the half-life ( $t_{1/2}$ ) of the hydrolysis of Gly-X dipeptides catalyzed by Zr-WD 1:2 with respect to the residue volume of amino acid X with different side chains containing: aliphatic (■), hydroxyl (▲), carbonyl (●), or charged (▼) groups. Adapted from ref. 79 with permission from the Royal Society of Chemistry. (c) Equilibrium between the monomeric [W<sub>5</sub>O<sub>18</sub>Zr(H<sub>2</sub>O)<sub>3</sub>]<sup>2-</sup> (Zr-L 1:1) and the dimeric [(Zr(μ-OH)(W<sub>5</sub>O<sub>18</sub>))<sub>2</sub>]<sup>6-</sup> (Zr-L 2:2) Lindqvist M-POMs.

coordination.<sup>83</sup> Increasing the concentration of Zr-K 2:2 also resulted in a decrease in the rate of Gly-Gly hydrolysis due to the conversion of Zr-K 2:2 into Zr-K 1:2. Therefore, while the speciation of polyoxotungstates is less complex in comparison to molybdates and vanadates, the equilibria between monomeric and dimeric species, which is affected by the solution conditions, still plays an important role in their reactivity and needs to be carefully considered.

In order to circumvent the equilibrium between dimeric and monomeric species observed for the Wells-Dawson and Keggin Zr-POMs, the hydrolytic activity of the monomeric Lindqvist Zr-POM (Fig. 4(c)), [W<sub>5</sub>O<sub>18</sub>Zr(H<sub>2</sub>O)<sub>3</sub>]<sup>2-</sup> (Zr-L 1:1), was also investigated towards dipeptides, since this M-POM was reported to be more stable as a monomer in solution.<sup>86</sup> Zr-L 1:1 displayed good hydrolytic activity towards a range of dipeptides (Table 1), with the fastest reaction rate observed for the histidylserine dipeptide (His-Ser). Interestingly, the pH dependence of the hydrolysis of His-Ser by Zr-L 1:1 gave a similar bell-shaped curve to that of Zr-K 2:2, but with a maximum at pH 7.5. The lower rate at higher pH was attributed to the formation of the dimeric form (Fig. 4(c)): [(Zr(μ-OH)(W<sub>5</sub>O<sub>18</sub>))<sub>2</sub>]<sup>6-</sup> (Zr-L 2:2). Furthermore, the role of Zr-L 1:1 as a catalyst was demonstrated through the successful hydrolysis of His-Ser added after completion of the initial His-Ser hydrolysis, which could be repeated for three consecutive cycles, but its overall hydrolytic activity was not significantly higher compared to the Keggin Zr-POMs.

In accordance with the results obtained for the molybdate and vanadate oxoanions, the hydrolytic activity of M-POMs was found to depend on the side chain of the amino acid residues in the hydrolyzed dipeptides (Fig. 4(b) and Table 1).<sup>79–81,84</sup> Similarly, hydrolysis was observed to be fastest for X-Ser

dipeptides due to the *N,O*-acyl rearrangement discussed above.<sup>79,84,86,88</sup> The rate of hydrolysis was also faster for dipeptides with positively charged amino acid side chains due to their favorable electrostatic interactions with negatively charged M-POMs.<sup>84</sup> For peptides with aliphatic residues, the reaction rate decreased with increasing molecular volume of the side chains, due to steric effects.<sup>79,84,86,87</sup> Additionally, out of a series of Gly-X dipeptides, the glycine dipeptide containing glutamate (Gly-Glu) gave the lowest rate constant for peptide bond hydrolysis catalyzed by Zr-WD 2:2, which was proposed to be due to the competing coordination of the carboxylate group of Glu to Zr<sup>4+</sup>.<sup>79</sup> However, the Gly-X dipeptide containing aspartate (Gly-Asp), which also has a carboxylate group in the side chain, was hydrolyzed faster due to weaker coordination of the carboxylate as a result of the shorter side chain of Asp.<sup>79</sup> Interestingly, deamination of Asn to Asp and Gln to Glu was also observed during the hydrolysis of Gly-Asn or Gly-Gln catalyzed by Zr-K 2:2.<sup>84</sup> The sequence of the amino acids in the dipeptides also influenced the rate of hydrolysis. The rate of Ser-Gly hydrolysis was lower compared to Gly-Ser due to a less favorable four-membered ring transition state.<sup>79,84</sup> On the other hand, the rate of Ala-Gly hydrolysis was faster than that of Gly-Ala due to Ala-Gly being less prone to cyclization, which slows down the reaction as the cyclic side products need to be hydrolyzed back to the dipeptides to afford the individual amino acids as the final reaction products.<sup>79,86</sup> Hence, this demonstrates the potential selectivity of M-POMs towards the preferential peptide bond hydrolysis of certain combinations of amino acids in a specific order. These early studies with dipeptides have proven to be valuable as they provided molecular insight into factors that could influence the selectivity of M-POMs, as observed from their tendency to preferentially hydrolyze a particular sequence of amino acids in a polypeptide chain.

Following studies on dipeptides, the hydrolysis of larger oligopeptides – triglycine (Gly<sub>3</sub>), tetraglycine (Gly<sub>4</sub>), pentaglycine (Gly<sub>5</sub>), glycylglycylhistidine (Gly-Gly-His), glycylserylphenylalanine (Gly-Ser-Phe-Ala), and insulin chain B (30 amino acid polypeptide) – promoted by Zr-WD 1:2, Zr-WD 4:2, Zr-K 2:2, and Zr-L 1:1 was also investigated.<sup>80,82,89,90</sup> In the presence of Zr-WD 1:2, Gly<sub>4</sub> was hydrolyzed to form Gly, but intermediate products, Gly<sub>3</sub> and Gly-Gly, were observed as well, indicating that the reaction likely proceeds *via* sequential hydrolysis of the peptide bonds in the tetrapeptide, tripeptide, and dipeptide.<sup>89</sup> Out of all explored POMs, Zr-WD 1:2 displayed the best hydrolytic activity towards tri- and tetrapeptides. Moreover, NMR studies revealed that Keggin and Lindqvist POMs interact with Gly<sub>3</sub>, Gly<sub>4</sub>, and Gly-Ser-Phe-Ala through the amine nitrogen and the *N*-terminal amide oxygen, while Gly-Gly-His interacts *via* a different binding mode due to additional coordination of the imidazole nitrogen atom to Zr<sup>4+</sup>.<sup>82</sup> Slower hydrolysis was observed in the presence of Zr-L 1:1 and Zr-WD 4:2 species, which was attributed to the formation of less-active complexes in solution.<sup>82</sup> The hydrolytic rate was fastest in the case of the tetrapeptide Gly-Ser-Phe-Ala, where the Gly-Ser peptide bond was preferentially hydrolyzed thanks to the intramolecular





Table 2 Conditions and cleavage sites of selected reports on the hydrolysis of proteins promoted by different MOCs

| MOC              | Protein <sup>c</sup> | Reaction conditions                                 | No. of cleavages | Cleavage sites <sup>a</sup>  | Ref. |
|------------------|----------------------|---|------------------|--|------|
| Zr-WD 1 : 2      | HSA                  | pH 7.4, 60 °C, 10 eq. POM                           | 4                | Arg114–Leu115, Ala257–Asp258, Lys313–Asp314, Cys392–Glu393   | 106  |
|                  |                      | pH 7.4, 60 °C, 50 eq. POM + CHAPS                   | 7                | Cys62–Asp63, Gly71–Asp72, Asp107–Asp108, Lys313–Asp314, His367–Glu368, Ser470–Asp471, Ala511–Asp512  | 114  |
| Hf-WD 1 : 2      | HEWL                 | pH 5.0, 60 °C, 100 eq. POM                          | 6                | Asp18–Asn19, Asp48–Gly49, Thr51–Asp52, Asp52–Tyr53, Asp66–Gly67, Asp101–Gly102   | 96   |
|                  | Ova                  | pH 7.4, 60 °C, 100 eq. POM                          | 8                | Phe13–Asp14, Arg85–Asp86, Asn95–Asp96, Ala139–Asp140, Ser148–Trp149, Ala361–Asp362, Asp362–His363, Pro364–Phe365   | 98   |
| Zr-K 1 : 2       | Ova                  | pH 7.4, 60 °C, 50 eq. POM (+ SDS, CHAPS, or Zw3–12) | 6                |  | 115  |
|                  | Cyt c                | pH 4.1, 60 °C, 40 eq. POM                           | 3                | Asp3–Val4, Asp51–Ala52, Gly78–Thr79  | 95   |
|                  |                      | pH 7.4, 60 °C, 50 eq. POM + SDS, CHAPS, or Zw3–12   | 3                |  | 116  |
| Zr-K 2 : 2       | HHM                  | pH 5, 60 °C, 100 eq. POM                            | 6                | Asp4–Gly5, Asp20–Ile21, Asp44–Lys45, Asp60–Leu61, Asp126–Ala127, Asp141–Ile142   | 70   |
|                  |                      | pH 5.5, 60 °C, 100 eq. POM + SDS + CHAPS or Zw3–12  | 8                |  | 117  |
|                  | Hb                   | pH 5, 60 °C, 100 eq. POM                            | 6                | $\alpha$ -subunit: Asp6–Lys7, Asp74–Asp75, Asp75–Leu76, Asp85–Leu86, Asp94–Pro95, Asp116–Phe117<br>$\beta$ -subunit: Asp46–Leu47, Asp51–Ala52, Asp68–Ser69, Asp78–Asp79, Asp98–Pro99 | 97   |
|                  |                      |   | 11               |  |      |
| Ce-K 1 : 2       | Cyt c                | pH 4.1, 60 °C, 40 eq. POM                           | 3                | Asp3–Val4, Asp51–Ala52, Gly78–Thr79  | 95   |
|                  | Cyt c                | pH 4.1–7.4, 37–60 °C, 40 eq. POM                    | 2                |  | 95   |
|                  |                      | pH 7.4, 60 °C, 50 eq. POM + SDS or CHAPS + Zw3–12   | 2                | Trp60–Lys61, Gly78–Thr791  | 116  |
|                  | HEWL                 | pH 7.4, 37 °C, 10 eq. POM                           | None             | Trp28–Val29, Asn44–Arg45   | 85   |
|                  | Hb                   | pH 5, 60 °C, 100 eq. POM                            | 7                | $\alpha$ -subunit: Asp75–Leu76, Asp94–Pro95<br>$\beta$ -subunit: Asp51–Ala52, Asp68–Ser69, Asp78–Asp79, Asp98–Pro99, Asp128–Phe129   | 93   |
| Hf <sub>18</sub> | HHM                  | pH 7.4, 60 °C, 2 $\mu$ mol cluster                  | 6                | Asp5, Asp21, Asp45, Asp110, Asp123, Asp142 <sup>b</sup>  | 118  |

<sup>a</sup> Determined by Edman degradation. <sup>b</sup> Both Asp–X and X–Asp cleavage were observed at all sites. <sup>c</sup> HSA = Human Serum Albumin; HEWL = Hen Egg White Lysozyme; Ova = Ovalbumin; Cyt c = Cytochrome c; HHM = Horse Heart Myoglobin; Hb = Hemoglobin.

nucleophilic attack of the peptide carbonyl carbon by the hydroxyl group in the side chain of Ser, as observed for dipeptides. This highlights the beneficial role of Ser in peptide bond hydrolysis, even in larger peptides.<sup>82</sup>

**2.1.2. Hydrolysis of proteins.** The hydrolysis of proteins into peptide fragments is of particular importance in the field of proteomics as peptide fragments can be more easily characterized by mass spectrometry.<sup>91,92</sup> The data obtained from peptide fragments can then be combined to determine the amino acid sequence of a protein, including its post-translational modifications. Hence, based on the promising hydrolytic activity of POMs towards di-, tri-, and tetra- peptides discussed in Section 2.1.1., the catalytic activity of POMs was further investigated towards the hydrolysis of a wide range of proteins that differ in terms of isoelectric point (pI), structure, and size including hen egg white lysozyme (HEWL), human serum albumin (HSA), human serum holo-/apo-transferrin, ovalbumin, bovine blood hemoglobin, horse heart myoglobin, and bovine/horse heart cytochrome c (Table 2).<sup>70,85,88,90,93–99</sup> All reactions were performed under mild reaction conditions in acetate or phosphate buffer at 37 or 60 °C, and were followed by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE). In some cases, SDS-PAGE was coupled with Edman degradation and liquid chromatography tandem mass

spectrometry (LC-MS/MS) to determine the position of the hydrolyzed peptide bond in the protein sequence (Table 2). These studies have shown that POMs can be particularly interesting as artificial proteases for middle-down proteomics, which focuses on analyzing peptide fragments of around 3–15 kDa. Fragments of this size are ideal because they preserve structural information about the protein and reduce the complexity of the proteolytic mixture that is often obtained when using natural enzymes, such as trypsin, that produce many smaller fragments.<sup>100,101</sup> In order to understand the driving forces behind the hydrolytic activity, the interactions of POMs with proteins have also been investigated *in silico* by DFT calculations and molecular dynamics simulations, as well as experimentally using a range of complementary techniques, including multinuclear NMR, fluorescence, circular dichroism, and UV-Vis spectroscopy, along with Isothermal Titration Calorimetry (ITC) and Single Crystal X-ray Diffraction (SC-XRD).<sup>102–110</sup> These interaction studies have shown that POM-protein binding is mainly driven by charge-charge and H-bonding interactions, although hydrophobic interactions and coordinative binding can also occur, especially with POMs functionalized with organic moieties and with M-POMs, respectively.<sup>110–113</sup> However, since POM-protein interactions and the hydrolytic activity of POMs towards proteins have been





recently reviewed by our group,<sup>24,58,111</sup> this section will only briefly discuss the main conclusions obtained from these studies.

Similarly to peptides, studies on the hydrolytic activity of POMs towards proteins initially focused on using molybdates and then moved on to investigate M-POMs as catalysts.<sup>78</sup> These studies showed that  $[\text{MoO}_4]^{2-}$  was the catalytically active species in the hydrolysis of HEWL, which occurred preferentially at Asp-X peptide bonds. This was an intriguing finding because studies with dipeptides showed that X-Ser dipeptides were hydrolyzed the fastest. However, similar selectivity was also observed for the hydrolysis of different proteins catalyzed by  $\text{Zr}^{4+}$  and  $\text{Hf}^{4+}$  Wells-Dawson, Keggin, and Lindqvist M-POMs, regardless of the structure of the POM framework (Table 2).<sup>70,88,90,95–97</sup> Based on DFT calculations, this selectivity towards Asp-X bonds in proteins was proposed to result from the intramolecular attack of the metal-activated carbonyl carbon of the peptide bond by the carboxylate group in the side chain of Asp, leading to the formation of a five-membered cyclic hemiaminal that is then hydrolyzed (Fig. 5). Interestingly, such selectivity towards Asp-X bonds in proteins is extremely rare in nature, which makes POMs unique alternatives to natural enzymes. Furthermore, this shows that the reactivity towards simpler model systems is not always entirely representative of more complex biomolecules.

$\text{Ce}^{4+}$  M-POMs generally displayed slightly different selectivity than  $\text{Zr}^{4+}$  and  $\text{Hf}^{4+}$  M-POMs under the same conditions (Table 2).<sup>88,93–95</sup> Although the reasons for this are not yet fully understood, it was hypothesized that this difference originates from the redox properties of  $\text{Ce}^{4+}$  as the reduction of  $\text{Ce}^{4+}$  to  $\text{Ce}^{3+}$  was observed to occur in the presence of proteins, resulting in loss of the hydrolytic activity due to the lower Lewis acidity of  $\text{Ce}^{3+}$ .<sup>85,88,93,95</sup> Hence, the properties of the catalytically active metal center embedded in the POM cluster affect the overall reactivity.

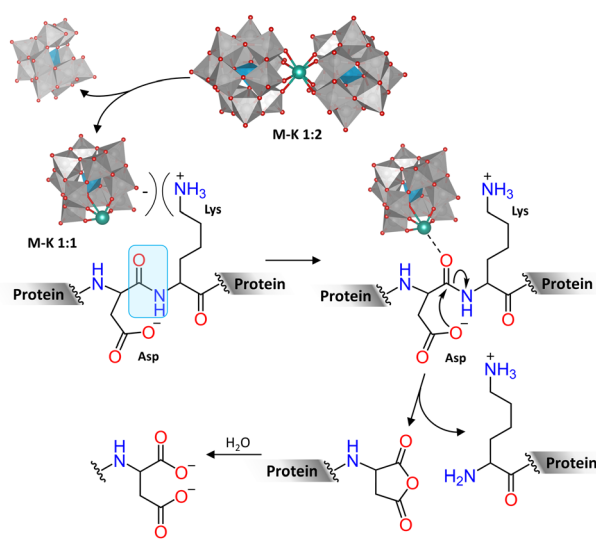


Fig. 5 Example mechanism of hydrolysis of an Asp-Lys peptide bond in a protein catalyzed by M-K 1:2, which dissociates into M-K 1:1 and a monolacunary Keggin POM at the protein surface.

Besides the crucial role of the Lewis acidic metal ion in the hydrolytic process, electrostatic interactions between negatively charged POMs and positively charged regions of proteins also have a significant influence on the outcome of the hydrolytic reaction. M-POM catalyzed hydrolysis typically occurs in positive regions of the protein due to favorable electrostatic interactions with the negatively charged POM, and, therefore, M-POMs with higher net negative charge have generally been reported to give rise to higher hydrolytic efficiencies. However, the catalytic activity of POMs has been found to also depend on the protein structure, as smaller proteins (*e.g.*, myoglobin and cytochrome *c*) displayed stronger interactions with smaller Keggin M-POMs, while larger proteins (*e.g.*, HSA and transferrin) interact more favorably with larger POMs, such as the Wells-Dawson M-POMs.<sup>70,88,95,99</sup> In addition, SC-XRD and DFT calculations indicated that the low dielectric constant at the surface of proteins facilitates the dissociation of M-POM dimers into the catalytically active monomeric species with more available coordination sites (Fig. 5).<sup>96</sup> Therefore, the interplay between the structure of the POM and that of the protein (*i.e.*, charge and size) is a determining factor in the hydrolysis reaction.

The reaction conditions also have an influence on the reactivity of M-POMs.<sup>58</sup> Acetate buffer (pH 5.5) has been determined to be more suitable for protein hydrolysis catalyzed by M-POMs than phosphate buffer (7.4) due to its lower pH and because of the inactivation of M-POMs by binding of phosphate to the embedded metal center.<sup>70,119</sup> Furthermore, the electrostatic nature of POM-protein interactions means that they are significantly affected by the pH, ionic strength, and temperature.<sup>58,111</sup> The reactivity also depends on the speciation of M-POMs, which in turn depends on the ionic strength, concentration, temperature, dielectric constant, and pH of the medium.<sup>58</sup> Hence, the conditions used need to be carefully considered to optimize the hydrolytic activity.

Recently, our group also demonstrated that M-POMs are mostly stable in the presence of surfactants and preserve their catalytic activity towards peptide bonds.<sup>114–116,120–123</sup> This is noteworthy since surfactants are essential in the study of many disease-related proteins and naturally water-insoluble proteins, such as membrane proteins. Studying these proteins is challenging and requires the use of surfactants to solubilize them. However, the presence of surfactants typically leads to denaturation of natural proteases causing them to lose their catalytic activity. In contrast, studies have shown that, even in the presence of surfactants, Zr-POMs are able to hydrolyze proteins such as  $\beta$ -casein, an unstructured and poorly soluble protein, and zein, a fully water-insoluble protein.<sup>120,122</sup> Therefore, M-POMs could aid in bridging the current gap in the study of disease-related proteins and naturally water-insoluble proteins that are difficult to hydrolyze with natural proteases.

Hydrolysis in the presence of surfactants can also be used to probe the structure of soluble proteins since protein unfolding induced by surfactants can lead to certain cleavage sites becoming more accessible to the POM catalyst depending on the protein structure. This was indeed observed for the hydrolysis



of myoglobin and HSA in the presence of anionic and/or zwitterionic surfactants as the addition of surfactants enhanced the hydrolytic efficiency of the M-POM and/or increased the number of peptide fragments (Table 2).<sup>114,117</sup> However, the presence of surfactant molecules interacting with residues on the surface of the protein may also hinder the binding of the POM to the protein surface. Therefore, compared to the reaction in the absence of surfactants, a decrease in the hydrolytic activity was observed in the presence of surfactants for the hydrolysis of ovalbumin, which has a high number of hydrophobic residues that can interact strongly with the hydrophobic tails of surfactants. Similarly, cytochrome *c* was hydrolyzed with a lower efficiency in comparison to the structurally similar myoglobin. This was attributed to the high pI of cytochrome *c*, which carries a higher overall positive charge compared to myoglobin, thus inducing stronger electrostatic interactions with zwitterionic and anionic surfactants, thereby blocking interactions between the protein and the M-POM. Accordingly, the hydrolytic activity of M-POMs towards proteins was observed to generally decrease with increasing concentration of surfactants, which hinders the exchange of surfactants with the M-POM at the protein surface and may induce dissociation of the M-POM.<sup>114,120,122</sup> In addition, the interaction of M-POMs with the ammonium groups of zwitterionic surfactants and their incorporation into micelles disrupted the equilibria between the dimeric and catalytically active monomeric species, which can negatively affect the hydrolytic activity of the M-POM.<sup>117</sup> Overall, these studies have shown that surfactants can be used as additives to tune the hydrolytic activity of POMs and the number of obtained peptide fragments. Moreover, the surfactant-protein interactions play an important role in the outcome of the hydrolytic reaction and depend on the structure of both the employed surfactant and the protein.

Despite the promising hydrolytic activity of POMs towards peptide bond cleavage, other MOCs have not been as extensively investigated for this type of reactivity. Nevertheless, a recent study by our group has shown that a water-insoluble neutral  $\text{Hf}^{4+}$  cluster (Fig. 6(a)),  $\text{Hf}_{18}\text{O}_{10}(\text{OH})_{26}(\text{SO}_4)_{13}(\text{H}_2\text{O})_{33}$  ( $\text{Hf}_{18}$ ), acted as an effective heterogeneous catalyst towards the hydrolysis of myoglobin in water and several buffers.<sup>118</sup> Interestingly,  $\text{Hf}_{18}$  hydrolyzed the protein at Asp-X and X-Asp cleavage sites in both positively and negatively charged regions

of the protein (Table 2), in contrast to the selectivity of M-POMs for Asp-X peptide bonds in positively charged protein regions. This was attributed to the synergistic effect of both Lewis and Brønsted acidity of the cluster. The Brønsted acidity due to labile water ligands bound to  $\text{Hf}^{4+}$  centers is distinct from the purely Lewis acidic POM catalysts and was essential for the hydrolytic activity of  $\text{Hf}_{18}$ . However, the protein was observed to adsorb strongly onto the solid MOC catalyst, especially at lower pH, and, as a result, the protein and peptide fragments could not be easily separated after the reaction without deactivating the solid catalyst. Therefore, further optimization of the separation protocol or the structure of the cluster is necessary in order to obtain a recyclable heterogeneous MOC catalyst for the hydrolysis of proteins.

**2.1.3. Peptide bond formation.** The cyclic by-products that were observed during the hydrolysis of dipeptides due to intramolecular peptide bond formation suggested that these POMs can also promote peptide bond formation. The formation of peptide bonds is important for producing many synthetic materials, such as pharmaceuticals and polymers, but remains relatively challenging as the removal of water and the use of additives to facilitate the reaction are often necessary.<sup>124,125</sup> Hence, the ability of Zr-POMs to promote amide bond formation in aqueous solutions is remarkable, and has been further explored by our group using POMs and Zr-oxo clusters to catalyze peptide bond formation. M-POMs were shown to effectively catalyze the cyclization of several different dipeptides in dimethyl sulfoxide (DMSO), with better activity being observed for  $\text{Zr}^{4+}$  and  $\text{Hf}^{4+}$  M-POMs than for M-POMs with other embedded metal centers ( $\text{Fe}^{3+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Ce}^{3+}$ , and  $\text{Ce}^{4+}$ ), similarly to peptide bond hydrolysis.<sup>126</sup>  $\text{Zr}^{4+}$  and  $\text{Hf}^{4+}$  M-POMs have also been shown to catalyze direct intermolecular amide bond formation in different organic solvents.<sup>127,128</sup> The catalytic activity of M-POMs towards both intra- and intermolecular peptide bond formation was found to depend on the POM structure and number of embedded metal centers.<sup>126,129</sup> Additionally, neutral hexanuclear and dodecanuclear  $\text{Zr}^{4+}$  clusters ( $[\text{Zr}_6(\text{OH})_4\text{O}_4(\text{OMc})_{12}]$  where Mc = methacrylate and  $[\text{Zr}_6(\text{OH})_4\text{O}_4(\text{OAc})_{12}]_2$  where Ac = acrylate) have also shown promising catalytic activity towards direct intermolecular amide bond formation (Fig. 1(f) and 6(b)).<sup>127,128</sup> These findings highlight the potential of both M-POMs and neutral  $\text{Zr}^{4+}$  clusters as highly promising water-tolerant catalysts for peptide bond formation that do not require harsh conditions or the use of additives.<sup>126–129</sup>

**2.1.4. Oxidative modifications and cleavage.** Investigating the POM-promoted oxidation of amino acids, peptides, and proteins is essential for potential biomedical applications of POMs as it can provide valuable insights into the underlying mechanisms behind their biological activity. While the biological activity of POMs is largely believed to be linked to their redox activity, the mechanism of action is still not well understood.<sup>130</sup> Furthermore, oxidative modifications of proteins have been linked to many diseases, such as Alzheimer's and cancer. Therefore, selectively reproducing similar modifications *in vitro* using POMs would provide a better

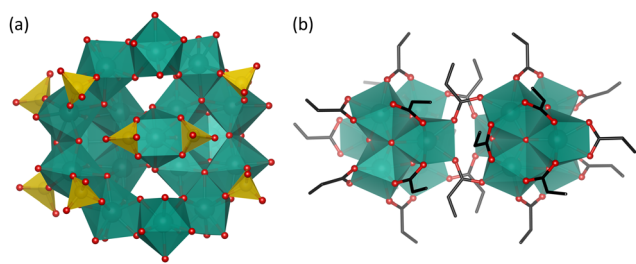


Fig. 6 Structures of (a)  $\text{Hf}_{18}\text{O}_{10}(\text{OH})_{26}(\text{SO}_4)_{13}(\text{H}_2\text{O})_{33}$  ( $\text{Hf}_{18}$ ) and (b)  $[\text{Zr}_6(\text{OH})_4\text{O}_4(\text{OAc})_{12}]_2$  (Ac = acrylate). Hafnium in teal, oxygen in red, sulfur in yellow, carbon in black.



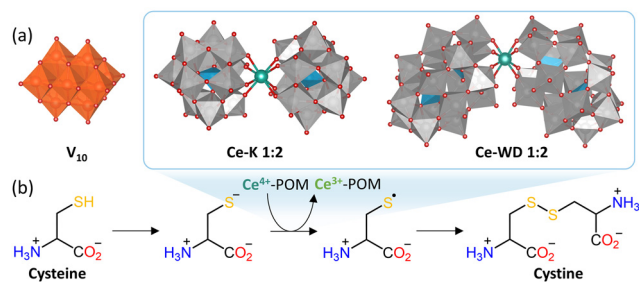


Fig. 7 (a) POM structures that have been reported to induce the oxidation of discrete amino acids or amino acids in peptides and/or proteins. (b) Proposed mechanism of the oxidation of cysteine to cystine via homocoupling of cysteinyl radicals.

understanding of the role of oxidation in the onset of these diseases.<sup>130–133</sup> Hence, the redox activity of POMs towards biomolecules could enable interesting opportunities in biomedicine.

POMs with redox-active metal centers have been shown to induce oxidative modifications in redox-active amino acids. Decavanadate ( $[H_xV_{10}O_{28}]^{(6-x)-}$ ;  $V_{10}$ ) has been proposed to oxidize certain cysteine (Cys) residues in actin proteins (Fig. 7(a)), which are present in muscle filaments, and in the  $Ca^{2+}$ -ATPase enzyme responsible for  $Ca^{2+}$  homeostasis and muscle relaxation.<sup>134,135</sup> This redox activity is likely linked to both the structure of  $V_{10}$  and the ease of  $V^{5+}$  reduction to  $V^{4+}$ . Vanadates with lower nuclearity did not oxidize Cys residues in actin and the isostructural decaniobate ( $[Nb_{10}O_{28}]^{6-}$ ;  $Nb_{10}$ ), which is kinetically inert and redox inactive, did not induce oxidation of Cys in  $Ca^{2+}$ -ATPase.<sup>134,135</sup> Furthermore, the redox activity of  $V_{10}$  is believed to be in part responsible for its ability to inhibit actin-stimulated myosin ATPase and  $Ca^{2+}$ -ATPase activity, which is of interest for the investigation and regulation of muscle function among other possible biomedical applications.<sup>134,136</sup> This illustrates how the redox activity of POMs likely plays an important role in their biological activity, which is especially relevant when exploring POMs as potential inorganic drugs.<sup>57</sup>

Our group has also recently shown that  $Ce^{4+}$ -substituted Keggin and Wells–Dawson POMs (Ce–POMs) displayed redox activity towards certain amino acids (Fig. 7(a)), which is likely driven by coordination of free carboxylate or amino groups to  $Ce^{4+}$ , as well as electrostatic and/or H-bonding interactions.<sup>137</sup> As with  $V_{10}$ , Cys was shown to be particularly susceptible to undergo oxidation into cystine in the presence of Ce–POMs, resulting in a concomitant reduction of  $Ce^{4+}$  to  $Ce^{3+}$ . Other redox-active amino acids – tryptophan (Trp), tyrosine (Tyr), histidine (His), and phenylalanine (Phe) – were also observed to cause reduction of  $Ce^{4+}$  in the M–POM to  $Ce^{3+}$ , but no oxidation products were detected.<sup>137</sup> This suggests that oxidation takes place *via* a radical mechanism in which the amino acids can be regenerated by the solvent or, in the case of Cys, cystine can form through homocoupling of cysteinyl radicals (Fig. 7(b)).

The rate of  $Ce^{4+}$  reduction in Ce–POMs by amino acids was found to depend on the nature of the amino acid side chain

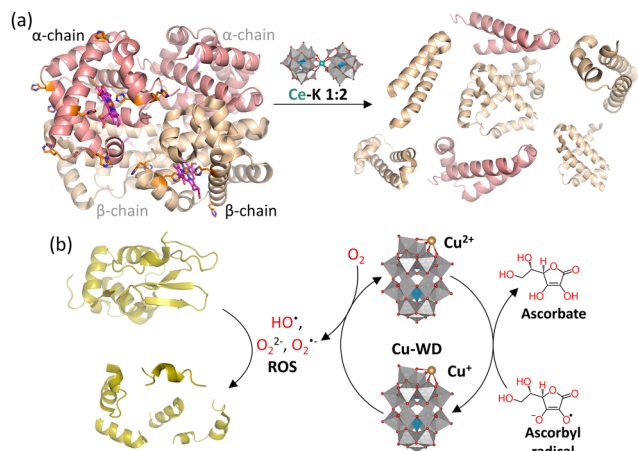
according to the following order: Cys  $\gg$  Trp  $>$  Tyr  $>$  Phe  $\approx$  His.<sup>137</sup> This corresponds to an inverse correlation with the redox potential of the amino acids, which means that amino acids that are oxidized more easily caused faster reduction of  $Ce^{4+}$ . The reaction rate of Cys oxidation was also observed to increase with the pH, temperature, and ionic strength. This suggests that cysteinyl radicals are formed from cysteinyl anions, which are favored at higher pH and whose coordination to the anionic M–POM is more favorable at higher ionic strengths due to shielding of repulsive electrostatic interactions. A similar influence of pH and ionic strength was also noted for Trp and Tyr, which can be attributed to a decrease in redox potential with increasing pH. In addition, due to the influence of the POM scaffold on the redox potential of  $Ce^{4+}$ , the rate of the reaction was also found to depend on the POM framework, with the Keggin Ce–POM giving a faster rate by one order of magnitude in comparison to the Wells–Dawson Ce–POM. Hence, several factors influence the ability of M–POMs to oxidize amino acids in solution, and the redox potential of both the amino acids and the  $Ce^{4+}$  center in the POM play an important role.

Oxidation of peptides was also observed in the presence of Ce–POMs, but the rate of reaction was affected by the incorporation of the amino acid into the peptide due to changes in the redox potential and additional steric effects.<sup>137</sup> Interestingly, the oxidation of peptides occurred at room temperature in the absence of any hydrolysis, indicating that the oxidation reaction is faster and the resulting formation of the  $Ce^{3+}$ -POM further prevents hydrolysis from occurring due to its lower Lewis acidity. In contrast, proteins have only been observed to induce reduction of the  $Ce^{4+}$  center in Ce–POMs upon incubation at higher temperatures. This was proposed to be due to amino acid residues within proteins being less accessible towards electron transfers, which makes reduction of  $Ce^{4+}$  by proteins less favorable at room temperature compared to peptides.<sup>93</sup> As a result, only partial reduction of  $Ce^{4+}$  to  $Ce^{3+}$  has been reported as a side-reaction during the hydrolysis of proteins by Ce–POMs.<sup>85,116</sup>

The redox activity of  $Ce^{4+}$  in Ce–POMs was found to have an impact on the efficiency and selectivity of the hydrolytic cleavage of hemoglobin, a tetrameric protein composed of two  $\alpha$ -chains and two  $\beta$ -chains (Fig. 8(a) and Table 2).<sup>93</sup> It was observed that, at pH 5, Asp–X peptide bonds were hydrolyzed on both the  $\alpha$ -chain (2 cleavage sites) and the  $\beta$ -chain (5 cleavage sites), but at pH 7.4 hydrolysis only occurred on the  $\beta$ -chain (1 cleavage site). This reduction in hydrolytic activity with increasing pH corresponds to weaker electrostatic interactions between the M–POM and the protein (pI = 6.8) and a higher rate of reduction of  $Ce^{4+}$  into the hydrolytically inactive  $Ce^{3+}$  at higher pH. Furthermore, the higher hydrolytic activity of the M–POM towards the  $\beta$ -chain is likely due to the higher number of redox-active residues on the  $\alpha$ -chain, particularly His, which are able to reduce hydrolytically active  $Ce^{4+}$  into the hydrolytically inactive  $Ce^{3+}$ . This was confirmed by the hydrolytic activity of the redox-inactive  $Zr^{4+}$ -POM towards the  $\alpha$ -chain (6 cleavage sites), which was higher compared to the  $Ce^{4+}$ -POM and did







**Fig. 8** (a) Hydrolysis of hemoglobin promoted by Ce–K 1:2 preferentially at the  $\beta$ -chain (beige) due to the higher number of redox active amino acids, such as histidine (orange), in the  $\alpha$ -chain (pink) that cause reduction of  $\text{Ce}^{4+}$  to the less hydrolytically active  $\text{Ce}^{3+}$ . Adapted from ref. 93 with permission from the Royal Society of Chemistry. (b) Oxidative cleavage of lysozyme due to the production of reactive oxygen species (ROS) induced by Cu–WD in combination with ascorbate.

not decrease as significantly upon increasing the pH.<sup>97</sup> Therefore, the redox activity of certain M–POMs can be leveraged to tune their hydrolytic activity and selectivity. In addition, fragmentation of proteins catalyzed by redox-active POMs can occur in a stepwise manner first involving oxidation of amino acid residues followed by rearrangement and hydrolytic cleavage. This was observed for the cleavage of  $\text{Ca}^{2+}$ -ATPase enzyme into two fragments catalyzed by  $\text{V}_{10}$  under UV irradiation, which occurred presumably *via* photo-oxidation of certain amino acid residues, such as serine.<sup>138</sup> Hence, the oxidation of amino acid residues by POMs may play an integral role in protein fragmentation that can be leveraged for bioanalytical applications.

Another strategy to achieve protein fragmentation is through oxidative cleavage of the peptide bond. Our group has recently demonstrated that a  $\text{Cu}^{2+}$ -substituted Wells–Dawson POM (Fig. 8(b)),  $[\text{Cu}(\text{H}_2\text{O})(\alpha_2\text{-P}_2\text{W}_{17}\text{O}_{61})]^{8-}$  (Cu-WD), can selectively cleave HEWL in the presence of ascorbate (Asc) *via* the formation of reactive oxygen species (ROS).<sup>130</sup> The proposed mechanism involves the Asc-induced reduction of  $\text{Cu}^{2+}$  to  $\text{Cu}^+$ , which then reduces  $\text{O}_2$  in the solution to form ROS in the vicinity of the protein (Fig. 8(b)), thereby producing radicals on the protein that subsequently break the peptide chain. Furthermore, as with the previously discussed hydrolytic reactions, binding of the POM to specific positively charged regions of the protein imposes the selectivity of the oxidative cleavage. However, protein fragmentation by oxidative cleavage is much faster than hydrolytic cleavage since fragments were obtained under physiological pH and temperature (37 °C; pH 7.5) after just 1 h. The cleavage efficiency was also observed to increase at more acidic pH, which was attributed to the enhanced production of ROS at lower pH. Hence, oxidative cleavage induced by POMs is a promising avenue for the selective oxidative modification and fragmentation of proteins, with many potential

industrial, biomedical, and bioanalytical applications, such as in redox proteomics.<sup>139</sup>

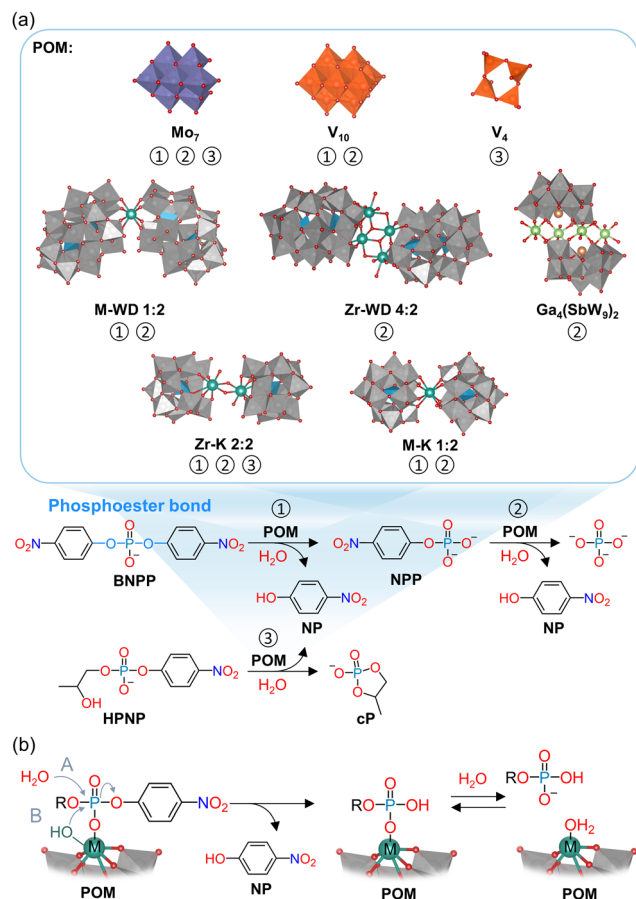
## 2.2. Nucleic acids and nucleotides

### 2.2.1. Hydrolysis of nucleic acids and their model systems.

The phosphoester bond, which forms the backbone of nucleic acids, is one of the most stable bonds present in biomolecules, with a half-life of  $\sim 100\,000$  years for DNA, which allows for the preservation of genetic material.<sup>140</sup> Nevertheless, the cleavage of phosphoester bonds is an essential step in a wide range of cellular processes, including DNA repair and excision.<sup>141,142</sup> In living organisms this is typically performed by metalloenzymes known as phosphatases or nucleases, most of which catalyze the hydrolysis of phosphoester bonds through two or more adjacent metal ions. These metal ions are typically positioned in such a way that activation of both the phosphate substrate and the water nucleophile is possible, which is necessary for the hydrolytic process to occur.<sup>143</sup> Hence, due to their inherent multinuclear composition, metallic clusters such as POMs, are particularly interesting for the development of artificial phosphatases. Investigating the hydrolytic activity of MOCs towards phosphoester bonds may also provide insights into the structure–activity relationships of metallic clusters in natural phosphatases. In addition, natural phosphatases are not easy to obtain in high purity and their enzymatic activity is often affected by the presence of other species in solution, such as amino acids. Therefore, there is a need for more accessible artificial phosphatases, which are of crucial importance in biology and medicine for the development of new therapeutic agents and biochemical tools.<sup>144,145</sup> In this respect, POMs have been shown to have promising antiviral, antibacterial, and antitumor activity that has often been linked to their phosphatase-like activity, making them attractive candidates for medical applications.<sup>56,57,146–149</sup> Hence, in order to better understand the biological activity of POMs and to develop them as artificial phosphatases, interaction and reactivity studies with nucleic acids and their model systems are of substantial interest.

Initial efforts to develop POMs as artificial phosphatases focused mostly on heptamolybdate ( $[\text{Mo}_7\text{O}_{24}]^{6-}$ ;  $\text{Mo}_7$ ), in part because this POM has shown considerable antitumor activity against different tumor cell lines, displaying comparable activity *in vivo* to some commercially used drugs.<sup>150–156</sup> Additionally, DNA fragmentation was detected for tumor cells treated with this POM.<sup>150</sup> In agreement with these observations,  $\text{Mo}_7$  was shown to catalyze the hydrolysis of (*p*-nitrophenyl)phosphate (NPP) and bis(*p*-nitrophenyl)phosphate (BNPP), which are commonly used as model systems for DNA, as well as 2-hydroxypropyl-4-nitrophenyl phosphate (HPNP), an RNA model substrate (Fig. 9).<sup>151–153,157</sup> Although multiple oxomolybdate clusters that differ in structure and nuclearity can form in aqueous solution depending on the pH and the presence of other species in solution (*e.g.*, templating phosphate ions),<sup>77</sup>  $\text{Mo}_7$  was proposed to be the catalytically active species since the rate of hydrolysis was highest at the pH where  $\text{Mo}_7$  is the major component (pH 5–6).<sup>151,153,157</sup> In contrast, hydrolysis was





**Fig. 9** (a) Hydrolysis of the phosphoester bond of DNA model systems, NPP and BNPP, catalyzed by  $[\text{Mo}_7\text{O}_{24}]^{6-}$  ( $\text{Mo}_7$ ),  $[\text{H}_x\text{V}_{10}\text{O}_{28}]^{(6-x)-}$  ( $\text{V}_{10}$ ),  $[\text{M}(\alpha_2\text{-P}_2\text{W}_{17}\text{O}_{61})_2]^{16-}$  ( $\text{M-WD } 1:2$ ),  $[\text{Zr}_4(\mu_3\text{-O})_2(\text{OH})_2(\text{H}_2\text{O})_4(\text{P}_2\text{W}_{16}\text{O}_{59})_2]^{14-}$  ( $\text{Zr-WD } 4:2$ ),  $[\text{Ga}_4(\text{H}_2\text{O})_{10}(\beta\text{-SbW}_9\text{O}_{33})_2]^{6-}$  ( $\text{Ga}_4(\text{SbW}_9)_2$ ),  $[\text{Zr}(\mu\text{-OH})(\text{H}_2\text{O})\alpha\text{-PW}_{11}\text{O}_{39}]_2^{8-}$  ( $\text{Zr-K } 2:2$ ), and  $[\text{M}(\alpha\text{-PW}_{11}\text{O}_{39})_2]^{10-}$  ( $\text{M-K } 1:2$ ) as well as of the RNA model system HPNP by  $\text{Mo}_7$ ,  $[\text{V}_4\text{O}_{12}]^{4-}$  ( $\text{V}_4$ ), and  $\text{Zr-K } 2:2$ . (b) Generalized mechanism of the hydrolysis of phosphate model systems by POMs through activation of the phosphoester bond towards nucleophilic attack by (A) solvent water or (B) metal-coordinated  $\text{HO}^-/\text{H}_2\text{O}$ .

barely observed at lower pH, where  $[\text{Mo}_8\text{O}_{26}]^{4-}$  ( $\text{Mo}_8$ ) is the main species (pH 2–6), or at higher pH (pH 8), where monomeric  $[\text{MoO}_4]^{2-}$  predominates.<sup>151,157</sup> Therefore, this demonstrates that the specific multimetallic structure of  $\text{Mo}_7$  is important for its catalytic activity towards phosphoester bond cleavage.

Even though the biological activity of POMs is often attributed to their redox properties, the cleavage of the phosphoester bond by  $\text{Mo}_7$  was established to be purely hydrolytic and not oxidative in nature since the reduction of  $\text{Mo}^{6+}$  to  $\text{Mo}^{5+}$  was not observed.<sup>151,153</sup> The ability of  $\text{Mo}_7$  to hydrolyze the phosphoester bond was suggested to be due to its lability and the partial detachment of at least one  $\{\text{MoO}_4\}$  unit.<sup>152,153</sup> This allows for the incorporation of a phosphate group into the POM structure, which can induce bond strain and polarization of the P–O ester bond, thereby activating it towards nucleophilic attack by water.<sup>152,153</sup> Hence, the isostructural heptatungstate POM  $[\text{W}_7\text{O}_{24}]^{6-}$  was reported to be hydrolytically inactive as it

is less labile.<sup>157,158</sup> Therefore, both the POM's structure and the composition of the POM in terms of the metal centers present have a large impact on its ability to promote the reaction.

Based on the proposed mechanism, the reaction of  $\text{Mo}_7$  with phosphate substrates should result in the formation of an intermediate complex consisting of an oxomolybdate species with the bound phosphate substrate. In the case of the hydrolysis of NPP, these intermediate structures were proposed to be  $[(\text{NPP})_2\text{Mo}_5\text{O}_{21}]^{4-}$  and  $[(\text{NPP})_2\text{Mo}_{12}\text{O}_{36}(\text{H}_2\text{O})_6]^{4-}$  based on diffusion ordered spectroscopy (DOSY) NMR.<sup>152</sup> These proposed structures are in agreement with the formation of a catalytically inactive phosphomolybdate species,  $[(\text{PO}_4)_2\text{Mo}_5\text{O}_{15}]^{6-}$  ( $\text{P}_2\text{Mo}_5$ ), after the hydrolysis of NPP and BNPP by  $\text{Mo}_7$ , as determined by both NMR and Extended X-ray Absorption Fine Structure (EXAFS) spectroscopy.<sup>152,153</sup> However, only a small amount of  $\text{P}_2\text{Mo}_5$  was formed after the hydrolysis of HPNP by  $\text{Mo}_7$ .<sup>151</sup> This is likely because the reaction does not result in the formation of phosphate as the cyclic phosphate ester is produced instead (Fig. 9(a)). More recently, DFT calculations and electrospray ionization mass spectrometry (ESI-MS) have suggested that  $\text{Mo}_7$  dissociates to form  $[\text{Mo}_2\text{O}_8]^{4-}$  ( $\text{Mo}_2$ ) and  $[\text{Mo}_5\text{O}_{15}]^0$  ( $\text{Mo}_5$ ). The binding of NPP to  $\text{Mo}_2$ , forming  $[\text{Mo}_2\text{O}_4(\text{OH})_4(\text{NPP})]^{2-}$ , was proposed to be responsible for promoting the cleavage of the phosphoester bond while the role of  $\text{Mo}_5$  is to capture the phosphate resulting from the phosphoester bond cleavage leading to the formation of  $\text{P}_2\text{Mo}_5$ .<sup>158</sup> Overall, these results highlight the challenging nature of identifying the intermediate species that can form in reactions catalyzed by highly dynamic POM structures.

The hydrolysis of phosphoester bonds was observed to be catalyzed by vanadates as well. The reaction was determined to also be hydrolytic in nature and to proceed *via* incorporation of the phosphate substrate into the POM framework to activate the P–O bond.<sup>159</sup> This was supported by the fact that the rate of hydrolysis of NPP and BNPP increased with the ionic strength of the solution due to shielding of repulsive electrostatic interactions between the negatively charged substrate and the anionic POM catalyst.<sup>160</sup> Interestingly, based on the pH dependence of the rate constant,  $\text{V}_{10}$  was identified to be the main catalytically active species in the hydrolysis of NPP and BNPP, while  $[\text{V}_4\text{O}_{12}]^{4-}$  ( $\text{V}_4$ ) was proposed to be responsible for promoting the hydrolysis of HPNP (Fig. 9(a)).<sup>159,160</sup> This difference could be attributed to the formation of vanadate ester intermediates with HPNP, which was determined to be deprotonated under the employed reaction conditions. However, the exact nature of the intermediate species formed upon binding of the phosphate substrate to  $\text{V}_{10}$  or  $\text{V}_4$  is unknown. Furthermore, the characterization of the species in solution is complicated by the dynamic exchange between oxovanadate species and/or the rapid association–dissociation equilibrium with the phosphate substrate in solution as observed by NMR.<sup>159,160</sup> Nevertheless, the catalytic activity of  $\text{V}_{10}$  and  $\text{V}_4$  could also be attributed to their lability, which allows for the incorporation of the substrate into a polyoxovanadate cluster thereby activating it towards nucleophilic attack.



Overall, these studies on the reactivity of molybdates and vanadates towards DNA and RNA model systems showcase that due to the dynamic and complex speciation of these POMs in aqueous solutions, the exact structures of the intermediate species formed upon binding of the substrate are difficult to determine.<sup>152,153,158–160</sup> Hence, a multi-technique approach is necessary to elucidate the nature of the species present throughout a reaction.

Tungstate M-POMs were also investigated as catalysts towards the hydrolysis of the phosphoester bond since they are less dynamic in solution. Moreover, the embedded Lewis acidic metal centers in M-POMs can also activate the phosphoester bond as well as coordinate H<sub>2</sub>O and/or HO<sup>−</sup>, which can serve as a nucleophile that attacks the phosphoester bond (Fig. 9(b)). Several Wells–Dawson M-POMs with different metal ions were screened and the Zr<sup>4+</sup> and Hf<sup>4+</sup> M-POMs ([M(α<sub>2</sub>-P<sub>2</sub>W<sub>17</sub>O<sub>61</sub>)<sub>2</sub>]<sup>16−</sup> where M = Zr<sup>4+</sup> or Hf<sup>4+</sup>) were found to display

better hydrolytic activity towards both NPP and BNPP while M-POMs with other embedded metal ions (M = Mn<sup>3+</sup>, Fe<sup>3+</sup>, Co<sup>2+</sup>, Ni<sup>2+</sup>, Y<sup>3+</sup>, La<sup>3+</sup>, Eu<sup>3+</sup>) displayed either no hydrolytic activity or were only able to hydrolyze NPP (Fig. 9). The better activity of Zr<sup>4+</sup> or Hf<sup>4+</sup> (Table 3) is due to their higher Lewis acidity, oxophilicity, redox-inactivity, fast ligand exchange as well as their ability to form complexes with high coordination numbers and flexible geometries.<sup>161</sup> In addition, similarly to the hydrolysis of peptide bonds (Section 2.1.1), it was determined that the 1 : 1 and 2 : 2 M-POM species were responsible for the hydrolytic activity since these species have vacant coordination sites that are available for the phosphate substrate to coordinate. This was confirmed by the faster rate of hydrolysis at lower pH values, which favors the formation of the monomeric 1 : 1 M-POM. The reaction was determined to proceed through a similar mechanism to that observed for molybdates and vanadates, involving activation of the P–O bond towards

**Table 3** Conditions and rate constants (*k*<sub>obs</sub>) of selected reports on the hydrolysis of phosphoester/phosphoanhydride bonds catalyzed by different POMs

| POM   | Substrate | Reaction conditions                                      | Conversion & <i>k</i> <sub>obs</sub> /×10 <sup>−6</sup> s <sup>−1</sup> | Ref. |
|---|-----------|--|---|------|
| Mo <sub>7</sub>   | NPP       | pD 5.5, 50 °C, 1 eq. POM                                 | 78.0  | 152  |
|   |           | pD 5.1, 50 °C, 7 eq. POM                                 | 27.3  | 153  |
|   | BNPP      | pD 5.5, 50 °C, 1 eq. POM                                 | 2.3   | 157  |
|   | HPNP      | pD 5.7, 50 °C, 0.1–1.0 eq. POM                           | 3.9 (0.1 eq.)–6.4 (1 eq.)   | 151  |
|   | ATP       | pH 2–6, r.t., 20 eq. Na <sub>2</sub> MoO <sub>4</sub>    | 3.7 (pH 6)–5.5 (pH 2) mM h <sup>−1</sup>                                | 162  |
| Mo <sub>8</sub> Pro   | ADP       | pH 4, r.t., 5 h, 20 eq. Na <sub>2</sub> MoO <sub>4</sub> | 12% yield of AMP  | 162  |
|   | ATP       | pH 3.4, r.t., 12 h, 3 eq. POM                            | 100% conversion   | 163  |
| Mo <sub>12</sub> (Mo <sub>4</sub> )                                     | ATP       | pH 7, r.t., 48 h, 3 eq. POM                              | 75% conversion  | 164  |
|   |           | pH 5, 25 °C, 10 days, 1 eq. POM                          | 10% conversion  |      |
|   |           | pH 5, 40 °C, 10 days, 1 eq. POM                          | 72% conversion  |      |
|   |           | pH 5, 40 °C, 10 days, 1 eq. POM, K <sup>+</sup>          | 86% conversion  |      |
|   |           | pH 7.5, 40 °C, 10 days, 1 eq. POM                        | 41% conversion  |      |
|   | ADP       | pH 5, 40 °C, 10 days, 1 eq. POM                          | 26% conversion  | 164  |
|   |           | pH 7.5, 40 °C, 10 days, 1 eq. POM                        | 6% conversion   |      |
| V <sub>10</sub>   | NPP       | pD 5.0, 50 °C, 10 eq. of Na <sub>3</sub> VO <sub>4</sub> | 17.0  | 160  |
|   | BNPP      | pD 5.0, 70 °C, 10 eq. of Na <sub>3</sub> VO <sub>4</sub> | 3.3   | 160  |
|   | ATP       | pH 2, 25 °C, 10 eq. Na <sub>3</sub> VO <sub>4</sub>      | 1130.0 ( <i>k</i> <sub>1</sub> ; ATP → ADP)                             | 165  |
|   |           |  | 46.7 ( <i>k</i> <sub>2</sub> ; ADP → AMP)                               |      |
|   |           |  | 10730.0 ( <i>k</i> <sub>1</sub> ; ATP → ADP)                            |      |
| V <sub>4</sub>  | HPNP      | pD 7.0, 37 °C, 0.66 eq. Na <sub>3</sub> VO <sub>4</sub>  | 383.3 ( <i>k</i> <sub>2</sub> ; ADP → AMP)                              | 159  |
|   |           |  | 4.3 ( <i>k</i> <sub>1</sub> ; ATP → ADP)                                |      |
| M-WD 1 : 2 (M = Y <sup>3+</sup> , La <sup>3+</sup> , Eu <sup>3+</sup> ) | NPP       | pD 7.2, 50 °C, 1 eq. POM                                 | 1.9   | 161  |
|   |           | pD 7.2, 50 °C, 1 eq. POM                                 | 0.3 (M = Y <sup>3+</sup> )  |      |
| Zr-WD 1 : 2   | NPP       | pD 7.2, 50 °C, 1 eq. POM                                 | 0.8 (M = La <sup>3+</sup> , Eu <sup>3+</sup> )                          | 161  |
|   | BNPP      | pD 7.2, 50 °C, 1 eq. POM                                 | 12.8  |      |
| Hf-WD 1 : 2   | NPP       | pD 7.2, 50 °C, 1 eq. POM                                 | 0.7   | 161  |
|   | BNPP      | pD 7.2, 50 °C, 1 eq. POM                                 | 19.7  |      |
| Zr-WD 4 : 2   | NPP       | pD 6.4, 50 °C, 0.2 eq. POM                               | 1.1   | 161  |
|   | BNPP      | pD 6.4, 50 °C, 0.2 eq. POM                               | 84.4  |      |
|   | HPNP      | pD 6.4, 50 °C, 0.2 eq. POM                               | 0.3   |      |
| Al <sub>4</sub> (SbW <sub>9</sub> ) <sub>2</sub>                        | NPP       | pD 6.4, 50 °C, 0.2 eq. POM                               | 5.1   | 166  |
| Ga <sub>4</sub> (SbW <sub>9</sub> ) <sub>2</sub>                        | NPP       | pD 6–7, 60 °C, 0.5 eq. POM                               | 6.9 (pD 7)–12.2 (pD 6)  | 167  |
|   | NPP       | pD 4–8, 60 °C, 0.5 eq. POM                               | 0.9 (pD 8)–23 (pD 4)  | 167  |
|   | ATP       | pD 5, 50 °C, 20 h, 0.1 eq. POM                           | 75% conversion  | 167  |
| Zr-K 1 : 2  | BNPP      | pD 6.4, 60 °C, 1 eq. POM                                 | 1.1   | 143  |
|   | HPNP      | pD 6.4, 50 °C, 1 eq. POM                                 | 9.2   | 168  |
| Zr-K 2 : 2  | BNPP      | pD 6.4, 60 °C, 1 eq. POM                                 | 4.8   | 143  |
|   | HPNP      | pD 6.4, 50 °C, 1 eq. POM                                 | 115.0   | 168  |
|   | ATP       | pD 6.4, 50 °C, 1 eq. POM                                 | 10.4  | 144  |
|   |           | pD 6.4, 50 °C, 0.15 eq. POM                              | 1.97  |      |
| Ce-K 1 : 2  | NPP       | pD 6.4, 50 °C, 1 eq. POM                                 | 5.3   | 169  |

r.t. = room temperature.





nucleophilic attack by a water molecule through coordination of the phosphate substrate to the embedded metal center, but without any structural rearrangement of the POM (Fig. 9(b)). Although the rate of hydrolysis was slower compared to Mo<sub>7</sub> (Table 3), the M-POMs could be used in catalytic amounts and under physiological pH conditions.<sup>161</sup>

The reactivity of M-POMs with multiple substituted metal centers was also investigated since the hydrolysis of phosphoester bonds is usually favored through adjacent metal centers, as observed in natural phosphatases. Accordingly, [Zr<sub>4</sub>(μ<sub>3</sub>-O)<sub>2</sub>(OH)<sub>2</sub>(H<sub>2</sub>O)<sub>4</sub>(P<sub>2</sub>W<sub>16</sub>O<sub>59</sub>)<sub>2</sub>]<sup>14-</sup> (Zr-WD 4:2), which has four Zr<sup>4+</sup> centers, gave rise to faster hydrolysis of NPP in comparison to Zr-WD 1:2 (Table 3), which has only one Zr<sup>4+</sup> center, and the catalyst remained active after at least 3 cycles.<sup>166</sup> For this reaction, it was proposed that NPP coordination occurs *via* the replacement of one of the waters coordinated to two Zr<sup>4+</sup> centers, {Zr(H<sub>2</sub>O)<sub>2</sub>}, and the phosphoester bond could be hydrolyzed by nucleophilic attack of the second coordinated water or solvent water (Fig. 9(b)). Furthermore, since there are two Zr<sup>4+</sup> centers with coordinated water ligands, coordination and hydrolysis of NPP can potentially occur simultaneously at both sites. Moreover, Zr-WD 4:2 was found to be active towards BNPP and HPNP as well.<sup>166</sup> Similarly, the sandwich-type Krebs M-POMs ([M<sub>4</sub>(H<sub>2</sub>O)<sub>10</sub>(β-SbW<sub>9</sub>O<sub>33</sub>)<sub>2</sub>]<sup>6-</sup> where M = Ga<sup>3+</sup> or Al<sup>3+</sup>; M<sub>4</sub>(SbW<sub>9</sub>)<sub>2</sub>), containing four metallic centers with available coordination sites, also showed hydrolytic activity towards NPP (Fig. 9).<sup>167</sup> However, due to the poor solubility of these Krebs M-POMs, the mechanism of action could not be investigated.

The reaction mechanism for the hydrolysis of NPP, BNPP, and HPNP by M-POMs was investigated in more detail for the dimeric Keggin M-POM with two adjacent Zr<sup>4+</sup>, Zr-K 2:2, due to its higher solubility (Fig. 9).<sup>143,168</sup> Hydrolysis of BNPP was 5 times faster with Zr-K 2:2 in comparison to Zr-K 1:2 (Table 3) due to the presence of more Zr<sup>4+</sup> centers with free coordination sites in Zr-K 2:2 that allow for easier nucleophilic activation. However, the monomeric Zr-K 1:1, which is in

equilibrium with both Zr-K 1:2 and Zr-K 2:2, was proposed to be the catalytically active form, in agreement with the results for peptide bond hydrolysis. The reaction mechanism for the hydrolysis of BNPP involves monodentate coordination of BNPP to Zr-K 1:1 followed by intramolecular nucleophilic attack of the phosphate group by an OH group coordinated to the Zr<sup>4+</sup> center, which leads to the release of *p*-nitrophenol (NP) and the formation of NPP (Fig. 9(b)). The formed NPP can also coordinate to Zr-K 1:1 and undergo hydrolysis *via* nucleophilic attack of a solvent water molecule. A similar reaction mechanism was also suggested for the hydrolysis of HPNP catalyzed by Zr-K 2:2.<sup>168</sup>

The reactivity of the dimeric Ce<sup>4+</sup> Keggin M-POM ([Ce{α-PW<sub>11</sub>O<sub>39</sub>}<sub>2</sub>]<sup>10-</sup>; Ce-K 1:2) towards NPP was also investigated *via* NMR experiments and DFT calculations to determine the influence of the embedded metal ion on the hydrolysis reaction (Fig. 9).<sup>168,169</sup> The monomeric Ce-K 1:1 species was suggested to be responsible for the hydrolytic activity in a comparable manner to what was established for Zr-POMs, even though Ce<sup>4+</sup> has a higher coordination number than Zr<sup>4+</sup> and Ce-K 1:2 is not coordinatively saturated. Due to the higher coordination number of Ce<sup>4+</sup>, two NPP molecules were proposed to coordinate to Ce<sup>4+</sup> at once, while only one coordinated NPP substrate was proposed for Zr<sup>4+</sup>. However, it was determined that only one of the coordinated NPP molecules can be hydrolyzed at a time during the reaction process. Therefore, changing the metal center to Ce<sup>4+</sup> did not have a major effect on the mechanism.

Based on the understanding gained from the studies on the model systems, the hydrolysis of supercoiled plasmid pUC19 DNA catalyzed by Zr-K 2:2 at physiological pH and temperature was also investigated.<sup>170</sup> Hydrolysis of one strand of DNA led to the relaxed form while hydrolysis at both strands was shown to result in the linear form (Fig. 10). Hydrolysis of calf thymus DNA (ctDNA) with the same M-POM was also attempted, but characterization of the fragmentation products was not successful since ctDNA is a complex mixture of linear

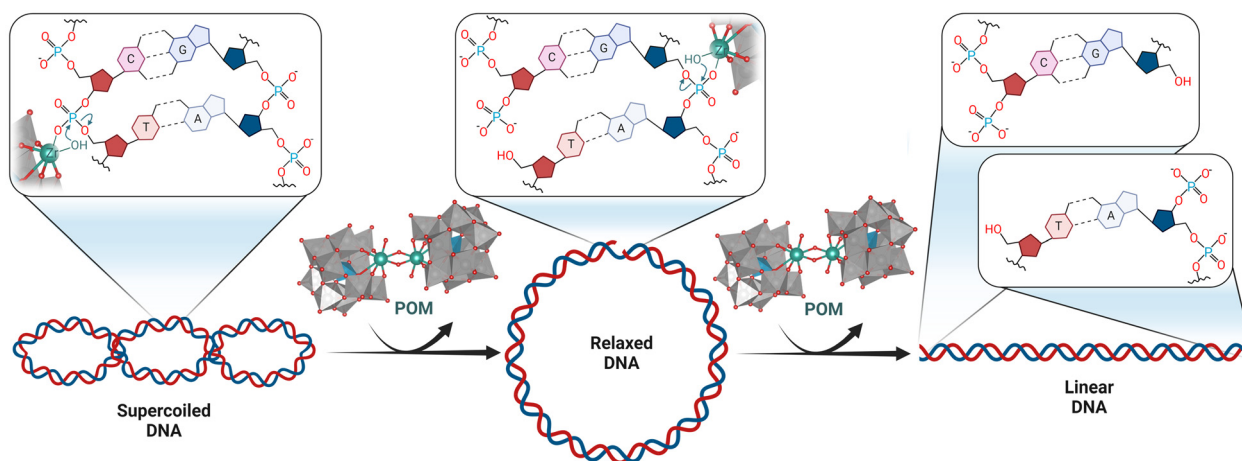


Fig. 10 Schematic representation of the proposed mechanism for phosphoester bond hydrolysis of supercoiled plasmid pUC19 DNA catalyzed by Zr-K 2:2, resulting in the relaxed form after cleavage of one strand of DNA and in the linear form after cleavage of both strands.



DNA fragments of different lengths.<sup>170</sup> To date, this remains the only reported study specifically on the hydrolysis of DNA catalyzed by POMs and no reports on the hydrolysis of RNA promoted by POMs have been published so far. Nevertheless, the antitumor activity of cyclopentadienyltitanium substituted Keggin M-POMs was proposed to be potentially linked to DNA hydrolysis, since agarose gel electrophoresis indicated that fragmentation of plasmid DNA occurred upon incubation with  $K_4H_3[(CpTi)_3SiW_9O_{37}]$ .<sup>171</sup> However, to the extent of our knowledge, this was not investigated further and the nature of the catalytically active species has not been confirmed since the stability of this M-POM after incubation with DNA was not reported. Therefore, the insights gained from the model systems still need to be implemented more extensively in the design of effective catalysts for the controlled hydrolysis of nucleic acids. This is essential in order to bridge the gap between simple model systems and more complex biomolecules, and, in doing so, gain a deeper understanding of the biological activity of POMs from a molecular point of view. Furthermore, this is also necessary for the development of POMs as genomic tools or therapeutic agents, with potential impact in biomedical sciences, biotechnology, and pharmacology.

**2.2.2. Hydrolysis of nucleotides.** Adenosine triphosphate (ATP) is an essential molecule in living organisms that provides the energy needed to drive many of the biochemical processes of life.<sup>143</sup> This transfer of energy is achieved through the energy released from the hydrolysis of the phosphoanhydride bond within the triphosphate group of ATP, to form adenosine diphosphate (ADP), which can be hydrolyzed further to form adenosine monophosphate (AMP) as shown in Fig. 11. In addition, ATP serves as a precursor for other important biomolecules such as RNA and DNA.<sup>172</sup> Furthermore, ATP is also a therapeutic target in diseases such as cancer which are known to result in consumption of ATP at a much higher unregulated rate than normal.<sup>173</sup> Therefore, catalysts that can hydrolyze ATP are of interest for biomedical applications.

As with phosphoester bond hydrolysis, initial studies into the hydrolysis of the phosphoanhydride bond catalyzed by POMs were in part inspired by the antitumor activity of  $Mo_7$ , which has also been attributed to the ability of molybdates to inhibit the formation of ATP.<sup>150,163</sup> At pH 6 and 4, ATP was hydrolyzed by  $Mo_7$  into ADP as the main product, whereas AMP was obtained as the major species at pH 2 and no hydrolysis was observed at pH 14.<sup>162</sup> The reaction mechanism was proposed to involve the incorporation of the phosphate groups into an oxomolybdate structure, resulting in different phosphomolybdate intermediate species as well as  $P_2Mo_5$ ,  $[(O_3POPO_3)Mo_6O_{18}(H_2O)_4]^{4-}$  ( $P_2Mo_6$ ), and Keggin-type phosphomolybdates as side-products.  $P_2Mo_5$  was mainly observed as a side product of the reactions where ADP was the major product, while only  $P_2Mo_6$  formed at lower pH when AMP was the main product. Hence, it was proposed that binding of ATP to a species similar to  $P_2Mo_5$  results in ADP while binding of ATP to a  $P_2Mo_6$ -like species results in the direct formation of AMP, with the speciation being determined by the pH. Interestingly, unlike in phosphoester bond hydrolysis, ATP

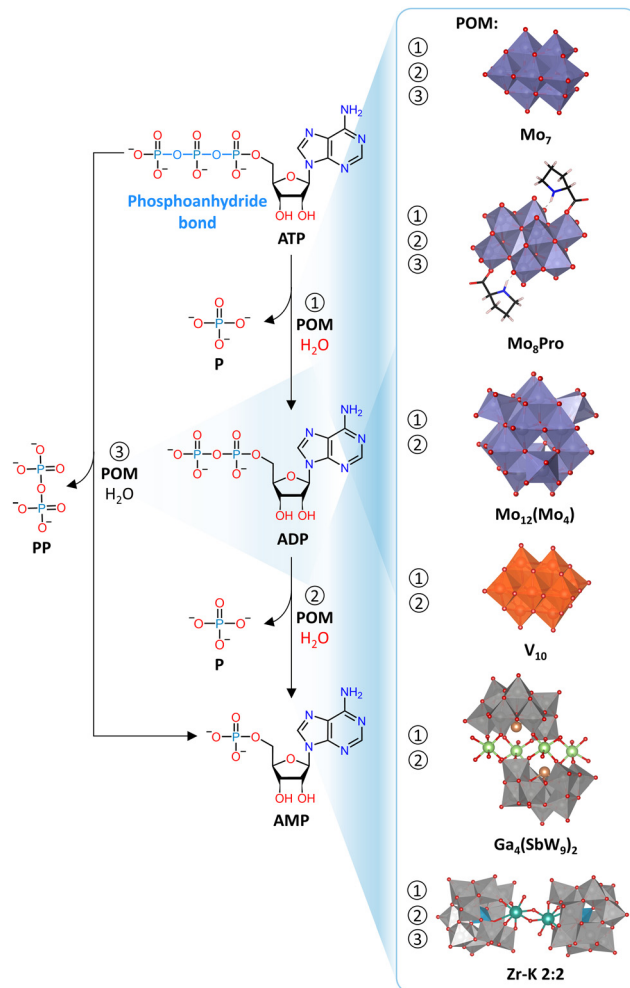


Fig. 11 Hydrolysis of the phosphoanhydride bond to form ADP from ATP (1) as well as AMP stepwise from ADP (1–2) or directly from ATP (3) catalyzed by  $Mo_7$ ,  $Mo_8Pro$ ,  $Mo_{12}(Mo_4)$ ,  $V_{10}$ ,  $Ga_4(SbW_9)_2$ , or  $Zr-K\ 2:2$ .

hydrolysis was observed in the presence of excess  $P_2Mo_5$ . Therefore, the speciation in solution should be further investigated to determine the reaction mechanism, which may differ from phosphoester bond hydrolysis.

Since octamolybdate POMs functionalized with amino acids have been shown to inhibit the growth of cancer cell lines,<sup>147</sup> the hydrolysis of ATP catalyzed by  $\gamma$ -octamolybdate covalently functionalized with two proline moieties ( $[Mo_8O_{24}(OCCC_4H_8N)_2]^{4-}$ ;  $Mo_8Pro$ ) was also investigated (Fig. 11).<sup>163</sup> A similar pH dependence and intermediate speciation to that observed with  $Mo_7$  was reported, with ADP and AMP being formed in equal amounts at pH 3.4, while ADP was favored at higher pH. This was linked to a structural rearrangement of  $Mo_8Pro$  to form  $Mo_7$  when the pH was increased to 5.7 and its decomposition into  $[MoO_4]^{2-}$  at physiological pH. Nevertheless, even at physiological pH hydrolysis of ca. 75% of ATP into ADP was still observed but at a lower rate (Table 3), which was suggested to be due to the small amount of the (rearranged)  $Mo_8Pro$  still present at this pH since  $[MoO_4]^{2-}$  is hydrolytically inactive. However, after 24 h at pH 3.4,  $[Mo_8O_{26}]^{4-}$  fully converted ATP to AMP while with  $Mo_8Pro$  significant



amounts of ADP were still present, suggesting that the proline functionality hinders the reactivity in this case. Nevertheless, this is one of the very few reported examples of the reactivity of an organic–inorganic hybrid POM towards a biomolecule. Therefore, further research should be performed to determine the role of the organic functionality and, in doing so, optimize the reactivity.

Another polyoxomolybdate with anticancer activity that has also been investigated as a catalyst for ATP hydrolysis is  $[\text{H}_2\text{Mo}^{5+}_{12}\text{O}_{28}(\text{OH})_{12}(\text{Mo}^{4+}\text{O}_3)_4]^{6-}$  ( $\text{Mo}_{12}(\text{Mo}_4)$ ), which consists of a 12-electron reduced  $\varepsilon$ -Keggin core capped at four triangular faces with  $\{\text{Mo}^{4+}\text{O}_3\}$  moieties and can be obtained from  $\text{Mo}_7$  by photoreduction (Fig. 11).<sup>164</sup> At 40 °C and pH 5 or pH 7.5,  $\text{Mo}_{12}(\text{Mo}_4)$  hydrolyzed ATP into ADP as the major product while producing AMP and  $\text{P}_2\text{Mo}_5$  as minor products, which remained present in low concentrations even after 10 days without any other byproducts. Furthermore, the rate of hydrolysis was faster at pH 5 than at pH 7.5 (Table 3). However, the rate of hydrolysis of ADP was much slower than for ATP, with only 26% conversion after 10 days at 40 °C (pH 5), and there was barely any hydrolysis of AMP, demonstrating the selectivity of  $\text{Mo}_{12}(\text{Mo}_4)$  for phosphoanhydride bond hydrolysis of ATP. Interestingly, the rate of hydrolysis was found to increase in the presence of  $\text{K}^+$  ions, which can shield repulsive interactions between the two negatively charged species (Table 3). In addition, the POM core remained relatively unchanged throughout the reaction as only partial release of  $\{\text{Mo}^{4+}\text{O}_3\}$  moieties was detected, which suggests that a  $\text{P}_2\text{Mo}_5$ -like species is not formed as an intermediate and the small amounts of  $\text{P}_2\text{Mo}_5$  are formed from the released  $\{\text{Mo}^{4+}\text{O}_3\}$  moieties. However, the exact mechanism has not been investigated further. Like for  $\text{Mo}_8\text{Pro}$ , this study stands out because it is one of the very few examples of the use of a reduced POM to promote biomolecular transformations and, therefore, incites further study of the reactivity of reduced POMs.

In addition to molybdates, several other POMs that were investigated as catalysts for phosphoester bond hydrolysis have been shown to be catalytically active towards ATP. For instance, the hydrolytic activity of vanadates has been recently reported.<sup>165</sup> In contrast to the results obtained for molybdates, the hydrolysis of ATP into ADP and AMP was seen to occur in succession, with AMP being formed from the hydrolysis of ADP rather than directly from ATP.<sup>165</sup> The mechanism was proposed to also involve complexation of the phosphate substrate to form an intermediate phosphovanadate species of unknown structure and composition. Furthermore, the rate of hydrolysis was highest at pH 2 (Table 3), where protonated  $\text{V}_{10}$  (Fig. 11) is the main species in solution. Therefore, it was proposed that  $\text{V}_{10}$  is more active than other vanadate species present at higher pH due to the lower negative charge of protonated  $\text{V}_{10}$  ( $[\text{H}_3\text{V}_{10}\text{O}_{28}]^{3-}$ ), resulting in less electrostatic repulsion towards ATP, as well as the higher kinetic lability of the oxo ligands of vanadates at lower pH.

M-POMs have also been reported to hydrolyze ATP. The Krebs-type M-POM  $\text{Ga}_4(\text{SbW}_9)_2$  was shown to catalyze the hydrolysis of ATP in a stepwise manner to produce ADP and then AMP (Fig. 11).<sup>167</sup> However, although NMR indicated that

complexation of the terminal phosphate of ATP to the M-POM likely occurred, the exact mechanism and the intermediate species formed are not known. A more detailed investigation into the mechanism was performed for the hydrolysis of ATP catalyzed by  $\text{Zr-K } 2:2$  (Fig. 11).<sup>144</sup> It was proposed that  $\text{Zr}^{4+}$  coordinates to either the terminal or the central phosphate groups of ATP, thereby activating them towards nucleophilic attack by coordinated  $\text{OH}^-$  or free  $\text{H}_2\text{O}$ . Hence, either stepwise formation of ADP and then AMP or direct formation of AMP could take place depending on where  $\text{Zr}^{4+}$  coordinates. Interestingly, phosphoester bond cleavage in AMP that would result in the formation of free triphosphate was not observed. Therefore, this illustrates the powerful selective nature of M-POM catalysts.

Overall, these studies provide valuable insights into the interactions and reactivity of POMs towards ATP. However, they mainly rely on NMR spectroscopy to obtain mechanistic insights. Therefore, additional studies employing complementary techniques, such as ESI-MS, and theoretical studies are necessary in order to gain a deeper understanding of how POMs catalyze the hydrolysis of ATP and how the selectivity of hydrolysis can be controlled.

### 2.3. Carbohydrates

Polysaccharides are polymeric chains of monosaccharides that are linked together *via* glycosidic bonds (Fig. 12) and are used by organisms for energy storage (*e.g.*, starch) or as structural elements (*e.g.*, cellulose).<sup>174</sup> Since polysaccharides represent a major component of biomass, their breakdown into monosaccharides, such as glucose, is the first step in the conversion of biomass into renewable and sustainable fuels.<sup>175,176</sup> This is an important type of reactivity to address since the environmentally damaging effects of the overconsumption of fossil fuels have led to the exploration of sustainable alternatives, and the conversion of biomass is a particularly attractive option because it is the only renewable carbon reservoir for the

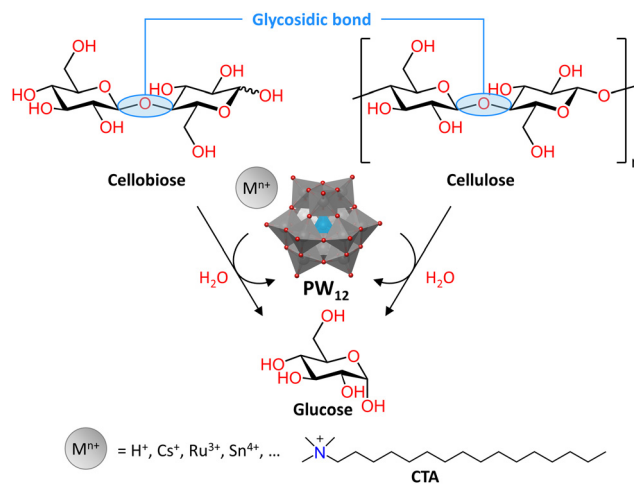


Fig. 12 Hydrolysis of the glycosidic bond in cellobiose and cellulose into glucose catalyzed by  $(\text{M}^{n+})_3/[\text{PW}_{12}\text{O}_{40}]^{3-}$  ( $\text{PW}_{12}$ ) with different counter-cations.





production of fuels and chemicals.<sup>177</sup> However, commonly employed techniques for biomass valorization (*i.e.*, thermochemical, enzymatic, and bacterial degradation) suffer from several drawbacks.<sup>178–181</sup> Therefore, the development of efficient catalysts is critical for the synthesis of biofuels as well as bulk and commodity chemicals, altogether contributing to the development of a sustainable chemical industry.

**2.3.1. Hydrolysis of biomass sugars.** Cleavage of glycosidic bonds in the polysaccharides that compose cellulosic biomass is typically achieved by employing mineral acids or natural enzymes.<sup>182–184</sup> However, both methods have disadvantages. Controlling the selectivity, rate, and efficiency of the hydrolysis reaction in the presence of mineral acids is very challenging, and the subsequent separation of the obtained products is difficult.<sup>176,185,186</sup> Moreover, corrosion of the reactor and high equipment cost associated with using mineral acids, together with waste disposal problems, represent major obstacles.<sup>185</sup> Additionally, such processes require very high pressures and temperatures (100–300 °C), which can result in side reactions and degradation of the obtained products.<sup>176,185,186</sup> Enzymatic processes, on the other hand, are less harmful and operate under very mild reaction conditions, but the high cost of the currently used enzymes and their limited recyclability, stability, and substrate selectivity hinder their use in industry.<sup>176,186</sup> Therefore, there is a great need for developing more environmentally conscious and cost-effective biomass conversion catalysts that can operate under mild reaction conditions.

Heteropolyacids (HPAs), a subclass of protonated POMs, have been investigated as homogeneous and heterogeneous catalysts due to their potential reusability, strong Brønsted acidity, proton mobility, and excellent stability at high temperatures, making them potentially very effective catalysts for biomass conversion, as has been recently reviewed in detail.<sup>175,187–189</sup> For instance, the Keggin-type HPAs  $H_3[PW_{12}O_{40}]$  and  $H_4[SiW_{12}O_{40}]$  were shown to catalyze the selective hydrolysis of cellobiose – a glucose disaccharide model for cellulose – as well as the more complex polysaccharides cellulose and lignocellulose (Fig. 12).<sup>175,185</sup> These studies showed that HPAs give better yields than traditional mineral acids due to their higher Brønsted acidity, with  $H_3[PW_{12}O_{40}]$  displaying slightly better reactivity than  $H_4[SiW_{12}O_{40}]$  (Table 4).<sup>185</sup> Brønsted acidity is needed for the hydrolysis of carbohydrates since the reaction mechanism proceeds first *via* protonation of the ether oxygen in the glycosidic bond, making it a good leaving group. This is in contrast to the hydrolysis of peptides/proteins, nucleic acids, and nucleotides where the mechanism involves Lewis acidic activation of C=O or P=O bonds towards nucleophilic attack. As a result, HPAs rather than M-POMs have been predominantly investigated for the hydrolysis of sugars.

In order to optimize the hydrolytic activity of  $H_3[PW_{12}O_{40}]$ , different parameters have been investigated, such as the catalyst loading, reaction time, and temperature. In doing so, better selectivity was observed at elevated temperatures, which could be exploited due to the high hydrothermal stability of the HPA.<sup>175</sup> However, extended hydrolysis at higher temperatures

resulted in reduced glucose yield due to the formation of by-products.<sup>175,202</sup> Additionally, due to their high thermal stability and high Brønsted acidity, HPAs ( $H_3[PW_{12}O_{40}]$ ,  $H_3[PMo_{12}O_{40}]$ ,  $H_4[SiW_{12}O_{40}]$ ,  $H_4[SiMo_{12}O_{40}]$ ,  $H_4[PVMO_{11}O_{40}]$ ,  $H_5[PV_2Mo_{10}O_{40}]$ ) were also shown to be suitable catalysts for the thermal decomposition of lignocellulosic biomass in pine wood as they lowered the required temperature by 100 °C.<sup>203</sup> Furthermore, the reusability of  $H_3[PW_{12}O_{40}]$  through extraction with diethyl ether or recrystallization was also reported.<sup>175</sup> More recently, it was found that microwave-assisted conversion of cellulose to glucose can be performed in the presence of a Wells–Dawson HPA,  $H_6[P_2W_{18}O_{60}]$ .<sup>190</sup> However, Keggin-type HPAs have shown better performance (Table 4).<sup>190</sup> Therefore,  $H_3[PW_{12}O_{40}]$  has been identified as a promising catalyst for biomass conversion.

The reactivity and selectivity of  $H_3[PW_{12}O_{40}]$  can be tuned through the replacement of the protons by counter-cations. A systematic study of 11 different metal counter-cations ( $M_{3/n}[PW_{12}O_{40}]$ ;  $M^{n+} = Ag^+$ ,  $Ca^{2+}$ ,  $Co^{2+}$ ,  $Y^{3+}$ ,  $Sn^{4+}$ ,  $Sc^{3+}$ ,  $Ru^{3+}$ ,  $Fe^{3+}$ ,  $Hf^{4+}$ ,  $Ga^{3+}$ , and  $Al^{3+}$ ) established that the Lewis acidity of the counter-cation has an influence on the hydrolytic activity of the POM towards cellobiose and cellulose. POMs with counter-cations having moderate Lewis acidities, such as  $Sn^{4+}$  and  $Ru^{3+}$ , gave higher conversion rates and even better turnover frequencies for the formation of glucose compared to  $H_3[PW_{12}O_{40}]$ .<sup>185</sup> Furthermore, in order to develop HPAs as suitable heterogeneous catalysts with a high-surface area, the hydrolytic activity of microporous and insoluble  $Cs^+$  salts of  $H_3[PW_{12}O_{40}]$  ( $Cs_xH_{3-x}[PW_{12}O_{40}]$ ;  $x = 1–3$ ) towards cellulose has also been investigated.<sup>192,204</sup> By varying the number of available surface acid sites through the partial substitution of protons by  $Cs^+$ ,  $Cs_1H_2[PW_{12}O_{40}]$  was shown to have the best catalytic activity due to its higher Brønsted acidity (Table 4). Contrastingly,  $Cs_{2.2}H_{0.8}[PW_{12}O_{40}]$  showed the highest selectivity, which was attributed to its microporous structure.<sup>192</sup> Similar results were reported for the hydrolytic activity of  $H_3[PW_{12}O_{40}]$  with the cationic surfactant cetyltrimethylammonium ( $C_{16}H_{33}N(CH_3)_3^+$ ; CTA), which forms a supramolecular micellar assembly with the POM that was shown to efficiently and selectively hydrolyze sucrose, starch, and cellulose.<sup>193</sup> By varying the ratio of protons to CTA ( $(CTA)_xH_{3-x}[PW_{12}O_{40}]$ ), the conversion and selectivity of the hydrolysis of the disaccharide sucrose into glucose and fructose was found to be highest for  $(CTA)_1H_2[PW_{12}O_{40}]$  due to its greater Brønsted acidity (Table 4). Furthermore,  $(CTA)_1H_2[PW_{12}O_{40}]$  produced relatively higher yields of the monosaccharides compared to  $H_3[PW_{12}O_{40}]$ , which was attributed to the accumulation of sucrose around the POM due to the formation of micelles. Moreover, the micellar catalyst could be removed through centrifugation and reused without any significant loss of catalytic activity. These reports are noteworthy since, although the hydrolytic activity is mainly accredited to the Brønsted acidity of the HPA, they tackle reaction optimization through the structure–activity relationship of the counter-cation rather than that of the POM framework itself. This is in stark contrast to many of the previously mentioned studies on the reactivity of POMs towards biomolecules, where the role of the counter-cation has been scarcely discussed.



Table 4 Selected conditions and percentage conversion of various sugars catalyzed by different POMs

| POM  | Substrate      | Reactivity                              | Reaction conditions                                     | Conversion/%     | Ref. |
|--|----------------|---|---|------------------|------|
| H <sub>3</sub> [PW <sub>12</sub> O <sub>40</sub> ]   | Cellulose      | Hydrolysis                              | H <sub>2</sub> O, 180 °C, 2 h, 0.08 mmol POM            | 51% glucose      | 175  |
|  |                |   | H <sub>2</sub> O, 120 °C, 24 h, 6 mol% POM              | 18               | 185  |
|  |                |   | H <sub>2</sub> O, 160 °C, 0.25 h, 0.14 mmol POM         | 90 (77% glucose) | 190  |
| H <sub>4</sub> [SiW <sub>12</sub> O <sub>40</sub> ]  | Cellobiose     | Hydrolysis                              | H <sub>2</sub> O, 120/150 °C, 24 h, 0.07 mmol POM       | 53 (51% glucose) | 185  |
|  | Lignocellulose | Hydrolysis                              | H <sub>2</sub> O, 120/150 °C, 24 h, 6 mol% POM          | 39               | 185  |
|  | Cellobiose,    | Hydrolysis                              | H <sub>2</sub> O, 120 °C, 24 h, 0.07 mmol POM           | 61 (53% glucose) | 185  |
|  | Cellulose      | Hydrolysis                              | H <sub>2</sub> O, 120 °C, 24 h, 6 mol% POM              | 17               | 185  |
|  | Chitin         | Hydrolysis, isomerization & dehydration | DMSO:H <sub>2</sub> O (1:3), 200 °C, 3 min 0.2 mmol POM | 23% HMF          | 191  |
| H <sub>6</sub> [P <sub>2</sub> W <sub>18</sub> O <sub>60</sub> ]<br>Cs <sub>1</sub> H <sub>2</sub> [PW <sub>12</sub> O <sub>40</sub> ] | Cellulose      | Hydrolysis                              | H <sub>2</sub> O, 160 °C, 5 min, 0.1 mmol POM           | 47 (37% glucose) | 190  |
|  | Cellulose      | Hydrolysis, isomerization & dehydration | H <sub>2</sub> O, 160 °C, 6 h, 0.06 mmol POM            | 27% glucose      | 192  |
| (CTA) <sub>1</sub> H <sub>2</sub> [PW <sub>12</sub> O <sub>40</sub> ]  | Sucrose        | Hydrolysis, isomerization & dehydration | H <sub>2</sub> O, 80 °C, 1 h, 0.08 mmol POM             | 100              | 193  |
|  | Starch         | Hydrolysis, isomerization & dehydration | H <sub>2</sub> O, 120 °C, 5 h, 0.07 mmol POM            | 96 (82% glucose) | 193  |
|  | Cellulose      | Hydrolysis, isomerization & dehydration | H <sub>2</sub> O, 170 °C, 8 h, 0.07 mmol POM            | 44               | 193  |
| Cr((DS)H <sub>2</sub> [PW <sub>12</sub> O <sub>40</sub> ]) <sub>3</sub>  | Cellulose      | Hydrolysis, isomerization & dehydration | H <sub>2</sub> O, 150 °C, 2 h, 0.06 mmol POM            | 77 (53% HMF)     | 194  |
| Sn <sub>0.1</sub> H <sub>2.6</sub> [PW <sub>12</sub> O <sub>40</sub> ]   | Cassava starch | Hydrolysis, isomerization & dehydration | DMSO/THF/NaCl, 160 °C, 1 h, 0.1 mmol POM                | 91 (54% HMF)     | 195  |
| KH <sub>3</sub> [CrPW <sub>11</sub> O <sub>39</sub> ]  | Glucose        | Dehydration                             | H <sub>2</sub> O, 130 °C, 2 h, 0.1 mmol POM             | 76 (33% HMF)     | 196  |
|  | Fructose       | Dehydration                             | H <sub>2</sub> O, 130 °C, 1.5 h, 0.1 mmol POM           | 91 (33% HMF)     | 196  |
| (CTA)H <sub>3</sub> [CrPW <sub>11</sub> O <sub>39</sub> ]  | Glucose        | Dehydration                             | H <sub>2</sub> O, 130 °C, 2 h, 0.1 mmol POM             | 84 (42% HMF)     | 196  |
|  | Fructose       | Dehydration                             | H <sub>2</sub> O, 130 °C, 1.5 h, 0.1 mmol POM           | 90 (41% HMF)     | 196  |
| Fe[PW <sub>12</sub> O <sub>40</sub> ]  | Fructose       | Dehydration                             | DMSO, 120 °C, 2 h, 6.9 μmol POM, 0.1 MPa                | 100 (97% HMF)    | 197  |
| Cs <sub>2.5</sub> H <sub>0.5</sub> [PW <sub>12</sub> O <sub>40</sub> ]   | Fructose       | Dehydration                             | MIBK, 115 °C, 2 h, 0.45 mmol POM                        | 95 (78% HMF)     | 198  |
| Ag <sub>3</sub> [PW <sub>12</sub> O <sub>40</sub> ]  | Fructose       | Dehydration                             | H <sub>2</sub> O/MIBK, 120 °C, 1 h, 0.02 mmol POM       | 78% HMF          | 199  |
|  | Glucose        | Isomerization & dehydration             | H <sub>2</sub> O/MIBK, 130 °C, 4 h, 0.1 mmol POM        | 76% HMF          | 199  |
| [MimAM]H <sub>2</sub> [PW <sub>12</sub> O <sub>40</sub> ]  | Glucose        | Dehydration                             | THF/H <sub>2</sub> O/NaCl, 160 °C, 7.5 h, 30 μmol POM   | 100 (54% HMF)    | 200  |
| TBA <sub>6</sub> [P <sub>2</sub> W <sub>17</sub> -SO <sub>3</sub> H]   | Fructose       | Dehydration                             | 1,4-Dioxane, 100 °C, 2 h, 150 mg POM                    | 99% HMF          | 201  |

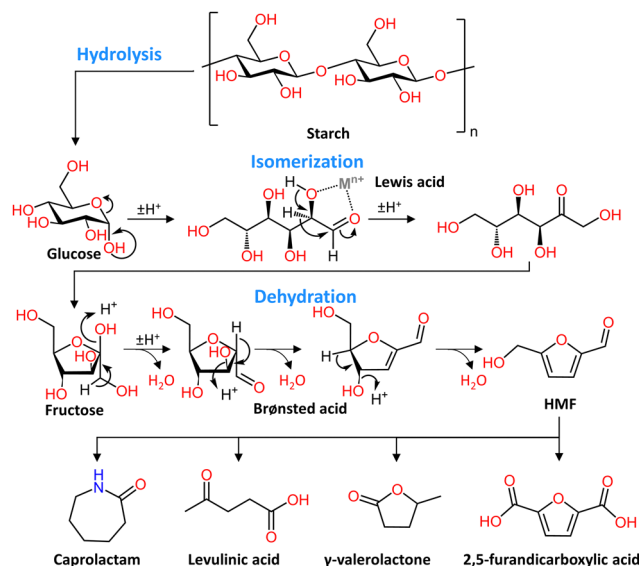
Hence, inspiration can be drawn from these results to apply the same strategies of systematic counter-cation variation to the optimization of reactivity in other areas.

**2.3.2. Dehydration of sugars into platform chemicals.** POMs have been shown to be effective catalysts not only in the hydrolysis of polysaccharides into monosaccharides but also for their conversion into platform chemicals such as 5-hydroxymethylfurfural (HMF). HMF is a useful, versatile, and key intermediate platform in the formation of industrially relevant value-added chemicals such as levulinic acid (LA),  $\gamma$ -valerolactone (GVL), caprolactam, adipic acid, and furan derivatives, such as 2,5-furandicarboxylic acid (Fig. 13). These value-added chemicals can in turn be used for manufacturing fuel additives, biofuels, polymers, and pharmaceuticals.<sup>191,193,195,204,205</sup> HMF can be obtained directly from fructose *via* an acid-catalyzed dehydration process. However, due to the high cost of fructose, glucose is a more attractive starting material for the synthesis of HMF due to its higher abundance in lignocellulosic biomass.<sup>205</sup> Hence, HMF can be obtained from cellulosic biomass through its hydrolysis into glucose, usually followed by isomerization to fructose,

which is further followed by dehydration (Fig. 13). HPAs can catalyze all these reactions allowing for the one-pot conversion of biomass into HMF.

Conversion of cellulose, as well as more complex mixtures of holocellulose and lignocellulose, into HMF under relatively mild conditions has been reported using H<sub>3</sub>[PW<sub>12</sub>O<sub>40</sub>] and a micellar HPA, Cr((OSO<sub>3</sub>C<sub>12</sub>H<sub>25</sub>)H<sub>2</sub>[PW<sub>12</sub>O<sub>40</sub>])<sub>3</sub> (Cr(DS)H<sub>2</sub>PW<sub>12</sub> where DS = dodecyl sulfate), which has both Brønsted and Lewis acidic sites due to the Cr<sup>3+</sup> counter-cation.<sup>194,203</sup> This synergistic combination of Brønsted and Lewis acidity is key for biomass conversion to HMF since Lewis acid sites favor the isomerization of glucose to fructose while the Brønsted acid sites promote the dehydration of fructose to HMF (Fig. 13). As discussed above, these micellar catalysts display enhanced reactivity due to the accumulation of the substrate onto the micellar catalyst. Therefore, Cr(DS)H<sub>2</sub>PW<sub>12</sub> promoted higher conversion into HMF than Cr(H<sub>3</sub>[PW<sub>12</sub>O<sub>40</sub>])<sub>3</sub>, Cr(OSO<sub>3</sub>C<sub>12</sub>H<sub>25</sub>)<sub>3</sub>, CrCl<sub>3</sub>, or H<sub>3</sub>[PW<sub>12</sub>O<sub>40</sub>] (Table 4). It also selectively produced HMF due to its Lewis acidity and micellar structure, as the decomposition of HMF into LA and furfural was slower compared to Cr(H<sub>3</sub>[PW<sub>12</sub>O<sub>40</sub>])<sub>3</sub>. Moreover, the catalyst was highly





**Fig. 13** Proposed mechanism for the conversion of cellulose to HMF via hydrolysis to glucose, which is then isomerized to fructose (catalyzed by a Lewis acid) that in turn then undergoes dehydration to HMF (catalyzed by a Brønsted acid). Some of the value-added chemicals that can be made from HMF are also shown at the bottom.

stable under the examined reaction conditions and could be reused for up to 6 reaction cycles with minimal leaching. In addition, HPAs generally lowered the decomposition temperature of pine wood by 100 °C for its conversion into furfural, thereby making the formation of feedstock chemicals from non-edible sources more accessible.<sup>203</sup>

More recently, it has been shown that HPAs also have promising catalytic activity for the conversion of other sources of biomass into HMF. Chitin, an abundant aminopolysaccharide composed of *N*-acetylglucosamines that is present in the exoskeleton of crustaceans, has been converted into HMF *via* a microwave-assisted process employing different Keggin HPAs ( $\text{H}_3[\text{PW}_{12}\text{O}_{40}]$ ,  $\text{H}_3[\text{PMo}_{12}\text{O}_{40}]$ ,  $\text{H}_4[\text{SiW}_{12}\text{O}_{40}]$ , and  $\text{H}_4[\text{SiMo}_{12}\text{O}_{40}]$ ) after pre-treatment of chitin with acid/base followed by ball milling to break the H-bond network and decrease its crystallinity.<sup>191</sup> Among all tested HPAs,  $\text{H}_4[\text{SiW}_{12}\text{O}_{40}]$  displayed the best catalytic performance (Table 4), which even surpassed all previously reported catalysts for chitin conversion into HMF. This was attributed to the greater softness of this HPA, which may enable the stabilization of organic intermediates. Cassava starch has also been transformed into HMF by using Brønsted–Lewis acid bifunctional catalysts ( $\text{Sn}_x\text{H}_{3-4x}[\text{PW}_{12}\text{O}_{40}]$ ,  $x = 0.10\text{--}0.75$ ).<sup>195</sup> The mechanism was proposed to first involve hydrolysis of starch to form glucose, followed by isomerization into fructose, which undergoes dehydration by elimination of three water molecules to form HMF (Fig. 13). Hydrolysis of starch to glucose and the dehydration of fructose to HMF are promoted by the Brønsted acidity of the POM while the isomerization of glucose to fructose is favored by the Lewis acidity of  $\text{Sn}^{4+}$ .  $\text{Sn}_{0.1}\text{H}_{2.6}[\text{PW}_{12}\text{O}_{40}]$  presented the best selectivity and HMF yield compared to  $\text{H}_3[\text{PW}_{12}\text{O}_{40}]$  and  $\text{Sn}_x\text{H}_{3-4x}[\text{PW}_{12}\text{O}_{40}]$  for  $x > 0.1$  due to the presence of Lewis acid sites without significantly

decreasing the Brønsted acidity. Therefore, these results show the tunability that can be achieved through changing the composition of POMs or the counter-cation.

Although biomass is a more accessible source of platform chemicals, the formation of HMF directly from fructose and glucose *via* dehydration catalyzed by heterogeneous and homogeneous POM catalysts has also been investigated. Similarly to the reports for biomass sources, this has been achieved using a micellar and Brønsted–Lewis acidic catalyst. More specifically, a  $\text{Cr}^{3+}$ -substituted Keggin M–POM with CTA as a counter-cation ( $(\text{CTA})_x\text{H}_{4-x}[\text{CrPW}_{11}\text{O}_{39}]$  where  $x = 1$  or 2) was prepared and characterized.<sup>196</sup> Although the incorporation of CTA as a counter-cation resulted in a decrease in the conversion of fructose, the selectivity towards the formation of HMF increased, which was proposed to be due to the formation of micelles preventing decomposition of HMF as previously discussed for  $\text{Cr}[(\text{DS})\text{H}_2\text{PW}_{12}\text{O}_{40}]$ . In addition, when starting from glucose, purely Brønsted acid catalysts, such as  $\text{H}_3[\text{PW}_{12}\text{O}_{40}]$ , typically produce LA and formic acid instead of HMF. However,  $\text{KH}_3[\text{CrPW}_{11}\text{O}_{39}]$  and  $(\text{CTA})\text{H}_3[\text{CrPW}_{11}\text{O}_{39}]$  produced HMF with yields of around 30% and 40%, respectively (Table 4).<sup>196</sup> This demonstrates how the reactivity can be tuned by embedding a Lewis acid metal center and using a cationic surfactant as counter-cation. Furthermore, these catalysts were active in water, while heterogeneous catalysts that have also been shown to catalyze the dehydration of fructose to HMF are typically less active in water due to undesired side reactions and poisoning of active sites.<sup>197–199</sup> However, high yields of HMF have also been reported using  $\text{Fe}[\text{PW}_{12}\text{O}_{40}]$  and  $\text{Cs}_{2.5}\text{H}_{0.5}[\text{PW}_{12}\text{O}_{40}]$  in dimethyl sulfoxide (DMSO) while removing water by continuous mild evacuation (Table 4).<sup>197</sup> Additionally, it was found that using a biphasic system of water and methyl isobutyl ketone (MIBK) significantly increased the conversion and selectivity for HMF catalyzed by  $\text{Cs}_{2.5}\text{H}_{0.5}[\text{PW}_{12}\text{O}_{40}]$  or  $\text{Ag}_3[\text{PW}_{12}\text{O}_{40}]$ , due to MIBK preventing decomposition of HMF (Table 4).<sup>198,199</sup>  $\text{Cs}_{2.5}\text{H}_{0.5}[\text{PW}_{12}\text{O}_{40}]$  in this biphasic system even gave a higher conversion and selectivity for HMF than other heterogeneous catalysts, such as Amberlyst-15, sulfated zirconia, and zeolites.<sup>198,199</sup> In addition, all of the above-mentioned catalysts could be recycled and their catalytic activity was generally preserved over several cycles.

The incorporation of ionic liquid (IL) cations with HPAs has also been explored to produce heterogeneous catalysts for the formation of HMF from sugars. Such catalysts can be obtained by simply combining HPAs with an IL cation. This has mainly been reported for HPAs with  $\text{SO}_3\text{H}$ -functionalized IL cations, such as 1-(3-sulfonic acid)propyl-3-methyl imidazolium ([MIMPS]) or poly(1-vinyl-3-propane sulfonate imidazolium) (poly(VMPS)), which have been demonstrated to act as effective heterogeneous catalysts for the dehydration of fructose to HMF.<sup>202,206,207</sup> Furthermore, HPAs with  $\text{SO}_3\text{H}$ -functionalized IL cations have been shown to catalyze the one-pot formation of HMF as well as levulinic acid and alkyl levulinates, which are also important platform chemicals.<sup>202,208,209</sup> This was achieved not only from fructose but also from sucrose and inulin, a polysaccharide typically obtained from chicory, as well as from





glucose, cellobiose, and cellulose. However, the conversion of glucose, cellobiose, and cellulose generally gave lower yields, which is likely due to the lack of Lewis acidic sites needed for the effective formation of HMF directly from glucose and its derivatives. A yield of only 26% was obtained for the conversion of glucose to HMF using  $\text{H}_3[\text{PW}_{12}\text{O}_{40}]$  and poly(VMPs) under optimized conditions.<sup>202,206</sup> However, conversion of glucose into HMF in good yields (Table 4) has been recently reported using HPA-IL catalysts formed through combining  $\text{H}_3[\text{PW}_{12}\text{O}_{40}]$  with the IL cation 1-aminoethyl-3-methylimidazolium ( $[\text{MimAM}]^+$ ):  $[\text{MimAM}]_x\text{H}_{3-x}[\text{PW}_{12}\text{O}_{40}]$  ( $x = 1-3$ ).<sup>200</sup> The reaction mechanism was proposed to involve (i) ring opening of glucose due to protonation of the bridging oxygen along with (ii) formation and stabilization of a 1,2-enediol intermediate followed by (iii) dehydration of the 1,2-enediol in a series of chain reactions *via* synergistic contributions of  $[\text{MimAM}]^+$  and the HPA (Fig. 14). Hence, the mechanism does not involve isomerization to fructose due to the formation of the HPA-stabilized 1,2-enediol intermediate. Interestingly, this suggests that amino-functionalized IL cations, such as  $[\text{MimAM}]^+$ , may serve as alternatives to using toxic Lewis acidic metal centers (*e.g.*,  $\text{Cr}^{3+}$ ). In addition, while  $[\text{MimAM}]\text{H}_2[\text{PW}_{12}\text{O}_{40}]$  displayed a slightly higher yield of HMF than  $\text{H}_3[\text{PW}_{12}\text{O}_{40}]$ , increasing the amount of  $[\text{MimAM}]^+$  counter-cations decreased the yield of HMF. This is due to the importance of the synergy between  $[\text{MimAM}]^+$  and the Brønsted acidity of the HPA, as evidenced by the low catalytic activity of  $[\text{MimAM}]\text{Br}$  and of the HPA with 1-ethyl-3-methylimidazolium ( $[\text{EMim}]\text{H}_2[\text{PW}_{12}\text{O}_{40}]$  where  $[\text{EMim}]^+$  lacks the amino functionality). Likewise, lower activity was observed for  $[\text{MimAM}]\text{H}_2[\text{PMo}_{12}\text{O}_{40}]$ ,  $[\text{MimAM}]\text{H}_3[\text{SiW}_{12}\text{O}_{40}]$ , and  $[\text{MimAM}]\text{H}_2[\text{PV}_2\text{Mo}_{10}\text{O}_{40}]$ . Furthermore, the catalytic activity decreased according to their Brønsted acid strength:  $\text{H}_3[\text{PW}_{12}\text{O}_{40}] > \text{H}_3[\text{PMo}_{12}\text{O}_{40}] > \text{H}_4[\text{SiW}_{12}\text{O}_{40}] > \text{H}_5[\text{PV}_2\text{Mo}_{10}\text{O}_{40}]$ . However, while a similar trend has also been

observed for the dehydration of fructose with different ILs, the influence of the structure of the IL cation has not been extensively investigated. Nevertheless, the differences in the reactivity of HPAs with  $\text{SO}_3\text{H}$ -functionalized IL cations *vs.* with amino-functionalization indicate a promising possible avenue for tuning the catalytic activity and selectivity of HPAs.

Phase transfer catalysts, which combine the benefits of both homogenous and heterogeneous catalysts, have also been recently proposed as an alternative to heterogeneous catalysts, which often suffer from reduced acidity with respect to homogeneous HPAs. Exceptionally, a tetrabutylammonium (TBA) salt of a Wells–Dawson POM covalently functionalized with sulfonic acid groups ( $\text{TBA}_6[\text{P}_2\text{W}_{17}\text{SO}_3\text{H}]$ ) has been recently reported as a phase transfer catalyst for the conversion of fructose to HMF with 99% yield after 2 h at 100 °C in 1,4-dioxane (Table 4).<sup>201</sup> It was shown to also catalyze the conversion of glucose, sucrose, and inulin into HMF in reasonable yields, but failed to convert cellulose. Moreover, this is one of the few examples where a covalently functionalized hybrid POM was used as a catalyst in biomolecular transformations. Hence, phase transfer catalysts and hybrid POMs should be investigated further to obtain better catalysts for biomass conversion.

**2.3.3. Hydrolysis of glycosidic bonds in glycoproteins.** As well as being important sources of energy and added-value chemicals, monosaccharides can also form shorter glycan chains that are typically attached to glycolipids or glycoproteins. As such, they serve as ligands in receptor-mediated intracellular interactions as part of several key processes such as cell recognition, adhesion, communication, proliferation, differentiation, and even viral infection.<sup>210</sup> Sugars known as sialic acids (Fig. 15) are often found at the terminal position of glycan chains, and they play particularly important roles in these biological processes.<sup>211</sup> Hence, sialic acids are used as biomarkers for several diseases, including cancer, and their

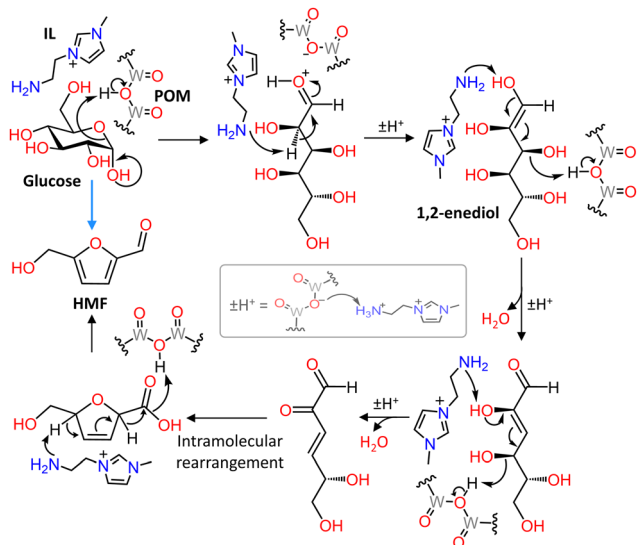


Fig. 14 Proposed mechanism for the conversion of glucose to HMF catalyzed by an HPA with an IL cation *via* formation of a 1,2-enediol intermediate that then undergoes dehydration to HMF through synergistic contributions of the IL cation and the HPA.

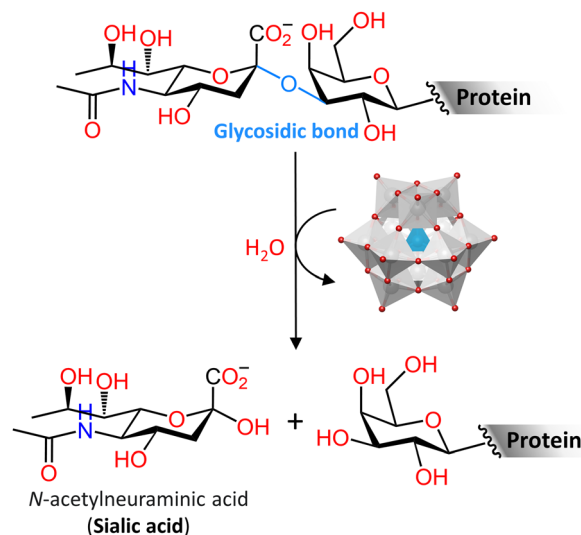
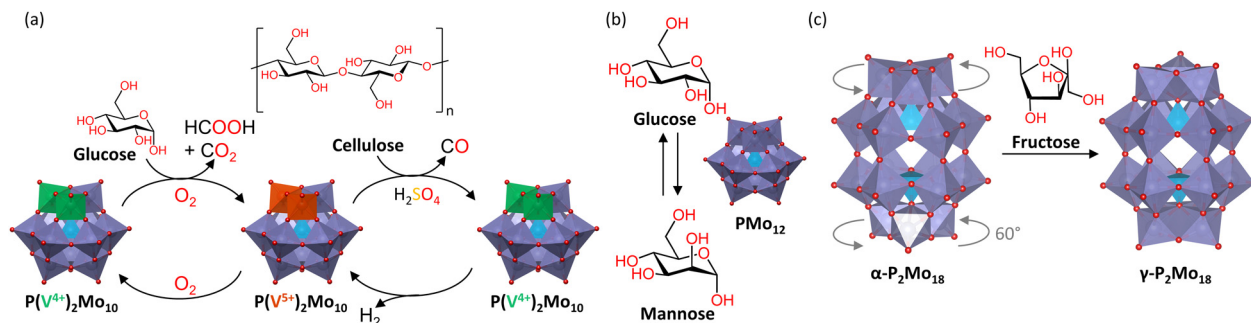


Fig. 15 Hydrolysis of the terminal glycosidic bond of an example glycan chain of a glycoprotein resulting in the release of a sialic acid catalyzed by  $\text{M}_3[\text{EA}_{12}\text{O}_{40}]^{3-}$  ( $\text{M} = \text{H}^+, \text{Na}^+, \text{or } \text{K}^+$ ;  $\text{E} = \text{P}^{5+} \text{ or } \text{Si}^{4+}$ ;  $\text{A} = \text{W}^{6+} \text{ or } \text{Mo}^{6+}$ ).





**Fig. 16** (a) Catalytic activity of  $\text{H}_5[\text{PV}_2\text{Mo}_{10}\text{O}_{40}]$  ( $\text{PV}_2\text{Mo}_{10}$ ) towards the decomposition of glucose into  $\text{HCOOH}$  and  $\text{CO}_2$  under an  $\text{O}_2$  pressurized atmosphere (left) or towards the decomposition of cellulose into  $\text{CO}$  and  $\text{H}_2$  (right). (b) Epimerization reaction between glucose and mannose catalyzed by  $[\text{PMo}_{12}\text{O}_{40}]^{3-}$  ( $\text{PMo}_{12}$ ). (c) Isomerization and reduction of  $[\alpha\text{-P}_2\text{Mo}_{18}\text{O}_{62}]^{6-}$  ( $\alpha\text{-P}_2\text{Mo}_{18}$ ) into 6-electron reduced  $[\gamma\text{-P}_2\text{Mo}_{18}\text{O}_{62}]^{12-}$  ( $\gamma\text{-P}_2\text{Mo}_{18}$ ), with its top and bottom  $\{\text{Mo}\}_3$  caps rotated by  $60^\circ$ , catalyzed by fructose.

inclusion in therapeutic agents has a large impact on their efficacy.<sup>210–212</sup> Therefore, being able to monitor the sialic acids on glycan chains is of crucial importance. This typically requires hydrolysis of the glycosidic bond using enzymes or acid hydrolysis, both of which face similar problems to those discussed for the hydrolysis of biomass polysaccharides.<sup>213</sup> Moreover, the less expensive acid hydrolysis method often leads to the destruction of the protein and the sialic acids, thereby complicating the interpretation of the obtained hydrosylates.<sup>213</sup> By leveraging the hydrolytic activity of HPAs, our research group recently explored the application of Keggin and Wells–Dawson type HPAs ( $\text{H}_3[\text{PW}_{12}\text{O}_{40}]$ ,  $\text{H}_4[\text{SiW}_{12}\text{O}_{40}]$ ,  $\text{H}_3[\text{PMo}_{12}\text{O}_{40}]$ ) and metal salts ( $\text{Na}_3[\text{PW}_{12}\text{O}_{40}]$ ,  $\text{Na}_6[\text{H}_2\text{W}_{12}\text{O}_{40}]$ ,  $\text{K}_6[\text{P}_2\text{W}_{18}\text{O}_{62}]$ ) towards the cleavage of glycosidic bonds in glycoproteins (Fig. 15). The release of sialic acids under mild reaction conditions was investigated on two glycoproteins with a high content of sialic acids:  $\alpha$ -2-HS-glycoprotein (fetuin) from bovine fetal serum and 1-acid glycoprotein (AGP) from bovine plasma.<sup>214,215</sup> The obtained results have shown that all investigated HPAs enable the selective release of sialic acids with minimal incubation time and protein destruction in comparison with conventional acids (*e.g.*,  $\text{HCl}$ ,  $\text{H}_2\text{SO}_4$ ,  $\text{CH}_3\text{COOH}$ ,  $\text{CF}_3\text{COOH}$ ) under the same reaction conditions. Although the composition, size, shape, and charge of the POM did not seem to have any major influence on the reactivity, their enhanced activity has been proposed to result from their negative charge which can stabilize the positively charged glycosyl cation and oxocarbenium ion intermediates.<sup>214</sup> Moreover, the structure of sialic acids during the hydrolytic process is preserved, which facilitates the structural analysis of sialic acids.<sup>214,215</sup> Hence, HPAs are promising catalysts that could facilitate the monitoring of sialic acids in glycoproteins for diagnostic and therapeutic applications. However, since only a limited number of studies have been performed so far, further research is required in order to determine the feasibility of their implementation and widespread use across different types of glycoproteins.

**2.3.4. Redox activity towards sugars.** The combined redox activity and Brønsted acidity of certain HPAs make them particularly suitable for the conversion of biomass into biofuels

and industrially relevant chemicals, such as formic acid.<sup>216</sup> For example,  $\text{H}_5[\text{PV}_2\text{Mo}_{10}\text{O}_{40}]$  has been shown to catalyze the selective decomposition of sugars (*e.g.*, glucose, cellobiose, sucrose, *etc.*) into formic acid and  $\text{CO}_2$  under a pressurized  $\text{O}_2$  atmosphere (Fig. 16(a)).<sup>217,218</sup> This has been proposed to take place *via* an electron transfer-oxygen transfer mechanism, whereby the HPA catalyzes the oxidative cleavage of carbon–carbon bonds in vicinal diols and primary alcohols through the insertion of oxygen.<sup>217,219</sup> The fully oxidized POM is then regenerated by  $\text{O}_2$ , which was proposed to be the rate-limiting step at low  $\text{O}_2$  pressures.<sup>220</sup> A systematic study of  $\text{V}^{5+}$ -substituted Keggin-type phosphomolybdate HPAs ( $\text{H}_{3+n}[\text{PV}_n\text{Mo}_{12-n}\text{O}_{40}]$  where  $n = 0\text{--}6$ ) showed that  $\text{H}_8[\text{PV}_5\text{Mo}_7\text{O}_{40}]$  had better selectivity for formic acid, which was attributed to the formation of  $\text{VO}_2^+$  with increasing  $\text{V}^{5+}$  centers.<sup>220</sup> In addition, when the decomposition of cellulose and hemicellulose was performed with  $\text{H}_5[\text{PV}_2\text{Mo}_{10}\text{O}_{40}]$  and  $\text{H}_2\text{SO}_4$  under anaerobic conditions,  $\text{HCOOH}$  was dehydrated into  $\text{CO}$  and the oxidized POM could be regenerated by electrolysis with the release of  $\text{H}_2$  (Fig. 16(a)).<sup>217</sup> The resulting mixture of  $\text{CO}$  and  $\text{H}_2$ , known as synthesis gas, can be used for the formation of hydrocarbon fuels as well as other compounds.<sup>217</sup> Nb- and Ta-substituted Keggin polyoxomolybdates have also been shown to catalyze the oxidation of glucose into formic acid as the major liquid-phase product, with their effective activation energy ( $55 \text{ kJ mol}^{-1}$ ) being lower compared to the non-substituted  $\text{H}_3[\text{PMo}_{12}\text{O}_{40}]$  ( $80 \text{ kJ mol}^{-1}$ ).<sup>216</sup> Therefore, these studies show that by varying the POM's composition and the reaction conditions, it is possible to optimize the reaction and even alter its outcome. However, although mechanistic studies have been performed, the mode of interaction of these POMs with sugars remains unclear.

In a different study, structurally related Keggin-type phosphomolybdates ( $\text{H}_3[\text{PMo}_{12}\text{O}_{40}]$ ,  $\text{Ag}_3[\text{PMo}_{12}\text{O}_{40}]$  and  $\text{Sn}_{0.75}[\text{PMo}_{12}\text{O}_{40}]$ ) were demonstrated to catalyze the epimerization of D-glucose to D-mannose (Fig. 16(b)), as well as the reverse reaction (at a slower rate), which is a relevant process in the production of certain pharmaceuticals.<sup>217</sup> This was proposed to occur due to binding of the sugar to molybdate, resulting in an electron transfer from glucose to the POM and a C1–C2 intramolecular carbon shift. The



counter-cation did not seem to have any influence on the reaction and no conversion was observed with the more inert isostructural phosphotungstate ( $\text{H}_3[\text{PW}_{12}\text{O}_{40}]$ ). However, the structure of the sugar-bound molybdate intermediate remains unknown and the role of the electron transfer event resulting in reduction of  $\text{Mo}^{6+}$  into  $\text{Mo}^{5+}$  is not well understood. Therefore, further research must be performed to shed light on this type of reactivity. Nevertheless, the use of  $^{13}\text{C}$ -labeled glucose in this study provided valuable insights into the reaction mechanism. This approach is noteworthy since it is not often used for investigating the reactivity of POMs towards biomolecules, but could potentially be applied to other systems to reveal additional mechanistic details.

In contrast to most of the reactions described in the literature, where the aim is the controlled modification of a biomolecule, the redox activity of sugars towards POMs has also been exploited to reduce and modify a POM structure. This is of interest because reduced POM species have been shown to have potential applications in several fields, but their controlled synthesis can be challenging.<sup>221</sup> Hence, fructose was used as a reducing agent in the selective conversion of  $[\alpha\text{-P}_2\text{Mo}_{18}\text{O}_{62}]^{6-}$ , the most stable isomer of the Wells–Dawson phosphomolybdate structure, into the less stable 6-electron reduced  $\gamma$ -isomer (Fig. 16(c)).<sup>222</sup> In contrast, glucose was less effective, which was ascribed to its weaker bidentate coordination to  $\text{Mo}^{6+}$ , while fructose can act as a tridentate ligand due to having three OH groups in close proximity that could promote better interaction and efficient reduction.<sup>222</sup> This binding is potentially responsible for the selectivity since a mixture of isomers was obtained with standard reducing agents. However, further studies are required to determine the exact role of the sugar and the nature of the sugar-bound intermediate. Furthermore, whether or not the sugars are transformed in the process was not reported, even though this reactivity suggests that phosphomolybdates may be of interest for the selective oxidation of sugars. Moreover, this study appears to be the other side of the coin to the previously discussed study on the epimerization of glucose and, therefore, highlights the importance of approaching the investigation of reactivity from both the point of view of the catalyst and that of the substrate.

## 2.4. Lipids

Lipids are a vast family of hydrophobic and amphiphilic biomolecules that, are typically used by living organisms as essential components of cell membranes, for signal transduction, and for energy storage as an alternative to carbohydrates.<sup>223</sup> Several studies have been reported on the interactions of POMs with lipid mono- and bi-layers in the context of the effect of POMs on cell membranes for biomedical applications, such as the development of antimicrobial drugs.<sup>224–228</sup> However, the reactivity of POMs towards lipids has mainly been investigated in the context of biodiesel production, which typically consists of fatty methyl or ethyl esters formed by transesterification of triglycerides in oils, such as vegetable oil and waste cooking oil, using mineral acids or bases.<sup>229–233</sup> Although base catalysts (e.g., KOH, NaOH,  $\text{NaOCH}_3$ ) are commonly used due to their higher

activity, they are corrosive and prone to form soaps if the oil contains a significant amount of water or free fatty acids, which leads to loss of catalyst and requires expensive isolation of the desired biodiesel products.<sup>234–236</sup> On the other hand, mineral acids (e.g., HCl,  $\text{H}_2\text{SO}_4$ ) can catalyze the transesterification of triglycerides and the esterification of free fatty acids simultaneously, but they are also highly corrosive and give rise to waste-disposal problems.<sup>234–236</sup> Therefore, alternative acid catalysts are needed to make the production of biodiesel more viable while also ensuring it is environmentally safe.

As previously discussed, the high Brønsted acidity and stability of HPAs make them good alternatives to mineral acids. Accordingly, it was shown that anhydrous Keggin-type HPAs ( $\text{H}_x[\text{EA}_{12}\text{O}_{40}]^{(n-x)-}$  where  $\text{E} = \text{P}^{5+}$  or  $\text{Si}^{4+}$  and  $\text{A} = \text{Mo}^{6+}$  or  $\text{W}^{6+}$ ) could catalyze the transesterification of rapeseed oil with methanol or ethanol resulting in better percentage conversion compared to  $\text{H}_2\text{SO}_4$  and  $\text{H}_3\text{PO}_4$  (Table 5).<sup>232</sup> Interestingly, greater triglyceride conversion was obtained with ethanol over methanol, and also when using anhydrous over hydrated HPAs, or when using polyoxomolybdates over polyoxotungstates (Table 5). The latter is contrary to the expected trend based on HPA acid strength, which is highest for  $\text{H}_3[\text{PW}_{12}\text{O}_{40}]$ , and this discrepancy was proposed to stem from the easier loss of crystallization waters from polyoxomolybdates as the presence of water affects the progress of the reaction. On the other hand, the transesterification of Macauba oil, a type of palm oil, was found to follow the expected trend according to the Brønsted acidity:  $\text{H}_3[\text{PW}_{12}\text{O}_{40}] > \text{H}_3[\text{PMo}_{12}\text{O}_{40}] > \text{H}_4[\text{SiW}_{12}\text{O}_{40}] > \text{H}_2\text{SO}_4$  (Table 5).<sup>237</sup> A more recent study has also shown that for the methanolic esterification of palmitic acid, one of the most commonly found saturated fatty acids in animal and vegetable oil,<sup>238</sup>  $\text{H}_5[\text{BW}_{12}\text{O}_{40}]$  was significantly more active as a catalyst than  $\text{H}_3[\text{PW}_{12}\text{O}_{40}]$  under the same conditions (Table 5).<sup>239</sup> This was attributed to the higher proton number and, therefore, higher Brønsted acidity of  $\text{H}_5[\text{BW}_{12}\text{O}_{40}]$ .<sup>239</sup> Nevertheless, the Keggin-type HPA  $\text{H}_3[\text{PW}_{12}\text{O}_{40}]$  has been most extensively investigated in the context of biodiesel production, which is likely due to both its relatively high Brønsted acidity and its commercial availability.<sup>239</sup> For instance, this HPA was shown to be an effective and recyclable catalyst for the methanolic and ethanolic esterification of palmitic acid and of other saturated and unsaturated fatty acids (e.g., myristic, stearic, oleic, and linoleic acids), resulting in similar or increased activity towards all fatty acids in comparison to  $\text{H}_2\text{SO}_4$  (Table 5).<sup>230,233,240</sup>  $\text{H}_3[\text{PW}_{12}\text{O}_{40}]$  was also shown to be active in the methanolic transesterification of triglycerides in waste cooking oil, which is particularly unsuitable for base-catalyzed transesterification due to its high content of free fatty acids and water.<sup>230,241,242</sup> In these esterification and transesterification reactions, HPAs have been proposed to catalyze the reaction through protonation of the ester carbonyl group, thereby activating it towards nucleophilic attack by an alcohol (Fig. 17).<sup>230,243</sup> In the case of triglycerides this proceeds in a stepwise manner *via* di- and mono-glycerides to produce fatty acid esters and glycerol (Fig. 17).<sup>230,243</sup> Hence, the high Brønsted



**Table 5** Selected conditions and percentage conversion reported for the esterification of free fatty acids (FA) and triglycerides (TG) catalyzed by different POMs

| POM  | Substrate  | Reaction conditions  | Conversion/%                                       | Ref.                      |
|--|--|--|--|---------------------------|
| H <sub>3</sub> [PW <sub>12</sub> O <sub>40</sub> ]                     | Palmitic acid  | 2.5 : 1 MeOH : FA, 65 °C, 12 h, 0.1 mmol POM                   | 97   | 230                       |
|  |  | 50 : 1 MeOH : FA, reflux, 10 h, 0.4 mmol POM                   | 97   | 233                       |
|  |  | 12 : 1 MeOH : FA, 90 °C, 3 h, 6 wt% POM                        | 87   | 229                       |
|  |  | 10 : 1 MeOH : FA, 65 °C, 3 h, 10.9 mM POM                      | 52   | 239                       |
|  |  | 20 : 1 MeOH : FA, 65 °C, 6 h, 243 : 1 FA : POM                 | 94   | 244                       |
|  | Myristic, stearic, oleic, linoleic acids<br>Waste cooking oil<br>Rapeseed oil  | 50 : 1 MeOH : FA, reflux, 10 h, 0.4 mmol POM                   | 93 (linoleic), 95 (oleic), 97 (myristic & stearic) | 233                       |
|  |  | 70 : 1 MeOH : oil, 65 °C, 14 h, 10 wt% POM                     | 87–89  | 230,241                   |
|  |  | MeOH, 80 °C, 5 h, 3 mol% POM,                                  | 11   | 232                       |
|  |  | 6 : 1 EtOH : oil, 85 °C, 3 h, 1.7 mol% POM                     | 27   |                           |
|  |  | 12 : 1 EtOH : oil, 90 °C, 8 h, 2 mol% POM                      | 68   | 237                       |
| H <sub>3</sub> [PMo <sub>12</sub> O <sub>40</sub> ]                    | Macauba oil  | 12 : 1 EtOH : oil, 90 °C, 8 h, 2 mol% POM                      | 65   | 232                       |
|  | Rapeseed oil   | 6 : 1 EtOH : oil, 85 °C, 3 h, 1.6 mol% POM                     | 55   | 237                       |
| H <sub>4</sub> [SiW <sub>12</sub> O <sub>40</sub> ]                    | Macauba oil  | 12 : 1 EtOH : oil, 90 °C, 8 h, 2 mol% POM                      | 65   | 232                       |
|  | Rapeseed oil   | 6 : 1 EtOH : oil, 85 °C, 3 h, 2 mol% POM                       | 20   | 237                       |
| H <sub>4</sub> [SiMo <sub>12</sub> O <sub>40</sub> ]                   | Macauba oil  | 12 : 1 EtOH : oil, 90 °C, 8 h, 2 mol% POM                      | 68   | 232                       |
|  | Rapeseed oil   | 6 : 1 EtOH : oil, 85 °C, 3 h, 2 mol% POM                       | 45   | 239                       |
| H <sub>5</sub> [BW <sub>12</sub> O <sub>40</sub> ]                     | Palmitic acid  | 10 : 1 MeOH : FA, 65 °C, 3 h, 10.9 mM POM                      | 99   | 244                       |
| H <sub>5</sub> [TiPW <sub>11</sub> O <sub>40</sub> ]                   | Palmitic acid  | 20 : 1 MeOH : FA, 65 °C, 6 h, 243 : 1 FA : POM                 | 94   | 239                       |
| [CTA]H <sub>4</sub> [BW <sub>12</sub> O <sub>40</sub> ]                | Palmitic acid  | 18 : 1 MeOH : FA, 65 °C, 5 h, 4.48 mM POM                      | 91   | 72                        |
|  |  | 18 : 1 EtOH : FA, 65 °C, 5 h, 4.48 mM POM                      | 91   |                           |
|  |  | 18 : 1 ButOH : FA, 65 °C, 5 h, 4.48 mM POM                     | 94   | 94 (stearic) & 97 (oleic) |
|  |  | 18 : 1 MeOH : FA, 65 °C, 5 h                                   | 95   | 244                       |
|  |  | 20 : 1 MeOH : FA, 65 °C, 6 h, 243 : 1 FA : POM                 | 95   | 229                       |
|  | Oleic & stearic acid<br>Palmitic acid<br>Palmitic acid   | 12 : 1 MeOH : FA, 90 °C, 3 h, 6 wt% POM                        | 81% (x = 3)–93% (x = 1) yield                      | 234                       |
|  |  | 30 : 1 MeOH : FA, 60 °C, 6 h, 50 mg POM                        | 9 (x = 3)–100 (x = 2.3)                            | 234                       |
|  |  | 30 : 1 MeOH : TG, 60 °C, 6 h, 50 mg POM                        | 3 (x = 0.9)–50 (x = 2.3)                           | 234                       |
|  |  | 30 : 1 MeOH : TG, 60 °C, 6 h, 50 mg POM (x = 2.3)              | 100 (FA) & 50 (tributyrin)                         | 234                       |
|  |  | 14 : 1 MeOH : FA, 65 °C, 3 h, 41 mg mL <sup>−1</sup> POM       | 2 (x = 3)–92 (x = 1)                               | 245                       |
| Cs <sub>x</sub> H <sub>3−x</sub> [PW <sub>12</sub> O <sub>40</sub> ]   | Palmitic acid<br>Tributyrin<br>Palmitic acid + tributyrin (50 : 1)<br>Rice bran FAs<br><i>Eruca sativa</i> Gars. oil | 5.3 : 1 MeOH : oil, 20 °C, 1 h, 20 μmol POM (x = 2.5)          | 40   | 235                       |
|  |  | 5.3 : 1 MeOH : oil, 55 °C, 0.75 h, 541 : 1 oil : POM (x = 2.5) | 99   |                           |
|  |  | 12 : 1 MeOH : oil, 60 °C, 0.2 h, 1 wt% POM (x = 2.5)           | 96% yield  | 246                       |
|  |  | 30 : 1 MeOH : oil, 140 °C, 8 h, 3 wt% POM (x = 2)              | 86   | 236                       |
|  |  | 30 : 1 MeOH : oil, 140 °C, 8 h, 3 wt% POM                      | 72   | 236                       |
|  | Soybean oil<br>Soybean oil<br>Soybean oil<br>Free FAs (20% oleic acid)<br>Palmitic acid                              | 30 : 1 MeOH : oil, 140 °C, 8 h, 3 wt% POM                      | 25 (x = 5)–93 (x = 1)                              | 236                       |
|  |  | 30 : 1 MeOH : oil, 140 °C, 4 h, 3 wt% POM                      | 95   | 236                       |
|  |  | 5 : 1 MeOH : FA, 65 °C, 4 h, 0.06 mmol POM                     | 96   | 247                       |
|  |  | 28 : 1 MeOH : FA, 65 °C, 12 h, 2.5 wt% POM                     | > 97   | 247                       |
|  |  | 12 : 1 EtOH : oil, 90 °C, 8 h, 2 mol% POM                      | 100% yield   | 237                       |
| Sn <sub>1.2</sub> H <sub>0.6</sub> [PW <sub>12</sub> O <sub>40</sub> ] | Macauba oil  | 12 : 1 EtOH : oil, 90 °C, 8 h, 2 mol% POM                      | 83   | 248                       |
|  | [H-pyrazine-SO <sub>3</sub> H] <sub>1.5</sub> [PW <sub>12</sub> O <sub>40</sub> ]                                    | 1 : 1 MeOH : FA, 25 °C, 24 h, 200 mg POM                       | 92% yield  | 249                       |
|  | [MIM-PSH] <sub>2</sub> H[PW <sub>12</sub> O <sub>40</sub> ]  | 13 : 1 EtOH : FA, 80 °C, 5 h, 7 wt% POM                        | 99% yield  | 201                       |
|  | TBA <sub>6</sub> [P <sub>2</sub> W <sub>17</sub> -SO <sub>3</sub> H]   | 10 : 1 MeOH : FA, 70 °C, 0.3 h, 10 wt% POM                     |  |                           |

MIM-PSH = 3-(1-methylimidazolium-3-yl)propane-1-sulfonate.

acidity of HPAs makes them promising catalysts for the production of biodiesel.

It has also been shown that the catalytic activity of HPAs in biodiesel production can be tuned by exchanging some of the protons for different counter-cations, as described for the hydrolysis of polysaccharides. For instance, the catalytic activity of H<sub>3</sub>[PW<sub>12</sub>O<sub>40</sub>] towards the esterification of palmitic acid could be varied by using glycine (Gly) counterions. [GlyH]<sub>1</sub>H<sub>2</sub>[PW<sub>12</sub>O<sub>40</sub>] was found to be ideal for this purpose since the Gly counter ion enabled better mass transport while leaving two available Brønsted acid protons.<sup>229</sup> This is in agreement with the ideal exchange ratio reported for polysaccharide hydrolysis in Section 2.3.1. Moreover, this homogeneous POM catalyst was reported to be highly water-tolerant, stable, and non-toxic.<sup>229</sup> The POM also self-separated since the

product formed an organic layer in which the POM was immiscible, allowing for the POM catalyst to be recycled and reused for several cycles.<sup>229</sup> Similarly, micellar catalysts for the esterification of fatty acids were obtained by exchanging one of the protons in H<sub>5</sub>[BW<sub>12</sub>O<sub>40</sub>] or H<sub>5</sub>[TiPW<sub>11</sub>O<sub>40</sub>] with the surfactant CTA.<sup>239,244</sup> These micellar catalysts could be easily recycled without significant loss of activity or loss of the catalyst and exhibited higher catalytic activity than the HPAs alone (Table 5). This was proposed to be due to the fatty acid and alcohol substrates being concentrated around the catalyst within micelles.<sup>239,244</sup> The catalytic activity was also found to increase with increasing chain length of the surfactant counter-cation up to around C<sub>16</sub>, which corresponds to the chain length of CTA and palmitic acid. Therefore, in accordance with the studies on biomass conversion (Section 2.3.1), the catalytic activity of





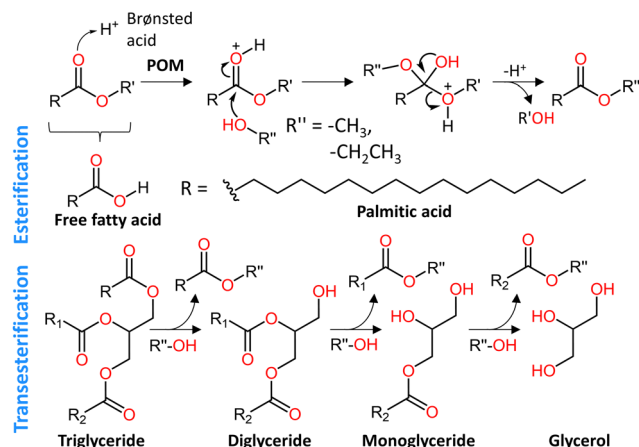


Fig. 17 General mechanism for the (trans)esterification of free fatty acids (e.g., palmitic acid) and triglycerides (via di- and monoglycerides) catalyzed by Brønsted acidic HPA catalysts.

HPAs towards biodiesel production can be enhanced by varying the counter-cation.

Cation exchange can also facilitate the separation of the POM after the reaction by forming insoluble salts that can act as heterogeneous catalysts.<sup>189</sup> For instance, a fully heterogeneous catalyst towards the esterification of palmitic acid was obtained by exchanging the protons in  $\text{H}_3[\text{PW}_{12}\text{O}_{40}]$  by  $\text{NH}_4$ , and partial exchange was shown to result in higher activity than the fully exchanged ammonium salt  $(\text{NH}_4)_3[\text{PW}_{12}\text{O}_{40}]$ .<sup>250</sup> Similarly, exchanging the protons with  $\text{Cs}^+$  was used to form insoluble salts of  $\text{H}_3[\text{PW}_{12}\text{O}_{40}]$  that could catalyze the esterification of palmitic acid and the transesterification of tributyrin, a triglyceride, with methanol (both separately and simultaneously).<sup>234</sup> Furthermore,  $\text{Cs}_x\text{H}_{3-x}[\text{PW}_{12}\text{O}_{40}]$  with  $x = 2.0\text{--}2.3$  gave higher conversion due to having more accessible surface acid sites (Table 5).<sup>234</sup> Moreover,  $\text{Cs}_{2.5}\text{H}_{0.5}[\text{PW}_{12}\text{O}_{40}]$  has been shown to display higher catalytic activity towards the transesterification of vegetable oil into biodiesel than  $\text{H}_2\text{SO}_4$ , while being tolerant towards water content and the presence of free fatty acids as well as being easily recyclable.<sup>235,246,251</sup> However,  $\text{Cs}_1\text{H}_2[\text{PW}_{12}\text{O}_{40}]$  was shown to be more active than salts with a higher content of  $\text{Cs}^+$  in the esterification of rice bran fatty acids with methanol (Table 5), which was proposed to be due to the higher Brønsted acidity of  $\text{Cs}_1\text{H}_2[\text{PW}_{12}\text{O}_{40}]$ .<sup>245</sup> Similarly, the catalytic activity of Cs salts of a mixed metal Keggin HPA  $(\text{PV}_3\text{MoW}_8\text{O}_{40})^{6-}$ , prepared by cation exchange, was recently reported to decrease with increasing amounts of  $\text{Cs}^+$  (Table 5) due to a drop in the Brønsted acidity. However, since  $\text{Cs}_1\text{H}_5[\text{PV}_3\text{MoW}_8\text{O}_{40}]$  was partially soluble,  $\text{Cs}_2\text{H}_4[\text{PV}_3\text{MoW}_8\text{O}_{40}]$  was determined to be the best heterogeneous catalyst.<sup>236</sup> In addition,  $\text{Cs}_2\text{H}_4[\text{PV}_3\text{MoW}_8\text{O}_{40}]$  was shown to be an effective and recyclable heterogeneous catalyst in the simultaneous transesterification of soybean oil and esterification of free fatty acids with superior conversion to  $\text{Cs}_2\text{H}_2[\text{SiW}_{12}\text{O}_{40}]$  and  $\text{Cs}_2\text{H}[\text{PW}_{12}\text{O}_{40}]$  (Table 5).<sup>236</sup> Mixed ammonium and Cs salts  $(\text{NH}_4)_x\text{Cs}_{2.5-x}\text{H}_{0.5}[\text{PW}_{12}\text{O}_{40}]$ ;  $x = 0.5, 1, 1.5, 2$  were also investigated and  $(\text{NH}_4)_2\text{Cs}_{0.5}\text{H}_{0.5}[\text{PW}_{12}\text{O}_{40}]$  gave the

highest turnover frequency for the esterification of oleic acid with ethanol, which was determined to be due to it having a higher surface area, pore size, and number of mesopores.<sup>252</sup> Consequently, exchanging the counter-cation to form insoluble salts can produce highly effective heterogeneous catalysts for biodiesel production with their catalytic activity depending on the structure and acid strength of the solid catalyst imparted by the cation exchange.

HPAs with several other organic and inorganic counter-cations have also been explored as heterogeneous catalysts for biodiesel production. For example,  $\text{Sn}^{2+}$  and  $\text{Zr}^{4+}$  HPA salts have been reported to form heterogeneous catalysts that benefit from the synergistic combination of the Lewis acidic cations with the Brønsted acidic HPAs.<sup>237,247</sup> Moreover, crystals of  $[\text{PW}_{12}\text{O}_{40}]^{3-}$  with sulfonic acid functionalized pyrazinium ( $[\text{H-pyrazine-SO}_3\text{H}]^{2+}$ ) displayed exceptional activity towards the esterification of free fatty acids using only equimolar amounts of methanol at room temperature, in contrast to typically reported conditions involving excess methanol and elevated temperatures (Table 5).<sup>248</sup> Such combinations of  $[\text{PW}_{12}\text{O}_{40}]^{3-}$  with IL cations also allow for facile recyclability of the catalyst.<sup>248,249</sup> Due to the advantages of heterogeneous catalysis, particularly regarding the ease of recovery and reuse of the catalyst, research focus has shifted in recent years towards the heterogenization of HPAs.

The TBA salt of a Wells–Dawson POM covalently functionalized with sulfonic acid groups,  $\text{TBA}_6[\text{P}_2\text{W}_{17}\text{O}_{61}\text{SO}_3\text{H}]$ , which was discussed in Section 2.3.2, was also reported to act as a phase transfer catalyst for the esterification of free fatty acids that displayed higher activity than  $\text{H}_3[\text{PW}_{12}\text{O}_{40}]$ , the monolacunary Wells–Dawson POM  $([\text{P}_2\text{W}_{17}\text{O}_{61}]^{10-})$ , and  $\text{H}_2\text{SO}_4$  (Table 5).<sup>201</sup> This higher catalytic activity was proposed to be due to the formation of a catalytically active emulsion, as a result of the use of TBA as the counter-cation, because it allowed for better interactions between the substrates and the POM active sites.<sup>201</sup> As the reaction progressed, a phase separation took place and the POM catalyst precipitated out of solution, allowing easy recovery of both the catalyst and products.<sup>201</sup> As for biomass conversion, this is also the only example of the use of a covalently functionalized hybrid POM for the production of biodiesel and suggests that such hybrid systems should be investigated further.

In contrast to the extensive research on the use of POMs for biodiesel production due to the Brønsted acidity of HPAs, the redox activity of POMs towards lipids has been scarcely explored. However, different salts of  $\text{M}_{3/n}[\text{PMO}_{12}\text{O}_{40}]$  ( $\text{M}^{n+} = \text{H}^+, \text{K}^+, \text{Zn}^{2+}, \text{Cu}^{2+}, \text{Cr}^{3+}, \text{Fe}^{3+}, \text{and Al}^{3+}$ ) have been explored as catalysts for the oxidation of glycerol, the major by-product of biodiesel production, into added-value platform chemicals such as lactic acid.<sup>253</sup> The oxidation of glycerol into lactic acid takes place in the presence of  $\text{O}_2$  via a radical chain mechanism involving the formation of dihydroxyacetone and glyceraldehyde, which are interconvertible isomers (Fig. 18).<sup>253</sup> Then, the dehydration of glyceraldehyde produces pyruvaldehyde, which reacts with water to form lactic acid. The activity and selectivity for these different reaction products were found to depend on



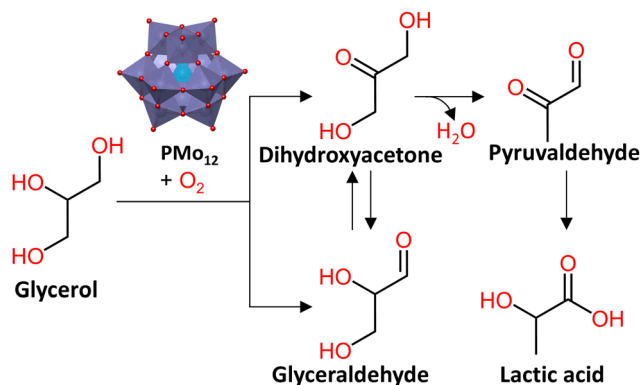


Fig. 18 Oxidation of glycerol by [PMo<sub>12</sub>O<sub>40</sub>]<sup>3−</sup> (PMo<sub>12</sub>) under an O<sub>2</sub> atmosphere to form dihydroxyacetone or glyceraldehyde, which interconvert, leading to the dehydration of dihydroxyacetone into pyruvaldehyde that is then converted into lactic acid.

the counter-cation. The counter-cation with the highest charge (Al<sup>3+</sup>) resulted in the highest conversion and selectivity for lactic acid, while Cu<sup>2+</sup> completely inhibited the reaction.<sup>253</sup> This was proposed to be due to the influence of the counter-cation on the generation of radicals, and the role of Al<sup>3+</sup> in catalyzing the dehydration reaction. Yet, oxidation of fatty acids or other lipids has not been significantly explored. Nevertheless, a recent report has shown that (NH<sub>4</sub>)<sub>6</sub>[Mo<sub>7</sub>O<sub>24</sub>] can induce the peroxidation of phospholipids in the cell membranes of glioblastoma cells, which may be linked to its anti-cancer activity.<sup>254</sup> Therefore, the redox activity of POMs towards lipids should be explored further.

### 3. Reactivity of MOC-based materials

In the previous sections, the focus was predominantly on MOCs acting as homogeneous catalysts in biomolecular transformations. Although this facilitates the study of their reactivity, it presents several disadvantages for practical applications, especially in the separation and reuse of the catalyst. In contrast, heterogeneous catalysts allow for easy separation and reuse of the catalyst since the catalytically active sites and the substrates are in separate phases, thereby providing an efficient solution for reliable catalyst durability and product isolation.<sup>255</sup> As a result, inherently insoluble MOCs and insoluble hybrid materials containing MOCs have been developed as alternatives that can catalyze biomolecular reactions heterogeneously.

The most notable and well-studied examples of these types of materials are MOC-based MOFs, which can function as heterogeneous catalysts for a diverse array of biologically relevant reactions, owing to their three-dimensional network of MOCs interconnected by organic linker molecules. Moreover, the composition, morphology, and architecture of MOFs can be tailored to specific reactions, making them highly versatile catalysts that offer high potential for designing efficient catalytic processes.<sup>256</sup> In addition, MOFs and MOF-based hybrid materials (*e.g.*, POM@MOF) can outperform traditional porous materials like zeolites in terms of their ability to control

porosity, backbone functionalization, guest immobilization, and active site modification. The porosity of MOFs can be fine-tuned by selecting appropriate organic linkers or by adjusting synthesis parameters to induce missing-linker or missing-cluster defects to further facilitate the diffusion of substrate molecules and favor the accessibility of active sites. Additionally, MOFs can serve as supports or hosts for the uniform immobilization or encapsulation of a large number of reactive species, preventing them from leaching, agglomerating, or deactivating during reactions. Hence, MOFs are highly versatile materials that provide many avenues to create highly promising efficient catalysts.<sup>45,50–52,257–259</sup> Moreover, MOFs have been shown to catalyze a wide range of biomolecular transformations as has been recently described in detail in a recent review.<sup>59</sup> Therefore, this section will only focus on the reactivity towards biomolecules of MOFs based on MOCs, especially to draw comparisons with the studies on discrete clusters. Other hybrid materials based on MOCs will also be discussed where relevant to highlight where the limits of MOFs lie and where improvements are still needed.

#### 3.1. Amino acids, peptides and proteins

**3.1.1. Hydrolysis of peptides.** Inspired by the hydrolytic activity of Zr-POMs towards peptides and proteins, the catalytic activity of MOFs based on hexanuclear Zr<sup>4+</sup> clusters, {Zr<sub>6</sub>O<sub>8</sub>}, was also investigated by our group. The hydrolysis of simple peptide bond model systems, such as Gly-Gly and other Gly-X dipeptides, with MOFs was first investigated by Ly *et al.*<sup>260</sup> This study employed MOF-808, which consists of {Zr<sub>6</sub>O<sub>8</sub>} nodes connected by six benzene-1,3,5-tricarboxylate (BTC) linkers to form a framework with a pore size of 18.4 Å (Fig. 19(a)).<sup>260</sup> MOF-808 was shown to hydrolyze Gly-Gly to form Gly and cGly-Gly as a byproduct. It reduced the activation energy by approximately 22 kJ mol<sup>−1</sup> and enhanced the rate of hydrolysis by almost 3 orders of magnitude compared to Zr-POMs under the same reaction conditions to give nearly 100% conversion after just 3 h (Table 6), while the Zr-POMs required several days to reach 100% conversion.<sup>80,83</sup> The hydrolysis of Gly-Gly under the same conditions was also attempted with Zr<sub>6</sub>-based clusters capped with benzoate (Zr<sub>6</sub>(OH)<sub>4</sub>O<sub>4</sub>(C<sub>6</sub>H<sub>5</sub>CO<sub>2</sub>)<sub>12</sub>) or methacrylate (Zr<sub>6</sub>(OH)<sub>4</sub>O<sub>4</sub>(CH<sub>2</sub>=C(CH<sub>3</sub>)CO<sub>2</sub>)<sub>12</sub>) ligands (Fig. 1(f)). However, the discrete MOCs produced no hydrolytic products after 5 h, which was attributed to all Zr<sup>4+</sup> centers being coordinatively saturated.<sup>260</sup> Consequently, Zr-based MOFs present distinct advantages over Zr-POMs and even discrete Zr<sub>6</sub> clusters.

The mechanism of MOF-808 reactivity towards peptide bonds was proposed to be similar to that observed for M-POMs. The reaction was catalyzed *via* coordination of the dipeptide to the redox inactive and Lewis acidic Zr<sup>4+</sup> centers, which occurs either through exchange with formate ligands (often present in MOFs from the use of formic acid during their synthesis) or directly at defect sites with missing BTC linkers. Coordination likely takes place at two adjacent Zr<sup>4+</sup> centers *via* an N-terminal N atom and a peptide carbonyl O atom of Gly-Gly, thereby activating the carbonyl group of the peptide bond towards a nucleophilic attack, as suggested by computational



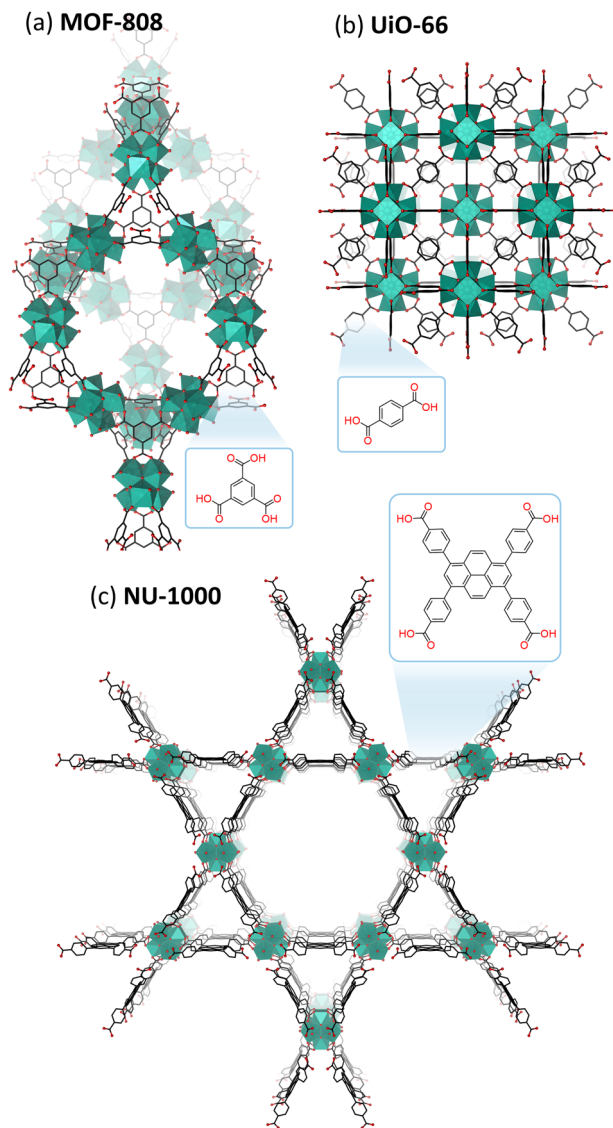


Fig. 19 Structures of  $\{M_6O_8\}$ -based MOFs: (a) MOF-808, (b) UiO-66, and (c) NU-1000.

modeling. Therefore, more missing linker defects result in an acceleration of the reaction rate, which motivates the use of MOFs with a higher number of missing linker defects, provided that MOF stability is not adversely affected. Overall, this shows that MOF-808 is an effective heterogeneous catalyst for the hydrolysis of peptide bonds due to the Lewis acidity of  $Zr^{4+}$  centers in available active sites.

The reaction rate of hydrolysis catalyzed by MOF-808 was found to depend on the nature of the dipeptide's structure in a similar way to what was observed for M-POMs. It was found that the C-terminal carboxylate group is important in the hydrolytic reaction since modifying it resulted in a slower reaction.<sup>260</sup> Gly-Gly hydrolysis in the presence of non-reactive multidentate carboxylic acids also caused a decrease in the reaction rate due to their ability to competitively coordinate to  $Zr^{4+}$  centers instead of Gly-Gly. The greatest decrease was

observed upon the addition of acids with short aliphatic chains since their interaction with the MOF is not sterically hindered. Similarly, conversion of Gly-X dipeptides decreased with increasing side chain bulkiness (Gly-Gly > Gly-Ala > Gly-Asp > Gly-Asn > Gly-Lys  $\approx$  Gly-Arg) due to steric hindrance.<sup>260</sup> On the other hand, Gly-Met and Gly-Phe displayed higher reaction rates, which was proposed to be due to their thiol and aromatic functionalities that facilitated interactions with the linker molecules, while Gly-X dipeptides with hydrophobic residues exhibited lower conversion rates due to the lack of favorable interactions with both the linkers and the  $\{Zr_6O_8\}$  cluster core.<sup>260</sup> Hence, the hydrolytic activity of MOF-808 depends on the composition of the peptide because of the role of terminal carboxylate groups in the hydrolysis reaction and the interactions of side chains with the linker molecules.

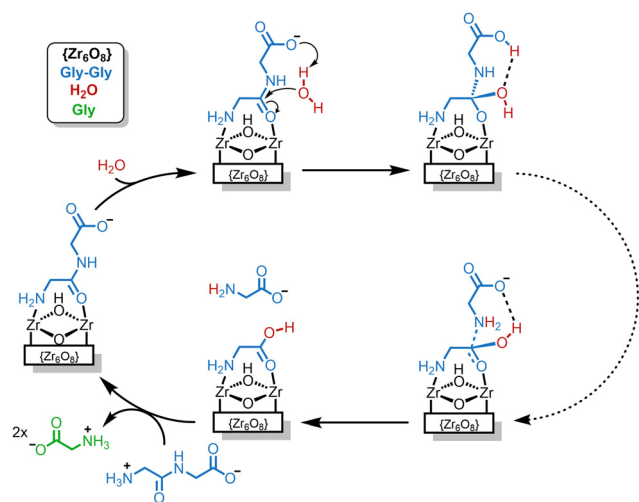
While Ly *et al.* briefly investigated the binding mode of Gly-Gly to the  $\{Zr_6O_8\}$  cluster core using DFT calculations,<sup>260</sup> Conic *et al.* went a step further by elucidating the full reaction mechanism of Gly-Gly hydrolysis after binding to the  $\{Zr_6O_8\}$  cluster.<sup>267</sup> More specifically, DFT calculations were used to gain a deeper understanding of the composition and arrangement of the dangling ligands surrounding the  $\{Zr_6O_8\}$  cluster under the hydrolysis reaction conditions and to elucidate the mechanism underlying efficient hydrolysis of Gly-Gly catalyzed by MOF-808. Under the conditions used for hydrolysis reactions, it was determined that the active sites of MOF-808 likely consist of protonated  $\{Zr_6O_8H_4\}$  clusters with negatively charged ligands (*i.e.*, bridging BTC and formate ligands) to reach charge-neutrality. Some of the formate ligands were found to be exchangeable with water and hydroxide, resulting in  $Zr_6O_4(OH)_4(BTC)_2(HCOO)_6$  and  $Zr_6O_4(OH)_4(BTC)_2(HCOO)_5-(OH)(H_2O)$  being predicted to be the most favorable structures. Binding of Gly-Gly was proposed to occur through the replacement of one of the formate ligands and it was found that: (i) formation of a neutral complex is preferred, (ii) binding to  $Zr^{4+}$  is strongest in the order carboxylate > carbonyl > amine, and (iii) complex stability is influenced by the relative coordination geometry of the dipeptide on the  $\{Zr_6O_8\}$  cluster core, especially with respect to the  $\mu_3$ -OH groups due to potentially stabilizing hydrogen bonding.<sup>267</sup> However, while binding of the carboxylate group in bidentate bridging mode is slightly more favorable, only bridging coordination to the amide carbonyl oxygen and N-terminal nitrogen results in activation towards hydrolysis (Fig. 20). In addition, hydrolysis was proposed to involve nucleophilic attack by a free external water molecule (Fig. 20), as the attack by core oxygens, coordinated  $OH^-$ , or coordinated  $H_2O$  is not viable due to their position on the cluster with respect to coordinated Gly-Gly. This external  $H_2O$  molecule can be activated towards nucleophilic attack by the neighboring carboxylate group, in a similar mechanism to that established for a Zr-POM,<sup>83</sup> which is followed by multiple proton shuffling steps and conformational changes (Fig. 20). The preferred reaction pathway involves two main transition states: nucleophilic attack on the amide carbon and breaking of the C-N bond with either one serving as the rate-determining step. However, the proton shuffling and conformational changes





**Table 6** Conditions and rate constants ( $k_{\text{obs}}$ ) for selected reports on MOF-catalyzed peptide hydrolysis

| MOF                        | Substrate               | Reaction conditions             | Conversion/%           | $k_{\text{obs}}/\times 10^{-6} \text{ s}^{-1}$ | Ref. |
|----------------------------|-------------------------|---------------------------------|------------------------|--|------|
| MOF-808(Zr)                | Gly-Gly                 | pD 7.4, 60 °C, 4 h, 1 eq. MOF   | 92                     | 269  | 260  |
|                            | Gly-Gly-NH <sub>2</sub> | pD 7.4, 60 °C, 1 eq. MOF        | —                      | 55   | 260  |
|                            | Gly-Gly-Ala             | pD 7.4, 60 °C, 1 eq. MOF        | —                      | 206  | 260  |
| MOF-808(Zr)<br>(35–850 nm) | Gly-Gly                 | pD 7.6, 60 °C, 20 h, 1 eq. MOF  | 71 (850 nm)–94 (35 nm) | 17.3 (850 nm)–38.7 (35 nm)                     | 261  |
| UiO-66(Zr)                 | Gly-Gly                 | pD 7.8, 60 °C, 360 h, 1 eq. MOF | 64                     | 0.8  | 262  |
| UiO-66(Zr)-NO <sub>2</sub> | Gly-Gly                 | pD 7.8, 60 °C, 1 eq. MOF        | —                      | 8.4  | 262  |
| UiO-66(Zr)-NH <sub>2</sub> | Gly-Gly                 | pD 7.8, 60 °C, 1 eq. MOF        | —                      | 9.0  | 262  |
| UiO-66(Zr)-TFA             | Gly-Gly                 | pD 7.8, 60 °C, 1 eq. MOF        | —                      | 8.0  | 262  |
| UiO-66(Zr/Ce)              | Gly-Gly                 | pD 7.4, 60 °C, 1 eq. MOF        | —                      | 0.6–1.35 (depending on %Ce)                    | 263  |
| NU-1000(Zr)                | Gly-Gly                 | pD 7.4, 60 °C, 72 h, 1 eq. MOF  | 53                     | 1.6  | 264  |
| NU-1000(Hf)                | Gly-Gly                 | pD 7.4, 60 °C, 24 h, 1 eq. MOF  | 7                      | 0.8  | 265  |
| MIP-201                    | Gly-Gly                 | pD 7.4, 60 °C, 24 h, 1 eq. MOF  | 80                     | 18.5   | 266  |

**Fig. 20** Proposed mechanism of Gly-Gly hydrolysis by a {Zr<sub>6</sub>O<sub>8</sub>}-based MOF. Adapted from ref. 267.

after the nucleophilic attack were not investigated in detail as they were assumed to have low energy barriers and the implicit solvent model used is too inaccurate for these low-energy calculations.<sup>267</sup> Therefore, additional computational and experimental studies are required to confirm these findings.

Following these mechanistic studies, the structure–activity relationships of MOFs were investigated further to optimize them as catalysts for protein hydrolysis. This was achieved by studying the impact of varying the linker functionality and the MOF framework on the hydrolysis of dipeptides. In this context, the catalytic activity of {Zr<sub>6</sub>O<sub>8</sub>}-based UiO-66 MOFs (Fig. 19(b)) with different linkers – the typical 1,4-benzenedicarboxylate (BDC) linker and BDC derivatives with electron-withdrawing (NO<sub>2</sub>) or electron-donating (NH<sub>2</sub>) groups – were investigated, all of which bind to the cluster at twelve sites.<sup>262</sup> The modified UiO-66(Zr)-NO<sub>2</sub> and UiO-66(Zr)-NH<sub>2</sub> variants were both observed to provide a 10-fold increase in catalytic activity with respect to standard UiO-66(Zr) (Table 6). The high reactivity of UiO-66(Zr)-NH<sub>2</sub> is surprising since the amino group's electron-donating ability should reduce the Lewis acidity of {Zr<sub>6</sub>O<sub>8</sub>}, thus reducing its ability to activate peptide bonds towards

nucleophilic attack by water. However, since the particle size of all three UiO-66 variants was practically identical, it was suggested that the reason for the higher activity was the greater porosity and larger number of missing linkers in UiO-66(Zr)-NH<sub>2</sub>, which makes the internal catalytic sites more accessible to the dipeptide.<sup>262</sup> This was confirmed by inducing missing linker defects using trifluoroacetic acid (TFA) to form a UiO-66 structure with higher porosity (UiO-66(Zr)-TFA) that hydrolyzed Gly-Gly with a similar reaction rate to UiO-66(Zr)-NO<sub>2</sub> and UiO-66(Zr)-NH<sub>2</sub> (Table 6). Furthermore, the different UiO-66 variants displayed distinct selectivities in the hydrolysis of Gly-X dipeptides, with UiO-66(Zr)-TFA showing preference for dipeptides with nucleophilic groups susceptible to intramolecular *N,O*-rearrangement (Gly-Ser and Gly-Asp). This was suggested to result from a greater contribution of Brønsted acid sites that aid the hydrolysis of the formed ester intermediate. In contrast, UiO-66(Zr)-NO<sub>2</sub> showed a greater contribution of Lewis acid sites, likely linked to the electron-withdrawing effect of the –NO<sub>2</sub> group, since it did not show increased reactivity toward any dipeptide in particular.<sup>262</sup> Therefore, this study demonstrates that the reactivity and selectivity of MOFs can be controlled through the number and functionality of the linkers.

The structure of MOFs also plays a key role in the hydrolysis reaction partly because of how it affects the adsorption of dipeptides onto the MOF. Therefore, Loosen *et al.* investigated the factors that influence peptide adsorption onto different MOF structures: MOF-808, UiO-66, and NU-1000 where MOF NU-1000 is composed of {M<sub>6</sub>O<sub>8</sub>} clusters with eight 1,3,6,8-(*p*-benzoate)pyrene linkers per cluster (Fig. 19(c)).<sup>268</sup> Adsorption of Gly (–OOCCH<sub>2</sub>NH<sub>3</sub><sup>+</sup>) and Gly-Gly (–OOCCH<sub>2</sub>NHOCCH<sub>2</sub>NH<sub>3</sub><sup>+</sup>) was observed to be significantly affected by the repulsion between the ammonium (–NH<sub>3</sub><sup>+</sup>) group and the positively charged surface of the MOFs. Gly is generally adsorbed less by all MOFs than Gly-Gly since the –NH<sub>3</sub><sup>+</sup> group in Gly-Gly is further away from the cluster when Gly-Gly is coordinated *via* its carboxylate group (Zr–OOCCH<sub>2</sub>NH<sub>3</sub><sup>+</sup> vs. Zr–OOCCH<sub>2</sub>NHOCCH<sub>2</sub>NH<sub>3</sub><sup>+</sup>). Furthermore, the absorption of Gly-Gly was found to increase for all MOFs with increasing pH (pH 3 and pH 7), which was attributed to the MOF surface charge becoming less positive, resulting in a decrease in repulsion between





the cluster and the  $\text{-NH}_3^+$  group of the peptide. Moreover, MOF-808 displayed greater adsorption of Gly-Gly compared to UiO-66 and NU-1000 under all investigated reaction conditions, which was proposed to be due to the presence of more available  $\text{Zr}^{4+}$  sites, since MOF-808 has six linkers per cluster while UiO-66 and NU-1000 have a higher linker connectivity.<sup>268</sup> Interestingly, N- and C-terminal protected peptides displayed distinct adsorption behavior for the different MOF architectures. From this, it was suggested that adsorption of peptides by hydrophilic MOFs, such as MOF-808 and UiO-66, is primarily driven by C-terminal carboxylate coordination to  $\{\text{Zr}_6\text{O}_8\}$  clusters. In contrast, peptide adsorption to the hydrophobic NU-1000 was mainly influenced by repulsive electrostatic interactions of the N-terminal  $\text{-NH}_3^+$  group of the dipeptide and hydrophobic interactions of the side chain of the dipeptide with the large aromatic linker of the MOF.<sup>268</sup> Because of these interactions, Gly-X dipeptides with more hydrophilic side chains were adsorbed more favorably by MOF-808 while NU-1000 highly favored the adsorption of dipeptides with hydrophobic/aromatic side chains, and the extent of adsorption increased with increasing hydrophobicity of the dipeptide. However, dipeptides with functional groups that allowed for coordination to the  $\{\text{Zr}_6\text{O}_8\}$  cluster – such as carboxylate, amide, and hydroxide groups – also resulted in more favorable adsorption on NU-1000. On the other hand, despite these systematic studies, a clear trend could not be established for the adsorption of Gly-X dipeptides to UiO-66. Nevertheless, the adsorption did not seem to depend on differences in particle size or pore size between the MOFs. Smaller particle size and higher external surface area did, however, favor adsorption of Gly-Gly by MOF-808, which suggests that hydrolysis most likely occurs on the external surface of the MOF, even though its pore size is sufficiently large to allow for permeation of dipeptides and other small oligopeptides.<sup>261</sup> In addition to particle size, porosity, and connectivity, the three-dimensional features of the MOF architecture can influence reactivity as well since the entropic contribution to the Gibbs free energy of hydrolysis is relatively large, as observed when comparing Zr-based NU-1000 and MOF-808.<sup>264</sup> Thus, while more detailed analysis remains necessary, the intricate MOF network allows for tunability of both the reactivity and selectivity.

Another factor that can impact reactivity, is the nature of the metal atom in the  $\{\text{M}_6\text{O}_8\}$  cluster. This was investigated by comparing NU-1000 MOFs (Fig. 19(c)) based on either  $\text{Zr}^{4+}$  or  $\text{Hf}^{4+}$ , both of which gave similar results for the hydrolysis of Gly-Gly (Table 6).<sup>265</sup> However, similar reaction rates were observed for the hydrolysis of Gly-Gly, Gly-Ala, Gly-Asp, and Gly-Ile catalyzed by NU-1000(Hf) despite the size differences of the dipeptides, which is distinctly different from the results of NU-1000(Zr) and indicates that NU-1000(Hf) is relatively less sensitive to steric hindrance of the peptide substrate.<sup>265</sup> Furthermore, while reaction rates for dipeptide and protein hydrolysis were similar, NU-1000(Zr) consistently performed better. This cannot result from differences in the intrinsic Brønsted acidity of  $\text{Zr}^{4+}$  and  $\text{Hf}^{4+}$ , since both MOFs perform best at pH 7–8, nor can it result from differences in enthalpy

and entropy contributions, since the same  $\Delta G^\ddagger$  was observed for the reaction with both MOFs. The differences in the reactivity most likely arise as a result of small structural variations as opposed to differences in the intrinsic reactivity of both metals.<sup>265</sup> Hence, even though  $\text{Zr}^{4+}$  and  $\text{Hf}^{4+}$  are similar, they can influence the MOF structure, and thus the reactivity, differently.

To enhance the reactivity and structural diversity of the MOFs used for peptide bond hydrolysis, different methods have been employed. While linker substitution and substitution of  $\text{Zr}^{4+}$  by  $\text{Hf}^{4+}$  both improved hydrolysis in different ways, no additional functionality was introduced.<sup>262,265</sup> For this purpose, a series of bimetallic UiO-66 MOFs with different Ce/Zr ratios were synthesized and employed for peptide bond hydrolysis.<sup>263</sup> Due to the superior reactivity of  $\text{Ce}^{4+}$  compared to  $\text{Zr}^{4+}$ , UiO-66(Ce) has shown a higher catalytic activity compared to UiO-66(Zr). However, the stability of UiO-66(Ce) under the conditions typically used for hydrolysis is lower compared to UiO-66(Zr) due to the facile reduction of  $\text{Ce}^{4+}$  to  $\text{Ce}^{3+}$ , which is less Lewis acidic. To circumvent the low stability of pure  $\text{Ce}^{4+}$ -based MOFs and still benefit from the increased reactivity of  $\text{Ce}^{4+}$ , bimetallic UiO-66(Ce/Zr) MOFs were synthesized containing different ratios of  $\text{CeZr}_5$  and  $\text{Ce}_6$  clusters based on the overall Ce/Zr ratio (28, 43, 61, and 87 mol% Ce). The reactivity towards the hydrolysis of Gly-Gly was similar for all variants (Table 6) and their stability was confirmed to be similar to that of UiO-66(Zr). Only a slight increase in the reactivity was observed upon increasing the number of  $\text{Ce}_6$  clusters, while increasing the content of  $\text{CeZr}_5$  clusters caused a slight decrease in the reactivity, which was attributed to the dipeptide's coordination to two different metal centers (Ce and Zr) being less favorable.<sup>263</sup> While the introduction of  $\text{Ce}^{4+}$  into MOFs did not significantly increase reactivity in peptide bond hydrolysis, the scope of reactions could be expanded.  $\text{Ce}^{4+}$  can function as a one-electron oxidizing agent, which endows UiO-66(Zr/Ce) MOFs with redox activity that could be leveraged to catalyze oxidation as well as hydrolytic reactions. As a result, the oxidation of cysteine to cystine was successfully catalyzed by a UiO-66(Zr/Ce) MOF. Moreover, the combined oxidation and hydrolysis of tripeptide glutathione was investigated, and a preference for oxidation was observed even though it was substantially slower than cysteine oxidation owing to steric hindrance or unfavorable coordination.<sup>263</sup> These studies show that the reaction scope can be successfully expanded by incorporating metals with different properties into the same MOF structure. Therefore, the effect of other metals should be investigated further, as it could enable one-pot reactions combining multiple reaction steps.

**3.1.2. Hydrolysis of proteins.** The hydrolytic activity of MOC-based MOFs towards peptides indicates that MOFs are potentially promising heterogeneous catalysts for the hydrolysis of proteins. Indeed, several MOC-based MOFs have been shown to catalyze protein hydrolysis.

The first example of protein hydrolysis catalyzed by MOC-based MOFs was the hydrolysis of HEWL by MOF-808(Zr) with 55% hydrolysis after 25 h at 60 °C.<sup>260</sup> Only four fragments of *ca.*



**Table 7** Conditions and number of peptide fragments of selected reports based on SDS-PAGE for the MOF-catalyzed hydrolysis of various proteins

| MOF                     | Protein <sup>a</sup> | Reaction conditions  | No. of fragments | Proposed cleavage sites  | Ref.    |
|-------------------------|----------------------|--|------------------|--|---------|
| MOF-808(Zr)             | HEWL                 | Phosphate buffer, pH 7.4, 60 °C, 3.1 mg mL <sup>-1</sup> MOF       | 4                | Asp18-Asn19, Asp52-Tyr53, Asp119-Val120  | 260,264 |
|                         | HHM                  | H <sub>2</sub> O, pH 7.4, 60 °C, 2 μmol MOF + SDS                  | 4                | Asp20-Ile21, Asp44-Lys45, Asp126-Ala127 <sup>b</sup>   | 270     |
|                         |                      |  | 7                | Asp44-Lys45 <sup>b</sup> , Asp60-Leu61 <sup>b</sup> , Asp109-Ala110, Asp122-Phe123, Asp126-Ala127 <sup>b</sup>               |         |
|                         |                      | + CHAPS  | 4                | Asp20-Ile21, Asp44-Lys45, Asp126-Ala127 <sup>b</sup>   |         |
|                         |                      | + Zw3-12   | 2                | Asp126-Ala127 <sup>b</sup>   |         |
| MOF-808(Zr) (35–850 nm) | HEWL                 | + TX-100   | 4                | Asp60-Leu61, Asp126-Ala127 <sup>b</sup>  | 261     |
|                         |                      | H <sub>2</sub> O, pH 7.2, 60 °C, 2 μmol MOF                        | 5                | Asp18-Asn19 <sup>b</sup> , Asp52-Tyr53, Asp66-Gly67, Asp101-Gly102, Asp119-Val120  |         |
| NU-1000(Zr)             | HEWL                 | H <sub>2</sub> O, pH 7.0, 60 °C, 2 μmol MOF                        | 3                | Asp18-Asn19 <sup>b</sup> , Asp52-Tyr53, Asp119-Val120  | 264     |
| NU-1000(Hf)             | HEWL                 | H <sub>2</sub> O, pH 7.0, 60 °C, 2 μmol MOF                        | 3                | Asp18-Asn19 <sup>b</sup> , Asp52-Tyr53, Asp119-Val120  | 265     |
| MIP-201                 | HHM                  | HEPES/H <sub>2</sub> O, pH 7.4, 60 °C, 2.8 mg mL <sup>-1</sup> MOF | 6                | Asp20-Ile21 <sup>b</sup> , Asp60-Leu61 <sup>b</sup> , Asp109-Ala110, Asp122-Phe123 <sup>b</sup> , Asp126-Asp127 <sup>b</sup> | 266     |

<sup>a</sup> HEWL = Hen Egg White Lysozyme; HHM = Horse Heart Myoglobin. <sup>b</sup> More than one peptide fragment may be assigned to this cleavage site.

12.2, 10.7, 8.2, and 6.4 kDa were observed using SDS-PAGE (Table 7), indicating selective peptide bond cleavage. This is much faster than what was described for various Zr-POMs under similar reaction conditions,<sup>70,88</sup> which can be attributed to a higher density of Zr<sup>4+</sup> ions that function as the active sites for hydrolysis. In regard to the hydrolysis mechanism, and the interaction mode between MOF-808 and HEWL, the following hypotheses have been suggested: (i) partial unfolding of HEWL at 60 °C could enable the protein to diffuse into the MOF's pores to access internal {Zr<sub>6</sub>O<sub>8</sub>} active sites,<sup>269</sup> or (ii) initial hydrolysis of HEWL occurs most likely in the half-spherical pockets on the surface of MOF-808 that are similar in size to the protein, thereby facilitating diffusion of hydrolytic fragments into the pores for further cleavage. Although no unambiguous evidence could be obtained for either scenario, considering the limited number of {Zr<sub>6</sub>O<sub>8</sub>} active sites (~1%) on the MOF's external surface area, hydrolysis of initially formed fragments within the MOF is plausible, especially given the high efficiency of the hydrolytic reaction.<sup>260</sup>

Even though MOF-808 showed promising hydrolytic activity, it was synthesized in a highly explosive and dangerous mixture of dimethylformamide (DMF) and formic acid at an elevated temperature, like many other MOFs.<sup>260</sup> Therefore, adjusting this synthesis procedure to ensure its safety and a lower environmental impact would be an important step in their development as nanozymes. Dai *et al.* accomplished this by synthesizing the {Zr<sub>6</sub>O<sub>8</sub>} clusters first in isopropanol and water and then using them to form the MOF-808 structure in a safer mixture of water and formic acid.<sup>261</sup> In addition to a greener synthesis, more monodisperse MOF-808 nanocrystals were obtained with a tunable size, thereby ensuring that the reactivity is identical throughout the whole synthetic batch. This is important because the hydrolytic activity of MOFs towards proteins is mainly affected by the MOF's macro characteristics, such as the particle size and external/internal surface areas, instead of the linker identity, which is much less important considering the vast size of the substrate.<sup>264</sup> Hence, varying the particle size and, therefore, the MOF's external surface allowed tuning of the number and concentration of peptide fragments,

as observed for the hydrolysis of HEWL by a series of MOF-808 with different sizes (Table 7).<sup>261</sup> Moreover, reproducibility, scalability, and low variability are essential for the implementation of MOFs in industrial applications and can be achieved through safer synthetic strategies, such as the one reported by Dai *et al.*

Hydrolysis of HEWL using NU-1000(Zr) and NU-1000(Hf) was also investigated, which allows for the influence of the nature of the metal center to be examined (Table 7). Both MOFs produced fragments of *ca.* 12.5, 10.3, and 8.5 kDa, which indicates that both Zr- and Hf-based NU-1000 cleave the protein with the same selectivity,<sup>264,265</sup> and that the selectivity is not affected by the MOF structure since similar fragments were also observed with MOF-808.<sup>260</sup> This also shows that the MOF structure likely has a direct impact on the number of peptide fragments produced, since more fragments were observed for MOF-808.<sup>260</sup> However, visualization of the hydrolytic fragments produced by NU-1000 using SDS-PAGE was very difficult, which was attributed to strong adsorption at the liquid-solid interface. In order to recover the peptide fragments from the MOF, various elution protocols were attempted by incubation with established protein eluents. While this was unsuccessful for glycine-HCl and guanidine-HCl eluents, the intact protein band could be visualized after incubation in 1% aqueous ammonia at ambient temperature, which also destroyed the MOF due to the low stability of NU-1000 under highly basic conditions.<sup>264</sup> Nevertheless, the observed hydrolytic activity of MOF-808 and NU-1000 shows that MOFs can selectively cleave proteins to produce peptide fragments suitable for middle-down proteomics. The cleavage was postulated to occur at Asp-X and X-Asp peptide bonds, similarly to Zr-POMs. Although the reactivity is still governed by the Lewis acidic metal centers, the MOF structure clearly plays a role, most likely by influencing the interactions between the MOF and the protein and/or the peptide fragments.

While several examples have indicated the hydrolytic potential of Zr- and Hf-based MOFs,<sup>260,264–266</sup> the detailed elucidation of the interaction mechanism remains an issue owing to the heterogeneous nature of these materials.



Furthermore, the adsorption of proteins and their hydrolytic fragments onto MOFs makes studying the reaction particularly challenging. Therefore, to increase the applicability of MOFs for protein hydrolysis, issues related to strong substrate adsorption need to be addressed, which starts by finding a suitable elution protocol that is not detrimental to the MOF's structure. Alternatively, more stable MOF structures can be used to withstand elution conditions. For instance, MIP-201, consisting of  $\{Zr_6O_8\}$  building units coordinated to six 3,3',5,5'-tetracarboxydiphenylmethane ( $H_4mdip$ ) linkers (Fig. 21(a)), was used for the cleavage of myoglobin and was successfully recycled and regenerated for several hydrolysis reaction cycles (Table 7).<sup>266</sup> This was possible because of the very high stability of this MOF, as it remained stable, even after prolonged incubation in boiling water under highly acidic conditions or under basic conditions (pH 10), which would allow for elution of protein fragments using basic media without destruction of the MOF.<sup>266</sup> Hence, finding better elution protocols or using more stable MOFs would enable more detailed studies of their reactivity and would make the implementation of MOFs in bio-technological applications more feasible.

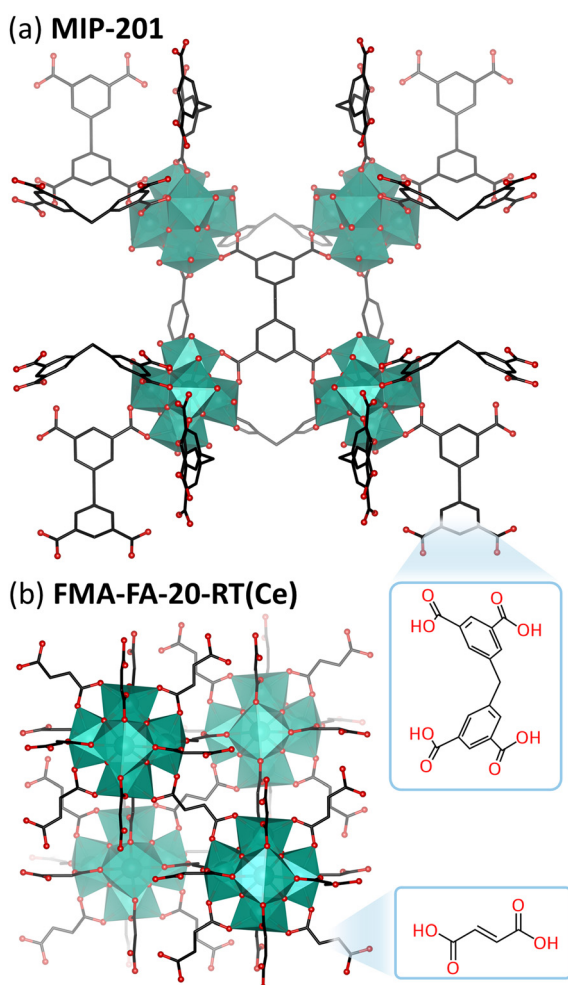


Fig. 21 Structures of (a) MIP-201, consisting of  $\{Zr_6O_8\}$  clusters, and (b) FMA-FA-20-RT(Ce), consisting of  $\{Ce_6O_8\}$  clusters.

MOF-808(Zr) has also been shown to remain active towards the hydrolysis of myoglobin in the presence of surfactants, making it an interesting catalyst for proteomics applications alongside M-POMs, especially for studying water-insoluble proteins that must be solubilized with surfactants. MOF-808(Zr) cleaved myoglobin producing fragments of *ca.* 4.6, 13.7, 12.2, and 11.1 kDa (Table 7), with a hydrolysis efficiency of around 63% after 8 days in the absence of any surfactant.<sup>270</sup> However, addition of surfactants affects both the protein structure and the MOF-protein interactions in a rather complex manner and, therefore, the structure of the surfactant used was determined to play an important role in the reaction outcome. The anionic surfactant SDS promoted protein hydrolysis by causing larger protein unfolding ( $\pm 65\%$  hydrolysis efficiency after 8 days) whereas zwitterionic CHAPS and neutral TX-100 surfactants inhibited hydrolysis by interacting with the MOF's active sites or forming micelles on its surface: around 39% and 31% hydrolysis efficiency after 8 days, respectively. Interestingly, while hydrolysis of myoglobin in the presence of CHAPS merely reduced the hydrolysis efficiency, hydrolysis in the presence of TX-100 caused a different cleavage selectivity.<sup>270</sup> Therefore, the addition of specific types of surfactants can influence both the efficiency and selectivity of protein hydrolysis by MOFs. This is extremely valuable for the development of MOFs as catalysts for protein hydrolysis as it shows that the reactivity of MOFs can be tuned through using additives, such as surfactants, and that the MOF catalyst does not completely lose its activity in the presence of surfactants, unlike natural proteases.

Besides  $Zr^{4+}$  and  $Hf^{4+}$ , other metal centers have also been described for protein hydrolysis. For instance, an FMA(Ce) MOF, which is homologous to UiO-66 with fumaric acid (FMA) ligands instead of BDC linkers (Fig. 21(b)), was used for the hydrolysis of bovine serum albumin (BSA) in a study on MOF-catalyzed hydrolysis of amide, phosphoester, phosphoanhydride, and glycosidic bonds.<sup>44</sup> The study showed that the hydrolytic activity could be increased by increasing the Lewis acidity of the incorporated metals, either by choosing a different metal or tuning the Lewis acidity by linker exchange, and by improving the active site density. The latter can easily be accomplished by selecting short linkers such as FMA. However, this also decreases pore size/volume, thus direct improvement using this strategy is less straightforward. Multiple MOF architectures were synthesized using acetic (AA), formic (FA), and TFA as modulators that can affect the morphology, size, yield, degree of crystallinity, and number of defects.<sup>271,272</sup> Out of all the tested MOFs, FMA-FA-20-RT(Ce) (synthesized at 25 °C using a molar ratio of 20:1 FA:FMA to introduce missing linker defects) was shown to be most catalytically active in initial model reactions, and was further investigated for the hydrolysis of BSA.<sup>44</sup> The reaction was followed by gel permeation chromatography (GPC), which indicated 100% BSA hydrolysis after 36 h at 60 °C, or after 7 days at 37 °C. While the efficiency of FMA-FA-20-RT(Ce) is 7 times lower in comparison to the commonly employed protease trypsin, this MOF is also 10 000 times more economical than trypsin. Moreover, FMA-FA-20-RT(Ce)





produces hydrolytic fragments of similar length compared to trypsin, even though it has considerable preference towards cleavage of bonds involving Arg, Lys, and Asp residues while trypsin only cleaves at Arg and Lys amino acids. Furthermore, MOF-808(Zr) was also investigated toward BSA hydrolysis, but performed considerably slower, with only 50% BSA hydrolysis after 24 h at 60 °C. Since the surface area of MOF-808 is much larger compared to FMA-FA-20-RT(Ce), this highlights the superior performance of Ce<sup>4+</sup> compared to Zr<sup>4+</sup> active sites.

In summary, several MOFs have shown great potential for the hydrolysis of proteins. However, while the mechanism for peptide bond hydrolysis and oxidative cleavage of proteins catalyzed by various MOCs has been elucidated in great detail,<sup>70,80,87,130,273</sup> the studies with MOFs are still to be developed further. Although initial reports yielded some insights,<sup>260,264,266,267</sup> the exact mechanism of MOF-catalyzed peptide bond hydrolysis in larger protein substrates, and especially in insoluble protein substrates, needs to be studied more extensively.

**3.1.3. Peptide bond formation.** As previously discussed for POMs, it is important to note that peptide bond hydrolysis is a reversible process, and the reverse reaction (peptide bond formation) can be achieved in the absence of water. While there have been a few reports of amide bond formation in model systems using MOC-based MOF catalysts,<sup>274,275</sup> peptide bond formation between amino acids has only been reported once to the best of our knowledge. In this report, de Azambuja *et al.* assessed the reactivity of Zr-based MOF-808, NU-1000, and UiO-66 towards dipeptide cyclization and intermolecular peptide formation between amino acids in organic solvents as model reactions for amide bond formation.<sup>276</sup> UiO-66 was found to be the most suitable MOF catalyst since the recovery of the products was easier and, unlike with MOF-808, hydrolysis products were not detected. Furthermore, these MOFs were shown to be highly water-tolerant and could catalyze the reaction under ambient conditions without the need for additives, unlike other catalysts commonly used for peptide bond formation. Therefore, the use of MOC-based MOFs for this type of reactivity could potentially be exploited for the formation of synthetic peptides and bio-inspired molecules that may be of interest for biotechnological or pharmaceutical applications.

## 3.2. Nucleic acids and nucleotides

### 3.2.1. Hydrolysis of nucleic acids and their model systems.

For the hydrolytic or oxidative cleavage of phosphoester bonds in various substrates, mainly gold and metal-oxide nanoparticles have been employed as (photo)catalysts.<sup>277</sup> Contrastingly, the cleavage of phosphoester bonds using cluster-based materials has been underreported, with the cleavage of DNA by MOFs having been described only recently.<sup>278</sup> To understand both what has been accomplished and what still needs to be addressed, this section will cover examples of phosphoester bond cleavage using cluster-based materials and discuss the hydrolysis mechanism for relevant substrates.

In 2012, Han *et al.* reported on the use of a phosphotungstate POM embedded into several Zn- and lanthanide-based MOFs, for hydrolysis of DNA model substrate BNPP.<sup>279</sup> While the heterogeneous catalysis of phosphoester bond cleavage was successful due to the reactivity of the well-dispersed POMs and the insolubility of the larger framework, the reaction rate was only moderate ( $t_{1/2}$  = 161.7–286.1 h) and the mechanism was not fully elucidated. A more comprehensive investigation was later carried out using MOC-based MOFs, in which the activation process, active site, and reaction mechanism were explored using Zr-based UiO-66, UiO-67, PCN-700, PCN-701, and PCN-703 frameworks (Fig. 22 and Table 8).<sup>280</sup> Out of all these MOFs, UiO-66 displayed the highest reactivity ( $K_M$  =  $1.55 \times 10^{-4}$  M), which was comparable to that of alkaline phosphatase. Furthermore, the lower activity of UiO-67 with respect to UiO-66 (Table 8) was proposed to be due to UiO-66 having inherently more linker vacancies. This was confirmed by following the reactivity of PCN-type MOFs, which increased in the order of PCN-703 < PCN-701 < PCN-700 corresponding to their increasing number of coordinatively unsaturated sites (CUS): 1, 2, and 4, respectively (Fig. 22 and Table 8). This demonstrates the importance of CUS for catalyzing phosphoester bond hydrolysis.

The catalytic activity was significantly boosted if the MOF was activated under strong acidic (pH 2–3) or basic (pH 8) conditions because monocarboxylate ligands bound to the {Zr<sub>6</sub>O<sub>8</sub>} clusters can be more easily exchanged at more extreme pH values.<sup>280</sup> As a result, more catalytically effective defects are introduced. In addition, the influence of pH on the hydrolysis of NPP catalyzed by UiO-66 was investigated and the best

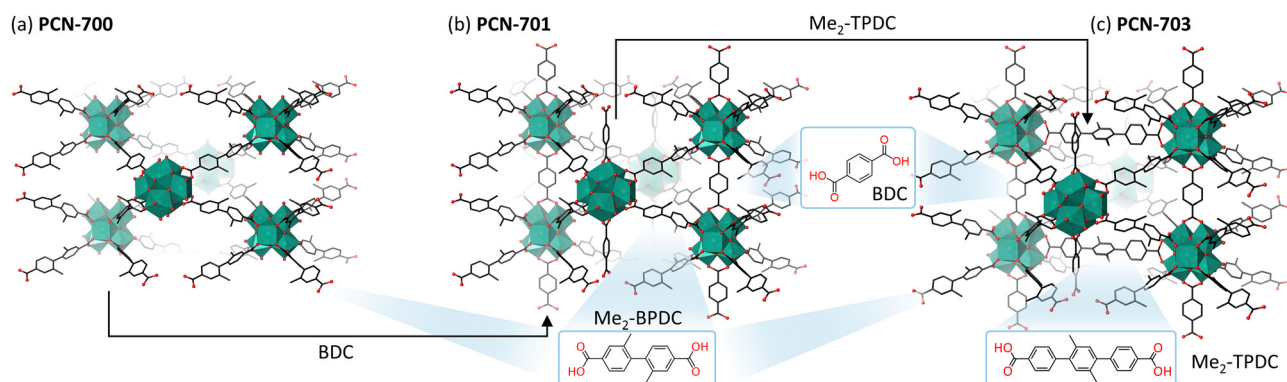


Fig. 22 Structures of MOFs (a) PCN-700, (b) PCN-701, and (c) PCN-703, which differ in terms of the linkers connecting the {Zr<sub>6</sub>O<sub>8</sub>} clusters.





**Table 8** Selected conditions and percentage conversion/rate constants reported for MOF-catalyzed phosphoester/phosphoanhydride bond hydrolysis

| MOF              | Substrate      | MOF connectivity                              | Reaction conditions  | Conversion & rate constant | Ref. |
|------------------|----------------|---|--|----------------------------|------|
| UiO-66(Zr)       | NPP            | 12 (many missing linker defects)              | 100 mM HEPES, pH 8.0, 37 °C, 0.08 mg mL <sup>-1</sup> MOF      | 108 μM h <sup>-1</sup>     | 280  |
| UiO-67(Zr)       | NPP            | 12 (limited number of missing linker defects) | 100 mM HEPES, pH 8.0, 37 °C, 0.08 mg mL <sup>-1</sup> MOF      | 60 μM h <sup>-1</sup>      | 280  |
| PCN-700(Zr)      | NPP            | 8   | 100 mM HEPES, pH 8.0, 37 °C, 1.5 mg mL <sup>-1</sup> MOF       | 73.2 μM h <sup>-1</sup>    | 280  |
| PCN-701(Zr)      | NPP            | 10  | 100 mM HEPES, pH 8.0, 37 °C, 1.5 mg mL <sup>-1</sup> MOF       | 57.6 μM h <sup>-1</sup>    | 280  |
| PCN-703(Zr)      | NPP            | 12  | 100 mM HEPES, pH 8.0, 37 °C, 1.5 mg mL <sup>-1</sup> MOF       | 25.8 μM h <sup>-1</sup>    | 280  |
| MOF(Hf/Ni)       | NPP            | 12  | 50 mM HEPES, pH 7.4, r.t., 0.2 eq. MOF                         | 194 μM h <sup>-1</sup>     | 281  |
|                  | BNPP           | 12  | 50 mM HEPES, pH 7.4, r.t., 0.2 eq. MOF                         | 18.9 μM h <sup>-1</sup>    | 281  |
|                  | NPA            | 12  | 50 mM HEPES, pH 7.4, r.t., 0.2 eq. MOF                         | 5.8 μM h <sup>-1</sup>     | 281  |
|                  | NPS            | 12  | 50 mM HEPES, pH 7.4, r.t., 0.2 eq. MOF                         | 1.5 μM h <sup>-1</sup>     | 281  |
| HMUiO-66(Ce)     | ssDNA          | 12 (assembled with macro-porous template)     | 20 mM HEPES, pH 7.4, 60 °C, 6 h, 0.35 mg mL <sup>-1</sup> MOF  | 100% conversion            | 278  |
|                  | Plasmid pBR322 | 12 (assembled with macro-porous template)     | 20 mM HEPES, pH 7.4, 60 °C, 24 h, 0.35 mg mL <sup>-1</sup> MOF | 74% conversion             | 278  |
| FMA-FA-20-RT(Ce) | NPP            | 12  | pH 9.0, 37 °C, 0.5 mg mL <sup>-1</sup> MOF                     | $t_{1/2} \leq 2$ min       | 44   |
|                  | ATP            | 12  | pH 10.0, 37 °C, 12 h, 0.5 mg mL <sup>-1</sup> MOF              | 21.5% conversion           | 44   |
|                  | ADP            | 12  | pH 10.0, 37 °C, 12 h, 0.5 mg mL <sup>-1</sup> MOF              | 18.5% conversion           | 44   |
|                  | AMP            | 12  | pH 7.0, 37 °C, 12 h, 0.5 mg mL <sup>-1</sup> MOF               | 5.0% conversion            | 44   |
| PCN(Fe)          | NPP            | 8   | 20 mM HEPES, pH 7.2, 65 °C, 0.75 mg mL <sup>-1</sup> MOF       | 186 μM h <sup>-1</sup>     | 282  |
| UiO-66(Ce)       | ATP            | 12  | 20 mM HEPES, pH 7.4, 37 °C, 1 h, 4.2 mg mL <sup>-1</sup> MOF   | 58.2% conversion           | 283  |

hydrolysis rate was observed at pH 8.<sup>280</sup> This was explained by the effect that pH has on Zr-bound water molecules and hydroxyl ions, which results in different reaction pathways depending on their protonation state. Based on DFT calculations and Diffuse Reflectance Infrared Fourier Transform Spectroscopy (DRIFTS), the most likely pathway at pH 8 was proposed to involve: (i) indirect coordination of NPP through H-bonding with a Zr-bound hydroxide; (ii) stabilization by a water molecule that is hydrogen bonded to both the substrate and a Zr-bound hydroxide; (iii) nucleophilic attack by the H-bonded water; and finally (iv) dissociation of the formed *p*-nitrophenoxide ion to yield H<sub>2</sub>PO<sub>4</sub><sup>-</sup> (Fig. 23(a)).<sup>280</sup> Furthermore, higher pH favored the reaction since this gave a lower  $\Delta G^\ddagger$  for the conformation of {Zr<sub>6</sub>O<sub>8</sub>}. However, above pH 8, the reaction rate was observed to decrease again, which was postulated to occur because of electrostatic repulsion between NPP and the {Zr<sub>6</sub>O<sub>8</sub>} cluster due to changes in the protonation state at high pH values. Therefore, although the  $\Delta G^\ddagger$  of {Zr<sub>6</sub>O<sub>8</sub>} at pH 8 was not optimal, the reaction rate was the highest due to lower repulsion.

Dong *et al.* further expanded this research by using Hf/Ni mixed-metal MOFs to hydrolyze NPP with near-complete position-selectivity, where phosphomonoesters were preferably hydrolyzed over other phosphoesters, and type-selectivity, where P–O rather than C–O or S–O bonds were cleaved.<sup>281</sup> They were assembled using isonicotinic and 3-aminoisonicotinic

acid ligands, to form MOF(Hf/Ni) and MOF(Hf/Ni)-NH<sub>2</sub> (Fig. 23(b)), respectively, with micropores of approximately 7.4 Å, and a large BET surface (1040 m<sup>2</sup> g<sup>-1</sup>). These MOFs were shown to display a high thermostability and solution stability towards leaching of Hf<sup>4+</sup> and Ni<sup>2+</sup> in HEPES buffer due to the high connectivity and elevated charge density exhibited by the Hf<sub>6</sub> cluster.<sup>281</sup> Furthermore, phosphatase-like hydrolysis of NPP was observed for both MOF(Hf/Ni) and MOF(Hf/Ni)-NH<sub>2</sub> (Table 8). Both MOFs significantly outperformed UiO-66(Hf), which suggests that the presence of Ni<sup>2+</sup> ions notably improved the catalytic activity. This probably results from the impact of Ni<sup>2+</sup> on the polarizability of the phosphoryl group and modification of transition state structures. In addition, MOF(Hf/Ni)-NH<sub>2</sub> displayed a higher reaction rate than MOF(Hf/Ni), which likely stems from the role of the amino functionalities as either base- or proton-transfer agents. The positive impact of both Ni<sup>2+</sup> and -NH<sub>2</sub> groups was confirmed by the drastically reduced catalytic activity of other MOFs without Ni and/or without the -NH<sub>2</sub> group (MOF(Hf/Cu), MOF(Hf/Cu)-NH<sub>2</sub>, MOF(Zr/Ni), MOF(Zr/Ni)-NH<sub>2</sub>, MOF(Zr/Cu), MOF(Zr/Cu)-NH<sub>2</sub>). However, while MOF(Hf/Ni) displayed excellent hydrolytic activity towards NPP, its activity in the hydrolysis of BNPP, 4-nitrophenyl acetate (NPA), and 4-nitrophenyl sulfate (NPS) was significantly lower or non-existent (Fig. 23(c) and (d)). This was rationalized based on the structure of the other substrates: steric hindrance of BNPP impedes diffusion through MOF



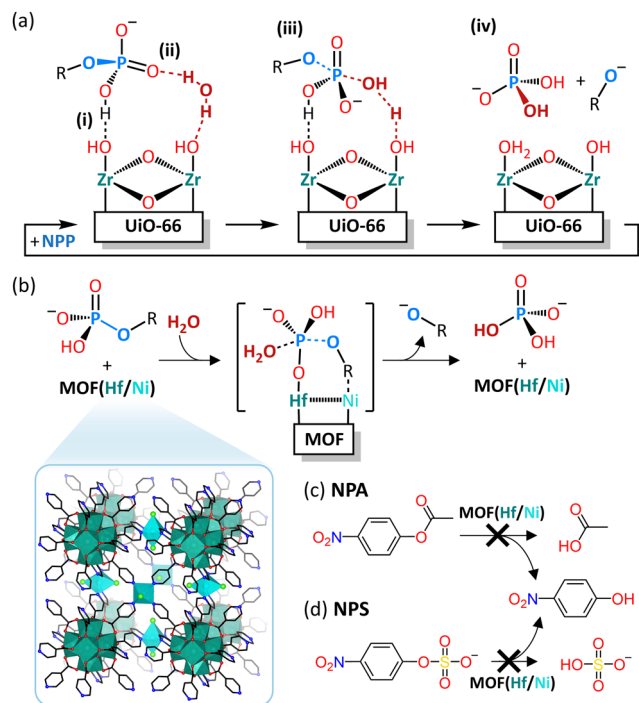


Fig. 23 Proposed mechanism of phosphoester bond hydrolysis of model systems (a) catalyzed by UiO-66(Zr) or (b) catalyzed by MOF(Hf/Ni), with the structure of MOF(Hf/Ni) shown in the inset (hafnium in teal, oxygen in red, carbon in black, nickel in light blue, nitrogen in blue, chlorine in light green). The unsuccessful hydrolysis of (c) 4-nitrophenyl acetate (NPA) and (d) 4-nitrophenyl sulfate (NPS) demonstrates the selectivity of MOF(Hf/Ni) towards hydrolysing P–O bonds over C–O and S–O bonds.

pores, the different geometry of NPA causes the formation of an unstable complex, and hydrolysis of NPS requires an expansive transition state with a much smaller solvent effect that is sterically unfavorable. Based on DFT calculations, complex stability with the four different substrates decreases in accordance with the experimental results:  $\text{NPP} \gg \text{BNPP} > \text{NPA} > \text{NPS}$  (Table 8). DFT calculations were also used to define the hydrolysis mechanism for NPP, which proceeds *via* (i) coordination of the nitril group of the substrate to  $\text{Ni}^{2+}$ , followed by (ii) stabilization of the complex through  $\pi$ – $\pi$  stacking between the MOF's linker and the substrate's nitrophenol group, and (iii)  $\text{P}=\text{O} \cdots \text{Hf}$  coordination between the substrate's phosphate group and the  $\{\text{Hf}_6\text{O}_8\}$  cluster (stabilized by hydrogen bonding), thereby enabling hydrolysis by nucleophilic attack of water (Fig. 23(b)). Finally, the cytotoxicity and biocompatibility of the MOFs were explored for future biomedical applications, which revealed that all investigated MOFs have very low cytotoxicity and that MOF(Hf/Ni) was the least cytotoxic, most probably due to its high stability.<sup>281</sup> Therefore, MOF(Hf/Ni) is a promising candidate for biomedical applications.

While successful hydrolysis of nucleic acid model systems, including the hydrolysis of structurally similar chemical warfare agents, has often been reported,<sup>48,284–289</sup> examples of phosphoester bond cleavage in larger substrates, such as DNA and RNA, remain very scarce. Li *et al.* hydrolyzed phosphoester bonds in NPP and BNPP using FMA-FA-20-RT(Ce) (Fig. 21(b)

and Table 8), which was also used for peptide bond hydrolysis, but DNA hydrolysis was not successful.<sup>44</sup> This is probably the result of the larger size of DNA prohibiting it from entering the MOF pores. In contrast, Yang *et al.* reported the hydrolysis of single-stranded DNA (ssDNA) using a hierarchically macro-microporous UiO-66(Ce) framework with a maximum pore size of 110 nm (HMUiO-66(Ce)), which completely hydrolyzed an 88-mer ssDNA within 6 h under physiological conditions (37 °C, pH 7.4).<sup>278</sup> HMUiO-66(Ce) was synthesized using a microemulsion-guided assembly, which allowed for large macropores that facilitate diffusion of large substrates, such as DNA, and thin microporous walls with a lot of solvent-exposed  $\{\text{Ce}_6\text{O}_8\}$  clusters for easy substrate access. Furthermore, HMUiO-66(Ce) structure is an excellent artificial nuclease due to the presence of  $\text{Ce}^{4+}$  and its abundance of highly accessible  $\text{Ce}^{4+}$ –OH sites (Table 8). Moreover, the ssDNA hydrolysis yield could be increased by raising the temperature (40–80 °C) and decreasing/increasing the pH (pH 3–9) because HMUiO-66(Ce) has excellent thermal and chemical stability, which is sorely lacking in natural enzymes. The efficiency of the catalyst was further confirmed by successfully cleaving double-stranded DNA in the form of a supercoiled plasmid. Finally, comparative analysis between the  $^{31}\text{P}$  NMR signal of free DNA (–1.21 ppm) and DNA in HMUiO-66(Ce) (–7.55 ppm) revealed the monodentate coordination of P–OH to  $\text{Ce}^{4+}$ –OH, making the phosphoester bond more susceptible to nucleophilic attack of water.<sup>278</sup>

The reported ability of certain MOFs to catalyze the cleavage of phosphoester bonds in DNA has potential implications for many biomedical applications, such as genetic engineering as well as biological imaging and sensing.<sup>46,290,291</sup> For example, Wang *et al.* reported the double role of PCN-8908, composed of  $\{\text{Zr}_6\text{O}_8\}$  clusters and methane-tetrakis(*p*-terphenyl carboxylate) linkers, as a delivery vehicle for Cpf1-encoding plasmids and as a catalyst for phosphoester bond cleavage.<sup>290</sup> While hydrolysis of DNA was not directly observed (hydrolytic activity was only demonstrated with diethyl phosphate), the MOF did increase the efficacy of the developed protocol, where DNA was cleaved using Cpf1/crRNA, with the editing efficiency of two genes increasing by 5-fold or more. Additionally, PCN-224(Zr), which is constructed from  $\{\text{Zr}_6\text{O}_8\}$  clusters that are each linked by 10 tetrakis(4-carboxyphenyl)porphyrin ligands, was reported to cause quenching of fluorescently labeled DNA *via* fluorescence resonance energy transfer (FRET), likely through the formation of Zr–O–P bonds between the ssDNA and the MOF. Since the formation of Zr–O–P bonds can be carefully tuned by occupying  $\text{Zr}^{4+}$  with free phosphate, the fluorescence behavior can be made to respond to various stimuli. This has been explored further in the creation of molecular sensing and logic systems capable of detecting multiple molecules.<sup>291</sup> Hence, the hydrolytic activity of MOFs towards phosphoester bonds has promising biotechnological applications and should be explored further.

**3.2.2. Hydrolysis of ATP.** Hydrolysis of phosphoester and phosphoanhydride bonds, found in molecules such as ATP, require specialized hydrolases optimized for one specific cleavage reaction (Fig. 24(a)). However, some of the phosphatase-mimicking MOFs discussed above possess sufficient Lewis



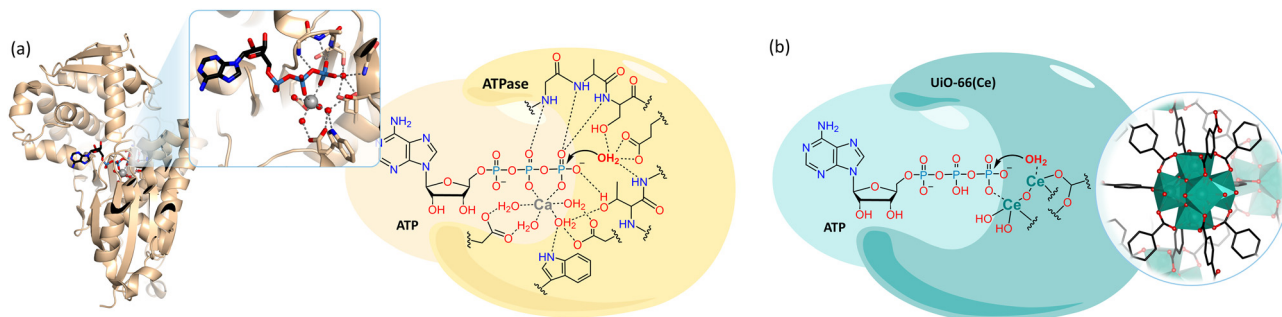


Fig. 24 Schematic representation of (a) the activation of ATP towards hydrolysis by an ATPase, based on a crystal structure of the protein (left; PDB code 3CJA), compared with (b) the proposed mechanism of activation by UiO-66(Ce). Adapted from ref. 283.

acidity for the activation of both bonds. This was established by Li *et al.*, who employed FMA-FA-20-RT(Ce) (Fig. 21(b)) to hydrolyze phosphoester bonds in model substrates, NPP and BNPP, as well as phosphoanhydride bonds in ATP and ADP.<sup>44</sup> In addition, it was found that both ATP and ADP were hydrolyzed by FMA-FA-20-RT(Ce) at similar rates, while AMP was hydrolyzed significantly slower. For all substrates, the observed mechanism aligned with cleavage of the P–O bond through Lewis acid activation by  $\text{Ce}^{4+}$  and nucleophilic addition of  $\text{Ce}^{3+}$ –OH, leading to a pentacoordinated intermediate that subsequently decomposes to the respective reaction products. The catalytic activity provided by the synergistic combination of different functionalities was also reported for another MOF consisting of  $\{\text{Zr}_6\text{O}_8\}$  clusters and iron porphyrin linkers (PCN(Fe)), which has the same crystal structure as the well-established PCN-222.<sup>282</sup> PCN(Fe) was shown to be active towards an array of reactions, owing to its oxidase-, peroxidase-, and phosphatase-like activity, including the dephosphorylation of ATP with  $\{\text{Zr}_6\text{O}_8\}$  clusters acting as the active site.<sup>282</sup> However, while ATP dephosphorylation was indeed observed, this study did not yield information on the mechanism or the kinetics of the reaction.<sup>282</sup>

A more detailed mechanistic study into the reactivity of MOFs towards nucleotides was presented in the first example of phosphoanhydride bond cleavage by a MOF reported in 2020 by Yang *et al.*, which involved the hydrolysis of ATP to ADP/AMP by UiO-66(Ce) (Table 8).<sup>283</sup> The mechanism of dephosphorylation for UiO-66(Ce) was found to be similar to that of the natural ATPase enzyme apyrase (Fig. 24), with coordination of the metal cation ( $\text{Ce}^{4+}$  in the case of the MOF and  $\text{Ca}^{2+}$  for apyrase) to one or two phosphate groups of the substrate followed by nucleophilic attack of water (coordinated to  $\text{Ce}^{3+}$  in the MOF or bound to the side chains of protein residues in the ATPase). While the mechanism itself is almost identical to that reported by Li *et al.*,<sup>44</sup> the enzyme-mimetic character of the proposed mechanism and the significance of both  $\text{Ce}^{3+}$  and  $\text{Ce}^{4+}$  were elaborated on in more detail.<sup>283</sup> Furthermore, while FMA-FA-20-RT(Ce) hydrolyzed ATP, ADP, and AMP, UiO-66(Ce) did not hydrolyze the phosphoester bond in AMP, which is an interesting demonstration of the selectivity of UiO-66(Ce) towards phosphoanhydride bonds. Furthermore, UiO-66(Ce) exhibited lower apparent  $K_M$  values for ATP and ADP substrates

compared to apyrase, with the sequential ADP hydrolysis being remarkably slower. This indicates that UiO-66(Ce) has higher substrate affinity due to its greater active site density, even though the hydrolysis mechanism is nearly identical to that of apyrase.<sup>283</sup> This bio-inspired approach where MOFs are compared to natural enzymes can provide valuable insights into how MOFs catalyze certain reactions, but also about the function of enzymes that catalyze the same reactions. However, despite these recent advances, further experimental and computational research is required to make MOFs, and other cluster-based hybrid systems, ready for potential applications.

### 3.3. Carbohydrates

**3.3.1. Hydrolysis of polysaccharides.** To convert biomass waste into biofuels or platform chemicals, the large polysaccharide structures must be hydrolyzed into monomeric units first. This bioconversion process requires the extraction of cellulose and hemicellulose components, followed by depolymerization of the carbohydrate polymers through hydrolysis of glycosidic bonds to produce free sugar monomers. As previously discussed in Section 2.3.1, the breakdown of lignocellulosic biomass into sugar monomers is typically performed *via* acid hydrolysis or enzymatic hydrolysis, both of which have several drawbacks.<sup>292–295</sup> Therefore, other catalytic routes are being explored, including the use of MOCs (Section 2.3.1) and MOC-based materials, especially as alternative heterogeneous catalysts for biomass valorization into platform chemicals that can be upgraded into various chemical compounds using an array of reaction mechanisms,<sup>180,296</sup> which will be discussed in the next section.

The previously described FMA-FA-20-RT(Ce) framework (Fig. 21(b)), which was studied for its hydrolytic activity toward an extensive array of biologically relevant bonds, was also investigated toward hydrolysis of glycosidic bonds.<sup>44</sup> Initial studies on the hydrolysis of 2-nitrophenyl  $\beta$ -D-galactopyranoside and 4-nitrophenyl *N*-acetyl- $\beta$ -D-glucosaminide were both successful, with the latter being cleaved more efficiently. However, the sugar dimers maltose, which contains an  $\alpha$ -1,4-glycosidic bond, and lactose, which contains the same  $\beta$ -1,4-glycosidic bond that is found in cellulose, could not be hydrolyzed into sugar monomers. This indicates that the nitrophenyl leaving groups assist in the hydrolysis. Therefore, carboxymethyl chitosan was chosen as a



model polymer instead of cellulose because the carboxymethyl group can favorably interact with the MOF's Ce clusters, thus assisting the hydrolysis reaction. Indeed, through the use of GPC, it was found that FMA-FA-20-RT(Ce) can successfully cleave carboxymethyl chitosan at 37 or 60 °C and pH 8 with limited recyclability.<sup>44</sup> Consequently, it was concluded that FMA-FA-20-RT(Ce) requires substrates with specific functional groups to lower the sorption energy for successful glycosidic bond hydrolysis, which means it cannot be used for biomass valorization as such, and further optimization is needed.

More recently, Han *et al.* reported cellulose hydrolysis using a hybrid catalyst (PW<sub>12</sub>@MIL-101-X) based on MOF MIL-101(Fe) that acts both as carrier for the hydrolytically active H<sub>3</sub>[PW<sub>12</sub>O<sub>40</sub>] and as a means to activate cellulose for hydrolysis by modifying its BDC linkers with electron-withdrawing groups (X = -Br, -NH<sub>2</sub>, -Cl, -NO<sub>2</sub>).<sup>297</sup> The addition of electron-withdrawing moieties on the MOF caused it to selectively adsorb cellulose by forming hydrogen bonds *via* its hydroxyl groups, which further weakens the existing hydrogen bonds within cellulose and thereby makes hydrolysis more efficient. Hence, the greatest glucose yield (16.2%) was obtained for PW<sub>12</sub>@MIL-101-NO<sub>2</sub> at 180 °C after 11 h due to the strong electron-withdrawing character of -NO<sub>2</sub> (Table 9).<sup>297</sup> However, the recyclability was sub-optimal with a loss of 4.3% in glucose yield after 3 cycles due to the weak interaction between POM and MOF, which led to a reduced catalyst loading within MIL-101-X for X = Br, Cl, NO<sub>2</sub>. PW<sub>12</sub>@MIL-101-NH<sub>2</sub>, on the other

hand, did not experience a significant loss in glucose yield after repeated cycles owing to stabilization of the POM through electrostatic interactions with -NH<sub>3</sub><sup>+</sup>, formed by protonation of the -NH<sub>2</sub> groups.<sup>297</sup> This highlights the importance of creating catalyst materials that are both reactive and stable, which can be achieved through careful design of the structure. Moreover, this shows the promising potential of combining POMs and MOFs, which has been recently explored for a wide range of reactions in organic chemistry and should be explored further with biomolecules.<sup>47</sup>

Overall, few examples of cellulose hydrolysis using MOC-based MOFs have been reported. However, this reaction could benefit greatly from MOF-related advantages, such as their heterogeneous nature facilitating catalyst separation after the reaction, since the current processes (*i.e.*, acid hydrolysis) are not environmentally conscious and, therefore, further research should be stimulated. Along with designing MOFs as a new class of catalysts for biomass conversion, MOFs have also been used to improve existing enzymatic catalysis. By using MOFs as protective coatings, biomolecules are effectively sheltered from harsh reaction conditions in which their performance is negatively impacted or in which they degrade entirely.<sup>302</sup> Various MOF structures have already been used to encapsulate and stabilize  $\beta$ -glucosidase for hydrolysis of cellulose in conditions that would otherwise be detrimental to the protein structure (*e.g.*, high temperature), with the benefit of enzyme heterogenization.<sup>49,303,304</sup> However, to the best of our

Table 9 Selected conditions and product yields reported for the conversion of sugars catalyzed by MOFs

| MOF   | Substrates | Reactivity    | Reaction conditions   | Product yield | Ref. |
|---|------------|---------------|---|---------------|------|
| PW <sub>12</sub> @MIL-101(Fe)-NO <sub>2</sub> | Cellulose  | Hydrolysis    | H <sub>2</sub> O, 180 °C, 3–11 h, 12 mg mL <sup>-1</sup> MOF  | 16.2% glucose | 297  |
| PW <sub>12</sub> @MIL-101(Fe)-NH <sub>2</sub> | Cellulose  | Hydrolysis    | H <sub>2</sub> O, 180 °C, 3–11 h, 12 mg mL <sup>-1</sup> MOF  | 12.9% glucose | 297  |
| UiO-66(Zr)-SO <sub>3</sub> H                  | Fructose   | Dehydration   | DMSO, 120 °C, 1 h, 60 mg mL <sup>-1</sup> MOF                 | 85% HMF       | 298  |
| UiO-66(Zr)                                    | Glucose    | Isomerization | H <sub>2</sub> O, 100 °C, 24 h, 25 mg mL <sup>-1</sup> MOF    | 5.0% fructose | 299  |
| UiO-66(Zr)-NH <sub>2</sub>                    | Glucose    | Epimerization | H <sub>2</sub> O, 100 °C, 24 h, 25 mg mL <sup>-1</sup> MOF    | 6.3% mannose  | 299  |
|   |            | Isomerization |   | 4.0% fructose |      |
|   |            | Epimerization |   | 3.3% mannose  |      |
|   |            | Dehydration   |   | 1.0% HMF      |      |
| NU-1000(Zr)                                   | Glucose    | Isomerization | H <sub>2</sub> O, 140 °C, 5 h, 10 mg mL <sup>-1</sup> MOF     | 19% fructose  | 300  |
|   |            | Dehydration   |   | 2.3% HMF      |      |
| NU-1000(Zr)-PO <sub>4</sub>                   | Glucose    | Isomerization | H <sub>2</sub> O/THF, 140 °C, 5 h, 10 mg mL <sup>-1</sup> MOF | 5.0% fructose | 300  |
|   |            | Dehydration   |   | 25% HMF       |      |
| UiO-66(Zr)                                    | Glucose    | Hydrolysis    | H <sub>2</sub> O, 140 °C, 3 h, 0.03 eq. MOF                   | 1.6% fructose | 301  |
|   |            | Isomerization |   | 0.5% mannose  |      |
|   |            | Dehydration   |   | 3.5% HMF      |      |
| Nano UiO-66(Zr)                               | Glucose    | Hydrolysis    | H <sub>2</sub> O, 140 °C, 3 h, 0.03 eq. MOF                   | 4.2% fructose | 301  |
|   |            | Isomerization |   | 5.3% mannose  |      |
|   |            | Dehydration   |   | 4.0% HMF      |      |
| Ultra-nano UiO-66(Zr)                         | Glucose    | Hydrolysis    | H <sub>2</sub> O, 140 °C, 3 h, 0.03 eq. MOF                   | 5.0% fructose | 301  |
|   |            | Isomerization |   | 6.1% mannose  |      |
|   |            | Dehydration   |   | 4.4% HMF      |      |
| MSBDC(50)-UiO-66(Zr)                          | Glucose    | Hydrolysis    | H <sub>2</sub> O, 140 °C, 3 h, 0.03 eq. MOF                   | 8.1% fructose | 301  |
|   |            | Isomerization |   | 7.3% mannose  |      |
|   |            | Dehydration   |   | 6.5% HMF      |      |
| Nano MSBDC(50)-UiO-66(Zr)                     | Glucose    | Hydrolysis    | H <sub>2</sub> O, 140 °C, 3 h, 0.03 eq. MOF                   | 6.4% fructose | 301  |
|   |            | Isomerization |   | 6.8% mannose  |      |
|   |            | Dehydration   |   | 7.7% HMF      |      |
| Naphtha(100)-UiO-66                           | Glucose    | Hydrolysis    | H <sub>2</sub> O, 140 °C, 3 h, 0.03 eq. MOF                   | 6.6% fructose | 301  |
|   |            | Isomerization |   | 1.2% mannose  |      |
|   |            | Dehydration   |   | 3.7% HMF      |      |

MSBDC = 2-monosulfo-benzene-1,4-dicarboxylate.





knowledge, MOC-based MOFs have not been reported for this purpose. Nevertheless, Zr- and Hf-based MOCs could be interesting for these applications as the synergy between the reactivity of the MOCs and that of the encapsulated enzyme may produce particularly effective catalysts.

### 3.3.2. Dehydration of sugars into platform chemicals.

After depolymerization, glucose can be processed into other useful reagents, such as HMF, which are obtained from glucose through isomerization into fructose and subsequent dehydration, as discussed in Section 2.3.2. HMF can then undergo further reactions to produce an array of useful platform chemicals. For example, it can be oxidized into 2,5-furandicarboxylic acid, which can replace terephthalic acid in polyethylene terephthalate (PET) production. Hence, the dehydration of glucose to HMF has been catalyzed with various MOF architectures.

In 2014, Chen *et al.* reported the dehydration of fructose with > 90% HMF yield in DMSO catalyzed by MIL-101(Cr)-SO<sub>3</sub>H consisting of trimeric Cr<sup>3+</sup> units, which are composed of {CrO<sub>6</sub>} octahedra linked by a central μ<sub>3</sub>-O ligand and are interconnected by Brønsted acidic 2-sulfoterephthalate linkers (Fig. 25).<sup>298</sup> The addition of sulfonic acid creates the Brønsted acidity needed for dehydration, while the Lewis acidity of Cr<sup>3+</sup> can promote the isomerization of glucose to fructose. However, the Cr metal used in this MOF is known to be toxic and harmful to the environment,<sup>31</sup> and the substitution of Cr in MIL-101 with other trivalent cations has been found to render the material unstable.<sup>305</sup> Consequently, alternative MOF architectures have been investigated as a means to gradually eliminate reliance on Cr. With this purpose, Luo *et al.* studied the isomerization of glucose using UiO-66(Zr) and compared it to MIL-101(Cr).<sup>299</sup> MIL-101(Cr) promoted glucose isomerization in both water and methanol at 100 °C and favored glucose epimerization to mannose (Fig. 25) at higher temperatures. In contrast, UiO-66(Zr) displayed simultaneous glucose isomerization and epimerization in aqueous media (Fig. 25), with the latter occurring through an intramolecular carbon skeleton rearrangement facilitated by UiO-66(Zr) that was not observed for MIL-101(Cr). This difference in behavior can be attributed to the larger ionic radius and increased polarizability of the Zr<sup>4+</sup> sites, leading to the formation of an eight-coordinate mononuclear complex that could promote carbon skeleton rearrangement (Fig. 25). On the other hand, MIL-101(Cr) with two Cr<sup>3+</sup> sites formed a hexa-coordinate binuclear complex due to the lower polarizability and steric hindrance of the trimeric units, resulting in glucose isomerization without carbon skeleton rearrangement (Fig. 25).<sup>299</sup> This contrasting behavior between MIL-101(Cr) and UiO-66(Zr) confirms the importance of both the structure and metal choice within MOFs. Furthermore, these studies show that the reactivity and selectivity are influenced by the nature of the active sites and their local environment within the framework, which must be considered when designing a MOF for a particular application.

Various Zr-based MOFs have been employed for the conversion of glucose, such as NU-1000(Zr) with {Zr<sub>6</sub>O<sub>8</sub>} cluster nodes that possess phosphate capping ligands in addition to the

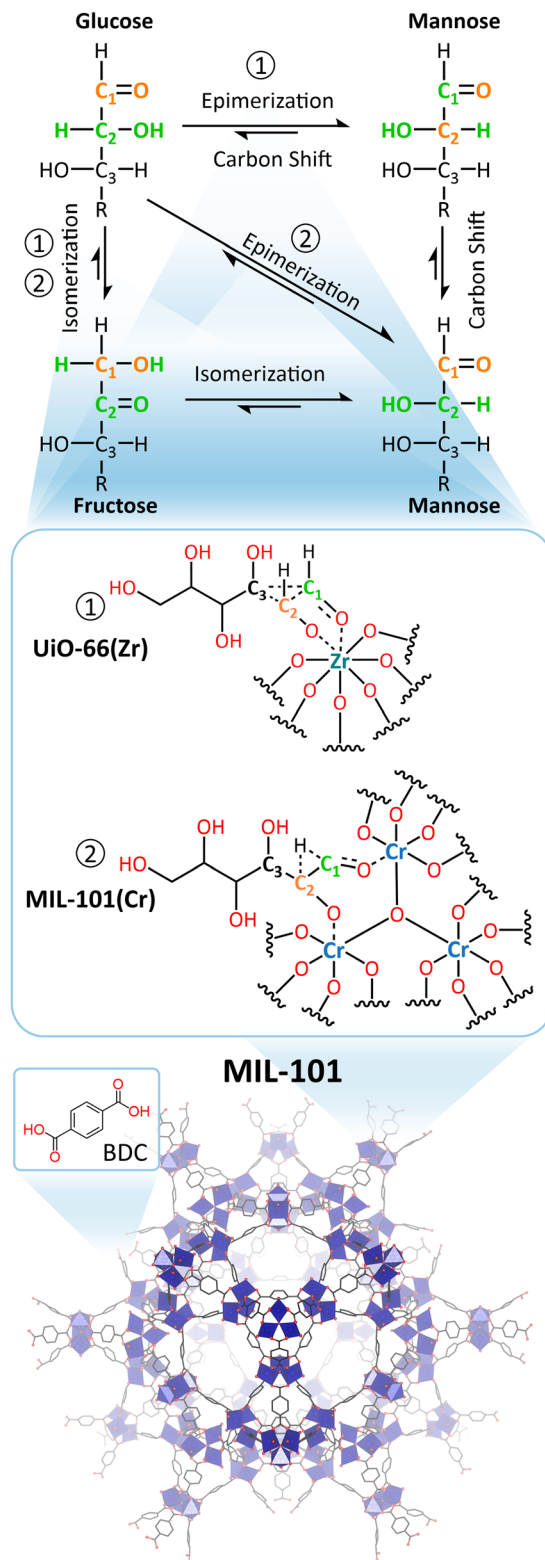


Fig. 25 Interconversion between glucose, fructose, and mannose depending on the MOF used, either UiO-66(Zr) or MIL-101, with the structure of MIL-101 with BDC linkers shown at the bottom. Adapted from ref. 299.

formate capping ligands to tune their Lewis acidity and avoid unwanted side reactions (Table 9).<sup>300</sup> Additionally, UiO-66(Zr)



modified with sulfonate-containing linkers was shown to catalyze the isomerization of glucose to fructose and dehydration to HMF in alcohol media because of its inherent defect-induced Lewis acidity (Table 9).<sup>306</sup> Furthermore, Oozeerally *et al.* showed that nano and ultra-nano UiO-66(Zr) MOFs, produced by slightly modifying the synthesis conditions, displayed increased catalytic performance (Table 9).<sup>301</sup> This can be attributed to two aspects of the reaction: (i) reduced particle size is accomplished by the introduction of more defects, which increases the number of accessible  $\text{Zr}^{4+}$  sites and the Lewis acidity, and (ii) the reaction can only occur on the external surface area because glucose is unable to enter the UiO-66 pores, which makes smaller particles with a larger surface-to-volume ratio more suitable. Secondly, partial substitution of the linkers in UiO-66(Zr) with 2-sulfoterephthalate resulted in much better catalytic performance owing to their electron-withdrawing effect, which enhances Lewis acidity of nearby defective  $\text{Zr}^{4+}$  sites. Thirdly, the hydrophobicity of UiO-66 was increased by functionalization with naphthalene, resulting in drastically enhanced selectivity towards fructose isomerization, and subsequent dehydration to HMF, instead of mannose epimerization. Finally, the introduction of free carboxylate groups increased HMF yield due to their mild Brønsted acidity, while leaving glucose conversion unaffected (Table 9). Because of these insights, the work by Oozeerally *et al.* serves as a roadmap for the future design of Zr-based MOFs for the conversion of glucose and as inspiration for other catalytic processes.<sup>301</sup>

HMF can further undergo a very wide range of different reactions, including condensation,<sup>307,308</sup> hydrogenation,<sup>308–312</sup> reduction,<sup>308</sup> oxidation,<sup>307,313</sup> esterification,<sup>314,315</sup> and etherification,<sup>316</sup> to functionalize it for specific downstream applications. These reactions are often suitable for both cascade and tandem one-pot reactions and while the scope should be expanded by further investigating these multi-step reactions, various examples are already known.<sup>187,256,307</sup> Moreover, MOFs are often regarded as greener alternatives compared to current practices that require large amounts of solvent or water, highly acidic/basic conditions, or precious metal catalysts, which further stimulates their development within this field. However, since these subsequent reactions are more relevant for industrial processes and because they have been discussed in great detail before,<sup>187,256,259</sup> they will not be discussed further here.

### 3.4. Lipids

Similarly to POMs, MOFs and other heterogeneous MOC-based hybrid materials have been explored as catalysts for the production of biofuels from lipids. To achieve this, increasing the Lewis and Brønsted acidity is a priority because they are necessary to increase the reactivity toward the esterification of fatty acids and transesterification of triglycerides during this process. However, while research on lipid (trans)esterification with POMs has been more extensively explored, using MOC-based materials and MOFs has not been studied to the same extent yet, especially with respect to the reaction mechanism. In

this regard, the group of W. Yang has recently made progress using MOFs for biodiesel production from microalgal lipids, which are especially interesting substrates since microalgae have a fast growth rate, high photosynthetic efficiency, and very high lipid content, making them the most abundant natural source for biodiesel.<sup>317,318</sup> Two distinct paths were chosen to increase the reactivity and solve problems related to other porous catalysts for these types of reactions. Enhanced access to active sites was postulated by opting for a two-dimensional MOF structure and enhanced Brønsted acidity was achieved by immobilizing an ionic liquid cation on a MOF.

In the first approach, the two-dimensional MOF TATAB(Zr), consisting of  $\{\text{Zr}_6\text{O}_8\}$  clusters connected by 4,4',4''-s-triazin-1,3,5-triyltri-*p*-aminobenzoic acid (TATAB) linkers (Fig. 26(a)), was employed for the esterification of free fatty acids and transesterification of triglycerides in a microalgal lipid mixture (*i.e.*, palmitic, palmitoleic, stearic, oleic, and linoleic acid) to synthesize the main component of biodiesel: fatty acid methyl esters (FAMES).<sup>317</sup> The microalgal lipid mixture was converted to FAME using TATAB(Zr) with an efficiency of 98.2% in methanol after 2 h at 195 °C (Table 10). This is not only the most effective catalyst based on MOCs reported for this reaction, but it also benefits from an easy synthesis procedure and economical catalyst use. Three possible reaction mechanisms were postulated (Fig. 26(a)): (i) methanol coordinated to  $\text{Zr}^{4+}$  releases a proton that causes the formation of water from a Zr-bound hydroxide ( $\text{Zr-OH}$ ), after which nucleophilic attack of the formed  $\text{Zr-OCH}_3$  on a triglyceride produces an intermediate that further decomposes to form FAME; (ii) the carbonyl oxygen of the triglyceride coordinates to  $\text{Zr}^{4+}$  and a nucleophilic attack by free methanol causes the formation of an intermediate that further decomposes to form FAME; or (iii) both the triglyceride and methanol are coordinated to the same  $\{\text{Zr}_6\text{O}_8\}$  cluster, thus both carbocations can directly react to form FAME.<sup>317</sup> These reaction pathways are also possible for the methanolic (trans)-esterification of monoglycerides, diglycerides, and free fatty acids. Furthermore, when compared with three-dimensional MOF-808(Zr), TATAB(Zr) performed significantly better. This is

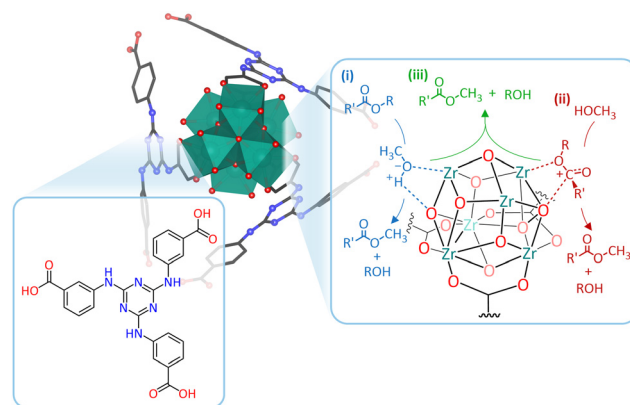


Fig. 26 Structural representation of the  $\text{Zr}_6$  cluster in TATAB(Zr) and TATAB linker along with a scheme for the three proposed reaction pathways for the transesterification of triglyceride. Adapted from ref. 317.



**Table 10** Selected conditions and percentage conversion reported for MOF-catalyzed (trans)esterification of free fatty acids (FA) and oil mixtures

| Heterogeneous hybrid catalyst  | Substrates        | Reactivity          | Reaction conditions                        | Conversion/% | Ref. |
|--|-------------------|---------------------|--|--------------|------|
| H <sub>3</sub> [PMo <sub>12</sub> O <sub>40</sub> ]/BTC(Fe)  | Oleic acid        | Esterification      | 16 : 1 MeOH : FA, 160 °C, 3 h, 10 wt% MOF  | 72           | 319  |
| Sn <sub>1.5</sub> [PW <sub>12</sub> O <sub>40</sub> ]/BTC(Cu)  | Oleic acid        | Esterification      | 20 : 1 MeOH : FA, 160 °C, 4 h, 0.2 g MOF   | 88           | 320  |
| TATAB(Zr)  | Microalgal lipids | Transesterification | 20 : 1 MeOH : oil, 195 °C, 2 h, 2 wt% MOF  | 98           | 317  |
| IL/H <sub>3</sub> [PMo <sub>12</sub> O <sub>40</sub> ]/MIL-101(Fe)   | Soybean oil       | Transesterification | 30 : 1 MeOH : oil, 120 °C, 8 h, 9 wt% MOF  | 93           | 321  |
| IL/POM/UiO-66(Zr)-2COOH  | Soybean oil       | Transesterification | 35 : 1 MeOH : oil, 110 °C, 6 h, 10 wt% MOF | 92–96        | 322  |
| IL/H <sub>3</sub> [PMo <sub>12</sub> O <sub>40</sub> ]/CoFe <sub>2</sub> O <sub>4</sub> /MIL-88B(Fe)-NH <sub>2</sub> | Soybean oil       | Transesterification | 30 : 1 MeOH : oil, 140 °C, 8 h, 8 wt% MOF  | 96           | 323  |
| Arg <sub>2</sub> H <sub>3</sub> [PW <sub>12</sub> O <sub>40</sub> ]/ZIF-8  | Insect lipids     | Transesterification | 9 : 1 MeOH : oil, 60 °C, 4 h, 3 wt% MOF    | 94           | 324  |

mainly attributed to the inaccessibility of the internal active Zr<sup>4+</sup> sites of MOF-808, but it can also be correlated to the higher degree of unsaturation and stronger overall acidity of TATAB(Zr).<sup>317</sup> Consequently, two-dimensional MOFs should be explored further, not only for this reaction but for all the other reactions discussed in this review.

The second approach for increasing reactivity in the (trans)-esterification of microalgal lipid mixtures to FAME, was explored using the isorecticular metal-organic framework-3 (IRMOF-3), consisting of tetranuclear Zn<sup>2+</sup> units connected by six 2-amino-1,4-benzenedicarboxylic acid (NH<sub>2</sub>-BDC) linkers, combined with the Brønsted acidic IL 1-butylsulfonate-3-methylimidazolium bisulfate ([BSO<sub>3</sub>HmIm][HSO<sub>4</sub>]).<sup>318</sup> Therefore, the catalyst has both Lewis (Zn<sup>2+</sup>) and Brønsted acid (HSO<sub>4</sub><sup>−</sup>) sites that can be exploited, resulting in similar FAME conversion as for TATAB(Zr) (Table 10).<sup>318</sup> The reaction mechanism was proposed to also be similar to that of TATAB(Zr), but in this case, the Lewis basic sites (uncoordinated −NH<sub>2</sub> and imidazole) are primarily responsible for the transesterification of triglycerides.<sup>318</sup> Similar systems, where the Lewis acidity of various MOFs was combined with the Brønsted acidity of ILs, have also been described for the esterification of oleic acids.<sup>325–327</sup> Furthermore, MOF systems that incorporate both ILs and POMs, which add to the Brønsted acidity, have been utilized for (trans)esterification of single fatty acids and other oil mixtures, such as vegetable oil and soybean oil (Table 10).<sup>319–323</sup> However, these have a lower conversion efficiency compared to both the TATAB(Zr) and IRMOF-3/IL systems because the accessibility to the active sites or the overall acidity is not optimized to the same extent. Therefore, there is still significant potential for exploring this second route for achieving better catalytic activity in biodiesel production using MOC-based materials.

Compared to completely MOF-based catalysts for lipid (trans)esterification, more examples of POM-based hybrids have been explored. Initially, the focus of research was driven by the heterogenization of POMs due to the benefits associated with heterogeneous catalysis. In that regard, POMs have been immobilized on various supports ranging from solid zirconia and zirconium phosphate to mesoporous materials (*i.e.*, zeolites and MOFs) and nanoparticles.<sup>243,324,328–332</sup> However, recent developments with MOFs have provided valuable insights about the reaction mechanism and clarified that the reaction is not significantly influenced by specific MOF parameters, but rather that it depends on the active site accessibility and the acidity/basicity of the catalyst.

## 4. Conclusions and outlook

MOCs and MOC-based materials have been established to be a powerful family of catalysts for biomolecular transformations with potential applications in many fields, such as medicine, biotechnology, and biofuel production. MOCs, as discrete clusters or as part of MOFs and other materials, have been shown to effectively catalyze a wide range of reactions (*e.g.*, hydrolysis, amidation, oxidation, isomerization, dehydration, (trans)esterification, *etc.*) of an even wider range of biomolecules: amino acids, peptides, (glyco)proteins, nucleic acids, nucleotides, sugars, fatty acids, triglycerides, *etc.* Furthermore, the diversity of this family of clusters in terms of metal composition and structure means that they can be tuned to optimize the reactivity for the desired application. However, this requires an in-depth understanding of structure–activity relationships, which can be gained from the multiple mechanistic studies presented in this review. These studies show that the reactivity of MOCs is generally linked to their Lewis acidity, Brønsted acidity, and/or redox activity. For instance, highly Lewis acidic metal centers are needed for peptide, phosphoester, and phosphoanhydride bond hydrolysis as well as for peptide bond formation since coordination to C=O or P=O bonds activates them towards nucleophilic attack. On the other hand, Brønsted acidity is needed to promote the hydrolysis of glycosidic bonds as well as the (trans)esterification of fatty acids and triglycerides. In addition, by combining Lewis and Brønsted acidity the direct conversion of biomass to HMF can be achieved *via* isomerization and dehydration reactions. Similarly, through a combination of Lewis acidity and redox activity, the hydrolysis of peptide bonds can be tuned. However, the redox properties of MOCs and MOC-based materials have not been as extensively investigated for biomolecular transformations, despite having been explored in many organic reactions.<sup>29</sup> Therefore, there is still immense untapped potential for the design of MOCs as highly active and selective catalysts in biochemical reactions, and investigating their mechanism of action is the first step in this direction.

Many mechanistic insights have been obtained from investigating the reactivity of discrete MOCs, particularly POMs. Their relatively simple atomically precise structures and their solubility in different solvents make them more amenable to spectroscopic and computational studies compared to MOC-based heterogeneous catalysts. However, POMs can have complex speciation in solution, which makes it difficult to determine the catalytically active species. As a result, less labile



polyoxotungstate M-POMs have been most extensively investigated. While the solubility of certain discrete MOCs as homogeneous catalysts is advantageous for studying their reactivity, it makes catalyst recycling challenging. Therefore, several strategies for the heterogenization of POMs have been explored, especially for HPAs that could be used in biofuel production. One way of achieving this for POMs is through ion exchange with surfactants, ionic liquid cations, or large monovalent ions (e.g.,  $\text{NH}_4^+$ ,  $\text{Cs}^+$ , etc.) to form insoluble salts. This also allows for the structural and catalytic properties of the counter-cation to be exploited to enhance the reactivity. Alternatively, MOCs can be heterogenized by incorporation into solid structures, such as MOFs. In this respect, MOFs have been shown to be particularly suitable due to their porosity and high surface area. Furthermore, the reactivity of MOFs can be tuned through careful choice of their structure, the number of defects, the nature of the MOC, and the functionality of the organic linker. Therefore, the reactivity of multiple MOC-based MOFs towards biomolecules has been increasingly explored in recent years.

When transitioning towards heterogeneous catalysis, the knowledge gained from the reactivity of discrete MOCs can be translated to MOC-based materials. This can be most clearly seen from the studies on the reactivity of MOCs towards peptide bonds in the context of proteomics analysis, as Zr/Hf-based soluble POMs, neutral MOCs, and MOC-based MOFs catalyzed the hydrolysis reaction in similar ways with differences in their reactivity likely linked to their structure. For instance,  $\{\text{Zr}_6\text{O}_8\}$ -based MOF-808 gave rise to a faster reaction rate for the hydrolysis of the dipeptide Gly-Gly than Zr-POMs and discrete  $\text{Zr}_6$  clusters, but a similar selectivity was likely achieved as for Zr-POMs.<sup>260</sup> However, the study of the reactivity of discrete MOCs towards biomolecules has been mainly limited to POMs even though there are several other classes of MOCs that have potential as catalysts, such as the  $\text{Zr}^{4+}$ ,  $\text{Hf}^{4+}$ , and  $\text{Ce}^{4+}$  clusters that have been used extensively as building blocks for MOFs. Moreover,  $\text{Zr}^{4+}$ ,  $\text{Hf}^{4+}$ , and  $\text{Ce}^{4+}$  have been shown to be particularly interesting due to their high Lewis acidity, high coordination number, and oxophilicity. Furthermore, the redox activity of  $\text{Ce}^{4+}$  makes it promising for oxidation reactions, either through direct oxidation of biomolecules or indirectly *via* the formation of ROS. Therefore, further studies on the reactivity of these discrete MOCs towards biomolecules could open new avenues of research and help direct the development of new related heterogeneous materials, such as 2D MOFs, which have not been as extensively explored so far.

Further research into the reactivity of metal-oxo clusters and cluster-based materials should ideally build upon the insights obtained until now from simple model systems to tackle more complex biomolecules, such as DNA or membrane proteins, while bearing in mind the limitations of such model systems for predicting the reactivity towards large biomolecules. In addition, the cytotoxicity and pharmacokinetics of MOCs need to be considered for their use in biomedical applications.<sup>57,333–336</sup> Therefore, additional studies on the impact of MOCs on living organisms and on their stability in biological systems are needed to develop them for these applications.<sup>337,338</sup> These studies could

benefit from the molecular-level insights discussed in this review by establishing a link between reactivity and cytotoxicity, which has been scarcely investigated in detail. Additionally, the study of the cytotoxicity of POMs and the development of biocompatible POMs with low off-target toxicity is of primordial importance for their implementation in biomedical applications since clinical trials of POM-based inorganic drugs have been largely hampered by their cytotoxicity.<sup>57</sup> Moreover, while this review covers the reactivity of MOCs towards the main types of biomolecules present in organisms, the lessons learned from these studies can be applied to other biomolecules, such as terpenes and their derivatives,<sup>234,339–342</sup> as well as in synthetic chemistry, especially in the production of pharmaceuticals.<sup>343</sup>

## Author contributions

The manuscript was written through contributions of all authors. All authors have given approval to the final version of the manuscript.

## Conflicts of interest

There are no conflicts to declare.

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